



UNIVERSITI PUTRA MALAYSIA

***ASSESSMENT OF *Enterococcus hirae* STRAIN LAB3 ISOLATED FROM
ASIAN SEABASS, *Lates calcarifer* (Bloch, 1970) AS PROBIOTIC
AGAINST *Vibrio harveyi* INFECTION***

NUR FATHIAH BINTI MASDUKI

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By

NUR FATHIAH BINTI MASDUKI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirement for the degree Master of Science**

May 2018

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DEDICATION

To my beloved father and mother

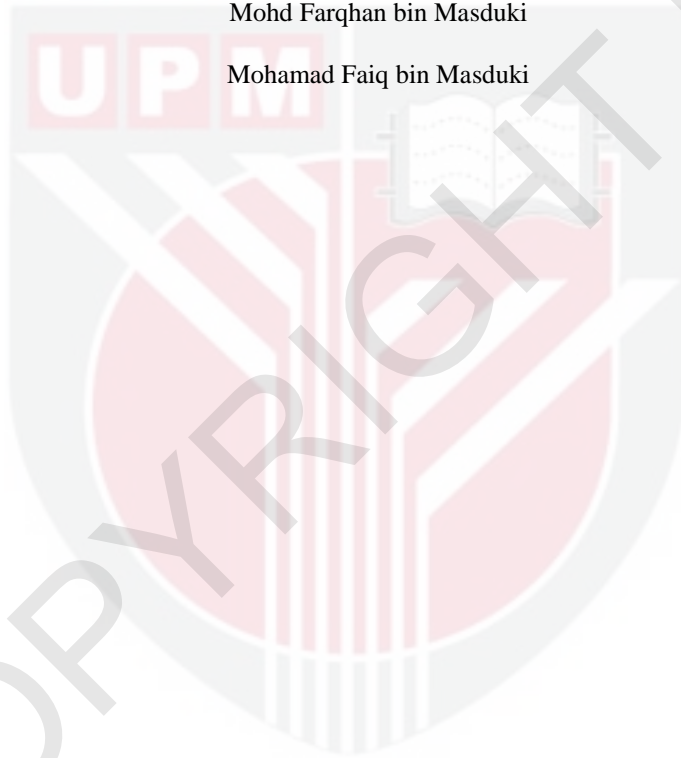
En. Masduki Bin Kasbi and Pn. Siti binti Kamin

Family members:

Nur Farqhana binti Masduki

Mohd Farqhan bin Masduki

Mohamad Faiq bin Masduki



Abstract of thesis prepared to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master Science

ASSESSMENT OF *Enterococcus hirae* STRAIN LAB3 ISOLATED FROM ASIAN SEABASS, *Lates calcarifer* (Bloch, 1970) AS PROBIOTIC AGAINST *Vibrio harveyi* INFECTION

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May 2018

Chair : Murni Marlina Abd Karim, PhD
Faculty : Institute of Bioscience

Seabass farming is one of significant important contributors to the annual world aquaculture production including Malaysia. However, vibriosis has been one of the main diseases problems often cause high mortality and reducing the production. Probiotics are now become subject of interest as new alternative in preventing vibriosis in fish farming culture. This study was undertaken to discover new potential probiotics strain from the group of lactic acid bacteria (LAB) isolated from seabass (*Lates calcarifer*).

Seven potential of LABs were successfully isolated from the intestine and liver of 15 healthy seabass. In *in vitro* screening test using series of plate assays, co-culture assay and pathogenicity test on TCBS agar, one isolate was showed potential as probiotics. The potential probiont was identified as *Enterococcus hirae* (LAB3) using 16S rRNA. This strain able to grow at pH 2 to 10 with the best growth at pH 7 within 3 h incubation period and grew best at 4% of NaCl in de Man Rogosa and Sharp (MRS) broth. Antibiotic susceptibility test revealed that, this strain was resistant to: kanamycin, penincilin, gentamycin, tetracycline and streptomycin as well as able to secrete lipase enzyme. In addition, this strain was able to produce biofilm up to 30 h of incubation period.

In a preliminary *in vivo* assay using *Artemia salina*, result demonstrated a significant survival of *Artemia* treated with *E. hirae* LAB3 at 10^6 CFUmL⁻¹ and challenged with *V. harveyi* (70.00±3.06%) compared to the control *Artemia* with pathogen only (*V. harveyi* 13.33±1.45%). This strain was proven able to recude the number of Vibrios load in both *Artemia* and culture water. Similar results were observed using seabass larvae as a host. Larvae treated with *E. hirae* LAB3 at 10^6 CFUmL⁻¹ had significant high survival after challenged with the pathogen *V. harveyi* (68.33±0.88%) compared with group with pathogen only (*V. harveyi* 16.67± 3.33%).

Vibrios counts were significantly lower in larvae and culture water treated with *E. hirae* LAB3 at the end of challenged assay. In term of growth rate, larvae treated with *E. hirae* LAB3 at concentration of 10^6 CFUmL⁻¹ showed slightly higher but with no

significant different in term of growth rate and length increment compared with control without probiotic added. This study suggests that *E. hirae* LAB3 has potential to be one of the probiotic candidates in aquaculture industry specifically for seabass hatchery system.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

PENILAIAN *Enterococcus hirae* STRAIN LAB3 YANG DIPENCIL DARIPADA SIAKAP, *Lates calcarifer* (Bloch, 1970) SEBAGAI PROBIOTIK TERHADAP SERANGAN *Vibrio harveyi*

Oleh

NUR FATHIAH BINTI MASDUKI

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Perternakan ikan siakap adalah salah satu penyumbang penting kepada pengeluaran tahunan akuakultur dunia, termasuk Malaysia. Walau bagaimanapun, penyakit vibriosis menjadi salah satu masalah utama kepada kadar kematian tinggi dan mengurangkan pengeluarannya. Probiotik kini menjadi subjek perhatian sebagai alternatif baru bagi mencegah vibriosis dalam budaya penternakan ikan. Kajian ini dijalankan untuk menemukan potensi baru probiotik dari kumpulan bakteria asid laktik (LAB) yang diasingkan dari ikan siakap (*Lates calcarifer*) bagi memerangi vibriosis.

Sejumlah tujuh LAB berpotensi telah berjaya diasingkan dari usus dan hati anak ikan siakap yang sihat. Dalam ujian penyaringan *in vitro* menggunakan siri ujian plat, ujian cerakan kultur dan ujian patogenik pada agar TCBS, satu bakteria berpotensi sebagai probiotik. Bakteria ini dikenal pasti sebagai *Enterococcus hirae* (LAB3) menggunakan 16S rRNA. *E. hirae* LAB3 dapat berkembang pada pH 2 hingga 10 dengan pertumbuhan terbaik pada pH 7 dalam tempoh masa inkubasi 3 jam dan berkembang dengan baik pada 4% daripada NaCl dalam stok Man Rogosa dan Sharp (MRS). Ujian kerentanan antibiotik mendedahkan bahawa, LAB3 ini mempunyai pertahanan terhadap: kanamicin, penicilin, gentamicin, tetracilin dan streptomisin, malah dapat merembeskan enzim lipase. Di samping itu, LAB3 ini dapat menghasilkan biofilm sehingga 30 jam selama tempoh inkubasi.

Pada prosedur awal *in vivo* menggunakan *Artemia salina*, hasil menunjukkan kelangsungan hidup *Artemia* yang dirawat dengan *E. hirae* LAB3 pada 10^6 CFU mL^{-1} dan dicabar oleh *V. harveyi* ($70.00 \pm 3.06\%$) berbanding dengan *Artemia* terkawal dengan patogen sahaja (*V. harveyi* : $13.33 \pm 1.45\%$). Terdapat pengurangan vibrios yang signifikan dalam *Artemia* dan kultur air yang dirawat dengan *E. hirae* LAB3 berbanding dengan kumpulan kawalan (*V. harveyi* sahaja). Hasil yang sama diperolehi dengan menggunakan larva ikan siakap sebagai hos. Larva yang dirawat dengan *E. hirae* LAB3 pada 10^6 CFU mL^{-1} mempunyai kadar kemandirian signifikan yang tinggi selepas dicabar dengan patogen *V. harveyi* ($68.33 \pm 0.88\%$) berbanding dengan kumpulan dengan patogen sahaja (*V. harveyi* : $16.67 \pm 3.33\%$).

Angka Vibrios jauh lebih rendah pada larva dan kultur air yang dirawat dengan *E. hirae* LAB3 pada akhir penilaian yang dijalankan. Dari segi kadar pertumbuhan, larva yang dirawat dengan *E. hirae* LAB3 pada kepekatan 10^6 CFU mL^{-1} menunjukkan sedikit tinggi tetapi tiada perbezaan yang signifikan dalam jangka masa pertumbuhan dan pertambahan kepanjangan berbanding dengan kawalan tanpa ditambah probiotik. Kajian ini menunjukkan bahawa *E. hirae* LAB3 berpotensi menjadi salah satu probiotik yang dicadangkan dalam industri akuakultur khususnya bagi sistem penetasan ikan siakap.



