



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF NATURALLY OCCURRING PHENOLIC COMPOUND ON
CELL
WALL DEGRADING ENZYMES AND SUPPRESSION OF *Ganoderma
boninense* INFECTION IN OIL PALM SEEDLINGS***

ARTHY SURENDRAN

IPTSM 2018 4



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By

ARTHY SURENDRAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
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Philosophy**

January 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

EFFECTS OF NATURALLY OCCURRING PHENOLIC COMPOUNDS ON CELL WALL DEGRADING ENZYMES AND THE SUPPRESSION OF *Ganoderma boninense* INFECTED OIL PALM SEEDLINGS

By

ARTHY SURENDRAN

January 2018

Chairman: Yasmeen Siddiqui, PhD

Faculty: Institute of Tropical Agriculture and Food Security

Palm oil is one of the major sources of edible oil in the world with 85% of it being produced by Malaysia and Indonesia. However, the production is greatly hindered by the basal stem rot (BSR) disease. The causal pathogen of BSR disease is *Ganoderma* sp. *Ganoderma boninense*, the causal pathogen of BSR in oil palm is white rot basidiomycetes. This pathogen infects oil palm primarily via roots by degrading the lignin and cellulose components. Therefore, understanding the mode of infection of *G. boninense* in oil palm would be advantageous. An alternative solution to control the emergence of BSR is to inhibit the lignolytic and the cellulolytic enzymes of *G. boninense*. The phenolic compounds present naturally in the plants play a critical role in the pathogen elimination, signaling, increasing the resistance and the lignin biosynthesis. Hence, ten naturally occurring phenolic compounds namely, benzoic acid, coumaric acid, 2,6-dimethoxyl benzoic acid, 2,6-dimethoxyl phenol, guaiacol, ferulic acid, pyrocatechol, salicylic acid, syringic acid, and vanillic acid were selected to evaluate their potential to inhibit *G. boninense*. In this study, the phenolic compounds were tested for their ability to inhibit the growth of *G. boninense* and their inhibitory effect towards the production of lignolytic and cellulolytic enzymes. Further, their efficacy was tested to suppress the BSR infection in oil palm seedlings. The ten selected phenolic compounds were able to inhibit the growth of *G. boninense* with different degrees depending on their concentrations. Microscopic observations revealed that mycelia growing on media containing phenolic compounds showed deterioration. A significant ($p \leq 0.05$) decrease in the production of lignolytic enzymes, as well as cellulase, amylase and xylanase of about 40-100% was identified. Except benzoic acid all the other phenolic compounds increased the secretion of lignolytic and cellulolytic enzymes at 1 mM concentration. However, as the concentrations increased more than 1 mM the inhibition

increased. The enzyme inhibitions have been further quantified and the type of inhibition was analysed along with their physicochemical properties. Most of the selected phenolic compounds inhibited the enzymes as uncompetitive or noncompetitive inhibitors. The lignolytic enzymes were active over a wide range of temperature from 40-80°C but sensitive to pH at 6. The cellulolytic enzymes are more stable in a wide range of pH from 3 to 8 and temperature 40-80°C when compared to the lignolytic enzymes. The wood degradation assay suggested that the *G. boninense* is a sequential degrader of lignin and cellulose components. The selected phenolic compounds significantly reduced the degradation rate upto 100% when compared to the control. This was due to their ability to inhibit the lignolytic and cellulolytic enzymes of *G. boninense*. Significant reductions in the disease progression upto 100% were observed in the oil palm seedlings treated with benzoic and salicylic acid. The oil palm seedling treated with benzoic and salicylic acid increased the growth parameters such as height, diameter of the stem, chlorophyll content, root and shoot weight. In addition, the oil palm seedlings treated with phenolic compounds showed increased lignification of four percentages. This was due to the increased activity of phenylalanine amino- lyases, peroxidase and polyphenol oxidase by 1.5 folds. These enzymes are known to be involved in lignin biosynthesis pathway. The benzoic acid and salicylic acid tested are the processors for the synthesis of lignin. The findings of this study could be useful for developing new strategies in controlling the spread of disease, which may reduce the BSR disease severity in oil palm areas of production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN-KESAN SEBATIAN SEMULAJADI FENOLIK PADA ENZIM YANG
MENGURAIKAN DINDING SEL DAN PENINDASAN TERHADAP
JANGKITAN *Ganoderma boninense* DALAM BENIH KELAPA SAWIT**

Oleh

ARTHY SURENDRAN

Januari 2018

Pengerusi: Yasmeen Siddiqui, PhD

Fakulti: Institut Pertanian Tropika Dan Sekuriti Makanan

Minyak sawit ialah salah satu sumber utama minyak makan di dunia dan 85% di antaranya dihasilkan oleh Malaysia dan Indonesia. Walau bagaimanapun, pengeluarannya amat terjejas akibat penyakit reput pangkal batang (BSR) sawit yang disebabkan oleh *Ganoderma* sp. *Ganoderma boninense* adalah patogen penyebab penyakit BSR dalam sawit dan ia merupakan sejenis basidiomycetes reput putih. Patogen ini menjangkiti sawit terutamanya melalui akar dengan menguraikan komponen lignin dan selulosa. Oleh kerana itu, memahami mod jangkitan *G. boninense* dalam kelapa sawit adalah agak penting. Antara penyelesaian alternatif untuk mengawal emergem BSR adalah dengan menyekat enzim lignolitik dan selulolisis *G. boninense*. Sebatian fenol hadir secara semulajadi dalam tumbuhan memainkan peranan kritikal dalam penyingkiran patogen, pengisyratan, menambahkan rintangan dan juga lignin biosintesis. Maka sepuluh sebatian fenol yang berlaku secara semulajadi iaitu asid benzoik, asid kumarik, asid benzoik 2,6-dimethoxyl, fenol 2,6-dimethoxyl, guaiakol, asid ferulik, pirokatekol, asid salisilik, asid syringik dan asid vanillik telah terpilih untuk menilai kecekapan mereka dalam mengawal *G. boninense*. Dalam kajian ini, sebatian fenol diuji untuk kemampuan mereka menghalang pertumbuhan *G. boninense* dan kesan bertentangan mereka ke arah pengeluaran lignolitik dan enzim selulolisis. Tambahan, keberkesanan mereka dalam menghalang jangkitan BSR di dalam anak benih sawit. Sepuluh sebatian fenol terpilih mampu menyekat pertumbuhan *G. boninense* dengan efikasi berbeza bergantung pada kepekatan mereka. Pemerhatian-pemerhatian mikroskopik mendedahkan bahawa mycelia tumbuh di atas media mengandungi sebatian fenol menunjukkan kemerosotan. Satu penurunan penting ($p \leq 0.05$) dalam pengeluaran enzim-enzim lignolitik, serta selulosa, amilase dan xilanase telah dikenal pasti. Kecuali asid benzoik semua sebatian fenol yang lain menambah rembesan lignolitik dan enzim selulolisis di

kepekatan 1 mM. Bagaimanapun, apabila kepekatan menambah mereka bertindak sebagai satu perencat. Perencatan enzim selanjutnya dijumlahkan dan jenis perencatan dianalisis bersama dengan sifat fisiko kimia mereka. Kebanyakan daripada sebatian fenol terpilih menghalang enzim sebagai perencat tiada tandingan atau tidak melibatkan persaingan. Enzim-enzim lignolitik aktif atas julat suhu 40-80°C luas tetapi sensitif kepada pH 6. Enzim selulolisis lebih stabil dalam julat pH yang lebih luas 3-8 dan suhu 40-80°C apabila dibandingkan dengan enzim-enzim lignolitik. Cerakinan degradasi kayu mencadangkan, *G. boninense* ialah degradasi berjujukan komponen lignin dan selulosa. Sebatian fenol terpilih mengurangkan kadar penguraian sehingga 100% apabila dibandingkan dengan kawalan. Ini adalah disebabkan kemampuan mereka menghalang lignolitik dan enzim selulolisis *G. boninense*. Pengurangan signifikan dalam kemaraan penyakit telah diperhatikan dalam anak-anak benih pokok kelapa sawit dirawat dengan asid benzoik dan salisilik. Anak benih sawit dirawat dengan asid benzoik dan salisilik menambah parameter pertumbuhan seperti ketinggian, garis pusat batang, kandungan klorofil, berat akar dan pucuk. Selain parameter pertumbuhan, anak-anak benih pokok kelapa sawit dirawat dengan sebatian fenol menunjukkan pengligninan bertambah. Peningkatan dalam pengligninan adalah disebabkan aktiviti phenylalanine amino-lyases, peroxidase dan polyphenol oxidase yang meningkat dengan 1.5 kali ganda. Enzim-enzim ini terkenal dalam penglibatan laluan biosintesis lignin. Asid benzoik dan salisilik yang diuji ialah pemproses untuk sintesis lignin. Penemuan kajian ini boleh digunakan untuk membangunkan strategi baru dalam mengawal penyebaran penyakit yang boleh mengurangkan penyakit BSR dalam estet kelapa sawit.

