



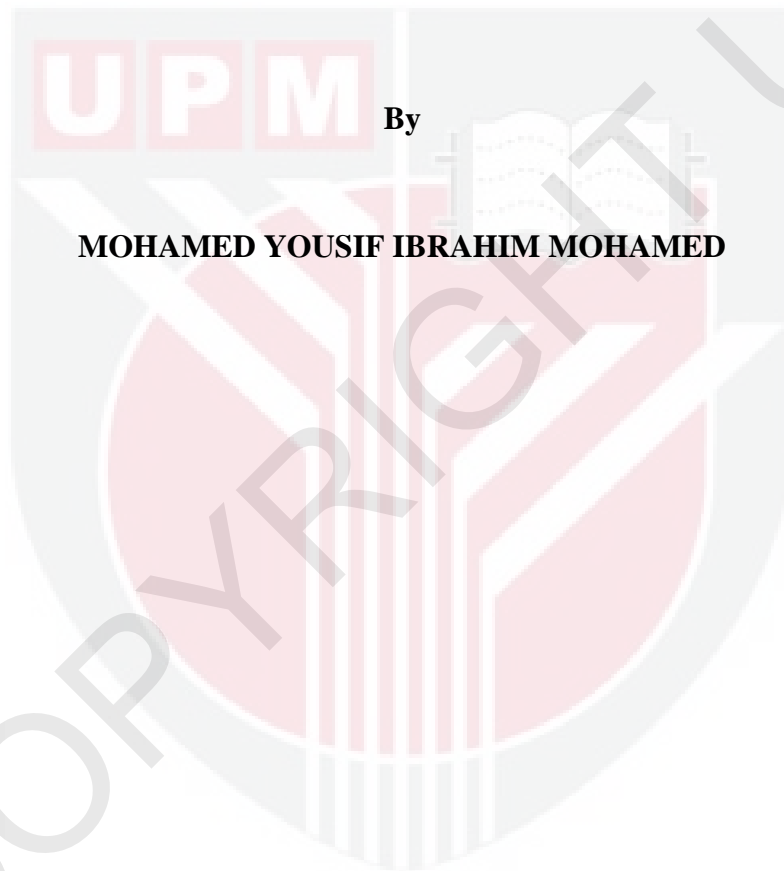
**UNIVERSITI PUTRA MALAYSIA**

***OCCURRENCE OF CAMPYLOBACTER IN WILD BIRD, AND CHICKENS  
AND DUCKS IN SELECTED MALAYSIAN FARMS***

**MOHAMED YOUSIF IBRAHIM MOHAMED**

**FPV 2014 35**

**OCCURRENCE OF *CAMPYLOBACTER* IN WILD BIRD, AND CHICKENS  
AND DUCKS IN SELECTED MALAYSIAN FARMS**



**This thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfilment of the Requirements for the Degree of Master of Veterinary  
Science.**

**October 2014**

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Veterinary Science.

## **OCCURRENCE OF CAMPYLOBACTER IN WILD BIRD, AND CHICKENS AND DUCKS IN SELECTED MALAYSIAN FARMS**

By

**MOHAMED YOUSIF IBRAHIM MOHAMED**

**October 2014**

**Chairman: Professor Dr. Saleha Abdul Aziz, PhD**

**Faculty: Veterinary Medicine**

Poultry, in particular chickens and wild birds are reported to be frequently infected with *Campylobacter*. It is well recognized that *Campylobacter jejuni* is one of the main causes of gastroenteritis in humans, and poultry meat is reported to be the main source. A number of studies in the country had shown the occurrence of *Campylobacter* in chicken and chicken meat. Wild birds were observed in abundance at market places and farms. It was reported that wild birds may harbor and transmit *Campylobacter* to farm animals. However, very few studies had been carried out on *Campylobacter* in wild birds in Malaysia and the antibiotic resistance status of *Campylobacter* in these birds was not known. Thus, this study had three main objectives: the first objective was to determine the occurrence of *Campylobacter* in the wild birds in open environment away from poultry farms and those near poultry farms; secondly, to determine the prevalence of *Campylobacter* in poultry (chickens and ducks) in the farms and thirdly, to determine the antibiotic resistance patterns among *Campylobacter* isolates. The birds in open environment in five locations (>5 km away from poultry farms) (92) and near six poultry farms (66) were humanely trapped using mist net. The photograph of each bird was taken and the birds were given an identification mark; once the cloacal swab was taken, the birds were released. Six poultry farms were visited and cloacal swabs were taken from 101 chickens from three farms and 103 were taken from ducks in three duck farms. In the laboratory, the swabs were directly streaked onto selective agar media and incubated under microaerophilic condition for isolation of *Campylobacter*. Suspected colonies of *Campylobacter* were subjected to biochemical tests for phenotypic identification. The confirmation of the isolates was done using multiplex polymerase chain reaction (mPCR) assay. From 66 birds belonging to nine species trapped near the farms environment, 20% were positive for *Campylobacter* and out of these, 92.3% was *C. jejuni*. Eurasian Tree Sparrow (39.3%) followed by Rock Pigeon (34.8%) were frequently trapped. A total of 92 birds belonging to 12 species were caught in the open environment. The most frequent bird species identified positive to *C. jejuni* and *C. coli* was Eurasian Tree Sparrow (62%), followed by Rock Pigeon (25.8%). Twenty seven percent (27%) of the birds in open environment were found positive for *Campylobacter* and most of the isolates were identified as *C. jejuni* (60%). In the

farms, a total of 40% of poultry were *Campylobacter*-positive; of these 29% of the ducks were positive for *Campylobacter* spp. of which 80% were *C. jejuni*, while 20% were *C. coli*. In chickens, a high prevalence of *Campylobacter* spp. was isolated at 60.3% and all 100% were *C. jejuni*. The *Campylobacter* isolates were subjected to antibiotic susceptibility test using disc diffusion method. The isolates were tested against 12 antibiotics, namely clindamycin, erythromycin, tetracycline, streptomycin, cefotaxime, ampicillin, gentamicin, nalidixic acid, enrofloxacin, ciprofloxacin, sulfamethoxazole-trimethoprim and chloramphenicol. The *Campylobacter* isolates from wild birds in open environment showed resistance to 2-9 antibiotics. The highest resistance was to cefotaxime and clindamycin (92.9% each), followed by nalidixic acid and streptomycin (78.6% each), tetracycline (64.3%) and chloramphenicol (57.1%). Those birds around poultry farms showed resistance to 1-4 antibiotics and the highest resistance was to clindamycin (66.7%) and less than 50% to the other four antibiotics. The isolates from poultry showed resistance to 1-9 antibiotics and highest resistance was to clindamycin (87.7%), followed by erythromycin (69.2%) and tetracycline (63.1%). From this study, it was found that *Campylobacter* was prevalent in wild birds in the open environment and near poultry farms and in chickens and ducks in the farms. The *Campylobacter* showed high resistance to antibiotics. Therefore, the birds could play a role in the dispersal and spread of antibiotic resistant *Campylobacter* in the environment and poultry farms. Thus, the presence of *Campylobacter* in wild birds, poultry and the environment may cause health hazard to human upon exposure to the organisms.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains Veterinar.

## **KEHADIRAN *CAMPYLOBACTER* PADA BURUNG LIAR DAN AYAM ITIK DI LADANG**

Oleh

**MOHAMED YOUSIF IBRAHIM MOHAMED**

**Oktober 2014**

**Pengerusi: Profesor Saleha Abdul Aziz, PhD**

**Fakulti: Perubatan Veterinar**

Unggas, khususnya ayam, dan burung liar telah dilaporkan kerap dijangkiti *Campylobacter*. Adalah diakui bahawa *Campylobacter jejuni* adalah salah satu punca utama gastroenteritis pada manusia, dan daging ayam dilaporkan menjadi punca utama. Banyak kajian dalam negara menunjukkan kehadiran *Campylobacter* pada ayam dan daging ayam. Banyak burung liar dijumpai di sekitar kawasan pasar dan ladang. Telah dilaporkan bahawa terdapat *Campylobacter* pada burung liar yang dapat disebarkan kepada haiwan ternakan. Terdapat sedikit sekali kajian mengenai kehadiran *Campylobacter* pada burung liar dan status kerintangan antibiotik *Campylobacter* tersebut tidak diketahui. Oleh itu, kajian ini mempunyai tiga objektif utama: Objektif pertama adalah untuk menentukan kehadiran *Campylobacter* pada burung liar di persekitaran terbuka jauh dari ladang ternakan; kedua, untuk mengesan kehadiran *Campylobacter* pada unggas liar di sekitar ladang ternakan dan daging ayam dan ketiga, untuk menuju pola rintangan antibiotik di kalangan pencilan *Campylobacter*. Burung liar dari lima persekitaran terbuka (>5 km jauh dari ladang ayam) (92) dan berhampiran enam ladang ternakan (66) telah diperangkap secara berperikemanusiaan menggunakan "mist net". Gambar setiap burung diambil dan burung-burung diberi tanda pengenalan; setelah swab kloaka diambil, burung-burung dibebaskan. Enam ladang ternakan dilawati dan swab kloaka telah diambil daripada 101 ayam daripada tiga ladang dan 103 telah diambil daripada itik dalam tiga ladang itik. Dalam makmal, swab disaput terus ke atas media agar terpilih dan dieram di bawah keadaan mikroaerofilik untuk pengasingan *Campylobacter*. Ujian biokimia telah dijalankan ke atas isolat yang disyaki *Campylobacter* untuk pengenalpastian fenotip. Pengesahan kepada pencilan dilakukan dengan menggunakan PCR asej multipleks. Daripada 66 burung dalam kalangan sembilan spesies yang didapati berhampiran ladang, 20% adalah *Campylobacter* spp. yang mana 92.3% terdiri daripada *C. jejuni*. Eurasian Tree Sparrow (39.3%) diikuti oleh Rock Pigeon (34.8%) telah kerap terperangkap. Sebanyak 104 burung daripada 14 spesies telah terperangkap dalam persekitaran terbuka. Spesies burung yang paling kerap dikenalpasti adalah Eurasian Tree Sparrow (62%), diikuti oleh Rock Pigeon (25.8%). Dua puluh tujuh peratus (27%) burung dalam persekitaran terbuka didapati positif untuk *Campylobacter* dan kebanyakan pencilan telah dikenalpasti sebagai *C. jejuni*