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I certify that a Thesis Examination Committee has met on 17 May 2018 to conduct the final examination of Nur Fathiah binti Masduki on her thesis entitled "Assessment of *Enterococcus hirae* Strain LAB3 Isolated from Asian Seabass, *Lates calcarifer* (Bloch, 1970) as Probiotic Against *Vibrio harveyi* Infection" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

DOC	Day of culture
NaCl	Sodium chloride
TCBS	Thiosulfate citrate bile salt sucrose
TSB	Trypticase soy broth
TSA	Trypticase soy agar
MRS	Man Rogosa and Sharp
FSSW	Filtered sterile seawater
CFU	Colony forming unit
CFU _{mL} ⁻¹	Colony forming unit per milliliter
rpm	Revolution per minute
μl	Microliter
h	Hour
Kb	Kilobase
bp	Basepair
16S rRNA	16 subunit ribosomal ribonucleic acid
PCR	Polymerase chain reaction
DNA	Deoxyribonucleic acid
sp. or spp.	Species (for singular or plural term)
SGR	Specific growth rate

CHAPTER 1

INTRODUCTION

1.1 Background of Study

One of the world's greatest challenges to be faced is how to feed more than 9.7 billion people by 2050. Gladly, in 2014, aquaculture sectors have contributed to the supply of fish for human consumption that represent one third of the world fisheries production (FAO, 2016). According to (FAO, 2015), global aquaculture production in 2014 was recorded at 73.8 million tonnes with Asian region as a whole has been produced more farmed fish than wild catch fish since 2008, with total production reached to 44.1 percent in 2014, up from 42.1 percent in 2012 and 31.1 percent in 2004.

Among 600 aquatic species that have been cultured worldwide for production, fish such as seabass (*Lates calcarifer*) seems to be the most cultured in a variety of farming systems (FAO, 2014). Moreover, fish provided more than 3.1 billion people due to their valuable nutritional properties. Considering to the global population growth, clearly, the demand for fish consumption will increase each year. Hence, the current trend in aquaculture is towards more intensive and commercialized of aquatic production.

As in aquaculture industry, presence of pathogenic microorganism in the culture system is a major concern to the industry. Vibriosis due to *Vibrio* sp. posed a threat to the most marine aquaculture production including seabass culture which causing major economic losses (Wei and Wee, 2014). This infectious disease cost the global losses of aquaculture industry in billions of dollars annually (Lafferty *et al.*, 2015).

With concerns regarding antibiotic resistance case among pathogens as well as many chemical are now being banned, environmental friendly alternatives are become subject of interest among researchers. Probiotic is one of the several alternative approaches that gaining popularity in aquaculture. Probiotic influence the composition of gut microbiota and confer beneficial health effect to their host (Nayak, 2010; Newaj-Fyzul *et al.*, 2014). Additionally, probiotics isolated from the host gut or their environment are more compatible to be commercialized due to their ability to colonize the host gut as well as prevent a threat to the surrounding ecosystem (Verschuere, 2000; Sayes *et al.*, 2018).

Desirable characteristics for the selection of probiotic include harmless to the host, should be accepted by the host and actually work *in vivo* as opposed to *in vitro* finding (Chandrakala and Soundharanayaki, 2017). Moreover, microorganism that used as probiotics in aquaculture should be safe not only for the host yet also for their environment also human (Munoz-Atienza *et al.*, 2013).

The diversity of probiotics in the host gut is correlated with their habitats and closely interacts with these microbes. For instance, probiotic of the genera *Bacillus*, *Lactobacillus* and *Enterococcus* have the potential to influence the immune system of the host (Sayes *et al.*, 2018). Recently, lactic acid bacteria (LABs) are gaining acceptance for human and animal purposes. It's known to be present in the intestine of healthy fish (Pandiyani *et al.*, 2013). In addition, *Lactobacillus* is one of the genera of LAB that most widely used in aquaculture due to their better yield in feed conversion, growth rate, weight gain (Dawood *et al.*, 2016) as well as increase the growth performance of the fish and antagonistic activity against *Vibrios* (Afrilasari and Meryandini, 2016; Gao *et al.*, 2017). Thus, it is interesting to isolate probiotic either from the wide diversity bacteria in either host or their culture environment and examine its potentially effects as potential probiotics for commercialize.

1.2 Problem Statement

Asian seabass (*Lates calcarifer*) was one of the leading species for the previous five years in Malaysia that become main cultured species comprises 37 percent over the marine total production. However, problems related to disease outbreaks have caused serious mortalities and lowered the aquaculture production (FAO, 2015). Vibriosis causes severe economical losses in shrimp, finfish and mollusk cultivation worldwide (Austin and Zhang, 2006; Defoird *et al.*, 2014).

Vibrios belong to the *Harveyi* clade is among the major pathogens of aquatic organism (Yang and Defoirdt, 2015). Asian seabass is also susceptible to *Vibrio harveyi* infections that include vasculitis, eye-lesions and luminous vibriosis (Austin and Zhang, 2006). In Malaysia, Ransangan *et al.* (2012) have stated that, *V. harveyi* infection in Asian seabass spreads rapidly among fish stocked in the same cage since in year 2008 in open cage in Sabah. In addition, *V. harveyi* also a pathogenic organism associated with luminous vibriosis that contributed mortality in penaeid shrimp farm (Wang *et al.*, 2015).

Vibriosis is highly infectious to the early stages of fish and introducing the beneficial and healthy microbial in aquatic environment in larval rearing tanks can positively influence the well-being of the fish during larviculture (Banerjee and Ray, 2017). Hence, it is interesting to develop potential probiotic bacteria isolated from the host itself for improvement of their health.

1.3 Significant of the Study

In the past few years, application of probiotics in aquaculture has been one of the major interesting research subjects due to their capability to control diseases in aquatic farms. It could be expressed by probiotic in ability to provide nutritional substances as well as enzymes, which aids in digestion process, improve, water quality and immune response and also confer resistance towards diseases (Qi *et al.*, 2009; Tuan *et al.*, 2013).

Recently, probiotic in aquaculture have been studied widely and lactic acid bacteria (LAB) have aroused as an important scientific interest in recent years (Alonso *et al.*, 2018). LAB has their own historical role as it present in the gut microbiota of freshwater fishes with beneficial effects on growth performance, nutrient digestibility and immune system modulator (Mohapatra *et al.*, 2011). Common LABs that have been used in aquaculture are *Lactobacillus lactis*, *Enterococcus* spp and *Lactococcus* spp (Lin *et al.*, 2017).

Despite being resistance to bile salts and acidity, LAB strain is indigenous population regarded as safe status adding a merit to be probiotic in aquaculture. Based on these criteria, it was possible to isolate LAB from the gut of local marine fish and develop new local probiotic especially for Asian seabass larviculture.

1.4 Objectives of Study

It is necessary to discover local strain of probiotics that can be used in Asian seabass cultures in order to increase the production of quality and healthy fish. Hence, the objectives of this research were:

1. To isolate, screen and identify lactic acid bacteria (LAB) as probiotic isolated from juvenile seabass (*Lates calcarifer*) against pathogenic *Vibrio* spp.
2. To determine the properties of selected LAB strain as potential probiont.
3. To evaluate the protective efficacy of selected potential probiotics on *Artemia salina* and Asian seabass (*Lates calcarifer*) larvae in *in vivo* challenge assay.

1.5 Hypothesis of Study

The hypothesis of the study:

Null hypothesis: Isolated LAB unable to inhibit the growth of pathogen in *in vitro* assay, did not potray any probiotic characteristics and incapable in protecting *Artemia salina nauplii* and seabass larvae from *V. harveyi* infections.

Alternative hypothesis: Isolated LAB able to inhibit the growth of pathogen in *in vitro* assay, showed positive characteristics as probiotics as well as able to confer protection to *Artemia salina nauplii* and seabass larvae against *V. harveyi* infection.

REFERENCES

- Adams, C. A. (2010). The probiotic paradox: live and dead cells are biological response modifiers. *Nutrition research reviews*, 23(1), 37-46.
- Adnan, A. F. M., & Tan, I. K. (2007). Isolation of lactic acid bacteria from Malaysian foods and assessment of the isolates for industrial potential. *Bioresource Technology*, 98(7), 1380-1385.
- Adnan, M., Patel, M., & Hadi, S. (2017). Functional and health promoting inherent attributes of *Enterococcus hirae* F2 as a novel probiotic isolated from the digestive tract of the freshwater fish *Catla catla*. *Peer Journal* (5), 3085.
- Akinjogunla, O.J., Eghafona, N.O., Enabulele, I.O., Mboti, C.I., & Ogbemudia, F.O. (2010). Antibacterial activity of ethanolic extracts of *Phyllanthus amarus* against extended spectrum β -lactamase producing *Escherichia coli* isolated from stool samples of HIV sero-positive patients with or without diarrhoea. *African Journal of Pharmacy and Pharmacology*, 4(6), 402-407.
- Ali A. 1987 a. Sea bass (*Lates calcarifer*) larvae and fry production in Malaysia, pp. 144-147. In: Copland JW, Grey DL (eds) Management of Wild and Cultured Sea Bass/ Barramundi (*Lates calcarifer*). ACIAR Proceeding No. 20, 210pp. Australian Centre for International Agricultural Research, Canberra.
- Ali A. 1987 b. Status of sea bass (*Lates calcarifer*) culture in Malaysia, pp. 165-167. In: Copland JW, Grey DL (eds) Management of Wild and Cultured Sea Bass/ Barramundi (*Lates calcarifer*). ACIAR Proceeding No. 20, 210pp. Australian Centre for International Agricultural Research, Canberra.
- Allameh, S. K., Daud, H., Yusoff, F. M., Saad, C. R., & Ideris, A. (2012). Isolation, identification and characterization of *Leuconoctos mesenteroides* as a new probiotic from intestine of snakehead fish (*Channa striatus*). *African Journal of Biotechnology*, 11, 3810-3816.
- Allameh, S. K., Ringø, E., Yusoff, F. M., Daud, H. M., & Ideris, A. (2014). Properties of *Enterococcus faecalis*, a new probiotic bacterium isolated from the intestine of snakehead fish (*Channa striatus* Bloch). *African Journal of Microbiology Research*, 8(22), 2215-2222.
- Aly, S. M., Abd-El-Rahman, A. M., John, G., & Mohamed, M. F. (2008). Characterization of some bacteria isolated from *Oreochromis niloticus* and their potential use as probiotics. *Aquaculture*, 277(1), 1-6.
- Ammor, M. S., Flórez, A. B., & Mayo, B. (2007). Antibiotic resistance in non-enterococcal lactic acid bacteria and bifidobacteria. *Food microbiology*, 24(6), 559-570.
- Askarian, F., Kousha, A., Salma, W., & Ringø, E. (2011). The effect of lactic acid bacteria administration on growth, digestive enzyme activity and gut

microbiota in Persian sturgeon (*Acipenser persicus*) and Beluga (*Huso huso*) fry. *Aquaculture Nutrition*, 17(5), 488-497.

