



UNIVERSITI PUTRA MALAYSIA

***ISOLATION AND CHARACTERIZATION OF PATHOGENIC
LEPTOSPIRES FROM RODENTS AND SMALL MAMMALS CAPTURED
IN HUMAN LEPTOSPIROSIS SUSPECTED AREAS IN SELANGOR,
MALAYSIA***

NURUL NATASYA BINTI AZHARI

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By

NURUL NATASYA BINTI AZHARI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of Master of
Science**

November 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

ISOLATION AND CHARACTERIZATION OF PATHOGENIC LEPTOSPIRES FROM RODENTS AND SMALL MAMMALS CAPTURED IN HUMAN LEPTOSPIROSIS SUSPECTED AREAS IN SELANGOR, MALAYSIA

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November 2018

Chair : Assoc. Prof. Vasantha Kumari Neela, PhD
Faculty : Medicine and Health Sciences

Leptospirosis, previously known to be a neglected zoonotic disease in the tropical region is now re-emerging as a threat to public health in both urban and rural settings. It is known that rodents are the major carrier of pathogenic *Leptospira* sp. In Malaysia, the endemicity of leptospirosis in human is concentrated in areas where rats were highly populated such as in residential and recreational areas with improper trash management and poor sanitation. The statistics in Malaysia has shown an increasing trend of suspected leptospirosis cases and reported deaths since it has been gazetted as a notifiable disease in 2010. This makes an urgent call to study the carrier status of pathogenic leptospires infecting the human through the study of *Leptospira* sp. in rodents and small mammals in Malaysia. The objective of this study is to identify and characterize the predominant pathogenic *Leptospira* sp. circulating in the leptospirosis suspected areas in the Selangor state of Malaysia. The study was carried out from January 2016 to April 2017 in six suspected areas comprising of urban, semi-urban and recreational forest areas. The study sites were identified by the Selangor State Health Department as outbreak or hotspot areas. Rodents trapping was performed in all six study sites. The trapped rodents were dissected and kidneys harvested. The rodent kidneys were subjected to *Leptospira* isolation by culture and dark-field microscopy. The identification and pathogenic strain of the isolated leptospires were determined by PCR approach. The characterization included *secY* and *lipL32* PCR and multi-locus sequence typing (MLST). A total of 14 small mammals species were identified from the 266 captured small mammals with *Rattus norvegicus* (66%, n=100) being the dominant rat species in the urban area while

Maxomys whiteheadi (30%, n=23) dominated the recreational forest area. Among the 266 rodents captured, 217 kidney samples were cultured, while for the remaining 49 samples, DNA was directly extracted from the kidney. From 217 samples cultured 55 (25.3%) were positive for spirochetes examined under the dark-field microscope (DFM). Of the 55 culture and 49 DNA samples, 38/266 (14.3%) were identified to be positive for pathogenic *Leptospira* confirmed by *secY* and *lipL32* PCR. Phylogenetic analysis by *secY* PCR showed unique clusters for each species and all isolates clustered according to the respective species. *Leptospira interrogans* dominated all studied sites followed by *L. kirschneri* (n=5), *L. borgpetersenii* (n=2) and *L. weilii* (n=1). However, no significant association was shown between the infection rate in small mammals with three different sites category (urban, semi-urban and recreational forest) (chi-square, χ^2 0.5296; $p=0.767$). From MLST analysis, three clones of *Leptospira* sp. were found to dominate the study sites; *L. interrogans* serovar Bataviae ST50, *L. kirschneri* serovar Grippotyphosa ST110 and *L. borgpetersenii* serovar Javanica ST143. While with the help of curator of MLST database, a new ST number (ST238, ST242 and ST243) representing new *L. interrogans* and *L. weilii* species was curated for samples isolated mostly from the recreational forest area sites. From the present study, *R. norvegicus* was identified as the common pathogenic *Leptospira* host dominating the urban area followed by *R. rattus*. However although *M. whiteheadi* dominated the recreational forest area, *S. muelleri* was the major carrier of the pathogenic *Leptospira*. In conclusion two species of rats (*R. norvegicus* and *S. muelleri*) were identified to play a vital role in environmental contamination in all study sites. This study also confirms for the first time, carriers of *Leptospira* locus ST110 *L. kirschneri*, ST242 *L. weilii*, ST238 *L. interrogans* and ST243 *L. interrogans* among small mammals in Malaysia. Thus further research and attention on the serovar status, of all isolates should be carried out as these species may potentially become highly pathogenic serovars contributing to increased risk of severe leptospirosis in Malaysia. In addition, these strains should be included into MAT panels as to improve the diagnosis in the future.

Keywords: Leptospirosis, *Leptospira* sp., rodents,

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**ISOLASI DAN PENCIRIAN BAKTERIA *LEPTOSPIRA* DARI TIKUS DAN
MAMALIA KECIL DI KAWASAN YANG DISYAKI PUNCA WABAK
LEPTOSPIROSIS DI SELANGOR, MALAYSIA**

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Leptospirosis dahulunya merupakan penyakit zoonotik yang terabai di kawasan tropik di Malaysia dan muncul semula sebagai ancaman kepada kesihatan awam dalam persekitaran bandar dan luar bandar. Secara umumnya tikus adalah pembawa utama bakteria *Leptospira* yang merupakan punca penyakit leptospirosis. Di Malaysia, penyebaran wabak penyakit ini tular di kawasan persekitaran yang kotor terutamanya kawasan perumahan yang mempunyai jumlah penempatan tertinggi dan kawasan rekreasi yang sering dilawati. Statistik di Malaysia juga turut mendedahkan kenaikan kadar pesakit yang disyaki menghidap penyakit leptospirosis dan kadar kematian juga turut meningkat sejak tahun 2012. Justeru, kajian ini dijalankan bagi mengetahui spesies bakteria *Leptospira* yang manakah menjadi punca utama kepada penyakit leptospirosis yang dibawa dikalangan pelbagai spesies mamalia kecil di Malaysia. Objektif kajian ini dijalankan adalah untuk mengenalpasti spesies bakteria *Leptospira* yang menjadi punca utama kepada penularan wabak leptospirosis di kalangan manusia terutamanya di kawasan Selangor. Kajian ini telah dijalankan pada Januari 2016 hingga April 2017 di enam kawasan yang disyaki penularan wabak yang dilabel sebagai kawasan urban, semi-urban dan hutan rekreasi. Keenam-enam kawasan ini telah disyorkan sebagai kawasan disyaki wabak oleh Kementerian Kesihatan Malaysia dan Jabatan Kesihatan Negeri Selangor. Penangkapan sampel tikus dan mamalia kecil telah dilakukan di enam lokasi seperti yang telah disenaraikan. Isolasi bakteria hidup *Leptospira* telah dilakukan dengan menggunakan teknik pengkulturan dan pengimejan melalui mikroskop lapangan gelap (DFM). Identifikasi status pembawa penyakit telah dilakukan dengan menggunakan teknik PCR. Identifikasi PCR menggunakan beberapa primer telah dilakukan termasuklah *secY* and *lipL32* dan kombinasi primer dengan teknik MLST telah dijalankan. Sebanyak 14 spesies tikus dijumpai daripada 266 jumlah mamalia kecil yang telah disampelkan di mana spesies dari *Rattus norvegicus* (66%, n=100) merupakan populasi terbanyak di kawasan urban manakala *Maxomys whiteheadi* (30%,

