



UNIVERSITI PUTRA MALAYSIA

***BACTERIAL CELLULOSE FROM A. XYLINUM FERMENTATION USING  
HYDROLYSATE OF BANANA-PEEL WASTE AS CARBON SOURCE***

MUSTAPHA BIN AKIL

FK 2011 165

BACTERIAL CELLULOSE FROM *A. XYLINUM* FERMENTATION USING  
HYDROLYSATE OF BANANA-PEEL WASTE AS CARBON SOURCE

by

MUSTAPHA BIN AKIL



Thesis Submitted to the School of Graduates Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirement for the Degree of Master of Science

December 2011

Abstract of thesis presented to the Senate Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science.

**BACTERIAL CELLULOSE FROM *A. XYLINUM* FERMENTATION USING HYDROLYSATE OF BANANA-PEEL WASTE AS CARBON SOURCE**

By

**MUSTAPHIA BIN AKIL**

**December 2011**

**Chair:** **Dayang Radiah Bt. Awang Biak, PhD**

**Faculty:** **Faculty of Engineering**

The high cost of utilizing commercial sugar as carbon source in bacterial cellulose (BC) production has become an issue in the industry. Discovery of alternative carbon source from cheap sources, e.g. agricultural (food) waste is needed to overcome this problem. Two key objectives are set in this study, namely (i) to investigate the reducing sugar yield obtained from the diluted acid hydrolysis of agricultural (food) waste and (ii) to synthesize and characterize BC using hydrolysate obtained in (i) as the carbon source.

Sago effluent, potato peel and banana peel were selected as raw material. The dilute acid hydrolysis method was used to hydrolyze these wastes. Three hydrolysis parameters, sulphuric acid concentration (2%, 4% and 6%), reaction time (20, 40 and 60 minutes) and temperature (105°C and 121°C) were studied. The highest glucose yield for each waste's hydrolysate was used in the fermentation experiment to screen the best carbon source for BC production.

Hydrolysis of potato produced the highest glucose yield, *i.e.* 43.44 mg/ml using 4% (v/v) acid at 20 minutes reaction time and 121°C reaction temperature. The highest glucose yield, *i.e.* 19.15°C from banana peel hydrolysis was obtained after 20 minutes using 6% (v/v) of acid at 121°C whilst for sago effluent, as reaction time was extended to 40 minutes using 4% (v/v) acid at 105°C, only 2.64 mg/ml of glucose was obtained.

Banana peel hydrolysate was selected as the carbon source used in this study because it produced the highest yield of BC, *i.e.* 2.89 g/l after 21 days of incubation period compared to other hydrolysate. For the selection of the suitable initial pH and initial carbon source concentration, it was found that initial pH 6 and 2.0% of carbon source concentration produced 2.81 g/l of BC after 16 days incubation. SEM analysis shows that BC has ultrafine fibrils and a pore-like reticulated structure. TGA result shows that BC was rapidly degraded at 250°C. The existence of common functional group *i.e.* OH group, C=O group, C-H bending, C-H stretching, C-H wagging, C-C stretching, C-O-C vibration and C-O stretching confirms the purity of BC. From BET and BJH analysis, BC has mesopore range of porosity with maximum pore size of 24.629Å during adsorption and 19.070Å for desorption process. The total surface area of the BC was 10.891 m<sup>2</sup>/g. Investigation of tensile strength shows that BC has maximum tensile stress and Young's modulus values at 55.84 MPa and 733.61 MPa, respectively.

In conclusion, this study provides information on converting agricultural (food) waste into fermentable sugar via dilute acid hydrolysis where it can be utilized for BC production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains.

**SELULOSA BAKTERIA DARIPADA FERMENTASI *A. XYLINUM*  
MENGGUNAKAN HIDROLISAT SISA KULIT PISANG SEBAGAI SUMBER  
KARBON**  
**Oleh**

**MUSTAPHA BIN AKIL**

**Disember 2011**

**Pengerusi: Dayang Radiah Bt. Awang Biak, PhD**

**Fakulti: Fakulti Kejuruteraan**

Kos yang tinggi apabila menggunakan gula komersial sebagai sumber karbon dalam penghasilan selulosa bakteria (BC) menjadi satu isu kepada industri. Pencarian sumber karbon alternatif daripada sumber yang murah seperti sisa tanaman (makanan) adalah perlu untuk mengatasi masalah ini. Dua objektif utama telah ditetapkan dalam kajian ini iaitu (i) mengkaji hasil gula terturun yang diperoleh daripada hidrolisis asid cair terhadap sisa tanaman (makanan) dan (ii) mensintesis BC menggunakan hidrolisat yang diperoleh daripada objektif (i) sebagai sumber karbon.

Efluen sagu, kulit kentang dan kulit pisang telah dipilih sebagai bahan mentah. Kaedah hidrolisis asid cair telah digunakan untuk menghidrolisis sisa-sisa tersebut. Tiga parameter hidrolisis iaitu kepekatan asid sulfurik (2%, 4% dan 6%), masa tindak balas

(20, 40 dan 60 minit) dan suhu ( $105^{\circ}\text{C}$  dan  $121^{\circ}\text{C}$ ) telah dikaji. Kandungan glukosa tertinggi bagi setiap hidrolisat daripada sisa telah digunakan dalam eksperimen penapaian untuk menyaring sumber karbon terbaik untuk penghasilan BC.

Hidrolisis bagi kulit kentang menghasilkan glukosa tertinggi iaitu 43.44 mg/ml dengan menggunakan asid berkepekatan 4% (v/v) selama 20 minit masa tindak balas dengan suhu tindak balas pada  $121^{\circ}\text{C}$ . Nilai glukosa tertinggi iaitu 19.15mg/ml diperoleh daripada hidrolisis kulit pisang setelah 20 minit masa tindak balas berlaku menggunakan asid berkepekatan 6% (v/v) pada suhu  $121^{\circ}\text{C}$  sementara bagi efluen sagu, setelah masa tindak balas dilanjutkan ke 40 minit menggunakan asid berkepekatan 4% (v/v) pada suhu  $105^{\circ}\text{C}$ , hanya 2.64mg/ml glukosa telah diperoleh.

Hidrolisat kulit pisang telah dipilih sebagai sumber karbon yang digunakan dalam kajian ini kerana ia menghasilkan BC dengan hasil tertinggi iaitu 2.89 g/l selepas 21 hari tempoh inkubasi berbanding hidrolisat lain. Oleh itu, hidrolisat kulit pisang telah dipilih sebagai sumber karbon dalam kajian ini. Dalam pemilihan pH permulaan dan kepekatan sumber karbon permulaan, pH 6 dan sumber karbon berkepekatan 2.0% menghasilkan 2.81 g/l BC selepas 16 hari tempoh inkubasi. Analisis Mikroskop Elektron Pengimbas (SEM) menunjukkan bahawa BC mempunyai fibril yang sangat halus dan struktur liang yang bersaring. Analisis TGA menunjukkan BC didegradasi dengan cepat pada suhu  $250^{\circ}\text{C}$ . Kehadiran kumpulan bersfungsi yang biasa seperti kumpulan O-H, kumpulan C=O, lenturan C-H, regangan C-H, goyangan C-H, regangan C-C, getaran C-O-C dan regangan C-O memastikan keaslian BC. Daripada analisis BET dan BJH, BC mempunyai keliangan pada julat mesoliang dengan saiz liang maksimum adalah

24.629 Å semasa proses penjerapan dan 19.070 Å bagi proses penyahjerapan. Jumlah luas permukaan BC adalah  $10.891\text{m}^2/\text{g}$ . Kajian terhadap kekuatan tegangan menunjukkan BC mempunyai nilai tegasan tengangan maksimum dan modulus Young pada 55.84 MPa dan 733.61 MPa masing-masing.

