

UNIVERSITI PUTRA MALAYSIA

IMPROVED REPRODUCTIVE PERFORMANCE THROUGH EDIBLE BIRD'S NEST AND ITS AMELIORATING PROPERTIES IN LEAD ACETATE TOXICITY OF REPRODUCTIVE SYSTEM IN FEMALE RATS

ABDULLA AAID HADI ALBISHTUE

FPV 2018 17



IMPROVED REPRODUCTIVE PERFORMANCE THROUGH EDIBLE BIRD'S NEST AND ITS AMELIORATING PROPERTIES IN LEAD ACETATE TOXICITY OF REPRODUCTIVE SYSTEM IN FEMALE RATS



ABDULLA AAID HADI ALBISHTUE

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfillment of the requirements for the Degree of Doctor of Philosophy

May 2018

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to my beloved parents; Aaid Hadi and Hameedah Sabr, my wife Hala Abdulrazzaq and my children; Morsaleen, Baqer and Sajjad.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

IMPROVED REPRODUCTIVE PERFORMANCE THROUGH EDIBLE BIRD'S NEST AND ITS AMELIORATING PROPERTIES IN LEAD ACETATE TOXICITY OF REPRODUCTIVE SYSTEM IN FEMALE RATS

By

ABDULLA AAID HADI ALBISHTUE

May 2018

Chairman Faculty Nurhusien Yimer Degu, PhD Veterinary Medicine

Edible Bird's Nest (EBN) is an animal product from the salivary secretion of male swiftlet birds (*Aerodramus fuciphagus and Aerodramus maximus*). It is traditionally consumed Asians for its nutritional and medicinal values. Although enhancing reproductive functions is among the traditionally claimed benefits of consuming EBN there is a dearth of scientific evidence in this regard. The aims of this study were to determine the effects of EBN supplement on the reproductive functions of cycling female rats, on pregnant female rats, and subsequently to evaluate the ameliorating effect of EBN supplement against toxic effect of lead acetate (LA) to the female reproductive system and pituitary gland.

To address the first objective of this study (evaluation of the effects of EBN supplement on the ovarian, uterine and pituitary gland activities of cycling female rats), histomorphometric analysis and assessment of expressions of epidermal growth factor (EGF), its receptor (REGF), proliferating cell nuclear antigen (PCNA), vascular endothelial growth factor (VEGF), steroid receptors on ovaries and uteri as well as measurement of steroid hormones, prolactin (P) and growth hormone (GH) from plasma, were employed. Twenty four Sprague Dawley rats were divided into 4 equal groups (n=6): G1 as control group while G2, G3 and G4 were treated groups with EBN at graded concentrations of 30, 60 and 120 mg/kg of body weight per day respectively for 8 weeks. The EBN was administered orally using gavage tube. At the proestrus stage all rats were sacrificed to remove ovaries, uteri, pituitary glands and liver for histological and immunohistochemical analyses. Results showed significant ovarian structural and histological changes such as numbers of interstitial cells and growing follicles in EBN treated groups, with significant increase in endocrine cells and vascularization of pars distalis of the pituitary glands as well as increased uterine

 \bigcirc

epithelium and number of uterine glands. There was no histomorphological change of liver among groups. Samples from G3 and G4 demonstrated significant expressions of EGF on ovarian surface epithelium, interstitial cells, uterine surface epithelium and uterine stromal cells as well as higher expressions of PCNA and VEGF, and estrogen receptor (E_2R) compared with G1 and G2. No staining for progesterone receptor (P_4R) was observed in the treated groups. In addition, immunohistochemistry of the uterus showed significantly higher expressions of EGF, REGF, PCNA, E_2R and P_4R (p < 0.05) in G4. The plasma levels of estrogen (E_2) (ng/mL) and progesterone (P_4) (ng/mL) in G4 (18000±1786; 168±17) were significantly higher than G3 (11000±3670; 84.04±9.56), G2 (6300±1566; 63.66±9.06), and G1 (1100±143; 50.03±4.18). Moreover, concentrations of prolactin P and GH were observed to be significantly higher (p < 0.05) in G4. These findings suggest that EBN supplement enhances ovarian follicular growth, uterine structures, expressions of E_2R , P_4R EGF, REGF, VEGF, and PCNA and subsequent rise in plasma E₂, P₄, P and GH levels. These imply the strong enhancing effect of EBN on the reproductive system of cycling female rats.

In order to determine effect of EBN supplement on embryo implantation rate and associated changes in the uterus, plasma steroids and oxidative stress biomarkers (2nd objective), a total 24 female adult rats underwent similar treatment for 8 weeks. In the last week of treatment, however, intact fertile male rats were introduced into each group (three per group) with proestrous stage overnight for mating. On day 7 postmating (expected days of implantation; peri- and post-implantation), the animals were sacrificed for assessment of implantation rate, histological and electron microscopic examination of the uterus, oxidative stress biomarkers (OSB) and antioxidant (AO) assay, GH, P, steroid hormones analysis, and expressions of steroid receptors, EGF, REGF, PCNA, and VEGF on the uterus. Results showed that as the concentrations of EBN increases, the pregnancy rate, embryonic implantations rate and development of microvilli with pinopodes in uterine epithelium were also increased. There was an increased level of superoxide dismutase (SOD) and total antioxidant capacity (TAC) (p < 0.05) in the G4, with lower (p < 0.05) concentrations of thiobarbituric acid reactive substance (TBARS) compared to control. All results of the hormones assay and immunohistochemistry showed significantly higher concentrations and expressions of steroid receptors, EGF and REGF, PCNA, and VEGF (p < 0.05) in G4 compared to the other groups. These findings imply that almost all the factors important for embryo implantation and development are enhanced in concentrations and expressions with EBN supplement in a dose dependant manner subsequently resulting in increased embryo-implantation and pregnancy rate.

 \bigcirc

The last objective in the present study was to evaluate the protective effect of EBN supplement to the reproductive system (ovaries and uteri) and pituitary glands of female rats against lead acetate (LA) toxicity. LA is a toxic compound that has harmful effects on the female reproductive system such as altered uterine and ovarian histology, size and function, and low oestrogen production. There were five treatment groups: Group 1 - control (C) was given normal saline, group 2 (T0) was administered with LA (10 mg/kg bwt), while groups 3 (T1), 4 (T2) and 5 (T3) were given LA (10

mg/kg bwt) and graded concentrations of 30, 60, and 120 mg/kg bwt of EBN, respectively. Rats were euthanized at day 30 for collection of blood plasma, ovary and uterus. Organ tissues were fixed in 10 % buffered formalin and subjected to histological analyses and immunohistochemistry for expression of steroid receptors, EGF and REGF, PCNA, and VEGF on the ovary and uterus. Plasma was used to determine concentrations of E2, oxidative stress biomarker (OSB) and antioxidant (AO). Results showed that the level of E_2 was lower (p < 0.05) in the T0 group while the T3 group had the highest E2 concentration. There was a decreased level of SOD and TAC (p < 0.05) in the T0 group and an increased SOD and TAC level in the T3 group, while T0 had higher (p < 0.05) concentrations of TBARS compared to treated groups, indicating oxidative stress. There was a reduced number of primordial follicles and increased numbers of atretic follicles in the ovary as well as significant damage in the uterus as evidenced by reduction in uterine glands and decrease in height of columnar cells in the T0 group compared with the treatment groups. Moreover, histological examination of pituitary gland of LA exposed rats without EBN supplement showed degenerative changes in endocrine cells of pars distalis such as non-uniform arrangement of the cells and decrease in cell number and size. Interestingly, histological examination of pituitary glands, ovaries and uteri of EBN treated groups showed significant protection as evidenced by a significant increase in endocrine cells of pars distalis, growing follicles and CL, but decrease in number of atretic follicles on ovaries as well as increase in uterine glands and height of columnar cells. All results of immunohistochemistry showed significantly higher expression of steroid receptors, EGF, PCNA, and VEGF (p < 0.05) in T3 compared to other groups. This part of the experiment reaffirmed the detrimental effects of LA on the reproductive system and revealed novel findings on the ameliorating effect of the oral supplementation of EBN against LA toxicity damage to the reproductive system, achieved best at 120mg/kg body weight.

In general, this study suggests that EBN supplement enhances functions of the pituitary gland, ovary, uterus, expression of steroid receptors, EGF, REGF, VEGF, and PCNA on ovaries and uteri along with a rise in serum E_2 , P_4 , P and GH levels. Moreover, EBN has showed a promoting effect on fertility indexes by increasing pregnancy and embryo implantation rates. Furthermore, the present study revealed that EBN supplement at oral dose of 60 - 120mg/kg body weight is capable of protecting and preventing alterations in the pituitary gland and the reproductive system due to lead toxicity through an integrated mechanism of maintaining antioxidant – reactive oxygen species (ROS) balance. Overall findings of the present study provide scientific evidence in support of the traditional claim of EBN's benefit to reproduction and being one of the reasons for consumption among humans.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PRESTASI PEMBIAKAN TERTINGKAT MELALUI PENGGUNAAN SANGKAR BURUNG BOLEH MAKAN DAN CIRI PEMBAIKANNYA DALAM KETOKSIKAN PLUMBUM ASETAT SISTEM PEMBIAKAN TIKUS BETINA

Oleh

ABDULLA AAID HADI ALBISHTUE

Mei 2018

Pengerusi: Nurhusien Yimer Degu, PhDFakulti: Perubatan Veterinar

Sarang burung yang boleh dimakan (EBN) merupakan produk haiwan dari rembesan air liur burung walit jantan (*Aerodramus fuciphagus* dan *Aerodramus maximus*). Ianya secara tradisi dimakan oleh orang Asia kerana nilai pemakanan dan perubatannya. Walaupun meningkatkan fungsi pembiakan adalah antara dakwaan manfaat tradisional memakan EBN, terdapat kekurangan bukti saintifik di dalam hal ini. Tujuan kajian ini adalah untuk menentukan kesan penambahan EBN ke atas fungsi pembiakan tikus betina dalam kitaran, tikus betina hamil, dan seterusnya menilai kesan pembaikan EBN terhadap kerosakan sistem pembiakan betina dan kelenjar pituitari yang disebabkan oleh toksik plumbum asetat (LA).

Bagi mencapai objektif pertama kajian ini (i.e penilaian kesan penambahan EBN ke atas aktiviti ovari, uterus dan kelenjar pituitari tikus betina dalam kitaran), analisis histomorfometri dan penilaian ekspresi faktor pertumbuhan epidermis (EGF), reseptornya (REGF), antigen nuklear sel membiak (PCNA), faktor pertumbuhan endotelium vaskular (VEGF), reseptor steroid pada ovari dan uterus serta pengukuran hormon steroid, prolaktin (P) dan hormon pertumbuhan (GH) dari plasma telah dijalankan. Dua puluh empat ekor tikus Sprague Dawley telah dibahagikan kepada 4 kumpulan (n=5): G1 adalah kumpulan kawalan yang tidak dirawat, manakala G2, G3 dan G4 adalah kumpulan yang dirawat dengan EBN pada kadar kepekatan yang berbeza masing-masing sebanyak 30, 60 dan 120 mg/kg berat badan setiap hari selama 8 minggu. Pemberian EBN adalah melalui mulut degan menggunakan tiub gavaj. Pada peringkat proestrus, semua tikus telah dikorbankan untuk mengeluarkan ovari, uterus, kelenjar pituitari dan hati untuk analisis histologi dan imunohistokimia. Hasil kajian menunjukkan perubahan struktur dan histologi ovari yang signifikan seperti jumlah



sel celahan dan folikel bertumbuhan dalam kumpulan yang dirawat dengan EBN, ditambah pula dengan peningkatan yang ketara bagi sel gonadotrof dan vaskularisasi kelenjar pituitari serta peningkatan epitelium uterus dan bilangan kelenjar uterus. Tiada perubahan histomorfologi bagi hati dalam semua kumpulan. Sampel dari G3 dan G4 menunjukkan ekspresi EGF yang ketara pada epitelium permukaan ovari, selsel celahan, epitelium permukaan uterus dan sel-sel stroma uterus serta ekspresi PCNA dan VEGF dan reseptor estrogen (E2R) yang lebih tinggi berbanding dengan G1 dan G2. Tiada pewarnaan untuk reseptor progesteron (P4R) diperhatikan dalam kumpulan yang dirawat kecuali pewarnaan minimum bagi epitelium ovari dalam G3 dan G4. Di samping itu, kajian imunohistokimia uterus menunjukkan ekspresi EGF, REGF, PCNA, E2R dan P4R yang lebih tinggi (p < 0.05) di dalam G4. Kadar plasma estrogen (E2) (ng/mL) dan progesteron (P4) (ng/mL) di dalam G4 (18000±1786; 168±17) lebih tinggi (p < 0.05) daripada G3 (11000±3670; 84.04±9.56), G2 (6300±1566; 63.66±9.06), dan G1 (1100±143; 50.03±4.18). Selain itu, kepekatan prolaktin P dan GH juga lebih tinggi (p < 0.05) dalam G4. Hasil kajian ini mencadangkan bahawa penambahan EBN meningkatkan pertumbuhan folikel ovari, struktur uterus, ekspresi ER, EGF, REGF, VEGF dan PCNA dan seterusnya kenaikan paras plasma E2, P4, P dan GH. Ini menunjukkan EBN mempunyai kesan peningkatan yang kuat ke atas sistem pembiakan tikus betina yang mengalami kitaran

Untuk menentukan kesan EBN ke atas kadar implantasi embrio dan perubahan yang berkaitan di dalam uterus, steroid plasma dan biomarker tekanan oksidatif (objektif ke 2), sejumlah 24 ekor tikus betina dewasa menjalani rawatan yang sama untuk 8 minggu. Walaubagaimanapun, pada minggu terakhir rawatan, tikus jantan yang subur telah dimasukkan ke dalam setiap kumpulan (tiga ekor setiap kumpulan) di peringkat proestrus semalaman untuk mengawan. Pada hari ke 6 dan 7 selepas mengawan (hari yang dijangka berlaku implantasi; semasa- dan pasca-implantasi), haiwan-haiwan tersebut dikorbankan untuk penilaian kadar implantasi, pemeriksaan histologi dan mikroskop elektron pada uterus, biomarker tekanan oksidatif (OSB) dan cerakin antioksidan (AO), GH, PR, analisis hormon steroid, dan ekspresi reseptor steroid, EGF, REGF, PCNA dan VEGF pada uterus. Hasil kajian menunjukkan apabila kepekatan EBN meningkat, kadar kehamilan, kadar implantatasi embrio dan pembangunan mikrovilus dengan pinopod dalam epitelium uterus juga meningkat. Terdapat peningkatan paras superoksida dismutase (SOD) dan jumlah kapasiti antioksidan (TAC) (p < 0.05) di dalam G4, dan lebih rendah (p < 0.05) kepekatan bahan reaktif asid tiobarbiturik (TBARS) berbanding dengan kawalan. Semua keputusan cerakin hormon dan imunohistokimia menunjukkan kepekatan dan ekspresi reseptor steroid EGF dan REGF, PCNA dan VEGF yang lebih tinggi (p < 0.05) dalam G4 berbanding dengan kumpulan lain. Keputusan ini memberi implikasi bahawa hampir semua faktor yang penting untuk implantasi dan pembangunan embrio telah dipertingkatkan dalam kepekatan dan ekspresi dengan penambahan EBN yang seterusnya akan menyebabkan peningkatan implantasi-embrio dan kadar kehamilan. Objektif terakhir di dalam kajian ini adalah untuk menilai kesan perlindungan penambahan EBN ke atas sistem pembiakan (ovari dan uterus) dan kelenjar pituitari tikus betina terhadap ketoksikan LA. Plumbum asetat adalah sebatian toksik yang mempunyai kesan buruk terhadap sistem pembiakan betina seperti perubahan histologi uterus dan ovari, saiz dan fungsi, dan pengeluaran estrogen yang rendah.

