



UNIVERSITI PUTRA MALAYSIA

***CHARACTERISATION OF RED PALM OIL MICROCAPSULE WITH
SOLUTION-ENHANCED DISPERSION BY SUPERCRITICAL CARBON
DIOXIDE TECHNOLOGY***

LEE WAN JUN

FSTM 2018 31



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By

LEE WAN JUN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

April 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

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Chairman: Associate Professor Chong Gun Hean, PhD
Faculty: Food Science and Technology

The carotenes and vitamin E (vE) in red palm oil (RPO) are highly susceptible to degradation which limits the utilisation of RPO as functional ingredient. The objective of this research was to enhance the stability of RPO and to slow down the degradation of bioactive compounds by microencapsulation using the solution-enhanced dispersion by supercritical carbon dioxide (SEDS) technology without the aid of organic solvents. This study began with the determination of RPO solubility and the concentration of carotenes, tocopherols and tocotrienols from the solubilised RPO in supercritical carbon dioxide (scCO₂) using the dynamic flow-through method, at different pressures (8.5– 25.0 MPa) and temperatures (40– 60°C), at a pre-determined scCO₂ flow rate of 2.9 g/min. The RPO had solubility of 0.5– 11.3 mg/g CO₂ and was significantly affected by pressure and temperature. The β -carotene recorded higher concentration in scCO₂ ($3 \times 10^{-3} \mu\text{mol/mol}$ to $17 \times 10^{-3} \mu\text{mol/mol}$) compared to α -carotene ($2 \times 10^{-3} \mu\text{mol/mol}$ to $15 \times 10^{-3} \mu\text{mol/mol}$); and tocotrienols were more soluble ($0.4 \mu\text{mol/mol}$ to $26 \mu\text{mol/mol}$) than tocopherols ($5 \times 10^{-2} \mu\text{mol/mol}$ to $3 \mu\text{mol/mol}$) following the preference of $\gamma\text{-T}_3 > \delta\text{-T}_3 > \alpha\text{-T}_3 > \alpha\text{-T} > \beta\text{-T} > \gamma\text{-T} > \delta\text{-T}$. Adachi-Lu model was the best fitting model for the solubility of RPO and the concentration of bioactive compounds in scCO₂. The RPO was then microencapsulated with SEDS at 10.0– 15.0 MPa, 40– 60°C, and feed injection flow rate 2.5 mL/min. The microcapsules were characterised in terms of their physical, structural, and thermal properties and also their oxidative stability. Microcapsules with microencapsulation efficiencies (ME) of oil (64– 92%), retention efficiencies (RE) of carotenes (50– 82%), RE of vE (64– 94%), particle size (4– 9 μm) were obtained. Comparing to the SEDS microcapsules (SEDS-M), the spray dried microcapsules (SD-M) as the control had wrinkled surfaces with lower ME (79%), similar RE (carotene= 85%, vE= 93%), larger particle size (17 μm) and larger particle size distribution. The fatty acid composition, chemical structures and thermal properties of the oil were not altered by the SEDS process. The thermal stability, oxidative stability and retention of carotenes and vE of RPO were improved after SEDS encapsulation. Colour change and degradation in carotenes and vE concentration for microcapsules and bulk oil at different storage temperatures (25°C, 45°C, 65°C, and 85°C) for up to 28 weeks was investigated and the degradation

kinetics using rate law and Arrhenius equation were studied. The degradation of carotenes and vE was well fitted into the first-order kinetic model. A negative correlation was found with the change in colour parameters to the degradation of carotenes, and two equations to correlate the relationship were established, recording R^2 of 0.8152 for SEDS-M and 0.8283 for SD-M. In summary, the SEDS process was able to produce RPO microcapsules containing high concentration of carotenes and vE, high oil encapsulation efficiency whilst able to provide protection against the deterioration of the bioactive components and oil quality.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN MIKROKAPSUL MINYAK SAWIT MERAH DENGAN
TEKNOLOGI PERTINGKATAN PENYEBARAN LARUTAN DENGAN
KARBON DIOKSIDA SUPERKRITIKAL**

Oleh

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Karoten dan vitamin E (vE) dalam minyak sawit merah (RPO) sangat mudah terdedah kepada kemerosotan yang menghalang penggunaan RPO sebagai bahan berfungsi. Objektif penyelidikan ini adalah untuk meningkatkan kestabilan RPO dan melambatkan kemerosotan komponen bioaktif dengan kaedah pengkapsulan menggunakan penyebaran larutan dengan teknologi superkritikal karbon dioksida (SEDS) tanpa bantuan pelarut organik. Kajian ini bermula dengan penentuan kelarutan RPO dan kepekatan karoten, tocopherols dan tocotrienols dari RPO yang terlarut dalam karbon dioksida superkritikal (scCO₂) menggunakan kaedah aliran dinamik, pada tekanan (8.5– 25.0 MPa) dan suhu (40– 60°C), pada kadar arus scCO₂ 2.9 g/min. RPO mempunyai kelarutan 0.5–11.3 mg/g CO₂ dan terjejas dengan ketara oleh tekanan dan suhu. β -karoten menghasilkan kepekatan yang lebih tinggi dalam scCO₂ (3×10^{-3} $\mu\text{mol/mol}$ hingga 17×10^{-3} $\mu\text{mol/mol}$) berbanding dengan α -karoten (2×10^{-3} $\mu\text{mol/mol}$ hingga 15×10^{-3} $\mu\text{mol/mol}$); dan tocotrienols mempunyai larut lebih tinggi (0.4 $\mu\text{mol/mol}$ hingga 26 $\mu\text{mol/mol}$) daripada tocopherols (5×10^{-2} $\mu\text{mol/mol}$ hingga 3 $\mu\text{mol/mol}$) mengikut keutamaan $\gamma\text{-T}_3 > \delta\text{-T}_3 > \alpha\text{-T}_3 > \alpha\text{-T} > \beta\text{-T} > \gamma\text{-T} > \delta\text{-T}$. Model Adachi-Lu adalah model terbaik untuk kelarutan RPO dan kepekatan komponen bioaktif dalam scCO₂. RPO itu kemudiannya dikapsulkan dengan SEDS pada 10.0–15.0 MPa, 40–60°C, dan kadar aliran sampel pada 2.5 mL/min. Mikrokapsul dicirikan dari segi sifat fizikal, struktur, dan terma dan juga kestabilan oksidatifnya. Mikrokapsul dengan kecekapan mikroencapsulasi (ME) minyak (64– 92%), kecekapan pengekalan (RE) karotene (50– 82%), RE vE (64–94%), saiz zarah (4–9 μm) telah diperolehi. Berbanding dengan mikrokapsul SEDS (SEDS-M), mikrokapsul kering semburan (SD-M) sebagai sampel kawalan mempunyai permukaan yang berkedut, dengan ME yang lebih rendah (79%), RE (karotena= 85%, vE= 93%), saiz (17 μm) dan taburan saiz zarah yang besar. Komposisi asid lemak, struktur kimia dan sifat termal minyak tidak diubah oleh proses SEDS. Kestabilan haba, kestabilan oksidatif dan pengekalan karoten dan vE RPO telah diperbaiki selepas pengkapsulan SEDS. Perubahan warna dan degradasi dalam karoten dan kepekatan vE untuk mikrokapsul dan minyak pada suhu penyimpanan yang berlainan (25°C, 45°C, 65°C, dan 85°C) selama 28 minggu serta kinetik degradasi dikaji menggunakan persamaan Arrhenius. Degradasi karoten

dan vE telah dikaitkan dengan baik dengan model kinetik pertama. Satu korelasi negatif didapati dengan perubahan parameter warna dengan kerosotan karoten, dan dua persamaan untuk mengaitkan hubungan telah ditubuhkan, merakamkan R^2 0.8152 untuk SEDS-M dan 0.8283 untuk SD-M. Secara ringkasnya, proses SEDS dapat menghasilkan mikrokapsul RPO yang mengandungi kepekatan karoten dan vE yang tinggi, kecekapan enkapsulasi minyak tinggi dan dapat memberikan perlindungan terhadap kerosotan komponen bioaktif dan kualiti minyak.



ACKNOWLEDGEMENTS

The thesis was completed with the kind support and help from many individuals and therefore, I would like to extend my sincere appreciation to all of them.

Foremost, I would like to express my sincere gratitude to my supervisor, Associate Professor Dr. Chong Gun Hean for his patience, motivation and willingness to impart his knowledge and expertise for me to complete this research.

Also, I would like to thank the supervisory committee members, Professor Tan Chin Ping and Dr. Rabiha Sulaiman for their insightful comments and help to improve this research.

I thank all the staffs from Faculty of Food Science and Technology and the Supercritical Fluid Centre, UPM for their assistance on using the laboratory equipment.

I thank my fellow labmates and friends for their insights. In particular, I am grateful to Dr. Hee Yen Yi for her encouragement and valuable suggestions for me to complete this work.

Last but not the least; I would like to thank my family; especially to my mother, my brothers and Chow Chang Yik for their unconditional love and support.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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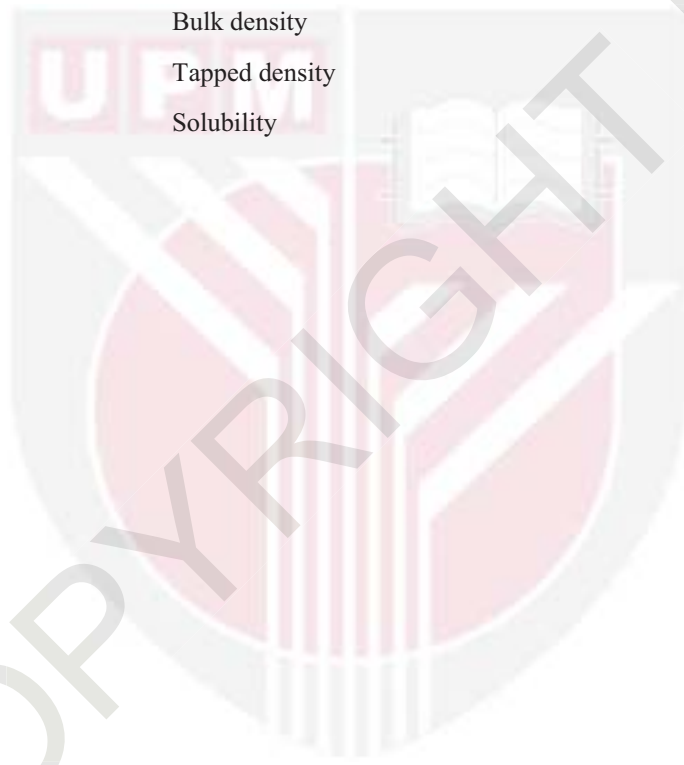
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LIST OF ABBREVIATIONS

AARD	Average absolute relative deviation
CO ₂	Carbon dioxide
C	Concentration per amount of CO ₂
C	Chroma
DTG	Derivative thermogravimetric
DE	Dextrose equivalent
DSC	Differential scanning calorimetry
FTIR	Fourier transform infrared spectroscopy
GC-FID	Gas chromatography-flame ionization detector
HPLC	High performance liquid chromatography
MC	Moisture content
MD	Maltodextrin
ME	Microencapsulation efficiency
MUFA	Monounsaturated fatty acid
NaCas	Sodium caseinate
o/w	Oil-in-water
PKO	Palm kernel oil
PO	Palm oil
PPO	Palm-pressed mesocarp fibre oil
PS	Particle size
PSD	Particle size distribution
PUFA	Polyunsaturated fatty acid
PV	Peroxide value
RE	Retention efficiency
RPO	Red palm oil
scCO ₂	Supercritical carbon dioxide
SD	Spray drying
SD-M	Spray dried microcapsules
SEDS	Solution-enhanced dispersion by supercritical carbon dioxide

SEDS-M	Microcapsules produced from the SEDS process
SFA	Saturated fatty acid
TGA	Thermogravimetric analysis
Totox	Total oxidation value
UFA	Unsaturated fatty acid
UV	Ultraviolet
vE	Vitamin E
<i>p</i> -AV	<i>p</i> -anisidine value
ρ_B	Bulk density
ρ_T	Tapped density
y_2	Solubility



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CHAPTER 1

INTRODUCTION

1.1 Background

Palm oil no longer remains anonymous to the people around the globe as it is widely used in the food industry as well as in the oleochemical industry. Malaysia is currently the second largest palm oil producer and exporter, accounting for approximately 30% and 37%, respectively, with recorded export revenue of RM 64.58 billion as of year 2016 (Malaysian Palm Oil Board, 2017). Red palm oil (RPO) is well known for its high concentration of bioactive components such as carotenes and vitamin E (vE). These bioactive components are reported to provide beneficial health effects because of their antioxidant properties (Amorim-Carrilho et al., 2014; Seppanen et al., 2010).

