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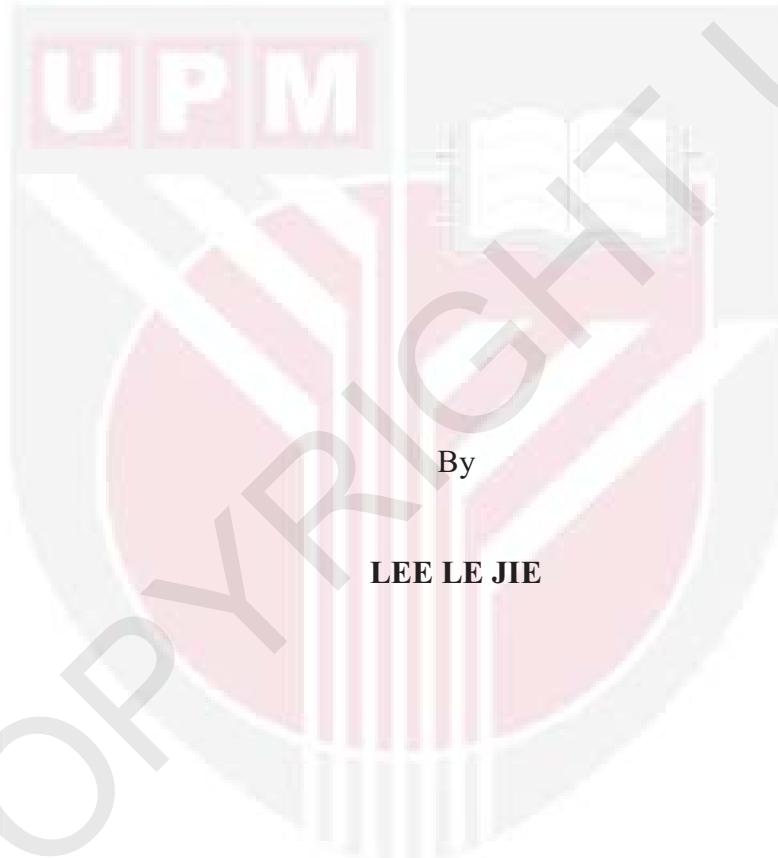
***INFLUENCE OF GENDER AND GENETIC POLYMORPHISM OF
NATURAL KILLER CELL GENES ON ACUTE LYMPHOBLASTIC
LEUKAEMIA***

LEE LE JIE

FPSK(P) 2018 17



**INFLUENCE OF GENDER AND GENETIC POLYMORPHISM OF
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LEUKAEMIA**



**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

September 2017

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Doctor of Philosophy

**INFLUENCE OF GENDER AND GENETIC POLYMORPHISM OF
NATURAL KILLER CELL GENES ON ACUTE LYMPHOBLASTIC
LEUKAEMIA**

By

LEE LE JIE

September 2017

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Acute lymphoblastic leukaemia (ALL) is characterized by an over-proliferation of immature lymphoblasts in the bone marrow. It is the most common cancer in children but affects adults as well. The majority of ALL are B cell lineage however approximately 15% are of T-cell origin. ALL predominates among males, from 1.2 times in B-cell lineage to 2.2 in T-cell ALL. Natural killer (NK) cells are significant players of the innate immune response characterised by cytolytic activity against virus-infected and tumour cells. The role of NK cells in immune surveillance and the tumour microenvironment are supported by earlier studies. Furthermore, like ALL it appears to be increased in the male gender. The activity of NK cells is regulated by groups of inhibitory and activating receptors. The role of NK cells in the leukemogenesis is still unclear. This study hypothesise that variation in NK cell associated genes may affect functional killing activities of NK cells and influence leukemogenesis. The objective of this study is to investigate genetic polymorphisms and variations in NK cell associated genes in of ALL patients and contrast between sexes.

The percentage of NK cells (CD56+) in peripheral blood mononuclear cells were compared between gender in B-cell and T-cell ALL and compared with healthy controls. The average percentage of NK cells in mononuclear cells was higher in male subjects including normal, B-ALL and T-ALL male compared to female subjects. Gene expression frequencies of activating and inhibitory killer cell immunoglobulin-like receptors (*KIR*), commonly studied were also compared between these samples. Significant absence in expression of *KIR2DL1-L4*, *3DL3*, *2DS2* and *S4* ($p<0.04$) were observed in ALL patients. The expression of *KIR2DL1*, *2DL3*, *2DL4*, *3DL3* and *2DS4* were significantly absent in T-ALL male patients

when compared with normal male controls. Genetic mutation and polymorphism, particularly single nucleotide polymorphism (SNPs) and small variants were then examined for NK-related genes in ALL samples via next-generation sequencing (NGS). Significant small variants present in specific gender and groups were identified. The consequences of these small variants to amino acid coding were predicted. These small variants have potential effects to the NK cells of individuals. The effects of selected small variants identified by NGS on gene expression levels were also investigated using qPCR. Small variants present in *KIR3DL2*, *KLRC1*, *KLRC2* and *LILRB1* were found to be significantly associated with the downregulation of gene expressions. Gene copy number variations (CNVs) defined by deletion or amplification in NK cells associated genes were also identified following next-generation sequencing and further validated using real-time quantitative polymerase chain reaction (qPCR). In ALL patients, amplification was detected in *LAIR2*, *KLRC1*, *NCR1*, *NCR3*, *KLRC2*, *KLRC3* and *KLRC4* while deletion was detected in *CD69* and *KLRD1*. Gene copy numbers of *KLRC2* and *KLRD1* were positively and significantly correlated with gene expression level. Lastly, the most common mechanism in epigenetic modification that is DNA methylation was investigated in the promoter regions of KIR. Hypermethylation or higher methylation index of KIR genes were observed in most ALL patients. Of these genes, *KIR2DL4* showed significant negative correlation between methylation index and gene expression ($r_s = -0.610$, $p < 0.001$). This study showed significant differences in genetics of NK cell related genes between ALL patients and healthy controls. There were also significant differences when genders were compared. These implied differences in function and activity of NK cells in the various groups and influence of gender from the variation in NK cell related genes may potentially affect leukaemogenesis on ALL. All the data from this fundamental study will lead to a better understanding on the innate immunity particularly the NK cell's arm in ALL patients and normal individuals on gender basis. The new insight on the genetic polymorphism, gene copy number variation and DNA methylation in ALL may be used to associate ALL with leukaemogenesis for better diagnosis and therapy in future. This study showed influence of gender in terms of genetic polymorphism, DNA methylation and gene copy number variations on ALL.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENGARUH JANTINA DAN POLIMORFISME GENETIK BAGI GEN-GEN SEL PEMUSNAH SEMULAJADI TERHADAP LEUKEMIA LIMFOSITIK AKUT

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Leukemia limfositik akut (ALL) dicirikan oleh proliferasi sel progenitor lymphoid secara melampau dalam sumsum tulang. Ia adalah kanser yang paling biasa ditemui di kalangan kanak-kanak. Kebanyakkan ALL adalah keturunan dari jenis sel-B (B-ALL) walaubagaimanapun kira-kira 15% adalah berasal dari sel-T (T-ALL). ALL lebih kerap terjadi di kalangan lelaki, 1.2 kali dalam ALL jenis sel-B dan 2.2 kali dalam ALL jenis sel-T. Sel pemusnah semulajadi (sel NK) adalah sejenis sel immune yang dapat dicirikan oleh aktiviti pembunuhan sel-sel yang dijangkiti oleh virus dan juga tumor. Peranan sel-sel NK dalam pengawasan imun dan persekitaran mikro tumor disokong oleh kajian terdahulu. Tambahan pula, seperti dalam ALL ia ternyata meningkat di golongan jantina lelaki. Aktiviti sel-sel NK dikawal oleh kumpulan-kumpulan reseptor penghalang dan pengaktif. Peranan sel NK dalam pembentukkan ALL masih kurang dikaji. Hipotesis kajian ini adalah variasi dalam gen-gen sel NK yang berkaitan berkeupayaan mempengaruhi fungsi kemusnahan sel NK dan berperanan dalam proses pembentukkan ALL. Objektif kajian ini adalah menyelidik poliformisme dan variasi genetic dalam gen-gen sel NK yang berkaitan dan membeza akan jantina.

Peratus sel NK (CD56+) dalam sel-sel mononuklear darah periferi dibandingkan antara jantina dalam sel-B dan sel-T ALL dan juga kumpulan kawalan yang sihat. Purata peratus sel-sel NK dalam sel-sel mononuklear adalah lebih tinggi dalam subjek lelaki termasuk dalam kumpulan kawalan, B-ALL dan T-ALL berbanding dengan subjek wanita. Frekuensi ekspresi gen-gen penghalang dan pengaktif reseptor killer cell immunoglobulin-like receptor (*KIR*) juga dibandingkan antara sampel-sampel. Ketakhadiran yang signifikan dalam ekspresi *KIR2DL1-L4*, *3DL3*, *2DS2* dan *S4* ($p<0.04$) diperhatikan dalam pesakit ALL. Ekspresi *KIR2DL1*, *2DL3*,

2DL4, *3DL3* dan *2DS4* juga lebih rendah secara signifikan di pesakit lelaki T-ALL berbanding dengan kawalan normal. Mutasi genetik dan polimorfisme, terutamanya polimorfisme nukleotida tunggal (SNP) dan varian kecil dalam gen-gen sel NK telah diperiksa melalui penjujukan generasi baru (NGS). Varian kecil yang signifikan dalam jantina dan kumpulan tertentu telah dikenal pasti. Akibat daripada varian kecil dalam pengkodan asid amino telah diperiksa. Varian kecil mempunyai potensi berkesan kepada sel-sel NK dalam individu. Kesan varian kecil yang dikenal pasti daripada NGS di peringkat ekspresi gen telah disiasat menggunakan qPCR. Varian kecil di dalam *KIR3DL2*, *KLRC1*, *KLRC2* dan *LILRB1* didapati berkait secara signifikan dengan pegawalaturan menurun ekspresi gen. Variasi salinan gen (CNVs) yang ditakrifkan sebagai penghapusan atau amplifikasi dalam gen-gen sel NK juga disiasat dengan cara NGS dan seterusnya disahkan menggunakan qPCR. Dalam pesakit ALL, amplifikasi dikesan di *LAIR2*, *KLRC1*, *NCR1*, *NCR3*, *KLRC2*, *KLRC3* dan *KLRC4* manakala penghapusan dikesan di *CD69* dan *KLRD1*. CNV dalam gen *KLRC2* dan *KLRD1* berhubung secara positif dan signifikan dengan ekspresi gen. Akhir sekali, mekanisma yang paling biasa dalam pengubahsuai epigenetik iaitu metilasi DNA telah disiasat di kawasan promoter gen-gen *KIR*. Hypermetilasi atau metilasi indeks gen *KIR* yang tinggi diperhatikan dalam kebanyakan pesakit ALL. Gen *KIR2DL4* menunjukkan hubungan negatif yang signifikan antara indeks metilasi dan ekspresi gen ($r_s = -0.610$, $p < 0.001$). Kajian ini menunjukkan perbezaan yang signifikan dalam genetic gen yang berkaitan dengan sel NK antara pesakit ALL dan kawalan yang sihat. Terdapat juga perbezaan yang signifikan apabila jantina dibandingkan. Perbezaan tersirat dalam fungsi dan aktiviti sel-sel NK dalam pelbagai kumpulan dan juga jantina akibat variasi dalam gen-gen sel NK, maka berpotensi mempengaruhi pembentukkan ALL. Semua data dari kajian asas ini akan membawa kepada pemahaman yang lebih mendalam mengenai imuniti semula jadi terutamanya dari lengan sel NK dalam pesakit ALL dan individu normal mengikut jantina. Wawasan baru dari polimorfisme genetik, variasi salinan gen dan metilasi DNA dalam ALL boleh digunakan untuk mengaitkan ALL dengan leukaemogenesis untuk menambahbaik diagnosis yang lebih baik dan terapi pada masa depan. Kajian ini menunjukkan pengaruh jantina dari segi polimorfisme genetik dan variasi salinan gene pada ALL.

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I certify that a Thesis Examination Committee has met on 20 September 2017 to conduct the final examination of Lee Le Jie on his thesis entitled "Influence of Gender and Genetic Polymorphism of Natural Killer Cell Genes on Acute Lymphoblastic Leukaemia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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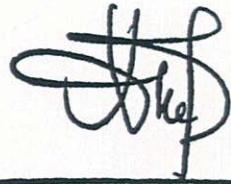
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TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xix
 CHAPTER	
1 INTRODUCTION	1
 2 LITERATURE REVIEW	3
2.1 Leukaemia	3
2.1.1 Epidemiology	3
2.1.2 Incidences	3
2.1.3 Gender Differences	4
2.2 Acute Leukaemia	4
2.2.1 Acute Lymphoblastic Leukaemia	5
2.2.2 Classification of Acute Leukaemia	6
2.3 Clinical Management	8
2.3.1 Risk Stratification	8
2.3.2 Minimal Residual Disease	8
2.3.3 General Treatments of ALL	9
2.3.4 Haematopoietic Stem Cell Transplant	9
2.4 The Immune System	10
2.4.1 Natural Killer Cell	10
2.4.1.1 Mechanism of NK cells Activity	11
2.4.1.2 Gender Difference in NK cell	12
2.4.1.3 Receptors of NK Cell	12
2.4.1.4 Other System and Genes Related to NK Cells	18
2.4.1.5 Effect of Sex Hormone on NK Cells	19
2.4.1.6 NK Cells and Tumour Microenvironment	19
2.4.1.7 NK Cells and Leukaemia	20
2.5 Epigenetics – DNA Methylation	20
2.5.1 Epigenetics in Leukaemia	21
2.6 Genetic Polymorphism	21
2.6.1 Impact of SNPs on Gene Function	22
2.6.2 SNPs and Leukaemia	22