n=23) merupakan spesies terbanyak di kawasan hutan rekreasi. Daripada 266 sampel mamalia kecil yang ditangkap, 217 sampel telah diproses bagi isolasi bakteria melalui teknik kultur, manakala pengekstrakan DNA bakteria *Leptospira* daripada baki 49 sampel lagi telah dilakukan dan tiada kultur dilakukan bagi sampel ini. Daripada 217 sampel organ yang dikultur, hanya 55/217 (25.3%) sahaja yang menunjukkan penampakan bakteria *Leptospira* di bawah DFM, manakala keputusan PCR yang telah dijalankan ke atas 55 sampel positif kultur dan 49 sampel DNA menunjukkan sebanyak 38/266 (14.3%) sampel positif terhadap *Leptospira* bakteria bahaya penyebab leptospirosis. Analisis filogenetik dengan penggunaan primer *secY* menunjukkan keunikan dan kepelbagaian di dalam genetik sampel yang terisolat dan majoriti spesies *Leptospira* dari *L. interrogans* (n=30) mendominasi semua kawasan kajian diikuti oleh *L. kirschneri* (n=5), *L. borgpetersenii* (n=2) dan *L. weilii* (n=1). Walaubagaimanapun, tiada hubungan kait ditunjukkan daripada analisa statistik di antara haiwan pembawa bakteria *Leptospira* di tiga kategori kawasan persampelan (urban, semi-urban dan hutan rekreasi) (chi-square, χ^2 0.5296; $p=$ 0.767). Daripada analisa MLST, tiga serovar bakteria *Leptospira* yang mendominasi kawasan kajian dikenali sebagai *L. interrogans* serovar Bataviae (ST50), *L. kirschneri* serovar Grippotyphosa (ST110) dan *L. borgpetersenii* serovar Javanica (ST143). Manakala id (ST) baru (ST238, ST242 dan ST243) telah diberikan kepada isolat bakteria *L. interrogans* dan *L. weilii* yang tidak didapati di dalam pangkalan data MLST. Kesimpulan daripada kajian ini mendapati, spesies mamalia kecil dari tikus yang dikenali sebagai *R. norvegicus* merupakan spesies yang utama pembawa bakteria bahaya *Leptospira* di kawasan urban diikuti dengan *R. rattus*. Manakala, spesies tikus dari *S. muelleri* merupakan pembawa utama bakteria bahaya *Leptospira* di kawasan hutan rekreasi walaupun populasi terbanyak yang mendominasi kawasan tersebut adalah dari spesies *M. whiteheadi*. Justeru daripada pendapatan kajian ini, dua species tikus ini merupakan spesies pembawa utama yang menjadi punca dalam penularan wabak leptospirosis. Kajian ini adalah yang pertama menjumpai isolat ST110 *L. kirschneri* dan ST242 *L. weilii*, ST238 *L. interrogans* dan ST243 *L. interrogans* di kalangan haiwan mamalia kecil di Malaysia. Justeru, pengesahan kajian selanjutnya dan perhatian mengenai status serovar bagi setiap isolat haruslah dijalankan kerana serovar dari isolat ini mungkin boleh menjadi punca penyebab jangkitan *Leptospira* yang serius di Malaysia. Selain itu, kepentingan menambah senarai panel serovar terutamanya isolat dari Malaysia bagi meningkatkan keberkesanan diagnosis penyakit leptospirosis di masa hadapan.

Kata kunci: Leptospirosis, spesies *Leptospira*, tikus

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

bp	Basepair
BSA	Bovine Serum Albumin
Buffer AL	DNA Binding Buffer
Buffer ATL	Tissue Lysis Buffer
Buffer W1	Wash Buffer
Buffer W2	Desalting Buffer
Buffer AE	Elution Buffer
CAAT	Cross Agglutination Absorption Test
D	Simpson's Diversity Index,
DFM	Dark-Field Microscope
dH ₂ O	Distilled water
DNA	Deoxyribonucleic acid
DMSO	Dimethylsulfoxide
EMJH Harris	Liquid Ellinghausen, Mccullough, Johnson,
g	Gram
<i>gyrB</i>	DNA <i>gyrase</i> subunit B
L	Litre
LPS	Lipopolysaccharide
M	Molar
MAT	Microscopic Agglutination Test
mg	Miligram
mg/ml	Milligram per millilitre
min	Minute
mL	Millilitre

MLST	Multi Locus Sequence Typing
mM	Milimolar
NCBI	National Center of Biotechnology Information
ng	Nanogram
PBS	Phosphate Buffered Saline
PCR	Polymerase chain reaction
PFGE	Pulse Field Gel Electrophoresis
pH	Potential of hydrogen
<i>rpoB</i>	RNA polymerase- β -subunit
rRNA	Ribosomal ribonucleic acid
<i>secY</i>	Preprotein translocase subunit
<i>rrs</i>	16S Ribosomal ribonucleic acid
SPSS	Statistical Package for the Social Sciences
ST	Sequence Type
TBE	Tris-Borate-EDTA
UV	Ultraviolet
WHO	World Health Organization
μ g	Microgram
μ g/mL	Microgram per millilitre
μ L	Microlitre
5-FU	5-Fluorouracil

CHAPTER 1

INTRODUCTION

1.1 Study Background

Leptospirosis locally known as rat urine disease is a bacterial infection caused by the corkscrew-shaped spirochete *Leptospira* (Bharti et al., 2003). Rodents serve as the main reservoir for *Leptospira* although all mammals do carry the bacteria in their kidney. *Leptospira* replicates in the renal tubules of rodents, domestic and wild animals, which are then shed in the environment through their urine (Sumanta et al., 2015). In rodents, *Leptospira* has the ability to escape from the host immune system and then establish persistent renal colonization by forming an amorphous, biofilm-like structure in the renal tubules (Agudelo-Flórez et al., 2013). Human gets infected either through contact with contaminated animal urine or body fluids or from water and soil contaminated with leptospiral spp. (Levett, 2001; Matthias et al., 2008). Human leptospirosis is reported to be concentrated in areas where rats are highly populated due to improper trash management and poor sanitation (Barcellos & Sabroza, 2001; Bharti et al., 2003). The spirochete *Leptospira* enter the human body through the skin (cuts or wound) or mucous membranes (eyes, nose, or mouth). The disease is usually vulnerable among poverty in middle and low-income countries and considered as occupational disease, and this includes people who work in a paddy field, sewer, gold mining, food markets and farmland (Schneider et al., 2013; WHO, 2003a).

Leptospirosis is known to be highly endemic in tropical and subtropical regions (Bharti et al., 2003), nevertheless a wide geographical distribution with global annual cases of 1.03 million and 58,999 deaths are reported (Costa et al., 2015). A systematic review revealed Oceania region (Australia, New Zealand and pacific island countries and territories) to be the most affected by leptospirosis with morbidity (150.68 cases/100,000 populations per year) and mortality (9.61 deaths/ 100,000 populations per year) (Costa et al., 2015). Recreational activities and ecotourism served as the leading cause of leptospirosis in the younger generation (Mayer-Scholl et al., 2014; Sejvar et al., 2003; Van Crevel et al., 1994). Few outbreaks in Malaysia that is associated with recreational activities include: outbreak associated with swimming in Beaufort (Koay et al., 2004), outbreak of melioidosis co-infection with leptospirosis (Sapian et al., 2012), and a death case of a national service trainee died of suspected leptospirosis (Thayaparan et al., 2013). Then followed by a few recent sporadic cases reported in national daily news and the press release by the officials of the ministry of health include: two death cases of the students swimming in contaminated rivers in Jeram and waterfall in Gunung Pulai in the year 2017 respectively and another case involving student infected with leptospirosis upon visitation to the recreational park in Jelebu (Garba et al., 2017).

