

UNIVERSITI PUTRA MALAYSIA

FUNCTIONAL CHARACTERISATION OF TRANSCRIPTION FACTORS WITH POTENTIAL AS KEY REGULATORS OF PALM OIL PRODUCTION

CHIN MEI-YEE

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FUNCTIONAL CHARACTERISATION OF TRANSCRIPTION FACTORS WITH POTENTIAL AS KEY REGULATORS OF PALM OIL PRODUCTION



Thesis submitted to the School of Graduate Studies, University Putra Malaysia, in fulfilment of the Requirements for the Degree of Master of Science

June 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

FUNCTIONAL CHARACTERISATION OF TRANSCRIPTION FACTORS WITH POTENTIAL AS KEY REGULATORS OF PALM OIL PRODUCTION

By

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June 2016

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Transcription factors play important and diverse roles in regulating gene expression. Each transcription factor binds to a specific DNA sequence (cis-elements) in the promoter region and can either activate or suppress the downstream gene expression. They have been known to coordinate expression of genes involved in the same biosynthetic pathway or physiological process. Three transcription factors EgNFYB3,EgLEC2 and EgWRI1-ANT were the target for studying transcriptional regulation of fatty acid biosynthesis in the oil palm. The sequences of the open reading frame were verified based on homology with known transcription factor sequences from other plant species. Recombinant vector constructs, each harbouring one of the selected transcription factors were constructed via Gateway cloning system. The oil palm genes driven by the cauliflower mosaic virus 35S constitutive promoter were introduced separately into 12 WAA (weeks after anthesis) oil palm mesocarp tissue slices, which is the period preceding oil synthesis through particle bombardment system. Each one was co-bombarded with a green fluorescent protein (GFP) reporter vector. Mesocarp tissue slices which expressed GFP protein at 48 hours postbombardment were selected for total RNA extraction. High throughput sequencing (RNA-Seq) and transcript profiling of transiently transformed mesocarp slices were performed to determine genes whose expression is affected by transient overexpression of each transcription factor. RNA-Seq results were analysed for differentially expressed genes that are involved in fatty acid biosynthesis, glycolysis and TAG assembly pathways. EgNFYB3 bombarded tissues upregulated transcripts encoding enzymes for glycolysis, EgWRI1-ANT bombarded tissues upregulated transcripts encoding enzymes in fatty acid biosynthesis and glycolysis, while EgLEC2 bombarded tissues upregulated a transcript encoding an enzyme from fatty acid biosynthesis pathway in both biological replicates. RT-qPCR validation of the differentially expressed genes were performed and the results showed correlation to the RNA-Seq data. EgWRI1-ANT was chosen for subsequent DNA-protein interaction in vivo with yeast one-hybrid assay and EMSA for in vitro study. RNA-Seq results reveal the role of EgWRI1-ANT transcription factor in regulating genes related to glycolysis and fatty acid biosynthesis. Yeast one-hybrid assay confirmed that EgWRI1-ANT transcription factor binds to the AW-box 1*cis*-element to activate transcription of the genes it upregulated. These results demonstrate that oil palm transcription factors EgNFYB3, EgLEC2 and EgWRI1-ANT are important transcriptional regulator of carbohydrate metabolism and fatty acid biosynthesis in oil palm mesocarp, a non-seed oil-rich tissue, thus making them candidates for future development of molecular tools for genetic improvement of the oil palm.



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PENCIRIAN BERFUNGSI FAKTOR-FAKTOR TRANSKRIPSI YANG BERPOTENSI SEBAGAI PENGAWAL SELIA PROSES PENGELUARAN MINYAK KELAPA SAWIT

Oleh

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Pengerusi: Profesor Datin Siti Nor Akmar Abdullah, PhDFakulti: Institut Pertanian Tropika dan Sekuriti Makanan

Faktor-faktor transkripsi memainkan peranan yang penting dan pelbagai dalam pengawalan selia ekspresi gen. Setiap faktor transkripsi mengikat kepada jujukan DNA spesifik (cis-elemen) di rantau penganjur dan boleh mengaktifkan atau menyekat ungkapan gen hiliran. Faktor-faktor transkripsi dikenal dalam menyelaraskan ekspresi gen yang terlibat dalam laluan biosintetik yang sama atau proses fisiologi yang sama. Tiga faktor transkripsi iaitu EgNFYB3, EgLEC2 dan EgWRI1-ANTadalah sasaran untuk mengkaji peraturan transkripsi biosintesis asid lemak dalam pokok kelapa sawit. Urutan rangka bacaan terbuka telah disahkan berdasarkan homologi dengan urutan faktor transkripsi diketahui dari spesis tumbuhan yang lain melalui penjajaran jujukan berbilang dan analisis filogenetik. Konstruk vektor rekombinan, setiap salah satu daripadanya mengandungi faktor-faktor transkripsi terpilih dari kelapa sawit dibina melalui sistem pengklonan Gateway. Gen faktor transkripsi kelapa sawit dibawah kawalan promoter constitutive cauliflower mosaic virus 35S diperkenalkan secara berasingan ke dalam tisu hirisan mesokarp kelapa sawit berusia 12 WAA (minggu selepas berbunga), iaitu tempoh sebelumnya sintesis minyak melalui sistem pembedilan zarah. Masing-masing telah dibedil bersama vektor Green fluorescent protein (GFP) yang merupakan gen pelapor. Hirisan tisu mesocarp yang mengekspresi protein GFP pada masa 48 jam selepas pembedilan telah dipilih untuk pengekstrakan RNA. Penjujukan kendalian tinggi (Penjujukan-RNA) dan pemprofilan transkrip hirisan mesokarp yang telah ditransformasi secara transient dijalankan untuk menentukan ungkapan gen yang terjejas hasil daripada ungkapan transien setiap faktor transkripsi. Keputusan RNA-Seq telah dianalisa bagi menentukan gen yang mengalami pembezaan pengeskpresan dan juga terlibat dalam biosintesis asid lemak, glikolisis dan laluan pemasangan TAG. Didapati bahawa faktor transkripsi EgNYFB3 menyebabkan peningkatan pengekspresan tiga transkrip yang mengekod enzim untuk glikolisis. EgWRI1-ANT menyebabkan peningkatan pengekspresan tiga transkrip yang biosintesis mengekod enzim untuk asid lemak glikolisis dan manakala EgLEC2menyebabkan peningkatan pengekspresan transkrip yang mengekod enzim untuk biosintesis asid lemak dalam kedua-dua replikat biologi. Pengesahan RT-qPCR ke atas gen yang mengalami pembezaan pengeskpresan telah dijalankan dan hasil kajian menunjukkan korelasi dengan data RNA-Seq. Faktor transkripsi EgWRI1-ANT telah dipilih untuk kajian seterusnya iaitu interaksi DNA-protein secara in vivo dengan asai yis-satu hibrid dan EMSA secara in vitro. Keputusan RNA-Seq mendedahkan peranan faktor transkripsi EgWRI1-ANT dalam mengawal selia gen-gen yang berkaitan dengan glikolisis dan biosintesis asid lemak. Asai yis-satu hibrid mengesahkan bahawa faktor transkripsi EgWRI1-ANT mengikat kepada jujukan DNA-spesifik AW box-1 untuk mengaktifkan transkripsi gen yang tahap pengekspresannya meningkat. Keputusan ini menunjukkan bahawa faktor transkripsi kelapa sawit EgNFYB3, EgLEC2 dan EgWRI1-ANT adalah pengawal-selia transkripsi yang penting terhadap karbohidrat metabolisme dan biosintesis asid lemak dalam mesocarp kelapa sawit, salah tisu kaya minyak bukan biji. Faktor-faktor transkripsi kelapa sawit ini adalah calon-calon untuk pembangunan alat molekul masa hadapan bagi usaha pembaikan genetik kelapa sawit.