(60%). Di ladang, sebanyak 40% daripada ayam adalah *Campylobacter* positif; dua puluh sembilan peratus 29% daripada itik didapati positif untuk *Campylobacter* spp. yang mana 80% adalah *C. jejuni*, manakala 20% adalah *Campylobacter coli*. Pada ayam, *Campylobacter* spp telah diasingkan sebanyak 60.3%, dan semua 100% adalah *C. jejuni*. Isolat *Campylobacter* dijalani ujian kerintangan antibiotik menggunakan kaedah cakera penyebaran. Isolat telah diuji terhadap 12 antibiotik, iaitu clindamycin, erythromycin, tetracycline, streptomycin, cefotaxime, ampicillin, gentamicin, asid nalidixic, enrofloxacin, ciprofloxacin, sulfamethoxazole-trimethoprim dan chloramphenicol. *Campylobacter* yang diasingkan daripada burung liar di persekitaran terbuka menunjukkan kerintangan terhadap 2-9 antibiotik. Rintangan tertinggi adalah untuk cefotaxime dan clindamycin (92.9% setiap satu), diikuti oleh asid nalidixic (78.6%), tetracycline (64.3%) dan chloramphenicol (57.1%). Unggas di sekitar ladang ternakan menunjukkan kerintangan kepada 1-4 antibiotik dan rintangan yang paling tinggi adalah untuk clindamycin (66.7%) dan kurang daripada 50% kepada empat antibiotik yang lain. Isolat *Campylobacter* yang diasingkan daripada ayam menunjukkan kerintangan kepada 1-9 antibiotik dan rintangan tertinggi adalah untuk clindamycin (87.7%), diikuti dengan erythromycin (69.2%) dan tetracycline (63.1%). Daripada kajian ini, didapati bahawa *Campylobacter* tersebar luas pada burung yang terbang di persekitaran terbuka dan berhampiran ladang ayam dan itik. *Campylobacter* menunjukkan rintangan yang tinggi terhadap antibiotik. Oleh itu, burung-burung boleh memainkan peranan dalam penyebaran *Campylobacter* tahan antibiotik dalam persekitaran dan ladang ayam. Oleh itu, kehadiran *Campylobacter* pada burung liar, ayam dan itik dan persekitaran boleh menimbulkan kemudharatan kepada manusia akibat terdedah kepada organisma tersebut.

## ACKNOWLEDGEMENTS

Firstly, my heartfelt thanks to Allah for giving me the power to complete this work. I offer my sincere gratitude to Prof. Dr. Saleha Abdul Aziz, the chairman of Supervisory Committee for her enthusiastic guidance, advice, support, assistance and patience during the course of the study and the preparation of this thesis. This great comradeship has made my study experience both enjoyable and rewarding. I am also grateful to Assoc. Prof. Dr. Siti Khairani Binti Bejo, Dr. Jalila Abu and Dr. Puan Chong Leong for their guidance, comments, suggestion and supervision during the course of the study. I am sincerely thankful to my supervisory committee for being generous with their time and helping me to smoothen the path way of this study.

I would like to use this opportunity to acknowledge my appreciation and deepest thanks to my beloved father and my mother (Yousif Ibrahim Mohamed Elnihi and Siham Taha Salim Hamed) for their continuous and generous assistance and always support and pray for me to be the best.

I would like to also express my appreciation and special gratitude to Dr. Abdulwahid, Dr. Faisal, Dr. Mohamed Alser Dr. Khyralsid, Dr. Ahmed Omer and Dr. Basher for their advice.

I would like also to thank the laboratory staff of Veterinary Public Health, especially Puan Fauziah Nordin for her contribution, advice and guidance. I am also thankful to the University Veterinary Hospital and Taman Pertanian Universiti (TPU) for providing horse blood to aid my research. I would like to acknowledge Veterinary Bacteriology lab staff for their kind assistance and corporation especially Azri for helping me in PCR assay and Krish and Faiz for helping me in the media preparations. Finally I would like to thank my friend Teguh for his help in my samples collection and also to Mohamed Rashed, Emelia, Ibrahim Jalo Muhammad, Mohamed Abderhman, Nazri Samad, Mian Khaqan Shah and Saiful Mizam.

Last but not the least, deepest thanks for my brothers and sisters for their kindly encouragement which gave me confident and courage to carry out my study. Without their support, surely I would not be able to concentrate on my study.



© COPYRIGHT UPM



Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of  
Chairman of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: \_\_\_\_\_

This thesis was submitted to Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Veterinary Science. The members of supervisory committee were as follows:

**Saleha Bt Abdul Aziz, PhD**

Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Chairman)

**Siti Khairani Binti Bejo, PhD**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

**Jalila Abu, PhD**

Associate Professor  
Faculty of Veterinary Medicine  
Universiti Putra Malaysia  
(Member)

**Puan Chong Leong PhD**

Lecturer  
Faculty of Forestry  
Universiti Putra Malaysia  
(Member)

---

**BUJANG BIN Kim HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustration and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar paper, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- this is no plagiarism or data falsification/ fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: \_\_\_\_\_

## TABLE OF CONTENTS

<b>ABSTRACT</b>	Page
<b>ABSTRAK</b>	i
<b>ACKNOWLEDGEMENTS</b>	iii
<b>APPROVAL</b>	v
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	vii
<b>LIST OF FIGURES</b>	xii
<b>LIST OF ABBREVIATIONS</b>	xiii
	xv

<b>CHAPTER</b>		<b>Page</b>
<b>1</b>	<b>INTRODUCTION</b>	1
<b>2</b>	<b>LITERATURE REVIEW</b>	3
2.1	The Organism	3
2.2	Epidemiology of <i>Campylobacter</i> in animals	4
2.2.1	Occurrence of <i>Campylobacter</i> in poultry (chickens and ducks)	4
2.2.2	<i>Campylobacter</i> in wild birds	5
2.2.3	Occurrence of <i>Campylobacter</i> in ruminants and pigs	5
2.2.4	Occurrence of <i>Campylobacter</i> in other animals	6
2.3	Prevalence of <i>Campylobacter</i> in Humans	6
2.3.1	Source and modes of transmission to humans	6
2.3.1.1	Water	7
2.3.1.2	Meat	7
2.3.1.3	Milk	8
2.3.2	Occurrence of <i>Campylobacter</i> in humans	8
2.3.3	Public health significance of <i>Campylobacter</i> infection	9
2.4	Isolation, Identification and Confirmation of <i>Campylobacter</i>	10
2.4.1	Isolation methods	10
2.4.2	Identification	10
2.4.3	Confirmation using Polymerase Chain Reaction (PCR)	11
2.5	Antibiotic resistance in <i>Campylobacter</i>	11
2.5.1	Occurrence of antibiotic resistance in poultry and wild birds	11
2.5.2	Public health importance of antibiotic resistance	12
<b>3</b>	<b>ISOLATION AND IDENTIFICATION OF <i>CAMPYLOBACTER</i> IN BIRDS IN OPEN ENVIRONMENT</b>	
3.1	Introduction	13
3.2	Materials and Methods	14

3.2.1	Samples collection	14
3.2.1.1	Wild birds	14
3.2.1.2	Poultry (chicken and duck)	14
3.2.2	Identification of birds	16
3.2.3	Isolation procedure	16
3.2.4	Identification of isolates	17
3.2.4.1	Cellular morphology	17
3.2.4.2	Biochemical tests	18
3.2.4.3	Confirmation of <i>Campylobacter</i> isolates using PCR assay	18
3.3	Results	21
3.3.1	Location of the birds and poultry farms sampled	21
3.3.1.1	Location of the birds sampled in open environment	21
3.3.1.2	Location of the poultry farms	21
3.3.2	Identification of the birds	23
3.3.2.1	Wild birds in open environment	23
3.3.2.2	Wild birds near poultry farms	24
3.3.3	Isolation and identification of <i>Campylobacter</i> in the birds and poultry	28
3.3.3.1	Wild birds in open environment	28
3.3.3.2	Wild birds near poultry farms	32
3.3.3.3	Poultry	32
3.3.4	Activities of the birds observed in the poultry farm	34
3.3.5	Statistical analysis	35
3.4	Discussion	36
<b>4</b>	<b>ANTIBIOTIC SUSCEPTIBILITY TEST ON <i>CAMPYLOBACTER</i> ISOLATES</b>	
4.1	Introduction	39
4.2	Materials and Methods	40
4.2.1	Bacteria isolates	40
4.2.2	Disc diffusion method	40
4.3	Results	41
4.4	Discussion	55
<b>5</b>	<b>SUMMARY AND RECOMMENDATION FOR FUTURE RESEARCH</b>	
5.1	Summary and General Conclusion	57
5.2	Recommendation for future research	59
	<b>REFERENCES</b>	61
	<b>APPENDICES</b>	79
	<b>BIODATA OF STUDENT</b>	89
	<b>LIST OF PUBLICATIONS</b>	90