Terdapat 5 kumpulan rawatan: Kumpulan 1 - kawalan (C) diberikan salina normal, kumpulan 2 (T0) telah diberi LA (10 mg/kg bwt), manakala kumpulan 3 (T1), 4 (T2) dan 5 (T3) diberikan LA (10mg/kg bwt) dan kepekatan EBN yang berbeza masingmasing sebanyak 30, 60, dan 120 mg/kg bwt. Semua tikus telah dimatikan pada hari ke 30 untuk pengambilan plasma darah, ovari dan uterus. Tisu organ telah diawetkan didalam 10% formalin berpenimbal untuk analisis histologi dan imunohistokimia bagi memeriksa ekspresi reseptor steroid, EGF dan REGF, PCNA dan VEGF pada ovari dan uterus. Plasma darah digunakan untuk menentukan kepekatan E2, OSB dan AO. Hasil kajian menunjukkan bahawa tahap E2 adalah lebih rendah (p < 0.05) dalam kumpulan T0 manakala kumpulan T3 mempunyai kepekatan E2 tertinggi. Terdapat penunurun tahap SOD dan TAC (p < 0.05) dalam kumpulan T0 dan peningkatan tahap TAC dalam kumpulan T3, manakala T0 mempunyai kepekatan TBARS lebih tinggi (p < 0.05) berbanding kumpulan dirawat, menunjukkan tekanan oksidatif. Terdapat pengurangan jumlah folikel primordium dan peningkatan bilangan folikel beratresia di dalam ovari serta kerosakan ketara di dalam uterus seperti yang dibuktikan oleh pengurangan kelenjar uterus dan pengurangan ketinggian sel turus di dalam kumpulan T0 berbanding dengan kumpulan rawatan. Selain itu, pemeriksaan histologi kelenjar pituitari tikus yang terdedah kepada LA tanpa penambahan EBN menunjukkan perubahan degeneratif di dalam sel endokrin pars distalis. Menariknya, pemeriksaan histologi kelenjar pituitari, ovari dan uterus kumpulan dirawat dengan EBN menunjukkan perlindungan yang signifikan seperti yang dibuktikan oleh peningkatan ketara sel gonadotrof, folikel yang membesar dan CL, tetapi bilangan folikel beratresia pada ovari serta peningkatan dalam kelenjar uterus dan ketinggian sel-sel turus adalah menurun. Semua keputusan imunohistokimia menunjukkan ekspresi yang lebih tinggi (p < 0.05) bagi reseptor steroid, EGF, PCNA dan VEGF dalam T3 berbanding kumpulan lain. Eksperimen ini mengesahkan kesan buruk LA ke atas sistem pembiakan dan mendedahkan penemuan baru kesan peningkatan penambahan EBN melalui mulut terhadap kerosakan yang disebabkan oleh ketoksikan LA kepada sistem pembiakan, dimana pencapaian terbaik adalah pada kadar 120mg/kg berat badan.

Secara umum, kajian ini menunjukkan bahawa penambahan EBN meningkatkan fungsi kelenhar pituitary, ovari, uterus, ekspresi reseptor steroid, EGF, REGF, VEGF dan PCNA ke atas ovari dan uterus bersama dengan peningkatan tahap serum E2, P4, P dan GH. Tambahan lagi, EBN telah menunjukkan kesan penggalakan pada indeks kesuburan dengan meningkatkan kadar kehamilan dan implantasi embrio. Juga, kajian ini menunjukkan bahawa penambahan EBN pada dos mulut 60 - 120mg/kg berat badan mampu melindungi dan mencegah perubahan dalam kelenjar pituitari dan sistem pembiakan akibat ketoksikan plumbum melalui suatu mekanisme bersepadu yang mengekalkan keseimbangan antioksidan-spesies oksigen reaktif (ROS). Dapatan keseluruhannya kajian ini menyediakan bukti saintifik yang menyokong tuntutan tradisional tentang manfaat EBN terhadap pembiakan yang menjadi salah satu sebab penggunaannya di kalangan manusia.

ACKNOWLEDGEMENTS

First of all, I want to thank the God Almighty for his grace and divine enablement to complete this work. Secondly, I wish to express my sincere gratitude to the following people without whom this study would not have become a reality:

My utmost gratitude goes to my supervisor Dr. Nurhusien Yimer for his generous advice and ongoing assistance and encouragement throughout the duration of this work. May God bless you richly.

The members of my supervisory committee, Professor Dr. Abd Wahid Haron, Professor. Dr. Md. Zuki Bin Abu Bakar and the late Associate Professor Dr. Rosnina Bt Hj. Yusoff for all their guidance and support during this study. Truly, they have been my guardian angels!

My father, mother, brothers and sisters, thank you all for the support that gave me throughtout this journey. Special thanks goes to my family: wife and children, you stand by me with all you have got in all situations 'under the rain and in the sun' giving me hope and motivations.may Allah reward you abundantly.

The Ministry of Higher Education and Scientific Research, Iraq for selecting me as a scholarly candidate and funding my study here in Malaysia. Acknowledgement also goes to Department of Anatomy and Histology, Faculty of Veterinary Medicine, University of Kufa, Najaf, Iraq for granting me leave during the tenure of my study. I would also like to express my appreciation to the academic, technical staff who contributed immensely to the success of this great accomplishment: Dr. Bahaa Almahanawi, Dr. Adamu Abdul Abubakar, Dr. Kareem Hendol, Mr. Jamil, Mrs. Jimilah and Mrs. Latifah.

Lastly, I am not forgetting, Professor Dr. Abdul Salam Babji for providing edible bird's nest to us during experimental period. Almighty God rewards you abundantly.

I certify that a Thesis Examination Committee has met on 7 May 2018 to conduct the final examination of Abdulla Aaid Hadi Albishtue on his thesis entitled "Improved Reproductive Performance Through Edible Bird's Nest and its Ameliorating Properties in Lead Acetate Toxicity of Reproductive System in Female Rats" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Abdul Aziz bin Saharee, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Faez Firdaus Jesse bin Abdullah, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Tengku Azmi bin Tengku Ibrahim, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Internal Examiner)

Mohamad Agus Setiadi, PhD

Professor Bogor Agricultural University Indonesia (External Examiner)

RUSLI HAJI ABDULLAH, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 30 July 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Nurhusien Yimer Degu, PhD

Senior Lecturer Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Abd Wahid Haron, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

Md Zuki Abu Bakar, PhD

Professor Faculty of veterinary Medicine Universiti Putra Malaysia (Member)

Rosnina bt Hj. Yusoff, PhD

Associate Professor Faculty of veterinary Medicine Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:	Date:

Name and Matric No: Abdulla Aaid Hadi, GS42373

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: Name of Chairman of Supervisory Committee:	Dr. Nurhusien Yimer Degu
Signature: Name of Member of Supervisory Committee:	Professor Dr. Abd Wahid Haron
Signature: Name of Member of Supervisory Committee:	Professor Dr. Md Zuki Abu Bakar
Signature: Name of Member of Supervisory Committee:	Associate Professor Dr. Rosnina bt Hj.Yusoff

TABLE OF CONTENTS

Page

ABS ACK APP DEC LIST LIST	ROVA CLARA F OF TA F OF FI	LEDGEMENTS L TION ABLES IGURES	i iv vii viii xiii xvii xix xxiii
CHA	APTER		
1	INTI	RODUCTION	1
2	LITE	ERATURE REVIEW	3
-	2.1	The reproductive anatomy and physiology of female rats	3
		2.1.1 Estrus cycle	4
		2.1.2 Mating	
		2.1.3 Pregnancy detection	5 5 5 5
		2.1.4 Fertilization and embryonic development	5
	2.2	Embryonic implantation	
		2.2.1 Uterine changes during implantation	6
		2.2.2 Uterine luminal closure for blastocyst apposition	7
		2.2.3 Implantation and steroid hormones	7
		2.2.4 Epidermal growth factors	8
		2.2.5 Vascular endothelial growth factor	9
		2.2.6 Cytokines	10
	2.3	2.2.7 Basic fibroblast growth factor Edible bird's nest	10 11
	2.3	2.3.1 Nutritional composition	13
		2.3.2 Bioactive compounds	13
		2.3.2.1 Effect of hormonal compositions of edible	15
		bird's nest on reproductive system and fertility	
		r in the state of	14
		2.3.2.2 Role of edible bird's nest on proliferating and	
		treatments of diseases	14
	2.4	Lead acetate toxicity	17
		2.4.1 Lead acetate effect on female reproductive system	18
		2.4.2 Effects of lead acetate on pituitary glands	20
	2.5	Liver	21

GENI	ERAL MATERIALS AND METHODS
3.1	Animals and experimental design
3.2	Preparation of edible bird's nest
3.3	Preparation of lead acetate solution
3.4	Determination of estrous cycle phases and synchronization
3.5	Ovary and pituitary tissues sampling and macroscopic and
	microscopic examination
3.6	Macroscopic and microscopic examinations of the uterus and
	liver
3.7	Measurement of ovarian or uterine weight: body weight ratio
3.8	Immunohistochemistry analyses of growth factors and their
	receptors, proliferating cell nuclear antigen and steroid receptors
	in ovarian and uterine tissues
3.9	Blood plasma collection and hormone assays
3.10	Oxidative stress biomarkers and antioxidant assay
	Statistical analysis

EDID	DLE DIRD'S NEST SUPPLEMENT WODULATES RATS	
HYP	OTHALAMIC-PITUITARY-OVARIAN AXIS AND	
EXPI	RESSIONS OF EPIDERMAL GROWTH FACTOR,	
VASC	CULAR ENDOTHELIAL GROWTH FACTOR AND	
PRO	LIFERATING CELL NUCLEAR ANTIGEN ON OVARY	29
4.1	Introduction	29
4.2	Materials and methods	30
	4.2.1 Animals and experimental design	30
	4.2.2 Statistical analysis	31
4.3	Results	31
	4.3.1 Effects of edible bird's nest treatment on ovarian	
	histomorphology and follicular development	31
	4.3.2 Effects of edible bird's nest treatment on	
	histomorphology of pituitary gland	34
	4.3.3 Expressions of epidermal growth factor, proliferating cell	
	nuclear antigen and vascular endothelial growth factor in	
	the ovary	36
	4.3.4 Expression of estrogen receptors in ovarian tissues	40
4.4	Discussion	42
4.5	Conclusion	44

5

EFFECT OF EDIBLE BIRD'S NEST ON UTERUS AND PLASMA CONCENTRATIONS OF STEROIDS, GROWTH HORMONES AND PROLACTIN OF CYCLING FEMALE RATS 5.1 Introduction

RAT	S		45
5.1	Intro	duction	45
5.2	Mate	Materials and methods	
	5.2.1	Animals and experimental design	46
	5.2.2	Statistical analysis	47

5.3	Result	ts	48
	5.3.1	Effect edible bird's nest on vaginal cytology	48
	5.3.2		
		uterine body weight ratio	50
	5.3.3		52
	5.3.4		53
	5.3.5		
		epidermal growth factor, proliferating cell nuclear	
		antigen, and vascular endothelial growth factor in the	
		uterus	55
	5.3.6		60
		Hormonal concentrations	63
		Oxidative stress biomarker and antioxidant assay	64
5.4	Discu		65
5.5	Concl		68
EFFI	ECTS O	F EBN SUPPLEMENTATION ON EMBRYO	
		TION, GENE EPRESSIONS, REPRODUCTIVE	
		S AND OXIDATIVE STRESS IN RATS	69
6.1		luction	69
6.2	Mater	ials and methods	71
		Animals and experimental design	71
	6.2.2		
		uterine tissues	72
	6.2.3		72
6.3	Result		73
		Effect of EDN on body weight overion and utering	

6.2.3	Statistical analysis	72
Result	S	73
6.3.1	Effect of EBN on body weight, ovarian and uterine	
	body weight ratios	73
6.3.2	Uterine examinations using light microscope and	
	scanning electronic microscope	76
6.3.3	Fertility index and embryo implantation rate	79

6.3.3 Fertility index and embryo implantation rate 6.3 c

3.4	Expressions of epidermal growth factor, receptor of
	epidermal growth factor, proliferating cell nuclear
	antigen, and vascular endothelial growth factor in the
	uterus
35	Expression of staroid hormone recentors in utaring

81

6.3.5 Expression of steroid hormone receptors in uterine tissues 87 6.3.6 Hormonal concentrations 90

6.3.7 Oxidative stress biomarkers and Antioxidant assay	91
Discussion	93
Conclusion	95

6.5 Conclusion

6.4

6

7

9

AMELIORATING EFFECT OF EDIBLE BIRD'S NEST AGAINST LEAD ACETATE TOXICITY ON RATS' OVARIES AND PITUITARY GLANDS

7.1	Introd	uction	96
7.2	Materi	als and Methods	97
	7.2.1	Animals and experimental design	97
	7.2.2	Statistical analysis	97
7.3	Result	S	98
	7.3.1	Effects of edible bird's nest treatment on ovarian	
		histomorphology and follicular development	98
	7.3.2	Histopathological findings in the female pituitary glands	103
	7.3.3	Expressions of epidermal growth factor and vascular	
		endothelial growth factor in the ovary	105
	7.3.4	Treatment effect of edible bird's nest on hormones	
		concentrations	109
7.4	Discus	ssion	110
7.5	Conch	ision	113

