



**UNIVERSITI PUTRA MALAYSIA**

**PHYTOCHEMICAL AND GLUCOSE LOWERING POTENTIAL OF  
SEAWEED *Eucheuma denticulatum* (N.L. Burman) F.S. Collins & A.B.  
Hervey  
*IN VITRO***

**B.VIMALA A/P R.M.T. BALASUBRAMANIAM**

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By

**B.VIMALA A/P R.M.T. BALASUBRAMANIAM**

Thesis Submitted to the School of Graduate Studies,  
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**Doctor of Philosophy**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of  
the requirement for the degree of Doctor of Philosophy

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**September 2017**

**Chair : Professor Amin Ismail, PhD**  
**Faculty: Medicine and Health Sciences**

Type 2 Diabetes Mellitus (T2DM) represents a serious global epidemic of the 21<sup>st</sup> century as reported by World Health Organisation (WHO). T2DM causes its own direct adverse effects as well as predisposes patients to the development of other chronic metabolic diseases such cardiovascular complications which lead to premature mortality. Thus, continuous search for remedies with minimum side effects were paramount. Scores of studies have demonstrated the health benefits derived from eating seaweed which promotes seaweed as a nutritional foodstuff. Moreover, research is advancing into using marine macroalgae also known as seaweed for production of novel foods and nutraceuticals. Modulating digestion with natural compounds has been shown to be a fruitful approach to the treatment of diabetes. The objectives of this study were to assess Malaysian seaweed species for their potential to regulate postprandial hyperglycaemia which is a pivotal feature of T2DM. Three species of Malaysian edible seaweed (*Eucheuma denticulatum*, *Sargassum polycystum* and *Caulerpa lentillifera*) found in coastal area of Sabah were selected and subjected to evaluation of their anti-diabetic potential *in vitro* in terms of their inhibition towards digestive enzymes that involve in hydrolysis of dietary carbohydrates ( $\alpha$ -amylase and  $\alpha$ -glucosidase). The seaweed were further subjected to other analyses related to glucose lowering properties such as antioxidant capacity, anti-inflammatory, adipogenesis, lipase enzyme inhibition and glucose uptake activities. Initially, the seaweed were screened and characterized for the presence of natural functional bioactive compounds which can be related to its health benefits. Following that, dinitrosalicylic acid assay was adapted in microplate to assess the inhibition of  $\alpha$ -amylase activity while colorimetry method for  $\alpha$ -glucosidase inhibition assay. Antioxidant capacity was evaluated for their free radical-scavenging capacity using 1, 1-diphenyl-2-picrylhydrazyl (DPPH) assay and oxygen radical absorbance capacity (ORAC). The anti-inflammatory potential and cytotoxic effects of the seaweed samples were evaluated by the inhibitory activity of nitric oxide (NO), interleukin-6 (IL-6), interleukin-1 (IL-1), tumor necrosis factor-alpha (TNF- $\alpha$ ), and monocyte chemoattractant protein-1 (MCP-1) on interferon-gamma/ lipopolysaccharide (IFN- $\gamma$ /LPS) stimulated murine macrophage cell line (RAW 264.7) and adipocytes (3T3-L1) using Griess reaction, immunoassays and MTS assay. Adipogenesis and glucose uptake

in 3T3-L1 were measured using commercial kits while lipase assay was measured by turbidimetric method. HPTLC, UHPLC and LC-MS/MS methods were established and validated for the quantitative determination of the identified compounds. The ethanolic extracts of the three species of seaweed showed the presence of carotenoids and the most notable being fucoxanthin, lutein, zeaxanthin, astaxanthin, canthaxanthin,  $\beta$ -cryptoxanthin,  $\beta$ -carotene and fatty acids such as palmitoleic acid (PA), docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA). Fucoxanthin was the major carotenoid detected in the brown seaweed (*S. polycystum*) (2,740 mg/ 100 g DW). In the case of red seaweed (*E. denticulatum*), lutein (88 mg/ 100 g DW) and zeaxanthin (21 mg/ 100 g DW) were the major carotenoids, apart from small amount of fucoxanthin, while for the green seaweed (*C. lentillifera*),  $\beta$ -carotene (20 mg/ 100 g DW) and canthaxanthin (15 mg/ 100 g DW) were detected as major carotenoids. The method exhibited (a) linear calibration curves ( $R^2>0.97$ ), (b) satisfactory recoveries for most of the pigments (between 74 and 104%), and (d) low detection (from 0.001 to 0.01 ng/ $\mu$ l) and quantification limits (from 0.004 to 0.02 ng/ $\mu$ l) (LOD and LOQ, respectively). Ethanolic and methanolic extracts from the studied seaweed were found to display inhibitory effects against  $\alpha$ -amylase (11-67%; n=3), but have no effect on  $\alpha$ -glucosidase activity *in vitro*. Amongst the 3 genera, *E. denticulatum* ethanolic extract was found to be most effective *in vitro* inhibitors of  $\alpha$ -amylase with IC<sub>50</sub> of 0.14 mg/ml. Thus, this seaweed was selected for further solvent fractionation and bioactivity assays. Among the five investigated fractions (hexane, ethyl acetate, acetone, butanol and water), the hexane, ethyl acetate and acetone fractions exhibited good inhibition with a mean of 42%. The brown seaweed, *S. polycystum* displayed the highest DPPH radical scavenging and TPC (20%; 400 mg GAE/ 100 g respectively) whilst for ORAC analysis, *E. denticulatum* exhibited the highest activity at 112,762  $\mu$ mol TE/ 100 g. *E. denticulatum* ethanol extract and fractions (1–100  $\mu$ g/ml), also exhibited anti-inflammatory activity without showing any cytotoxic effect to RAW 264.7 cells. The crude and fractions seem to inhibit adipogenesis and enhances glucose uptake in the 3T3-L1 cell model while ethanolic extract showed the highest lipase enzyme inhibitory (83%) compared to other sample preparation. The presence of fatty acids such as PA and EPA, antioxidant compounds such as polyphenols and carotenoids may probably contributed to the glucose lowering efficacy of these seaweed. In conclusion, this study demonstrated that *E. denticulatum* was able to exert bioactive actions such as anti-diabetic, antioxidant, immune modulating and anti-obesity properties *in vitro* which suggesting its potential source as functional ingredient that can be further exploited to generate new valuable products for various commercial applications.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai  
memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN FITOKIMIA DAN POTENSI  
PENURUNAN GLUKOS OLEH RUMPAI LAUT  
*Eucheuma denticulatum* (N.L. Burman) F.S. Collins & A.B. Hervey SECARA *IN  
VITRO***

Oleh

**B.VIMALA A/P R.M.T. BALASUBRAMANIAM**

**September 2017**

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**Fakulti : Perubatan dan Sains Kesihatan**

Diabetes Melitus- Jenis 2 (T2DM) merupakan satu wabak global yang serius pada abad ke-21 menurut Pertubuhan Kesihatan Sedunia (WHO). T2DM bukan sahaja menyebabkan kesan-kesan buruk secara langsung tetapi juga mempengaruhi pesakit untuk mendapat penyakit kronik yang lain seperti penyakit jantung dan penyakit buah pinggang. Oleh itu, usaha berterusan untuk mencari penawar dengan kesan sampingan yang minimum adalah paling utama. Kajian telah menunjukkan manfaat kesihatan daripada pemakanan rumpai laut. Selain itu, penyelidikan rumpai laut telah maju dan semakin berkembang dalam bidang pengeluaran makanan baharu atau unik dan nutraceutical. Modulasi pencernaan makanan dengan sebatian semula jadi telah terbukti sebagai salah satu pendekatan yang bermanfaat untuk rawatan kencing manis. Objektif kajian ini adalah untuk mengumpul, menyaring dan menilai spesies rumpai laut Malaysia sebagai potensi untuk mengawal hiperglisemia atau peningkatan gula dimana ia merupakan ciri penting untuk mengawal T2DM. Tiga spesies rumpai laut (*Eucheuma denticulatum*, *Sargassum polycystum* dan *Caulerpa lentillifera*) yang terdapat di kawasan pesisiran pantai Sabah telah dipilih bagi penilaian potensi anti-diabetes mereka dalam kajian *in vitro* iaitu dari segi perencutan terhadap enzim pencernaan yang terlibat dalam hidrolisis karbohidrat ( $\alpha$ -amylase dan  $\alpha$ -glucosidase) dan seterusnya menilai kapasiti antioksidan, aktiviti anti-inflamatori, perencutan lemak di dalam sel, pengambilan glukosa oleh tisu dan perencutan enzim lemak lipase. Rumpai laut juga telah dianalisis untuk mengetahui kehadiran sebatian bioaktif berfungsi semula jadi yang boleh dikaitkan dengan manfaat kesihatan. Analisis asid dinitrosalicylic telah dijalankan untuk menilai aktiviti  $\alpha$ -amylase manakala kaedah kolorimetri untuk  $\alpha$ -glucosidase. Kapasiti antioksidan dinilai dengan menggunakan 1, 1-Diphenyl-2-pricrylhydrazyl (DPPH) assay dan analisis ORAC. Kesan-kesan anti-radang dan sifat keracunan oleh sampel rumpai laut telah dinilai dari segi penurunan biomarker seperti nitrik oksida (NO), interleukin-6 (IL-6), interleukin-1 (IL-1), tumor nekrosis faktor-alpha (TNF- $\alpha$ ), dan monocyte chemoattractant protein-1 (MCP-1) di dalam sel makrofaj (RAW 264,7) dan lemak (3T3-L1) yang telah di aktifkan oleh interferon gamma / lipopolysaccharide (IFN- $\gamma$ / LPS) menggunakan analisis Griess,

*immunoassays* dan kit MTS. Adipogenesis dan penyerapan glukosa dalam sel lemak, 3T3-L1 diukur menggunakan kit komersial manakala analisis perencutan enzim lemak lipase menggunakan kaedah turbidimetric. Kaedah HPTLC, UHPLC dan LC-MS/MS telah digunakan untuk penentuan kandungan sebatian atau komponen yang dikenal pasti di dalam ekstrak rumpai laut. Ekstrak ethanol ketiga-tiga rumpai laut yang di kaji mengandungi karotenoid seperti fucoxanthin, lutein, zeaxanthin, astaxanthin, canthaxanthin,  $\beta$ -cryptoxanthin,  $\beta$ -carotene dan asid lemak seperti asid palmitoleic (PA), asid docosahexaenoic (DHA) dan asid eicosapentaenoic (EPA). Fucoxanthin adalah karotenoid utama dikesan dalam rumpai laut *S. polycystum* (2,740 mg/ 100 g DW). Dalam kes rumpai laut merah (*E. denticulatum*), lutein (88 mg/ 100 g DW) dan zeaxanthin ( 21 mg/ 100 g DW) merupakan karotenoid utama, selain fucoxanthin, manakala bagi rumpai laut hijau,  $\beta$  -carotene (20 mg/ 100 g DW) dan canthaxanthin (15 mg/ 100 g DW) dikesan sebagai karotenoid utama. Kaedah yang digunakan mempamerkan (a) keluk penenturan linear ( $R^2 > 0.97$ ), (b) *recovery* yang memuaskan bagi kebanyakan pigmen (antara 74 dan 104%), dan (d) pengesanan rendah (0.001-0.01 ng/ $\mu$ l) dan had kuantifikasi (0.004-0.02 ng/ $\mu$ l) (LOD dan LOQ, masing-masing). Ekstrak ethanol dan metanol dari rumpai laut yang dikaji didapati memaparkan kesan pengurangan terhadap  $\alpha$ -amylase (11-67%; n=3), tetapi tidak menunjukkan kesan ke atas aktiviti  $\alpha$ -glucosidase secara *in vitro*. Di antara 3 jenis rumpai laut, ekstrak ethanol *E. denticulatum* didapati paling berkesan merentang  $\alpha$ -amylase dengan IC<sub>50</sub> 0.14 mg/ml. Oleh itu, rumpai laut ini telah dipilih untuk kajian selanjutnya. Antara lima *fraction* rumpai laut *E. denticulatum* (hexane, ethyl acetate, acetone, butanol dan air), hexane, ethyl acetate dan aceton menunjukkan kadar perencutan enzim yang baik dengan purata penurunan sebanyak 42%. *S. polycystum* menunjukkan aktiviti DPPH dan TPC yang tinggi (20%; 400 mg/ GAE 100 g masing-masing) manakala bagi analisis ORAC, *E. denticulatum* mencatatkan aktiviti tertinggi iaitu 112.762  $\mu$ mol TE / 100 g. Ekstrak etanol *E. denticulatum* dan *fractions* (1-100  $\mu$ g/ml), juga menunjukkan aktiviti anti-radang tanpa menunjukkan apa-apa kesan sitotoksik kepada sel-sel RAW 264.7. Ekstrak rumpai laut yang sama juga menurunkan aktiviti adipogenesis dan meningkatkan penyerapan glukosa dalam model sel 3T3-L1 manakala penurunan enzim lipase sebanyak 83%. Kehadiran asid lemak EPA dan PA, sebatian antioksidan seperti *polyphenol* dan karotenoid mungkin menyumbang kepada bioaktiviti keberkesanan penurunan glukosa oleh rumpai laut ini. Kesimpulannya, kajian ini menunjukkan bahawa rumpai laut yang dikaji menunjukkan ciri-ciri kesihatan seperti anti-diabetes, anti-obesiti, anti-oksida dan modulasi imun dalam kajian *in vitro*. Kesimpulan yang dapat dibuat daripada keputusan kajian ialah rumpai laut ini berpotensi untuk menjadi sumber makanan berfungsi yang boleh terus dibangunkan untuk aplikasi komersial.

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I certify that a Thesis Examination Committee has met on 11 September 2017 to conduct the final examination of B.Vimala a/p R.M.T Balasubramaniam on her thesis entitled "Phytochemical and Glucose Lowering Potential of Seaweed *Eucheuma denticulatum* (N.L.Burman) F.S.Collins and A.B.Harvey *In Vitro*" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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**5**

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## LIST OF ABBREVIATIONS

APCI	Atmospheric pressure chemical ionization
DMEM	Dulbecco's modified Eagle's medium
FBS	Fetal bovine serum
DMSO	Dimethyl sulfoxide
DPPH	2,2-diphenyl-1-picrylhydrazyl
ESI	Electrospray ionization
GAE	Gallic acid equivalents
GC-MS	Gas chromatography-mass spectrometry
HPTLC	High performance thin layer chromatography
LC	Liquid chromatography
LOD	Limit of detection
LOQ	Limit of quantitation
MS	Mass spectrometry
MS/MS	Tandem mass spectrometry
2-NBDG	2-deoxy-2-[(7-nitro-2, 1, 3-benzoxadiazol-4-yl) amino]-D-glucose
SD	Standard deviation
TIC	Total ion chromatogram
MCP-1	Monocyte chemoattractant-1
IL-6	Interleukin-6
IL-1 $\beta$	Interleukin -1 $\beta$
TAG	Triacylglycerol
T1DM	Type I Diabetes Mellitus
T2DM	Type II Diabetes Mellitus
TNF- $\alpha$	Tumour necrosis factor- $\alpha$



# CHAPTER 1

## INTRODUCTION

### 1.1 Background

Diabetes Mellitus (DM) is a metabolic disorder which occurs due to insulin resistance and impaired insulin secretion in the body. DM mainly falls into two broad etiopathogenetic categories; insulin dependent DM (type 1 diabetes) and non-insulin dependent DM (type 2 diabetes). Type 1 diabetes (T1DM) is generally treated through insulin replacement therapy while type 2 diabetes (T2DM) is treated with oral hypoglycemic agents (Rengasamy, Aderogba, Amoo, Stirk, & Van Staden, 2013).