## LIST OF TABLES

Table	Page	
3.1	Differentiating <i>Campylobacter</i> species using biochemical tests	18
3.2	The primers used in PCR assay to confirm <i>Campylobacter</i> isolates	19
3.3	Location and number of the birds sampled	21
3.4	Location and number of birds caught and poultry sampled in the poultry farms	22
3.5	The species of birds in different locations	24
3.6	Number and species of wild birds caught near poultry farms	25
3.7	The species of birds in different farms	25
3.8	The <i>Campylobacter</i> positive birds in open environment	31
3.9	<i>Campylobacter</i> species isolated from birds in open environment	31
3.10	<i>Campylobacter</i> species isolated from wild birds	32
3.11	<i>Campylobacter</i> species isolated from poultry (chickens and ducks)	33
3.12	Variable associated by weighed multivariable logistic regression with <i>Campylobacter</i> prevalence in wild bird faecal samples.	35
4.1	Breakpoints to determine antimicrobial susceptibility of <i>Campylobacter</i> isolates of using the disc diffusion method	41
4.2	Antibiotic resistance of <i>Campylobacter</i> isolated from poultry in different farms	43
4.3	Antibiotic resistance of <i>Campylobacter</i> isolated from wild birds in different locations	44
4.4	Antibiotic resistance in <i>Campylobacter</i> isolates in poultry and wild birds according to farms and locations	45
4.5	Resistance patterns of <i>Campylobacter</i> isolates from poultry	52
4.6	Resistance pattern of <i>Campylobacter</i> isolates from wild birds near poultry farm	53
4.7	Resistance pattern of <i>Campylobacter</i> isolates from wild birds in open environment	53
4.8	Summary of antibiotic resistance in <i>Campylobacter</i> isolates	54

## LIST OF FIGURES

Figure		Page
3.1	Mist net setup in an open environment (beside Faculty of Veterinar Medicine)	15
3.2	A bird caught and a red band (A) was put around one leg	15
3.3	Eurasian Tree Sparrow caught inside the duck house using mist net	22
3.4	A pigeon trapped just outside a duck house	23
3.5	Wild birds caught and sampled	26
3.6A	Oxidase test	29
3.6B	Catalase test	29
3.6C	Indoxyl acetate hydrolysis test	29
3.6D	Hippurate hydrolysis hydrolysis test	30
3.7	Modified multiplex PCR to detect <i>Campylobacter</i> from wild birds	30
3.8	Modified multiplex PCR on representative isolates from poultry	33
3.9	Pigeons inside the house	34
4.1	The percentage of <i>Campylobacter</i> isolates from poultry resistant to number of antibiotics	46
4.2	The percentage of <i>Campylobacter</i> isolates from wild birds near a poultry farm resistant to number of antibiotics	47
4.3	The percentage of <i>Campylobacter</i> isolates in wild birds in open environment showing resistant to number of antibiotics	48
4.4	The percentage of <i>Campylobacter</i> isolates from poultry resistant to of antibiotics	49
4.5	The percentage of <i>Campylobacter</i> isolates from wild birds near pou farm resistant to type of antibiotics	50

- 4.6 The percentage of *Campylobacter* isolates from wild birds in open environment resistant to type of antibiotics 51





## LIST OF ABBREVIATIONS

ATCC	: American Type Culture Collection
<sup>0</sup> C	: Degree Celsius
CCUG	: Culture Collection, University of Goteborg
CDC	: Centers for Disease Control and Prevention
DNA	: Deoxyribonucleic acid
ECDC	: European Centre for Disease Prevention and Control
EFSA	: Community Zoonoses Reports of the European Food Safety Authority
FAO	: Food and Agriculture Organization
FoodNet	: Foodborne Diseases Active Surveillance Network
km	: Kilometer
MDR	: Multidrug drug resistance
ml	: Milliliter
m-PCR	: Multiplex polymerase chain reaction
PCR	: Polymerase chain reaction
PFGE	: Pulsed field gel electrophoresis
rRNA	: Ribosomal ribonucleic acid
V	: Volt

## **DEDICATION**

This thesis is specially dedicated to:

**My beloved parents,**

YOUSIF IBRAHIM MOHAMED ELNOHI

and

SIHAM TAHA SALIM HAMED

**My beloved siblings,**

AHLAM YOUSIF IBRAHIM MOHAMED,

ESRAA YOUSIF IBRAHIM MOHAMED,

OMER YOUSIF IBRAHIM MOHAMED

and

ABEDELNASSER YOUSIF IBRAHIM MOHAMED

Who always supported and encourage me to do the best.

All material contained within the thesis, including without limitation text, logos, icons, photograph and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, writer permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## CHAPTER 1

### INTRODUCTION

Campylobacteriosis is a foodborne disease in humans caused by thermophilic *Campylobacter* spp., mainly *Campylobacter jejuni* and *Campylobacter coli* (Bahrndorff et al., 2013). *Campylobacter* are slender, spirally curved, gram-negative rods with a characteristic corkscrew-like darting motility. In comparison with other food-borne bacterial pathogens, *Campylobacter* are more fragile and require microaerobic conditions for growth (Park, 2002). The two species of *Campylobacter* are widespread among livestock and poultry, usually colonizing the intestines without causing clinical diseases (Doyle and Erickson, 2006). Animals harboring *Campylobacter* consistently shed the bacteria through feces, spreading within and among animal species on farms (Doyle and Erickson, 2006). Wild mammals and wild birds have been identified as possible reservoirs for *Campylobacter* transmission to agricultural animals because they carry *Campylobacter* spp. in their gut and also, these animals could be possible potential reservoirs for human (Horrocks et al., 2009; Kwan et al., 2008; Colles et al., 2003; Broman et al., 2002; Craven et al., 2000). Thus far, campylobacteriosis has been reported in wild birds and mammals.

In human, *Campylobacter* infection could either be severe or mild depending on the individuals infected; for example, in young, old or people with weak immunity, it is known to have serious prolonged effect while in the middle aged people with strong immunity it is easily controlled without antibiotic administration (Smith and Fratamico, 2010). The common symptoms of *Campylobacter* infection include fever, diarrhea and abdominal pain (Butzler, 2004). In few cases, the infection can lead to severe complications, such as the Guillain-Barré syndrome which is an acute autoimmune neuropathy with ascending paralysis (Zilbauer et al., 2008). Human may acquire *Campylobacter* from sources such as untreated raw milk (Heuvelink et al., 2009) or water (Abe et al., 2008), but the consumption of poultry meat, particularly raw or inadequately cooked meat, constitutes the main risk factor for infection (Wingstrand et al., 2006). Potential sources and vectors for environmental contamination are infected livestock and free-living animals (Zweifel et al., 2008; Ellis-Iversen et al., 2009), rodents and flies (Hazeleger et al., 2008), contaminated surface water (Messens et al., 2009) and personnel and farm equipment (Ramabu et al., 2004) at the farm.

There is scarcity of information on the epidemiology of *Campylobacter* in wild birds compared to poultry. A study on *Campylobacter* spp. in black-headed gull found the prevalence ranged from 27.9 to 95.5%, depending on the species sampled (Broman et al., 2002). Studies on *Campylobacter* spp. in broiler chickens found the prevalence ranged from 40 to 60% (Broman et al., 2002).

For clinical therapy of campylobacteriosis, erythromycin (a macrolide) is adjudged the drug of preference, but fluoroquinolones (e.g., ciprofloxacin) are also commonly used

owing to their expansive continuum of activity against enteric pathogens (Luangtongkum et al., 2009). The results from a study on *Campylobacter* by Chen et al. (2010) showed that a vast majority of the isolates from birds appeared to be resistant to fluoroquinolones by more than 98%. In the same study, the *C. jejuni* was resistant to gentamicin at 27.2%. Importantly, previous study from The Netherlands revealed a substantial increase in fluoroquinolone resistance among human cases since the advent of enrofloxacin in veterinary medicine (Endtz et al., 1991). According to Zendeabad et al. (2013), *Campylobacter* spp. isolates from chicken, quail, partridge, and turkey meat in Mashhad (Iran) showed low resistance to streptomycin at 4.9%, 7.4% and 5.4%, respectively.

Alternative drugs used in cases of systemic infection with *Campylobacter* include tetracyclines and gentamicin (Ge et al., 2002). However, the growing resistance to *Campylobacter* is of a serious concern globally. *Campylobacter* is continuously exposed to antibiotics used in animal production and human medicine thus, causing it to become resistant to antibiotics which can create limited therapeutic options (Luangtongkum et al., 2009).

A number of studies in the country had shown the occurrence of *Campylobacter* in chicken and chicken meat. Wild birds were observed in abundance at market places and farms. Stald et al (2003) reported that wild birds may harbor and transmit *Campylobacter* to farm animals. However, very few studies had been carried out on *Campylobacter* in wild birds in Malaysia and the antibiotic resistance status of *Campylobacter* in these birds was not known.

The hypotheses of this study were:

- i) There is a high occurrence of *Campylobacter* in wild birds, chickens and ducks.
- ii) *Campylobacter* isolates showed high resistance to a number of antibiotics.

Thus, the objectives of this study were:

- i. to determine the occurrence of *Campylobacter* in wild birds in open environment, poultry farms and in poultry (chickens and ducks), and
- ii. to determine the antibiotic resistance patterns among *Campylobacter* isolates from wild birds and poultry.

## REFERENCES

- Abe, T., Haga, S., Yokoyama, K. and Watanabe, N. 2008. An outbreak of *Campylobacter jejuni* subsp. *jejuni* infection via tap water. *Japanese Journal of Infectious Diseases* 61: 327.
- Abulreesh, H. H., Paget, T. A. and Goulder, R. 2006. *Campylobacter* in waterfowl and aquatic environments: incidence and methods of detection. *Environmental Science and Technology* 40(23): 7122-7131.
- Adams, J. B., Leah J. J., Linda D. P., Quig, D. and Rubin, R. A. 2011. Gastrointestinal flora and gastrointestinal status in children with autism--comparisons to typical children and correlation with autism severity. *BMC Gastroenterology* 11: 22. doi:10.1186/1471-230X-11-22.
- Adhikari, B., Madie, P., Connolly, J., Davies, P., Layland, M., and Rogers, L. 2002. Wild birds, flies, and rodents as reservoirs of *Campylobacter* spp. on dairy farm. <http://maxa.maf.govt.nz/mafnet/rural-nz/research-and-development/pestcontrol/campylobacter-on-a-dairy-farm/wild-birds-technical-paper-200218.pdf> [Accessed on 15 December 2013].
- Adzitey, F., Rusul, G., Huda, N., Cogan, T. and Corry, J. 2012. Prevalence, antibiotic resistance and RAPD typing of *Campylobacter* species isolated from ducks, their rearing and processing environments in Penang, Malaysia. *International Journal of Food Microbiology* 154: 197-205.
- Alanis, A. J. 2005. Resistance to antibiotics: are we in the post-antibiotic era?. *Archives of Medical Research* 36: 697-705.
- Allos, B. M. 2001. *Campylobacter jejuni* infections: update on emerging issues and trends. *Clinical Infectious Diseases* 32(8): 1201-1206.
- Altekruse, S. F., Stern, N. J., Fields, P. I., Swerdlow, D. L. 1999. *Campylobacter jejuni* an emerging foodborne pathogen. *Emerging Infectious Diseases Journal* 5:28-35.
- Andres, S., Vico, J. P., Garrido, V., Grillo, M. J., Samper, S., Gavin, P., Herrere-Leon, S. and Mainer-Jaime, R. C. 2012. Epidemiology of subclinical salmonellosis in wild birds from an area of high prevalence of pig salmonellosis: phenotypic and genetics profiles of *Salmonella* isolates. *Zoonoses and Public Health* 60(5): 355-65. doi: 10.1111/j.1863-2378.2012.01542.x. Epub 2012 Aug.
- Andrzejewska, M., Szczepańska, B., Klawe, J. J., Śpica, D. and Chudzińska, M. 2013. Prevalence of *Campylobacter jejuni* and *Campylobacter coli* species in cats and dogs from Bydgoszcz (Poland) region. *Polish Journal of Veterinary Sciences* 16(1): 115-120.

