



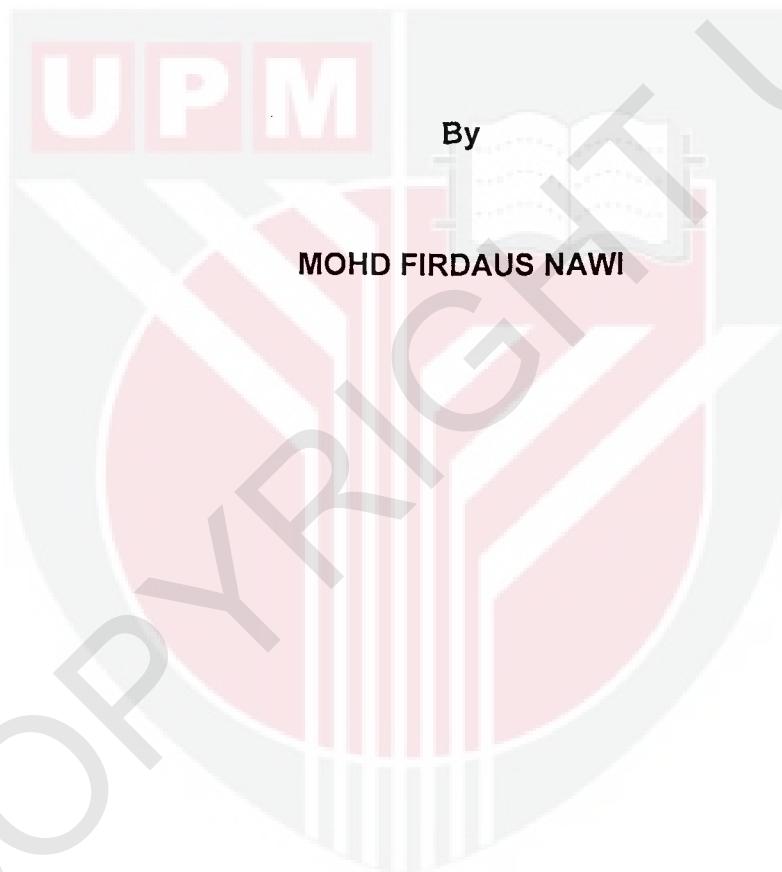
UNIVERSITI PUTRA MALAYSIA

***IMMUNE RESPONSES TO Streptococcus agalactiae IN RED TILAPIA,
Oreochromis spp. FOLLOWING VACCINATION WITH NON-
ADJUVANTED AND ADJUVANTED VACCINE INCORPORATED FEED
PELLETS***

MOHD FIRDAUS NAWI

FPV 2011 13

**IMMUNE RESPONSES TO *Streptococcus agalactiae* IN RED TILAPIA,
Oreochromis spp. FOLLOWING VACCINATION WITH NON-
ADJUVANTED AND ADJUVANTED VACCINE INCORPORATED FEED
PELLETS**



**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of Requirement for the Degree of Master of
Science**

August 2011

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Master of Science

**IMMUNE RESPONSES TO *Streptococcus agalactiae* IN RED TILAPIA,
Oreochromis spp. FOLLOWING VACCINATION WITH NON-
ADJUVANTED AND ADJUVANTED VACCINE INCORPORATED FEED
PELLETS**

By

MOHD FIRDAUS NAWI

August 2011

Chairman : Md. Sabri Mohd Yusoff, DVM, MSc, PhD
Faculty : Veterinary Medicine

Streptococcosis is an important bacterial disease in tilapia in many countries, including Malaysia. The two common *Streptococcus* species causing the disease are *S. agalactiae* and *S. iniae*. In Malaysia, outbreaks of streptococcosis in red tilapia (*Oreochromis* spp.) were due to *Streptococcus agalactiae* while infection by *Streptococcus iniae* has never been reported. Mass mortalities among red tilapia in cage-culture system were reported to occur usually during the dry months of the year, between April and August when water temperature was high.

This study was conducted to investigate the effect of high water temperature on the susceptibility of red tilapia to infection by *Streptococcus agalactiae*, the immune response by red tilapia following exposure to live *S. agalactiae* and the protection following vaccination against streptococcosis. To achieve the first objective, eight groups of red tilapias of approximately 100 g were infected intraperitoneally with 0.5 mL of live *S. agalactiae* at 6.3×10^6 CFU/mL, 6.3×10^7 CFU/mL, 6.3×10^8 CFU/mL and 6.3×10^9 CFU/mL. Four groups were kept at normal water temperature of $27^\circ\text{C} \pm 2^\circ\text{C}$ while the rest were kept in high water temperature of $33^\circ\text{C} \pm 2^\circ\text{C}$ for a period of one week. The high water temperature caused an increased in the susceptibility of red tilapia to *S. agalactiae* infection as indicated by the rapid rate of mortality at lethal dose 50 (LD₅₀). The period to achieve 100% mortality in the high temperature group was faster than the normal water temperature group, in four days compared to seven days. The lethal dose 50 (LD₅₀) for groups that were kept in high water temperature was 5.68×10^6 CFU/mL, significantly ($p<0.05$) lower than those that were kept in normal water temperature (2.29×10^7 CFU/mL). The clinical signs included loss of appetite, lethargy, unilateral or bilateral exophthalmia, cloudy eyes, erratic swimming and inflammation of the skin.

Once the susceptibility was determined, formalin killed whole-cell *S. agalactiae* at the concentration of 6.7×10^6 CFU/mL was incorporated homogenously into fish pellet as feed vaccine (FNV vaccine). The vaccine was then administered orally to the red tilapia in three different vaccination regimes; once (F1D), thrice (F3D) and five times a week (F5D). Body mucus and blood serum were sampled every week for eight weeks to analyze the

mucosal and systemic antibody responses by using ELISA. Immunization by FNV vaccine resulted in significant ($p<0.05$) increase in the serum and mucus antibody levels (IgM) as early as week 2 in all vaccinated groups, while the control unvaccinated group (FC) showed insignificant ($p>0.05$) increase of the serum and mucus antibody levels. Group F5D showed the highest antibody levels, followed by groups F3D and F1D. At the end of the experiment, twenty fishes from each group were challenged by immersion while another twenty were challenged intra-peritoneal. Survival rate was low indicating poor protective immune response in all groups of tilapia tested. Gut samples were obtained at the end of the experiment and subjected to histological analyses to examine the presence of gut-associated lymphoid tissue (GALT). According to the analyses, exposure at the rate of once a week to FNV vaccine was sufficient to stimulate GALT and skin mucus antibody responses. However, more frequent exposures stimulated better responses by GALT as observed in red tilapias of groups F3D and F5D. As expected, unvaccinated red tilapias failed to stimulate any GALT development. In conclusion, vaccination using FNV vaccine stimulated mucosal and systemic immunities but the protection provided was unsatisfactory.

Following the episode of poor protection provided by oral administration of FNV vaccine, the vaccine was further modified by adding Freund's Incomplete Adjuvant (FIA) into the vaccine, a known potent immune response enhancer at both humoral and cellular levels. The feed adjuvanted vaccine (FAV) was found to improve the mucosal immune response and elicited excellent systemic immune response. Apart from higher body mucus,

both gut lavage fluid and blood serum antibody titers were also higher than the FNV vaccine. The FAV vaccine also provided 100% protection following challenged with 3.4×10^9 CFU/mL of live *S. agalactiae*, significantly ($p \geq 0.05$) higher than protection provided by FNV vaccine. Similarly, the size of GALT and the number of lymphocytes in the FAV-vaccinated group were significantly ($p < 0.05$) greater compared to the FNV-vaccinated group.