T1DM is an auto-immune mediated impairment of pancreatic  $\beta$ -cells which lead to lack of insulin production while T2DM, is a metabolic disorder that is distinguished by a high level of glucose in the blood due to a defect in insulin production and/or action, or both (American Diabetes Association, 2014; Low, 2010). Improper glucose metabolism occurs during onset and development of T2DM which subsequently affects the lipid metabolism (Chang et al. 2002; Cheplick, Kwon, Bhowmik, & Shetty, 2010). Pathological changes like nephropathy, retinopathy and cardiovascular complications start occurring with the progression of disease in the body which led to increased morbidity and mortality (Rother, 2007).

Postprandial phase in diabetes is portrayed by a sudden increase in blood glucose levels, and evidence suggest that the postprandial “hyperglycemic spikes” may be linked to the pathophysiological conditions of late diabetes (Bonora & Muggeo, 2001; Ceriello, 2005). Therefore, the control of postprandial hyperglycemia is suggested to be paramount in the treatment of diabetes and its complications.

Diabetes mellitus (DM) is termed as the emerging pandemics in the 21st century and declared as a major health problem rising rapidly throughout the world (Maeda, Tsukui, Sashima, Hosokawa, & Miyashita, 2008) which is projected to reach 552 million people by 2030 (Whiting, Guariguata, Weil, & Shaw, 2011). T2DM previously referred as “non–insulin-dependent diabetes” or “adult-onset diabetes,” accounts for 90–95% of all diabetes (American Association Diabetes, 2016) and is responsible for 85–95% of the cases globally (Zimmet, 1999) with Asian countries contribute to more than 60% of the world’s diabetic population (Ramachandran, 2012). As a matter of fact, for Malaysia alone an estimated 3.5 million adults (18 years and above) or 17.5% are living with diabetes (NHMS, 2015). Socio-economic growth and industrialisation are rapidly occurring in many of these countries. Increased rates of urbanization, modernization, readily available fast foods and sedentary habits have altered the lifestyle of the population, mainly among the youth were the main causes of prevalence of T2DM (Ramachandran, 2012).

In diabetes, oral hypoglycemic and anti-hyperglycemic drugs are administered to maintain blood glucose level near the normal range. However, the DM drugs have limitation such as undesirable side effects like weight gain, stomach discomfort and vomiting (Egan, Bulotta, Hui, & Perfetti, 2003), thus more researches are diverting to the use of natural products as an alternative measure for management of T2DM. Medicinal herbs as a substitute and complementary medicine have been long practised due to its rich source of functional metabolites which ascribed as DM remedies. Rios, Francini, & Schinella (2015) reviewed the use of natural products as a source of therapeutic agent for the treatment of Type 2 Diabetes Mellitus. More than 1200 species of organisms have been reported for the use to treat symptoms of DM. They represent more than 725 genera in 183 families, ranging from seaweed and fungi to advanced plants. *Galega officinalis* L. (Fabaceae) is the first medicinal plant reported with a definite anti-diabetic effect (Marles & Farnsworth, 1995).

Edible seaweed products have been used in many Asian countries as a food item since ancient times and currently considered with high economic potential in food and pharmaceutical industry as well as human health (Dhargalkar & Pereira, 2005). In general, seaweed-derived compounds have shown to contribute to vast biological activities including anti-diabetes (Chin et al., 2014; Sharifuddin, Chin, Lim, & Phang, 2015). Furthermore, an increasing number of scientific papers published correlating seaweed and health benefits from diet replacements or extracts (Holdt & Kraan, 2011) and these activities may be attributed to antioxidants, polyunsaturated fatty acids (PUFA), pigments or the unique mineral contents present in the seaweed (Bocanegra et al. 2009; Ortega-Calvo et al. 1993; Rupérez 2001). Since, seaweed exhibited promising source of natural agent with potential biological effects, thus more efforts were taken to isolate the bioactive compounds and explore its action mechanisms (Holdt & Kraan, 2011; Heo et al., 2009). Seaweed are the largest and most complex marine source with unique bioactive ingredients. The seaweed are characterized as photosynthetic like plants that occupy a wide range of ecological niches. They are important living resources of the world oceans which form a basic biomass in the intertidal zone and contribute ecologically and economically. The seaweed are categorised as two major types; microalgae which are found in both benthic and littoral habitats and also throughout the ocean waters as phytoplankton and the macroalgae or seaweed which occupy the littoral zone area up to a certain depth where very little photosynthetic light is available (Braune and Guiry, 2011).

Seaweed belong to three different groups, distinguished on the basis of thallus colour: brown seaweed (phylum Ochrophyta; Class Phaeophyceae), red seaweed (phylum Rhodophyta) and green seaweed (phylum Chlorophyta; Classes Bryopsidophyceae, Chlorophyceae, Dasycladophyceae, Prasinophyceae and Ulvophyceae) (Braune and Guiry, 2011).

Seaweed consist of key ingredients such as fibres, proteins, minerals, vitamins, antioxidants, phytochemicals, and polyunsaturated fatty acids with low caloric values. However, their nutrient composition are affected by external factors such as the salinity, geographic location, environmental, harvesting season, sampling methods (Rohani-Ghadikolaei, Abdulalian, & Ng, 2012; Dawes, Kovach, & Friedlander, 1993). High essential nutrients and phytochemical composition complimented with high rate

growths of seaweed make it as a sustainable functional ingredient for complementary and alternative therapy.

## **1.2 Problem statements**

Seaweed industry has been identified by the government as a high potential sector which could contribute to the country's economy and therefore could increase the income of Malaysians, especially at the coastal areas (Lunkapis & Danny, 2016; Galid, 2003). This industry has commercial values based on its use as raw material for pharmaceutical, cosmeceutical and nutraceuticals product development. Currently, the seaweed researches in Malaysia mostly focus on food and non-food based product development (Phang, 2010). However, this potential marine source has remained not investigated or dearth in research especially for its possible glucose lowering properties. On the other hand, T2DM prevalence in Malaysia is at an alarming rate (17.5%) and expected to increase further (NHMS, 2015). Due to limited number of anti-diabetic drugs as well as high cost of medication with potentially hazardous side effects, there is a need and high demand for a therapeutically potent, yet safe, anti-hyperglycemia agent derived from natural products, such as marine macroalgae (seaweed). Thus, finding effective ways to treat or prevent these problems is paramount.

## **1.3 Significance of the study**

The increasing number of scientific evidences in the last two decades relating diet and health has substantiate the remarkable potential of foods to promote and enhance health. The findings correlate the intake of certain food and their modulation of chronic diseases (Willett, Koplan, Nugent, Puska, & Gaziano, 2006; Simopoulos, 2008). As a result, there is now a huge interest on products that can promote health and well-being among consumers and the food industry. Interestingly, the marine world represents a largely untapped reserve of bioactive ingredients and as reported in numerous studies of late, *Eucheuma* species have been commercially introduced throughout mostly tropical parts of the world for cultivation, serving as a means of livelihood for the locals, and as an additional source of revenue for the economy of the country. Many recent studies have associated seaweed with anti-diabetic properties (Sharifuddin et al., 2015; Chin et al., 2014; Motshakeri, Ebrahimi, Goh, Matanjun & Mohamed, 2013). Hence, this study will further enhance the scientific information and knowledge on the health-promoting properties of our local seaweed especially as anti-hyperglycemia agent which will further strengthen the claims of seaweed as a healthy food product as well as uplifting our local seaweed industry.

## **1.4 Objectives**

### **1.4.1 General objective**

To evaluate phytochemical content and glucose lowering potential of selected Malaysian seaweed.

#### **1.4.2 Specific objectives**

- To identify and characterize the bioactive components present in the studied seaweed
- To evaluate the regulatory effects of seaweed extracts on the starch digestive enzymes ( $\alpha$ -amylase and  $\alpha$ -glucosidase) and antioxidant capacity of the seaweed
- To determine the effect of the seaweed on the inflammatory biomarkers associated with hyperglycemia using RAW 264.7 and 3T3-L1 cell line models
- To determine the effect of the seaweed extract(s) / fraction(s) on the lipase enzymes, adipogenesis activity and glucose uptake in 3T3-L1 cell line

## REFERENCES

- Agardh, C.A. (1820). The well-known species of algae: with synonyms, diagnoses and succinct descriptions of the species. Lund, Berling
- Abidov, M., Ramazanov, Z., Seifulla, R., & Grachev, S. (2010). The effects of Xanthigen™ in the weight management of obese premenopausal women with non-alcoholic fatty liver disease and normal liver fat. *Diabetes, Obesity and Metabolism*, 12(1), 72–81
- Ahmad, F., Sulaiman, M. R., Saimon, W., Yee, C. F., & Matanjun, P. (2012). Proximate Compositions and Total Phenolic Contents of Selected Edible Seaweed From Semporna, Sabah , Malaysia. *Borneo Science*, 31, 74–83
- Ahn, G. N., Kim, K. N., Cha, S. H., Song, C. B., Lee, J., Heo, M. S., Yeo, I.K., Lee, N.H., Jee, Y.H., Kim, J.S., Heu, M.S., & Jeon, Y. J. (2007). Antioxidant activities of phlorotannins purified from Ecklonia cava on free radical scavenging using ESR and H<sub>2</sub>O<sub>2</sub>-mediated DNA damage. *European Food Research and Technology*, 226(1-2), 71–79
- Ahn, M.J., Yoon, K.D., Min, S.Y., Lee, J. S., Kim, J. H., Kim, T. G., Kim, S.H., Kim, N.G., Huh, H., & Kim, J. (2004). Inhibition of HIV-1 reverse transcriptase and protease by phlorotannins from the brown alga Ecklonia cava. *Biological & Pharmaceutical Bulletin*, 27(4), 544–547
- Aihara, M.S. & Yamamoto, H.Y. (1968). Occurrence of antheraxanthin in two Rhodophyceae Acanthophora spicifera and Gracilaria lichenoides. *Phytochemistry*, 7, 497 – 499
- Alam, M. N., Bristi, N. J., & Rafiquzzaman, M. (2013). Review on in vivo and in vitro methods evaluation of antioxidant activity. *Saudi Pharmaceutical Journal*, 21(2), 143–152
- Alexandraki, K., Piperi, C., Kalofoutis, C., Singh, J., Alaveras, A., & Kalofoutis, A. (2006). Inflammatory process in type 2 diabetes: The role of cytokines. *Annals of the New York Academy of Sciences*, 1084, 89–117
- Ali, H., Houghton, P. J., & Soumyanath, A. (2006).  $\alpha$ -Amylase inhibitory activity of some Malaysian plants used to treat diabetes; with particular reference to Phyllanthus amarus. *Journal of Ethnopharmacology*, 107(3), 449–455
- Allen, N. E., Sauvaget, C., Roddam, A. W., Appleby, P., Nagano, J., Suzuki, G., Key, T.J., Koyama, K. (2004). A prospective study of diet and prostate cancer in Japanese men. *Cancer Causes & Control*, 15(9), 911–920
- Amano, S. U., Cohen, J. L., Vangala, P., Tencerova, M., Nicoloro, S. M., Yawe, J. C., Shen, Y., Czech, M.P., & Aouadi, M. (2014). Local proliferation of macrophages contributes to obesity-associated adipose tissue inflammation. *Cell Metabolism*, 19(1), 162–171
- American Association Diabetes (2016). Classification and Diagnosis of Diabetes.

*Diabetes Care*, 39(Supplement 1), S13–S22

American Diabetes Association. (2014). *Diabetes Care*, 37 (Suppl.(January), 81–90

American Diabetes Association (2006). Standards of Medical Care in Diabetes. *Diabetes Care*, 29 (Supplement 1), S4-S42

American Diabetes Association (2004). Standards of Medical Care in Diabetes. *Diabetes Care*, 27, (Supplement 1), S15-S35

Andersson, M., Schubert, H., Pedersén, M., & Snoeijs, P. (2006). Different patterns of carotenoid composition and photosynthesis acclimation in two tropical red algae. *Marine Biology*, 149(3), 653–665

Aneiros, A., & Garateix, A. (2004). Bioactive peptides from marine sources: pharmacological properties and isolation procedures. *Journal of Chromatography. B, Analytical Technologies in the Biomedical and Life Sciences*, 803(1), 41–53

Apostolidis, E., & Lee, C. M. (2010). *In vitro* potential of *ascophyllum nodosum* phenolic antioxidant-mediated  $\alpha$ -glucosidase and  $\alpha$ -amylase inhibition. *Journal of Food Science*, 75(3)

Apostolidis, E., Li, L., Lee, C., & Seeram, N. P. (2011). *In vitro* evaluation of phenolic-enriched maple syrup extracts for inhibition of carbohydrate hydrolyzing enzymes relevant to type 2 diabetes management. *Journal of Functional Foods*, 3(2), 100–106

Arasaki, S., & Arasaki, T.(1983). Low Calorie, High Nutrition Vegetables from the Sea to Help You Look and Feel Better (Vol. 60). Japan Publications, Tokyo

Arnold, C. E., Whyte, C. S., Gordon, P., Barker, R. N., Rees, A. J., & Wilson, H. M. (2014). A critical role for suppressor of cytokine signalling 3 in promoting M1 macrophage activation and function *in vitro* and *in vivo*. *Immunology*, 141(1), 96–110

Ashikin, N. (2014). Antioxidant and  $\alpha$ -glucosidase inhibitory activities of the leaf and stem of selected traditional medicinal plants. *International Food Research Journal*, 21(1), 165–172

Ashok, K. B., Lakshman, K., Jayaveera, K., Sheshadri Shekar, D., Narayan Swamy, V. B., Khan, S., & Velumurga, C. (2011). In vitro  $\alpha$ -amylase inhibition and antioxidant activities of methanolic extract of *Amaranthus caudatus* Linn. *Oman Medical Journal*, 26(3), 166–170

Attie, A. D., & Scherer, P. E. (2009). Adipocyte metabolism and obesity. *Journal of Lipid Research*, 50, S395–399

Awang, A. N., Ng, J. L., Matanjun, P., Sulaiman, M. R., Tan, T. S., & Ooi, Y. B. H. (2013). Anti-obesity property of the brown seaweed, *Sargassum polycystum* using an *in vivo* animal model. *Journal of Applied Phycology*, 26(2),1043-1048