96

8 PROPHYLACTIC EFFECT OF EDIBLE BIRD'S NEST AGAINST LEAD ACETATE TOXICITY INDUCED UTERINE CHANGES

CHA	NGES		114
8.1	Introd	luction	114
8.2	Mater	ials and methods	115
	8.2.1	Animals and experimental design	115
	8.2.2	Statistical analysis	115
8.3	Result	ts	116
	8.3.1	Effect of edible bird's nest on estrus cycle, body weight	
		and uterine to body weight ratio of rats that were	
		exposed to lead acetate	116
	8.3.2	Effect of edible bird's nest on histopathological findings	
		in the uteri of rats that were exposured to lead acetate	119
	8.3.3		
		proliferating cell nuclear antigen, and vascular	
		endothelial growth factor in the uteri of rats that were	
		exposured to lead acetate	121
	8.3.4	1 1	
		uterine tissues of rats that were exposured to lead acetate	125
		Oxidative stress biomarker and antioxidant assay	128
8.4	Discus		129
8.5	Concl	usion	133
~~~			
		DISCUSION, CONCLUSION AND	10.1
RH(C	' AN MAR	TNDATIONS	13/

9.1	General discusion	134	
9.2	General conclusion	141	
9.3	Recommendations	143	

REFERENCES	144
APPENDICES	167
BIODATA OF STUDENT	175
LIST OF PUBLICATIONS	176



# LIST OF TABLES

Table		Page
4.1	Animal grouping and treatment regime of EBN administered by gavage tube	30
4.2	Morphometric parameters of the rat ovaries measured during the pro- estrus stage of the estrous cycle	34
4.3	Histomorphometric parameters in the pituitary glands of rats assessed at the pro-estrus stage of the estrous cycle	35
4.4	Expressions of EGF, VEGF and PCNA in the OSE and interstitial cells of ovary of rats treated with different doses of EBN and sacrificed at pro-estrus stage of the estrous cycle	40
4.5	Evaluation of $E_2R$ expression on rat ovary as measured during sacrificed at sacrificed at pro-estrus stage of the estrous cycle	42
5.1	Histomorphometric parameters evaluated in the rat uterus, measured during the proestrus phase	53
5.2	Histomorphometric parameters in the liver of rats assessed at the pro- estrus stage of the estrous cycle	55
5.3	Expressions of EGF, REGF, VEGF and PCNA in the LE, GE and stromal cells of uteri of rats treated with different doses of EBN	60
5.4	Expressions of $E_2R$ and $P_4R$ in the LE, GE and stromal cells of uterus of rats treated with different doses of EBN and sacrificed at seventh day of suspected pregnancy	63
6.1	Expressions of EGF, REGF, VEGF and PCNA in the LE, GE and stromal cells of uteri of rats treated with different doses of EBN.	86
6.2	Expressions of $E_2R$ and $P_4R$ in the LE, GE and stromal cells of uterus of rats treated with different doses of EBN	90
7.1	Animal grouping and toxicity of LA and treatment regime of EBN administered by gavage tube	97
7.2	Effects of EBN on morphometric parameters assessed in the rat LA exposed ovaries of rats, measured during the pro estrus stage of the estrous cycle	103

7.3	Effects of EBN on histomorphometric parameters in LA exposed the pituitary glands of rats assessed at the pro-estrus stage of the estrous cycle	105
7.4	Expressions of EGF in the ovary of rats treated with different doses of EBN and sacrificed at pro-estrus stage of the estrous cycle	107
7.5	Expressions of VEGF in the ovary of rats treated with different doses of EB and sacrificed at pro-estrus stage of the estrous cycle	109
8.1	Histomorphometric parameters evaluated in the rat and uterus, measured during the proestrus phase	121
8.2	Expressions of EGF, REGF, VEGF and PCNA in the LE, GE and stromal cells of uteri of rats treated with different doses of EBN after exposure LA	125
8.3	Expressions of E2R and P4R in the LE, GE and stromal cells of uteri of rats treated with different doses of EBN after exposure LA	128

C

# LIST OF FIGURES

Figure		Page
2.1	Female rat reproductive tract across the hormonal cycle and plan for dissection of tissue	3
2.2	Ovarian sections from the female normal rat	4
2.3	Flow chart of swiftlet farming activities	12
3.1	Morphometric parameters of the rat ovaries measured during the pro- estrus stage of the estrous cycle	25
3.2	Histological section of rat ovary. Ovarian surface epithelium and interstitial cells	25
3.3	Lightmicrographs of histological sections of adult rat pituitary glands	26
4.1	Histological section of rat ovary showing follicles at different stages	31
4.2	Histological sections of rat ovaries showing cuboidal type of epithelium in all groups	32
4.3	Comparative assessment of ovarian follicular development and corpora lutea in the different groups of rats	33
4.4	Photomicrographs of histological sections of adult rat pituitary glands treated with different doses of EBN supplement	35
4.5	Photomicrograph sections of the ovaries of rats of different experimental groups (G1, G2, G3 and G4) treated with different doses of EBN showing expressions of EGF	37
4.6	Photomicrograph sections of the ovaries of rats of different experimental groups (G1, G2, G3 and G4) treated with different doses of EBN showing expressions of VEGF	38
4.7	Photomicrograph sections of the ovaries of rats of different experimental groups (G1, G2, G3 and G4) treated with different doses of EBN showing expressions of PCNA	39
4.8	Photomicrograph sections of rat ovaries of different experimental groups (G1, G2, G3 and G4). First group (E2R) shows higher expression in G3 and G4	41
5.1	Histological section of rat uterus	47

5.2	Stages of the estrous cycle in the rat as monitored by vaginal cytology	
5.3	Effect of EBN on body weight of non-pregnant rats	51
5.4	Effect of EBN on ovarian and uterine body weight ratio and length change of non-pregnant rats	52
5.5	Photomicrograph reveals that histological structures of rat liver at the 8 th week after treatment of rats with different dosages of EBN	54
5.6	Photomicrograph sections of the uteri of rats of different experimental groups (G1, G2, G3 and G4) treated with different doses of EBN showing expressions of EGF	56
5.7	Photomicrograph sections of the uteri of rats of different experimental groups (G1, G2, G3 and G4) treated with different doses of EBN showing expressions of REGF	57
5.8	Photomicrograph sections of the uteri of rats of different experimental groups (G1, G2, G3 and G4) treated with different doses of EBN showing expressions of VEGF	58
5.9	Photomicrograph sections of the uteri of rats of different experimental groups (G1, G2, G3 and G4) treated with different doses of EBN showing expressions of PCNA	59
5.10	Photomicrograph sections of rat uteri of different experimental groups (G1, G2, G3 and G4)	61
5.11	Photomicrograph sections of rat uteri of different experimental groups (G1, G2, G3 and G4)	62
5.12	Plasma concentrations of estradiol hormone (A), progesterone hormone (B), growth hormone (C) and prolactin (D) in rats supplemented with different doses of EBN	64
5.13	Effect of EBN on oxidative stress and antioxidant biomarkers in plasma	65
6.1	(A) Vaginal plug . Vaginal smear at day 1 pregnancy	72
6.2	Effect of EBN on body weight of rats	74
6.3	Effect of EBN on ovarian and uterine to body weight ratio of pregnant rats	75
6.4	H and E stain of histological sections of rat gravid uteri	77

6.5	Scanning electronic microscope of histological sections of uteri of pregnant rats	
6.6	Representative photograph of implanted blastocysts (black arrow) in uteri	79
6.7	Effect of EBN on Percentage of fertility index and number embryo implantation sites (EIS)	80
6.8	Photomicrograph sections of the uteri of rats of different experimental groups (G1, G2, G3 and G4) treated with different doses of EBN showing expressions of EGF	82
6.9	Photomicrograph sections of the uteri of rats of different experimental groups treated with different doses of EBN showing expressions of REGF	83
6.10	Photomicrograph sections of the uteri of rats of different experimental groups treated with different doses of EBN showing expressions of VEGF	84
6.11	Photomicrograph sections of the uteri of rats of different experimental groups treated with different doses of EBN showing expressions of PCNA	85
6.12	Photomicrograph sections of rat uteri of different experimental groups (G1, G2, G3 and G4). Estrogen receptor was observed in all groups with highest expression in G4	88
6.13	Photomicrograph sections of rat uteri of different experimental groups (G1, G2, G3 and G4) showing P4R expressions. Higher expression of P4R in G3 and G4	89
6.14	Plasma concentrations of estradiol hormone, progesterone hormone growth hormone and prolactin in rats supplemented with different doses of EBN	91
6.15	Effect of EBN on oxidative stress and antioxidant biomarkers in plasma	92
7.1	Gross appearance of rat ovary is grape-like structure. Gross appearance of pituitary gland	98
7.2	Effect of EBN on ovarian- body weight ratio of rats exposed to LA toxicity	99
7.3	Histological section of rat ovary showing follicles at different stages	99

7.4	Histologic sections from LA exposed ovaries rat showing cuboidal type of epithelium in all groups	100
7.5	Effects of EBN on follicular development after female rats exposed LA toxicity	102
7.6	Effects of EBN on histopathological sections from LA exposed pituitary glands of female rats	104
7.7	Photomicrograph sections of LA exposed the ovaries of rats of different experimental groups (C, T0, T1, T2 and T3) treated with different doses of EBN showing expression of EGF	106
7.8	Photomicrograph sections of LA exposed the ovaries of rats of different experimental groups (C, T0, T1, T2 and T3) treated with different doses of EBN showing expression of VEGF	108
7.9	Effect of EBN on concentration of estradiol hormone, progesterone hormone growth hormone and prolactin hormones in plasma of LA exposed rats	110
8.1	Effect of EBN on body weight of rats exposed to LA toxicity	117
8.2	Effect of EBN on uterine-body weight ratio and length of rats exposed to LA toxicity	118
8.3	Histologic sections from adult rat uteri exposed to LA	120
8.4	Histologic sections from adult rat uteri exposed to LA	122
8.5	Histologic sections from adult rat uteri exposed to LA	123
8.6	Histologic sections from adult rat uteri exposed to LA. Note remarkably higher PCNA expression is visible in T2 and T3	124
8.7	Photomicrograph sections of LA exposed the uteri of rats of different experimental groups (C, T0, T1, T2 and T3) treated with different doses of EBN showing expression of $E_2R$	126
8.8	Photomicrograph sections of LA exposed the uteri of rats of different experimental groups (C, T0, T1, T2 and T3) treated with different doses of EBN showing expression of $P_4R$	127
8.9	Effect of EBN on Oxidative stress, total antioxidant capacity (TAC) and antioxidant enzyme activities in plasm of LA exposed rats	129

# LIST OF ABBREVIATIONS

	μg	micro gram
	μl	Microliter
	μm	Micrometer
	μmol	Micromole
	AF	Atretic follicle
	ALP	Alkaline phosphatase
	ALT	Alanine transaminase
	ANOVA	Analysis of variance
	AO	Antioxidant
	AR	Amphiregulin
	AST	Asparate aminotransferase
	ATSDR	Agency for Toxic Substances and Disease Registry
	bFGF	Basic fibroblast growth factor
	BW	Body weight
	Caco-2 cells	Human colonic adenocarcinoma cell line
	CFTR	Cystic fibrosis transmembrane conductance regulator
	CL	Corpus luteum
	CO ₂	Carbon dioxide
	cpm	Counts per minute
	CREA	Creatinine
	DAB	3,3-diaminobenzidine
	DNA	Deoxyribonucleic acid
	$E_2$	Estrogen
	$E_2R$	Receptor of estrogen

EBN	Edible bird's nest
EGF	Epidermal growth factor
EIS	Embryonic implantation sites
ENaC	Epithelial Na+ channel
ETP	Economic Transformation Programme
F	Follicular unit
FGFS	Fibroblast growth factors
FSH	Follicle-stimulating hormone
FSH	Follicular stimulating hormone
GE	Glandular epithelium
GH	Growth hormone
GHRH	Growth hormone releasing hormone
GSH	Glutathione
H and E	Using hematoxylin and eosin
H ₂ O ₂	Hydrogen peroxide
hADSCs	Human adipose-derived stem cells
HB-EGF	Heparin-binding epidermal growth factor
HGF	Hepatocyte growth factor
IC	Interstitial cell
ICM	Inner cell mass
IGF	Insulin-like growth factor
LA	Lead acetate
LE	The luminal epithelium
LH	Luteinizing hormone
MDA	malondialdehyde

min	Minute
ml	Milliliter
mmol	Millimole
MSCs	Placenta-derived multi potent mesenchymal stem cells
NC	Control stain without antibody
NF	Antral follicle
NF-κB	Nuclear factor kappa-light-chain-enhancer of activated B cells
NO	Nitrogen oxides
NOS	Nitrogen oxide species
O ₂ -	Superoxide
OBWR	Ovarian body weight ratio
°C	degree Celsius
OD	optical density
OS	Oxidative stress
OSB	Oxidative stress biomarkers
OSE	Ovarian surface epithelium
Р	Prolactin
P4	Progesterone
P4R	Receptor of Progesterone
Pb	Lead
PCNA	Proliferating cell nuclear antigen
REGF	Receptor of epidermal growth factor
ROS	Reactive oxygen species
S	Stromal cells
SA	Sialic acid

SE	Standard error
SEM	Scanning electron microscope
SOD	Superoxide dismutase
Т	Testosterone
TAC	Total antioxidant capacity
TBARS	Thiobarbituric acid reactive substance
TNF-α	Tumour necrosis factor-alpha
UBWR	Uterine body weight ratio
UE	Uterine endothelium
un	Number of uterine glands in endothelium
VEGF	Vascular endothelial growth factor
WHO	World Health Organization

C