- Austin, B., & Zhang, X. H. (2006). *Vibrio harveyi*: a significant pathogen of marine vertebrates and invertebrates. *Letters in applied microbiology*, 43(2), 119-124.
- Austin, B., Austin, D., Sutherland, R., Thompson, F., & Swings, J. (2005). Pathogenicity of vibrios to rainbow trout (*Oncorhynchus mykiss*, Walbaum) and *Artemia* nauplii. *Environmental microbiology*, 7(9), 1488-1495.
- Bairagi, A., Ghosh, K. S., Sen, S. K., & Ray, A. K. (2002). Enzyme producing bacterial flora isolated from fish digestive tracts. *Aquaculture International*, 10(2), 109-121.
- Balan, S. S., Nethaji, R., Sankar, S., & Jayalakshmi, S. (2012). Production of gelatinase enzyme from *Bacillus* spp isolated from the sediment sample of Porto Novo Coastal sites. *Asian Pacific Journal of Tropical Biomedicine*, 2(3), S1811-S1816.
- Balcázar, J. L. (2003). Evaluation of probiotic bacterial strains in *Litopenaeus vannamei*. Final Report. *National Center for Marine and Aquaculture Research, Guayaquil, Ecuador*.
- Balcázar, J. L., De Blas, I., Ruiz-Zarzuela, I., Cunningham, D., Vendrell, D., & Muzquiz, J. L. (2006). The role of probiotics in aquaculture. *Veterinary microbiology*, 114(3), 173-186.
- Balcázar, J. L., De Blas, I., Ruiz-Zarzuela, I., Vendrell, D., Calvo, A. C., Márquez, I., & Muzquiz, J. L. (2007). Changes in intestinal microbiota and humoral immune response following probiotic administration in Brown trout (*Salmo trutta*). *British journal of nutrition*, 97(03), 522-527.
- Balcázar, J. L., Vendrell, D., de Blas, I., Ruiz-Zarzuela, I., Muzquiz, J. L., & Girones, O. (2008). Characterization of probiotic properties of lactic acid bacteria isolated from intestinal microbiota of fish. *Aquaculture*, 278(1), 188-191.
- Baldassarri, L., Cecchini, R., Bertuccini, L., Ammendolia, M. G., Iosi, F., Arciola, C. R., & Orefici, G. (2001). *Enterococcus* spp. produces slime and survives in rat peritoneal macrophages. *Medical microbiology and immunology*, 190(3), 113-120.
- Banerjee, S., Khatoon, H., Shariff, M., & Yusoff, F. M. (2010). Enhancement of *Penaeus monodon* shrimp postlarvae growth and survival without water exchange using marine *Bacillus pumilus* and periphytic microalgae. *Fisheries Science*, 76(3), 481-487.
- Barman, P., Banerjee, A., Bandyopadhyay, P., Mondal, K. C., & Das Mohapatra, P. K. (2011). Isolation, identification and molecular characterization of potential probiotic bacterium, *Bacillus subtilis* PPP 13 from *Penaeus monodon*. *Biotechnol Bioinf Bioeng*, 1(4), 473-482.

- Bauer, A. W., Kirby, W. M., Sherris, J. C., & Turck, M. (1966). Antibiotic susceptibility testing by a standardized single disk method. *American journal of clinical pathology*, 45(4), 493.
- Bennani, M., Amarouch, H., Oubrim, N., & Cohen, N. (2012). Identification and antimicrobial resistance of fecal enterococci isolated in coastal Mediterranean environments of Morocco. *European Journal Science Research*, 70(2), 266-275.
- Bentzon-Tilia, M., Sonnenschein, E. C., & Gram, L. (2016). Monitoring and managing microbes in aquaculture—Towards a sustainable industry. *Microbial biotechnology*, 9(5), 576-584.
- Birkbeck, T. H., & Ringø, E. (2005). Pathogenesis and the gastrointestinal tract of growing fish. *Biology of Growing Animals*, 2, 208-234.
- Bogino, P. C., Oliva, M. D. L. M., Sorroche, F. G., & Giordano, W. (2013). The role of bacterial biofilms and surface components in plant-bacterial associations. *International journal of molecular sciences*, 14(8), 15838-15859.
- Bondad-Reantaso, M. G., Subasinghe, R. P., Arthur, J. R., Ogawa, K., Chinabut, S., Adlard, R., & Shariff, M. (2005). Disease and health management in Asian aquaculture. *Veterinary parasitology*, 132(3), 249-272.
- Brown, M. (2011). Modes of action of probiotics: recent developments. *Journal of animal and Veterinary Advances*, 10(14), 1895-1900.
- Bruhn, J. B., Gram, L., & Belas, R. (2007). Production of antibacterial compounds and biofilm formation by *Roseobacter* species are influenced by culture conditions. *Applied and environmental microbiology*, 73(2), 442-450.
- Bucio, A., Hartemink, R., Schrama, J. W., Verreth, J., & Rombouts, F. M. (2006). Presence of *lactobacilli* in the intestinal content of freshwater fish from a river and from a farm with a recirculation system. *Food microbiology*, 23(5), 476-482.
- Buntin, N., Chanthachum, S., & Hongpattarakere, T. (2008). Screening of lactic acid bacteria from gastrointestinal tracts of marine fish for their potential use as probiotics. *Sonklanakarin Journal of Science and Technology*, 30(1), 141.
- Cai, Y., Suyanandana, P., Saman, P., & Benno, Y. (1999). Classification and characterization of lactic acid bacteria isolated from the intestines of common carp and freshwater prawns. *The Journal of general and applied microbiology*, 45(4), 177-184.
- Caipang, C. M. A., Pakingking Jr, R. V., & Apines-Amar, M. J. S. (2012). Screening of vibriosis in Asian seabass, *Lates calcarifer* using loop-mediated isothermal amplification (LAMP) assay. *Human & Veterinary Medicine*, 4(2).
- Calo-Mata, P., Arlindo, S., Boehme, K., de Miguel, T., Pascoal, A., & Barros-Velazquez, J. (2008). Current applications and future trends of lactic acid

- bacteria and their bacteriocins for the biopreservation of aquatic food products. *Food and Bioprocess Technology*, 1(1), 43-63.
- Carnevali, O., Zamponi, M. C., Sulpizio, R., Rollo, A., Nardi, M., Orpianesi, C., & Cresci, A. (2004). Administration of probiotic strain to improve sea bream wellness during development. *Aquaculture International*, 12(4-5), 377-386.
- Cebeci, A., & Gürakan, C. (2003). Properties of potential probiotic *Lactobacillus plantarum* strains. *Food Microbiology*, 20(5), 511-518.
- Chang, C. I., & Liu, W. Y. (2002). An evaluation of two probiotic bacterial strains, *Enterococcus faecium* SF68 and *Bacillus toyoi*, for reducing *edwardsiellosis* in cultured European eel, *Anguilla anguilla* L. *Journal of Fish Diseases*, 25(5), 311-315.
- Charteris, W. P., Kelly, P. M., Morelli, L., & Collins, J. K. (1998). Antibiotic susceptibility of potentially probiotic *Lactobacillus* species. *Journal of food protection*, 61(12), 1636-1643.
- Chatterjee, S., & Haldar, S. (2012). *Vibrio* related diseases in aquaculture and development of rapid and accurate identification methods. *Journal of Marine Science Research and Development S, 1*.
- Chen, H., Liu, S., Xu, X. R., Liu, S. S., Zhou, G. J., Sun, K. F., & Ying, G. G. (2015). Antibiotics in typical marine aquaculture farms surrounding Hailing Island, South China: occurrence, bioaccumulation and human dietary exposure. *Marine pollution bulletin*, 90(1), 181-187.
- Chong, C. I., and Liu, W. Y. (2000). An evaluation of two probiotic bacterial strains, *Enterococcus faecium* SF68 and *Bacillus toyoi*, for reducing *edwardsiellosis* in cultured European eel, *Anguilla anguilla* L. *Journal of Fish Diseases*, 25(5), 311-315.
- Chu, W. H. (2007). Optimization of extracellular alkaline protease production from species of *Bacillus*. *Journal of industrial microbiology & biotechnology*, 34(3), 241-245.
- Creeper, J. H., & Buller, N. B. (2006). An outbreak of *Streptococcus iniae* in barramundi (*Lates calcarifera*) in freshwater cage culture. *Australian veterinary journal*, 84(11), 408-411.
- Clements, K. D. (1997). Fermentation and gastrointestinal microorganisms in fishes. *Gastrointestinal microbiology*, 1, 156-198.
- Conway, P. L. (1996). Selection criteria for probiotic microorganisms. *Asia Pacific Journal of Clinical Nutrition*, 5, 10-14.
- Das, P., Mandal, S. C., Bhagabati, S. K., Akhtar, M. S., & Singh, S. K. (2012). Important live food organisms and their role in aquaculture. *Frontiers in Aquaculture*, 5, 69-86.

