



**UNIVERSITI PUTRA MALAYSIA**

***INTROGRESSION OF HEAT SHOCK PROTEIN GENES FOR  
DEVELOPMENT OF HEAT TOLERANT CHILLI (*Capsicum annuum* L.)  
GENOTYPES THROUGH MARKER-ASSISTED BACKCROSSING***

**USMAN MAGAJI**

**IPTSM 2018 6**



**INTROGRESSION OF HEAT SHOCK PROTEIN GENES FOR  
DEVELOPMENT OF HEAT TOLERANT CHILLI (*Capsicum annuum* L.)  
GENOTYPES THROUGH MARKER-ASSISTED BACKCROSSING**

**By**

**USMAN MAGAJI**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirement for the Degree of Doctor of Philosophy**

**April 2018**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## DEDICATION

To my loving parents; Late Alhaji Wa`alamu Magaji, Hajiya Fatima Abubakar Kurfi  
and Alhaji Aminu Abubakar



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**INTROGRESSION OF HEAT SHOCK PROTEIN GENES FOR  
DEVELOPMENT OF HEAT TOLERANT CHILLI (*Capsicum annuum* L.)  
GENOTYPES THROUGH MARKER-ASSISTED BACKCROSSING**

By

**USMAN MAGAJI**

**April 2018**

**Supervisor: Professor Mohd Rafii Yusop, PhD**  
**Institute: Institute of Tropical Agriculture and Food Security**

Domestic production of chilli is insufficient (52% self-sufficiency level) and can hardly meet 70% of local demand due to some factors including the poor performance of local varieties under high temperatures above 42 °C. The optimum growing temperature for chilli ranged from 20 to 30 °C. Climate change especially high temperature is projected to negatively impact future agricultural production worldwide. According to reports, Malaysia will lose about 10% of major staples food by 2030 due to climate change, which is projected to rise average temperature by 0.3 °C to 4.5 °C and this warmer climate will cause a rise in sea level, and this will reduce crop yield. Development of improved heat-tolerant chilli varieties will contribute to self-sufficiency in chilli in Malaysia. Backcrossing together with simple sequence repeat marker strategy was adopted to improve popular Malaysian Kulai 907 (*Capsicum annuum* L.) for heat tolerance. The use of molecular markers in backcross breeding and selection contributes significantly to overcome the main drawbacks such as increase linkage drag and time consumption, in the conventional breeding approach and to speed up the genome recovery of the recurrent parent. The approach was adopted to introgress heat shock protein gene(s) from AVPP0702, a heat-tolerant variety, into the genetic profile of Kulai 907, a high-yielding chilli but heat sensitive. Introgression of heat shock proteins (Hsps) genes has shown considerable success in improvement of crop plants such as maize and rice against heat stress. However, no study has been reported on the introgression of Hsps genes in chilli for the improvement of chilli heat tolerance. The main objective of this study was to develop heat tolerant variety of chilli with high yielding potential and while the specific objectives were; to introgress Hsp genes from heat tolerant (AVPP0702) to high yielding Kulai 907 variety, to identify polymorphic molecular markers for heat tolerant characteristics and recipient parent genome recoveries (RPG) and to validate the backcross progenies for heat tolerance (Hsp loci). Local Kulai 907 variety was used as the recurrent parent and AVPP0702 was used as the donor parent. The parents were grown on seed trays and parental screening was carried out with 252 simple sequence repeat markers (SSR). DNA of young fresh leaves was extracted using CTAB method. Out of the 252 SSR markers, 27% showed clear polymorphism between heat sensitive and tolerant parent. Sixty-eight markers

appeared to be polymorphic and used to estimate the recovery of the recurrent parent in the backcross generations; BC<sub>1</sub>F<sub>1</sub>, BC<sub>2</sub>F<sub>1</sub>, BC<sub>3</sub>F<sub>1</sub> and BC<sub>3</sub>F<sub>2</sub>. The average RPG of the selected four BC<sub>1</sub>F<sub>1</sub> plants was 80.75% which were used to produce the BC<sub>2</sub>F<sub>1</sub> generation. BC<sub>1</sub>-P<sub>7</sub> plant was the best in BC<sub>1</sub>F<sub>1</sub> generation having the highest recovery 83.40% and positive to Hsp-linked markers (Hsp70-u2 and AGi42). After three successive generations of backcrossing, the average genome recovery (RPG) of the recurrent parent in the selected plants in BC<sub>3</sub>F<sub>1</sub> population was 95.37% and BC<sub>3</sub>F<sub>2</sub> population was 97.90%. Hsp gene expression analysis was carried out on BC<sub>1</sub>F<sub>1</sub>, BC<sub>2</sub>F<sub>1</sub>, BC<sub>3</sub>F<sub>1</sub> and BC<sub>3</sub>F<sub>2</sub> selected genotypes with high recovery of the recurrent parent. The Hsp genes are found to be up-regulated with more than 10.9-, 18.4-, 8.8- and 22.2-fold increase when exposed to heat treatment. The pattern of Hsp expression in the backcross generations was similar with the donor parent (up-regulated). This confirms the successful introgression of stress responsive gene (Hsp) into Kulai 907 variety. Twelve improved heat-tolerant chilli genotypes, namely; BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>1</sub>-P<sub>2</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>1</sub>-P<sub>9</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>1</sub>-P<sub>11</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>3</sub>-P<sub>5</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>1</sub>-P<sub>13</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>3</sub>-P<sub>4</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>7</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>9</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>14</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>3</sub>-P<sub>16</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>15</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>18</sub> were selected from the BC<sub>3</sub>F<sub>2</sub> population that had homozygous Hsp alleles from AVPP0702 and recurrent genome recovery of Kulai 907 (average RPG 97.9%). Most of the morphological and agronomical traits were recovered in the selected improved-heat tolerant genotypes from Kulai 907 such as plant height (75.94 cm), number of days to 50% flowering (56.5%), number of fruits (91.6), stem length (22.3 cm), stem diameter (6 cm), fruit length (13.3 cm) and weight (17.5 cm) and total fruit yield per plant (862.3 g).

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**INTROGRESI GEN PROTEIN RENJATAN HABA UNTUK PEMBANGUNAN  
VARIETI CILI (*Capsicum annuum* L.) KETAHANAN HABA MELALUI  
KACUKBALIK BANTUAN PENANDA**

Oleh

**USMAN MAGAJI**

**April 2018**

**Penyelia: Profesor Mohd Rafii Yusop, PhD**  
**Institut: Institut Pertanian Tropika dan Sekuriti Makanan**

Pengeluaran domestik cili adalah tidak mencukupi (tahap kecukupan 52%) dan ianya sangat sukar untuk memenuhi 70% permintaan tempatan disebabkan beberapa faktor termasuk prestasi varieti tempatan yang rendah di bawah suhu tinggi di atas 42 °C. Suhu pertumbuhan yang optima untuk cili adalah dari 20 hingga 30 °C. Perubahan iklim terutama suhu tinggi dijangka memberikan impak negatif terhadap pengeluaran pertanian masa depan di seluruh dunia. Menurut laporan, Malaysia akan kehilangan kira-kira 10% daripada makanan ruji menjelang 2030 ekoran perubahan iklim, yang dijangkakan berlaku peningkatan suhu purata sebanyak 0.3 °C hingga 4.5 °C dan iklim panas ini akan menyebabkan kenaikan paras laut, dan ini akan mengurangkan hasil pengeluaran tanaman. Pembangunan varieti maju cili toleran haba akan menyumbang kepada tahap kecukupan cili di Malaysia. Kacukbalik bersama dengan strategi penanda jujukan berulang mudah telah digunakan untuk pembiakbakaan ketahanan terhadap haba ke atas varieti cili popular di Malaysia, varieti Kulai 907 (*Capsicum annuum* L.). Penggunaan penanda molekul dalam pembiakbakaan kacukbalik dan pemilihan telah memberi sumbangan untuk mengatasi kekangan utama seperti hambatan pautan dan tempuh masa yang diperlukan dalam kaedah pembiakbakaan konvensional, dan untuk mempercepatkan pemulihan genom induk penerima. Kaedah ini digunakan dengan mengintrogressi gen protein renjatan haba dari AVPP0702, satu varieti yang toleran haba ke dalam profil genetik Kulai 907 yang berhasil tinggi tetapi sensitif kepada haba. Introgressi gen protein renjatan haba (Hsps) telah menunjukkan hasil yang memberasangkan dalam pembiakbakaan tanaman seperti jagung dan padi terhadap tekanan haba. Walau bagaimanapun, tiada kajian yang telah dilaporkan berkaitan penggabungan gen Hsps ke cili untuk penambahbaikan toleransi ketahanan haba. Objektif utama kajian ini adalah untuk membangunkan genotip toleran haba serta mempunyai potensi hasil tinggi, dan objektif spesifik adalah; untuk mengintrogressi gen-gen Hsp dari varieti toleran haba (AVPP0702) kepada varieti Kulai 907 yang memberikan penghasilan tinggi, untuk mengenalpasti penanda molekul polimorfik bagi ciri toleran haba dan pemulihan genom induk penerima (RPG), untuk mengesahkan progeni kacukbalik yang toleransi haba (lokus Hsp). Varieti tempatan Kulai 907 telah digunakan sebagai induk penerima dan AVPP0702 sebagai induk penderma. Induk-

induk tersebut telah ditanam dalam bekas percambahan biji benih dan induk tersebut disaring menggunakan 252 penanda berulang jujukan mudah (SSR). DNA dari daun muda yang segar diekstrak menggunakan kaedah CTAB. Dari 252 penanda SSR, 27% telah menunjukkan polimorfik yang jelas antara induk sensitif dan toleran haba. Enam puluh lapan penanda memberikan polimorfik dan telah digunakan untuk menganggarkan pemulihan induk penerima dalam generasi kacukbalik BC<sub>1</sub>F<sub>1</sub>, BC<sub>2</sub>F<sub>1</sub>, BC<sub>3</sub>F<sub>1</sub> dan BC<sub>3</sub>F<sub>2</sub>. Purata RPG untuk empat pokok BC<sub>1</sub>F<sub>1</sub> terpilih adalah 80.75% yang telah digunakan untuk menghasilkan generasi BC<sub>2</sub>F<sub>1</sub>. BC<sub>1</sub>-P<sub>7</sub> adalah pokok generasi BC<sub>1</sub>F<sub>1</sub> terbaik yang mempunyai pemulihan tertinggi iaitu 83.40% dan positif kepada penanda-pautan Hsp (Hsp70-u2 dan AGi42). Selepas tiga generasi kacukbalik, purata RPG bagi pokok terpilih populasi BC<sub>3</sub>F<sub>1</sub> adalah 95.37% dan BC<sub>3</sub>F<sub>2</sub> adalah 97.90%. Analisis pengekspresan gen Hsp telah dijalankan ke atas genotip terpilih BC<sub>1</sub>F<sub>1</sub>, BC<sub>2</sub>F<sub>1</sub>, BC<sub>3</sub>F<sub>1</sub> dan BC<sub>3</sub>F<sub>2</sub> yang mempunyai pemulihan induk berulang yang tinggi. Gen Hsp didapati menunjukkan regulasi-keatas yang melebihi 10.9-, 18.4-, 8.8-, dan 22.2-ganda peningkatan apabila didedah kepada rawatan haba. Corak ekspresi Hsp dalam generasi-generasi kacukbalik tersebut adalah sama dengan induk penderma (regulasi-keatas). Ini mengesahkan kejayaan introgresi gen ketahanan haba (Hsp) ke varieti cili Kulai 907. Dua belas genotip maju cili toleran haba iaitu BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>1</sub>-P<sub>2</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>1</sub>-P<sub>9</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>1</sub>-P<sub>11</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>3</sub>-P<sub>5</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>1</sub>-P<sub>13</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>3</sub>-P<sub>4</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>7</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>9</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>14</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>3</sub>-P<sub>16</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>15</sub>, BC<sub>1</sub>-P<sub>7</sub>-P<sub>10</sub>-P<sub>4</sub>-P<sub>18</sub> telah dipilih daripada populasi BC<sub>3</sub>F<sub>2</sub> yang mempunyai alel homozaigus Hsp dari AVPP0702 serta mempunyai pemulihan ciri morfologi dan agronomi cili Kulai 907 (purata RPG 97.9%). Kebanyakan ciri-ciri morfologi dan agronomi telah pulih dalam genotip toleran haba terpilih dari Kulai 907 seperti ketinggian pokok (75.94 cm), bilangan hari hingga 50% berbunga (56.5%), bilangan buah (91.6), panjang batang pokok (22.3 cm), diameter batang (6 cm), panjang (13.3 cm) dan berat buah (17.5 cm), dan jumlah hasil buah per pokok (862.3 g).