2.7	Copy Number Variation	23
3	MATERIALS AND METHODS	24
3.1	Experimental Design	24
3.2	Sample Collection	25
3.3	Study Group	25
3.4	Isolation of Mononuclear Cells	25
3.4.1	Viability, Cell Count and Cryopreservation	26
3.5	DNA Extraction	26
3.6	Total RNA Extraction	27
3.6.1	DNase Treatment	27
3.6.2	RNA Qualification and Quantification	28
3.6.3	Reverse Transcription	28
3.6.4	Polymerase Chain Reaction (PCR) with Reference Genes	28
3.7	Killer cell Immunoglobulin-like Receptors Gene Expression	29
3.7.1	Duplex PCR	29
3.8	Immunophenotyping	31
3.9	Next-generation Sequencing Library Preparation	31
3.9.1	Designation of Probes for Target Enrichment	31
3.9.2	Qubit dsDNA BR Assay	32
3.9.3	DNA Shearing	32
3.9.4	Sample Purification after Shearing	32
3.9.5	Quality Assessment	33
3.9.6	Repair of Ends	33
3.9.7	Addition of 'A' bases to the 3' End of the DNA Fragment	33
3.9.8	Ligation of the Indexing-specific Paired-end Adaptor	34
3.9.9	Amplification of Adaptor-ligated Library	34
3.9.10	Hybridisation	34
3.9.11	Preparation of Magnetic Beads	35
3.9.12	Selection of Hybrid Capture	35
3.10	Addition of Index Tags by Post-hybridisation Amplification	36
3.10.1	Amplification of the Captured Library to Add Index Tags	36
3.10.2	Quantity Assessment of Index-tagged Library by qPCR	37
3.10.3	Pooling of Samples for Multiplexed Sequencing	37
3.10.4	Cluster Amplification and Sequencing	37
3.11	Bioinformatics Analysis	38
3.11.1	Pre-alignment Quality Inspection	38
3.11.2	Raw Reads Alignment	38
3.11.3	Post-alignment Quality Control and Filters	39
3.11.4	Small Variants Analysis	39
3.11.4.1	Significant SNPs	39
3.11.4.2	SNP Effect Analysis	40
3.11.5	Copy Number Analysis	40

3.11.5.1	Validation of Copy Number Variations	40
3.12	Gene Expression	42
3.12.1	Real-time PCR	42
3.13	Bisulfite Conversion	43
3.13.1	Quantitative Methylation Specific PCR	43
3.13.2	Analysis of Real-time PCR Data	45
3.14	Statistical Analysis	46
4	RESULTS	47
4.1	Sample Selection	47
4.2	DNA Extraction	49
4.2.1	DNA Quality and Integrity	49
4.2.2	Quality Assessment after DNA Shearing	49
4.2.3	Quality Assessment after Amplification of Adaptor-ligated Library	50
4.2.4	Quality Assessment of Library after Addition of Index Tags by Post-hybridisation	51
4.2.5	Quantity Assessment of each Index-tagged Library by qPCR	51
4.2.5.1	Standard Curve	51
4.2.5.2	Concentration of Index-tagged Captured Library	52
4.3	Next-generation Sequencing	53
4.3.1	Raw Reads Quality Control	53
4.3.2	Pre-alignment Quality Inspection	53
4.3.2.1	Base Quality by Position	53
4.3.2.2	Base Composition by Position	54
4.3.2.3	Base and Read Quality Distributions	54
4.3.2.4	GC-content	56
4.3.2.5	Alignment Quality by Tile	57
4.3.2.6	Matching Status	58
4.3.2.7	Filters	59
4.3.2.8	Targeted Region Quality Control	59
4.4	Small Variants Detection in NK Cell Receptors	59
4.5	Small Variants in Genes Related to Sex Hormone	65
4.6	Small Variants in Other Genes Related to NK Cells	67
4.7	Association of Small Variants with Gene Expression	69
4.8	Copy Number Variation	70
4.8.1	Validation of CNV by qPCR	70
4.8.1.1	Standard Curves and Melting Curves Analysis	70
4.8.1.2	Copy Number Variations Detection	72
4.9	Total RNA Isolation	74
4.9.1	RNA Integrity	74
4.9.2	RNA Purity and Quantity	75
4.9.3	RT-PCR with reference gene GAPDH and beta-actin	75
4.10	Percentages of NK cells in Lymphocyte Population	76

4.10.1	Percentage of CD56+CD19+ Cells	78
4.11	KIR Gene Expression	78
4.11.1	Gender Differences in Frequency of KIR Gene Expressed	80
4.12	Gene Expression of NK Cell Receptors	81
4.13	Methylation Specific PCR	85
4.13.1	Methylation Status	85
4.14	Correlation of KIR Gene Expressions with Methylation Status	86
4.15	Correlation of Gene Expressions with Gene Copy Number Values	87
5	DISCUSSION	88
5.1	DNA Quality and Integrity	88
5.2	Quality Assessment after Shearing and Sequencing	89
5.3	Small Variant Analysis	90
5.4	Copy Number Variation	93
5.5	RNA Extraction and Quality Controls	95
5.6	Percentage of NK Cells	95
5.7	Expression of CD19+56+ Double Positive Population	96
5.8	KIR Gene Expression Frequency	96
5.9	Gene Expression of NK Cell Receptors and <i>GP6</i>	98
5.10	DNA Methylation of KIR Genes	100
5.11	Limitations of study	101
6	CONCLUSION AND FUTURE RECOMMENDATIONS	103
6.1	Conclusion	103
6.2	Future Recommendations	104
REFERENCES		105
APPEDICES		132
BIODATA OF STUDENT		149
LIST OF PUBLICATIONS		150

LIST OF TABLES

Table	Page
2.1 New cases of leukaemia in major ethnic groups	4
2.2 FAB morphological classification of ALL	6
2.3 WHO classification of acute leukaemia	7
2.4 Immunophenotypic classification and leukocyte antigen expression (% of cases positive) in acute lymphoblastic leukaemias	7
3.1 Primer sequences for reference genes of PCR	29
3.2 Primer sequences of beta-actin and KIR primers	30
3.3 Nucleotide sequences of each of the SureSelectXT indexes for Illumina sequencing	36
3.4 PCR programme for amplification of index tags by post-hybridisation amplification	36
3.5 PCR profiles for qPCR assessment of index-tagged library	37
3.6 Primer sequences of genes to validate CNVs using quantitative PCR	41
3.7 Primer sequences of genes for gene expression analysis	42
3.8 Primer sequences of reference and target genes used for methylation specific quantitative PCR	45
4.1 Characteristic of normal controls (NC) and ALL samples used for different methods in this study	48
4.2 Concentration of index tagged captured library	52
4.3 Raw reads quality control of each library after sequencing	53
4.4 Protein effects caused by SNPs or MNPs in genes coding for NK cell receptors	60
4.5 Comparison of small variants detected within and between different combinations of group	61
4.6 Small variants present exclusively in T-ALL patients (n=6). Fisher's exact test was used for statistical test, p<0.05 was considered significant	62

4.7	Small variants present exclusively in normal male controls (n=4). Fisher's exact test was used for statistical test, p<0.05 was considered significant	63
4.8	Small variants present exclusively in normal female controls (n=4). Fisher's exact test was used for statistical test, p<0.05 was considered significant	64
4.9	Protein effects caused by small variants in sex hormone related genes	65
4.10	Comparison of small variants detected between and within different combinations of group	65
4.11	Small variants present only in a group. Fisher's exact test was used for statistical test, p<0.05 was considered significant	66
4.12	Protein effects caused by small variants in NK cell related genes	67
4.13	Comparison of small variants detected between and within different combinations of group	67
4.14	Small variants present only in a group. Fisher's exact test was used for statistical test, p<0.05 was considered significant	68
4.15	Association of genetic variants with mRNA expression	69

LIST OF FIGURES

Figure		Page
2.1	Schematic diagram of inhibition and activation of NK cells	12
2.2	Inhibitory and activating signaling of NK cell receptors	13
2.3	Examples of inhibitory and activating NK cell receptors and the effects of their interactions with ligands	14
2.4	Inhibitory KIRs, activating KIRs and their identified ligands	17
2.5	Genomic organization of the KIR gene cluster	18
3.1	Flow chart on summary of methods used in the study	24
3.2	Haemacytometer grid to determine the cell concentrations (cells/mL) and viability (%) of isolated mononuclear cells. Four quadrants (4x4 boxes) at the edge of the grid were calculated to obtain the average	26
4.1	Gel image of 10 samples (lane 1 to 10) on 2% agarose gel	49
4.2	Overlaid electropherograms of seven representative sheared DNA samples with peak incident power 75 W	50
4.3	Analysis of amplified prepped library DNA using a DNA 100 assay	50
4.4	Analysis of amplified captured DNA using the High Sensitivity DNA Kit	51
4.5	Standard curve $y = -3.425x + 10.85$	52
4.6	Base quality plot of normal male control, M1 by position	54
4.7	Base composition plot of normal control, M1 by position	55
4.8	Base quality distribution of normal male control, M1	55
4.9	Read quality distribution of normal male control, M1	56
4.10	GC-content analysis of a representative sample, normal male control, M1	57
4.11	Alignment quality by tile plot of a representative normal male control, M1	58

4.12	Chart pie showing matching status of sample M1 after alignment	59
4.13	Venn diagram for number of significant small variants in genes coding for NK cell receptors	61
4.14	Copy number variations in T-ALL patient (n=6), normalised with normal male controls (n=4)	70
4.15	Standard curve for gene <i>KLRC2</i> , $y = -3.298x + 2122$, $R^2 = 0.997$	71
4.16(a)	Melting curves of PCR products from <i>KLRC1</i> amplification	71
4.16(b)	Gel electrophoresis of PCR products	71
4.17	Copy number variations in inhibitory genes <i>LAIR2</i> and <i>KLRC1</i>	72
4.18	Copy number variations in activating genes <i>NCR1</i> , <i>NCR3</i> , <i>KLRC2</i> and <i>KLRC3</i>	73
4.19	Copy number variations in gene <i>KLRC4</i> , <i>CD69</i> and <i>KLRD1</i>	74
4.20	Representative image of inverted gel electrophoresis photograph of extracted RNA from samples	75
4.21	Representative of inverted gel image of amplified PCR products for <i>GAPDH</i> (left) and <i>beta-actin</i> (right)	76
4.22	Region gating strategy on flow cytometry plots	77
4.23	Average percentages of NK cells were higher in all the male subjects	77
4.24	$CD56^+19^+$ double expression plots	78
4.25	Representative gel electrophoresis for <i>KIR2DL2</i> and <i>beta-actin</i>	79
4.26	Frequency of KIR gene expressions in normal controls (n=16), B-ALL (n=16) and T-ALL (n=8) patients	79
4.27	Frequency of inhibitory KIR gene expressed in normal male control (n=8), normal female control (n=8), B-ALL male (n=8), B-ALL female (n=8) and T-ALL (n=8) patients	80
4.28	Frequency of activating KIR gene expressed in normal male control (n=8), normal female control (n=8), B-ALL male (n=8), B-ALL female (n=8) and T-ALL (n=8) patients	81
4.29	mRNA expression of NK cell receptors in normal controls (n=24), B-ALL patients (n=24) and T-ALL patients (n=14) determined by qPCR	82

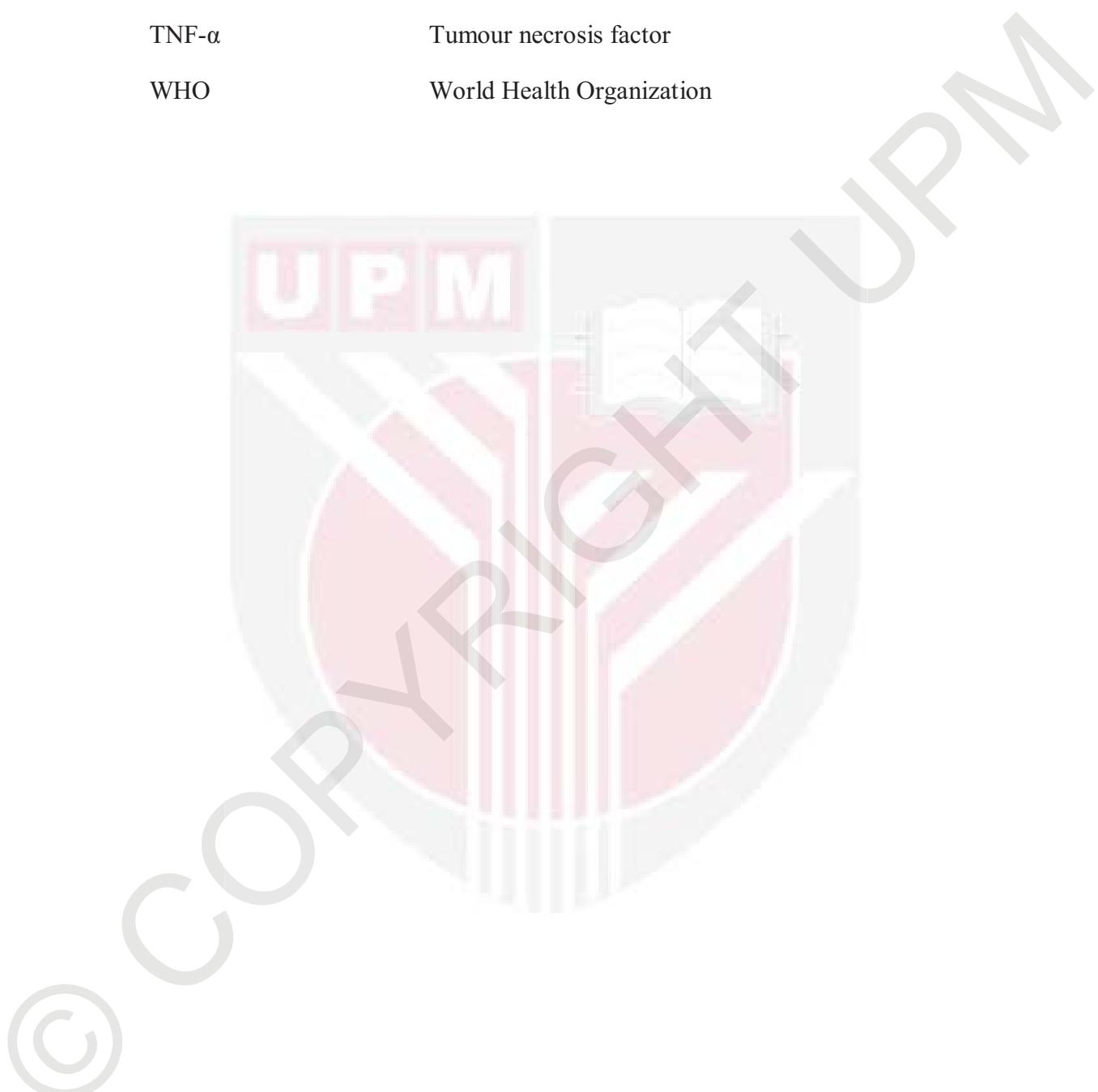
4.30	mRNA expression of activating receptors in normal male controls (n=12), normal female controls (n=12), B-ALL male patients (n=12), B-ALL female patients (n=12) and T-ALL male patients (n=14) using qPCR	83
4.31	mRNA expression of inhibitory receptors in normal male controls (n=12), normal female controls (n=12), B-ALL male patients (n=12), B-ALL female patients (n=12) and T-ALL male patients (n=14)	84
4.32	mRNA expression of <i>KLRC4</i> , <i>KLRD1</i> and <i>GP6</i> in normal male controls (n=12), normal female controls (n=12), B-ALL male patients (n=12), B-ALL female patients (n=12) and T-ALL male patients (n=14)	84
4.33	Methylation indexes in the promoter regions of inhibitory KIR genes in normal male controls (n=12), normal female controls (n=12), B-ALL male patients (n=12), B-ALL female patients (n=12) and T-ALL male patients (n=10)	85
4.34	Methylation indexes in the promoter regions of activating gene KIR2DS2 and 2DS4 in normal male controls (n=12), normal female controls (n=12), B-ALL male patients (n=12), B-ALL female patients (n=12) and T-ALL male patients (n=10)	86