In conclusion, this study demonstrated that adjuvanted vaccine (FAV) was more effective compared to the non-adjuvanted vaccine (FNV). The FAV vaccine effectively stimulated both mucosal and systemic immunity and enhanced protection against *S. agalactiae* infection in red tilapia. Therefore, FAV vaccine is the best candidate for control of streptococcosis in red tilapia.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan ijazah Master Sains

**TINDAKBALAS IMUN TERHADAP *Streptococcus agalactiae* DIDALAM
IKAN TILAPIA MERAH, *Oreochromis spp.* SETELAH DIVAKSINASI
DENGAN PELLET BERCOMPUR VAKSIN TANPA ADJUVANT (FNV) DAN
PELLET BERCOMPUR VAKSIN DENGAN ADJUVANT**

Oleh

MOHD FIRDAUS NAWI

Ogos 2011

Pengerusi : Md. Sabri Mohd Yusoff, DVM, MSc, PhD

Fakulti : Perubatan Veterinar

Streptococcosis merupakan penyakit bakteria yang utama pada ikan tilapia di seluruh dunia, termasuklah Malaysia. Dua jenis spesis Streptococcus yang biasa menyebabkan penyakit streptococcosis adalah *Streptococcus agalactiae* and *Streptococcus iniae*. Di Malaysia, wabak streptococcosis di kalangan ikan tilapia merah (*Oreochromis spp*) telah di kenal pasti berpunca dari jangkitan *S. agalactiae* manakala jangkitan oleh *S. iniae* tidak pernah dilaporkan. Kematian secara besar-besaran ikan tilapia merah yang diternak dalam sistem sangkar biasanya dilaporkan berlaku semasa musim kemarau di antara bulan April hingga Ogos dimana suhu air adalah sangat tinggi pada waktu itu.

Kajian telah dilakukan untuk mengenalpasti kesan suhu air yang tinggi terhadap kerentanan ikan tilapia merah kepada jangkitan *S. agalactiae*, tindakblas imun tilapia merah setelah didedahkan kepada *S. agalactiae* hidup dan perlindungan yang diberi apabila dicabar setelah diberi vaksinasi untuk melawan streptococcosis. Untuk mencapai objektif pertama, lapan kumpulan ikan tilapia seberat kira-kira 100 g telah dijangkitkan melalui suntikan intraperitonium dengan 0.5 mL *S. agalactiae* hidup yang mengandungi 6.3×10^6 CFU/mL, 6.3×10^7 CFU/mL, 6.3×10^8 CFU/mL dan 6.3×10^9 CFU/mL. Empat kumpulan ikan tilapia merah dibela pada suhu air biasa ($27^{\circ}\text{C} \pm 2^{\circ}\text{C}$) dan yang selebihnya pada suhu air yang tinggi ($33^{\circ}\text{C} \pm 2^{\circ}\text{C}$) selama seminggu. Suhu air yang tinggi telah menyebabkan peningkatan kerentanan ikan tilapia merah terhadap jangkitan *S. agalactiae* seperti mana ditunjukkan melalui keputusan kadar cepatnya kematian dan dos yang mematikan 50 (LD_{50}). Jangkamasa kadar kematian 100% untuk kumpulan ikan yang dibela pada suhu air yang tinggi adalah lebih cepat berbanding kumpulan ikan yang dibela pada suhu air biasa. Kadar dos yang mematikan 50 (LD_{50}) bagi kumpulan ikan yang dibela pada suhu air yang tinggi adalah 5.68×10^6 CFU/mL, secara signifikannya lebih rendah ($p<0.05$) berbanding kumpulan ikan yang dibela pada suhu air biasa, 2.29×10^7 CFU/mL. Tanda-tanda klinikal yang diamati sepanjang kajian perkembangan penyakit ini termasuk hilangnya selera makan, kelihatan lesu, unilateral dan bilateral exophthalmia, mata beselaput, berenang tak menentu dan inflamasi pada kulit. Tanda-tanda yang diamati sangat berguna untuk mengenalpasti wabak apabila ia berlaku.

Setelah kerentanan ditentukan, sel-sel bakteria *S. agalactiae* yang dimatikan dengan formalin pada kepekatan 6.7×10^6 CFU/mL dicampurkan sehingga rata bersama pelet makanan ikan tilapia merah (Vaksin FNV) untuk menentang streptococcosis. Vaksin FNV kemudiannya diberi makan kepada ikan tilapia merah dalam tiga rejim vaksinasi berlainan, sekali (F1D), tiga kali (F3D) dan lima kali dalam seminggu (F5D). Sampel lendir badan dan serum darah diambil setiap minggu sehingga minggu kelapan untuk menganalisis aras antibodi ikan tilapia merah selepas divaksinasi menggunakan teknik ELISA. Pemvaksinan dengan vaksin FNV pada ikan tilapia merah telah menghasilkan peningkatan yang signifikan ($p<0.05$) pada aras antibodi serum dan mukus seawal minggu 2 dalam kumpulan-kumpulan yang divaksinasi, sementara tiada peningkatan signifikan ($p>0.05$) dalam kumpulan kawalan (FC). Aras antibodi dalam kumpulan F5D merupakan yang tertinggi, diikuti oleh kumpulan F3D dan F1D. Pada penghujung eksperimen, 20 ekor ikan tilapia merah dari setiap kumpulan dicabar secara rendaman untuk menilai perlindungan yang diperolehi daripada keimunan mukosa, sementara 20 ekor ikan tilapia yang selebihnya dari setiap kumpulan dicabar secara suntikan intraperitonium untuk menilai perlindungan yang diperolehi daripada keimunan sistemik. Kadar survival adalah rendah dan ini menandakan perlindungan hasil daripada tindak balas keimunan yang lemah dalam kumpulan-kumpulan yang diuji. Sampel usus ikan tilapia merah diambil pada minggu terakhir eksperimen dan dianalisis secara histologi untuk mengesan kehadiran "Sel Limfoid Berkaitan Usus" atau "Gut-Associated Lymphoid Tissue" (GALT). Berdasarkan analisis tersebut, pendedahan pada vaksin untuk sekali sahaja sudah mencukupi

untuk merangsang penghasilan GALT dan tindakbalas antibodi mukus. Walau bagaimanapun, lebih kerap pendedahan yang diberi merangsang tindakbalas GALT yang lebih bagus seperti apa yang dapat dilihat dalam kumpulan F3D dan F5D. Seperti dijangka, ikan tilapia merah yang tidak divaksinasi gagal untuk menghasilkan tindak balas GALT. Kesimpulannya, pemvaksinan menggunakan vaksin FNV dapat merangsang tindak balas keimunan mukosa dan sistemik tetapi perlindungan yang dihasilkan adalah tidak memuaskan. Oleh yang demikian, pemvaksinan perlu dipertingkatkan lagi untuk mendapatkan perlindungan keimunan sistemik yang berkesan pada ikan tilapia merah untuk menentang jangkitan dari *S. agalactiae*.

