



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION AND PRESENCE OF *ica* GENES IN COAGULASE
NEGATIVE STAPHYLOCOCCI ISOLATES IN A PUBLIC HOSPITAL IN
MALAYSIA***

SUBASHINI A/P VNAYAKAM

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NEGATIVE STAPHYLOCOCCI ISOLATES IN A PUBLIC HOSPITAL IN
MALAYSIA**

By

SUBASHINI A/P VNAYAKAM

**Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfilment of the
Requirements for the Master of Science**

October 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

CHARACTERIZATION AND PRESENCE OF *ica* GENES IN COAGULASE-NEGATIVE STAPHYLOCOCCI ISOLATES IN A PUBLIC HOSPITAL IN MALAYSIA

By

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October 2017

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Coagulase-negative staphylococci (CoNS) are often considered as contaminants or normal skin flora but in the past few decades, they had emerged as pathogenic bacteria causing serious infections especially in patients with prosthetic medical devices. Their ability to produce biofilm on polymer surfaces results in treatment failure as it increases the resistance to antibiotics and host defense. Biofilms formation are mediated by polysaccharide intercellular adhesin (PIA), encoded by intercellular adhesion (*ica*) genes and it has been postulated that the presence of *ica* genes in CoNS is associated with antimicrobial resistance and responsible for catheter and medical device-related sepsis. The aim of this study is to determine the species distribution, antibiotic susceptibility pattern and to detect the presence of *ica* genes (*icaA*, *icaD*, *icaB*, and *icaC*) among CoNS isolated from blood culture. This cross-sectional study was conducted between January 2015 and December 2015, with CoNS isolates obtained from the blood cultures of patients at Hospital Serdang. Coagulase negative staphylococci were identified by gram staining, catalase and coagulase test, followed by species identification by Analytical Profile Index (API) Staph identification strips. Antimicrobial susceptibility testing was performed using Kirby Bauer method interpreted according to the Clinical and Laboratory Standards Institute guidelines. The presence of *ica* genes were detected using multiplex polymerase chain reaction. *Staphylococcus epidermidis* dominated the total number of species isolated (n=64, 40.0%). A total of 160 CoNS, 72.5% were resistant to penicillin and 60% were methicillin-resistant (MR) CoNS. Majority of CoNS harbored *icaD* (59.3%) and in 4 (2.5%) strains, all *icaADBC* genes were observed. Detection of *ica* genes indicates CoNS are able to produce biofilm that causes serious nosocomial infections and should not always be reported as colonizer or contaminants.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**KARAKTERISASI DAN KEHADIRAN GENE *ica* DALAM ISOLASI
COAGULASE-NEGATIF STAPHYLOCOCCI DARIPADA HOSPITAL AWAM
DI MALAYSIA**

Oleh

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Staphylococci-koagulase negatif (CoNS) sering dianggap sebagai kontaminasi atau flora kulit tetapi dalam beberapa dekad yang lalu, ia telah muncul sebagai bakteria patogenik yang menyebabkan jangkitan serius terutamanya pada pesakit yang menggunakan peranti perubatan palsu. Keupayaan mereka menghasilkan biofilem pada permukaan polimer menyebabkan kegagalan rawatan kerana ia meningkatkan daya tahan terhadap antibiotik dan imunitasi pesakit. Pembentukan biofilem dibantu oleh *polysaccharide intercellular adhesion* (PIA), yang dikodkan oleh gen perekatan (*ica*) antara sel dan telah dikenalpasti bahawa kehadiran gen *ica* pada CoNS dikaitkan dengan rintangan antimikrob dan bertanggungjawab untuk sepsis yang berkaitan dengan peranti perubatan. Tujuan kajian ini adalah untuk menentukan taburan spesies, corak kerintangan antibiotik dan untuk mengesan kehadiran gen *ica* (*icaA*, *icaD*, *icaB*, dan *icaC*) di kalangan CoNS dalam sampel darah. Kajian rentas keratan ini dijalankan antara Januari 2015 dan Disember 2015, dengan isolasi CoNS yang diperolehi dari kultur darah pesakit di Hospital Serdang. Staphylococci negatif koagulase telah dikenalpasti oleh ujian pewarnaan gram, catalase dan koagulase, diikuti dengan identifikasi spesies oleh Strip Pengenalan Staph Analisis Profil (API). Ujian kerintangan antimikrob dilakukan dengan menggunakan kaedah Kirby Bauer yang ditafsirkan mengikut garis panduan Institut Standardisasi Klinikal dan Makmal. Kehadiran gen *ica* dikesan menggunakan tindak balas rantaian polimerase multiplex. *Staphylococcus epidermidis* menguasai jumlah spesies yang terpencil (n = 64, 40.0%). Sebanyak 160 CoNS, 72.5% menunjukkan rintangan terhadap penicillin dan 60% adalah *methicillin-resistant* (MR) CoNS. Majoriti CoNS mempunyai *icaD* (59.3%) dan dalam 4 (2.5%) strain, semua gen *icaADBC* dikenalpasti. Pengesanan gen *ica* memutuskan bahawa CoNS dapat menghasilkan biofilem yang menyebabkan jangkitan nosokomial yang serius dan tidak seharusnya dilaporkan sebagai kolonizer atau kontaminasi.