### **CHAPTER 1**

#### **INTRODUCTION**

There has been an increase in interest towards natural substances and their bioactive components in the last two decades. One of the primary motives was hormone replacement therapy that showed their dangerous side effects over time, whereas natural substances have been used for centuries without side effects (Hanafy and Hatem, 1991). Edible bird's nest (EBN) is a natural product of the salivary secretion of two male distinct swiftlets; *Aerodramus fucifagus* and *Aerodramus maximus*. Swiftlet's are insectivorous birds naturally found inhabiting mainly limestone caves in Asian countries, such as Malaysia, Indonesia, Thailand, Vietnam, Philippines and China (Marcone, 2005). Indonesia is the major resource, while the Malaysian Borneo provinces of Sarawak and Sabah (East Malaysia) being the second major sources (Hobbs, 2004). Edible bird's nest is traditionally considered as effective medicine by the Chinese community for centuries and has been used to alleviate many ailments.

Research on various aspects of EBN, including its nutritional and health benefit, has gained momentum recently and scientific explanations are being provided mainly supporting the traditional beliefs. Enhancement of reproductive performance and behaviours such as libido are among the traditionally believed benefits of EBN which are yet to be explained and proven scientifically. Emerging research findings on EBN in areas other than reproduction have shown that EBN possesses many biological properties, including the ability to stimulate growth and proliferation of stem cells, epidermal growth factor (EGF) – like activity, enhance production of reproductive hormones like estrogen and act as an antioxidant. Bioactive compounds reported to be found in EBN include sialic acid, glycoproteins, hormones, minerals and vitamins (Ma and Liu, 2012b). Thus, it is understood that all these biological properties of EBN would have a potential influence on the process of reproduction. According to Spencer (2013) pregnancy rates from a single insemination per animal is known as fertility, which is considered high in sheep, moderate in beef cattle and low in dairy and humans. Therefore, infertility and sub fertility are major problems in humans. Early pregnancy losses, endometria are responsible for about two- third (Ledee-Bataille et al., 2002). Successful implantation requires regulation of maternal hormones such as ovarian estrogen and progesterone and their receptors in endometrium that lead to production of signaling molecules which include proliferating cell nuclear antigen (PCNA) and growth factors such as vascular endothelial growth factor (VEGF), epidermal growth factor (EGF) and receptor of epidermal growth factor (REGF) (Dey et al., 2004). Looking for strategies to enhance reproductive success, which is a multifactorial issue, has been an active research area. Considering all the potential bioactive ingredients and functions of EBN reviewed above, we hypothesized that EBN would also confer enhancing effect to reproduction as well as be able to ameliorate toxic damages caused by exposure toxic heavy materials like Lead (pb) among major growing environmental pollutants that affect body organs through exposure from air, water and food sources .Band lead acetate(LA) was one of the

mainly environmental pollutants environmental levels of lead have increased more than 1000-fold over the past three centuries as result of human activity and due to increasing worldwide use of leaded gasoline(Agency for Toxic Substances and Disease Registry - ATSDR, 2007). Several studies showed the adverse effect of Pb toxicity on the liver, pituitary gland and reproduction associated with damage to surviving ovarian follicles and the reproductive system including the ovary and uterus and ovarian follicles. One of the important mechanisms underlying LA toxicity is the induction of oxidative stress as a result of production of reactive oxygen species, depletion of the antioxidant defense system and LA crosses the biomembranes to reach the soft tissue cells, and thus precipitates in the ovary, and placenta (Flora *et al.*,2011). Although EBN has been praised as a potent antioxidant (Yida *et al.*, 2015a), its role in mitigating the effect of LA toxicity on the reproductive system is unknown.

Therefore, the objectives of the study were:

- 1. To determine effects of EBN supplementation on cycling female rats with expect to its reproductive and pituitary gland histomorphology, hormones and expressions of their recepters, proliferating cell nuclear antigen (PCNA) and growth factors such as EGF, REGF, and VEGF as well as on oxidative stress biomarkers (OSB) and antioxidants (AO).
- 2. To determine effects of EBN supplementation on rat's uterine histomorphology and ultrastructure, embryo implantation, endometrial expression of P₄R and E₂R, PCNA and growth factors as well as levels of steroid hormones, OSB and AOs.
- 3. To evaluate ameliorating effect of EBN supplementation on LA toxicity effect on the reproductive system of female rats through assessment of histopathology of ovary, uterus, pituitary gland, ovarian and endometrial expression of E₂R, P₄R and PCNA and growth factors as well as plasma steroid hormones, OSB and AOs.

From the above objectives it is hypothesized that EBN supplement improves fertility of female rats through enhancing the functions of uterus, ovary and pituitary gland as well as minimizing oxidative stress without causing toxic effect on liver. In addition, EBN plays a prophylactic role in protecting the reproductive functions of rats from toxic effect of lead acetate.

#### REFERENCES

- Abidin, F. Z., Hui, C. K., Luan, N. S., Ramli, E. S. M., Hun, L. T., and Ghafar, N. A. (2011). Effects of edible bird's nest (EBN) on cultured rabbit corneal keratocytes. *BMC Complementary and Alternative Medicine*, 11 (1), 94:1-10.
- Adler, N., and Zoloth, S. (1970). Copulatory behavior can inhibit pregnancy in female rats. *Science*, *168* (3938), 1480-1482.
- Aebi H (1974). Catalase, 673 p. In: Bergmeyer HU (Ed.). Methods in enzymatic analysis, 2, New York, Academic Press.
- Agarwal, A., Gupta, S., and Sharma, R. K. (2005). Role of oxidative stress in female reproduction. *Reproductive Biology and Endocrinology*, *3* (1), 28:1-21.
- Agca, Y., and Critser, J. K. (2006). Assisted reproductive technologies and genetic modifications in rats. *The Laboratory Rat (Second Edition)* (pp. 165-189): Elsevier.
- Aghajanova, L., Bjuresten, K., Altmäe, S., Landgren, B.-M., and Stavreus-Evers, A. (2008). HB-EGF but not amphiregulin or their receptors HER1 and HER4 is altered in endometrium of women with unexplained infertility. *Reproductive Sciences*, *15* (5), 484-492.
- Aitken, R. J., and Krausz, C. (2001). Oxidative stress, DNA damage and the Y chromosome. *Reproduction*, 122 (4), 497-506.
- Alan, E., Liman, N., and Sağsöz, H. (2015). The profile of the epidermal growth factor system in rat endometrium during postpartum involution period. *Veterinary Research Communications*, 39 (2), 115-135.
- Alchalabi, A. S., Rahim, H., Aklilu, E., Al-Sultan, I. I., Aziz, A. R., Malek, M. F., Ronald, S. H., and Khan, M. A. (2016). Histopathological changes associated with oxidative stress induced by electromagnetic waves in rats' ovarian and uterine tissues. *Asian Pacific Journal of Reproduction*, 5 (4), 301-310.
- Anasti, J., Kalantaridou, S., Kimzey, L., George, M., and Nelson, L. (1998). Human follicle fluid vascular endothelial growth factor concentrations are correlated with luteinization in spontaneously developing follicles. *Human Reproduction (Oxford, England), 13* (5), 1144-1147.
- Aplin, J. D., and Kimber, S. J. (2004). Trophoblast-uterine interactions at implantation. *Reproductive Biology and Endocrinology*, 2 (1), 48:1-12.
- Arii, S., and Imamura, M. (2000). Physiological role of sinusoidal endothelial cells and Kupffer cells and their implication in the pathogenesis of liver injury. *Journal of Hepato-Biliary-Pancreatic Sciences*, 7(1), 40-48.

- Armelin, H. A. (1973). Pituitary extracts and steroid hormones in the control of 3T3 cell growth. *Proceedings of the National Academy of Sciences*, 70 (9), 2702-2706.
- Assennato, G., Paci, C., Baser, M. E., Molinini, R., Candela, R. G., Altamura, B. M., and Giorgino, R. (1986). Sperm count suppression without endocrine dysfunction in lead-exposed men. *Archives of Environmental Health: An International Journal*, 41 (6), 387-390.
- Assi, M. A., Hezmee, M. N. M., Abba, Y., Yusof, M. S. M., Haron, A. W., Rajion, M. A., and Al-Zuhairy, M. A. (2016). Prophylactic effect of Nigella sativa against lead acetate induced changes in spermiogram, reproductive hormones and gonadal histology of rats. *Veterinary World*, 9 (11), 1305–1311.
- Assi, M., Hezmee, M., Abba, Y., Rajion, M., Wahid, H., and Yusof, M. (2017a). Assessment of therapeutic effects of Nigella sativa against chronic lead acetate-induced reproductive dysfunction in male Sprague-Dawley rats. *Comparative Clinical Pathology*, 26 (1), 87-97.
- Assi, M., Hezmee, M., Abba, Y., Sabri, M., Haron, A., Baiee, F., and Rajion, M. (2017b). Effect of Nigella sativa Pre-Treatment on Sub-Chronic Lead Acetate Induced Hematological and Biochemical Alterations. *Journal of Computational and Theoretical Nanoscience*, 14 (6), 2752-2758.
- Atsdr, (Agency for Toxic Substances and Disease Registry). (2007) . Toxicological Profile for Lead. (Draft for Public Comment) Agency for Toxic Substances and Disease Registry, Public Health Service, United State Department of Health and Human Services, Atlanta, GA.
- Auersperg, N., Wong, A. S., Choi, K.-C., Kang, S. K., and Leung, P. C. (2001). Ovarian surface epithelium: biology, endocrinology, and pathology. *Endocrine Reviews*, 22 (2), 255-288.
- Aziz, F., Maulood, I., and Chawsheen, M. (2012). Effects of melatonin, vitamin Cand E alone or in combination on lead-induced injury in liver and kidney organs of rats. *IOSR Journal of Parmacy*, 2(5),13-18.
- Baird, A., and Walicke, P. A. (1989). Fibroblast growth factors. *British Medical Bulletin*, 45 (2), 438-452.
- Baker, D. E. (1979). Reproduction and breeding. *In: the laboratory rat* (First Edition), (pp. 153-168) : Elsevier.
- Baker, M. A., and Aitken, R. J. (2005). Reactive oxygen species in spermatozoa: methods for monitoring and significance for the origins of genetic disease and infertility. *Reproductive Biology and Endocrinology*, *3* (1), 67:1-9.
- Baranowska-Bosiacka, I., Gutowska, I., Rybicka, M., Nowacki, P., and Chlubek, D. (2012). Neurotoxicity of lead. Hypothetical molecular mechanisms of synaptic function disorders. *Neurologia i Neurochirurgia Polska*, 46 (6), 569-578.

- Bazer, F. W. (1975). Uterine protein secretions: relationship to development of the conceptus. *Journal of Animal Science*, 41 (5), 1376-1382.
- Bazer, F. W., Wu, G., Spencer, T. E., Johnson, G. A., Burghardt, R. C., and Bayless, K. (2009). Novel pathways for implantation and establishment and maintenance of pregnancy in mammals. *MHR: Basic Science of Reproductive Medicine*, 16 (3), 135-152.
- Been, J. V., Nurmatov, U., van Schayck, C. P., and Sheikh, A. (2013). The impact of smoke-free legislation on fetal, infant and child health: a systematic review and meta-analysis protocol. *BMJ Journal*, *3* (2), e002261.1-5.
- Bergemann, N., Mundt, C., Parzer, P., Jannakos, I., Nagl, I., Salbach, B., Klinga, K., Runnebaum, B., and Resch, F. (2005). Plasma concentrations of estradiol in women suffering from schizophrenia treated with conventional versus atypical antipsychotics. *Schizophrenia Research*, 73 (2), 357-366.
- Bivin WS. The rat. 1986. In: Morrow DA, editor. Current therapy in theriogenology
  2. Diagnosis, treatment, and prevention of reproductive diseases in small and large animals, p 1015–1021.London (UK): WB Saunders.
- Bjersing, L., and Cajander, S. (1975). Ovulation and the role of the ovarian surface epithelium. *Cellular and Molecular Life Sciences*, *31* (5), 605-608.
- Bolin, C. M., Basha, R., Cox, D., Zawia, N. H., Maloney, B., Lahiri, D. K., and Cardozo-Pelaez, F. (2006). Exposure to lead and the developmental origin of oxidative DNA damage in the aging brain. *The FASEB Journal*, 20 (6), 788-790.
- Böttcher, R. T., and Niehrs, C. (2005). Fibroblast growth factor signaling during early vertebrate development. *Endocrine Reviews, 26* (1), 63-77.
- Bradshaw, H. B., and Allard, C. (2011). Endogenous cannabinoid production in the rat female reproductive tract is regulated by changes in the hormonal milieu. *Pharmaceuticals*, 4 (6), 933-949.
- Brigstock, D., Heap, R., and Brown, K. (1989). Polypeptide growth factors in uterine tissues and secretions. *Journal of Reproduction and Fertility*, 85 (2), 747-758.
- Bussmann, U. A., Bussmann, L. E., and Barañao, J. L. (2006). An aryl hydrocarbon receptor agonist amplifies the mitogenic actions of estradiol in granulosa cells: evidence of involvement of the cognate receptors. *Biology of Reproduction*, 74 (2), 417-426.
- Byun, H.-S., Lee, G.-S., Lee, B.-M., Hyun, S.-H., Choi, K.-C., and Jeung, E.-B. (2008). Implantation-related expression of epidermal growth factor family molecules and their regulation by progesterone in the pregnant rat. *Reproductive Sciences*, *15* (7), 678-689.