Susulan episod perlidungan keimunan sistemik yang tidak memuaskan diperolehi dari vaksinasi oral FNV, vaksin tersebut telah diberi penambahbaikan dengan memasukkan Freund Incomplete Adjuvant (FIA), penggalak tindak balas imun yang kuat pada peringkat humoral dan selular ke dalam formulasi vaksin. Keputusan kajian menunjukkan vaksin yang ditambah dengan adjuvant (FAV vaksin) tersebut telah meningkatkan respon keimunan mukosa dan juga berjaya menghasilkan respon keimunan sistemik yang baik. Disamping memberi aras antibodi yang lebih tinggi didalam lendir badan, cairan usus dan serum darah, vaksin FAV juga memberikan 100% perlindungan setelah dicabar dengan 3.4×10^9 *S. agalactiae* hidup, dimana secara signifikan ($p<0.05$) lebih tinggi berbanding dengan perlindungan yang diberikan oleh vaksin FNV. Saiz GALT dan jumlah limfosit yang dikira dalam kumpulan ikan yang diberi vaksin FAV juga secara signifikannya ($p<0.05$) adalah lebih tinggi berbanding dengan kumpulan ikan yang diberikan vaksin FNV. Kesimpulannya, kajian ini menunjukkan vaksin FAV adalah lebih efektif

berbanding vaksin FNV. Vaksin FAV secara efektifnya merangsang kedua-dua sistem keimunan, mukosa dan sistemik, dan meningkatkan perlindungan dari jangkitan *S. agalactiae* pada ikan tilapia merah. Oleh yang demikian, vaksin FAV merupakan calon vaksin terbaik dalam mengawal streptococcosis pada ikan tilapia merah.



ACKNOWLEDGEMENTS

First and foremost praises to ALLAH S.W.T, THE MOST COMPASSIONATE AND MERCIFUL for giving me the strength and courage to complete this thesis.

I would like to express my sincere gratefulness and appreciation to my supervisor Dr. Md. Sabri Mohd Yusoff for his invaluable guidance, advice, constructive suggestion, tolerant supervision and support towards completion of this study.

I wish to express my gratitude to my co-supervisors, Prof. Dr. Mohd Zamri Saad and Dr. Siti Zahrah Abdullah for their ideas, advice, encouragement, unfailing help and offered insightful suggestion throughout the course of this study. Sincere thanks also for Zulkafli Rashid, Ramley Abu Bakar and Misri Samingin for their kindness in helping me in statistical analysis. Grateful wish also goes to Hanan Mohd Yusoff for his brilliant ideas, technical assistance and invaluable time spent.

I wish to thank all my former and current colleagues in Histopathology Lab, Faculty of Veterinary Medicine, University Putra Malaysia, Dr. Shafarin Shamsudin, Dr. Ina Salwany Mohd Yasin, Dr. Didik Handijatno, Dr. Sriyanto, Dr. Yulianna Puspitasari, Dr. Khin Myat Nwe, Dr. Trang, Dr. Hani Plumeriastuti, Dr. Sharom Salisi, Dr. Rafidah Othman, Dr. Abu Bakar Salisu, Dr. Nurrul Shaqinah Nasrudin, Dr. Noor Amal Azmai, Nur Nazifah Mansur, Saidatul Atyah Apendai, Noraini Isa, Illazuwa Mohd Yusoff, and Nur Hazwani

Oslan for their friendship and help. Sincere thank also goes to lab staffs, Jamilah Jahari, Latifah Hanan and Mohd Jamil Samad for their technical help throughout this study.

I would like to thank all National Fish Health Research Centre (NaFisH) members especially Shahidan Hashim, Wan Norazlan Ghazali, Aziel Sukiman, Fahmi Sudirwan, Faizul Helmi Hasmi and Norazian Rashid for their helpful hands during completing this study.

A special dedication to my wife, Mrs. Wan Nadilah Adibah Wan Ahmad, my son, Wan Amsyar Nufail Mohd Firdaus, my father, Nawi Abdullah, my mother Norjan Abdullah, my late brother, Mohd Faizul Nawi, my sisters, Nurul Hidayah Nawi, Nuradlin Syafini Nawi, Nuraimi Shahira Nawi and my father and mother in law, Wan Ahmad Wan Hussin and Wan Rahani Yunus and other family members, Wan Haspinah Wan Hussin, Wan Mohd Azam Datuk Wan Najib, Wan Nurkhaizan Wan Ahmad, Wan Amirul Amin Wan Ahmad, Wan Najwa Arifah Wan Ahmad, Wan Afifah Mardhiah Wan Ahmad and Wan Najiah Bahirah Wan Ahmad for their love, patience, understanding and encouragement throughout the course of my study.

Finally I would like to express my gratitude and appreciation to my best friends, Azrul Lokman, Che Ku Dahlia Che Ku Daud and Muhammed Irwan Mansoor for their support and purest friendship.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Md. Sabri bin Mohd Yusoff, DVM, MVSc, PhD

Senior Lecturer

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Chairman)

Mohd Zamri bin Saad, PhD

Professor

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Member)

Siti Zahrah binti Abdullah, M.Sc

Director

Fish Disease Division

Fisheries Research Institute

(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	vi
ACKNOWLEDGEMENT	x
APPROVAL	xii
DECLARATION	xiv
LIST OF TABLES	xx
LIST OF FIGURES	xxi
LIST OF ABBREVIATIONS	xxiii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 World Aquaculture	4
2.2 Aquaculture Industry in Malaysia	5
2.3 Status of Aquaculture Industry in Malaysia	6
2.4 Types of Aquaculture Management in Malaysia	8
2.5 Tilapia	9
2.6 Streptococcus	11
2.7 <i>Streptococcus agalactiae</i>	12
2.8 Streptococcosis	13
2.9 Predisposing Factors	16
2.10 Immune Response Against Streptococcosis	17
2.11 The Cell-Mediated Immunity	17
2.12 The Humoral Immunity	18
2.13 Vaccination	19

2.14	Fish Vaccination	21
2.14.1	Inject Vaccination	22
2.14.2	Immersion Vaccination	23
2.14.3	Oral Vaccination	24
3	PATHOGENICITY OF <i>Streptococcus agalactiae</i> IN RED TILAPIA (<i>Oreochromis</i> spp.) KEPT AT NORMAL AND HIGH WATER TEMPERATURE	
3.1	Introduction	26
3.2	Materials and Methods	
3.2.1	Pathogenicity of <i>Streptococcus agalactiae</i> in red tilapia kept in normal and high water temperature water.	27
3.2.1.1	Bacteria	27
3.2.1.2	Preparation of <i>Streptococcus agalactiae</i> Inoculum	28
3.2.1.3	Fish	28
3.2.1.4	Experimental Design	29
3.2.1.5	Sampling and Samples Processing	30
3.2.1.6	Statistical Analysis	31
3.3	Results	
3.3.1	Pathogenecity of <i>Streptococcus agalactiae</i> in red tilapia.	31
3.3.1.1	Mortality	31
3.3.1.2	Clinical Signs	36
3.3.1.3	Bacterial Isolation	38
3.4	Discussion	40
4	DEVELOPMENT OF AN ORAL DELIVERY FEED VACCINE AGAINST STREPTOCOCCOSIS IN RED TILAPIA	
4.1	Introduction	43

4.2	Materials and Methods	
4.2.1	Fish	45
4.2.2	Bacterial and Growth Condition	45
4.2.3	Formalin-Killed Bacteria (FKB) Preparation	45
4.2.4	Vaccine Incorporated Pellet (FNV) Preparation	46
4.2.5	Experimental Design	46
4.2.6	Preparation of <i>Streptococcus agalactiae</i> inoculum For Challenge.	47
4.2.7	Challenge Trial	48
4.2.8	Enzyme-Linked Immunosorbent Assay (ELISA)	49
4.2.9	Preparation of Gut Samples for Histological Analysis	51
4.2.10	Statistical Analysis	52
4.3	Results	
4.3.1	Antibody Response	53
4.3.1.1	Serum Antibody Response	53
4.3.1.2	Mucus Antibody Response	56
4.3.2	Challenge Trial	59
4.3.2.1	Antibody Level	59
4.3.2.2	Clinical Signs	59
4.3.2.3	Gross Findings	61
4.3.2.4	Bacterial Isolation	62
4.3.3	Histological Analysis	62
4.4	Discussion	66