- Davey, M. E., & O'toole, G. A. (2000). Microbial biofilms: from ecology to molecular genetics. *Microbiology and molecular biology reviews*, 64(4), 847-867.
- Davis, T. L. O. (1985). The food of barramundi, *Lates calcarifer* (Bloch), in coastal and inland waters of Van Diemen Gulf and the Gulf of Carpentaria, Australia. *Journal of fish biology*, 26(6), 669-682.
- Devriese, L. A., Van de Kerckhove, A., Kilpper-Bälz, R., & Schleifer, K. H. (1987). Characterization and identification of *Enterococcus* species isolated from the intestines of animals. *International Journal of Systematic and Evolutionary Microbiology*, 37(3), 257-259.
- Dicks, L. M. T., & Botes, M. (2010). Probiotic lactic acid bacteria in the gastrointestinal tract: health benefits, safety and mode of action. *Beneficial Microbes*, 1, 11-29.
- Djajadiredja, R., T.H. Panjaitan, A. Rukyani, A. Saron, D. Satyani, H. Supriyadi, (1983). Country reports: Indonesia. In: Davy, F.B., Chouinard, A. (Eds.). Fish quarantine and fish diseases in Southeast Asia, pp: 19-30. Report of workshop held in Jakarta, Indonesia, 7-10 December 1982. International Development Research Centre Publication IDRC-210e, Ottawa.
- Drancourt, M., Bollet, C., Carlouz, A., Martelin, R., Gayral, J. P., & Raoult, D. (2000) 16S ribosomal DNA sequence analysis of a large collection of environmental and clinical unidentifiable bacterial isolates. *Journal Clinical. Microbiology*, 38, 3623-3630.
- Duan, Y., Tan, Z., Wang, Y., Li, Z., Li, Z., Qin, G., & Cai, Y. (2008). Identification and characterization of lactic acid bacteria isolated from Tibetan Qula cheese. *The Journal of general and applied microbiology*, 54(1), 51-60.
- Dunstan, D. J & CSIRO. Division of Fisheries and Oceanography (1959). The barramundi *Lates calcarifer* (Bloch) in Queensland waters. CSIRO, Melbourne.
- Eaton, T. J., & Gasson, M. J. (2001). Molecular screening of *Enterococcus* virulence determinants and potential for genetic exchange between food and medical isolates. *Applied and environmental microbiology*, 67(4), 1628-1635.
- Edward D. 2011. The health benefits of lipase. Global health centre. Version June 2011. Available at [http:// www.globalhealingcenter.com/ natural-health/ lipase/](http://www.globalhealingcenter.com/natural-health/lipase/) (accessed on 21 January 2018).
- El- Haroun, E. R., Goda, A. S., & Chowdhury, K. (2006). Effect of dietary probiotic Biogen® supplementation as a growth promoter on growth performance and feed utilization of Nile tilapia *Oreochromis niloticus* (L.). *Aquaculture Research*, 37(14), 1473-1480.
- F.A.O. (2007). The State of World Fisheries and Aquaculture 2006. Rome: Food and Agriculture Organization of United Nations.

- F.A.O. (2012). The State of World Fisheries and Aquaculture 2012. Global Aquaculture Production Statistics for the year.
- F.A.O. (2014). The State of World Fisheries and Aquaculture 2014. The opportunities and challenges in aquaculture. Rome, 2014.
- F.A.O. (2015). Food and Agricultural Organization. The State of Food and Agriculture.
- F.A.O. (2016). The State of World Fisheries and Aquaculture 2016. Contributing to food security and nutrition. Rome. 200p.p.
- Fajardo, P., Atanassova, M., Garrido-Maestu, A., Wortner-Smith, T., Cotterill, J., Cabado, A. G. (2014). Bacteria isolated from shellfish digestive gland with anti-pathogenic activity as candidates to increase the efficiency of shellfish depuration process. *Food Control*, 46, 272-281.
- FAO/WHO. (2001). Evaluation of health and nutritional properties of powder milk and live lactic acid bacteria. Food and Agriculture Organization of the United Nations and World Health Organization Expert Consultation Report. FAO, Rome, Italy.
- Farrow, J. A., & Collins, M. D. (1985). *Enterococcus hirae*, a new species that includes amino acid assay strain NCDO 1258 and strains causing growth depression in young chickens. *International journal of systematic bacteriology*, 35(1), 73-75.
- Farzanfar, A. (2006). The use of probiotics in shrimp aquaculture. *Pathogens and Disease*, 48(2), 149-158.
- Franz, C. M., Huch, M., Abriouel, H., Holzapfel, W., & Gálvez, A. (2011). Enterococci as probiotics and their implications in food safety. *International journal of food microbiology*, 151(2), 125-140.
- Fuller, R. (1989). Probiotics in man and animals. *Journal of applied bacteriology*, 66, 365-378.
- Gatesoupe, F. J. (1991). The effect of three strains of lactic bacteria on the production rate of rotifers, *Brachionus plicatilis*, and their dietary value for larval turbot, *Scophthalmus maximus*. *Aquaculture*, 96(3-4), 335-342.
- Gatesoupe, F. J. (1994). Lactic acid bacteria increase the resistance of turbot larvae, *Scophthalmus maximus*, against pathogenic *Vibrio*. *Aquatic living resources*, 7(4), 277-282.
- Gatesoupe, F. J. (1999). The use of probiotics in aquaculture. *Aquaculture*, 180(1), 147-165.
- Gatesoupe, F. J. (2007). Updating the importance of lactic acid bacteria in fish farming: natural occurrence and probiotic treatments. *Journal of molecular microbiology and biotechnology*, 14(1-3), 107-114.
- German, D. P., Nagle, B. C., Villeda, J. M., Ruiz, A. M., Thomson, A. W., Contreras Balderas, S., & Evans, D. H. (2009). Evolution of herbivory in a carnivorous

clade of minnows (*Teleostei: Cyprinidae*): effects on gut size and digestive physiology. *Physiological and Biochemical Zoology*, 83(1), 1-18.

- Ghosh, K., & Ray, A. K. (2011). Tannins in plant feed ingredients: Facts and Probable Consequences in Fish Nutrition. *Tannins: Types, Foods Containing, and Nutrition*, 265-280.
- Gildberg, A., Mikkelsen, H., Sandaker, E., & Ringø, E. (1997). Probiotic effect of lactic acid bacteria in the feed on growth and survival of fry of Atlantic cod (*Gadus morhua*). *Hydrobiologia*, 352(1), 279-285.
- Giri, S. S., Sukumaran, V., Sen, S. S., & Jena, P. K. (2014). Effects of dietary supplementation of potential probiotic *Bacillus subtilis* VSG1 singularly or in combination with *Lactobacillus plantarum* VSG3 or/and *Pseudomonas aeruginosa* VSG2 on the growth, immunity and disease resistance of Labeo rohita. *Aquaculture Nutrition*, 20(2), 163-171.
- Gismondo, M.R., Drago, L. & Lombardi, A. (1999). Review of probiotics available to modify gastrointestinal flora. *International Journal Antimicrobiology Agents*, 12, 287-292.
- Gómez, N. C., Ramiro, J. M., Quecan, B. X., & de Melo Franco, B. D. (2016). Use of potential probiotic lactic acid bacteria (LAB) biofilms for the control of *Listeria monocytogenes*, *Salmonella Typhimurium*, and *Escherichia coli* O157: H7 biofilms formation. *Frontiers in microbiology*, 7.
- Gomez-Gil, B., Roque, A. Velasco, G. (2002). Culture of the bacterial strain C7b, a potential probiotic bacterium, with the microalgae *Chaetoceros muelleri*. *Aquaculture*, 211, 43-48.
- Gómez-León, J., Villamil, L., Lemos, M. L., Novoa, B., & Figueras, A. (2005). Isolation of *Vibrio alginolyticus* and *Vibrio splendidus* from aquacultured carpet shell clam (*Ruditapes decussatus*) larvae associated with mass mortalities. *Applied and environmental microbiology*, 71(1), 98-104.
- Gonzalez, C. J., Encinas, J. P., Garcia-López, M. L., & Otero, A. (2000). Characterization and identification of lactic acid bacteria from freshwater fishes. *Food Microbiology*, 17(4), 383-391.
- Gopalakannan, A., & Rathnakumar, K. (2013). The Role of Probiotics in Aquaculture.
- Grisez, L., & Ollevier, F. (1995). *Vibrio (Listonella) anguillarum* infection in marine fish larviculture. *Larvi*, 91, 497.
- Grisez, L., Sorgeloos, P., & Ollevier, F. (1996). Mode of infection and spread of *Vibrio anguillarum* in turbot *Scophthalmus maximus* larvae after oral challenge through live feed. *Diseases of Aquatic Organisms*, 26(3), 181-187.
- Gulig, P. A., Bourdage, K. L., & Starks, A. M. (2005). Molecular pathogenesis of *Vibrio vulnificus*. *The Journal of Microbiology*, 43(1), 118-131.