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I certify that a Thesis Examination Committee has met on 23 October 2017 to conduct the final examination of Subashini a/p Vnayakam on her thesis entitled "Characterization and Presence of *ica* Genes in Coagulase-Negative Staphylococci Isolates in a Public Hospital in Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

°C	degree Celcius
%	Percentage
α	Alpha
β	Beta
Aap	Accumulation of associated protein
API	Analytical Profile Index
ATCC	American Type Culture Collection
Bap	Biofilm-associated protein
BLAST	basic local alignment search tool
CAPD	Continuous ambulatory peritoneal dialysis
CDC	Centre for Disease Control
CLSI	Clinical and Laboratory Standards Institute
CoNS	coagulase negative <i>staphylococci</i>
CRBSI	catheter-related blood stream infection
CRF	coagulase-reacting factor
DNA	deoxyribonucleic acid
EPS	Extracellular Polymeric Substance
E-test	epsilometer test
H ₂ O ₂	hydrogen peroxide
<i>ica</i>	intercellular adhesion
MIC	minimum inhibitory concentration
MR-CoNS	Methicillin-Resistant coagulase negative <i>Staphylococci</i>
NVE	Native valve endocarditis

PCR	polymerase chain reaction
PIA	polysaccharide intercellular adhesion
PNAG	Poly-N-acetylglucosamine
RNA	ribonucleic acid
UTI	urinary tract infection



CHAPTER 1

INTRODUCTION

1.1 Background of the study

The staphylococcus is an aerobic gram-positive spherical cell which are arranged in grapelike irregular clusters. The genus *Staphylococcus* is currently divided into 39 species (Euzebay, 2007). The staphylococci are the most frequently isolated pathogen in microbiology laboratory which causes a range of diseases in hospital-related infections. They are further classified generally into two major groups which are coagulase-positive staphylococci (e.g. *Staphylococcus aureus*) and coagulase-negative staphylococci (e.g. *Staphylococcus epidermidis*). Coagulase-negative staphylococci comprises of a variety of species and many different strains and among the predominant species isolated in these infections are *Staphylococcus epidermidis*, *Staphylococcus lugdunensis*, *Staphylococcus warneri*, *Staphylococcus hominis* and *Staphylococcus saprophyticus* (Garza-Gonzalez *et al.*, 2010).

For a long time, coagulase-negative staphylococci were considered as non-pathogenic as it is present on outer body surfaces as normal micro flora which naturally colonizes humans. Meanwhile, isolated CoNS were not further investigated as they were grown as a result of contamination (Majumder *et al.*, 2014). Nowadays, they are often associated with chronic infections such as urinary tract infection, osteomyelitis, native and prosthetic valve endocarditis and intravenous catheter infection (Becker *et al.*, 2014).

Coagulase-negative staphylococci emerged as pathogens due to rise in use of invasive procedures both temporarily inserted and permanently implanted such as implanted medical devices, joint prostheses, shunts and intravascular catheters especially in immunosuppressed patients, infants, transplant patients, intravenous drug abusers and geriatric patients (Al Wohoush *et al.*, 2011). The use of indwelling medical devices is important in the treatment of critically and chronically ill patients, however bacterial colonization of implanted foreign material can cause major nosocomial infection (Mathur *et al.*, 2006). Thus, biomaterial-associated infections (BAI) contribute to the increase of hospital-acquired infections.

Regardless of low virulence, CoNS are considered clinically significant due to their ability to adhere to and to form biofilms on the surface of medical devices (Los *et al.*, 2010). In addition, production of biofilm serves as a protection of bacteria from the effects of antibiotics which results in antimicrobial resistance (Bozkurt *et al.*, 2009).