One of the problem that RPO faces is the susceptibility of the bioactive compounds such as carotenes and vE towards degradation when exposed to the environment, especially to heat (Cardoso et al., 2015; Zeb and Murkovic, 2013). The sensitivity of carotenes and vE had limited the application of RPO as functional ingredients. Even at temperature below 100°C, the degradation of carotenes and vitamin E is evident. The exposure of carotenes to heat at 37°C for 72 hours caused 64– 68% of β -carotene and 36– 50% of α -carotene to degrade whereas at 60°C, 90– 95% of β -carotene and 85– 90% of α -carotene were loss (Anguelova and Warthesen, 2000). As for the degradation of vE, Sabliov et al. (2009) reported the degradation of α -tocopherol at temperature as low as 40°C and Nissiotis and Tasioula-Margari (2002) reported a complete degradation of α -tocopherol in virgin olive oil stored at 60°C for 30 days. The deterioration of these bioactive components not only cause the nutritional value in the RPO to be lowered, but also bring about quality changes such as lipid peroxidation, discolouration and off flavours (Soukoulis and Bohn, 2015). These issues have created limitations to the wide applications of RPO and this setback could be solved by protecting the RPO with a layer of solid matrix using the microencapsulation technology.

Different encapsulation techniques have been explored by researchers to microencapsulate different types of oil. For instance, spray drying (Binsi et al., 2017; Tolve et al., 2017), freeze drying (Fioramonti et al., 2017; Gonzalez et al., 2016), co-extrusion (Chew and Nyam, 2016), complex coacervation (Timilsena et al., 2016) and fluidised-bed agglomeration (Fuchs et al., 2006). Several methods have been used to microencapsulate palm oil, such as spray drying (Ferreira et al., 2016; Kelly et al., 2014) and complex coacervation (Rutz et al., 2017). However, there are certain drawbacks with the aforementioned encapsulation technologies. The spray-dried microcapsules are reported to have wide particle size distribution and have wrinkled surface morphology (Domian et al., 2017; Umesha et al., 2013). Besides, the high operating temperature needed for drying could potentially deteriorate the bioactive components and also, the presence of oxygen could lead to higher oxidation in oil (Ferreira et al., 2016; Heinzlmann et al., 2000). As for the complex coacervation microencapsulation, it is difficult to optimise the process because there are numerous factors and parameters to

be considered (Siow, 2012). Microencapsulation with supercritical fluid (SCF) technology, the solution-enhanced dispersion by supercritical carbon dioxide (SEDS) technology is therefore introduced as an alternative to overcome the shortcomings of the aforementioned technologies. The SEDS particle formation could be carried out at mild temperatures, attributed to the low critical pressure and low critical temperature of the carbon dioxide. Hence, the degradation of thermolabile components could be potentially lowered (Nunes and Duarte, 2011). Besides, the process parameters such as pressure, temperature, feed and supercritical carbon dioxide (scCO₂) flow rate could be easily adjusted to produce microcapsules with desired properties (Chen et al., 2012; Yan et al., 2015; Zhao et al., 2015). Particle formation or encapsulation of different food bioactive components has been carried out using the SEDS technology (Aguilar et al., 2016; Machado et al., 2014; Zhao et al., 2015), however, the encapsulation of edible oil using the SEDS technology is lacking. Also, the reported SEDS process required the use of organic solvent which is compatible with food products (ethanol) and or incompatible (dimethyl sulfoxide, dichloromethane and acetone) to aid in the encapsulation process. The organic solvents were needed to increase the solubility of the solute (Chemat et al., 2017) with their traits of having high solubility and high expansion rate in the scCO₂. The use of water as the sole solvent for SEDS process has not been reported.

Hence, this work contributes by introducing the use of SEDS as an alternating technique to microencapsulate RPO without the aid from organic solvents. The SEDS microencapsulation technology is proposed to produce microcapsules with comparable or even better characteristics compared to the conventional technique (spray drying).

1.2 Objective

The main objective of this research was to enhance the oxidative stability of RPO and to slow down the degradation of bioactive components in RPO by microencapsulation using the SEDS technology without the aid of organic solvents. Emphasis was placed on studying the physical, chemical, thermal and storage properties of the RPO microcapsules produced.

The specific objectives of this study are:

- i. To determine the solubility of RPO in supercritical carbon dioxide and the concentration of carotenes (α -, and β -carotene) and vE (tocopherols and tocotrienols) in the solubilised RPO per amount of carbon dioxide and to correlate the solubility data with mathematical models.
- ii. To investigate the effect of SEDS operating pressure and temperature on the characteristics of RPO microcapsules.
- iii. To determine the chemical structure, thermal properties, thermal stability and oxidative stability of RPO microcapsules produced from SEDS.
- iv. To study degradation kinetics of carotenes and vitamin E and colour change in RPO, SEDS-M and SD-M.

REFERENCES

- Achir, N., Randrianatoandro, V.A., Bohuon, P., Laffargue, A., Avallone, S., 2010. Kinetic study of β -carotene and lutein degradation in oils during heat treatment. *Eur. J. Lipid Sci. Technol.* 112, 349–361.
- Adachi, Y., Lu, B.C.-Y., 1983. Supercritical fluid extraction with carbon dioxide and ethylene. *Fluid Phase Equilib.* 14, 147–156.
- Aghbashlo, M., Mobli, H., Madadlou, A., Rafiee, S., 2013. Influence of wall material and inlet drying air temperature on the microencapsulation of fish oil by spray drying. *Food Bioprocess Technol.* 6, 1561–1569.
- Aghbashlo, M., Mobli, H., Madadlou, A., Rafiee, S., 2012. The correlation of wall material composition with flow characteristics and encapsulation behavior of fish oil emulsion. *Food Res. Int.* 49, 379–388.
- Aguiar, G.P.S., Boschetto, D.L., Chaves, L.M.P.C., Arcari, B.D., Piato, A.L., Oliveira, J.V., Lanza, M., 2016. Trans-resveratrol micronization by SEDS technique. *Ind. Crops Prod.* 89, 350–355.
- Al-Darmaki, N., Lu, T., Al-Duri, B., Harris, J.B., Favre, T.L.F., Bhaggan, K., Santos, R.C.D., 2011. Solubility measurements and analysis of binary, ternary and quaternary systems of palm olein, squalene and oleic acid in supercritical carbon dioxide. *Sep. Purif. Technol.* 83, 189–195.
- Al-Saqer, J.M., Sidhu, J.S., Al-Hooti, S.N., Al-Amiri, H.A., Al-Othman, A., Al-Haji, L., Ahmed, N., Mansour, I.B., Minal, J., 2004. Developing functional foods using red palm olein. IV. Tocopherols and tocotrienols. *Food Chem.* 85, 579–583.
- Albanese, D., Adiletta, G., D'Acunto, M., Cinquanta, L., Di Matteo, M., 2014. Tomato peel drying and carotenoids stability of the extracts. *Int. J. Food Sci. Technol.* 49, 2458–2463.
- Amorim-Carrilho, K.T., Cepeda, A., Fente, C., Regal, P., 2014. Review of methods for analysis of carotenoids. *Trends Anal. Chem.* 56, 49–73.
- Andreu-Sevilla, A.J., Hartmann, A., Burlo, F., Poquet, N., Carbonell-Barrachina, A.A., 2009. Health benefits of using red palm oil in deep-frying potatoes: Low Acrolein emissions and high intake of carotenoids. *Food Sci. Technol. Int.* 15, 15–22.
- Anguelova, T., Warthesen, J., 2000. Degradation of lycopene, α -carotene, and β -carotene during lipid peroxidation. *J. Food Sci.* 65, 71–75.
- Anwar, S.H., Kunz, B., 2011. The influence of drying methods on the stabilization of fish oil microcapsules: Comparison of spray granulation, spray drying, and freeze drying. *J. Food Eng.* 105, 367–378.
- Aparicio-Ruiz, R., Mínguez-Mosquera, M.I., Gandul-Rojas, B., 2011. Thermal

- degradation kinetics of lutein, β -carotene and β -cryptoxanthin in virgin olive oils. *J. Food Compos. Anal.* 24, 811–820.
- Araus, K.A., Canales, R.I., del Valle, J.M., de la Fuente, J.C., 2011. Solubility of β -carotene in ethanol- and triolein-modified CO₂. *J. Chem. Thermodyn.* 43, 1991–2001.
- Ayeleso, A., Brooks, N., Oguntibeju, O., 2014. Modulation of antioxidant status in streptozotocin-induced diabetic male wistar rats following intake of red palm oil and/or rooibos. *Asian Pac. J. Trop. Med.* 7, 536–544.
- Bahrami, M., Ranjbarian, S., 2007. Production of micro- and nano-composite particles by supercritical carbon dioxide. *J. Supercrit. Fluids* 40, 263–283.
- Bakry, A.M., Abbas, S., Ali, B., Majeed, H., Abouelwafa, M.Y., Mousa, A., Liang, L., 2016. Microencapsulation of oils: A comprehensive review of benefits, techniques, and applications. *Compr. Rev. Food Sci. Food Saf.* 15, 143–182.
- Bartle, K.D., Clifford, A.A., Jafar, S.A., Shilstone, G.F., 1991. Solubilities of solids and liquids of low volatility in supercritical carbon dioxide. *J. Phys. Chem. Ref. Data* 20, 713–756.
- Baseri, S., Karimi, M., Morshed, M., 2012. Study of structural changes and mesomorphic transitions of oriented poly(ethylene terephthalate) fibers in supercritical CO₂. *Eur. Polym. J.* 48, 811–820.
- Beta, T., Hwang, T., 2018. Influence of heat and moisture treatment on carotenoids, phenolic content, and antioxidant capacity of orange maize flour. *Food Chem.* 246, 58–64.
- Bezerra, F.M., Carmona, O.G., Carmona, C.G., Lis, M.J., de Moraes, F.F., 2016. Controlled release of microencapsulated citronella essential oil on cotton and polyester matrices. *Cellulose* 23, 1459–1470.
- Binsi, P.K., Nayak, N., Sarkar, P.C., Jeyakumari, A., Muhamed Ashraf, P., Ninan, G., Ravishankar, C.N., 2017. Structural and oxidative stabilization of spray dried fish oil microencapsulates with gum arabic and sage polyphenols: Characterization and release kinetics. *Food Chem.* 219, 158–168.
- Birtigh, A., Johannsen, M., Brunner, G., Nair, N., 1995. Supercritical-fluid extraction of oil-palm components. *J. Supercrit. Fluids* 8, 46–50.
- Boon, C.S., McClements, D.J., Weiss, J., Decker, E.A., 2010. Factors influencing the chemical stability of carotenoids in foods. *Crit. Rev. Food Sci. Nutr.* 50, 515–532.
- Boschetto, D.L., Aranha, E.M., de Souza, A.A.U., Souza, S.M.A.G.U., Ferreira, S.R.S., Priamo, W.L., Oliveira, J.V., 2014. Encapsulation of bixin in PHBV using SEDS technique and in vitro release evaluation. *Ind. Crop. Prod.* 60, 22–29.
- Boschetto, D.L., Dalmolin, I., de Cesaro, A.M., Rigo, A.A., Ferreira, S.R.S., Meireles, M.A.A., Batista, E.A.C., Oliveira, J.V., 2013. Phase behavior and process

- parameters effect on grape seed extract encapsulation by SEDS technique. *Ind. Crop. Prod.* 50, 352–360.
- Botrel, D.A., Borges, S.V., Yoshida, M.I., Feitosa, J.P. de A., Fernandes, R.V. de B., de Souza, H.J.B., de Paula, R.C.M., 2017. Properties of spray-dried fish oil with different carbohydrates as carriers. *J. Food Sci. Technol.* 1–8.
- Boyère, C., Jérôme, C., Debuigne, A., 2014. Input of supercritical carbon dioxide to polymer synthesis: An overview. *Eur. Polym. J.* 61, 45–63.
- Bramley, P.M., Elmadfa, I., Kafatos, A., Kelly, F.J., Manios, Y., Roxborough, H.E., Schuch, W., Sheehy, P.J.A., Wagner, K.-H., 2000. Vitamin E. *J. Sci. Food Agric.* 80, 913–938.
- Brandt, L., Elizalde-Solis, O., Galicia-Luna, L.A., Gmehling, J., 2010. Solubility and density measurements of palmitic acid in supercritical carbon dioxide+alcohol mixtures. *Fluid Phase Equilib.* 289, 72–79.
- Brunner, G., 2005. Supercritical fluids: Technology and application to food processing. *J. Food Eng.* 67, 21–33.
- Bustos C, R.O., Alberti R, F. V., Matiacevich, S.B., 2016. Edible antimicrobial films based on microencapsulated lemongrass oil. *J. Food Sci. Technol.* 53, 832–839.
- Butstraen, C., Salaün, F., Devaux, E., 2015. Sol – gel microencapsulation of oil phase with Pickering and nonionic surfactant based emulsions. *Powder Technol.* 284, 237–244.
- Calvo, P., Castaño, Á.L., Lozano, M., González-Gómez, D., 2012. Influence of the microencapsulation on the quality parameters and shelf-life of extra-virgin olive oil encapsulated in the presence of BHT and different capsule wall components. *Food Res. Int.* 45, 256–261.
- Calvo, P., Hernández, T., Lozano, M., González-Gómez, D., 2010. Microencapsulation of extra-virgin olive oil by spray-drying: Influence of wall material and olive quality. *Eur. J. Lipid Sci. Technol.* 112, 852–858.
- Cao, W.-J., Ou, S.-Y., Lin, W.-F., Tang, C.-H., 2016. Food protein-based phytosterol nanoparticles: fabrication and characterization. *Food Funct.* 7, 3973–3980.
- Cardoso, L.D.M., Pinheiro, S.S., Da Silva, L.L., De Menezes, C.B., De Carvalho, C.W.P., Tardin, F.D., Queiroz, V.A.V., Martino, H.S.D., Pinheiro-Sant’Ana, H.M., 2015. Tocochromanols and carotenoids in sorghum (*Sorghum bicolor* L.): Diversity and stability to the heat treatment. *Food Chem.* 172, 900–908.
- Carneiro, H.C.F., Tonon, R. V., Grosso, C.R.F., Hubinger, M.D., 2013. Encapsulation efficiency and oxidative stability of flaxseed oil microencapsulated by spray drying using different combinations of wall materials. *J. Food Eng.* 115, 443–451.
- Çaylı, G., Küsefoğlu, S., 2008. Increased yields in biodiesel production from used cooking oils by a two step process: Comparison with one step process by using

TGA. Fuel Process. Technol. 89, 118–122.