- Balasubramaniam, V., Lee, J. C., Noh, M. F. M., Ahmad, S., Brownlee, I. A., & Ismail, A. (2015). Alpha-amylase, antioxidant, and anti-inflammatory activities of Eucheuma denticulatum (N.L. Burman) F.S. Collins and Hervey. *Journal of Applied Phycology*, 28(3), 1965–1974
- Balasubramaniam, V., Mustar, S., Mustafa Khalid, N., Abd Rashed, A., Mohd Noh, M. F., Wilcox, M. D., Chater, P. I., Brownlee, I. A., & Pearson, J. P. (2013). Inhibitory activities of three Malaysian edible seaweeds on lipase and  $\alpha$ -amylase. *Journal of Applied Phycology*, 25(5), 1405–1412
- Balboa, E. M., Conde, E., Moure, A., Falqué, E., & Domínguez, H. (2013). *In vitro* antioxidant properties of crude extracts and compounds from brown algae. *Food Chemistry*, 138(2-3), 1764–1785
- Ballinger, A., & Peikin, S. R. (2002). Orlistat: its current status as an anti-obesity drug. *European Journal of Pharmacology*, 440(2-3), 109–117
- Bao, J. X., Li, K. H., Yi, N. Z., Jeong, H. L., & Chang, K. S. (2005). *In vitro* inhibitory effect of triterpenoidal saponins from platycodi radix on pancreatic lipase. *Archives of Pharmacal Research*, 28(2), 180–185
- Betteridge, D. J. (1989). Diabetes, lipoprotein metabolism and atherosclerosis. *British Medical Bulletin*, 45 (1), 285–311
- Bhandari, M. R., Jong-Anurakkun, N., Hong, G., & Kawabata, J. (2008).  $\alpha$ -Glucosidase and  $\alpha$ -amylase inhibitory activities of Nepalese medicinal herb Pakhanbhed (*Bergenia ciliata*, Haw.). *Food Chemistry*, 106(1), 247–252
- Bijtebier, S., D'Hondt, E., Noten, B., Hermans, N., Apers, S., & Voorspoels, S. (2014). Ultra high performance liquid chromatography versus high performance liquid chromatography: Stationary phase selectivity for generic carotenoid screening. *Journal of Chromatography A*, 1332, 46–56
- Birari, R. B., & Bhutani, K. K. (2007). Pancreatic lipase inhibitors from natural sources: unexplored potential. *Drug Discovery Today*, 12(19-20), 879–889
- Bitou, N., Ninomiya, M., Tsujita, T., & Okuda, H. (1999). Screening of lipase inhibitors from marine algae. *Lipids*, 34(5), 441–445
- Bixler, H. J., & Porse, H. (2011). A decade of change in the seaweed hydrocolloids industry. *Journal of Applied Phycology*, 23(3), 321–335
- Bjørnland, T. (1983). Chlorophyll and carotenoids of five isolates of the red alga *Antithamnion plumula*. *Biochemical Systematics and Ecology*, 11(2), 73–76
- Block, M. L., Zecca, L., & Hong, J.S. (2007). Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nature Reviews Neuroscience*, 8(1), 57–69
- Blois, M. S. (1958). Antioxidant determination by the use of a stable free radical. *Nature*, 181, 1199–1200

- Bocanegra, A., Bastida, S., Benedi, J., Rodenas, S., & Sanchez-Muniz, F. J. (2009). Characteristics and nutritional and cardiovascular-health properties of seaweeds. *Journal of Medicinal Food*, 12(2), 236–258
- Bonora, E., & Muggeo, M. (2001). Postprandial blood glucose as a risk factor for cardiovascular disease in Type II diabetes: the epidemiological evidence. *Diabetologia*, 44(12), 2107–2114
- Borchardt, S. A., Allain, E. J., Michels, J. J., Stearns, G. W., Kelly, R. F., & McCoy, W. F. (2001). Reaction of acylated homoserine lactone bacterial signaling molecules with oxidized halogen antimicrobials. *Applied and Environmental Microbiology*, 67(7), 3174–3179
- Borges de Melo, E., da Silveira Gomes, A., & Carvalho, I. (2006).  $\alpha$ - and  $\beta$ -Glucosidase inhibitors: chemical structure and biological activity. *Tetrahedron*, 62(44), 10277–10302
- Braune, W. & Guiry, M.D. (2011). Seaweeds. A colour guide to common benthic green, brown and red algae of the world's oceans. A.R.G. Gantner, Ruggell.
- Brereton, R. G. (2007). *Applied chemometrics for scientists*. John Wiley & Sons
- Brown, E. M., Allsopp, P. J., Magee, P. J., Gill, C. I., Nitecki, S., Strain, C. R., & McSorley, E. M. (2014). Seaweed and human health. *Nutrition Reviews*, 72(3), 205–216
- Brownlee, M. (2005). The pathobiology of diabetic complications. *Diabetes*, 54(6), 1615
- Bruun, J. M., Lihn, A. S., Pedersen, S. B., & Richelsen, B. (2005). Monocyte chemoattractant protein-1 release is higher in visceral than subcutaneous human adipose tissue (AT): implication of macrophages resident in the AT. *The Journal of Clinical Endocrinology and Metabolism*, 90(4), 2282–2289
- Burtin, P. (2003). Nutritional Value of Seaweeds. *Quality*, 2(4), 498–503
- Bustanji, Y., Mohammad, M., Hudaib, M., Tawaha, K., Al-Masri, I. M., AlKhatib, H. S., Issa, A., & Alali, F. Q. (2011). Screening of some medicinal plants for their pancreatic lipase inhibitory potential. *Jordan Journal of Pharmaceutical Sciences*, 4(2), 81–88
- Cann, S. A., van Netten, J. P., & van Netten, C. (2000). Hypothesis: iodine, selenium and the development of breast cancer. *Cancer Causes & Control*, 11(2), 121–127
- Cao, G., & Prior, R. (1998). Comparison of different analytical methods for assessing total antioxidant capacity of human serum. *Clinical Chemistry*, 1315, 1309–1315
- Careri, M., Elviri, L., & Mangia, A. (1999). Liquid chromatography-electrospray mass spectrometry of  $\beta$ -carotene and xanthophylls: Validation of the analytical method. *Journal of Chromatography. A*, 854(1-2), 233–244
- Cavalcante-Silva, L. H. A., Da Matta, C. B. B., De Araújo, M. V., Barbosa-Filho, J.

- M., De Lira, D. P., De Oliveira Santos, B. V., de Miranda, G.E.C., & Alexandre-Moreira, M. S. (2012). Antinociceptive and anti-inflammatory activities of crude methanolic extract of red alga *Bryothamnion triquetrum*. *Marine Drugs*, 10(9), 1977–1992
- Cavaliere, H., Floriano, I., & Medeiros-Neto, G. (2001). Gastrointestinal side effects of orlistat may be prevented by concomitant prescription of natural fibers (psyllium mucilloid). *International Journal of Obesity and Related Metabolic Disorders*, 25(7), 1095–1099
- Cengiz, S., Cavas, L., & Yurdakoc, K. (2010). Alpha-amylase inhibition kinetics by caulerpenyne. *Mediterranean Marine Science*, 11, 93–103
- Ceriello, A. (2005). Postprandial hyperglycemia and diabetes complications. *Diabetes*, 54, 1–7
- Chan, C.X., Ho, C.L., & Phang, S.M. (2006). Trends in seaweed research. *Trends in Plant Science*, 11, 165–166.
- Chang, H.B., Kim, S.H., Kwon, Y.I., Choung, D.H., Choi, W.K., Kang, T. W., Lee, S., Kim, J.G., Chun, H.S., Ahn, S.K., Hong, C.I., & Han, K.H. (2002). Novel  $\alpha$ -Glucosidase Inhibitors, CKD-711 and CKD-711a Produced by Streptomyces sp. CK-4416. *The Journal of Antibiotics*, 55(5), 467–471
- Chater, P. I., Wilcox, M., Cherry, P., Herford, A., Mustar, S., Wheater, H., Brownlee, I., Seal, C., & Pearson, J. (2015). Inhibitory activity of extracts of Hebridean brown seaweeds on lipase activity. *Journal of Applied Phycology*, 28(2), 1303–1313
- Chater, P. I., Wilcox, M. D., Houghton, D., & Pearson, J. P. (2015). The role of seaweed bioactives in the control of digestion: implications for obesity treatments. *Food Function*, 6(11), 3420–3427
- Chauveau-Duriot, B., Doreau, M., Noziere, P., & Graulet, B. (2010). Simultaneous quantification of carotenoids, retinol, and tocopherols in forages, bovine plasma, and milk: validation of a novel UPLC method. *Analytical and Bioanalytical Chemistry*, 397(2), 777–790
- Chee, S. Y., Wong, P. K., & Wong, C. L. (2011). Extraction and characterisation of alginate from brown seaweeds (Fucales, Phaeophyceae) collected from Port Dickson, Peninsular Malaysia. *Journal of Applied Phycology*, 23(2), 191–196
- Chen, K.J., Tseng, C.K., Chang, F.R., Yang, J.I., Yeh, C.C., Chen, W.C., Wu, S.F., Chang, H.W., & Lee, J.C. (2013). Aqueous extract of the edible *Gracilaria tenuistipitata* inhibits hepatitis C viral replication via cyclooxygenase-2 suppression and reduces virus-induced inflammation. *PloS One*, 8(2), e57704
- Chen, H. (2006). Cellular inflammatory responses: novel insights for obesity and insulin resistance. *Pharmacological Research*, 53(6), 469–477
- Chensusue, S. W., Warmington, K. S., Ruth, J. H., Sanghi, P. S., Lincoln, P., & Kunkel, S. L. (1996). Role of monocyte chemoattractant protein-1 (MCP-1) in Th1

- (mycobacterial) and Th2 (schistosomal) antigen-induced granuloma formation: relationship to local inflammation, Th cell expression, and IL-12 production. *The Journal of Immunology*, 157(10), 4602–4608
- Cheplick, S., Kwon, Y. I., Bhowmik, P., & Shetty, K. (2010). Phenolic-linked variation in strawberry cultivars for potential dietary management of hyperglycemia and related complications of hypertension. *Bioresource Technology*, 101(1), 404–413
- Chew, Y. L., Lim, Y. Y., Omar, M., & Khoo, K. S. (2008). Antioxidant activity of three edible seaweeds from two areas in South East Asia. *LWT - Food Science and Technology*, 41(6), 1067–1072
- Chin, Y. X., Lim, P. E., Maggs, C. A., Phang, S. M., Sharifuddin, Y., & Green, B. D. (2014). Anti-diabetic potential of selected Malaysian seaweeds. *Journal of Applied Phycology*, 27(5), 2137–2148
- Christie, W.W., (1989). Preparation of methyl ester and other derivatives. In: Christie, W.W. (Ed.), *Gas Chromatography and Lipids. A Practical Guide*, 1st ed., Oily Press: Glasgow, UK, pp. 36–47
- Christensen M.B. (2016). Glucose-dependent insulinotropic polypeptide: effects on insulin and glucagon secretion in humans. *Danish Medical Journal*, 63(4), B5230
- Chung, H.J., Lee, H.S., Shin, J.S., Lee, S.H., Park, B.M., Youn, Y.S., & Lee, S. K. (2010). Modulation of acute and chronic inflammatory processes by a traditional medicine preparation GCSB-5 both in vitro and in vivo animal models. *Journal of Ethnopharmacology*, 130(3), 450–9
- Clissold, S. P., & Edwards, C. (2012). Acarbose. *Drugs*, 35(3), 214–243
- Colombo, M. L., Risè, P., Giavarini, F., De Angelis, L., Galli, C., & Bolis, C. L. (2006). Marine Macroalgae as Sources of Polyunsaturated Fatty Acids. *Plant Foods for Human Nutrition*, 61(2), 64–69
- Connors, K. A. (1974). Use of multiple Rf values for identification by paper and thin-layer chromatography. *Analytical Chemistry*, 46(1), 53–58
- Cooper, R., Dragar, C., Elliot, K., Fitton, J. H., Godwin, J., & Thompson, K. (2002). GFS, a preparation of Tasmanian Undaria pinnatifida is associated with healing and inhibition of reactivation of Herpes. *BMC Complementary and Alternative Medicine*, 2, 11
- Coura, C. O., Souza, R. B., Rodrigues, J. A. G., Vanderlei, E. de S. O., de Araújo, I. W. F., Ribeiro, N. A. et al. (2015). Mechanisms Involved in the Anti-Inflammatory Action of a Polysulfated Fraction from Gracilaria cornea in Rats. *PloS One*, 10(3), 1–18
- Cowherd, R. M., Lyle, R. E., & McGehee, R. E. J. (1999). Molecular regulation of adipocyte differentiation. *Seminars in Cell & Developmental Biology*, 10(1), 3–10

- Cullberg, K. B., Larsen, J. Ø., Pedersen, S. B., & Richelsen, B. (2014). Effects of LPS and dietary free fatty acids on MCP-1 in 3T3-L1 adipocytes and macrophages in vitro. *Nutrition & Diabetes*, 4, e113
- Dandona, P., Aljada, A., & Bandyopadhyay, A. (2004). Inflammation: The link between insulin resistance, obesity and diabetes. *Trends in Immunology*, 25(1), 4–7
- Dawes, C., Kovach, C. & Friedlander, M. (1993). Exposure of Gracilaria to Various Environmental Conditions. II. The Effect on Fatty Acid Composition. *Botanica Marina*. 36(4):289-296
- Dayer, J. M. (2003). The pivotal role of interleukin-1 in the clinical manifestations of rheumatoid arthritis. *Rheumatology (Oxford, England)*, 42 (Supplement 2), ii3–ii10
- de Ruiter, G. A., & Rudolph, B. (1997). Carrageenan biotechnology. *Trends in Food Science and Technology*, 8(12), 389–395
- de Sales, P. M., de Souza, P. M., Simeoni, L. A., Magalhães, P. D. O., & Silveira, D. (2012).  $\alpha$ -amylase inhibitors: A review of raw material and isolated compounds from plant source. *Journal of Pharmacy and Pharmaceutical Sciences*, 15(1), 141–183
- Delgado-Lista, J., Perez-Martinez, P., Lopez-Miranda, J., & Perez-Jimenez, F. (2012). Long chain omega-3 fatty acids and cardiovascular disease: a systematic review. *The British Journal of Nutrition*, 107, S201–213
- Dhargalkar, V. K., & Pereira, N. (2005). Seaweed : Promising Plant of the Millennium. *Source*, 71(Table 2), 60–66
- Di Carli, M. F., Janisse, J., Grunberger, G., & Ager, J. (2003). Role of chronic hyperglycemia in the pathogenesis of coronary microvascular dysfunction in diabetes. *Journal of the American College of Cardiology*, 41(8), 1387–1393
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of Limnophila aromatica. *Journal of Food and Drug Analysis*, 22(3), 296–302
- Dong, H. Q., Li, M., Zhu, F., Liu, F. L., & Huang, J. B. (2012). Inhibitory potential of trilobatin from Lithocarpus polystachyus Rehd against  $\alpha$ -glucosidase and  $\alpha$ -amylase linked to type 2 diabetes. *Food Chemistry*, 130(2), 261–266
- Dudonné, S., Vitrac, X., Coutière, P., Woillez, M., & Mérillon, J.M. (2009). Comparative Study of Antioxidant Properties and Total Phenolic Content of 30 Plant Extracts of Industrial Interest Using DPPH, ABTS, FRAP, SOD, and ORAC Assays. *Journal of Agricultural and Food Chemistry*, 57(5), 1768–1774
- Egan, J. M., Bulotta, A., Hui, H., & Perfetti, R. (2003). GLP-1 receptor agonists are growth and differentiation factors for pancreatic islet beta cells. *Diabetes/Metabolism Research and Reviews*, 19(2), 115–123