Kesimpulannya, kajian ini memberi informasi dalam menukarkan sisa tanaman (makanan) kepada gula boleh tapai melalui hidrolisis asid cair yang mana ia boleh digunakan dalam penghasilan BC.

## **ACKNOWLEDGEMENT**

“In the name of Allah S.W.T., the most Benevolent and Merciful”

Alhamdulillah, finally I managed to finish my study. First and foremost, I would like to express my sincere gratitude to the beloved chairperson of my supervisory committee, Dr. Dayang Radiah Awang Biak, for the continuous support of my study. Her patience, enthusiasm, kindness, immense knowledge and encouragement helped me in all the time of research and writing of this thesis. She gave me the confidence to start my study and taught me for solution of any problem related to my work. Big thanks for you, Dr.

To the member of my supervisory committee, Prof. Madya Dr. Rosfarizan Mohamad and Dr. Intan Salwani Ahamad, your encouragement, support and idea led me to finish my study here. No words can I say but million thanks for your kindness.

To the first person whom introduces me back to this field, Madam Suryani Kamaruddin, your guidance and help are much appreciated. Thank you doesn't seem sufficient but it is said with appreciation and respect.

The kindness of Universiti Putra Malaysia for giving me chance and opportunity to further my study here and financing my study with scholarship is much appreciated. I would like to express my gratitude to all the staffs especially from Chemical and Environmental Engineering Department, Faculty of Engineering for their contribution and full support.

A special thanks to someone special, Jie, your support and patience throughout my study are immeasurable. To my fellow labmates and friends, Meen, Kak Za, Usop, Meg, Amir, Taha, Wan, Zaza, Fadh, Kak Zila, Kak Yan, Mustika, Toybah, Nadh, Pqa, Maziah, Yana and Nik, our routine as research student with a lot of joy, happiness and sadness is a very nice moment in my life.

Sincere and warmest gratitude to my beloved mother, Hjh. Jaharia Bte Akkuahi and my father, Hj. Akil Hamaruddin for giving birth to me at first place and to all my family members especially my brothers, Boby, Nizam and Jasni for their payers, love and generosity throughout my life. Last but not least, million thanks to my best friend and housemate, Fizi and Ditha for their courage and support.

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	ii
<b>ABSTRAK</b>	iv
<b>ACKNOWLEDGEMENTS</b>	vii
<b>APPROVAL</b>	ix
<b>DECLARATION</b>	xi
<b>LIST OF TABLES</b>	xiv
<b>LIST OF FIGURES</b>	xvi
<b>LIST OF ABBREVIATION</b>	xix
 <b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	 1
1.1 Background	1
1.2 Problem statements	2
1.3 Objectives	3
1.4 Scope of work	4
1.5 Thesis outline	4
<b>2 LITERATURE REVIEW</b>	 5
2.1 Introduction	5
2.2 Wastes in Malaysia	5
2.3 Pretreatment of waste	8
2.4 Acid hydrolysis	9
2.5 Bacterial Cellulose	13
2.6 <i>Acetobacter xylinum</i>	21
2.7 Carbon source for BC production	24
2.8 Applications of bacterial cellulose	27
<b>3 METHODOLOGY</b>	 30
3.1 Introduction	30
3.2 Sampling	31
3.3 Diluted acid hydrolysis	32
3.4 Sugar analysis	33
3.6 Preparation for seed culture of <i>Acetobacter xylinum</i>	34
3.6 Bacterial cellulose production culture	35
3.6.1 Screening test	35
3.6.2 Selection of parameter	35

3.7	Harvesting method	36
3.8	Analytical method	37
3.8.1	Sugar Content and pH analysis	37
3.8.2	Cell mass	37
3.8.3	Thickness	38
3.8.4	Wet weight and dry weight	38
3.8.5	Scanning Electron Microscope (SEM)	38
3.8.6	Thermogravimetric Analysis (TGA)	39
3.8.7	Fourier Transform Infrared Analaysis (FTIR)	39
3.8.8	Surface area and porosity analysis	39
3.8.9	Tensile strength	40
<b>4</b>	<b>RESULTS AND DISCUSSIONS</b>	<b>41</b>
4.1	Introduction	41
4.2	Sugar analysis for sago effluent hydrolysate	42
4.3	Sugar analysis for potato peel hydrolysate	44
4.4	Sugar analysis for banana peel hydrolysate	49
4.5	Seed culture	54
4.6	Screening of carbon source from agro waste hydrolysate	55
4.7	Selection of parameters	57
4.8	Cell growth	59
4.9	Product formation and substrate utilization	61
4.10	Product characterization	64
4.10.1	Morphology of BC	64
4.10.2	Thermal stability of BC	66
4.10.3	FTIR analysis	67
4.10.4	Total surface area and porosity analysis	68
4.10.5	Tensile strength	70
<b>5</b>	<b>CONCLUSIONS AND RECOMMENDATIONS</b>	<b>73</b>
5.1	Conclusions	73
5.2	Recommendations for future research	75
<b>REFERENCES</b>		<b>76</b>
<b>APPENDICES</b>		<b>87</b>
<b>BIODATA OF STUDENT</b>		<b>95</b>
<b>LIST OF RELATED PUBLICATIONS</b>		<b>96</b>

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
2.1	A few hydrolysis processes from different raw material, different process and different purpose	10
2.2	Common functional group for BC	19
2.3	The carbon source percentage and pH for the growth of <i>Acetobacter xylinum</i> to produce BC	22
2.4	Effects of composition ratios of carbon source mixtures on the cellulose production by <i>A. xylinum</i> BRC5 (adapted from Yang <i>et al.</i> , 1998)	26
3.1	Parameters chosen for acid hydrolysis operation	32
3.2	HS medium recipe for seed culture	34
3.3	Preliminary test to find the suitable pH range for different concentration of banana peel hydrolysate used	36
B1	OD reading for cell mass determination without cellulase enzyme	89
C1	Sago effluent hydrolysate obtained using 4% of acid concentration reacted for 40 minutes at 105°C. (Initial pH 6)	90
C2	Potato peel hydrolysate obtained using 4% of acid concentration reacted for 20 minutes at 121°C. (Initial pH 6)	90
C3	Banana peel hydrolysate obtained using 6% of acid concentration reacted for 20 minutes at 121°C. (Initial pH 6)	90
D1	Preliminary test using banana peel hydrolysate at 0.5% concentration of glucose	91
D2	Preliminary test using banana peel hydrolysate at 1.0% concentration of glucose	91
D3	Preliminary test using banana peel hydrolysate at 2.0%	

concentration of glucose	91
E1 BC production for 0.5% initial glucose concentration	92
E2 BC production for 1.0% initial glucose concentration	92
E3 BC production for 2.0% initial glucose concentration	92



## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1	Mechanism of acid-catalyzed hydrolysis of $\beta$ -1-4 glucan (Xiang <i>et al.</i> , 2004)	12
2.2	Formation of bacterial cellulose (Klemm <i>et al.</i> , 2001)	15
2.3	The pathway of cellulose synthesis from glucose and fructose (modified from Chawla <i>et al.</i> , 2009)	16
2.4	Chemical structure of BC	17
2.5	BC microfibril structure under SEM observation (Krystynowicz <i>et al.</i> , 2002).	18
2.6	Schematic diagram of BC degradation using TGA	18
3.1	Summary of the methodology used in this work	31
4.1	HPLC chromatogram for sago effluent hydrolysate	42
4.2	Glucose concentration in sago effluent (a) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (b) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (c) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (d) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (e) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (f) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C.	44
4.3	HPLC chromatogram for potato peel hydrolysate	45
4.4	Glucose concentration in potato peel hydrolysate (a) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (b) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (c) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (d) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (e) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (f) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C.	47
4.5	Xylose concentration in potato peel hydrolysate (a) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (b) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (c) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (d) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (e) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (f) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C	48
4.6	Arabinose concentration in potato peel hydrolysate (a) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (b) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (c) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (d) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (e) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (f) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C	49

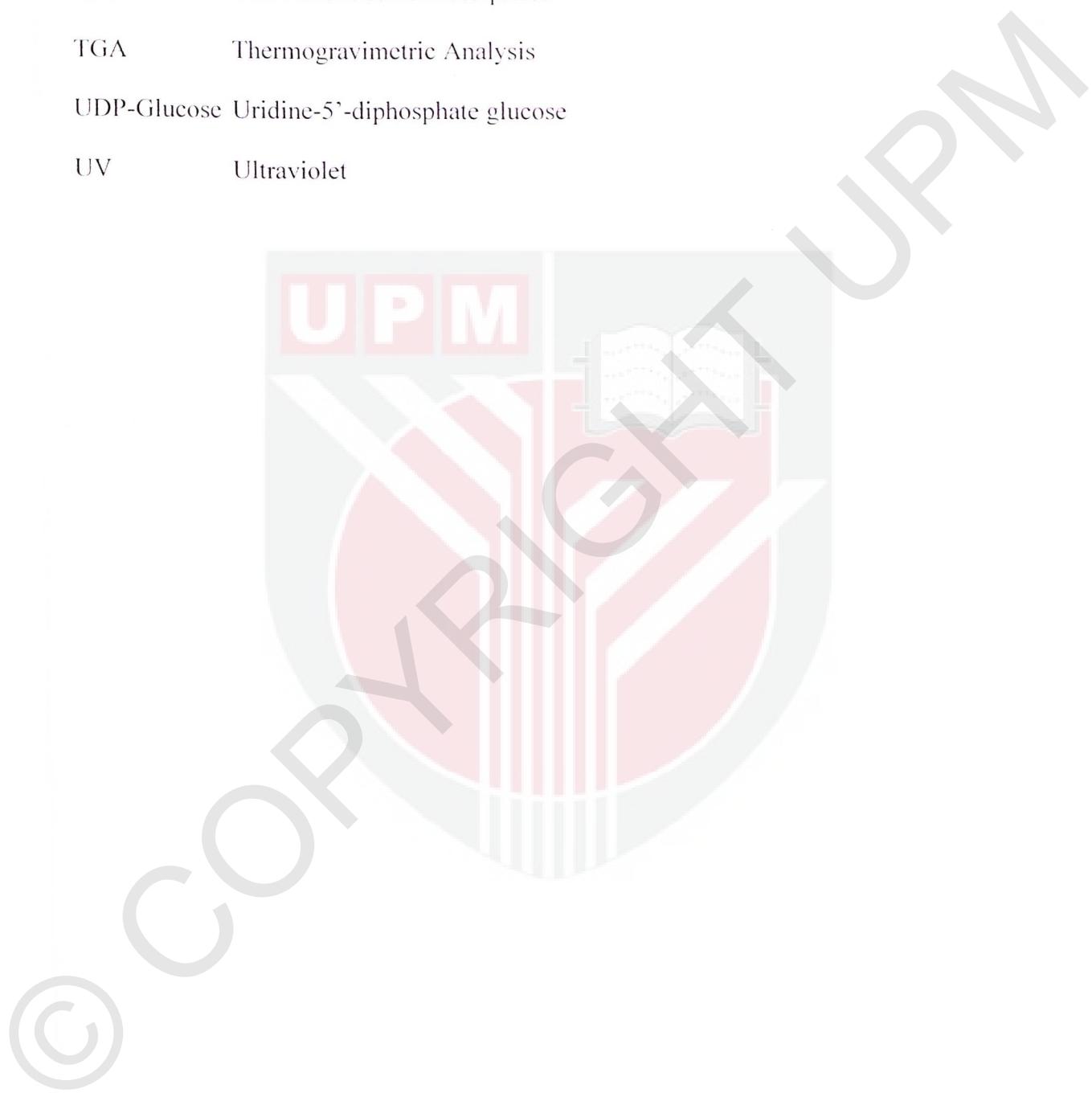
4.7	HPLC chromatogram for banana peel hydrolysate	50
4.8	Glucose concentration in banana peel hydrolysate (a) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (b) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (c) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (d) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (e) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (f) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C	51
4.9	Xylose concentration in banana peel hydrolysate (a) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (b) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (c) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (d) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (e) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (f) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C.	52
4.10	Fructose concentration in banana peel hydrolysate (a) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (b) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (c) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 105°C, (d) 2% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (e) 4% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C, (f) 6% (v/v) H <sub>2</sub> SO <sub>4</sub> , 121°C.	54
4.11	Growth pattern of seed culture	55
4.12	Dry weight (g/l) of BC fermented from different alternative carbon sources.	57
4.13	Dry weight of BC at different initial pH for (●) 0.5 %, (■) 1.0 % and (▲) 2.0 % carbon source concentration after 21 days of incubation.	59
4.14	Growth curve of <i>Acetobacter xylinum</i>	60
4.15	Logarithm graph of <i>Acetobacter xylinum</i> growth	61
4.16	Dry weight of BC using 2.0% glucose concentration and initial pH 6	62
4.17	Time course of BC production and sugar utilization with initial pH 6, (■) glucose, (●) fructose, (▲) xylose, (○) BC production and (—) cell mass	63
4.18	BC structure under SEM. (a) 0.5% carbon source, pH 7, (b) 1.0% carbon source, pH 7, (c) 2.0% carbon source, pH 6 and (d) BC from HS medium by Krystynowicz <i>et al.</i> , (2002).	65
4.19	The TGA curve of BC for (▲) 0.5% carbon source concentration and pH 7, (■) 1.0% carbon source concentration and pH 7 and (●) 2.0%	

carbon source concentration and pH 6 after 16 days of incubation period. Inset is the DTG curve for each condition.	67
4.20 FTIR spectra of BC	68
4.21 The isotherm linear of (●) adsorption and (■) desorption	69
4.22 Pore size distribution for BC (a) adsorption and (b) desorption	71
4.23 Stress-strain curve of BC from banana peel hydrolysate medium	71
4.24 Stress-strain curve of GC from HS medium	69
A1 Glucose calibration curve for HPLC	87
A2 Xylose calibration curve for HPLC	87
A3 Fructose calibration curve for HPLC	88
A4 Arabinose calibration curve for HPLC	88