- Chandrakant, K.K., Pravin, D.J., Bharat, H.S., Sachin, K., Amol, K.P., 2011. An overview on supercritical fluid extraction for herbal drugs. *Pharmacologyonline* 2, 575–596.
- Chang, F., Jin, J., Zhang, N., Wang, G., Yang, H.-J., 2012. The effect of the end group, molecular weight and size on the solubility of compounds in supercritical carbon dioxide. *Fluid Phase Equilib.* 317, 36–42.
- Chemat, F., Rombaut, N., Meullemiestre, A., Turk, M., Perino, S., Fabiano-Tixier, A.-S., Abert-Vian, M., 2017. Review of green food processing techniques. Preservation, transformation, and extraction. *Innov. Food Sci. Emerg. Technol.* 41, 357–377.
- Chen, A.-Z., Li, L., Wang, S.-B., Zhao, C., Liu, Y.-G., Wang, G.-Y., Zhao, Z., 2012. Nanonization of methotrexate by solution-enhanced dispersion by supercritical CO₂. *J. Supercrit. Fluids* 67, 7–13.
- Chen, A.Z., Li, Y., Chau, F.T., Lau, T.Y., Hu, J.Y., Zhao, Z., Mok, D.K. wah, 2009. Application of organic nonsolvent in the process of solution-enhanced dispersion by supercritical CO₂ to prepare puerarin fine particles. *J. Supercrit. Fluids* 49, 394–402.
- Chen, C.-C., Chang, C.J., Yang, P., 2000. Vapor–liquid equilibria of carbon dioxide with linoleic acid, α -tocopherol and triolein at elevated pressures. *Fluid Phase Equilib.* 175, 107–115.
- Chen, F., Yin, G., Liao, X., Yang, Y., Huang, Z., Gu, J., Yao, Y., Chen, X., Gao, H., 2013. Preparation, characterization and in vitro release properties of morphine-loaded PLLA-PEG-PLLA microparticles via solution enhanced dispersion by supercritical fluids. *J. Mater. Sci. Mater. Med.* 24, 1693–1705.
- Chen, H., Zhong, Q., 2015. Thermal and UV stability of β -carotene dissolved in peppermint oil microemulsified by sunflower lecithin and Tween 20 blend. *Food Chem.* 174, 630–636.
- Chen, P.-Y., Chen, W.-H., Lai, S.-M., Chang, C.-M.J., 2011. Solubility of *Jatropha* and *Aquilaria* oils in supercritical carbon dioxide at elevated pressures. *J. Supercrit. Fluids* 55, 893–897.
- Chen, W., Wang, H., Zhang, K., Gao, F., Chen, S., Li, D., 2016. Physicochemical properties and storage stability of microencapsulated DHA-rich oil with different wall materials. *Appl. Biochem. Biotechnol.* 1129–1142.
- Cheng, Y., Xu, W., Chen, Z., Wang, Z., Huang, D., 2016. Micronization of etoposide using solution-enhanced dispersion by supercritical CO₂. *J. Supercrit. Fluids* 115, 10–16.
- Cheng, Y.S., Lu, P.M., Huang, C.Y., Wu, J.J., 2017. Encapsulation of lycopene with lecithin and α -tocopherol by supercritical antisolvent process for stability

- enhancement. *J. Supercrit. Fluids* 130, 246–252.
- Chew, S.C., Nyam, K.L., 2016. Microencapsulation of kenaf seed oil by co-extrusion technology. *J. Food Eng.* 175, 43–50.
- Chia, S.L., Boo, H.C., Muhamad, K., Sulaiman, R., Umanan, F., Chong, G.H., 2015. Effect of subcritical carbon dioxide extraction and bran stabilization methods on rice bran oil. *J. Am. Oil Chem. Soc.* 92, 393–402.
- Chinnarasu, C., Montes, A., Ponce, M.T.F.-, Casas, L., Mantell, C., Pereyra, C., de la Ossa, E.J.M., 2015. Precipitation of antioxidant fine particles from *Olea europaea* leaves using supercritical antisolvent process. *J. Supercrit. Fluids* 97, 125–132.
- Chinta, D.D., Graves, R.A., Pamujula, S., Praetorius, N., Bostanian, L.A., Mandal, T.K., 2009. Spray-dried chitosan as a direct compression tableting excipient. *Drug Dev. Ind. Pharm.* 35, 43–48.
- Chrastil, J., 1982. Solubility of solids and liquids in supercritical gases. *J. Phys. Chem.* 86, 3016–3021.
- Coates, J., 2000. Interpretation of infrared spectra, A practical approach. *Encycl. Anal. Chem.*
- Cocero, M.J., Martín, Á., Mattea, F., Varona, S., 2009. Encapsulation and co-precipitation processes with supercritical fluids: Fundamentals and applications. *J. Supercrit. Fluids* 47, 546–555.
- Codex Alimentarius, 2013. Codex standard for named vegetable oils. *Codex Stan210* 1–16.
- Coimbra, P., Duarte, C.M.M., De Sousa, H.C., 2006. Cubic equation-of-state correlation of the solubility of some anti-inflammatory drugs in supercritical carbon dioxide. *Fluid Phase Equilib.* 239, 188–199.
- Colombo, M.L., 2010. An update on Vitamin E, Tocopherol and Tocotrienol—Perspectives. *Molecules* 15, 2103–2113.
- Comunian, T.A., Chaves, I.E., Thomazini, M., Moraes, I.C.F., Ferro-Furtado, R., de Castro, I.A., Favaro-Trindade, C.S., 2017. Development of functional yogurt containing free and encapsulated echium oil, phytosterol and sinapic acid. *Food Chem.* 237, 948–956.
- Cortés-Camargo, S., Cruz-Olivares, J., Barragán-Huerta, B.E., Dublán-García, O., Román-Guerrero, A., Pérez-Alonso, C., 2017. Microencapsulation by spray drying of lemon essential oil: Evaluation of mixtures of mesquite gum–nopal mucilage as new wall materials. *J. Microencapsul.* 34, 395–407.
- Craft, N.E., Craft, N.E., 2001. Chromatographic techniques for carotenoid separation, in: *Current Protocols in Food Analytical Chemistry*. John Wiley & Sons, Inc., Hoboken, NJ, USA, p. F:F2:F2.3.

- Cunha, V.M.B., Silva, M.P., Raul N. Carvalho Jr., M.A.A.M., Machado, N.T., Araújo, M.E., 2016. Lauric acid rich oil supercritical extraction and methodology to predict solubility. *Food Public Heal.* 6, 26–32.
- da Silva, J., 2016. DNA damage induced by occupational and environmental exposure to miscellaneous chemicals. *Mutat. Res. Mutat. Res.* 770, 170–182.
- Dal Magro, C., Aguiar, G.P.S., Veneral, J.G., dos Santos, A.E., de Chaves, L.M.P.C., Oliveira, J.V., Lanza, M., 2017. Co-precipitation of trans-resveratrol in PHBV using solution enhanced dispersion by supercritical fluids technique. *J. Supercrit. Fluids*.
- Danielski, L., Campos, L.M.A.S., Bresciani, L.F.V., Hense, H., Yunes, R.A., Ferreira, S.R.S., 2007. Marigold (*Calendula officinalis* L.) oleoresin: Solubility in scCO₂ and composition profile. *Chem. Eng. Process. Process Intensif.* 46, 99–106.
- Danlami, J.M., Zaini, M.A.A., Arsad, A., Yunus, M.A.C., 2015. Solubility assessment of castor (*Ricinus communis* L.) oil in supercritical CO₂ at different temperatures and pressures under dynamic conditions. *Ind. Crops Prod.* 76, 34–40.
- Dauqan, E., Sani, H.A., Abdullah, A., 2011. Vitamin E and Beta Carotene Composition in four different vegetable oils. *Am. J. Appl. Sci.* 8, 407–412.
- Davarnejad, R., Ahmad, Z., Sata, S.A., Moraveji, M.K., Ahmadloo, F., 2010. Mutual solubility study in supercritical fluid extraction of tocopherols from crude palm oil using CO₂ solvent. *Int. J. Mol. Sci.* 11, 3649–59.
- Davarnejad, R., Kassim, K.M., Ahmad, Z., Sata, S.A., 2009. Solubility of β -Carotene from crude palm oil in high-temperature and high-pressure carbon dioxide. *J. Chem. Eng. Data* 54, 2200–2207.
- De Aguiar, A.C., Silva, L.P.S., Alves De Rezende, C., Barbero, G.F., Martínez, J., 2016. Encapsulation of pepper oleoresin by supercritical fluid extraction of emulsions. *J. Supercrit. Fluids* 112, 37–43.
- de Barros Fernandes, R.V., Botrel, D.A., Silva, E.K., Borges, S.V., de Oliveira, C.R., Yoshida, M.I., de Andrade Feitosa, J.P., de Paula, R.C.M., 2016. Cashew gum and inulin: New alternative for ginger essential oil microencapsulation. *Carbohydr. Polym.* 153, 133–142.
- De Leonardis, A., Macciola, V., Niro, S., Nag, A., Panfili, G., 2017. Limits and potentials of African red palm oils purchased from European ethnic food stores. *Eur. Food Res. Technol.* 243, 1239–1248.
- de Lucas, A., Gracia, I., Rincón, J., García, M.T., 2007. Solubility determination and model prediction of olive husk oil in supercritical carbon dioxide and cosolvents. *Ind. Eng. Chem. Res.* 46, 5061–5066.
- De Paz, E., Martín, Á., Cocero, M.J., 2012. Formulation of β -carotene with soybean lecithin by PGSS (Particles from Gas Saturated Solutions)-drying. *J. Supercrit. Fluids* 72, 125–133.

- de Paz, E., Martín, Á., Duarte, C.M.M., Cocero, M.J., 2012. Formulation of β -carotene with poly-(ϵ -caprolactones) by PGSS process. *Powder Technol.* 217, 77–83.
- de Souza, V.B., Thomazini, M., de Carvalho Balieiro, J.C., Fávaro-Trindade, C.S., 2015. Effect of spray drying on the physicochemical properties and color stability of the powdered pigment obtained from vinification byproducts of the Bordo grape (*Vitis labrusca*). *Food Bioprod. Process.* 93, 39–50.
- Del Valle, J.M., Aguilera, J.M., 1988. An improved equation for predicting the solubility of vegetable oils in supercritical carbon dioxide. *Ind. Eng. Chem. Res.* 27, 1551–1553.
- del Valle, J.M., de la Fuente, J.C., Uquiche, E., 2012. A refined equation for predicting the solubility of vegetable oils in high-pressure CO₂. *J. Supercrit. Fluids* 67, 60–70.
- Demiray, E., Tulek, Y., 2015. Color degradation kinetics of carrot (*Daucus carota* L.) slices during hot air drying. *J. Food Process. Preserv.* 39, 800–805.
- Demiray, E., Tulek, Y., Yilmaz, Y., 2013. Degradation kinetics of lycopene, β -carotene and ascorbic acid in tomatoes during hot air drying. *LWT - Food Sci. Technol.* 50, 172–176.
- Dima, C., Cotârlet, M., Alexe, P., Dima, S., 2014. Microencapsulation of essential oil of pimento [*Pimenta dioica* (L) Merr.] by chitosan/k-carrageenan complex coacervation method. *Innov. Food Sci. Emerg. Technol.* 22, 203–211.
- Dima, C., Pătraşcu, L., Cantaragiu, A., Alexe, P., Dima, Ş., 2015. The kinetics of the swelling process and the release mechanisms of *Coriandrum sativum* L. essential oil from chitosan/alginate/inulin microcapsules. *Food Chem.* 195, 39–48.
- do Prado, I.M., Giufrida, W.M., Alvarez, V.H., Cabral, V.F., Quispe-Condori, S., Saldaña, M.D.A., Cardozo-Filho, L., 2011. Phase equilibrium measurements of Sacha Inchi oil (*Plukenetia volubilis*) and CO₂ at high pressures. *J. Am. Oil Chem. Soc.* 88, 1263–1269.
- Domian, E., Brynda-Kopytowska, A., Marzec, A., 2017. Functional properties and oxidative stability of flaxseed oil microencapsulated by spray drying using legume proteins in combination with soluble fiber or trehalose. *Food Bioprocess Technol.* 10, 1374–1386.
- Domian, E., Cenkier, J., Górska, A., Brynda-Kopytowska, A., 2018. Effect of oil content and drying method on bulk properties and stability of powdered emulsions with OSA starch and linseed oil. *LWT - Food Sci. Technol.* 88, 95–102.
- Domian, E., Sulek, A., Cenkier, J., Kerschke, A., 2014. Influence of agglomeration on physical characteristics and oxidative stability of spray-dried oil powder with milk protein and trehalose wall material. *J. Food Eng.* 125, 34–43.