LIST OF ABBREVIATIONS

ALL	Acute lymphoblastic leukaemia
AML	Acute myeloid leukaemia
AR	Androgen receptor
BLAST	Basic local alignment search tool
bp	Base pair
CD	Clusters of differentiation
cDNA	Complementary DNA
CLEC	C-type lectin domain
CLL	Chronic lymphoblastic leukaemia
CML	Chronic myeloid leukaemia
CNV	Copy number variations
CR	Complete remission
DEPC	Diethylpyrocarbonate
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
dsDNA	Double stranded DNA
EDTA	Ethylenediaminetetraacetic
ESR	Estrogen receptor
FAB	French-American-British
FISH	Fluorescence <i>in situ</i> hybridisation
FU	Fluorescent unit
gDNA	Genomic DNA
HLA	Human leukocyte antigen
HSCT	Haematopoietic stem cell transplantation

IFN- γ	Interferon gamma
IL	Interleukin
ITAM	Immunoreceptor tyrosine-based activation motifs
ITGAL	Integrin alpha L
ITIM	Immunoreceptor tyrosine-based inhibitory motifs
KIR	Killer cell immunoglobulin-like receptors
KLR	C-type lectin-like receptors
LAIR	Leucocyte-associated inhibitory receptor
LCR	Leucocyte receptor complex
LILR	Leucocyte immunoglobulin-like receptor
M-MLV	Moloney murine leukaemia virus
MNP	Multiple nucleotide polymorphism
MOH	Ministry of Health
MRD	Minimal residual disease
mRNA	Messenger RNA
NCBI	National Center for Biotechnology Information
NCR	Natural cytotoxic receptors
NGS	Next-generation sequencing
NK cell	Natural killer cell
NTC	No-template control
PAS	Period acid-Schiff
PBS	Phosphate buffered saline
PGR	Progesterone receptor
qPCR	Quantitative polymerase chain reaction
RNA	Ribonucleic acid
RPMI	Roswell Park Memorial Institute

RT	Room temperature
SEM	Standard error of mean
SNP	Single nucleotide polymorphism
TNF- α	Tumour necrosis factor
WHO	World Health Organization



CHAPTER 1

INTRODUCTION

According to National Cancer Registry year 2011, leukaemia is the 7th most common cancer in Malaysia. Leukaemia is the haematological malignancy of blood or bone marrow, characterised by the production of large number of abnormal and immature white blood cells. Acute leukaemia is the aggressive type of leukaemia where the primary diagnosis and classification of acute leukaemia relied on cytomorphological, cytochemical and immunophenotype of leukaemic cells. Acute lymphoblastic leukaemia (ALL) is characterised by the over proliferation of immature and cancerous lymphocytes known as lymphoblast. ALL is a common pediatric leukaemia with peak incidence around two to five years old while another peak incidence appears in old age. There are generally few types of ALL which include B-cell lineage and T-cell lineage. The higher incidence of acute lymphoblastic leukaemia in male has been reported in Malaysia and other studies across the world.

The etiology of acute leukaemia is still unclear. Immune surveillance protects the body from cancer formation. Mechanisms in which tumour cells evade immune eradication include hijacking immune suppressive pathways as well as maintaining immune ignorance via both innate and adaptive immune systems (Gajewski et al., 2013).

Natural killer (NK) cells are significant players of the innate immune response characterized by cytolytic activity against virus-infected and tumour cells (Lorenzo, 2007). The major function of NK cells in fighting cancer is to be in surveillance and elimination of cells that become malignant. NK cell function and detection systems are regulated by a group of inhibitory and activating receptors and also their ligand. Major NK cell receptors include killer cell immunoglobulin-like receptors (KIRs), natural cytotoxic receptors (NCRs) and C-type lectin-like receptors (KLRs). The ligands for NK cell receptors were reported be HLA class I, however, majority of the specific ligands for NK cells receptors are yet to be identified. Apart from specific NK cells receptors, other system such as Fas and Fas ligand and sex hormones are also important factors in cytotoxicity mediated by NK cells (Chua et al., 2004). NK cells were reported to be predominant in male (Oshimi et al., 1980; Santoli et al., 1976; Kennichi et al., 2003; Reichert et al., 1991). For instance, in a study of gender effect on *in vitro* lymphocyte subset levels, Abdullah et al. (2012) reported a higher percentage and absolute count of NK cells in blood of healthy male.

As the incidence of ALL was reported to be higher in male and NK cells were also found to be higher percentage and activity in male. However, the association between the higher incidence of ALL and NK cells activity in male remains unknown. It is hypothesised NK cell variations at genetic level are present in ALL cases and is gender biased.

There are several potential factors causing the dysfunction of NK cells related genes in ALL including genetic factors such as mutations, genetic polymorphisms, and gene copy number variations (CNV). The sources of genetic polymorphism include single nucleotide polymorphism (SNPs), sequence repeats, insertions, deletions and recombination. SNP is the most common sequence variation in human genome (Risch, 2001). Associations of small variants with leukaemia prognosis, survival, therapy and relapse have been described in some studies. However, the association between small variants in genes coded for NK cell receptors and with its function has yet to be identified. Other than small variant, CNV has emerged as another crucial research area in leukaemia and other diseases as CNVs are present in human population with high frequency and potentially explain more variation than SNPs (Pang et al., 2010). This is because large scale variations with possible deletions of tumour suppressor genes or amplifications of oncogenes are often found in cancer genome. Lastly, epigenetic modification such as DNA methylation may be one of the factors causing irregular gene expression of NK cell receptors. DNA methylation in the promoter region of a gene can suppress transcription events and thus reduce gene expression (Stirzaker et al., 2004). Cancer-associated hypomethylation and hypermethylation in different extent, frequency and types of cancers had been described; however, more studies on the role of DNA methylation in tumourgenesis and tumour progression need to be carried out (Ehrlich, 2009). Study involved all four aspects (genetic polymorphism, gene expression, gene copy number and DNA methylation) in NK cells and ALL has not been substantiated by others.

The general objective of this study is to investigate the association between genetic variations in NK cell related genes in ALL patients and gender bias.

The specific objectives of this study are:

1. To compare gene expression of NK cell receptors and percentage of NK cells in ALL patients and healthy controls of different genders.
2. To investigate genetic polymorphisms and gene copy number variation in NK cell related genes in ALL patients and healthy controls of different genders.
3. To study the effects of genetic polymorphism on NK cell genes expressions in ALL patients and healthy controls of different genders.
4. To identify DNA methylation and its correlation with gene expression levels of NK cell related genes in ALL patients and healthy controls of different genders.

REFERENCES

- Abdullah, M., Chai, P-S., Chong, M-Y., Mohd Tohit, E.R., Ramasamy, R., Pei, C.P. and Vidyadarshan, S. 2012. Gender effect on in vitro lymphocyte subset levels of healthy individuals. *Cellular Immunology*, 272:214-219
- Abel, H.J. and Duncavage, E.J. 2014. Detection of structural DNA variation from next generation sequencing data: a review of informatics approaches. *Cancer Genetics*, 206:432-440
- Adamaki, M., Lambrou, G.I., Athanasiadou, A., Tzanoudaki, M. and Vlahopoulos, S. 2013. Implication of IRF4 aberrant gene expression in the acute leukemias of childhood. *PLoS ONE*, 8(8): e72326
- Agiannitopoulos, K., Kyriarissi, A., Manginas, A., Papamzelopoulos, S. and Lamnisso, K. 2014. Genetic variant in the CYP17 gene and risk of premature coronary artery disease. *Hellenic Journal of Cardiology*, 55:126-131
- Albers, C.A., Lunter, G., MacArthur, D.G., McVean, G., Ouwehand, W.H. and Durbin, R. 2011. Dindel: accurate indel calls from short-read data. *Genome Research*, 21(6):961-973
- Albertsson, P.A., Basse, P.H., Hokland, M., Goldfarb, R.H., Nagelkerke, J.F., Nannmark, U. and Kuppen, P.J. 2003. Nk cells and tumour microenvironment: implications for NK-cell function and anti-tumour activity. *Trends in Immunology*, 24:603-609
- Aldred, P.M., Hollox, E.J. and Armour, J.A. 2005. Copy number polymorphism and expression level variation of the human alpha-defensin gene DEFA1 and DEFA3. *Human Molecular Genetics*, 14:2045-2052
- Almalte, Z., Samarani, S., Iannello, A., Debbeche, O., Amre, D.K., Sinnet, D. and Ahmad, A. 2011. Novel associations between activating killer-cell immunoglobulin-like receptor genes and childhood leukemia. *Blood*, 118:1323-1328.
- Al-Mawali, A., Gillis, D., Hissaria, P. and Lewis, I. 2008. Incidence, sensitivity, and specificity of leukemia-associated phenotypes in acute myeloid leukemia using specific five-color multparameter flow cytometry. *American Journal of Clinical Pathology*, 129:934-945.
- Amy, F., Tobi, B., Suvankar, M., David, E., Dana, J. and Gail, M. 2010. Gender differences in incidence rates of childhood B-precursor acute lymphocytic leukemia in Mississippi. *Journal of Pediatric Oncology Nursing*, 27:164-167
- Anderson, K.J. and Allen, R.L. 2009. Regulation of T-cell immunity by leucocyte immunoglobulin-like receptors: innate immune receptors for self on antigen-presenting cells. *Immunology*, 127(1):8-17

- Anfossi, N., Andre, P., Guia, S., Falk, C.S., Roetynck, S., Stewart, C.A., Breso, B., Frassati, C., Reviron, D., Middleton, D., Romagne, F., Ugolini, S. and Vivier, E. 2006. Human NK cell education by inhibitory receptors for MHC class I. *Immunity*, 25(2):331-342
- Annino, L., Vegna, M.L., Camera, A., Specchia, G., Visani, G., Fioritoni, G., Ferrara, F., Peta, A., Coilli, S., Deplano, W., Fabbiano, F., Sica, D., Di Raimondo, F., Cascavilla, N., Tabillio, A., Leoni, P., Invernizzi, R., Baccarani, M., Rotoli, B., Amadori, S., Mandelli, F. and GIMEMA Group. 2002. Treatment of adult acute lymphoblastic leukemia (ALL): long-term follow-up of the GIMEMA ALL 0288 randomized study. *Blood*, 99: 863-871
- Aretz, S., Stienen, D., Uhlhaas, S., Pagenstecher, C., Mangold, E., Caspari, R., Propping, P. and Friedl, W. 2005. Large submicroscopic genomic APC deletions are a common cause of typical familial adenomatous polyposis. *Journal of Medical Genetics*, 42:185-192
- Assaf, C., Gellrich, S., Whittaker, S., Robson, A., Cerroni, L., Massone, C., Kerl, H., Rose, C., Chott, A., Chimenti, S., Hallermann, C., Petrella, T., Wechsler, J., Bagot, M., Hummel, M., Bullani-Kerl, K., Bekkenk, M.W., Kempf, W., Meijer, C.J., Willemze, R. and Sterry, W. 2007. CD56-positive haematological neoplasms of the skin: a multicentre study of the Cutaneous Lymphoma Project Group of the European Organisation for Research and Treatment of Cancer. *Journal of Clinical Pathology*, 60:981-989.
- Babor, F., Manser, A., Schonberg, K., Enczmann, J., Borkhardt, A., Meisel, R. and Uhrberg, M. 2012. Lack of association between KIR genes and acute lymphoblastic leukemia in children. *Blood*, 120:2770-2772
- Bainbridge, M.N., Wang, M., Burgess, D.L., Kovar, C., Rodesch, M.J., D'Ascenzo, M., Kitzman, J., Wu, Y.Q., Newsham, I., Richmond, T.A., Jeddelloh, J.A., Muzny, D., Albert, T.J. and Gibbs, R.A. 2010. Whole exome capture in solution with 3Gbp of data. *Genome Biology*, 11:R62
- Baker, D.A., Hameed, C., Tejani, N., Milch, P., Thomas, J., Monheit, A.G. and Dattwyler, R.J. 1985. Lymphocyte subsets in women on low dose oral contraceptives. *Contraception*, 32:377-382
- Barber, D.F., Faure, M. and Long, E.O. 2004. LFA-1 contributes an early signal for NK cell cytotoxicity. *Journal of Immunology*, 173(6):3653-3659
- Barten, R., Torkar, M., Haude, A., Trowsdale, J. and Wilson, M.J. 2001. Divergent and convergent evolution of NK-cell receptors. *Trends in Immunology*, 22:52-57
- Bene, M. C. 2005. Immunophenotyping of acute leukaemias. *Immunology Letters*, 98:9-21

- Bene, M.C., Castoldi, G., Knapp, W., Ludwig, W.D., Matutes, E., Orfao, A. and van't Veer, M.B. 1995. Proposals for the immunological classification of acute leukemias. European Group for the Immunological Characterization of Leukemias (EGIL). *Leukemia*, 9(10):1783-1786
- Bennett, J.M., Catovsky, D., Daniel, M.T., Flandrin, G., Galton, D.A., Gralnick, H.R. and Sultan, C. 1976. Proposals for the classification of the acuteleukaemias. French-American-British (FAB) co-operative group. *British Journal of Haematology*, 33:451-458
- Bennett, J.M., Catovsky, D., Daniel, M-T., Flandrin, G., Galton, D.A., Gralnick, H.R. and Sultan, C. 1991. Proposal for the recognition of minimally differentiated acute myeloid leukaemia (AML-M0). *British Journal of Haematology*, 78(3):325-329
- Berg, L., Riise, G.C., Cosman, D., Bergstrom, T., Olofsson, S., Karre, K. and Carbone, E. 2003. LIR-1 expression on lymphocytes, and cytomegalovirus disease in lung-transplant recipients. *Lancet*, 361:1099-1101
- Beziat, V., Traherne, J., Liu, L., Jayaraman, J., Engvist, M., Larsson, S., Trowsdale, J. and Malberg, K. 2013. Influence of KIR gene copy number on natural killer cell education. *Blood*, 121(23):4703-4707
- Biassoni, R., Cantoni, C., Pende, D., Sivori, S., Parolini, S., Vitale, M., Bottino, C. and Moretta, A. 2001. Human natural killer cell receptors and co-receptors. *Immunology Reviews*, 181:203-214
- Biassoni, R., Pessino, A. and Malaspina, A. 1997. Role of amino acid position 70 in the binding affinity of p50.1 and p58.1 receptors for HLA-Cw4 molecules. *European Journal of Immunology*, 27:3095-3099
- Bird, A. 2002. DNA methylation patterns and epigenetic memory. *Genes and Development*, 16:6-21
- Biron, C.A., Nguyen, K.B., Pien, G.C., Cousens, L.P. and Salazar-Mather, T.P. 1999. Natural killer cells in antiviral defense: function and regulation by innate cytokines. *Annual Review of Immunology*, 17:189-220
- Bolufer, P., Barragan, E., Collado, M., Cervera, J., Lopez, J.A. and Sanz, M.A. 2006. Influence of genetic polymorphisms on the risk of developing leukemia and on disease progression. *Leukemia Research*, 30(12):1471-1491
- Botstein, D. and Risch, N. 2003. Discovering genotypes underlying human phenotypes: past successes for Mendelian disease, future approaches for complex disease. *Nature Genetics*, 33:228-237
- Bouman, A., Schipper, M., Heineman, M.J. and Fass, M.M. 2004. Gender difference in the non-specific and specific immune response in human. *American Journal of Reproductive Immunology*, 52:19-26