## LIST OF ABBREVIATION

ATR	Attenuated total reflectance
BC	Bacterial cellulose
BET	Brunauer-Emmett-Teller
BH	Banana peel hydrolysate
BJH	Barret-Joyner-Halenda
Ca	Calcium
$\text{CaCO}_3$	Calcium carbonate
$\text{CaSO}_4$	Calcium sulphate
CM-BC	Carboxymethylated-bacterial cellulose
CMC	Carboxymethylated cellulose
DTG	Derivative Thermogravimetric
EABC	Diethylenetriamine-bacterial cellulose
EDX	Energy Dispersive X-ray
ELSD	Evaporative Light Scanning Detector
FTIR	Fourier Transform Infrared
$\text{H}_2\text{SO}_4$	Sulphuric acid
$\text{H}_3\text{COOH}$	Acetic acid
HPLC	High Performance Liquid Chromatography
HS	Hestrin and Schramm
MARDI	Malaysian Agricultural Research and Development Institute
Na	Sodium
NaOH	Sodium hydroxide

OD	Optical density
PES	Millipore Express PLUS
SEM	Scanning Electron Microscope
SME	Small and medium enterprises
TGA	Thermogravimetric Analysis
UDP-Glucose	Uridine-5'-diphosphate glucose
UV	Ultraviolet



## CHAPTER 1

### INTRODUCTION

#### 1.1 Background

Cellulose mainly is produced by plant as one of the core component of cell walls (Somerville, 2006). However, to obtain cellulose from plants, tree and woods are sacrificed and it costs adverse impact to the environment. Hestrin & Schramm, (1954) had found an alternative way to obtained cellulose, *i.e.* which produced by *Acetobacter xylinum* and named it bacterial cellulose (BC). Since then, BC became an interesting biomaterial to be studied by researchers all over the world either to increase its production or to be used in a few applications as reviewed later.

Production of BC requires carbon source as the main nutrient to be used by *Acetobacter xylinum*. Initially, glucose was used as the main carbon source for BC production (Hestrin & Schramm, 1954). Later, researchers had shifted to other saccharides and alcohol (Masaoka *et al.*, 1993; Keshk & Sameshima, 2005). Abundant amount of waste biomass in Malaysia might be utilized as a new energy source for BC production. Malaysia has many natural resources available especially from forestry and agriculture sectors (Tock *et al.*, 2010). In addition, municipal solid waste also generated about 1.7

kg/person/day where about 60 % was contributed by food and organic waste (Kathirvale *et al.*, 2004). Those wastes have potentials to be converted into valuable material, e.g. bioethanol (Del Campo *et al.*, 2006), biofuel (Lenihan *et al.*, 2010) and new carbon source that can be used to produce BC. For example, in sago starch production, the factory generated approximately 1000 m<sup>3</sup> effluent per day in the process (Vikineswary *et al.*, 1994). Since the effluent contains sago pith and also starch, it has potential to be used as carbon source for BC production. Banana peel also can be used as carbon source in BC production since it is a lignocellulosic material and has high amount of starch. The peel can be purchased or obtained from various places like food processing plants, markets, food and beverages shops and etc (Tock *et al.*, 2010). In addition, potato peel can also be used as carbon sources to produce BC. Since Malaysia has a few products that are potato-based product, e.g. Twist Potato and Jacker, the peels supplies exist. The main hypothesis of this work is that those materials are expected to have potentials to become alternative carbon source due to they consist complex sugar that can be treated. Treatment of those materials is needed so that it is suitable and can be used by *Acetobacter xylinum* for BC production.

## 1.2 Problem statements

In order to obtain fermentable sugar by utilizing agricultural (food) wastes, hydrolysis of those wastes is needed. Diluted acid hydrolysis is widely used and preferable to treat wastes compared to enzymatic hydrolysis due to its low cost and effective process for energy production (Aguilar *et al.*, 2002; Sun and Cheng, 2002; Herrera *et al.*, 2004; Del Campo *et al.*, 2006; Lenihan *et al.*, 2010). The high cost of enzyme, the need for specialized labor and the requirement for sophisticated laboratories (Tasic *et al.*, 2009)

become a major problem to use enzymatic hydrolysis. Agro waste such as potato peel, banana peel and sago effluent which has high content of carbohydrate such as cellulose and starch that can be manipulated. The hydrolysis of wastes may give two benefits *i.e.* production of new alternative of fermentable sugar and at the same time elimination of the wastes.

Investigation of carbon source content to improve BC production was conducted by using a few type of biochemical (Keshk & Sameshima, 2005). The high cost of using commercial sugar, *i.e.* glucose, fructose, mannitol etc limit its industrial production. Discovery of new carbon source had been done by Hong and Qiu (2008) by preparing alternative carbon source from konjac powder to replace the commercial sugar. In addition, a few researchers had used fruit juices in the production of BC (Kongruang, 2008; Kurosumi *et al.*, 2009). Hence, hydrolysate from agro waste especially from sago effluent, banana peel and potato peel which is not available in literature is investigated in this study to produce BC.

### **1.3 Objectives**

The main objective of this study is to synthesize BC from alternative carbon source. To accomplish this objective, two key objectives were set, namely:

- i. To investigate the reducing sugar yield obtained from the diluted acid hydrolysis of agricultural (food) waste.
- ii. To synthesize and characterize BC using hydrolysate obtained in (i) as the carbon source.

## **1.4 Scope of work**

The scope of this work can be divided into three main sections:

- i. Hydrolyze and characterize the agricultural (food) wastes that will be used as carbon source
- ii. Synthesize the BC using the selected carbon source
- iii. Analyze BC and media of fermentation

## **1.5 Thesis Outline**

The thesis consists of five chapters. Chapter 1 covers the background, the problem statement and the research objectives of the study.

The literature review is covered in Chapter 2 where the basis to the production of fermentable sugar and BC production are elucidated. Published works are also described, summarized and discussed.

Chapter 3 briefly describes the research approach of this work. The methodology is explained in detail. Here, the method used to convert waste into fermentable sugar and production of BC from the selected alternative carbon source is presented.

In Chapter 4, the results and discussions were explained in detail. Findings for first and second objective were discussed.

Chapter 5 concludes the major finding of this work and provides recommendation for future work.

