



**UNIVERSITI PUTRA MALAYSIA**

***DEVELOPMENT OF AN ELECTROCHEMICAL SENSOR FOR  
DETECTION OF SECONDARY METABOLITE QUINOLINE IN  
Ganoderma boninense INFECTED OIL PALMS***

**FOWOTADE SULAYMAN AKANBI**

**FS 2018 84**



**DEVELOPMENT OF AN ELECTROCHEMICAL SENSOR FOR  
DETECTION OF SECONDARY METABOLITE QUINOLINE IN  
*Ganoderma boninense* INFECTED OIL PALMS**

By

**FOWOTADE SULAYMAN AKANBI**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

**May 2018**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**DEVELOPMENT OF AN ELECTROCHEMICAL SENSOR FOR  
DETECTION OF SECONDARY METABOLITE QUINOLINE IN  
*Ganoderma boninense* INFECTED OIL PALMS**

By

**FOWOTADE SULAYMAN AKANBI**

**May 2018**

**Chairman : Professor Nor Azah Binti Yusof. PhD**

**Faculty : Science**

The devastating effect of *Ganoderma boninense* (G. b) infections in oil palms, which leads to low-income revenues, due to the low yield of diseased palms, has driven researchers to look for early diagnostic techniques. The secondary metabolite, quinoline which was reported to be excreted from the oil palms when attacked by G. b can be used to detect the pathogenic fungus. In order to facilitate an indirect early detection of G. b, a new electrode based on functionalized multi-walled carbon nanotube was developed in this study. The development of the new electrode is based on layer-by-layer self-assembly method using activated multi-walled carbon nanotubes (aMWCNTs) as a backbone, for the attachment of other nanomaterials such as gold nanoparticles (AuNPs) and low molecular chitosan nanoparticle (ChTSNPs). The synthesized gold nanoparticles dispersion was characterized using Zetasizer nano series, UV-visible spectroscopy and cyclic voltammetric (CV) technique. The aMWCNTs and prepared nanohybrid materials (AuNPs-aMWCNTs), were characterized with the aid of field emission scanning electron microscope (FESEM), energy dispersive X-Ray (EDX), while ChTSNPs-aMWCNTs and aMWCNTs were characterized utilizing Fourier-transform infrared (FTIR) spectroscopy. The electrode modification process was monitored by FESEM and voltammetric techniques. Secondary metabolites were extracted from healthy and infected oil palm extracts, using ultrasound-assisted extraction (UAE) method. The performance of the developed electrode was optimized and characterized in quinoline using CV and linear sweep voltammetric (LSV) methods. The developed electrode was characterized in the leaves and root extract secondary metabolites using LSV technique under optimized conditions. The results showed that AuNPs of size 49.27nm and polydispersity index (PDI) of approximately 46% was chosen for electrode modification. This is because the PDI is below 50%. The FESEM micrographs show distinction among the pristine MWCNTs, aMWCNTs AuNPs-

aMWCNTs, bare electrode and the modified electrode. Also, the attachment of the carboxylic group (-COOH) to the walls of MWCNTs and the loading of the ChTSNPs onto the aMWCNTs were confirmed by the FT-IR spectral. The optimized conditions are as follow: 0.20 M citrate buffer, pH 5.5, accumulation potential, -0.52 V, accumulation time, 180 s and scan rate, 0.06 V/s. Under the measured optimal conditions, the anodic peak current ( $I_{pa}$ ) is directly proportional to concentration of quinoline, giving rise to the linear regression equation,  $I_{pa} (\mu A) = 0.7684 + 43.197 [\text{Quinoline}] / (\mu M)$ , coefficient of correlation,  $R^2 = 0.9949$ , with linear range 0.0004 and 0.10  $\mu M$ , limit of detection (LOD) 3.75 nM and limit of quantification (LOQ) 12.5 nM. The relative standard deviation (RSD) of  $I_{pa}$  of quinoline with single repeatedly used developed electrode was 2.33%, while it retained 91.7% of the current after being kept for twenty days. The evaluated reproducibility RSD for the between developed electrode anodic peak current response to quinoline oxidation was 3.52%. In addition, no apparent interference was observed in the presence of 1000-fold excess inorganic ions and 500-fold excess organics in 10.0  $\mu M$  quinoline as all the percentage interferences are below  $\pm 10\%$ . Furthermore, the newly developed electrode revealed satisfactory  $I_{pa}$  extract secondary metabolite response over the concentration range of 0.1 to 0.5 ppm with the limit of detection (3 S/N) ranging from 7.87 ppb to 18.54 ppb. The RSD value for reproducibility of  $I_{pa}$  across all the secondary metabolites ranges from 0.73% to 29.35%. The 500-fold excess of interfering organic species in 100 ppm extract secondary metabolite averagely exhibited insignificant interference in the detection process. The proposed sensor stands a brighter future in providing a point of care service in the management of BSR disease of oil palms in South East Asia, especially in Malaysia.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PEMBANGUNAN SENSOR ELEKTROKIMIA UNTUK PENGESANAN  
METABOLIT SEKUNDER DALAM *Ganoderma boninense* YANG  
MENJANGKITI KELAPA SAWIT**

Oleh

**FOWOTADE SULAYMAN AKANBI**

**Mei 2018**

**Pengerusi : Profesor Nor Azah Binti Yusof, PhD**  
**Fakulti : Sains**

Kesan buruk jangkitan *Ganoderma boninense* (G.b) pada kelapa sawit menyebabkan pulangan yang rendah disebabkan oleh hasil yang berkurangan daripada sawit yang dijangkiti, ini menyebabkan para pengkaji mencari kaedah diagnostik awalan. Metabolit sekunder, kuinolina yang telah dilaporkan dirembes dari pokok sawit apabila diserang oleh G.b boleh digunakan untuk mengesan kulat patogenik ini. Untuk menjalankan pengesanan awal G.b, sebuah elektrod baharu yang difungsi dengan karbon nanotub multidinding telah dihasilkan dalam kajian ini. Penghasilan elektrod baharu ini berasaskan kaedah swa pasang lapisan demi lapisan menggunakan karbon nanotub multidinding teraktif (aMWCNTs) sebagai tulang belakang, untuk pelekatan bahan nano lain seperti nanopartikel emas (AuNPs) dan nanopartikel kitosan rendah molekular (ChTSNPs). Ampaian nanopartikel emas yang disintesis telah dicirikan menggunakan siri Zetasizer nano, spektroskopi UV - tampak dan teknik voltammetrik kitaran (CV). aMWCNTs dan bahan nanohibrid yang disediakan (AuNPs-aMWCNT, dicirikan dengan bantuan mikroskop elektron pengimbasan pengaruh medan (FESEM), dan X-Ray penyebaran tenaga (EDX). Manakala ChTSNPs-aMWCNTs dan aMWCNTs dicirikan menggunakan spektroskopi inframerah Fourier-transform (FTIR). Proses pengubahsuaian elektrod dipantau menggunakan FESEM dan kaedah voltammetrik. Metabolit sekunder diekstrak daripada pokok kelapa sawit yang sihat dan terjangkit, menggunakan kaedah pengekstrakan berbantu ultrabunyi. Keupayaan elektrod yang dibina dioptimumkan dan dicirikan dalam quinoline menggunakan CV dan kaedah volumetrik sapuan linear (LSV). Elektrod yang dibina dicirikan pada ekstrak metabolit sekunder dedaun dan akar menggunakan teknik LSV dibawah persekitaraan yang optimum. Hasilnya menunjukkan AuNP bersaiz 49.27 nm dan indeks polidispersi (PDI) hampir 46% dipilih untuk pengubahsuaian elektrod. Ini kerana PDI hendaklah di bawah 50%. Mikrograf FESEM menunjukkan perbezaan

ketara diantara MCWCNTs, aMWCNTs AuNPs-aMWCNTs elektrod terdedah dan elektrod diubahsuai. Pelekatan kumpulan karboksil (-COOH) pada permukaan MWCNTs dan pemuatan MWCNTs pada aMWCNTs dikenal pasti menggunakan FTIR. Keadaan optimum adalah seperti berikut : 0.20 M penimbal sitrat, pH 5.5, pengumpulan keupayaan, -0.52 V, masa pengumpulan, 180 s dan kadar imbas, (0.06 V/s. di bawah keadaan optimum, puncak arus anodic ( $I_{pa}$ ) adalah berkadar langsung kepada kepekatan kuinolina, meningkatkan persamaan regresi linear,  $I_{pa} (\mu A) = 0.7684 + 43.197 [\text{kuinolina}] / (\mu M)$ , pekali korelasi,  $R^2 = 0.9949$ , dengan julat linear 0.0004 dan 0.10  $\mu M$ , had pengesanan (LOD) 3.75 dan had kuantifikasi (LOQ) 12.5 nM. Sisihan piawai relative (RSD)  $I_{pa}$  kuinolina dengan satu elektrod ulang pakai adalah 2.33%, dan mengekalkan 91.7% arus selepas disimpan untuk 20 hari. Nilai penghasilan-semula (RSD) untuk diantara tindakbalas arus puncak elektrod bertindak balas kepada pengoksidaan kuinolina adalah 3.52 %. Tambahan, tiada gangguan yang jelas diperhatikan dengan kehadiran lebih ion tidak organik 1000 ganda dan lebih ion organik 500 ganda, pada 10.0  $\mu M$  kuinolina, kerana peratusan gangguan keduanya adalah di bawah  $\pm 10\%$ . Adapun, elektrod yang baharu dibangunkan ini menunjukkan tindakbalas  $I_{pa}$  ekstrak metabolit sekunder yang memuaskan pada julat kepekatan 0.1 hingga 0.5 ppm dengan had pengesanan (3 S/N) berjulat dari 7.87 ppb to 18.54 ppb. Nilai RSD untuk penghasilan semula  $I_{pa}$  diantara kesemua metabolit sekunder berjulat dari 0.73% hingga 29.35%. Lebihan spesies organik gangguan di dalam ekstrak metabolit sekunder 100 ppm secara purata mempamerkan gangguan yang tidak signifikan dalam proses pengesanan. Sensor yang dicadangkan memiliki masa depan yang cerah dalam menangani penyakit BSR miyak sawit di Asia Tenggara, terutama di Malaysia.

## ACKNOWLEDGEMENTS

Bismillahir Rahmanir Raheem

The zenith of gratification belongs only and alone to Allah tabaraka wa ta'ala. The Lord of the worlds. The creator of everything. Then, to the seal of prophets, the Amir of the prophets, the mercy for the entire creation, Muhammad Habibullah Mustapha salallahu alayhi wa salam.

My sincerest gratitude goes to my Supervisor: Professor (Dr) Nor Azah binti Yusof, who by Allahu ta'ala ordering has been with me every step of the way and has helped nurture my love for diagnostics and sensors, for being there since the commencement of my postgraduate career, and for the invaluable advice and effort put into the realization of this work. She is a mentor, a guide, a companion, an instructor, a sister, a mother. A very big thank you, Ma.

To my families, the Fowotades, the Akinbiyis and the Oyenirans, most especially my loving wives Shakirat and Kifayat and dearing kids, Sufyan, Muhammad, Khadijat, Maymuna, Maryam, Dawood and Zulfiqar thank you for being the support and guidance that I have required throughout, for instilling a sense of pride and perseverance, I am forever indebted.

To my Shaykh AbdulQadr Naqshbandi (Damat Barakatuh), I express my reverence for your eminence personality and your duas and priceless words of Remembrance.

To the Abu sumayyah, Mallam Monko family and othe family friends I extend a special thank you, for your support and care. I am forever grateful for the supporting role you played throughout my PhD.