Leptospira which are classified into pathogens, intermediates and saprophytes are distributed to 23 species to date comprising ten pathogenic (*Leptospira interrogans*, *L. kirschneri*, *L. borgpetersenii*, *L. santarosai*, *L. noguchii*, *L. weilii*, *L. alexanderi*, *L. kmetyi*, *L. alstonii* and *L. mayottensis*), six intermediates (*L. inadai*, *L. broomii*, *L. fainei*, *L. wolffii*, *L. licerasiae* and *L. venezuelensis*) and seven saprophytes (*L. biflexa*, *L. wolbachii*, *L. meyeri*, *L. vanthiellii*, *L. terpstrae*, *L. idonii* and *L. yanagawae*) (Bourhy et al., 2014). At least 20 serogroups and more than 300 serovars have been described (Mayer-Scholl et al., 2014; Picardeau, 2013). Pathogenic species are usually carried by rodents, while intermediates and saprophytes are commonly seen in the environment (water or soil). To date based on serological or bacteriological diagnosis, at least thirty-seven serovars have been isolated and characterized with majority of them carried by rodents (Benacer et al., 2013).

In Malaysia, leptospirosis is an emerging public health problem affecting both urban and rural population hence has been gazetted as a notifiable disease since 2010 by the Ministry of Health, Malaysia. According to them, the incidence of leptospirosis showed an increasing trend from 263 in 2004 to 5370 in 2015 with mortality of 20 to 96 cases per annum. Eventhough the reported number of clinical leptospirosis cases in Malaysia is high, the number of laboratory-confirmed cases is low. One of the contributing factors could be the information on the circulating serovars or infecting strains are limited. Updating the serovar panel is utmost important in diagnostic assays for leptospirosis. The epidemiology of the leptospirosis is not clear in Malaysia despite rodents were identified as the major reservoirs. Hence, the present study is undertaken to perform a systemic investigation on the *Leptospira* strains circulating among small mammals with primary focus on rodents in the urban, semi-urban and recreational forest sites where human leptospirosis are frequently reported in the Selangor state of Malaysia. The study was aim to identify the circulating pathogenic species and their associated animal reservoirs. This will guide public health professionals in further epidemiological investigations to determine the risk factors associated with leptospirosis in Malaysia and also to improve the diagnostic panel with more pathogenic species.

1.2 Problem Statement

In Malaysia, leptospirosis is an emerging public health problem affecting both urban and rural population. The listing of leptospirosis as a notifiable disease since 2010 has shown increasing trends of infection incidence and mortality rates. Rodents especially rats serve as the main reservoir for supplying pathogenic leptospiral strains for human infection in Malaysia as elsewhere. In Malaysia, a majority of the cases are either due to vacationing at recreational forest waterfalls or from poorly maintained non-hygienic residential premises. Although most clinical symptoms and laboratory indications suspected for human leptospirosis, most cases do not achieve a confirmed diagnosis. One of the main reasons is the lack of comprehensive data on the circulating strains, which is not only essential to understand the potential zoonosis, but also very crucial to be included in the diagnostic panel especially the gold standard microscopic

agglutination test (MAT). Secondly, for any public health measures (awareness, control, and prevention) to be taken, the knowledge on the ecological niche of the reservoir animals or maintenance host, the prevalence, and type of the leptospiral species and characterization of strains is utmost important. Hence, the present study is exceptionally important to obtain the baseline data on circulating and prevailing strains, reservoir host and their ecological niche, and most importantly live strains to be included in MAT panel. This study may also help unravel the mystery behind the suspected leptospirosis cases frequently reported from these geographical niches.

1.3 Objectives

1.3.1 General Objective

To obtain a comprehensive understanding of the leptospiral species circulating among rodents and small mammals inhabiting selected residential and recreational sites suspected of human leptospirosis in the Selangor state of Malaysia.

1.3.2 Specific Objectives

1. To determine leptospires carrying small mammals and to determine the reservoir animal species in the study population.
2. To identify the prevailing pathogenic *Leptospira* species.
3. To determine the genetic relationship among the *Leptospira* strains and identify the circulating genotypes.

REFERENCES

- Adler, B. (2015). History of leptospirosis and leptospira. In B. Adler (ed), *Leptospira and Leptospirosis* (pp. 1-9). Berlin: Springer.
- Agudelo-Flórez, P., Murillo, V. E., Londoño, A. F., and Rodas, J. D. (2013). Histopathological kidney alterations in rats naturally infected with Leptospira. *Biomédica* 33, 82-88.
- Alston, J. M., Broom, J. C., and Doughty, C. J. A. (1958). *Leptospirosis in man and animals* (Vol. 5). Edinburgh: Livingstone.
- Ahmed, A., Engelberts, M. F., Boer, K. R., Ahmed, N., and Hartskeerl, R. A. (2009). Development and validation of a real-time PCR for detection of pathogenic Leptospira species in clinical materials. *PLoS One* 4(9), e7093.
- Ahmed, N., Devi, S. M., De los Á Valverde, M., Vijayachari, P., Machang'u, R. S., Ellis, W. A., and Hartskeerl, R. A. (2006). Multilocus sequence typing method for identification and genotypic classification of pathogenic Leptospira species. *Annals of clinical microbiology and antimicrobials* 5(1): 28.
- Amarasekera, J., Agampodi, S., and Kodituwakku, M. (2013). Risk factors and reservoir species for leptospirosis in Sri Lanka. *J South Asia Regionsymposirum* 54:55.
- Amran, F., Khalid, M. K. N. M., Mohamad, S., Ripen, A. M., Ahmad, N., Goris, M. G., Muhammad, A. H. and Halim, N. A. N. (2016). Draft genome sequence of Leptospira interrogans serovar Bataviae strain LepIMR 22 isolated from a rodent in Johor, Malaysia. *Genome announcements* 4(5):e00956-16.
- Audy, J. R. (1958). The localization of disease with special reference to the zoonoses. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 52(4): 308-28.
- Bahaman, A. R., and Ibrahim, A. L. (1988). A review of leptospirosis in Malaysia. *Veterinary research communications* 12(2-3): 179-189.
- Balamurugan, V., Gangadhar, N. L., Mohandoss, N., Thirumalesh, S. R. A., Dhar, M., Shome, R., Krishnamoorthy, P., Prabhudas, K., et al. (2013). Characterization of leptospira isolates from animals and humans: phylogenetic analysis identifies the prevalence of intermediate species in India. *Springerplus* 2(1): 362.
- Barcellos, C., and Sabroza, P. C. (2001). The place behind the case: leptospirosis risks and associated environmental conditions in a flood-

related outbreak in Rio de Janeiro. *Cadernos de Saúde Pública* 17:S59-S67.