5 PROTECTIVE CAPACITY OF THE ENHANCED FEED-BASED VACCINE WITH INCOMPLETE FREUND'S ADJUVANT AGAINST STREPTOCOCCOSIS IN RED TILAPIA

5.1	Introduction	70
5.2	Materials and Methods	

5.2.1	Fish	72
5.2.2	Bacterial and Growth Condition	73
5.2.3	Formalin-Killed Bacteria (FKB) Preparation	73
5.2.4	Vaccine Incorporated Pellet (FNV) Preparation	73
5.2.5	Adjuvant Vaccine Incorporated Pellet (FAV) Preparation	73
5.2.6	Experimental Design	74
5.2.7	Challenge Trial	74
5.2.8	Enzyme-Linked Immunosorbent Assay (ELISA)	75
5.2.9	Preparation of Gut Samples for Histological Analysis	77
5.2.10	Statistical Analysis	77
5.3	Results	
5.3.1	Antibody Response	77
5.3.1.1	Serum Antibody Response	77
5.3.1.2	Mucus Antibody Response	78
5.3.1.3	Gut Lavage Fluid Antibody Response	80
5.3.2	Challenge Trial	82
5.3.2.1	Antibody Level	82
5.3.2.2	Clinical Signs	82
5.3.2.3	Gross Findings	84
5.3.2.4	Bacterial Isolation	84
5.3.3	Histological Analysis	85
5.4	Discussion	90
GENERAL DISCUSSION		94
BIBLIOGRAPHY		101
APPENDICES		
Appendix A		111

Appendix B	114
Appendix C	115
BIODATA OF STUDENT	116



LIST OF TABLES

Table		Page
3.1	The inoculum dosages and treatments used in pathogenecity study.	30
3.2	The number and percentage of mortalities in red tilapia infected with various concentrations of live <i>S. agalactiae</i> kept in normal water temperature.	33
3.3	The number and percentage of mortalities in red tilapia infected with various concentrations of live <i>S. agalactiae</i> kept in high water temperature.	35
4.1	The treatments and the routes of challenged applied in oral vaccination study.	49
4.2	Average size of the GALT and the number of lymphoid cells in red tilapia following different frequency of oral exposures of FNV vaccine. Group F1D was orally vaccinated once in a week, Group F3D was vaccinated for three continuously days and Group F5D for five continuously days. Group FC remained control unexposed.	63
5.1	The number and percentage of survival in red tilapia after challenge with 3.4×10^9 live <i>S. agalactiae</i> by intraperitoneal injection.	83

LIST OF FIGURES

Figure		Page
3.1	Lethal dose 50 (Ld_{50}) for red tilapia infected with various concentrations of live <i>S. agalactiae</i> via i.p injection kept in normal water temperature is 2.2919×10^7 within 12h.	34
3.2	Lethal dose 50 (Ld_{50}) for red tilapia infected with various concentrations of live <i>S. agalactiae</i> via i.p injection kept in high water temperature is 5.6806×10^6 within 12h.	36
3.3	Unilateral exophthalmia and cloudiness of eye was observed in all inoculated fish after day-1 post-inoculation.	37
3.4	Reddish skin was observed at the operculum (yellow circle) of the inoculated fish after day-1 post-inoculation.	37
3.5	Percentage of <i>S. agalactiae</i> isolated from brain, eye and kidney of red tilapia infected with various bacterial concentrations kept in normal water temperature.	39
3.6	Percentage of <i>S. agalactiae</i> isolated from brain, eye and kidney of red tilapia infected with various bacterial concentrations kept in high water temperature.	40
4.1	Immunoglobulin M (IgM) level (ELISA OD) in serum of vaccinated (group F1D, F3D &F5D) and unvaccinated (group FC) adult tilapia monitored weekly before and after immunization. First immunization was conducted at week 1 and booster two weeks after first immunization followed by challenge with 3.4×10^9 cfu/mL ⁻¹ live <i>S. agalactiae</i> via immersion route for 24h at week 5.	54

4.2	Immunoglobulin M (IgM) level (ELISA OD) in serum of vaccinated (group F1D,F3D &F5D) and unvaccinated (group FC) adult tilapia monitored weekly before and after immunization. First immunization was conducted at week 1 and booster two weeks after first immunization followed by challenge with 3.4×10^9 cfu mL ⁻¹ live <i>S. agalactiae</i> via i.p injection route at week 5.	55
4.3	Immunoglobulin M (IgM) level (ELISA OD) in body mucus of vaccinated (group F1D,F3D &F5D) and unvaccinated (group FC) adult tilapia monitored weekly before and after immunization. First immunization was conducted at week 1 and booster two weeks after first immunization followed by challenge with 3.4×10^9 cfu mL ⁻¹ live <i>S. agalactiae</i> via immersion route for 24h at week 5.	57
4.4	Immunoglobulin M (IgM) level (ELISA OD) in body mucus of vaccinated (group F1D,F3D &F5D) and unvaccinated (group FC) adult tilapia monitored weekly before and after immunization. First immunization was conducted at week 1 and booster two weeks after first immunization followed by challenge with 3.4×10^9 cfu mL ⁻¹ live <i>S. agalactiae</i> via i.p injection at week 5.	58
4.5	The percentage of survival among groups of adult tilapia orally vaccinated and unvaccinated. All groups were challenged with 3.4×10^9 cfu mL ⁻¹ live <i>S. agalactiae</i> via immersion route for 24h.	60
4.6	The percentage of survival among groups of adult tilapia orally vaccinated and unvaccinated. All groups were challenged with 3.4×10^9 cfu mL ⁻¹ live <i>S. agalactiae</i> via i.p injection route.	61
4.7	Cross-section of the gut of red tilapia fed with FNV vaccine from Group F1D. Aggregation of lymphoid cells or GALT formed in the lamina propria as marked with yellow circle. HE x200.	64

4.8	Cross-section of the gut of red tilapia fed with FNV vaccine from Group F3D. Aggregation of lymphoid cells or GALT formed in the lamina propria as marked with yellow circle. HE x200.	64
4.9	Cross-section of the gut of red tilapia fed with FNV vaccine from Group F5D. Aggregation of lymphoid cells or GALT formed in the lamina propria as marked with yellow circle. HE x200.	65
4.10	Cross-section of the gut of fish fed with commercial feed from Group FC. No aggregation of lymphoid cells or GALT formed in the lamina propria. HE x200.	65
5.1	The serum IgM response following vaccination with vaccine incorporated pellet (FNV) and adjuvant vaccine incorporated pellet (FAV). The control group (FC) fishes were fed with commercial pellet.	78
5.2	The mucus IgM response following vaccination with vaccine incorporated pellet (FNV) and adjuvant vaccine incorporated pellet (FAV). The control group (FC) fishes were fed with commercial pellet.	79
5.3	The gut lavage fluid IgM response following vaccination with vaccine incorporated pellet (FNV) and adjuvant vaccine incorporated pellet (FAV). The control group (FC) fishes were fed with commercial pellet.	81
5.4	The percentage of survival in red tilapia after challenge with 3.4×10^9 live <i>S. agalactiae</i> by intraperitoneal injection.	84
5.5	The diameter of gut-associated lymphoid tissue (GALT) observed in red tilapia following oral vaccination of FNV and FAV vaccine.	86
5.6	The number of lymphocytes counted in gut-associated lymphoid tissue (GALT) observed in red tilapia following oral vaccination of FNV and FAV vaccine.	87
5.7	Cross-section of the gut of red tilapia fed with FAV vaccine. Three aggregations of lymphoid cells or GALTs formed in the lamina propria as marked with yellow circles. HE x200.	88