- Elisia, I., Hu, C., Popovich, D. G., & Kitts, D. D. (2006). Antioxidant assessment of an anthocyanin-enriched blackberry extract. *Food Chemistry*, 101(3), 1052–1058
- Ermakova, S., Sokolova, R., Kim, S. M., Um, B. H., Isakov, V., & Zvyagintseva, T. (2011). Fucoidans from brown seaweeds *Sargassum horneri*, *Ectonia cava*, *Costaria costata*: Structural characteristics and anticancer activity. *Applied Biochemistry and Biotechnology*, 164(6), 841–850
- Eskin, B. A., Grotkowski, C. E., Connolly, C. P., & Ghent, W. R. (1995). Different tissue responses for iodine and iodide in rat thyroid and mammary glands. *Biological Trace Element Research*, 49(1), 9–19
- Emenheiser, C., Sander, L. C., & Schwartz, S. J. (1995). Capability of a polymeric {C30} stationary phase to resolve cis-trans carotenoid isomers in reversed-phase liquid chromatography. *Journal of Chromatography A*, 707(2), 205–216
- Etxeberria, U., de la Garza, A. L., Campión, J., Martínez, J. A., & Milagro, F. I. (2012). Anti-diabetic effects of natural plant extracts via inhibition of carbohydrate hydrolysis enzymes with emphasis on pancreatic alpha amylase. *Expert Opinion on Therapeutic Targets*, 16(3), 269–297
- FAO, (2009). [www.fao.org](http://www.fao.org); searched on 10 March 2016.
- Fasshauer, M., Klein, J., Lossner, U., & Paschke, R. (2003). Interleukin (IL)-6 mRNA expression is stimulated by insulin, isoproterenol, tumour necrosis factor alpha, growth hormone, and IL-6 in 3T3-L1 adipocytes. *Hormone and Metabolic Research*, 35(3), 147–152
- Fasshauer, M., Kralisch, S., Klier, M., Lossner, U., Bluher, M., Klein, J., & Paschke, R. (2003). Adiponectin gene expression and secretion is inhibited by interleukin-6 in 3T3-L1 adipocytes. *Biochemical and Biophysical Research Communications*, 301(4), 1045–1050
- Fayaz, M., Namitha, K. K., Murthy, K. N. C., Swamy, M. M., Sarada, R., Khanam, S., Subbarao, P.V., & Ravishankar, G. A. (2005). Chemical composition, iron bioavailability, and antioxidant activity of *Kappaphycus alvarezii* (Doty). *Journal of Agricultural and Food Chemistry*, 53(3), 792–797
- Fedor, D., & Kelley, D. S. (2009). Prevention of insulin resistance by n-3 polyunsaturated fatty acids. *Current Opinion in Clinical Nutrition and Metabolic Care*, 12(2), 138–146
- Felber, J.P., Ferrannini, E., Golay, A., Meyer, H. U., Theibaud, D., Curchod, B., Maeder, E., Jequier, E., & DeFronzo, R. A. (1987). Role of Lipid Oxidation in Pathogenesis of Insulin Resistance of Obesity and Type II Diabetes. *Diabetes*, 36(11), 1341–1350
- Frankel, E. N., & Meyer, A. S. (2000). The problems of using one-dimensional methods to evaluate multifunctional food and biological antioxidants. *Journal of the Science of Food and Agriculture*, 80(13), 1925–1941

- Frestedt, J. L., Kuskowski, M. A., & Zenk, J. L. (2009). A natural seaweed derived mineral supplement (Aquamin F) for knee osteoarthritis: A randomised, placebo controlled pilot study. *Nutrition Journal*, 8(1), 7
- Frestedt, J. L., Walsh, M., Kuskowski, M. A., & Zenk, J. L. (2008). A natural mineral supplement provides relief from knee osteoarthritis symptoms: a randomized controlled pilot trial. *Nutrition Journal*, 7, 9
- Friedman, J. (2002). Diabetes: Fat in all the wrong places. *Nature*, 415(6869), 268–269
- Galid, R.S., (2003). Investment Opportunities in the Aquaculture Industry in sabah, malaysia. Paper presented at the 7<sup>th</sup> BIMPEAGA Working Group on Fisheries Cooperation Meeting. 10-12 June 2003. Palawan, Philippines
- Galland-Irmouli, A. V., Pons, L., Lucon, M., Villaume, C., Mrabet, N. T., Gueant, J. L., & Fleurence, J. (2000). One-step purification of R-phycerythrin from the red macroalga *Palmaria palmata* using preparative polyacrylamide gel electrophoresis. *Journal of Chromatography. B, Biomedical Sciences and Applications*, 739(1), 117–123
- Ganesan, P., Kumar, C. S., & Bhaskar, N. (2008). Antioxidant properties of methanol extract and its solvent fractions obtained from selected Indian red seaweeds. *Bioresource Technology*, 99(8), 2717–2723
- Garside, C., & Riley, J. P. (1968). The absorptivity of fucoxanthin. *Deep Sea Research and Oceanographic Abstracts*, 15(5), 627
- Gavino, C., & Trono, J. (1992). Eucheuma and Kappaphycus: taxonomy and cultivation. *Bulletin of Marine Science Fishing*, 12(12), 51–65
- Geronikaki, A. A., & Gavalas, A. M. (2006). Antioxidants and inflammatory disease: synthetic and natural antioxidants with anti-inflammatory activity. *Combinatorial Chemistry & High Throughput Screening*, 9(6), 425–442
- Gerwick, W.H. & Bernart, M.W. (1993). Eicosanoids and related com-pounds from marine algae. In Attaway DH, Zaborsky OR (eds),Marine Biotechnology, Vol.1, Pharmaceutical and Bioactive Natural Products, Plenum Press, NY, pp. 101–152
- Giorgino, F., Leonardini, A., Laviola, L., Perrini, S., & Natalicchio, A. (2009). Cross-Talk between PPAR $\gamma$  and insulin signaling and modulation of Insulin Sensitivity. *PPAR Research 2009*, 818945
- Giri, L., Andola, H. C., Kant Purohit, V., Rawat, M. S. M., Rawal, R. S., & Bhatt, I. D. (2010). Chromatographic and spectral fingerprinting standardization of traditional medicines: An overview as modern tools. *Research Journal of Phytochemistry*, 4(4), 234–241
- Go, H., Hwang, H.J., & Nam, T.J. (2010). A glycoprotein from *Laminaria japonica* induces apoptosis in HT-29 colon cancer cells. *Toxicology in vitro : An International Journal Published in Association with BIBRA*, 24(6), 1546–1553
- Godbout, A., & Chiasson, J.L. (2007). Who should benefit from the use of alpha-

- glucosidase inhibitors? *Current Diabetes Reports*, 7(5), 333–9
- Gondoin, A., Grussu, D., Stewart, D., & McDougall, G. J. (2010). White and green tea polyphenols inhibit pancreatic lipase in vitro. *Food Research International*, 43(5), 1537–1544
- Goodwin, T.W. (Ed.), 1980. The Biochemistry of Carotenoids. Plants. Chapman and Hall, New York
- Grasa-López, A., Miliar-García, Á., Quevedo-Corona, L., Paniagua-Castro, N., Escalona-Cardoso, G., Reyes-Maldonado, E., & Jaramillo-Flores, M.-E. (2016). *Undaria pinnatifida* and Fucoxanthin Ameliorate Lipogenesis and Markers of Both Inflammation and Cardiovascular Dysfunction in an Animal Model of Diet-Induced Obesity. *Marine Drugs*, 14(8), 148
- Guiry, M.D. & Guiry, G.M. (2016). AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. <http://www.algaebase.org>; searched on 17 August 2016
- Gupta, S., & Abu-Ghannam, N. (2011). Bioactive potential and possible health effects of edible brown seaweeds. *Trends in Food Science and Technology*, 22(6), 315–326
- Hafting, J. T., Craigie, J. S., Stengel, D. B., Loureiro, R. R., Buschmann, A. H., Yarish, C., Edwards, M.D., & Critchley, A. T. (2015). Prospects and challenges for industrial production of
- Hammoud, T., Tanguay, J.-F., & Bourassa, M. G. (2000). Management of coronary artery disease: therapeutic options in patients with diabetes. *Journal of the American College of Cardiology*, 36(2), 355–365
- Hanefeld, M. (1998). The role of acarbose in the treatment of non-insulin-dependent diabetes mellitus. *Journal of Diabetes and Its Complications*, 12(4), 228–237
- Hao, Z., Parker, B., Knapp, M., & Yu, L. (2005). Simultaneous quantification of alpha-tocopherol and four major carotenoids in botanical materials by normal phase liquid chromatography-atmospheric pressure chemical ionization-tandem mass spectrometry. *Journal of Chromatography. A*, 1094(1-2), 83–90
- Hara, K., Boutin, P., Mori, Y., Tobe, K., Dina, C., Yasuda, K., Yamauchi, T., Otabe, S., Okada, T., Eto, K., & Kadokawa, T. (2002). Genetic variation in the gene encoding adiponectin is associated with an increased risk of type 2 diabetes in the Japanese population. *Diabetes*, 51(2), 536–540
- Hara, Y., & Honda, M. (1990). The inhibition of alpha -amylase by tea polyphenols. *Agricultural and Biological Chemistry*, 54(8), 1939–1945
- Hasnain, S. Z., Borg, D. J., Harcourt, B. E., Tong, H., Sheng, Y. H., Ng, C. P., Das, I., Wang, R., Chen, A.C., Loudovaris, T., Kay, T.W., Thomas, H.E., Whitehead, J.P., Forbes, J.M., Prins, J.B., & McGuckin, M. A. (2014). Glycemic control in diabetes is restored by therapeutic manipulation of cytokines that regulate beta cell stress. *Nature Medicine*, 20(12), 1417–1426

Hegazi, M. M. I. (2002). Separation, identification and quantification of photosynthetic pigments from three Red Sea seaweeds using reversed-phase high-performance liquid chromatography Muhammad M. I. Hegazi. *Egyptian Journal of Biology*, 4(1998), 1–6

Heo, S. J., Yoon, W. J., Kim, K. N., Ahn, G. N., Kang, S. M., Kang, D. H., Affan, A., Oh, C., Jung, W. K., & Jeon, Y. J. (2010). Evaluation of anti-inflammatory effect of fucoxanthin isolated from brown algae in lipopolysaccharide-stimulated RAW 264.7 macrophages. *Food and Chemical Toxicology*, 48(8-9), 2045–2051

Heo, S. J., Hwang, J. Y., Choi, J. I., Han, J. S., Kim, H. J., & Jeon, Y. J. (2009). Diphlorethohydroxycarmalol isolated from Ishige okamurae, a brown algae, a potent alpha-glucosidase and alpha-amylase inhibitor, alleviates postprandial hyperglycemia in diabetic mice. *European Journal of Pharmacology*, 615(1-3), 252–256

Hegazi, M. M., Pérez-Ruzafa, A., Almela, L., & Candela, M. E. (1998). Separation and identification of chlorophylls and carotenoids from *Caulerpa prolifera*, *Jania rubens* and *Padina pavonica* by reversed-phase high-performance liquid chromatography. *Journal of Chromatography A*, 829(1-2), 153–159

Holdt, S. L., & Kraan, S. (2011). Bioactive compounds in seaweed: Functional food applications and legislation. *Journal of Applied Phycology*, 23(3), 543–597

Hong, G., Park, H., Kim, J., Nagappan, A., Zhang, J., Kang, S., & Won, C. (2012). Anti-oxidant and anti-inflammatory effects of Fraxinus rhynchophylla on lipopolysaccharide (LPS) -induced murine Raw 264 . 7 cells. *Journal of Biomedical Research*, 13(4), 331–338

<http://www.globinmed.com>, Accessed on 20 March 2017

Hu, X., Tao, N., Wang, X., Xiao, J., & Wang, M. (2016). Marine-derived bioactive compounds with anti-obesity effect : A review. *Journal of Functional Foods*, 21, 372–387

Huang, D., Ou, B., Hampsch-Woodill, M., Flanagan, J. A., & Prior, R. L. (2002). High-Throughput Assay of Oxygen Radical Absorbance Capacity (ORAC) Using a Multichannel Liquid Handling System Coupled with a Microplate Fluorescence Reader in 96-Well Format. *Journal of Agricultural and Food Chemistry*, 50(16), 4437–4444

Huang, Y. W., Liu, Y., Dushenkov, S., Ho, C. T., & Huang, M. T. (2009). Anti-obesity effects of epigallocatechin-3-gallate, orange peel extract, black tea extract, caffeine and their combinations in a mouse model. *Journal of Functional Foods*, 1(3), 304–310

Huck, C. W., Popp, M., Scherz, H., & Bonn, G. K. (2000). Development and evaluation of a new method for the determination of the carotenoid content in selected vegetables by HPLC and HPLC-MS-MS. *Journal of Chromatographic Science*, 38(10), 441–449

Huheihel, M., Ishanu, V., Tal, J., & Arad, S. M. (2002). Activity of Porphyridium sp.