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I certify that a Thesis Examination Committee has met on 11 January 2018 to conduct the final examination of Arthy Surendran on her thesis entitled "Effect of Naturally Occurring Phenolic Compounds on Cell Wall Degrading Enzymes and Suppression of *Ganoderma boninense*" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Mui-Yun Wong, PhD

Associate Professor
Faculty of Agriculture
University Putra Malaysia
(Chairman)

Ganesan Vadamalai, PhD

Associate Professor
Faculty of Agriculture
University Putra Malaysia
(Internal Examiner)

Kamaruzaman Sijam, PhD

Associate Professor
Faculty of Agriculture
University Putra Malaysia
(Internal Examiner)

Silvia Bautista Baños, PhD

Professor
Postharvest Technology
Instituto Politécnico Nacional Mexico
(External Examiner)

NOR AINI AB. SHUKOR, PhD

Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia
Date

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Yasmeen Siddiqui, PhD

Senior lecturer
Institute of Tropical Agriculture and Food security
University Putra Malaysia
(Chairman)

Halimi Bin Mohd Saud, PhD

Associate Professor
Faculty of Agriculture
University Putra Malaysia
(Member)

Nusaibah Syd Ali, PhD

Senior lecturer Faculty of Agriculture
University Putra Malaysia
(Member)

Sivakumar Manickam, PhD

Professor
Faculty of engineering
University of Nottingham
(Member)

ROBIAH BINTI YUNUS, PHD

Professor and Dean
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Universiti Putra, Malaysia
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Signature: _____

Name of Chairman of
Supervisory
Committee:

Dr. Yasmeen Siddiqui

Signature: _____

Name of Member of
Supervisory
Committee:

Assoc. Prof. Dr. Halimi Bin Mohd Saud

Signature: _____

Name of Member of
Supervisory
Committee:

Dr. Nusaibah Syd Ali

Signature: _____

Name of Member of
Supervisory
Committee:

Prof. Dr. Sivakumar Manickam

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scale of 0-4.



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LIST OF ABBREVIATIONS

Ø	Diameter
%	Percentage
ABTS	2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonicacid) diammonium salt
AIL	Acid insoluble lignin
ANOVA	Analysis of variance
ASL	Acid soluble lignin
AUDPC	Area under disease progress curve
BSR	Basal stem rot
Cfu	Colony forming unit
Cm	Centimeter
CMC	Carboxymethyl cellulose
DR	Disease reduction
DS	Disease severity
EDTA	Ethylenediaminetetraacetic acid
FT-IR	Fourier transform infrared spectroscopy
g	Gram
h	Hours
K _m	Substrate concentration
L	liter
LB	Luria-Bertani broth
M	Molar
MAI	Moth after inoculation
MEA	Malt extract agar
mg	Milligram
Min	minutes
mm	Millimeter
mM	millimolar
MPOB	Malaysian Palm Oil Board
OD	Optical density
PAL	Phenyl ammonia lyase
PDA	Potato Dextrose Agar
PDB	Potato Dextrose Broth
PIRG	Percentage Inhibition of Radial Growth
POD	peroxidase
PPO	polyphenol oxidase
RBBR	Remazol Brilliant Blue R
RPM	Revolution per minute
SEM	Scanning electron microscope
SDS	Sodium dodecyl sulfate
TGA	Thermogravimetry analysis
USR	Upper stem rot
V _{max}	Velocity of the reaction
w ⁻¹	Volume per volume
wv ⁻¹	Weight per volume

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CHAPTER 1

INTRODUCTION

1.1 Background

Oil palm (*Elaeis guineensis* Jacq.) is an important cash crop of Malaysia, which bolsters the economy of the country. Oil palm is a native crop to Africa and it was introduced to Malaysia by the British in 1870 as an ornamental plant (Corley and Teo, 1976). It has been first commercially planted at Selangor state in 1970 (Hartley, 1967). Palm oil is gaining fast recognition as it contributes 30% of world's edible oil demand. The oil palm input to output outpaces other oil bearing crops such as soybean, rapeseed and sunflower (MPOC 2015). The palm oil contributes about 8% of total income of Malaysia and hence it is called as the "golden crop" of Malaysia.

The statistics from the past indicates that the strongest period of growth was in 1989-2008. During this period, the production was raised by 4% every year. In the last decade, an unexpected break in the growth rate was observed with 2% decremented yield loss annually (MPOB, 2015), due to various factors such as adverse climatic condition, labour cost, tree age, low yielding crop variety and diseases. Hence, Malaysia is compelled to increase the production of palm oil to meet the world's oil demand. The improvement in palm oil is not only governed by implementing new plantation, using high yield varieties and replanting the old trees with new ones or by increasing the total plantation area but also a more efficient pest and disease control measures to fill the production gap (Jalani et al., 2002).

Many pests and diseases from the seed hinder the growth of oil palm. Some of the oil palm pests and diseases are brown germ, bagworm, upper stem rot disease (USR) and basal stem rot disease (BSR). However, the palm oil production is majorly affected by the BSR disease.

1.2 Problem statement

The BSR disease in oil palm is caused by a white rot basidiomycete fungus *Ganoderma boninense* and is considered as the most virulent strain (Rees et al., 2009). Basal stem rot disease was first identified by Thomson in 1931. This disease causes serious economic impact on Malaysia's oil palm industry today. The disease incidence increases/accumulates over the successive planting cycles (Susanto et al., 2005). The high incidence of BSR results in the drastic reduction in weight and number of bunches produced, they can reach at times to zero percentage yield (Chong et al., 2012). Initially, the *Ganoderma* reported

to infect only aged plants but in the last decade infection was observed even in seedlings less than six months old (Rees et al., 2007). The most accepted mode of infection is considered when there is a contact of infected roots with healthy oil palm roots (Rees et al., 2009; Singh et al., 1991). Another mode of infection is by the dispersal of basidiospore (Paterson, 2007).

To date, the status of disease management remains the same, since the method to address the disease effectively is scanty. Various methods have been introduced to the industry to prolong the life span of the tree and to increase the yield of the infected oil palm. These include soil mounding, trunk injection with fungicide, clean and clear practices. So far the hexaconazol applied via trunk injection is effective in controlling the BSR disease in oil palm to up to 70% (Idris et al., 2004 a). However the chemical control method is found to be cost forbidding.

The salient feature of white rot fungus (WRF) is the complete degradation of lignin and cellulose components in the plant. White rot fungus spends energy to degrade lignin in order to get access for itself in cellulose components. The degradation of lignin is to be the rate limiting step in the infection process (Paterson et al., 2009). Nevertheless, the pattern of lignin degradation of *Ganoderma* has not been explored till date. Two types of patterns were observed in the WRF degradation, the first one is the degradation of lignin followed by cellulose whereas second type is the concurrent degradation of both lignin and cellulose (Hatakka, 1994). To degrade the cellular structure WRF employs lignolytic and cellulolytic enzymes. Hence, the lignocellulolytic enzymes may also play an important role in the BSR disease progression.

Therefore, it is desirable to find a biomolecule that could inhibit the lignolytic and the cellulolytic enzymes of *Ganoderma boninense* to control the BSR disease in oil palm. The biomolecule should be of small in size so that it can travel towards the site of infection and is resistant to auto-oxidation. If this compound can induce resistance and there by leverage the oil palm seedlings over the BSR disease.