- Benacer, D., Mohd Zain, S. N., Amran, F., Galloway, R. L., and Thong, K. L. (2013). Isolation and molecular characterization of *Leptospira interrogans* and *Leptospira borgpetersenii* isolates from the urban rat populations of Kuala Lumpur, Malaysia. *The American Journal of Tropical Medicine and Hygiene* 88(4): 704–9.
- Benacer, D., Mohd Zain, S. N., Sim, S. Z., Mohd Khalid, M. K. N., Galloway, R. L., Souris, M., and Thong, K. L. (2016). Determination of *Leptospira borgpetersenii* serovar Javanica and *Leptospira interrogans* serovar Bataviae as the persistent *Leptospira* serovars circulating in the urban rat populations in Peninsular Malaysia. *Parasites and Vectors* 9(1): 117.
- Benacer, D., Thong, K. L., Ooi, P. T., Souris, M., Lewis, J. W., Ahmed, A. A., and Mohd Zain, S. N. (2017). Serological and molecular identification of *Leptospira* spp. in swine and stray dogs from Malaysia. *Tropical Biomedicine* 34(1): 89–97.
- Bharti, A. R., Nally, J. E., Ricaldi, J. N., Matthias, M. A., Diaz, M. M., Lovett, M. A., Levett, P. N., Gilman, R. H., et al. (2003). Leptospirosis: a zoonotic disease of global importance. *The Lancet infectious diseases* 3(12): 757-771.
- Boonsilp, S., Thaipadungpanit, J., Amornchai, P., Wuthiekanun, V., Bailey, M. S., Holden, M. T. G., Zhang, C., Jiang, X., et al. (2013). A Single Multilocus Sequence Typing (MLST) Scheme for Seven Pathogenic *Leptospira* Species. *PLoS Neglected Tropical Diseases* 7(1): e1954.
- Bourhy, P., Collet, L., Brisse, S., and Picardeau, M. (2014). *Leptospira mayottensis* sp. nov., a pathogenic species of the genus *Leptospira* isolated from humans. *International Journal of Systematic and Evolutionary Microbiology* 64(12): 4061-4067.
- Bourhy, P., Storck, C. H., Theodose, R., Olive, C., Nicolas, M., Hochedez, P., Lamaury, I., Zinini, F., et al. (2013). Serovar diversity of pathogenic *Leptospira* circulating in the French West Indies. *PLoS neglected tropical diseases* 7(3): e2114.
- Brendle, J. J., Rogul, M., and Alexander, A. D. (1974). Deoxyribonucleic acid hybridization among selected leptospiral serotypes. *International Journal of Systematic and Evolutionary Microbiology* 24(2): 205-214.
- Brenner, D. J., Kaufmann, A. F., Sulzer, K. R., Steigerwalt, A. G., Rogers, F. C., and Weyant, R. S. (1999). Further determination of DNA relatedness between serogroups and serovars in the family Leptospiraceae with a proposal for *Leptospira alexanderi* sp. nov. and four new *Leptospira* genomospecies. *International Journal of Systematic Bacteriology* 49(2): 839–858.

- Bunnell, J. E., Hice, C. L., Watts, D. M., Montrueil, V. I. C. T. O. R., Tesh, R. B., and Vinetz, J. M. (2000). Detection of pathogenic *Leptospira* spp. infections among mammals captured in the Peruvian Amazon basin region. *The American journal of tropical medicine and hygiene* 63(5): 255-258.
- Carbrey, E. A. (1960). The relative importance of variable factors in the agglutination-lysis test. *Annual Proceedings of the United States Livestock Sanitary Associations* 64: 130-142.
- Cerqueira, G. M., and Picardeau, M. (2009). A century of *Leptospira* strain typing. *Infection, Genetics and Evolution* 9(5): 760–768.
- Chiani, Y., Jacob, P., Varni, V., Landolt, N., Schmeling, M. F., Pujato, N., Caimi, K., Vanasco, B. (2016). Isolation and clinical sample typing of human leptospirosis cases in Argentina. *Infection, Genetics and Evolution* 37: 245–251.
- Chung, H. L., Ts'ao, W. C., and Chih, Y. (1963). Transplacental or Congenital Infection of Leptospirosis. Clinical and Experimental Observations. *Chinese medical journal* 82(12): 777-82.
- Coghlan, J. D., and Bain, A. D. (1969). Leptospirosis in human pregnancy followed by death of the foetus. *British Medical Journal* 1(5638): 228-230.
- Cole, J. R., Sulzer, C. R., and Pursell, A. R. (1973). Improved Microtechnique for the Leptospiral Microscopic Agglutination Test1. *Applied Microbiology* 25(6): 976–980.
- Cosson, J. F., Picardeau, M., Mielcarek, M., Tatard, C., Chaval, Y., Suputtamongkol, Y., Buchy, P., Jittapalapong, S., et al. (2014). Epidemiology of *Leptospira* transmitted by rodents in Southeast Asia. *PLoS Neglected Tropical Diseases* 8(6): e2902.
- Costa, F., Hagan, J. E., Calcagno, J., Kane, M., Torgerson, P., Martha, S., Stein, C., et al. (2015). Global morbidity and mortality of leptospirosis: a systematic review. *PLoS neglected tropical diseases* 9(9): e0003898.
- Costa, F., Wunder, E. A., de Oliveira, D., Bisht, V., Rodrigues, G., Reis, M. G., Albert, I. K., Begon, M., et al. (2015). Patterns in *Leptospira* shedding in Norway rats (*Rattus norvegicus*) from Brazilian slum communities at high risk of disease transmission. *PLoS Neglected Tropical Diseases* 9(6): 1–14.
- Cox, T. E., Smythe, L. D., and Leung, L. P. (2005). Flying foxes as carriers of pathogenic *Leptospira* species. *Journal of wildlife diseases* 41(4): 753-757.
- Dias, J. P., Teixeira, M. G., Costa, M. C. N., Mendes, C. M. C., Guimarães, P., Reis, M. G., Ko, A. and Barreto, M. L. (2007). Factors associated with

Leptospira sp infection in a large urban center in northeastern Brazil. *Revista da Sociedade Brasileira de Medicina Tropical* 40(5): 499-504.

Dikken, H., and Kmety, E. (1978). Chapter VIII serological typing methods of leptospire. In T. Bergan and J.R. Norris, *Methods in microbiology* (Vol. 11, pp. 259-307). London: Academic Press.

Dunn, M. (Ed.). (1989). *Exploring your world: The adventure of geography*. National Geographic Society.

El Jalii, I. M., and Bahaman, A. R. (2004). A review of human leptospirosis in Malaysia. *Tropical Biomedicine* 21(2): 113–119.

Enright, M. C., and Spratt, B. G. (1999). Multilocus sequence typing. *Trends in microbiology* 7(12): 482-487.

Evangelista, K., and Coburn, J. (2010). Leptospira as an emergin pathogen: a review of its biology, pathogenesis and host immune responses. *Future Microbiol* 5(9): 1413–1425.

Faine, S., and Stallman, N. D. (1982). Amended descriptions of the genus Leptospira Noguchi 1917 and the species L. interrogans (Stimson 1907) Wenyon 1926 and L. biflexa (Wolbach and Binger 1914) Noguchi 1918. *International Journal of Systematic and Evolutionary Microbiology* 32(4): 461-463.

Faine, S., Adler, B., Christopher, W., and Valentine, R. (1984). Fatal congenital human leptospirosis. *Zentralblatt fur Bakteriologie, Mikrobiologie, und Hygiene. Series A, Medical microbiology, infectious diseases, virology, parasitology* 257(4): 548.

Faisal, S. M., McDonough, S. P., and Chang, Y. F. (2012). Leptospira: Invasion, pathogenesis and persistence. In *The Pathogenic Spirochetes: strategies for evasion of host immunity and persistence* (pp.143-172). Boston:Springer.

de Faria, M. T., Athanazio, D. A., Ramos, E. G., Silva, E. F., Reis, M. G. D., and Ko, A. I. (2007). Morphological alterations in the kidney of rats with natural and experimental Leptospira infection. *Journal of comparative pathology* 137(4): 231-238.

de Faria, M. T., Calderwood, M. S., Athanazio, D. A., McBride, A. J., Hartskeerl, R. A., Pereira, M. M., Ko, A. I. and Reis, M. G. (2008). Carriage of Leptospira interrogans among domestic rats from an urban setting highly endemic for leptospirosis in Brazil. *Acta tropica* 108(1): 1-5.