Ang, C. W., Teunis, P. F. M., Herbrink, P., Keijser, J., Duynhoven, Y. H. T. P. V., Visser, C. E. and Pelt, W. V. 2011. seroepidemiological studies indicate frequent and repeated exposure to *Campylobacter* spp. during childhood. *Epidemiology and Infection* 139(9): 1361-8.

Anonymous. 2007. Emerging infections program. [http://cdc.gov/foodnet/news/2007/October2007\\_foodnet\\_news.pdf](http://cdc.gov/foodnet/news/2007/October2007_foodnet_news.pdf) [Accessed on 20 September 2013].

Antilles, N., Sanglas, A. and Cerda-Cuellar, M. 2013. Free-living waterfowl as a source of zoonotic bacteria in a dense wild bird population area in northeastern Spain. *Transboundary and Emerging Diseases*. doi:10.1111/tbed.12169.

Axelsson-Olsson, D., Olofsson, J., Svensson, L., Griekspoor, P., Waldenström, J., Patrik Ellström, P. and Olsen, B. 2010. Amoebae and algae can prolong the survival of *Campylobacter* species in co-culture. *Experimental Parasitology* 126(1): 59–64.

Bahrndorff, S., Rangstrup-Christensen, L., Nordentoft, S. and Hald, B. 2013. Foodborne disease prevention and broiler chickens with reduced *Campylobacter* infection. *Emerging Infectious Diseases* 19(3): 425-430.

Beth, K. D., David, T. R. 2011. Update on human *Campylobacter jejuni* infections. *Gastroenterology and Hepatology Journal* 27(1): 1-7.

Bogaard, A. E. V. D. and Stobberingh, E. E. 2000. Epidemiology of resistance to antibiotics links between animals and human. *International Journal of Antimicrobial Agents* 14: 327–335.

Boonmar, S., Yingsakmongkon, S., Songserm, T., Hanhaboon, P. and Passadurak, W. 2007. Detection of *Campylobacter* in duck using standard culture method and multiplex polymerase chain reaction. *Southeast Asian Journal of Tropical Medicine* 38(4): 728-731.

Boysen, L., Nauta, M., Duarte, A. S. R. and Rosenquist, H., 2013. Human risk from thermotolerant *Campylobacter* on broiler meat in Denmark. *International Journal of Food Microbiology* 162: 129-134.

Briffett, C. 1993. The Birds of Singapore. Collins: Kuala Lumpur. Oxford University Press.

Broman, T., Palmgren, H., Bergström, S., Sellin, M., Waldenström, J., Danielsson-Tham M. L. and Olsen, B. 2002. *Campylobacter jejuni* in black-headed gulls (*Larus ridibundus*): prevalence, genotypes, and influence on *C. jejuni* epidemiology. *Journal of Clinical Microbiology* 40(12): 4594-4602.

Buhr, R. J., Cox, N. A., Stern, N. J., Musgrove, M. T., Wilson, J. L. and Hiatt, K. L. 2002. Recovery of *Campylobacter* from segments of the reproductive tract of broiler breeder hens. *Avian Diseases* 46:919-924.

- Bull, S. A., Allen, V. M., Domingue, G., Jørgensen, F., Frost, J. A., Ure, R., Whyte, R., Tinker, D., Corry, J. E. L., Gillard-King, J. and Humphrey, T. J. 2006. Sources of *Campylobacter* spp. colonizing housed broiler flocks during rearing. *Applied and Environmental Microbiology* 72(1): 645-652.
- Burns, B. M, Fordyce, G. and Holroyd, R. G. 2010. A review of factors that impact on the capacity of beef cattle females to conceive, maintain a pregnancy and wean a calf-implications for reproductive efficiency in northern Australia. *Animal Reproduction Science* 122 (1-2): 1–22.
- Butzler, J. P., 2004. *Campylobacter*, from obscurity to celebrity. *Clinical Microbiology and Infection*. 10: 868-876.
- Camarda, A., Newell, D. G., Nasti, R. and Modugno, G. D. 2000. Genotyping *Campylobacter jejuni* strains isolated from the gut and oviduct of laying hens. *Avian Diseases* 44:907-912.
- Carrique-Mas, J. J., Bryant, J. E., Cuong, N. V., Hoang, N. V. M., Campbell, J., Hoang, N. V., Dung, T. T. N., Duy, D.T., Hoa, N. T., Thompson, C., Hien, V. V., Phat, V. V., Farrar, J. and Baker, S. 2013. An epidemiological investigation of *Campylobacter* in pig and poultry farms in the Mekong delta of Vietnam. *Epidemiology Infection*. doi:10.1017/S0950268813002410.
- Carter, A. M., Pacha, R. E., Clark, G. W. and Williams, E. A. 1987. Seasonal occurrence of *Campylobacter* spp. in surface waters and their correlation with standard indicator bacteria. *Applied and Environmental Microbiology* 53(3): 523-526.
- Cawthraw, S. A., Wassenaar, T. M., Ayling, R. and Newell, D. G. 1996. Increased colonization potential of *Campylobacter jejuni* strain 81116 after passage through chickens and its implication on the rate of transmission within flocks. *Epidemiology and Infection* 117(1): 213-215.
- Centers for Disease Control and Prevention. 2010. Preliminary FoodNet Data on the incidence of infection with pathogens transmitted commonly through food – 10 states, 2009. *MMWR Morbidity Mortal Weekly Report journal* 59(14):418-22.
- Champlot, S., Berthelot, C., Pruvost, M., Bennett, E. A., Grange, T. and Geigl, E. 2010. An efficient multistrategy DNA decontamination procedure of PCR reagents for hypersensitive PCR applications. *PloS One* 5(9): 4–15.
- Chen, X., Naren, G. W., Wu, C. M., Wang, Y., Dai, L., Xia, L. N., Luo, P. J., Zhang, Q. and Shen, J. Z. 2010. Prevalence and antimicrobial resistance of *Campylobacter* isolates in broilers from China. *Veterinary Microbiology* 144: 133-139.
- Chon, J. W., Hyeon, J. Y., Yim, J. H., Kim, J. H., Song, K. S. and Seo, K. H. 2011. Improvement of modified charcoal-cefoperazone-deoxycholate agar by supplementation with a high concentration of polymyxin b for detection of *C.*



*jejuni* and *C. coli* in chicken carcass rinses. *Applied and Environmental Microbiology* 78(5): 1624-1626.

Clinical and Laboratory Standards Institute (CLSI). 2010. Performance standards for antimicrobial susceptibility testing: twentieth information Supplement M100-S20 Wayne, PA. USA.

Cody, A. J., Maiden, M. J. C. and Dingle, K. E. 2009. Genetic diversity and stability of the pora allele as a genetic marker in human *Campylobacter* infection. *Journal of Microbiology* 155(12): 4145-4154.

Cokal, Y., Caner, V., Sen, A., Cetin, C. and Telli, M. 2011. The presence of *Campylobacter jejuni* in broiler houses: Results of a longitudinal study. *African Journal of Microbiology Research* 5(4): 389-393.

Colles, F. M., Ali, J. S., Sheppard, S. K., McCarthy, N. D. and Maiden, M. C. J. 2011. *Campylobacter* populations in wild and domesticated Mallard ducks (*Anas platyrhynchos*). *Environmental Microbiology Reports* 3(5): 574-580.

Colles, F. M., Dingle, K. E., Cody, A. J. and Maiden, M. C. J. 2008. Comparison of *Campylobacter* populations in wild geese with those in starlings and free-range poultry on the same farm. *Applied and Environmental Microbiology* 74(11): 3583-3590.

Colles, F. M., Jones, K., Harding, R. M. and Maiden, M. C. 2003. Genetic diversity of *Campylobacter jejuni* isolates from farm animals and the farm environment. *Applied and Environmental Microbiology*. 69: 7409-7413.

Collignon, P., Powers, J. H., Chiller, T. M., Aidara-Kane, A. and Aarestrup, F. M. 2009. World Health Organization ranking of antimicrobials according to their importance in human medicine: a critical step for developing risk management strategies for the use of antimicrobials in food production animals. *Journal of Food Safety* 49: 132-141.

Connell, S., Meade, K. G., Allan, B., Lloyd, A. T., Kenny, E., Cormican, P., Morris, D. W., Bradley, D. G. and O'Farrelly, C. 2012. Avian resistance to *Campylobacter jejuni* colonization is associated with an intestinal immunogene expression signature identified by MRNA sequencing. *PloS One* 7(8): e40409.

Cox, N. A., Richardson, L. J., Maurer, J. J., Berrang, M. E., Fedorka-Cray, P. J., Buhr, R. J., Byrd, J. A., Lee, M. D., Hofacre, C. L., O'Kane, P. M., Lammerding, A. M., Clark, A. G., Thayer, S. G. and Doyle, M. P. 2012. Evidence for horizontal and vertical transmission in *Campylobacter* passage from hen to her progeny. *Journal of Food Protection* (10): 1728-1902.