- Carlone, D. L., and Rider, V. (1993). Embryonic modulation of basic fibroblast growth factor in the rat uterus. *Biology of Reproduction, 49* (4), 653-665.

- Carmeliet, P., Ferreira, V., Breier, G., Pollefeyt, S., Kieckens, L., Gertsenstein, M., Fahrig, M., Vandenhoeck, A., Harpal, K., and Eberhardt, C. (1996). Abnormal blood vessel development and lethality in embryos lacking a single VEGF allele. *Nature*, 380 (6573), 435-439.
- Carpenter, K. D., and Korach, K. S. (2006). Potential biological functions emerging from the different estrogen receptors. *Annals of the New York Academy of Sciences, 1092* (1), 361-373.
- Carson DD. (2008).Molecular and cell biology of embryo-uterine interactions mammalian embryo implantation. *Seminars in Cell & Developmental Biology* .19(2):160. [PubMed: 18272413].
- Cavagna, M., and Mantese, J. (2003). Biomarkers of endometrial receptivity—a review. *Placenta, 24*, S39-S47.
- Chakraborty, I., Das, S., and Dey, S. (1995). Differential expression of vascular endothelial growth factor and its receptor mRNAs in the mouse uterus around the time of implantation. *Journal of Endocrinology*, *147* (2), 339-352.
- Chan, L., Tsang, L., Rowlands, D., Rochelle, L., Boucher, R., Liu, C., and Chan, H. (2002). Distribution and regulation of ENaC subunit and CFTR mRNA expression in murine female reproductive tract. *Journal of Membrane Biology*, 185 (2), 165-176.
- Chau, Q., Cantor, S. B., Caramel, E., Hicks, M., Kurtin, D., Grover, T., and Elting, L. S. (2003). Cost-effectiveness of the bird's nest filter for preventing pulmonary embolism among patients with malignant brain tumors and deep venous thrombosis of the lower extremities. *Supportive Care in Cancer*, 11 (12), 795-799.
- Chen, J., Cheng, J.-G., Shatzer, T., Sewell, L., Hernandez, L., and Stewart, C. (2000). Leukemia inhibitory factor can substitute for nidatory estrogen and is essential to inducing a receptive uterus for implantation but is not essential for subsequent embryogenesis. *Endocrinology*, 141 (12), 4365-4372.
- Chobotova, K., Spyropoulou, I., Carver, J., Manek, S., Heath, J. K., Gullick, W. J., Barlow, D. H., Sargent, I. L., and Mardon, H. J. (2002). Heparin-binding epidermal growth factor and its receptor ErbB4 mediate implantation of the human blastocyst. *Mechanisms of Development*, 119 (2), 137-144.
- Christenson, L. K., and Stouffer, R. L. (1997). Follicle-stimulating hormone and luteinizing hormone/chorionic gonadotropin stimulation of vascular endothelial growth factor production by macaque granulosa cells from pre-and periovulatory follicles. *The Journal of Clinical Endocrinology and Metabolism, 82* (7), 2135-2142.

- Chua, K.-H., Lee, T.-H., Nagandran, K., Yahaya, N. H. M., Lee, C.-T., Tjih, E. T. T., and Aziz, R. A. (2013). Edible Bird's nest extract as a chondro-protective agent for human chondrocytes isolated from osteoarthritic knee: in vitro study. *BMC Complementary and Alternative Medicine*, *13* (1), 19:1-9.
- Chua, L. S., and Zukefli, S. N. (2016). A comprehensive review of edible bird nests and swiftlet farming. *Journal of Integrative Medicine*, 14 (6), 415-428.
- Clifford, A., Lang, L., and Chen, R. (2012). Effects of maternal cigarette smoking during pregnancy on cognitive parameters of children and young adults: a literature review. *Neurotoxicology and Teratology*, *34* (6), 560-570.
- Colombo, J. P., Garcia- Rodenas, C., Guesry, P., and Rey, J. (2003). Potential effects of supplementation with amino acids, choline or sialic acid on cognitive development in young infants. *Acta Paediatrica*, *92* (s442), 42-46.
- Couse, J. F., Dixon, D., Yates, M., Moore, A. B., Ma, L., Maas, R., and Korach, K. S. (2001). Estrogen receptor-α knockout mice exhibit resistance to the developmental effects of neonatal diethylstilbestrol exposure on the female reproductive tract. *Developmental Biology*, 238 (2), 224-238.
- Cullinan-Bove, K., and Koos, R. D. (1993). Vascular endothelial growth factor/vascular permeability factor expression in the rat uterus: rapid stimulation by estrogen correlates with estrogen-induced increases in uterine capillary permeability and growth. *Endocrinology*, *133* (2), 829-837.
- Darbre, P. (2006). Metalloestrogens: an emerging class of inorganic xenoestrogens with potential to add to the oestrogenic burden of the human breast. *Journal of Applied Toxicology*, 26 (3), 191-197.
- Das, S. K., Wang, X.-N., Paria, B. C., Damm, D., Abraham, J. A., Klagsbrun, M., Andrews, G. K., and Dey, S. K. (1994). Heparin-binding EGF-like growth factor gene is induced in the mouse uterus temporally by the blastocyst solely at the site of its apposition: a possible ligand for interaction with blastocyst EGF-receptor in implantation. *Development*, 120 (5), 1071-1083.
- Das, S., Das, N., Wang, J., Lim, H., Schryver, B., Plowman, G., and Dey, S. (1997).
   Expression of betacellulin and epiregulin genes in the mouse uterus temporally by the blastocyst solely at the site of its apposition is coincident with the "window" of implantation. *Developmental Biology*, 190 (2), 178-190.
- Dechanet C, Brunet C, Anahory T, Hamamah S, Hedon B, Dechaud H(2011).Effect of cigarette smoking of embryo implantation and placentation and analysis of factors interfering with cigarette smoke effects (part II).*Gynecologie Obstetrique and Fertilite* 39(10): 567–574.
- Dey, S., Lim, H., Das, S. K., Reese, J., Paria, B., Daikoku, T., and Wang, H. (2004). Molecular cues to implantation. *Endocrine Reviews*, 25 (3), 341-373.

- Dickson, S. E., and Fraser, H. M. (2000). Inhibition of early luteal angiogenesis by gonadotropin-releasing hormone antagonist treatment in the primate. *The Journal of Clinical Endocrinology & Metabolism, 85* (6), 2339-2344.
- Dormandy, T. (1978). Free-radical oxidation and antioxidants. *The Lancet, 311* (8065), 647-650.
- Dorostghoal, M., Dezfoolian, A., and Sorooshnia, F. (2011). Effects of maternal lead acetate exposure during lactation on postnatal development of testis in offspring wistar rats. *Iranian Journal of Basic Medical Sciences, 14* (2), 122-131.
- Doumouchtsis, K., Doumouchtsis, S., Doumouchtsis, E., & Perrea, D. (2009). The effect of lead intoxication on endocrine functions. Journal of endocrinological investigation, 32(2), 175-183.
- Drummond, A. E. (2006). The role of steroids in follicular growth. *Reproductive Biology and Endocrinology*, 4 (1), 16:1-11.
- Elgawish, R. A. R., and Abdelrazek, H. M. (2014). Effects of lead acetate on testicular function and caspase-3 expression with respect to the protective effect of cinnamon in albino rats. *Toxicology Reports*, 1, 795-801.
- El-Tantawy, W. H. (2016). Antioxidant effects of Spirulina supplement against lead acetate-induced hepatic injury in rats. *Journal of Traditional and Complementary Medicine*, 6 (4), 327-331.
- Enders, A. C. (2000). Trophoblast-uterine interactions in the first days of implantation: models for the study of implantation events in the human. Paper presented at the Seminars in reproductive medicine.
- Enders, A. C., and Nelson, D. M. (1973). Pinocytotic activity of the uterus of the rat. *Developmental Dynamics*, 138 (3), 277-299.
- Eugenia, D., Alexandra, T., Argherie, D., and Cristina, R. (2009). The consequences of in utero exposure to lead acetate on exposure and integrity biomarkers of reproductive system in female rats at sexual maturity. *Lucr Şt Med Vet Timişoara Luc Stii Medi Veteri, 42*, 295-300.
- Farrag, A.-R. H. (2007). Protective Effect of Nigella sativa Seeds Against Leadinduced Hepatorenal Damage in Male Rats' Abdel-Razik H. Farrag, "Karam A. Mahdy, 'Gamal H. Abdel Rahman and "Mostafa M. Osfor" Departments of Pathology, "Department of Medical Biochemistry. *Pakistan Journal of Biological Sciences*, 10 (17), 2809-2816.
- Ferdous, A., Morris, J., Abedin, M. J., Collins, S., Richardson, J. A., and Hill, J. A. (2011). Forkhead factor FoxO1 is essential for placental morphogenesis in the developing embryo. *Proceedings of the National Academy of Sciences*, 108 (39), 16307-16312.

- Ferrara, N., and Davis-Smyth, T. (1997). The biology of vascular endothelial growth factor. *Endocrine Reviews*, 18 (1), 4-25.
- Ferriani, R., Charnock-Jones, D., Prentice, A., Thomas, E., and Smith, S. (1993). Immunohistochemical localization of acidic and basic fibroblast growth factors in normal human endometrium and endometriosis and the detection of their mRNA by polymerase chain reaction. *Human Reproduction*, 8 (1), 11-16.
- Flohé, L., and Zimmermann, R. (1970). The role of GSH peroxidase in protecting the membrane of rat liver mitochondria. *Biochimica et Biophysica Acta (BBA)-Bioenergetics*, 223 (1), 210-213.
- Fukuda, M. N., Sato, T., Nakayama, J., Klier, G., Mikami, M., Aoki, D., and Nozawa, S. (1995). Trophinin and tastin, a novel cell adhesion molecule complex with potential involvement in embryo implantation. *Genes & Development*, 9 (10), 1199-1210.
- Gerhard, I., Waibel, S., Daniel, V., and Runnebaum, B. (1998). Impact of heavy metals on hormonal and immunological factors in women with repeated miscarriages. *Human Reproduction Update*, 4 (3), 301-309.
- Ghassem, M., Arihara, K., Mohammadi, S., Sani, N. A., and Babji, A. S. (2017). Identification of two novel antioxidant peptides from edible bird's nest (Aerodramus fuciphagus) protein hydrolysates. *Food & Function*, 8 (5), 2046-2052.
- Gheri, G., Vichi, D., Thyrion, G. Z., Bonaccini, L., Vannelli, G., Marini, M., and Sgambati, E. (2009). Sialic acid in human testis and changes with aging. *Reproduction, Fertility and Development, 21* (5), 625-633.
- Gillet, J., Maillet, R., and Gautier, C. (1980). Blood and lymph supply of the ovary *Biology of the Ovary* .(pp. 86-98): Martinus Nijhoff London.
- Gray, C., Burghardt, R., Johnson, G., Bazer, F., and Spencer, T. (2002). Evidence that absence of endometrial gland secretions in uterine gland knockout ewes compromises conceptus survival and elongation. *Reproduction, 124* (2), 289-300.
- Gupta, A., Bazer, F. W., and Jaeger, L. A. (1997). Immunolocalization of acidic and basic fibroblast growth factors in porcine uterine and conceptus tissues. *Biology of Reproduction, 56* (6), 1527-1536.
- Gupta, Ramesh. C. (2011). Arsenic, cadmium and lead. In Swaran J. S. Flora, Vidhu Pachauri and Geetu Saxena, editor .Reproductive and Developmental Toxicology (p415-438). Kentucky, USA: Academic Press.
- Guzeloglu-Kayisli, O., Kayisli, U. A., and Taylor, H. S. (2009). The role of growth factors and cytokines during implantation: endocrine and paracrine interactions. Paper presented at the Seminars in Reproductive Medicine.

- Halliwell, B., and Whiteman, M. (2004). Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean? *British Journal of Pharmacology*, *142* (2), 231-255.
- Hamadouche, N. A., Nesrine, S., and Abdelkeder, A. (2013). Lead toxicity and the hypothalamic-pituitary-testicular axis. *Notulae Scientia Biologicae*, 5 (1), 1-6.
- Hamatani, T., Daikoku, T., Wang, H., Matsumoto, H., Carter, M. G., Ko, M. S., and Dey, S. K. (2004). Global gene expression analysis identifies molecular pathways distinguishing blastocyst dormancy and activation. *Proceedings of the National Academy of Sciences of the United States of America*, 101 (28), 10326-10331.
- Hamid, HudaYahya. (2012). Morphological and molecular changes in embryomaternal interactions in a rat model at optimal and elevated environmental temperatures. Doctoral thesis, Universiti Putra Malaysia. (FPV 2012 4).
- Hammed, M. S. (2015). Evaluation of Performance of Date Palm Pollen on Urea and Creati-nine Levels in Adult Female Rats Exposed to Lead Acetate Intoxication. *International Journal of Biomedical and Advance Research*, 6, 20-24.
- Hanafy, M., and Hatem, M. (1991). Studies on the antimicrobial activity of Nigella sativa seed (black cumin). *Journal of Ethnopharmacology*, 34 (2-3), 275-278.
- Hartwig, A. (2010). Mechanisms in cadmium-induced carcinogenicity: recent insights. *Biometals*, 23 (5), 951-960.
- He, Q., Tsang, L. L., Ajonuma, L. C., and Chan, H. C. (2010). Abnormally upregulated cystic fibrosis transmembrane conductance regulator expression and uterine fluid accumulation contribute to Chlamydia trachomatis-induced female infertility. *Fertility and Sterility*, 93 (8), 2608-2614.
- Heinrich, P. C., Behrmann, I., Müller-Newen, G., Schaper, F., and Graeve, L. (1998). Interleukin-6-type cytokine signalling through the gp130/Jak/STAT pathway. *Biochemical Journal, 334* (2), 297-314.
- Herbst, R. S., Giaccone, G., Schiller, J. H., Natale, R. B., Miller, V., Manegold, C., Scagliotti, G., Rosell, R., Oliff, I., and Reeves, J. A. (2004). Gefitinib in combination with paclitaxel and carboplatin in advanced non–small-cell lung cancer: a phase III trial—INTACT 2. *Journal of Clinical Oncology, 22* (5), 785-794.
- Hillier, S. G., and De zwart, F. A. (1981). Evidence that granulosa cell aromatase induction/activation by follicle-stimulating hormone is an androgen receptor-regulated process in-vitro. *Endocrinology*, *109* (4), 1303-1305.