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I certify that a Thesis Examination Committee has met on 8 June 2016 to conduct the final examination of Chin Mei - Yee on her thesis entitled "Functional Characterisation of Transcription Factors with Potential as Key Regulators of Palm Oil Production" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

AP2/EREBP	APETALA2 Ethylene Response Element Binding Protein
RAV	Related to ABI3/VP1
OPTF	Oil Palm Transcription Factor
NFYB3	Nuclear Factor Y Subunit B3
LEC2	LEAFY COTYLEDON2
WRI1-ANT	APETALA2 Ethylene Response Element Binding Protein-ANT
CDS	Coding Region
ORF	Open Reading Frame
cDNA	Complementary Deoxyribonucleic Acid
LiCl	Lithium Chloride
EMSA	Electrophoretic Mobility Shift Assay
RT-PCR	Reverse Transcriptase Polymerase Chain Reaction
RT-qPCR	Reverse transcription quantitative Real-Time PCR
NCBI	National Center for Biotechnology Information
LiAc	Lithium Acetate
PEG	Polyethylene Glycol
mRNA	Messenger RNA
CaMV	Cauliflower Mosaic Virus
EVC	Empty Vector Control
DNase	Deoxyribonuclease
RPM	Revolutions per minute
OD	Optical Density
WAA	Weeks After Anthesis
ALDO-P	fructose-bisphosphate aldolase, chloroplast precursor, putative
	glyceraldehyde-3-phosphatedehydrogenase A, chloroplast
GAPDH-A	precursor, putative
	glyceraldehyde-3-phosphatedehydrogenase B chloroplast
GAPDH-B	precursor, putative
FatB	acyl-ACP thioesterase, chloroplast precursor, putative
HK2	Hexokinase-2
Fad7	omega-3 fatty acid desaturase, chloroplast precursor, putative
GFP	Green Fluorescent Protein
MCS	Multiple Cloning Site
TAG	Triacylglycerol

CHAPTER 1

INTRODUCTION

The oil palm (*Elaeis guineensis* Jacq.) is an economically important oil crop originating from central and West Africa (Corley and Tinker, 2003). It is the most productive amongst all oil crops, producing an average yield of about 4 t ha⁻¹ year⁻¹ of palm oil. The oil palm produces two different types of oils. Crude palm oil, also known as mesocarp oil, is extracted from the mesocarp and accounts for 95% of the total oil of the fruit, whereas the other 5% of crude palm kernel oil (CPKO), which is a non-edible oil, comes from the kernel. As global demand for palm oil continues to increase, from 17.7 million tonnes in 1997 to 61.1 million tonnes in 2015 (Oil World 2015), it is thus critical to meet this demand by increasing the oil palm yield. To meet this demand, the yield per unit area needs to be increased in the efforts to raise productivity without jeopardizing the environment by opening more lands for agriculture (Parveez et al., 2015).

Transcription factors are known to coordinate the expression of genes involved in the same biosynthetic pathway or physiological process, thus providing an interesting alternative to single-enzyme based solutions to improve complex traits such as oil accumulation in crops (Broun et al., 2004; Grotewold et al., 2008). However, the mechanism of regulation by these transcription factors are complex and may involve a network of different transcription factors interacting with different cis-acting elements working in synergy to produce the desired output.

Nuclear factor Y (NF-Y) is a heterotrimeric complex composed of NF-YA, NF-YB (HAP3), and NF-YC subunits which can bind to the CCAAT promoter elements (Dolfini et al., 2012) Based on sequence similarity of the conserved B-domain region, HAP3 transcription factors are divided into LEAFY COTYLEDON1 (LEC1) type and Non-LEC1 type (Kwong et al., 2003; Lee et al., 2003). Functional studies in Arabidopsis showed that LEC1-type are mostly expressed specifically in developing seeds, while most non-LEC1 type subunits are expressed in various other tissues (Zimmerman et al., 2004).

LEAFY COTYLEDON2 (LEC2) belongs to the B3 domain superfamily of plant specific transcription factors (Suzuki and McCarty, 2008). It is part of the closely related AFL clade which comprises ABSCISIC ACID INSENSITIVE3 (ABI3), FUSCA3 (FUS3), and LEC2, playing important role in seed development and maturation (Stone et al., 2001). The conserved structure of the B3 domain allows LEC2 to regulate the expression of downstream genes possessing the specific RY recognition motif "CATGCA", enriched in the promoter region of seed-specific genes (Stone et al., 2008). Due to partial functional redundancy and the overlapping expression patterns of the B3 AFL genes, the specific function of each AFL is hard to be inferred from genetic analyses (Roscoe et al., 2015).