- Cox, N., Stern, N., Wilson, J., Musgrove, M., Buhr, R. and Hiatt, R. 2001. Isolation of *Campylobacter* spp. from semen samples of commercial broiler breeder roosters. *Avian Diseases* 46(3): 717-720.
- Craven, S. E., Stern, N. J., Line, E., Bailey, J. S., Cox, N. A. and Fedorka-Cray, P. 2000. Determination of the Incidence of *Salmonella* spp., *Campylobacter jejuni*, and *Clostridium perfringens* in wild birds near broiler chicken houses by sampling intestinal droppings. *Avian Diseases* 44(3): 715-720.
- Craven, S.E., Stern, N. J., Line, E., Bailey, J. S., Cox, N. A., Fedorka-Cray, P. 2000. Determination of the incidence of *Salmonella* spp., *Campylobacter jejuni*, and *Clostridium perfringens* in wild birds near broiler chicken houses by sampling intestinal droppings. *Avian Diseases*. 44: 715-720.
- Dasti, J. I., A. Tareen, M., Lugert, R., Zautner, A. E. and Gross, U. 2010. *Campylobacter jejuni*: a brief overview on pathogenicity-associated factors and disease-mediating mechanisms. *International Journal of Medical Microbiology* (4): 205-211.
- Denis, M., Tanguy, M., Chidaine, B., Laisney, M. J., Mégraud, F. and Fravallo, P. 2011. Description and sources of contamination by *Campylobacter* spp. of river water destined for human consumption in brittany, france. *Pathologie Biologie* 59(5): 256-263.
- Dhama, K., M. M. and Tomar, S. 2008. Pathogens transmitted by migratory birds: Threat perceptions to poultry health and production. *International Journal of poultry science* 7(6): 516-525.
- Doyle, M. P. and Erickson, M. C. 2006. Reducing the carriage of foodborne pathogens in livestock and poultry. *Journal of Poultry Science*. 85: 960-973.
- Dyke, M. I. V., Morton, V. K., McLellan, N. L. and Huck, P. M. 2010. The occurrence of *Campylobacter* in river water and waterfowl within a watershed in southern Ontario, Canada. *Journal of Applied Microbiology* 109: 1364-5072.
- Edens, F.W. 2003. An alternative for antibiotic use in poultry: probiotics. *Rev. Bras. Cienc. Avic.* 5(2). <http://dx.doi.org/10.1590/S1516-635X2003000200001>.
- EFSA., 2007. The community summary report on trends and sources of zoonoses, zoonotic agents, antimicrobial resistance and foodborne outbreaks in the European union in 2006. *EFSA Journal* 130: 130-155.
- EFSA., 2009. The community summary report on trends and sources of zoonoses and zoonotic. Agents in the European union in 2007. *EFSA Journal* 223: 223-440.

- EFSA., 2010. The community summary report on trends and sources of zoonoses, zoonotic agents and foodborne outbreaks in the European union in 2008. *EFSA Journal* 8: 1496-1906.
- Ellis-Iversen, J., Jorgensen, F., Bull, S., Powell, L., Cook, A. J. and Humphrey, T. J. 2009. Risk factors for *Campylobacter* colonisation during rearing of broiler flocks in great britain. *Veterinary Journal*. 89: 178–184.
- Endtz, H. P., Ruijs, G. J., Klinger B. V., Jansen W. H., Reyden, T. V. D. and Mouton R. P. 1991. Quinolone resistance in *Campylobacter* isolated from man and poultry following the introduction of fluoroquinolones in veterinary medicine. *Journal of Antimicrobial Chemotherapy* 27: 199– 208.
- Engberg, J., Aarestrup, F. M., Taylor, D. E., Gerner-Smidt, P. and Nachamkin, I. 2001. Quinolone and macrolide resistance in *Campylobacter jejuni* and *C. coli*: resistance mechanisms and trends in human isolates. *Emerging Infectious Diseases* 7(1): 24-34.
- Faiza, N., S., Saleha, A.A., Jalila, A. and Fauziah, N., 2013. Occurrence of *Campylobacter* and *Salmonella* in ducks and duck eggs in Selangor, Malaysia. *Tropical Biomedicine* 30(1): 155-158.
- Fearnley, C., Manning, G., Bagnall, M., Javed, M. A., Wassenaar, T. M. and Newell, D. G. 2008. Identification of hyperinvasive *Campylobacter jejuni* strains isolated from poultry and human clinical sources. *Journal of Medical Microbiology* 57(5): 570-580.
- Fernández, H., Gesche, W., Montefusco, A. and Schlatter, R. 1996. Wild birds as reservoir of thermophilic enteropathogenic *Campylobacter* species in southern chile. *Memórias do Instituto Oswaldo Cruz* 91(6): 699-700.
- Figuroa, G., Troncoso, M., López, C., Rivas, P. and Toro, M. 2009. Occurrence and enumeration of *Campylobacter* spp. during the processing of chilean broilers. *BMC Microbiology* 9: 94.
- French, N. P., Midwinter, A. Holland, B., Collins-Emerson, J., Pattison, R., Colles, F. and Carter, P. 2009. Molecular epidemiology of *Campylobacter jejuni* isolates from wild-bird fecal material in children’s playgrounds. *Applied and Environmental Microbiology* 75: 779-783.
- Friis, C., Wassenaar, T. M., Javed, M. A., Snipen, L., Lagesen, K., Hallin, P. F., Newell, D. G., Toszeghy, M., Ridley, A., Manning, G. and Ussery, D. W. 2010. Genomic characterization of *Campylobacter jejuni* strain M1. *PLoS ONE* 5(8): e12253. doi:10.1371/journal.pone.0012253.

- Ganapathy, K., Saleha, A. A., Jaganathan, M., Tan, C. G., Chong, C. T., Tang, S. C., Ideris, A. Dare, C. M. and Bradbury, J. M. (2007). Survey of *Campylobacter*, *Salmonella* and *Mycoplasmas* in house crows (*Corvus splendens*) in Malaysia. *Journal of the British Veterinary Association* 160 (18): 622-624.
- Ge, B., Bodeis, S., Walker, R. D., White, D. G., Zhao, S., McDermott, P. F. and Meng, J. 2002. Comparison of the Etest and agar dilution for in vitro antimicrobial susceptibility testing of *Campylobacter*. *Journal of Antimicrobial Chemotherapy*. 50: 487-494.
- Gilpin, B. J., Robson, B., Scholes, P., Nourozi, F. and Sinton, L. W. 2009. Survival of *Campylobacter* spp. in bovine faeces on pasture. *Letters in Applied Microbiology* 48(2): 162-166.
- Gilpin, B. J., Thorrold, B., Scholes, P., Longhurst, R. D., Devane, M., Nicol, C., Walker, S., Robson, B. and Savill, M. 2008. Comparison of *Campylobacter jejuni* genotypes from dairy cattle and human sources from the matamata-piako district of New Zealand. *Journal of Applied Microbiology* 105(5):1354-1360.
- Goldman, C. G., Matteo, M. J., Loureiro, J. D., Degrossi, J., Teves, S., Heredia, S. R., Alvarez, K., Gonzalez, A. B. N., Catalano, M., Boccio, J., Cremaschi, G., Solnick, J. V., Zubillaga, M. B. 2009. Detection of helicobacter and *Campylobacter* spp. from the aquatic environment of marine mammals. *Veterinary Microbiology* 133(3): 287-291.
- Granato, P. A., Chen, L., Holiday, I., Rawling, R. A., Novak-Weekley, S. M., Quinlan, T. and Musser, K. A. 2010. Comparison of premier CAMPY enzyme immunoassay (EIA), ProSpecT *Campylobacter* EIA, and ImmunoCard STAT! CAMPY tests with culture for laboratory diagnosis of *Campylobacter* enteric infections. *Journal of Clinical Microbiology* 48(11): 4022-4027.
- Gras, L. M., Smid, J. H., Wagenaar, J. A., Koene, M. G. J., Havelaar, A. H., Friesema, I. H. M., French, N. P., Flemming, C., Galson, J. D., Graziani, C., Busani L. and Pelt, W. V. 2013. Increased risk for *Campylobacter jejuni* and *C. coli* infection of pet origin in dog owners and evidence for genetic association between strains causing infection in humans and their pets. *Epidemiology and Infection Journal* 141(12): 2526-2535.
- Gupta, A., Nelson, J. M., Barrett, T. J., Tauxe, R. V., Rossiter, S. P., Friedman, Joyce, C. R. K. W., Smith, K. E., Jones, T. F., Hawkins, M. A., Shiferaw, Beebe, B. J. L., Vugia, D. J. V., Rabatsky-Ehr, T., Benson, J. A., Root, T. P. and Angulo, F. J. 2004. Antimicrobial Resistance among *Campylobacter* Strains, United States, 1997–2001. *Emerging Infectious Diseases* 10(6): 1102-1109.
- Hakkinen, M., Nakari, U. M. and Siitonen, A. 2009. Chickens and cattle as sources of sporadic domestically acquired *Campylobacter jejuni* infections in finland. *Applied and Environmental Microbiology* 75(16): 5244-5249.