- Hagi, T., Tanaka, D., Iwamura, Y., & Hoshino, T. (2004). Diversity and seasonal changes in lactic acid bacteria in the intestinal tract of cultured freshwater fish. *Aquaculture*, 234(1), 335-346.
- Hai, N. V. (2015). The use of probiotics in aquaculture. *Journal of applied microbiology*, 119(4), 917-935.
- Haq, M. B., Ali, H. A., Nazar, A. R., & Shalini, S. (2011). Assessment of *Artemia franciscana* as a probable vector for WSSV transmission to *Macrobrachium idella idella* (Hilgendorf 1898). *International Journal of Chemical and Analytical Science*, 2(9), 1159-1170.
- Halami, P. M., Chandrashekar, A., & Nand, K. (2000). *Lactobacillus farciminis* MD, a newer strain with potential for bacteriocin and antibiotic assay. *Letters in applied microbiology*, 30(3), 197-202.
- Harzevilli, A. R. S., Van Duffel, H., Defoort, T., Dhert, P. H., Sorgeloos, P., & Swings, J. (1997). The influence of a selected bacterial strain *Vibrio anguillarum* TR27 on the growth rate of rotifers in different culture conditions. *Aquacul Int*, 5, 183-188.
- Herrera, B., & Toranzo, A. E. (2005). Vaccination strategies to prevent emerging diseases for Spanish aquaculture. *Development Biology. Basel*, 121, 85-95.
- Holzapfel, W. H., & Wood, B. J. B. (1995). Lactic acid bacteria in contemporary perspective. In *The genera of lactic acid bacteria* (pp. 1-6). Springer, Boston, MA.
- Holzapfel, W. H., Geisen, R., & Schillinger, U. (1995). Biological preservation of foods with reference to protective cultures, bacteriocins and food-grade enzymes. *International journal of food microbiology*, 24(3), 343-362
- Hoppe, H.G. 1991. Microbial extracellular enzyme activity: a new key parameter in aquatic ecology. In *Microbial Enzymes in Aquatic Environments*, ed. R.J. Chrost, pp.60-83. New York: Springer-Verlag.
- Huang, Y., Zhang, L., Tiu, L., & Wang, H. H. (2015). Characterization of antibiotic resistance in commensal bacteria from an aquaculture ecosystem. *Frontiers in microbiology*, 6.
- Ibrahim, M. D. (2015). Evolution of probiotics in aquatic world: potential effects, the current status in Egypt and recent perspectives. *Journal of advanced Research*, 6(6), 765-791.
- Ikeogu, F. C., Nsofor, C. I., & Ikpeze, O. O. (2010). A review of risk factors for fish diseases in aquatic environments. Proceedings of the 6th National Conference of the Society for Occupational Safety and Environmental Health.
- Irianto, A., & Austin, B. (2002). Use of probiotics to control furunculosis in rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of fish diseases*, 25(6), 333-342.

- Irianto, A., Robertson, P. A. W., & Austin, B. (2000). The use of probiotics in aquaculture. *Recent research developments in microbiology*, 4(2), 557-567.
- Jöborn, A., Olsson, J. C., Westerdahl, A., Conway, P. L., & Kjelleberg, S. (1997). Colonization in the fish intestinal tract and production of inhibitory substances in intestinal mucus and faecal extracts by *Carnobacterium* sp. strain K1. *Journal of Fish Diseases*, 20(5), 383-392.
- Kennedy, S. B., Tucker, J. W., Thoresen, M., & Sennett, D. G. (1998). Current methodology for the use of probiotic bacteria in the culture of marine fish larvae. *Aquaculture*, 98, 286.
- Kesarcodi-Watson, A., Kaspar, H., Lategan, M. J., & Gibson, L. (2008). Probiotics in aquaculture: the need, principles and mechanisms of action and screening processes. *Aquaculture*, 274(1), 1-14.
- Khan, R. A. (2012). Host-parasite interactions in some fish species. *Journal of parasitology research*, 2012.
- Kim, D. H., & Austin, B. (2006). Innate immune responses in rainbow trout (*Oncorhynchus mykiss*, Walbaum) induced by probiotics. *Fish & shellfish immunology*, 21(5), 513-524.
- Kim, D. H., & Austin, B. (2008). Characterization of probiotic carnobacteria isolated from rainbow trout (*Oncorhynchus mykiss*) intestine. *Letters in applied microbiology*, 47(3), 141-147.
- Kim, D.H. & B, Austin. (2008). Innate immune responses in rainbow trout (*Oncorhynchus mykiss*, Walbaum) induced by probiotics. *Fish and Shellfish Immunology*, 21, 513-524.
- Klaenhammer, T. R., Barrangou, R., Buck, B. L., Azcarate-Peril, M. A., & Altermann, E. (2005). Genomic features of lactic acid bacteria effecting bio-processing and health. *FEMS Microbiology Reviews*, 29(3), 393-409.
- Klein, G. (2003). Taxonomy, ecology and antibiotic resistance of enterococci from food and the gastro-intestinal tract. *International journal of food microbiology*, 88(2), 123-131.
- Klewicki, R., & Klewicka, E. (2004). Antagonistic activity of lactic acid bacteria as probiotics against selected bacteria of the Enterobacteriaceae family in the presence of polyols and their galactosyl derivatives. *Biotechnology letters*, 26(4), 317-320.
- Kozasa, M. (1986). Toyocerin (*Bacillus toyoi*) as growth promoter for animal feeding. *Microbiol. Aliment. Nutrition*, 4, 121-135.
- Kristich, C. J., Li, Y. H., Cvitkovitch, D. G., & Dunny, G. M. (2004). Esp-independent biofilm formation by *Enterococcus faecalis*. *Journal of bacteriology*, 186(1), 154-163.

- Krkosek, M. (2009). Host density thresholds and disease control for fisheries and aquaculture. *Aquaculture Environment Interactions*, 1(1), 21-32.
- Kubota, H., Senda, S., Nomura, N., Tokuda, H., & Uchiyama, H. (2008). Biofilm formation by lactic acid bacteria and resistance to environmental stress. *Journal of bioscience and bioengineering*, 106(4), 381-386.
- Lafferty, K. D., Harvell, C. D., Conrad, J. M., Friedman, C. S., Kent, M. L., Kuris, A. M., & Saksida, S. M. (2015). Infectious diseases affect marine fisheries and aquaculture economics. *Annual review of marine science*, 7, 471-496.
- Lamari, F., Sadok, K., Bakhrouf, A., & Gatesoupe, F. J. (2014). Selection of lactic acid bacteria as candidate probiotics and *in vivo* test on *Artemia* nauplii. *Aquaculture international*, 22(2), 699-709.
- Lara-Flores, M., Aguirre-Guzman, G. (2009). The use of probiotic in fish and shrimp aquaculture. A review. In: N.P Guerra and L.P. Castro (Eds.) *Probiotics: Production, evaluation and uses in animal feed*. Research Signpost 37/661 (2), Fort P.O., Trivandrum-695 023, Kerala, India.
- Lara-Flores, M., Olvera-Novoa, M. A., Guzmán-Méndez, B. E., & López-Madrid, W. (2003). Use of the bacteria *Streptococcus faecium* and *Lactobacillus acidophilus*, and the yeast *Saccharomyces cerevisiae* as growth promoters in Nile tilapia (*Oreochromis niloticus*). *Aquaculture*, 216(1), 193-201.
- Lazado, C. C., Caipang, C. M. A., & Kiron, V. (2012). Enzymes from the gut bacteria of Atlantic cod, *Gadus morhua* and their influence on intestinal enzyme activity. *Aquaculture Nutrition*, 18(4), 423-431.
- Lazado, C. C., Lacsamana, J. I., & Caipang, C. M. (2015). 5. Mechanisms of probiotic actions in shrimp: Implications to tropical aquaculture. In *Biotechnological Advances in Shrimp Health Management in the Philippines* (pp. 89-114). Research Signpost.
- Lee Seong Wei & Wendy Wee. (2014). Diseases in Aquaculture. *Research. Journal. Animal. & Veterinar. Science.*, 7(1): 1-6.
- Lee, Y. K., & Salminen, S. (2009). *Handbook of probiotics and prebiotics*. John Wiley & Sons.
- Lee, Y. K., Lim, C. Y., Teng, W. L., Ouwehand, A. C., Tuomola, E. M., & Salminen, S. (2000). Quantitative approach in the study of adhesion of lactic acid bacteria to intestinal cells and their competition with enterobacteria. *Applied and environmental microbiology*, 66(9), 3692-3697.
- Lésel, R. (1990). Thermal effect on bacterial flora in the gut of rainbow trout and African catfish. *Microbiology in Poecilotherms*, 33-38.
- Liasi, S. A., Azmi, T. I., Hassan, M. D., Shuhaimi, M., Rosfarizan, M., & Ariff, A. B. (2009). Antimicrobial activity and antibiotic sensitivity of three isolates of lactic