To my brothers Ali Akbar, Salman Jan, Kashif, Sufyan, Faisal (Jakel mall), Faisal (UM), Munir (Pakistani), Dr. Muhammad Nazrin, Ilyas, Azewan (Malaysia) Mahfuz, Mahmud, Arif rahman, Dr Ahsan, Dr Kamal, Dr Sohel Masud (Bangladesh) Muhammad Amin (The Gambia), Ahmad Tall, Abdullahi, Shuaib, Abdulkani, AbdurRazaq, Wilos (Somalis), Abdulwaheed, Isyaku Saleh, Dr Saheed Olajide (Nigerians), Dr. Muhammad Muhammad (Saudi Arabia, Egypt) I would like to say a special thank you for your contribution to the successful completion of this programme.



To the elderly friends Hajj Amir, Hajj Saleh, Hajj Umar, and all members of surau Al-hidayah the imam, pengurussi and secretary inclusive, please do accept my token appreciation for your concern and support through my stay in Malaysia.

To the laboratory mates of the Sensor and Biosensor Research Group: Dr. Muhammad Aliyu, Dr. Ibrahim Birma, Dr. Dayah, Dr. Diana, Salihu Suleiman, Boshra, Hamizah, Zulaiha, Nazifah, Zarif, Zila, Ben and others that spent some time in the group.

I wish to thank Boshra and Zarif for the fast editing of the bahasa malayu version of the abstract.

Also my appreciation goes to Prof Ali and Prof. Zurkarnial for discussing 40 ahadiths of Imam an-Nawawi with the Nigerian muslim brothers.

In the same vein is Muhammad of UPM masjid for sharing the jumu'a khutubah with me on weekly basis.

Lastly, to every other person that I could not be able to mention their names, I say Jazakumullah bikhayr kathiran.

I certify that a Thesis Examination Committee has met on 7 May 2018 to conduct the final examination of Fowotade Sulayman Akanbi on his thesis entitled "Development of an Electrochemical Sensor for Detection of Secondary Metabolite Quinoline in *Ganoderma boninense* Infected Oil Palms" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Mohamed Ibrahim bin Mohamed Tahir, PhD**

Senior Lecturer  
Faculty of Science  
Universiti Putra Malaysia  
(Chairman)

**Zulkarnain bin Zainal, PhD**

Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Lim Hong Ngee, Janet, PhD**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Bambang Kuswandi, PhD**

Professor  
University of Jember  
Indonesia  
(External Examiner)



---

**RUSLI HAJI ABDULLAH, PhD**  
Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 30 July 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Nor Azah Binti Yusof, PhD**

Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Chairman)

**Yusran Sulaiman, PhD**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

**Ja'afar Abdullah, PhD**

Senior Lecturer  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Fowotade Sulayman Akanbi, GS 43321

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_  
Name of Chairman  
of Supervisory  
Committee: Professor Dr. Nor Azah Binti Yusof

Signature: \_\_\_\_\_  
Name of Member  
of Supervisory  
Committee: Associate Professor Dr. Yusran Sulaiman

Signature: \_\_\_\_\_  
Name of Member  
of Supervisory  
Committee: Dr. Ja'afar Abdullah

## TABLE OF CONTENTS

		Page
<b>ABSTRACT</b>		i
<b>ABSTRAK</b>		iii
<b>ACKNOWLEDGEMENTS</b>		v
<b>APPROVAL</b>		vii
<b>DECLARATION</b>		ix
<b>LIST OF TABLES</b>		xv
<b>LIST OF FIGURES</b>		xvi
<b>LIST OF ABBREVIATIONS</b>		xx
<b>CHAPTER</b>		
<b>1</b>	<b>INTRODUCTION</b>	1
	1.1 Background of the study	1
	1.2 Problem statement	5
	1.3 Objective of the study	6
<b>2</b>	<b>LITERATURE REVIEW</b>	8
	2.1 The oil palm ( <i>Elaeis guineensis</i> )	8
	2.2 <i>Ganoderma boninense</i>	8
	2.2.1 Sources of <i>G. boninense</i>	9
	2.2.1.1 The other host plants for <i>G. boninense</i>	9
	2.2.1.2 The airborne basidiospores	9
	2.2.1.3 The spread via root to root contact	10
	2.2.2 Stages of the <i>G. boninense</i> infection	10
	2.3 Secondary metabolites	11
	2.3.1 The concept of secondary metabolites (SMs)	11
	2.3.2 The functional roles of secondary metabolites	11
	2.3.2.1 Defensive and resistant secondary metabolites	12
	2.3.2.2 Recognizing invasive pathogen or herbivore	13
	2.3.2.3 Types of phytoalexins	13
	2.4 Quinoline	14
	2.5 Electrochemical detection of secondary metabolites	15
	2.6 Electrochemical Sensors	16
	2.7 Voltammetry	17
	2.8 Screen printed electrodes (SPEs)	18
	2.8.1 Screen printed carbon electrodes (SPCEs)	19
	2.9 Layer-by-layer Assembly approach	20
	2.10 Applications of nanomaterial in electrochemical sensing devices	21

<b>3</b>	<b>MATERIALS AND METHODS</b>	<b>23</b>
3.1	Chemicals and Reagents	23
3.1.1	Preparation of solutions from reagents	26
3.1.1.1	Milky solution for activating pristine MWCNTS	26
3.1.1.2	Hydrogen tetrachloroaurate trihydrate solution	26
3.1.1.3	Sodium citrate solution	26
3.1.1.4	Chitosan nanoparticles solution	26
3.1.1.5	Quinoline solution	27
3.1.1.6	Hydrochloric acid solution	27
3.1.1.7	Potassium ferricyanide solution	27
3.1.1.8	Buffer solutions	27
3.1.2	Surface modifiers	28
3.1.2.1	Activation of pristine Multi-walled carbon nanotubes (pMWCNTs)	29
3.1.2.2	Preparation of gold nanoparticles (AuNPs)	29
3.1.2.3	Preparation of nanocomposites	29
3.1.3	Extraction of secondary metabolites from healthy and artificially inoculated samples	30
3.1.3.1	Plant materials and cultivation	30
3.1.3.2	Sample preparation	30
3.1.4	Extraction of secondary metabolites from field sampled leaves	31
3.2	Apparati and Instrumentation	31
3.2.1	Voltammetric Analyzer	31
3.2.2	Malvern Nano-sizer	32
3.2.3	Fourier-transform infrared (FT-IR) spectrometer	33
3.2.4	Field emission scanning electron microscope (FESEM)	33
3.2.5	Energy Dispersive X-ray (EDX) spectroscopy	34
3.2.6	Ultraviolet-visible (UV-Vis) Spectroscopy	34
3.3	Procedures	35
3.3.1	Pre-treatment of Screen printed carbon Electrode (SPCE)	35
3.3.2	Electrochemical optimization of different sizes of AuNPs	35
3.3.3	Voltammetric transduction	35
3.3.4	Modification of Screen printed carbon electrodes (SPCEs)	35
3.3.4.1	Electrode modification using Layer-by-Layer self-assembly approach	36
3.3.4.2	Electrochemical characterization for the active surface area determination of bare and modified electrodes	37
3.3.5	Optimization of the working conditions for modified electrode in quinoline	37
3.3.5.1	Effect of bilayers of nanocomposite surface modifiers on SPCEs	38
3.3.5.2	Influence of supporting electrolytes	38
3.3.5.3	Consequence of pH	39
3.3.5.4	Accumulation potential study	39
3.3.5.5	Accumulation time study	39
3.3.5.6	Scan rate study	39

3.3.6	Electrochemical characterization of Developed electrode for quinoline detection	40
3.3.6.1	Repeatability study	40
3.3.6.2	Reproducibility study	40
3.3.6.3	Stability study	40
3.3.6.4	Interference study	40
3.3.6.5	Sensitivity study	41
3.3.6.6	Recovery study	41
3.3.7	Characterization of developed electrode in healthy and artificially inoculated samples (Roots and leaves)	41
3.3.8	Characterization of developed electrode in healthy and Ganoderma treated on-site leaves samples	42
<b>4</b>	<b>RESULTS AND DISCUSSIONS</b>	<b>43</b>
4.1	Physicochemical Characterization of AuNPs, pMWCNTs, aMWCNTs, ChTSNPs-aMWCNTs	43
4.1.1	Zetasizer nano series	43
4.1.2	UV-Visible Spectrophotometry	45
4.1.3	Fourier Transform Infrared (FT-IR) spectroscopy	46
4.1.4	Energy Dispersive X-ray (EDX) Spectroscopy	48
4.2	Morphological Characterization of MWCNTs, AuNPs-aMWCNTs	49
4.2.1	Field emission scanning electron microscopy (FESEM)	49
4.3	Characterization of bare and modified bilayer electrodes	51
4.3.1	FESEM	51
4.4	Electrochemical characterization of different sizes of AuNPs	52
4.5	Electrochemical characterization of bare and modified SPCEs	54
4.5.1	aMWCNTs/SPCE, AuNPs/SPCE, and ChTSNPs/SPCE electrodes	54
4.5.2	AuNPs-aMWCNTs/SPCE and ChTSNPs-aMWCNTs/SPCE electrodes	55
4.5.3	ChTSNPs-aMWCNTs/ AuNPs-aMWCNTs/SPCE (BL1/SPCE) electrode	56
4.6	Electrochemical optimization of bilayer modified SPCEs for Quinoline Detection	56
4.6.1	Effect of thickness of bilayer of nanocomposite films of surface modifiers	56
4.6.2	Determination of effective electro-active surface area, $A_{eff}$ (cm <sup>2</sup> )	59
4.6.3	Influence of supporting electrolytes	61
4.6.4	pH study	62
4.6.5	Accumulation potential study	63
4.6.6	Accumulation time study	64
4.6.7	Scan rates study	64
4.7	Electrochemical characterization of Developed electrode for Quinoline Detection	67
4.7.1	Repeatability study	67



4.7.2	Stability study	68
4.7.3	Reproducibility study	68
4.7.4	Sensitivity Study	69
4.7.5	Interference Study	71
4.7.6	Field application of the developed electrode	73
4.8	Electrochemical response of bare and modified electrodes in different media	75
4.9	Electrochemical characterization of modified electrode for secondary metabolites Detection in healthy and artificially inoculated oil palms	77
4.9.1	Characterization in Leaf extracts secondary metabolites (leaf-SMs)	77
4.9.1.1	Reproducibility study	77
4.9.1.2	Sensitivity Study	79
4.9.1.3	Interference study	80
4.9.2	Characterization in Root extracts secondary metabolites (root-SMs)	81
4.9.2.1	Reproducibility study	82
4.9.2.2	Sensitivity Study	83
4.9.2.3	Interference study	84
4.10	Electrochemical characterization of modified electrode for secondary metabolites detection in on-site plantation oil palms	85
4.10.1	Characterization in 6 months on-site plant extract secondary metabolites	85
4.10.1.1	Reproducibility study	85
4.10.1.2	Sensitivity Study	87
4.10.1.3	Selectivity study	88
<b>5</b>	<b>SUMMARY, CONCLUSIONS AND RECOMMENDATIONS</b>	<b>89</b>
5.1	Summary	89
5.2	Conclusions	91
5.3	Recommendations for future work	91
	<b>REFERENCES</b>	<b>93</b>
	<b>APPENDICES</b>	<b>106</b>
	<b>BIODATA OF STUDENT</b>	<b>128</b>
	<b>LIST OF PUBLICATIONS</b>	<b>129</b>