- 5.8 Cross-section of the gut of red tilapia fed with FNV vaccine. Aggregations of lymphoid cells or GALT formed in the lamina propria as marked with yellow circle. HE x200. 89
- 5.9 Cross-section of the gut of fish fed with commercial feed from Group FC. No aggregation of lymphoid cells or GALT formed in the lamina propria. HE x200. 90



LIST OF ABBREVIATIONS

BHI	Brain Heart infusion
BSA	Bovine Serum albumin
CFU	Colony Forming Units (bacteria)
ELISA	Enzyme-Linked Immunosorbent Assay
FNV	Vaccine Incorporated Pellet
FAV	Adjuvant Vaccine Incorporated Pellet
GALT	Gut-associated Lymphoid Tissue
H&E	Hematoxylin and Eosin
M	Molar
PBS	Phosphate-Buffered Saline
rpm	Revolutions per minute
RM	Malaysian Ringgit
OD	Optical Density
<i>S. agalactiae</i>	<i>Streptococcus agalactiae</i>
<i>S. iniae</i>	<i>Streptococcus iniae</i>
USD	United States Dollar

CHAPTER 1

INTRODUCTION

Streptococcus agalactiae is an important pathogenic bacterium that infects various species of animals. *Streptococcus agalactiae* has a broad host range and is pathogenic to mammals, reptiles, amphibians and fish (Elliott *et al.*, 1990). *Streptococcus agalactiae* causes bovine mastitis in cow, characterized by inflammation of the parenchyma of the mammary gland in the presence of significantly increased leukocyte content in milk (Meiri-Bendek *et al.*, 2002). In human, *S. agalactiae* causes sepsis in neonates and their mothers, may lead to overwhelming infection without much localization (septicemia) or predominantly localized in the lung causing pneumonia or the brain causing meningitis (Fanrong *et al.*, 2005). *S. agalactiae* infection or streptococcosis was reported to cause significant mortalities among numerous wild and cultured fish species, including menhaden (*Brevoortia patronus*) (Plumb *et al.*, 1974), bullminnows (*Fundulus grandis*) (Rasheed and Plumb, 1984), striped bass (*Morone saxatilis*) (Baya *et al.*, 1990), mullet (*Liza klunzingeri*), and tilapia (*Oreochromis niloticus*) (Evans *et al.*, 2002).

Streptococcosis is a septicaemic disease affecting both captive and wild fish in all water bodies (Evans *et al.*, 2002). It affects large-sized tilapias, particularly those weighing at least 100 grams (Amal *et al.*, 2009). Currently, the disease has become a major problem in many tilapia farms and there is still no effective control measure. The two common *Streptococcus* sp. found

to infect tilapia were *S. agalactiae* (Pasnik *et al.*, 2005) and *S. iniae* (Intervet, 2006).

Streptococcosis has proven to be very difficult to control with antibiotics. According to Agriculture Research Service (ARS), United States Department of Agriculture (USDA), no antibiotic is available to treat the pathogen at this time (Agriculture Research Service, 2010). Moreover, the infection is usually transmitted from fish to fish where the bacteria were released from dead and dying fish that was considered as the most important source of infection (Kitao, 1993). Hence, vaccination is considered as the effective way to control the disease. In Malaysia, high incidences of *S. agalactiae* infection in red tilapia were reported during dry months between April and July, where water temperature was high (Siti-Zahrah *et al.*, 2005; Amal *et al.*, 2008).

Vaccination of fish by intraperitoneal injection and bath immersion against bacterial infections has been proven to be a commercial success (Lamers, 1985). However, oral vaccination, with antigen being incorporated into a feed, is potentially the most appropriate method for mass vaccination of fishes of all sizes. This route of vaccination avoids the stress and reduces time and labour cost whilst allowing a more flexible approach to the formulation of immunization regime, particularly important during handling large numbers of fish (Hart *et al.*, 1987). Oral vaccination is also easy to apply on all production facilities and is effective on all sizes of fish, making it the most preferred method by fish farmers (Le Breton, 2009). Immunization through oral or anal routes resulted in antigen-specific antibodies in skin mucus, bile and intestine of several fish species (Fletcher and White, 1973; Roumbout *et al.*, 1986).

The significant response by the mucosal immune system makes oral vaccination more promising since the first contact with pathogens occurs usually through mucosal surfaces (Manganaro *et al.*, 1994).

Thus, the objectives of this study was:

1. to determine the pathogenecity of *S. agalactiae* in normal and temperature-stressed red tilapia (*Oreochromis* spp).
2. to evaluate the protective capacity provided by an orally delivered incorporated feed vaccine (FNV) against streptococcosis in red tilapia (*Oreochromis* spp).
3. to enhance the protection provided by the newly developed oral vaccine (FAV) against streptococcosis in red tilapia (*Oreochromis* spp).

Hypotheses of this study was:

1. *S. agalactiae* is more virulence in temperature-stressed red tilapia rather than in non-stressed red tilapia.
2. orally delivered incorporated feed vaccine (FNV) are able to give protection against streptococcosis in red tilapia (*Oreochromis* spp.).
3. Newly developed oral vaccine (FAV) are able to enhance the protection against streptococcosis in red tilapia (*Oreochromis* spp.).

BIBLIOGRAPHY

- Acton, R.T., Weinheimer, P.F., Hall, S. J., Niedermaier, W., Shelton, E. and Bennett, J.C. (1971). Tetrameric immune macroglobulins in three orders of bony fishes. *Proceedings of the National Academy of Sciences, USA*. 68: 107–111.
- AC Tropical Fish. (2008). Streptococcus in tilapia. (Accessed on August 21, 2008 at <http://www.aquaticcommunity.com/tilapia/Streptococcus.php>.)
- Anderscn, D., Dixon, O. W. and Roherson, B. S. (1979). Kinetics of the primary immune response in rainbow trout after flush exposure to *Yersinia ruckeri* O-antigen. *Developmental and Comparative Immunology*. 3: 739-744.
- Agriculture Research Service (ARS), United States Department of Agriculture (USDA). (2010). New Streptococcal Vaccine. (Accessed on August 1, 2010 at <http://www.ars.usda.gov/research/patents/patents.htm?serialnum=10807575>.)
- Alcamo, I. E. (1997). In: *Fundamentals of Microbiology*. 5th edn.,(Addison Wesley Longman, Inc., California), 766.
- Amal, A.M.N., Siti-Zahrah, A., Zulkafli, R., Misri, S., Ramley, A. and Zamri-Saad, M. (2008). The effect of water temperature on the incidence of *Streptococcus agalactiae* infection in cage cultured tilapia. *International Seminar on Management Strategies on Animal Health and Production Control in Anticipation of Global Warming*, Surabaya, Indonesia. 48-51.
- Anderson, D., Dixon, O. W. and Roherson, B. S. (1979). Kinetics of the primary immune response in rainbow trout after flush exposure to *Yersinia ruckeri* O-antigen. *Developmental and Comparative Immunology*. 3: 739-744.
- Anon. (2004). Annual Fisheries Statistics Volume 1, (2003). Department of Fisheries Malaysia, Ministry of Agriculture and Agro-Based Industry.
- Audibert, F. M. and Lise, L. D. (1993). Adjuvants: Current status, clinical perspectives and future prospects. *Immunology Today*. 14: 281-284.
- Balarin, J. D. and Haller, R. D. (1982). The intensive culture of tilapia in tanks, raceways and cages. In: Muir, J. F. and Roberts, R. J. (eds) *Recent Advances in Aquaculture*. Croom Helm, London and Canberra, and Westview Press, Boulder, Colorado. 267-355.