In agreement with that, the capacity of certain strains to form a biofilm consists of multi-layered cell clusters or intercellular adhesion (*ica*) embedded in a matrix of extracellular

polysaccharide, which facilitates the adherence of these microbial to biomedical surfaces made it recognized as a potential pathogen (Piette and Verschraegen, 2009). This multi-step mechanism is one of the major factors involved in the pathogenicity of catheter-related infections caused by CoNS. Investigation of the second stage of biofilm formation has demonstrated that cell aggregation and biofilm accumulation are mediated by the products of the chromosomal *ica* gene locus, which comprises four intercellular adhesion genes (*icaA*, *icaB*, *icaC* and *icaD*) and one regulator gene (*icaR*) (Cafiso *et al.*, 2004). These genes influence the production of polysaccharide intercellular adhesion (PIA), which acts as an intercellular adhesion on hydrophilic surfaces.

In the clinical laboratories, identification of coagulase-negative staphylococcus is often limited because distinction between pathogenic and contaminating isolates is difficult to perform (Huebner and Goldmann, 1999). The severity of infections differs in different species which emphasizes the need of species identification for a better choice of treatment. However, rapid screening tests could only distinguish *S.aureus*, while non *S.aureus* isolates are reported as CoNS (Hirota *et al.*, 2011). Therapeutically, CoNS infections remains challenging to clinician due to the large proportion of isolates as limited clinical trials are available to determine optimal therapy.

1.2 Need and Significance

This study could be able to fill in the gap of knowledge in understanding the significance of CoNS which contribute to nosocomial infections especially in prosthetic devices and indwelling catheters.

1.3 Research Question

Does presence of intercellular adhesion (*ica*) genes determine the clinical significance of CoNS isolated from blood culture?

1.4 Objectives

1.4.1 General objective

To determine the characterization of coagulase-negative staphylococci isolated from blood culture.

1.4.2 Specific objectives

- i. To determine the distribution of species of coagulase-negative staphylococci isolated from blood culture.

- ii. To study the antibiotic susceptibility patterns of coagulase-negative staphylococci species.
- iii. To detect the intercellular adhesion (*ica*) genes among the isolated coagulase-negative staphylococci species.

1.5 Hypothesis

There is presence of intercellular adhesion (*ica*) genes in coagulase-negative staphylococci which makes it clinically significant in nosocomial infections.



REFERENCES

- Al Wohoush, I., Rivera, J., Cairo, J., Hachem, R., & Raad, I. (2011). Comparing clinical and microbiological methods for the diagnosis of true bacteraemia among patients with multiple blood cultures positive for coagulase-negative staphylococci. *Clinical Microbiology and Infection*, 17(4), 569-571.
- Angeles Argudín, M., Vanderhaeghen, W., Vandendriessche, S., Vandecandelaere, I., Denis, O., Coenye, T., & Butaye, P. (2015). Biofilm formation of *ica* operon positive *Staphylococcus epidermidis* from different sources. *Apmis*, 123(12), 1081-1089.
- Archer, G. L., & Climo, M. W. (1994). Antimicrobial susceptibility of coagulase negative staphylococci. *Antimicrobial Agents and Chemotherapy*, 38(10), 2231.
- Arciola, C. R., Baldassarri, L., & Montanaro, L. (2001). Presence of *ica* A and *ica* D genes and slime production in a collection of Staphylococcal strains from catheter-associated infections. *Journal of Clinical Microbiology*, 39(6), 2151-2156.
- Bannerman, T. L. (2003). Staphylococcus, Micrococcus, and other catalase positive cocci that grow aerobically. *Manual of Clinical Microbiology, American Society Microbiology*, 384-404.
- Becker, K., Heilmann, C., & Peters, G. (2014). Coagulase-negative staphylococci. *Clinical microbiology reviews*, 27(4), 870-926.
- Bozkurt, H., Kurtoglu, M. G., Bayram, Y., Keşli, R., & Berktaş, M. (2009). Correlation of slime production investigated via three different methods in coagulase-negative staphylococci with crystal violet reaction and antimicrobial resistance. *Journal of International Medical Research*, 37(1), 121-128.
- Brooks, G.F., Carroll, K.C., Butel, J.S., Morse, S.A. and Mietzner, T.A. (2010). In Jawetz, Melnick and Adelbergs, 25th Edition, Medical Microbiology McGraw Hill, (pp. 185-191).
- Cafiso, V., Bertuccio, T., Santagati, M., Campanile, F., Amicosante, G., Perilli, M. G. Perilli, L. Selan, M. Artini, G. Nicoletti, and S. Stefani, S. (2004). Presence of the *ica* operon in clinical isolates of *Staphylococcus epidermidis* and its role in biofilm production. *Clinical Microbiology and Infection*, 10(12), 1081-1088.
- Camposcia, D., Montanaro, L., & Arciola, C. R. (2013). A review of the biomaterials technologies for infection-resistant surfaces. *Biomaterials*, 34(34), 8533-8554.