## ACKNOWLEDGEMENTS

All praise and thanks be to Allah, the compassionate, most exalted with humility. My most sincere thanks goes to my academic and research supervisory committee; Prof. Dr. Mohd Rafii bin Yusop (Chairman), Prof. Dr. Mohd Razi bin Ismail and Dr. Martini binti Mohammad Yusoff, who by the way of appreciation of ideas, encouragement, useful suggestions and constructive criticisms guided me throughout the period of the research.

My special appreciation goes to the entire staff of Laboratory of Climate-Smart Food Crop Production, Institute of Tropical Agriculture and Food Security, Universiti Putra Malaysia especially able and capable science officers particularly Rafidah Mohd Yusoff, Nooraishah Abdul Aziz and Muhammad Adzan Mastor among others.

Gratitude is also extended to my parents as well as my brothers, sisters and relatives especially Hafsat Abdullahi (Auta), Khadijah Magaji, Amina Magaji and Husna Auwal, Abubakar Magaji and Aminu Magaji for their moral support and understanding. Among friends and colleagues, I consider it necessary to acknowledge and appreciate the efforts of Oladosu Yusuff, Tijjani Ahmadu and Raihana Ridzuan among others.

Finally, appreciation also goes to Ministry of Education, Malaysia, for adequately funding of the research through the Fundamental Research Grant Scheme (FRGS/1/2012/STWN03/UPM/02/2:07-01-13-1240FR) and Universiti Putra Malaysia towards the success of the research through the Geran Putra (GP – IBT/2013/9421000).

I certify that a Thesis Examination Committee has met on 10 April 2018 to conduct the final examination of Usman Magaji on his thesis entitled "Introgression of Heat Shock Protein Genes for Development of Heat Tolerant Chilli (*Capsicum annuum* L.) Genotypes Through Markerassisted Backcrossing" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Mahmud bin Tengku Muda Mohamed, PhD**

Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Nor Aini binti Ab Shukor, PhD**

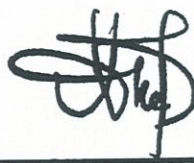
Professor  
Faculty of Forestry  
Universiti Putra Malaysia  
(Internal Examiner)

**Khairulmazmi bin Ahmad, PhD**

Associate Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Internal Examiner)

**Sanjeet Kumar, PhD**

Senior Lecturer  
World Vegetable Center  
Taiwan  
(External Examiner)



---

**NOR AINI AB. SHUKOR, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 24 May 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the Degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Mohd Rafii Bin Yusop, PhD**

Professor  
Institute of Tropical Agriculture and Food Security  
Universiti Putra Malaysia  
(Chairman)

**Mohd Razi Ismail, PhD**

Professor  
Institute of Tropical Agriculture and Food security  
Universiti Putra Malaysia  
(Member)

**Martini Mohammad Yusoff, PhD**

Senior Lecturer  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by Graduate Student

I hereby confirmed that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: Usman Magaji (GS42009)

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision,
- supervision responsibilities as slated in Rule 41 in Rules 2003(Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_

Name of

Chairman

of supervisory

committee: Prof. Dr. Mohd Rafii Yusop

Signature: \_\_\_\_\_

Name of

Member

of supervisory

committee: Professor Dr. Mohd Razi bin Ismail

Signature: \_\_\_\_\_

Name of

Member

of supervisory

committee: Dr. Martini Mohammad binti Yusoff



	2.8.1	Conventional Breeding	24
	2.8.2	Marker Assisted Backcrossing (MAB)	25
<b>3</b>		<b>PARENTAL POLYMORPHISM BETWEEN POPULAR CHILLI VARIETY KULAI AND HEAT TOLERANT DONOR (AVPP0702) AND THEIR INHERITANCE PATTERNS</b>	39
	3.1	Introduction	27
	3.2	Materials and Methods	28
	3.2.1	Planting Material, growth condition and crossing procedure	28
	3.2.2	SSR Marker	29
	3.2.3	DNA isolation, PCR conditions and electrophoresis	34
	3.3	Results and Discussion	35
	3.3.1	Parental polymorphism for Hsp genes	35
	3.3.2	Parental polymorphism survey between recipient parent, chilli Kulai and the donor parent, AVPP0702	36
	3.3.3	F <sub>1</sub> confirmation using Tightly-Linked Hsp Markers	41
	3.4	Conclusions	42
<b>4</b>		<b>ANALYSIS OF RECURRENT PARENT GENOME RECOVERY THROUGH MARKER ASSISTED BACKCROSSING IN BC<sub>1</sub>F<sub>1</sub>, BC<sub>2</sub>F<sub>1</sub>, AND BC<sub>3</sub>F<sub>1</sub> POPULATIONS</b>	43
	4.1	Introduction	43
	4.2	Materials and Methods	43
	4.2.1	Plant material and breeding approach	43
	4.2.2	Leaf samples	45
	4.2.3	Molecular marker analysis	45
	4.2.3.1	Foreground selection	45
	4.2.3.2	Background selection	45
	4.2.4	DNA extraction, polymerase chain reaction and gel electrophoresis	45
	4.2.5	Phenotypic selection	46
	4.2.6	Data analysis	46
	4.3	Results and Discussion	46
	4.3.1	Marker genotyping of BC <sub>1</sub> F <sub>1</sub> generation	46
	4.3.1.1	Marker assisted foreground selection	46
	4.3.1.2	Marker assisted background selection	47
	4.3.2	Marker genotyping of BC <sub>2</sub> F <sub>1</sub> generation	51
	4.3.2.1	Marker assisted foreground selection	51
	4.3.2.2	Marker assisted background selection	52
	4.3.3	Marker genotyping of BC <sub>3</sub> F <sub>1</sub> generation	54
	4.3.3.1	Marker assisted foreground selection	54
	4.3.3.2	Marker assisted background selection	56
	4.3.3.3	Agro-morphological performance of the BC <sub>3</sub> F <sub>1</sub> generation	59
	4.4	Conclusion	61

<b>5</b>	<b>HEAT SHOCK PROTEIN GENE CONFIRMATION AND EXPRESSION ANALYSIS IN BC<sub>1</sub>F<sub>1</sub>, BC<sub>2</sub>F<sub>1</sub> and BC<sub>3</sub>F<sub>1</sub> GENERATION</b>	62
5.1	Introduction	62
5.2	Materials and Methods	62
5.2.1	Planting Material	62
5.2.2	Screening of precision introgression lines for heat tolerance	62
5.2.3	Total RNA Extraction, RNA integrity and cDNA synthesis	64
5.2.4	Quantitative Real Time PCR (qRT-PCR) analysis	64
5.2.5	Primer Design	64
5.2.6	Statistical Analysis	65
5.3	Results and Discussion	65
5.3.1	Validation of target and reference genes	65
5.3.2	Amplification and dissociation curve analysis of Hsp70 and OsHsp24 gene in the candidate BC <sub>1</sub> F <sub>1</sub> , BC <sub>2</sub> F <sub>1</sub> and BC <sub>3</sub> F <sub>1</sub> generations	67
5.3.3	Expression profiling of Hsp70 and OsHsp24 genes under heat stress in BC <sub>1</sub> F <sub>1</sub> , BC <sub>2</sub> F <sub>1</sub> and BC <sub>3</sub> F <sub>1</sub> generations	69
5.3.4	Recovery analysis	75
5.4	Conclusion	76
<b>6</b>	<b>SELECTION OF HOMOZYGOUS LINES AND PHENOTYPIC ASSESSMENT OF THE BC<sub>3</sub>F<sub>2</sub> IMPROVED HEAT-TOLERANT CHILLIES</b>	77
6.1	Introduction	77
6.2	Materials and Methods	77
6.2.1	Planting materials	77
6.2.2	DNA extraction, Polymerase Chain Reaction and Marker analysis	77
6.2.3	Heat treatment and morph physiological parameters	78
6.2.3.1	Cell membrane stability index (CMSI)	78
6.2.3.2	Measurement of canopy temperature depression (CTD)	78
6.2.3.3	Relative chlorophyll content (SPAD) and chlorophyll stability index (CSI)	79
6.2.3.4	Morphological parameters	79
6.2.4	Hsp gene expression profile in BC <sub>3</sub> F <sub>2</sub> population	79
6.2.4.1	Heat Stress Treatment	79
6.2.4.2	RNA extraction and quantitative Real Time PCR (qRT PCR)	79
6.2.5	Data Analysis	80
6.3	Results and Discussion	80
6.3.1	Genotyping of BC <sub>3</sub> F <sub>2</sub> population	80
6.3.1.1	Confirmation and selection of homozygous improved heat-tolerant chilli lines	80
6.3.1.2	Background Selection	82



6.3.2	Phenotyping of BC <sub>3</sub> F <sub>2</sub> population	86
6.3.2.1	Cell membrane stability index (CMSI)	86
6.3.2.2	Canopy temperature depression	88
6.3.2.3	Relative chlorophyll content (SPAD) and chlorophyll stability index (CSI)	89
6.3.2.4	Correlation between CMSI, CTD and CSI	91
6.3.3	Analysis of fold difference in BC <sub>3</sub> F <sub>2</sub> improved heat-tolerant chilli lines	92
6.3.4	Morphological and physiological comparison of selected homozygous BC <sub>3</sub> F <sub>2</sub> improved heat-tolerant chilli lines with Kulai (Recurrent parent) under normal growth condition	93
6.5	Conclusion	98
<b>7</b>	<b>SUMMARY, CONCLUSION AND RECOMMENDATIONS</b>	<b>99</b>
7.1	Summary	99
7.2	Conclusion	100
7.3	Recommendation	101
	<b>REFERENCES</b>	<b>102</b>
	<b>APPENDICES</b>	<b>118</b>
	<b>BIODATA OF STUDENT</b>	<b>131</b>
	<b>LIST OF PUBLICATIONS</b>	<b>132</b>

## LIST OF TABLES

<b>Table</b>	<b>Page</b>
2.1 Comparison between conventional backcross breeding and marker-assisted backcrossing (MAB)	26
3.1 Polymorphic tightly-linked Hsp gene and background SSR marker information	30
3.2 Polymorphic primers with their polymorphic percentage obtained from the sixty-eight simple sequence repeat markers	38
3.3 Characteristics of polymorphic simple sequence repeat markers	40
4.1 RPG recovery in percentage and heterozygous segment of selected individuals in BC <sub>1</sub> F <sub>1</sub> population	48
4.2 Proportion of sensitive and tolerant individuals in BC <sub>2</sub> F <sub>1</sub> generation using foreground marker	51
4.3 RPG recovery and heterozygous segment of chosen individuals in BC <sub>2</sub> F <sub>1</sub> population	52
4.4 Proportion of sensitive and tolerant individuals in BC <sub>3</sub> F <sub>1</sub> generation of the foreground marker using Chi square test of association	56
4.5 Descriptive t-test and homogeneity variance test statistics for the mean comparison between the recurrent (recipient) parent and BC <sub>3</sub> F <sub>1</sub> lines using independent t-test	60
4.6 Morphological performance of the recipient (recurrent) parent and improved BC <sub>3</sub> F <sub>1</sub> lines using Tukey's HSD test	60
5.1 List of primers used for quantitative real time (qRT-PCR) amplification of Heat Shock Protein 70 (Hsp70) and OsHsp24 in chilli	65
5.2 Gene regulation and probability value of the analysis of Hsp70 and OsHsp24 under differential heat stress condition among the parents and backcross generations	71
6.1 Genotypic marker segregation analysis in BC <sub>3</sub> F <sub>2</sub> population using chi square test of association to test the expected 1:2:1 ratio in a single gene model	81
6.2 Background and introgressed segment analysis in selected BC <sub>3</sub> F <sub>2</sub> populations	83