Phenolic compounds are the compounds which can address the above demand. The phenolic compounds are naturally present in plants at normal conditions. It gets elevated during biotic and abiotic stresses. Although, these compounds are involved in the signal transduction pathway in the stress mode, they primarily function in production lignin and facilitate cell wall synthesis. They are tightly bound to the lignin structure and play an important role in protecting and strengthening the cell wall structure (Hammerschmidt, 2005).

1.3 Hypothesis

Inhibition of lignocellulolytic enzymes (the key enzymes in the infection process) utilizing phenolic compounds could be an effective method to control the BSR disease of oil palm.

1.4 Research objectives

The general objective is to examine the effect of phenolic compounds in inhibiting BSR disease in oil palm. The specific objectives of the study are as follow:

1. To determine antagonistic effect of phenolic compounds on the growth and secretion of lignolytic and cellulolytic enzymes of *G. boninense*.
2. To characterize and determine the inhibition of lignolytic and cellulolytic enzyme secretion by *G. boninense* utilizing naturally occurring phenolic compounds.
3. To characterize and assess the pattern of oil palm wood degradation by *G. boninense* in the presence of phenolic compounds.
4. To determine the effect of benzoic and salicylic acid in the immunization of oil palm seedlings- challenged by *G. boninense*.

REFERENCES

- Abu Seman, I.b. (1999). Basal stem rot (BSR) of oil palm (*Elaeis guineensis* Jacq) in Malaysia factors associated with variation in disease severity. *Imperial college London* (University of London).
- Adaskaveg, J. E., Gilbertson, R. L., and Blanchette, R. A. (1990). Comparative studies of delignification caused by *Ganoderma* species. *Applied and environmental microbiology*, 56(6):1932-1943.
- Ahmad, H., and Salmah, J. (1998, May). Palm oil as diesel fuel: field trial on cars with Elsbett engine. In *Proceeding of the 1998 PORIM international biofuel and lubricant conference*, PORIM, Bangi 165-174.
- Ahn, M. Y., Zimmerman, A. R., Martínez, C. E., Archibald, D. D., Bollag, J. M., and Dec, J. (2007). Characteristics of *Trametes villosa* laccase adsorbed on aluminium hydroxide. *Enzyme and Microbial Technology*, 41(1):141-148.
- Aist, J. R., Gold, R. E., Bayles, C. J., Morrison, G. H., Chandra, S., and Israel, H. W. (1988). Evidence that molecular components of papillae may be involved in ml-o resistance to barley powdery mildew. *Physiological and molecular plant pathology*, 33(1):17-32.
- Akhtar, M., and Malik, A. (2000). Roles of organic soil amendments and soil organisms in the biological control of plant-parasitic nematodes: a review. *Bioresource Technology*, 74(1):35-47.
- Alfredsen, G., Bader, T. K., Dibdiakova, J., Filbakk, T., Bollmus, S., and Hofstetter, K. (2012). Thermogravimetric analysis for wood decay characterisation. *European Journal of Wood and Wood Products*, 70(4):527-530.
- Amborabé, B. E., Fleurat-Lessard, P., Chollet, J. F., and Roblin, G. (2002). Antifungal effects of salicylic acid and other benzoic acid derivatives towards *Eutypa lata*: structure–activity relationship. *Plant Physiology and Biochemistry*, 40(12):1051-1060.
- Ander, P., Hatakka, A., and Eriksson, K. E. (1980). Vanillic acid metabolism by the white-rot fungus *Sporotrichum pulverulentum*. *Archives of Microbiology*, 125(3):189-202.
- Andrews, J.H., Harris, R.F., 2000. The ecology and biogeography of microorganisms on plant surfaces. *Annual review of phytopathology* 38:145-180.
- Anterola, A. M., and Lewis, N. G. (2002). Trends in lignin modification: a comprehensive analysis of the effects of genetic manipulations/mutations on lignification and vascular integrity. *Phytochemistry*, 61(3): 221-294.

- Arif, M. S., Roslan, A., and Idris, A. S. (2011). Economics of OP pests and *Ganoderma* disease and yield losses. In *Proceedings of the Third MPOB-IOPRI International Seminar: Integrated OP Pests and Diseases Management*
- Ariffin, D., Idris, A. S., and Singh, G. (2000). Status of *Ganoderma* in oil palm. *Ganoderma diseases of perennial crops*, (200):49-68.
- Arora, D. S., and Gill, P. K. (2001). Comparison of two assay procedures for lignin peroxidase. *Enzyme and microbial technology*, 28(7):602-605.
- Atalla, M. M., Zeinab, H. K., Eman, R. H., Amani, A. Y., and Abeer, A. A. E. A. (2013). Characterization and kinetic properties of the purified *Trematosphaeria mangrovei* laccase enzyme. *Saudi journal of biological sciences*, 20(4):373-381.
- Azadeh, B. F., Sariah, M., and Wong, M. Y. (2010). Characterization of *Burkholderia cepacia* genomovar I as a potential biocontrol agent of *Ganoderma boninense* in oil palm. *African Journal of Biotechnology*, 9(24): 3542-3548.
- Bagga, P. S., Sandhu, D. K., and Sharma, S. (1990). Purification and characterization of cellulolytic enzymes produced by *Aspergillus nidulans*. *Journal of Applied Microbiology*, 68(1): 61-68.
- Baldrian, P. (2006). Fungal laccases—occurrence and properties. *FEMS microbiology reviews*, 30(2):215-242.
- Barber, M. S., and Mitchell, H. J. (1997). Regulation of phenylpropanoid metabolism in relation to lignin biosynthesis in plants. *International review of cytology*, 172:243-293.
- Bari, E., Nazarnezhad, N., Kazemi, S. M., Ghanbary, M. A. T., Mohebbi, B., Schmidt, O., and Clausen, C. A. (2015). Comparison between degradation capabilities of the white rot fungi *Pleurotus ostreatus* and *Trametes versicolor* in beech wood. *International Biodeterioration & Biodegradation*, 104:231-237.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology*, 109(4):289-295.
- Bhuiyan, N. H., Selvaraj, G., Wei, Y., and King, J. (2008). Gene expression profiling and silencing reveal that monolignol biosynthesis plays a critical role in penetration defence in wheat against powdery mildew invasion. *Journal of Experimental Botany*, 60(2): 509-521.
- Bilal, M., Asgher, M., and Ramzan, M. (2015). Purification and biochemical characterization of extracellular manganese peroxidase from

Ganoderma lucidum IBL-05 and its application. *Scientific research and Essays*, 10(14):456-464.

Blanchette, R. A. (2000). A review of microbial deterioration found in archaeological wood from different environments. *International Biodeterioration & Biodegradation*, 46(3): 189-204.

Boerjan, W., Ralph, J., and Baucher, M. (2003). Lignin biosynthesis. *Annual review of plant biology*, 54(1):519-546.

Bolwell, G. P., Robbins, M. P., & Dixon, R. A. (1985). Metabolic changes in elicitor-treated bean cells. *The FEBS Journal*, 148(3):571-578.

Bucher, V. V. C., Hyde, K. D., Pointing, S. B., and Reddy, C. A. (2004). Production of wood decay enzymes, mass loss and lignin solubilization in wood by marine ascomycetes and their anamorphs. *Fungal Diversity*, 5:1-14.

Buswell, J., Ander, P., Pettersson, B., and Eriksson, K.E., 1979. Oxidative decarboxylation of vanillic acid by *Sporotrichum pulverulentum*. *FEBS letters* 103: 98-101.

Campbell, W. H. (1999). Nitrate reductase structure, function and regulation: bridging the gap between biochemistry and physiology. *Annual review of plant biology*, 50(1):277-303.

Carrier, M., Loppinet-Serani, A., Denux, D., Lasnier, J. M., Ham-Pichavant, F., Cansell, F., and Aymonier, C. (2011). Thermogravimetric analysis as a new method to determine the lignocellulosic composition of biomass. *Biomass and Bioenergy*, 35(1):298-307.

Carver, T. L. W., Robbins, M. P., Thomas, B. J., Troth, K., Raistrick, N., and Zeyen, R. J. (1998). Silicon deprivation enhances localized autofluorescent responses and phenylalanine ammonia-lyase activity in oat attacked by *Blumeria graminis*. *Physiological and Molecular Plant Pathology*, 52(4):245-257.