AP2-like ethylene-responsive binding protein (AP2/EREBP) is one of the largest families of plant specific transcription factors, with roles in the control of primary and secondary metabolism, plant growth and development and responses to biotic and abiotic stresses (Licausi et al., 2013). Members of the AP2 subfamily contain conserved double AP2/ EREBP domains composed of 60-70 amino acid residues joined by a linker sequence of about 25 amino acids (Riechmann and Meyerowitz, 1998). Based on phylogenetic analyses, AP2 subfamily is further divided into two clades, APETALA2-like (AP2-like) and AINTEGUMENTA-like (ANT-like) groups (Shigyo & Ito, 2004). The ANT group is characterized by lineage-specific motifs or amino acid insertions in the two AP2 domains (Kim et al., 2006) with functions in lateral organ development by controlling cell number and growth (Elliot et al., 1996). Another member of the AP2 subfamily, WRINKLED1 (WRI1), regulates oil accumulation in seed tissues and its direct targets are late glycolysis and fatty acid biosynthesis genes (Cernac and Benning, 2004; Baud et al., 2009). Sequence analysis of its putative target gene promoters led to the identification of the AW-box DNAbinding site with the consensus sequence "[CnTnG](n)7[CG)]" (Maeo et al., 2009).

Recent studies have found that genes involved in de novo fatty acid biosynthetic pathway share the same temporal transcription pattern. Before the start of seed maturation in Arabidopsis thaliana, coordinated upregulation of genes encoding different enzymes were observed, suggesting that the pathway may undergo a system of global transcriptional regulation. This was further supported by the discovery of master regulators of seed maturation such as LEC2 and WRI1 transcription factors, which have been shown to be involved in a regulatory cascade determining the induction of fatty acid biosynthetic genes before the start of seed maturation (Baud and Lepiniec, 2010). LEC2 induction was shown to correlate with the onset of oil accumulation in maturing seeds while its ectopic expression triggered the TAG accumulation in rosette leaves (Stone et al., 2001; Santos-Mendoza et al., 2005). Furthermore, it has been shown that in *Arabidopsis thaliana*, LEC1 positively regulates WRI1 expression and various genes from fatty acid biosynthetic pathway (Mu et al., 2008). WRI1 is well studied in oil synthesis but the other members of AP2 subfamily including ANT, which have a common function related to flowering (Li et al., 2015) and fruit development (Omidvar et al., 2013) is unknown in relation to oil synthesis. Flower development is important for fruit growth and proper development of oil palm fruit is crucial as the source of palm oil is from the mesocarp and kernel of the fruit (Sarpan et al., 2015).

Extensive research on oil biosynthesis has been done on seeds in which triacylglycerols (TAGs) accumulated during maturation (Baud et al., 2010) but the molecular mechanisms of lipid metabolism in the mesocarp of oil palm, which accumulate high levels of TAG, and the mesocarp of similar fleshy fruits such as olive and avocado are still unclear. An understanding of the transcriptional control of oil synthesis in mesocarp tissues of the oil palm is crucial in the efforts to produce high yielding oil palm genotypes. Therefore, in this study, we are interested to determine if OPTF members from NF-YB family (EgNFYB3), B3 domain family (EgLEC2) and AP2 subfamily (EgWRI1-ANT) play a role in the regulation of gene expression of glycolysis, fatty acid biosynthesis and TAG assembly pathways through identification of their putative target genes.

1.1 Research Objectives

Below are the objectives of this research project:

- 1) To produce recombinant vector constructs, each harbouring one of the selected transcription factors (NFYB3, LEC2 and WRI1-ANT) from oil palm and to introduce into oil palm mesocarp tissue slices
- 2) To determine genes whose expression are affected by transient over expression of each transcription factor in the mesocarp tissue slices by high throughput sequencing and transcript profiling method.
- 3) To validate the interaction with corresponding cis-acting elements of a selected transcription factor which may have a role in regulating oil production, using yeast one-hybrid assay and electrophoretic mobility shift assay (EMSA).

REFERENCES

- Rival A (2007) Oil Palm. In: Pua EC, Davey MR (eds), Transgenic crops VI. Springer Berlin Heidelberg, pp 59–80.
- AOCS Lipid Library. (2012) Role of transcription factors in storage lipid accumulation in plants.

http://lipidlibrary.aocs.org/Biochemistry/content.cfm?ItemNumber=40316.

- Baranowskij N, Frohberg C, Prat S, Willmitzer L (1994) A novel DNA binding protein with homology to Myb oncoproteins containing only one repeat can function as a transcriptional activator. The EMBO journal 13:5383–5392.
- Baud S, Mendoza MS, To A, et al (2007) WRINKLED1 specifies the regulatory action of LEAFY COTYLEDON2 towards fatty acid metabolism during seed maturation in Arabidopsis. The Plant Journal 50:825–838.
- Baud S, Dubreucq B, Miquel M, et al (2008) Storage reserve accumulation in arabidopsis: metabolic and developmental control of seed filling. The Arabidopsis Book 6:e0113.
- Baud S, Lepiniec L (2010) Physiological and developmental regulation of seed oil production. Progress in Lipid Research 49:235–249.
- Beattie KL, Wiegand RC, Radding CM (1977) Uptake of homologous single-stranded fragments by superhelical DNA II. Characterisation of the reaction. Journal of Molecular Biology 166:783–803.
- Benfey PN and Chua NH (1990) The cauliflower mosaic virus 35S promoter: combinatorial regulation of transcription in plants. Science 250: 959-966.

Bewley J (1997) Seed germination and dormancy. The Plant Cell 9:1055-1066.

- Birch RG (1997) Plant transformation: problems and strategies for practical application. Annual Review of Plant Physiology and Plant Molecular Biology 48:297–326.
- Bouché N, Bouchez D (2001) Arabidopsis gene knockout: phenotypes wanted. Current Opinion in Plant Biology 4:111–117.