6.3	Genome recovery of the background and introgressed segment analysis in chosen BC <sub>3</sub> F <sub>2</sub> populations across the 12-chromosome of chilli	83
6.4	Analysis of variance showing the Mean Squares of percentage damage and cell membrane stability index (%) of the parental as well as the improved heat-tolerant chilli lines exposed to high temperature stress (40 and 50 °C) experimental water bath	86
6.5	Relative percentage damage (PD) of the parental as well as the improved heat-tolerant chilli lines exposed to different temperature regimes	87
6.6	Canopy Temperature Depression measured among the parents and improved heat-tolerant lines	89
6.7	SPAD chlorophyll content mean comparison among the parental lines and improved heat-tolerance chilli lines exposed to high temperature stress (40°C)	90
6.8	Mean Squares of chlorophyll stability index (%) of the parental (AVPP0702 and Kulai) as well as the improved heat-tolerant chili lines exposed to high temperature stress (40 °C) at different exposure time in experimental plant growth chamber	90
6.9	Pearson correlation matrix showing the relationship among chlorophyll stability index (CSI), cell membrane stability index (CMSI) and canopy temperature depression (CTD) of the parent and improved heat-tolerant chilli lines	92
6.10	Analysis of Hsp70 gene under differential heat stress condition among the parents and improved heat-tolerant chili lines revealed by qRT-PCR at 4 h	93
6.11	Descriptive statistics for the ten dependent variables of the improved heat-tolerant lines (BC <sub>3</sub> F <sub>2</sub> populations) and chilli Kulai (recurrent parent)	96
6.12	Descriptive t-test and homogeneity variance test statistics for the mean improved heat-tolerant chilli lines compared with chilli Kulai (recurrent parent)	97

## LIST OF FIGURES

Figure		Page
2.1	Genetic lineage of the Capsicum genus (Bosland and Votava, 2012)	7
2.2	Chilli Kulai plant at six weeks after transplanting	9
2.3	Relative normalized expression distribution of Hsp70 among four different chilli genotypes with different genetic backgrounds when exposed to a 42 °C heat treatment for 2 h. The Hsp70 transcript was differentially expressed among the peppers Culled from Usman et al. (2015)	10
2.4	Chilli production in states Malaysia (DoA, 2016)	12
2.5	Structural model of Hsp70. Hsp70s consist of three domains: an N-terminal ATPase domain (N); a Substrate Binding Domain (SBD); and a C-terminal “lid” domain (C). ATP bound in the N Domain which opens the lid of Hsp70 substrate-binding domain and allows substrate to bind. Catalyzed by various co-chaperones, ATP is hydrolyzed which leads to closure of the SBD lid, trapping substrate in the substrate binding pocket. Nucleotide exchange factors exchange ADP with ATP and releases the bound substrate and a new cycle begins	18
2.6	Schematic process of the Hsp70 response to stress in plants under abiotic stress	19
2.7	The contribution of the donor parent genome is reduced by half with each generation of backcrossing. Percentages of recurrent parent (light purple) are expressed as a ratio to percentages of donor parent (dark red-purple) (Byrne and Richardson, 2005)	25
3.1	Selected male flower during anthesis at 48-days after transplanting	29
3.2	Chilli fresh young leaves at four weeks after sowing	34
3.3	Polymorphism between parental lines, AVPP0702 (1) and recipient parent Kulai 907 (2), for two target genes using Hsp gene specific primer pairs (Hsp70-u2 and AGi42). M: 50bp ladder	36
3.4	Screening of parental lines (A: AVPP0702 and K: Kulai) for polymorphism using some of the SSR marker. Running on 2.5% metaphor agarose gel stained with midori green. M: 50bp Ladder	36
3.5	Frequencies of total 68 hyper variable SSR markers with different repeat motifs covering chromosome 1 to 12	39
3.6	Verified F <sub>1</sub> plants grown to generate BC <sub>1</sub> F <sub>1</sub> backcross populations	41

4.1	Breeding scheme of the study. It takes 12 weeks for a complete life cycle	44
4.2	Genotyping with markers Hsp70-u2 (a) and AGi42 (b) linked to Hsp genes in BC <sub>1</sub> F <sub>1</sub> population of chilli derived from K: Kulai × A: AVPP0702. H: indicates heterozygous individuals. Running on 2.5% metaphor agarose gel stained with midori green, only 14 samples plus the two parents for each marker are shown (M=50 bp ladder).	47
4.3	Banding pattern of some background markers in BC <sub>1</sub> F <sub>1</sub> population; A: donor parent; K: recipient parent; H: heterozygous pattern; only 13 individuals and the two parents were shown out of 20 individual plants.	48
4.4	Chromosome-wise recurrent parent genome recovery of 20 selected BC <sub>1</sub> F <sub>1</sub> plants. Red colour indicates homozygous region for AVPP0702 alleles, Purple colour indicates homozygous regions for Kulai alleles, and Light green colour indicates heterozygous region	49
4.5	Chromosome-wise recurrent parent genome recovery of the selected best plant BC <sub>1</sub> -P <sub>7</sub> . Red colour indicates homozygous region for AVPP0702 alleles, Purple colour indicates homozygous regions for Kulai alleles, and Light green colour indicates heterozygous region	50
4.6	Genotyping BC <sub>2</sub> F <sub>1</sub> (derived from K: Kulai × A: AVPP0702) using foreground markers Hsp70-u2 (23 individuals) and AGi42 (23 individuals) plus the two parents H: indicates heterozygous individuals. Running on 2.5% metaphor agarose gel stained with midori green. M: 50bp Ladder	51
4.7	Selected BC <sub>2</sub> F <sub>1</sub> plants overview 6 weeks after transplanting	52
4.8	Chromosome-wise recurrent parent genome recovery of the selected best plant BC <sub>1</sub> -P <sub>7</sub> -P <sub>10</sub> . Red colour indicates homozygous region for AVPP0702 alleles, Dark blue colour indicates homozygous regions for Kulai alleles, and Black colour indicates heterozygous region	53
4.9	Some background markers screened in BC <sub>2</sub> F <sub>1</sub> . A: donor parent; K: recipient parent, number of individuals: the progenies. M = 1kb 50bp ladder	54
4.10	BC <sub>3</sub> F <sub>1</sub> plants overview 4 weeks after transplanting	54
4.11	BC <sub>3</sub> F <sub>1</sub> confirmation using foreground markers Hsp70-u2 (9 individuals) and AGi42 (8 individuals). A: AVPP0702 and K: Kulai; H: heterozygous region. Only 8 individual samples were shown plus the two parents. M: 1kb 50bp ladder	55
4.12	Individual plant wise recurrent parent genome recovery in BC <sub>3</sub> F <sub>1</sub> generation of selected plants with an average of 95.37%. H = Heterozygous	57

4.13	Chromosome-wise recurrent parent genome recovery of the selected best plant BC <sub>1</sub> -P <sub>7</sub> -P <sub>10</sub> -P <sub>3</sub>	58
4.14	Graphical representation of normal mean distribution of genotypes (recipient and BC <sub>3</sub> F <sub>1</sub> ) in plant height	61
5.1	Backcross progenies and parental lines grown in small plastic pots at four weeks after sowing (A) and experimental plant growth chamber (B)	63
5.2	Amplification curve of the 5-point serial dilution series of Hsp70-u2	66
5.3	Standard curve with the C <sub>T</sub> plotted against the log of the starting quantity of cDNA template for Hsp70-u2	66
5.4	Amplification levels of candidate target Hsp70-u2 gene in the different parents and backcross generations under differential heat treatment and exposure time (2 h and 4 h), C <sub>q</sub> values: mean of triplicate samples	67
5.5	Accumulation of candidate target OsHsp24 gene in the different parents and backcross generations under differential heat treatment and exposure time (2 h and 4 h), C <sub>q</sub> values: mean of triplicate samples	68
5.6	Melt curve analysis of Hsp70-u2 (red) and OsHsp24 (green) at all temperature and exposure time	69
5.7	The expression levels of Hsp70 at different temperature treatments and duration of exposure	72
5.8	The expression levels of OsHsp24 at different temperature treatments and duration of exposure.	74
5.9	Relative normalized expression curves of Hsp70-u2 (A) and OsHsp24 (B) genes under recovery analysis	76
6.1	Improved homozygous heat-tolerant chilli lines screening using foreground makers (Hsp70-u2 and AGi42) in selected plant population P <sub>1</sub> (A) and P <sub>4</sub> (B). Running on 2.5% metaphor agarose gel stained with midori green. M: 50bp Ladder	81
6.2	Chromosome-wise recurrent parent genome recovery of the selected best plant (BC <sub>1</sub> -P <sub>7</sub> -P <sub>10</sub> -P <sub>1</sub> -P <sub>7</sub> ) with introgressed gene. A (Red): donor parent, K (Blue): recurrent parent, H (Light green): heterozygous segments from GGT software 2.0v	84
6.3	Chromosome-wise recurrent parent genome recovery of the selected best plant (BC <sub>1</sub> -P <sub>7</sub> -P <sub>10</sub> -P <sub>1</sub> -P <sub>16</sub> ) with introgressed gene. A (Red): donor parent, K (Blue): recurrent parent, H (Light green): heterozygous segments from GGT software 2.0v	85

6.4	Cell membrane stability index of the parental (AVPP0702 and Kulai) as well as the improved heat-tolerant chilli lines exposed to high temperature stress (40 and 50°C). The improved heat-tolerant chilli lines are significantly similar with donor AVPP0702 at $p < 0.05$	88
6.5	Chlorophyll stability index of the parental (AVPP0702 and Kulai) as well as the improved heat-tolerant chilli lines exposed to high temperature stress (40 °C) at different exposure time in experimental plant growth chamber	91
6.6	Correlation matrix among chlorophyll stability index, CSI; cell membrane stability index, CMSI; canopy temperature depression, CTD of the parents and improved heat-tolerant chilli lines	92
6.7	BC <sub>3</sub> F <sub>2</sub> plants overview (A) at 8 weeks after transplanting, plant height (B) and fruit length (C)	95

## LIST OF APPENDICES

Appendix		Page
A	Temperature records during the experimental period (February – April 2016)	118
B	Fertilizer Formulations	120
C	Crossing procedure to generate backcross generations	121
D	Typical spectral pattern for Nucleic acid (DNA and RNA) of some extracted DNA and RNA samples showing their purity	122
E	Amplification Curves of OsHsp24 and Hsp70-u2 when exposed to different temperature treatment in the experimental plant growth chamber using qRT-PCR system	124
F	Graphical genotypes of the selected introgression lines in chilli Kulai genetic make-up developed in this study. A (Red): donor parent, K (Blue): recurrent parent, H (Light green): heterozygous segments from GGT software 2.0v	125



## LIST OF SYMBOLS AND ABBREVIATIONS

ANOVA	Analysis of Variance
ATP	Adenosine Triphosphate
AVRDC	Asian Vegetable Research and Development Centre
bp	Base pair
cm	Centimeter
cDNA	Complementary Deoxyribonucleic Acid
CMSI	Cell Membrane Stability Index
CVMV	Cucumber Mosaic Virus
°C	Degree centigrade
DEPC	Diethylpyrocarbonate
EC	Electrical Conductivity
et al	et alia
gm	Grams
h	hours
HSC	Heat Shock Cognate
HSD	Honest Significant Difference
HSE	Heat Shock Element
HSF	Heat Shock Factor
HSP	Heat Shock Protein
kDa	Kilo Dalton
Kg	Kilogram
μ	micro
μS	microSiemens
MAB	Marker-assisted backcrossing
ml	millilitre
Mol	mole
MPa	Megapascal
%	Percentage
PD	Percentage Damage
PCR	Polymerase Chain Reaction
qRT-PCR	Quantitative Real Time PCR
RNA	Ribonucleic Acid
SE	Standard Error
Sec	Seconds
spp	Species
σ <sup>2</sup>	Variance

# CHAPTER 1

## GENERAL INTRODUCTION

### 1.1 Introduction

Chilli (*Capsicum annuum* L.) varieties are among the high-valued important vegetable widely cultivated mostly as a spice crop (Naik *et al.*, 2017). *Capsicum* belongs to the family Solanaceae together with other vegetable such as tomato, potato, eggplant, tobacco and petunia. Chilli has multiple uses and functions due to its large variability and great geographical distribution worldwide. It can be consumed fresh, cooked or dried. Its production is seasonal due to lack of appropriate varieties and techniques. Optimum day temperatures for chili pepper growth range from 20-30 °C (Naik *et al.*, 2017) and day time temperatures rise above 30°C year round in Malaysia (Sabiha *et al.*, 2015). Such conditions are the important factors limiting the production of chili. Though chili is an important spice crop, still Malaysian domestic production cannot meet the 70% demand due to the poor performance of local varieties under high temperatures (DOSM, 2014). According to DOSM (2014), the self-sufficiency level in chilli is 52% and import dependency ratio is 52.9% (45, 000 metric tonnes per year).

