



**UNIVERSITI PUTRA MALAYSIA**

***CHARACTERIZATION OF EXOPOLYSACCHARIDES FROM  
PROBIOTIC *Lactobacillus* STRAINS ISOLATED FROM TRADITIONAL  
MALAYSIAN FOODS***

**EILAF SULIMAN KHALIL SULIMAN**

**FSTM 2018 30**



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By

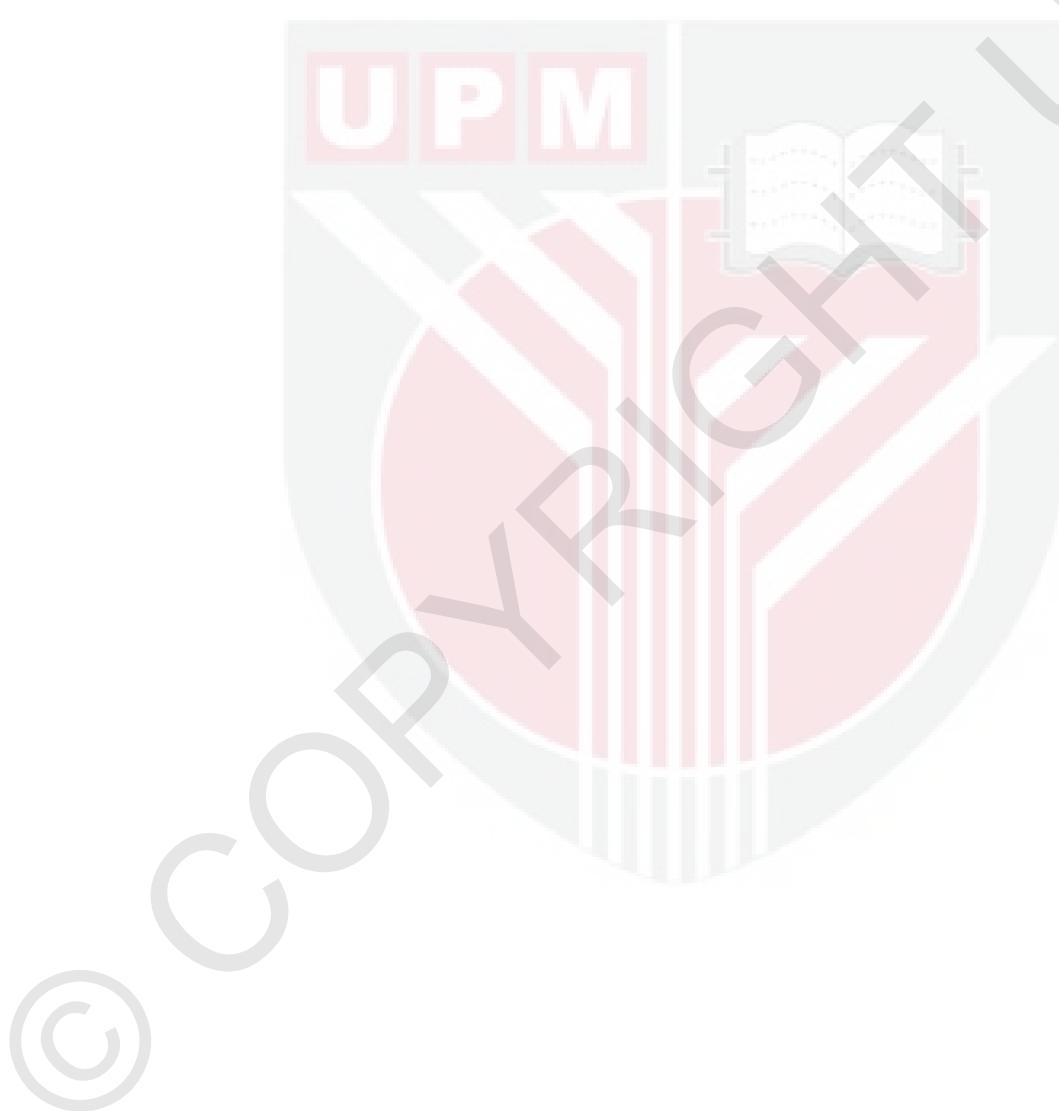
**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

**April 2018**

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## **DEDICATION**

*This work is dedicated to the soul of my late mother  
(May Allah rest her soul and forgive her)*

*My father who always prays for my success*

*My brothers and sisters for their constant encouragement and support*

*My husband and children for being my inspiration*

*My friends and colleagues*



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment  
of the requirement for the degree of Doctor of Philosophy

**CHARACTERIZATION OF EXOPOLYSACCHARIDES FROM  
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MALAYSIAN FOODS**

By

**EILAF SULIMAN KHALIL SULIMAN**

**April 2018**

**Chairman : Professor Mohd Yazid B Abd Manap, PhD**  
**Faculty : Food Science and Technology**

Lactic acid bacteria (LAB) are generally recognized as safe (GRAS) microorganisms, and play an important role in food and animal feed as potential probiotics. LAB are used for the production of several industrially interesting metabolites such as bioactive peptides, antibacterial compounds, aroma compounds, and cells wall component like exopolysaccharides (EPS). EPS are generally related to all forms of polysaccharides that are present outside of the microbial cell wall. Although, EPS produced by LAB have multifunctional, technological, and health benefits, their industrial applications are hindered by their low yield. The present study was conducted to isolate and identify varieties of LAB from some traditional Malaysian foods and to investigate their probiotic characteristics, functional properties, ability to produce EPS, and to study their bioactivities. One hundred and twenty LAB strains were isolated from five different traditional Malaysian foods. The preliminary selection carried out optically by screening the strains' resistance to low pH 2.0 and bile salts 0.3% as well as ability to produce EPS, and a total of 13 isolates were accordingly selected. The 13 isolates were identified using the carbohydrates fermentation profile and 16S rRNA gene sequences. Six of the *Lactobacillus* strains were associated with the species *Lb. fermentum* (DUR18, TAP1, PIC7, BU11, BU14, and TAP16) and four strains belonged to *Lb. plantarum* (DUR2, DUR5, DUR8, and TEMP9). Besides, *Lb. crispatus* (DUR4) *Lb. reuteri* (DUR12) and *Lb. pentosus* (DUR20) were also identified. Two commercial *Lactobacillus* strains namely: *Lb. rhamnosus* (ATCC53103) and *Lb. plantarum* (ATCC8014) were used as reference strains. The isolated strains exhibited high tolerance to acid (<80%) and bile (<65 %) as well as high survival rate in the gastrointestinal tract. The results of the cells wall properties showed that auto-aggregation and cell surface hydrophobicity were ranged between 53-80 and 16-80%, respectively, whereas, the highest co-aggregation value (66%) was reported by *Lb. plantarum* (DUR8) with *Pseudomonas aeruginosa*. The in vitro assay

for the functional properties of the isolated strains showed antioxidant activity equivalent to trolox (30-70%), high cholesterol assimilated. Also, the isolated strains exhibited good inhibitory activity against some tested pathogens due to organic acids production. The EPS production by the 13 *Lactobacillus* isolated strains were ranged between 50 and 850 mg/L. Based on that, two strains; *Lb. pentosus* and *Lb. reuteri* which showed the highest EPS yield were screened out for further characterization. The purified EPS were composed of glucose, arabinose, and rhamnose. FTIR analysis indicated a specific spectrum of neutral polysaccharides in the region of 1577 cm<sup>-1</sup> which attributed to C=O stretching. An extracellular glucosytransferase (GTF) enzyme corresponding to EPS from the selected strains were produced with the molecular weight falls between 120 and 150 kDa. The GTF exhibited high activity under static condition at pH 5 and incubation temperature 28°C. EPS yield from *Lb. reuteri* and *Lb. pentosus* were optimized using response surface methodology (RSM). The maximum EPS yields attained were 1.0 g/L and 1.07 g/L produced by *Lb. reuteri* and *Lb. pentosus*, respectively. Moreover, the optimal values for the tested variables were: 41 and 39.5 °C for incubation temperature, pH 6.4 and 6.5, and the incubation times were 33.7 and 48 h for *Lb. reuteri* and *Lb. pentosus*, respectively. The RSM displayed a significant ( $p < 0.05$ ) response fit for the studied variables with a high coefficient of determination ( $R^2$ ). The predicted and observed values displayed no significant ( $p > 0.05$ ) differences. Both incubation time and the pH significantly ( $p < 0.05$ ) affected the EPS production in both strains. The optimum points were practically validated. The EPS samples revealed potent antioxidant activity to scavenging DPPH (65.8 and 68.0% for *Lb. pentosus* and *Lb. reuteri*, respectively) and antibiofilm activity against some pathogens (above 50%). It can be concluded that *Lactobacillus* strains isolated from traditional Malaysian foods in this study are promising probiotics which have the potential to produce EPS with functional attributes.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN EKSOPOLISAKARIDA DARIPADA STRAIN  
LAKTOBASILUS PROBIOTIK TERASING DARIPADA MAKANAN  
TRADISIONAL MALAYSIA**

Oleh

**EILAF SULIMAN KHALIL SULIMAN**

April 2018

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**Fakulti** : Sains dan Teknologi Makanan

Secara umumnya, bakteria asid laktik (LAB) telah diakui sebagai mikroorganisma selamat (GRAS) dan ia memainkan peranan penting dalam makanan dan makanan haiwan sebagai probiotik yang berpotensi. LAB telah digunakan untuk penghasilan beberapa metabolit menarik berunsurkan industri seperti peptida bioaktif, kompoun antibakteria, kompoun aroma dan komponen dinding sel seperti eksopolisakarida (EPS). EPS secara umumnya berkaitan dengan semua bentuk polisakarida yang wujud di luar dinding sel mikroba. Walaupun, EPS yang dihasilkan oleh LAB mempunyai pelbagai manfaat multifungsi, teknologikal dan kesihatan, aplikasi perindustrian telah dihalang oleh hasil rendah mereka. Kajian ini telah dijalankan untuk mengasas dan mengenal pasti pelbagai jenis LAB daripada beberapa makanan tradisional Malaysia dan juga bertujuan untuk menyelidiki ciri probiotik mereka, sifat fungsian dan kemampuan untuk menghasilkan EPS dan mengkaji bioaktiviti mereka. Seratus dua puluh strain LAB telah diasingkan daripada lima makanan tradisional Malaysia yang berbeza. Pemilihan awal telah dijalankan secara optik dengan menyaring ketahanan strain pada pH rendah 2.0 dan garam hemedu 0.3% di samping kemampuan untuk menghasilkan EPS, dan sejumlah 13 isolat telah dipilih sewajarnya. 13 isolat telah dikenal pasti menggunakan profil fermentasi karbohidrat dan susunan gen rRNA 16S. Enam strain Laktobasilus telah dikaitkan dengan spesis *Lb. fermentum* (DUR18, TAP1, PIC7, BU11, BU14 dan TAP16) dan empat strain tergolong dalam *Lb. plantarum* (DUR2, DUR5, DUR8 dan TEMP9). Selain itu, *Lb. crispatus* (DUR4), *Lb. reuteri* (DUR12), dan *Lb. pentosus* (DUR20) juga telah dikenal pasti. Dua strain *Lb.* yang dihasilkan secara komersial, iaitu *Lb. rhinnosus* (ATCC53103) dan *Lb. plantarum* (ATCC8014) telah digunakan sebagai strain rujukan. Strain yang diasingkan itu menunjukkan sifat toleransi yang tinggi terhadap asid (<80%) dan hemedu (<65%) di samping kadar kelangsungan yang tinggi dalam saluran