Chan Cupul, W., Heredia Abarca, G., RodriguezVazquez, R., Salmenes, D., Gaitan, Hernandez, R., and Alarcon Gutierrez, E., (2014). Repuesta de macrohongos lignoliticos al herbicida: bioensayos, dosis-resputa. *Revista argentina de microbiologia*46:348-357.

Chappell, J., and Hahlbrock, K. (1984). Transcription of plant defence genes in response to UV light or fungal elicitor. *Nature*, 311(5981):76-78.

ChengTuck, H., and Hashim, K. (1997). Usefulness of soil mounding treatments in prolonging productivity of prime-aged *Ganoderma* infected palms. *Planter*, (73): 239-244.

- Chong, J., Pierrel, M. A., Atanassova, R., Werck-Reichhart, D., Fritig, B., and Saindrenan, P. (2001). Free and conjugated benzoic acid in tobacco plants and cell cultures. Induced accumulation upon elicitation of defense responses and role as salicylic acid precursors. *Plant Physiology*, 125(1):318-328.
- Chong, K. P., Markus, A., and Rossall, S. (2012). The susceptibility of different varieties of oil palm seedlings to *Ganoderma boninense* infection. *Pak. Journal of Botany*, 44(6): 2001-2004.
- Chong, K. P., Rossall, S., and Atong, M. (2009). In vitro antimicrobial activity and fungitoxicity of syringic acid, caffeic acid and 4-hydroxybenzoic acid against *Ganoderma boninense*. *Journal of Agricultural Science*, 1(2):15-21.
- Choo, Y. M., Yap, S. C., Ooi, C. K., Ma, A. N., Goh, S. H., and Ong, A. S. H. (1996). Recovered oil from palm-pressed fiber: a good source of natural carotenoids, vitamin E, and sterols. *Journal of the American Oil Chemists' Society*, 73(5):599-602.
- Choquer, M., Fournier, E., Kunz, C., Levis, C., Pradier, jm., Simon, A., and Viaud, M. (2007). *Botrytis cinerea* virulence factors: new insights into a necrotropic and polypathogen. *FEMS Microbiology Letters*, 277, 1-10.
- Collins, P. J., and Dobson, A. (1997). Regulation of laccase gene transcription in *Trametes versicolor*. *Applied and Environmental Microbiology*, 63(9):3444-3450.
- Corley R.H.V and Teo C. (1976). Disbudding of mature oil palm as a method of controlling yield fluctuation. *MARDI Research Bulletin*, 4: 1-6.
- Carver, T. L. W., Robbins, M. P., Thomas, B. J., Troth, K., Raistrick, N., and Zeyen, R. J. (1998). Silicon deprivation enhances localized autofluorescent responses and phenylalanine ammonia-lyase activity in oat attacked by *Blumeria graminis*. *Physiological and Molecular Plant Pathology*, 52(4):245-257.
- Cragg, S. M., Beckham, G. T., Bruce, N. C., Bugg, T. D., Distel, D. L., Dupree, P., and McGeehan, J. E. (2015). Lignocellulose degradation mechanisms across the Tree of Life. *Current Opinion in Chemical Biology*, 29:108-119
- Darus, A., Seman, I.A., and Hassan, A.H., (1989). Significance of the black line within oil palm tissue decayed by *Ganoderma boninense*. *Elaeis* 1:11-16.
- Dashtban, M., Schraft, H., Syed, T. A., and Qin, W. (2010). Fungal biodegradation and enzymatic modification of lignin. *International journal of biochemistry and molecular biology*, 1(1):36-50.

- Dat, J. F., Lopez-Delgado, H., Foyer, C. H., and Scott, I. M. (1998). Parallel changes in H₂O₂ and catalase during thermotolerance induced by salicylic acid or heat acclimation in mustard seedlings. *Plant Physiology*, 116(4):1351-1357.
- Dayou, J., Alexander, A., Sipaut, C. S., Chong, K. P., and Lee, P. C. (2014). On the possibility of using FTIR for detection of *Ganoderma boninense* in infected oil palm tree. *International Journal of Advances in Agricultural and Environmental Engineering*, 1(1):161-163.
- De Ascensao, A. R., and Dubery, I. A. (2000). Panama disease: cell wall reinforcement in banana roots in response to elicitors from *Fusarium oxysporum* f. sp. cubense race four. *Phytopathology*, 90(10):1173-1180.
- De Jong, E. D., Field, J. A., and de Bont, J. A. (1994). Aryl alcohols in the physiology of ligninolytic fungi. *FEMS Microbiology Reviews*, 13(2-3):153-187.
- Dekker, R. F., Barbosa, A. M., and Sargent, K. (2002). The effect of lignin-related compounds on the growth and production of laccases by the ascomycete, *Botryosphaeria* sp. *Enzyme and microbial technology*, 30(3):374-380.
- Delmer, D. P. (1999). Cellulose biosynthesis: exciting times for a difficult field of study. *Annual review of plant biology*, 50(1):245-276.
- Dewick, P. M. (1995). The biosynthesis of shikimate metabolites. *Natural product reports*, 12(2):101-133.
- Dharmaputra, O.S., Tjitrosomo, H., and Abadi, A., (1989). Antagonistic effect of four fungal isolates to *Ganoderma boninense*, the causal agent of basal stem rot of oil palm. *Biotropia*, (3):41-49.
- Dixon, R. A., and Paiva, N. L. (1995). Stress-induced phenylpropanoid metabolism. *The plant cell*, 7(7):1085-1097.
- Do, T. T., Quyen, D. T., and Dam, T. H. (2012). Purification and characterization of an acid-stable and organic solvent-tolerant xylanase from *Aspergillus awamori* VTCC-F312. *ScienceAsia*, 38(2012):157-165.
- Ducros, V., Brzozowski, A. M., Wilson, K. S., Brown, S. H., Østergaard, P., Schneider, P., and Davies, G. J. (1998). Crystal structure of the type-2 Cu depleted laccase from *Coprinus cinereus* at 2.2 Å resolution. *Nature Structural & Molecular Biology*, 5(4):310-316.
- Durrand-Gasselien, T., Asmady, H., Flori, A., Jacquemard, J., Breton, f., and De Franqueville, H. (2005). Possible sources of genetic resistance in oil

palm (*Elaeis guineensis* Jacq.) to basal stem rot caused by *Ganoderma boninense* –prospects for future breeding. *Mycopathologia*, 159: 93-100.

- Dutta, T., Sahoo, R., Sengupta, R., Ray, S. S., Bhattacharjee, A., and Ghosh, S. (2008). Novel cellulases from an extremophilic filamentous fungi *Penicillium citrinum*: production and characterization. *Journal of industrial microbiology & biotechnology*, 35(4):275-282.
- Elisashvili, V., Kachlishvili, E., and Penninckx, M. (2008). Effect of growth substrate, method of fermentation, and nitrogen source on lignocellulose-degrading enzymes production by white-rot basidiomycetes. *Journal of industrial microbiology & biotechnology*, 35(11):1531-1538.
- Enyedi, A. J., Yalpani, N., Silverman, P., and Raskin, I. (1992). Localization, conjugation, and function of salicylic acid in tobacco during the hypersensitive reaction to tobacco mosaic virus. *Proceedings of the National Academy of Sciences*, 89(6):2480-2484.
- Eriksson, K. E., Gupta, J. K., Nishida, A., and Rao, M. (1984). Syringic acid metabolism by some white-rot, soft-rot and brown-rot fungi. *Microbiology*, 130(10): 2457-2464.
- Eriksson, K.-E.L., Blanchette, R., and Ander, P., 2012. Microbial and enzymatic degradation of wood and wood components. Springer Science & Business Media.
- Fariduddin, Q., Hayat, S., and Ahmad, A. (2003). Salicylic acid influences net photosynthetic rate, carboxylation efficiency, nitrate reductase activity and seed yield in *Brassica juncea*. *Phytosynthetica*.41:281-284.
- Farmer, E. E. (1985). Effects of fungal elicitor on lignin biosynthesis in cell suspension cultures of soybean. *Plant Physiology*, 78(2), 338-342.
- Ferrari, S., Plotnikova, J. M., De Lorenzo, G., and Ausubel, F. M. (2003). Arabidopsis local resistance to *Botrytis cinerea* involves salicylic acid and camalexin and requires EDS4 and PAD2, but not SID2, EDS5 or PAD4. *The Plant Journal*, 35(2):193-205.
- Flood, J., and Hasan, Y. (2004). Basal stem rot-taxonomy, biology, epidemiology, economic status and control in South East Asia and Pacific Islands. Malaysian Palm Oil Board. (5):117-113.
- Flood, J., Hasan, Y., and Foster, H. (2002). *Ganoderma* diseases of oil palm-an interpretation from Bah Lias Research Station. *Planter*, 78(921):689-710.
- Forootanfar, H., Faramarzi, M. A., Shahverdi, A. R., and Yazdi, M. T. (2011). Purification and biochemical characterization of extracellular laccase