- Dong, D., Qi, Z., Hua, Y., Chen, Y., Kong, X., Zhang, C., 2015. Microencapsulation of flaxseed oil by soya proteins-gum arabic complex coacervation. *Int. J. Food Sci. Technol.* 50, 1785–1791.
- Durante, M., Lenucci, M., Mita, G., 2014. Supercritical carbon dioxide extraction of carotenoids from pumpkin (*Cucurbita* spp.): A review. *Int. J. Mol. Sci.* 15, 6725–6740.
- Dutta, D., Dutta, A., Raychaudhuri, U., Chakraborty, R., 2006. Rheological characteristics and thermal degradation kinetics of beta-carotene in pumpkin puree. *J. Food Eng.* 76, 538–546.
- Dutta, S., Bhattacharjee, P., 2017. Nanoliposomal encapsulates of piperine-rich black pepper extract obtained by enzyme-assisted supercritical carbon dioxide extraction. *J. Food Eng.* 201, 49–56.
- Edem, D.O., 2002. Palm oil: biochemical, physiological, nutritional, hematological, and toxicological aspects: A review. *Plant foods Hum. Nutr.* 57, 319–41.
- Ewing, A. V., Gabrienko, A.A., Semikolenov, S. V., Dubkov, K.A., Kazarian, S.G., 2015. How do intermolecular interactions affect swelling of polyketones with a differing number of carbonyl groups? An in situ ATR-FTIR spectroscopic study of CO₂ sorption in polymers. *J. Phys. Chem. C* 119, 431–440.
- Fang, Z., Bhandari, B., 2011. Effect of spray drying and storage on the stability of bayberry polyphenols. *Food Chem.* 129, 1139–1147.
- Fayyad, E.M., Almaadeed, M.A., Jones, A., 2016. Preparation and characterization of urea – formaldehyde microcapsules filled with paraffin oil. *Polym. Bull.* 73, 631–646.
- Felix, P.H.C., Birchal, V.S., Botrel, D.A., Marques, G.R., Borges, S.V., 2016. Physicochemical and thermal stability of microcapsules of cinnamon essential oil by spray drying. *J. Food Process. Preserv.* 1–9.
- Ferreira, C.D., da Conceição, E.J.L., Machado, B.A.S., Hermes, V.S., de Oliveira Rios, A., Druzian, J.I., Nunes, I.L., 2016. Physicochemical characterization and oxidative stability of microencapsulated crude palm oil by spray drying. *Food Bioprocess Technol.* 9, 124–136.
- Fioramonti, S.A., Rubiolo, A.C., Santiago, L.G., 2017. Characterisation of freeze-dried flaxseed oil microcapsules obtained by multilayer emulsions. *Powder Technol.* 319, 238–244.
- Flory, P.J., 1945. Thermodynamics of high-polymer solutions. *J. Chem. Phys.* 1, 25–30.
- Foerster, M., Liu, C., Gengenbach, T., Woo, M.W., Selomulya, C., 2017. Reduction of surface fat formation on spray-dried milk powders through emulsion stabilization with λ -carrageenan. *Food Hydrocoll.* 70, 163–180.
- Follegatti-Romero, L.A., Piantino, C.R., Grimaldi, R., Cabral, F.A., 2009. Supercritical CO₂ extraction of omega-3 rich oil from Sacha inchi (*Plukenetia volubilis* L.)

- seeds. *J. Supercrit. Fluids* 49, 323–329.
- Franceschi, E., De Cesaro, A.M., Ferreira, S.R.S., Vladimir Oliveira, J., 2009. Precipitation of β -carotene microparticles from SEDS technique using supercritical CO₂. *J. Food Eng.* 95, 656–663.
- Frascareli, E.C., Silva, V.M., Tonon, R.V., Hubinger, M.D., 2012. Effect of process conditions on the microencapsulation of coffee oil by spray drying. *Food Bioprod. Process.* 90, 413–424.
- Fuchs, M., Turchiuli, C., Bohin, M., Cuvelier, M.E., Ordonnaud, C., Peyrat-Maillard, M.N., Dumoulin, E., 2006. Encapsulation of oil in powder using spray drying and fluidised bed agglomeration. *J. Food Eng.* 75, 27–35.
- Gadkari, P.V., Balaraman, M., 2015. Solubility of caffeine from green tea in supercritical CO₂: a theoretical and empirical approach. *J. Food Sci. Technol.* 52, 8004–8013.
- Gallardo, G., Guida, L., Martinez, V., López, M.C., Bernhardt, D., Blasco, R., Pedroza-Islas, R., Hermida, L.G., 2013. Microencapsulation of linseed oil by spray drying for functional food application. *Food Res. Int.* 52, 473–482.
- Gaspar, F., Lu, T., Marriott, R., Mellor, S., Watkinson, C., Al-Duri, B., Santos, R., Seville, J., 2003. Solubility of Echium, Borage, and Lunaria Seed Oils in Compressed CO₂. *J. Chem. Eng. Data* 48, 107–109.
- Giménez, P.J., Fernández-López, J.A., Angosto, J.M., Obón, J.M., 2015. Comparative thermal degradation patterns of natural yellow colorants used in foods. *Plant Foods Hum. Nutr.* 70, 380–387.
- Girardi, N.S., García, D., Passone, M.A., Nesci, A., Etcheverry, M., 2017. Microencapsulation of *Lippia turbinata* essential oil and its impact on peanut seed quality preservation. *Int. Biodeterior. Biodegradation* 116, 227–233.
- Girardi, N.S., García, D., Robledo, S.N., Passone, M.A., Nesci, A., Etcheverry, M., 2016. Microencapsulation of *Peumus boldus* oil by complex coacervation to provide peanut seeds protection against fungal pathogens. *Ind. Crops Prod.* 92, 93–101.
- Gómez-Prieto, M.S., Caja, M.M., Santa-María, G., 2002. Solubility in supercritical carbon dioxide of the predominant carotenes of tomato skin. *J. Am. Oil Chem. Soc.* 79, 897–902.
- Gonalves, E.M., Pinheiro, J., Abreu, M., Brandão, T.R.S., Silva, C.L.M., 2011. Kinetics of quality changes of pumpkin (*Curcubita maxima* L.) stored under isothermal and non-isothermal frozen conditions. *J. Food Eng.* 106, 40–47.
- Gonçalves, E.M., Pinheiro, J., Abreu, M., Brandão, T.R.S., Silva, C.L.M., 2007. Modelling the kinetics of peroxidase inactivation, colour and texture changes of pumpkin (*Cucurbita maxima* L.) during blanching. *J. Food Eng.* 81, 693–701.

- Gonzalez, A., Martinez, M.L., Paredes, A.J., Leon, A.E., Ribotta, P.D., 2016. Study of the preparation process and variation of wall components in chia (*Salvia hispanica* L.) oil microencapsulation. *Powder Technol.* 301, 868–875.
- Gordillo, M.D., Blanco, M.A., Molero, A., Martinez De La Ossa, E., 1999. Solubility of the antibiotic Penicillin G in supercritical carbon dioxide. *J. Supercrit. Fluids* 15, 183–190.
- Gouin, S., 2004. Microencapsulation: Industrial appraisal of existing technologies and trends, in: *Trends in Food Science and Technology*. Elsevier, pp. 330–347.
- Goyal, A., Sharma, V., Sihag, M.K., Tomar, S.K., Arora, S., Sabikhi, L., Singh, A.K., 2015. Development and physico-chemical characterization of microencapsulated flaxseed oil powder: A functional ingredient for omega-3 fortification. *Powder Technol.* 286, 527–537.
- Guamán-Balcázar, M.C., Montes, A., Pereyra, C., de la Ossa, E.M., 2017. Precipitation of mango leaves antioxidants by supercritical antisolvent process. *J. Supercrit. Fluids* 128, 218–226.
- Guan, Y., Wu, J., Zhong, Q., 2016. Eugenol improves physical and chemical stabilities of nanoemulsions loaded with β -carotene. *Food Chem.* 194, 787–796.
- Guclu-Ustundag, O., Temelli, F., 2000. Correlating the solubility behavior of fatty acids, mono-, di-, and triglycerides, and fatty acid esters in supercritical carbon dioxide. *Ind. Eng. Chem. Res.* 39, 4756–4766.
- Güçlü-Üstündağ, Ö., Temelli, F., 2004. Correlating the solubility behavior of minor lipid components in supercritical carbon dioxide. *J. Supercrit. Fluids* 31, 235–253.
- Gupta, R.B., Shim, J.-J., 2007. *Solubility in supercritical carbon dioxide*. CRC Press.
- Habibi, A., Keramat, J., Hojjatoleslami, M., Tamjidi, F., 2017. Preparation of fish oil microcapsules by complex coacervation of gelatin–gum arabic and their utilization for fortification of pomegranate juice. *J. Food Process Eng.* 40, 1–11.
- Haq, R. ul, Prasad, K., 2017. Antioxidant activity, phenolic, carotenoid and color changes in packaged fresh carrots stored under refrigeration temperature. *J. Food Meas. Charact.* 11, 1542–1549.
- Hategekimana, J., Masamba, K.G., Ma, J., Zhong, F., 2015. Encapsulation of vitamin E: Effect of physicochemical properties of wall material on retention and stability. *Carbohydr. Polym.* 124, 172–179.
- Hategekimana, J., Zhong, F., 2015. Degradation of vitamin E in nanoemulsions during storage as affected by temperature, light and darkness. *Int. J. Food Eng.* 11, 199–206.
- He, W.Z., Suo, Q.L., Jiang, Z.H., A, S., Hong, H.L., 2004. Precipitation of ephedrine by SEDS process using a specially designed prefilming atomizer. *J. Supercrit. Fluids* 31, 101–110.

- Heinzelmann, K., Franke, K., Velasco, J., Márquez-Ruiz, G., 2000. Microencapsulation of fish oil by freeze-drying techniques and influence of process parameters on oxidative stability during storage. *Eur. Food Res. Technol.* 211, 234–239.
- Hemery, Y.M., Fontan, L., Moench-Pfanner, R., Laillou, A., Berger, J., Renaud, C., Avallone, S., 2015. Influence of light exposure and oxidative status on the stability of vitamins A and D3 during the storage of fortified soybean oil. *Food Chem.* 184, 90–98.
- Hezave, A.Z., Aftab, S., Esmailzadeh, F., 2012. Solubility of sulindac in the supercritical carbon dioxide: Experimental and modeling approach. *J. Supercrit. Fluids* 68, 39–44.
- Hidalgo, A., Brandolini, A., Pompei, C., 2009. Kinetics of tocopherols degradation during the storage of einkorn (*Triticum monococcum* L. ssp. *monococcum*) and breadwheat (*Triticum aestivum* L. ssp. *aestivum*) flours. *Food Chem.* 116, 821–827.
- Hildebrand, J.H., 1970. Regular and related solutions; the solubility of gases, liquids, and solids, in: *Regular and Related Solutions; the Solubility of Gases, Liquids, and Solids*. Van Nostrand Reinhold Co., New York.
- Hong, S.-A., Kim, J.-D., Kim, J., Kang, J.W., Kang, I.-J., 2010. Phase equilibria of palm oil, palm kernel oil, and oleic acid+supercritical carbon dioxide and modeling using Peng–Robinson EOS. *J. Ind. Eng. Chem.* 16, 859–865.
- Hornero-Méndez, D., Pérez-Gálvez, A., Mínguez-Mosquera, I., 2001. A rapid spectrophotometric method for the determination of peroxide value in food lipids with high carotenoid content. *J. Am. Oil Chem. Soc.* 78, 1151–1155.
- Hsieh, C.-M., Vrabec, J., 2015. Vapor–liquid equilibrium measurements of the binary mixtures CO₂ + acetone and CO₂ + pentanones. *J. Supercrit. Fluids* 100, 160–166.
- Hu, D., Lin, C., Liu, L., Li, S., Zhao, Y., 2012. Preparation, characterization, and in vitro release investigation of lutein/zein nanoparticles via solution enhanced dispersion by supercritical fluids. *J. Food Eng.* 109, 545–552.
- Hu, L., Zhang, J., Hu, Q., Gao, N., Wang, S., Sun, Y., Yang, X., 2016. Microencapsulation of *Brucea javanica* oil: Characterization, stability and optimization of spray drying conditions. *J. Drug Deliv. Sci. Technol.* 36, 46–54.
- Hu, Q., Gerhard, H., Upadhyaya, I., Venkitanarayanan, K., Luo, Y., 2016. Antimicrobial eugenol nanoemulsion prepared by gum arabic and lecithin and evaluation of drying technologies. *Int. J. Biol. Macromol.* 87, 130–140.
- Hwang, E.S., Stacewicz-Sapuntzakis, M., Bowen, P.E., 2012. Effects of heat treatment on the carotenoid and tocopherol composition of tomato. *J. Food Sci.* 77, C1109–C1114.
- Ifeduba, E.A., Akoh, C.C., 2016. Microencapsulation of stearidonic acid soybean oil in Maillard reaction-modified complex coacervates. *Food Chem.* 199, 524–532.