- polysaccharide against herpes simplex viruses in vitro and in vivo. *Journal of Biochemical and Biophysical Methods*, 50(2–3), 189–200
- Hurtado, A. Q., Neish, I. C., & Critchley, A. T. (2015). Developments in production technology of Kappaphycus in the Philippines: more than four decades of farming. *Journal of Applied Phycology*, 27(5), 1945–1961
- IDF, International Diabetes Federation (2003). Diabetes Atlas. Executive Summary, 2nd edn. Brussels
- Ionov, V. A., & Basova, M. M. (2003). Use of blue-green micro-seaweed Spirulina platensis for the correction of lipid and hemostatic disturbances in patients with ischemic heart disease. *Voprosy pitaniia*, 72(6), 28–31
- Issa, A., Mohammad, M., Hudaib, M., Tawah, K., Abu Rjai, T., Oran, S., & Bustanji, Y. (2011). A potential role of Lavandula angustifolia in the management of diabetic dyslipidemia. *Journal of Medicinal Plants Research*, 5(16), 3876–3882
- Ito, K., & Hori, K. (1989). Seaweed: Chemical composition and potential food uses. *Food Reviews International*, 5(1), 101–144
- Iwai, K. (2008). Anti-diabetic and Antioxidant Effects of Polyphenols in Brown Alga Ecklonia stolonifera in Genetically Diabetic KK-A<sup>y</sup> Mice. *Plant Foods Human Nutrition*, 63, 163–169
- Jabatan Perikanan Malaysia (Sabah) (2009). Pengenalan kepada industri rumpai laut. Jabatan Perikanan Negeri Sabah
- Jayaraj, J., Wan, A., Rahman, M., & Punja, Z. K. (2008). Seaweed extract reduces foliar fungal diseases on carrot. *Crop Protection*, 27(10), 1360–1366
- Jayaraman, J., Norrie, J., & Punja, Z. K. (2011). Commercial extract from the brown seaweed Ascophyllum nodosum reduces fungal diseases in greenhouse cucumber. *Journal of Applied Phycology*, 23(3), 353–361
- Je, J.Y., Park, P.J., Kim, E.K., Park, J.S., Yoon, H.D., Kim, K.R., & Ahn, C.B. (2009). Antioxidant activity of enzymatic extracts from the brown seaweed Undaria pinnatifida by electron spin resonance spectroscopy. *LWT - Food Science and Technology*, 42(4), 874–878
- Jensen, A. (1993). Present and future needs for algae and algal products. *Hydrobiologia*, 260(1), 15–23
- Jiang, Z., Okimura, T., Yokose, T., Yamasaki, Y., Yamaguchi, K., & Oda, T. (2010). Effects of sulfated fucan, ascophyllan, from the brown Alga Ascophyllum nodosum on various cell lines: A comparative study on ascophyllan and fucoidan. *Journal of Bioscience and Bioengineering*, 110(1), 113–117
- Jiménez-Escrig, A., Jiménez-Jiménez, I., Pulido, R., & Saura-Calixto, F. (2001). Antioxidant activity of fresh and processed edible seaweeds. *Journal of the Science of Food and Agriculture*, 81(5), 530–534

- Jin, E., Polle, J. E. W., Lee, H. K., Hyun, S. M., & Chang, M. (2003). Xanthophylls in microalgae: From biosynthesis to biotechnological mass production and application. *Journal of Microbiology and Biotechnology*, 13(2), 165–174
- Kang, M.C., Kang, N., Kim, S.Y., Lima, I. S., Ko, S.C., Kim, Y.T., Kim, Y.B., Jeung H.D., Choi, K.S. & Jeon, Y.J. (2016). Popular edible seaweed, *Gelidium amansii* prevents against diet-induced obesity. *Food and Chemical Toxicology*, 90, 181–187
- Kang, M.C., Cha, S. H., Wijesinghe, W. A. J. P., Kang, S.M., Lee, S.H., Kim, E.A., Song, C.B., & Jeon, Y.J. (2013). Protective effect of marine algae phlorotannins against AAPH-induced oxidative stress in zebrafish embryo. *Food Chemistry*, 138(2-3), 950–5.
- Kang, S.I., Jin, Y.J., Ko, H.C., Choi, S.Y., Hwang, J.H., Whang, I., Kim M.H., Shin H.S., Jeong H.B., & Kim, S.J. (2008). Petalonia improves glucose homeostasis in streptozotocin-induced diabetic mice. *Biochemical and Biophysical Research Communications*, 373(2), 265–269
- Karalis, K. P., Giannogonas, P., Kodela, E., Koutmani, Y., Zoumakis, M., & Teli, T. (2009). Mechanisms of obesity and related pathology: linking immune responses to metabolic stress. *The FEBS Journal*, 276(20), 5747–5754
- Karamadoukis, L., Shivashankar, G. H., Ludeman, L., & Williams, A. J. (2009). An unusual complication of treatment with orlistat. *Clinical Nephrology*, 71(4), 430–432
- Kaur, Cheryl, Rita, Ang, & Margeret, 2009. Seaweed Culture and Utilisation in Malaysia: Status, Challenges, and Economic Potential. Retreived from: <http://www.mima.gov.my/v2/data/pdf/presentation/30.cheryl%20%20margaret.pdf> Accessed on 12 September, 2017
- Kellogg, J., Grace, M., & Lila, M. (2014). Phlorotannins from Alaskan Seaweed Inhibit Carbolytic Enzyme Activity. *Marine Drugs*, 12(10), 5277–5294
- Kern, P. A., Ranganathan, S., Li, C., Wood, L., & Ranganathan, G. (2001). Adipose tissue tumor necrosis factor and interleukin-6 expression in human obesity and insulin resistance. *American Journal of Physiology. Endocrinology and Metabolism*, 280(5), E745–51
- Keyrouz, R., Abasq, M. L., Bourvellec, C. Le, Blanc, N., Audibert, L., ArGall, E., & Hauchard, D. (2011). Total phenolic contents, radical scavenging and cyclic voltammetry of seaweeds from Brittany. *Food Chemistry*, 126(3), 831–836
- Khairy, H. M., & El-Shafay, S. M. (2013). Seasonal variations in the biochemical composition of some common seaweed species from the coast of Abu Qir Bay, Alexandria, Egypt. *Oceanologia*, 55(2), 435–452
- Khoo, H. E., & Ismail, A. (2009). Stability of Carotenoids from Hexane Fractions of 12 Malaysian Underutilised Tropical Fruits during Low Temperature Storage. *Food*, 3(1), 43–46

- Kim, S. K., & Himaya, S. W. (2011). Medicinal effects of phlorotannins from marine brown algae. *Advances in Food and Nutrition Research*, 64, 97–109
- Kim, Y. S., Lee, Y. M., Kim, J. H., & Kim, J. S. (2013). *Polygonum cuspidatum* inhibits pancreatic lipase activity and adipogenesis via attenuation of lipid accumulation. *BMC Complementary and Alternative Medicine*, 13(1), 282
- Kim, S. M., Jung, Y. J., Kwon, O. N., Cha, K. H., Um, B. H., Chung, D., & Pan, C. H. (2012). A potential commercial source of fucoxanthin extracted from the microalga *Phaeodactylum tricornutum*. *Applied Biochemistry and Biotechnology*, 166(7), 1843–1855
- Kim, T. H., & Bae, J.S. (2010). Ecklonia cava extracts inhibit lipopolysaccharide induced inflammatory responses in human endothelial cells. *Food and Chemical Toxicology : An International Journal Published for the British Industrial Biological Research Association*, 48(6), 1682–1687
- Kim, K. Y., Nguyen, T. H., Kurihara, H., & Kim, S. M. (2010). Alpha-glucosidase inhibitory activity of bromophenol purified from the red alga *Polyoppe lancifolia*. *Journal of Food Science*, 75(5), H145–50
- Kim, K. Y., Nam, K. A., Kurihara, H., & Kim, S. M. (2008). Potent  $\alpha$ -glucosidase inhibitors purified from the red alga *Gratelouzia elliptica*. *Phytochemistry*, 69(16), 2820–2825
- Kondo, H., Shimomura, I., Matsukawa, Y., Kumada, M., Takahashi, M., Matsuda, M. et al. (2002). Association of adiponectin mutation with type 2 diabetes. *Diabetes*, 51, 2325–2328
- Kong, C.S., Kim, J.A., Yoon, N.Y., & Kim, S.K. (2009). Induction of apoptosis by phloroglucinol derivative from Ecklonia Cava in MCF-7 human breast cancer cells. *Food and Chemical Toxicology : An International Journal Published for the British Industrial Biological Research Association*, 47(7), 1653–1658
- Kumar, S., & Brown, L. (2013). Seaweeds as potential therapeutic interventions for the metabolic syndrome. *Reviews in Endocrine and Metabolic Disorders*, 14(3), 299–308
- Kumar, M., Kumari, P., Trivedi, N., Shukla, M. K., Gupta, V., Reddy, C. R. K., & Jha, B. (2011). Minerals, PUFAs and antioxidant properties of some tropical seaweeds from Saurashtra coast of India. *Journal of Applied Phycology*, 23(5), 797–810
- Kumar, A., Ilavarasan, R., Jayachandran, T., Deecaraman, M., Aravindan, P., Padmanabhan, N., & Krishan, M. R. V. (2008). Anti-diabetic activity of *Syzygium cumini* and its isolated compound against streptozotocin-induced diabetic rats. *Journal of Medicinal Plants Research*, 2(9), 246–249
- Kurihara, H., Mitani, T., Kawabata, J., & Takahashi, K. (1999). Inhibitory Potencies of Bromophenols from Rhodomelaceae Algae against Alpha-Glucosidase Activity. *Fisheries Science*, 2, 300–303

- Kwon, Y.I., Apostolidis, E., & Shetty, K. (2008). In vitro studies of eggplant (*Solanum melongena*) phenolics as inhibitors of key enzymes relevant for type 2 diabetes and hypertension. *Bioresource Technology*, 99(8), 2981–2988
- Lann, K. Le, Ferret, C., VanMee, E., Spagnol, C., Lhuillery, M., Payri, C., & Stiger-Pouvreau, V. (2012). Total phenolic, size-fractionated phenolics and fucoxanthin content of tropical Sargassaceae (Fucales, Phaeophyceae) from the South Pacific Ocean: Spatial and specific variability. *Phycological Research*, 60(1), 37–50
- Lebovitz, H. E. (1997). alpha-Glucosidase inhibitors. *Endocrinology and Metabolism Clinics of North America*, 26(3), 539–551
- Lee, S. H., Kang, S. M., Ko, S. C., Lee, D. H., & Jeon, Y. J. (2012). Octaphlorethol A, a novel phenolic compound isolated from a brown alga, *Ishige foliacea*, increases glucose transporter 4-mediated glucose uptake in skeletal muscle cells. *Biochemical and Biophysical Research Communications*, 420(3), 576–581
- Lee, D.S., Park, W. S., Heo, S.J., Cha, S.H., Kim, D., Jeon, Y.J. et al. (2011). Polyopes affinis alleviates airway inflammation in a murine model of allergic asthma. *Journal of Biosciences*, 36(5), 869–877
- Lee, H. J., Kim, H. C., Vitek, L., & Nam, C. M. (2010). Algae consumption and risk of type 2 diabetes: Korean National Health and Nutrition Examination Survey in 2005. *Journal of Nutritional Science and Vitaminology*, 56, 13–18
- Lee, S.H., Yong, L., Karadeniz, F., Kim, M.M., & Kim, S.K. (2009).  $\alpha$ -Glucosidase and  $\alpha$ -amylase inhibitory activities of phloroglucinal derivatives from edible marine brown alga, *Ecklonia cava*. *Journal of the Science of Food and Agriculture*, 89(9), 1552–1558
- Lee, Y. S., Shin, K. H., Kim, B.K., & Lee, S. (2004a). Anti-diabetic activities of fucosterol from *Pelvetia siliquosa*. *Archives of Pharmacal Research*, 27(11), 1120–1122
- Lee, J. Y., Zhao, L., Youn, H. S., Weatherill, A. R., Tapping, R., Feng, L., Lee, W. H., Fitzgerald, K. A., & Hwang, D. H. (2004b). Saturated fatty acid activates but polyunsaturated fatty acid inhibits Toll-like receptor 2 dimerized with Toll-like receptor 6 or 1. *The Journal of Biological Chemistry*, 279(17), 16971–16979
- Lee, S., & Jeon, Y. (2013). Fitoterapia Anti-diabetic effects of brown algae derived phlorotannins , marine polyphenols through diverse mechanisms. *Fitoterapia*, 86, 129–136
- Li, H., Tyndale, S. T., Heath, D. D., & Letcher, R. J. (2005). Determination of carotenoids and all-trans-retinol in fish eggs by liquid chromatography-electrospray ionization-tandem mass spectrometry. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 816(1–2), 49–56
- Li, Y., Wen, S., Kota, B. P., Peng, G., Li, G. Q., Yamahara, J., & Roufogalis, B. D. (2005). *Punica granatum* flower extract, a potent  $\alpha$ -glucosidase inhibitor, improves postprandial hyperglycemia in Zucker diabetic fatty rats. *Journal of Ethnopharmacology*, 99(2), 239–244

- Lim, C. S., Jin, D.Q., Sung, J.Y., Lee, J. H., Choi, H. G., Ha, I., & Han, J.S. (2006). Antioxidant and anti-inflammatory activities of the methanolic extract of Neorhodomela aculeata in hippocampal and microglial cells. *Biological & Pharmaceutical Bulletin*, 29(6), 1212–6
- Lin, H. Y., Juan, S. H., Shen, S. C., Hsu, F. L., & Chen, Y. C. (2003). Inhibition of lipopolysaccharide-induced nitric oxide production by flavonoids in RAW 264.7 macrophages involves heme oxygenase-1. *Biochemical Pharmacology*, 66(9), 1821–1832
- Lin, J., Della-fera, M. A., Baile, C. A., & Clifton, A. (2005). Inhibits Adipogenesis and Induces Apoptosis in 3T3-L1 Adipocytes. *Obesity Research*, 13(6), 982–990
- Liu, L., Heinrich, M., Myers, S., & Dworjanyn, S. A. (2012). Towards a better understanding of medicinal uses of the brown seaweed Sargassum in Traditional Chinese Medicine: A phytochemical and pharmacological review. *Journal of Ethnopharmacology*, 142(3), 591–619
- Loescher, C. M., Morton, D. W., Razic, S., & Agatonovic-Kustrin, S. (2014). High performance thin layer chromatography (HPTLC) and high performance liquid chromatography (HPLC) for the qualitative and quantitative analysis of Calendula officinalis-Advantages and limitations. *Journal of Pharmaceutical and Biomedical Analysis*, 98, 52–59
- Lordan, S., Smyth, T. J., Soler-Vila, A., Stanton, C., & Ross, R. P. (2013). The  $\alpha$ -amylase and  $\alpha$ -glucosidase inhibitory effects of Irish seaweed extracts. *Food Chemistry*, 141(3), 2170–2176
- Lordan, S., Ross, R. P., & Stanton, C. (2011). Marine bioactives as functional food ingredients: Potential to reduce the incidence of chronic diseases. *Marine Drugs*, 9(6), 1056–1100
- Lorente-Cebrian, S., Costa, A. G. V., Navas-Carretero, S., Zabala, M., Laiglesia, L. M., Martinez, J. A., & Moreno-Aliaga, M. J. (2015). An update on the role of omega-3 fatty acids on inflammatory and degenerative diseases. *Journal of Physiology and Biochemistry*, 71(2), 341–349
- Low, L. (2010). The epidemic of type 2 diabetes mellitus in the Asia-Pacific region. *Pediatric Diabetes*, 11, 212–215
- Lunkapis, G.J. & Danny, H.W. (2016). Preliminary Impact Assessment of Seaweed Cultivation by the Coastal Communities in Sabah, Malaysia. *Transactions on Science and Technology*, 3(2-2), 413 - 420
- MacArtain, P., Gill, C. I. R., Brooks, M., Campbell, R., & Rowland, I. R. (2007). Nutritional value of edible seaweeds. *Nutrition Reviews*, 65(12 Pt 1), 535–543
- Maeda, H., Tsukui, T., Sashima, T., Hosokawa, M., & Miyashita, K. (2008). Seaweed carotenoid, fucoxanthin, as a multi-functional nutrient. *Asia Pacific Journal of Clinical Nutrition*, 17(Supplement 1), 196–199
- Maeda, H., Hosokawa, M., Sashima, T., & Miyashita, K. (2007). Dietary combination

of fucoxanthin and fish oil attenuates the weight gain of white adipose tissue and decreases blood glucose in obese/diabetic KK-Ay mice. *Journal of Agricultural and Food Chemistry*, 55(19), 7701–7706