Fletcher, W. (1928). Recent Work on Leptospirosis, Tsutsugamusfai Disease, and Tropical Typhus in the Federated Malay States. *Transactions of the Royal Society of Tropical medicine and Hygiene* 21(4): 265-268.

- Francis, C. M., and Barrett, P. (2008). *A guide to the mammals of Southeast Asia*. Princeton University Press.
- Galloway, R. L., and Levett, P. N. (2008). Evaluation of a modified pulsed-field gel electrophoresis approach for the identification of *Leptospira* serovars. *American Journal of Hygiene and Tropical Medicine* 78(4): 628–632.
- Ganoza, C. A., Matthias, M. A., Saito, M., Cespedes, M., Gotuzzo, E., and Vinetz, J. M. (2010). Asymptomatic renal colonization of humans in the peruvian Amazon by *Leptospira*. *PLoS neglected tropical diseases* 4(2): e612.
- Garba, B., Bahaman, A. R., Bejo, S. K., Zakaria, Z., Mutalib, A. R., and Bande, F. (2018). Major epidemiological factors associated with leptospirosis in Malaysia. *Acta Tropica* 178: 242–247.
- Garba, B., Bahaman, A. R., Khairani-Bejo, S., Zakaria, Z., and Mutalib, A. R. (2017). Retrospective Study of Leptospirosis in Malaysia. *EcoHealth* 14(2): 389–398.
- Goldstein, S. F., and Charon, N. W. (1988). Motility of the spirochete leptospira. *Cell Motility and the Cytoskeleton* 9(2): 101–110.
- Gravekamp, C., Van de Kemp, H., Franzen, M., Carrington, D., Schoone, G. J., Van Eys, G. J. J. M., Everard, C. O. R., Hartskeerl, R. A., et al. (1993). Detection of seven species of pathogenic leptospires by PCR using two sets of primers. *Journal of General Microbiology* 139(8): 1691–1700.
- Haake, D. A., and Levett, P. N. (2015). Leptospirosis in humans. In *Leptospira and leptospirosis* (pp. 65-97). Berlin: Springer.
- Harper, G. A., and Bunbury, N. (2015). Invasive rats on tropical islands: Their population biology and impacts on native species. *Global Ecology and Conservation* 3: 607–627.
- Hovind-Hougen, K. A. R. I. (1979). Leptospiraceae, a new family to include *Leptospira* Noguchi 1917 and *Leptonema* gen. nov. *International Journal of Systematic and Evolutionary Microbiology* 29(3): 245-251.
- Hovind-Hougen, K., Ellis, W. A., and Birch-Andersen, A. (1981). *Leptospira parva* sp. nov.: some morphological and biological characters. *Zentralblatt für Bakteriologie, Mikrobiologie und Hygiene. 1. Abt. Originale. A, Medizinische Mikrobiologie, Infektionskrankheiten und Parasitologie* 250(3): 343-354.
- Inada, R., Ido, Y., Hoki, R., Kaneko, R., and Ito, H. (1916). The etiology, mode of infection, and specific therapy of Weil's disease (spirochaetosis icterohaemorrhagica). *The Journal of experimental medicine* 23(3): 377.
- Johnson, R. C., and Harris, V. G. (1967). Differentiation of pathogenic and saprophytic letospires. I. Growth at low temperatures. *Journal of*

Bacteriology 94(1): 27–31.

- Kariv, R., Klempfner, R., Barnea, A., Sidi, Y., and Schwartz, E. (2001). The Changing Epidemiology of Leptospirosis in Israel. *Emerging Infectious Diseases* 7(6): 990–992.
- Kajdacsí, B., Costa, F., Hyseni, C., Porter, F., Brown, J., Rodrigues, G., Farias, H., Reis, M. G., et al. (2013). Urban population genetics of slum-dwelling rats (*Rattus norvegicus*) in Salvador, Brazil. *Molecular Ecology* 22(20): 5056–5070.
- Khairani-Bejo, S., Oii, S. S., and Bahaman, A. R. (2004). Rats: leptospirosis reservoir in Serdang Selangor residential area. *Journal of Animal and Veterinary Advances* 3(2): 66-69.
- Kit, L. S. (2002). Emerging and Re-Emerging Diseases in Malaysia. *Asia Pacific Journal of Public Health* 14(1): 6–8.
- Ko, A. I., Galvão Reis, M., Ribeiro Dourado, C. M., Johnson, W. D., and Riley, L. W. (1999). Urban epidemic of severe leptospirosis in Brazil. Salvador Leptospirosis Study Group. *Lancet (London, England)* 354(9181): 820–825.
- Ko, A. I., Goarant, C., and Picardeau, M. (2009). Leptospira: The Dawn of the Molecular Genetics Era for an Emerging Zoonotic Pathogen. *Nature Reviews Microbiology* 7(10): 736–747.
- Koay, T. K., Nirmal, S., Noitie, L., and Tan, E. (2004). An epidemiological investigation of an outbreak of leptospirosis associated with swimming, Beaufort, Sabah. *Medical Journal of Malaysia* 59(4): 455–459.
- Koizumi, N., Muto, M., Tanikawa, T., Mizutani, H., Sohmura, Y., Hayashi, E., Akao, N., Hoshino, M., et al. (2009). Human leptospirosis cases and the prevalence of rats harbouring *Leptospira interrogans* in urban areas of Tokyo, Japan. *Journal of Medical Microbiology* 58(9): 1227–1230.
- Kumar, S., Stecher, G., and Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution* 33(7):1870–1874.
- Levett, P. N. (2001). Leptospirosis. *Clinical Microbiology* 14(2): 296–326.
- Levett, P. N., Morey, R. E., Galloway, R., Steigerwalt, A. G., and Ellis, W. A. (2005). Reclassification of *Leptospira parva* Hovind-Hougen et al. 1982 as *Turneriella parva* gen. nov., comb. nov. *International journal of systematic and evolutionary microbiology* 55(4): 1497-1499.
- Levett, P. N., Morey, R. E., Galloway, R. L., and Steigerwalt, A. G. (2006). *Leptospira broomii* sp. nov., isolated from humans with

leptospirosis. *International journal of systematic and evolutionary microbiology* 56(Pt 3): 671-673.

Levett, P. N. (2015). Systematics of leptospiraceae. In *Leptospira and Leptospirosis* (pp. 11-20). Berlin: Springer.

Lim, J. K., Murugaiyah, V. A., Ramli, A. S., Rahman, H. A., Mohamed, N. S. F., Shamsudin, N. N., and Tan, J. C. (2011). A case study: leptospirosis in Malaysia. *WebmedCentral INFECTIOUS DISEASES* 2(12): WMC002764.

Maiden, M. C., Bygraves, J. A., Feil, E., Morelli, G., Russell, J. E., Urwin, R., Zhang, Q., Zhou, J., et al. (1998). Multilocus sequence typing: a portable approach to the identification of clones within populations of pathogenic microorganisms. *Proceedings of the National Academy of Sciences* 95(6): 3140-3145.

Manual perhutanan., 2003, Jilid I. 2005. Jabatan Perhutanan Semenanjung Malaysia: Ampang Press Sdn. Bhd.

Mason, M. R., Encina, C., Sreevatsan, S., and Muñoz-Zanzi, C. (2016). Distribution and diversity of pathogenic *Leptospira* species in peri-domestic surface waters from south central Chile. *PLoS neglected tropical diseases* 10(8): e0004895.