- Boulikas T (1994) Putative nuclear localization signals(NLS) in protein transcription factors. Journal of Cellular Biochemestry 55:32-58.
- Bourgis F, Kilaru A, Cao X, et al (2011) Comparative transcriptome and metabolite analysis of oil palm and date palm mesocarp that differ dramatically in carbon partitioning. Proceedings of the National Academy of Sciences 108:18186– 18186.
- Bonaventure G, Salas JJ, Pollard MR, Ohlrogge JB (2003) Disruption of the FATB gene in Arabidopsis demonstrates an essential role of saturated fatty acids in plant growth. The Plant Cell 15:1020–33.
- Braybrook SA, Stone SL, Park S, et al (2006) Genes directly regulated by LEAFY COTYLEDON2 provide insight into the control of embryo maturation and somatic embryogenesis. Proceedings of the National Academy of Sciences 103:3468–3473.
- Broun P (2004) Transcription factors as tools for metabolic engineering in plants. Current Opinion in Plant Biology 7:202–209.
- Brown A, Slabas A, Rafferty J (2010) Fatty acid biosynthesis in plants—metabolic pathways, structure and organization. In: Wada H, Norio Murata (eds) Lipids in photosynthesis. Springer Netherlands, pp 11–34
- Bustin SA, Benes V, Garson JA, et al (2009) The MIQE guidelines: minimum information for publication of quantitative real-tme PCR experiments. Clinical Chemistry 55:611–622.
- Carey MF, Peterson CL, Smale ST (2012) Confirming the functional importance of a protein-DNA interaction. Cold Spring Harbor Protocols 7:733–757.
- Cernac A, Benning C (2004) WRINKLED1 encodes an AP2/EREB domain protein involved in the control of storage compound biosynthesis in Arabidopsis. The Plant Journal 40:575–585.
- Chapman K, Ohlrogge J (2012) Compartmentation of Triacylglycerol Accumulation in Plants. Journal of Biological Chemistry 287: 2288-2294.
- Cheah SC, Sambanthamurthi R, Siti Nor Akmar A, Abrizah O, Manaf MAA, Umi Salamah R, Parveez GKA (1995) Towards genetic engineering of oil palm (Elaeis guineensis Jacq.). In: Kader JC,Mazliak P (eds) Plant lipid metabolism. Kluwer Academic, Dordrecht, pp 570–572

- Cheetham J, Dehne F, Pitre S, et al (2003) Parallel CLUSTAL W for PC clusters. Proceedings of the 2003 international conference on Computational science and its applications: PartII 300–309.
- Chen L, Lee JH, Weber H, et al (2013) Arabidopsis BPM proteins function as substrate adaptors to a cullin3-based E3 ligase to affect fatty acid metabolism in plants. The Plant cell 25:2253–64.
- Chern MS, Bobb AJ, Bustos MM (1996) The regulator of MAT2(ROM2) protein binds to early maturation promoters and repressors PvALF activated transcription. Plant Cell 8:305.
- Claeyssen É and Rivoal J (2007) Isozymes of plant hexokinase: occurrence, properties and functions. Phytochemistry 68:709-731.

Corley RHV, Tinker PB (2003) The oil palm. Blackwell Science, Oxford.

- Corley RHV, Tinker PB (2016) The oil palm, 5th edn. WILEY Blackwell, West Sussex
- Curtis MD, Grossniklaus U (2003) A Gateway cloning vector set for high-throughput functional analysis of genes in planta [w]. Plant physiology 133:462–469.
- D'Aoust M-A, Lavoie P-O, Belles-Isles J, et al (2009) Transient expression of antibodies in plants using syringe agroinfiltration. In: Faye L, Gomord V (eds) Recombinant proteins from plants. Humana Press, pp 41–50
- De Cleene M, De Ley J (1976) The host range of crown gall. The Botanical Review 42: 389-466.
- Dare AP, Schaffer RJ, Lin-Wang K, et al (2008) Identification of a cis-regulatory element by transient analysis of co-ordinately regulated genes. Plant Methods 10:1746– 4811.
- Dietz KJ, Vogel MO, Viehhauser A (2010) AP2/EREBP transcription factors are part of gene regulatory networks and integrate metabolic, hormonal and environmental signals in stress acclimation and retrograde signalling. Protoplasma 245:3–14.
- Dinh TT, Girke T, Liu X, et al (2012) The floral homeotic protein APETALA2 recognizes and acts through an AT-rich sequence element. Development (Cambridge, England) 139:1978–86.

- Ditt RF, Kerr KF, de Figueiredo P, et al (2006) The Arabidopsis thaliana transcriptome in response to Agrobacterium tumefaciens. Molecular plant-microbe interactions : MPMI 19:665–81.
- Dolfini D, Gatta R, Mantovani R (2012) NF-Y and the transcriptional activation of CCAAT promoters. Critical Reviews in Biochemistry and Molecular Biology47:29–49.
- Dussert S, Guerin C, Andersson M, et al (2013) Comparative transcriptome analysis of three oil palm fruit and seed tissues that differ in oil content and fatty acid composition. Plant Physiology 162:1337–1358.
- Ebrahimi M, Abdullah SNA, Aziz MA, Namasivayam P (2015) A novel CBF that regulates abiotic stress response and the ripening process in oil palm (Elaeis guineensis) fruits. Tree Genetics & Genomes 11:56.
- Egan AN, Schlueter J, Spooner DM (2012) Applications of next-generation sequencing in plant biology. American Journal of Botany 99:175–185.
- Elliott RC, Betzner AS, Huttner E, Oakes MP, Tucker WQ, Gerentes D, Perez P, Smyth DR (1996) AINTEGUMENTA, an APETALA2-like gene of Arabidopsis with pleiotropic roles in ovule development and floral organ growth. Plant Cell8:155–168.
- Eshed Y, Baum SF, Perea JV, Bowman JL (2001) Establishment of polarity in lateral organs of plants. Current Biology 11:1251–1260.
- Finer JJ, Beck S, Buenrostro-Nava, et al (2006) Monitoring gene expression in plant tissues. In: Gupta SD, Ibaraki Y (eds) Plant Tissue Culture Engineering. Springer Netherlands, pp 31–46
- Focks N, Benning C (1998) wrinkled1: A novel, low-seed-oil mutant of Arabidopsis with a deficiency in the seed-specific regulation of carbohydrate metabolism. Plant physiology 118:91–101.
- Franco-Zorrilla JM, López-Vidriero I, Carrasco JL, et al (2014) DNA-binding specificities of plant transcription factors and their potential to define target genes. Proceedings of the National Academy of Sciences of the United States of America 111:2367–72.
- Fried MG and Bromberg JL (1997) Factors that affect the stability of protein-DNA complexes during gel electrophoresis. Electrophoresis 18:6–11.