Maeda, H., Hosokawa, M., Sashima, T., Takahashi, N., Kawada, T., & Miyashita, K. (2006). Fucoxanthin and its metabolite, fucoxanthinol, suppress adipocyte differentiation in 3T3-L1 cells. *International Journal of Molecular Medicine*, 18(1), 147–152

Mariutti, L. R. B., Pereira, D. M., Mercadante, A. Z., Valentão, P., Teixeira, N., & Andrade, P. B. (2012). Further insights on the carotenoid profile of the echinoderm *Marthasterias glacialis* L. *Marine Drugs*, 10(7), 1498–1510

Marles, R. J., & Farnsworth, N. R. (1995). Anti-diabetic plants and their active constituents. *Phytomedicine*, 2(2), 137–189

Marquardt, J., & Hanelt, D. (2004). and Other Marine Red Algae From Polar and Temperate Habitats. *European Journal of Phycology*, 39(3), 285–292

Marsham, S., Scott, G. W., & Tobin, M. L. (2007). Comparison of nutritive chemistry of a range of temperate seaweeds. *Food Chemistry*, 100(4), 1331–1336

Martins, A., Vieira, H., Gaspar, H., & Santos, S. (2014). Marketed Marine Natural Products in the Pharmaceutical and Cosmeceutical Industries: Tips for Success. *Marine Drugs*, 12(2), 1066–1101

Mary, J. S., Vinotha, P., & Pradeep, A. M. (2012). Screening for in vitro cytotoxic activity of seaweed, *Sargassum* sp. against Hep-2 and MCF-7 cancer cell lines. *Asian Pacific Journal of Cancer Prevention*, 13(12), 6073–6076

Matanjun, P., Mohamed, S., Mustapha, N. M., & Muhammad, K. (2009). Nutrient content of tropical edible seaweeds, *Eucheuma cottonii*, *Caulerpa lentillifera* and *Sargassum polycystum*. *Journal of Applied Phycology*, 21(1), 75–80

Matanjun, P., Mohamed, S., Mustapha, N. M., Muhammad, K., & Ming, C. H. (2008). Antioxidant activities and phenolics content of eight species of seaweeds from north Borneo. *Journal of Applied Phycology*, 20(4), 367–373

Mathew, B., Sankaranarayanan, R., Nair, P. P., Varghese, C., Somanathan, T., Amma, B. P., Amma, N.S., & Nair, M. K. (1995). Evaluation of chemoprevention of oral cancer with *Spirulina fusiformis*. *Nutrition and Cancer*, 24(2), 197–202

Matsui, T., Tanaka, T., Tamura, S., Toshima, A., Tamaya, K., Miyata, Y., Tanaka, K., & Matsumoto, K. (2007). Alpha-Glucosidase inhibitory profile of catechins and theaflavins. *Journal of Agricultural and Food Chemistry*, 55(1), 99–105

Mattio, L., Payri, C. E., & Verlaque, M. (2009). Taxonomic revision and geographic distribution of the subgenus sargassum (FUCALES, PHAEOPHYCEAE) in the western and central pacific islands based on morphological and molecular analyses. *Journal of Phycology*, 45(5), 1213–1227

Maury, E., & Brichard, S. M. (2010). Adipokine dysregulation, adipose tissue

- inflammation and metabolic syndrome. *Molecular and Cellular Endocrinology*, 314(1), 1–16
- Mayer, A. M. S., & Hamann, M. T. (2002). Marine Pharmacology in 2000 : Marine Compounds Antituberculosis , and Antiviral Activities; Affecting the Cardiovascular , Immune , and Nervous Systems and Other Miscellaneous Mechanisms of Action. *Comparative Biochemistry and Physiology Part C*, 132, 315–339
- McHugh, D. J. (2003). A Guide to the Seaweed Industry. *FAO Fisheries Technical Paper*.
- Melendez-Martinez, A. J., Britton, G., Vicario, I. M., & Heredia, F. J. (2005). Identification of isolutein (lutein epoxide) as cis-antheraxanthin in orange juice. *Journal of Agricultural and Food Chemistry*, 53(24), 9369–9373
- Mikami, K., & Hosokawa, M. (2013). Biosynthetic pathway and health benefits of fucoxanthin, an algae-specific xanthophyll in brown seaweeds. *International Journal of Molecular Sciences*, 14(7), 13763–13781
- Miller, G. L. (1959). Use of Dinitrosalicylic Acid Reagent for Determination of Reducing Sugar. *Analytical Chemistry*, 31(3), 426–428
- Min, K.H., Kim, H.J., Jeon, Y.J., & Han, J.S. (2011). *Ishige okamurae* ameliorates hyperglycemia and insulin resistance in C57BL/KsJ-db/db mice. *Diabetes Research and Clinical Practice*, 93(1), 70–76
- Mishra, K., Ojha, H., & Chaudhury, N. K. (2012). Estimation of antiradical properties of antioxidants using DPPH - assay: A critical review and results. *Food Chemistry*, 130(4), 1036–1043
- Miyashita, K. (2009). The carotenoid fucoxanthin from brown seaweed affects obesity. *Lipid Technology*, 21(8-9), 186–190
- Miyashita, K., Nishikawa, S., Beppu, F., Tsukui, T., Abe, M., & Hosokawa, M. (2011). The allenic carotenoid fucoxanthin, a novel marine nutraceutical from brown seaweeds. *Journal of the Science of Food and Agriculture*, 91(7), 1166–1174
- Mohamed, S., Hashim, S. N., & Rahman, H. A. (2012). Seaweeds: A sustainable functional food for complementary and alternative therapy. *Trends in Food Science and Technology*, 23(2), 83–96
- Morgan, N. G., & Dhayal, S. (2010). Unsaturated fatty acids as cytoprotective agents in the pancreatic beta-cell. *Prostaglandins, Leukotrienes, and Essential Fatty Acids*, 82(4-6), 231–236
- Motshakeri, M., Ebrahimi, M., Goh, Y. M., Matanjun, P., & Mohamed, S. (2013). *Sargassum polycystum* reduces hyperglycaemia, dyslipidaemia and oxidative stress via increasing insulin sensitivity in a rat model of type 2 diabetes. *Journal of the Science of Food and Agriculture*, 93(7), 1772–1778
- Motshakeri, M., Ebrahimi, M., Goh, Y. M., Othman, H. H., Hair-Bejo, M., &

- Mohamed, S. (2014). Effects of brown seaweed (*Sargassum polycystum*) extracts on kidney, liver, and pancreas of type 2 diabetic rat model. *Evidence-Based Complementary and Alternative Medicine*, 2014, Article ID 379407, 11
- Mowles, J. (1990). Mycoplasma detection. In J. Pollard & J. Walker (Eds.), Methods in molecular biology, animal cell culture (pp. 65–74). New Jersey: Humana Press
- Munro MHG & Blunt JW (2005) MarinLit, a marine chemical literature database. Available from: University of Canterbury, Christchurch, New Zealand,
- Murata, M., & Nakazoe, J. (2001). Production and Use of Marine Algae in Japan. *Jarp-Japan Agricultural Research Quarterly*, 35(4), 281–290
- Murugan, A. C., Karim, M. R., Yusoff, M. B. M., Tan, S. H., Asras, M. F. B. F., & Rashid, S. S. (2015). New insights into seaweed polyphenols on glucose homeostasis. *Pharmaceutical Biology*, 1–11
- Nagappan, T., & Vairappan, C. S. (2013). Nutritional and bioactive properties of three edible species of green algae, genus Caulerpa (Caulerpaceae). *Journal of Applied Phycology*, 1–9
- Nathan, D. M., Davidson, M. B., DeFronzo, R. A., Heine, R. J., Henry, R. R., Pratley, R., & Zinman, B. (2007). Impaired fasting glucose and impaired glucose tolerance: Implications for care. *Diabetes Care*, 30(3), 753–759
- NHMS, 1986–1987(National Health Morbidity Survey 1986–1987). Diabetes Mellitus. Volume 4. Institute for Public Health, Ministry of Health, Malaysia
- NHMS, 1996 (National Health Morbidity Survey 1996). Diabetes. Volume 9. Institute for Public Health, Ministry of Health, Malaysia
- NHMS, 2011(National Health and Morbidity Survey 2011). Volume 2: Non-communicable diseases; 2011. Institute for Public Health, Ministry of Health, Malaysia
- NHMS, 2015 (National Health and Morbidity Survey 2015). Volume 2: Non-Communicable Diseases, Risk Factors & Other Health Problems . Institute for Public Health, Ministry of Health, Malaysia
- Nickavar, B., & Mosazadeha, G. (2009). Influence of Three Morus Species Extracts on  $\alpha$ -Amylase Activity. *Iranian Journal of Pharmaceutical Research*, 8, 115–119
- Nielsen, L. L., Young, A. A., & Parkes, D. G. (2004). Pharmacology of exenatide (synthetic exendin-4): a potential therapeutic for improved glycemic control of type 2 diabetes. *Regulatory Peptides*, 117(2), 77–88
- Nomura, T., Kikuchi, M., Kubodera, A., & Kawakami, Y. (1997). Proton-donative antioxidant activity of fucoxanthin with 1,1-Diphenyl-2-Picrylhydrazyl (DPPH). *IUBMB Life*, 42(2), 361–370
- Nor, A. M., Gray, T. S., Caldwell, G. S., & Stead, S. M. (2016). Is a cooperative approach to seaweed farming effectual? An analysis of the Seaweed Cluster

- Project ( SCP ), Is a cooperative approach to seaweed farming effectual ? An analysis of the seaweed cluster project (SCP), Malaysia. *Journal of Applied Phycology*. <http://doi.org/10.1007/s10811-016-1025-y>
- Norziah, M. H., & Ching, C. Y. (2000). Nutritional composition of edible seaweed Gracilaria changgi. *Food Chemistry*, 68(1), 69–76
- Ntambi, M. J., & Kim, Y.-C. (2000). Symposium: Adipocyte Function, Differentiation and Metabolism Regulation of Leptin Production in Humans. *Journal Nutrition*, 130, 3127–3131
- Nugent, C., Prins, J. B., Whitehead, J. P., Wentworth, J. M., Chatterjee, V. K., & O'Rahilly, S. (2001). Arachidonic acid stimulates glucose uptake in 3T3-L1 adipocytes by increasing GLUT1 and GLUT4 levels at the plasma membrane. Evidence for involvement of lipoxygenase metabolites and peroxisome proliferator-activated receptor gamma. *The Journal of Biological Chemistry*, 276(12), 9149–9157
- Nwosu, F., Morris, J., Lund, V. a., Stewart, D., Ross, H. a., & McDougall, G. J. (2011). Anti-proliferative and potential anti-diabetic effects of phenolic-rich extracts from edible marine algae. *Food Chemistry*, 126(3), 1006–1012
- O'Connor, C. J., Sun, D., Smith, B. G., & Melton, L. D. (2003). Effect of Soluble Dietary Fibers on Lipase-catalyzed Hydrolysis of Tributyrin. *Journal of Food Science*, 68(3), 1093–1099
- Oh, J.H., Kim, J., & Lee, Y. (2016). Anti-inflammatory and anti-diabetic effects of brown seaweeds in high-fat diet-induced obese mice. *Nutrition Research and Practice*, 10(1), 42–48
- O'Sullivan, A. M., O'Callaghan, Y. C., O'Grady, M. N., Queguineur, B., Hanniffy, D., Troy, D. J., O'Brien, N. M. (2011). In vitro and cellular antioxidant activities of seaweed extracts prepared from five brown seaweeds harvested in spring from the west coast of Ireland. *Food Chemistry*, 126(3), 1064–1070
- Ono, Y., Hattori, E., Fukaya, Y., Imai, S., & Ohizumi, Y. (2006). Anti-obesity effect of Nelumbo nucifera leaves extract in mice and rats. *Journal of Ethnopharmacology*, 106(2), 238–244
- Ortega-Calvo, J. J., Mazuelos, C., Hermosin, B., & Saiz-Jimenez, C. (1993). Chemical composition of Spirulina and eukaryotic algae food products marketed in Spain. *Journal of Applied Phycology*, 5(4), 425–435
- Oumaskour, K., Boujaber, N., Etahiri, S., & Assobhei, O. (2013). Anti-inflammatory and antimicrobial activities of twenty-tree marine red algae from the coast of Sidi Bouzid (El Jadida-Morocco). *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(3), 145–149
- Palanisamy, U., Manaharan, T., Teng, L. L., Radhakrishnan, A. K. C., Subramaniam, T., & Masilamani, T. (2011). Rambutan rind in the management of hyperglycemia. *Food Research International*, 44(7), 2278–2282

- Pangestuti, R., & Kim, S. K. (2011). Biological activities and health benefit effects of natural pigments derived from marine algae. *Journal of Functional Foods*, 3(4), 255–266
- Patankar, M. S., Oehninger, S., Barnett, T., Williams, R. L., & Clark, G. F. (1993). A revised structure for fucoidan may explain some of its biological activities. *The Journal of Biological Chemistry*, 268(29), 21770–21776
- Paul, N. a., Neveux, N., Magnusson, M., & de Nys, R. (2013). Comparative production and nutritional value of “sea grapes” — the tropical green seaweeds *Caulerpa lentillifera* and *C. racemosa*. *Journal of Applied Phycology*, 1833–1844
- Paxman, J. R., Richardson, J. C., Dettmar, P. W., & Corfe, B. M. (2008). Daily ingestion of alginic reduces energy intake in free-living subjects. *Appetite*, 51(3), 713–719
- Pedersén, M., Collen, J., Abrahamsson, K., & Ekdahl, A. (1996). Production of halocarbons from seaweeds: an oxidative stress reaction. *Scientia Marina*, 60(Supplement 1), 257–263
- Peng, J., Yuan, J. P., Wu, C. F., & Wang, J. H. (2011). Fucoxanthin, a marine carotenoid present in brown seaweeds and diatoms: Metabolism and bioactivities relevant to human health. *Marine Drugs*, 9(10), 1806–1828
- Pereira, H., Barreira, L., Figueiredo, F., Custódio, L., Vizotto-Duarte, C., Polo, C., Rešek, E., Engelen, A., & Varela, J. (2012). Polyunsaturated fatty acids of marine macroalgae: Potential for nutritional and pharmaceutical applications. *Marine Drugs*, 10(9), 1920–1935
- Peterson, T. R., Sengupta, S. S., Harris, T. E., Carmack, A. E., Kang, S. A., Balderas, E., Guertin, D.A., Madden, K.L., Carpenter, A.E., Finck, B.N., & Sabatini, D. M. (2011). mTOR complex 1 regulates lipin 1 localization to control the SREBP pathway. *Cell*, 146(3), 408–420
- Phang, S. M. (2006). Seaweed resources in Malaysia: Current status and future prospects. *Aquatic Ecosystem Health & Management*, 9(2), 185–202
- Phang, S.M., Lewmanomont, K. & Lim, P.E. (eds) (2008). Taxonomy of Southeast Asian Seaweeds. Institute of Ocean and Earth Sciences, University of Malaya Monograph Series 2. ISBN: 978-967-5148-13-2
- Phang, S. M. (2010). Potential products from tropical algae and seaweeds, especially with reference to Malaysia. *Malaysian Journal of Science*, 29(2), 160–166.
- Pillay, R.P. & Lim E.H. (1960). Incidence of Diabetes Mellitus in Malaya. *Medical Journal of Malaya*, 16, 242-244
- Pires, K. M., de Alencar, D. B., de Sousa, M. B., Sampaio, A. H., & Saker-Sampaio, S. (2008). Levels of  $\alpha$ -carotene and  $\beta$ -carotene in dehydrated marine macroalgae. *Revista Ciência Agronómica*, 39(2), 257-262
- Pires-Cavalcante, K. M. D. S., de Alencar, D. B., de Sousa, M. B., Sampaio, A. H., &