Plant growth and development is the product of the interaction between the genotype (genetic potential) and the environment in which the plant grows (Blum, 2018). Plant growth and development depends on biochemical processes (e.g. photosynthesis) that in turn depends on factors in the environment in order to proceed optimally (Blum, 2018). When the environmental condition is less than the optimum requirement (20-30 °C) chilli plant experiences stress which adversely affects its growth and development and ultimately, its productivity and economic value (Pessarakli, 2016). The common abiotic stresses that plants may be exposed to include; heat, drought, salinity, and mineral toxicity (Zinn *et al.*, 2010; Hall, 2011). Among the abiotic stresses, temperature increment (transient 10 – 15 °C above ambient) due to changing climatic conditions is a serious threat which affects crop production (Jones *et al.*, 1999). Heat stress occurs when temperatures are high enough (above 40 °C) to cause irreversible damage to plant function (Hall, 2011). Heat stress affects various physiological and metabolic processes in plants such as uptake of water and ions, translocation of solutes, photosynthesis and respiration, and produce inactivation of enzymes, accumulation of unprocessed peptides, and is detrimental in terms of growth and productivity (Larkindale and Vierling, 2008; Frank *et al.*, 2009; Snider *et al.*, 2009; Saha *et al.*, 2010; Ahmed and Hassan, 2011). A single hot day (above 38 °C) can be fatal to reproductive success for many plant species including chilli (Kelly *et al.*, 2010). Most physiological processes of plants remain normal at temperatures ranging from approximately 0 to 40 °C. Well-known responses of plants to extreme temperatures are cold acclimatization and vernalization (Sheldon *et al.*, 2000). The molecular mechanisms of these responses have recently been determined through intensive studies (Sharma *et al.*, 2005; Kuwabara and Imai, 2009). Under increased temperature (above 30 °C) flowering asynchrony, decreased pollen fertility and, abscission of flower buds, flowers and fruits are more common in chilli (Barnabas *et al.*, 2008; Hedhly *et al.*, 2009; Craufurd and Wheeler, 2009; Thakur *et al.*, 2010). The

commercial vegetable growers are quite aware about the importance of new varieties due to having high yield potential with uniformity in maturity, having tolerance to abiotic and biotic stresses, and better quality as compared to the standard open pollinated varieties (Wilson *et al.*, 2012). Suhana *et al* (2014) reported that hybrid chilli 'Ch5' exhibited better performance, with heaviest mean yield per plot (13317 kg) and fruit weight (15.86 g) compared with the MC12 indicating good prospect for commercial cultivation. A hybrid tomato resistant to *Helicoverpa armigera* damage recorded higher yield per plant compared with the local variety (Degri and Sani, 2015).

Stress resistance is an inherent part of all cultivar development programs. Typical response to environmental stress conditions is established by the induction of a set of stress proteins that protects the organism from cellular damage. A basic response of plants to heat stress that is very common is the heat shock proteins (Hsps) expression, which is known as a mechanism in plant tolerance to heat stress (Feder and Hoffman, 1999). Incorporation of heat shock proteins has shown considerable success in improvement of crop plants against heat stress. In breeding heat-tolerant maize, a 45 kDa Hsp was found in F<sub>2</sub> population produced from a cross between ZPBL 1304 (tolerant to heat) that produced a 45 kDa Hsp and ZPL 389 line that was not tolerant to heat and does not produce this protein. These F<sub>2</sub> plants become more resistant to heat stress (Ristic *et al.*, 1998, Ristic *et al.*, 1991). In whatever way, screening genetic materials against high temperature depends on field and whole-plant techniques, which as a result of environmental interaction effect their efficiency is less and sensitive (Hall, 2011). Over expression of a rice chloroplast sHsp (Oshsp26) gene showed more tolerance to high temperature and oxidative stresses in *E. coli* (Lee *et al.*, 2000). Research have been intensified in determining efficient, suitable and accurate strategies that allow screening large number of genetic materials at the same time so as to breed chilli for heat tolerance in hot and humid areas (Gajanayake *et al.*, 2011). However, no work has yet been reported on the incorporation of Hsps in chilli for the improvement of chilli tolerance to heat.

Ancient manual breeding approach contributed substantially to the hereditary change of chilli germplasm in the most recent century (Reddy *et al.*, 2014). Ancient manual breeding has additionally been utilized to purposefully grow new heat tolerant genotypes (Driedonks *et al.*, 2016). For instance, an assortment of broccoli has an enhanced head quality on account of early development, since this attribute counteracts hot days after the fact in season to influence the warmth touchy blossom start formative stage (Farnham and Bjorkman 2011). In potato breeding a hereditary pick up was gotten after three cycles of intermittent determination for warm resilience prompting solid increment in yield up to 37.8 % (Benites and Pinto, 2011). Albeit regular "yield" breeding has prevailed with regards to creating heat-tolerant lines, a definitive hereditary and physiological base of the changes stay hazy. This keeps the advancement of molecular or different biomarkers, which would help germplasm screening for enhanced heat resistance and take into account productive breeding of the intricate characteristic. Another disadvantage of ancient manual breeding is that the projects are frequently in view of crossing moderately propelled starting material, which has just been utilized as a part of the specific breeding zones particularly identified with the market segment that is focused on. This suggests the potential pick up in heat tolerance level is constrained by the low genetic diversity (Paran and Van

Der Knaap, 2007). Backcross breeding approach can be employed to introduce a specific trait, such as heat tolerance, from one line, often an unimproved line, to another line that is typically an elite breeding line (Hain and Lee, 2005). The backcrossing process can often be accelerated using marker-assisted backcrossing, also known as background selection using molecular markers (Byrne and Richardson, 2005).

Molecular markers are effective in cultivar identification for protecting proprietary rights as well as authenticating plant cultivars. Molecular markers of necessity are applied based on two basic chemical procedures - protein and DNA markers. There are many DNA markers used which include Restriction Fragment Length Polymorphism (RFLP), Simple Sequence Repeats (SSRs), Amplified Fragment Length Polymorphism (AFLP), and Single Nucleotide Polymorphism (SNPs) (Dhaliwal *et al.*, 2013). In general, improved varieties have better yield characteristics and suffer less from different stresses than the traditional ones. Therefore, no research have been reported so far for the incorporation of Hsps in chilli to improve or develop heat tolerant genotypes.

## 1.2 Justification

Malaysian average temperature is projected to increase from 0.3 to 4.5 °C due to greenhouse emission (Alam *et al.*, 2011) and the warmer temperature will cause a rise in sea level and thus will reduce crop yield (Alam *et al.*, 2011). With a population of about 31.19 Million, Malaysia is ranked 26<sup>th</sup> largest greenhouse gas emitter in the world (Alam *et al.*, 2011; Hosseini *et al.*, 2013) and its likely to increase due to the growth rate of the emission. This will lead to fluctuation in rainfall and ultimately reduce crop yield and increase the risk of drought (Chong and Mathews, 2001). In Malaysia, the estimated annual production of chilli for 2016 was 43,738 metric tonnes (Mt) planted over 4,020 ha cultivated land area (DOA, 2016). While the estimated total chilli consumption is 62,380 Mt (DOSM, 2014), indicating that domestic supplies is insufficient and can hardly meet 70% of the raising demand, thus Malaysia imports approximately 45,000 metric tonnes per year. Moreover, recently chilli output in Malaysia dropped drastically from 47,015 Mt in 2015 to 43,738 Mt in 2016 (DoA, 2016) due to El Nino, leading to the shortage of supply. El Nino is a complex series of climatic changes that occurs irregularly and affects sea surface temperature in most tropics and subtropics. To increase production there is the need to improve/produce varieties having high yielding potential as well as tolerance against high temperature that will be suitable for lowland cultivation. Under this context, it is imperative to identify and incorporate Hsps in chilli for the development of heat tolerant as well as high yielding chilli variety. The experience on the possibility of exploiting the hybrid vigor in chilli has shown considerable promise and as such manoeuvres to control Hsp genes production help in breeding chilli genotypes tolerance to high temperature.

### 1.3 Objectives

**The main objective of this study was:**

To develop heat tolerant variety of chilli with high yielding potential

**The Specific objectives were:**

- i. To introgress heat shock protein genes from AVPP0702 variety, donor to Kulai 907, recipient parent
- ii. To identify polymorphic molecular markers for chilli heat tolerant characteristics and recipient genome
- iii. To validate the backcross progenies for heat tolerance with the polymorphic molecular markers (Hsp loci) and to determine the percentage recovery of Kulai 907 genome in BC<sub>3</sub>F<sub>2</sub> population.
- iv. To determine the homozygous lines that are positive to Hsp genes and phenotypic performance of the BC<sub>3</sub>F<sub>2</sub> improved heat-tolerant chilli lines similar with Kulai 907.

## REFERENCES

- Ahmed, J.U. & Hassan, M.A. (2011). Evaluation of seedling proline content of wheat genotypes in relation to heat tolerance. *Bangladesh Journal of Botany*, 40(1): 17-22.
- Åkerfelt, M., Morimoto, R.I. & Sistonen, L. (2010). Heat shock factors: integrators of cell stress, development and lifespan. *Nature Reviews Molecular Cell Biology*, 11(8): 545–555.
- Alam, M., Siwar, C., Murad, W. & bin Toriman, M.E. (2011). Farm level assessment of climate change, agriculture and food security issues in Malaysia. *World Applied Sciences Journal*, 14(3): 431-442.
- Ali, A.M., & Kelly, W. C. (1993). Effect of Pre-anthesis Temperature on the Size and the Shape of Sweet Pepper (*Capsicum spp.* L.) Fruit. *Scientia Horticulturae*, 54: 97-105
- Alvim, F.C., Carolino, S.M., Cascardo, J.C., Nunes, C.C., Martinez, C.A, Otoni, W.C., & Fontes, E.P. (2001). Enhanced accumulation of BiP in transgenic plants confers tolerance to water stress. *Plant Physiology*, 126(7): 1042–1054.
- Amarawathi, Y., Singh, R., Singh A.K., Singh, V.P., Mohapatra, T., Sharma, T.R. & Singh, N.K. (2008). Mapping of quantitative trait loci for basmati quality traits in rice (*Oryza sativa* L.). *Molecular Breeding*, 21: 49-65
- Allard, R.W. (1999). *Principles of Plant Breeding*, 2nd edn. Wiley and Sons New York.
- Ashraf, M., Akram, N.A., Rahman, M. & Foolad M.R. (2012). Marker-assisted selection in plant breeding for salinity tolerance. *Plant Salt Tolerance: Methods and Protocols, Methods in Molecular Biology*, 305-333.
- AVRDC, IPGRI & CATE, (1995). *Descriptors for Capsicum (Capsicum spp)*. International Plant Genetic Resources Institute, Rome, Italy.
- AVRDC, IPGRI & CATE, (2001). *Descriptors for Capsicum (Capsicum spp)*. International Plant Genetic Resources Institute, Rome, Italy. Revised.
- Ayres, N.M., McClung, A.M., Larkin, P.D., Bligh, H.F., Jones, C.A. & Park, W.D. (1997). Microsatellites and a single-nucleotide polymorphism differentiate apparent amylose classes in an extended pedigree of US rice germplasm. *Theoretical and Applied Genetics*, 9: 773-781.
- Baba, V.Y., Rocha, K.R., Gomes, G.P., de Fátima Ruas, C., Ruas, P.M., Rodrigues, R., & Gonçalves, L.S.A. (2016). Genetic diversity of *Capsicum chinense* accessions based on fruit morphological characterization and AFLP markers. *Genetic resources and crop evolution*, 63(8): 1371-1381.
- Babu, R., Nair, S.K., Kumar, A., Venkatesh, S., Sekhar, J.C., Singh, N.N., Srinivasan, G. & Gupta, H.S. (2005). Two-generation marker-aided backcrossing for rapid

conversion of normal maize lines to quality protein maize (QPM). *Theoretical and Applied Genetics*, 111: 888-897.