- Imsanguan, P., Pongamphai, S., Douglas, S., Teppaitoon, W., Douglas, P.L., 2010. Supercritical antisolvent precipitation of andrographolide from *Andrographis paniculata* extracts: Effect of pressure, temperature and CO₂ flow rate. *Powder Technol.* 200, 246–253.
- ISO 6885:2016 Animal and vegetable fats and oils - Determination of anisidine value, n.d. ISO 6885:2016.
- Jafari, S.M., Assadpoor, E., He, Y., Bhandari, B., 2008. Encapsulation efficiency of food flavours and oils during spray drying. *Dry. Technol.* 26, 816–835.
- Jaromír Klemeš, J., Yen Liew, P., Shin Ho, W., Shiun Lim, J., Diana Hassan, N., Idayu Muhamad, I., 2017. Drying and preservation of phytochemicals from *Elateriospermum Tapos* seed oil using combination of freeze drying and microwave technique. *Chem. Eng. Trans.* 56, 1003–1008.
- Jia, J., Wang, J., Zhang, K., Zhou, D., Ge, F., Zhao, Y., 2017. Aescin nanoparticles prepared using SEDS: Composition stability and dissolution enhancement. *J. Supercrit. Fluids* 130, 267–272.
- Jiménez-Martín, E., Antequera Rojas, T., Gharsallaoui, A., Ruiz Carrascal, J., Pérez-Palacios, T., 2016. Fatty acid composition in double and multilayered microcapsules of ω -3 as affected by storage conditions and type of emulsions. *Food Chem.* 194, 476–486.
- Jiménez-Martín, E., Gharsallaoui, A., Pérez-Palacios, T., Ruiz Carrascal, J., Antequera Rojas, T., 2015. Volatile compounds and physicochemical characteristics during storage of microcapsules from different fish oil emulsions. *Food Bioprod. Process.* 96, 52–64.
- Joback, K.G., Reid, R.C., 2007. Estimation of pure-component properties from group-contributions. *Chem. Eng. Commun.* 57, 233–243.
- Johannsen, M., Brunner, G., 1997. Solubilities of the fat-soluble Vitamins A, D, E, and K in supercritical carbon dioxide. *J. Chem. Eng. Data* 42, 106–111.
- Jokić, S., Sudar, R., Svilović, S., Vidović, S., Bilić, M., Velić, D., Jurković, V., 2013. Fatty acid composition of oil obtained from soybeans by extraction with supercritical carbon dioxide. *Czech J. Food Sci* 31, 116–125.
- Jokić, S., Svilović, S., Zeković, Z., Vidović, S., 2011a. Mathematical modelling of soybean oil solubility in supercritical carbon dioxide. *Int. J. Food Sci. Technol.* 46, 1031–1037.
- Jokić, S., Svilović, S., Zeković, Z., Vidović, S., Velić, D., 2011b. Solubility and kinetics of soybean oil and fatty acids in supercritical CO₂. *Eur. J. Lipid Sci. Technol.* 113, 644–651.
- Jouyban, A., Chan, H.K., Foster, N.R., 2001. Mathematical representation of solute solubility in supercritical carbon dioxide using empirical expressions. *J. Supercrit. Fluids* 24, 19–35.

- Joyner, N.T., McIntyre, J.E., 1938. The oven test as an index of keeping quality. *Oil Soap* 15, 184–186.
- Jung, J., Perrut, M., 2001. Particle design using supercritical fluids: Literature and patent survey. *J. Supercrit. Fluids* 20, 179–219.
- Kagami, Y., Sugimura, S., Fujishima, N., Matsuda, K., Kometani, T., Matsumura, Y., 2003. Oxidative stability, structure, and physical characteristics of microcapsules formed by spray drying of fish oil with protein and dextrin wall materials. *J. Food Sci.* 68, 2248–2255.
- Karthik, P., Anandharamakrishnan, C., 2013. Microencapsulation of docosahexaenoic acid by spray-freeze-drying method and comparison of its stability with spray-drying and freeze-drying methods. *Food Bioprocess Technol.* 6, 2780–2790.
- Kaushik, P., Dowling, K., Barrow, C.J., Adhikari, B., 2015. Microencapsulation of omega-3 fatty acids: A review of microencapsulation and characterization methods. *J. Funct. Foods* 19, 868–881.
- Kaushik, P., Dowling, K., McKnight, S., Barrow, C.J., Adhikari, B., 2016. Microencapsulation of flaxseed oil in flaxseed protein and flaxseed gum complex coacervates. *Food Res. Int.* 86, 1–8.
- Kelly, G.M., O'Mahony, J.A., Kelly, A.L., O'Callaghan, D.J., 2014. Physical characteristics of spray-dried dairy powders containing different vegetable oils. *J. Food Eng.* 122, 122–129.
- Kha, T.C., Nguyen, M.H., Roach, P.D., Stathopoulos, C.E., 2015. A storage study of encapsulated gac (*Momordica cochinchinensis*) oil powder and its fortification into foods. *Food Bioprod. Process.* 96, 113–125.
- Khimeche, K., Alessi, P., Kikic, I., Dahmani, A., 2007. Solubility of diamines in supercritical carbon dioxide: Experimental determination and correlation. *J. Supercrit. Fluids* 41, 10–19.
- Kim, S., Kim, Y., Park, B.H., Lee, J.H., Kang, J.W., 2012. Measurement and correlation of solubility of water in carbon dioxide-rich phase. *Fluid Phase Equilib.* 328, 9–12.
- Klaypradit, W., Huang, Y.-W., 2008. Fish oil encapsulation with chitosan using ultrasonic atomizer. *LWT - Food Sci. Technol.* 41, 1133–1139.
- Klinkesorn, U., Sophanodora, P., Chinachoti, P., Decker, E.A., McClements, D.J., 2006. Characterization of spray-dried tuna oil emulsified in two-layered interfacial membranes prepared using electrostatic layer-by-layer deposition. *Food Res. Int.* 39, 449–457.
- Ko, S.N., Kim, C.J., Kim, C.T., Kim, Y., Kim, I.H., 2010. Effects of tocopherols and tocotrienols on the inhibition of autoxidation of conjugated linoleic acid. *Eur. J. Lipid Sci. Technol.* 112, 496–501.

- Kong, H., Teng, C., Liu, X., Zhou, J., Zhong, H., Zhang, Y., Han, K., Yu, M., 2014. Simultaneously improving the tensile strength and modulus of aramid fiber by enhancing amorphous phase in supercritical carbon dioxide. *RSC Adv.* 4, 20599.
- Kua, Y.L., Gan, S., Morris, A., Ng, H.K., 2016. Ethyl lactate as a potential green solvent to extract hydrophilic (polar) and lipophilic (non-polar) phytonutrients simultaneously from fruit and vegetable by-products. *Sustain. Chem. Pharm.* 4, 21–31.
- Kumar, S., Sangwan, S., Yadav, R., Langyan, S., Singh, M., 2014. Maize carotenoid composition and biofortification for provitamin a activity, in: *Maize: Nutrition Dynamics and Novel Uses*. Springer India, New Delhi, pp. 83–91.
- Kumar, S.K., Johnston, K.P., 1988. Modelling the solubility of solids in supercritical fluids with density as the independent variable. *J. Supercrit. Fluids* 1, 15–22.
- Laidler, K.J., 1984. The development of the Arrhenius equation. *J. Chem. Educ.* 61, 494.
- Laohasongkram, K., Mahamaktudsanee, T., Chaiwanichsiri, S., 2011. Microencapsulation of Macadamia oil by spray drying. *Procedia Food Sci.* 1, 1660–1665.
- Lau, H.L.N., Choo, Y.M., Ma, A.N., Chuah, C.H., 2007. Production of refined carotene-rich palm oil from palm mesocarp (*Elaeis guineensis*) using supercritical carbon dioxide. *J. Food Lipids* 14, 396–410.
- Jerma-García, M.J., Ramis-Ramos, G., Herrero-Martínez, J.M., Simó-Alfonso, E.F., 2010. Authentication of extra virgin olive oils by Fourier-transform infrared spectroscopy. *Food Chem.* 118, 78–83.
- Li, Y., Yu, Y., Wang, H., Zhao, F., 2016. Effect of process parameters on the recrystallization and size control of puerarin using the supercritical fluid antisolvent process. *Asian J. Pharm. Sci.* 11, 281–291.
- Lik Nang Lau, H., Choo, Y.M., Ma, A.N., Chuah, C.H., 2008. Selective extraction of palm carotene and vitamin E from fresh palm-pressed mesocarp fiber (*Elaeis guineensis*) using supercritical CO₂. *J. Food Eng.* 84, 289–296.
- Lim, S.-B., Jwa, M.-K., 1995. Extraction of beta-carotenes from carrot by supercritical carbon dioxide. *Korean J. Food Sci. Technol.* 27, 414–419.
- Lin, K.-L., Chng, L.-M., Lin, J.C.-T., Hsu, S.-L., Young, C.-C., Shieh, C.-J., Chang, C.-M.J., 2014. Effect of anti-solvent conditions on low density supercritical fluids precipitation of Zeaxanthin Palmitates from *Lycium barbarum* fruits. *J. Supercrit. Fluids* 87, 104–110.
- Liu, F., Cao, X., Wang, H., Liao, X., 2010. Changes of tomato powder qualities during storage. *Powder Technol.* 204, 159–166.
- Liu, T.T., Yang, T.S., 2011. Optimization of emulsification and microencapsulation of

- evening primrose oil and its oxidative stability during storage by response surface methodology. *J. Food Qual.* 34, 64–73.
- Lopes, B.L.F., Sánchez-Camargo, A.P., Ferreira, A.L.K., Grimaldi, R., Paviani, L.C., Cabral, F.A., 2012. Selectivity of supercritical carbon dioxide in the fractionation of fish oil with a lower content of EPA + DHA. *J. Supercrit. Fluids* 61, 78–85.
- Machado, F.R.S., Reis, D.F., Boschetto, D.L., Burkert, J.F.M., Ferreira, S.R.S., Oliveira, J.V., Burkert, C.A. V., 2014. Encapsulation of astaxanthin from *Haematococcus pluvialis* in PHBV by means of SEDS technique using supercritical CO₂. *Ind. Crops Prod.* 54, 17–21.
- Machado, F.R.S., Trevisol, T.C., Boschetto, D.L., Burkert, J.F.M., Ferreira, S.R.S., Oliveira, J.V., Burkert, C.A. V., 2016. Technological process for cell disruption, extraction and encapsulation of astaxanthin from *Haematococcus pluvialis*. *J. Biotechnol.* 218, 108–114.
- Machado, L.C., Pelegati, V.B., Oliveira, A.L., 2016. Study of simple microparticles formation of limonene in modified starch using PGSS - Particles from gas-saturated suspensions. *J. Supercrit. Fluids* 107, 260–269.
- MacHmudah, S., Zakaria, Winardi, S., Sasaki, M., Goto, M., Kusumoto, N., Hayakawa, K., 2012. Lycopene extraction from tomato peel by-product containing tomato seed using supercritical carbon dioxide. *J. Food Eng.* 108, 290–296.
- Maddinedi, S.B., Mandal, B.K., Vankayala, R., Kalluru, P., Tammina, S.K., Kiran Kumar, H.A., 2014. Casein mediated green synthesis and decoration of reduced graphene oxide. *Spectrochim. Acta - Part A Mol. Biomol. Spectrosc.* 126, 227–231.
- Mahfoudhi, N., Hamdi, S., 2015. Kinetic degradation and storage stability of β -carotene encapsulated by freeze-drying using almond gum and gum arabic as wall materials. *J. Food Process. Preserv.* 39, 896–906.
- Malacrida, C.R., Ferreira, S., Zuanon, L.A.C., Nicoletti Telis, V.R., 2015. Freeze-drying for microencapsulation of turmeric oleoresin using modified starch and gelatin. *J. Food Process. Preserv.* 39, 1710–1719.
- Malaysian Palm Oil Board, 2017. Overview of the Malaysian oil palm industry 2016 [WWW Document]. *Econ. Ind. Dev. Div. Malaysian Palm Oil Board*. URL http://bepi.mpob.gov.my/images/overview/Overview_of_Industry_2016.pdf (accessed 11.30.17).
- Mao, L., Wang, D., Liu, F., Gao, Y., 2017. Emulsion design for the delivery of β -carotene in complex food systems. *Crit. Rev. Food Sci. Nutr.* 1–15.
- Marcela, F., Lucía, C., Eva, B., David, G., Ángeles, B.M., Luis, B., 2015. Microencapsulation of essential oils by interfacial polymerization using polyurea as a wall material. *J. Encapsulation Adsorpt. Sci.* 05, 165–177.
- Markom, M., Singh, H., Hasan, M., 2001. Supercritical CO₂ fractionation of crude

palm oil. *J. Supercrit. Fluids* 20, 45–53.