gastrosus. Dapatan mengenai sifat dinding sel sel dinding menunjukkan bahawa autoagregasi dan hidrofobisiti permukaan sel adalah masing-masing antara 53-80 dan 16-80%, manakala, nilai koagregasi tertinggi ( 66%) telah dilaporkan oleh *Lb. plantarum* (DUR8) dengan *Pseudomonas aeruginosa*. Cerakinan in vitro bagi sifat fungsian strain terasing menunjukkan aktiviti antioksidan bersamaan dengan troloks (30-70%), kolesterol tinggi berasimilasi. Di samping itu, strain terasing tersebut memperlihatkan aktiviti penentangan yang kuat terhadap beberapa patogen yang diuji akibat penghasilan asid organik. Penghasilan EPS oleh 13 strain *Lactobacillus* terasing berjulat antara 50 dan 850 mg/L. Berdasarkan julat tersebut, dua strain; *Lb. pentosus* dan *Lb. reuteri* yang menunjukkan hasil EPS tertinggi telah disaring bagi pencirian selanjutnya. EPS tulen terdiri daripada glukosa, arabinosa, dan rhamnosa. Analisis FTIR menunjukkan spektrum khusus bagi polisakarida neutral dalam lingkungan  $1577\text{ cm}^{-1}$  yang ditentukan pada peluasan C=O. Enzim glukositransferas ekstraselular (GTF) sepadan dengan EPS daripada strain terpilih telah dihasilkan dengan berat molekular turun antara 120 dan 150 kDa. GTF memperlihatkan aktiviti yang tinggi pada pH 5 dan suhu inkubasi 28°C. Hasil EPS daripada *Lb. reuteri* dan *Lb. pentosus* telah dioptimumkan menggunakan metodologi permukaan tindak balas (RSM). Hasil EPS maksimum yang diperoleh ialah 1.0 g/L bagi *Lb. reuteri* dan 1.07 g / L bagi *Lb. pentosus* yang dihasilkan, masing-masing oleh *Lb. reuteri* dan *Lb. Pentosus*. Tambahan pula, nilai optimum bagi pemboleh ubah yang diuji ialah: 41 dan 39.5 °C untuk suhu inkubasi, pH 6.4 dan 6.5, dan masa inkubasi ialah masing-masing 33.7 dan 48 bagi *Lb. reuteri* dan *Lb. pentosus*. RSM menunjukkan tindak balas ( $p \leq 0.05$ ) yang signifikan sesuai bagi pemboleh ubah yang dikaji dengan pekali penentuan yang tinggi ( $R^2$ ). Nilai yang diramal dan diperhatikan memperlihatkan tiada perbezaan yang signifikan ( $p > 0.05$ ). Kedua-dua masa inkubasi dan pH ( $p \leq 0.05$ ) secara signifikan memberi kesan pada penghasilan EPS dalam kedua-dua strain. Titik optimum secara praktikal telah disahkan. Sampel EPS menunjukkan aktiviti antioksidan poten pada menghapus-sisa DPPH (masing-masing 65.8 dan 68.0% untuk *Lb. pentosus* dan *Lb. reuteri*) dan aktiviti antibiofilem terhadap beberapa patogen (melebihi 50%). Kesimpulannya, strain *Lactobacillus* terasing daripada makanan tradisional Malaysia dalam kajian ini merupakan probiotik yang menggalakkan yang mempunyai potensi untuk menghasilkan EPS dengan atribut fungsian.

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This thesis is submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
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## LIST OF ABBREVIATIONS

$\mu\text{L}$	Micro liter
$\mu\text{M}$	microMolar
Amp	Ampicillin
ANOVA	Analysis of Variance
ATCC	American Type Culture Collection
ATP	Adenosine triphosphate
BHI	Brain heart infusion
BLAST	Basic local alignment search tool
bp	Base pairs
BSA	Bovine serum albumin
CaCl <sub>2</sub>	Calcium Chloride
CCD	Central composite design
CFU	Colony Forming Unit
DNA	Deoxyribonucleic acid
DSC	Differential Scanning Calorimetry
EPS	Exopolysaccharides
Ery	Erythromycin
FAO	Food and Agriculture Organization
FTF	Fructosyltransferase
FT-IR	Fourier Transmission Infrared Spectroscopy
G	Gram
GIT	Gastrointestinal tract
GRAS	Generally recognized as safe
GTFs	Glycosyltransferase(s)

H	Hour
$\text{H}_2\text{SO}_4$	Sulphuric acid
HCl	Hydrochloride acid
HPLC	High Performance Liquid Chromatography
$\text{K}_2\text{HPO}_4$	Potassium dihydrogen phosphate
kb	Kilo Base
kDa	Kilo Dalton
$\text{KH}_2\text{PO}_4$	Potassium phosphate
KOH	Potassium hydroxide
L	Liter
LAB	Lactic Acid Bacteria
MIC	Minimum Inhibitory Concentration
Log	Logarithm
min	Minutes
mL	Milliliter
MRS	de Man, Rogosa Sharpe
NaCl	Sodium Chloride
$\text{NaHCO}_3$	Sodium Bicarbonate
NaOH	Sodium Hydroxide
nm	Nanometre
P	Probability
PBS	Phosphate buffered saline
PCR	Polymerase chain reaction
RNA	Ribonucleic acid
rpm	Revolutions per minute
rRNA	Ribosomal RNA

RSM	Response Surface Methodology
SDS	Sodium Dodecyl Sulphate
SD	Standard deviation
SE	Standard error of mean
Tm	Melting Temperature
UV	Ultraviolet
UV	Ultra violet
w/v	Weight Per Volume
WHO	World Health Organization
$\mu\text{g}$	Microgram

## CHAPTER 1

### GENERAL INTRODUCTION

In the last decade, polysaccharides have drawn increasing attention for their technological properties and biological activities. Polysaccharides can be derived from plants, animals, algae, and microorganisms (fungi and bacteria). Plant and algal polysaccharides are the most predominant sources (Badel *et al.*, 2011). Microbial polysaccharides have many merits compared to other sources of polysaccharides; these advantages include (1) the production of polysaccharides is simple and less susceptible to the effect of environmental conditions – they are of high-quality with a stable supply and a short production cycle (Wang *et al.*, 2010); (2) diversity – the structure of EPS depends on the species of microorganism and the culture conditions (growth factors and growth conditions) (Harutoshi, 2013); (3) microbial EPS have unique physiochemical features that add functional, technological and economic value when used as food additives or functional foods (Li *et al.*, 2014a); (4) the low-cost possibility when the industrial wastes, such as glycerol, whey, molasses, hydrocarbon residue and CO<sub>2</sub>, are exploited as carbon substrates (Zannini *et al.*, 2016).

Among the microbial polysaccharides, the exopolysaccharides (EPS) produced by lactic acid bacteria (LAB), in particular, the *Lactobacillus* species, are the most widely used groups of bacteria that have obtained the status of Generally Recognized As Safe (GRAS) (Bourdichon *et al.*, 2012). EPS produced from LAB have received considerable attention due to their importance in the pharmaceutical, medical and food industries. The health benefits of EPS have been claimed to include the following properties: antioxidant activity, immune modulation, aiding in tolerance to acid/bile salts, facilitating adherence and colonization within the host, lowering the blood cholesterol, immunostimulatory activity, and antitumor effects (Ruas-Madiedo *et al.*, 2007; Hidalgo-Cantabrana *et al.*, 2012; Li *et al.*, 2014b). The technological functions of EPS in the food industry include their use as viscosifying agents, stabilizers, emulsifiers, gelling agents, and water-binding agents (Torino *et al.*, 2015).

Based on their monomeric composition, lactic acid bacteria EPS can be classified into homopolysaccharides (HoPS) and heteropolysaccharides (HePS). HoPS contain only one type of monosaccharide repeating unit (e.g., glucans and fructans made of glucose and fructose, respectively), whereas HePS consist of two or more types of monosaccharides (De Vuyst and Degeest, 1999). As reported by Abid *et al.* (2017) the chemical characterization is crucial for determining their potential application, sugar composition, chain length, and sugar linkage, as the presence of repeated units affect the biotechnological properties of EPS. The optimum technological and functional proprieties of EPS require sufficient production of these molecules to improve their yields and to obtain a particular functionality. As compared to some other bacteria, *Lactobacillus* is not a very efficient producer of EPS (Leroy and De Vuyst, 2016). Recently, EPS from various *Lactobacillus* species have been studied and characterized (Wang *et al.*, 2010; Li *et al.*, 2014a; Wang *et al.*, 2015; Wang *et al.*, 2016), but to date

no work has been carried out on EPS produced by *Lactobacillus* isolated from traditional Malaysian.

In order to achieve maximum EPS production, optimization is required, which includes various production factors starting from the selection of an appropriate strain and optimization of the culture conditions to the protocol used for EPS isolation. Furthermore, there is great interest in identifying new EPS that are suitable for special applications, or that have potential industrial relevance, either by applying different culture conditions or by using novel bacterial strains (Aslim *et al.*, 2005).

To ensure that the EPS produced from *Lactobacillus* act as a natural antioxidant or antibiofilm or displays another bioactive attribute, adequate research on the improvement of the yield and characterization of physiochemical and bioactivity properties is necessary. This study assumes that LAB isolated from traditional Malaysian foods have a potential probiotic characteristics and have the ability to produce EPS with functional attributes under optimal conditions.

Based on the above-mentioned hypothesis the specific objectives of the study in this thesis were:

- i. To isolate and identify LAB producing EPS from several traditional Malaysian fermented foods and evaluating their characteristics as potential probiotics.
- ii. To optimize incubation temperature, pH, and time for the production of EPS from *Lb. pentosus* and *Lb. reuteri*.
- iii. To investigate the production of EPS from *Lb. pentosus* and *Lb. reuteri*, and to study the activity of glucosyltransferase (GTF) enzyme.
- iv. To determine the antioxidant activity and antibiofilm activity of EPS from *Lb. pentosus* and *Lb. reuteri*.

## REFERENCES

- Abdi, R., Sheikh-Zeinoddin, M., and Soleimanian-Zad, S. (2012). Efficiency of modified skimmed milk base media to achieve high exopolysaccharide/cell ratios by *Lactobacillus delbrueckii* subsp. *bulgaricus* SZ2 in optimised conditions defined by the response surface methodology. *International Journal of Food Science and Technology*, **47**(4): 768-775.
- Abid, Y., Casillo, A., Gharsallah, H., Joulak, I., Lanzetta, R., Corsaro, M. M., Attia, H., and Azabou, S. (2017). Production and structural characterization of exopolysaccharides from newly isolated probiotic lactic acid bacteria. *International Journal of Biological Macromolecules*.
- Ahotupa, M., Saxelin, M., and Korpela, R. (1996). Antioxidative Properties of *Lactobacillus GG*. *Nutrition Today*, **31**(6): 51S.
- Alander, M., Korpela, R., Saxelin, M., Vilpponen-Salmela, T., Mattila-Sandholm, T., and von Wright, A. (1997). Recovery of *Lactobacillus rhamnosus* GG from human colonic biopsies. *Letters in Applied Microbiology*, **24**(5): 361-364.
- Ammor, S., Tauveron, G., Dufour, E., and Chevallier, I. (2006). Antibacterial activity of lactic acid bacteria against spoilage and pathogenic bacteria isolated from the same meat small-scale facility: 1—Screening and characterization of the antibacterial compounds. *Food Control*, **17**(6): 454-461.
- Anandharaj, M., Sivasankari, B., Santhanakaruppu, R., Manimaran, M., Rani, R. P., and Sivakumar, S. (2015). Determining the probiotic potential of cholesterol-reducing *Lactobacillus* and *Weissella* strains isolated from gherkins (fermented cucumber) and south Indian fermented koozh. *Research in Microbiology*, **166**(5): 428-439.
- Andersson, H., Asp, N., Bruce, Å., Roos, S., Wadström, T., and Wold, A. (2016). Health effects of probiotics and prebiotics A literature review on human studies. *Näringsforskning*, **45**(1): 58-75.
- Anita, S. K., Kalpana, M., and Bhavanath, J. (2007). Bacterial exopolysaccharides—a perception. *Journal of Basic Microbiology*, **47**(2): 103-117.
- Anwar, M. A., Kralj, S., van der Maarel, M. J., and Dijkhuizen, L. (2008). The probiotic *Lactobacillus johnsonii* NCC 533 produces high-molecular-mass inulin from sucrose by using an inulosucrase enzyme. *Applied and Environmental Microbiology*, **74**(11): 3426-3433.
- Aslim, B., Yüksekdağ, Z. N., Beyatlı, Y., and Mercan, N. (2005). Exopolysaccharide production by *Lactobacillus delbrueckii* subsp. *bulgaricus* and *Streptococcus thermophilus* strains under different growth conditions. *World Journal of Microbiology and Biotechnology*, **21**(5): 673-677.