- Fukuda N, Ikawa Y, Aoyagi T, Kozaki A (2013) Expression of the genes coding for plastidic acetyl-CoA carboxylase subunits is regulated by a location-sensitive transcription factor binding site. Plant Molecular Biology 82:473–483.
- Goldfarb AN & Lewandowska K (1994) Nuclear redirection of a cytoplasmic helix-loophelix protein via heterodimerization with a nuclear localizing partner. Experimental Cell Research 214: 481–485.
- Griffith M, Griffith OL, Mwenifumbo J, Goya R, Morrissy AS, et al. (2010) Alternative expression analysis by RNA Sequencing. Nature Methods 7: 843–847.
- Grimberg Å, Carlsson AS, Marttila S, et al (2015) Transcriptional transitions in Nicotiana benthamiana leaves upon induction of oil synthesis by WRINKLED1 homologs from diverse species and tissues. BMC Plant Biology 15:192.
- Grotewold E (2008) Transcription factors for predictive plant metabolic engineering: are we there yet?. Current Opinion in Biotechnology 19:138–144.
- Guerin C, Joët T, Serret J, Lashermes P, Vaissayre V, Agbessi M, Beulé T, Severac D, Amblard P, Tregear J, Durand-Gasselin T, Morcillo F, Dussert S (2016) Gene coexpression network analysis of oil biosynthesis in an interspecific backcross of oil palm. The Plant Journal87: 423-441.
- Guiltinan MJ & Miller L (1994) Molecular characterization of the DNA-binding and dimerization domains of the bZIP transcription factor, EmBP-1. Plant Molecular Biology 26:1041–1053.
- Guo L, Ma F, Wei F, et al (2014) Cytosolic phosphorylating glyceraldehyde-3-phosphate dehydrogenases affect Arabidopsis cellular metabolism and promote seed oil accumulation. Plant cell 26:1–14.
- Hadi NAA, Abdullah SNA (2015) Effects of over-expressing ethylene responsive transcription factor on expression of selected fruit ripening-related genes in oil palm (Elaeis guineensis Jacq.) mesocarp. Pertanika Journal of Tropical Agricultural Science 38:143–159.
- Harada JJ (1997). Seed maturation and control of germination. In: Larkins BA and Vasi IK (eds), Advances in Cellular and Molecular Biology of Plants, Volume 4, Cellular and Molecular Biology of Seed Development. Dordrecht: Kluwer Academic Publishers, pp. 545–592

- Harwood JL (2005). Fatty acid biosynthesis. In: Murphy DJ (ed), Plant Lipids: biology, utilisation and manipulation. Blackwell Publishing, Oxford, pp. 27-66.
- Hashim AT, Ishak Z, Ooi SE, Rosli SK, Chan PL, Rohani O et al. (2011) "Forging ahead with clones," In: Wahid MB., Choo YM, Chan KW (eds) Further Advances in Oil Palm Research. Kuala Lumpur: Malaysian Palm Oil Board, pp 102–140.
- Hellman LM, Fried MG (2007) Electrophoretic mobility shift assay (EMSA) for detecting protein-nucleic acid interactions. Nature protocols 2:1849–61.
- Helwa R, Hoheisel JD (2010) Analysis of DNA-protein interactions: from nitrocellulose filter binding assays to microarray studies. Analytical and Bioanalytical Chemistry 398:2551–2561.

Hughes TR (2011) A handbook of transcription factors.

- J.Galas D and Schmitz A (1978) DNAase footprinting : a simple method for the detection of protein-DNA binding specificity. Nucleic Acids Research 5:3157–3170.
- Jofuku KD, Den Boer BG, Van Montagu M, Okamuro JK (1994) Control of Arabidopsis flower and seed development by the homeotic gene APETALA2. The Plant cell 6:1211–1225.
- Kagaya Y, Ohmiya K, Hattori T (1999) RAV1, a novel DNA-binding protein, binds to bipartite recognition sequence through two distinct DNA-binding domains uniquely found in higher plants. Nucleic Acids Research 27:470–478.
- Kamath RS, Fraser AG, Dong Y, Poulin G, Durbin R, Gotta M, Kanapin, A., Le Bot N, Moreno S, Sohrmann M. et al. (2003) Systematic functional analysis of the Caenorhabditis elegans genome using RNAi. Nature, 421: 231-237.
- Kikkert JR, Vidal JR, Reisch BI (2005) Stable transformation of plant cells by particle bombardment/biolistics. In: Leandro Peña (ed) Transgenic Plants: Methods and Protocols. Totowa, NJ, pp 61–78
- Kilaru A, Cao X, Dabbs PB, et al (2015) Oil biosynthesis in a basal angiosperm: transcriptome analysis of Persea Americana mesocarp. BMC Plant Biology 15:203.
- Kim S, Soltis PS, Wall K & Soltis DE (2006) Phylogeny and domain evolution in the APETALA2-like gene family. Molecular Biology Evolution 23: 107–120.

- Klucher KM, Chow H, Reiser L, Fischer RL (1996) The AINTEGUMENTA gene of Arabidopsis required for ovule and female gametophyte development is related to the floral homeotic gene APETALA2. The Plant cell 8:137–53.
- Koressaar T, Remm M (2007) Enhancements and modifications of primer design program Primer3. Bioinformatics 23:1289-91.
- Krizek BA (2003) AINTEGUMENTA utilizes a mode of DNA recognition distinct from that used by proteins containing a single AP2 domain. Nucleic Acids Research 31:1859–1868.
- Kroj T, Savino G, Valon C, et al (2003) Regulation of storage protein gene expression in Arabidopsis. Development 130:6065–73.
- Kumaran MK, Bowman JL, Sundaresan V (2002) YABBY polarity genes mediate the repression of KNOX homeobox genes in Arabidopsis. The Plant cell 14:2761–2770.
- Kwong RW, Bui AQ, Lee H, Kwong LW, Fischer RL, Goldberg RB and Harada JJ (2003) LEAFY COTYLEDON1-LIKE Defines a Class of Regulators Essential for Embryo Development. The Plant Cell 15:5–18.
- Leblanc BP, Moss T (eds) (2015) In vitro DNase I footprinting. In: DNA-Protein Interactions: Principles and Protocols, Methods in Molecular Biology. Springer New York, pp 17–27.
- Lee H, Fischer RL, Goldberg RB, Harada JJ (2003) Arabidopsis LEAFY COTYLEDON1 represents a functionally specialized subunit of the CCAAT binding transcription factor. Proceedings of the National Academy of Sciences of the United States of America 100:2152–6.
- Li-Beisson Y, Shorrosh B, Beisson F, et al (2010) Acyl-lipid metabolism. The Arabidopsis book / American Society of Plant Biologists 8:e0133.
- Licausi F, Ohme-Takagi M, Perata P (2013) APETALA2/Ethylene Responsive Factor (AP2/ERF) transcription factors: Mediators of stress responses and developmental programs. New Phytologist 199:639–649.
- Liljegren SJ, Ditta GS, Eshed Y, Savidge B, Bowman JL, Yanofsky MF (2000) SHATTERPROOF MADS-box genes control seed dispersal in Arabidopsis. Nature. 404:766-770.