Marques da Silva, M., Paese, K., Stanisçuaski Guterres, S., Raffin Pohlmann, A., Kuhn Rutz, J., Fernando Flores Cantillano, R., Nora, L., de Oliveira Rios, A., 2017. Thermal and ultraviolet–visible light stability kinetics of co-nanoencapsulated carotenoids. *Food Bioprod. Process.* 105, 86–94.

Marszałek, K., Skąpska, S., Woźniak, Ł., Sokołowska, B., 2015. Application of supercritical carbon dioxide for the preservation of strawberry juice: Microbial and physicochemical quality, enzymatic activity and the degradation kinetics of anthocyanins during storage. *Innov. Food Sci. Emerg. Technol.* 32, 101–109.

Martín, Á., Pham, H.M., Kilzer, A., Kareth, S., Weidner, E., 2010. Micronization of polyethylene glycol by PGSS (Particles from Gas Saturated Solutions)-drying of aqueous solutions. *Chem. Eng. Process. Process Intensif.* 49, 1259–1266.

Mattea, F., Martín, Á., Matías-Gago, A., Cocero, M.J., 2009. Supercritical antisolvent precipitation from an emulsion: β -Carotene nanoparticle formation. *J. Supercrit. Fluids* 51, 238–247.

Mba, O.I., Dumont, M.-J., Ngadi, M., 2015. Palm Oil: Processing, Characterization and Utilization in the Food Industry – A Review. *Food Biosci.* 10, 26–41.

Mba, O.I., Dumont, M.J., Ngadi, M., 2017. Thermostability and degradation kinetics of tocochromanols and carotenoids in palm oil, canola oil and their blends during deep-fat frying. *LWT - Food Sci. Technol.* 82, 131–138.

McClements, D.J., 2012. Nanoemulsions versus microemulsions: terminology, differences, and similarities. *Soft Matter* 8, 1719–1729.

Mellado-Ortega, E., Hornero-Méndez, D., 2016. Carotenoid evolution during short-storage period of durum wheat (*Triticum turgidum* conv. *durum*) and tritordeum (\times *Tritordeum Ascherson* et *Graebner*) whole-grain flours. *Food Chem.* 192, 714–723.

Mendes, R.L., Nobre, B.P., Coelho, J.P., Palavra, A.F., 1999. Solubility of β -carotene in supercritical carbon dioxide and ethane. *J. Supercrit. Fluids* 16, 99–106.

Méndez-Santiago, J., Teja, A.S., 1999. The solubility of solids in supercritical fluids. *Fluid Phase Equilib.* 158–160, 501–510.

Mezzomo, N., Paz, E. de, Maraschin, M., Martín, Á., Cocero, M.J., Ferreira, S.R.S., 2012. Supercritical anti-solvent precipitation of carotenoid fraction from pink shrimp residue: Effect of operational conditions on encapsulation efficiency. *J. Supercrit. Fluids* 66, 342–349.

Miao, H., Chen, Z., Xu, W., Wang, W., Song, Y., Wang, Z., 2018. Preparation and characterization of naringenin microparticles via a supercritical anti-solvent process. *J. Supercrit. Fluids* 131, 19–25.

Mihalcea, L., Turturică, M., Ghinea, I.O., Barbu, V., Ioniță, E., Cotârleț, M., Stănciuc,

- N., 2017. Encapsulation of carotenoids from sea buckthorn extracted by CO₂ supercritical fluids method within whey proteins isolates matrices. *Innov. Food Sci. Emerg. Technol.* 42, 120–129.
- Min, J., Li, S., Hao, J., Liu, N., 2010. Supercritical CO₂ Extraction of *Jatropha* oil and solubility correlation. *J. Chem. Eng. Data* 55, 3755–3758.
- Mohd Nasir, H., Md Salleh, L., Ismail, A.R., Machmudah, S., 2017. Solubility correlation of gall (*Quercus infectoria*) extract in supercritical CO₂ using semi-empirical equations. *Asia-Pacific J. Chem. Eng.* 12, 790–797.
- Montes, A., Pereyra, C., de la Ossa, E.J.M., 2015. Screening design of experiment applied to the supercritical antisolvent precipitation of quercetin. *J. Supercrit. Fluids* 104, 10–18.
- Moshashaée, S., Bisrat, M., Forbes, R.T., Quinn, éilís Á., Nyqvist, H., York, P., 2003. Supercritical fluid processing of proteins: lysozyme precipitation from aqueous solution. *J. Pharm. Pharmacol.* 55, 185–192.
- Mustapa, A.N., Manan, Z.A., Mohd Azizi, C.Y., Setianto, W.B., Mohd Omar, A.K., 2011. Extraction of β -carotenes from palm oil mesocarp using sub-critical R134a. *Food Chem.* 125, 262–267.
- Mutsokoti, L., Panozzo, A., Tongonya, J., Kebede, B.T., Van Loey, A., Hendrickx, M., 2017. Carotenoid stability and lipid oxidation during storage of low-fat carrot and tomato based systems. *LWT - Food Sci. Technol.* 80, 470–478.
- N. Nguyen, H., . Gaspillo, P.D., B. Maridable, J., M. Malaluan, R., Hinode, H., Salim, C., Huynh, H.K.P., 2011. Modeling of *Moringa oleifera* oil solubility in supercritical carbon dioxide. *J. Chem. Eng. Process Technol.* 02.
- Nagendran, B., Unnithan, U.R., Choo, Y.M., Sundram, K., 2000. Characteristics of red palm oil, α -Carotene and vitamin E-rich refined oil for food uses. *Food Nutr. Bull.* 21, 189–194.
- Nagesha, G.K., Manohar, B., Udaya Sankar, K., 2003. Enrichment of tocopherols in modified soy deodorizer distillate using supercritical carbon dioxide extraction. *Eur. Food Res. Technol.* 217, 427–433.
- Nalawade, S.P., Picchioni, F., Janssen, L.P.B.M., 2006. Supercritical carbon dioxide as a green solvent for processing polymer melts: Processing aspects and applications. *Polym. Sci* 31, 19–43.
- Nejadmansouri, M., Hosseini, S.M.H., Niakosari, M., Yousefi, G.H., Golmakani, M.T., 2016. Physicochemical properties and storage stability of ultrasound-mediated WPI-stabilized fish oil nanoemulsions. *Food Hydrocoll.* 61, 801–811.
- Nerome, H., Machmudah, S., Wahyudiono, Fukuzato, R., Higashiura, T., Youn, Y.S., Lee, Y.W., Goto, M., 2013. Nanoparticle formation of lycopene/ β -cyclodextrin inclusion complex using supercritical antisolvent precipitation. *J. Supercrit. Fluids* 83, 97–103.

- Ng, S.K., Jessie, L.Y.L., Tan, C.P., Long, K., Nyam, K.L., 2013. Effect of accelerated storage on microencapsulated kenaf seed oil. *JAOCS, J. Am. Oil Chem. Soc.* 90, 1023–1029.
- Nissiotis, M., Tasioula-Margari, M., 2002. Changes in antioxidant concentration of virgin olive oil during thermal oxidation. *Food Chem.* 77, 371–376.
- Nogala-Kalucka, M., Korczak, J., Elmadfa, I., Wagner, K.H., 2005. Effect of α - and δ -tocopherol on the oxidative stability of a mixed hydrogenated fat under frying conditions. *Eur. Food Res. Technol.* 221, 291–297.
- Nunes, A.V.M., Duarte, C.M.M., 2011. Dense CO₂ as a solute, co-solute or co-solvent in particle formation processes: A review. *Materials (Basel)*. 4, 2017–2041.
- O'Brien, R.D., 2009. *Fats and oils: formulating and processing for applications*. CRC Press.
- Oguntibeju, O.O., Esterhuyse, A.J., Truter, E.J., 2010. Possible role of red palm oil supplementation in reducing oxidative stress in HIV / AIDS and TB patients: A Review. *J. Med. plants Res.* 4, 188–196.
- Oliveira, D.A., Mezzomo, N., Gomes, C., Ferreira, S.R.S., 2017. Encapsulation of passion fruit seed oil by means of supercritical antisolvent process. *J. Supercrit. Fluids* 129, 96–105.
- Özkal, S.G., Salgın, U., Yener, M.E., 2005. Supercritical carbon dioxide extraction of hazelnut oil. *J. Food Eng.* 69, 217–223.
- Özkal, S.G., Yener, M.E., Bayindirli, L., 2006. The solubility of apricot kernel oil in supercritical carbon dioxide. *Int. J. Food Sci. Technol.* 41, 399–404.
- Partanen, R., Raula, J., Seppänen, R., Buchert, J., Kauppinen, E., Forssell, P., 2008. Effect of relative humidity on oxidation of flaxseed oil in spray dried whey protein emulsions. *J. Agric. Food Chem.* 56, 5717–22.
- Pedro, A.S., Villa, S.D., Caliceti, P., De Melo, S.A.B.V., Albuquerque, E.C., Bertucco, A., Salmaso, S., 2016. Curcumin-loaded solid lipid particles by PGSS technology. *J. Supercrit. Fluids* 107, 534–541.
- Peng, D.-Y., Robinson, D.B., 1976. A New Two-Constant Equation of State. *Ind. Eng. Chem. Fundam.* 15, 59–64.
- Pereira Souza, A.C., Deyse Gurak, P., Damasceno Ferreira Marczak, L., 2017. Maltodextrin, pectin and soy protein isolate as carrier agents in the encapsulation of anthocyanins-rich extract from *Jaboticaba pomace*. *Food Bioprod. Process.* 102, 186–194.
- Plöcker, U., Knapp, H., Prausnitz, J., 1978. Calculation of high-pressure vapor-liquid equilibria from a corresponding-states correlation with emphasis on asymmetric mixtures. *Ind. Eng. Chem. Process Des. Dev.* 17, 324–332.

- Podsędek, A., 2007. Natural antioxidants and antioxidant capacity of Brassica vegetables: A review. *LWT - Food Sci. Technol.* 40, 1–11.
- Pourashouri, P., Shabanpour, B., Razavi, S.H., Jafari, S.M., Shabani, A., Aubourg, S.P., 2014. Oxidative stability of spray-dried microencapsulated fish oils with different wall materials. *J. Aquat. Food Prod. Technol.* 23, 567–578.
- Priamo, W.L., De Cezaro, A.M., Benetti, S.C., Oliveira, J.V., Ferreira, S.R.S., 2011. In vitro release profiles of β -carotene encapsulated in PHBV by means of supercritical carbon dioxide micronization technique. *J. Supercrit. Fluids* 56, 137–143.
- Priamo, W.L., De Cezaro, A.M., Ferreira, S.R.S., Oliveira, J.V., 2010. Precipitation and encapsulation of β -carotene in PHBV using carbon dioxide as anti-solvent. *J. Supercrit. Fluids* 54, 103–109.
- Przybylski, R., Wu, J., Eskin, N.A.M., 2013. A rapid method for determining the oxidative stability of oils suitable for breeder size samples. *J. Am. Oil Chem. Soc.* 90, 933–939.
- Puah, C.W., Choo, Y.M., Ma, A.N., Chuah, C.H., 2007. Solubility of tocopherol and tocotrienols from palm oil in supercritical carbon dioxide. *J. Food Lipids* 14, 377–385.
- Puah, C.W., Choo, Y.M., Ngan, M.A., Chuah, C.H., 2005. Supercritical fluid extraction of palm carotenoids. *Am. J. Environ. Sci.* 1, 264–269.
- Qian, M.C., 2010. Gas Chromatography, in: *Food Analysis Laboratory Manual*. Springer Science & Business Media, pp. 159–163.
- QUAN, C., Johan, C., Charlotta, T., 2009. Carotenoids particle formation by supercritical fluid technologies. *Chinese J. Chem. Eng.* 17, 344–349.
- Quispe-Condori, S., Saldaña, M.D.A., Temelli, F., 2011. Microencapsulation of flax oil with zein using spray and freeze drying. *LWT - Food Sci. Technol.* 44, 1880–1887.
- Rahman, N.N.A., Al-Rawi, S.S., Ibrahim, A.H., Ben Nama, M.M., Ab Kadir, M.O., 2012. Supercritical carbon dioxide extraction of the residual oil from palm kernel cake. *J. Food Eng.* 108, 166–170.
- Raitio, R., Orlén, V., Skibsted, L.H., 2011. Storage stability of cauliflower soup powder: The effect of lipid oxidation and protein degradation reactions. *Food Chem.* 128, 371–379.
- Raksa, A., Sawaddee, P., Raksa, P., Aldred, A.K., 2017. Microencapsulation, chemical characterization, and antibacterial activity of *Citrus hystrix* DC (Kaffir Lime) peel essential oil. *Monatshefte für Chemie* 148, 1229–1234.
- Rantakylä, M., Jäntti, M., Aaltonen, O., Hurme, M., 2002. The effect of initial drop size on particle size in the supercritical antisolvent precipitation (SAS) technique.