Masuzawa, T., Okamoto, Y., Une, Y., Takeuchi, T., Tsukagoshi, K., Koizumi, N., Kawabata, H., Ohta, S., et al. (2006). Leptospirosis in squirrels imported from United States to Japan. *Emerging Infectious Diseases* 12(7): 1153–1155.

Marshall, and RB. (1992). International Committee on Systematic Bacteriology Subcommittee on the Taxonomy of *Leptospira*: Minutes of the Meetings, 13 and 15 September 1990, Osaka, Japan. *International Journal of Systematic Bacteriology* 42(2): 330–334.

Matthias, M. A., Ricaldi, J. N., Cespedes, M., Diaz, M. M., Galloway, R. L., Saito, M., Steigerwalt, A. G., Patra, K. P., et al (2008). Human leptospirosis caused by a new, antigenically unique *Leptospira* associated with a *Rattus* species reservoir in the Peruvian Amazon. *PLoS neglected tropical diseases* 2(4): e213.

Mayer-Scholl, A., Hammerl, J. A., Schmidt, S., Ulrich, R. G., Pfeffer, M., Woll, D., Scholz, H. C., Thomas, A., et al (2014). *Leptospira* spp. in rodents and shrews in Germany. *International Journal of Environmental Research and Public Health* 11(8): 7562–7574.

Meeus, S. J., and Gulinck, H. (2008). Semi-urban areas in landscape research: A review. *Living Reviews in Landscape Research* 2(3): 1-45.

Mgode, G. F., Machang'u, R. S., Mhamphi, G. G., Katakweba, A., Mulungu, L. S., Durnez, L., Leirs, H., Hartskeerl, R. A., et al (2015). *Leptospira*

serovars for diagnosis of leptospirosis in humans and animals in Africa: common *Leptospira* isolates and reservoir hosts. *PLoS neglected tropical diseases* 9(12): e0004251.

Mohamed-Hassan, S. N., Bahaman, A. R., Mutalib, A. R., and Khairani-Bejo, S. (2010). Serological prevalence of leptospiral infection in wild rats at the national service training centres in Kelantan and Terengganu. *Tropical Biomedicine* 27(1): 30–32.

Mohamed-Hassan, S. N., Bahaman, A. R., Mutalib, A. R., and Khairani-Bejo, S. (2012). Prevalence of pathogenic leptospires in rats from selected locations in peninsular Malaysia. *Research Journal of Animal Sciences* 6(1): 12-25.

Monahan, A. M., Miller, I. S., and Nally, J. E. (2009). Leptospirosis: risks during recreational activities. *Journal of Applied Microbiology* 107(3): 707–716.

Morey, R. E., Galloway, R. L., Bragg, S. L., Steigerwalt, A. G., Mayer, L. W., and Levett, P. N. (2006). Species-specific identification of Leptospiraceae by 16S rRNA gene sequencing. *Journal of clinical microbiology* 44(10): 3510-3516.

Mortimer, R. B. (2005). Leptospirosis in a caver returned from Sarawak, Malaysia. *Wilderness and Environmental Medicine* 16(3): 129–31.

Murray, G. L. (2013). The lipoprotein LipL32, An enigma of leptospiral biology. *Veterinary Microbiology* 162(2–4): 305–314.

Murray, G. L. (2015). The molecular basis of leptospiral pathogenesis. In *Leptospira and Leptospirosis* (pp. 139-185). Berlin: Springer.

Nally, J. E., Monahan, A. M., Miller, I. S., Bonilla-Santiago, R., Souda, P., and Whitelegge, J. P. (2011). Comparative proteomic analysis of differentially expressed proteins in the urine of reservoir hosts of leptospirosis. *PLoS ONE* 6(10): 1–9.

Noguchi, H. (1917). Spirochæta Icterohæmorrhagiæ in American Wild Rats And Its Relation To The Japanese And European Strains: First Paper. *The Journal of experimental medicine* 25(5): 755.

Oliveira, M. A. A., Caballero, O. L., Vago, A. R., Harskeerl, R. A., Romanha, A. J., Pena, S. D. J., Simpson, A. J. and Koury, M. C. (2003). Low-stringency single specific primer PCR for identification of *Leptospira*. *Journal of Medical Microbiology* 52(2): 127–135.

Other Spirochetoses. (2017). In *Tropical Dermatology* (2nd ed., pp. 346–358). Elsevier.

- Palaniappan, R. U., Ramanujam, S., and Chang, Y. F. (2007). Leptospirosis: pathogenesis, immunity, and diagnosis. *Current opinion in infectious diseases* 20(3): 284-292.
- Pappas, G., Papadimitriou, P., Siozopoulou, V., Christou, L., and Akritidis, N. (2008). The globalization of leptospirosis: worldwide incidence trends. *International Journal of Infectious Diseases* 12(4): 351–357.
- Perez, J., Brescia, F., Becam, J., Mauron, C., and Goarant, C. (2011). Rodent abundance dynamics and leptospirosis carriage in an area of hyper-endemicity in New Caledonia. *PLoS neglected tropical diseases* 5(10): e1361.
- Perolat, P., Chappel, R. J., Adler, B., Baranton, G., Bulach, D. M., Billingham, M. L., Letocart, M., Merien, F., et al. (1998). *Leptospira fainei* sp. nov., isolated from pigs in Australia. *International Journal of Systematic Bacteriology* 48 Pt 3: 851–8.
- Picardeau, M. (2013). Diagnosis and epidemiology of leptospirosis. *Médecine et maladies infectieuses* 43(1): 1-9.
- Picardeau, M., Bulach, D. M., Bouchier, C., Zuerner, R. L., Zidane, N., Wilson, P. J., Creno, S., Kuczek, E. S., et al. (2008). Genome sequence of the saprophyte *Leptospira biflexa* provides insights into the evolution of *Leptospira* and the pathogenesis of leptospirosis. *PLoS ONE* 3(2): 1–9.
- Postic, D., Riquelme-Sertour, N., Merien, F., Perolat, P., and Baranton, G. (2000). Interest of partial 16S rDNA gene sequences to resolve heterogeneities between *Leptospira* collections: Application to *L. meyeri*. *Research in Microbiology* 151(5): 333–341.
- Priya, C. G., Hoogendijk, K. T., Berg, M. V. D., Rathinam, S. R., Ahmed, A., Muthukkaruppan, V. R., and Hartskeerl, R. A. (2007). Field rats form a major infection source of leptospirosis in and around Madurai, India. *Journal of postgraduate medicine* 53(4): 236-40.
- Puche, R., Ferrés, I., Caraballo, L., Rangel, Y., Picardeau, M., Takiff, H., and Iraola, G. (2018). *Leptospira venezuelensis* sp. nov., a new member of the intermediate group isolated from rodents, cattle and humans. *International Journal of Systematic and Evolutionary Microbiology* 68: 513-517.
- Pui, C. F., Bilung, L. M., Apun, K., and Su'ut, L. (2017). Diversity of *Leptospira* spp. in Rats and Environment from Urban Areas of Sarawak, Malaysia. *Journal of Tropical Medicine* 2017(ID3760674): 1–8.
- Pui, C. F., Bilung, L. M., Su'ut, L., and Apun, K. (2015). Prevalence of *Leptospira* Species in Environmental Soil and Water from National Parks in Sarawak, Malaysia. *Borneo Journal of Resource Science and Technology* 5(1): 49–57.