- Chandran, A. U., & Rennie, R. (2005). Routine antimicrobial susceptibility testing of coagulase- negative staphylococci isolated from blood cultures: is it necessary? *Clinical Microbiology and Infection*, 11(12), 1037-1040.
- Cherifi, S., Byl, B., Deplano, A., Nagant, C., Nonhoff, C., Denis, O., & Hallin, M. (2014). Genetic characteristics and antimicrobial resistance of *Staphylococcus epidermidis* isolates from patients with catheter-related bloodstream infections and from colonized healthcare workers in a Belgian hospital. *Annals of Clinical Microbiology and Antimicrobials*, 13(1), 1.
- Clinical and Laboratory Standards Institute (CLSI): Methods for Antimicrobial Dilution and Disk Susceptibility Testing of Infrequently Isolated or Fastidious Bacteria; Approved Guideline. Second Edition. *CLSI document M45-A2*. Wayne, PA 2010.
- Clinical and Laboratory Standards Institute (CLSI): Performance Standards for Antimicrobial Disk Susceptibility Tests. Twelfth Edition. *CLSI document M02-A12*. Wayne, PA 2015.
- Cogen, A.L., Nizet, V., Gallo, R.L. (2008) Skin microbiota: A source of disease or defence? *British Journal of Dermatology*, 158, 442–455.
- Cunha, M. D. L. R., Sinzato, Y. K., & Silveira, L. V. (2004). Comparison of methods for the identification of coagulase negative staphylococci. *Memórias do Instituto Oswaldo Cruz*, 99(8), 855-860.
- Daniel, W. W., & Cross, C. L. (1995). Biostatistics: a foundation for analysis in the health sciences.
- Daniel, B., Saleem, M., Naseer, G., & Fida, A. (2014). Significance of *Staphylococcus haemolyticus* in hospital acquired infections. *J Pioneer Med Sci*, 4, 119-25.
- de la Mària, C. G., Cervera, C., Pericàs, J. M., Castañeda, X., Armero, Y., Soy, D., Almela, M., Ninot, S., Falces, C., Mestres, C.A. and Gatell, J. M. (2015). Epidemiology and Prognosis of Coagulase-Negative Staphylococcal Endocarditis: Impact of Vancomycin Minimum Inhibitory Concentration. *PloS one*, 10(5), e0125818.
- Edmond, M. B., Wallace, S. E., McClish, D. K., Pfaller, M. A., Jones, R. N., & Wenzel, R. P. (1999). Nosocomial bloodstream infections in United States hospitals: a three- year analysis. *Clinical infectious diseases*, 29(2), 239-244.
- El- Mahallawy, H. A., Loutfy, S. A., El- Wakil, M., El- Al, A. K. A., & Morcos, H. (2009). Clinical implications of *ica A* and *ica D* genes in coagulase negative staphylococci and *Staphylococcus aureus* bacteremia in febrile neutropenic pediatric cancer patients. *Pediatric blood & cancer*, 52(7), 824-828.

- Esteban, J., Molina-Manso, D., Spiliopoulou, I., Cordero-Ampuero, J., Fernández Roblas, R., Foka, A., & Gómez-Barrena, E. (2010). Biofilm development by clinical isolates of *Staphylococcus spp.* from retrieved orthopedic prostheses. *Acta orthopaedica*, 81(6), 674-679.
- Euzeby, J. P. (2007). List of prokaryotic names with standing in nomenclature. Retrieved from: <http://www.bacterio.cict.fr/>.
- Fey, P. D., & Olson, M. E. (2010). Current concepts in biofilm formation of *Staphylococcus epidermidis*. *Future microbiology*, 5(6), 917-933.
- Fiebelkorn, K. R., Crawford, S. A., McElmeel, M. L., & Jorgensen, J. H. (2003). Practical disk diffusion method for detection of inducible clindamycin resistance in *Staphylococcus aureus* and coagulase-negative staphylococci. *Journal of Clinical Microbiology*, 41(10), 4740-4744.
- Fredheim, E. G. A., Klingenberg, C., Rohde, H., Frankenberger, S., Gaustad, P., Flægstad, T., & Sollid, J. E. (2009). Biofilm formation by *Staphylococcus haemolyticus*. *Journal of Clinical Microbiology*, 47(4), 1172-1180.
- Gad, G. F. M., El-Feky, M. A., El-Rehewy, M. S., Hassan, M. A., Abolella, H., & El Baky, R. M. A. (2009). Detection of *icaA*, *icaD* genes and biofilm production by *Staphylococcus aureus* and *Staphylococcus epidermidis* isolated from urinary tract catheterized patients. *The Journal of Infection in Developing Countries*, 3(05), 342-351.
- Garza-González, E., Morfin-Otero, R., Macedo, P., Gonzalez, G. M., Llaca-Díaz, J. M., Perez-Gómez, R., & Rodriguez Noriega, E. (2010). Evaluation of Sensititre plates for identification of clinically relevant coagulase-negative staphylococci. *Journal of Clinical Microbiology*, 48(3), 963-965.
- Geraghty, L., Booth, M., Rowan, N., & Fogarty, A. (2013). Investigations on the efficacy of routinely used phenotypic methods compared to genotypic approaches for the identification of staphylococcal species isolated from companion animals in Irish veterinary hospitals. *Irish Veterinary Journal*, 66(1), 1.
- Gerke, C., Kraft, A., Süßmuth, R., Schweitzer, O., & Götz, F. (1998). Characterization of the Acetyl-glucosaminyl transferase Activity Involved in the Biosynthesis of the *Staphylococcus epidermidis* Polysaccharide Intercellular Adhesin. *Journal of Biological Chemistry*, 273(29), 18586-18593.
- Gill, V. J., Selepak, S. T., & Williams, E. C. (1983). Species identification and antibiotic susceptibilities of coagulase negative staphylococci isolated from clinical specimens. *Journal of clinical microbiology*, 18(6), 1314-1319.
- Götz, F. (2002). *Staphylococcus* and biofilms. *Molecular microbiology*, 43(6), 1367-1378.