## REFERENCES

- Abd-Aziz, S. (2002). Sago starch and its utilisation. *Journal of Bioscience and Bioengineering*. 94(6): 526-529.
- Adisa, V., Okey, E. (1987). Carbohydrate and protein composition of banana pulp and peel as influenced by ripening and mold contamination. *Food Chemistry*. 25(2): 85-91.
- Aguilar, R., Ramirez, J., Garrote, G., Vazquez, M. (2002). Kinetic study of the acid hydrolysis of sugar cane bagasse. *Journal of Food Engineering*. 55(4): 309-318.
- Anhwange, B., Ugye, T., Nyiaatagher, T. (2008). Chemical composition of *Musa sapientum* (banana) peels. *Journal of Food Technology*. 6(6): 263-266.
- Asia Research News. (2008). Retrieved from <http://www.researchsea.com/html/article.php/aid/3069/eid/2>: 15 April 2011
- Awg-Adeni, D.S., Abd-Aziz, S., Bujang, K. and Hassan, M.A. (2010). Bioconversion of sago residue into value added products. *African Journal of Biotechnology*. 9(14): 2016-2051.
- Bae, S., Shoda, M. (2005). Production of bacterial cellulose by *Acetobacter xylinum* BPR2001 using molasses medium in a jar fermentor. *Applied Microbiology and Biotechnology*. 67(1): 45-51.
- Bae, S., Shoda, M. (2005). Statistical optimization of culture conditions for bacterial cellulose production using Box Behnken design. *Biotechnology and Bioengineering*. 90(1): 20-28.
- Bae, S., Sugano, Y., Shoda, M. (2004). Improvement of bacterial cellulose production by addition of agar in a jar fermentor. *Journal of Bioscience and Bioengineering*. 97(1): 33-38.
- Bodin, A., Bäckdahl, H., Fink, H., Gustafsson, L., Risberg, B., Gatenholm, P. (2007). Influence of cultivation conditions on mechanical and morphological properties of bacterial cellulose tubes. *Biotechnology and Bioengineering*. 97(2): 425-434.
- Bottom, R. (2008). Principles and applications of thermal analysis, UK. Blackwell Publishing.
- Bower, S., Wickramasinghe, R., Nagle, N.J., Schell, D.J. (2008). Modeling sucrose hydrolysis in dilute sulfuric acid solutions at pretreatment conditions for lignocellulosic biomass. *Bioresource Technology*. 99(15): 7354-7362.

- Brown, A.J. (1886). The chemical action of pure cultivation of *Bacterium aceti*. *J. Chem. Soc.* 49(432-439).
- Budhiono, A., Rosidi, B., Taher, H., Iguchi, M. (1999). Kinetic aspects of bacterial cellulose formation in nata-de-coco culture system. *Carbohydrate Polymers*. 40(2): 137-143.
- Bungay, H.R., Serafica, G., Mormino, R. (1997). Environmental implications of microbial cellulose. *Studies in Environmental Science*. 66: 691-700.
- Camire, M.E., Violette, D., Dougherty, M.P., McLaughlin, M.A. (1997). Potato peel dietary fiber composition: effects of peeling and extrusion cooking processes. *Journal of Agricultural and Food Chemistry*. 45(4): 1404-1408.
- Cannon, R.E., Anderson, S.M. (1991). Biogenesis of bacterial cellulose. *Critical Reviews in Microbiology*. 17(6): 435-447.
- Chawla, P.R., Bajaj, I.B., Survase, S.A., Singhal, R.S. (2009). Microbial cellulose: fermentative production and applications. *Food Technology and Biotechnology*. 47(2): 107-124.
- Chen, S., Zou, Y., Yan, Z., Shen, W., Shi, S., Zhang, X., Wang, H. (2009). Carboxymethylated-bacterial cellulose for copper and lead ion removal. *Journal of Hazardous Materials*. 161(2-3): 1355-1359.
- Chen, P., Kim, H-S., Kwon, S-M., Yun, Y.S. and Jin, H-J. (2009). Regenerated bacterial cellulose/multi-walled carbon nanotubes composite fibers prepared by wet-spinning. *Current Applied Physics* 9: 96-99.
- Cheng, H.P., Wang, P.M., Chen, J.W., Wu, W.T. (2002). Cultivation of *Acetobacter xylinum* for bacterial cellulose production in a modified airlift reactor. *Biotechnology and Applied Biochemistry*. 35(2): 125-132.
- Cook, K.E., Colvin, J.R. (1980). Evidence for a beneficial influence of cellulose production on growth of *Acetobacter xylinum* in liquid medium. *Current Microbiology*. 3(4): 203-205.
- Czaja, W., Krystynowicz, A., Bielecki, S., Brown, R.M. (2006). Microbial cellulose--the natural power to heal wounds. *Biomaterials*. 27(2): 145-151.
- Del Campo, I., Alegría, I., Zazpe, M., Echeverría, M., Echeverría, I. (2006). Diluted acid hydrolysis pretreatment of agri-food wastes for bioethanol production. *Industrial Crops and Products*. 24(3): 214-221.
- Delgado, R., Castro, A., Vázquez, M. (2009). A kinetic assessment of the enzymatic hydrolysis of potato (*Solanum tuberosum*). *LWT-Food Science and Technology*. 42(4): 797-804.

- Dobre, L.M., Stoica, A., Stroescu, M., S., J., Jipa, I., Dobre, T. (2010). Characterization of composite materials based on biocellulose membranes impregnated with silver particles as antimicrobial agent. *U.P.B. Sci. Bull., Series B.* 72(4): 55-64.
- Embuscado, M.E., Marks, J.S., BeMiller, J.N. (1994). Bacterial cellulose. II. Optimization of cellulose production by *Acetobacter xylinum* through response surface methodology. *Food Hydrocolloids.* 8(5): 419-430.
- Evans, B.R., O'Neill, H.M., Malyvanh, V.P., Lee, I., Woodward, J. (2003). Palladium-bacterial cellulose membranes for fuel cells. *Biosensors and Bioelectronics.* 18(7): 917-923.
- Fontana, J., De Souza, A., Fontana, C., Torriani, I., Moreschi, J., Gallotti, B., De Souza, S., Narciso, G., Bichara, J., Farah, L.F.X. (1990). Acetobacter cellulose pellicle as a temporary skin substitute. *Applied Biochemistry and Biotechnology.* 24(1): 253-264.
- Gamez, S., Gonzalez-Cabriales, J.J., Ramirez, J.A., Garrote, G., Vazquez, M. (2006). Study of the hydrolysis of sugar cane bagasse using phosphoric acid. *Journal of Food Engineering.* 74(1): 78-88.
- George, J., Sajeevkumar, V.A., Kumar, R., Ramana, K.V., Sabapathy, S.N. and Bawa, S. (2008). Enhancement of thermal stability associated with the chemical treatment of bacterial (*Gluconacetobacter xylinus*) cellulose. *Journal of Applied Polymer Science.* 108: 1845–1851.
- Global Environmental Centre. (2011). Retrieved from <http://www.geenet.info/index.cfm?&menuid=83&parentid=30> : 24 March 2011.
- Goelzer, F.D.E., Faria-Tischer, P.C.S., Vitorino, J.C., Sierakowski, M.R. and Tischer, C.A. (2009). Production and characterization of nanospheres of bacterial cellulose from *Acetobacter xylinum* from process rice bark. *Material Science and Engineering C29.* 546-551.
- Gorinstein, S., Oates, C., Chang, S. (1994). Enzymatic hydrolysis of sago starch. *Food Chemistry.* 49(4): 411-417.
- Hamelinck, C.N., Hooijdonk, G., and Faaij, A.P.C. (2005). Ethanol from lignocellulosic biomass: techno-economic performance in short-, middle- and long-term. *Biomass and Bioenergy.* 28: 384–410
- Happi Emaga, T., Andrianaivo, R.H., Wathélet, B., Tchango, J.T., Paquot, M. (2007). Effects of the stage of maturation and varieties on the chemical composition of banana and plantain peels. *Food Chemistry.* 103(2): 590-600.
- Henriksson, M. Berglund, L.A., Isaksson, P., Lindstrom T., and Nishino, T. (2008). Cellulose nanopaper structures of high toughness. *Biomacromolecules* 9:1579–1585