- Hald, B., Madsen, J. J., Rahbek, C., Chriel, M., Nielsen, E. M., Bang, D. D., Lodal, J., Jespersen, J. B., Wainø, M., Dietz, H. H., Jørgensen, J. C., Baggesen, D. L., Skov, M. N. and Madsen M. 2003. *Campylobacter* carriage by wild birds, rodents, insects and other animals in the immediate environment of cattle, pig and poultry farms in Denmark. <http://pure.au.dk/portal/files/603192/campylobacterposter.pdf> [Accessed on 15 May 2013].
- Hald, B., Rattenborg, E. and Madsen, M. 2001. Role of batch depletion of broiler houses on the occurrence of *Campylobacter* spp. in chicken flocks. *Letters in Applied Microbiology* 32: 253-256.
- Hald, B., Skovgård, H. and Madsen, M. 2004. Flies and *Campylobacter* Infection of Broiler Flocks. *Emerging Infectious Diseases* 10(8): 1490-1492.
- Hansson, I., Engvall, E. O., Vågsholm, I. and Nyman, A. 2010. Risk factors associated with the presence of *Campylobacter*-positive broiler flocks in Sweden. *Preventive Veterinary Medicine* 96(1-2): 114–121.
- Hazeleger, W. C., Bolder, N. M., Beumer, R. R., Jacobs-Reitsma, W. F. 2008. Darkling beetles (*Alphitobius diaperinus*) and their larvae as potential vectors for the transfer of *Campylobacter jejuni* and *Salmonella enterica* serovar paratyphi B variant java between successive broiler flocks. *Applied and Environmental Microbiology*. 74: 6887–6891.
- Henry, I., Reichardt, J., Denis, M. and Cardinale, E., 2011. Prevalence and risk factors for *Campylobacter* spp. in chicken broiler flocks in Reunion Island (Indian Ocean). *Preventive Veterinary Medicine* 100: 64-70.
- Hermans, D., Pasmans, F., Messens, W., Martel, A., Immerseel, F. V., Rasschaert, G., Heyndrickx, M., Deun, K. V. and Haesebrouck, F. 2012. Poultry as a host for the zoonotic pathogen *Campylobacter jejuni*. *Vector Borne and Zoonotic Diseases* 12(2): 89-98.
- Heuvelink, A. E., Heerwaarden, C. V., Ans Zwartkruis-Nahuis, A., Tilburg, J. J. H. C., Bos, M. H., Heilmann, F. G. C., Hofhuis, A., Hoekstra, T. and Boer, E. D. 2009. Two outbreaks of Campylobacteriosis associated with the consumption of raw cows' milk. *International Journal of Food Microbiology* 134(1-2): 70-4.
- Hiatt, K. L., Cox, N. A., Buhr, R. J. and Stern, N. J. 2002. Genotype analyses of *Campylobacter* isolated from distinct segments of the reproductive tracts of broiler breeder hens. *Current microbiology* 45:400-404.
- Hjartardottir, S., Fridriksdottir, V., Gunnarsson, E., Reiersen, J., Andresdottir, V., Bjarnadottir, S., Jonsdottir, G. and Astradsdottir, K. 2013. Comparison of sensitivity of a Campy-Cefex dilution method and PCR in detecting *Campylobacter* in broilers. *Icelandic Agricultural Sciences. Science Citation Index* 26: 11-19.

- Horrocks, S. M., Anderson, R. C., Nisbet, D. J., Ricke, S. C. 2009. Incidence and ecology of *Campylobacter jejuni* and *coli* in animals. *Anaerobe* 15(1-2): 18-25.
- Iovine, N. M. and Blaser, M. J. 2004. Antibiotics in Animal Feed and Spread of Resistant *Campylobacter* from Poultry to Humans. *Emerging Infectious Diseases* 10(6): 1158-1189.
- Jacobs-Reitsma, W. F., Kan, C. A. and Bolder, N. M. 1994. The induction of quinolone resistance in *Campylobacter* bacteria in broilers by quinolone treatment. *Applied Microbiology* 19(4): 228-231.
- Jasass, B. 2010. Application of polymerase chain reaction to differentiate between strains of *Campylobacter jejuni* and *Campylobacter coli*. *African Journal of Biotechnology* 9(10): 1412-1415.
- Jay-Russell, M. T., Mandrell, R. E., Yuan, J., Bates, A., Manalac, R., Mohle-Boetani, J., Kimura, A., Lidgard, J. and Miller, W. G. 2013. Using major outer membrane protein typing as an epidemiological tool to investigate outbreaks caused by milk-borne *Campylobacter jejuni* isolates in California. *Journal of Clinical Microbiology* 51(1): 195. DOI: 10.1128/JCM.01845-12.
- Jore, S., Viljugrein, H., Brun, E., Heier, B. T., Borck, B., Ethelberg, S., Hakkinen, M., Kuusi, M., Reiersen, J., Hansson, I., Olsson E., Engvall, Løfdahl, M., Wagenaar, J. A., Pelt, W. V., Hofshagen, M. 2010. Trends in *Campylobacter* incidence in broilers and humans in six european countries, 1997-2007. *Preventive Veterinary Medicine* 93(1): 33-41.
- Jun, W., Chang, G. Y., and Ning, L. 2013. Prevalence and risk assessment of *Campylobacter jejuni* in chicken in China. *Journal of Environmental Science* 26(4): 243-248.
- Kasrazadeh, M. and Genigeorgis, C. 1987. Origin and prevalence of *Campylobacter jejuni* in ducks and duck meat at the farm and processing level. *Journal of Food Protection* 50: 321-326.
- Kaur, T., Singh, J., Huffman, M. A., Petrželková, K. J., Taylor, N. S., Xu, S., Dewhirst, F. E., Paster, B. J., Debruyne, L., Vandamme, P. and Fox, J. G. 2011. *Campylobacter troglodytis* sp. nov., isolated from feces of human-habituated wild chimpanzees (*Pan troglodytes schweinfurthii*) in Tanzania. *Applied and Environmental Microbiology* 77(7): 2366-2373.
- Keller, J. I., Shriver, W. G., Waldenstrom, J., Griekspoor, P. and Olsen, B. 2011. Prevalence of *Campylobacter* in wild birds of the Mid-Atlantic region, USA. *Journal of Wildlife Diseases* 47(3): 750-754.
- Kendall, J. J., Barrero-Tobon, A. M., Hendrixson, D. R. and Kelly, D. J. 2013. Hemerythrins in the microaerophilic bacterium *Campylobacter jejuni* help

protect key iron–sulphur cluster enzymes from oxidative damage. *Environmental Microbiology* 16(4): 1105-1121.

Keramas, G., Bang, D. D., Lund, M., Madsen, M., Bunkenborg, H., Telleman, P. and Christensen, C. B. V. 2004. Use of culture, PCR analysis, and DNA microarrays for detection of *Campylobacter jejuni* and *Campylobacter coli* from chicken feces. *Journal of Clinical Microbiology* 42(9): 3985-3991.

Kienesberger, S., Fauster, C. S. T. A., Lang, S., Sprenger, H., Gorkiewicz, G. and Zechner, E. L. 2011. Interbacterial macromolecular transfer by the *Campylobacter fetus* subsp. *venerealis* type iv secretion system. *Journal of Bacteriology* 193(3): 744-758.

King, B.F. and E.C. Dickinson. 1975. A Field Guide to the Birds of South-East Asia. Collins: London.

Koene, M. G. J., Houwers, D. J., Dijkstra, J. R., Duim, B. B. and Wagenaar, A. J. 2009. Strain variation within *Campylobacter* species in fecal samples from dogs and cats. *Veterinary Microbiology* 133(1-2): 199-205.

Koziel, M., Lucid, A., Bullman, S., Corcoran, G. D., Brigid Lucey, B. and Sleator, R. D. 2014. Draft genome sequence of *Campylobacter corcagiensis* strain CIT045T, a representative of a novel *Campylobacter* species isolated from lion-tailed macaques (*Macaca silenus*). *American Society for Microbiology* 2(2): e00248-14.

Kumarasamy, K. K., Toleman, M. A., Walsh, T. R., Bagaria, J., Butt, F., Balakrishnan, R., Chaudhary, U., Doumith, M., Giske, C. G., Irfan, S., Krishnan, P., Kumar, A. V., Maharjan, S., Mushtaq, S., Noorie, T., Paterson, D. L., Pearson, A., Perry, C., Pike, R., Rao, B., Ray, U., Sarma, J. B., Sharma, M., Sheridan, E., Thirunarayan, M. A., Turton, J., Upadhyay, S., Warner, M., Welfare, W., Livermore, D. M. and Woodford, N. 2010. Emergence of a new antibiotic resistance mechanism in India, Pakistan, and the UK: a molecular, biological, and epidemiological study. *Lancet Infectious Diseases* 10: 597-602.

Kwan, P. S., Barrigas, M., Bolton, F. J., French, N. P., Gowland, P., Kemp, R., Leatherbarrow, H., Upton, M. and Fox, A. J. (2008). Molecular epidemiology of *Campylobacter jejuni* populations in dairy cattle, wildlife, and the environment in a farmland area. *Applied and Environmental Microbiology* 74: 5130-5138.

Lentzsch, P., Moser, P. S. I. and Rieksneuw, B. 2001. Genomic heterogeneity and o-antigenic diversity of *Campylobacter upsaliensis* and *Campylobacter helveticus* strains isolated from dogs and cats in Germany. *Journal of clinical microbiology* 39(7): 2548-2557.