- from the ascomycete *Paraconiothyrium variabile*. *Bioresource Technology*, 102(2):1808-1814.
- GaitFee, C., (2011). Management of Ganoderma diseases in oil palm plantations. *Planter*, 87:325-339.
- Giatti Marques De Souza, C., Kirst Tychanowicz, G., Farani De Souza, D., and Peralta, R. M. (2004). Production of laccase isoforms by *Pleurotus pulmonarius* in response to presence of phenolic and aromatic compounds. *Journal of Basic Microbiology*, 44(2):129-136.
- Grant, M. R., and Jones, J. D. (2009). Hormone (dis) harmony moulds plant health and disease. *Science*, 324(5928):750-752.
- Gonçalves, M. L. F., and Steiner, W. (1996). Purification and characterization of laccase from a newly isolated wood-decaying fungus, 1996:258-266.
- Guillén, F., Martínez, M. J., Gutiérrez, A., and Del Rio, J. C. (2005). Biodegradation of lignocellulose: microbial, chemical, and enzymatic aspects of the fungal attack of lignin. *International Microbiology*, 8:195-204.
- HakWan, H., (2007). *Ganoderma* disease of oil palm in Sabah. *Planter*, 83 (974):299-313.
- Hammerschmidt, R. (2005). Phenols and plant-pathogen interactions: the saga continues. *Physiological and Molecular Plant Pathology*, 66(3):77-78.
- Hartley, C. W. S. (1967). The oil palm. *The oil palm. : Longmans, Green and Co. Ltd.* London
- Hashim, R., Nadhari, W. N. A. W., Sulaiman, O., Kawamura, F., Hiziroglu, S., Sato, M., and Tanaka, R. (2011). Characterization of raw materials and manufactured binderless particleboard from oil palm biomass. *Materials & Design*, 32(1):246-254.
- Hatakka, A. (1994). Lignin-modifying enzymes from selected white-rot fungi: production and role from in lignin degradation. *FEMS microbiology reviews*, 13(2-3):125-135.
- Hayat, Q., Hayat, S., Alyemeni, M. N., and Ahmad, A. (2012). Salicylic acid mediated changes in growth, photosynthesis, nitrogen metabolism and antioxidant defense system in *Cicer arietinum* L. *Plant Soil Environ*, 58(9):417-423.
- Hayat, Q., Hayat, S., Irfan, M., and Ahmad, A. (2010). Effect of exogenous salicylic acid under changing environment: a review. *Environmental and experimental botany*, 68(1):14-25.

- Heinzkill, M., Bech, L., Halkier, T., Schneider, P., and Anke, T. (1998). Characterization of laccases and peroxidases from wood-rotting fungi (family Coprinaceae). *Applied and Environmental Microbiology*, 64(5):1601-1606.
- Higuchi, T. (1990). Lignin biochemistry: biosynthesis and biodegradation. *Wood Science and Technology*, 24(1):23-63.
- Ho, Y. W., and Khairudin, H. (1995). Pathogenicity and histopathology of *Ganoderma boninense* on oil palm seedlings. *Journal of Bioscience(Penang)*, 6(2):155-164.
- Ho, Y. W., and Nawawi, A. (1986). Isolation, growth and sporophore development of *Ganoderma boninense* from oil palm in Malaysia. *Pertanika*, 9(1):69-73.
- Idris, A. S., Ariffin, H., and D Ismail, S. (2004 a). *Prolonging the productive life of Ganoderma-infected palms with Hexaconazole*. MPOB bulletin, 241:1-4.
- Idris, A., Kushairi, A., Ismail, S., and Ariffin, D. (2004b). Selection for partial resistance in oil palm progenies to *Ganoderma* basal stem rot. *J Oil Palm Res*, 16(2):12-18.
- Idris, A., Kushairi, D., Ariffin, D., and Basri, M., 2006. Technique for inoculation of oil palm germinated seeds with *Ganoderma*. *MPOB Inf Ser* 314:1-4.
- Idris, A. S., Rajinder, S., Madihah, A. Z., and Mohd, B. W. (2010). Multiplex PCR-DNA kit for early detection and identification of *Ganoderma* species in oil palm. *MPOB Inf Ser, MPOB TS*, (73).
- Jalani, B., Basiron, Y., Darus, A., Chan, K., and Rajanaidu, N., (2002). Prospects of elevating national oil palm productivity: a Malaysian perspective. *Oil Palm Industry Economic J* 2:1-9.
- Johnson, G., and Schaal, L. A. (1952). Relation of chlorogenic acid to scab resistance in potatoes. *Science*, 115(2997):627-629.
- Jourdan, C., and Rey, H. (1997). Architecture and development of the oil-palm (*Elaeis guineensis* Jacq.) root system. *Plant and Soil*, 189(1):33-48.
- Jung, H. W., Tschaplinski, T. J., Wang, L., Glazebrook, J., and Greenberg, J. T. (2009). Priming in systemic plant immunity. *Science*, 324(5923):89-91.
- Kessmann, H., Staub, T., Hofmann, C., Maetzke, T., Herzog, J., Ward, E and Ryals, J. (1994). Induction of systemic acquired disease resistance in plants by chemicals. *Annual review of phytopathology*, 32(1):439-459.

- Khairudin, H., 1993. Ganoderma disease of oil palm, Workshop on Plantation Management Update, 29. International oil palm conference, *Palm Oil Research Institution*, 418-423.
- Khalil, A. (2002). Production and characterization of cellulolytic and xylanolytic enzymes from the lignolytic white-rot fungus *Phanerochate chrysosporium* grown on sugar bagasse. *World Journal of Microbiology and Biotechnology* 18:753-759.
- Kinge, T. R., and Mih, A. M. (2011). *Ganoderma ryvardense* sp. nov. associated with basal stem rot (BSR) disease of oil palm in Cameroon. *Mycosphere*, 2(2):179-188.
- Kirk, T. K., and Farrell, R. L. (1987). Enzymatic" combustion": the microbial degradation of lignin. *Annual Reviews in Microbiology*, 41(1):465-501.
- Koyama, M., Helbert, W., Imai, T., Sugiyama, J., and Henrissat, B. (1997). Parallel-up structure evidences the molecular directionality during biosynthesis of bacterial cellulose. *Proceedings of the National Academy of Sciences*, 94(17):9091-9095.
- Kumari, M., Yadav, R. S. S., and Yadav, K. D. S. (2002). Secretion of ligninperoxidase by *Penicillium citrinum*, *Fusarium oxysporum* and *Aspergillus terreus*. *Indian Journal of Experimental boilohy*, (40):802-806.
- Kurosaki, F., Tashiro, N., and Nishi, A. (1986). Induction of chitinase and phenylalanine ammonia-lyase in cultured carrot cells treated with fungal mycelial walls. *Plant and cell physiology*, 27(8):1587-1591.
- Lam, T. B. T., Iiyama, K., and Stone, B. A. (2003). Hot alkali-labile linkages in the walls of the forage grass *Phalaris aquatica* and *Lolium perenne* and their relation to in vitro wall digestibility. *Phytochemistry*, 64(2):603-607.
- Lamaming, J., Hashim, R., Sulaiman, O., Leh, C. P., Sugimoto, T., and Nordin, N. A. (2015). Cellulose nanocrystals isolated from oil palm trunk. *Carbohydrate polymers*, 127:202-208.
- Langcake, P., Irvine, J. A., and Jeger, M. J. (1981). Alternative chemical agents for controlling plant disease. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*:83-101.
- Lattanzio, V., Lattanzio, V. M., and Cardinali, A. (2006). Role of phenolics in the resistance mechanisms of plants against fungal pathogens and insects. *Phytochemistry: Advances in research*, 66:1-7.
- Law, K. N., Kokta, B. V., and Mao, C. B. (2001). Fibre morphology and soda-sulphite pulping of switchgrass. *Bioresource technology*, 77(1):1-7.