- Herrera, A., Tellez-Luis, S.J., Gonzalez-Cabriales, J.J., Ramirez, J.A., Vazquez, M. (2004). Effect of the hydrochloric acid concentration on the hydrolysis of sorghum straw at atmospheric pressure. *Journal of Food Engineering*. 63(1): 103-109.
- Hesse, S., Kondo, T. (2005). Behavior of cellulose production of *Acetobacter xylinum* in <sup>13</sup>C-enriched cultivation media including movements on nematic ordered cellulose templates. *Carbohydrate Polymers*. 60(4): 457-465.
- Hestrin, S., Schramm, M. (1954). Synthesis of cellulose by *Acetobacter xylinum*. 2. Preparation of freeze-dried cells capable of polymerizing glucose to cellulose. *Biochemical Journal*. 58(2): 345.
- Hidalgo, C., Vegas, C., Mateo, E., Tesfaye, W., Cerezo, A., Callejón, R., Poblet, M., Guillamón, J., Mas, A., Torija, M. (2010). Effect of barrel design and the inoculation of *Acetobacter pasteurianus* in wine vinegar production. *International Journal of Food Microbiology*. 141(1-2): 56-62.
- Hoffman, R.V. (2004). *Organic Chemistry: An Intermediate Text*. 2nd ed. New Jersey: John Wiley & Sons Inc..
- Hong, F., Qiu, K. (2008). An alternative carbon source from konjac powder for enhancing production of bacterial cellulose in static cultures by a model strain *Acetobacter aceti* subsp. *xylinus* ATCC 23770. *Carbohydrate Polymers*. 72(3): 545-549.
- Hwang, J.W., Yang, Y.K., Hwang, J.K., Pyun, Y.R., Kim, Y.S. (1999). Effects of pH and dissolved oxygen on cellulose production by *Acetobacter xylinum* BRC5 in agitated culture. *Journal of Bioscience and Bioengineering*. 88(2): 183-188.
- Iguchi, M., Yamanaka, S., Budhiono, A. (2000). Bacterial cellulose - a masterpiece of nature's arts. *Journal of Materials Science*. 35(2): 261-270.
- Ishihara, M., Matsunaga, M., Hayashi, N., Tiler, V. (2002). Utilization of xylose as carbon source for production of bacterial cellulose. *Enzyme and Microbial Technology*. 31(7): 986-991.
- Jonas, R., Farah, L.F. (1998). Production and application of microbial cellulose. *Polymer Degradation and Stability*. 59(1-3): 101-106.
- Kapdan, I.K., Kargi, F. (2006). Bio-hydrogen production from waste materials. *Enzyme and Microbial Technology*. 38(5): 569-582.
- Karimi, K., Kheradmandinia, S., Taherzadeh, M.J. (2006). Conversion of rice straw to sugars by dilute-acid hydrolysis. *Biomass and Bioenergy*. 30(3): 247-253.

- Karthikeyan, A., Sivakumar, N. (2010). Citric acid production by Koji fermentation using banana peel as a novel substrate. *Bioresource Technology*. 101(14): 5552-5556.
- Kashima, Y., Iijima, M., Nakano, T., Tayama, K., Koizumi, Y., Ueda, S., Yanagida, F. (2000). Role of intracellular esterases in the production of esters by *Acetobacter pasteurianus*. *Journal of Bioscience and Bioengineering*. 89(1): 81-83.
- Kathiravale, S., Muhd Yunus, M. (2008). Waste to wealth. *Asia Europe Journal*. 6(2): 359-371.
- Kathirvale, S., Muhd Yunus, M.N., Sopian, K., Samsuddin, A.H. (2004). Energy potential from municipal solid waste in Malaysia. *Renewable Energy*. 29(4): 559-567.
- Keshk, S., Sameshima, K. (2005). Evaluation of different carbon sources for bacterial cellulose production. *African Journal of Biotechnology*. 4(6): 478-482.
- Keshk, S., Sameshima, K. (2006). Influence of lignosulfonate on crystal structure and productivity of bacterial cellulose in a static culture. *Enzyme and Microbial Technology*. 40: 4-8
- Kilzer, F.J. & Broido, A. (1965). Speculations on the nature of cellulose pyrolysis. *Pyrolydynamics* 2: 151-163.
- Kim, J., Cai, Z., Chen, Y. (2010). Biocompatible bacterial cellulose composites for biomedical application. *Journal of Nanotechnology in Engineering and Medicine*. 1(1): 01006-7.
- Kim, K.I., Kim, W.K., Seo, D.K., Yoo, I.S., Kim, E.K., Yoon, H.H. (2003). Production of lactic acid from food wastes. *Applied Biochemistry and Biotechnology*. 107(1): 637-647.
- Klemm, D., Schumann, D., Udhardt, U., Marsch, S. (2001). Bacterial synthesized cellulose--artificial blood vessels for microsurgery. *Progress in Polymer Science*. 26(9): 1561-1603.
- Kongruang, S. (2008). Bacterial cellulose production by *Acetobacter xylinum* strains from agricultural waste products. *Applied Biochemistry and Biotechnology*. 148(1): 245-256.
- Kouda, T., Naritomi, T., Yano, H., Yoshinaga, F. (1997). Effects of oxygen and carbon dioxide pressures on bacterial cellulose production by Acetobacter in aerated and agitated culture. *Journal of Fermentation and Bioengineering*. 84(2): 124-127.
- Krishna, C. (1999). Production of bacterial cellulases by solid state bioprocessing of banana wastes. *Bioresource Technology*. 69(3): 231-239.