- Aween. (2012). Evaluation on Antibacterial Activity of *Lactobacillus acidophilus* Strains Isolated from Honey. *American Journal of Applied Sciences*, **9**(6): 807-817.
- Axelsson, L., and Ahrné, S. (2000). Lactic acid bacteria *Applied microbial systematics* (pp. 367-388): Springer.
- Ayala-Hernández, I., Hassan, A., Goff, H. D., Mira de Orduña, R., and Corredig, M. (2008). Production, isolation and characterization of exopolysaccharides produced by *Lactococcus lactis* subsp. *cremoris* JFR1 and their interaction with milk proteins: Effect of pH and media composition. *International Dairy Journal*, **18**(12): 1109-1118.
- Ayeni, F. A., Sanchez, B., Adeniyi, B. A., de Los Reyes-Gavilan, C. G., Margolles, A., and Ruas-Madiedo, P. (2011). Evaluation of the functional potential of *Weissella* and *Lactobacillus* isolates obtained from Nigerian traditional fermented foods and cow's intestine. *International Journal of Food Microbiology*, **147**(2): 97-104.
- Badel, S., Bernardi, T., and Michaud, P. (2011). New perspectives for *Lactobacilli* exopolysaccharides. *Biotechnology Advances*, **29**(1): 54-66.
- Bajpai, V. K., Rather, I. A., and Park, Y. H. (2016). Partially Purified Exopolysaccharide from *Lactobacillus Sakei* Probio 65 with Antioxidant,  $\alpha$ -Glucosidase and Tyrosinase Inhibitory Potential. *Journal of Food Biochemistry*, **40**(3): 264-274.
- Bao, Y., Zhang, Y., Zhang, Y., Liu, Y., Wang, S., Dong, X., Wang, Y., and Zhang, H. (2010). Screening of potential probiotic properties of *Lactobacillus fermentum* isolated from traditional dairy products. *Food Control*, **21**(5): 695-701.
- Barrow, G. I., and Cowan, F. R. (1993). *Steel's manual for the identification of medical bacteria*: Cambridge, UK: Cambridge University Press.
- Begley, M., Hill, C., and Gahan, C. G. (2006). Bile salt hydrolase activity in probiotics. *Applied and Environmental Microbiology*, **72**(3): 1729-1738.
- Bejar, W., Gabriel, V., Amari, M., Morel, S., Mezghani, M., Maguin, E., Fontagné-Faucher, C., Bejar, S., and Chouayekh, H. (2013). Characterization of glucansucrase and dextran from *Weissella* sp. TN610 with potential as safe food additives. *International Journal of Biological Macromolecules*, **52**: 125-132.
- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., and Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, **76**(5): 965-977.
- Blandino, A., Al-Aseeri, M. E., Pandiella, S. S., Cantero, D., and Webb, C. (2003). Cereal-based fermented foods and beverages. *Food Research International*, **36**(6): 527-543.

- Bosch, M., Rodriguez, M., Garcia, F., Fernández, E., Fuentes, M., and Cuñé, J. (2012). Probiotic properties of *Lactobacillus plantarum* CECT 7315 and CECT 7316 isolated from faeces of healthy children. *Letters in Applied Microbiology*, **54**(3): 240-246.
- Bothara, S. B., and Singh, S. (2012). Thermal studies on natural polysaccharide. *Asian Pacific Journal of Tropical Biomedicine*, **2**(2): S1031-S1035.
- Bottazzi, V. (1986). Absorption of cholesterol by intestinal lactic acid bacteria. *Annal Microbiology*, **36**: 1-5.
- Bounaix, M., Gabriel, V. r., Morel, S., Robert, H., Rabier, P., Remaud-Siméon, M., Gabriel, B., and Fontagné-Faucher, C. (2009). Biodiversity of exopolysaccharides produced from sucrose by sourdough lactic acid bacteria. *Journal of Agricultural and Food Chemistry*, **57**(22): 10889-10897.
- Bounaix, M. S., Gabriel, V., Robert, H., Morel, S., Remaud-Simeon, M., Gabriel, B., and Fontagne-Faucher, C. (2010a). Characterization of glucan-producing Leuconostoc strains isolated from sourdough. *International Journal of Food Microbiology*, **144**(1): 1-9.
- Bounaix, M. S., Robert, H., Gabriel, V., Morel, S., Remaud-Simeon, M., Gabriel, B., and Fontagne-Faucher, C. (2010b). Characterization of dextran-producing Weissella strains isolated from sourdoughs and evidence of constitutive dextransucrase expression. *FEMS Microbiology Letters*, **311**(1): 18-26.
- Bourdichon, F., Casaregola, S., Farrokh, C., Frisvad, J. C., Gerds, M. L., Hammes, W. P., Harnett, J., Huys, G., Laulund, S., Ouwehand, A., Powell, I. B., Prajapati, J. B., Seto, Y., Ter Schure, E., Van Boven, A., Vankerckhoven, V., Zgoda, A., Tuijtelaars, S., and Hansen, E. B. (2012). Food fermentations: microorganisms with technological beneficial use. *International Journal of Food Microbiology*, **154**(3): 87-97.
- Bouzar, F., Cerning, J., and Desmazeaud, M. (1996). Exopolysaccharide Production in Milk by *Lactobacillus delbrueckii* ssp. *bulgaricus* CNRZ 1187 and by Two Colonial Variants. *Journal of Dairy Science*, **79**(2): 205-211.
- Buddington, R. (2009). Using probiotics and prebiotics to manage the gastrointestinal tract ecosystem *Prebiotics and Probiotics Science and Technology* (pp. 1-31): Springer.
- Caggianiello, G., Kleerebezem, M., and Spano, G. (2016). Exopolysaccharides produced by lactic acid bacteria: from health-promoting benefits to stress tolerance mechanisms. *Applied Microbiology and Biotechnology*, **100**(9): 3877-3886.

- Casas López, J. L., Sánchez Pérez, J. A., Fernández Sevilla, J. M., Acién Fernández, F. G., Molina Grima, E., and Chisti, Y. (2003). Production of lovastatin by *Aspergillus terreus*: effects of the C:N ratio and the principal nutrients on growth and metabolite production. *Enzyme and Microbial Technology*, **33**(2-3): 270-277.
- Castro-Bravo, N., Sánchez, B., Margolles, A., and Ruas-Madiedo, P. (2018). Biological Activities and Applications of Bifidobacterial Exopolysaccharides: From the Bacteria and Host Perspective *The Bifidobacteria and Related Organisms* (pp. 177-193): Elsevier.
- Cerning, J. (1990). Exocellular polysaccharides produced by lactic acid bacteria. *FEMS Microbiology Reviews*, **7**(1-2): 113-130.
- Cerning, J. (1995). Production of exopolysaccharides by lactic acid bacteria and dairy propionibacteria. *Le Lait*, **75**(4-5): 463-472.
- Chen, Z.-Y., Ma, K. Y., Liang, Y., Peng, C., and Zuo, Y. (2011). Role and classification of cholesterol-lowering functional foods. *Journal of Functional Foods*, **3**(2): 61-69.
- Chen, Z., Shi, J., Yang, X., Nan, B., Liu, Y., and Wang, Z. (2015). Chemical and physical characteristics and antioxidant activities of the exopolysaccharide produced by Tibetan kefir grains during milk fermentation. *International Dairy Journal*, **43**: 15-21.
- Chuah, L. O., Shamila-Syuhada, A. K., Liong, M. T., Rosma, A., Thong, K. L., and Rusul, G. (2016). Physio-chemical, microbiological properties of tempoyak and molecular characterisation of lactic acid bacteria isolated from tempoyak. *Food Microbiology*, **58**: 95-104.
- Coetzee, J. C. J. (2007). *Increased production of bacST4SA by Enterococcus mundtii in an industrial-based medium with pH-control*. Stellenbosch: University of Stellenbosch.
- Collado, M. C., Meriluoto, J., and Salminen, S. (2007). Adhesion and aggregation properties of probiotic and pathogen strains. *European Food Research and Technology*, **226**(5): 1065-1073.
- Cote, G. L., Skory, C. D., Unser, S. M., and Rich, J. O. (2013). The production of glucans via glucansucrases from *Lactobacillus satsumensis* isolated from a fermented beverage starter culture. *Applied Microbiology and Biotechnology*, **97**(16): 7265-7273.
- D'Aimmo, M. R., Modesto, M., and Biavati, B. (2007). Antibiotic resistance of lactic acid bacteria and *Bifidobacterium* spp. isolated from dairy and pharmaceutical products. *International Journal of Food Microbiology*, **115**(1): 35-42.

- Dabour, N., and LaPointe, G. (2005). Identification and molecular characterization of the chromosomal exopolysaccharide biosynthesis gene cluster from *Lactococcus lactis* subsp. *cremoris* SMQ-461. *Applied and Environmental Microbiology*, **71**(11): 7414-7425.
- Danielsen, M., and Wind, A. (2003). Susceptibility of *Lactobacillus* spp. to antimicrobial agents. *International Journal of Food Microbiology*, **82**(1): 1-11.
- Das, D., and Goyal, A. (2014). Isolation, purification and functional characterization of glucansucrase from probiotic *Lactobacillus plantarum* DM5. *Annals of Microbiology*, **64**(4): 1715-1724.
- De Palencia, P. F., López, P., Corbí, A. L., Peláez, C., and Requena, T. (2008). Probiotic strains: survival under simulated gastrointestinal conditions, in vitro adhesion to Caco-2 cells and effect on cytokine secretion. *European Food Research and Technology*, **227**(5): 1475-1484.
- De Vuyst, L., and Degeest, B. (1999). Heteropolysaccharides from lactic acid bacteria. *FEMS Microbiology Reviews*, **23**(2): 153-177.
- Dec, M., Urban-Chmiel, R., Gnat, S., Puchalski, A., and Wernicki, A. (2014). Identification of *Lactobacillus* strains of goose origin using MALDI-TOF mass spectrometry and 16S-23S rDNA intergenic spacer PCR analysis. *Research in Microbiology*, **165**(3): 190-201.
- Degeest, B., Janssens, B., and De Vuyst, L. (2001). Exopolysaccharide (EPS) biosynthesis by *Lactobacillus sakei* 0-1: production kinetics, enzyme activities and EPS yields. *Journal of Applied Microbiology*, **91**(3): 470-477.
- Del Piano, M., Morelli, L., Strozzi, G., Allesina, S., Barba, M., Deidda, F., Lorenzini, P., Ballare, M., Montino, F., and Orsello, M. (2006). Probiotics: from research to consumer. *Digestive and Liver Disease*, **38**: S248-S255.
- Del Re, B., Sgorbati, B., Miglioli, M., and Palenzona, D. (2000). Adhesion, autoaggregation and hydrophobicity of 13 strains of *Bifidobacterium longum*. *Letters in applied microbiology*, **31**(6): 438-442.
- Dertli, E., Colquhoun, I. J., Gunning, A. P., Bongaerts, R. J., Le Gall, G., Bonev, B. B., Mayer, M. J., and Narbad, A. (2013). Structure and biosynthesis of two exopolysaccharides produced by *Lactobacillus johnsonii* FI9785. *Journal of Biological Chemistry*, **288**(44): 31938-31951.
- Dertli, E., Mercan, E., Arıcı, M., Yılmaz, M. T., and Sağdıç, O. (2016). Characterisation of lactic acid bacteria from Turkish sourdough and determination of their exopolysaccharide (EPS) production characteristics. *LWT - Food Science and Technology*, **71**: 116-124.
- Desai, A. R., Shah, N. P., and Powell, I. B. (2006). Discrimination of dairy industry isolates of the *Lactobacillus casei* group. *Journal of Dairy Science*, **89**(9): 3345-3351.