- Baya. A. M., Lupiani, B., Hetrick, F. M., Roberson. B. S., Lukacovic, R., May, E. and Poukish. C. (1990). Association of *Streptococcus* sp, with fish mortalities in Chesapeake Bay and its tributaries. *Journal of Fish Diseases*. 13: 251-253.
- Berg, A., Rodseth O. M. and Hansen, T. (2007). Fish size at vaccination influence the development of side-effects in Atlantic salmon (*Salmo salar* L.). *Aquaculture*. 265: 9–15.
- Black, J.G. (2002). Microbiology: Principles and Explorations 5th Edition. John Wiley and Son Inc. United States. 455-456.
- Boomker, J., Imes, G. D. Jr, Cameron, C. M., Naude, T. W. and Schoonbee, H. J. (1979). Trout mortalities as a result of *Streptococcus* infection. *Onderstepoort Journal of Veterinary Research*. 46: 71-77.
- Bowden, T. J., Lester, K., MacLachlan, P. and Bricknell, I. R. (2000). Preliminary study into the short term effects of adjuvants on Atlantic halibut (*Hippoglossus hippoglossus* L.). *Bulletin of The European Association of Fish Pathologists*. 20: 148–152.
- Bragg, R. R., Oosthuizen, J. H. and Lordan, S. M. (1989). The leech *Batracobdelloides tricarinata* (Blanchard, 1897) (Hirudinea: Glossiphoniidae) as a possible reservoir of the rainbow trout pathogenic *Streptococcus* species. *Onderstepoort Journal of Veterinary Research*. 56: 203-204.
- Bravo, S. and Midtlyng, P. J. (2007). The use of fish vaccines in the Chilean salmon industry 1999–2003. *Aquaculture*. 270: 36–42.
- Brooks, G. F., Butel, J. S. and Morse, S. A. (2001). Jawetz, Melnick, & Adelberg's Medical Microbiology. 22nd Edition. Appleton & Lange.
- Brugere, C. and Ridler, N. (2004). Global Aquaculture Outlook In The Next Decades: An analysis of National Aquaculture Production Forecasts 2030. *Food and Agriculture Organization of The United Nations, Rome*.
- Buchmann, K. and Lindenstrom, T. (2002) Interactions between monogenean parasites and their fish hosts. *International Journal for Parasitology*. 32: 309–319.
- Carson, J., Gudkovs, N. and Austin, B. (1993). Characteristics of an *Enterococcus*-like bacterium from Australia and South Africa, pathogenic for rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of Fish Diseases*. 16: 381-388.
- Cook, D. W. and Lofton, S. R. (1975). Pathogenecity studies with a *Streptococcus* sp. isolated from fishes in an Alabama-Florida fish kill. *Transactions of the American Fisheries Society*. 2: 286-288.

- Coykendall, A. L. (1989). Classification and identification of the viridans streptococci. *Clinical Microbiology Reviews*. 2: 315-328.
- Dalmo, R. A., Ingebrightsens, K. and Bogwald, J. (1997). Non-specific defense mechanisms in fish, with particular reference to the reticuloendothelial system (RES). *Journal of Fish Diseases*. 20: 241-273.
- Davidson, G. A., Lin, S.-H., Secombes, C. J. & Ellis, A. E. (1997). Detection of specific and 'constitutive' antibody secreting cells in the gills, head kidney and peripheral blood leucocytes of dab (*Limanda limanda*). *Veterinary Immunology and Immunopathology*. 58: 363-374.
- Davina, J.H.M., Parmentier, H.K. and Timmermans, L.P.M. (1982). Effect of oral administration of vibrio bacterin on the intestine of cyprinid fish. *Development and Comparative Immunology Supplement*, 2, 166.
- Department of Fisheries Malaysia, Ministry of Agriculture and Agro-Based Industry. (2010). Senario Industri Perikanan Malaysia. (Accessed on August 25, 2010 at http://www.dof.gov.my/18?p_p_id=56_INSTANCE_3Fyn&p_p_lifecycle=0&p_p_state=normal&p_p_mode=view&p_p_col_id=column-2&p_p_col_count=1&page=7).
- Duremdez, R., Al-Marzouk, A., Qasem, J. A., Al-Harbi, A. and Gharabally, H. 2004. Isolation of *Streptococcus agalactiae* from cultured silver pomfret, *Pampus argenteus* (Euphrasen), in Kuwait. *Journal of Fish Diseases*. 27: 307-310.
- Eldar, A., Bejerano, Y., Livoff, A., Hurvitz, A. and Bercovier, H. (1995). Experimental meningo-encephalitis in cultured fish. *Veterinary Microbiology*. 43: 33-40.
- Eldridge, J.H., Hammond, C. J., Meulbroek, J. A., Staas, J. K., Gilley, R.M. and Tice, T. R. (1990). Controlled vaccine release in the gut-associated lymphoid tissues. I. Orally administered biodegradable microspheres target the Peyer's patches. *Journal of Controlled Release*. 11: 205-214.
- Elliott, J.A., Facklam, R.R. and Richter, C. B. (1990). Whole-cell protein patterns of non-hemolytic group B, type1b, streptococci isolated from humans, mice, cattle, frogs, and fish. *Journal of Clinical Microbiology*. 28: 628-630.
- Ellis, A. E., Stapleton, K. J. and Hastings, T. S. (1988). The humoral immune response of rainbow trout (*Salmo gairdneri*). *Veterinary Immunology and Immunopathology*. 19: 153-164.

El-Sayed, A. -F. M. (2006). Tilapia Culture. CABI Publication, Oxfordshire, United Kingdom.

Evans, J. J., Klesius, P. H., Gilbert, P. M., Shoemaker, C. A., Al Sarawi, M. A., Landsberg, J., Duremdez, R., Al Marzouk, A. and Al Zenki, S. (2002). Characterization of β -haemolytic Group B *Streptococcus agalactiae* in cultured seabream, *Sparus auratus* L., and wild mullet, *Liza klunzingeri* (Day), in Kuwait. *Journal of Fish Diseases*. 25: 505-513.

Evans J.J., Klesius P.H. and Shoemaker C.A. (2006). An overview of *Streptococcus* in warm water fish. *Aquaculture Health International*. 7: 10-14.

Evensen, O. (2009). Development in fish vaccinology with focus on delivery methodologies, adjuvants and formulations. *Options Méditerranéennes*. 86: 177-186.

Fanrong, K., Lin, M and Gwendolyn, L. G. (2005). Simultaneous detection and serotype identification of *Streptococcus agalactiae* using multiplex PCR and reverse line blot hybridization. *Journal of Medical Microbiology*. 54: 1133-1138.

FAO. (2000). World Review of Fisheries and Aquaculture. *Fisheries Resources: Trends In Production, Utilization and Trade*. Rome: Food and Agriculture Organization of the United Nations.

FAO. (2003). Aquaculture Production Statistics 1988–1997. Rome, FAO.

FAO. (2004). *Fishstat Plus*. FAO, Rome.

FAOSTAT. (2004). Fisheries Data. Available online: <http://faostat.external.fao.org/faostat/collections?subset=fisheries>, last updated in February 2004. Rome, FAO.

FAO. (2010). Text by Rohana Subasinghe. Aquaculture topics and activities. State of World Aquaculture, 2005-2010. In: *FAO Fisheries and Aquaculture Department* [online]. Rome. Updated 27 May 2005. [Cited 11 September 2010].