- Hillier, S. G., Knazek, R. A., and ROSS, G. T. (1977). Androgenic stimulation of progesterone production by granulosa cells from preantral ovarian follicles: further in vitro studies using replicate cell cultures. *Endocrinology*, 100 (6), 1539-1549.

- Hirano, T., Ishihara, K., and Hibi, M. (2000). Roles of STAT3 in mediating the cell growth, differentiation and survival signals relayed through the IL-6 family of cytokine receptors. *Oncogene, 19* (21), 2548-2556.
- Hobbs, J. J. (2004). Problems in the harvest of edible birds' nests in Sarawak and Sabah, Malaysian Borneo. *Biodiversity & Conservation, 13* (12), 2209-2226.
- Horiguchi, H., Oguma, E., Sasaki, S., Okubo, H., Murakami, K., Miyamoto, K., Hosoi, Y., Murata, K., and Kayama, F. (2013). Age-relevant renal effects of cadmium exposure through consumption of home-harvested rice in female Japanese farmers. *Environment International*, 56, 1-9.
- Hsu, P.-C., and Guo, Y. L. (2002). Antioxidant nutrients and lead toxicity. *Toxicology*, *180* (1), 33-44.
- Huet, Y. M., Andrews, G., and Dey, S. (1989). Modulation of c-myc protein in the mouse uterus during pregnancy and by steroid hormones. *Progress in Clinical and Biological Research*, 294, 401-412.
- Huet-hudson, Y. M., Andrews, G. K., and Dey, S. K. (1989). Cell type-specific localization of c-myc protein in the mouse uterus: modulation by steroid hormones and analysis of the periimplantation period. *Endocrinology*, *125* (3), 1683-1690.
- Irusta, G., Abramovich, D., Parborell, F., and Tesone, M. (2010). Direct survival role of vascular endothelial growth factor (VEGF) on rat ovarian follicular cells. *Molecular and Cellular Endocrinology*, *325* (1), 93-100.
- Ismail M. Hou, Z.P. Stanslas, J. Iman, M.U. Zhang, Y.D. Ideris A. and Mahmud R. Edible Bird's Nest Pretreatment Prevents Ovariectomy Induced Cognitive Aging in Morris Water Maze. Proceedings of the Edible Bird Nest Industry Conference, 25-26 November 2014, Putrajaya, Malaysia.
- Järup, L. (2003). Hazards of heavy metal contamination. *British Medical Bulletin, 68* (1), 167-182.
- Jessmon, P., Leach, R. E., and Armant, D. R. (2009). Diverse functions of HBEGF during pregnancy. *Molecular Reproduction and Development*, 76 (12), 1116-1127.
- Jin, K., Mao, X., Batteur, S., McEachron, E., Leahy, A., and Greenberg, D. (2001). Caspase-3 and the regulation of hypoxic neuronal death by vascular endothelial growth factor. *Neuroscience*, 108 (2), 351-358.
- Johnson, D. E., and Williams, L. T. (1992). Structural and functional diversity in the FGF receptor multigene family. *Advances in Cancer Research*, *60*, 1-41.
- Jomova, K., and Valko, M. (2011). Advances in metal-induced oxidative stress and human disease. *Toxicology*, 283 (2), 65-87.

- Jones, R., Bulmer, J., and Searle, R. (1995). Immunohistochemical characterization of proliferation, oestrogen receptor and progesterone receptor expression in endometriosis: comparison of eutopic and ectopic endometrium with normal cycling endometrium. *Human Reproduction*, *10* (12), 3272-3279.
- Jory, P., and Saengthong, J. (2007). Birds' Nests: Secrets of a Billion-Dollar Business. Thailand Research Fund. *Regional Studies Program, Walailak University, Nakhon Si Thammarat*
- Joswig, A., Gabriel, H.-D., Kibschull, M., and Winterhager, E. (2003). Apoptosis in uterine epithelium and decidua in response to implantation: evidence for two different pathways. *Reproductive Biology and Endocrinology*, 1 (1), 44:1-9.
- Juliani, C., Silva-Zacarin, E., Santos, D., and Boer, P. (2008). Effects of atrazine on female Wistar rats: morphological alterations in ovarian follicles and immunocytochemical labeling of 90kDa heat shock protein. *Micron, 39* (5), 607-616.
- Kakuta, H., Tanaka, M., Chambon, P., Watanabe, H., Iguchi, T., and Sato, T. (2012). Involvement of gonadotropins in the induction of hypertrophy-hyperplasia in the interstitial tissues of ovaries in neonatally diethylstilbestrol-treated mice. *Reproductive Toxicology*, 33 (1), 35-44.
- Kamijo, T., Rajabi, M. R., Mizunuma, H., and Ibuki, Y. (1998). Biochemical evidence for autocrine/paracrine regulation of apoptosis in cultured uterine epithelial cells during mouse embryo implantation in vitro. *Molecular Human Reproduction*, 4 (10), 990-998.
- Kelman, Z. (1997). PCNA: structure, functions and interactions. Oncogene, 14 (6),629-640.
- Kimmel, C. A., L. D. Grant, C. S. Sloan and B. C. Gladen, 1980: Chronic low-level lead toxicity in the rat: I. Maternal toxicity and perinatal effects. *Toxicology* and Applied Pharmacology, 56, 28-41.
- Klejewski, A., Urbaniak, T., Pisarska-Krawczyk, M., and Sobczyk, K. (2012). Influence of smoking on pregnancy. *Przeglad Lekarski, 69* (10), 929-933.
- Knecht, M., and Catt, K. (1983). Modulation of cAMP-mediated differentiation in ovarian granulosa cells by epidermal growth factor and platelet-derived growth factor. *Journal of Biological Chemistry*, 258 (5), 2789-2794.
- Kodaman, P. H., and Taylor, H. S. (2004). Hormonal regulation of implantation. *Obstetrics and gynecology clinics of North America, 31* (4), 745-766.
- Kolios, G., Valatas, V., and Kouroumalis, E. (2006). Role of Kupffer cells in the pathogenesis of liver disease. *World Journal of Gastroenterology*: WJG, 12(46), 7413-7420.

- Kondoh, E., Okamoto, T., Higuchi, T., Tatsumi, K., Baba, T., Murphy, S. K., Takakura, K., Konishi, I., and Fujii, S. (2008). Stress affects uterine receptivity through an ovarian-independent pathway. *Human Reproduction*, 24 (4), 945-953.
- Kong, Y., Keung, W., Yip, T., Ko, K., Tsao, S., and Ng, M. (1987). Evidence that epidermal growth factor is present in swiftlet's (Collocalia) nest. *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry*, 87 (2), 221-226.
- Kong, Y., Tsao, S., Song, M., Ng, M., and Lin, Z. (1989). Potentiation of mitogenic response by extracts of the swiftlet's (Apus) nest collected from Huai-ji. Acta Zoologica Sinica, 35 (4), 429-435.
- Kumar, T. R., Wang, Y., Lu, N., and Matzuk, M. M. (1997). Follicle stimulating hormone is required for ovarian follicle maturation but not male fertility. *Nature Genetics*, 15 (2), 201-204.
- Lakshmi, B., Sudhakar, M., and Aparna, M. (2013). Protective potential of Black grapes against lead induced oxidative stress in rats. *Environmental Toxicology and Pharmacology*, *35* (3), 361-368.
- Lanzone, A., Fortini, A., Fulghesu, A. M., Soranna, L., Caruso, A., and Mancuso, S. (1996). Growth hormone enhances estradiol production follicle-stimulating hormone-induced in the early stage of the follicular maturation. *Fertility and Sterility*, 66 (6), 948-953.
- Lau, A. S., and Melville, D. S. (1994). *International trade in swiftlet nests with special reference to Hong Kong*: Traffic International.
- Laughlin, N. K., Bowman, R. E., Franks, P. A., and Dierschke, D. J. (1987). Altered menstrual cycles in rhesus monkeys induced by lead. *Toxicological Sciences*, 9 (4), 722-729.
- Lawton, L. J., and Donaldson, W. (1991). Lead-induced tissue fatty acid alterations and lipid peroxidation. *Biological Trace Element Research*, 28 (2), 83-97.
- Lecce, G., Meduri, G., Ancelin, M., Bergeron, C., and Perrot-applanat, M. (2001). Presence of estrogen receptor  $\beta$  in the human endometrium through the cycle: expression in glandular, stromal, and vascular cells. *The Journal of Clinical Endocrinology & Metabolism, 86* (3), 1379-1386.
- Ledee-Bataille, N., Lapree-Delage, G., Taupin, J.-L., Dubanchet, S., Frydman, R., and Chaouat, G. (2002). Concentration of leukaemia inhibitory factor (LIF) in uterine flushing fluid is highly predictive of embryo implantation. Human Reproduction, 17 (1), 213-218.
- Li, D., Wang, G.-Y., Dong, B.-H., Zhang, Y.-C., Wang, Y.-X., and Sun, B.-C. (2007). Biological characteristics of human placental mesenchymal stem cells and their proliferative response to various cytokines. *Cells Tissues Organs*, 186 (3), 169-179.

- Lim, C. K., Cranbrook, G. G.-H., Cranbrook, G. G.-H., Zoologiste, G. B., Cranbrook, G. G.-H., and Zoologist, G. B. (2002). Swiftlets of Borneo: builders of edible nests: Natural History Publications (Borneo) SDN., BHD, 1-171.
- Lim, H. J., and Dey, S. (2009). HB-EGF: a unique mediator of embryo-uterine interactions during implantation. *Experimental Cell Research*, 315 (4), 619-626.
- Liu, C.-M., Ma, J.-Q., and Sun, Y.-Z. (2012). Puerarin protects the rat liver against oxidative stress-mediated DNA damage and apoptosis induced by lead. *Experimental and Toxicologic Pathology*, 64 (6), 575-582.
- Liu, M.-Y., Cheng, Y.-J., Chen, C.-K., and Yang, B.-C. (2005). Coexposure of leadand lipopolysaccharide-induced liver injury in rats: involvement of nitric oxide-initiated oxidative stress and TNF-α. *Shock*, 23 (4), 360-364.
- Lonergan, P. (2011). Influence of progesterone on oocyte quality and embryo development in cows. *Theriogenology*, 76 (9), 1594-1601.
- Lopata, A., Bentin-Ley, U., and Enders, A. (2002). "Pinopodes" and implantation. *Reviews in Endocrine & Metabolic Disorders, 3* (2), 77-86.
- Luo, L., Huang, J., Fu, Y., Xu, J., and Qian, Y. (2008). Effects of tea polyphenols on ovarian development in rats. *Journal of Endocrinological Investigation*, 31 (12), 1110-1118.
- Ma, F., and Liu, D. (2012a). Extraction and determination of hormones in the edible bird's nest. *Asian Journal of Chemistry*, 24 (1), 117-120.
- Ma, F., and Liu, D. (2012b). Sketch of the edible bird's nest and its important bioactivities. *Food Research International, 48* (2), 559-567.
- Maeda,K-L.,Ohkura, S. and Tsukamura,H. (2000). physiology of reproduction.In .G.Krinke.The Laboratory Rat(pp.145-176).New York:Academic Press.
- Mahesh, V. B., Brann, D. W., and Hendry, L. B. (1996). Diverse modes of action of progesterone and its metabolites. *The Journal of Steroid Biochemistry and Molecular Biology*, 56 (1), 209-219.
- Makrigiannakis, A., Karamouti, M., Petsas, G., Makris, N., Nikas, G., and Antsaklis, A. (2009). The expression of receptivity markers in the fallopian tube epithelium. *Histochemistry and Cell Biology*, *132* (2), 159-167.
- Marcone, M. F. (2005). Characterization of the edible bird's nest the "Caviar of the East". *Food Research International, 38* (10), 1125-1134.
- Martin, L., Finn, C., and CARTER, J. (1970). Effects of progesterone and oestradiol-17β on the luminal epithelium of the mouse uterus. *Journal of Reproduction and Fertility, 21* (3), 461-469.

- Matsukawa, N., Matsumoto, M., Bukawa, W., Chiji, H., Nakayama, K., Hara, H., and Tsukahara, T. (2011). Improvement of bone strength and dermal thickness due to dietary edible bird's nest extract in ovariectomized rats. *Bioscience, Biotechnology, and Biochemistry*, *75* (3), 590-592.
- Matsumoto,H.,Sakai,K. and I washita,M.(2008). Insulin-like growth factor binding protein -1 induced decidualization of human endometriAL stromal cellS via α5β1 integrin.*Molecular Human Reproduction and Development*.14:485-489.
- McGee, E. A., and Hsueh, A. J. (2000). Initial and cyclic recruitment of ovarian follicles. *Endocrine Reviews, 21* (2), 200-214.
- McLean, A. C., Valenzuela, N., Fai, S., and Bennett, S. A. (2012). Performing vaginal lavage, crystal violet staining, and vaginal cytological evaluation for mouse estrous cycle staging identification. *Journal of Visualized Experiments: JoVE*(67), e4389:1-6.
- Meisel,R.L and SachS,B.D.In E.Knobil and J.D.Neil.(1994). physiology of reproduction(pp3-105).New York.Raven.
- Milovanov, A., Erofeeva, L., Zolotukhina, I., and Aleksandrovich, N. (2010). Morphogenesis of human placenta in the first trimester of gestation. *Morfologiia* (Saint Petersburg, Russia), 139(2), 72-76.
- Minas, V., Loutradis, D., and Makrigiannakis, A. (2005). Factors controlling blastocyst implantation. *Reproductive Biomedicine Online*, 10 (2), 205-216.
- Mondschein, J. S., and Schomberg, D. W. (1981). Growth factors modulate gonadotropin receptor induction in granulosa cell cultures. *Science*, 211 (4487), 1179-1180.
- Moneim, A. E. A., Dkhil, M. A., and Al-Quraishy, S. (2011). The protective effect of flaxseed oil on lead acetate-induced renal toxicity in rats. *Journal of Hazardous Materials*, 194, 250-255.
- Montgomery, V., Loutradis, D., Tulchinsky, D., and Kiessling, A. (1988). FSHinduced ovulation in intact and hypophysectomized mice. *Journal of Reproduction and Fertility*, 84 (1), 1-6.
- Muhammad, S. I., Ismail, M., Mahmud, R. B., Salisu, A. M., and Zakaria, Z. A. (2013). Germinated brown rice and its bioactives modulate the activity of uterine cells in oophorectomised rats as evidenced by gross cytohistological and immunohistochemical changes. *BMC Complementary and Alternative Medicine*, 13 (1), 198:1-9.
- Mujaibel, L. M., and Kilarkaje, N. (2015). Mitogen- activated protein kinase signaling and its association with oxidative stress and apoptosis in lead- exposed hepatocytes. *Environmental Toxicology*, *30* (5), 513-529.

- Nakade, U. P., Garg, S. K., Sharma, A., Choudhury, S., Yadav, R. S., Gupta, K., and Sood, N. (2015). Lead-induced adverse effects on the reproductive system of rats with particular reference to histopathological changes in uterus. *Indian Journal of Pharmacology*, 47 (1), 22-26.