- Burshtyn, D.N., Scharenberg, A.M., Wagtmann, N., Rajagopalan, S., Berrada, K., Yi, T., Kinet, J.P. and Long, E.O. 1996. Recruitment of tyrosine phosphatase HCP by the killer cell inhibitory receptor. *Immunity*, 4:77-85
- Cabanski, C.R., Cavin, K., Bizon, C., Wilkerson, M.D., Parker, J.S., Wilhelmsen, K.C., Perou, C.M., Marron, J.S. and Hayes, D.N. 2012. ReQON: a Bioconductor package for recalibrating quality scores from next-generation sequencing data. *BMC Bioinformatics*, 13:221 DOI: 10.1186/1471-2105-13-221
- Cai, H., Shu, X.O., Egan, K.M., Cai, Q., Long, J-R., Gao, Y-T. and Zheng, W. 2008. Association of genetic polymorphisms in CYP19A1 and blood level of sex hormones among postmenopausal Chinese women. *Pharmacogenetics and Genomics*, 18(8):657-664
- Campana, D. and Behm, F.G. 2000. Immunophenotyping of leukemia. *Journal of Immunological Methods*, 243:59-75
- Campana, D. and Coustan-Smith, E. 2004. Minimal residual disease studies by flow cytometry in acute leukemia. *Acta Haematologica*, 112:8-15
- Campbell, K.S., Dessing, M., Lopez-Botet, M., Calla, M and Collona, M. 1996. Tyrosine phosphorylation of a human killer inhibitory receptor recruits protein tyrosine phosphatase 1C. *Journal of Experimental Medicine*, 184:93-100
- Cantoni, C., Ponassi, M., Biassoni, R., Conte, R., Spallarossa, A., Moretta, A., Moretta, L., Bolognesi, M. and Bordo, D. 2003. The three-dimensional structure of the human NK cell receptor NKp44, a triggering partner in natural cytotoxicity. *Structure*, 11:725-734
- Carlson, C.S., Eberle, M.A., Rieder, M.J., Smith, J.D., Kruglyak, L. and Nuckerson, D.A. 2003. Additional SNPs and linkage-disequilibrium analyses are necessary for whole-genome association studies in human. *Nature Genetics*, 33:518-521
- Carr, W.H., Pando, M.J. and Parham, P. 2005. KIR3DL1 polymorphisms that affect NK cell inhibition by HLA-Bw4 ligand. *Journal of Immunology*, 175:5222-5229
- Chaix, J., Tessmer, M.S., Hoebe, K., Fuseri, N., Ryffel, B., Dalod, M., Alexopoulou, L., Beutler, B., Brossay, L., Vivier, E. and Walzer, T. 2008. Cutting edge: Priming of NK cells by IL-18. *Journal of Immunology*, 181(3)1627-1631
- Chan, H., Kurago, Z.B., Steward, C.A., Wilson, M.J., Martin, M.P., Mace, B.E., Carrington, M., Trowsdale, J. and Lutz, C.T. 2003. DNA methylation maintains allele-specific KIR gene expression in human natural killer cells. *The Journal of Experimental Medicine*, 197:245-255

- Chan, H., Miller, J.S., Moore, M.N. and Lutz, C.T. 2005. Epigenetic control of highly homologous killer Ig-like receptor gene alleles. *The Journal of Immunology*, 175:5966-5974
- Chang, H., Brandwein, J., Yi, Q., Chun, K., Patterson, B. and Brien, B. 2004. Extramedullary infiltrates of AML are associated with CD56 expression, 11q23 abnormalities and inferior clinical outcome. *Leukemia Research*, 28:1007-1011.
- Chang, J.S., Wiemels, J.L., Chokkalingam, A.P., Metayer, C., Barcellos, L.F., Hansen, H.M., Aldrich, M.C., Guha, N., Urayama, K.Y., Scelo, G., Green, J., May, S.L., Kiley, V.A., Wiencke, J.K. and Buffler, P.A. 2010. Genetic polymorphisms in adaptive immunity genes and childhood acute lymphoblastic leukemia. *Cancer Epidemiology, Biomarkers and Prevention*, 19:2153-2163
- Chen, B.A., Huang, Z.H., Zhang, X.P., Ou-Yang, J., Li, J.Y., Zhai, Y.P., Sun, X.M., Xu, Y.L., Lu, Q., Wang, J.M., Li, D., Liao, H., Shen, Z.X., Wang, Y.Y., Yu, X.J., Ye, H. and Zhuang, L.Y. 2010. An epidemiological investigation of leukemia incidence between 2003 and 2007 in Nanjing, China. *Journal of Hematology and Oncology*, 3:21
- Chen, Y., Breeze, C.E., Zhen, S., Beck, S. and Teschendorff, A.E. 2016. Tissue-independent and tissue-specific patterns of DNA methylation alteration in cancer. *Epigenetic and Chromatin*, 9:10
- Chen, Z.T., Wang, I., Liao, Y., Shih, Y. and Lin, L.L. 2011. Polymorphisms in steroidogenesis genes, sex steroid levels and high myopia in the Taiwanese population. *Molecular Vision*, 17:2297-2310
- Choi, J., Lee, S.J., Lee, Y.A., Maeng, H.G., Lee, J.K. and Kang, Y.W. 2014. Reference values for peripheral blood lymphocyte subsets in healthy Korean population. *Immune Network*, 14(6):289-295
- Chua, H.L., Serov, Y. and Brahmi, Z. 2004. Regulation of FasL expression in natural killer cells. *Human Immunology*, 65(4):317-327
- Colovai, A.I., Tsao, L., Wang, Su., Lin, H., Wang, C., Seki, T., Fisher, J.G., Menes, M., Bhagat, G., Alobeid, B. and Suciu-Foca, N. 2007. Expression of inhibitory receptor ILT3 on neoplastic B cells is associated with lymphoid tissue involvement in chronic lymphocytic leukemia. *Clinical Cytometry Society*, 72B:354-362
- Cooley, S., Weisdorf, D.J., Guethlein, L.A., Klein, J.P., Wang, T. and Le, C.T. 2010. Donor selection for natural killer cell receptor genes leads to superior survival after unrelated transplantation of acute myelogenous leukemia. *Blood*, 116:2411-2419

- Costello, R.T., Sivori, S., Marcenaro, E., Lafage-Pochitaloff, M., Mozziconacc, M., Reviron, D., Gastaut, J., Pende, D., Olive, D. and Moretta, A. 2002. Defective expression and function of natural killer cell-triggering receptors in patients with acute myeloid leukemia. *Blood*, 99:3661-3667
- Cuttner, J., Mick, R., Budman, D.R., Mayer, R.J., Lee, E.J., Henderson, E.S., Weiss, R.B., Paciucci, P.A., Sobol, R. and Davey, F. 1991. Phase III trial of brief intensive treatment of adult acute lymphocytic leukemia comparing daunorubicin and miroxantrone: a CALGB Study. *Leukemia*, 5:425-431
- D'Andrea, A., Chang, C., Franz-Bacon, K., McClanahan, T., Philips, J.H. and Lanier, L.L. 1995. Molecular cloning of NKB1. A natural killer cell receptor for HLA-B allotypes. *Journal of Immunology*, 155:2306-2310
- Deaton, A.M., Webb, S., Kerr, A.R., Illingworth, R.S., Guy, J., Andrews, R. and Bird, A. 2011. Cell type-specific DNA methylation at intragenic GpG islands in the immune system. *Genome Research*, 21(7):1074-1086
- DePristo, M.A., Banks, E., Poplin, R., Garimella, K.V., Maguire, J.R., Hartl, C., Philippakis, A.A., del Angel, G., Rivas, M.A., Hanna, M., McKenna, A., Fennell, T.J., Kernytsky, A.M., Sivachenko, A.Y., Cibulskis, K., Gabriel, S.B., Altshuler, D. and Daly, M.J. 2011. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nature Genetics*, 43(5):491-498
- Derek, M., Martin, C. and Lynne, M. 2002. Natural killer cells and their receptors. *Transplant Immunology*, 10:147-164
- Diaz-Pena, R., Blanco-Gelaz, M.A., Suarez-Alvarez, B., Martinez-Borra, J., Lopez-Vazquez, A., Alonso-Arias, R., Bruges-Armas, J., Vidal-Castineira, J.R. and Lopez-Larrea, C. 2008. Activatiy KIR genes are associated with ankylosing spondylitis in Asian population. *Human Immunology*, 69:437-442
- Dores, G.M., Devesa, S.S., Curtis, R.E., Linet, M.S. and Morton, L.M. 2012. Acute leukemia incidence and patient survival among children and adults in the United States, 2001-2007. *Blood*, 119:34-43
- Doukas, C., Saltiki, K., Mantzou, A., Cimponeriu, A., Terzidis, K., Sarika, L., Mavrikakis, M., Sfikakis, P. and Alevizaki, M. 2013. Hormonal parameters and sex hormone receptor gene polymorphisms in men with autoimmune diseases. *Rheumatology International*, 33(3):575-582,
- Duan, J., Wainwright, M.S., Comeron, J.M., Saitou, N., Sanders, A.R., Gelernter, J. and Gejman, P.V. 2003. Synonymous mutations in the human dopamine receptor D2 (D2D2) affect mRNA stability and synthesis of the receptor. *Human Molecular Genetics*, 12:205-216
- Duan, J., Zhang, J.G., Deng, H.W. and Wang, Y.P. 2013. Comparative studies of copy number variation detection methods for next-generation sequencing technologies. *PLoS One*, 8(3):e59126 DOI:10.1371/journal.pone.0059128

- Ehrlich, M. 2002. DNA methylation in cancer: too much, but also too little. *Oncogene*, 21:5400-5413
- Ehrlich, M. 2009. DNA hypomethylation in cancer cells. *Epigenomics*, 1(2):239-259
- Elghetany, M.T. 1998. Surface marker abnormalities in myelodysplastic syndromes. *Hematologica*, 83:1104-1115.
- Fehniger, T.A., Cooper, M.A., Nuovo, G.J., Cella, M., Facchetti, F., Colonna, M. and Caligiuri, M.A. 2003. CD56bright natural killer cells are present in human lymph nodes and are activated by T cell-derived IL-2: a potential new link between adaptive and innate immunity. *Blood*, 101:3052-3057
- Feinberg, A.P. and Tycko, B. 2004. The history of cancer epigenetics. *Nature Reviews Cancer*, 4:143-153
- Figueroa, M.E., Lugthart, S., Li, Y., Deng, X., Christos, P.J., Schifano, E., Booth, J., Skrabanek, L., Campagne, F., Mazumdar, M., Valk, P.J., Delwel, R. and Melnick, A. 2010. DNA methylation signatures identify biologically distinct subtypes in acute myeloid leukemia. *Cancer Cell*, 17:13-27
- Fisher, L., Gokbuget, N., Schwartz, S., Burmeister, T., Rieder, H., Bruggemann, M., Hoelzer, D. and Thiel, E. 2009. CD56 expression in T-cell acute lymphoblastic leukemia is associated with non-thymic phenotype and resistance to induction therapy but no inferior survival after risk-adapted therapy. *Hematologica*, 94:224-229.
- Forbes, S.A., Beare, D., Gunasekaran, P., Leung, K., Bindal, N., Boutselakis, H., Ding, M., Bamford, S., Cole, C., Ward, S., Kok, C.Y., Jia, M., De, T., Teague, J.W., Stratton, M.R., McDermott, U. and Campbell, P.J. 2014. COSMIC: exploring the world's knowledge of somatic mutations in human cancer. *Nucleic Acids Research*, 43:805-811
- French, D., Yang, W., Hamilton, L.H., Neale, G., Fan, Y., Downing, J.R., Cox, N.J., Pui, C., Evans, W.E. and Relling, M.V. 2008. Concordant gene expression in leukemia cells and normal leukocytes is associated with germline cis-SNPs. *PLoS ONE* 3(5):e2144 DOI:10.1371/journal.pone.0002144
- Gajewski, T., Schreiber, H. and F, Y-X. 2013. Innate and adaptive immune cells in tumor microenvironment. *Nature Immunology*, 14(10):1014-1022
- Gamal, A-H. 2011. Classification of acute leukemia, Acute Leukemia – The Scientist's Perspective and Challenge, Prof. Mariastefania Antica (Ed.), ISBN:978-953-307-553-2
- Garcia-Dabrio, M.C., Pujol-Moix, N., Martinez-Perez, A., Fontcuberta, J., Souto, J.C., Soria, J.M. and Nomdedeu, J.F. 2012. Influence of age, gender and lifestyle in lymphocyte subsets: report from the Spanish Gait-2 study. *Acta Haematologica*, 127:244-249

Garcia-Manero, G., Yang, H., Kuang, S., O'Brien, S., Thomas, D. and Kantarjian, H. 2009. Epigenetics of acute lymphocytic leukemia. *Seminars in Hematology*, 46(1) DOI:10.1053/j.seminhematol.2008.09.008

Gattazzo, C., Teramo, A., Miorin, M., Scquizzato, E., Cabrelle, A., Balsamo, M., Agostin, C., Vendrame, E., Facco, M., Albergoni, M.P., Trentin, L., Vitale, M., Semenzato, G. and Zambello, R. 2010. Lack of expression of inhibitory KIR3DL1 receptor in patients with natural killer cell-type lymphoproliferative disease of granular lymphocytes. *Haematologica*, 95(10):1722-1729

Ghosh, S., Yates, A.J., Eruwald, M.C., Miecznikowski, J.C., Plass, C. and Smiraglia, D. 2010. Tissue specific DNA methylation of CpG islands in normal human adult somatic tissues distinguishes neural from non-neural tissues. *Epigenetics*, 5(6):527-538

Giglio, T., Imro, M.A., Filaci, G., Scudeletti, M., Puppo, F., De Cecco, L., Indiveri, F. and Costantini, S. 1994. Immune cell circulating subsets are affected by gonadal function. *Life Science*, 54:1305-1312

Giltay, E.J., Fonk, J.C., von Blomberg, B.M., Drexhage, H.A., Schalkwijk, C. and Gooren, L.J. 2000. Consistent production of a higher TH1:TH2 cytokine ratio by stimulated T cells in men compared with women. *European Journal of Endocrinology*, 143:31-36

Gismondi, A., Jacobelli, J., Strippoli, R., Maineiro, F., Soriani, A., Cifaldi, L., Piccoli, M., Frati, L. and Santoni, A. 2003. Proline-rich tyrosine kinase 2 and Rac activation by chemokine and integrin receptors controls NK cell transendothelial migration. *Journal of Immunology*, 170(6):3065-3073

Godal, R., Bachanova, V., Gleason, M., McCullar, V., Yun, G.H., Cooley, S., Verneris, M.R., McGlave, P.B. and Miller, J.S. 2010. Natural killer cell killing of acute myelogenous leukemia and acute lymphoblastic leukemia blasts by killer cell immunoglobulin-like receptor-negative natural killer cells after NKG2A and LIR-1 blockade. *Biology Blood Marrow Transplant*, 16:612-621.