- Backonja, M.M., Malan, T.P., Vanhove, G.F. & Tobias, J.K. (2010). NGX-4010, a high-concentration capsaicin patch, for the treatment of postherpetic neuralgia: A randomized, double-blind, controlled study with an open-label extension. *Pain Medicine*, 11: 600–608.
- Baral, J. & Bosland, P.W. (2002). Genetic diversity of a Capsicum germplasm collection from Nepal as determined by randomly amplified polymorphic DNA markers. *Journal of the American Society for Horticultural Science*, 127(3): 318-324.
- Barnabás, B., Jäger, K., & Fehér, A. (2008). The effect of drought and heat stress on reproductive processes in cereals. *Plant, Cell and Environment*, 31: 11–38.
- Barone, A., Ercolano, M.R., Langella, R., Monti, L. & Frusciante, L. (2005). Molecular marker-assisted selection for pyramiding resistance genes in tomato. *Advances in Horticultural Science*, 19: 147-152
- Basavaraj, S.H., Singh, V.K., Singh, A.T., Singh, A.S., Singh, A.N., Anand, D., Yadav, S., Ellur, R.K., Singh, D., Gopalakrishnan, S., Nagarajan, M., Mohapatra, T., Prabhu, K.V. & Singh, A.K. (2010). Marker assisted improvement of bacterial blight resistance in parental lines of Pusa RH10, a superfine grain aromatic rice hybrid. *Molecular Breeding*, 26: 293-305.
- Benites, F.R.G., & Pinto, C.A.B.P. (2011). Genetic gains for heat tolerance in potato in three cycles of recurrent selection. *Crop breeding and applied biotechnology*, 11(2): 133-140.
- Bin, W.S., Wei, L.K., Ping, D.W., Li, Z., Wei, G., Bing, L.J. & Feng, C.J. (2012). Evaluation of appropriate reference genes for gene expression studies in pepper by quantitative real-time PCR. *Molecular breeding*, 30(3): 1393-1400.
- Bjørnstad, Å., Patil, V., Tekauz, A., Marøy, A.G., Skinnes, H., Jensen, A., Magnus, H., MacKey, J. (2002). Resistance to scald (*Rhynchosporium secalis*) in barley (*Hordeum vulgare*) studied by near-isogenic lines: I. Markers and differential isolates. *Phytopathology*, 92: 710-720.
- Bonnett, D.G., Rebetzke, G. & Spielmeyer, W. (2005). Strategies for efficient implementation of molecular markers in wheat breeding. *Molecular Breeding*, 15: 75-85.
- Blum, A. (1988). *Plant breeding for stress environments*. CRC Press, Inc.
- Blum, A. (2018). *Plant breeding for stress environments*. CRC press, New Edition.
- Bosland, P.W. & Votava, E.J. (2012). *Peppers: Vegetable and Spice Capsicums* J. Atherton and A. Rees (eds), CABI Publishing, pp: 199.

- Bosland, P.W., Votava, E.J. & Votava, E.M. (2012). *Crop Production Science in Horticulture*. In Peppers: Vegetable and spice capsicums, 2nd ed. pp. 105-116. Cabi, United Kingdom.
- Bosland, P.W. (1996). *Capsicums: Innovative uses of an ancient crop*. Progress in new crops. ASHS Press, Arlington, VA, 479-487.
- Bukau, B., Weissman, J. & Horwich, A. (2006). Molecular chaperones and protein quality control. *Cell*, 125: 443-451.
- Byrne, P. & Richardson, K. (2005). Marker-assisted backcrossing for a single gene [Online lesson]. Marker-assisted selection. Plant and Soil Sciences eLibrary, University of Nebraska – Lincoln. Available at: [http://plantandsoil.unl.edu/croptechology2005/pages/index.jsp?what=topic\\_sD&informationModuleId=1087488148&topicOrder=7&max=10&min=0&verified=23 Sept 2010](http://plantandsoil.unl.edu/croptechology2005/pages/index.jsp?what=topic_sD&informationModuleId=1087488148&topicOrder=7&max=10&min=0&verified=23%20Sept%202010)
- Chamikara, M.D.M., Ishan, M., Karunadasa, S.S., Perera, M.K.D.I., Rajapaksha, P.I., Lelwala, R.V., & Sooriyapathirana, S.D.S.S. (2015). Morphological and microsatellite marker analysis of fruit size and shape in selected accessions and commercial cultivars of capsicum species in Sri Lanka. *International Journal of multidisciplinary Studies*, 2(1): 11-32
- Chen, Z.W., Guan H.Z., Wu W.R., Zhou Y.C. & Han, Q.D. (2005). The screening of molecular markers closely linked to rice blast-resistant gene Pi-1 and their application. *Journal of Fujian Agricultural and Forestry University* (Natural Science Edition). 34: 74–77.
- Cheng, J., Qin, C., Tang, X., Zhou, H., Hu, Y., Zhao, Z., Cui, J., Li, B., Wu, Z., Yu, J. & Hu, K. (2016). Development of a SNP array and its application to genetic mapping and diversity assessment in pepper (*Capsicum* spp.). *Scientific Reports*, 6: 1-11
- Chomczynski, P. & Sacchi, N. (1987). Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Analytical Biochemistry*, 162: 156-159.
- Chong, A.L. & Mathews, P. (2001). *National Response Strategies to Climate Change*. In. Putrajaya: Ministry of Science, Technology and the Environment pp 389–422. MOSTE publisher, Putrajaya Malaysia.
- Craufurd, P., Prasad, P.V., Kakani, V., Wheeler, T. & Nigam, S. (2003). Heat tolerance in groundnut. *Field Crops Research*, 80:63-77.
- Craufurd, P.Q. & Wheeler, T.R. (2009). Climate change and the flowering time of annual crops. *Journal of Experimental Botany*, 60(9): 2529–2539.
- Cuc, I.M., Huyen, L.T.N., Hien, P.T.M., Hang, V.T.T., Dam, N., Mui, P.T., Quang, V.D., Ismail, A.M. & Ham, L.H. (2012). Application of marker assisted backcrossing to introgress the submergence tolerance QTL SUB1 into the Vietnam elite rice variety-AS996. *American Journal of Plant Sciences*, 3: 528-536.



- Degri, M.M., & Sani, A.A. (2015). Comparison of Tomato (*Lycopersicon lycopersicum* Mill.) Varieties to Field Insect Pest Infestations. *American Research Journal of Agriculture*, 1(2): 14-17
- De Masi, L., Siviero, P., Esposito, C., Castaldo, D., Siano, F. & Laratta, B. (2006). Assessment of agronomic, chemical and genetic variability in common basil (*Ocimum basilicum* L.). *European Food Research and Technology*, 223(2): 273-281
- De La Peña, R. & Hughes, J. (2007). Improving vegetable productivity in a variable and changing climate. *Journal of SAT Agricultural Research*, 4(1): 1-22.
- Dhaliwal, M.S., Jindal, S.K., Gaikwad, A.K. & Singh, K. (2013). Genetic Diversity Analysis and DNA Fingerprinting of Elite Chilli Pepper Lines Using SSR Markers. *International Journal of Vegetable Science*, 19: 207-216.
- DoA (Department of Agriculture) (2014). Ministry of Agriculture. Malaysia: Kuantiti (MT) dan Nilai (RM) Export Sayur sayuran mengikut Negara dituju@Destinasi bagi tahun 2014.
- DOSM (Department of Statistics, Malaysia) (2014). Supply and Utilization Accounts Selected Agricultural Commodities, Malaysia 2010-2014. Agrofood Statistics 2014, Ministry of Agriculture and Agro-Based Industry, Malaysia.
- Doyle, J.J. & Doyle, J.L. (1990). Isolation of plant DNA from fresh tissue. *Focus* (San Francisco, Calif.) 12: 13-15.
- Dragovic, Z., Broadley S.A., Shomura, Y., Bracher, A. & Hartl, F.U. (2006). Molecular chaperones of the Hsp110 family act as nucleotide exchange factors of Hsp70s. *EMBO Journal*, 25(11): 2519–2528.
- Driedonks, N., Rieu, I., & Vriezen, W.H. (2016). Breeding for plant heat tolerance at vegetative and reproductive stages. *Plant reproduction*, 29(1-2): 67-79.
- Erickson, A.N., & Markhart, A.H. (2002). Flower developmental stage and organ sensitivity of bell pepper (*Capsicum annuum* L.) to elevated temperature. *Plant, Cell & Environment*, 25(1): 123-130.
- Farokhzadeh, S., & AliFakheri, B. (2014). Marker-assisted selection for disease resistance: applications in breeding. *International Journal of Agriculture and Crop Sciences*, 7(14): 1392.
- Farnham, M.W. & Bjorkman, T. (2011). Breeding vegetables adapted to high temperatures: a case study with broccoli. *HortScience*, 46(8): 1093-1097.
- Feder, M.E. & Hoffman G.E. (1999). Heat shock proteins, molecular chaperones, and the Stress response: evolutionary and ecological physiology. *Annual Review of Physiology*, 61: 243-282.
- Fei, J., Wang, Y.S., Zhou, Q. & Gu, J.D. (2015). Cloning and expression analysis of HSP70 gene from mangrove plant *Kandelia obovata* under cold stress. *Ecotoxicology*, 24(7-8): 1677–1685.