- Nardo, L., Sabatini, L., Rai, R., and Nardo, F. (2002). Pinopode expression during human implantation. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 101 (2), 104-108.
- Narkar, M., Kholkute, S., Chitlange, S., and Nandedkar, T. (2006). Expression of steroid hormone receptors, proliferation and apoptotic markers in primate endometrium. *Molecular and Cellular Endocrinology, 246* (1), 107-113.
- Ng, M., Chan, K., and Kong, Y. (1986). Potentiation of mitogenic response by extracts of the swiftlet's (Collocalia) nest. *Biochemistry International*, 13 (3), 521-531.
- Nikas, G., Develioglu, O. H., Toner, J. P., and Jones Jr, H. W. (1999). Endometrial pinopodes indicate a shift in the window of receptivity in IVF cycles. *Human Reproduction*, 14 (3), 787-792.
- Nobuzane, T., Tashiro, S., and Kudo, Y. (2008). Morphologic effects of epithelial ion channels on the mouse uterus: differences between raloxifene analog (LY117018) and estradiol treatments. *American Journal of Obstetrics and Gynecology*, 199 (4), 363: e1-e6.
- Odenbro, A., and Kihlström, J. E. (1977). Frequency of pregnancy and ova implantation in triethyl lead-treated mice. *Toxicology and Applied Pharmacology, 39* (3), 359-363.
- Ohkawa, H., Ohishi, N., and Yagi, K. (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Analytical Biochemistry*, 95 (2), 351-358.
- Omobowale, T. O., Oyagbemi, A. A., Akinrinde, A. S., Saba, A. B., Daramola, O. T., Ogunpolu, B. S., and Olopade, J. O. (2014). Failure of recovery from lead induced hepatoxicity and disruption of erythrocyte antioxidant defence system in Wistar rats. *Environmental Toxicology and Pharmacology*, *37* (3), 1202-1211.
- Ostrowska, Z., Kos-Kudla, B., Swietochowska, E., Marek, B., Kajdaniuk, D., and Ciesielska-Kopacz, N. (2001). Influence of pinealectomy and long-term melatonin administration on GH-IGF-I axis function in male rats. *Neuroendocrinology Letters*, 22 (4), 255-262.
- Oyagbemi, A. A., Omobowale, T. O., Akinrinde, A. S., Saba, A. B., Ogunpolu, B. S., and Daramola, O. (2015). Lack of reversal of oxidative damage in renal tissues of lead acetate- treated rats. *Environmental Toxicology*, *30* (11), 1235-1243.
- Pallares, P., and Gonzalez-Bulnes, A. (2009). A new method for induction and synchronization of oestrus and fertile ovulations in mice by using exogenous hormones. *Laboratory Animals, 43* (3), 295-299.

- Panzan, M. Q., Mattar, R., Maganhin, C. C., dos Santos Simoes, R., Rossi, A. G. Z., da Motta, E. L. A., Baracat, E. C., and Soares-Jr, J. M. (2013). Evaluation of FAS and caspase-3 in the endometrial tissue of patients with idiopathic infertility and recurrent pregnancy loss. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 167 (1), 47-52.
- Paria, B., Huet-Hudson, Y., and Dey, S. (1993). Blastocyst's state of activity determines the" window" of implantation in the receptive mouse uterus. *Proceedings of the National Academy of Sciences*, 90 (21), 10159-10162.
- Prelle, K., Stojkovic, M., Boxhammer, K., Motlik, J., Ewald, D., Arnold, G. J., and Wolf, E. (2001). Insulin-like growth factor I (IGF-I) and long R3IGF-I differently affect development and messenger ribonucleic acid abundance for IGF-binding proteins and type I IGF receptors in in vitro produced bovine embryos. *Endocrinology*, 142 (3), 1309-1316.
- Prüss-Üstün, A., and Corvalán, C. (2006). Preventing disease through healthy environments. Towards an estimate of the environmental burden of disease. Geneva: *World Health Organization*
- Qureshi, N., and Sharma, R. (2012). Lead toxicity and infertility in female Swiss mice: a review. *Journal of Chemical, Biological and Physical Sciences (JCBPS), 2* (4), 72-82.
- Rabinowitz, M. B. (1991). Toxicokinetics of bone lead. *Environmental Health Perspectives*, 91, 33-37.
- Raden Fadzilah A'ini Abdul Kadir, Noor Aini Mohd Ain, Aini Ideris and Abdul Rahman Omar (2017). Edible Bird Nest Oncolytic Effects on Human Cancer Cell Lines In Vitro. Proceeding of 29th Congress,6-8 October 2017,Shah Alam,Selangor,Malaysia.
- Rahman, S., and Sultana, S. (2006). Chemopreventive activity of glycyrrhizin on lead acetate mediated hepatic oxidative stress and its hyperproliferative activity in Wistar rats. *Chemico-Biological Interactions*, *160* (1), 61-69.
- Rajaram, S., Baylink, D. J., and Mohan, S. (1997). Insulin-like growth factor-binding proteins in serum and other biological fluids: regulation and functions. *Endocrine Reviews*, 18 (6), 801-831.
- Rajendren, G., and Gibson, M. J. (2001). A confocal microscopic study of synaptic inputs to gonadotropin-releasing hormone cells in mouse brain: regional differences and enhancement by estrogen. *Neuroendocrinology*, *73* (2), 84-90.
- Ramathal, C. Y., Bagchi, I. C., Taylor, R. N., and Bagchi, M. K. (2010). Endometrial decidualization: of mice and men. Paper presented at the Seminars in Reproductive Medicine.

- Rehman, J., Traktuev, D., Li, J., Merfeld-Clauss, S., Temm-Grove, C. J., Bovenkerk, J. E., Pell, C. L., Johnstone, B. H., Considine, R. V., and March, K. L. (2004). Secretion of angiogenic and antiapoptotic factors by human adipose stromal cells. *Circulation*, 109 (10), 1292-1298.
- Reynolds, L. P., Kirsch, J. D., Kraft, K. C., and Redmer, D. A. (1998). Time-course of the uterine response to estradiol-17 $\beta$  in ovariectomized ewes: expression of angiogenic factors. *Biology of Reproduction*, 59 (3), 613-620.
- Rider, V., Carlone, D., and Foster, R. (1997). Oestrogen and progesterone control basic fibroblast growth factor mRNA in the rat uterus. *Journal of Endocrinology*, 154 (1), 75-84.
- Robb, L., Li, R., Hartley, L., Nandurkar, H. H., Koentgen, F., and Begley, C. G. (1998). Infertility in female mice lacking the receptor for interleukin 11 is due to a defective uterine response to implantation. *Nature Medicine*, 4 (3), 303-308.
- Rogers, P. A., Donoghue, J. F., Walter, L. M., and Girling, J. E. (2009). Endometrial angiogenesis, vascular maturation, and lymphangiogenesis. *Reproductive Sciences*, *16* (2), 147-151.
- Roh, K.-B., Lee, J., Kim, Y.-S., Park, J., Kim, J.-H., Lee, J., and Park, D. (2012). Mechanisms of edible bird's nest extract-induced proliferation of human adipose-derived stem cells. *Evidence-Based Complementary and Alternative Medicine*, 2012, 1-11
- Romeu, L. R. G., da Motta, E. L. A., Maganhin, C. C., Oshima, C. T., Fonseca, M. C., Barrueco, K. F., Simoes, R. S., Pellegrino, R., Baracat, E. C., and Soares-Junior, J. M. (2011). Effects of melatonin on histomorphology and on the expression of steroid receptors, VEGF, and PCNA in ovaries of pinealectomized female rats. *Fertility and Sterility*, 95 (4), 1379-1384.
- Ruan, Y. C., Guo, J. H., Liu, X., Zhang, R., Tsang, L. L., Da Dong, J., Chen, H., Yu, M. K., Jiang, X., and Zhang, X. H. (2012). Activation of the epithelial Na+ channel triggers prostaglandin E2 release and production required for embryo implantation. *Nature Medicine*, 18 (7), 1112-1117.
- Rzymski, P., Rzymski, P., Tomczyk, K., Niedzielski, P., Jakubowski, K., Poniedziałek, B., and Opala, T. (2014). Metal status in human endometrium: relation to cigarette smoking and histological lesions. *Environmental Research*, 132, 328-333.
- Rzymski, P., Tomczyk, K., Poniedzialek, B., Opala, T., and Wilczak, M. (2015). Impact of heavy metals on the female reproductive system. *Annals of Agricultural and Environmental Medicine*, 22 (2),259-264.
- Saengkrajang, W., Matan, N., and Matan, N. (2013). Nutritional composition of the farmed edible bird's nest (Collocalia fuciphaga) in Thailand. *Journal of Food Composition and Analysis*, 31 (1), 41-45.

- Sakr, S., Bayomy, M., & El-Morsy, A. (2015). Rosemary extract ameliorates cadmium-induced histological changes and oxidative damage in the liver of albino rat. *The Journal of Basic & Applied Zoology*, 71, 1-9.
- Salleh, N., Baines, D., Naftalin, R., and Milligan, S. (2005). The hormonal control of uterine luminal fluid secretion and absorption. *The Journal of Membrane Biology*, 206 (1), 17-28.
- Sarkar, D., Chiappa, S. A., Fink, G., and SHERWOOD, N. M. (1976). Gonadotropinreleasing hormone surge in pro-oestrous rats. *Nature*, 264 (5585), 461-463.
- Satarug, S., Baker, J. R., Reilly, P. E., Moore, M. R., and Williams, D. J. (2002). Cadmium levels in the lung, liver, kidney cortex, and urine samples from Australians without occupational exposure to metals. *Archives of Environmental Health: An International Journal*, 57 (1), 69-77.
- Schauer, R. (2000). Achievements and challenges of sialic acid research. *Glycoconjugate Journal*, 17 (7-9), 485-499.
- Schlafke, S., and Enders, A. C. (1975). Cellular basis of interaction between trophoblast and uterus at implantation. *Biology of Reproduction*, 12 (1), 41-65.
- Schlessinger, J., Schreiber, A. B., Levi, A., Lax, I., Libermann, T., and Yarden, Y. (1983). Regulation of cell proliferation by epidermal growth factor. *Critical Reviews in Biochemistry*, 14 (2), 93-111.
- Schmidt, C. M., Blount, J. D., and Bennett, N. C. (2014). Reproduction is associated with a tissue-dependent reduction of oxidative stress in eusocial female Damaraland mole-rats (Fukomys damarensis). *Plos one*, 9 (7), e103286.
- Schwarzkopf, M., Knobeloch, K.-P., Rohde, E., Hinderlich, S., Wiechens, N., Lucka, L., Horak, I., Reutter, W., and Horstkorte, R. (2002). Sialylation is essential for early development in mice. *Proceedings of the National Academy of Sciences, 99* (8), 5267-5270.
- Sen, S., Chowdhury, G., and Chowdhury, M. (2001). Sialic acid binding protein of human endometrium: Its regulation by steroids. *Molecular and Cellular Biochemistry*, 221 (1), 17-23.
- Sferruzzi- Perri, A., Owens, J., Pringle, K., and Roberts, C. (2011). The neglected role of insulin- like growth factors in the maternal circulation regulating fetal growth. *The Journal of Physiology*, 589 (1), 7-20.
- Sharma, R., Garu, U., and Panwar, K. (2012). Developing gonads and lead exposure. *World Journal of Environmental Biosciences*, *1*, 30-37.
- Sharma, S., and Singh, B. (2014). Effects of acute and chronic lead exposure on kidney lipid peroxidation and antioxidant enzyme activities in BALB-C mice (Mus musculus). *International Journal of Science and Research*, *3*, 1564-1566.

- Shweiki, D., Itin, A., Neufeld, G., Gitay-Goren, H., and Keshet, E. (1993). Patterns of expression of vascular endothelial growth factor (VEGF) and VEGF receptors in mice suggest a role in hormonally regulated angiogenesis. *Journal of Clinical Investigation*, 91 (5), 2235-2243.
- Silva, N., Peiris- John, R., Wickremasinghe, R., Senanayake, H., and Sathiakumar, N. (2012). Cadmium a metalloestrogen: are we convinced? *Journal of Applied Toxicology*, 32 (5), 318-332.
- Simmons, R. M., Erikson, D. W., Kim, J., Burghardt, R. C., Bazer, F. W., Johnson, G. A., and Spencer, T. E. (2009). Insulin-like growth factor binding protein-1 in the ruminant uterus: potential endometrial marker and regulator of conceptus elongation. *Endocrinology*, 150 (9), 4295-4305.
- Singh, M. K., Dwivedi, S., Yadav, S. S., Sharma, P., and Khattri, S. (2014). Arsenicinduced hepatic toxicity and its attenuation by fruit extract of Emblica officinalis (amla) in mice. *Indian Journal of Clinical Biochemistry*, 29 (1), 29-37.
- Singh, M., Chaudhry, P., and Asselin, E. (2011). Bridging endometrial receptivity and implantation: network of hormones, cytokines, and growth factors. *Journal of Endocrinology, 210* (1), 5-14.
- Song, H., and Lim, H. (2006). Evidence for heterodimeric association of leukemia inhibitory factor (LIF) receptor and gp130 in the mouse uterus for LIF signaling during blastocyst implantation. *Reproduction*, 131 (2), 341-349.
- Song, H., Han, K., and Lim, H. (2007). Progesterone supplementation extends uterine receptivity for blastocyst implantation in mice. *Reproduction*, 133 (2), 487-493.
- Song, H., Lim, H., Das, S. K., Paria, B. C., and Dey, S. K. (2000). Dysregulation of EGF family of growth factors and COX-2 in the uterus during the preattachment and attachment reactions of the blastocyst with the luminal epithelium correlates with implantation failure in LIF-deficient mice. *Molecular Endocrinology*, 14 (8), 1147-1161.
- Spanos, S., Becker, D. L., Winston, R. M., and Hardy, K. (2000). Anti-apoptotic action of insulin-like growth factor-I during human preimplantation embryo development. *Biology of Reproduction, 63* (5), 1413-1420.
- Spencer, T. E. (2013). Early pregnancy: Concepts, challenges, and potential solutions. *Animal Frontiers*, *3* (4), 48-55.
- Spencer, T. E., Sandra, O., and Wolf, E. (2008). Genes involved in conceptus– endometrial interactions in ruminants: insights from reductionism and thoughts on holistic approaches. *Reproduction*, 135 (2), 165-179.
- Spencer, T., and Bazer, F. (2004). Uterine and placental factors regulating conceptus growth in domestic animals. *Journal of Animal Science, 82* (13_suppl), E4-E13.

- Speroff, L., and Fritz, M. A. (2005). *Clinical gynecologic endocrinology and infertility*: Lippincott Williams & Wilkins.