- Saker-Sampaio, S. (2011). Seasonal changes of alpha-tocopherol in green marine algae (*Caulerpa* genus). *Journal of Food Science*, 76(5), C775–81
- Pop, R. M., Weesepoel, Y., Socaciu, C., Pintea, A., Vincken, J. P., & Gruppen, H. (2014). Carotenoid composition of berries and leaves from six Romanian sea buckthorn (*Hippophae rhamnoides* L.) varieties. *Food Chemistry*, 147, 1–9
- Pradhan, A. D., Manson, J. E., Rifai, N., Buring, J. E., & Ridker, P. M. (2001). C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *The Journal of the American Medical Association*, 286(3), 327–334
- Prior, R. L., & Cao, G. (1999). In vivo total antioxidant capacity: comparison of different analytical methods. *Free Radical Biology & Medicine*, 27(11-12), 1173–1181
- Prior, R. L., Wu, X., & Schaich, K. (2005). Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal of Agricultural and Food Chemistry*, 53(10), 4290–4302
- Rajauria, G., Jaiswal, A. K., Abu-Gannam, N., & Gupta, S. (2013). Antimicrobial, antioxidant and free radical-scavenging capacity of brown seaweed *himanthalia elongata* from western coast of Ireland. *Journal of Food Biochemistry*, 37(3), 322–335
- Ramachandran, A. (2012). Trends in prevalence of diabetes in Asian countries. *World Journal of Diabetes*, 3(6), 110
- Rangel-Yagui, C., Danesi, E., de Carvalho, J., & Sato, S. (2004). Chlorophyll production from *Spirulina platensis*: cultivation with urea addition by fed-batch process. *Bioresource Technology*, 92(2), 133–141
- Rangwala, S. M., & Lazar, M. A. (2000). Transcriptional Control Of Adipogenesis. *Annual Review of Nutrition*, 20(1), 535–559
- Rasmussen, R. S., & Morrissey, M. T. (2007). Marine biotechnology for production of food ingredients. *Advances in Food and Nutrition Research*, 52, 237–292
- Rebah, F., Smaoui, S., Frikha, F., Gargouri, Y., & Miled, N. (2008). Inhibitory Effects of Tunisian Marine Algal Extracts on Digestive Lipases. *Applied Biochemistry and Biotechnology*, 151(1), 71–79
- Reich, E., Schibli, A., & DeBatt, A. (2008). Validation of high-performance thin-layer chromatographic methods for the identification of botanicals in a cGMP environment. *Journal of AOAC International*, 91(1), 13–20
- Rengasamy, K. R. R., Aderogba, M. a, Amoo, S. O., Stirk, W. a, & Van Staden, J. (2013). Potential antiradical and alpha-glucosidase inhibitors from *Ecklonia maxima* (Osbeck) Papenfuss. *Food Chemistry*, 141(2), 1412–5
- Rhein-Knudsen, N., Ale, M., & Meyer, A. (2015). Seaweed Hydrocolloid Production: An Update on Enzyme Assisted Extraction and Modification Technologies. *Marine Drugs*, 13(6), 3340–3359

- Rios, J. L., Francini, F., & Schinella, G. R. (2015). Natural Products for the Treatment of Type 2 Diabetes Mellitus. *Planta Medica*, 2015, 1–20
- Rieusset, J., Touri, F., Michalik, L., Escher, P., Desvergne, B., Niesor, E., Wahli, W. (2002). A new selective PPAR $\gamma$  antagonist with antiobesity and anti-diabetic activity. *Molecular Endocrinology*, 16, 2628–2644
- Rivera, S. M., & Canela-Garayoa, R. (2012). Analytical tools for the analysis of carotenoids in diverse materials. *Journal of Chromatography A*, 1224, 1–10
- Rivera, S. M., Vilardo, F., Zhu, C., Bai, C., Farre, G., Christou, P., & Canela-garayoa, R. (2013). Fast Quantitative Method for the Analysis of Carotenoids in Transgenic Maize. *Journal of Agricultural and Food Chemistry*, 61(22) 5279–5285
- Rivera, S. M., Christou, P., & Canela-Garayoa, R. (2014). Identification of carotenoids using mass spectrometry. *Mass Spectrometry Reviews*, 33(5), 353–372.
- Rmiki, N.-E., Brunet, C., Cabioch, J., & Lemoine, Y. (1996). Xanthophyll-cycle and photosynthetic adaptation to environment in macro- and microalgae. In S. C. Lindstrom & D. J. Chapman (Eds.), *Fifteenth International Seaweed Symposium: Proceedings of the Fifteenth International Seaweed Symposium held in Valdivia, Chile, in January 1995* (pp. 407–413). Dordrecht: Springer Netherlands
- Rock, C. L. (2003). Carotenoid update. *Journal of the American Dietetic Association*, 103(4), 423–425
- Rodríguez-Bernaldo de Quirós, A., Lage-Yusty, M. A., & López-Hernández, J. (2010). Determination of phenolic compounds in macroalgae for human consumption. *Food Chemistry*, 121(2), 634–638
- Rohani-Ghadikolaei, K., Abdulalian, E., & Ng, W.K. (2012). Evaluation of the proximate, fatty acid and mineral composition of representative green, brown and red seaweeds from the Persian Gulf of Iran as potential food and feed resources. *Journal of Food Science and Technology*, 49(6), 774–780
- Roncari, D. A., Lau, D. C., & Kindler, S. (1981). Exaggerated replication in culture of adipocyte precursors from massively obese persons. *Metabolism: Clinical and Experimental*, 30(5), 425–427.
- Rosen, E. D. (2002). The Molecular Control of Adipogenesis, with Special Reference to Lymphatic Pathology. *Annals of the New York Academy of Sciences*, 979(1), 143–158
- Rosenfeld, Y., & Shai, Y. (2006). Lipopolysaccharide (Endotoxin)-host defense antibacterial peptides interactions: Role in bacterial resistance and prevention of sepsis. *Biochimica et Biophysica Acta - Biomembranes*, 1758(9), 1513–1522
- Rosiak, M., Grzeszczak, S., Kosior, D. a, & Postuła, M. (2014). Emerging treatments in type 2 diabetes: focus on canagliflozin. *Therapeutics and Clinical Risk Management*, 10, 683–9

- Rother, K. (2007). Diabetes Treatment -Bridging the Divide. *The New England Journal of Medicine*, 356(15), 1499 – 1501
- Rubio-Rodriguez, N., Beltrin, S., Jaime, I., de Diego, S. M., Sanz, M. T., & Carballido, J. R. (2010). Production of omega-3 polyunsaturated fatty acid concentrates: A review. *Innovative Food Science and Emerging Technologies*, 11(1), 1–12
- Ruperez, P., & Saura-Calixto, F. (2001). Dietary fibre and physicochemical properties of edible Spanish seaweeds. *European Food Research and Technology*, 212(3), 349–354
- Rutten, G.E.H.M., De Grauw, W.J.C., Nijpels, G., et al. (2006). Dutch College of General Practitioners: guidelines on Type 2 diabetes, second revision (in Dutch). *Huisarts Wet*, 49, 137–52
- Sabeena Farvin, K. H., & Jacobsen, C. (2013). Phenolic compounds and antioxidant activities of selected species of seaweeds from Danish coast. *Food Chemistry*, 138(2-3), 1670–1681
- Sachindra, N. M., Sato, E., Maeda, H., Hosokawa, M., Niwano, Y., Kohno, M., & Miyashita, K. (2007). Radical scavenging and singlet oxygen quenching activity of marine carotenoid fucoxanthin and its metabolites. *Journal of Agricultural and Food Chemistry*, 55(21), 8516–8522
- Saito, N., Sakai, H., Suzuki, S., Sekihara, H., & Yajima, Y. (1998). Effect of an alpha-glucosidase inhibitor (voglibose), in combination with sulphonylureas, on glycaemic control in type 2 diabetes patients. *The Journal of International Medical Research*, 26(5), 219–232
- Sánchez-Machado, D. I., López-Cervantes, J., López-Hernández, J., & Paseiro-Losada, P. (2004). Fatty acids, total lipid, protein and ash contents of processed edible seaweeds. *Food Chemistry*, 85(3), 439–444
- Saw, C. L. L., Yang, A. Y., Guo, Y., & Kong, A.N. T. (2013). Astaxanthin and omega-3 fatty acids individually and in combination protect against oxidative stress via the Nrf2-ARE pathway. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*, 62, 869–75.
- Schmid, M., Guihéneuf, F., & Stengel, D. B. (2014). Fatty acid contents and profiles of 16 macroalgae collected from the Irish Coast at two seasons. *Journal of Applied Phycology*, 26(1), 451–463
- Schubert, N., García-Mendoza, E., & Pacheco-Ruiz, I. (2006). Carotenoid Composition of Marine Red Algae. *Journal of Phycology*, 42(6), 1208–1216
- Seo, M. J., Lee, O. H., Choi, H. S., & Lee, B. Y. (2012). Extract from edible red seaweed (*Gelidium amansii*) inhibits lipid accumulation and ROS production during differentiation in 3T3-L1 cells. *Preventive Nutrition and Food Science*, 17(2), 129–135

- Sharifuddin, Y., Chin, Y. X., Lim, P. E., & Phang, S. M. (2015). Potential bioactive compounds from seaweed for diabetes management. *Marine Drugs*, 13(8), 5447–549
- Sharma, B. R., & Rhyu, D. Y. (2014). Anti-diabetic effects of *Caulerpa lentillifera*: stimulation of insulin secretion in pancreatic  $\beta$ -cells and enhancement of glucose uptake in adipocytes. *Asian Pacific Journal of Tropical Biomedicine*, 4(7), 575–80
- Shibata, T., Ishimaru, K., Kawaguchi, S., Yoshikawa, H., & Hama, Y. (2008). Antioxidant activities of phlorotannins isolated from Japanese Laminariaceae. *Journal of Applied Phycology*, 20(5), 705–711
- Shiratori, K., Ohgami, K., Ilieva, I., Jin, X.H., Koyama, Y., Miyashita, K., Yoshida, K., Kase, S. & Ohno, S. (2005). Effects of fucoxanthin on lipopolysaccharide-induced inflammation in vitro and in vivo. *Experimental Eye Research*, 81(4), 422–428
- Shu, M.H., Appleton, D., Zandi, K., & Abu Bakar, S. (2013). Anti-inflammatory, gastroprotective and anti-ulcerogenic effects of red algae *Gracilaria changii* (Gracilariales, Rhodophyta) extract. *BMC Complementary and Alternative Medicine*, 13(1), 61
- Sievanen, L., Crawford, B., Pollnac, R., & Lowe, C. (2005). Weeding through assumptions of livelihood approaches in ICM: Seaweed farming in the Philippines and Indonesia. *Ocean & Coastal Management*, 48(3–6), 297–313
- Simopoulos, A. P. (2008). The importance of the omega-6/omega-3 fatty acid ratio in cardiovascular disease and other chronic diseases. *Experimental Biology and Medicine*, 233(6), 674–688
- Simopoulos, A. P. (2002). The importance of the ratio of omega-6/omega-3 essential fatty acids. *Biomedicine and Pharmacotherapy*, 56(8), 365–379
- Singleton, V. L., & Rossi, J. A. (1965). Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. *American Journal of Enology and Viticulture*, 16, 144–158
- Sivagnanam, S. P., Yin, S., Choi, J. H., Park, Y. B., Woo, H. C., & Chun, B. S. (2015). Biological Properties of Fucoxanthin in Oil Recovered from Two Brown Seaweeds Using Supercritical CO<sub>2</sub> Extraction. *Marine Drugs*, 13(6), 3422–42
- Sivathanu, B., & Palaniswamy, S. (2012). Purification and characterization of carotenoids from green algae *Chlorococcum humicola* by HPLC-NMR and LC-MS-APCI. *Biomedicine and Preventive Nutrition*, 2(4), 276–282
- Smit, A. J. (2004). Medicinal and pharmaceutical uses of seaweed natural products: A review. *Journal of Applied Phycology*, 16(4), 245–262
- Sokolova, E. V., Barabanova, a. O., Bogdanovich, R. N., Khomenko, V. a., Solov'eva, T. F., & Yermak, I. M. (2011). In vitro antioxidant properties of red algal polysaccharides. *Biomedicine & Preventive Nutrition*, 1(3), 161–167

- Song, Y., Manson, J. E., Buring, J. E., Sesso, H. D., & Liu, S. (2005). Associations of dietary flavonoids with risk of type 2 diabetes, and markers of insulin resistance and systemic inflammation in women: a prospective study and cross-sectional analysis. *Journal of the American College of Nutrition*, 24(5), 376–384
- Sorensen, S. H., Noren, O., Sjostrom, H., & Danielsen, E. M. (1982). Amphiphilic Pig Intestinal Microvillus Maltase/Glucoamylase. Structure and Specificity. *European Journal of Biochemistry*, 126(3), 559–568
- Sousa, M. B. de, Pires, K. M. dos S., Alencar, D. B. de, Sampaio, A. H., & Saker-Sampaio, S. (2008). Alfa-carotene and beta-carotene contents in dried marine macroalgae. *Food Science and Technology (Campinas)*, 28, 953–958
- Souza, B. W. S., Cerqueira, M. A., Bourbon, A. I., Pinheiro, A. C., Martins, J. T., Teixeira, J. A., Coimbra M.A., & Vicente, A. A. (2012). Food Hydrocolloids Chemical characterization and antioxidant activity of sulfated polysaccharide from the red seaweed Gracilaria birdiae. *Food Hydrocolloids*, 27(2), 287–292
- Souza, P. R., & Norling, L. V. (2015). Implications for eicosapentaenoic acid- and docosahexaenoic acid-derived resolvins as therapeutics for arthritis. *European Journal of Pharmacology*, 1–9
- Spangenberg, B., Poole, C. F., & Weins, C. (2011). Quantitative thin-layer chromatography: A practical survey. Springer-Verlag Berlin Heidelberg
- Stefanov, K., Konaklieva, M., Brechany, E. Y., & Christie, W. W. (1988). Fatty acid composition of some algae from the black sea. *Phytochemistry*, 27(11), 3495–3497
- Stirk, W. (2006). World seaweed resources. *South African Journal of Botany*, 72(4), 666–666
- Straub, O., & Pfander, H. (1987). Key to Carotenoids. Basel: Birkhauser, ISBN: 0817618600
- Su, L., Yin, J.J., Charles, D., Zhou, K., Moore, J., & Yu, L. (Lucy). (2007). Total phenolic contents, chelating capacities, and radical-scavenging properties of black peppercorn, nutmeg, rosehip, cinnamon and oregano leaf. *Food Chemistry*, 100(3), 990–997
- Suarez, Y., Gonzalez, L., Cuadrado, A., Berciano, M., Lafarga, M., & Munoz, A. (2003). Kahalalide F, a new marine-derived compound, induces oncosis in human prostate and breast cancer cells. *Molecular Cancer Therapeutics*, 2(9), 863–872.
- Suhaimi Md. Yasir & Ramlan Ali@Ally. (2012). Sistem Mini Estet Industri Rumpai Laut. PutrajayaJabatan Perikanan Malaysia
- Sullivan, O., & Naim, A. M. (2013). Cellular and *In vitro* Models to Assess Antioxidant Activities of Seaweed Extracts and the Potential Use of the Extracts as Ingredients. PhD Thesis, University College Cork