- Le Bris, C., Paillard, C., Stiger-Pouvreau, V., and Guérard, F. (2013). Laccase-like activity in the hemolymph of *Venerupis philippinarum*: characterization and kinetic properties. *Fish & shellfish immunology*, 35(6):1804-1812.
- Lee, K. C., Arai, T., Ibrahim, D., Prawitwong, P., Deng, L., Murata, Y., and Kosugi, A. (2015). Purification and Characterization of a Xylanase from the Newly Isolated *Penicillium rolfsii* c3-2 (1) IBRL. *BioResources*, 10(1):1627-1643.
- Lewis, N. G., and Yamamoto, E. (1990). Lignin: occurrence, biogenesis and biodegradation. *Annual review of plant biology*, 41(1):455-496.
- Lim, K., Chuah, J., and Ho, C. (1993). Effects of soil heaping on *Ganoderma* infected oil palms. In *Proceedings of the 735-738*.
- Lorenz, A. J., Anex, R. P., Isci, A., Coors, J. G., De Leon, N., and Weimer, P. J. (2009). Forage quality and composition measurements as predictors of ethanol yield from maize (*Zea mays* L.) stover. *Biotechnology for biofuels*, 2(1):5-13.
- Luna, M. L., Murace, M. A., Keil, G. D., and Otaño, M. E. (2004). Patterns of decay caused by *Pycnoporus sanguineus* and *Ganoderma lucidum* (Aphyllophorales) in poplar wood. *IAWA Journal*, 25(4):425-433.
- Machado, K. M., Matheus, D. R., and Bononi, V. L. (2005). Lignolytic enzymes production and Remazol Brilliant Blue R decolorization by tropical Brazilian basidiomycetes fungi. *Brazilian Journal of Microbiology*, 36(3):246-252.
- Maldonado, A.M., Doerner, P., Dixon, R.A., Lamb, C.J., and Cameron, R.K., (2002). A putative lipid transfer protein involved in systemic resistance signalling in *Arabidopsis*. *Nature*, 419:399-403.
- Mandels, M., and Reese, E. T. (1963). Inhibition of cellulases and β -glucosidases. *Advances In enzymic hydrolysis of cellulose and related materials*. Pergamon, London, 115.
- Mandels, M., and Reese, E. T. (1965). Inhibition of cellulases. *Annual review of Phytopathology*, 3(1):85-102.
- Mann, D. G., Labbé, N., Sykes, R. W., Gracom, K., Kline, L., Swamidoss, I. M., and Stewart, C. N. (2009). Rapid assessment of lignin content and structure in switchgrass (*Panicum virgatum* L.) grown under different environmental conditions. *BioEnergy Research*, 2(4):246-256.
- Martinez, D., Larrondo, L. F., Putnam, N., Gelpke, M. D. S., Huang, K., Chapman, J., and Coutinho, P. M. (2004). Genome sequence of the

- lignocellulose degrading fungus *Phanerochaete chrysosporium* strain RP78. *Nature biotechnology*, 22(6):695-899.
- Matern, U., and Kneusel, R. E. (1988). Phenolic compounds in plant disease resistance. *Phytoparasitica*, 16(2):153-170.
- Mauch-Mani, B., and Slusarenko, A. J. (1996). Production of salicylic acid precursors is a major function of phenylalanine ammonia-lyase in the resistance of Arabidopsis to *Peronospora parasitica*. *The Plant Cell*, 8(2):203-212.
- McNeil, M., Davill, A. G., Fry, S. C., and Albersheim, P. (1984). Structure and function of the primary cell walls of plants. *Annual review of biochemistry*, 53(1):625-663.
- Métraux, J., Singer, H., Ryals, J., Ward, E., and Wyss-Benz, M. (1990). Increase in salicylic acid at the onset of systemic acquired resistance in cucumber, *Science*, 250:1004-1110.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical chemistry*, 31(3):426-428.
- Miller, R. N. G., Holderness, M., Bridge, P. D., Chung, G. F., and Zakaria, M. H. (1999). Genetic diversity of *Ganoderma* in oil palm plantings. *Plant Pathology*, 48(5):595-603.
- Mohd, Z., and Faridah, A. (2008). Disease suppression in *Ganoderma*-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science*, 44(3):101-107.
- Mohammed, C. L., Rimbawanto, A., and Page, D. E. (2014). Management of basidiomycete root-and stem-rot diseases in oil palm, rubber and tropical hardwood plantation crops. *Forest pathology*, 44(6):428-446.
- More, S. S., Renuka, P.S., and Malini, S. (2011). Isolation, purification, and characterization of fungal laccase from *Pleurotus* sp. *Enzyme research*, 2011.1-7.
- Moreira, F. G., Lenartovicz, V., and Peralta, R. M. (2004). A thermostable maltose-tolerant α -amylase from *Aspergillus tamarii*. *Journal of basic microbiology*, 44(1):29-35.
- MPOB (2015). Production of crude oil the month of February 2015, www.mpob.gov.my/index.php/statistics/production. (accessed Mac 17, 2015).
- MPOC (2015). Monthly production of oil palm products summary for the month of May 2015. Retrieved 03 July 2014, from

<http://bebi.mpob.gov.my/index.php/statistics/production-2014/660-production-of-oil-palm-products-2015.html>

- Mooney, C. A., Mansfield, S. D., Touhy, M. G., and Saddler, J. N. (1998). The effect of initial pore volume and lignin content on the enzymatic hydrolysis of softwoods. *Bioresource Technology*, 64(2):113-119.
- Moura-Sobczak, J., Souza, U., and Mazzafera, P. (2011, September). Drought stress and changes in the lignin content and composition in Eucalyptus. *In BMC proceedings* (Vol. 5, No. 7, p. P103). BioMed Central. 5(7):103.
- Moura, J. C. M. S., Bonine, C. A. V., De Oliveira Fernandes Viana, J., Dornelas, M. C., and Mazzafera, P. (2010). Abiotic and biotic stresses and changes in the lignin content and composition in plants. *Journal of integrative plant biology*, 52(4):360-376.
- Murai, K., Uchida, R., Okubo, A., and Kondo, R. (2009). Characterization of the oil palm (*Elaeis guineensis*) trunk as a material for bio-ethanol production. *Journal of the Japan Wood Research Society (Japan)*.
- Naidu, Y., Meon, S., and Siddiqui, Y. (2013). Foliar application of microbial-enriched compost tea enhances growth, yield and quality of muskmelon (*Cucumis melo L.*) cultivated under fertigation system. *Scientia Horticulturae*, 159:33-40.
- Narayanasamy, P. (2011). Detection of fungal pathogens in plants. In *Microbial Plant Pathogens-Detection and Disease Diagnosis*: Springer Netherlands:5-199.
- Návarová, H., Bernsdorff, F., Döring, A. C., and Zeier, J. (2012). Pipecolic acid, an endogenous mediator of defense amplification and priming, is a critical regulator of inducible plant immunity. *The Plant Cell*, 24(12):5123-5141.
- Nazir, A., Soni, R., Saini, H. S., Manhas, R. K., and Chadha, B. S. (2009). Purification and characterization of an endoglucanase from *Aspergillus terreus* highly active against barley β -glucan and xyloglucan. *World Journal of Microbiology and Biotechnology*, 25(7):1189-1197.
- Nicholson, R. L., and Hammerschmidt, R. (1992). Phenolic compounds and their role in disease resistance. *Annual review of phytopathology*, 30(1):369-389.
- Niemann, G. J., van der Kerk, A., Niessen, W. M., and Versluis, K. (1991). Free and cell wall-bound phenolics and other constituents from healthy and fungus-infected carnation (*Dianthus caryophyllus L.*) stems. *Physiological and molecular plant pathology*, 38(6):417-432.