## LIST OF TABLES

Table		Page
2.1	Electroanalytical techniques and electrode materials used in the detection and determination of secondary metabolites	16
3.1	Chemicals and reagents utilized in this study	24
4.1	Particle size distribution of synthesized AuNPs from Zetasizer Nano series using DLS technique	45
4.2	EDX elemental results for carbon nanotubes and nanocomposites	49
4.3	Ipa and Epa for repeatedly determined 10.0M Quinoline at BL4/SPCE	67
4.4	Effect of developed electrode storage on the Ipa of 10.0 $\mu$ M quinolone	68
4.5	Electroanalytical approaches used to detect metabolites with the electrode materials, sensitivity and LOD of the method used	71
4.6	Determination of quinoline in oil palm leaf extract secondary metabolites of healthy (Samples 1, 2, 3) and infected (Samples 4, 5, 6) trees	73
4.7	Determination of quinoline in oil palm root extract secondary metabolites of healthy (Samples 1, 2, 3) and infected (Samples 4, 5, 6) trees	74
4.8	Oxidation anodic peak current and potential in various media at bare and modified electrodes	77

## LIST OF FIGURES

Figure		Page
2.1	Molecular structure of Quinoline	14
2.2	(a) Electrodes in ECS. (b) Scheme of typical electrochemical sensor	17
2.3	A screen printed carbon electrode used in this study made by MIMOS, berhad Sdn	20
2.4	A Layer-by-Layer technique with gold nanoparticles-multi-walled carbon nanotube composite (AuNPs-aMWCNTs) and chitosan- multi-walled carbon nanotube composite (ChTSNPs-aMWCNTs)	21
3.1	Laboratory schematic representation of LBL self-assembly for the newly developed electrode	37
4.1	A plot of Mean volume (%) against diameter (nm) for PSD of synthesized AuNPs, 30/0.5 AuNPs (curve a), 30/0.4 AuNPs (curve b), 30/0.6 AuNPs (curve c), 30/0.2 AuNPs (curve d), 30/0.3 AuNPs (curve e)	44
4.2	UV- visible absorption spectra of spherical AuNPs prepared with different amounts of 1 wt.% trisodium citrate solution and fixed amount of 0.01% (wt/vol %) gold salt solution: 30/0.3 (a), 30/0.2(b), 30/0.4(c), 30/0.5(d), and 30/0.6(e) ml	46
4.3	FT-IR spectral of pristine MWCNTs (A); activated MWCNTs (B) and ChTSNPs-aMWCNTs (C) showing the various functional groups present in each sample	47
4.4	EDX spectral for pMWCNTs (A), activated MWCNTs (B) and AuNPs-aMWCNTs (C) exhibiting the compositional elements in each sample	48
4.5	FESEM micrographs of pMWCNTs (A), aMWCNTs (B), AuNPs-aMWCNTs (C) showing the surface morphology of various MWCNTs	50
4.6	The FESEM images of bare electrode, Bare/SPCE (A); modified electrodes, BL1/SPCE (B); BL4/SPCE (C) displaying the surface topography of each electrode	52
4.7	Cyclic voltammograms of 30/0.2AuNPs/SPCE (A), 30/0.3AuNPs/SPCE (B), 30/0.4AuNPs/SPCE (C), 30/0.5AuNPs/SPCE (D), 30/0.6 AuNPs/ SPCE (E) and overlay forms of all the voltammograms for	

	all the different sizes of AuNPs/SPCE in comparison with the bare/SPCE (F) in 1mM $K_3Fe(CN)_6$ /0.10 M KCl at a scan rate of 0.05 V/s	53
4.8	Comparing cyclic voltammograms of singly modified SPCEs and bare/ SPCE (A), compositely modified SPCEs and bare/SPCE (B) and BL1/ SPCE and bare/SPCE (C) in 1mM $K_3Fe(CN)_6$ /0.10 M KCL at 0.05 V/s	55
4.9	Cyclic voltammograms of bare/SPCE, BL1/SPCE, BL2/SPCE, BL3/SPCE BL4/SPCE, BL5/SPCE and BL6/SPCE in 1.0 mM $K_3Fe(CN)_6$ and 0.10 M KCl (A) and in buffered 10 $\mu$ M Quinoline (B) at a scan rate of 0.05 $Vs^{-1}$	57
4.10	Cyclic voltammograms and plots of peak current (A) against square root of scan rate ( $Vs^{-1}$ ) <sup>1/2</sup> (0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.09, 0.10 $Vs^{-1}$ ) of (A) bare/SPCE (B) BL1/SPCE (C) BL4/SPCE in 1.0 mM $K_3Fe(CN)_6$ and 0.1 M KCl	60
4.11	CVs of BL4/SPCE in 10 $\mu$ M quinoline with different buffers at scan rate 0.05 $Vs^{-1}$ (A), a plot of average peak current versus supporting electrolytes (B)	62
4.12	A plot of mean peak current, $I_{pa}$ ( $\mu$ A) [replicates (n = 3)] against pH showing the $I_{pa}$ of 10.0 $\mu$ M quinoline in 0.20M citrate buffer at scan rate 0.05 $Vs^{-1}$	63
4.13	Schematic representation of the oxidation of quinoline in acidic medium	63
4.14	Effect of accumulation potential on the anodic oxidation peak current of 10.0 $\mu$ M quinoline(A), Influence of accumulation time on the anodic oxidation peak current of 10.0 $\mu$ M quinoline (B) [Error bar represents the standard deviation of triplicate measurements (n = 3)]	64
4.15	CVs of 10.0 $\mu$ M quinoline at BL4/SPCE in scan rate 0.01 – 0.10 V/s (A), Plot of anodic peak current versus scan rate (B), Plot of log $I_{pa}$ versus log scan rate (C), Plot of $E_{pa}$ versus log scan rate (D) Error bar represents the standard deviation of triple measurements (n = 3)	66
4.16	Cyclic voltammograms of reused BL4/SPCE in 0.20 M citrate buffer (pH 5.5) at scan rate 0.06V/s, -0.52 V and 180s	67
4.17	CVs of seven independently produced BL4/SPCEs in 10.0 $\mu$ M quinoline (A); a plot of anodic peak current versus regenerated BL4/SPCEs (B)	69
4.18	Linear sweep voltammograms of BL4/SPCE in different concentrations of Quinoline in 0.2 M CBS (pH 5.5) Inset a: LSV of	

	0.0004 - 0.001 ( $\mu\text{M}$ ) Quinoline, Inset b: LSV of 0.002 - 0.006 ( $\mu\text{M}$ ) Quinoline, Inset c: linear relationship of $I_{pa}$ and concentrations	70
4.19	Cyclic voltammograms of 10.0 $\mu\text{M}$ quinoline in the presence of interfering species, inorganics A (i) and organics B (i) at BL4/SPCE under optimum conditions and the plots of percent interference against interfering species, inorganics A (ii) and organics B (ii)	72
4.20	Cyclic voltammograms of bare electrode (curve, a) and modified electrode (curve, b) in 100ppm leaf-SMs (A), 100ppm root-SMs (B) and 0.20 M CBS (pH 5.5), 10.0 $\mu\text{M}$ quinoline and 0.10 M methanol (C); Arrow direction indicates increase in $I_{pa}$ (a b)	75
4.21	LSVs of five (5) independently produced BL4/SPCEs in 100ppm leaf- SMs 14DHLSMs (A <sub>i</sub> ), 14DILSMs (B <sub>i</sub> ), 30DHLSMs (C <sub>i</sub> ), 30DILSMs D <sub>i</sub> ); A plot of anodic peak current versus regenerated BL4/SPCEs 14DHLSMs (A <sub>i</sub> ), 14DILSMs (B <sub>i</sub> ), 30DHLSMs (C <sub>i</sub> ), 30DILSMs D <sub>i</sub> )	78
4.22	Linear sweep voltammograms of BL4/SPCE in different concentrations of leaf-SMs 14DHLSMs (A <sub>i</sub> ), 14DILSMs (B <sub>i</sub> ), 30DHLSMs (C <sub>i</sub> ), 30DILSMs D <sub>i</sub> ), and linear relationship of anodic peak current, $I_{pa}$ and concentrations 14DHLSMs (A <sub>i</sub> ), 14DILSMs (B <sub>i</sub> ), 30DHLSMs (C <sub>i</sub> ), 30DILSMs D <sub>i</sub> )	79
4.23	A plot of percent interference in anodic peak current ( $I_{pa}$ ) response of leaf-SMs 14DHLSMs (A), 14DILSMs (B), 30DHLSMs (C) and 30DILSMs (D) against interfering species at modified electrode under optimized conditions (-0.53 V, 180 s, 0.06 V/s)	81
4.24	LSVs of five (5) independently produced BL4/SPCEs in 100ppm root- SMs 14DHRSMs (A <sub>i</sub> ), 14DIRSMs (B <sub>i</sub> ), 30DHRSMs (C <sub>i</sub> ), 30DIRSMs D <sub>i</sub> ); A plot of anodic peak current versus regenerated BL4/SPCEs 14DHRSMs (A <sub>ii</sub> ), 14DIRSMs (B <sub>ii</sub> ), 30DHRSMs (C <sub>ii</sub> ), 30DIRSMs D <sub>ii</sub> ) under optimized conditions (-0.53 V, 180 s, 0.06 V/s)	82
4.25	Linear sweep voltammograms of BL4/SPCE in different concentrations of root-SMs, 14DHRSMs (A <sub>i</sub> ), 14DIRSMs (B <sub>i</sub> ), 30DHRSMs (C <sub>i</sub> ), 30DIRSMs D <sub>i</sub> ), and linear relationship of anodic peak current, $I_{pa}$ and concentrations 14DHRSMs (A <sub>ii</sub> ), 14DIRSMs (B <sub>ii</sub> ), 30DHRSMs (C <sub>ii</sub> ), 30DIRSMs D <sub>ii</sub> ) under optimized conditions (-0.53 V, 180 s, 0.06 V/s)	83
4.26	A plot of percent interference in anodic peak current ( $I_{pa}$ ) response of root-SMs 14DHRSMs (A), 14DIRSMs (B), 30DHRSMs (C) and 30DIRSMs (D) against interfering species at modified electrode under optimized conditions (-0.53 V, 180 s, 0.06 V/s)	84

- 4.27 LSVs of six (6) independently produced BL4/SPCEs in 100ppm on-site leaf-SMs (6MHLSMs A<sub>i</sub> - 6MILSMs B<sub>i</sub>); A plot of anodic peak current versus regenerated BL4/SPCEs (6MHLSMs A<sub>ii</sub> - 6MILSMs B<sub>ii</sub>) 86
- 4.28 Linear sweep voltammograms of BL4/SPCE in different concentrations of on-site leaf-SMs [6MHLSMs (A<sub>i</sub>), 6MILSMs (B<sub>i</sub>)] and linear relationship of anodic peak current, I<sub>pa</sub> and concentrations [6MHLSMs (A<sub>ii</sub>), 6MILSMs (B<sub>ii</sub>)] 87
- 4.29 A plot of percent interference in anodic current response of on-site leaf-SMs 6MHLSMs (A), 6MILSMs (B) against interfering species at modified electrode under optimized conditions (-0.53 V, 180 s, 0.06 V/s) 88



## LIST OF ABBREVIATIONS

6MHLSMs	6 months healthy leaf secondary metabolites
6MILSMs	6 months infected leaf secondary metabolites
14DHLSMs	14 days healthy leaf secondary metabolites
14DHRSMs	14 days healthy root secondary metabolites
14DILSMs	14 days infected leaf secondary metabolites
14DIRSMs	14 days infected root secondary metabolites
30DHLSMs	30 days healthy leaf secondary metabolites
30DHRSMs	30 days healthy root secondary metabolites
30DILSMs	30 days infected leaf secondary metabolites
30DIRSMs	30 days infected root secondary metabolites
ABS	Acetate buffer solution
AdSV	Adsorptive stripping voltammetry
AMP	Amperometry
aMWCNTs	Activated multi-walled carbon nanotubes
Au	Gold
AuNPs	Gold nanoparticles
AuNPs- aMWCNTs	Gold nanoparticles- activated multi-walled carbon nanotubes
BBS	Borate buffer solution
BL	Bilayer
BL1/SPCE	Bilayer one screen printed carbon electrode
BSR	Basal stem rot
CaBS	Carbonate buffer solution
CBS	Citrate buffer solution