Goldstone, A.H., Richards, S.M., Lazarus, H.M., Tallman, M.S., Buck, G., Fielding, A.K., Chopra, R., Wiernik, P.H., Foroni, L., Litzow, M.R., Marks, D.I., Durrant, J., McMillan, A., Franklin, I.M., Luger, S., Ciobanu, N. and Rowe, J.M. 2008. In adults with standard-risk acute lymphoblastic leukemia, the greatest benefit is achieved from a matched sibling allogeneic transplantation in first complete remission, and an autologous transplantation is less effective than conventional consolidation/maintenance chemotherapy in all patients: final results of the International ALL Trial (MRC UKALL XII/ECOG E2993). *Blood*, 111(4):1827-1833

- Gorlov, I.P., Byun, J., Gorlova, O., Aparicio, A.M., Efstatheiu, E. and Logothetis, C.J. 2009. Candidate pathways and genes for prostate cancer: a meta-analysis of gene expression data. *BMC Medical Genomics*, 2:48 DOI:10.1186/1755-8794-2-48
- Gottlieb, A.J., Weinberg, V., Ellson, R.R., Henderson, E.S., Terebelo, H., Rafla, S., Silver, R.T., Carey R.W. and Levy, R.N. 1984 Efficacy of daunorubicin in the therapy of adult acute lymphocytic leukemia: a prospective randomized trial by cancer and leukemia group B. *Blood*, 64:267-274
- Gumperz, J.E., Barber, L.D., Valiante, N.M., Percival, L., Philips, J.H. and Lanier, L.L. 1997. Conserved and variable residues within the Bw4 motif of HLA-B make separable contributions to recognition by the NKB1 killer cell-inhibitory receptor. *Journal of Immunology*, 158:5237-5241
- Gupta, R., Bhaskar, A., Kumar, L., Sharma, A. and Jain, P. 2009. Flow cytometry immunophenotyping and minimal residual disease analysis in multiple myeloma. *American Journal of Clinical Pathology*, 132:728-732
- Guryev, V., Saar, K., Adamovic, T., Verheul, M., van Heesch, S.A., Cook, S., Pravenec, M., Aitman, T., Jacob, H. and Shull, J.D. 2008. Distribution and functional impact of DNA copy number variation in the rat. *Natural Genetics*, 40:538-545
- Harel-Bellan, A., Bertoglio, J., Quillet, A., Marchiol, C., Wakasugi, H., Mishall, Z. and Fradelizi, D. 1986. Interleukin 2 (IL2) up-regulated its own receptor on a subset of human unprimed peripheral blood lymphocytes and triggers their proliferation. *Journal of Immunology*, 136(7):2463-2469
- Harmon, Q.E., Engel, S.M., Olshan, A.F., Moran, T., Stuebe, A.M., Luo, J., Wu, M.C. and Avery, C.L. 2013. Association of polymorphisms in natural killer cell-related genes with preterm birth. *American Journal of Epidemiology*, DOI:10.1093/aje/kwt108
- Harrington, A.M., Hari, P. and Kroft, S.H. 2009. Utility of CD56 immunohistochemical studies in follow-up plasma cell myeloma. *American Journal of Clinical Pathology*, 132:60-66.
- Harris, N.L., Jaffe, E.S., Diebold, J., Flandrin, G., Muller-Hermelink, H.K., Vardiman, J., Lister, T.A. and Bloomfield, C.D. 1999. World Health Organization classification of neoplastic diseases of the hematopoietic and lymphoid tissues: report of the Clinical Advisory Committee meeting-Airlie House, Virginia, November 1997. *Journal of Clinical Oncology*, 17(12):3835-3849
- Hashimoto, S., Toba, K., Aoki, S., Tsuchiyama, J., Tsukada, N., Takahashi, H., Takahashi, M. and Aizawa, Y. 2002. Acute T-lymphoblastic leukemia relapsed with the character of myeloid/natural killer cell precursor phenotype: a case report. *Leukemia Research*, 26:215-219.

- Head, S.R., Komori, H.K., LaMere, S.A., Whisenant, T., Nieuwerburgh, F.V., Salomon, D.R. and Ordoukhalian, P. 2014. Library construction for next-generation sequencing: overviews and challenges. *BioTechniques* 56, 61-77
- Heap, G.A., Trynka, G., Jansen, R.C., Bruinenberg, M., Swertz, M.A., Dinese, L.G., Hunt, K.A., Wijmenga, C., vanHell, D.A. and Franke, L. 2009. Complex nature of SNP genotype effects on gene expression in primary human leucocytes. *BMC Medical Genomics*, 2:1 DOI:10.1186/1755-87940-2-1
- Hellmann, I., Lim, S., Gelman, R. and Letvin, N. 2011. Association of activating KIR copy number variation of NK cells with containment of SIV replication in rhesus monkeys. *PLoS Pathology*, 7(12): e1002436
- Hembruff, S.L., Villeneuve, D.J. and Parissenti, A.M. 2005. The optimization of quantitative reverse transcription PCR for verification of cDNA microarray data. *Analytical Biochemistry*, 345:237-249
- Henrichsen, C.N., Chaignat, E. and Reymond, A. 2009b. Copy number variants, diseases and gene expression. *Human Molecular Genetics*, DOI:10.1093/hmg/ddp011
- Henrichsen, C.N., Vinckenbosch, N., Zollner, S., Chaignat, E., Pradervand, S., Schutz, F., Ruedi, M., Kaessmann, H. and Reymond, A. 2009a. Segmental copy number variation shapes tissue transcriptome. *Nature Genetics*, DOI:10.1038/ng.345
- Herman, J.G., Graff, J.R., Myohanen, S., Nelkin, B.D. and Baylin, S.B. 1996. Methylation-specific PCR: A novel PCR assay for methylation status of GpG islands. *Proceedings of the National Academy of Sciences*, 93:9821-9826
- Hernandez, H.G., Tse, M.Y., Pang, S.C., Arboleda, H. and Forero, D.A. 2013. Optimizing methodologies for PCR-based DNA methylation analysis. *BioTechniques*, 55:181-197
- Hodkinson, B. and Grice, E.A. 2015. Next-generation Sequencing: A review of technologies and tools for wound microbio research. *Advance in Wound Care*, 4(1):50-58
- Homer, N. and Nelson, S.F. 2010. Improved variant discovery through local re-alignment of short-read next-generation sequencing data using SRMA. *Genome Biology*, 11(10):R99 DOI: 10.1186/gb-2010-11-10-r99
- Huntington, N.D., Vosshenrich, C.A. and Di Santo, J.P. 2007. Developmental pathways that generate natural-killer-cell diversity in mice and humans. *Nature Review Immunology*, 7(9):503-714
- Iafrate, A.J., Feuk, L., Rivera, M., Listewnik, M., Donahoe, P., Qi, Y., Scherer, S. and Lee, C. 2004. Detection of large-scale variation in the human genome. *Nature Genetics*, 36:949-951

- Ibrahim, K., Daud, S.S., Seah, Y-L., Yeoh, A.E. and Ariffin, H. 2008. Rapid detection of prognostically important childhood acute lymphoblastic leukemia chimeric transcripts using multiplex SYBR green real-time reverse transcription PCR. *Annals of Clinical and Laboratory Science*, 38(4):338-343
- Igartua, C., Turner, E.H., Ng, S.B., Hodge, E., Hannon, G.J., Bhattacharjee, A., Rieder, M.J., Nickerson, D.A. and Shendure, J. 2010. Targeted enrichment of specific regions in the human genome by array hybridization. *Current Protocols in Human Genetics*, 66:18.3.1-18.3.14
- Impola, U., Turpeinen, H., Alakulppi, N., Linjama, T., Volin, L. and Niittyvuopio, R. 2014. Donor haplotype B of NK KIR receptor reduces the relapse risk in HLA-identical sibling haematopoietic stem cell transplantation of AML patients. *Frontiers in Immunology*, 5:405 DOI:10.3389/fimmu.2014.00405
- Inoshita, M., Numata, S., Tajima, A., Konoshita, M., Umehara, H., Yamamori, H., Hashimoto, R., Imoto, I. and Ohmori, T. 2015. Sex differences of leukocytes DNA methylation adjusted for estimated cellular proportions. *Biology of Sex Differences*, 6:11, DOI:10.1186/s13293-0150-0029-7
- Ishida, Y., Zhao, D., Ohkuchi, A., Kuwata, T., Yoshitake, H., Yuge, K., Takizawa, T., Matsubara, S., Suzuki, M., Saito, S. and Takizawa, T. 2015. Maternal peripheral blood natural killer cells incorporate placenta-associated microRNAs during pregnancy. *International Journal of Molecular Medicine*, 35(6):1511-1524
- Ismail, S.I., Naffa, R.G., Yousef, A.F. and Ghanim, M.T. 2014. Incidence of bcr-abl fusion transcripts in healthy individuals. *Molecular Medicine Reports*, 9:1271-1276
- Jarduli, L.R., Alves, H.V., Souza-Santana, F.C., Marcos, E.V., Pereira, A.C., Dias-Baptista, I.M., Fava, V.M., Mira, M.T., Moraes, M.O., Virmond, M.C. and Visentainer, J.E.. 2014. Influence of KIR genes and their HLA ligands in the pathogenesis of leprosy in a hyperendemic population of Rondonopolis, Southern Brazil. *BMC Infectious Diseases*, 14:438
- Jiang, K., Zhu, F.M., Lv, Q.F. and Yan, L.X. 2005. Distribution of killer cell immunoglobulin-like receptor genes in the Chinese Han population. *Tissue Antigens*, 65:556
- Kang, J.E., Hwang, S.H., Lee, J.H., Park, D.Y. and Kim, H.H. 2011. Effect of RBC removal and TRIzol of peripheral blood samples on RNA stability. *Clinica Chimica Acta*, 412:1883-1885
- Katz, G., Markel, G., Mizrahi, S., Arnon, T.I. and Mandelboim, O. 2001. Recognition of HLA-Cw4 but not HLA-Cw6 by the NK cell receptor killer cell Ig-like receptor two-domain short tail number 4. *Journal of Immunology*, 166:7260-7267

- Kelly, J., Walter, L. and Trowsdale, J. 2005. Comparative genomics of natural killer cell receptor gene clusters. *PLoS Genetics*, DOI: 10.1371/journal.pgen.0010027
- Kennichi, C.D., Daniel, J.C., Erlinda, K. and Stephen, A.S. 2003. NK cells regulate CD4 responses prior to antigen encounter. *The Journal of Immunology*, 171:234-239
- Khong, H.T. and Restifo, N.P. 2002. Natural selection of tumor variants in the generation of “tumor escape” phenotypes. *Nature Immunology*, 3:999-1005
- Kikuchi-Maki, A., Yusa, S., Catina, T.L. and Campbell, K.S. 2004. KIR2DL4 is an IL-2-regulated NK cell receptor that exhibits limited expression in humans but triggers strong IFN-gamma production. *Journal of Immunology*, 171:3415-3425
- Kim, S., Cho, H., Lee, D. and Webster, M.J. 2012. Association between SNPs and gene expression in multiple regions of the human brain. *Translational Psychiatry*, 2:e113 DOI:10.1038/tp.2012.42
- Kim, S., Poursine-Laurent, J., Truscott, S.M., Lybarger, L, Song, Y-J., Yang, L., French, A.R., Sunwoo, J.B., Leumieux, S., Hansen, T.H. and Yokoyama, W.M. 2005. Licensing of natural killer cells by host major histocompatibility complex class I molecules. *Nature*, 436:709-713
- Kim-Schulze, D., Scotto, L., Vlad, G., Piazza, F., Lin, H., Liu, Z., Cortesini, R. and Suciu-Foca, N. 2006. Recombinant Ig-like transcript 3-Fc modulates T cell responses via induction of Th anergy and differentiation of CD8+ T suppressor cells. *Journal of Immunology*, 176:2790-2798
- Kontogianni, K., Nocholson, A.G., Butcher, D. and Sheppard, M.N. 2005. CD56: a useful tool for the diagnosis of small cell lung carcinomas on biopsies with extensive crush artefact. *Journal of Clinical Pathology*, 58:970-980.
- Kopp, H.G., Placke, T. and Salih, H.R. 2009. Platelet-derived transforming growth factor-beta down-regulates NKG2D thereby inhibiting natural killer cell antitumor reactivity. *Cancer Research*, 69(19):7775-5583
- Kroger, N., Binder, T., Zabelina, T., Wolschke, C., Schieder, H. and Renges, H. 2006. Low number of donor activating killer immunoglobulin-like receptors (KIR) genes but not KIR-ligand mismatch prevents relapse and improves disease-free survival in leukemia patients after in vivo T-cell depleted unrelated stem cell transplantation. *Transplantation*, 82:1024-1030
- Kruglyak, L. and Nickerson, D.A. 2001. Variation is the spice of life. *Nature Genetics*, 27:234-236
- Krzysztof, M., David, P., Harper. and Peter, D. 2009. Cytogenetics and molecular genetics of acute lymphoblastic leukemia. *Hematology/Oncology Clinics of North America*, 23:991-1010