- Krystynowicz, A., Bielecki, S., Czaja, W., Rzyska, M. (2000). Application of bacterial cellulose for clarification of fruit juices. *Progress in Biotechnology*. 17: 323-327.
- Kuan-Chen, C., Ali, D., Jeff M, C. (2009). Enhanced production of bacterial cellulose by using a biofilm reactor and its material property analysis. *Journal of Biological Engineering*. 3(12).
- Kumoro, A., Ngoh, G., Hasan, M., Ong, C., Teoh, E. (2008). Conversion of fibrous sago (*Metroxylon sagu*) waste into fermentable sugar via acid and enzymatic hydrolysis. *Asian Journal of Scientific Research*. 1(4): 412-420.
- Kurosumi, A., Sasaki, C., Yamashita, Y., Nakamura, Y. (2009). Utilization of various fruit juices as carbon source for production of bacterial cellulose by *Acetobacter xylinum* NBRC 13693. *Carbohydrate Polymers*. 76(2): 333-335..
- Lee, H.C., Zhao, X. (1999). Effects of mixing conditions on the production of microbial cellulose by *Acetobacter xylinum*. *Biotechnology and Bioprocess Engineering*. 4(1): 41-45.
- Lenihan, P., Orozco, A., O'Neill, E., Ahmad, M., Rooney, D., Walker, G. (2010). Dilute acid hydrolysis of lignocellulosic biomass. *Chemical Engineering Journal*. 156(2): 395-403.
- Liimatainen, H., Kuokkanen, T., Kaariainen, J., 2004. Development of bio-ethanol production from waste potatoes. *Proceedings of the Waste Minimization and Resources Use Optimization Conference*, Oulu University Press, University of Oulu, Finland. 123-129.
- Marlida, Y., Saari, N., Karim, R., Mohamed, S., Karim, A. (2007). Improvement of Glucose Production by Raw Starch Degrading Enzyme Utilizing Acid-Treated Sago Starch as Substrate. *ASEAN Food Journal*. 14(2): 83-90.
- Masaoka, S., Ohe, T., Sakota, N. (1993). Production of cellulose from glucose by *Acetobacter xylinum*. *Journal of Fermentation and Bioengineering*. 75(1): 18-22.
- Memon, J.R., Memon, S.Q., Bhanger, M.I., Khuhawar, M.Y. (2008). Banana peel: a green and economical sorbent for Cr (III) removal. *Pak. J. Anal. Environ. Chem.* 9(1): 20-25.
- Mikkelsen, D., Flanagan, B., Dykes, G., Gidley, M. (2009). Influence of different carbon sources on bacterial cellulose production by *Gluconacetobacter xylinus* strain ATCC 53524. *Journal of Applied Microbiology*. 107(2): 576-583.
- Mosier, N., Wyman, C., dale, B., Elander, R., Lee, Y.Y., Holtzapple, M. and Landisch, M. (2005) Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technology* 96(6): 673-686.

- Mosier, N., Hendrickson, R., Ho, N., Sedlak, M., and Ladisch, M. R. (2005) Optimization of pH controlled liquid hot water pretreatment of corn stover. *Bioresource Technology*. 96: 1986–1993.
- Neureiter, M., Danner, H., Thomasser, C., Saidi, B., Braun, R. (2002). Dilute-acid hydrolysis of sugarcane bagasse at varying conditions. *Applied Biochemistry and Biotechnology*. 98(1): 49-58.
- Ochaikul, D., Chotirittikrai, K., Chantra, J., Wutigornsombatkul, S. (2006). Studies on fermentation of *Monascus purpureus* TISTR 3090 with bacterial cellulose from *Acetobacter xylinum* TISTR 967. *KMITL Sci Technol J.* 6: 13-17.
- Okeke, B.C., Frankenberger, W.T. (2005). Use of starch and potato peel waste for perchlorate bioreduction in water. *Science of the Total Environment*. 347(1-3): 35-45.
- Okiyama, A., Motoki, M., Yamanaka, S. (1992). Bacterial cellulose II. Processing of the gelatinous cellulose for food materials. *Food Hydrocolloids*. 6(5): 479-487.
- Okuda, N., Soneura, M., Ninomiya, K., Katakura, Y., Shioya, S. (2008). Biological detoxification of waste house wood hydrolysate using *Ureibacillus thermosphaericus* for bioethanol production. *Journal of Bioscience and Bioengineering*. 106(2): 128-133.
- Oshima, T., Kondo, K., Ohto, K., Inoue, K., Baba, Y. (2008). Preparation of phosphorylated bacterial cellulose as an adsorbent for metal ions. *Reactive and Functional Polymers*. 68(1): 376-383.
- Pertile, R.A.N., Andrade, F.K., Alves Jr, C., Gama, M. (2010). Surface modification of bacterial cellulose by nitrogen-containing plasma for improved interaction with cells. *Carbohydrate polymers*. 82(3): 692-698.
- Perunding Good Earth Sdn Bhd, (2011). E-Waste Inventory Project in Malaysia
- Phisalaphong, M., Jatupaiboon, N. (2008). Biosynthesis and characterization of bacteria cellulose-chitosan film. *Carbohydrate Polymers*. 74(3): 482-488.
- Plazl, I., Leskovsek, S. and Koloini, T. (1995). Hydrolysis of sucrose by conventional and microwave heating in stirred tank reactor. *The Chemical Engineering Journal and the Biochemical Engineering Journal*. 59(3): 253-257.
- Pourramezan, G., Roayaei, A., Qezelbash, Q. (2009). Optimization of Culture Conditions for Bacterial Cellulose Production by *Acetobacter* sp. 4B-2. *Biotechnology*. 8(1): 150-154.
- Rahman, S., Choudhury, J., Ahmad, A., Kamaruddin, A. (2007). Optimization studies on acid hydrolysis of oil palm empty fruit bunch fiber for production of xylose. *Bioresource Technology*. 98(3): 554-559.

- Rambo, C., Recouvreux, D., Carminatti, C., Pitlovanciv, A., Antônio, R., Porto, L. (2008). Template assisted synthesis of porous nanofibrous cellulose membranes for tissue engineering. *Materials Science and Engineering: C*. 28(4): 549-554.
- Rodriguez-Chong, A., Alberto Ramirez, J., Garrote, G., Vazquez, M. (2004). Hydrolysis of sugar cane bagasse using nitric acid: a kinetic assessment. *Journal of Food Engineering*. 61(2): 143-152.
- Rouessac, F. and Rouessac, A. *Chemical Analysis: Modern Instrumentation Methods and Techniques*. 2<sup>nd</sup> ed. England: John Wiley & Sons Ltd., 2007.
- Saxena, I.M., Brown Jr, R. (2001). Biosynthesis of cellulose. *Progress in Biotechnology*. 18: 69-76.
- Scionti, G. (2010). Mechanical properties of bacterial cellulose implants. Published Master Thesis. Chalmers University of Technology, Sweden.
- Sharma, N., Kalra, K., Oberoi, H.S., Bansal, S. (2007). Optimization of fermentation parameters for production of ethanol from kinnow waste and banana peels by simultaneous saccharification and fermentation. *Indian Journal of Microbiology*. 47(4): 310-316.
- Shen, W., Chen, S., Shi, S., Li, X., Zhang, X., Hu, W., Wang, H. (2009). Adsorption of Cu (II) and Pb (II) onto diethylenetriamine-bacterial cellulose. *Carbohydrate Polymers*. 75(1): 110-114.
- Shukla, J., Kar, R. (2006). Potato peel as a solid state substrate for thermostable - amylase production by thermophilic *Bacillus* isolates. *World Journal of Microbiology and Biotechnology*. 22(5): 417-422.
- Shuler, M.L. and Kargi, F. (2002). *Bioprocess engineering basic concept*. USA, Prentice Hall.
- Singhal, R.S., Kennedy, J.F., Gopalakrishnan, S.M., Kaczmarek, A., Knill, C.J., Akmar, P.F. (2008). Industrial production, processing, and utilization of sago palm-derived products. *Carbohydrate Polymers*. 72(1): 1-20.
- Skinner, P.O.N., Cannon, R.E. (2000). Acetobacter xylinum: An inquiry into cellulose biosynthesis. *The American Biology Teacher*. 442-444.
- Smith, F., Srivastava, H. (1959). Constitutional studies on the glucomannan of konjak Flour. *Journal of the American Chemical Society*. 81(7): 1715-1718.
- Sokolnicki, A.M., Fisher, R.J., Harrah, T.P., Kaplan, D.I.. (2006). Permeability of bacterial cellulose membranes. *Journal of Membrane Science*. 272(1-2): 15-27.
- Somerville, C. (2006). Cellulose synthesis in higher plants. *Annual Review of Cell and Developmental Biology*. 22: 53-78.