- Liu L, White MJ, Macrae TH (1999) Transcription factors and their genes in higher plants. Eur j Biochem 257:247–257.
- Liu Q, Zhang GY, Chen SY (2001) Structure and regulatory function of plant transcription factors. Chinese Science Bulletin 46:271–278.
- Liu Y, Zhou J, White KP (2014) RNA-seq differential expression studies: More sequence or more replication? Bioinformatics 30:301–304.
- Loei H, Lim J, Tan M, Lim TK, Lin QS, Chew FT, Kulaveerasingam H, Chung MC (2013) Proteomic analysis of the oil palm fruit mesocarp reveals elevated oxidative phosphorylation activity is critical for increased storage oil production. Journal of Proteome Research. 12:5096-5109
- Lotan T, Ohto M, Yee KM, et al (1998) Arabidopsis LEAFY COTYLEDON1 is sufficient to induce embryo development in vegetative cells. Cell 93:1195–1205.
- Lü S, Gu H, Yuan X, et al (2007) The GUS reporter-aided analysis of the promoter activities of a rice metallothionein gene reveals different regulatory regions responsible for tissue-specific and inducible expression in transgenic Arabidopsis. Transgenic Research 16:177–191.
- Ma W, Kong Q, Arondel V, et al (2013) WRINKLED1, a ubiquitous regulator in oil accumulating tissues from Arabidopsis embryos to oil palm mesocarp. PLoS One 8: e68887
- Maeo K, Tokuda T, Ayame A, et al (2009) An AP2-type transcription factor, WRINKLED1, of Arabidopsis thaliana binds to the AW-box sequence conserved among proximal upstream regions of genes involved in fatty acid synthesis. The Plant Journal 60:476–487.
- Maity, S. and de Crombrugghe, B (1998) Role of the CCAAT-binding protein CBF/NF-Y in transcription. Trends in Biochemical Sciences. 23:174-178.
- Marchive C, Nikovics K, To A, et al (2014) Transcriptional regulation of fatty acid production in higher plants: Molecular bases and biotechnological outcomes. European Journal of Lipid Science and Technology 116:1332–1343.
- Marioni JC, Mason CE, Mane SM, et al (2008) comparison with gene expression arrays RNA-seq: An assessment of technical reproducibility and comparison with gene expression arrays. Genome Research 18:1509–1517.

- Meinke DW, Franzmann LH, Nickle TC, Yeung EC (1994) Leafy cotyledon mutants of Arabidopsis. The Plant Cell 6:1049–1064.
- Mizukami Y & Fischer RL (2000) Plant organ size control: AINTEGUMENTA regulates growth and cell numbers during organogenesis. Proceedings of the National Academy of Sciences of the United States of America 97:942–947.
- MPOB (2015) Malaysian oil palm statistics. Planted area and yield 2015. http://bepi.mpob.gov.my/images/overview/Overview_of_Industry_2015.pdf
- Mortazavi A, Williams BA, McCue K, et al (2008) Mapping and quantifying mammalian transcriptomes by RNA-Seq. Nature Methods 5:621–628.
- Mu J, Tan H, Zheng Q, et al (2008) LEAFY COTYLEDON1 is a key regulator of fatty acid biosynthesis in Arabidopsis. Plant physiology 148:1042–1054.
- Muñoz-Bertomeu J, Cascales-Miñana B, Mulet JM, et al (2009) Plastidial glyceraldehyde-3-phosphate dehydrogenase deficiency leads to altered root development and affects the sugar and amino acid balance in Arabidopsis. Plant physiology 151:541–558.
- Murashige, T and Skoog F (1962) A revised medium for rapid growth and bio assays with tobacco tissue cultures. Physiologia Plantarum 15:473-497.
- Murphy DJ (2003) Working to improve the oil palm crop. INFORM International News on Fats, Oils and Related Materials 14:670–671.
- Murphy DJ (2014) The future of oil palm as a major global crop: opportunities and challenges. Journal of Oil Palm Research 26:1–24.
- Nakano T, Suzuki K, Fujimura T, Shinshi H (2006) Genome-wide analysis of the ERF gene family. Plant Physiology 140:411–432.
- Neuhaus H and Emes M (2000) Non photosynthetic metabolism in plastids. Annual Review of Plant Physiology and Plant Molecular Biology 51: 111-140.
- Nishiuchi T (1999) Wound-induced expression of the FAD7 Gene Is mediated by different regulatory domains of its promoter in leaves/stems and roots. Plant Physiology 121:1239–1246.

Oil World (2015). Available from http://www.oilworld.biz/

Oil World (2016). Available from http://www.oilworld.biz/

Ohlrogge J, Browse J (1995) Lipid biosynthesis. The Plant Cell 7:957–970.