- Kulkarni, S., Martin, M.P. and Carrington, M. 2008. The Yin and Yang of HLA and KIR in human disease. *Seminars in Immunology*, 20:343-352
- Kusnierszyk, P. 2013. Killer cell immunoglobulin-like receptor gene associations with autoimmune and allergic diseases, recurrent spontaneous abortion, and neoplasms. *Frontiers in Immunology*, 4:8 DOI: 10.3389/fimmu.2013.00008
- Lanier, L.L. 2005. NK cell recognition. *Annual Review of Immunology*, 23:225-274
- Lanier, L.L. 2007. Back to future – defining NK cells and T cells. *European Journal of Immunology*, 37:1424-1426
- Larson, R.A., Dodge, R.K., Burns, C.P., Lee, E.J., Stone, R.M., Schulman, P., Duggan, D., Davey, F.R., Sobel, R.E. and Frankel, S.R. 1995. A five-drug remission induction regimen with intensive consolidation for adults with acute lymphoblastic leukemia: cancer and leukemia group B study 8811. *Blood*, 85:2025-2037
- Le, Q.H., Thomas, X., Ecochard, R., Iwaz, J., Lheritier, V., Michallet, M. and Fiere, D. 2006. Initial and late prognostic factors to predict survival in adult acute lymphoblastic leukaemia. *European Journal of Haematology*, 77:471-479
- Lee, L.J. and Abdullah, M. 2014. Optimization of genomic DNA shearing by sonication for next-generation sequencing library preparation. *Asia Pacific Journal of Molecular Biology and Biotechnology*, 22(3):200-208
- Lesueur, F., de Lichy, M., Barrois, M., Durand, G., Bombled, J., Avril, M.F., Chompret, A., Boitier, F., Lenoir, G.M., Bressac-de Paillerets, B., Baccard, M., Bacholle, B., Berthet, P., Bonadona, V., Bonnetblanc, J.M., Caron, O., Chevrant-Breton, J., Cuny, J.F., Dalle, S., Delaunay, M., Demange, L., De Quatrebarbes, J., Dore, J.F., Frenay, M., Fricker, J.P., Gautheir-Villars, M., Gestal, P., Giraud, S., Gorry, P. and Grange F. 2008. The contribution of large genomic deletions at the CDKN2A locus to the burden of familial melanoma. *British Journal of Cancer*, 99:364-370.
- Li, R., Li, Y., Fang, X., Yang, H., Wang, J., Kristiansen, K. and Wang, J. 2009. SNP detection for massively parallel whole-genome resequencing. *Genome Research*, 19(6):1124-1132
- Li, W. and Olivier, M. 2012. Current analysis platforms and methods for detecting copy number variation. *Physiological Genomics*, 45:1-16
- Linsley, P.S., Speake, C., Whalen, E. and Chaussabel, D. 2014. Copy number loss of the interferon gene cluster in melanomas is linked to reduced T cell infiltrate and poor patient prognosis. *PLoS ONE* 9(10): e109760
- Liu, J., Morgan, M., Hutchison, K. and Calhoun, V.D. 2010. A study of the influence of sex on genome wide methylation. *PLoS ONE* 5(4): e10028

- Liu, Y., Kuich, R., Hanash, S. and Richardson, B. 2008. DNA methylation inhibition increases T cell KIR expression through effects on both promoter methylation and transcription factors. *Clinical Immunology*, 130:213-224
- Ljunggren, H.G. and Karrer, K. 1990. In search of the ‘missing self’: MHC molecules and NK cell recognition. *Immunology Today*, 11(7):237-244
- Long, E.O. 1999. Regulation of immune responses through inhibitory receptors. *Annual Review of Immunology*, 17:875-904
- Lorenzo, M. 2007. NK cell-mediated immune response against cancer. *Surgical Oncology*, 16:3-5
- Lutterbach, B. and Hiebert, S.W. 2000. Role of the transcription factor AML-1 in acute leukemia and hematopoietic differentiation. *Gene*, 245(2):224-235
- Ma, W., Wang, M., Wang, Z.Q., Sun, L., Graber, D., Matthews, J., Champlin, R., Yi, Q., Orlowski, R.Z., Kwak, L.W., Weber, D.M., Thomas, S.K., Shah, J., Kormblau, S. and Davis, R.E. 2010. Effect of long-term storage in TRIzol on microarray-based gene expression profiling. *Cancer Epidemiology, Biomarkers and Prevention*, 19:2445-2452
- Manchester, K.L. 1996. Use of UV methods for measurement of protein and nucleic acid concentrations. *Biotechniques* 20, 968-970
- Mandelboim, O., Lieberman, N., Lev, M., Paul, L., Arnon, T.I., Bushkin, Y., Davis, J.L., Strominger, J.W. and Yewdell, A. 2001. Recognition of haemagglutinins on virus-infected cells by NKp46 activates lysis by human NK cells. *Nature*, 409:1055-1060
- Marshall-Gradisnik, S., Huth, T., Chacko, A., Johnston, S., Smith, P. and Staines, D. 2016 Natural killer cells and single nucleotide polymorphisms of specific ion channels and receptor genes in myalgic encephalomyelitis/chronic fatigue syndrome. *The Application of Clinical Genetics*, 9:39-47
- Martin, A.M., Kulski, J.K., Witt, C., Pontarotti, P. and Christiansen, F.T. 2002. Leukocyte Ig-like receptor complex (LRC) in mice and men. *Trends in Immunology*, 23:81-88
- Mathot, L., Wallin, M. and Sjöblom, T. 2013. Automated serial extraction of DNA and RNA from biobanked tissue specimens. *BMC Biotechnology* 13:66
- McEwen-Smith, R.M., Salio, M. and Cerundolo, V. 2015. The regulatory role of invariant NKT cells in tumour immunity. *Cancer Immunology Research*, 3(5):425-435
- Melki, J.R., Vincent, P.C. and Clark, S.J. 1999. Concurrent DNA hypermethylation of multiple genes in acute myeloid leukemia. *Cancer Research*, 59:3730-3740

- Michaud, A., Dardari, R., Charrier, E., Cordeiro, P., Herblot, S. and Duval, M. 2010. IL-7 enhances survival of human CD56bright NK cells. *Journal of Immunotherapy*, 33(4):382-390
- Michou, L., Cornelis, F., Baron, M., Bombardieri, S., Balsa, A., Westhovens, R., Barrera, P., Alves, H., Radstake, T.R., Migliorini, P., Bardin, T., Petit-Teixeira, E. and Boillard, E. 2013. Association study of the platelet collagen receptor glycoprotein VI gene with rheumatoid arthritis. *Clinical and Experimental Rheumatology*, 31(5):770-772
- Middleton, D., Curran, M. and Maxwell., L. 2002. Natural killer cells and their receptors. *Transplant Immunology*, 10:147-164
- Miller, D.W., Hague, S.M., Clarimon, J., Baptista, M., Gwinn-Hardy, K., Cookson, M.R. and Singleton, A.B. 2004. Alpha-synuclein in blood and brain from familial Parkinson disease with SNCA locus triplication. *Neurology*, 62:1835-1838
- Monsivais-Urenda, A., Nino-Moreno, P., Abud-Mendoza, C., Baranda, L., Layseca-Espinosa, E., Lopez-Botet, M. and Gonzalez-Amaro, R. 2007. Analysis of expression and function of the inhibitory receptor ILT2 (CD85j/LILRB1/LIR-1) in peripheral blood mononuclear cells from patients with systemic lupus erythematosus (SLE). *Journal of Autoimmunity*, 29:97-105
- Montagna, M., Dalla Palma, M., Menin, C., Agata, S., De Nicolo, A., Chieco-Bianchi, L. and D'Andrea, E. 2003. Genomic rearrangements account for more than one-third of the BRCA1 mutations in northern Italian breast/ovarian cancer families. *Human Molecular Genetics*, 12:1055-1061
- Moretta, A., Biasssoni, R., Bottino, C., Mingari, M.C. and Moretta, L. 2000. Natural cytotoxicity receptors that trigger human NK-mediated cytolysis. *Immunology Today*, 21:228-234
- Moretta, A., Pende, D., Locatelli, F. and Moretta, L. 2009. Activating and inhibitory killer immunoglobulin-like receptors (KIR) in haploidentical haemopoietic stem cell transplantation to cure high-risk leukaemias. *Clinical and Experimental Immunology*, 157:325-337
- Moretta, A., Tambussi, G., Bottino, C.C., Tripodi, G., Merli, A., Ciccone, E., Pantaleo, G. and Moretta, L. 1990. A novel surface antigen expressed by a subset of human CD3-CD16+ natural killer cells: role in cell activation and regulation of cytolytic function. *Journal of Experimental Medicine*, 171:695
- Moretta, A., Vitale, M., Bottino, C., Orengo, A.M., Morelli, L., Augugliaro, R., Barbaresi, M., Ciccone, E. and Moretta, L. 1993. p58 molecules as putative receptors for major histocompatibility complex (MHC) class I molecules in human natural killer (NK) cells: anti-p58 antibodies reconstitute lysis of MHC class I-protected cells in NK clones displaying different specificities. *Journal of Experimental Medicine*, 178:597-604

- Moretta, L., Locatelli, F., Pende, D., Sivori, S., Falco, M., Bottino, C., Mingari, M.C. and Moretta, A. 2011. Human NK receptors: from the molecules to the therapy of high risk leukemias. *FEBS Letters*, 585:1563-1567
- Mori, R., Wang, Q., Danenberg, K.D., Pinski, J.K. and Danenberg, P.V. 2008. Both beta-actin and GAPDH are useful reference genes for normalization of quantitative RT-PCR in human FFPE tissue samples of prostate cancer. *Prostate*, 68(14):1555-1560
- Morice, W.G., Kurtin, P.J., Leibson, P.J., Tefferi, A. and Hanson, C.A. 2003. Demonstration of aberrant T-cell and natural killer-cell antigen expression in all cases of granular lymphocytic leukaemia. *British Journal of Haematology*, 120(6):1026-1036
- Mullighan, C.G., Goorha, S., Radtke, I., Miller, C.B., Coustan-Smith, E., Dalton, J.D., Girtman, K., Mathew, S., Ma, J., Pounds, S.B., Su, X., Pui, C.H., Relling, M.V., Evans, W.E., Shurtliff, S.A. and Downing, J.R. 2007. Genome-wide analysis of genetic alterations in acute lymphoblastic leukaemia. *Nature*, 446:758-764
- Naluyima, P., Eller, L.A., Ouma, B.J., Kyabaggu, D., Kataaha, P., Guwatudde, D., Kibuuka, H., Wabwire-Mangen, F., Robb, M.L., Michael, N.L., Souza, M.S., Sandberg, J.K. and Eller, M.A. 2016. Sex and urbanicity contribute to variation in lymphocyte distribution across Ugandan populations. *PLOS one*, DOI: 10.1371/journal.pone.0146196
- Narayanan, D. and Shami, P.J. 2012. Treatment of acute lymphoblastic leukemia in adults. *Critical Reviews in Oncology/Hematology*, 81(1):94-102
- Natarajan, K., Dimasi, N., Wang, J., Mariuzza, R.A. and Margulies, D.H. 2002. Structure and function of natural killer cell receptors: multiple molecular solutions to self, nonself discrimination. *Annual Review of Immunology*, 20:853-885
- Ng, P.C. and Kirkness, E.F. 2010. Whole genome sequencing. *Methods in Molecular Biology*, 628:215-226
- Nielsen, R., Paul, J.S., Albrechtsem, A. and Song, Y.S. 2011. Genotype and SNP calling from next-generation sequencing data. *Nature Reviews Genetics*, 12(6):443-451
- Norman, P., Stephens, H.A., Verity, D.H., Chandanayyingying, D. and Vaughan R.W.J. 2001. Distribution of natural killer cell immunoglobulin-like receptor sequences in three ethnic groups. *Immunogenetics*, 52:195
- Norris, S., Doherty, D.G., Curry, M., McEntee, G., Traynor, O., Hegarty, J.E. and O'Farrelly, C. 2003. Selective reduction of natural killer cells and T cells expressing inhibitory receptors for MHC class I in the livers of patients with hepatic malignancy. *Cancer Immunology, Immunotherapy*, 52(1):53-58

- Nowakowski, G.S., Morice, W.G., Phyliky, R.L., Li, C.Y. and Tefferi, A. 2005. Human leucocyte antigen class I and killer immunoglobulin-like receptor expression pattern in T-cell large granular lymphocyte leukaemia. *British Journal of Haematology*, 128(4):490-492
- Nugent, B.M., Wright, C.L., Shetty, A.C., Hodes, G.E., Lenz, K.M., Mahurkar, A., Russo, S.J. Devine, S.E. and McCarthy, M.M. 2015. Brain feminization requires active repression of masculinization via DNA methylation. *Nature Neuroscience*, 18:690-697
- O'Connor, G.M., Holmes, A., Mulcahy, F. and Gardiner, C.M. 2007. Natural killer cells from long-term non-progressor HIV patients are characterized by altered phenotype and function. *Clinical Immunology*, 124:277-283
- Ogata, K., An, E., Shioi, Y., Nakamura, K., Luo, S., Yokose, N. and Dan, K. 2001. Association between natural killer cell activity and infection in immunologically normal elderly people. *Clinical & Experimental Immunology*, 124:392-397
- Okamoto, R., Ogawa, S., Nowak, D., Kawamata, N., Akagi, T., Kato, M., Sanada, M., Weiss, T., Haferlach, C., Dugas, M., Ruckert, C., Haferlach, T. and Koeffler, H.P. 2010. Genomic profiling of adult acute lymphoblastic leukemia by single nucleotide polymorphism oligonucleotide microarray and comparison to pediatric acute lymphoblastic leukemia. *Haematologica*, 95:1481-1488
- Olivier, M. 2004. From SNPs to function: the effect of sequence variation on gene expression. Focus on "A survey of genetic and epigenetic variation affecting human gene expression. *Physiological Genomics*, 16(2):182-183
- Omar, Z.A. and Ibrahim Tamam, N.S. 2011. National Cancer Registry Report, Malaysian Cancer Statistics – Data and Figure National Cancer, *Ministry of Health, Malaysia* page 1-126
- Orange, J.S. and Ballas, Z.K. 2006. Natural killer cells in human health and disease. *Clinical Immunology*, 118:1-10
- Orange, J.S., Fassett, M.S., Koopman, L.A., Boyson, J.E. and Strominger, J.L. 2002. Viral evasion of natural killer cells. *Nature Immunology*, 3:1006-1012
- Oshimi, K., Gonda, N., Sumiya, M. and Kano, S. 1980. Effects of corticosteroids on natural killer cell activity in systemic lupus erythematosus. *Clinical and Experimental Immunology*, 40:83-88
- Owens, M.A., Vall, H.G., Hurley, A.A. and Wormsley, S.B. 2000. Validation and quality control of immunophenotyping in clinical flow cytometry. *Journal of Immunological Methods*, 243:33-50