- Deveau, H., and Moineau, S. (2003). Use of RFLP to characterize *Lactococcus lactis* strains producing exopolysaccharides. *Journal of Dairy Science*, **86**(4): 1472-1475.
- Dimitonova, S. P., Bakalov, B. V., Aleksandrova-Georgieva, R. N., and Danova, S. T. (2008). Phenotypic and molecular identification of lactobacilli isolated from vaginal secretions. *Journal of Microbiology, Immunology and Infection*, **41**(6): 469-477.
- Doyle, R. J., and Rosenberg, M. (1995). Measurement of microbial adhesion to hydrophobic substrata. *Methods in Enzymology*, **253**: 542-550.
- Dubernet, S. g. n., Desmases, N., and GuÃ©guen, M. (2002). A PCR-based method for identification of lactobacilli at the genus level. *FEMS Microbiology Letters*, **214**(2): 271-275.
- DuBois, M., Gilles, K. A., Hamilton, J. K., Rebers, P., and Smith, F. (1956). Colorimetric method for determination of sugars and related substances. *Analytical Chemistry*, **28**: 350-356.
- Dunne, C., O'Mahony, L., Murphy, L., Thornton, G., Morrissey, D., O'Halloran, S., Feeney, M., Flynn, S., Fitzgerald, G., and Daly, C. (2001). In vitro selection criteria for probiotic bacteria of human origin: correlation with in vivo findings. *American Journal of Clinical Nutrition*, **73**(2): 386-392.
- Duskova, M., Sedo, O., Ksicova, K., Zdrahal, Z., and Karpiskova, R. (2012). Identification of lactobacilli isolated from food by genotypic methods and MALDI-TOF MS. *International Journal of Food Microbiology*, **159**(2): 107-114.
- Ehrmann, M. A., and Vogel, R. F. (2005). Molecular taxonomy and genetics of sourdough lactic acid bacteria. *Trends in Food Science & Technology*, **16**(1-3): 31-42.
- Fan, Y., He, X., Zhou, S., Luo, A., He, T., and Chun, Z. (2009). Composition analysis and antioxidant activity of polysaccharide from *Dendrobium denneanum*. *International Journal of Biological Macromolecules*, **45**(2): 169-173.
- Ferreira, C. L., Grzeskowiak, L., Collado, M. C., and Salminen, S. (2011). In vitro evaluation of *Lactobacillus gasseri* strains of infant origin on adhesion and aggregation of specific pathogens. *Journal of food protection*, **74**(9): 1482-1487.
- Frengova, G. I., Simova, E. D., Beshkova, D. M., and Simov, Z. I. (2000). Production and monomer composition of exopolysaccharides by yogurt starter cultures. *Canadian Journal of Microbiology*, **46**(12): 1123-1127.
- Fuller, R. (1992). History and development of probiotics *Probiotics* (pp. 1-8): Springer, Dordrecht.

- Fung, W.-Y., Lye, H.-S., Lim, T.-J., Kuan, C.-Y., and Lioung, M.-T. (2011). Roles of Probiotic on Gut Health *Probiotics* (pp. 139-165): Springer.
- Gancel, F., and Novel, G. (1994). Exopolysaccharide Production by *Streptococcus salivarius* ssp. *thermophilus* Cultures. 1. Conditions of Production. *Journal of Dairy Science*, **77**(3): 685-688.
- Gassem, M. A., Schmidt, K. A., and Frank, J. F. (1997). Exopolysaccharide production from whey lactose by fermentation with *Lactobacillus delbrueckii* ssp. *bulgaricus*. *Journal of Food Science*, **62**(1): 171-173.
- Gevers, D., Huys, G., and Swings, J. (2001). Applicability of rep-PCR fingerprinting for identification of *Lactobacillus* species. *FEMS Microbiology Letters*, **205**(1): 31-36.
- Gilliland, S. E., Staley, T. E., and Bush, L. J. (1984). Importance of bile tolerance of *Lactobacillus acidophilus* used as a dietary adjunct. *Journal of Dairy Science*, **67**(12): 3045-3051.
- Goswami, G., Bora, S. S., Parveen, A., Boro, R. C., and Barooah, M. (2017). Identification and functional properties of dominant lactic acid bacteria isolated from Kahudi, a traditional rapeseed fermented food product of Assam, India. *Journal of Ethnic Foods*, **4**(3): 187-197.
- Granito, M., Frias, J., Doblado, R., Guerra, M., Champ, M., and Vidal-Valverde, C. (2002). Nutritional improvement of beans (*Phaseolus vulgaris*) by natural fermentation. *European Food Research and Technology*, **214**(3): 226-231.
- Gueimonde, M., Sánchez, B., de los Reyes-Gavilán, C. G., and Margolles, A. (2013). Antibiotic resistance in probiotic bacteria. *Frontier Microbiology*, **4**: 202.
- Haj-Mustafa, M., Abdi, R., Sheikh-Zeinoddin, M., and Soleimanian-Zad, S. (2015). Statistical study on fermentation conditions in the optimization of exopolysaccharide production by *Lactobacillus rhamnosus* 519 in skimmed milk base media. *Biocatalysis and Agricultural Biotechnology*, **4**(4): 521-527.
- Hall, T. A. (1999). *BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT*. Paper presented at the Nucleic Acids Symposium Series.
- Haller, D., Colbus, H., Gänzle, M., Scherenbacher, P., Bode, C., and Hammes, W. (2001). Metabolic and functional properties of lactic acid bacteria in the gastrointestinal ecosystem: a comparative in vitro study between bacteria of intestinal and fermented food origin. *Systematic and Applied Microbiology*, **24**(2): 218-226.
- Harutoshi, T. (2013). Exopolysaccharides of lactic acid bacteria for food and colon health applications *Lactic Acid Bacteria-R & D for Food, Health and Livestock Purposes*: InTech.

- Harzallah, D., and Belhadj, H. (2013). Lactic acid bacteria as probiotics: characteristics, selection criteria and role in immunomodulation of human GI mucosal barrier. In M. Kongo (Ed.), *Lactic Acid Bacteria – R & D for Food, Health and Livestock Purposes* (pp. 197-216). Rijeka, Croatia: InTech.
- Heller, K. J. (2001). Probiotic bacteria in fermented foods: product characteristics and starter organisms. *American Journal of Clinical Nutrition*, **73**(2): 374-379.
- Herreros, M. A., Fresno, J. M., González Prieto, M. J., and Tornadijo, M. E. (2003). Technological characterization of lactic acid bacteria isolated from Armada cheese (a Spanish goats' milk cheese). *International Dairy Journal*, **13**(6): 469-479.
- Hidalgo-Cantabrana, C., López, P., Gueimonde, M., Clara, G., Suárez, A., Margolles, A., and Ruas-Madiedo, P. (2012). Immune modulation capability of exopolysaccharides synthesised by lactic acid bacteria and bifidobacteria. *Probiotics and Antimicrobial Proteins*, **4**(4): 227-237.
- Hor, Y. Y., and Lioung, M. T. (2014). Use of extracellular extracts of lactic acid bacteria and bifidobacteria for the inhibition of dermatological pathogen *Staphylococcus aureus*. *Dermatologica Sinica*, **32**(3): 141-147.
- Horn, N., Wegmann, U., Dertli, E., Mulholland, F., Collins, S. R., Waldron, K. W., Bongaerts, R. J., Mayer, M. J., and Narbad, A. (2013). Spontaneous mutation reveals influence of exopolysaccharide on *Lactobacillus johnsonii* surface characteristics. *PloS One*, **8**(3): e59957.
- Hosono, A., and Tono-Oka, T. (1995). Binding of cholesterol with lactic acid bacterial cells. *Milchwissenschaft (Germany)*.
- Hugenholtz, J. (2008). The lactic acid bacterium as a cell factory for food ingredient production. *International Dairy Journal*, **18**(5): 466-475.
- Ibarburu, I., Puertas, A. I., Berregi, I., Rodriguez-Carvajal, M. A., Prieto, A., and Duenas, M. T. (2015). Production and partial characterization of exopolysaccharides produced by two *Lactobacillus suebicus* strains isolated from cider. *International Journal of Food Microbiology*, **214**: 54-62.
- Imran, M. Y., Reehana, N., Jayaraj, K. A., Ahamed, A. A., Dhanasekaran, D., Thajuddin, N., Alharbi, N. S., and Muralitharan, G. (2016). Statistical optimization of exopolysaccharide production by *Lactobacillus plantarum* NTMI05 and NTMI20. *International Journal of Biological Macromolecules*, **93**(Pt A): 731-745.
- Ismail, B., and Nampoothiri, K. M. (2010). Production, purification and structural characterization of an exopolysaccharide produced by a probiotic *Lactobacillus plantarum* MTCC 9510. *Archives of Microbiology*, **192**(12): 1049-1057.