- Ferguson, D.L., Guikema, J.A. & Paulsen, G.M. (1990). Ubiquitin pool modulation and protein degradation in wheat roots during high temperature stress. *Plant Physiology*, 92:740-746.
- Frank, G., Pressman, E., Ophir, R., Althan, L., Shaked, R., Freedman, M., Shen, S., & Firon, N. (2009). Transcriptional profiling of maturing tomato (*Solanum lycopersicum* L.) microspores reveals the involvement of heat shock proteins, ROS scavengers, hormones, and sugars in the heat stress response. *Journal Experimental Botany*, 60: 3891–3908.
- Gajanayake, B., Trader, B.W., Reddy, K.R. & Harkess, R.L. (2011). Screening ornamental pepper cultivars for temperature tolerance using pollen and physiological parameters. *Hortscience*, 46: 878-884.
- Geleta, L.F. Labuschagne, M.T. & Viljoen, C.D. (2004). Relationship between heterosis and genetic distance based on morphological traits and AFLP markers in pepper. *Plant Breeding*, 123: 467-473.
- Ghasemi, M., Babaeian Jelodar, N., Modarresi, M., Bagheri, N., & Jamali, A. (2016a). Increase of chamazulene and  $\alpha$ -bisabolol contents of the essential oil of german chamomile (*Matricaria chamomila* L.) using salicylic acid treatments under normal and heat stress conditions. *Foods*, 5: 56-70
- Ghasemi, M., Modarresi, M., Babaeian Jelodar, N., Bagheri, N. & Jamali, A. (2016b). The evaluation of exogenous application of salicylic acid on physiological characteristics, proline and essential oil content of chamomile (*Matricaria chamomila* L.) under normal and heat stress conditions. *Agriculture*, 6: 31-46
- Gorovits, R., Moshe, A., Ghanim, M. & Czosnek, H. (2013). Recruitment of the Host Plant Heat Shock Protein 70 by Tomato Yellow Leaf Curl Virus Coat Protein Is Required for Virus Infection. *PLoS One*, 8(7): 1-11
- Guo, M., Zhai, Y., Lu, J., Chai, L., Chai, W. & Gong, Z. (2014). Characterization of CaHsp70-1, a Pepper Heat-Shock Protein Gene in Response to Heat Stress and Some Regulation Exogenous Substances in *Capsicum annuum* L. *International Journal of Molecular Science*, 15: 19741–19759.
- Gupta, S.C., Sharma, A., Mishra, M., Mishra, R.K. & Chowdhuri, D.K. (2010). Heat shock proteins in toxicology: How close and how far? *Life Sciences*, 86: 377-384.
- Guy, C. (1999). Molecular responses of plants to cold shock and cold acclimation. *Journal of Molecular Microbiology and Biotechnology*, 1(2): 231-242.
- Hain, P. & Lee, D. (2005). Backcross breeding 2 - the backcrossing process [Online lesson]. Plant and Soil Sciences eLibrary, University of Nebraska – Lincoln. Available at:<http://plantandsoil.unl.edu/croptechnology2005/pages/index.jsp?what=topics&D&topicOrder=1&informationModuleId=959009357> (verified 23 Sept 2010).
- Hall, A.E. (1992). Breeding for heat tolerance. *Plant Breeding Review*, 10:129-168.

- Hall, A.E. (2011). *Crop Responses to Environment*. CRC press LLC, Boca Raton, Florida.
- Han, K., Jeong, H. J., Sung, J., Keum, Y. S., Cho, M. C., Kim, J. H., ... & Kang, B. C. (2013). Biosynthesis of capsinoid is controlled by the *Pun1* locus in pepper. *Molecular breeding*, 31(3): 537-548.
- Hanáček, R., Vyhnaněk, T., Rohrer, M., Cieslarová, J. & Stavěliková, H. (2009). DNA polymorphism in genetic resources of red pepper using microsatellite markers. *Horticultural Science*, 36(4): 127-132.
- Hanson, P.M., Chen, J.T. & Kuo, C.G. (2002). Gene action and heritability of high temperature fruit set in tomato line CL5915. *HortScience*, 37:172-175.
- Hartl, F.U, Andreas, B. & M.H.-H (2011). Molecular chaperones in protein folding and proteostasis. *Nature*, 475(7356): 324–332.
- Hedhly, A., Hormaza, J.I., & Herrero, M. (2009). Global warming and sexual plant reproduction. *Trends in Plant Science*, 14: 30–36.
- Hospital, F. & Charcosset, A. (1997). Marker-assisted introgression of quantitative trait loci. *Genetics*, 147: 1469–1485
- Hosseini, S. E., Wahid, M. A., & Aghili, N. (2013). The scenario of greenhouse gases reduction in Malaysia. *Renewable and Sustainable Energy Reviews*, 28: 400-409.
- Hu, W., Hu, G. & Han, B. (2009). Genome-wide survey and expression profiling of heat shock proteins and heat shock factors revealed overlapped and stress specific response under abiotic stresses in rice. *Plant Science*, 176:583-590.
- Hua, J. (2009). From freezing to scorching, transcriptional responses to temperature variations in plants. *Current opinion in plant biology*, 12(5): 568-573.
- Hurni, S., Brunner, S., Stirnweis, D., Herren, G., Peditto, D. & McIntosh, R.A. (2014). The powdery mildew resistance gene Pm8 derived from rye is suppressed by its wheat ortholog Pm3. *Plant Journal*, 79: 904-913.
- IBPGR (1983). *Genetic resources of Capsicum*. International Board for Plant Genetic Resources, Rome, Italy.
- Iftekharruddaula, K.M. (2008). *Comparison of new selection strategies or marker assisted backcrossing for a submergence tolerant gene rice*. Doctoral dissertation. Bangladesh Agricultural University Mymensingh, Bangladesh, pp 1-205
- Ilbi, H. (2003). RAPD markers assisted varietal identification and genetic purity test in pepper, *Capsicum annum*. *Scientia Horticulturae*, 97:211–218.
- Ince, A.G., Karaca, M. & Onus, A.N. (2010). Polymorphic microsatellite markers transferable across *Capsicum* species. *Plant Molecular Biology Reporter*, 28(2): 285-291.

- Islam, M.S., Ahmad, S., Uddin, M.N., Rafii, M.Y., Ismail, M.R. & Malek, M.A. (2014). Identification of tomato inbred lines for heat tolerance through agronomic and physiological approaches. *Journal of Food, Agriculture and Environment* 12(1):281-284.
- Jagtap, A.B., Sujatha, R., Nazeem, P.A., Meena, O.P. & Pathania, S. (2016). Morpho-molecular characterization of putative interspecific crosses in black pepper (*Piper nigrum* L. and *Piper colubrinum*). *Plant Omics*, 9(1): 73-80.
- Jeong, H.S., Jang, S., Han, K., Kwon, J.K. & Kang, B.C. (2015). Marker-assisted backcross breeding for development of pepper varieties (*Capsicum annuum*) containing capsaicinoids. *Molecular Breeding*, 35(12): 226 – 236.
- Jones, P.D. New, M., Parker, D.E., Martin, S. & Rigor, I.G. (1999). Surface air temperature and its changes over the past 150years. *Reviews of Geophysics*, 37(2): 173–199
- Jungkunz, I., Link, K., Vogel, F., Voll, L.M., Sonnewald, S. & Sonnewald, U. (2011). AtHsp70-15-deficient Arabidopsis plants are characterized by reduced growth, a constitutive cytosolic protein response and enhanced resistance to TuMV. *Plant Journal*, 66(6): 983–995
- Katiyar-Agarwal, S., Agarwal, M. & Grover, A. (2003). Heat-tolerant basmati rice engineered by over-expression of hsp101. *Plant Molecular Biology*, 51(5): 677-686.
- Kelly, E.Z., Meral, T.O. & Jeffrey, F.H. (2010). Review paper: Temperature stress and plant sexual reproduction: uncovering the weakest links. *Journal of Experimental Botany*, 61(7): 1959-1968.
- Khanh, T.D., Linh, L.H., Linh, T.H., Ham, I.H. & Uan, T.D. (2013). Rapid and high-precision marker assisted backcrossing to introgress the SUB1 QTL into the Vietnamese elite rice variety. *Journal of Plant Breeding and Crop Science*, 5: 26-33
- Kim, S., Park, M., Yeom, S.I., Kim, Y.M., Lee, J.M., Lee, H.A., ... & Jung, K. (2014). Genome sequence of the hot pepper provides insights into the evolution of pungency in *Capsicum* species. *Nature genetics*, 46(3): 270-279
- Koizumi, S., Ohama, N., Mizoi, J., Shinozaki, K. & Yamaguchi-Shinozaki, K. (2014). Functional analysis of the Hikeshi-like protein and its interaction with HSP70 in Arabidopsis. *Biochemical and Biophysical Research Communications*, 450(1): 396–40
- Kong, Q., Zhang, G., Chen, W., Zhang, Z. & Zou, X. (2012). Identification and development of polymorphic EST-SSR markers by sequence alignment in pepper, *Capsicum annuum* (Solanaceae). *American Journal of Botany*, 99(2): 59-61
- Krishnamurthy, S. L., Prashanth, Y., Rao, A. M., Reddy, K. M., & Ramachandra, R. (2015). Assessment of AFLP marker based genetic diversity in chilli

(*Capsicum annuum* L. & *C. baccatum* L.). *Indian Journal of Biotechnology*, 14: 49-54

- Kumar, G.S., Kumari, K.A., Rani, C.V.D., Sundaram, R.M., Vanisree, S., Jamaloddin, M. & Swathi, G. (2013). Study of simple sequence repeat (SSR) polymorphism for biotic stress resistance in elite rice variety JGL 1798. *African Journal of Biotechnology*, 12(40): 5833.
- Kuwabara, C. & Imai, R. (2009). Molecular basis of disease resistance acquired through cold acclimation in overwintering plants. *Journal of Plant Biology*, 52: 19-26.
- Kwon, Y.S., Lee, J.M., Yi, G.B., Yi, S.I., Kim, K.M., Soh, E.H. & Kim, B.D. (2005). Use of SSR markers to complement tests of distinctiveness, uniformity, and stability (DUS) of pepper (*Capsicum annuum* L.) varieties. *Molecular Cells*, 19(3): 428-435.
- Lagercrantz, U., Ellegren, H. & Andersson, L. (1993). The abundance of various polymorphic microsatellite motifs differs between plants and vertebrates. *Nucleic Acids Research*, 21(5): 1111-1115.
- Lang, N.T., Ha, P.T.T., Tru, P.C., Toan, T.B., Buu, B.C. & Cho, Y.C. (2015). Breeding for Heat Tolerance Rice Based on Marker-Assisted Backcrossing in Vietnam. *Plant Breeding and Biotechnology*, 3(3): 274-281.
- Lang, Y., Kisaka, H., Sugiyama, R., Nomura, K., Morita, A., Watanabe, T., ... & Miwa, T. (2009). Functional loss of *pAMT* results in biosynthesis of capsinoids, capsaicinoid analogs, in *Capsicum annuum* cv. CH-19 Sweet. *The Plant Journal*, 59(6): 953-961.
- Larkindale, J. & Vierling, E. (2008). Core genome responses involved in acclimation to high temperature. *Plant Physiology*, 146:748-761.
- Lau, W.C.P., Rafii, M.Y., Ismail, M.R., Puteh, A., Latif, M.A., Asfaliza, R. & Miah, G. (2017). Development of advanced fragrant rice lines from MR269 3 × Basmati 370 through marker-assisted backcrossing. *Euphytica*, 213: 11 doi: 10.1007/s10681-016-1794-z.
- Lawson, D.M., Lunde, C.F. & Mutschler, M.A. (1997). Marker-assisted transfer of acylsugar-mediated pest resistance from the wild tomato, *Lycopersicon pennellii*, to the cultivated tomato, *Lycopersicon esculentum*. *Molecular Breeding*, 3: 307-317.
- Leborgne-Castel, N., Jelitto-Van Doorena, E.P., Crofts, J. & Denecke, J. (1999). Overexpression of BiP in tobacco alleviates endoplasmic reticulum stress. *Plant Cell*, 11(3): 459-470.
- Lecomte, L., Duffé, P., Buret, M., Servin, B., Hospital, F. & Causse, M. (2004). Marker-assisted introgression of five QTLs controlling fruit quality traits into three tomato lines revealed interactions between QTLs and genetic backgrounds. *Theoretical and Applied Genetics*, 109: 658-668.