- acid bacteria from fermented fish product, Budu. *Malaysian Journal of Microbiology*, 5(1), 33-37.
- Lowther, A. (2005). Highlights from the FAO Database on Aquaculture Production Statistics—Fishery Information, Data and Statistics Unit FAO Fisheries Department. *Food and Agriculture Organization of the United Nations, Rome*.
- Luna, S. (2008). *Lates calcarifer*, barramundi: fisheries, aquaculture, gamefish, aquarium:(on-line).
- Marques, A., François, J. M., Dhont, J., Bossier, P., & Sorgeloos, P. (2004). Influence of yeast quality on performance of gnotobiotically grown *Artemia*. *Journal of Experimental Marine Biology and Ecology*, 310(2), 247-264.
- Marshall, B. M., & Levy, S. B. (2011). Food animals and antimicrobials: impacts on human health. *Clinical microbiology reviews*, 24(4), 718-733.
- Martínez Cruz, P., Ibáñez, A. L., Monroy Hermosillo, O. A., & Ramírez Saad, H. C. (2012). Use of probiotics in aquaculture. *ISRN microbiology*, 2012.
- Mazurkiewicz, J., Przybyl, A., Sip, A., & Grajek, W. (2007). Effect of *Carnobacterium divergens* and *Enterococcus hirae* as probiotic bacteria in feed for common carp, *Cyprinus carpio* L. *Archiwum Rybactwa Polskiego*, 15(2), 93.
- Merrifield, D. L., Dimitroglou, A., Foey, A., Davies, S. J., Baker, R. T., Bøggwald, J. & Ringø, E. (2010). The current status and future focus of probiotic and prebiotic applications for salmonids. *Aquaculture*, 302(1), 1-18.
- Meyer, F. P. (1991). Aquaculture disease and health management. *Journal of animal science*, 69(10), 4201-4208.
- Miller, R. A. (2016). Antimicrobial susceptibility testing guidelines as a necessary tool to guide chemotherapeutic interventions in aquaculture. *Microbiology Australia*, 37(3), 104-107.
- Mohamed, J. A., & Huang, D. B. (2007). Biofilm formation by enterococci. *Journal of medical microbiology*, 56(12), 1581-1588.
- Mohapatra, S., Chakraborty, T., Prusty, A. K., Das, P., Paniprasad, K., & Mohanta, K. N. (2012). Use of different microbial probiotics in the diet of rohu, *Labeo rohita* fingerlings: effects on growth, nutrient digestibility and retention, digestive enzyme activities and intestinal microflora. *Aquaculture Nutrition*, 18(1), 1-11.
- Mohideen, M.M., Kader, T.S., Mohan, S.P., Mohamed and Hussain, M.I.Z.(2010). Effect of Probiotic Bacteria on the Growth rate of Fresh Water Fish, *Catla catla*. *International Journal of Biological Technology*. 1(2),113-117.
- Mondal, S., Roy, T., & Ray, A. K. (2010). Characterization and identification of enzyme- producing bacteria isolated from the digestive tract of bata, *Labeo bata*. *Journal of the World Aquaculture Society*, 41(3), 369-377.

- Montero, A. B., & Austin, B. (1999). Characterization of extracellular products from an isolate of *Vibrio harveyi* recovered from diseased post-larval *Penaeus vannamei* (Bonne). *Journal of Fish Diseases*, 22(5), 377-386.
- Moore, R. (1982). Spawning and early life history of burramundi, *Lates calcarifer* (Bloch), in Papua New Guinea. *Marine and Freshwater Research*, 33(4), 647-661.
- Moriarty, D. J. W. (1998). Control of luminous *Vibrio* species in penaeid aquaculture ponds. *Aquaculture*, 164(1), 351-358.
- Muñoz-Atienza, E., Gómez-Sala, B., Araújo, C., Campanero, C., Del Campo, R., Hernández, P. E., ... & Cintas, L. M. (2013). Antimicrobial activity, antibiotic susceptibility and virulence factors of lactic acid bacteria of aquatic origin intended for use as probiotics in aquaculture. *BMC microbiology*, 13(1), 15.
- Muthukumar, P., & Kandeepan, C. (2015). Isolation, identification and characterization of probiotic organisms from intestine of fresh water fishes. *International Journal Current Microbiology Applied Science*, 4, 607-616.
- Nadell, C. D., Xavier, J. B., & Foster, K. R. (2008). The sociobiology of biofilms. *FEMS microbiology reviews*, 33(1), 206-224.
- Nashima, K., Santhiya, P., & Palanisamy, A. (2012). Production and optimization of lipase from wild and mutant strains of *Bacillus* sp. and *Pseudomonas* sp. *Journal. Academic. Industrial. Reserch*, 1, 97-100.
- Nayak, S. K. (2010). Probiotics and immunity: a fish perspective. *Fish & shellfish immunology*, 29(1), 2-14.
- Newaj-Fyzul, A., Al-Harbi, A. H., & Austin, B. (2014). Developments in the use of probiotics for disease control in aquaculture. *Aquaculture*, 431, 1-11.
- Nikoskelainen, S., Ouwehand, A., Salminen, S., Bylund, G. (2001). Protection of rainbow trout (*Oncorhynchus mykiss*) from furunculosis by *Lactobacillus rhamnosus*. *Aquaculture*, 198 (3-4), 229-236.
- Ninawe, A. S., & Selvin, J. (2009). Probiotics in shrimp aquaculture: avenues and challenges. *Critical reviews in microbiology*, 35(1), 43-66.
- Noriega, L., Gueimonde, M., Sánchez, B., Margolles, A., & de los Reyes-Gavilán, C. G. (2004). Effect of the adaptation to high bile salts concentrations on glycosidic activity, survival at low pH and cross-resistance to bile salts in *Bifidobacterium*. *International journal of food microbiology*, 94(1), 79-86.
- Okaeme, A. N (2010). *Managing Aquatic Environment for best practices in Fish Farming*. Professional Continuing Education Manual. Veterinary Council of Nigeria (VCN). Abuja, Nigeria.
- Olafsen, J. A. (2001). Interactions between fish larvae and bacteria in marine aquaculture. *Aquaculture*, 200(1), 223-247.

- Pai, S. S. (2006). Biocontrol of *Vibrio harveyi* in *Penaeus monodon* larval rearing systems employing probiotics and vibriophages.
- Pan, X., Wu, T., Zhang, L., Song, Z., Tang, H., & Zhao, Z. (2008). *In vitro* evaluation on adherence and antimicrobial properties of a candidate probiotic *Clostridium butyricum* CB2 for farmed fish. *Journal of applied microbiology*, 105(5), 1623-1629.
- Pandiyan, P., Balaraman, D., Thirunavukkarasu, R., George, E. G. J., Subaramaniyan, K., Manikkam, S., & Sadayappan, B. (2013). Probiotics in aquaculture. *Drug Invention Today*, 5(1), 55-59.
- Patra, S. K., & Mohamed, K. S. (2003). Enrichment of *Artemia nauplii* with the probiotic yeast *Saccharomyces boulardii* and its resistance against a pathogenic *Vibrio*. *Aquaculture international*, 11(5), 505-514.
- Pfeiler, E. A., & Klaenhammer, T. R. (2007). The genomics of lactic acid bacteria. *Trends in microbiology*, 15(12), 546-553.
- Prieur, D., Mevel, G., Nicolas, J. L., Plusquellec, A., & Vigneulle, M. (1990). Interactions between bivalve molluscs and bacteria in the marine environment. *Oceanography. Marine. Biology. Annual. Review*, 28, 277-352.
- Qi, Z., Zhang, X. H., Boon, N., & Bossier, P. (2009). Probiotics in aquaculture of China—current state, problems and prospect. *Aquaculture*, 290(1), 15-21.
- Qian, P. Y., Lau, S. C., Dahms, H. U., Dobretsov, S., & Harder, T. (2007). Marine biofilms as mediators of colonization by marine macroorganisms: implications for antifouling and aquaculture. *Marine Biotechnology*, 9(4), 399-410.
- RabanaJ, H.R., and Soesanto. V. 1982. Introduction to the taxonomy, biology and fishery of the giant seaperch or sea-bass *Lates calcarifer*. In: Report of Training Course on Seabass Spawning and Larval Rearing, Songkhla, Thailand June 1982. FAO SCS/GEN/82/39. 105 p,
- Ransangan, J., & Manin, B. O. (2010). Mass mortality of hatchery-produced larvae of Asian seabass, *Lates calcarifer* (Bloch), associated with viral nervous necrosis in Sabah, Malaysia. *Veterinary microbiology*, 145(1), 153-157.
- Ransangan, J., & Mustafa, S. (2009). Identification of *Vibrio harveyi* isolated from diseased Asian Seabass *Lates calcarifer* by use of 16S ribosomal DNA sequencing. *Journal of aquatic animal health*, 21(3), 150-155.
- Ransangan, J., Lal, T. M., & Al-Harbi, A. H. (2012). Characterization and experimental infection of *Vibrio harveyi* isolated from diseased Asian seabass (*Lates calcarifer*). *Malaysian Journal of Microbiology*, 8(2), 104-115.
- Rawn, D. F., Krakalovich, T., Forsyth, D. S., & Roscoe, V. (2009). Analysis of fin and non- fin fish products for azamethiphos and dichlorvos residues from the Canadian retail market. *International journal of food science & technology*, 44(8), 1510-1516.