- Son, H.J., Kim, H.G., Kim, K.K., Kim, H.S., Kim, Y.G., Lee, S.J. (2003). Increased production of bacterial cellulose by *Acetobacter* sp. V6 in synthetic media under shaking culture conditions. *Bioresource Technology*. 86(3): 215-219.
- Spiridon, I., Teaca, C.A., Bodirliu, R. (2011). Structural changes evidenced by FTIR spectroscopy in cellulose materials after pre-treatment with ionic liquid and enzymatic hydrolysis. *BioResources*. 6(1): 400-413.
- Sun, D., Yang, J., Wang, X. (2009). Bacterial cellulose/TiO<sub>2</sub> hybrid nanofibers prepared by the surface hydrolysis method with molecular precision. *Nanoscale*. 2(2): 287-292.
- Sun, Y., Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review\*. *1. Bioresource Technology*. 83(1): 1-11.
- Surma-Slusarska, B., Danielewicz, D., Presler, S. (2008). Characteristics of bacterial cellulose obtained from *Acetobacter xylinum* culture for application in papermaking. *Fibres & Textiles in Eastern Europe*. 16(4): 69.
- Suwannapinunt, N., Burakorn, J., Thaenthanee, S. (2007). Effect of culture conditions on bacterial cellulose (BC) production from acetobacter xylinum Tistr976 and physical properties of BC parchment paper. *Suranaree J. Sci. Technol.* 14(4): 357-365.
- Tasic, M.B., Konstantinovic, B.V., Lazic, M.L., Veljkovic, V.B. (2009). The acid hydrolysis of potato tuber mash in bioethanol production. *Biochemical Engineering Journal*. 43(2): 208-211.
- Tewari, H., Marwaha, S., Rupal, K. (1986). Ethanol from banana peels. *Agricultural wastes*. 16(2): 135-146.
- Thompson, N.S., Carlson, J.A., Kaustinen, H.M. and Uhlin, K.I. (1988). Tunnel structure in *Acetobacter xylinum*. *Int. J. Biol. Macromol.* 10: 126-127.
- Tomita, Y. and Kondo, T. (2009). Influential factors to enhance the moving rate of *Acetobacter xylinum* due to its nanofiber secretion on oriented templates. *Carbohydrate Polymers*. 77: 754-759.
- Tock, J.Y., Lai, C.L., Lee, K.T., Tan, K.T., Bhatia, S. (2010). Banana biomass as potential renewable energy resource: A Malaysian case study. *Renewable and Sustainable Energy Reviews*. 14(2): 798-805.
- Vandamme, E., De Baets, S., Vanbaelen, A., Joris, K., De Wulf, P. (1998). Improved production of bacterial cellulose and its application potential. *Polymer Degradation and Stability*. 59(1-3): 93-99.
- Verschuren, P.G., Cardona, T.D., Nout, M., De Gooijer, K.D., Van den Heuvel, J.C. (2000). Location and limitation of cellulose production by *Acetobacter xylinum*

established from oxygen profiles. *Journal of Bioscience and Bioengineering*. 89(5): 414-419.

Vikineswary, S., Shim, Y., Thambirajah, J., Blakebrough, N. (1994). Possible microbial utilization of sago processing wastes. *Resources, Conservation and Recycling*. 11(1-4): 289-296.

Wang, W., Powell, A., Oates, C. (1996). Sago starch as a biomass source: Raw sago starch hydrolysis by commercial enzymes. *Bioresource Technology*. 55(1): 55-61.

Williams, W.S., Cannon, R.E. (1989). Alternative environmental roles for cellulose produced by *Acetobacter xylinum*. *Applied and Environmental Microbiology*. 55(10): 2448.

Wu, S.C., Lia, Y.K. (2008). Application of bacterial cellulose pellets in enzyme immobilization. *Journal of Molecular Catalysis B: Enzymatic*. 54(3-4): 103-108.

Xiang, Q., Lee, Y.Y. and Torget R.W. (2004). Kinetics of glucose decomposition during dilute-acid hydrolysis of lignocellulosic biomass. *Applied Biochemistry and Biotechnology*. 113–116.

Yamanaka, S., Watanabe, K., and Kitamura, N. (1989). The structure and mechanical properties of sheets prepared from bacterial cellulose. *Journal of Material Science*. 24: 3141-3145.

Yan, S., Li, J., Chen, X., Wu, J., Wang, P., Ye, J., Yao, J. (2011). Enzymatical hydrolysis of food waste and ethanol production from the hydrolysate. *Renewable Energy*. 36(4): 1259-1265.

Yan, Z., Chen, S., Wang, H., Wang, B., Wang, C., Jiang, J. (2008). Cellulose synthesized by *Acetobacter xylinum* in the presence of multi-walled carbon nanotubes. *Carbohydrate Research*. 343(1): 73-80.

Yang, Y.K., Park, S.H., Hwang, J.W., Pyun, Y.R., Kim, Y.S. (1998). Cellulose production by *Acetobacter xylinum* BRC5 under agitated condition. *Journal of Fermentation and Bioengineering*. 85(3): 312-317.

Zhang, P., Whistler, R.L., BeMiller, J.N., Hamaker, B.R. (2005). Banana starch: production, physicochemical properties, and digestibility-a review. *Carbohydrate Polymers*. 59(4): 443-458.

Zheng, Y., Pan, Z., Zhang, R. (2009). Overview of biomass pretreatment for cellulosic ethanol production. *International Journal of Agricultural and Biological Engineering*. 2(3): 51-68.

Zhou, L., Sun, D., Hu, L., Li, Y., Yang, J. (2007). Effect of addition of sodium alginate on bacterial cellulose production by *Acetobacter xylinum*. *Journal of Industrial Microbiology and Biotechnology*. 34(7): 483-489.