- Itsaranuwat, P., Al-Haddad, K. S., and Robinson, R. (2003). The potential therapeutic benefits of consuming ‘health-promoting’ fermented dairy products: a brief update. *International Journal of Dairy Technology*, **56**(4): 203-210.
- Jahromi, M. F., Liang, J. B., Abdullah, N., Goh, Y. M., Ebrahimi, R., and Shokryazdan, P. (2015). Extraction and characterization of oligosaccharides from palm kernel cake as prebiotic. *BioResources*, **11**(1): 674-695.
- Jb Prajapat, A. P. (2013). Food and Health Applications of Exopolysaccharides produced by Lactic acid Bacteria. *Advances in Dairy Research*, **01**(02): 1-8.
- Jin, L. Z., Ho, Y. W., Abdullah, N., Ali, M., and Jalaludin, S. (1996). Antagonistic effects of intestinal *Lactobacillus* isolates on pathogens of chicken. *Letters in Applied Microbiology*, **23**(2): 67-71.
- Joint, F. (2002). WHO working group report on drafting guidelines for the evaluation of probiotics in food. *London, Ontario, Canada*, **30**.
- Jolly, L., Newell, J., Porcelli, I., Vincent, S. J., and Stingele, F. (2002). *Lactobacillus helveticus* glycosyltransferases: from genes to carbohydrate synthesis. *Glycobiology*, **12**(5): 319-327.
- Kanmani, P., Satish Kumar, R., Yuvaraj, N., Paari, K., Pattukumar, V., and Arul, V. (2013). Probiotics and its functionally valuable products—A review. *Critical Reviews in Food Science and Nutrition*, **53**(6): 641-658.
- Kanmani, P., Satish kumar, R., Yuvaraj, N., Paari, K. A., Pattukumar, V., and Arul, V. (2011). Production and purification of a novel exopolysaccharide from lactic acid bacterium *Streptococcus phocae* PI80 and its functional characteristics activity in vitro. *Bioresource Technology*, **102**(7): 4827-4833.
- Kaushik, J. K., Kumar, A., Duary, R. K., Mohanty, A. K., Grover, S., and Batish, V. K. (2009). Functional and probiotic attributes of an indigenous isolate of *Lactobacillus plantarum*. *PloS One*, **4**(12): e8099.
- Kilinc, B., Cakli, S., Tolasa, S., and Dincer, T. (2006). Chemical, microbiological and sensory changes associated with fish sauce processing. *European Food Research and Technology*, **222**(5-6): 604-613.
- Kim, Y., Kim, J. U., Oh, S., Kim, Y. J., Kim, M., and Kim, S. H. (2008a). Technical optimization of culture conditions for the production of exopolysaccharide (EPS) by *Lactobacillus rhamnosus* ATCC 9595. *Food Science and Biotechnology*, **17**(3): 587-593.
- Kim, Y., Whang, J. Y., Whang, K. Y., Oh, S., and Kim, S. H. (2008b). Characterization of the cholesterol-reducing activity in a cell-free supernatant of *Lactobacillus acidophilus* ATCC 43121. *Bioscience, Biotechnology, and Biochemistry*, **72**(6): 1483-1490.

- Klaver, F. A., and Van Der Meer, R. (1993). The assumed assimilation of cholesterol by Lactobacilli and Bifidobacterium bifidum is due to their bile salt-deconjugating activity. *Applied and Environmental Microbiology*, **59**(4): 1120-1124.
- Korakli, M., and Vogel, R. F. (2006). Structure/function relationship of homopolysaccharide producing glycansucrases and therapeutic potential of their synthesised glycans. *Applied Microbiology and Biotechnology*, **71**(6): 790-803.
- Kormin, S., Rusul, G., Radu, S., and Ling, F. H. (2001). Bacteriocin-producing lactic Acid bacteria isolated from traditional fermented food. *The Malaysian journal of medical sciences: MJMS*, **8**(1): 63.
- Kos, B., Šušković, J., Vuković, S., Šimpraga, M., Frece, J., and Matosić, S. (2003). Adhesion and aggregation ability of probiotic strain *Lactobacillus acidophilus* M92. *Journal of Applied Microbiology*, **94**(6): 981-987.
- Kralj, S., van Geel-Schutten, G., Dondorff, M., Kirsanovs, S., Van Der Maarel, M., and Dijkhuizen, L. (2004a). Glucan synthesis in the genus *Lactobacillus*: isolation and characterization of glucansucrase genes, enzymes and glucan products from six different strains. *Microbiology*, **150**(11): 3681-3690.
- Kralj, S., van Geel-Schutten, G., Van Der Maarel, M., and Dijkhuizen, L. (2004b). Biochemical and molecular characterization of *Lactobacillus reuteri* 121 reuteransucrase. *Microbiology*, **150**(7): 2099-2112.
- Ku, S., Park, M. S., Ji, G. E., and You, H. J. (2016). Review on *Bifidobacterium bifidum* BGN4: Functionality and Nutraceutical Applications as a Probiotic Microorganism. *International journal of molecular sciences*, **17**(9): 1544.
- Ku, S., You, H. J., and Ji, G. E. (2009). Enhancement of anti-tumorigenic polysaccharide production, adhesion, and branch formation of *Bifidobacterium bifidum* BGN4 by phytic acid. *Food Science and Biotechnology*, **18**(3): 749-754.
- Kullisaar, T., Zilmer, M., Mikelsaar, M., Vihalemm, T., Annuk, H., Kairane, C., and Kilk, A. (2002). Two antioxidative lactobacilli strains as promising probiotics. *International Journal of Food Microbiology*, **72**(3): 215-224.
- Kumar, A. S., and Mody, K. (2009). Microbial exopolysaccharides: variety and potential applications. In B. H. A. Rehm (Ed.), *Microbial production of biopolymers and polymer precursors: applications and perspectives* (pp. 229-253). Norfolk, UK: Caister Academic Press.
- Kumar, M., Nagpal, R., Kumar, R., Hemalatha, R., Verma, V., Kumar, A., Chakraborty, C., Singh, B., Marotta, F., and Jain, S. (2012). Cholesterol-lowering probiotics as potential biotherapeutics for metabolic diseases. *Experimental Diabetes Research*, **2012**.

- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, **227**(5259): 680-685.
- Lai, Y.-J., Tsai, S.-H., and Lee, M.-Y. (2014). Isolation of exopolysaccharide producing *Lactobacillus* strains from sorghum distillery residues pickled cabbage and their antioxidant properties. *Food Science and Biotechnology*, **23**(4): 1231-1236.
- Lane, D. J. (1991). *16S/23S rRNA sequencing. In 'Nucleic acid techniques in bacterial systematics'*. (Eds E Stackebrandt, M Goodfellow) pp. 115–175: John Wiley and Sons: Chichester, UK.
- Laws, A., Gu, Y., and Marshall, V. (2001). Biosynthesis, characterisation, and design of bacterial exopolysaccharides from lactic acid bacteria. *Biotechnology Advances*, **19**(8): 597-625.
- Lee, B.-J., Kim, J.-S., Kang, Y. M., Lim, J.-H., Kim, Y.-M., Lee, M.-S., Jeong, M.-H., Ahn, C.-B., and Je, J.-Y. (2010). Antioxidant activity and  $\gamma$ -aminobutyric acid (GABA) content in sea tangle fermented by *Lactobacillus brevis* BJ20 isolated from traditional fermented foods. *Food Chemistry*, **122**(1): 271-276.
- Lee, C.-H. (1997). Lactic acid fermented foods and their benefits in Asia. *Food Control*, **8**(5-6): 259-269.
- Lee, J., Hwang, K. T., Heo, M. S., Lee, J. H., and Park, K. Y. (2005). Resistance of *Lactobacillus plantarum* KCTC 3099 from Kimchi to oxidative stress. *Journal of Medicinal Food*, **8**(3): 299-304.
- Lee, Y.-K., and Salminen, S. (1995). The coming of age of probiotics. *Trends in Food Science and Technology*, **6**(7): 241-245.
- Leemhuis, H., Pijning, T., Dobruchowska, J. M., van Leeuwen, S. S., Kralj, S., Dijkstra, B. W., and Dijkhuizen, L. (2013). Glucansucrases: Three-dimensional structures, reactions, mechanism,  $\alpha$ -glucan analysis and their implications in biotechnology and food applications. *Journal of Biotechnology*, **163**(2): 250-272.
- Leisner, J. J., Pot, B., Christensen, H., Rusul, G., Olsen, J. E., Wee, B. W., Muhamad, K., and Ghazali, H. M. (1999). Identification of lactic acid bacteria from chili bo, a Malaysian food ingredient. *Applied and Environmental Microbiology*, **65**(2): 599-605.
- Leisner, J. J., Vancanneyt, M., Lefebvre, K., Vandemeulebroecke, K., Hoste, B., Vilalta, N. E., Rusul, G., and Swings, J. (2002). *Lactobacillus durianis* sp. nov., isolated from an acid-fermented condiment (tempoyak) in Malaysia. *Applied and Environmental Microbiology*, **52**(3): 927-931.
- Leisner, J. J., Vancanneyt, M., Rusul, G., Pot, B., Lefebvre, K., Fresi, A., and Tee, L. K. (2001). Identification of lactic acid bacteria constituting the predominating microflora in an acid-fermented condiment (tempoyak) popular in Malaysia. *International Journal of Food Microbiology*, **63**(1): 149-157.

- Leroy, F., and De Vuyst, L. (2004). Lactic acid bacteria as functional starter cultures for the food fermentation industry. *Trends in Food Science & Technology*, **15**(2): 67-78.
- Leroy, F., and De Vuyst, L. (2016). Advances in production and simplified methods for recovery and quantification of exopolysaccharides for applications in food and health. *Journal of Dairy Science*, **99**(4): 3229-3238.
- Levett, P. (1985). Identification of *Clostridium difficile* using the API ZYM system. *European Journal of Clinical Microbiology*, **4**(5): 505-507.
- Li, S., Zhao, Y., Zhang, L., Zhang, X., Huang, L., Li, D., Niu, C., Yang, Z., and Wang, Q. (2012). Antioxidant activity of *Lactobacillus plantarum* strains isolated from traditional Chinese fermented foods. *Food Chemistry*, **135**(3): 1914-1919.
- Li, W., Ji, J., Chen, X., Jiang, M., Rui, X., and Dong, M. (2014a). Structural elucidation and antioxidant activities of exopolysaccharides from *Lactobacillus helveticus* MB2-1. *Carbohydrate Polymers*, **102**: 351-359.
- Li, W., Ji, J., Tang, W., Rui, X., Chen, X., Jiang, M., and Dong, M. (2014b). Characterization of an antiproliferative exopolysaccharide (LHEPS-2) from *Lactobacillus helveticus* MB2-1. *Carbohydrate Polymers*, **105**: 334-340.
- Li, W., Mutuvulla, M., Chen, X., Jiang, M., and Dong, M. (2012). Isolation and identification of high viscosity-producing lactic acid bacteria from a traditional fermented milk in Xinjiang and its role in fermentation process. *European Food Research and Technology*, **235**(3): 497-505.
- Li, Y., Li, Q., Hao, D., Jiang, D., Luo, Y., Liu, Y., and Zhao, Z. (2015). Production, purification, and antibiofilm activity of a novel exopolysaccharide from *Arthrobacter* sp. B4. *Preparative Biochemistry and Biotechnology*, **45**(2): 192-204.
- Liang, T. W., Tseng, S. C., and Wang, S. L. (2016). Production and Characterization of Antioxidant Properties of Exopolysaccharide(s) from *Peanibacillus mucilaginosus* TKU032. *Marine Drugs*, **14**(2): 40.
- Lilly, D. M., and Stillwell, R. H. (1965). Probiotics: growth-promoting factors produced by microorganisms. *Science*, **147**(3659): 747-748.
- Lin, E. S., and Chen, Y. H. (2007). Factors affecting mycelial biomass and exopolysaccharide production in submerged cultivation of *Antrodia cinnamomea* using complex media. *Bioresource Technology*, **98**(13): 2511-2517.
- Liong, M. T., and Shah, N. P. (2006). Effects of a *Lactobacillus casei* symbiotic on serum lipoprotein, intestinal microflora, and organic acids in rats. *Journal of Dairy Science*, **89**(5): 1390-1399.