ChTSN	Chitosan
ChTSNP-aMWCNTs	Chitosan nanoparticles- activated multi-walled carbon nanotubes
cMWCNT	Carboxylated multi-walled carbon nanotubes
CNTs	Carbon nanotubes
CSSTM	Chemically selective scanning tunneling microscopy
CV	Cyclic voltammogram / Cyclic voltammetry
DLS	Dynamic light scattering
DVP	Differential pulse voltammetry
ECS	Electrochemical sensors
EDXS	Energy Dispersive X-Ray spectroscopy
ELISA	Enzyme-linked immunosorbent assay
Epa	Anodic peak potential
Epc	Cathodic peak potential
FDL	Functional Devices Laboratory
FESEM	Field emission electron microscopy
FPs	Fallen palms
<i>G.b</i>	<i>Ganoderma boninense</i>
GCE	Glassy carbon electrode
GSM	Ganoderma selective medium
IC	Indirect competitive assay
Ipa	Anodic peak current
Ipc	Cathodic peak current
issAP:-Ver.A:	Single stranded DNA
ITMA	Institute of Advanced Technology



ITO	Indium tin oxide electrode
Lac	Laccase
LbL	Layer-by-layer
LOD	Limit of detection
LOQ	Limit of quantification
LSV	Linear sweep voltammetry/ voltammogram
MBs	Magnetic beads
MOFs	Metal organic frameworks
mRNA	Molecular ribonucleic acid
NPs	Nanoparticles
PBS	Phosphate buffer solution
PCR	Polymerase chain reaction
PCS	Photon correlation spectroscopy
PDDA	Poly(diallyldimethylammonium chloride)
PEDOT	Poly (3,4-ethylene dioxythiophene)
PMs	Primary metabolites
pMWCNTs	Pristine Multi-walled carbon nanotubes
PSMs	Plant secondary metabolites
PSS	Poly(sodium 4-styrenesulfonate)
QELS	Quasi elastic light scattering
QY	Quinoline yellow
RSD	Relative standard deviation
SECM	Scanning electrochemical microscopy
SEM	Scanning electron microscopy
SMs	Secondary metabolites

SOPs	Standing oil palms
SPCEs	Screen printed carbon electrodes
SPEs	Screen printed electrodes
SWV	Square wave voltammetry
TEM	Transmission electron microscopy
TGA	Thermogravimetric analysis
UPM	Universiti Putra Malaysia
USR	Upper stem rot
UV-Vis	Ultraviolet-visible
$\beta$ -CD	$\beta$ -cyclodextrin



© COPYRIGHT UPM

# CHAPTER 1

## INTRODUCTION

### 1.1 Background of the study

The incapability of plants to successfully undergo translational motion has indeed limit their escaping potentials from invading living and non-living components of the ecosystem. Despite this fact, they are able to checkmate a number of attacks by brewing up highly functional biological defense weaponry, termed plant secondary metabolites (PSMs) or simply secondary metabolites (SMs). Examples of some selected SMs that have been studied include phenylpropanoid (López-Gresa *et al.*, 2011), glucosinolates (Pedras & Hossain, 2011), phytosterols (Ekade & Manik, 2014), eugenol and 1,8-cineole (Pavarini *et al.*, 2012), indole (Ahuja *et al.*, 2012). SMs are also referred to as volatile organics or volatile organic compounds (Pavarini *et al.*, 2012; Sankaran *et al.*, 2010), bioactive compounds (Ekade & Manik, 2014; Pavarini *et al.*, 2012), antimicrobial compounds (Ahuja *et al.*, 2012). They are, therefore, biochemical molecules capable of restraining the adverse environmental effects on the standing plants (López-Gresa *et al.*, 2011). SMs are low-molecular-mass biochemical species endowed with the natural propensity to guard the plants from all kinds of stressor factors emanating from the ecosystem (Ahuja *et al.*, 2012; Ekade & Manik, 2014; López-Gresa *et al.*, 2011; Pavarini *et al.*, 2012). These groups of compounds are known to significantly exhibit two opposing traits, which include being beneficial as well as being destructive (Pedras & Hossain, 2011).

However, the concern of this present study is on their protective responsibilities for the plant kingdom. SMs protect and guard the plants against devastatingly destructive agents like insects, pests, microorganisms (Ekade & Manik, 2014; López-Gresa *et al.*, 2011; Pedras & Hossain, 2011) and drought, salinity, erosion, humidity (Ekade & Manik, 2014; López-Gresa *et al.*, 2011). SMs employed in plant defense strategies are of two classes, phytoanticipins (constitutive) and phytoalexins (inducible) (Pedras & Hossain, 2011). The focal point of this study is phytoalexins. Phytoalexins are a combination of inter-related substances displaying efficient biochemical workforce against different disease-causing microbes, whose idea was conceived seventy years ago. They are also low molecular mass secondary metabolites bearing antimicrobial property and stress induced. Based on their effective check and impedence to the spread of pathogenic invasion, they are tagged disease resistance molecular markers or simply disease resistance markers (Ahuja *et al.*, 2012). Quite unfortunate, lots of crops are lost to these devastating, non-compromising pathogens (viral, bacterial or fungal) either on the fields or off the fields (storage). The economic importance of these pathogens, no doubt has precipitated a universal headache, the world over (Ahuja *et al.*, 2012). Researchers, scientists and other stakeholders are on their toes looking for the right remedy. In this regard, the oil palms in Malaysia and Indonesia have suffered many losses from the pathogenic fungus, *Ganoderma boninense* (G.b) trading a disease called basal

stem rot (BSR) disease. According to Nusaibah and co-workers (2016), quinoline is SMs produced in oil palms at the site of the attack by G.b within 24hours.

This tree plant species belong to the genus *Elaeis* and family *Palmae* (Bivi *et al.*, 2010). Its scientific nomenclature is *Elaeis guineensis* Jacq. The genesis of this plant is from West African sub-continent, the Gulf of Guinea to be precise.

The Portuguese transported oil palm to Brazil in the 15th century, while the Dutch catapulted it to Indonesia in the 18th century and in 1878, it was embraced as a decorative plant in Malaya (Naher *et al.*, 2013). It was also reported that the British introduced the oil palm to south-east Asia in the early part of the 1870s (Hushiarian *et al.*, 2013). The fruit of the oil palm is a drupe, with three-layered pericarp comprising of the outer skin (exocarp), outer pulp bearing palm oil (mesocarp) and inner hardcore circumscribing the kernel bearing the kernel oil (endocarp) (Naher *et al.*, 2013). The oil palm trees commence fruit procreation after thirty months of cultivation and are blessed with a proactive lifespan of twenty to thirty years (Hushiarian *et al.*, 2013). A quarter of a hectare of land is required to yield a tone of palm oil. They are topmost edible oil generating crop globally (Hushiarian *et al.*, 2013; Naher *et al.*, 2013; Tee *et al.*, 2013) Other products obtainable from oil palms include biofuels (Naher *et al.*, 2013) , kernel oil, kernel cake, oleochemicals, biodiesel and other up-stream products (Ho & Tan, 2015). Reference to the foreign exchange earnings, Indonesia and Malaysia are the leading nations in palm oil exportation around the globe (Ho & Tan, 2015; Hushiarian *et al.*, 2013) as over several billions of US dollars were pocketed by both countries in the year 2012. According to Bivi and co-workers (2010), oil palm is known as the Golden crop of Malaysia as it stands out as the only crop used as means of poverty alleviation in the country. However, this economic crop is faced with a great threat of drastic low yield which may result to proportionate low foreign exchange income on the part of exporting countries and subsequently low-quality products by the importing countries.

The oil palm is challenged by a pathogenic fungus, called *Ganoderma boninense*, causative organism for basal stem rot (BSR) disease (Bivi *et al.*, 2010; Ho & Tan, 2015; Hushiarian *et al.*, 2013; Naher *et al.*, 2013; Zain *et al.*, 2013; Nusaibah *et al.*, 2016; Tee *et al.*, 2013).

This destructively devastating disease had killed and still killing lots of oil palms, yet to be remedied. Economic losses to the tune of millions of US dollars have largely been reported as result of the lethal BSR disease (Hushiarian *et al.*, 2013; Nusaibah *et al.*, 2016; Zain *et al.*, 2013).

The uncompromising pathogenic fungus, launching an assault on oil palms specifically in Malaysia and Indonesia and other tropical regions of Thailand, Papua New Guinea and Africa have been uncovered to be *G. boninense* (Kok *et al.*, 2013).

It is of the genus *Ganoderma* Karst and classified as a higher fungus. It is a member of the large family of *Ganodermataceae*, in the order *Aphyllphorales*, in subclass *Hymenomycetes* and in the class *Basidiomycetes* (Idris *et al.*, 2000). It has been affirmed that it is the causative organism for Upper stem rot (USR) and basal stem rot (BSR) diseases of oil palms (Rakib *et al.*, 2014a, 2014b). Out of the 15 *Ganoderma* species responsible for stem rot disease world over, 7 are documented in Malaysia, as follows *G. boninense*, *G. zonatum*, *G. miniatocinctum* and *G. tornatum*, *G. applanatum*, *G. chalceum*, *G. lucidum* and *G. pseudoferreum*. The first three are highly pathogenic, while *G. tornatum* is non-pathogenic. The format of disease spread is precipitated on two pathways, namely, root contact and basidiospore (Rakib *et al.*, 2014b). BSR is propagated in a very latent mode. The exploit of *G. boninense* being necrotrophic is not easily deciphered at the on-set of its attack on oil palms. In the process, the fungi destroyed cell wall barrier of the oil palms through the activity of the destructive enzymes released by the fungal pathogens, thus enabling the formation of regular infection hyphae within the oil palms. Mostly, the basidiomata are revealed after the demise of the cell of the oil palms.

The mycelia of these fungi often sensitize the protective defense response in oil palms (Nusaibah *et al.*, 2016).

Both youthful and aged standing palms are prone to BSR with high severity documented in old palms (Bivi *et al.*, 2010; Kok *et al.*, 2013). Some of the symptoms include deterioration of bole, closed spear leaves, fractured frond petioles, and appearance of basidiomata around the stem, root or bole regions (Kok *et al.*, 2013), eventuality paving a way to the killing the oil palm in the process. Eighty percent of palm stands can be laid to rest by BSR, usually midway through their productive lifespan (Bivi *et al.*, 2010; Chong *et al.*, 2012; Priwiratama & Susanto, 2014). A variety of control measures have been undertaken, such as, agrophysical methods like soil drenching, crop rotation, clean clearing, chemical methods like use of fungicides, biological methods like use of endophytic bacteria (Bivi *et al.*, 2010; Chong *et al.*, 2012) or saprophytic fungi (Priwiratama & Susanto, 2014), series of molecular approaches, lignin content alteration in oil palm root. Summarily, till date, no certified cure has been adduced to *G. boninense* as most of the controls are limited in their efficacies.