- J. Supercrit. Fluids 24, 251–263.
- Rao, A. V, Rao, L.G., 2007. Carotenoids and human health. *Pharmacol. Res.* 55, 207–16.
- Razmkhah, S., Tan, C.P., Long, K., Nyam, K.L., 2013. Quality changes and antioxidant properties of microencapsulated Kenaf (*Hibiscus cannabinus* L.) seed oil during accelerated storage. *J. Am. Oil Chem. Soc.* 90, 1859–1867.
- Redlich, O., Kwong, J.N.S., 1949. On the thermodynamics of solutions. V. An equation of state. Fugacities of gaseous solutions. *Chem. Rev.* 44, 233–244.
- Reverchon, E., Lamberti, G., Antonacci, A., 2008. Supercritical fluid assisted production of HPMC composite microparticles. *J. Supercrit. Fluids* 46, 185–196.
- Rice, A.L., Burns, J.B., 2010. Moving from efficacy to effectiveness: Red palm oil's role in preventing vitamin a deficiency. *J. Am. Coll. Nutr.* 29, 302S–313S.
- Rodrigues, J.E., Araújo, M.E., Azevedo, F.F.M., Machado, N.T., 2005. Phase equilibrium measurements of Brazil nut (*Bertholletia excelsa*) oil in supercritical carbon dioxide. *J. Supercrit. Fluids* 34, 223–229.
- Rodrigues, M.A., Li, J., Almeida, A.J., Matos, H.A., Gomes De Azevedo, E., 2006. Efficiency of water removal from water/ethanol mixtures using supercritical carbon dioxide. *Brazilian J. Chem. Eng.* 23, 205–212.
- Rohman, A., Man, Y.B.C., 2010. Fourier transform infrared (FTIR) spectroscopy for analysis of extra virgin olive oil adulterated with palm oil. *Food Res. Int.* 43, 886–892.
- Romero, N., Robert, P., Masson, L., Ortiz, J., González, K., Tapia, K., Dobarganes, C., 2007. Effect of α -tocopherol, α -tocotrienol and *Rosa mosqueta* shell extract on the performance of antioxidant-stripped canola oil (*Brassica* sp.) at high temperature. *Food Chem.* 104, 383–389.
- Romero, N., Robert, P., Masson, L., Ortiz, J., Pavez, J., Garrido, C., Foster, M., Dobarganes, C., 2004. Effect of α -tocopherol and α -tocotrienol on the performance of Chilean hazelnut oil (*Gevuina avellana* Mol) at high temperature. *J. Sci. Food Agric.* 84, 943–948.
- Rutz, J.K., Borges, C.D., Zambiasi, R.C., Crizel-Cardozo, M.M., Kuck, L.S., Noreña, C.P.Z., 2017. Microencapsulation of palm oil by complex coacervation for application in food systems. *Food Chem.* 220, 59–66.
- Sabliov, C.M., Fronczek, C., Astete, C.E., Khachatryan, M., Khachatryan, L., Leonardi, C., 2009. Effects of temperature and UV light on degradation of α -Tocopherol in free and dissolved form. *J. Am. Oil Chem. Soc.* 86, 895–902.
- Saldaña, M.D.A., Temelli, F., Guigard, S.E., Tomberli, B., Gray, C.G., 2010. Apparent solubility of lycopene and β -carotene in supercritical CO₂, CO₂+ethanol and CO₂+canola oil using dynamic extraction of tomatoes. *J. Food Eng.* 99, 1–8.

- Sampaio, K.A., Ayala, J. V, Silva, S.M., Ceriani, R., Verhe, R., Meirelles, A.J.A., 2013. Thermal degradation kinetics of carotenoids in palm oil. *J. Am. Oil Chem. Soc.* 90, 191–198.
- Sanchez-Reinoso, Z., Gutiérrez, L.F., 2017. Effects of the emulsion composition on the physical properties and oxidative stability of Sacha Inchi (*Plukenetia volubilis* L.) oil microcapsules produced by spray drying. *Food Bioprocess Technol.* 10, 1354–1366.
- Sánchez, M.D.R.H., Cuvelier, M.-E., Turchiuli, C., 2016. Effect of α -tocopherol on oxidative stability of oil during spray drying and storage of dried emulsions. *Food Res. Int.* 88, 32–41.
- Sanguansri, L., Udabage, P., Bhail, S., Ying, D.Y., Cheng, L.J., Shen, Z., Augustin, M.A., 2016. Microencapsulated fish oil powder formulation with improved resistance to oil leakage during powder compression. *JAOCS, J. Am. Oil Chem. Soc.* 93, 701–710.
- Santos, D.T., Martín, Á., Meireles, M.A.A., Cocero, M.J., 2012. Production of stabilized sub-micrometric particles of carotenoids using supercritical fluid extraction of emulsions. *J. Supercrit. Fluids* 61, 167–174.
- Santos, D.T., Meireles, M.A.A., 2013. Micronization and encapsulation of functional pigments using supercritical carbon dioxide. *J. Food Process Eng.* 36, 36–49.
- Schaffer, S., Muller, W.E., Eckert, G.P., 2005. Tocotrienols: Constitutional Effects in Aging and Disease. *Am. Soc. Nutr. Sci.* 135, 151–154.
- Secuianu, C., Feroiu, V., Gean̄, D., 2008. Phase behavior for carbon dioxide + ethanol system: Experimental measurements and modeling with a cubic equation of state. *J. Supercrit. Fluids* 47, 109–116.
- Sen, C.K., Khanna, S., Roy, S., 2007. Tocotrienols in health and disease: the other half of the natural vitamin E family. *Mol. Aspects Med.* 28, 692–728.
- Seppanen, C.M., Song, Q., Saari Csallany, A., 2010. The antioxidant functions of tocopherol and tocotrienol homologues in oils, fats, and food systems. *J. Am. Oil Chem. Soc.* 87, 469–481.
- Serfert, Y., Drusch, S., Schwarz, K., 2009. Chemical stabilisation of oils rich in long-chain polyunsaturated fatty acids during homogenisation, microencapsulation and storage. *Food Chem.* 113, 1106–1112.
- Setianto, W.B., Atmaji, P., Anggoro, D.D., 2012. Solubility examination of palm kernel oil in supercritical CO₂ and its correlation with solvent density based model, in: *Proceedings of International Conference on Chemical and Material Engineering*, p. SPE.10-1-6.
- Shamaei, S., Seiiedlou, S.S., Aghbashlo, M., Tsotsas, E., Kharaghani, A., 2017. Microencapsulation of walnut oil by spray drying: Effects of wall material and drying conditions on physicochemical properties of microcapsules. *Innov. Food*

- Sci. Emerg. Technol. 39, 101–112.
- Shao, D., Venkitasamy, C., Li, X., Pan, Z., Shi, J., Wang, B., Teh, H.E., McHugh, T.H., 2015. Thermal and storage characteristics of tomato seed oil. *LWT - Food Sci. Technol.* 63, 191–197.
- Shariati, A., Peters, C.J., 2003. Recent developments in particle design using supercritical fluids. *Curr. Opin. Solid State Mater. Sci.* 7, 371–383.
- Sharif, H.R., Goff, H.D., Majeed, H., Liu, F., Nsor-Atindana, J., Haider, J., Liang, R., Zhong, F., 2017. Physicochemical stability of β -carotene and α -tocopherol enriched nanoemulsions: Influence of carrier oil, emulsifier and antioxidant. *Colloids Surfaces A* 529, 550–559.
- Shen, L., Chen, J., Bai, Y., Ma, Z., Huang, J., Feng, W., 2016. Physical properties and stabilization of microcapsules containing thyme oil by complex coacervation. *J. Food Sci.* 81, N2258–N2262.
- Shi, J., Mittal, G., Kim, E., Xue, S.J., 2007. Solubility of carotenoids in supercritical CO₂. *Food Rev. Int.* 23, 341–371.
- Silva, E.K., Azevedo, V.M., Cunha, R.L., Hubinger, M.D., Meireles, M.A.A., 2016. Ultrasound-assisted encapsulation of annatto seed oil: Whey protein isolate *versus* modified starch. *Food Hydrocoll.* 56, 71–83.
- Siow, L.-F., 2012. Effect of pH on garlic oil encapsulation by complex coacervation. *J. Food Process. Technol.* 04, 1–5.
- Soares, B.M.C., Gamarra, F.M.C., Paviani, L.C., Gonçalves, L.A.G., Cabral, F.A., 2007. Solubility of triacylglycerols in supercritical carbon dioxide. *J. Supercrit. Fluids* 43, 25–31.
- Soave, G., 1972. Equilibrium constants from a modified Redlich-Kwong equation of state. *Chem. Eng. Sci.* 27, 1197–1203.
- Sosa, M.V., Rodríguez-Rojo, S., Mattea, F., Cismondi, M., Cocero, M.J., 2011. Green tea encapsulation by means of high pressure antisolvent coprecipitation. *J. Supercrit. Fluids* 56, 304–311.
- Sosnik, A., Seremeta, K.P., 2015. Advantages and challenges of the spray-drying technology for the production of pure drug particles and drug-loaded polymeric carriers. *Adv. Colloid Interface Sci.* 223, 40–54.
- Soukoulis, C., Bohn, T., 2015. A comprehensive overview on the micro- and nano-technological encapsulation advances for enhancing the chemical stability and bioavailability of carotenoids. *Crit. Rev. Food Sci. Nutr.* 1–36.
- Sovová, H., Stateva, R.P., Galushko, A.A., 2001a. Solubility of β -carotene in supercritical CO₂ and the effect of entrainers. *J. Supercrit. Fluids* 21, 195–203.
- Sovová, H., Zarevúcka, M., Vacek, M., Stránský, K., 2001b. Solubility of two

- vegetable oils in supercritical CO₂. *J. Supercrit. Fluids* 20, 15–28.
- Spada, J.C., Pelayo, C., Nor Na, Z., Damasceno, L., Marczak, F., Tessaro, I.C., 2012. Study on the stability of β -carotene microencapsulated with *pinhão* (*Araucaria angustifolia* seeds) starch. *Carbohydr. Polym.* 89, 1166–1173.
- Sparks, D.L., Hernandez, R., Estévez, L.A., 2008. Evaluation of density-based models for the solubility of solids in supercritical carbon dioxide and formulation of a new model. *Chem. Eng. Sci.* 63, 4292–4301.
- Sparks, D.L., Hernandez, R., Estévez, L.A., Meyer, N., French, T., 2007. Solubility of Azelaic acid in supercritical carbon dioxide. *J. Chem. Eng. Data* 52, 1246–1249.
- Stievano, M., Elvassore, N., 2005. High-pressure density and vapor–liquid equilibrium for the binary systems carbon dioxide–ethanol, carbon dioxide–acetone and carbon dioxide–dichloromethane. *J. Supercrit. Fluids* 33, 7–14.
- Sui, X., Bary, S., Zhou, W., 2016. Changes in the color, chemical stability and antioxidant capacity of thermally treated anthocyanin aqueous solution over storage. *Food Chem.* 192, 516–524.
- Sun-Waterhouse, D., Zhou, J., Miskelly, G.M., Wibisono, R., Wadhwa, S.S., 2011. Stability of encapsulated olive oil in the presence of caffeic acid. *Food Chem.* 126, 1049–1056.
- Sun, M., Temelli, F., 2006. Supercritical carbon dioxide extraction of carotenoids from carrot using canola oil as a continuous co-solvent. *J. Supercrit. Fluids* 37, 397–408.
- Sundram, K., Sambanthamurthi, R., Tan, Y.A., 2003. Palm fruit chemistry and nutrition. *Asia Pac. J. Clin. Nutr.* 12, 355–362.
- Sung, H. Do, Shim, J.J., 1999. Solubility of C. I. Disperse Red 60 and C. I. Disperse Blue 60 in supercritical carbon dioxide. *J. Chem. Eng. Data* 44, 985–989.
- Taberero, A., Martín Del Valle, E.M., Galán, M.A., 2012a. Supercritical fluids for pharmaceutical particle engineering: Methods, basic fundamentals and modelling. *Chem. Eng. Process.* 60, 9–25.
- Taberero, A., Martín Del Valle, E.M., Galán, M.A., 2012b. Precipitation of tretinoin and acetaminophen with solution enhanced dispersion by supercritical fluids (SEDS). Role of phase equilibria to optimize particle diameter. *Powder Technol.* 217, 177–188.
- Tang, Z., Jin, J., Zhang, Z., Liu, H., 2012. New experimental data and modeling of the solubility of compounds in supercritical carbon dioxide. *Ind. Eng. Chem. Res.* 51, 5515–5526.
- Timilsena, Y.P., Adhikari, R., Barrow, C.J., Adhikari, B., 2016. Microencapsulation of chia seed oil using chia seed protein isolate-chia seed gum complex coacervates. *Int. J. Biol. Macromol.* 91, 347–357.