- Harris, L. G., Murray, S., Pascoe, B., Bray, J., Meric, G., Mageiros, L., Wilkinson, T.S., Jeeves, R., Rohde, H., Schwarz, S. and De Lencastre, H. (2016). Correction: Biofilm morphotypes and population structure among *Staphylococcus epidermidis* from commensal and clinical samples. *PLoS one*, 11(4), e0154510.
- Healthcare-associated Infections. (2010, November 24). Retrieved April 18, 2017, from <https://www.cdc.gov/hai/settings/lab/labdetectioncoagulase-negative.html>
- Health Protection Agency (2007). Identification of *Staphylococcus* species, *Micrococcus* species and *Rothia* species. National Standard Method BSOP ID, 7(2.1). Retrieved from http://www.hpa.standardmethods.org.uk/pdf_sops.asp.
- Hedayati, S., Eftekhari, F., & Hosseini, S. (2015). Biofilm Formation by Bacteria Isolated from Intravenous Catheters. *Journal of Medical Bacteriology*, 3(3-4), 26-31.
- Herwaldt, L. A., Geiss, M., Kao, C., & Pfaller, M. A. (1996). The positive predictive value of isolating coagulase-negative staphylococci from blood cultures. *Clinical infectious diseases*, 22(1), 14-20.
- Hirota, S., Sasaki, T., Kuwahara-Arai, K., & Hiramatsu, K. (2011). Rapid and accurate identification of human associated staphylococci by use of multiplex PCR. *Journal of Clinical Microbiology*, 49(10), 3627-3631.
- Hitzenbichler, F., Simon, M., Salzberger, B., & Hanses, F. (2017). Clinical significance of coagulase-negative staphylococci other than *S. epidermidis* blood stream isolates at a tertiary care hospital. *Infection*, 45(2), 179-186.
- Huebner J., Goldman DA. (1999). Coagulase-negative staphylococci: role as pathogens. *Ann. Rev. Med* 50, 223-236.
- Karlowsky, J. A., Jones, M. E., Draghi, D. C., Thornsberry, C., Sahm, D. F., & Volturo, G. A. (2004). Prevalence and antimicrobial susceptibilities of bacteria isolated from blood cultures of hospitalized patients in the United States in 2002. *Annals of clinical microbiology and antimicrobials*, 3(1), 1.
- Kim, J. H., Kim, C. H., Hacker, J., Ziebuhr, W., Lee, B. K., & Cho, S. H. (2008). Molecular characterization of regulatory genes associated with biofilm variation in a *Staphylococcus aureus* strain. *Journal of Microbiology Biotechnology*, 18(1), 28-34.
- Kim, S. D., McDonald, L. C., Jarvis, W. R., McAllister, S. K., Jerris, R., Carson, L. A., & Miller, J. M. (2000). Determining the Significance of Coagulase Negative Staphylococci Isolated from Blood Cultures at a Community Hospital A Role for Species and Strain Identification. *Infection Control & Hospital Epidemiology*, 21(03), 213-217.