- Ohme-Takagi M, Shinshi H (1995) Ethylene-inducible DNA binding proteins that interact with an ethylene-responsive element. The Plant cell 7:173–82.
- Okamuro JK, Caster B, Villarroel R, et al (1997) The AP2 domain of APETALA2 defines a large new family of DNA binding proteins in Arabidopsis. Proceedings of the National Academy of Sciences of the United States of America 94:7076–81.
- Omidvar V, Siti Nor Akmar a., Marziah M, Maheran a. a. (2008) A transient assay to evaluate the expression of polyhydroxybutyrate genes regulated by oil palm mesocarp-specific promoter. Plant Cell Reports 27:1451–1459.
- Parveez GKA, Rasid OA, Masani MYA, Sambanthamurthi R (2015) Biotechnology of oil palm: strategies towards manipulation of lipid content and composition. Plant Cell Reports 34:533–543.
- Pellegrini L, Tan S, Richmond TJ (1995) Structure of serum response factor core bound to DNA. Nature 376:490–498.
- Peng J, Richards DE, Hartley NM, Murphy GP, Devos KM, Flintham JE, Beales J, Fish LJ, Worland AJ, Pelica F, et al (1999) 'Green revolution' genes encode mutant gibberellin response modulators. Nature 400:256-261.
- Pohanke J, Saleh J, Plieth C. Ballistic bombardment allows transient co-expression to 100%. Poster presented at Botanikertagung conference, Kiehl, Germany.
- Pouvreau B, Baud S, Vernoud V, et al (2011) Duplicate maize wrinkled1 transcription factors activate target genes involved in seed oil biosynthesis. Plant Physiology 156:674–686.
- Ramli Z, Nor S, Abdullah A (2003) Development of a transient promoter assay system for oil palm. Journal of Oil Palm Research 15:62–69.
- Rapaport F, Khanin R, Liang Y, et al (2013) Comprehensive evaluation of differential gene expression analysis methods for RNA-seq data. Genome Biology 14:3158

- Reece-Hoyes JS, Walhout AJM (2012) Yeast one-hybrid assays: a historical and technical perspective. Methods 57:441–447.
- Riechmann JL, Meyerowitz EM (1998) The AP2/EREBP family of plant transcription factors. Biological Chemistry. 379:633–646.
- Riechmann JL, Ratcliffe OJ (2000) A genomic perspective on plant transcription factors. Current Opinion in Plant Biology 3:423–434.
- Riggs AD, Bourgeois S, Newby RF, Cohn M (1968) DNA binding of the lac repressor. Journal of Molecular Biology 34:365–368.
- Riggs AD, Bourgeois S, Cohn M (1970) The lac repressor- operator interaction. 3. Kinetic studies. Journal of Molecular Biology 53:401–417
- Roscoe TT, Guilleminot J, Bessoule JJ, Berger F, Devic M (2015) Complementation of seed maturation phenotypes by ectopic expression of ABSCISIC ACID INSENSITIVE3, FUSCA3 and LEAFY COTYLEDON2 in Arabidopsis. Plant& Cell Physiology56: 1215–1228.
- Ruuska SA (2002) Contrapuntal networks of gene expression during Arabidopsis seed filling. The Plant Cell 14:1191–1206.
- Sainz MB, Grotewold E, Chandler VL (1997) Evidence for direct activation of an anthocyanin promoter by the maize C1 protein and comparison of DNA binding by related Myb domain proteins. The Plant cell 9:611–625.
- Sakuma Y, Liu Q, Dubouzet JG, et al (2002) DNA-binding specificity of the ERF/AP2 domain of Arabidopsis DREBs, transcription factors involved in dehydrationand cold-inducible gene expression. Biochemical and biophysical research communications 290:998–1009.

Saleh A, Pagés M (2003) Plant AP2/ERF transcription factors. Genetika 35:37-50.

- Sambanthamurthi R, Parveez GKA, Cheah SC (2000) Genetic engineering of oil palm. In: Basiron Y, Jalani BS, Chan KW (eds) Advances in oil palm research. Malaysian Palm Oil Board, Kuala Lumpur, pp 284–331
- Sanford JC, Smith FD, Russell JA (1993) Optimizing the biolistic process for different biological applications. Methods in Enzymology 217: 483-509

- Santos Mendoza M, Dubreucq B, Miquel M, et al (2005) LEAFY COTYLEDON 2 activation is sufficient to trigger the accumulation of oil and seed specific mRNAs in Arabidopsis leaves. FEBS Letters 579:4666–4670.
- Santos-Mendoza M, Dubreucq B, Baud S, et al (2008) Deciphering gene regulatory networks that control seed development and maturation in Arabidopsis. The Plant Journal 54:608–620.
- Sarpan N, Kok SY, Chai SK, Fitrianto A, Nuraziyan A, Zamzuri I, Ong-Abdullah M and Ooi SE (2015) A model for predicting flower development in Elaeis guineensis Jacq. Journal of Oil Palm Research 27: 315-325.
- Sawant SS, Singh PK, Tuli R. (2000) Pretreatment of microprojectiles to improve the delivery of DNA in plant transformation. BioTechniques 29: 246–248.
- Schwechheimer C and Bevan M (1998) The regulation of transcription factor activity in plants. Trend in Plant Science 3(10): 378-383.
- Schwender J, Ohlrogge JB, Shachar-Hill Y (2003) A flux model of glycolysis and the oxidative pentose phosphate pathway in developing Brassica napus embryos. Journal of Biological Chemistry 278:29442–29453.
- Seifert GJ (2004) Nucleotide sugar interconversions and cell wall biosynthesis: how to bring the inside to the outside. Current Opinion in Plant Biology 7:277–284.
- Shen B, Allen WB, Zheng P, et al (2010) Expression of ZmLEC1 and ZmWRI1 increases seed oil production in maize. Plant physiology 153:980–987.
- Shi Y, He M (2014) Differential gene expression identified by RNA-Seq and qPCR in two sizes of pearl oyster (Pinctada fucata). Gene 538:313–322.
- Shigyo M, Ito M (2004) Analysis of gymnosperm two-AP2-domain-containing genes. Development Genes and Evolution 214:105–114.
- Shigyo M, Hasebe M, Ito M (2006) Molecular evolution of the AP2 subfamily. Gene 366:256–265.
- Sinha S, Kim IS, Sohn KY, et al (1996) Three classes of mutations in the A subunit of the CCAAT-binding factor CBF delineate functional domains involved in the three-step assembly of the CBF-DNA complex. Molecular and cellular biology 16:328–37.