- Tochigi, K., Namae, T., Suga, T., Matsuda, H., Kurihara, K., Carolina Dos Ramos, M., McCabe, C., 2010. Measurement and prediction of high-pressure vapor–liquid equilibria for binary mixtures of carbon dioxide + n-octane, methanol, ethanol, and perfluorohexane. *J. Supercrit. Fluids* 55, 682–689.
- Tolve, R., Condelli, N., Can, A., Tchuenbou-Magaia, F.L., 2017. Development and characterization of phytosterol-enriched oil microcapsules for foodstuff application. *Food Bioprocess Technol.* 1–12.
- Tomita, K., Machmudah, S., Fukuzato, R., Kanda, H., Quitain, A.T., Sasaki, M., Goto, M., 2014. Extraction of rice bran oil by supercritical carbon dioxide and solubility consideration. *Sep. Purif. Technol.* 125, 319–325.
- Tomita, K., Machmudah, S., Quitain, A.T., Sasaki, M., Fukuzato, R., Goto, M., 2013. Extraction and solubility evaluation of functional seed oil in supercritical carbon dioxide. *J. Supercrit. Fluids* 79, 109–113.
- Topal, U., Sasaki, M., Goto, M., Hayakawa, K., 2006. Extraction of lycopene from tomato skin with supercritical carbon dioxide: Effect of operating conditions and solubility analysis. *J. Agric. Food Chem.* 54, 5604–5610.
- Torres, A., Ilabaca, E., Rojas, A., Rodriguez, F., Galotto, M.J., Guarda, A., Villegas, C., Romero, J., 2017. Effect of processing conditions on the physical, chemical and transport properties of polylactic acid films containing thymol incorporated by supercritical impregnation. *Eur. Polym. J.* 89, 195–210.
- Tsai, W.-C., Rizvi, S.S.H., 2016. Measurement and correlation of Citronellal and Methyl Anthranilate solubilities in supercritical carbon dioxide. *J. Chem. Eng. Data* 61, 182–187.
- Tsutsumi, C., Watanabe, R., Tokumaru, A., Kuwaoka, N., Nakayama, Y., Shiono, T., 2016. Effects of supercritical carbon dioxide treatment on the morphology of poly(L-lactide). *J. Appl. Polym. Sci.* 133, 1–9.
- Turasan, H., Sahin, S., Sumnu, G., 2015. Encapsulation of rosemary essential oil. *LWT - Food Sci. Technol.* 64, 112–119.
- Turchiuli, C., Fuchs, M., Bohin, M., Cuvelier, M.E., Ordonnaud, C., Peyrat-Maillard, M.N., Dumoulin, E., 2005. Oil encapsulation by spray drying and fluidised bed agglomeration. *Innov. Food Sci. Emerg. Technol.* 6, 29–35.
- Ubeyitogullari, A., Ciftci, O.N., 2017. Generating phytosterol nanoparticles in nanoporous bioaerogels via supercritical carbon dioxide impregnation: Effect of impregnation conditions. *J. Food Eng.* 207, 99–107.
- Umesha, S.S., Monahar, B., Naidu, K.A., 2013. Microencapsulation of α -linolenic acid-rich garden cress seed oil: Physical characteristics and oxidative stability. *Eur. J. Lipid Sci. Technol.* 115, 1474–1482.
- Van Rooyen, J., Esterhuysen, A.J., Engelbrecht, A.M., Du Toit, E.F., 2008. Health benefits of a natural carotenoid rich oil: A proposed mechanism of protection

against ischaemia/reperfusion injury. *Asia Pac. J. Clin. Nutr.* 17, 316–319.

- Varona, S., Kareth, S., Martín, Á., Cocero, M.J., 2010. Formulation of lavandin essential oil with biopolymers by PGSS for application as biocide in ecological agriculture, in: *Journal of Supercritical Fluids*. pp. 369–377.
- Verleyen, T., Verhe, R., Huyghebaert, A., Dewettinck, K., De Greyt, W., 2001. Identification of α -tocopherol oxidation products in triolein at elevated temperatures. *J. Agric. Food Chem.* 49, 1508–1511.
- Vicente, J., de Souza Cezarino, T., Pereira, L.J.B., da Rocha, E.P., Sá, G.R., Gamallo, O.D., de Carvalho, M.G., Garcia-Rojas, E.E., 2017. Microencapsulation of sachai oil using emulsion-based delivery systems. *Food Res. Int.* 99, 612–622.
- Villanueva Bermejo, D., Ibáñez, E., Reglero, G., Turner, C., Fornari, T., Rodríguez-Meizoso, I., 2015. High catechins/low caffeine powder from green tea leaves by pressurized liquid extraction and supercritical antisolvent precipitation. *Sep. Purif. Technol.* 148, 49–56.
- Visentin, A., Rodríguez-Rojo, S., Navarrete, A., Maestri, D., Cocero, M.J., 2012. Precipitation and encapsulation of rosemary antioxidants by supercritical antisolvent process. *J. Food Eng.* 109, 9–15.
- Wagner, K.-H., Wotruba, F., Elmadfa, I., 2001. Antioxidative potential of tocotrienols and tocopherols in coconut fat at different oxidation temperatures. *Eur. J. Lipid Sci. Technol.* 103, 746–751.
- Wang, S., Shi, Y., Tu, Z., Zhang, L., Wang, H., Tian, M., Zhang, N., 2017. Influence of soy lecithin concentration on the physical properties of whey protein isolate-stabilized emulsion and microcapsule formation. *J. Food Eng.* 207, 73–80.
- Wang, W., Waterhouse, G.I.N., Sun-Waterhouse, D., 2013. Co-extrusion encapsulation of canola oil with alginate: Effect of quercetin addition to oil core and pectin addition to alginate shell on oil stability. *Food Res. Int.* 54, 837–851.
- Wang, Y.F., Jia, J.X., Tian, Y.Q., Shu, X., Ren, X.J., Guan, Y., Yan, Z.Y., 2018. Antifungal effects of clove oil microcapsule on meat products. *LWT - Food Sci. Technol.* 89, 604–609.
- Wardhani, D.H., Fuciños, P., Vázquez, J.A., Pandiella, S.S., 2013. Inhibition kinetics of lipid oxidation of model foods by using antioxidant extract of fermented soybeans. *Food Chem.* 139, 837–844.
- Waterhouse, G.I.N., Wang, W., Sun-Waterhouse, D., 2014. Stability of canola oil encapsulated by co-extrusion technology: effect of quercetin addition to alginate shell or oil core. *Food Chem.* 142, 27–38.
- Wibowo, S., Vervoort, L., Tomic, J., Santiago, J.S., Lemmens, L., Panozzo, A., Grauwet, T., Hendrickx, M., Van Loey, A., 2015. Colour and carotenoid changes of pasteurised orange juice during storage. *Food Chem.* 171, 330–340.

- Wong, C.W., Wijayanti, H.B., Bhandari, B.R., 2015. Maillard reaction in limited moisture and low water environment, in: *Water Stress in Biological, Chemical, Pharmaceutical and Food Systems*. pp. 41–63.
- Woodside, J. V, McGrath, A.J., Lyner, N., McKinley, M.C., 2015. Carotenoids and health in older people. *Maturitas* 80, 63–68.
- Xia, Q., Wang, B., Akanbi, T.O., Li, R., Yang, W., Adhikari, B., Barrow, C.J., 2017. Microencapsulation of lipase produced omega-3 concentrates resulted in complex coacervates with unexpectedly high oxidative stability. *J. Functional Foods*, 35, 499-506.
- Xie, M., Fan, D., Zhao, Z., Li, Z., Li, G., Chen, Y., He, X., Chen, A., Li, J., Lin, X., Zhi, M., Li, Y., Lan, P., 2015. Nano-curcumin prepared via supercritical: Improved anti-bacterial, anti-oxidant and anti-cancer efficacy. *Int. J. Pharm.* 496, 732–740.
- Yahia, E.M., Ornelas-Paz, J. de J., 2009. Chemistry, stability, and biological actions of carotenoids, in: *Fruit and Vegetable Phytochemicals: Chemistry, Nutritional Value, and Stability*. Wiley-Blackwell, Oxford, UK, pp. 177–222.
- Yan, T., Cheng, Y., Wang, Z., Huang, D., Miao, H., Zhang, Y., 2015. Preparation and characterization of baicalein powder micronized by the SEDS process. *J. Supercrit. Fluids* 104, 177–182.
- Yang, G., Li, Z., Shao, Q., Feng, N., 2017. Measurement and correlation study of silymarin solubility in supercritical carbon dioxide with and without a cosolvent using semi-empirical models and back-propagation artificial neural networks. *Asian J. Pharm. Sci.* 12, 456–463.
- Yang, G., Zhao, Y., Feng, N., Zhang, Y., Liu, Y., Dang, B., 2015. Improved dissolution and bioavailability of silymarin delivered by a solid dispersion prepared using supercritical fluids. *Asian J. Pharm. Sci.* 10, 194–202.
- Yang, J., Ciftci, O.N., 2017. Encapsulation of fish oil into hollow solid lipid micro- and nanoparticles using carbon dioxide. *Food Chem.* 231, 105–113.
- Yang, J., Ciftci, O.N., 2016. Development of free-flowing peppermint essential oil-loaded hollow solid lipid micro- and nanoparticles via atomization with carbon dioxide. *Food Res. Int.* 87, 83–91.
- Yang, X., Gao, N., Hu, L., Li, J., Sun, Y., 2015. Development and evaluation of novel microcapsules containing poppy-seed oil using complex coacervation. *J. Food Eng.* 161, 87–93.
- Yeo, S.-D., Kiran, E., 2005. Formation of polymer particles with supercritical fluids: A review. *J. Supercrit. Fluids* 34, 287–308.
- Young, T.J., Johnston, K.P., Mishima, K., Tanaka, H., 1999. Encapsulation of lysozyme in a biodegradable polymer by precipitation with a vapor-over-liquid antisolvent. *J. Pharm. Sci.* 88, 640–650.

- Yu, J.M., Lu, B.C.Y., 1987. A three-parameter cubic equation of state for asymmetric mixture density calculations. *Fluid Phase Equilib.* 34, 1–19.
- Yu, Z.R., Singh, B., Rizvi, S., Zollweg, J., 1994. Solubilities of fatty acids, fatty acid esters, triglycerides, and fats and oils in supercritical carbon dioxide. *J. Supercrit. Fluids* 7, 51–59.
- Yun, J.H., Lee, H.Y., Asaduzzaman, A.K.M., Chun, B.S., 2013. Micronization and characterization of squid lecithin/polyethylene glycol composite using particles from gas saturated solutions (PGSS) process. *J. Ind. Eng. Chem.* 19, 686–691.
- Yusuf, M., Sharma, V., Pathak, K., 2014. Nanovesicles for transdermal delivery of felodipine: Development, characterization, and pharmacokinetics. *Int. J. Pharm. Investig.* 4, 119–130.
- Zeb, A., Murkovic, M., 2013. Pro-oxidant effects of β -carotene during thermal oxidation of edible oils. *J. Am. Oil Chem. Soc.* 90, 881–889.
- Zhang, S., Chen, J., Yin, X., Wang, X., Qiu, B., Zhu, L., Lin, Q., 2017. Microencapsulation of tea tree oil by spray-drying with methyl cellulose as the emulsifier and wall material together with chitosan/alginate. *J. Appl. Polym. Sci.* 134.
- Zhang, Y., Tan, C., Abbas, S., Eric, K., Xia, S., Zhang, X., 2015. Modified SPI improves the emulsion properties and oxidative stability of fish oil microcapsules. *Food Hydrocoll.* 51, 108–117.
- Zhao, L., Temelli, F., 2017. Preparation of anthocyanin-loaded liposomes using an improved supercritical carbon dioxide method. *Innov. Food Sci. Emerg. Technol.* 39, 119–128.
- Zhao, S., Zhang, D., 2014. An experimental investigation into the solubility of *Moringa oleifera* oil in supercritical carbon dioxide. *J. Food Eng.* 138, 1–10.
- Zhao, Z., Xie, M., Li, Y., Chen, A., Li, G., Zhang, J., Hu, H., Wang, X., Li, S., 2015. Formation of curcumin nanoparticles via solution-enhanced dispersion by supercritical CO₂. *Int. J. Nanomedicine* 10, 3171–3181.
- Zhou, D., Pan, Y., Ye, J., Jia, J., Ma, J., Ge, F., 2017. Preparation of walnut oil microcapsules employing soybean protein isolate and maltodextrin with enhanced oxidation stability of walnut oil. *LWT - Food Sci. Technol.* 83, 292–297.
- Zhu, C., Cai, Y., Gertz, E.R., La Frano, M.R., Burnett, D.J., Burri, B.J., 2015. Red palm oil-supplemented and biofortified cassava gari increase the carotenoid and retinyl palmitate concentrations of triacylglycerol-rich plasma in women. *Nutr. Res.* 35, 965–974.
- Zou, Y., Jiang, Y., Yang, T., Hu, P., Xu, X., 2012. Minor constituents of palm oil: Characterization, processing, and application, in: *Palm Oil: Production, Processing, Characterization, and Uses*. Elsevier, pp. 471–526.

Zuknik, M.H., Nik Norulaini, N.A., Wan Nursyazreen Dalila, W.S., Ali, N.R., Omar, A.K.M., 2016. Solubility of virgin coconut oil in supercritical carbon dioxide. *J. Food Eng.* 168, 240–244.

Zunin, P., Boggia, R., Turrini, F., Leardi, R., 2015. Total and “free” lipids in commercial infant formulas: Fatty acid composition and their stability to oxidation. *Food Chem.* 173, 332–338.