- Kloos, W. E., & Schleifer, K. H. (1975). Simplified scheme for routine identification of human *Staphylococcus* species. *Journal of Clinical Microbiology*, 1(1), 82-88.
- Kloos, W. E., and D. W. Lambe. (1991). in A. Balows, W. J. Hausler, Jr., K. L. Herrmann, H. D. Isenberg, and H. J. Shadomy (ed.), *Manual of clinical microbiology*, 5th ed. American Society for Microbiology, Washington D.C. p.222–237.
- Koksal, F., Yasar, H., & Samasti, M. (2009). Antibiotic resistance patterns of coagulase-negative staphylococcus strains isolated from blood cultures of septicemic patients in Turkey. *Microbiological research*, 164(4), 404-410.
- Kresken, M., & Hafner, D. (1999). Drug resistance among clinical isolates of frequently encountered bacterial species in central Europe during 1975-1995. *Infection*, 27, S2-S8.
- Lachachi, M., Hassaine, H., Nayme, K., Bellifa, S., M'hamedi, I., Terki, I. K., & Timinouni, M. (2014). Detection of biofilm formation, *ica* ADBC gene and investigation of toxin genes in *Staphylococcus* spp. strain from dental unit waterlines, University Hospital Center (UHC) Tlemcen Algeria. *African Journal of Microbiology Research*, 8(6), 559-565.
- Lange, C. C., Brito, M. A. V. P., Reis, D. R. L., Machado, M. A., Guimara, A. S., Azevedo, A. L. S., Salles, É.B., Alvim, M.C., Silva, F.S. and Meurer, I. R. (2015). Species-level identification of staphylococci isolated from bovine mastitis in Brazil using partial 16S Rna sequencing. *Veterinary Microbiology*. doi:http://dx.doi.org/10.1016/j.vetmic.2015.01.024
- Laverty, G., Gorman, S. P., & Gilmore, B. F. (2013). Biomolecular mechanisms of staphylococcal biofilm formation. *Future microbiology*, 8(4), 509-524.
- Los, R., Sawicki, R., Juda, M., Stankevic, M., Rybojad, P., Sawicki, M., Malm, A. and Ginalska, G. (2010). A comparative analysis of phenotypic and genotypic methods for the determination of the biofilm-forming abilities of *Staphylococcus epidermidis*. *FEMS microbiology letters*, 310(2), 97-103.
- Mack, D., Davies, A. P., Harris, L. G., Rohde, H., Horstkotte, M.A., & Knobloch, J. K. M. (2007). Microbial interactions in *Staphylococcus epidermidis* biofilms. *Analytical and bioanalytical chemistry*, 387(2), 399-408.
- Majumder, S., Wolffs, P. F. G., J. P. A. Hoebe, C., & Rahmatullah, M. (2014). Rapid detection of methicillin resistance and biofilm formation in *Staphylococcal* species. *Journal of Chemical and Pharmaceutical Research*, 6(10), 148–154.
- Mangram, A. J., Horan, T. C., Pearson, M. L., Silver, L. C., Jarvis, W. R., & Hospital Infection Control Practices Advisory Committee. (1999). Guideline