Based on the foregoing, the sustainability of oil palms is greatly at stake, as no known technique has been discovered to tackle the devastating arsenal of *G. boninense* till date (Chong *et al.*, 2011). The urgent need for a proactive detective method is therefore non-negotiable. Methods employed so far include the use of *Ganoderma* selective medium (GSM) and polymerase chain reaction (Chong *et al.*, 2011). Some of these methods are not efficient enough or are technically deficient in the application. Santoso *et al.*, (2011) discovered and mapped oil palms with BSR disease using strongly resolving QuickBird satellite imagery in oil palm plantation located in North Sumatra, Indonesia. This imaging technique is a kind of remote sensing method. GanoSken Tomography Technology comprising of sound sensors

and tomography software has also been utilized to monitor deterioration in tree plants.

Though effective, it is not time-friendly. Considering the large expanse of oil palms plantation a relatively simple, easy to handle and cost-effective detection equipment will be a better option. On this premise the combination of the multispectral and thermal camera had been successfully implemented by Khairunniza-bejo *et al.*, (2015) to investigate BSR diseased and BSR disease free oil palms. These technologies are still deficient in that they only detect the BSR at an advanced stage, which left the planter no option of any curative approach. Anyway, the search for early diagnostic method continues. Kandan *et al.*, (2010) applied a serological method based on enzyme-linked immunosorbent assay (ELISA) with polyclonal and /or monoclonal antibodies to detect Ganoderma disease. The shortcoming of the ELISA technique is cross-reactivity among different Ganoderma spp and other saprophytic fungi, give rise to low sensitivity and specificity. Lelong *et al.*, (2010) made use of hyperspectral reflectance spectroscopy to uncover BSR disease and monitor different stages of the pathogen infection on oil palms. However, the method is limited in that canopy optical properties arising from the white rot fungus is yet to be ratified. Rakib *et al.*, (2014b) employed geographical information system and geostatistics to monitor the pattern of distribution of the disease in the field. Nasim *et al.*, (2010) used PCR method to detect *Ganoderma* disease, utilizing powerful genetic marker, internal transcribed spacers region of the ribosomal-DNA. A number of issues arose in this approach ranging from Inefficiencies of different DNA polymerases, the presence of PCR inhibitors in the sample matrix and variation in the performance of PCR thermal cyclers.

Unarguably, the search for quick, simple, sensitive, portable, cheap and point of care detective method is on-going. A look at electrochemical approaches may herald the much-needed antidote. The only limiting factor is the absolute dearth of studies on the electrochemical detection of *G. boninense* either laboratory wise or field wise.

However, Dutse and co-workers, (2012) had eventually detected *G. boninense* using electrochemical-based DNA approach. The advent of nanotechnology aided the team in mixing relevant nanomaterials to obtain nanocomposites of salient properties like improved biocompatibility, stability, penetration capacity, area to volume ratio, electrical conductance, catalytic ability, biocatalyst loading potential and structural feature. The team employed PEDOT-PSS blended with metallic silver nanoparticles to modify the electrode and applied a novel ruthenium complex as intercalating material for interaction and detection of *G. boninense* (Dutse & Yusof, 2011). Based on the success recorded by this team, it is hoped that pure electrochemical sensor could as well provide a functional approach for early detection of *G. boninense*.

It is a truism that many of the chemical and related research laboratories cannot do without the tool called electrochemical methods. A typical example of this method is the voltammetric method. These approaches have been successfully employed in

basic studies of redox processes to analyze reaction mechanisms. Studies involving the kinetics and thermodynamics of ion and electron transfer processes have equally been conducted via these essential tools. Courses on adsorption and crystallization events at the surfaces of an electrode are inclusive. The broad utilization of these techniques is ascribed to the following; inexpensive instrumentation, the excellent sensitivity with a lengthy linear range of concentration, quick response period and concurrent detection and determination of many analytes (Gulaboski & Pereira, 2008).

## 1.2 Problem statement

*Ganoderma boninense* is the principal pathogenic fungus responsible for the spread of BSR in every developmental stage of the oil palms, commencing from the nursery seedlings to adults on the plantation fields. BSR can be transmitted through airborne basidiospores and mycelia of *G. boninense* during root to root contact in oil palms fields. The dreadful arsenals from the fungi easily disintegrate cell wall components, the strongly protective lignin inclusive. BSR not only promulgates retarded oil yield, it also causes the stands to collapse thus causing severe economic loss to the oil palm industry. 1928 was the year BSR came to be recognized in the country (Bivi *et al.*, 2010). The ensuing economic loss accrued to the tune of millions US dollars, thereby increasing the anxiety of stakeholders in the oil palm sector (Hushiarian *et al.*, 2013). The worry of the stakeholders is precipitated by the fact that the oil palm, is a globally acclaimed economic tree, being portrayed as one of the world's major sources of edible oil and a momentous precursor of biodiesel fuel. Therefore, there is dire need to protect the palms from extinction by detecting the devastating fungus at the onset of infection in oil palms. Thus, this study aimed at early detection of *G. boninense*.

Although *G. boninense* had been successfully identified as the pathogenic fungi causing BSR in oil palms for the past few decades, it remained to be the most serious problem in many areas in Malaysia and Indonesia. Unfortunately, there is no single reliable procedure for curtailing the spread of this disease. Unarguably, a limiting factor in controlling the BSR disease is the lack of reliable diagnostic technique(s) for early diagnosis. This implies the severity of BSR detection at its earliest stage. The findings from previous methods produced little success.

Molecular method like polymerase chain reaction (PCR) is used to detect the presence of bacteria in a sample that might be negative in a routine culture and serology due to the trace amount. In the process, DNA fragments are isolated from a specific region of a genomic DNA. However, many uncertainties from PCR results were observed with regard to whether or not bacterial DNA detected is alive or dead. Inclusive are inefficiencies of different DNA polymerases, the presence of PCR inhibitors in the sample matrix and variation in the performance of PCR thermal cyclers (Chong *et al.*, 2011). The use of blotting hybridization requires coupling with a PCR procedure that makes it laborious and requires being operated by an expert.

On the other hand, enzyme-linked immunosorbent assay (ELISA) is another promising approach faced with cross-reactivity among different *Ganoderma spp* and other saprophytic fungi resulting in low sensitivity and specificity (Kandan *et al.*, 2010). Imagery techniques have equally been advanced such as GammaScorpion: mobile gamma-ray tomography system by Hamidon & Mukhlisin (2014). The challenges being Cumbersomeness, non-portability, long scanning period and harmful radiation effect of gamma rays emitted by the instrument. In the same vein, hyperspectral reflectance technique was employed by Khairunniza-bejo *et al.*, (2015) in oil palm plantation in Malaysia. The modus operandi of HSI can be summarized as follows, 50 – 300 images are acquirable in a given hyperspectral image cube at different wavelengths with the resolution of 1 – 10nm from a specific wavelength region. The problem encountered is the inability to image thick foliage. Anyway, research is on-going in various areas to factor out the much-needed technique to combat the menace of this pathogenic fungus on standing palms.

Despite the fact that the electrochemical pathway is convenient, simple, selective, speedy and environmentally friendly the challenge remains the development and production of a smart and efficient sensing system for the early detection of *G. boninense*, with the potential of providing a point of care service.

The current miniaturization of electrochemical detection with the advent of screen printing technology for environmental analysis may be effective in this regard. The current advancement in the technology of electrochemical sensor may be the answer to this problem. Thus, the development of a novel sensor is hereby embarked upon to facilitate the early detection of *G. boninense* in infected oil palms.

However, the present study hopes to advance an indirect approach in conciliating early detection of *G. boninense* infections in oil palms, by considering secondary metabolites (SMs) excreted during the attack of the lethargic fungi. The detection of these SMs is crucial to the early detection of *G. boninense*, the causative fungal of basal stem rot (BSR) disease in oil palms. These metabolites are often excreted by the oil palms as defensive soldiers whenever the pathogenic fungi launch a morbid attack on the economic trees. One of the SMs produced by the plants during such invasion within 24 hours of attack is quinoline.

### 1.3 Objective of the study

The focal point of this study is to develop a simple, efficient and cheap electrode system for the detection of secondary metabolites in *G. boninense* infected oil palms. This implies an indirect early detection of the troublesome fungus. The detection system consists of screen printed carbon electrode modified with nanohybrid materials, namely gold nanoparticles activated multi-walled carbon nanotubes (AuNPs-aMWCNTs) and chitosan activated multi-walled carbon nanotubes (ChTSNPs-aMWCNTs). The analytical performance of the developed system in the



detection of quinoline and secondary metabolites in both healthy and *G.boninense* infected leaf and root extracts of oil palm will be investigated.

However, the general objective of this study is to develop an electrochemical sensor for the detection of secondary metabolite quinoline in *G. boninense* infected oil palms. The specific objectives of the present study are:

- i. To prepare and characterize surface modifiers such as the gold nanoparticle, nanocomposites (AuNPs-aMWCNTs and ChTSNPs-aMWCNTs) and to modify the electrode and optimize the operating conditions of the modified electrode
- ii. To carry out the electrochemical characterization of the developed electrode for quinoline detection using cyclic voltammetry
- iii. To carry out the electrochemical characterization of the sensor for secondary metabolite detection in roots and leaves extracts from healthy and infected oil palms using linear sweep voltammetry
- iv. To verify the on-site efficacy of the sensor by carrying out the characterized detection using developed electrode in the main plantation 6-months old oil palms

## REFERENCES

- Ahmad, N. M. (2007). Chapter 12 Quinolines. *Tetrahedron Organic Chemistry Series*, 26(2), 511–539.
- Ahmad, T., & Khan, W. (2013). Size Variation of Gold Nanoparticles Synthesized Using Tannic Acid in Response to Higher Chloroauric Acid Concentrations. *World Journal of Nano Science and Engineering*, 3(3), 62–68.
- Ahuja, I., Kissen, R., & Bones, A. M. (2012). Phytoalexins in defense against pathogens. *Trends in Plant Science*, 17(2), 73–90.
- Alkhatib, M.F., Mohamed, E.S., Mirghani, I. Y., & Qudsieh, I. A. F. H. (2010). Immobilization of CTS on CNT for Pb removal.pdf. kuala lumpur: *Asian Network for scientific information*. Retrieved from <http://ansinet.com>
- Amiri, M., Ghaffari, S., Bezaatpour, A., & Marken, F. (2012). Carbon nanoparticle-chitosan composite electrode with anion, cation, and neutral binding sites: Dihydroxybenzene selectivity. *Sensors and Actuators, B: Chemical*, 162(1), 194–200.
- Baetz, U., & Martinoia, E. (2014). Root exudates: The hidden part of plant defense. *Trends in Plant Science*, 19(2), 90–98.
- Bataglion, G. A., Da Silva, F. M. A., Eberlin, M. N., & Koolen, H. H. F. (2015). Determination of the phenolic composition from Brazilian tropical fruits by UHPLC-MS/MS. *Food Chemistry*, 180, 280–287.
- Bednarek, P. (2012). Chemical Warfare Or Modulators Of Defence Responses - The Function Of Secondary Metabolites In Plant Immunity. *Current Opinion in Plant Biology*, 15(4), 407–414.
- Bivi, M. R., Farhana, M. S., Khairulmazmi, A., & Idris, A. (2010). Control of ganoderma boninense: A causal agent of basal stem rot disease in oil palm with endophyte bacteria in vitro. *International Journal of Agriculture and Biology*, 12(6), 833–839.
- Bonel, L., Vidal, J. C., Duato, P., & Castillo, J. R. (2011). An electrochemical competitive biosensor for ochratoxin A based on a DNA biotinylated aptamer. *Biosensors and Bioelectronics*, 26(7), 3254–3259.
- Bourgaud, F., Gravot, A., Milesi, S., & Gontier, E. (2001). Production of plant secondary metabolites : a historical perspective, *Plant Science*, 161, 839–851.
- Buang, N. A., Fadil, F., Majid, Z. A., & Shahir, S. (2012). Characteristic of mild acid functionalized multiwalled carbon nanotubes towards high dispersion with low structural defects. *Digest Journal of Nanomaterials and Biostructures*, 7(1), 33–