- Stavreus-Evers, A., Aghajanova, L., Brismar, H., Eriksson, H., Landgren, B.-M., and Hovatta, O. (2002). Co-existence of heparin-binding epidermal growth factorlike growth factor and pinopodes in human endometrium at the time of implantation. *MHR: Basic Science of Reproductive Medicine*, 8 (8), 765-769.
- Stewart, C. L., Kaspar, P., Brunet, L. J., Bhatt, H., Gadi, I., Köntgen, F., and Abbondanzo, S. J. (1992). Blastocyst implantation depends on maternal expression of leukaemia inhibitory factor. *Nature*, 359 (6390), 76-79.
- Su, G. L. (2002). Lipopolysaccharides in liver injury: molecular mechanisms of Kupffer cell activation. American Journal of Physiology-Gastrointestinal and Liver Physiology, 283(2), G256-G265.
- Suzuki, N., Zara, J., Sato, T., Ong, E., Bakhiet, N., Oshima, R. G., Watson, K. L., and Fukuda, M. N. (1998). A cytoplasmic protein, bystin, interacts with trophinin, tastin, and cytokeratin and may be involved in trophinin-mediated cell adhesion between trophoblast and endometrial epithelial cells. Proceedings of the National Academy of Sciences, 95 (9), 5027-5032.
- Syarmila, Etty., I.K., M.H. Nurfatin, M. Masitah, Z. Farahniza, A. Mohd. Khan, M.K. Zalifah and B. Abdul Salam (2014). Edible bird nest hydrolysates as natural antioxidative peptides (poster). Proceedings of the Edible Bird Nest Industry Conference, 25-26 November 2014, Putrajaya, Malaysia
- Tamura, H., Nakamura, Y., Korkmaz, A., Manchester, L. C., Tan, D.-X., Sugino, N., and Reiter, R. J. (2009). Melatonin and the ovary: physiological and pathophysiological implications. *Fertility and Sterility*, 92 (1), 328-343.
- Tamura, N., Sugihara, K., Akama, T. O., and Fukuda, M. N. (2011). Trophininmediated cell adhesion induces apoptosis of human endometrial epithelial cells through PKC-δ. *Cell Cycle*, 10 (1), 135-143.
- Tan, J., Paria, B. C., Dey, S. K., and Das, S. K. (1999). Differential uterine expression of estrogen and progesterone receptors correlates with uterine preparation for implantation and decidualization in the mouse. *Endocrinology*, 140 (11), 5310-5321.
- Thavamanithevi, S, R. Sarifah, C.G. Lim, M. Theanmalar, M.S. Aidawati, M. Durgah Devi and A.A. Saleha. (2014). Characterization And Standardization Of Edible Birds Nest (EBN)-Determination of Sialic Acid. Proceedings of the Edible Bird Nest Industry Conference, 25-26 November 2014, Putrajaya.
- Threadgill, D. W., Dlugosz, A. A., Hansen, L. A., Tennenbaum, T., Lichti, U., Yee, D., LaMantia, C., Mourton, T., Herrup, K., and Harris, R. C. (1995). Targeted disruption of mouse EGF receptor: effect of genetic background on mutant phenotype. *Science*, 230-234.

- Tilly, J. L., and Tilly, K. (1995). Inhibitors of oxidative stress mimic the ability of follicle-stimulating hormone to suppress apoptosis in cultured rat ovarian follicles. *Endocrinology*, 136 (1), 242-252.
- Tilly, J. L., Billig, H., Kowalski, K. I., and Hsueh, A. (1992b). Epidermal growth factor and basic fibroblast growth factor suppress the spontaneous onset of apoptosis in cultured rat ovarian granulosa cells and follicles by a tyrosine kinase-dependent mechanism. *Molecular Endocrinology*, *6* (11), 1942-1950.
- Tilly, J., Kowalski, K., Schomberg, D., and Hsueh, A. (1992a). Apoptosis in atretic ovarian follicles is associated with selective decreases in messenger ribonucleic acid transcripts for gonadotropin receptors and cytochrome P450 aromatase. *Endocrinology*, 131 (4), 1670-1676.
- Tranguch, S., Daikoku, T., Guo, Y., Wang, H., and Dey, S. (2005). Molecular complexity in establishing uterine receptivity and implantation. *Cellular and Molecular Life Sciences*, 62 (17), 1964-1973.
- Ullrich, A., and Schlessinger, J. (1990). Signal transduction by receptors with tyrosine kinase activity. *Cell*, *61* (2), 203-212.
- Utarabhand, P., Rittidach, W., Rattanaporn, O., Runsaeng, P., and Hedrick, J. L. (2017). Sialic acid-specific lectin participates in an immune response and ovarian development of the banana shrimp Fenneropenaeus merguiensis. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 203, 132-140.
- Vaziri, N., and Khan, M. (2007). Interplay of reactive oxygen species and nitric oxide in the pathogenesis of experimental lead- induced hypertension. *Clinical and Experimental Pharmacology and Physiology*, 34 (9), 920-925.
- Vermande-Van Eck, G. J., and Meigs, J. W. (1960). Changes in the ovary of the rhesus monkey after chronic lead intoxication. *Fertility and Sterility*, 11 (2), 223-234.
- Vértes, Z., Sándor, A., Kovács, K., Oszter, A., Környei, J., Kovács, S., and Vértes, M. (2000). Epidermal growth factor influenced by opioid peptides in immature rat uterus. *Journal of Endocrinological Investigation, 23* (8), 502-508.
- Vimala, B., Hussain, H., and Nazaimoon, W. W. (2012). Effects of edible bird's nest on tumour necrosis factor-alpha secretion, nitric oxide production and cell viability of lipopolysaccharide-stimulated RAW 264.7 macrophages. *Food And Agricultural Immunology, 23* (4), 303-314.
- Vitale, A. M., Gonzalez, O. M., Parborell, F., Irusta, G., Campo, S., and Tesone, M. (2002). Inhibin a increases apoptosis in early ovarian antral follicles of diethylstilbestrol-treated rats. *Biology of Reproduction*, 67 (6), 1989-1995.
- Walmer, D. K., Wrona, M. A., Hughes, C. L., and Nelson, K. G. (1992). Lactoferrin expression in the mouse reproductive tract during the natural estrous cycle: correlation with circulating estradiol and progesterone. *Endocrinology*, 131 (3), 1458-1466.

- Wang, B., and Brand-Miller, J. (2003). The role and potential of sialic acid in human nutrition. *European journal of clinical nutrition*, 57 (11), 1351-1369.
- Wang, H., and Dey, S. K. (2006). Roadmap to embryo implantation: clues from mouse models. *Nature Reviews Genetics*, 7 (3), 185-199.
- Wang, J., Mayernik, L., Schultz, J. F., and Armant, D. R. (2000). Acceleration of trophoblast differentiation by heparin-binding EGF-like growth factor is dependent on the stage-specific activation of calcium influx by ErbB receptors in developing mouse blastocysts. *Development*, 127 (1), 33-44.
- Wang, N., Luo, L.-L., Xu, J.-J., Xu, M.-Y., Zhang, X.-M., Zhou, X.-L.,Liu, W-J and Fu, Y.-C. (2014). Obesity accelerates ovarian follicle development and follicle loss in rats. Metabolism, 63(1), 94-103.
- Wang, X., Matsumoto, H., Zhao, X., Das, S. K., and Paria, B. C. (2004). Embryonic signals direct the formation of tight junctional permeability barrier in the decidualizing stroma during embryo implantation. *Journal of Cell Science*, 117 (1), 53-62.
- Wato, E., Asahiyama, M., Suzuki, A., Funyu, S., and Amano, Y. (2009). Collaborative work on evaluation of ovarian toxicity 9) Effects of 2-or 4-week repeated dose studies and fertility study of di (2-ethylhexyl) adipate (DEHA) in female rats. *The Journal of Toxicological Sciences, 34* (Special), SP101-SP109.
- Wdowiak, A., Lewicka, M., Plewka, K., and Bakalczuk, G. (2013). Nicotinism and quality of embryos obtained in in-vitro fertilization programmes. *Annals of Agricultural and Environmental Medicine*, 20 (1),82-85.
- Weiner, I. D., Mitch, W. E., and Sands, J. M. (2015). Urea and ammonia metabolism and the control of renal nitrogen excretion. *Clinical Journal of the American Society of Nephrology*, 10 (8), 1444-1458.
- Wiebe, J. P., and Barr, K. J. (1988). Effect of prenatal and neonatal exposure to lead on the affinity and number of estradiol receptors in the uterus. *Journal of Toxicology and Environmental Health, Part A Current Issues, 24* (4), 451-460.
- Williams, P. J., Bulmer, J. N., Innes, B. A., and Broughton Pipkin, F. (2011). Possible roles for folic acid in the regulation of trophoblast invasion and placental development in normal early human pregnancy. *Biology of Reproduction*, 84(6), 1148-1153.
- Wimsatt, W. A. (1975). Some comparative aspects of implantation. *Biology of Reproduction*, 12 (1), 1-40.
- Winwood, P. J., and Arthur, M. J. (1993). Kupffer cells: their activation and role in animal models of liver injury and human liver disease. Paper presented at the Seminars in liver disease.

- Wong, H.-L., Siu, W.-s., Shum, W.-t., Gao, S., Leung, P.-C., and Ko, C.-H. (2012). Application of Chinese herbal medicines to revitalize adult stem cells for tissue regeneration. *Chinese Journal of Integrative Medicine*, 18 (12), 903-908.
- Wordinger, R. J., Smith, K. J., Bell, C., and Chang, I.-F. C. (1994). The immunolocalization of basic fibroblast growth factor in the mouse uterus during the initial stages of embryo implantation. *Growth Factors, 11* (3), 175-186.
- Wu, R., and Zhou, F. (2004). Insulin-like growth factor II and its receptor gene expression in the endometrium of women with unexplained infertility. *Zhonghua fu Chan ke za zhi, 39* (4), 242-245.
- Xie, H., Wang, H., Tranguch, S., Iwamoto, R., Mekada, E., DeMayo, F. J., Lydon, J.
   P., Das, S. K., and Dey, S. K. (2007). Maternal heparin-binding-EGF deficiency limits pregnancy success in mice. Proceedings of the National Academy of Sciences, 104 (46), 18315-18320.
- Yanagimachi, R. (1994). Mammalian fertilization. *The Physiology of Reproduction*, 189-317.
- Yang, J. Z., Ajonuma, L. C., Tsang, L. L., Lam, S. Y., Rowlands, D. K., Ho, L. S., Zhou, C. X., Chung, Y. W., and Chan, H. C. (2004). Differential expression and localization of CFTR and ENaC in mouse endometrium during preimplantation. *Cell Biology International*, 28 (6), 433-439.
- Yarden, Y. (2001). The EGFR family and its ligands in human cancer: signalling mechanisms and therapeutic opportunities. *European Journal of Cancer, 37*, 3-8.
- Yepiz-Plascencia, G., Vargas-Albores, F., Jimenez-Vega, F., Ruiz-Verdugo, L. M., and Romo-Figueroa, G. (1998). Shrimp plasma HDL and β-glucan binding protein (BGBP): comparison of biochemical characteristics. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 121 (3), 309-314.
- Yew, M. Y., Koh, R. Y., Chye, S. M., Othman, I., and Ng, K. Y. (2014). Edible bird's nest ameliorates oxidative stress-induced apoptosis in SH-SY5Y human neuroblastoma cells. *BMC Complementary and Alternative Medicine*, *14* (1), 391:1-12.
- Yida, Z., Imam, M. U., Ismail, M., Hou, Z., Abdullah, M. A., Ideris, A., and Ismail, N. (2015a). Edible Bird's Nest attenuates high fat diet-induced oxidative stress and inflammation via regulation of hepatic antioxidant and inflammatory genes. *BMC Complementary and Alternative Medicine*, 15 (1), 310:1-7.
- Yida, Z., Imam, M. U., Ismail, M., Ooi, D.-J., Sarega, N., Azmi, N. H., Ismail, N., Chan, K. W., Hou, Z., and Yusuf, N. B. (2015b). Edible bird's nest prevents high fat diet-induced insulin resistance in rats. *Journal of Diabetes Research*, 2015,1-11.

- Yoshida, K., Chambers, I., Nichols, J., Smith, A., Saito, M., Yasukawa, K., Shoyab, M., Taga, T., and Kishimoto, T. (1994). Maintenance of the pluripotential phenotype of embryonic stem cells through direct activation of gp130 signalling pathways. *Mechanisms of Development*, 45 (2), 163-171.
- Yoshinaga, K. (2010). Research on blastocyst implantation essential factors (BIEFs). *American journal of reproductive immunology, 63* (6), 413-424.
- Yue, Z. P., Yang, Z. M., Li, S. J., Wang, H. B., and Harper, M. J. (2000). Epidermal growth factor family in rhesus monkey uterus during the menstrual cycle and early pregnancy. *Molecular Reproduction and Development*, 55 (2), 164-174.
- Zelikoff, J., Li, J., Hartwig, A., Wang, X., Costa, M., and Rossman, T. (1988). Genetic toxicology of lead compounds. *Carcinogenesis*, 9 (10), 1727-1732.
- Zhang, Q., and Paria, B. C. (2006). Importance of uterine cell death, renewal, and their hormonal regulation in hamsters that show progesterone-dependent implantation. *Endocrinology*, 147 (5), 2215-2227.
- Zhang, S., Lin, H., Kong, S., Wang, S., Wang, H., Wang, H., and Armant, D. R. (2013). Physiological and molecular determinants of embryo implantation. *Molecular Aspects of Medicine*, 34 (5), 939-980.
- Zheng, J., Redmer, D. A., Killilea, S. D., and Reynolds, L. P. (1998). Characterization of heparin-binding endothelial mitogen (s) produced by the ovine endometrium during early pregnancy. *Biochemistry and Cell Biology*, 76 (1), 89-96.
- Zheng, X.-Y., Chen, G.-A., and Wang, H.-Y. (2004). Expression of cystic fibrosis transmembrane conductance regulator in human endometrium. *Human Reproduction*, 19 (12), 2933-2941.
- Zhiping, H., Imam, M. U., Ismail, M., Ismail, N., Yida, Z., Ideris, A., Sarega, N., and Mahmud, R. (2015). Effects of edible bird's nest on hippocampal and cortical neurodegeneration in ovariectomized rats. *Food & Function*, 6 (5), 1701-1711.
- Zucchetto, A., Serraino, D., Polesel, J., Negri, E., De Paoli, A., Dal Maso, L., Montella, M., La Vecchia, C., Franceschi, S., and Talamini, R. (2009). Hormone-related factors and gynecological conditions in relation to endometrial cancer risk. *European Journal of Cancer Prevention*, 18 (4), 316-321.
- Żukowska-Arendarczyk, M. (1981). Effect of hypophyseal gonadotropins (FSH and LH) on the ovaries of the sand shrimp Crangon crangon (Crustacea: Decapoda). *Marine Biology*, *63* (3), 241-247.