- for prevention of surgical site infection, 1999. *American journal of infection control*, 27(2), 97-134.
- Marsik, F. J., & Brake, S. Y. L. V. I. A. (1982). Species identification and susceptibility to 17 antibiotics of coagulase-negative staphylococci isolated from clinical specimens. *Journal of Clinical Microbiology*, 15(4), 640-645.
- Mathur T, Singhal S, Khan S, Upadhyay D J, Fatma T, Rattan A (2006). Detection of biofilm formation among the clinical isolates of *staphylococci*: An evaluation of three different screening methods. *Indian Journal of Medical Microbiology*; 24:25(9)
- Mirrett, S., Weinstein, M. P., Reimer, L. G., Wilson, M. L., & Reller, L. B. (2001). Relevance of the number of positive bottles in determining clinical significance of coagulase negative staphylococci in blood cultures. *Journal of Clinical Microbiology*, 39(9), 3279-3281.
- Molina, J., Penuela, I., Lepe, J. A., Gutiérrez-Pizarra, A., Gómez, M. J., García Cabrera, E., Cordero, E., Aznar, J. and Pachón, J. (2013). Mortality and hospital stay related to coagulase negative Staphylococci bacteraemia in non-critical patients. *Journal of Infection*, 66(2), 155-162.
- Montanaro, L., C. R. Arciola, E. Borsetti, S. Collamati, L. Baldassarri, and L. Montanaro. (1999). Detection of fibronectin-binding protein genes in staphylococcal strains from periprostheses infections. *Journal of New Microbiology*. 22:331–336.
- Namvar, A. E., Asghari, B., Ezzatifar, F., Azizi, G., & Lari, A. R. (2013). Detection of the intercellular adhesion gene cluster (*ica*) in clinical *Staphylococcus aureus* isolates. *GMS hygiene and infection control*, 8(1).
- Ogston, A. (1984). On abscesses. *Review of Infectious Diseases*, 6(1), 122-128.
- Obajuluwa, A. F., Onaolapo, J. A., Olayinka, B. O., & Adeshina, G. O. (2017). Antibiotics Susceptibility Pattern of Coagulase Negative Staphylococci Isolates from Orthopaedic Patients. *International Educational Applied Scientific Research Journal*, 1(3).
- Oliveira, A. D., Sanches, P., Lyra, J. C., Bentlin, M. R., Rugolo, L. M., & Cunha, M. D. L. R. D. (2012). Risk Factors for Infection with coagulase-negative staphylococci in newborns from the neonatal Unit of a Brazilian University Hospital. *Clinical Medicine Insights. Pediatrics*, 1-9.
- Oliveira, C. F. D., Paim, T. G. D. S., Reiter, K. C., Rieger, A., & D'azevedo, P. A. (2014). Evaluation of four different DNA extraction methods in coagulase-negative staphylococci clinical isolates. *Revista do Instituto de Medicina Tropical de São Paulo*, 56(1), 29-33.
- Oufriid, S., Ghazlane, Z., Jamali, L., El Otmani, F., Talmi, M., Elmdaghri, N., Zerouali, K. and Timinouni, M. (2015). Correlation between *staphylococcal* biofilm formation in vitro and potential for catheter-related infections. *The Journal of Infection in Developing Countries*, 9(04), 368-372.

- Papadimitriou-Olivgeri, I., Giormezis, N., Papadimitriou-Olivgeris, M., Zotou, A., Kolonitsiou, F., Koutsileou, K., Fligou, F., Marangos, M., Anastassiou, E.D. and Spiliopoulou, I. (2016). Number of positive blood cultures, biofilm formation, and adhesin genes in differentiating true coagulase-negative staphylococci bacteremia from contamination. *European Journal of Clinical Microbiology & Infectious Diseases*, 35(1), 57-66.
- Pfaller, M. A., & Herwaldt, L. A. (1988). Laboratory, clinical, and epidemiological aspects of coagulase-negative staphylococci. *Clinical Microbiology Reviews*, 1(3), 281-299.
- Prasad, S., Nayak, N., Satpathy, G., Nag, H. L., Venkatesh, P., Ramakrishnan, S., Ghose, S. and Nag, T. C. (2012). Molecular & phenotypic characterization of *Staphylococcus epidermidis* in implant related infections. *The Indian Journal of Medical Research*, 136(3), 483.
- Piette, A., & Verschraegen, G. E. R. D. A. (2009). Role of coagulase-negative staphylococci in human disease. *Veterinary microbiology*, 134(1), 45-54.
- Pongruangporn, M., Ajenjo, M. C., Russo, A. J., McMullen, K. M., Robinson, C., Williams, R. C., & Warren, D. K. (2013). Patient-and Device-Specific Risk Factors for Peripherally Inserted Central Venous Catheter—Related Bloodstream Infections. *Infection Control & Hospital Epidemiology*, 34(02), 184-189.
- Poyart, C., Quesne, G., Boumaila, C., & Trieu-Cuot, P. (2001). Rapid and accurate species-level identification of coagulase negative staphylococci by using the *sodA* gene as a target. *Journal of Clinical Microbiology*, 39(12), 4296-4301.
- Rahman, Z. A., Hamzah, S. H., Asma'Hassan, S., Osman, S., & Noor, S. S. M. (2013). The significance of coagulase negative staphylococci bacteremia in a low resource setting. *The Journal of Infection in Developing Countries*, 7(06), 448-452.
- Rogers, K. L., Rupp, M. E., & Fey, P. D. (2008). The presence of *ica* ADBC is detrimental to the colonization of human skin by *Staphylococcus epidermidis*. *Applied and Environmental Microbiology*, 74(19), 6155-6157.
- Rohde, H., Kalitzky, M., Kröger, N., Scherpe, S., Horstkotte, M. A., Knobloch, J. K. M., Zander, A.R. and Mack, D. (2004). Detection of virulence-associated genes not useful for discriminating between invasive and commensal *Staphylococcus epidermidis* strains from a bone marrow transplant unit. *Journal of clinical microbiology*, 42(12), 5614-5619.
- Rupp, M. E., Cavalieri, R. J., Marolf, C., & Lyden, E. (2017). Reduction in Blood Culture Contamination Through Use of Initial Specimen Diversion Device. *Clinical Infectious Diseases*, cix304.
- Sani, N. A. M., Sapri, H. F., Noordin, A., Neoh, H. M., & Hussin, S. (2011). Species identification of Coagulase Negative Staphylococci (CoNS) isolates in

Universiti Kebangsaan Malaysia Medical Centre (UKMMC). *Asia-Pacific Journal of Molecular Medicine*, 1, 1-5.