Figueiredo, H.C.P., Carneiro, D.O., Faria, F.C., Costa, G.M. (2006). *Streptococcus agalactiae* associado à meningoencefalite e infecção sistêmica em tilápias-do-nilo (*Oreochromis niloticus*) no Brasil. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*. 8: 678–680.

Finegold, S. M. and Baron E. J. (1986). Bailey and Scott's Diagnostic Microbiology 7th Edition. The C. V. Mosby Company, United States of America.

FishStat Plus. (2004). Aquaculture Production: Quantities 1950–2002. Updated 30 April 2004. Rome, FAO.

Fletcher, T. C. and A. White, Antibody production in the plaice (*Pleuronectes platessa*) after oral and parenteral immunization with *Vibrio anguilarum* antigens. *Aquaculture*, 1 : p. 417.

Garcia, J.C., Klesius, P.H., Evans, J.J., Shoemaker, C.A., 2008. Non-infectivity of cattle *Streptococcus agalactiae* in Nile tilapia, *Oreochromis niloticus* and channel catfish, *Ictalurus punctatus*. *Aquaculture*. 281: 151–154.

Georgopoulou, U. and Vernier, J. M. (1986). Local immunological response in the posterior intestinal segment of the rainbow trout after oral administration of macromolecules. *Developmental and Comparative Immunology*. 10: 529-537.

Getachew, T. (1989). Stomach pH, feeding rhythm and ingestion rate in *Oreochromis niloticus* L. (Pisces: Cichlidae) in Lake Awasa, Ethiopia. *Hydrobiologia*. 174: 43-48.

Ghittino, C. and Prearo, M. (1992). Segnalazione di Streptococci nella trota iridea (*Onchorynkus mykiss*) in Italia: nota preliminare. *Boll. Soc. Ital. Pattol. Ittica (S.I.P.I.)*. 4: 4-11.

Gina, C. and David, A., C. (2006). Bacterial Haemorrhagic Septicaemia in Tilapias. *Aquaculture Health International*. 7: 7-8.

Gudding, R., Lillehaug, A. and Evensen, O. (1999). Recent developments in fish vaccinology. *Veterinary Immunology and Immunopathology*. 72: 203-212.

Iger, Y. & Wendelaar Bonga, A. F. (1994). Cellular aspects of the skin of carp exposed to acidified water. *Cell and Tissue Research*. 275, 481–492.

Intervet. (2006). Streptococcosis in Tilapia. (Accessed on August 22, 2008 at http://aqua.intervet.com/news/2006-06-20_streptococcosis_in_tilapia.asp).

Intervet. (2010). New Advances for Warmwater Aquaculture. (Accessed on October 10, 2010 at http://aqua.intervet.com/binaries/Aguirre_tcm127-207376.pdf).

Iwama, G. and Nakanishi, T. (1996). The Fish Immune System. Organ, Pathogen, and Environment. Academic Press, San Diego. 1-380.

Hoshina, T. (1956). An epidemic disease affecting rainbow trout in Japan. *Journal of the Tokyo University of Fisheries*. 42: 35-46.

- Humphrey, J. D., Lancaster, C. E., Gudkovs, N. and Copland, J. W (1987). The disease status of Australian salmonids: bacteria and bacterial diseases. *Journal of Fish Diseases*. 10: 403-410.
- Kaattari, S., Evans, D. and Klemer, J. (1998). Varied redox forms of teleost IgM: an alternative to isotopic diversity? *Immunological Reviews*. 166: 133–142.
- Kenneth, D. C., Darren, R. J. and Robert, L. R. (2000). Characterization of mucosal and systemic immune responses in rainbow trout (*Oncorhynchus mykiss*) using surface plasmon resonance. *Fish and Shellfish Immunology*. 10: 651-666.
- Kimura, H. and Kusuda, R. (1979). Studies on the pathogenesis of streptococcal infection in cultured yellowtails *Seriola* spp.: effect of cell free culture on experimental streptococcal infection. *Journal of Fish Diseases*. 2: 501-510.
- Kitao, T. (1993). Streptococcal infections. In: V. Inglis, R. J. Roberts and N. R. Bromage (Editors). *Bacterial Diseases of Fish*. Blackwell, Oxford, pp. 196-210.
- Kitao, T., Aoki, T. and Sakoh, R. (1981). Epizootic caused by beta haemolytic *Streptococcus* species in cultured freshwater fish. *Fish Pathology*. 15: 301-307.
- Klesius, P. H., Evans, J. J., Shoemaker, C. A. and Lim, C. (2006). A US perspective on selected biotechnological advancements in fish health. *Aquaculture Health International*. 5: 10- 12.
- Knox, K. W and Wicken, A. J. (1973). Immunological properties of teichoic acids.. *Bacteriological Reviews*. 37 (2): 215–57.
- Kobayashi, K., Tomonaga, S. and Kajii, T. (1984). A second class of immunoglobulin other than IgM present in the serum of cartilaginous fish, the skate, *Raja kenojei*: Isolation and characterization. *Molecular Immunology*. 21: 397–404.
- Komar, C., Enright, W. J., Grisez, L. and Tan, Z. (2004). Understanding Fish Vaccination. *Aquaculture Asia Pasific*. Nov/Dec: 24-26.
- Kusuda, R., Kawai, K., Jo, Y., Akizuki, T., Fukunaga M. & Kotake, N. (1978). Efficacy of oral vaccination for vibriosis in cultured ayu. *Bulletin of the Japanese Society of Scientific Fisheries*. 44: 21-26.
- Le Breton, A. D. (2009). Vaccines in Mediterranean aquaculture: Practice and needs. *Options Mediterraneennes*. 86: 147-154.

- Levinson, W. and Jawetz, E. (1998). Medical Microbiology & Immunology, Examination & Board Review 6th. McGraw-Hill International Editions, New York. Pp. 339-340.
- Lin, S.-H., Davidson, G. A., Secombes, C. J. & Ellis, A. E. (1998). A morphological study of cells isolated from the perfused gill of dab and Atlantic salmon. *Journal of Fish Biology*. 53: 560–568.
- Lindblad, E. B. (2000). Freund's adjuvants. In: O'Hagan DT, editor. Methods in molecular medicine: Vaccine adjuvants—preparation methods and research protocols. Totowa, New Jersey: Humana Press. 49–63.
- Lobb, C. J. (1987). Secretory immunity induced in catfish, *Ictalurus punctatus*, following bath immunization. *Developmental and Comparative Immunology* 11, 727–738.
- Lombard M, Pastoret PP, Moulin AM (2007). "A brief history of vaccines and vaccination". *Reviews. - Off. International Epizootiology*. 26 (1): 29–48.
- MacDonald, T.T. and Miller, R.D. (2005). Phylogeny of the Gut-Associated Lymphoid Tissue (GALT). In Ogra, Mestecky, Lamm, Strober, McGhee and Bienenstock (Eds.), *Mucosal immunology* (3rd edn.) (pp. 323-334). San Diego, Academic Press.
- Magnadottir, B. (1998). Comparison of immunoglobulin (IgM) from four fish species. *Iceland Agriculture Science*. 12: 47-59.
- McMillan, D. N. and Secombes, C. J. (1997). Isolation of rainbow trout (*Oncorhynchus mykiss*) intestinal intraepithelial lymphocytes (IEL) and measurement of their cytotoxic activity. *Fish and Shellfish Immunology*. 7:527-541.
- Meiri-Bendek, I., Lipkin, E., Friedman, A., Leitner, G., Saran, A., Friedman, S and Kashi, Y. (2002). A PCR based method for the detection *Streptococcus agalactiae* in milk. *Journal of Dairy Science*. 85 : 1717-1723.
- Meneguen, A. and Gohin, F. (2006). Observation and modelling of natural retention structures in the English Channel. *Journal of Marine System*. 63: 244-256.
- Midtlyng, P. J. and Lillehaug, A. (1998). Growth of Atlantic salmon *Salmo salar* after intraperitoneal administration of vaccines containing adjuvants. *Disease of Aquatic Organisms*. 32: 91–97.
- Miyazaki, T., Kubota, S. S., Kaige, N. and Miyashita, T. (1984). A histopathological study of streptococcal disease in tilapia. *Fish Pathology*. 19: 167-172.