- Ray, A. K., Ghosh, K., & Ringø, E. (2012). Enzyme- producing bacteria isolated from fish gut: a review. *Aquaculture Nutrition*, 18(5), 465-492.
- Ray, A. K., Roy, T., Mondal, S., & Ringø, E. (2010). Identification of gut- associated amylase, cellulase and protease- producing bacteria in three species of Indian major carps. *Aquaculture Research*, 41(10), 1462-1469.
- Rengpipat, S., Rueangruklikhit, T., & Piyatiratitivorakul, S. (2008). Evaluations of lactic acid bacteria as probiotics for juvenile seabass *Lates calcarifer*. *Aquaculture Research*, 39(2), 134-143.
- Rengpipat, S., Rukpratanporn, S., Piyatiratitivorakul, S., & Menasaveta, P. (2000). Immunity enhancement in black tiger shrimp (*Penaeus monodon*) by a probiont bacterium (Bacillus S11). *Aquaculture*, 191(4), 271-288.
- Ringø, E. (1993). Does dietary linoleic acid affect intestinal microflora in Arctic charr, *Salvelinus alpinus* (L.). *Aquaculture Research*, 24(1), 133-135.
- Ringø, E., & Birkbeck, T. H. (1999). Intestinal microflora of fish larvae and fry. *Aquaculture research*, 30(2), 73-93.
- Ringø, E., & Gatesoupe, F. J. (1998). Lactic acid bacteria in fish: a review. *Aquaculture*, 160(3), 177-203.
- Ringø, E., & Olsen, R. E. (1999). The effect of diet on aerobic bacterial flora associated with intestine of Arctic charr (*Salvelinus alpinus* L.). *Journal of Applied Microbiology*, 86(1), 22-28.
- Ringø, E., Jutfelt, F., Kanapathipillai, P., Bakken, Y., Sundell, K., Glette, J., & Olsen, R. E. (2004). Damaging effect of the fish pathogen *Aeromonas salmonicida* spp. salmonicida on intestinal enterocytes of Atlantic salmon (*Salmo salar* L.). *Cell and tissue research*, 318(2), 305-311.
- Ringø, E., Løvmo, L., Kristiansen, M., Bakken, Y., Salinas, I., Myklebust, R., & Mayhew, T. M. (2010). Lactic acid bacteria vs. pathogens in the gastrointestinal tract of fish: a review. *Aquaculture Research*, 41(4), 451-467.
- Ringø, E., Schillinger, U., & Holzapfel, W. (2005). Antimicrobial activity of lactic acid bacteria isolated from aquatic animals and the use of lactic acid bacteria in aquaculture. *Biology of growing animals*, 2, 418-453.
- Ringø, E., Seppola, M., Berg, A., Olsen, R. E., Schillinger, U., & Holzapfel, W. (2002). Characterization of *Carnobacterium divergens* strain 6251 isolated from intestine of Arctic charr (*Salvelinus alpinus* L.). *Systematic and applied microbiology*, 25(1), 120-129.
- Ringø, E., Wesmajervi, M. S., Bendiksen, H. R., Berg, A., Olsen, R. E., Johnsen, T., & Holzapfel, W. (2001). Identification and characterization of carnobacteria isolated from fish intestine. *Systematic and applied microbiology*, 24(2), 183-191.

- Romero, J., & Navarrete, P. (2006). 16S rDNA-based analysis of dominant bacterial populations associated with early life stages of coho salmon (*Oncorhynchus kisutch*). *Microbial ecology*, 51(4), 422-430.
- Romero, J., Feijoó, C. G., & Navarrete, P. (2012). Antibiotics in aquaculture—use, abuse and alternatives. In *Health and environment in aquaculture*. InTech.
- Roy, T., Mondal, S., & Ray, A. K. (2009). Phytase- producing bacteria in the digestive tracts of some freshwater fish. *Aquaculture research*, 40(3), 344-353.
- Safari, R., Adel, M., Lazado, C. C., Caipang, C. M. A., & Dadar, M. (2016). Host-derived probiotics *Enterococcus casseliflavus* improves resistance against *Streptococcus iniae* infection in rainbow trout (*Oncorhynchus mykiss*) via immunomodulation. *Fish & shellfish immunology*, 52, 198-205.
- Saha, S., Roy, R. N., Sen, S. K., & Ray, A. K. (2006). Characterization of cellulase- producing bacteria from the digestive tract of tilapia, *Oreochromis mossambica* (Peters) and grass carp, *Ctenopharyngodon idella* (Valenciennes). *Aquaculture Research*, 37(4), 380-388.
- Sakai, M., Yoshida, T., Atsuta, S., & Kobayashi, M. (1995). Enhancement of resistance to vibriosis in rainbow trout, *Oncorhynchus mykiss* (Walbaum), by oral administration of *Clostridium butyricum* bacterin. *Journal of Fish Diseases*, 18(2), 187-190.
- Salinas, I., Cuesta, A., Esteban, M. Á., & Meseguer, J. (2005). Dietary administration of *Lactobacillus delbrückii* and *Bacillus subtilis*, single or combined, on gilthead seabream cellular innate immune responses. *Fish & shellfish immunology*, 19(1), 67-77.
- Salinas, I., Díaz-Rosales, P., Cuesta, A., Meseguer, J., Chabrillón, M., Morinigo, M. A., & Esteban, M. A. (2006). Effect of heat-inactivated fish and non-fish derived probiotics on the innate immune parameters of a teleost fish (*Sparus aurata* L.). *Veterinary immunology and immunopathology*, 111(3), 279-286.
- Samuelsen, O. B. (2006). Pharmacokinetics of quinolones in fish: a review. *Aquaculture*, 255(1), 55-75.
- Schembri, M. A., & Klemm, P. (2001). Biofilm formation in a hydrodynamic environment by novel FimH variants and ramifications for virulence. *Infection and immunity*, 69(3), 1322-1328.
- Schillinger, U. & Lücke, F. K. (1989). Antibacterial activity of *Lactobacillus sake* isolated from meat. *Applied and Environmental Microbiology*, 55(8), 1901-1906.
- Seenivasan, C., Bhavan, P. S., Radhakrishnan, S., & Shanthy, R. (2012). Enrichment of *Artemia* nauplii with *Lactobacillus sporogenes* for enhancing the survival, growth and levels of biochemical constituents in the post-larvae of the

freshwater prawn *Macrobrachium rosenbergii*. Turkish Journal of Fisheries and Aquatic Sciences, 12(1).

- Seppola, M., Olsen, R. E., Sandaker, E., Kanapathippillai, P., Holzapfel, W., & Ringø, E. (2006). Random amplification of polymorphic DNA (RAPD) typing of carnobacteria isolated from hindgut chamber and large intestine of Atlantic cod (*Gadus morhua* L.). *Systematic and applied microbiology*, 29(2), 131-137.
- Shaklee, J. B., & Salini, J. P. (1985). Genetic variation and population subdivision in Australian barramundi, *Lates calcarifer* (Bloch). *Marine and Freshwater Research*, 36(2), 203-218.
- Shariff, M., 1995. Fish health: an odyssey through the Asia-Pacific region. Syarahan inaugural. Universiti. Pertanian Malaysia, Serdang, Malaysia, pp: 25.
- Silva, J., Carvalho, A. S., Teixeira, P., & Gibbs, P. A. (2002). Bacteriocin production by spray- dried lactic acid bacteria. *Letters in Applied Microbiology*, 34(2), 77-81.
- Siti Nur, (2016). *Preliminary In vivo of potential strain G87 as probiotic against Vibrio harveyi infection in Artemia Nauplii and Asian seabass (Lates calcarifer) larvae*. Universiti Putra Malaysia.
- Skea, G. L., Mountfort, D. O., & Clements, K. D. (2007). Contrasting digestive strategies in four New Zealand herbivorous fishes as reflected by carbohydrate activity profiles. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 146(1), 63-70.
- Soltanian, S., Dhont, J., Sorgeloos, P., & Bossier, P. (2007). Influence of different yeast cell-wall mutants on performance and protection against pathogenic bacteria (*Vibrio campbellii*) in gnotobiotically-grown *Artemia*. *Fish & shellfish immunology*, 23(1), 141-153.
- Son, V. M., Chang, C. C., Wu, M. C., Guu, Y. K., Chiu, C. H., & Cheng, W. (2009). Dietary administration of the probiotic, *Lactobacillus plantarum*, enhanced the growth, innate immune responses, and disease resistance of the grouper *Epinephelus coioides*. *Fish & shellfish immunology*, 26(5), 691-698.
- Sorroza, L., Padilla, D., Acosta, F., Román, L., Grasso, V., Vega, J., & Real, F. (2012). Characterization of the probiotic strain *Vagococcus fluvialis* in the protection of European sea bass (*Dicentrarchus labrax*) against vibriosis by *Vibrio anguillarum*. *Veterinary microbiology*, 155(2), 369-373.
- Stiles, M. E., & Holzapfel, W. H. (1997). Lactic acid bacteria of foods and their current taxonomy. *International journal of food microbiology*, 36(1), 1-29.
- Sugita, H., Kawasaki, J., & Deguchi, Y. (1997). Production of amylase by the intestinal microflora in cultured freshwater fish. *Letters in Applied Microbiology*, 24(2), 105-108.