- Somerville C and Browse J (1991) Plant Lipids: Metabolism, Mutants, And Membranes. Science 252:80-87.
- Stephenson TJ, McIntyre L, Collet C, Xue G-P (2007) Genome-wide identification and expression analysis of the NF-Y family of transcription factors in Triticum aestivum. Plant Molecular Biology 65:77–92.
- Šmídková M, Holá M, Angelis KJ (2010) Efficient biolistic transformation of the moss Physcomitrella patens. Biologia Plantarum 54:777–780.
- Stockinger EJ, Gilmour SJ, Thomashow MF (1997) Arabidopsis thaliana CBF1 encodes an AP2 domain-containing transcriptional activator that binds to the Crepeat/DRE, a cis-acting DNA regulatory element that stimulates transcription in response to low temperature and water deficit. Proceedings of the National Academy of Sciences of the United States of America 94:1035–1040.
- Stone SL, Kwong LW, Yee KM, et al (2001) LEAFY COTYLEDON2 encodes a B3 domain transcription factor that induces embryo development. Proceedings of the National Academy of Sciences of the United States of America 98:11806– 11811.
- Stone S, Braybrook S, Paula S, Kwong L, Meuser J, Pelletier J, Hsieh T, Fischer R, Goldberg R, Harada J (2008) Arabidopsis LEAFY COTYLEDON2 induces maturation traits and auxin activity: Implications for somatic embryogenesis. Proceedings of the National Academy of Sciences105: 3151-3156.
- Suzuki M, McCarty DR (2008) Functional symmetry of the B3 network controlling seed development. Current Opinion in Plant Biology 11:548–553.
- Swaminathan K, Peterson K, Jack T (2008) The plant B3 superfamily. Trends in Plant Science 13:647–655
- Tamura K, Peterson D, Peterson N, et al (2011) MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Molecular Biology and Evolution 28:2731–2739.
- Tan H, Yang X, Zhang F, et al (2011) Enhanced seed oil production in canola by conditional expression of Brassica napus LEAFY COTYLEDON1 and LEC1-LIKE in developing seeds. Plant physiology 156:1577–1588.

- To A, Joubès J, Barthole G, et al (2012) WRINKLED transcription factors orchestrate tissue-specific regulation of fatty acid biosynthesis in Arabidopsis. The Plant cell 24:5007–23.
- Towbin H, Staehelin T, Gordon J (1979) Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. Proceedings of the National Academy of Sciences of the United States of America 76:4350–4354.
- Tranbarger TJ, Dussert S, Joet T, et al (2011) Regulatory Mechanisms Underlying Oil Palm Fruit Mesocarp Maturation, Ripening, and Functional Specialization in Lipid and Carotenoid Metabolism. Plant Physiology 156:564–584.
- Trapnell C, Roberts A, Goff L, et al (2012) Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks. Nature protocols 7:562–78.
- Ueki S, Lacroix B, Krichevsky A, et al (2009) Functional transient genetic transformation of Arabidopsis leaves by biolistic bombardment. Nature protocols 4:71–77.
- Ueki S, Magori S, Lacroix B, Citovsky Vi (2013) Transient gene expression in epidermal cells of plant leaves by biolistic DNA delivery. Methods in Molecular Biology 940:17–26.
- Untergasser A, Cutcutache I, Koressaar T, Ye J, Faircloth BC, Remm M, Rozen SG (2012) Primer3 new capabilities and interfaces. Nucleic Acids Research 40(15):e115
- Vandesompele J, De Preter K, Pattyn F, et al (2002) Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. Genome biology 3:RESEARCH0034.
- Vanhercke T, El Tahchy A, Shrestha P, et al (2013) Synergistic effect of WRI1 and DGAT1 coexpression on triacylglycerol biosynthesis in plants. FEBS Letters 587:364–369.
- Viola IL, Gonzalez DH (2015) Methods to study transcription factor structure and function. In: Gonzalez DH (ed) Plant transcription factors: evolutionary, structural and functional aspects, 1st ed. Academic Press, pp 13-33

- Wang Z, Kenigsbuch D, Sun L, et al (1997) A Myb-related transcription factor is involved in the phytochrome regulation of an Arabidopsis Lhcb Gene. The Plant Cell 9:491–507.
- Wang RL, Stec A, Hey J, et al (1999) The limits of selection during maize domestication. Nature 398:236–239.
- Warden CD, Yuan Y-C, Wu X (2013) Optimal calculation of RNA-Seq fold-change values. International Journal of Computational Bioinformatics and In Silico Modeling 2:285–292.
- Wood C, Petrie J, Shrestha P, Mansour P, Nichols P, Green A, Singh S (2009): A leafbased assay using interchangeable design principles to rapidly assemble multistep recombinant pathways. Plant Biotechnology Journal 9:914-924.
- Woodbury CP Jr, von Hippel PH (1983) On the determination of deoxyribonucleic acidprotein interaction parameters using the nitrocellulose filter-binding assay. Biochemistry 22:4730–4737.
- Wu AR, Neff NF, Kalisky T, Dalerba P, Treutlein B, et al. (2014) Quantitative Assessment of Single-Cell RNA-Sequencing Methods. Nature Methods 11: 41–46.
- Yamasaki K, Kigawa T, Seki M, et al (2013) DNA-binding domains of plant-specific transcription factors: Structure, function, and evolution. Trends in Plant Science 18:267–276.
- Yamashita T, Iida A, Morikawa H (1991) Evidence that more than 90% of betaglucuronidase-expressing cells after particle bombardment directly receive the foreign gene in their nucleus. Plant physiology 97:829–31.
- Yang Y, Li R, Qi M (2000) In vivo analysis of plant promoters and transcription factors by agroinfiltration of tobacco leaves. The Plant Journal 22:543–551.
- Yazawa K, Kamada H (2007) Identification and characterization of carrot HAP factors that form a complex with the embryo-specific transcription factor C-LEC1. Journal of Experimental Botany 58:3819–3828.
- Xing Y, Fikes JD, Guarente L (1993) Mutations in yeast HAP2/HAP3 define a hybrid CCAAT box binding domain. Embo J 12:4647–4655.

- Zhang JZ (2003) Overexpression analysis of plant transcription factors. Current Opinion in Plant Biology 6:430–440.
- Zhang Y, Clemens A, Maximova SN, Guiltinan MJ (2014) The Theobroma cacao B3 domain transcription factor TcLEC2 plays a duel role in control of embryo development and maturation. BMC Plant Biology 14:1–16.
- Zhuang J, Cai B, Peng RH, et al (2008) Genome-wide analysis of the AP2/ERF gene family in Populus trichocarpa. Biochemical and Biophysical Research Communications 371:468–474.