- Liu, C. F., Tseng, K. C., Chiang, S. S., Lee, B. H., Hsu, W. H., and Pan, T. M. (2011). Immunomodulatory and antioxidant potential of *Lactobacillus* exopolysaccharides. *Journal of the Science of Food and Agriculture*, **91**(12): 2284-2291.
- Liu, W. J., Chen, Y. F., Kwok, L. Y., Li, M. H., Sun, T., Sun, C. L., Wang, X. N., Dan, T., Zhang, H. P., and Sun, T. S. (2013). Preliminary selection for potential probiotic *Bifidobacterium* isolated from subjects of different Chinese ethnic groups and evaluation of their fermentation and storage characteristics in bovine milk. *Journal of Dairy Science*, **96**(11): 6807-6817.
- Looijesteijn, P. J., Trapet, L., de Vries, E., Abee, T., and Hugenholtz, J. (2001). Physiological function of exopolysaccharides produced by *Lactococcus lactis*. *International Journal of Food Microbiology*, **64**(1): 71-80.
- Low, D., Ahlgren, J. A., Horne, D., McMahon, D. J., Oberg, C. J., and Broadbent, J. R. (1998). Role of *Streptococcus thermophilus* MR-1C capsular exopolysaccharide in cheese moisture retention. *Applied and Environmental Microbiology*, **64**(6): 2147-2151.
- Lundstedt, T., Seifert, E., Abramo, L., Thelin, B., Nyström, Å., Pettersen, J., and Bergman, R. (1998). Experimental design and optimization. *Chemometr. Intell. Lab. Syst.*, **42**(1-2): 3-40.
- Lye, H.-S., Rahmat-Ali, G. R., and Liong, M.-T. (2010). Mechanisms of cholesterol removal by lactobacilli under conditions that mimic the human gastrointestinal tract. *International Dairy Journal*, **20**(3): 169-175.
- Lye, H. S., Rusul, G., and Liang, M. T. (2010). Removal of cholesterol by lactobacilli via incorporation and conversion to coprostanol. *Journal of Dairy Science*, **93**(4): 1383-1392.
- Macedo, M., Lacroix, C., Gardner, N., and Champagne, C. (2002). Effect of medium supplementation on exopolysaccharide production by *Lactobacillus rhamnosus* RW-9595M in whey permeate. *International Dairy Journal*, **12**(5): 419-426.
- Macedo, M. G., Lacroix, C., and Champagne, C. P. (2002). Combined effects of temperature and medium composition on exopolysaccharide production by *Lactobacillus rhamnosus* RW-9595M in a whey permeate based medium. *Biotechnology Progress*, **18**(2): 167-173.
- Madani, G., Mirlohi, M., Yahay, M., and Hassanzadeh, A. (2013). How much in vitro cholesterol reducing activity of Lactobacilli predicts their in vivo cholesterol function? *International Journal of Preventive Medicine*, **4**(4).
- Maeda, H., Zhu, X., Omura, K., Suzuki, S., and Kitamura, S. (2004). Effects of an exopolysaccharide (kefiran) on lipids, blood pressure, blood glucose, and constipation. *BioFactors*, **22**(14): 197-200.

- Mainville, I., Arcand, Y., and Farnworth, E. R. (2005). A dynamic model that simulates the human upper gastrointestinal tract for the study of probiotics. *International Journal of Food Microbiology*, **99**(3): 287-296.
- Majumder, A., Mangtani, A., and Goyal, A. (2008). Purification, identification and functional characterization of glucansucrase from *Leuconostoc dextranicum* NRRL B-1146. *Current Trends in Biotechnology and Pharmacy*, **2**(4): 493-505.
- Malang, S. K., Maina, N. H., Schwab, C., Tenkanen, M., and Lacroix, C. (2015). Characterization of exopolysaccharide and ropy capsular polysaccharide formation by *Weissella*. *Food Microbiology*, **46**: 418-427.
- Marteau, P. M., Minekus, M., Havenga, R., and Huis, J. H. J. (1997). Survival of lactic acid bacteria in a dynamic model of the stomach and small intestine: validation and the effects of bile. *Journal of Dairy Science*, **80**(6): 1031-1037.
- Mathara, J. M., Schillinger, U., Guigas, C., Franz, C., Kutima, P. M., Mbugua, S. K., Shin, H.-K., and Holzapfel, W. H. (2008). Functional characteristics of *Lactobacillus* spp. from traditional Maasai fermented milk products in Kenya. *International Journal of Food Microbiology*, **126**(1): 57-64.
- Mazzoli, R., Bosco, F., Mizrahi, I., Bayer, E. A., and Pessione, E. (2014). Towards lactic acid bacteria-based biorefineries. *Biotechnology Advances*, **32**(7): 1216-1236.
- McDonald, I. R., Kenna, E. M., and Murrell, J. C. (1995). *Detection of methanotrophic bacteria in environmental samples with the PCR* (Vol. 61).
- Mende, S., Rohm, H., and Jaros, D. (2016). Influence of exopolysaccharides on the structure, texture, stability and sensory properties of yoghurt and related products. *International Dairy Journal*, **52**: 57-71.
- Metchnikoff, I. I. (2004). *The prolongation of life: optimistic studies*: Springer Publishing Company.
- Miao, M., Bai, A., Jiang, B., Song, Y., Cui, S. W., and Zhang, T. (2014). Characterisation of a novel water-soluble polysaccharide from *Leuconostoc citreum* SK24.002. *Food Hydrocolloids*, **36**: 265-272.
- Miao, M., Huang, C., Jia, X., Cui, S. W., Jiang, B., and Zhang, T. (2015). Physicochemical characteristics of a high molecular weight bioengineered  $\alpha$ -D-glucan from *Leuconostoc citreum* SK24.002. *Food Hydrocolloids*, **50**: 37-43.
- Mikelsaar, M., and Zilmer, M. (2009). *Lactobacillus fermentum* ME-3—an antimicrobial and antioxidative probiotic. *Microbial Ecology in Health and Disease*, **21**(1): 1-27.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*, **31**(3): 426-428.

- Miller, N. J., Rice-Evans, C., Davies, M. J., Gopinathan, V., and Milner, A. (1993). A novel method for measuring antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. *Clinical Science*, **84**(4): 407-412.
- Miremadi, F., Ayyash, M., Sherkat, F., and Stojanovska, L. (2014). Cholesterol reduction mechanisms and fatty acid composition of cellular membranes of probiotic Lactobacilli and Bifidobacteria. *Journal Functional Foods*, **9**: 295-305.
- Mirhosseini, H., and Tan, C. P. (2009). Response surface methodology and multivariate analysis of equilibrium headspace concentration of orange beverage emulsion as function of emulsion composition and structure. *Food Chemistry*, **115**(1): 324-333.
- Mishra, A., and Jha, B. (2009). Isolation and characterization of extracellular polymeric substances from micro-algae Dunaliellasalina under salt stress. *Bioresource Technology*, **100**(13): 3382-3386.
- Mohd Adnan, A. F., and Tan, I. K. (2007). Isolation of lactic acid bacteria from Malaysian foods and assessment of the isolates for industrial potential. *Bioresource Technology*, **98**(7): 1380-1385.
- Montgomery, D. C. (2001). Design and Analysis of Experiments, John Wiley & Sons. New York: 64-65.
- Moreira, J. L., Mota, R. M., Horta, M. F., Teixeira, S. M., Neumann, E., Nicoli, J. R., and Nunes, A. C. (2005). Identification to the species level of *Lactobacillus* isolated in probiotic prospecting studies of human, animal or food origin by 16S-23S rRNA restriction profiling. *BMC Microbiology*, **5**(1): 15.
- Morelli, L. (2007). In vitro assessment of probiotic bacteria: From survival to functionality. *International Dairy Journal*, **17**(11): 1278-1283.
- Naidu, A., Bidlack, W., and Clemens, R. (1999). Probiotic spectra of lactic acid bacteria (LAB). *Critical Reviews in Food Science and Nutrition*, **39**(1): 13-126.
- Nampoothiri, K. M., Beena, D. J., Vasanthakumari, D. S., and Ismail, B. (2017). Health benefits of exopolysaccharides in fermented foods. In J. Frias, C. Martinez-Villaluenga & E. Peñas (Eds.), *Fermented foods in health and disease prevention* (pp. 49-62): Elsevier.
- Nelson, N. (1944). A photometric adaptation of the Somogyi method for the determination of glucose. *Journal of Biological Chemistry*, **153**(2): 375-380.
- Ng, S. C., Hart, A. L., Kamm, M. A., Stagg, A. J., and Knight, S. C. (2009). Mechanisms of action of probiotics: recent advances. *Inflammatory Bowel Diseases*, **15**(2): 300-310.

- Nishimura, J. (2014). Exopolysaccharides produced from *Lactobacillus delbrueckii* subsp. *bulgaricus*. *Advances in Microbiology*, **4**(14): 1017.
- Nivoliez, A., Veisseire, P., Alaterre, E., Dausset, C., Baptiste, F., Camarès, O., Paquet-Gachinat, M., Bonnet, M., Forestier, C., and Bornes, S. (2015). Influence of manufacturing processes on cell surface properties of probiotic strain *Lactobacillus rhamnosus* Lcr35®. *Applied Microbiology and Biotechnology*, **99**(1): 399-411.
- Noriega, L., Gueimonde, M., Sanchez, B., Margolles, A., and de los Reyes-Gavilan, C. G. (2004). Effect of the adaptation to high bile salts concentrations on glycosidic activity, survival at low PH and cross-resistance to bile salts in *Bifidobacterium*. *International Journal of Food Microbiology*, **94**(1): 79-86.
- Ohba, T., Uemura, K., and Nabetani, H. (2017). Changes in biosynthesis of exopolysaccharide in *Lactococcus lactis* subspecies *cremoris* treated by moderate pulsed electric field treatment. *Bioscience, Biotechnology, and Biochemistry*, **81**(4): 724-734.
- Oleksy, M., and Klewicka, E. (2016). Exopolysaccharides produced by *Lactobacillus* sp.: Biosynthesis and applications. *Critical Reviews in Food Science and Nutrition*, **58**(3): 450-462.
- Palomba, S., Cavella, S., Torrieri, E., Piccolo, A., Mazzei, P., Blaiotta, G., Ventorino, V., and Pepe, O. (2012). Polyphasic screening, homopolysaccharide composition, and viscoelastic behavior of wheat Sourdough from a *Leuconostoc lactis* and *Lactobacillus curvatus* exopolysaccharide-producing starter culture. *Applied and Environmental Microbiology*, **78**(8): 2737-2747.
- Parker, R. (1974). The other half of the antibiotics story. *Animal Nutrition and Health*, **29**: 4-8.
- Patel, A., Lindström, C., Patel, A., Prajapati, J., and Holst, O. (2012). Probiotic properties of exopolysaccharide producing lactic acid bacteria isolated from vegetables and traditional Indian fermented foods. *International Journal of Fermented Foods*, **1**: 87-100.
- Patel, A., Lindström, C., Patel, A., Prajapati, J., and Holst, O. (2012). Screening and isolation of exopolysaccharide producing lactic acid bacteria from vegetables and indigenous fermented foods of Gujarat, India. *International Journal of Fermented Foods*, **1**: 87-101.
- Patel, S., Kothari, D., and Goyal, A. (2011). Purification and characterization of an extracellular dextranucrase from *Pediococcus pentosaceus* isolated from the soil of North East India. *Food Technology and Biotechnology*, **49**(3): 297.
- Patel, S., Majumder, A., and Goyal, A. (2012). Potentials of exopolysaccharides from lactic acid bacteria. *Indian journal of microbiology*, **52**(1): 3-12.