- Moore, J. D., Ototake, M. & Nakanishi, T. (1998). Particulate antigen uptake during immersion immunisation of fish: The effectiveness of prolonged exposure and the role of the skin and gills. *Fish & Shellfish Immunology*. 8: 393–407.
- Mutoloki, S., Alexandersen, S., Gravning, K. and Evensen, O. (2008). Time-course study of injection site inflammatory reactions following intraperitoneal injection of Atlantic cod (*Gadus morhua* L.) with oil-adjuvanted vaccines. *Fish and Shellfish Immunology*. 24: 386–393.
- Navot, N., Kimmel, E. and Avtalian, R. R. (2005). Immunisation of fish by bath immersion using ultrasound. *Developmental Biology (Basel)*. 121: 135-142.
- Pasnik, D. J., Evans, J. J., Klesius, P. H., Shelby, R. A and Shoemaker, C. A. (2005). Antigenicity of *Streptococcus agalactiae* extracellular product and vaccine efficacy. *Journal of Clinical Microbiology*. 35 : 2573-2579.
- Perera, R.P., Johnson, S.K. and Lewis, D.H. (1997). Epizootiological aspects of *Streptococcus iniae* affecting tilapia in Texas. *Aquaculture*. 152: 25-33.
- Pillay, T. V. R. (1990). Aquaculture principles and practices. Fishing New Book, Blackwell Science, Oxford, UK.
- Plotkin, S. (2003). Vaccines, vaccination and vaccinology. *Journal of Infectious Disease*. 187: 1347-1359.
- Plumb, J. A., Schachte, J. H., Gaines, J. L., Peltier, W. and Carroll, B. (1974). *Streptococcus* sp. from marine fishes along the Alabama and northwest Florida coast of the Gulf of Mexico. *Transactions of the American Fisheries Society*. 2: 358-361.
- Press, C. McL. and Evensen, O. (1999). The morphology of the immune system in teleost fishes. *Fish & Shellfish Immunology*. 9: 309-318.
- Pyle, S. W. and Dawe, D. L. (1985). Immune response of channel catfish, *Ictalurus punctatus* Rafinesque, to bacterial and protozoal antigens administered by three routes. *Aquaculture*. 46: 1-1
- Rasheed, V. and Plumb, J. A. (1984). Pathogenicity of a non-haemolytic group B *Streptococcus* sp. In gulf killifish, *Fundulus grandis*, Baird & Girard. *Aquaculture*. 37: 97–105.
- Robinson, J. A. and Meyer, F. P. (1966). Streptococcal fish pathogen. *Journal of Bacteriology*. 92: 51
- Ronald, R. J. (1978). Fish Pathology, Fifth Edition. Baillière Tindall of 1st Anne's Road, Eastbourne, East Sussex.

- Rombout, J. H. W. M., Blok, L. J., Lamers, C. H. J. and Engbert, E. (1986). Immunization of carp (*Cyprinus carpio*) with *Vibrio anguillarum* bacterin: indications for a common mucosal immune system. *Development and Comparative Immunology*. 10: 341.
- Rombout, J. H. W. M., Taverne, N., van de Kamp, M. and Taverne-Thiele, A. J. (1993). Differences in mucus and serum immunoglobulin of carp (*Cyprinus carpio* L.). *Development and Comparative Immunology*. 17: 309–317.
- Salerno-Goncalves, R. and Sztein, M. B. (2006). Cell-mediated immunity and the challenges for the vaccine development. *Trends in Microbiology*. 14: 536-542.
- Secombes, C. (2008). Will advances in fish immunology change vaccination strategies? *Fish and Shellfish Immunology*. 25: 409-416.
- Shelby, R. A., Klesius, P. H., Shoemaker, C. A. and Evans, J. J. (2002). Passive immunization of tilapia, *Oreochromis niloticus* (L.), with anti-*Streptococcus iniae* whole sera. *Journal of Fish Diseases*. 25: 1-6.
- Shoemaker, C. A., Evans, J. J. and Klesius, P. H. (2000). Density and dose: factors affecting mortality to *Streptococcus iniae*-infected tilapia (*Oreochromis niloticus*). *Aquaculture*. 188: 229-235.
- Siti-Zahrah, A., Padilah, B., Azila, A., Rimatulhana, R. and Shahidan, H. (2005). Multiple streptococcal species infection in cage-cultured red tilapia, but showing similar clinical signs. In: *Proceedings of the Sixth Symposium on Diseases in Asian Aquaculture*. MG Bondad-Reantaso, CV Mohan, M Crumlish, RP Subasinghe, Eds., Colombo, Sri Lanka. 332–339.
- Siti-Zahrah, A., S. Misri, B. Padilah, R. Zulkafli, B. C. Kua, A. Azila and R. Rimatulhana. (2004). Pre-disposing factors associated with outbreak of Streptococcal infection in floating cage-cultured red tilapia in reservoirs. *Abstracts of the 7th Asian Fisheries Forum 04, The Triennial Meeting of The Asian Fisheries Society 30th*. Nov-4th Dec 2004, Penang, Malaysia. 129.
- Spence, K. D., Fryer, J. L. and Pilcher, K. S. (1965). Active and passive immunization of certain salmonid fishes against *Aeromonas salmonicida*. *Canadian Journal of Microbiology*. 13: 397–405.
- Stills Jr, H. F. (2005). Adjuvants and antibody production: dispelling the myths associated with Freund's complete and other adjuvants. *ILAR Journal*. 46: 280–293.
- Takahashi, H. (2003). Antigen presentation in vaccine development. *Comparative Immunology, Microbiology and Infectious Diseases*. 26: 309–328.

- Toranzo, A. E., Devesa, S. P., Riaza, A., Nunez, S. and Barja, J. L. (1994). Streptococcosis in cultured turbot caused by an *Enterococcus*-like bacterium. *Bulletin of European Association of Fish Pathology*. 14: 19-24.
- Vandenberg, G. W. (2004). Oral vaccines for finfish: academic theory or commercial reality? *Animal Health Research Reviews*. 5 (2): 301-304.
- Van Heijenoort, J. (2001). Formation of the glycan chains in the synthesis of bacterial peptidoglycan. *Glycobiology*. 11 (3): 25-36.
- Yanong, R.P.E. and Floyd, R.F., 2002. Streptococcal infections of fish. Circular FA057, Florida Cooperative Extension Service, IFAS, University of Florida.
- Xu-dong, J., Shuang, C. and Yong-hua, H. (2010). Comparative study of the effects of aluminium adjuvants and Freund's incomplete adjuvant on the immune response to an *Edwardsiella tarda* major antigen. *Vaccine*. 28: 1832-1837.
- Zamri-Saad, M., Amal, M. N. A. and Siti-Zahrah, A. (2010). Pathological changes in red tilapias (*Oreochromis* spp.) naturally infected by *Streptococcus agalactiae*. *Journal of Comparative Pathology*. 143: 227-229.