- Nuemberger, T., and Lipka, V. (2005). Non-host resistance in plants: new insights into an old phenomenon. *Molecular plant pathology*, 6(3):335-345.
- Nun, N. B., Lev, A. T., Harel, E., and Mayer, A. M. (1988). Repression of laccase formation in *Botrytis cinerea* and its possible relation to phytopathogenicity. *Phytochemistry*, 27(8):2505-2509.
- Nurrashyeda, R., Idris, A. S., Madihah, A. Z., Ramle, M., and Kushairi, A. (2011). Hendersonia GanoEF1 granules for the control of *Ganoderma boninense* in oil palm. *MPOB Information Series*, (556).
- Nyanhongo, G. S., Gomes, J., Gübitz, G., Zvauya, R., Read, J. S., and Steiner, W. (2002). Production of laccase by a newly isolated strain of *Trametes modesta*. *Bioresource Technology*, 84(3):259-263.
- Ogawa, D., Nakajima, N., Seo, S., Mitsuhara, I., Kamada, H., and Ohashi, Y. (2006). The phenylalanine pathway is the main route of salicylic acid biosynthesis in Tobacco mosaic virus-infected tobacco leaves. *Plant biotechnology*, 23(4):395-398.
- Okino, L. K., Machado, K. M. G., Fabris, C., and Bononi, V. L. R. (2000). Lignolytic activity of tropical rainforest basidiomycetes. *World Journal of Microbiology and Biotechnology*, 16(8), 889-893.
- Ommelna, B. G., Jennifer, A. N., and Chong, K. P. (2012). The potential of chitosan in suppressing *Ganoderma boninense* infection in oil-palm seedlings. *J Sustain Sci Manage*, 7(2):186-192.
- Oostendorp, M., Kunz, W., Dietrich, B., and Staub, T. (2001). Induced disease resistance in plants by chemicals. *European Journal of Plant Pathology*, 107(1):19-28.
- Pal, S., Banik, S. P., Ghorai, S., Chowdhury, S., and Khowala, S. (2010). Purification and characterization of a thermostable intra-cellular β -glucosidase with transglycosylation properties from filamentous fungus *Termitomyces clypeatus*. *Bioresource technology*, 101(7):2412-2420.
- Palmieri, G., Giardina, P., Bianco, C., Fontanella, B., and Sannia, G. (2000). Copper induction of laccase isoenzymes in the ligninolytic fungus *Pleurotus ostreatus*. *Applied and Environmental Microbiology*, 66(3):920-924.
- Pantzaris, T. P. (2001). Techno-economic aspects of palm kernel oil. *INFORM-International News on Fats, Oils and Related Materials*, 12(1):69-80.

- Parihar, P. S., Prakash, O., and Punetha, H. (2012). Investigation on defensive enzymes activity of *Brassica juncea* genotypes during pathogenesis of *Alternaria* blight. *Nature and Science*, 10(2):63-68.
- Patel, H., Gupte, S., Gahlout, M., and Gupte, A. (2014). Purification and characterization of an extracellular laccase from solid-state culture of *Pleurotus ostreatus* HP-1. *Biotech*, 4(1):77-84.
- Paterson, R. R. M., and Bridge, P. D. (1994). *Biochemical techniques for filamentous fungi* (No. 1). CAB INTERNATIONAL.
- Paterson, R. R. M. (2007). *Ganoderma* disease of oil palm—A white rot perspective necessary for integrated control. *Crop protection*, 26(9), 1369-1376.
- Paterson, R. R., Meon, S., Abidin, M. Z., and Lima, N. (2008). Prospects for inhibition of lignin degrading enzymes to control *Ganoderma* white rot of oil palm. *Current Enzyme Inhibition*, 4(4):172-179.
- Paterson, R. R., Moen, S., and Lima, N. (2009). The feasibility of producing oil palm with altered lignin content to control *Ganoderma* disease. *Journal of Phytopathology*, 157(11-12):649-656.
- Pérez, J., Muñoz-Dorado, J., de la Rubia, T. D. L. R., and Martínez, J. (2002). Biodegradation and biological treatments of cellulose, hemicellulose and lignin: an overview. *International Microbiology*, 5(2):53-63.
- Picart, P., Diaz, P., and Pastor, F. I. J. (2007). Cellulases from two *Penicillium* sp. strains isolated from subtropical forest soil: production and characterization. *Letters in applied microbiology*, 45(1):108-113.
- Pointing, S. B. (1999). Qualitative methods for the determination of lignocellulolytic enzyme production by tropical fungi. *Fungal diversity*, 2:17-34.
- Popescu, C. M., Lisa, G., Manoliu, A., Gradinariu, P., and Vasile, C. (2010a). Thermogravimetric analysis of fungus-degraded lime wood. *Carbohydrate Polymers*, 80(1):78-83.
- Popescu, C. M., Popescu, M. C., and Vasile, C. (2010b). Structural changes in biodegraded lime wood. *Carbohydrate Polymers*, 79(2):362-372.
- Potlakayala, S. D., Reed, D. W., Covello, P. S., and Fobert, P. R. (2007). Systemic acquired resistance in canola is linked with pathogenesis-related gene expression and requires salicylic acid. *Phytopathology*, 97(7):794-802.

- Rahamah Bivi, M., Farhana, M. S. N., Khairulmazmi, A., Idris, A., Ahmed, O. H., Zamri, R., and Sariah, M. (2012). In vitro effects of salicylic acid, calcium and copper ions on growth and sporulation of *Ganoderma boninense*. *African Journal of Biotechnology*, 11(70): 13477-13489.
- Raseda, N., Hong, S., Kwon, O. Y., and Ryu, K. (2014). Kinetic Evidence for the Interactive Inhibition of Laccase from *Trametes versicolor* by pH and Chloride. *J. Microbiol. Biotechnol*, 24(12):1673-1678.
- Rees, R. W., Flood, J., Hasan, Y., and Cooper, R. M. (2007). Effects of inoculum potential, shading and soil temperature on root infection of oil palm seedlings by the basal stem rot pathogen *Ganoderma boninense*. *Plant Pathology*, 56(5):862-870.
- Rees, R. W., Flood, J., Hasan, Y., Potter, U., and Cooper, R. M. (2009). Basal stem rot of oil palm (*Elaeis guineensis*); mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant pathology*, 58(5):982-989.
- Riddell, R. W. (1950). Permanent stained mycological preparations obtained by slide culture. *Mycologia*, 42(2):265-270.
- Ride, J. P. (1983). Cell wall and other structural barriers in defense. *Biochemical plant pathology*, 215-236.
- Riou, C., Salmon, J. M., Vallier, M. J., Günata, Z., and Barre, P. (1998). Purification, characterization, and substrate specificity of a novel highly glucose-tolerant β -glucosidase from *Aspergillus oryzae*. *Applied and Environmental Microbiology*, 64(10):3607-3614.
- Risanto, L., Hermiati, E., and Sudiyani, Y. (2014). Properties of Lignin from Oil Palm Empty Fruit Bunch and Its Application for Plywood Adhesive. *Makara Journal of Technology*, 18(2):67-75.
- Ruiz-Dueñas, F. J., Martínez, M. J., and Martínez, A. T. (1999). Molecular characterization of a novel peroxidase isolated from the ligninolytic fungus *Pleurotus eryngii*. *Molecular microbiology*, 31(1):223-235.
- Saleem, A., and Ebrahim, M. K. (2014). Production of amylase by fungi isolated from legume seeds collected in Almadinah Almunawwarah, Saudi Arabia. *Journal of Taibah University for Science*, 8(2):90-97.
- Sanderson, F. R., Pilotti, C. A., and Bridge, P. D. (2000). Basidiospores: their influence on our thinking regarding a control strategy for basal stem rot of oil palm. *Ganoderma diseases of perennial crops*, 113-119.
- Sansone, G., Rezza, I., Fernández, G., Calvente, V., Benuzzi, D., and Sanz, M. I. (2011). Inhibitors of polygalacturonase and laccase of *Botrytis cinerea*

and their application to the control of this fungus. *International biodeterioration & biodegradation*, 65(1):243-247.