- Palmen, J., Smith, A.J.P., Dorfmeister, B., Putt, W., Humphries, S.E. and Talmud, P.J. 2008. The functional interaction on *in vitro* gene expression of APOA5 SNPs, defining haplotype APOA5*2, and their paradoxical association with plasma triglyceride but not plasma apoAV levels. *Biochimica et Biophysica Acta (BBA) – Molecular Basis of Disease*, 8:447-452
- Paltiel, O., Susan, H., Lisa, D., Ariella, K., Sausan, M., Efrat, T., Micha, B. and Yehiel, F. 2004. Birth weight and other risk factors for acute leukemia in the Jerusalem Perinatal Study Cohort. *Cancer Epidemiology, Biomarkers & Prevention*, 13:1057-1062
- Pang, A.W., MacDonald, J.R., Pinto, D., Wei, J., Rafiq, M.A., Conrad, D.F., Park, H., Hurles, M.E., Lee, C., Venter, J.C., Kirkness, E.F., Levy, S., Feuk, L. and Scherer, S.W. 2010. Towards a comprehensive structural variation map of an individual human genome. *Genome Biology*, 11:R52
- Pang, A.W., MacDonald, J.R., Pinto, D., Wei, J., Rafiq, M.A., Conrad, D.F., Park, H., Hurles, M.E., Lee, C., Venter, J.C., Kirkness, E.F., Levy, S., Feuk, L. and Scherer, S.W. 2010. Towards a comprehensive structural variation map of an individual human genome. *Genome Biology*, 11:R52
- Papaemmanuil, E., Hosking, F.J., Vijayakrishnan, J., Price, A., Olver, B., Sheridan, E., Kinsey, S.E., Lightfoot, T., Roman, E., Irving, J.A.E., Allan, J.M., Tomlinson, I.P., Taylor, M., Greaves, M. and Houlston, R.S. 2009. Loci on 7p12.2, 10q21.2 and 14q11.2 are associated with risk of childhood acute lymphoblastic leukemia. *Nature Genetics*, 41:1006-1010
- Parham, P. 2005. MHC class I molecules and KIRs in human history, health and survival. *Nature Review of Immunology*, 5:201-214
- Pastinen, T., Sladek, R., Gurd, S., Sammak, A., Ge, B., Lepage, P., Lavergne, K., Villeneuve, A., Gaudin, T., Brandstrom, H., Beck, A., Verner, A., Kingsley, J., Harmsen, E., Labuda, D., Morgan, K., Vohl, M.C., Naumova, A.K., Sinnott, D. and Hudson, T.J. 2004. A survey of genetic and epigenetic variation affecting human gene expression. *Physiological Genomics*, 16(2):184-193
- Paul, B., Mukiihi, J.M., Mandisodza, A., Levy, L. and Nkrumah, F.K. 1992. A three-year prospective study of 137 cases of acute leukaemia in Zimbabwe. *Central African Journal of Medicine*, 38:95-99
- Plaschke, J., Ruschoff, J. and Schackert, H.K. 2003. Genomic rearrangements of hMSH6 contribute to the genetic predisposition in suspected hereditary non-polyposis colorectal cancer syndrome. *Journal of Medical Genetics*, 40:597-600
- Plougastel, B. and Trowsdale, J. 1997. Cloning of NKG2-F, a new member of the NKG2 family of human natural killer cell receptor genes. *European Journal of Immunology*, 27:2835-2839

- Qin, Q., Zhao, B.R., Mao, Y.M., Cui, R.Z., Kou, L., Li, Y.L., Zhao, F.M. and Hui, R.T. 2005. Association of matrix metalloproteinase-9 and platelet membrane glycoprotein VI polymorphisms with acute coronary syndrome. *Zhonghua Xin Xue Guan Bing Za Zhi*, 33(7):622-626
- Rajagopalan, S. and Long, E.O. 1999. A human histocompatibility leukocyte antigen (HLA)-G-specific receptor expressed on all natural killer cells. *Journal of Experimental Medicine*, 189:1093-1100
- Rajalingam, R. 2002. Diversity of NK cell receptors and their HLA class I ligands.
- Raspadori, D., Damiani, D., Michieli, M., Stocchi, R., Gentili, S., Gozzetti, A., Masolini, P., Michelutti, A., Geromin, A., Fanin, R. and Lauria, F. 2002. CD56 and PGP expression in acute myeloid leukemia: impact on clinical outcome. *Haematologica*, 87:1135-1140.
- Raulet, D.H. 2006. Missing self recognition and self tolerance of natural killer (NK) cells. *Seminars in Immunology*, 18:145-150
- Raulet, D.H., Vance, R.E. and McMahon, C.W. 2001. Regulation of the natural killer cell receptor repertoire. *Annual Review of Immunology*, 19:291-330
- Reboucas, E., Costa, J., Passos, M.J., Passos, J.R.S., Hurk, R. and Silva, J.R. 2013. Real time PCR and importance of housekeepings genes for normalization and quantification of mRNA expression in different tissues. *Brazilian Archives of Biology and Technology*, 56(1):143-154
- Reichert, T., Debruyere, M., Deneys, V., Totterman, T., Lydyard, P., Yuksel, F., Chaper, H., Jewell, D., Hove, V., Linden, J. and Buchner, L. 1991. Lymphocyte subset reference ranges in adult Caucasians. *Clinical Immunology and Immunopathology*, 60:190-208
- Risch, N. 2000. Searching for genetic determinants in the new millennium. *Nature*, 405(6788):847-856
- Risch, N. 2001. The genetic epidemiology of cancer: interpreting family and twin studies and their implications for molecular genetic approaches. *Cancer Epidemiology Biomarkers Preview*, 10:733-741
- Roitt, I., Brostoff, J. and Male, D. 2001. *Immunology* (6th Eds.). pg 480. St. Louis: Mosby
- Rovelet-Lecrux, A., Hanneguin, D., Raux, G., Le Meur, N., Lagurriere, A., Vital, A., Dumanchin, C., Feuillette, S., Brice, A., Vercelletto, M., Dubas, F., Frebourg, T. and Campion, D. 2006. APP locus duplication causes autosomal dominant early-onset Alzheimer disease with cerebral amyloid angiopathy. *Nature Genetics*, 38:24-26

- Rowe, J.M., Buck, G., Burnett, A.K., Chopra, R., Wiernik, P.H., Richards, S.M., Lazarus, H.M., Franklin, I.M., Litzow, M.R., Ciobanu, N., Prentice, H.G., Durrant, J., Tallman, M.S. and Goldstone, A.H. 2005. Induction therapy for adults with acute lymphoblastic leukemia: results of more than 1500 patients from the international ALL trial: MRC UKALL XII/ECOG E2993. *Blood*, 106:3760-3767
- Ruggeri, L., Capanni, M., Urbani, E., Perruccio, K., Shlomchik, W.D. and Tosti, A. 2002. Effectiveness of donor natural killer cell alloreactivity in mis-matched hematopoietic transplants. *Science*, 295:2097
- Sakharkar, M.K., Chow, V.T. and Kangueane, P. 2004. Distributions of exons and introns in the human genome. *In Silico Biology* 4, 387-393
- Sambrook, J., Fritsh, E. and Maniatis, T. 2001. Molecular cloning: a laboratory manual 3rd edition. New York: *Cold Spring Harbor Press*
- Santoli, D., Trinchieri, G., Zmijewski, C.M. and Koprowski, H. 1976. HLA-related control of spontaneous and antibody-dependent cell-mediated cytotoxic activity in humans. *Journal of Immunology*, 117:765
- Santourlidis, S., Trompeter, H., Weinhold, S., Eisermann, B., Meyer, K.L., Wernet, P. and Uhrberg, M. 2002. Crucial role of DNA methylation in determination of clonally distributed killer cell Ig-like receptor expression patterns in NK cells. *The Journal of Immunology*, 169:4253-4261
- Santourlidis, S., Trompeter, H., Weinhold, S., Eisermann, B., Meyer, K.L., Wernet, P. and Uhrberg, M. 2002. Crucial role of DNA methylation in determination of clonally distributed killer cell Ig-like receptor expression patterns in NK cells. *The Journal of Immunology*, 169:4253-4261
- Scavino, H.F., George, J.N. and Sears, D.A. 1976. Remission induction in adult acute lymphocytic leukemia. Use of vincristine and prednisone alone. *Cancer*, 38(2):672-677
- Schonfeld, S.J., Neta, G., Sturgis, R.M., Pfeiffer, R.M., Hutchinson, A.A., Xu, L., Wheeler, W., Guenel, P., Rajaraman, P., de Vathaire, F., Ron, E., Tucker, M.A., Chanock, S.J., Sigurdson, A.J. and Brenner, A.V. 2012. Common genetic variants in sex hormone pathway genes and papillary thyroid cancer risk. *Thyroid*, 22(2):151-156
- Schumacher, H.R., Alvares, C.J., Blough, R.I. and Mazzella, F. 2002. Acute leukemia. *Clinics in Laboratory Medicine*, 22(1):153-192
- Schurks, M., Rist, P.M. and Kurth, T. 2010. Sex hormone receptor gene polymorphisms and migraine: a systematic review and meta-analysis. *Cephalgia*, 30(11):1306-1328

- Sebat, J., Lakshmi, B., Malhotra, D., Troge, J., Lese-Martin, C., Walsh, T., Yamrom, B., Yoon, S., Krasnitz, A., Kendall, J., Leotta, A., Pai, D., Zhang, R., Lee, Y.H., Hicks, J., Spence, S.J., Lee, A.T., Puura, K., Lehtimaki, T., Ledbetter, D., Gregersen, P.K., Bregman, J., Sutcliffe, J.S., Jobanputra, V., Chung, W., Warburton, D., King, M.C., Skuse, D., Geschwind, D.H., Gilliam, T.C., Ye, K. and Wigler, M. 2007. Strong association of de novo copy number mutations with autism. *Science*, 316:445-449
- Sebat, J., Lakshmi, B., Troge, J., Alexander, J., Young, J., Lundin, P., Maner, S., Massa, H., Walker, M., Chi, M., Navin, N., Lucito, R., Healy, J., Hicks, J., Ye, K., Reiner, A., Gilliam, T., Trask, B., Patterson, N., Zetterberg, A. and Wigler, M. 2004. Large-scale copy number polymorphism in the human genome. *Science*, 305:525-528
- Shaffer, J.R., Kammerer, C.M., Dorn, J., Ferrell, R.E., Iacoviello, L., Trevisan, M. and Donahue, R.P. 2011. Polymorphisms in the platelet-specific collagen receptor GP6 are associated with risk of nonfatal myocardial infarction in Caucasians. *Nutrition, Metabolism and Cardiovascular Diseases*, 21(8):546-552
- Shahsavar, F., Tajik, N., Entezami, K.Z., Fallah, R.M., Asadifar, B., Alimoghaddam, K., Ostadali, D.M., Jalali, A., Ghashqhaie, A. and Ghavamzadeh, A. 2010. KIR2DS3 is associated with protection against acute myeloid leukemia. *Iranian Journal of Immunology*, 7(1):8-17
- Shastry, B.S. 2009. SNPs: impact on gene function and phenotype. *Methods in Molecular Biology*, 578:3-22
- Shen, L. and Waterland, R.A. 2007. Methods of DNA methylation analysis. *Current Opinion in Clinical Nutrition and Metabolic Care*, 10:576-581
- Shokere, L.A., Holden, M.J. and Jenkins, G.R. 2009. Comparison of fluorometric and spectrophotometric DNA quantification for real-time quantitative PCR of degraded DNA. *Food Control* 20, 391-401
- Simbole, M., Gottardi, M., Corbo, V., Fassan, M., Mafficini, A., Malpeli, G., Lawlor, R.T. and Scarpa, A. 2013. DNA qualification workflow for next-generation sequencing of histopathological samples. *PLoS ONE*, 8:1-8
- Singh, N., Frey, N.V., Grupp, S.A. and Maude, S.L. 2016. CAR T cell therapy in acute lymphoblastic leukemia and potential for chronic lymphocytic leukemia. *Current Treatment Options in Oncology*, 17(6):28
- Single, R.M., Martin, M.P., Gao, X., Meyer, D., Yeager, M. and Kidd, J.R. 2007. Global diversity and evidence for coevolution of KIR and HLA. *Nature Genetics*, 39:1114-1119

- Singmann, P., Shem-Tov, D., Wahl, S., Grallert, H., Fiorito, G., Shin, S.Y., Schramm, K., Wolf, P., Kunze, S., Baran, Y., Guarnera, S., Vineis, P., Krogh, V., Panico, S., Tumino, R., Kretschmer, A., Gieger, C., Peters, A., Prokisch, H., Relton, C.L., Matullo, G., Illiq, T., Waldenberger, M. and Halperin, E. 2015. Characterization of whole-genome autosomal differences of DNA methylation between men and women. *Epigenetics and Chromatin*, 8:43, DOI:10.1186/s13072-015-0035-3
- Sivori, S., Pende, D., Bottino, C., Marcenaro, E., Pessino, A., Biassoni, R., Moretta, L. and Moretta, A. 1999. NKp46 is the major triggering receptor involved in the natural cytotoxicity of fresh or cultured human NK cells. Correlation between surface density of NKp46 and natural cytotoxicity against autologous, allogenic or xenogenic target cells. *European Journal of Immunology*, 29:1656-1666
- Sokol, J., Biringer, K., Skerenova, M., Hasko, M., Bartosova, L., Stasko, J., Danko, J. and Kubisz, P. 2012. Platelet aggregation abnormalities in patients with fetal losses: the GP6 gene polymorphism. *Fertility and Sterility*, 98(5):1170-1174
- Son, S.W., Kim, E.O., Ryu, E.S., Kim, T.J., Kim, J.N., Choi, J.E., Kye, Y.C. and Lee, K.M. 2009. Upregulation of Fas and downregulation of CD94/NKG2A inhibitory receptors on circulating natural killer cells in patients with new-onset psoriasis. *The British Journal of Dermatology*, 161(2):281-288
- Souza, S.S., Castro, F.A., Mendonca, H.C., Palma, P.V., Morais, F.R., Ferriani, R.A. and Volteralli, J.C. 2001. Influence of menstrual cycle on NK activity. *Journal of Reproductive Immunology*, 50:151-159
- Spinelli, O., Peruta, B., Tosi, M., Guerini, V., Salvi, A., Zanotti, M.C., Oldani, E., Grassi, A., Intermesoli, T., Mico, C., Rossi, G., Fabris, P., Lambertenghi-Deliliears, G., Angelucci, E., Barbui, T., Bassan, R. and Rambaldi, A. 2007. Clearance of minimal residual disease after allogeneic stem cell transplantation and the prediction of the clinical outcome of adult patients with high-risk acute lymphoblastic leukemia. *Haematologica*, 95:612-618
- Stankiewicz, P. and Lupski, J.R. 2010. Structural variation in the human genome and its role in disease. *Annual Review of Medicine*, 61:437-455
- Stefansson, H., Rujescu, D., Cichon, S., Pietilainen, O.P.H., Ingason, A., Steinberg, S., Fossdal, R., Sigurdsson, E. and Buizer-Voskamp, J.E. 2008. Larger recurrent microdeletions associated with schizophrenia. *Nature*, 455:232-236
- Stepp, S.E., Dufourcq-Lagelouse, R., Le Deist, F., Bhawan, S., Certain, S., Mathew, P.A., Henter, J.I., Bennett, M., Fischer, A., de Saint Basile, G. and Kumar, V. 1999. Perforin gene defects in familial hemophagocytic lymphogistiocytosis. *Science*, 286:1957-1959
- Stern, M., Paulussen, M., Rischewski, J., Tichelli, A. and Gratwohl, A. 2008. Missing ligand model in autologous stem cell transplantation. *British Journal of Cancer*, 98:852-853