- Patil, M. M., Pal, A., Anand, T., and Ramana, K. (2010). Isolation and characterization of lactic acid bacteria from curd and cucumber. *Indian Journal of Biotechnology*, **9**(2): 166-172.
- Pedersen, K., and Tannock, G. (1989). Colonization of the porcine gastrointestinal tract by lactobacilli. *Applied and Environmental Microbiology*, **55**(2): 279-283.
- Pereira, D. I., and Gibson, G. R. (2002). Cholesterol assimilation by lactic acid bacteria and bifidobacteria isolated from the human gut. *Applied and Environmental Microbiology*, **68**(9): 4689-4693.
- Peres, C. M., Alves, M., Hernandez-Mendoza, A., Moreira, L., Silva, S., Bronze, M. R., Vilas-Boas, L., Peres, C., and Malcata, F. X. (2014). Novel isolates of lactobacilli from fermented Portuguese olive as potential probiotics. *LWT - Food Science and Technology*, **59**(1): 234-246.
- Perez-Ramos, A., Mohedano, M. L., Lopez, P., Spano, G., Fiocco, D., Russo, P., and Capozzi, V. (2017). In Situ beta-Glucan Fortification of Cereal-Based Matrices by *Pediococcus parvulus* 2.6: Technological aspects and prebiotic potential. *International Journal of Molecular Science*, **18**(7): 1588.
- Pérez, P., Minnaard, Y., Disalvo, E. A., and De Antoni, G. L. (1998). Surface properties of bifidobacterial strains of human origin. *Applied and Environmental Microbiology*, **64**(1): 21-26.
- Petry, S., Furlan, S., Crepeau, M. J., Cerning, J., and Desmazeaud, M. (2000). Factors affecting exocellular polysaccharide production by *Lactobacillus delbrueckii* subsp. *bulgaricus* grown in a chemically defined medium. *Applied and Environmental Microbiology*, **66**(8): 3427-3431.
- Pham, P. L., Dupont, I., Roy, D., Lapointe, G., and Cerning, J. (2000). Production of exopolysaccharide by *Lactobacillus rhamnosus* R and analysis of its enzymatic degradation during prolonged fermentation. *Applied and Environmental Microbiology*, **66**(6): 2302-2310.
- Pidoux, M., Marshall, V. M., Zanoni, P., and Brooker, B. (1990). Lactobacilli isolated from sugary kefir grains capable of polysaccharide production and minicell formation. *Journal of Applied Microbiology*, **69**(3): 311-320.
- Pinto, M. G., Franz, C. M., Schillinger, U., and Holzapfel, W. H. (2006). *Lactobacillus* spp. with in vitro probiotic properties from human faeces and traditional fermented products. *International Journal of Food Microbiology*, **109**(3): 205-214.
- Pitino, I., Randazzo, C. L., Mandalari, G., Curto, A. L., Faulks, R. M., Le Marc, Y., Bisignano, C., Caggia, C., and Wickham, M. S. J. (2010). Survival of *Lactobacillus rhamnosus* strains in the upper gastrointestinal tract. *Food Microbiology*, **27**(8): 1121-1127.

- Polak-Berecka, M., Choma, A., Waśko, A., Górska, S., Gamian, A., and Cybulska, J. (2015). Physicochemical characterization of exopolysaccharides produced by *Lactobacillus rhamnosus* on various carbon sources. *Carbohydrate polymer*, **117**: 501-509.
- Polak-Berecka, M., Waśko, A., and Kubik-Komar, A. (2014). Optimization of culture conditions for exopolysaccharide production by a probiotic strain of *Lactobacillus rhamnosus* E/N. *Polish Journal of Microbiology*, **63**(2): 253-257.
- Prasanna, P. H. P., Grandison, A. S., and Charalampopoulos, D. (2012). Screening human intestinal *Bifidobacterium* strains for growth, acidification, EPS production and viscosity potential in low-fat milk. *International Dairy Journal*, **23**(1): 36-44.
- Provencher, C., LaPointe, G., Sirois, S., Van Calsteren, M., and Roy, D. (2003). Consensus-degenerate hybrid oligonucleotide primers for amplification of priming glycosyltransferase genes of the exopolysaccharide locus in strains of the *Lactobacillus casei* group. *Applied and Environmental Microbiology*, **69**(6): 3299-3307.
- Purama, R., and Goyal, A. (2008). Identification, effective purification and functional characterization of dextranucrase from *Leuconostoc mesenteroides* NRRL B-640. *Bioresource Technology*, **99**(9): 3635-3642.
- Remaud-Simeon, M., Willemot, R. M., Sarçabal, P., de Montalk, G. P., and Monsan, P. (2000). Glucansucrases: molecular engineering and oligosaccharide synthesis. *Journal of Molecular Catalysis B: Enzymatic*, **10**(1-3): 117-128.
- Ren, D., Li, C., Qin, Y., Yin, R., Du, S., Ye, F., Liu, C., Liu, H., Wang, M., and Li, Y. (2014). In vitro evaluation of the probiotic and functional potential of *Lactobacillus* strains isolated from fermented food and human intestine. *Anaerobe*, **30**: 1-10.
- Rendueles, O., Kaplan, J. B., and Ghigo, J. M. (2013). Antibiofilm polysaccharides. *Environmental Microbiology*, **15**(2): 334-346.
- Roca, C., Alves, V. D., Freitas, F., and Reis, M. A. (2015). Exopolysaccharides enriched in rare sugars: bacterial sources, production, and applications. *Frontier Microbiology*, **6**: 288.
- Rodríguez, E., Arqués, J. L., Rodríguez, R., Peirotén, Á., Landete, J. M., and Medina, M. (2012). Antimicrobial properties of probiotic strains isolated from breast-fed infants. *Journal of Functional Foods*, **4**(2): 542-551.
- Röling, W. F. M., Apriyantono, A., and Van Verseveld, H. W. (1996). Comparison between traditional and industrial soy sauce (kecap) fermentation in Indonesia. *Journal of Fermentation and Bioengineering*, **81**(3): 275-278.

- Ruas-Madiedo, P., and De Los Reyes-Gavilán, C. (2005). Invited review: methods for the screening, isolation, and characterization of exopolysaccharides produced by lactic acid bacteria. *Journal of Dairy Science*, **88**(3): 843-856.
- Ruas-Madiedo, P., Moreno, J. A., Salazar, N., Delgado, S., Mayo, B., Margolles, A., and de Los Reyes-Gavilan, C. G. (2007). Screening of exopolysaccharide-producing *Lactobacillus* and *Bifidobacterium* strains isolated from the human intestinal microbiota. *Applied and Environmental Microbiology*, **73**(13): 4385-4388.
- Ruas-Madiedo, P., Salazar, N., and de los Reyes-Gavilán, C. (2009). Exopolysaccharides produced by lactic acid bacteria in food and probiotic applications. *Microbial glycobiology. Structures, relevance and applications*. Academic Press Elsevier, London: 887-902.
- Ruas-Madiedo, P., Salazar, N., and de los Reyes-Gavilán, C. G. (2010). Exopolysaccharides produced by lactic acid bacteria in food and probiotic applications. *Microbial Glycobiology, Structures, Relevance and Applications*: 885-902.
- Ruhmann, B., Schmid, J., and Sieber, V. (2015). Methods to identify the unexplored diversity of microbial exopolysaccharides. *Frontier Microbiology*, **6**: 565.
- Ruiz, L., Margolles, A., and Sanchez, B. (2013). Bile resistance mechanisms in *Lactobacillus* and *Bifidobacterium*. *Frontier Microbiology*, **4**: 396.
- Ryan, P. M., Ross, R. P., Fitzgerald, G. F., Caplice, N. M., and Stanton, C. (2015). Sugar-coated: exopolysaccharide producing lactic acid bacteria for food and human health applications. *Food and Function*, **6**(3): 679-693.
- Saarela, M., Mogensen, G., Fonden, R., Matto, J., and Mattila-Sandholm, T. (2000). Probiotic bacteria: safety, functional and technological properties. *Journal of Biotechnology*, **84**(3): 197-215.
- Salazar, N., Gueimonde, M., De Los Reyes-Gavilán, C. G., and Ruas-Madiedo, P. (2015). Exopolysaccharides produced by lactic acid bacteria and bifidobacteria as fermentable substrates by the intestinal microbiota. *Critical Reviews in Food Science and Nutrition*, **56**(9): 1440- 1453.
- Salminen, S. (2016). Human studies on probiotics: Aspects of scientific documentation. *Näringsforskning*, **45**(1): 8-12.
- Salminen, S., Laine, M., Vonwright, A., Vuopio-Varkila, J., Korhonen, T., and Mattila-Sandholm, T. (1996). Development of selection criteria for probiotic strains to assess their potential in functional foods: A Nordic and European Approach. *Bioscience and Microflora*, **15**(2): 61-67.
- Salminen, S., Ouwehand, A., Benno, Y., and Lee, Y. K. (1999). Probiotics: how should they be defined? *Trends in Food Science and Technology*, **10**(3): 107-110.

- Salminen, S., von Wright, A., Morelli, L., Marteau, P., Brassart, D., de Vos, W. M., Fondén, R., Saxelin, M., Collins, K., and Mogensen, G. (1998). Demonstration of safety of probiotics—a review. *International Journal of Food Microbiology*, **44**(1): 93-106.
- Sanchez, J. I., Martinez, B., Guillen, R., Jimenez-Diaz, R., and Rodriguez, A. (2006). Culture conditions determine the balance between two different exopolysaccharides produced by *Lactobacillus pentosus* LPS26. *Applied and Environmental Microbiology*, **72**(12): 7495-7502.
- Sanz, Y. (2007). Ecological and functional implications of the acid-adaptation ability of *Bifidobacterium*: A way of selecting improved probiotic strains. *International Dairy Journal*, **17**(11): 1284-1289.
- Sartor, R. B. (2004). Therapeutic manipulation of the enteric microflora in inflammatory bowel diseases: antibiotics, probiotics, and prebiotics. *Gastroenterology*, **126**(6): 1620-1633.
- Saxelin, M., Tynkkynen, S., Mattila-Sandholm, T., and de Vos, W. M. (2005). Probiotic and other functional microbes: from markets to mechanisms. *Current Opinion in Biotechnology*, **16**(2): 204-211.
- Schrezenmeir, J., and de Vrese, M. (2001). Probiotics, prebiotics, and synbiotics—approaching a definition. *The American journal of clinical nutrition*, **73**(2): 361s-364s.
- Servin, A. L. (2004). Antagonistic activities of lactobacilli and bifidobacteria against microbial pathogens. *FEMS Microbiology Reviews*, **28**(4): 405-440.
- Seviour, R. J., McNeil, B., Fazenda, M. L., and Harvey, L. M. (2011). Operating bioreactors for microbial exopolysaccharide production. *Critical Reviews in Biotechnology*, **31**(2): 170-185.
- Shah, N. (2004). Probiotics and prebiotics. *Agro Food Industry Hi-Tech*.
- Shah, N. P., and Lankaputhra, W. E. (1997). Improving viability of *Lactobacillus acidophilus* and *Bifidobacterium* spp. in yogurt. *International Dairy Journal*, **7**(5): 349-356.
- Shang, N., Xu, R., and Li, P. (2013). Structure characterization of an exopolysaccharide produced by *Bifidobacterium animalis* RH. *Carbohydrate Polymers*, **91**(1): 128-134.
- Shokryazdan, P., Sieo, C. C., Kalavathy, R., Liang, J. B., Alitheen, N. B., Faseleh Jahromi, M., and Ho, Y. W. (2014). Probiotic potential of *Lactobacillus* strains with antimicrobial activity against some human pathogenic strains. *BioMed Research International*.