- Lee, S., Lee, D.W., Lee, Y., Mayer, U., Stierhof, Y.D., Lee, S., Jürgens, G. & Hwang, I. (2009). Heat shock protein cognate 70-4 and an E3 ubiquitin ligase, CHIP, mediate plastid-destined precursor degradation through the ubiquitin-26S proteasome system in Arabidopsis. *Plant Cell*, 21(12): 3984–4001.
- Lee, J.M., Nahm, S.H., Kim, Y.M. & Kim, B.D. (2004). Characterization and molecular genetics mapping of microsatellite loci in pepper. *Theoretical and Applied Genetics*, 108(4): 619-627.
- Lee, B.H., Won, S.H., Lee, H.S., Miyao, M., Chung, W.I., Kim, I.J. & Jo, J. (2000). Expression of the chloroplast-localized small heat shock protein by oxidative stress in rice. *Gene*, 245(2): 283-290.
- Lee, C. J., Yoo, E., Shin, J. H., Lee, J., Hwang, H. S., & Kim, B. D. (2005). Non-pungent Capsicum contains a deletion in the capsaicinoid synthetase gene, which allows early detection of pungency with SCAR markers. *Molecules & Cells* (Springer Science & Business Media BV), 19(2): 262-267.
- Lefebvre, V., Goffinet, B., Chauvet, J.C., Caromel, B., Signoret, P., Brand, R. & Palloix A. (2001). Evaluation of genetic distances between peppers inbred lines for cultivar protection purposes: comparison of AFLP, RAPD and phenotypic data. *Theoretical and Applied Genetics*, 102:741–750
- Li, W., Cheng, J., Wu, Z., Qin, C., Tan, S., Tang, X., Cui, J., Zhang, L. & Hu, K., (2015). An InDel-based linkage map of hot pepper (*Capsicum annuum*). *Molecular Breeding*, 35(1): p.32.
- Lin, B.L., Wang, J.S., Liu, H.C., Chen, R.W., Meyer, Y., Barakat, A. & Delseny, M. (2001). Genomic analysis of the Hsp70 superfamily in *Arabidopsis thaliana*. *Cell Stress and Chaperones*, 6: 201–208.
- Lindquist, S. (1986). The heat-shock response. *Annual Review of Biochemistry*, 55:1151-1191.
- Liu, Z.J. & Cordes, J.F. (2004). DNA marker technologies and their application in aquaculture genetics. *Aquaculture*, 238: 1-37.
- Livak, K. J., & Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 25: 402-408.
- Loss, C.R. & Bouzari, A. (2016). On food and chemesthesis–food science and culinary perspectives. *Chemesthesis: Chemical Touch in Food and Eating*, 250.
- Lu, R.C., Tan, M.S., Wang, H., Xie, A.M., Yu, J.T., & Tan, L. (2014). Heat shock protein 70 in Alzheimer’s disease. *BioMed Research International*, 2014. doi.org/10.1155/2014/435203
- Luders, E., Toga, A.W., Lepore, N. & Gaser, C. (2009). The underlying anatomical correlates of long-term meditation: Larger hippocampal and frontal volumes of gray matter. *Neuroimage*, 45(3): 672–678.

- Lutz, N. (1991). *Heat shock response*. Boca Raton, NW: CRC Press. p. 138.
- Magaji, U.G., Rafii, M.Y., Martini, M.Y., Yusuff, O.A. & Ismail M.R. (2016). Polymorphism between popular Kulai and heat tolerant donor AVPP0702 chilli peppers revealed through SSR markers. *Transactions of Persatuan Genetik Malaysia*, 3: 39-44.
- Mayer, M.P. (2010). Gymnastics of molecular chaperones. *Molecular Cell* 39(3): 321–331. Available at <http://dx.doi.org/10.1016/j.molcel.2010.07.012>.
- Mayer, M.P. & Bukau, B. (2005). Hsp70 chaperones: Cellular functions and molecular mechanism. *Cellular and Molecular Life Sciences*, 62(6): 670–684
- MARDI - Malaysian Agriculture Research and Development Institute (2009). Ingin Meningkatkan Hasil Dan Kualiti Produk Pertanian Anda? MARDI publisher, Selangor, Malaysia
- Miah, G., Rafii, M.Y., Ismail, M.R., Puteh, A.B., Rahim, H.A. & Latif, M.A. (2015). Recurrent parent genome recovery analysis in a marker-assisted backcrossing program of rice (*Oryza sativa* L.). *Comptes Rendus Biologies*, 338(2): 83-94.
- Mimura, Y., Inoue, T., Minamiyama, Y. & Kubo, N. (2012). An SSR-based genetic map of pepper (*Capsicum annuum* L.) serves as an anchor for the alignment of major pepper maps. *Breeding Science*, 62(1): 93-98.
- Minamiyama, Y., Tsuru, M. & Hirai, M. (2006). An SSR-based linkage map of *Capsicum annuum*. *Molecular Breeding*, 18(2): 157-169.
- Mirus, O. & Schleiff, E. (2009). The evolution of tetratricopeptide repeat domain containing receptors involved in protein translocation--review. *Endocytobiosis and Cell Research*, 19: 31–50
- MoA (Ministry of Agriculture), (2012). Malaysia: Kuantiti (MT) dan Nilai (RM) Export Sayur sayuran mengikut Negara dituju@Destinasi baagi tahun 2011.
- Mohan, M., Nair, S., Bhagwat, A., Krishna, T.G. & Yano, M. (2000). Genome mapping, molecular markers and marker-assisted selection in crop plants. *Molecular Breeding*, 3: 87-103.
- Muthusamy, V., Hossain, F., Thirunavukkarasu, N., Choudhary, M., Saha, S., Bhat, J.S., Prasanna, B.M. & Gupta, H.S. (2014). Development of beta-carotene rich maize hybrids through marker-assisted introgression of beta-carotene hydroxylase allele. *PLoS ONE*, 9: 1 – 22.
- Nagy, I., Stágel, A., Sasvári, Z., Röder, M. & Ganai, M. (2007). Development, characterization, and transferability to other Solanaceae of microsatellite markers in pepper (*Capsicum annuum* L.). *Genome*, 50(7): 668-688.
- Naik, P. S., Singh, M., & Ranjan, J. K. (2017). Impact of Climate Change on Vegetable Production and Adaptation Measures. In *Abiotic Stress Management for Resilient Agriculture* (pp. 413-428). Springer, Singapore.

- Nakamoto, H. & Víg, L. (2007). The small heat shock proteins and their clients. *Cellular and Molecular Life Sciences*, 64:294-306.
- Nicolaï, M., Cantet, M., Lefebvre, V., Sage-Palloix, A.M. & Palloix, A. (2013). Genotyping a large collection of pepper (*Capsicum spp.*) with SSR loci brings new evidence for the wild origin of cultivated *C. annuum* and the structuring of genetic diversity by human selection of cultivar types. *Genetic Resources and Crop Evolution*, 60(8): 2375-2390.
- Oh, S.J., Song, J.Y., Lee, J.R., Lee, G.A., Ko, H.C., Stoilova, T. & Ro, N.Y. (2012). Evaluation of Genetic Diversity of Red Pepper Landraces (*Capsicum annuum* L.) from Bulgaria Using SSR Markers. *The Journal of the Korean Society of International Agriculture*, 24(5): pp.547-556
- Opena, R.T., Green, S.K., Talekar, N.S. & Chen, J.T. (2009). *Genetic improvement of tomato adaptability to the tropics: Progress and future prospects*. In: Green SK (ed) *Tomato and pepper production in the tropics*. AVRDC, Shanhua, Taiwan pp 70-85.
- Opena, R.T. & SH, L.O. (1999). Breeding for heat tolerance in heading Chinese cabbage. In: Talekar NS, Griggs TD (eds). *Proceedings of the 1st International Symposium on Chinese cabbage*. AVRDC, Shanhua, Taiwan.
- Ottaviano, E., Gorla, M.S., Pe, E. & Frova, C. (1991). Molecular markers (RFLPs and HSPs) for the genetic dissection of thermo-tolerance in maize. *Theoretical and Applied Genetics*, 81(6): 713-719.
- Pagamas, P. & Nawata, E. (2008). Effect of Heat Stress Application to Flower and Fruit on Seed Quality of Chili Pepper. *Tropical Agriculture and Development*, 52: 82-87.
- Paran I., & Van Der Knaap E. (2007). Genetic and molecular regulation of fruit and plant domestication traits in tomato and pepper. *Journal of Experimental Botany*, 58: 3841–3852.
- Parsell, D. & Lindquist, S. (1993). The function of heat-shock proteins in stress tolerance: Degradation and reactivation of damaged proteins. *Annual Review of Genetics* 27: 437-496.
- Paulus, A.D. (2014). Strategies, progress and achievements of pepper breeding. In: *A Practical Compendium on Advances in Plant Breeding*. Second International Plant Breeding Seminar: Advanced Breeding Strategies in Crop Improvement, Genetic Society of Malaysia. pp 169-174.
- Pessarakli, M. (Ed.). (2016). *Handbook of plant and crop stress*. CRC press.
- Pfaffl, M.W. (2001). A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Research*, 29(9): 2002-2007.
- Pickersgill, B. (1997). Genetic resources and breeding of *Capsicum* spp. *Euphytica* 96:129-133.



- Purseglove, J. (1987). *Tropical crops: Dicotyledons* (low priced edn). England longmans book society, pp 719.
- Rapurohit, D., Kumar, R., Kumar, M., Paul, P., Awasthi, A.A., Basha, P.O., Puri, A., Jhang, T., Singh, K. & Dhaliwal, H.S. (2011). Pyramidin of two bacterial blight resistances and a semi dwarfing gene in Type 3 Basmati using marker-assisted selection. *Euphytica*, 178: 111-126.
- Rao, G.U, Ben Chaim, A., Borovsky, Y., & Paran, I. (2003). Mapping of yield-related QTLs in pepper in an interspecific cross of *Capsicum annuum* and *C. frutescens*. *Theoretical and Applied Genetics*, 106: 1457–1466
- Reddy, M.K., Srivastava, A., Kumar, S., Kumar, R., Chawda, N., Ebert, A.W. & Vishwakarma, M. (2014). Chilli (*Capsicum annuum* L.) breeding in India: an overview. *SABRAO Journal of Breeding and Genetics*, 46(2): 160-173.
- Reyes-Escogido, M.L., Gonzalez-Mondragon, E.G. & Vazquez-Tzompantzi, E. (2011). Chemical and Pharmacological Aspects of Capsaicin. *Molecules*, 16, 1253–1270.
- Reynolds, M.P., Singh, R.P., Ibrahim, A., Ageeb, O.A.A., Larque-Saavedra, A. & Quick, J.S. (1998). Evaluating the physiological traits to complement empirical selection for wheat in warm environments. *Euphytica*, 100: 85-94.
- Richter, K., Haslbeck, M. & Buchner, J. (2010). The heat shock response: Life on the verge of death. *Molecular Cell*, 40:253-266
- Ristic, Z., Gifford, D.J. & Cass, D.D. (1991). Heat Shock Proteins in Two Lines of Zea mays L. That Differ in Drought and Heat Resistance. *Plant Physiology*, 97(4): 1430–1434.
- Ristic, Z., Yang, G., Martin, B. & Fullerton, S. (1998). Evidence of association between specific heat-shock protein (s) and the drought and heat tolerance phenotype in maize. *Journal of Plant Physiology*, 153(3): 497-505.
- Rodriguez, J.M., Berke, T., Engle, L. & Nienhuis, J. (1999). Variation among and within *Capsicum* species revealed by RAPD markers. *Theoretical and Applied Genetics*, 99:147–156.
- Sabiha, M. A., Saidur, R., Hassani, S., Said, Z., & Mekhilef, S. (2015). Energy performance of an evacuated tube solar collector using single walled carbon nanotubes nanofluids. *Energy Conversion and Management*, 105, 1377-1388.
- Saha, S.R., Hossain, M.M., Rahman, M.M., Kuo, C.G. & Abdullah, S. (2010). Effect of high temperature stress on the performance of twelve sweet pepper genotypes. *Bangladesh Journal of Agricultural Research*, 35(3): 525-534.
- Saghai-Marooif, M.A., Soliman, K.M., Jorgensen, A.R. & Allard, R.W. (1984). Ribosomal DNA spacer length polymorphism in barley: Inheritance, chromosomal location and population dynamics. *Proceedings of the National Academy of Science USA*, 81: 8014–8018.