Sani, N. A. M., Sapri, H. F., Neoh, H. M., & Hussin, S. (2014). First report on the molecular epidemiology of Malaysian *Staphylococcus epidermidis* isolated from a University Teaching Hospital. *BMC research notes*, 7(1), 1.

Shittu, A., Lin, J., Morrison, D., & Kolawole, D. (2006). Identification and molecular characterization of mannitol salt positive, coagulase-negative staphylococci from nasal samples of medical personnel and students. *Journal of Medical Microbiology*, 55(3), 317-324.

Solati, S. M., Tajbakhsh, E., Khamesipour, F., & Gugnani, H. C. (2015). Prevalence of virulence genes of biofilm producing strains of *Staphylococcus epidermidis* isolated from clinical samples in Iran. *AMB Express*, 5(1), 1.

Soumya, K. R., Philip, S., Sugathan, S., Mathew, J., & Radhakrishnan, E. K. (2017). Virulence factors associated with Coagulase Negative Staphylococci isolated from human infections. *3 Biotech*, 7(2), 140.

Sydnor, E. R., & Perl, T. M. (2011). Hospital epidemiology and infection control in acute-care settings. *Clinical Microbiology Reviews*, 24(1), 141-173.

Szymanska, G., Szemraj, M., & Szewczyk, E. M. (2011). Species specific sensitivity of coagulase-negative staphylococci to single antibiotics and their combinations. *Polish Journal of Microbiology*, 60(2), 155-161.

Tashiro, M., Izumikawa, K., Ashizawa, N., Narukawa, M., & Yamamoto, Y. (2015). Clinical significance of methicillin resistant coagulase-negative staphylococci obtained from sterile specimens. *Diagnostic microbiology and infectious disease*, 81(1), 71-75.

Uyanik, M. H., Yazgi, H., Ozden, K., Erdil, Z., & Ayyildiz, A. (2014). Comparison of coagulase-negative staphylococci isolated from blood cultures as a true bacteremia agent and contaminant in terms of slime production and methicillin resistance. *The Eurasian journal of medicine*, 46(2), 115.

Vacheethasanee, K., Temenoff, J. S., Higashi, J. M., Gary A., Anderson, J. M., Bayston, R., & Marchant, R. E. (1998). Bacterial surface properties of clinically isolated *Staphylococcus epidermidis* strains determine adhesion on polyethylene. *Journal of biomedical materials research*, 42(3), 425-432.

Vijayalakshmi, P. (2015). Incidence of *Staphylococcus aureus* in surgical site infections in a teaching hospital. *International Journal of Current Microbiology and Applied Sciences*, 4(4), 32-34.

Vuong, C., Kocianova, S., Voyich, J. M., Yao, Y., Fischer, E. R., DeLeo, F. R., & Otto, M. (2004). A crucial role for exopolysaccharide modification in bacterial biofilm formation, immune evasion, and virulence. *Journal of Biological Chemistry*, 279(52), 54881-54886.

- Wojtyczka, R. D., Orlewska, K., Kępa, M., Idzik, D., Dzedzic, A., Mularz, T., Krawczyk, M., Mikłasińska, M. and Wąsik, T. J. (2014). Biofilm formation and antimicrobial susceptibility of *Staphylococcus epidermidis* strains from a hospital environment. *International Journal of Environmental Research and Public Health*, 11(5), 4619-4633.
- Yao, Y., Sturdevant, D. E., & Otto, M. (2005). Genomewide analysis of gene expression in *Staphylococcus epidermidis* biofilms: insights into the pathophysiology of *S. epidermidis* biofilms and the role of phenol-soluble modulins in formation of biofilms. *Journal of Infectious Diseases*, 191(2), 289-298.
- Zadoks, R. N., & Watts, J. L. (2009). Species identification of coagulase-negative staphylococci: genotyping is superior to phenotyping. *Veterinary microbiology*, 134(1), 20-28.
- Zhou, S., Chao, X., Fei, M., Dai, Y., & Liu, B. (2013). Analysis of *S. epidermidis ica A* and *ica D* genes by polymerase chain reaction and slime production: a case control study. *BMC infectious diseases*, 13(1), 1.