- Chen, J. Y., Zhou, P. J., Li, J. L., & Li, S. Q. (2007). Depositing Cu<sub>2</sub>O of different morphology on chitosan nanoparticles by an electrochemical method. *Carbohydrate Polymers*, 67(4), 623–629.
- Chikae, M., Idegami, K., Kerman, K., Nagatani, N., Ishikawa, M., Takamura, Y., & Tamiya, E. (2006). Direct fabrication of catalytic metal nanoparticles onto the surface of a screen-printed carbon electrode. *Electrochemistry Communications*, 8(8), 1375–1380.
- Chong, K.P., Lum, M.S., Foong, C.P., Wong, C.M.V.L., Atong, M. & Rossal, S. (2011). First Identification of *Ganoderma boninense* isolated from Sabah based on PCR and sequence homology. *African Journal of Biotechnology*, 10(66), 14718–14723.
- Chong, K. P., Markus, A., & Rossall, S. (2012). The susceptibility of different varieties of oil palm seedlings to *Ganoderma boninense* infection. *Pakistan Journal of Botany*, 44(6), 2001–2004.
- Choudhry, N. A., Kampouris, D. K., Kadara, R. O., Jenkinson, N., & Banks, C. E. (2009). Next generation screen printed electrochemical platforms: Non-enzymatic sensing of carbohydrates using copper(ii) oxide screen printed electrodes. *Analytical Methods*, 1(3), 183.
- Codognoto, L., Zuin, V. G., De Souza, D., Yariwake, J. H., Machado, S. A. S., & Avaca, L. A. (2004). Electroanalytical and chromatographic determination of pentachlorophenol and related molecules in a contaminated soil: A real case example. *Microchemical Journal*, 77(2), 177–184.
- Cooper, R. M., Flood, J & Rees, R. W. (2011). *Ganoderma boninense* in Oil Palm Plantations : Current Thinking on Epidemiology , Resistance and Pathology. *The Planter*, 87(1024), 515–526.
- Datsyuk, V., Kalyva, M., Papagelis, K., Parthenios, J., Tasis, D., Siokou, A., Kallitsis, I., & Galiotis, C. (2008). Chemical oxidation of multiwalled carbon nanotubes. *Carbon*, 46(6), 833–840.
- Davies, T. J., Hyde, M. E., & Compton, R. G. (2005). Nanotrench arrays reveal insight into graphite electrochemistry. *Angewandte Chemie - International Edition*, 44(32), 5121–5126.
- De Coninck, B., Timmermans, P., Vos, C., Cammue, B. P. A., & Kazan, K. (2015). What lies beneath: Belowground defense strategies in plants. *Trends in Plant Science*, 20(2), 91–101.
- de Oliveira-Roberth, A., Santos, D. I. V, Cordeiro, D. D., Lino, F. M. de A., Bara, M. T. F., & Gil, E. de S. (2012). Voltammetric determination of Rutin at

Screen-Printed carbon disposable electrodes. *Central European Journal of Chemistry*, 10(5), 1609–1616.

- Decher, G. (2012). Layer-by-Layer Assembly (Putting Molecules to Work). *Multilayer Thin Films: Sequential Assembly of Nanocomposite Materials: Second Edition*, 1, 1–21.
- Delgado-Zamarreño, M. M., Bustamante-Rangel, M., Sánchez-Pérez, A., & Carabias-Martínez, R. (2004). Pressurized liquid extraction prior to liquid chromatography with electrochemical detection for the analysis of vitamin e isomers in seeds and nuts. *Journal of Chromatography A*, 1056(1–2 SPEC.ISS.), 249–252.
- Di, J., & Zhang, F. (2003). Voltammetry determination of trace manganese with pretreatment glassy carbon electrode by linear sweep voltammetry. *Talanta*, 60(1), 31–36.
- Diaconu, M., Litescu, S. C., & Radu, G. L. (2010). Chemical Laccase – MWCNT – chitosan biosensor — A new tool for total polyphenolic content evaluation from in vitro cultivated plants, *Sensors and Actuators B*, 145, 800–806.
- Dontsova, E. A., Zeifman, Y. S., Budashov, I. A., Eremenko, A. V., Kalnov, S. L., & Kurochkin, I. N. (2011). Screen-printed carbon electrode for choline based on MnO<sub>2</sub> nanoparticles and choline oxidase/polyelectrolyte layers. *Sensors and Actuators, B: Chemical*, 159(1), 261–270.
- DropSens (2017). Ref. STAT8000. Product New Bronchure. [http://www.dropsens.com/en/pdfs\\_productos/new\\_brochures/stat8000.pdf](http://www.dropsens.com/en/pdfs_productos/new_brochures/stat8000.pdf). Date visited 28/2/2017
- Dutse, S. W., & Yusof, N. A. (2011). ). DNA-based Biosensor for Detection of Ganoderma boninense , an Oil Palm Pathogen Utilizing Newly Synthesized Ruthenium Complex [ Ru ( phen ) 2 ( qtpy ) ] 2 + Based on a PEDOT-PSS / Ag Nanoparticles Modified Electrode. *Sensors*, 8, 5754–5768.
- Dutse, S. W., Yusof, N. A., Ahmad, H., Hussein, M. Z., & Zainal, Z. (2012). An electrochemical DNA biosensor for ganoderma boninense pathogen of the oil palm utilizing a new ruthenium complex, [ru(dppz)2(qtpy)]Cl<sub>2</sub>. *International Journal of Electrochemical Science*, 7(9), 8105–8115.
- Ekade, P. P., & Manik, S. R. (2014). Investigations on Secondary Metabolites in Different Parts of Radermachera xylocarpa using GC-MS. *Journal of Pharmacognosy and Phytochemistry JPP*, 2(6), 39–47.
- Fan, Z., Ho, J. C., Takahashi, T., Yerushalmi, R., Takei, K., Ford, A. C., Chueh, Y., & Javey, A. (2009). Toward the Development of Printable Nanowire Electronics and Sensors. *Advanced Materials*, 21(37), 3730–3743.

- Fanjul-Bolado, P., Hernández-Santos, D., Lamas-Ardisana, P. J., Martín-Pernía, A., & Costa-García, A. (2008). Electrochemical characterization of screen-printed and conventional carbon paste electrodes. *Electrochimica Acta*, 53(10), 3635–3642.
- Farghaly, O. A., Hameed, R. S. A., Alhakeem, A., & Abu-Nawwas, H. (2014). Analytical Application Using Modern Electrochemical Techniques. *Int. J. Electrochem. Sci*, 9, 3287–3318.
- Fee, C. G. (2011). Management of Ganoderma Diseases in Oil Palm Plantations . *The Planter*, 87(39), 325–339.
- Feng, S., Zhang, Y., Zhong, Y., Li, Y., & Li, S. (2014). Simultaneous determination of hydroquinone and catechol using covalent layer-by-layer self-assembly of carboxylated-MWNTs. *Journal of Electroanalytical Chemistry*, 733, 1–5.
- Fu, L., Yu, S., Thompson, L., & Yu, A. (2015). Development of a novel nitrite electrochemical sensor by stepwise in situ formation of palladium and reduced graphene oxide nanocomposites. *RSC Adv.*, 5(50), 40111–40116.
- Gao, Y., Wang, M., Yang, X., Sun, Q., & Zhao, J. (2014). Rapid detection of quinoline yellow in soft drinks using polypyrrole/single-walled carbon nanotubes composites modified glass carbon electrode. *Journal of Electroanalytical Chemistry*, 735, 84–89.
- Gao, Y., Wu, L., Zhang, K., Xu, J., & Lu, L. (2015). Electroanalytical method for determination of shikonin based on the enhancement effect of cyclodextrin functionalized carbon nanotubes. *Chinese Chemical Letters*, 26(5), 613–618.
- Ghalkhani, M., & Shahrokhian, S. (2010). Application of carbon nanoparticle/chitosan modified electrode for the square-wave adsorptive anodic stripping voltammetric determination of Niclosamide. *Electrochemistry Communications*, 12(1), 66–69.
- Ghamouss, F., Tessier, P.-Y., Djouadi, A., Besland, M.-P., & Boujtita, M. (2007a). Screen-printed carbon electrode modified on its surface with amorphous carbon nitride thin film: Electrochemical and morphological study. *Electrochimica Acta*, 52, 5053–5061.
- Ghamouss, F., Tessier, P. Y., Djouadi, M. A., Besland, M. P., & Boujtita, M. (2007b). Examination of the electrochemical reactivity of screen printed carbon electrode treated by radio-frequency argon plasma. *Electrochemistry Communications*, 9(7), 1798–1804.
- Ghosh, D., & Chattopadhyay, N. (2013). Gold nanoparticles: acceptors for efficient energy transfer from the photoexcited fluorophores. *Optics and Photonics Journal*, 3(March), 18–26.

- Govindhan, B. M., Adhikari, B. & Chen, A. (2014). Nanomaterials-based electrochemical detection of chemical contaminants. *RSC Adv.*, *11*(109), 54–59.
- Guan, W.-J., Li, Y., Chen, Y.-Q., Zhang, X.-B., & Hu, G.-Q. (2005). Glucose biosensor based on multi-wall carbon nanotubes and screen printed carbon electrodes. *Biosensors and Bioelectronics*, *21*(3), 508–512.
- Gulaboski, R., & Pereira, C. M. (2008). Electroanalytical techniques and instrumentation in food analysis. *Handbook of Food Analysis Instruments*, 379–402.
- Gupta, P. N. and Rather, G. M. (1989). Electrochemical synthesis of quinolic acid from quinoline.pdf. *Asian Journal of Chemistry*, *1*(1), 52–56.
- Hamidon, N.A., & Mukhlisin, M. (2014). Jurnal Teknologi Full paper A Review of Application of Computed Tomography on Early Detection of Basal Stem Rot Disease. *Jurnal Teknologi*, *3*, 45–47.
- Hammerschmidt, R. (2012). Secondary metabolites and defense: The story continues. *Physiological and Molecular Plant Pathology*, *80*, iii-viii.
- Hayat, A., Haider, W., Rolland, M., & Marty, J.-L. (2013a). Electrochemical grafting of long spacer arms of hexamethyldiamine on a screen printed carbon electrode surface: application in target induced ochratoxin A electrochemical aptasensor. *The Analyst*, *138*(10), 2951.
- Hayat, A., Marty, J. L., & Radi, A. E. (2012). Novel Amperometric Hydrogen Peroxide Biosensor Based on Horseradish Peroxidase Azide Covalently Immobilized on Ethynyl-Modified Screen-Printed Carbon Electrode via Click Chemistry. *Electroanalysis*, *24*(6), 1446–1452.
- Hayat, A., Sassolas, A., Marty, J. L., & Radi, A. E. (2013b). Highly sensitive ochratoxin A impedimetric aptasensor based on the immobilization of azido-aptamer onto electrografted binary film via click chemistry. *Talanta*, *103*, 14–19.
- Ho, C. L., & Tan, Y. C. (2015). Molecular defense response of oil palm to Ganoderma infection. *Phytochemistry*, *114*, 168–177.
- Horiba. (2013). A Guidebook To Particle Size Analysis. *Horiba Instruments, Inc.* Retrieved from [http://www.horiba.com/fileadmin/uploads/Scientific/Documents/PSA/PSA\\_Guidebook.pdf](http://www.horiba.com/fileadmin/uploads/Scientific/Documents/PSA/PSA_Guidebook.pdf)
- Huang, J., Xing, X., Zhang, X., He, X., Lin, Q., Lian, W., & Zhu, H. (2011). A molecularly imprinted electrochemical sensor based on multiwalled carbon nanotube-gold nanoparticle composites and chitosan for the detection of tyramine. *Food Research International*, *44*(1), 276–281.