- Sutherland, J. E., & Hoehns, J. D. (2004). Treating type 2 diabetes: targeting the many causative factors. *The Journal of Family Practice*, 53(5), 376–388
- Suzuki, N., Fujimura, A., Nagai, T., Mizumoto, I., Itami, T., Hatake, H., Nozawa T., Kato N., Nomoto T., & Yoda, B. (2004). Antioxidative activity of animal and vegetable dietary fibers. *BioFactors*, 21(1-4), 329–333
- Swanson, D., Block, R., & Mousa, S. (2012). Omega-3 fatty acids EPA and DHA: health benefits throughout life. *Advances in Nutrition*, 3(1), 1–7
- Takaichi, S. (2013). Distributions, biosyntheses and functions of carotenoids in algae. *Agro Food Industry Hi-Tech*, 24(1), 55–58
- Takaichi, S. (2011). Carotenoids in Algae: Distributions, Biosyntheses and Functions. *Marine Drugs*, 9(6), 1101–1118
- Tan, J., Lim, P. E., & Phang, S. M. (2013). Phylogenetic relationship of *Kappaphycus Doty* and *Eucheuma J. Agardh* (Solieriaceae, Rhodophyta) in Malaysia. *Journal of Applied Phycology*, 25(1), 13–29
- Tannoury, M. Y., Elia, J. M., Saab, A. M., Makhlof, H. Y., Abboud, J. S., & Daou, R. J. (2016). Evaluation of Cytotoxic Activity of *Sargassum vulgare* From the Lebanese Coast Against Jurkat Cancer Cell Line, 6(06), 108–112
- Teixeira, V. L. (2013). Marine natural products from seaweeds. *Produtos Naturais de Algas Marinhas Bentônicas*, 5(3), 343–362
- Unno, Y., Akuta, T., Sakamoto, Y. I., Horiuchi, S., & Akaike, T. (2006). Nitric oxide-induced downregulation of leptin production by 3T3-L1 adipocytes. *Nitric Oxide - Biology and Chemistry*, 15(2), 125–132
- Vairappan, C. S., Kamada, T., Lee, W. W., & Jeon, Y. J. (2013). Anti-inflammatory activity of halogenated secondary metabolites of *Laurencia snackeyi* (Weber-van Bosse) Masuda in LPS-stimulated RAW 264.7 macrophages. *Journal of Applied Phycology*, 25(6), 1805–1813
- van Breemen, R. B., Dong, L., & Pajkovic, N. D. (2012). Atmospheric pressure chemical ionization tandem mass spectrometry of carotenoids. *International Journal of Mass Spectrometry*, 312, 163–172
- van de Laar, F. A., Lucassen, P. L., Akkermans, R. P., van de Lisdonk, E. H., Rutten, G. E., & van Weel, C. (2005). Alpha-glucosidase inhibitors for patients with type 2 diabetes: results from a Cochrane systematic review and meta-analysis. *Diabetes Care*, 28(1), 154–63
- van de Laar, F. A. (2008). Alpha-glucosidase inhibitors in the early treatment of type 2 diabetes. *Vascular Health and Risk Management*, 4(6), 1189–1195
- van Ginneken, V. J. T., Helsper, J. P. F. G., de Visser, W., van Keulen, H., & Brandenburg, W. A. (2011). Polyunsaturated fatty acids in various macroalgal species from North Atlantic and tropical seas. *Lipids in Health and Disease*, 10(1), 104

- Vishnu , P. C. N., Anjana, T., Banerji, A., & Gopalakrishnapillai, A. (2010). Gallic acid induces GLUT4 translocation and glucose uptake activity in 3T3-L1 cells. *FEBS Letters*, 584(3), 531–536
- Vogel, W. C., & Zieve, L. (1963). A rapid and sensitive turbidimetric method for serum lipase based upon differences between the lipases of normal and pancreatitis serum. *Clinical Chemistry*, 9, 168–181
- Wang, T., Jónsdóttir, R., Liu, H., Gu, L., Kristinsson, H. G., Raghavan, S., & Ólafsdóttir, G. (2012). Antioxidant capacities of phlorotannins extracted from the brown algae *Fucus vesiculosus*. *Journal of Agricultural and Food Chemistry*, 60(23), 5874–5883
- Wang, B. G., Zhang, W. W., Duan, X. J., & Li, X. M. (2009a). *In vitro* antioxidative activities of extract and semi-purified fractions of the marine red alga, *Rhodomela confervoides* (Rhodomelaceae). *Food Chemistry*, 113(4), 1101–1105
- Wang, T., Jónsdóttir, R., & Ólafsdóttir, G. (2009b). Total phenolic compounds, radical scavenging and metal chelation of extracts from Icelandic seaweeds. *Food Chemistry*, 116(1), 240–248
- Watanabe, J., Kawabata, J., Kurihara, H. & Niki, R. (1997) Isolation and Identification of  $\alpha$ -Glucosidase Inhibitors from Tochu-cha (*Eucommia ulmoides*). *Bioscience, Biotechnology, and Biochemistry*, 61(1), 177–178
- Weesepoel, Y., Gruppen, H., de Bruijn, W., & Vincken, J.P. (2014). Analysis of palmitoyl apo-astaxanthinals, apo-astaxanthinones, and their epoxides by UHPLC-PDA-ESI-MS. *Journal of Agricultural and Food Chemistry*, 62(42), 10254–10263
- Wei, Z., Wang, W., Chen, J., Yang, D., Yan, R., & Cai, Q. (2014). A prospective, randomized, controlled study of omega-3 fish oil fat emulsion-based parenteral nutrition for patients following surgical resection of gastric tumors. *Nutrition Journal*, 13(1), 1–6
- Weibel, E. K., Hadvary, P., Hochuli, E., Kupfer, E., & Lengsfeld, H. (1987). Lipstatin, an inhibitor of pancreatic lipase, produced by *Streptomyces toxytricini*. I. Producing organism, fermentation, isolation and biological activity. *The Journal of Antibiotics*, 40(8), 1081–1085
- Weisberg, S. P., McCann, D., Desai, M., Rosenbaum, M., Leibel, R. L., & Ferrante, A. W. J. (2003). Obesity is associated with macrophage accumulation in adipose tissue. *The Journal of Clinical Investigation*, 112(12), 1796–1808
- West, K. M., & Kalbfleisch, J. M. (1966). Glucose Tolerance, Nutrition, and Diabetes in Uruguay, Venezuela, Malaya, and East Pakistan. *Diabetes*, 15(1), 9–18
- Whiting, D. R., Guariguata, L., Weil, C., & Shaw, J. (2011). IDF Diabetes Atlas: Global estimates of the prevalence of diabetes for 2011 and 2030. *Diabetes Research and Clinical Practice*, 94(3), 311–321
- World Health Organization, (1999). Definition, diagnosis and classification of Diabetes

Mellitus and its complications. Part 1: Diagnosis and Classification of Diabetes Mellitus. Report of a WHO Consultation. Geneva: World Health Organization, 1999

Wijesekara, I., Pangestuti, R., & Kim, S.K. (2011). Biological activities and potential health benefits of sulfated polysaccharides derived from marine algae. *Carbohydrate Polymers*, 84(1), 14–21

Wilcox, M. D., Brownlee, I. A., Richardson, J. C., Dettmar, P. W., & Pearson, J. P. (2014). The modulation of pancreatic lipase activity by alginates. *Food Chemistry*, 146, 479–484

Willett, W. C., Koplan, J. P., Nugent, R., Puska, P., & Gaziano, T. A. (2006). Prevention of Chronic Disease by Means of Diet and Lifestyle Changes. *Disease Control Priorities in Developing Countries*, (44), 833–850

Wong, C. L., & Phang, S. M. (2004). Biomass production of two *Sargassum* species at Cape Rachado, Malaysia. *Hydrobiologia*, 512, 79–88

Woo, M. N., Jeon, S. M., Kim, H. J., Lee, M. K., Shin, S. K., Shin, Y. C., Park ,Y.B., & Choi, M. S. (2010). Fucoxanthin supplementation improves plasma and hepatic lipid metabolism and blood glucose concentration in high-fat fed C57BL/6N mice. *Chemico-Biological Interactions*, 186(3), 316–322

Wu, Z., Rosen, E. D., Brun, R., Hauser, S., Adelman, G., Troy, A. E., McKeon, C. Darlington, G.J., & Spiegelman, B. M. (1999). Cross-regulation of C/EBP alpha and PPAR gamma controls the transcriptional pathway of adipogenesis and insulin sensitivity. *Molecular Cell*, 3(2), 151–158

www.who.int, Accessed on 10 January, 2017

Xu, N., Fan, X., Yan, X., Li, X., Niu, R., & Tseng, C. K. (2003). Antibacterial bromophenols from the marine red alga *Rhodomela confervoides*. *Phytochemistry*, 62(8), 1221–1224

Xu, X., Bi, D. C., Li, C., Fang, W. S., Zhou, R., Li, S. M., Chi, L.L., Wan, M., & Shen, L. M. (2015). Morphological and proteomic analyses reveal that unsaturated guluronate oligosaccharide modulates multiple functional pathways in murine macrophage RAW 264.7 cells. *Marine Drugs*, 13(4), 1798–1818

Xu, X., Song, F., Wang, S., Li, S., Xiao, F., Zhao, J., Yang, Y., Shang, S., Yang, L., Shi, J. (2004). Dibenzyl Bromophenols with Diverse Dimerization Patterns from the Brown Alga *Leathesia nana*. *Journal of Natural Products*, 67(10), 1661–1666

Xu, X.L., Fan, X., Song, F.H., Zhao, J.L., Han, L.J., Yang, Y.C., & Shi, J.G. (2004). Bromophenols from the brown alga *Leathesia nana*. *Journal of Asian Natural Products Research*, 6(3), 217–221

Yan, X., Chuda, Y., Suzuki, M., & Nagata, T. (1999). Fucoxanthin as the major antioxidant in *Hijikia fusiformis*, a common edible seaweed. *Bioscience, Biotechnology, and Biochemistry*, 63(3):605-607

- Yang, E.J., Moon, J.Y., Kim, M.J., Kim, D. S., Kim, C.S., Lee, W. J., Lee, N.H., & Hyun, C.G. (2010). Inhibitory effect of Jeju endemic seaweeds on the production of pro-inflammatory mediators in mouse macrophage cell line RAW 264.7. *Journal of Zhejiang University. Science. B*, 11(5), 315–322
- Yang, E.J., Moon, J.Y., Kim, S. S., Yang, K.W., Lee, W. J., Lee, N. H., & Hyun, C.G. (2014). Jeju seaweeds suppress lipopolysaccharide-stimulated proinflammatory response in RAW 264.7 murine macrophages. *Asian Pacific Journal of Tropical Biomedicine*, 4(7), 529–537
- Yang, H., Zeng, M., Dong, S., Liu, Z., & Li, R. (2010). Anti-proliferative activity of phlorotannin extracts from brown algae *Laminaria japonica* Aresch. *Chinese Journal of Oceanology and Limnology*, 28(1), 122–130
- Yang, Z.H., Miyahara, H., & Hatanaka, A. (2011). Chronic administration of palmitoleic acid reduces insulin resistance and hepatic lipid accumulation in KK-Ay Mice with genetic type 2 diabetes. *Lipids in Health and Disease*, 10(1), 120
- Yeh, C.J., Chang, H.Y., & Pan, W.H. (2011). Time trend of obesity, the metabolic syndrome and related dietary pattern in Taiwan: from NAHSIT 1993-1996 to NAHSIT 2005-2008. *Asia Pacific Journal of Clinical Nutrition*, 20(2), 292–300
- Yoon, W.J., Heo, S.J., Han, S.C., Lee, H.J., Kang, G.J., Kang, H.K., Hyun, J.W., Koh, Y.S., & Yoo, E.S. (2012). Anti-inflammatory effect of sargachromanol G isolated from *Sargassum siliquastrum* in RAW 264.7 cells. *Archives of Pharmacal Research*, 35(8), 1421–1430
- Yoon, W., Ham, Y. M., Kim, K., Park, S., Lee, N. H., Hyun, G., & Lee, W. J. (2009). Anti-inflammatory activity of brown alga *Dictyota dichotoma* in murine macrophage RAW 264.7 cells. *Journal of Medicinal Plants Research*, 3(1), 1–8
- Young, A.J., Phillip, D. & Savill, J. (1997). Carotenoids in higher plant photosynthesis. In *Handbook of Photosynthesis*, 575–596. New York, USA
- Yuan, Y.V. (2007). Antioxidants from Edible Seaweeds. In *Antioxidant Measurement and Applications*, 268-301. American Chemical Society.
- Yudkin, J.S., Kumari, M., Humphries, S.E., & Mohamed Ali, V. (2000). Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? *Atherosclerosis*, 148(2), 209–214
- Zemke-White, W.L., & Smith, J.E. (2006) Environmental impacts of seaweed farming in the tropics, In: A.T. Critchley, M. Ohno and D. Largo (eds.) *World Seaweed Resources*. Expert Centre for Taxonomic Identification (ETI), University of Amsterdam (CD-ROM series)
- Zhang, J., Tiller, C., Shen, J., Wang, C., Girouard, G. S., Dennis, D., Barrow, C.J., Miao, M., & Ewart, H. S. (2007). Anti-diabetic properties of polysaccharide- and polyphenolic-enriched fractions from the brown seaweed *Ascophyllum nodosum*. *Canadian Journal of Physiology and Pharmacology*, 85(11), 1116–1123

Zhao, Y., Joshi-Barve, S., Barve, S., & Chen, L. H. (2004). Eicosapentaenoic acid prevents LPS-induced TNF-alpha expression by preventing NF-kappa $\beta$  activation. *Journal of the American College of Nutrition*, 23(1), 71–78

Zimmet, P. Z., (1999). Diabetes epidemiology as a tool to trigger diabetes research and care. *Diabetologia*, 42, 499-518