- Stirzaker, C., Song, J.Z., Davidson, B. and Clark, S.J. 2004. Transcriptional gene silencing promotes DNA hypermethylation through a sequential change in chromatin modifications in cancer cells. *Cancer Research*, 6:3871-3877
- Stranger, B.E., Forrest, M.S., Dunning, M., Ingle, C.E., Beazley, C., Thorne, N., Redon, R., Bird, C.P., Grassi, A., Lee, C., Tyler-Smith, C., Carter, N., Scherer, S.W., Tavare, S., Deloukas, P., Hurles, M. E. and Dermitzakis, E.T. 2007. Relative impact of nucleotide and copy number variation on gene expression phenotypes. *Science*, 5813:848-853
- Stratton, M. 2008. Genome resequencing and genetic variation. *Nature Biotechnology*, 26:65-66
- Stringaris, K., Sekine, T., Khoder, A., Alsuliman, A., Razzaghi, B., Sargeant, R., Pavlu, J., Brisley, G., Lavallade, H., Sarvaria, A., Marin, D., Mielke, S., Apperley, J.F., Shpall, E.J., Barrett, J. and Rezvani, K. 2014. Leukemia-induced phenotypic and functional defects in natural killer cells predict failure to achieve remission in acute myeloid leukemia. *Haematologica*, 99(5):836-847
- Su, S.T., Lin, S.H. and Chen, Y.C. 2011. Association of sex hormone receptor gene polymorphisms with recurrent pregnancy loss: a systematic review and meta-analysis. *Fertility and Sterility*, 96(6):1435-1444
- Suciufoca, N., Feirt, N., Zhang, Q.Y., Vlad, G., Liu, Z., Lin, H., Chang, C.C., Ho, E.K., Colovai, A., Kaufman, H., D'Agati, V.D., Thaker, H.M., Remotti, H., Galluzzo, S., Cinti, P., Rabitti, C., Allendorf, J., Chabot, J., Caricato, M., Coppola, R., Berloco, P. and Cortesini, R. 2007. Soluble Ig-like transcript 3 inhibits tumor allograft rejection in humanized SCID mice and T cell responses in cancer patients. *Journal of Immunology*, 178(11):7432-7441
- Sun, Y., Liu, J., Gao, P., Wang, Y. and Liu, C. 2008. Expression of Ig-like transcript 4 inhibitory receptor in human non-small cell lung cancer. *Chest*, 134:783-788
- Suttmann, H., Jocham, D., Bohle, A. and Brandau, S. 2002. The role of LFA-1 in the lysis of bladder cancer cells by bacillus Calmette-Guerin and interleukin 2-activated killer cells. *Urological Research*, 30(4):233-239
- Szczepanski, T., van der, V. and van Dongen, J.J.M. 2003. Classification systems for acute and chronic leukaemias. *Best Practice and Research Clinical Haematology*, 16(4):561-582
- Takei, F., McQueen, K.L., Maeda, M., Willhelm, B.T., Lohwasser, S., Lian, R.H. and Mager, D.L. 2001. Ly49 and CD94/NKG2: developmentally regulated expression and evolution. *Immunological Reviews*, 181:90-103

- Talkowski, M.E., Ernst, C., Heilbut, A., Chiang, C., Hanscom, C., Lindgren, A., Kirby, A., Liu, S., Muddukrishna, B., Ohsumu, T.K., Shen, Y., Borowsky, M., Daly, M.J., Morton, C.C. and Gusella, J.F. 2011. Next-generation sequencing strategies enable routine detection of balanced chromosome rearrangements for clinical diagnostics and genetic research. *The American Journal of Human Genetics*, 88:469-481
- Tay, C.H., Szomolanyi-Tsuda, E. and Welsh, R.M. 1998. Control of infections by NK cells. *Current Topics in Microbiology and Immunology*, 230:193-220
- Teitel, M.A. and Pandolfi, P.P. 2009. Molecular genetics of acute lymphoblastic leukemia. *Annual Review of Pathology: Mechanisms of Disease*, 4:175-198
- Toneva, M., Lepage, V. and Michaylov, A. 2001. Genomic diversity of natural killer cell receptor genes in three populations. *Tissue Antigens*, 57:358-362.
- Torkar, M., Haude, A., Milne, S., Beck, S. and Trowsdale, J. 2000. Arrangement of the ILT gene cluster: A common null allele of the ILT6 gene results from a 6.7-kbp deletion. *European Journal of Immunology*, 30:3655-3662
- Trigunaite, A., Dimo, J. and Jorgensen, T.N. 2015. Suppressive effects of androgens on the immune system. *Cellular Immunology*, 294:87-94
- Trundley A., Frebel, H., Jones, D., Chang, C. and Trowsdale, J. 2007. Allelic expression patterns of KIR3DS1 and 3DL1 usin the Z27 and DX9 antibodies. *European Journal of Immunology*, 37:780-787
- Turner, P., McLennan, A. and Bates, A. 2005. Molecular Biology 3rd. London: Taylor and Francis Ltd
- Uhrberg, M. 2005. Shaping the human NK cell repertoire: an epigenetic glance at KIR gene regulation. *Molecular Immunology*, 42:471-475
- Uhrberg, M., Parham, P. and Wernet, P. 2002. Definition of gene content for nine common group B haplotypes of the Caucasoid population: KIR haplotypes contain between seven and eleven KIR genes. *Immunogenetics*, 54(4):221-229
- Uhrberg, M., Valiante, N.M. and Shum, B.P. 1997. Human diversity in killer cell inhibitory receptor genes. *Immunity*, 7(6):753-763.
- Uhrberg, M., Valiante, N.M., Shum, B.P., Shiling, H.G., Liener-Weidenbach, K., Corliss, B., Tyan, D., Lanier, L.L. and Parham, P. 1997. Human diversity in killer cell inhibitory receptor genes. *Immunity*, 7:753
- Vales-Gomez, M., Reyburn, H.T., Erskine, R.A., Lopez-Botet, M. and Strominger, J.L. 1999. Kinetics and peptide dependency of the binding of the inhibitory NK receptor CD94/NKG2-A and activating receptor CD94/NKG2-C to HLA-E. *The EMBO Journal*, 18:4250-4260

- Valiante, N.M., Uhrberg, M., Shilling, H.G., Lienert-Weidenbach, K., Arnett, K.L., D'Andrea, A., Phillips, J.H., Lanier, L.L. and Parham P. 1997. Functionally and structurally distinct NK cell receptor repertoires in the peripheral blood of two human donors. *Immunity*, 7(6):739-751
- van Dongen, J.J., Seriu, T., Panzer-Grumayer, E.R., Biondi, A., Pongers-Willemse, M.J., Corral, L., Stolz, F., Schrappe, M., Masera, G., Kamps, W.A., Gadner, H., van Wering, E.R., Ludwig, W.D., Basso, G., de Bruijn, M.A., Cazaniga, G., Hettinger, K., van der Does-van den Berg, A., Hop, W.C., Riehm, H. and Bartram, C.R. 1998. Prognostic value of minimal residual disease in acute lymphoblastic leukaemia in childhood. *The Lancet*, 352:1731-1738
- Vardiman, J.W. 2010. The World Health Organization (WHO) classification of tumors of the hematopoietic and lymphoid tissues: an overview with emphasis on the myeloid neoplasms. *Chemico-Biological Interaction*. 184(1-2):16-20
- Vardiman, J.W., Harris, N.L. and Brunning, R.D. 2002. The World Health Organization (WHO) classification of the myeloid neoplasms. *Blood*, 100:2292-2302
- Venstrom, J., Pittari, G., Gooley, T.A., Chewming, J.H., Spellman, S., Haagenson, M. and Katharine, C.H. 2012. HLA-C-dependent prevention of leukemia relapse by donor activating KIR2DS1. *The New England Journal of Medicine*, 367:805-816
- Verheyden, S., Bernier, M. and Demanet, C. 2004. Identification of natural killer cell receptor phenotypes associated with leukemia. *Leukemia*, 18:2002-2007
- Vivier, E., Nunes, J.A. and Vely, F. 2004. Natural killer cell signaling pathways. *Science*, 306:1517-1519
- Vivier, E., Tomasello, E., Baratin, M., Walzer, T. and Ugolini, S. 2008. Functions of natural killer cells. *Nature Immunology*, 9:503-510
- Volpi, L., Roversi, G., Colombo, E.A., Leijsten, N., Concolino, D., Calabria, A., Mencarelli, M.A., Fimiani, M., Macciardi, F., Pfundt, R., Schoenmakers, E.F. and Larizza, L. 2010. Targeted next-generation sequencing appoints c16orf57 as clericuzio-type poikiloderma with neutropenia gene. *The American Journal of Human Genetics*, 86:72-76
- Wang, L., Kang, N., Zhou, J., Guo, Y., Zhang, X., Cui, L., Ba, D. and He, W. 2012. Downregulation of CD94/NKG2A inhibitory receptor on decreased $\gamma\delta$ T cells in patients with systemic lupus erythematosus. *Scandinavian Journal of Immunology*, 76(1):62-69
- Wang, R., Jaw, J.J., Stutzman, N.C., Zou, Z. and Sun, P.D. 2012. Natural killer cell-produced IFN- γ and TNF- α induce target cell cytosis through up-regulation of ICAM-1. *Journal of Leukocyte Biology*, 91(2):299-309

- Wauquier, N., Padilla, C., Becquart, P., Leroy, E. and Vieillard, V. 2010. Association of KIR2DS1 and KIR2DS3 with fatal outcome in Ebola virus infection. *Immunogenetics*, 62(11-12):767-771
- Weiss, M.A., Heffner, L. and Lamanna, N. 2005. A randomized trial of cytarabine compared to a standad vincristine/prednisone-based regime as induction therapy. *Journal of Clinical Oncology*, 23:6516
- Weisser, M., Haferlach, T., Schoch, C., Hiddemann, W. and Schnittger, S. 2004. The use of housekeeping genes for real-time PCR-based quantification of fusion gene transcripts in acute leukemia. *Leukemia*, 18:1551-1553
- Westgaard, I.H., Berg, S.F., Orstavik, S., Fossum, S. and Dissen, E. 1998. Identification of a human member of the Ly-49 multigene family. *European Journal of Immunology*, 28: 1839-1846
- White, D.L., Liu, Y., Garcia, J., El-Serag, H.B., Jiao, L., Tsavachidis, S., Franco, L.M., Lee, J-S., Tavakoli-Tabasi, S., Moore, D., Goldman, R., Kuzniarek, J., Ramsey, D.J., Kanwal, F. and Marcelli, M. 2014. Sex hormone pathway gene polymorphisms are associated with risk of advanced hepatitis C-related liver disease in males. *International Journal of Molecular Epidemiology and Genetics*. 5(3):164-176
- Wiendl, H., Feger, U., Mittelbronn, M., Jack, C., Schreiner, B., Stadelmann, C., Antel, J., Brueck, W., Meyermann, R., Bar-Or, A., Kieseier, B.C. and Weller, M. 2005. Expression of the immune-tolerogenic major histocompatibility molecule HLA-G in multiple sclerosis: implications for CNS immunity. *Brain*, 128(11):2689-2704
- Winter, C.C. and Long., E. 1997. A single amino acid in the p58 killer cell inhibitory receptor controls the ability of natural killer cells to discriminate between the two groups of HLA-C allotypes. *Journal of Immunology*, 158:4026-4028
- Wu, G., Huang, S., Nastiuk, K.L., Li, J., Gu, J., Wu, M., Zhang, Q., Lin, H. and Wu, D. 2015. Variant allele of HSD3B1 increases progression to castration-resistant prostate cancer. *The Prostate*, 75(7):777-782
- Wu, J., Song, Y., Bakker, A.B., Bauer, S., Spies, T., Lanier, L.L. and Phillips, J.H. 1999. An activating immunoreceptor complex formed by NKG2D and DAP10. *Science*, 285(5428):730-732
- Yang-Fang, T., Dong, W., Li, P., Wen-Li, Z., Jun, L. and Jian, P. 2012. Analyzing the gene expression profile of pediatric acute myeloid leukemia with real-time PCR arrays. *Cancer Cell International*, 12:40
- Yawata, M., Yawata, N., Draghi, M., Little, A-M., Partheniou, F. and Parham, P. 2006. Role for HLA and KIR polymorphisms in natural killer cell repertoire selection and modulation of effector function. *Journal of Experimental Medicine*, 203(3):633-654

- Yeoh, A. E-J., Lu, Y., Chan, J. Y-S., Chan, Y.H., Ariffin, H., Kham, S. K-Y. and Quah, T.C. 2010. Genetic susceptibility to childhood acute lymphoblastic leukemia shows protection in Malay boys: Results from the Malaysia-Singapore ALL study group. *Leukemia Research*, 34:276-283
- Yovel, G., Shakhar, K. and Ben-Eliyahu, S. 2001. The effects of sex, menstrual cycle, and oral contraceptives on the number and activity of natural killer cells. *Gynecologic Oncology*, 81:254-262
- Yu, J., Mao, H.C., Wei, M., Hughes, T., Zhang, J., Park, I.K., Liu, S., McClory, S., Marcucci, G., Trotta, R. and Caligiuri, M.A. 2010. CD94 surface density identifies a functional intermediary between the CD56bright and CD56dim human NK-cell subsets. *Blood*, 115(2):274-281
- Yuan, Y., Liu, L., Chen, H., Wang, Y., Xu, Y., Mao, H., Li, J., Mills, G.B., Shu, Y., Li, L. and Liang, H. 2016. Comprehensive characterization of molecular differences in cancer between male and female patients. *Cancer Cell*, 29:711-722
- Zambello, R., Falco, M., Della Chiesa, M., Trentin, L., Carollo, D. and Castriconi, R. 2003. Expression and function of KIR and natural cytotoxicity receptors in NK-type lymphoproliferative disease of granular lymphocytes. *Blood*, 102(5): 1797-1805
- Zeddou, M., Rahmouni, S., Vandamme, A., Jacobs, N., Frippiat, F., Leonard, P., Schaaf-Lafontaine, N., Vaira, D., Boniver, J. and Moustschén, M. 2007. Downregulation of CD94/NKG2A inhibitory receptors on CD8+ T cells in HIV infection is more pronounced in subjects with detected viral load than in their aviraemic counterparts. *Retrovirology*, 4:72
- Zhang, F.F., Cardarelli, R., Carroll, J., Fulda, K.G., Kaur, M., Gonzalez, K., Vishwanathan, J.K., Santella, R.M. and Morabia, A. 2011. Significant differences in global genomic DNA methylation by gender and race/ethnicity in peripheral blood. *Epigenetics*, 6(5):623-629
- Zhang, Y., Wang, B., Ye, S., Liu, S., Liu, M., Shen, C., Teng, Y. and Qi, J. 2010. Killer cell immunoglobulin-like receptor gene polymorphisms in patients with leukaemia: possible association with susceptibility to the disease. *Leukemia Research*, 34:55-58.