- Hushiarian, R., Yusof, N. A., & Dutse, S. W. (2013). Detection and control of *Ganoderma boninense*: strategies and perspectives. *SpringerPlus*, 2(1), 555.
- Idris A. S., Ariffin D., Swinburne T. R., and Watt. T. A. (2000). The identity of *Ganoderma* species Responsible for BSR disease of Oil Palm in Malaysia- Morphological Characteristics.pdf. MPOB Information series, MPOB TT No.77a ISSN 1511-7871.
- Iriti, M., & Faoro, F. (2009). Chemical diversity and defence metabolism: how plants cope with pathogens and ozone pollution. *International Journal of Molecular Sciences*, 10(8), 3371–99.
- Jiang, H., Liu, X., Qiu, Y., Yao, D., & Xie, C. (2015). Development of an aptasensor for the fast detection of Versicolorin A. *Food Control*, 56, 202–210.
- Kalantar-zadeh, K., Ou, J. Z., Daeneke, T., Mitchell, A., Sasaki, T., & Fuhrer, M. S. (2016). Two dimensional and layered transition metal oxides. *Applied Materials Today*, 5, 73–89.
- Kandan, A., Ramanathan, A., Raguchander, T., Balasubramanian, P., & Samiyappan, R. (2010). Development and evaluation of an enzyme-linked immunosorbent assay (ELISA) and dot immunobinding assay (DIBA) for the detection of *Ganoderma* infecting palms. *Archives of Phytopathology & Plant Protection*, 43(15), 1473–1484.
- Kaszuba, M., McKnight, D., Connah, M. T., McNeil-Watson, F. K., & Nobbmann, U. (2008). Measuring sub nanometre sizes using dynamic light scattering. *Journal of Nanoparticle Research*, 10(5), 823–829.
- Khairunniza-bejo, S., Yusoff, Y., Nik Yusoff, N.S., Seman, I. A., & Anuar, M. I. (2015). Identification of Healthy and BSR-Infected Oil Palm Trees Using Color Indices, *International Journal of Biological, Biomolecular, Agricultural Food and Biotechnological Engineering*, 9(8), 808–811.
- Khairy, M., Kadara, R. O., & Banks, C. E. (2010). Electroanalytical sensing of nitrite at shallow recessed screen printed microelectrode arrays. *Analytical Methods*, 2, 851.
- Kok, S. M., Goh, Y.K., Tung, H.J., Goh, K. J., Wong, W. C. and Goh, Y. K. (2013). In vitro growth of *Ganoderma boninense* isolates on novel palm extract medium and virulence on oil palm (*Elaeis guineensis*) seedlings. *Malaysian Journal of Microbiology*, 9(2), 166–175.
- Lehman, J. H., Terrones, M., Mansfield, E., Hurst, K. E., & Meunier, V. (2011). Evaluating the characteristics of multiwall carbon nanotubes. *Carbon*, 49(8), 2581–2602.

- Lelong, C.C.D., Jean-Michel Roger, J.M., Brégand, S., Dubertret, F., Mathieu Lanore, M., Sitorus, N.A., Raharjo, D.A., & Caliman, J. P. (2010). Evaluation of Oil-Palm Fungal Disease Infestation with Canopy Hyperspectral Reflectance Data. *Sensors*, *10*, 734–747.
- Li, Y., Feng, S., Li, S., Zhang, Y., & Zhong, Y. (2014). A high effect polymer-free covalent layer by layer self-assemble carboxylated MWCNTs films modified GCE for the detection of paracetamol. *Sensors and Actuators, B: Chemical*, *190*, 999–1005.
- Liang, S., Li, G., & Tian, R. (2016). Multi-walled carbon nanotubes functionalized with a ultrahigh fraction of carboxyl and hydroxyl groups by ultrasound-assisted oxidation. *Journal of Materials Science*, *51*(7), 3513–3524.
- Lin, Y., Lu, F., & Wang, J. (2004). Disposable Carbon Nanotube Modified Screen-Printed Biosensor for Amperometric Detection of Organophosphorus Pesticides and Nerve Agents. *Electroanalysis*, *16*(12), 145–149.
- Liu, X., Zhang, Y., Ma, D., Tang, H., Tan, L., Xie, Q., & Yao, S. (2013). Biocompatible multi-walled carbon nanotube-chitosan-folic acid nanoparticle hybrids as GFP gene delivery materials. *Colloids and Surfaces B: Biointerfaces*, *111*, 224–231.
- Long, N. N., Vu, L. Van, Kiem, C. D., Doanh, S. C., Nguyet, C. T., Hang, P. T., Thien, N.D., & Quynh, L. M. (2009). Synthesis and optical properties of colloidal gold nanoparticles. *Journal of Physics: Conference Series*, *187*.
- López-Gresa, M. P., Torres, C., Campos, L., Lisón, P., Rodrigo, I., Bellés, J. M., & Conejero, V. (2011). Identification of defence metabolites in tomato plants infected by the bacterial pathogen *Pseudomonas syringae*. *Environmental and Experimental Botany*, *74*(1), 216–218.
- Mallakpour, S., & Madani, M. (2015). Valine amino acid-functionalized multiwalled carbon nanotube/chitosan green nanocomposite membranes: Synthesis and characterization, *High Performance Polymers*, *27*(7), 793–801.
- Malvern (2017). Malvern Zetasizer Nano user manual, [http://www.biophysics.bioc.cam.ac.uk/files/Zetasizer\\_Nano\\_user\\_manual\\_Man0317-1.1.pdf](http://www.biophysics.bioc.cam.ac.uk/files/Zetasizer_Nano_user_manual_Man0317-1.1.pdf) Date visited 2/6/2017
- Manso, J., Agüí, L., Yáñez-Sedeño, P., & Pingarrón, J. M. (2004). Development and Characterization of Colloidal Gold-Cysteamine-Carbon Paste Electrodes. *Analytical Letters*, *37*(5), 37–41.
- Martelli, A. (2009). Ultra-violet Visible Spectroscopy I. Theoretical principles. Retrieved from <http://schepartzlab.yale.edu/intranet/protocols/Ultraviolet.pdf>



- Mazid, M., Khan, T. A., & Mohammad, F. (2011). Role of secondary metabolites in defense mechanisms of plants. *Biology and Medicine*, 3(2 SPECIALISSUE), 232–249.
- Medini, F., Fellah, H., Ksouri, R., & Abdelly, C. (2014). Total phenolic, flavonoid and tannin contents and antioxidant and antimicrobial activities of organic extracts of shoots of the plant *Limonium delicatulum*. *Journal of Taibah University for Science*, 8(3), 216–224.
- Mert-türk, F. (2002). Phytoalexins : Defence or just a response to stress ? *Journal of Cell and Molecular Biology*, 1, 1–6.
- Metters, J. P., Kadara, R. O., & Banks, C. E. (2011). New directions in screen printed electroanalytical sensors: an overview of recent developments. *The Analyst*, 136(6), 1067.
- Mirčeski, V., Gulaboski, R., Jordanoski, B., & Komorsky-Lovrić, Š. (2000). Square-wave voltammetry of 5-fluorouracil. *Journal of Electroanalytical Chemistry*, 490(1), 37–47.
- Mohammed, C. L., Rimbawanto, A., & Page, D. E. (2014). Management of basidiomycete root- and stem-rot diseases in oil palm, rubber and tropical hardwood plantation crops. *Forest Pathology*, 44(6), 428–446.
- Montes-Burgos, I., Walczyk, D., Hole, P., Smith, J., Lynch, I., & Dawson, K. (2010). Characterisation of nanoparticle size and state prior to nanotoxicological studies. *Journal of Nanoparticle Research*, 12(1), 47–53.
- Muhammad, A., Yusof, N. A., Hajian, R., & Abdullah, J. (2016). Construction of an electrochemical sensor based on carbon nanotubes/gold nanoparticles for trace determination of amoxicillin in bovine milk. *Sensors (Switzerland)*, 16(1), 1–13.
- Muhammad, N., Abdullah, J., Sulaiman, Y., & Ngee, L.H. (2017). Voltammetric Determination of Nitrophenol using PEDOT Decorated Graphene Oxide as Composite Film. *International Journal of Electrochemical Science*, 12, 9432–9444.
- Mulabagal, V., & Tsay, H. (2004). Plant Cell Cultures - An Alternative and Efficient Source for the Production of Biologically Important Secondary Metabolites. *International Journal of Applied Science and Engineering*, 2(1), 29–48.
- Naher, L., Yusuf, U. K., Ismail, A., Tan, S. G., & Mondal, M. M. A. (2013). Review Article Ecological status of *Ganoderma* and basal stem rot disease of oil palms (*Elaeis guineensis* Jacq.). *Australian Journal of Crop Science*, 7(11), 1723–1727.

- Nasim, G., Ali, M., & Mehmood, N. (2010). Molecular analysis of *Ganoderma lucidum* isolates from Lahore. *Pakistan Journal of Botany*, 42(5), 3307–3315.
- Neilson, E. H., Goodger, J. Q. D., Woodrow, I. E., & Møller, B. L. (2013). Plant chemical defense: At what cost? *Trends in Plant Science*, 18(5), 250–258.
- Njila, N. M. I., Mahdi, E., Lembe, D. M., Nde, Z., & Nyonseu, D. (2017). Review on Extraction and Isolation of Plant Secondary Metabolites. *7th Int'l Conference on Agricultural, Chemical, Biological and Environmental Sciences (ACBES-2017) May 22-24, 2017 Kuala Lumpur (Malaysia) Review*, 67–72.
- Nobbmann, U., Connah, M., Fish, B., Varley, P., Gee, C., Mulot, S., Chen, J., Zhou, L.L.,
- Yanling, S., Fei-Yi, J., & Harding, S. E. (2007). Dynamic light scattering as a relative tool for assessing the molecular integrity and stability of monoclonal antibodies. *Biotechnology and Genetic Engineering Reviews*, 24(1), 117–128.
- Nusaibah, S. A., Siti Nor Akmar, A., Idris, A. S., Sariah, M., & Mohamad Pauzi, Z. (2016). Involvement of metabolites in early defense mechanism of oil palm (*Elaeis guineensis* Jacq.) against *Ganoderma* disease. *Plant Physiology and Biochemistry*, 109, 156–165.
- Oksman-Caldentey, K. M., & Inzé, D. (2004). Plant cell factories in the post-genomic era: New ways to produce designer secondary metabolites. *Trends in Plant Science*, 9(9), 433–440.
- Okumura, L. L., & Stradiotto, N. R. (2007). Simultaneous determination of quinoline and pyridine compounds in gasoline and diesel by differential pulse voltammetry. *Electroanalysis*, 19(6), 709–716.
- Olafisoye, O. B., Oguntibeju, O. O., & Osibote, O. A. (2017). Trace elements and radionuclides in palm oil, soil, water, and leaves from oil palm plantations: A review. *Critical Reviews in Food Science and Nutrition*, 57(7), 1295–1315.
- Özcan, L., Sahin, M., & Sahin, Y. (2008). Electrochemical Preparation of a Molecularly Imprinted Polypyrrole-modified Pencil Graphite Electrode for Determination of Ascorbic Acid. *Sensors*, 8, 5792–5805.
- Pandeya, S. N., & Tyagi, A. (2011). Synthetic approaches for quinoline and isoquinoline. *International Journal of Pharmacy and Pharmaceutical Sciences*, 3(3), 52–61.
- Paterson, R. R. M. (2007). *Ganoderma* disease of oil palm—A white rot perspective necessary for integrated control. *Crop Protection*, 26(9), 1369–1376.
- Pavarini, D. P., Pavarini, S. P., Niehues, M., & Lopes, N. P. (2012). Exogenous influences on plant secondary metabolite levels. *Animal Feed Science and*

*Technology*, 176(1–4), 5–16.