Leonard, E. K., Pearl, D. L., Janecko, N., Weese, J. S., Reid-Smith, R. J., Peregrine A. S. and Finley, R. L. 2011. Factors related to *Campylobacter* spp. carriage in client-

- owned dogs visiting veterinary clinics in a region of Ontario, Canada. *Epidemiology and Infection* 139(10): 1531-1541.
- Li, X., Swaggerty, C. L., Kogut, M. H., Chiang, H. I., Wang, Y., Genovese, K. J., He, H. and Zhou, H. 2010. Gene expression profiling of the local cecal response of genetic chicken lines that differ in their susceptibility to *Campylobacter jejuni* colonization. *PLoS One* 5(7): e11827.
- Luangtongkum, T., Jeon, B., Han, J., Plummer, P., Logue, C. M. and Zhang, Q. 2009. Antibiotic resistance in *Campylobacter*: emergence, transmission and persistence. *Future Microbiology* 4(2): 189-200.
- Luangtongkum, T., Morishita, T. Y., Ison, A. J., Huang, S., McDermott, P. F. and Zhang, Q. 2006. Effect of conventional and organic production practices on the prevalence and antimicrobial resistance of *Campylobacter* spp. in poultry. *Applied and Environmental Microbiology* 72(5): 3600.
- Lynch, Ó. A., Cagney, C., McDowell, D. A. and Duffy, G. 2010. A method for the growth and recovery of 17 species of *Campylobacter* and its subsequent application to inoculated beef. *Journal of Microbiological Methods* 83: 1-7.
- Malher, X., Simon, M., Charnay, V., Déserts, R. D. D., Lehébel, A. and Belloc, C. 2011. Factors associated with carcass contamination by *Campylobacter* at slaughterhouse in cecal-carrier broilers. *International Journal of Food Microbiology* 150(1): 8-13.
- Man, S. M., Ming, S., Leach, S. T., Nahidi, L., Lu, H. K., Norman, J., Day, A. S., Zhang, L. and Mitchell, H. M. 2010. Host attachment, invasion, and stimulation of proinflammatory cytokines by *Campylobacter concisus* and other non-*Campylobacter jejuni* *Campylobacter* species. *Journal of Infectious Diseases* 202(12): 1855-1865.
- Mansouri-Najand, L., Saleha, A. A. and Wai, S. S. 2012. Prevalence of multidrug resistance *Campylobacter jejuni* and *Campylobacter coli* in chickens slaughtered in selected markets, Malaysia. *Tropical Biomedicine* 29(2): 231-238.
- Martinez, J. L., 2009. The role of natural environments in the evolution of resistance traits in pathogenic bacteria. *Proceedings of the Royal Society Biological Sciences* 276: 2521-2530.
- Matsuda, M., Kaneko, A., Stanley, T., Millar, B. C., Miyajima, M., Murphy, P. G. and Moore, J. E. 2003. Characterization of urease-positive thermophilic *Campylobacter* subspecies by multilocus enzyme electrophoresis typing. *Applied and environmental microbiology* 69(6): 3308-3310.



- McCrea, B. A., Tonooka, K. H., VanWorth, C., Atwill, E. R. and Schrader, J. S. 2006. Colonizing capability of *Campylobacter jejuni* genotypes from low- prevalence avian species in broiler chickens. *Journal of Food Protection* 69: 417-420.
- Messens, W., Herman, L., Zutter, L. D., Heyndrickx, M. 2009. Multiple typing for the epidemiological study of contamination of broilers with thermotolerant *Campylobacter*. *Veterinary Microbiology*. 138: 120-131.
- Miller, W. G., Parker, C. T., Rubenfield, M., Mendz, G. L., Wösten, M. M. S. M. Ussery, D. W., Stolz, J. F., Binnewies, T. T., Hallin, P. F., Wang, G., Malek, J. A., Rogosin, A., Stanker, L. H. and Mandrell, R. E. 2007. The complete genome sequence and analysis of the epsilonproteobacterium *arcobacter butzleri*. *PloS One* 2(12): e1358.
- Minihan, D., Whyte, P., O'Mahony, M., Fanning, S., McGill K. and Collins, J. D. 2004. *Campylobacter* spp. in irish feedlot cattle: a longitudinal study involving pre-harvest and harvest phases of the food chain. *Journal of Veterinary Medicine* 51(1): 28-33.
- Montwedi, M. and Ateba., C. N. 2012. Use of the *cdt* gene specific PCR in determining virulence properties of *Campylobacter jejuni* isolated from chicken meat samples obtained in some supermarkets in Mafikeng, NWP, South Africa. *Life Science Journal* 9(3): 2696-2701.
- Morris, C. N., Scully, B. and Garvey, G. J. 1998. *Campylobacter lari* associated with permanent pacemaker infection and bacteremia. *Clinical Infectious Diseases* 27(1): 220-221.
- Nachamkin I., Blaser M. J. 2000. *Campylobacter*, 2nd edition. Washington: American Society for Microbiology.
- Newell, D. G. and Fearnley, C. 2003. Sources of *Campylobacter* colonization in broiler chickens. *Applied and Environmental Microbiology* 69(8): 4343-4351.
- Nkuchia, M. M., Dettinger, L. A., Perry, A., Rogers, P., Reynolds, S. M. and Nachamkin, I. 2012. Culturing stool specimens for *Campylobacter* spp., pennsylvania, USA. *Emerging Infectious Diseases* 18(3): 484-487.
- Noppon, B., Asai, T., Kataoka, Y. and Sawada, T. 2011. Comparison of isolation rates of *Campylobacter* spp. isolated from chicken meats between Japan and Thailand. *Laos Journal on Applied Science* 2:464-467.
- Park, S. F. (2002). The physiology of *Campylobacter* species and its relevance to their role as foodborne pathogens. *International Journal of Food Microbiology*, 74: 177-188.

- Parsons, B. N., Porter, C. J., Ryvar, R., Stavisky, J., Williams, N. J., Pinchbeck, G. L., Birtles, R. J., Christley, R. M., German, A. J., Radford, A. D., Hart, C. A., Gaskell, R. M., Dawson, S. 2010. Prevalence of *Campylobacter* spp. in a cross-sectional study of dogs attending veterinary practices in the UK and risk indicators associated with shedding. *Veterinary Journal* 184(1): 66-70.
- Patel, A., Lloyd, D. H. and Lamport, A. I. 1999. Antimicrobial resistance of feline staphylococci in south-eastern England. *Veterinary Dermatology* 10: 257-261.
- Patriarchi, A., Fox, Á., Maunsell, B., Fanning, S. and Bolton, D. 2011. Molecular characterization and environmental mapping of *Campylobacter* isolates in a subset of intensive poultry flocks in Ireland. *Foodborne Pathogens and Disease* 8(1): 99-108.
- Persson, S. and Olsen, K. E. P. 2005. Multiplex PCR for identification of *Campylobacter coli* and *Campylobacter jejuni* from pure cultures and directly on stool samples. *Journal of Medical Microbiology* 54: 1043-7.
- Pezzotti, G., Serafin, A., Luzzi, I., Mioni, R., Milan, M. and Perin, R. 2003. Occurrence and resistance to antibiotics of *Campylobacter jejuni* and *Campylobacter coli* in animals and meat in northeastern Italy. *International Journal of Food Microbiology* 82: 281-287.
- Pielsticker, C., Glünder, G. and Rautenschlein, S. 2012. Colonization properties of *Campylobacter jejuni* in chickens. *European Journal of Microbiology and Immunology* 2(1): 61-65.
- Quessy, S. and Messier, S. 1992. Prevalence of *Salmonella* spp., *Campylobacter* spp., and *Listeria* spp. in ring-billed gulls (*Larus delawarensis*). *Journal of Wildlife Diseases* 28(4): 526-531.
- Rahimi, E., Chakeri, A. and Tajbakhsh, E. 2011. Detection of *Campylobacter* species in feces of persian sheepdogs, pigeons and squirrels. *Global Veterinaria* 7(4): 365-369.
- Ramabu, S. S., Boxall, N. S., Madie, P., Fenwick, S. G. 2004. Some potential sources for transmission of *Campylobacter jejuni* to broiler chickens. *Letters in Applied Microbiology* 39: 252-256.
- Ramos, R., Cerd'a-Cu'ellar, M., Ramírez, F., Jover, L. and Ruiz, X. 2010. Influence of Refuse Sites on the Prevalence of *Campylobacter* spp. and *Salmonella* Serovars in Seagulls. *Applied and Environmental Microbiology* 76(9): 3052-3056.
- Rapp, D. and Ross, C. M. 2012. Prevalence of six *Campylobacter* species in a New Zealand dairy goat herd. *New Zealand Journal of Agricultural Research* 55(3): 235-240.

- Rejab, S. B. M., Zessin, K. H., Fries, R. and Patchanee, P. 2012. *Campylobacter* in chicken carcasses and slaughterhouses in Malaysia. *Southeast Asian Journal of Tropical Medicine* 43(1): 96-104.
- Rodgers, J. D., Lawes, J. R., Vidal, A. B., Ellis-Iversen, J., Ridley, A., Pleydell, E. J., Powell, L. F., Toszeghy, M., Stapleton, K. and Clifton-Hadley, F. A. 2012. Characteristics and comparative performance of direct culture, direct PCR and enumeration methods for detection and quantification of *Campylobacter* spp. in broiler caeca. *Veterinary Microbiology* 159(3-4): 390-396.
- Rodrigues, L. C., Cowden, J. M., Wheeler, J. G., Sethi, D., Wall, P. G., Cumberland, P., Tompkins, D. S., Hudson, M. J., Roberts, J. A. and Roderick, P. J. 2001. The study of infectious intestinal disease in England: risk factors for cases of infectious intestinal disease with *Campylobacter jejuni* infection. *Epidemiology and Infection* 127: 185-193.
- Rodríguez, S. and Araujo, R. 2010. Occurrence of thermotolerant *Campylobacter* species in surface waters of a Mediterranean area and in its prevailing pollution sources. *Journal of Applied Microbiology* 109(3): 1027-1034.
- Rosef, O., Johnsen, G., Stølan, A. and Klæboe, H. 2008. Similarity of *Campylobacter lari* among human, animal, and water isolates in Norway. *Foodborne Pathogens and Disease* 5(1): 33-39.
- Saleha A.A. 2004. Epidemiological study on the colonization of chickens with *Campylobacter* in broiler farms in Malaysia: possible risk and management factors. *International Journal of Poultry Science* 3(2): 129-134.
- Saleha, A.A., Christopher, J.R.G., Aini, I and Ganapathy, K. 2001. Occurrence of *Campylobacter* in free flying birds. Proceeding of 2<sup>nd</sup> International Conference 13<sup>th</sup> VAM conference and CVA Australasia/Oceania regional Symposium, 27-30 August, Kuala Lumpur, .70-71.
- Salihu, M. D. A. A. M., Abdulkadir, J. U. and Kolawale, A. 2010. Survey of thermophilic *Campylobacter* species in cats and dogs in north-western Nigeria. *Veterinaria Italiana* 46(4): 425-430.
- Salihu, M. D., Junaidu, A. U., Oboegbulem, S. I. and Egwu, G. O. 2009. Prevalence and biotypes of *Campylobacter* species isolated from sheep in sokoto state, Nigeria. *International Journal of Animal and Veterinary Advances* 1(1): 6-9.
- Sallam, K. I., 2007. Prevalence of *Campylobacter* in chicken and chicken by-products retailed in Sapporo area, Hokkaido, Japan. *Food Control* 18(9): 1113-1120.
- Sanad, Y. M., Kassem, I. I., Abley, M., Gebreyes, W., LeJeune, J. T. and Rajashekara, G. 2011. Genotypic and phenotypic properties of cattle-associated

*Campylobacter* and their implications to public health in the USA. *PLoS ONE* 6(10): e25778.