- Suji, H. A., Palevesam, T. A., Immanuel, G., & Raj, S. (2014). Effect of different growth parameters on chitinase enzyme activity. *African Journal of Biotechnology*, 13(23).
- Sun, Y. Z., Yang, H. L., Ma, R. L., Song, K., & Li, J. S. (2012). Effect of *Lactococcus lactis* and *Enterococcus faecium* on growth performance, digestive enzymes and immune response of grouper *Epinephelus coioides*. *Aquaculture Nutrition*, 18(3), 281-289.
- Sutherland, I. W. (2001). Biofilm exopolysaccharides: a strong and sticky framework. *Microbiology*, 147(1), 3-9.
- Suzer, C., Çoban, D., Kamaci, H. O., Saka, Ş., Firat, K., Otgucuoğlu, Ö., & Küçüksari, H. (2008). *Lactobacillus* spp. bacteria as probiotics in gilthead sea bream (*Sparus aurata*, L.) larvae: effects on growth performance and digestive enzyme activities. *Aquaculture*, 280(1), 140-145.
- Tannock, G. W. (1995). Microecology of the gastrointestinal tract in relation to lactic acid bacteria. *International Dairy Journal*, 5(8), 1059-1070.
- Temmerman, R., Pot, B., Huys, G., & Swings, J. (2003). Identification and antibiotic susceptibility of bacterial isolates from probiotic products. *International journal of food microbiology*, 81(1), 1-10.
- Tendencia, E. A. (2002). *Vibrio harveyi* isolated from cage-cultured seabass *Lates calcarifer* Bloch in the Philippines. *Aquaculture Research*, 33(6), 455-458.
- Timmermans LPM (1987). Early development and differentiation in fish. *Sarsia* 72: 331-339.
- Tinh, N. T. N., Dierckens, K., Sorgeloos, P., & Bossier, P. (2008). A review of the functionality of probiotics in the larviculture food chain. *Marine Biotechnology*, 10(1), 1-12.
- Toranzo, A. E., Devesa, S., Romalde, J. L., Lamas, J., Riaza, A., Leiro, J., & Barja, J. L. (1995). Efficacy of intraperitoneal and immersion vaccination against *Enterococcus* sp. infection in turbot. *Aquaculture*, 134(1), 17-27.
- Touraki, M., Karamanlidou, G., Karavida, P., & Chrysi, K. (2012). Evaluation of the probiotics *Bacillus subtilis* and *Lactobacillus plantarum* bioencapsulated in *Artemia nauplii* against vibriosis in European sea bass larvae (*Dicentrarchus labrax*, L.). *World Journal of Microbiology and Biotechnology*, 28(6), 2425-2433.
- Tovar-Ramirez, D., Infante, J. Z., Cahu, C., Gatesoupe, F. J., & Vázquez-Juárez, R. (2004). Influence of dietary live yeast on European sea bass (*Dicentrarchus labrax*) larval development. *Aquaculture*, 234(1), 415-427.
- Tuan, T. N., Duc, P. M., & Hatai, K. (2013). Overview of the use of probiotics in aquaculture. *International Journal of Research in Fisheries and Aquaculture*, 3(3), 89-97.

- Vadstein, O. (1997). The use of immunostimulation in marine larviculture: possibilities and challenges. *Aquaculture*, 155, 404-417.
- Van Hai, N., Fotedar, R., & Buller, N. (2007). Selection of probiotics by various inhibition test methods for use in the culture of western king prawns, *Penaeus latissulcatus* (Kishinouye). *Aquaculture*, 272(1), 231-239.
- Vaseeharan, B. A. R. P., & Ramasamy, P. (2003). Control of pathogenic *Vibrio* spp. by *Bacillus subtilis* BT23, a possible probiotic treatment for black tiger shrimp *Penaeus monodon*. *Letters in applied microbiology*, 36(2), 83-87.
- Vaughan, E. E., & Mollet, B. (1999). Functionality of probiotics and intestinal lactobacilli: light in the intestinal tract tunnel. *Current Opinion in Biotechnology*, 10(5), 505-510.
- Vendrell, D., Balcazar, J. L., de Blas, I., Ruiz-Zarzuola, I., Gironés, O., & Muzquiz, J. L. (2008). Protection of rainbow trout (*Oncorhynchus mykiss*) from lactococcosis by probiotic bacteria. *Comparative immunology, microbiology and infectious diseases*, 31(4), 337-345.
- Venkat, H. K., Sahu, N. P., & Jain, K. K. (2004). Effect of feeding *Lactobacillus*- based probiotics on the gut microflora, growth and survival of postlarvae of *Macrobrachium rosenbergii* (de Man). *Aquaculture Research*, 35(5), 501-507.
- Verschuere, L., Rombaut, G., Sorgeloos, P., & Verstraete, W. (2000). Probiotic bacteria as biological control agents in aquaculture. *Microbiology and molecular biology reviews*, 64(4), 655-671.
- Villamil, L., Figueras, A., Planas, M., & Novoa, B. (2003). Control of *Vibrio alginolyticus* in *Artemia* culture by treatment with bacterial probiotics. *Aquaculture*, 219(1), 43-56.
- Vine, N. G., Leukes, W. D., & Kaiser, H. (2006). Probiotics in marine larviculture. *FEMS microbiology reviews*, 30(3), 404-427.
- Vine, N. G., Leukes, W. D., Kaiser, H., Daya, S., Baxter, J., & Hecht, T. (2004). Competition for attachment of aquaculture candidate probiotic and pathogenic bacteria on fish intestinal mucus. *Journal of fish diseases*, 27(6), 319-326.
- Wang, X., Li, H., Zhang, X., Li, Y., Ji, W., & Xu, H. (1999). Microbial flora in the digestive tract of adult penaeid shrimp (*Penaeus chinensis*). *Journal of Ocean University of Qingdao*, 30(3), 493-498.
- Wang, Y. B., Tian, Z. Q., Yao, J. T., & Li, W. F. (2008). Effect of probiotics, *Enterococcus faecium*, on tilapia (*Oreochromis niloticus*) growth performance and immune response. *Aquaculture*, 277(3), 203-207.
- Watnick, P. I., Fullner, K. J., & Kolter, R. (1999). A role for the mannose-sensitive hemagglutinin in biofilm formation by *Vibrio cholerae* El Tor. *Journal of bacteriology*, 181(11), 3606-3609.

- Webster, C., C. Lim. 2002. *Nutrient Requirements and Feeding of Finfish for Aquaculture*. Wallingford, Oxon; New York, NY: CABI Publishing. Accessed November 13, 2017 at https://books.google.com.my/books?lr=&id=9Vohcd0ISJQC&redir_esc=y
- Wei, Q., 2002. Social and economic impacts of aquatic animal health problems in aquaculture in China, pp. 55–61. In: Arthur, J.R., Phillips, M.J., Subasinghe, R.P., Reantaso, M.B., MacRae, I.H. (Eds.). *Primary Aquatic Animal Health Care in Rural, Small-Scale, Aquaculture Development*.
- Wilkenfeld, J. S. (1992). Commercial hatchery status report: an industry panel viewpoint. In *Proceedings of the special session on shrimp farming* (pp. 71-86).
- Wuertz, S., Okabe, S., & Hausner, M. (2004). Microbial communities and their interactions in biofilm systems: an overview. *Water Science and Technology*, 49(11-12), 327-336.
- Wyban, J., & Sweeney, J. N. (1991). *Intensive shrimp production technology: the Oceanic Institute shrimp manual*. The Institute.
- Yip, E. S., Geszvain, K., DeLoney-Marino, C. R., & Visick, K. L. (2006). The symbiosis regulator RscS controls the *syp* gene locus, biofilm formation and symbiotic aggregation by *Vibrio fischeri*. *Molecular microbiology*, 62(6), 1586-1600.
- Yusoff, A. (2015). Status of resource management and aquaculture in Malaysia. In *Resource Enhancement and Sustainable Aquaculture Practices in Southeast Asia: Challenges in Responsible Production of Aquatic Species: Proceedings of the International Workshop on Resource Enhancement and Sustainable Aquaculture Practices in Southeast Asia 2014 (RESA)* (pp. 53-65). Aquaculture Department, Southeast Asian Fisheries Development Center.
- Zapata, A. A., & Lara-Flores, M. (2012). Antimicrobial activities of lactic acid bacteria strains isolated from Nile Tilapia intestine (*Oreochromis niloticus*). *Journal of Biology and Life Science*, 4(1).
- Zhao, W. (2015). *Characterization of the probiotic mechanism of Phaeobacter gallaeciensis S4 against bacterial pathogens*. University of Rhode Island.
- Zhou, J. S., Pillidge, C. J., Gopal, P. K., & Gill, H. S. (2005). Antibiotic susceptibility profiles of new probiotic Lactobacillus and Bifidobacterium strains. *International journal of food microbiology*, 98(2), 211-217.
- Zorilla, I., Chabrillon, M., Arijo, S., Diaz-Rosales, P., Martinez-Manzanares, P., Balebona, M.C & Morinigo, M.A. (2003). Bacteria recovered from diseased cultured gilthead sea bream (*Sparus auratus* L.) in Southwestern Spain. *Aquaculture*, 218, 11-20.

Zorriehzahra, M. J., Delshad, S. T., Adel, M., Tiwari, R., Karthik, K., Dhama, K., & Lazado, C. C. (2016). Probiotics as beneficial microbes in aquaculture: An update on their multiple modes of action: A review. *Veterinary Quarterly*, 36(4), 228-241.