- Rahelinirina, S., Léon, A., Harstskeerl, R. A., Sertour, N., Ahmed, A., Raharimanana, C., Ferquel, E., Garnier, M., et al. (2010). First isolation and direct evidence for the existence of large small-mammal reservoirs of *Leptospira* sp. in Madagascar. *PloS one* 5(11): e14111.
- Rahmat, M. S., MIMLS, K. H., Paramasvaran, S., Azizah, M. R., and Imran, F. (2012). Prevalence of leptospiral DNA among wild rodents from a selected area in Beguk Dam Labis, Segamat, Johor, Malaysia. *The Malaysian journal of pathology* 34(2): 157.
- Rafizah, A. A. N., Aziah, B. D., Azwany, Y. N., Imran, M. K., Rusli, A. M., Nazri, S. M., Nikman, A. M., Nabilah, I., et al. (2013). Risk factors of leptospirosis among febrile hospital admissions in northeastern Malaysia. *Preventive Medicine*, 57 Suppl: S11-3.
- Ramadass, P., Jarvis, B. D. W., Corner, R. J., Penny, D., and Marshall, R. B. (1992). Genetic characterization of pathogenic *Leptospira* species by DNA hybridization. *International Journal of Systematic and Evolutionary Microbiology* 42(2): 215-219.
- Ridzlan, F. R., Bahaman, a. R., Khairani-Bejo, S., and Mutalib, a. R. (2010). Detection of pathogenic *Leptospira* from selected environment in Kelantan and Terengganu, Malaysia. *Tropical Biomedicine* 27(3): 632–638.
- Ridzuan, M. J., Aziah, B. D., and Zahiruddin, W. M. (2016). The Occupational Hazard Study for Leptospirosis among Agriculture Workers. *International Journal of Collaborative Research on Internal Medicine and Public Health* 8(3): MA13-MA22.
- Robins, J. H., Hingston, M., Matisoo-Smith, E. and Ross, H. A. (2007). Identifying *Rattus* species using mitochondrial DNA. *Molecular Ecology Notes* 7(5): 717-729.
- Romero, E. C., Blanco, R. M., and Galloway, R. L. (2011). Analysis of Multilocus Sequence Typing for identification of *Leptospira* spp. Isolates in Brazil. *Journal of clinical microbiology* 49(11): 3940-2.
- Rossetti, C. A., Liem, M., Samartino, L. E., and Hartskeerl, R. A. (2005). Buenos Aires, a new *Leptospira* serovar of serogroup Djasiman, isolated from an aborted dog fetus in Argentina. *Veterinary Microbiology* 107(3–4): 241–248.
- Ruppert, N. A. D. I. N. E., Asyraf, M., and Shahrul Anuar, M. S. (2012). One year mark-and-recapture study on small terrestrial mammals at Segari Melintang Forest Reserve, Perak: assessing trap height and bait preferences and efficiency of different tagging methods for longterm identification. Seminar on long-term ecological research in the EAP-region. 4th July.

- Russ, A., Jali, I., Bahaman, A., Tuen, A., and Ismail, G. (2003). Seroepidemiological study of leptospirosis among the indigenous communities living in the periphery of Crocker Range Park Sabah, Malaysia. *ASEAN Review of Biodiversity and Environmental Conservation* (pp. 1-5): 1175-5326.
- Saito, M., Villanueva, S. Y., Chakraborty, A., Miyahara, S., Segawa, T., Asoh, T., Ozuru, R., Gloriani, N.G., et al. (2013). Comparative analysis of *Leptospira* strains isolated from environmental soil and water in the Philippines and Japan. *Applied and Environmental Microbiology* 79(2): 601-609.
- Samsudin, S., Masri, S. N., Tengku Jamaluddin, T. Z. M., Saudi, S. N. S., Md Ariffin, U. K., Amran, F., and Osman, M. (2015). Seroprevalence of leptospiral antibodies among healthy municipal service workers in Selangor. *Advances in Public Health* 23(3): 327-333.
- Sapian, M., Khairi, M. T., How, S. H., Rajalingam, R., Sahhir, K., Norazah, A., ... Jamalludin, A. R. (2012). Outbreak of Melioidosis and Leptospirosis Co-infection following a rescue operation. *Med J Malaysia*. 67(3): 293-297.
- Schneider, M. C., Jancloes, M., Buss, D. F., Aldighieri, S., Bertherat, E., Najera, P., Galan, D. I., Durski, K., et al. (2013). Leptospirosis: A silent epidemic disease. *International Journal of Environmental Research and Public Health* 10(12): 7229-7234.
- Scola, B. La, Bui, L. T. M., Baranton, G., Khamis, A., and Raoult, D. (2006). Partial rpoB gene sequencing for identification of *Leptospira* species. *FEMS Microbiology Letters* 263(2): 142-147.
- Sejvar, J., Bancroft, E., Winthrop, K., Bettinger, J., Bajani, M., Bragg, Shutt, S. K., Kaiser, R., et al. (2003). Leptospirosis in "Eco-Challenge" athletes, Malaysian Borneo, 2000. *Emerging Infectious Diseases* 9(6): 702-707.
- Slack, A. T., Symonds, M. L., Dohnt, M. F., and Smythe, L. D. (2006). Identification of pathogenic *Leptospira* species by conventional or real-time PCR and sequencing of the DNA gyrase subunit B encoding gene. *BMC microbiology* 6(1): 95.
- Slack, A., Symonds, M., Dohnt, M., Harris, C., Brookes, D., and Smythe, L. (2007). Evaluation of a modified Taqman assay detecting pathogenic *Leptospira* spp. against culture and *Leptospira*-specific IgM enzyme-linked immunosorbent assay in a clinical environment. *Diagnostic Microbiology and Infectious Disease* 57(4): 361-366.
- Slack, A. T., Kalambaheti, T., Symonds, M. L., Dohnt, M. F., Galloway, R. L., Steigerwalt, A. G., Chaicumpa, W., Bunyaraksyotin, G., et al. (2008). *Leptospira wolffii* sp. nov., isolated from a human with suspected leptospirosis in Thailand. *International Journal of Systematic and Evolutionary Microbiology* 58(10): 2305-2308.

- Slack, A. T., Khairani-Bejo, S., Symonds, M. L., Dohnt, M. F., Galloway, R. L., Steigerwalt, A. G., Bahaman, A. R., Craig, S., et al. (2009). *Leptospira kmetyi* sp. nov., isolated from an environmental source in Malaysia. *International journal of systematic and evolutionary microbiology* 59(4): 705-708.
- Smith, C. E., Turner, L. H., Harrison, J. L., and Broom, J. C. (1961). Animal leptospirosis in Malaya: 1. Methods, zoogeographical background, and broad analysis of results. *Bulletin of the World Health Organization* 24(1): 5-21.
- Smythe, L., Adler, B., Hartskeerl, R. A., Galloway, R. L., Turenne, C. Y., and Levett, P. N. (2013). Classification of *Leptospira* genomospecies 1, 3, 4 and 5 as *Leptospira alstonii* sp. nov., *Leptospira vanthielii* sp. nov., *Leptospira terpstrae* sp. nov. and *Leptospira yanagawae* sp. nov., respectively. *International Journal of Systematic and Evolutionary Microbiology* 63(5): 1859-1862.
- Stimson, A. M. (1907). Note on an Organism Found in Yellow-Fever Tissue. *Public Health Reports (1896-1970)* 22(18): 541.
- Sumanta, H., Wibawa, T., Hadisusanto, S., Nuryati, A., and Kusnanto, H. (2015). Genetic variation of *Leptospira* isolated from rats caught in Yogyakarta Indonesia. *Asian Pacific Journal of Tropical Medicine* 8(9): 710-713.
- Suut, L., Mazlan, M. N.-A., Arif, M. T., Yusoff, H., Abdul Rahim, N.-A., Safii, R., and Suhaili, M. R. (2016). Serological Prevalence of Leptospirosis Among Rural Communities in the Rejang Basin, Sarawak, Malaysia. *Asia Pacific Journal of Public Health* 28(5): 450-457.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., and Kumar, S. (2011). MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution* 28(10): 2731-9.
- Tan, W. L., Soelar, S. A., Suan, M. A. M., Hussin, N., Cheah, W. K., Verasahib, K., and Goh, P. P. (2016). Leptospirosis incidence and mortality in Malaysia. *Southeast Asian Journal of Tropical Medicine and Public Health* 47(3): 434-440.
- Terpstra, W. J., Korver, H., Schoone, G. J., Leeuwen, J. v., Schönemann, C. E., De Jonge-Aglibut, S., and Kolk, A. H. J. (1987). Comparative classification of leptospira serovars of the Pomona group by monoclonal antibodies and restriction-endonuclease analysis. *Zentralblatt Fur Bakteriologie Mikrobiologie Und Hygiene - Abt. 1 Orig. A* 266(3-4): 412-421.
- Thayaparan, S., Robertson, I. A. N., Amraan, F., Ut, L. S. U., and Abdullah, M. T. (2013). Serological Prevalence of Leptospiral Infection in Wildlife in Sarawak, Malaysia. *Borneo Journal of Resource Science and Technology* 2: 71-74.