- Singh, R. P., Shukla, M. K., Mishra, A., Kumari, P., Reddy, C. R. K., and Jha, B. (2011). Isolation and characterization of exopolysaccharides from seaweed associated bacteria *Bacillus licheniformis*. *Carbohydrate Polymers*, **84**(3): 1019-1026.
- Singh, T. P., Kaur, G., Malik, R. K., Schillinger, U., Guigas, C., and Kapila, S. (2012). Characterization of intestinal *Lactobacillus reuteri* strains as potential probiotics. *Probiotics and Antimicrobial Proteins*, **4**(1): 47-58.
- Smibert, R., Buchanan, R., and Gibbons, N. (1974). Bergey's manual of determinative bacteriology. *Williams and Wilkins Co, Baltimore*, **1**: 207.
- Somogyi, M. (1945). A new reagent for the determination of sugars. *Journal of Biological Chemistry*, **160**: 61-68.
- Son, S., and Lewis, B. A. (2002). Free radical scavenging and antioxidative activity of caffeic acid amide and ester analogues: Structure–activity relationship. *Journal of Agricultural and Food Chemistry*, **50**(3): 468-472.
- Sornplang, P., and Piyadeatsoontorn, S. (2016). Probiotic isolates from unconventional sources: a review. *Journal of Animal Science and Technology*, **58**(1): 26.
- Sperti, G. S. (1971). *Probiotics*: AVI Publishing Company.
- Steinkraus, K. H. (2002). Fermentations in world food processing. *Comprehensive Reviews in Food Science and Food Safety*, **1**(1): 23-32.
- Stiles, M. E., and Holzapfel, W. H. (1997). Lactic acid bacteria of foods and their current taxonomy. *International Journal of Food Microbiology*, **36**(1): 1-29.
- Stoodley, P., Sauer, K., Davies, D. G., and Costerton, J. W. (2002). Biofilms as complex differentiated communities. *Annual Reviews in Microbiology*, **56**(1): 187-209.
- Suhartatik, N., Cahyanto, M., Rahardjo, S., Miyashita, M., and Rahayu, E. (2014). Isolation and identification of lactic acid bacteria producing  $\beta$  glucosidase from Indonesian fermented foods. *International Food Research Journal*, **21**(3).
- Sujaya, I., Amachi, S., Saito, K., Yokota, A., Asano, K., and Tomita, F. (2002). Specific enumeration of lactic acid bacteria in ragi tape by colony hybridization with specific oligonucleotide probes. *World Journal of Microbiology and Biotechnology*, **18**(3): 263-270.
- Sumner, J. B., and Howell, S. F. (1935). A method for determination of saccharase activity. *Journal of Biological Chemistry*, **108**(1): 51-54.

- Šušković, J., Kos, B., Beganović, J., Leboš Pavunc, A., Habjanič, K., and Matosić, S. (2010). Antimicrobial activity—the most important property of probiotic and starter lactic acid bacteria. *Food Technology and Biotechnology*, **48**(3): 296-307.
- Suzuki, C., Kobayashi, M., and Kimoto-Nira, H. (2013). Novel exopolysaccharides produced by *Lactococcus lactis* subsp. *lactis*, and the diversity of *epsE* genes in the exopolysaccharide biosynthesis gene clusters. *Bioscience, Biotechnology, and Biochemistry*, **77**(10): 2013-2018.
- Swain, M. R., Anandharaj, M., Ray, R. C., and Parveen Rani, R. (2014). Fermented fruits and vegetables of Asia: a potential source of probiotics. *Biotechnology Research International*, **2014**: 250424.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A., and Kumar, S. (2013). MEGA6: molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, **30**(12): 2725-2729.
- Tannock, G. W. (1999). Identification of lactobacilli and bifidobacteria. *Current Issues in Molecular Biology*, **1**(1-2): 53-64.
- Tieking, M., Korakli, M., Ehrmann, M. A., Gänzle, M. G., and Vogel, R. F. (2003). In situ production of exopolysaccharides during sourdough fermentation by cereal and intestinal isolates of lactic acid bacteria. *Applied and Environmental Microbiology*, **69**(2): 945-952.
- Todorov, S. D., Botes, M., Guigas, C., Schillinger, U., Wiid, I., Wachsman, M. B., Holzapfel, W. H., and Dicks, L. M. T. (2008). Boza, a natural source of probiotic lactic acid bacteria. *Journal of Applied Microbiology*, **104**(2): 465-477.
- Torino, M. I., Font de Valdez, G., and Mozzi, F. (2015). Biopolymers from lactic acid bacteria. Novel applications in foods and beverages. *Frontier Microbiology*, **6**: 834.
- Tulumoglu, S., Yuksekdag, Z. N., Beyatli, Y., Simsek, O., Cinar, B., and Yaşar, E. (2013). Probiotic properties of lactobacilli species isolated from children's feces. *Anaerobe*, **24**: 36-42.
- Valko, M., Rhodes, C. J., Moncol, J., Izakovic, M., and Mazur, M. (2006). Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions*, **160**(1): 1-40.
- Van der Meulen, R., Grosu-Tudor, S., Mozzi, F., Vanngelgem, F., Zamfir, M., de Valdez, G. F., and De Vuyst, L. (2007). Screening of lactic acid bacteria isolates from dairy and cereal products for exopolysaccharide production and genes involved. *International Journal of Food Microbiology*, **118**(3): 250-258.

- Vanengelgem, F., Zamfir, M., Adriany, T., and De Vuyst, L. (2004). Fermentation conditions affecting the bacterial growth and exopolysaccharide production by *Streptococcus thermophilus* ST 111 in milk-based medium. *Journal of Applied Microbiology*, **97**(6): 1257-1273.
- Vasiljevic, T., and Shah, N. P. (2008). Probiotics—from Metchnikoff to bioactives. *International Dairy Journal*, **18**(7): 714-728.
- Vastano, V., Perrone, F., Marasco, R., Sacco, M., and Muscariello, L. (2016). Transcriptional analysis of exopolysaccharides biosynthesis gene clusters in *Lactobacillus plantarum*. *Archives of Microbiology*, **198**(3): 295-300.
- Velasco, S., Arskold, E., Paese, M., Grage, H., Irastorza, A., Radstrom, P., and van Niel, E. W. (2006). Environmental factors influencing growth of and exopolysaccharide formation by *Pediococcus parvulus* 2.6. *International Journal of Food Microbiology*, **111**(3): 252-258.
- Vinderola, G., Perdigón, G., Duarte, J., Farnworth, E., and Matar, C. (2006). Effects of the oral administration of the exopolysaccharide produced by *Lactobacillus kefiranofaciens* on the gut mucosal immunity. *Cytokine*, **36**(5): 254-260.
- Vlková, E., Rada, V., Šmehilová, M., and Killer, J. (2008). Auto-aggregation and co-aggregation ability in bifidobacteria and clostridia. *Folia Microbiologica*, **53**(3): 263-269.
- Wang, A. N., Yi, X. W., Yu, H. F., Dong, B., and Qiao, S. Y. (2009). Free radical scavenging activity of *Lactobacillus fermentum* in vitro and its antioxidative effect on growing-finishing pigs. *Journal of Applied Microbiology*, **107**(4): 1140-1148.
- Wang, C.-Y., Lin, P.-R., Ng, C.-C., and Shyu, Y.-T. (2010). Probiotic properties of *Lactobacillus* strains isolated from the feces of breast-fed infants and Taiwanese pickled cabbage. *Anaerobe*, **16**(6): 578-585.
- Wang, J., Li, B., Zhou, L., Zhang, X., and Shi, P. (2016). Probiotic potential and function of a *Lactobacillus* strain L1 isolated from Silage. *Journal of Food Safety*.
- Wang, J., Zhang, H., Chen, X., Chen, Y., and Bao, Q. (2012). Selection of potential probiotic lactobacilli for cholesterol-lowering properties and their effect on cholesterol metabolism in rats fed a high-lipid diet. *Journal of Dairy Science*, **95**(4): 1645-1654.
- Wang, J., Zhao, X., Yang, Y., Zhao, A., and Yang, Z. (2015). Characterization and bioactivities of an exopolysaccharide produced by *Lactobacillus plantarum* YW32. *International Journal of Biological Macromolecules*, **74**: 119-126.
- Wang, X., Shao, C., Liu, L., Guo, X., Xu, Y., and Lu, X. (2017). Optimization, partial characterization and antioxidant activity of an exopolysaccharide from *Lactobacillus plantarum* KX041. *International Journal of Biological Macromolecules*, **103**: 1173-1184.

- Wang, Y., Li, C., Liu, P., Ahmed, Z., Xiao, P., and Bai, X. (2010). Physical characterization of exopolysaccharide produced by *Lactobacillus plantarum* KF5 isolated from Tibet Kefir. *Carbohydrate Polymers*, **82**(3): 895-903.
- WHO. (2001). Health and nutritional properties of probiotics in food including powdered milk with live lactic acid bacteria: a joint FAO/WHO expert consultation report. Retrieved 3 December 2016, from World Health Organization (WHO) Available from URL: <http://isappscience.org/wp-content/uploads/2015/12/FAO-WHO-2001-Probiotics-Report.pdf>
- Yang, Z., Li, S., Zhang, X., Zeng, X., Li, D., Zhao, Y., and Zhang, J. (2010). Capsular and slime-polysaccharide production by *Lactobacillus rhamnosus* JAAS8 isolated from Chinese sauerkraut: potential application in fermented milk products. *Journal of Bioscience and Bioengineering*, **110**(1): 53-57.
- Ye, L., Zhang, J., Yang, Y., Zhou, S., Liu, Y., Tang, Q., Du, X., Chen, H., and Pan, Y. (2009). Structural characterisation of a heteropolysaccharide by NMR spectra. *Food Chemistry*, **112**(4): 962-966.
- Yoon, E. J., Yoo, S. H., Cha, J., and Lee, H. G. (2004). Effect of levan's branching structure on antitumor activity. *International Journal of Biological Macromolecules*, **34**(3): 191-194.
- You, H. J., Oh, D. K., and Ji, G. E. (2004). Anticancerogenic effect of a novel chiroinositol-containing polysaccharide from *Bifidobacterium bifidum* BGN4. *FEMS Microbiology Letters*, **240**(2): 131-136.
- Yuliana, N., and Dizon, E. I. (2011). Phenotypic identification of lactic acid bacteria isolated from tempoyak (fermented durian) made in the Philippines. *International Journal of Biology*, **3**(2): 145-152.
- Zamfir, M., and Grosu-Tudor, S.-S. (2014). Stress response of some lactic acid bacteria isolated from Romanian artisan dairy products. *World Journal of Microbiology and Biotechnology*, **30**(2): 375-384.
- Zannini, E., Mauch, A., Galle, S., Gänzle, M., Coffey, A., Arendt, E. K., Taylor, J. P., and Waters, D. M. (2013). Barley malt wort fermentation by exopolysaccharide forming *Weissella cibaria* MG1 for the production of a novel beverage. *Journal of Applied Microbiology*, **115**(6): 1379-1387.
- Zannini, E., Waters, D. M., Coffey, A., and Arendt, E. K. (2016). Production, properties, and industrial food application of lactic acid bacteria-derived exopolysaccharides. *Applied Microbiology and Biotechnology*, **100**(3): 1121-1135.
- Zhang, J., Zhang, X., Zhang, L., Zhao, Y., Niu, C., Yang, Z., and Li, S. (2014). Potential probiotic characterization of *Lactobacillus plantarum* strains isolated from Inner Mongolia "Hurood" cheese. *Journal of Microbiology and Biotechnology*, **24**(2): 225-235.

Zhang, L., Liu, C., Li, D., Zhao, Y., Zhang, X., Zeng, X., Yang, Z., and Li, S. (2013). Antioxidant activity of an exopolysaccharide isolated from *Lactobacillus plantarum* C88. *International Journal of Biological Macromolecules*, **54**: 270-275.