- Sariah, M., Choo, C. W., Zakaria, H., and Norihan, M. S. (2005). Quantification and characterisation of *Trichoderma* spp. from different ecosystems. *Mycopathologia*, 159(1):113-117.
- Sariah, M., Hussin, M. Z., Miller, R. N. G., and Holderness, M. (1994). Pathogenicity of *Ganoderma boninense* tested by inoculation of oil palm seedlings. *Plant Pathology*, 43(3):507-510.
- Sasidhara, R., and Thirunalasundari, T. (2014). Lignolytic and Lignocellulosic enzymes of *Ganoderma lucidum* in liquid medium. *European Journal of Experimental Biology*, 4(2):375-379.
- Seo, G. S., and Kirk, P. M. (2000). *Ganodermataceae*: nomenclature and classification. *Ganoderma diseases of perennial crops*, 3-22.
- Sethi, B. K., Nanda, P. K., Sahoo, S., and Sena, S. (2016). Characterization of purified α -amylase produced by *Aspergillus terreus* NCF 4269.10 using pearl millet as substrate. *Cogent Food & Agriculture*, 2(1):1-11.
- Shabana, Y. M., Abdel-Fattah, G. M., Ismail, A. E., and Rashad, Y. M. (2008). Control of brown spot pathogen of rice (*Bipolaris oryzae*) using some phenolic antioxidants. *Brazilian Journal of Microbiology*, 39(3):438-444.
- Shah, J., and Klessig, D. F. (1999). Salicylic acid: signal perception and transduction. *New Comprehensive Biochemistry*, 33:513-541.
- Shah, J., and Zeier, J. (2013). Long-distance communication and signal amplification in systemic acquired resistance. *Frontiers in Plant Science*, 4(30):1-30.
- Shapiro, A. D., and Gutsche, A. T. (2003). Capillary electrophoresis-based profiling and quantitation of total salicylic acid and related phenolics for analysis of early signaling in *Arabidopsis* disease resistance. *Analytical biochemistry*, 320(2):223-233.
- Shirasu, K., Nakajima, H., Rajasekhar, V. K., Dixon, R. A., and Lamb, C. (1997). Salicylic acid potentiates an agonist-dependent gain control that amplifies pathogen signals in the activation of defense mechanisms. *The Plant Cell*, 9(2):261-270.
- Singh, G., Darus, A., and Sukaimi, J. (1991). *Ganoderma*-scourge of oil palm in the coastal area, Proceedings on *Ganoderma* workshop, Bangi, Selangor, Malaysia, Palm Oil Research Institute of Malaysia, 7-35.

- Silva, L. A. O., Terrasan, C. R. F., and Carmona, E. C. (2015). Purification and characterization of xylanases from *Trichoderma inhamatum*. *Electronic Journal of Biotechnology*, 18(4):307-313.
- Skidmore, A. M., and Dickinson, C. H. (1976). Colony interactions and hyphal interference between *Septoria nodorum* and phylloplane fungi. *Transactions of the British Mycological Society*, 66(1):57-64.
- Song, G.C., Choi, H.K., and Ryu, C.M. (2013). The folate precursor para-aminobenzoic acid elicits induced resistance against *Cucumber mosaic virus* and *Xanthomonas axonopodis*. *Annals of botany*, 111:925-934.
- Song, J. T., Koo, Y. J., Seo, H. S., Kim, M. C., Do Choi, Y., and Kim, J. H. (2008). Overexpression of AtSGT1, an Arabidopsis salicylic acid glucosyl transferase, leads to increased susceptibility to *Pseudomonas syringae*. *Phytochemistry*, 69(5):1128-1134.
- Srivastava, P., Andersen, P. C., Marois, J. J., Wright, D. L., Srivastava, M., and Harmon, P. F. (2013). Effect of phenolic compounds on growth and ligninolytic enzyme production in *Botryosphaeria* isolates. *Crop protection*, 43:146-156.
- Steyaert, R. L. (1967). Les *Ganoderma palmicoles*. *Bulletin du Jardin botanique national de Belgique/Bulletin van de National Plantentuin van België*, 37(4):465-492.
- Strasser, H., Tietjen, K. G., Himmelsbach, K., and Matern, U. (1983). Rapid effect of an elicitor on uptake and intracellular distribution of phosphate in cultured parsley cells. *Plant cell reports*, 2(3):140-143.
- Sun, R., and Tomkinson, J. (2001). Fractional separation and physico-chemical analysis of lignins from the black liquor of oil palm trunk fibre pulping. *Separation and Purification Technology*, 24(3):529-539.
- Sun, S., Zhang, Y., Que, Y., Liu, B., Hu, K., and Xu, L. (2013). Purification and characterization of fungal laccase from *Mycena purpureofusca*. *Chiang Mai J Sci*, 40:151-160.
- Sun, Y., Cheng, J. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource*, 83,1-11.
- Sundram, S., Meon, S., Seman, I. A., and Othman, R. (2015). Application of *arbuscular mycorrhizal* fungi with *Pseudomonas aeruginosa* UPMP3 reduces the development of *Ganoderma* basal stem rot disease in oil palm seedlings. *Mycorrhiza*, 25(5):387-397.
- Susanto, A., Sudharto, P. S., and Purba, R. Y. (2005). Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, 159(1), 153-157.

- Terashima, N., and Fukushima, K. (1989). Biogenesis and structure of macromolecular lignin in the cell wall of tree xylem as studied by microautoradiography. 160-168.
- Tey, C.C., and Mohd ahdy, A., (2007). Mitigating measures against *Ganoderma* basal stem rot of oil palm. Proc. PIPOC international Palm Oil Congress- Agriculture conference MPOB, Bangi 2, 866-880.
- Thompson, A., 1931. Stem rot of oil palm in Malaysia. Bulletin of the Department of Agriculture, Straits Settlements and F. M. S Science series 6.
- Tomlinson, P. B. (1961). Anatomy of the monocotyledons. II. Palmae. *Anatomy of the monocotyledons. II. Palmae.*
- Turner, P. D. (1981). *Oil palm diseases and disorders*. Oxford Univ. Press.
- Turner, P. D., and Gillbanks, R. A. (1974). Oil palm cultivation and management. *Oil palm cultivation and management.*
- Valle, E. (1957). On anti-fungal factors in potato leaves. *Acta Chemica Scandinavica*, 11(2):395-397.
- Vanholme, R., Morreel, K., Ralph, J., and Boerjan, W. (2008). Lignin engineering. *Current opinion in plant biology*, 11(3):278-285.
- Vio-Michaelis, S., Apablaza-Hidalgo, G., Gómez, M., Peña-Vera, R., and Montenegro, G. (2012). Antifungal activity of three Chilean plant extracts on *Botrytis cinerea*. *Bot. Sci*, 90(2):179-183.
- White, R. F. (1979). Acetylsalicylic acid (aspirin) induces resistance to tobacco mosaic virus in tobacco. *Virology*, 99(2):410-412.
- Widsten, p., and Kandelbaure, A. (2008). Laccase applications in the forest products industry:a review. *Enzyme and Microbiology Technology*, 42:293-307.
- Wijesekara, H. T. R., Wijesundera, R. L. C., and Rajapakse, C. N. K. (2013). Hyphal interactions between *Trichoderma viridae* and *Ganoderma boninense* Pat., the cause of coconut root and bole rot. *Journal of the National Science Foundation of Sri Lanka*, 24(3).
- Wildemuth, M. C., Dewdney, J., Wu, G., and Ausubel, F. M. (2001). Isochorismate synthase is required to synthesize salicylic acid for plant defense. *Nature*, 414:562-565.

- Ximenes, E., Kim, Y., Mosier, N., Dien, B., and Ladisch, M. (2011). Deactivation of cellulases by phenols. *Enzyme and microbial technology*, 48(1):54-60.
- Xu, F. (1996). Oxidation of phenols, anilines, and benzenethiols by fungal laccases: correlation between activity and redox potentials as well as halide inhibition. *Biochemistry*, 35(23):7608-7614.
- Yan, T. R., and Lin, C. L. (1997). Purification and characterization of a glucose-tolerant β -glucosidase from *Aspergillus niger* CCRC 31494. *Bioscience, biotechnology, and biochemistry*, 61(6):965-970.
- Yao, B., and Ji, Y., 2014. Lignin biodegradation with laccase-mediator systems. *Frontiers in Energy Research* 2, 12.
- Zhu, J. Y., and Pan, X. J. (2010). Woody biomass pretreatment for cellulosic ethanol production: technology and energy consumption evaluation. *Bioresource technology*, 101(13):4992-5002.