- Scarcelli, E., Piatti, R. M., Harakava, R., Miyashiro, S., Campos, F. R., Souza, M. C. A., Cardoso, M. V., Teixeira, S. R. and Genovez, M. E. 2009. Use of pcr-rflp of the *fla* a gene for detection and subtyping of *Campylobacter jejuni* strains potentially related to guillain–barré syndrome, isolated from humans and animals. *Brazilian Journal of Microbiology* 40: 952-959.
- Schwartz, T., Kohnen, W., Jansen, B. and Obst, U. 2003. Detection of antibiotic-resistant bacteria and their resistance genes in wastewater, surface water and drinking water biofilms. *FEMS Microbiology Ecology* 43: 325-335.
- Sensale, M., Cuomo, A., Dipineto, L., Santaniello, A., Calabria, M., Menna, L. F. and Fioretti, A. 2006. Survey of *Campylobacter jejuni* and *Campylobacter coli* in different taxa and ecological guilds of migratory birds. *Italian Journal of Animal Science* 5: 291-294.
- Silva, J., Leite, D., Fernandes, M., Mena, C., Gibbs, P. A. and Teixeira, P. 2011. *Campylobacter* spp. as a foodborne pathogen: a review. *Frontiers in Microbiology* 2: 200.
- Silva, J., Leite, D., Fernandes, M., Mena, C., Gibbs, P. A. and Teixeira, P. 2011. *Campylobacter* spp. as a foodborne pathogen: A Review. *Frontiers in Microbiology* 2: 1-12.
- Sippy, R., Sandoval-Green, C. M. J., Sahin, O., Plummer, P., Fairbanks, W. S., Zhang, Q. and Blanchong, J. A. 2012. Occurrence and molecular analysis of *Campylobacter* in wildlife on livestock farms. *Veterinary Microbiology* 157(3-4): 369-375.
- Skjøt-Rasmussen, L., Ethelberg, S., Emborg, H. D., Agersø, Y., Larsen, L. S., Nordentoft, S., Olsen, S. S., Ejlersen, T., Holt, H., Nielsen, E. M. and Hammerum, A. M. 2009. Trends in occurrence of antimicrobial resistance in *Campylobacter jejuni* isolates from broiler chickens, broiler chicken meat, and human domestically acquired cases and travel associated cases in Denmark. *International Journal of Food Microbiology* 131: 277-279.
- Smith, D. L., Harris, A. D., Johnson, J. A., Silbergeld, E. K. and Morris, J. G., Jr. 2002. Animal antibiotic use has an early but important impact on the emergence of antibiotic resistance in human commensal bacteria. *National Academy of Sciences* 99(9): 6434-6439.
- Smith, J. L., and Fratamico, P. M. (2010). Fluoroquinolone resistance in *Campylobacter*. *Journal of Food Protection* 73(6): 1141-1152.

- Sparks, N. H. C. 2009. The role of the water supply system in the infection and control of *Campylobacter* in chicken. *World's Poultry Science Journal* 65(03): 459.
- Sproston, E. L., Ogden, I. D., MacRae, M., Dallas, J. F., Sheppard, S. K., Cody, A. J., Colles, F. M., Wilson, M. J., Forbes, K. I. and Strachan, N. J. C. 2011. Temporal variation and host association in the *Campylobacter* population in a longitudinal ruminant farm study. *Applied and Environmental Microbiology* 77(18): 6579-86.
- Strother, K. O., Steelman, C. D. and Gbur, E. E. 2005. Reservoir Competence of Lesser Mealworm (Coleoptera: Tenebrionidae) for *Campylobacter jejuni* (Campylobacterales: Campylobacteraceae). *Journal of Medical Entomology* 42(1):42-47.
- Tang, J. Y. H., Ghazali, F. M., Saleha, A. A., Nishibuchi, M. and Son, R. 2009. Comparison of thermophilic *Campylobacter* spp. occurrence in two types of retail chicken samples. *International Food Research Journal* 16: 277-288.
- Tang, J. Y. H., Saleha, A. A., Abu, J., Ghazali, F. M., Chilek, T. Z. T., Ahmad, N., Sandra, A., Nishibuchi, M. and Radu, S. 2010. Thermophilic *Campylobacter* spp. Occurrence on Chickens at Farm, Slaughter House and Retail. *International Journal of Poultry Science* 9(2): 134-138.
- Taylor E. V, Herman K. M, Ailes E. C, Fitzgerald C., Yoder J. S., Mahon B. E. and Tauxe R. V. 2012. Common source outbreaks of *Campylobacter* infection in the USA, 1997–2008. *Epidemiology and Infection Journal* 15:1-10.
- Thakur, S. and W. A. Gebreyes. 2010. Phenotypic and genotypic heterogeneity of *Campylobacter coli* within individual pigs at farm and slaughter in the US. *Zoonoses and Public Health* 57:100-106.
- Thomas, D. K., Lone, A. G., Selinger, L. B., Taboada, E. N., Uwiera, R. R. E., Abbott, D. W. and Inglis, G. D. 2014. Comparative variation within the genome of *Campylobacter jejuni* nctc 11168 in human and murine hosts . *PLoS ONE* 9(2): e88229. doi:10.1371/journal.pone.0088229.
- Tsiodras, S., Kelesidis, T., Kelesidis, I., Bauchinger, U. and Falagas, M. E. 2008. Human infections associated with wild birds. *Journal of Infection* 56(2): 83-98.
- Vally, H. G. H., Scallan, E., Kirk, M. D. and Angulo, F. J. 2009. Higher rate of culture-confirmed *Campylobacter* infections in Australia than in the USA: Is this due to differences in healthcare-seeking behaviour or stool culture frequency?. *Epidemiology and Infection* 137(12): 1751-8.
- Vandamme, P. and Ley, J. D. 1991. Proposal for a new family, campylobacteraceae. *International journal of Systematic Bacteriology* 41(3): 451-455.

- Vandamme, P., Debruyne, L., Brandt, E. D. and Falsen, E. 2010. Reclassification of *Bacteroides ureolyticus* as *Campylobacter ureolyticus* Comb. Nov., and emended description of the genus *Campylobacter*. *International Journal of Systematic and Evolutionary Microbiology* 60(9): 2016-2022.
- Waage, A. S., Vardund, T., Lund, V. and Kapperud, G. 1999. Detection of small numbers of *Campylobacter jejuni* and *Campylobacter coli* cells in environmental water, sewage, and food samples by a seminested PCR assay. *Applied and Environmental Microbiology* 65(4): 1636-1643.
- Waldenström, J., Broman, T., Carlsson, I., Hasselquist, D., Achterberg, R. P., Wagenaar, J. A. and Olsen, B. 2002. Prevalence of *Campylobacter jejuni*, *Campylobacter lari*, and *Campylobacter coli* in different ecological guilds and taxa of migrating birds. *Applied and Environmental Microbiology* 68(12): 5911-5917.
- Wingstrand, A., Neimann, J., Engberg, J., Nielsen, E. M., Gerner-Smidt, P., Wegener, H. C., Molbak, K. 2006. Fresh chicken as main risk factor for campylobacteriosis, Denmark. *Emerging Infectious Diseases journal* 12(2): 280-285.
- Xavier, G., Winy, M., Dirk, H., Koen, G., Hartnett, Emma, H. and Jacques, V. 2008. Economics of reducing *Campylobacter* at different levels within the belgian poultry meat Chain. *Journal of Food Protection* (3): 468-661.
- Xu, X., Li, Y., Zhao, H., Wen, S. Y., Wang, S. Q., Huang, J., Huang, K. L. and Luo, Y. B., 2005. Rapid and reliable detection and identification of GM events using multiplex PCR coupled with oligonucleotide microarray. *Journal of Agricultural food chemistry* 53(10):3789-3794.
- Yamazaki-Matsune, W., Taguchi, M., Seto, K., Kawahara, R., Kawatsu, K., Kumeda, Y., Kitazato, M., Nukina, M., Misawa, N. and Tsukamoto, T. 2007. Development of a multiplex PCR assay for identification of *Campylobacter coli*, *Campylobacter fetus*, *Campylobacter hyointestinalis* subsp. *hyointestinalis*, *Campylobacter jejuni*, *Campylobacter lari* and *Campylobacter upsaliensis*. *Journal of Medical Microbiology* 56: 1467-1473.
- Zakaria, M., Chong Leong, P. and Chin Aik, Y., 2008. A photographic guide to birds of ayer hitam forest reserve, Selangor. Penerbit Universiti Putra Malaysia. Selangor.
- Zendehbad, B., Arian, A. A., Alipour, A. 2013. Identification and antimicrobial resistance of *Campylobacter* species isolated from poultry meat in Khorasan province, Iran. *Food Control* 32: 724-727.
- Zhao, S., Young, S. R., Tong, E., Abbott, J. W., Womack, N., Friedman S. L. and McDermott, P. F. 2010. Antimicrobial resistance of *Campylobacter* isolates from retail meat in the United States between 2002 and 2007. *Applied and Environmental Microbiology* 76(24): 7949-7956.

Zilbauer, M., Dorrell, N., Wren, B. W. and Bajaj-Elliott, M. 2008. *Campylobacter jejuni* mediated disease pathogenesis: an update. *Royal Society of Tropical Medicine and Hygiene* 102(2): 123-129.

Zweifel, C., Scheu, K. D., Keel, M., Renggli, F. and Stephan, R. 2008. Occurrence and genotypes of *Campylobacter* in broiler flocks, other farm animals and the environment during several rearing periods on selected poultry farms. *International Journal of Food Microbiology* 125(2): 182-187.