- Pchelintsev, N. A., & Millner, P. A. (2007). Development of Surface Activated Screen-Printed Carbon Transducers for Biosensors Application. *Analytical Letters*, 40(7), 1317–1332.
- Pedras, M. S. C., & Hossain, S. (2011). Interaction of cruciferous phytoanticipins with plant fungal pathogens: Indole glucosinolates are not metabolized but the corresponding desulfo-derivatives and nitriles are. *Phytochemistry*, 72(18), 2308–2316.
- Peng, Y., Liu, F., & Ye, J. (2006). Quantitative and qualitative analysis of flavonoid markers in *Fucus aurantii* of different geographical origin by capillary electrophoresis with electrochemical detection, *Journal of Chromatography B*, 830, 224–230.
- Piovesan, J. V., & Spinelli, A. (2014). Determination of Quercetin in a Pharmaceutical Sample by Square-Wave Voltammetry Using a Poly(vinylpyrrolidone)-Modified Carbon-Paste Electrode, *J. Braz. Chem. Soc.*, 25(3), 517–525.
- Polte, J., Ahner, T. T., Delissen, F., & Sokolov, S. (2010). Mechanism of Gold Nanoparticle Formation in the Classical Citrate Synthesis Method Derived from Coupled In Situ XANES and SAXS Evaluation. *Jacs*, (9), 1296–1301.
- Priwiratama, H., & Susanto, A. (2014). Utilization of Fungi for the Biological Control of Insect Pests and Ganoderma Disease in Indonesian Oil Palm Industry. *Journal of Agricultural Science and Technology A4*, 4, 103–111.
- Pundir, C. S., Sandeep Singh, B., & Narang, J. (2010). Construction of an amperometric triglyceride biosensor using PVA membrane bound enzymes. *Clinical Biochemistry*, 43(4–5), 467–472.
- Rakib, M. R. M., Bong, C. F. J., Khairulmazmi, A., & Idris, A. S. (2014a). Genetic and morphological diversity of Ganoderma species isolated from infected oil palms (*Elaeis guineensis*). *International Journal of Agriculture and Biology*, 16(4), 691–699.
- Rakib, M. R. M., Bong, C. F. J., Khairulmazmi, A., & Idris, A. S. (2014b). Occurrence and spatial distribution of Ganoderma species causing upper and basal stem rot in oil palm. *Journal of Food, Agriculture and Environment*, 12(2), 360–364.
- Rakib, M. R. M., Bong, C. F. J., Khairulmazmi, A., Idris, A. S., Jalloh, M. B., & Ahmed, O. H. (2017). Association of Copper and Zinc Levels in Oil Palm (*Elaeis guineensis*) to the Spatial Distribution of Ganoderma Species in the Plantations on Peat. *Journal of Phytopathology*, 165(4), 276–282.

- Raouf, J. B., Ojani, R., & Rashid-Nadimi, S. (2005). Voltammetric determination of ascorbic acid and dopamine in the same sample at the surface of a carbon paste electrode modified with polypyrrole/ ferrocyanide films. *Electrochimica Acta*, 50(24), 4694–4698.
- Rees, R. W., Flood, J., Hasan, Y., Wills, M. A. and Cooper, R. M. (2012). Ganoderma boninense Basidiospores In Oil Palm Plantations: Evaluation Of Their Possible Role In Stem Rots of Elaeis guineensis. *Plant Pathology*, 61(3), 567–578.
- Rees, W. R. (2006). Ganoderma Stem Rot of Oil Palm ( *Elaeis guineensis* ): Mode of Infection , Epidemiology and Biological Control. *PhD Thesis (University of Bath) Published by Proquest LLC*, 254.
- Ribera, A. E., & Zuñiga, G. (2012). Induced Plant Secondary Metabolites For Phytopatogenic Fungi Control: A Review. *J. Soil Sci. Plant Nutr.*, 12(4), 893–911.
- Rungjindamai, N., Pinruan, U., Choeyklin, R., Hattori, T. and Jones, E. B. G. (2008). Molecular characterization of basidiomycetous endophytes isolated from leaves , rachis and petioles of the oil palm , *Elaeis guineensis* , in Thailand. *Fungal Diversity*, (1991), 139–161.
- Sales, E. S., Schneider, J. M. F. M., Santos, M. J. L., Bortoluzzi, A. J., Cardoso, D. R., Santos, W. G., & Merlo, and A. A. (2015). Quinolines by Three-Component Reaction: Synthesis and Photophysical Studies. *J. Braz. Chem. Soc.*, 26(3), 562–571.
- Sankaran, S., Mishra, A., Ehsani, R., & Davis, C. (2010). A review of advanced techniques for detecting plant diseases. *Computers and Electronics in Agriculture*, 72(1), 1–13.
- Santoso, H., Gunawan, T., Jatmiko, R. H., Darmosarkoro, W., & Minasny, B. (2011). Mapping and identifying basal stem rot disease in oil palms in North Sumatra with QuickBird imagery. *Precision Agriculture*, 12(2), 233–248.
- Šljukić, B., Malakhova, N. A., Brainina, K. Z., Banks, C. E., & Compton, R. G. (2006). Screen printed electrodes and screen printed modified electrodes benefit from insonation. *Electroanalysis*, 18(9), 928–930.
- Soleimani, H., Yahya, N., Mk, B., Khodapanah, L., Sabet, M., Burda, M., Oechsner, A., & Awang, M. (2015). Synthesis of Carbon Nanotubes for Oil-water Interfacial Tension Reduction. *Oil and Gas Research*, 1(1), 1–5.
- Stobinski, L., Lesiak, B., Kövér, L., Tóth, J., Biniak, S., Trykowski, G., & Judek, J. (2010). Multiwall carbon nanotubes purification and oxidation by nitric acid studied by the FTIR and electron spectroscopy methods. *Journal of Alloys and Compounds*, 501(1), 77–84.

- Stradiotto, N. R., Yamanaka, H., & Zanoni, M. V. B. (2003). Review Electrochemical Sensors: A Powerful Tool in Analytical Chemistry. *J.Braz.Chem.Aoc.*, *14*(2), 159–173.
- Su, W. Y., Wang, S. M., & Cheng, S. H. (2011). Electrochemically pretreated screen-printed carbon electrodes for the simultaneous determination of aminophenol isomers. *Journal of Electroanalytical Chemistry*, *651*(2), 166–172.
- Tang, D., Zhong, Z., Niessner, R., & Knopp, D. (2009). Multifunctional magnetic bead-based electrochemical immunoassay for the detection of aflatoxin B1 in food. *The Analyst*, *134*(8),
- Tee, S. S., Tan, Y. C., Abdullah, F., Ong-Abdullah, M., & Ho, C. L. (2013). Transcriptome of oil palm (*Elaeis guineensis* Jacq.) roots treated with *Ganoderma boninense*. *Tree Genetics and Genomes*, *9*(2), 377–386.
- Tehrani, M. S., Azar, P. A., Namin, P. E., & Dehaghi, S. M. (2013). Removal of Lead Ions from Wastewater Using Functionalized Multiwalled Carbon Nanotubes with Tris ( 2-Aminoethyl ) Amine. *Journal of Environmental Protection*, *4*(June), 529–536.
- Tengoua, F. F., Hanafi, M. M., Idris, A. S., Jugah, K., Azwa, J. N. M., Hasmah, M., & Syed-Omar, S. R. (2014). Effect Of Micronutrients-Enriched Fertilizers On Basal Stem Rot Disease Incidence And Severity On Oil Palm (*Elaeis guineensis* Jacq.) Seedlings. *American Journal of Applied Sciences*, *11*(10), 1841–1859.
- Trykowski, G., Biniak, S., Stobinski, L., & Lesiak, B. (2010). Preliminary Investigations into the Purification and Functionalization of Multiwall Carbon Nanotubes. *Acta Physica Polonica A*, *118*(3), 5–8.
- Tsai, Y. C., Coles, B. A., Holt, K., Foord, J. S., Marken, F., & Compton, R. G. (2001). Microwave-enhanced anodic stripping detection of lead in a river sediment sample. A mercury-free procedure employing a boron-doped diamond electrode. *Electroanalysis*, *13*(10), 831–835.
- Vasconsuelo, A., & Boland, R. (2007). Molecular aspects of the early stages of elicitation of secondary metabolites in plants. *Plant Science*, *172*(5), 861–875.
- Völker, E., Calvo, E. J., & Williams, F. J. (2012). Formation, characterization and electrocatalytic activity of layer-by-layer self-assembled films containing polyoxomolybdate over Au surfaces. *Journal of Electroanalytical Chemistry*, *673*, 1–7.
- Wang, J., & Musameh, M. (2004). Carbon nanotube screen-printed electrochemical sensors. *The Analyst*, *129*(1), 1–2.

- Wattanapenpaiboon, N., & Wahlqvist, M. L. (2003). Phytonutrient deficiency: The place of palm fruit. *Asia Pacific Journal of Clinical Nutrition*, 12(3), 363–368.
- Wei, H., Sun, J. J., Xie, Y., Lin, C. G., Wang, Y. M., Yin, W. H., & Chen, G. N. (2007). Enhanced electrochemical performance at screen-printed carbon electrodes by a new pretreating procedure. *Analytica Chimica Acta*, 588(2), 297–303.
- Wepasnick, K. a., Smith, B. a., Schrote, K. E., Wilson, H. K., Diegelmann, S. R., & Fairbrother, D. H. (2011). Surface and structural characterization of multi-walled carbon nanotubes following different oxidative treatments. *Carbon*, 49(1), 24–36.
- Yan, D., Bazant, M. Z., Biesheuvel, P. M., Pugh, M. C., & Dawson, F. P. (2017). Theory of linear sweep voltammetry with diffuse charge: Unsupported electrolytes, thin films, and leaky membranes. *Physical Review E*, 95(3), 1–20.
- Zain, N., Seman, I. A. B. U., Kushairi, A., & Ramli, U. M. I. S. (2013). Metabolite Profiling Of Oil Palm Towards Understanding Basal Stem Rot ( BSR ) Disease, *Journal of Oil Palm Research*, 25, 58–71.
- Zhang, S., Shi, Z., & Wang, J. (2015). Sensitive and rapid determination of quinoline yellow in drinks using polyvinylpyrrolidone-modified electrode. *Food Chemistry*, 173, 449–453.
- Zhang, Z., & Xu, X. (2015). Nondestructive Covalent Functionalization of Carbon Nanotubes by Selective Oxidation of the Original Defects with  $K_2FeO_4$ . *Applied Surface Science*, 346, 520–527.
- Zhao, J., Davis, L. C., & Verpoorte, R. (2005). Elicitor signal transduction leading to production of plant secondary metabolites. *Biotechnology Advances*, 23(4), 283–333.
- Zheng, Y., Fu, L., Wang, A., & Cai, W. (2015). Electrochemical Detection of Quinoline Yellow in Soft Drinks Based on Layer-by-layer Fabricated Multi-Walled Carbon Nanotube, *Electrochimica Acta*, 10, 3530–3538.
- Zhu, S., Li, H., Niu, W., & Xu, G. (2009). Simultaneous electrochemical determination of uric acid, dopamine, and ascorbic acid at single-walled carbon nanohorn modified glassy carbon electrode. *Biosensors and Bioelectronics*, 25(4), 940–943.