- Thayaparan, S., Robertson, I. D., Fairuz, a., Suut, L., and Abdullah, M. T. (2013). Leptospirosis, an emerging zoonotic disease in Malaysia. *Malaysian Journal of Pathology* 35(2):123–132.
- Ungku Omer, A. (1967). Veterinary public health with particular reference to Malaysia. *Kajian Veterinier* 1: 54-62.
- Valverde, M. de los A., Ramírez, J. M., Oca, L. G. M. de, Goris, M. G. A., Ahmed, N., and Hartskeerl, R. A. (2008). Arenal, a new *Leptospira* serovar of serogroup Javanica, isolated from a patient in Costa Rica. *Infection, Genetics and Evolution* 8(5): 529–533.
- Van Crevel, R., Speelman, P., Gravekamp, C., and Terpstra, W. J. (1994). Leptospirosis in travelers. *Clinical Infectious Diseases* 19(1): 132-134.
- Van Thiel, P. H. (1948). *The Leptospiroses*. Universitaire Pers. Leiden; the Netherlands.
- Victoria, B., Ahmed, A., Zuerner, R. L., Ahmed, N., Bulach, D. M., Quinteiro, J., and Hartskeerl, R. A. (2008). Conservation of the S10-spc- α locus within otherwise highly plastic genomes provides phylogenetic insight into the genus *Leptospira*. *PLoS ONE* 3(7): 1–9.
- Victoriano, A. F. B., Smythe, L. D., Gloriani-Barzaga, N., Cavinta, L. L., Kasai, T., Limpakarnjanarat, K., Ong, B. L., Gongal, G., et al. (2009). Leptospirosis in the Asia Pacific region. *BMC Infectious Diseases* 9(1): 147.
- Vijayachari, P., Sugunan, A. P., and Shriram, A. N. (2008). Leptospirosis: an emerging global public health problem. *Journal of Biosciences* 33(4): 557–569.
- Wahab, Z. (2015). Epidemiology and Current Situation of Leptospirosis in Malaysia. *Jkt.Kpkt.Gov.My*. Retrieved from http://jkt.kpkt.gov.my/jkt/resources/PDF/Persidangan_2015/persidangan_kesihatan/Leptospirosis_in_Malaysia.pdf
- Waitkins, S. A. (1986). Leptospirosis as an occupational disease. *British Journal of Industrial Medicine* 43(11): 721.
- Walch-Sorgdrager, B. (1939). Leptospiroses. *Bulletin Health Organisation (League of Nations)* 8: 143–386.
- Watt, G., Alquiza, L. M., Padre, L. P., Tuazon, M. L., and Laughlin, L. W. (1988). The Rapid Diagnosis of Leptospirosis: A Prospective Comparison of the Dot Enzyme-Linked Immunosorbent Assay and the Genus-Specific Microscopic Agglutination Test at Different Stages of Illness. *The Journal of Infectious Diseases* 157(4): 840-842.

- Webster, J. P., Ellis, W. A., and Macdonald, D. W. (1995). Prevalence of *Leptospira* spp. in wild brown rats (*Rattus norvegicus*) on UK farms. *Epidemiology and Infection* 114(1): 195-201.
- WHO. (2003a). Human leptospirosis: guidance for diagnosis, surveillance and control. *WHO Library* 45(5): 1–109.
- WHO. (2003b). Human leptospirosis: guidance for diagnosis, surveillance and control. *WHO Library* 45(5): 1–109.
- Wolff, J. (1954). In The laboratory diagnosis of leptospirosis. *American lecture series, no. 183. (pp.xiv-99)*. Springfield.
- Xue, F., Dong, H., Wu, J., Wu, Z., Hu, W., Sun, A., Troxell, B., Yang, X. F., et al. (2010). Transcriptional responses of *Leptospira interrogans* to host innate immunity: Significant changes in metabolism, oxygen tolerance, and outer membrane. *PLoS Neglected Tropical Diseases* 4(10): e857.
- Yasuda, P. H., Steigerwalt, A. G., Sulzer, K. R., Kaufmann, A. F., Rogers, F., and Brenner, D. J. (1987). Deoxyribonucleic Acid Relatedness between Serogroups and Serovars in the Family Leptospiraceae with Proposals for Seven New *Leptospira* Species. *International Journal of Systematic Bacteriology* 37(4): 407–415.
- Zuerner, R. L., Hartskeerl, R. A., van de Kemp, H., and Bal, A. E. (2000). Characterization of the *Leptospira interrogans* S10-spc-alpha operon. *FEMS Microbiology Letters* 182(2): 303–308.

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LIST OF PUBLICATIONS

- Azhari, N. N.**, Ramli, S. N. A., Joseph, N., Philip, N., Mustapha, N. F., Ishak, S. N., Mohd-Taib, F. S., Md Nor, S., et al. (2018). Molecular characterization of pathogenic *Leptospira* sp. in small mammals captured from the human leptospirosis suspected areas of Selangor state, Malaysia. *Acta tropica*, 188, 68-77. (Published)
- Ramli, S. N. A., Joseph, N., Philip, N., **Azhari, N. N.**, Garba, B., Masri, S. N., Sekawi, Z., Neela, V. K. (2018). Diagnostic accuracy of rapid diagnostic tests for the early detection of leptospirosis. *Journal of Infection and Public Health*. S1876-0341(18): 30307-1. (Published)
- Yusof, M. A., Mohd-Taib, F. S., Ishak, S. N., Md-Nor, S., Md-Sah, S. A., Mohamed, N. Z., **Azhari, N. N.**, Neela, V. K. & Sekawi, Z. (2018). Microhabitat Factors Influenced Prevalence of Pathogenic *Leptospira* sp. in Small Mammals Host. *EcoHealth*. (Accepted)
- Neela, V. K., **Azhari, N. N.**, Joseph, N., Philip, N., Alia, S. N., Mustapha, N. F., Ishak, S. N., Mohd-Taib, F. S., et al. (2018). An outbreak of Leptospirosis among reserve military recruits, Hulu Perdik, Malaysia. *European Journal of Clinical Microbiology & Infectious Diseases*. (Published)



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