- Sanwen, H., Baoxi, Z., Milbourne, D., Cardle, L., Guimei, Y. & Jiazhen, G. (2001). Development of pepper SSR markers from sequence databases. *Euphytica*, 117(2): 163-167.
- Sarkar, N.K., Kundnani, P. & Grover, A. (2013). Functional analysis of Hsp70 superfamily proteins of rice (*Oryza sativa*). *Cell Stress and Chaperones*, 18(4): 427–437.
- Schmider, E., Ziegler, M., Danay, E., Beyer, L. & Bühner, M. (2010). Is it really robust?. *European Journal of Research Methods for the Behavioral and Social Sciences*. pp 6.
- Schramm, F., Ganguli, A., Kiehlmann, E., English, G., Walch, D. & von Koskull-Doring, P. (2006). The heat stress transcription factor HsfA2 serves as a regulatory amplifier of a subset of genes in the heat stress response in *Arabidopsis*. *Plant Molecular Biology*, 60: 759–772.
- Semagn, K., Bjørnstad, Å. & Ndjiondjop, M. N. (2006). Progress and prospects of marker assisted backcrossing as a tool in crop breeding programs. *African Journal of Biotechnology*, 5(25): 2588-2603
- Sikora, B. & Nowaczyk, P. (2014). Application of RAPD technique for identification of interspecific hybrids from genus *Capsicum*. *Acta Scientiarum Polonorum*, 13(1): 155-166.
- Silva, L.R., Azevedo, J., Pereira, M.J., Carro, L., Velazquez, E., Peix, A., & Andrade, P.B. (2014). Inoculation of the non-legume (*Capsicum annuum* L.) with *Rhizobium* strains. 1. Effect on bioactive compounds, antioxidant activity, and fruit ripeness. *Journal of agricultural and food chemistry*, 62(3): 557-564.
- Silva, L.R., Azevedo, J., Pereira, M.J., Valentão, P., & Andrade, P.B. (2013). Chemical assessment and antioxidant capacity of pepper (*Capsicum annuum* L.) seeds. *Food and chemical toxicology*, 53: 240-248.
- Singh, R.P., Prasad, P., Sunita, K., Giri, S. & Reddy, K.R. (2007). Influence of high temperature and breeding for heat tolerance in cotton: A review. *Advances in Agronomy*, 93:313-385.
- Singh, V.K., Singh, A., Singh, S.P., Ellur, R.K., Singh, D., Krihnan, S.G., Bhowmick, P.K., Nagarajan, M., Vinod, K.K., Singh, U.D., Mohapatra, T., Prabhu, K.V. & Singh, A.K. (2013). Marker assisted simultaneous but stepwise backcross breeding for pyramiding blast resistance genes Piz5 and Piz54 into an elite Basmati rice restorer line `PRR78` *Plant Breeding*, 132: 486-495
- Sharangi, A. B., & Acharya, S. K. (2018). Spices in India and Beyond: The Origin, History, Tradition and Culture. In *Indian Spices* (pp. 1-11). Springer, Cham
- Sharma, D. & Masison, D.C. (2009). Hsp70 structure, function, regulation and influence on yeast prions. *Protein and Peptide Letters*, 16(6): 571–581.

- Snider, J. L., Oosterhuis, D. M., Skulman, B. W., & Kawakami, E. M. (2009). Heat stress- induced limitations to reproductive success in *Gossypium hirsutum*. *Physiologia plantarum*, 137: 125-138.
- Stummel, J. & Bosland, P. (2007). *Ornamental pepper Capsicum annum*. Flower Breeding and Genetics. Springer, New York, (pp 561-599).
- Suhana, O., Melor, R., & Norfadzilah, A. F. (2014). Yield performance and stability of new promising chilli genotypes in different agro-ecological environments in Malaysia. In XXIX International Horticultural Congress on Horticulture: Sustaining Lives, Livelihoods and Landscapes (IHC2014): 1127 (pp. 485-490).
- Sundaram, R.M., Vishnupriya, M.R., Biradar, S.K., Laha, G.S., Reddy, G.A., ShobhaRani, N., Sarma, N.P. & Sonti, R.V. (2008). Marker-assisted introgression of bacterial blight resistance in Samba Mahsuri, an elite indica rice variety. *Euphytica*, 160: 411-422.
- Sung, D.Y. & Guy, C.L. (2003). Physiological and molecular assessment of altered expression of Hsc70-1 in Arabidopsis. Evidence for pleiotropic consequences. *Plant Physiology*, 132(2): 979-987.
- Supena, E., Suharsono, S., Jacobsen, E. & Custers, J. (2006). Successful development of a shed-microspore culture protocol for doubled haploid production in indonesian hot pepper (*Capsicum annum* L.). *Plant Cell Reports*, 25:1-10.
- Suwor, P., Sanitchon, J., Thummabenjapone, P., Kumar, S., & Techawongstien, S. (2017). Inheritance analysis of anthracnose resistance and marker-assisted selection in introgression populations of chili (*Capsicum annum* L.). *Scientia Horticulturae*, 220: 20-26.
- Tanweer, F.A., Rafii, M.Y., Sijam, K., Rahim, H.A., Ahmed, F., Ashkani, S. & Latif, M.A. (2015). Identification of suitable segregating SSR markers for blast resistance in rice using inheritance and disease reaction analysis in backcross families. *Australasian Plant Pathology*, 44(6): 619-627.
- Thabuis, A., Palloix, A., Servin, B., Daubeze, A. M., Signoret, P. & Lefebvre, V. (2004). Marker-assisted introgression of 4 *Phytophthora capsici* resistance QTL alleles into a bell pepper line: validation of additive and epistatic effects. *Molecular Breeding*, 14(1): 9-20.
- Thakur, P., Kumar, S., Malik, J. A., Berger, J. D. & Nayyar, H. (2010). Cold stress effects on reproductive development in grain crops: An overview. *Environmental and Experimental Botany*, 67: 429-443.
- Thomas Bjorkman, (2011). Breeding Vegetables Adapted to High Temperatures: A Case Study with Broccoli. *Hortscience*, 46(8): 1093-1097.
- Tian, J., Belanger, F.C., & Huang, B. (2009). Identification of heat stress-responsive genes in heat-adapted thermal *Agrostis scabra* by suppression subtractive hybridization. *Journal of plant physiology*, 166(6): 588-601.

- Tompa, P. & Kovacs, D. (2010). Intrinsically disordered chaperones in plants and animals This paper is one of a selection of papers published in this special issue entitled “Canadian Society of Biochemistry, Molecular & Cellular Biology 52nd Annual Meeting—Protein Folding: Principles and Diseases” and has undergone the Journal's usual peer review process. *Biochemistry and Cell Biology*, 88(2): 167-174.
- Tilahun, S., Paramaguru, P. & Babu, J.K. (2013). Genetic Diversity in Certain Genotypes of Chilli and Paprika as Revealed by RAPD and SSR Analysis. *Asian Journal of Agricultural Sciences*, 5(2): 25-31
- Tran L. & Thuy M. K. (2015). Effect of High Temperature on Fruit Productivity and Seed-Set of Sweet Pepper (*Capsicum annuum* L.) in the Field Condition. *Journal of Agricultural Science and Technology A and B*, 5(12): 515-520.
- Usman, M.G., Rafii, M.Y., Ismail, M.R., Malek, M.A. & Latif, M.A. (2014a). Capsaicin and Dihydrocapsaicin Determination in Chili Pepper Genotypes Using Ultra-Fast Liquid Chromatography. *Molecules*, 19(5):6474-6488.
- Usman, M.G., Rafii, M.Y., Ismail, M.R., Malek, M.A. & Abdul Latif, M. (2014b). Heritability and genetic advance among chili pepper genotypes for heat tolerance and morphophysiological characteristics. *Scientific World Journal* Article 308042. DOI: 10.1155/2014/308042.
- Usman, M.G., Rafii, M.Y., Ismail, M.R., Malek, M.A. & Latif, M.A. (2015). Expression of Target Gene *Hsp70* and Membrane Stability determine Heat Tolerance in Chili Pepper. *Journal of the American Society for Horticultural Science*, 140(2):144–150.
- Van Berloo, R. (2008). “GGT 2.0: Versatile Software for Visualization and Analysis of Genetic Data,” *Journal of Heredity*, 99(2): 232-236. [doi:10.1093/jhered/esm109](https://doi.org/10.1093/jhered/esm109)
- Vollenweider, P., & Günthardt-Goerg, M. S. (2005). Diagnosis of abiotic and biotic stress factors using the visible symptoms in foliage. *Environmental Pollution*, 137(3): 455-465.
- Wahid, A., Gelani, S., Ashraf, M. & Foolad, M. (2007). Heat tolerance in plants: An overview. *Environmental and Experimental Botany*, 61:199-223.
- Wan, H., Yuan, W., Ruan, M., Ye, Q., Wang, R., Li, Z., & Liu, S. (2011). Identification of reference genes for reverse transcription quantitative real-time PCR normalization in pepper (*Capsicum annuum* L.). *Biochemical and Biophysical Research Communications*, 416: 24-30.
- Wang, D. & Bosland, P.W. (2006). The genes of capsicum. *Hortscience*, 41:1169-1187.
- Wang, Y., Lin, S., Song, Q., Li, K., Tao, H., Huang, J., Chen, X., Que, S. & He, H. (2014). Genome-wide identification of heat shock proteins (Hsps) and Hsp interactors in rice: Hsp70s as a case study. *BMC Genomics*, 15(1): 344

- Wang, W., Vinocur, B., Shoseyov, O. & Altman, A. (2004). Role of plant heat-shock proteins and molecular chaperones in the abiotic stress response. *Trends in Plant Science*, 9: 244-252.
- Wang, Z., Weber, J.L., Zhong, G. & Tanksley, S.D. (1994). Survey of plant short tandem DNA repeats. *Theoretical and Applied Genetics*, 88(1): 1-6.
- Wehmeyer N. & Vierling E (2000). The expression of small heat shock proteins in seeds responds to discrete developmental signals and suggests a general protective role in desiccation tolerance. *Plant Physiology*, 122: 1099–1108
- Willits, D. & Peet, M. (1998). The effect of night temperature on greenhouse grown tomato yields in warm climates. *Agricultural and Forest Meteorology*, 92: 191-202.
- Wilson, H.P., Kuhar, T.P., Rideout, S.L., Freeman, J.H., Reiter, M.S., Straw, R.A. & Aigner, J.D. (2012). Commercial vegetable production Recommendations-Virginia. *Virginia Cooperative Extension, Blacksburg, VA. Pub. (Louisiana Cooperative Extension Service) (USA)*.
- Ye, G. & Smith, K.F. (2010). Marker assisted gene pyramiding for inbred line development: Basic principles and practical guidelines. *International Journal of Plant Breeding*, 2: 1-10.
- Ye, S., Yu, S., Shu, L., Wu, J., Wu, A. & Luo, L. (2012). Expression profile analysis of 9 heat shock protein genes throughout the life cycle and under abiotic stress in rice. *Chinese Science Bulletin*, 57: 336-343.
- Yi, G., Lee, J.M., Lee, S., Choi, D. & Kim, B.D. (2006). Exploitation of pepper EST–SSRs and an SSR-based linkage map. *Theoretical and Applied Genetics*, 114(1): 113-130.
- Young, J.C. (2010). Mechanisms of the Hsp70 chaperone system. *Biochemistry of Cellular Biology*, 88:291–300.
- Young, N.D. & Tanksley, S.D. (1989). Restriction fragment length polymorphism maps and the concept of graphical genotypes. *Theoretical and Applied Genetics*, 77: 95-101.
- Zinn, K.E., Tunc-Ozdemir, M., & Harper, J.F. (2010). Temperature stress and plant sexual reproduction: uncovering the weakest links. *Journal Experimental Botany*, 61: 1959–1968.
- Zou, J., Ailing, L., Xinbo, C., Xiaoyun, Z., Guofu, G., Wenfang, W. & Xianwen Z. (2009). Expression Analysis of Nine Rice Heat Shock Protein Genes Under Abiotic Stresses and ABA Treatment. *Journal of Plant Physiology*, 166(8):851-861.
- Zurbriggen, M.D., Hajirezaei, M.R. & Carrillo, N. (2010). Engineering the future. Development of transgenic plants with enhanced tolerance to adverse environments. *Biotechnology and Genetic Engineering Reviews*, 27:33–56.