



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION OF PEPTIDES AND PROTEINS FROM
EPIDERMAL MUCUS OF CLIMBING PERCH *Anabas testudineus*
(Bloch, 1972) WITH ANTIBACTERIAL AND IMMUNOMODULATORY
PROPERTIES***

AGHARID ALI HUSSEIN

FPV 2018 18



**CHARACTERIZATION OF PEPTIDES AND PROTEINS FROM
EPIDERMAL MUCUS OF CLIMBING PERCH *Anabas testudineus*
(Bloch, 1972) WITH ANTIBACTERIAL AND IMMUNOMODULATORY
PROPERTIES**

By

AGHARID ALI HUSSEIN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

April 2018

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

In the name of Allah, the most compassionate, the most merciful

In memory of my beloved parents, who unconditionally supported me throughout my academic career. Though they are no longer with me physically, their love was ever-present, and I have fulfilled their aspirations for me. May Allah (swt) reunite us in Paradise



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**CHARACTERIZATION OF PEPTIDES AND PROTEINS FROM
EPIDERMAL MUCUS OF CLIMBING PERCH *Anabas testudineus*
(Bloch, 1972) WITH ANTIBACTERIAL AND IMMUNOMODULATORY
PROPERTIES**

By

AGHARID ALI HUSSEIN

April 2018

Chairman : Associate Professor Hassan Hj Mohd Daud, PhD
Faculty : Veterinary Medicine

The skin of fish with its large mucosal surface contains a variety of biologically active compounds and antimicrobial peptides that are constitutively expressed to provide protection against potential pathogenic microbes. Climbing perch (*Anabas testudineus*) is an obligate air breather and tolerates extremely unfavourable water conditions. Its body is covered by a thick coat of mucus composed of mucopolysaccharides, lipids and proteins making the fish a suitable candidate to obtain bioactive compounds such as antibacterial proteins and peptides. Based on this background, the assessment of the antimicrobial and immunomodulatory effects of bioactive proteins and peptides in crude mucous extracts derived from epidermis of the climbing perch is an important step. Hence, the aim of this study was to determine the antimicrobial and immunomodulatory effects of the bioactive crude mucous extract from epidermis of climbing perch in order to test their efficacy both *in vivo* and *in vitro*.

The bioactive crude mucous extract was prepared in aqueous and acidified form by heating moderately, followed by assessing their immune components as well as evaluating their antibacterial activity and determining some immune-related enzymes using well agar diffusion test, respectively. Proteomic analysis was also conducted to estimate the protein concentration including SDS-PAGE, haemolytic activity of the acidic crude extract as well as histopathological analysis. The minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) as well as NaCl₂ tolerance of the bioactive crude were also conducted. Antibacterial activity was found to be statistically significant ($P < 0.05$) showing maximum activity against *P. aeruginosa* ATCC10145. However, no haemolytic

activity was observed for the acidified crude extract on horse RBCs. Total soluble protein concentration by OPA assay and SDS-PAGE profile revealed 13 bands (245-11 kDa) distinguished using NuPAGE 4-12% Bis-Tris (Novex Invitrogen) gel. The MIC and MBC of mucous crude extract as determined by resazurin-based microliter dilution assay was found to have a similar value (2.5mg/ml) against multidrug resistance (MDR) *P. aeruginosa* ATCC10145 in addition to salt tolerance at physiological solution. Assessment of the immune-related enzymes activities of protease, lysozyme and esterase in bioactive crude indicated significant roles in disease defence mechanism. Similarly, antimicrobial proteins and peptides isolated by sepPak C18 and Ultrafiltration 30kDa and Superdex™peptide showed that the fractions in the first peak refer to antibacterial properties against *P. aeruginosa* ATCC 10145. Furthermore, bioactive fractions were run through Q-TOFLC/MS with subsequent digestion by trypsin to identify the antimicrobial proteins and peptides. This was the first time to identify novel ApolipoproteinA-1 (Antimicrobial proteins) and Haemoglobin subunit beta (Fragment) as well as β -actin (Fragment) and Elongation factor 1- α (Fragment) and Cytochrome-C oxidase subunit1(Fragment) in mucous crude extract derived from epidermal scraping of climbing perch. The *de novo* peptide AAGPKGPLGPR was selected among other *de novo* peptides as it was predicted to be antibacterial peptide according on its net charge (+2), low residues (11), the hydrophobic ratio of 27% and richness in proline and glycine. Following the proteomic analysis of bioactive crude and estimation of immune related enzymes activities, the efficacy of antimicrobial proteins and bioactive potential peptide in the crude (AMPPC) against *P. aeruginosa* ATCC 10145 was applied in an animal model. The inoculum size for MDR of *P. aeruginosa* ATCC 1014 was determined at 2×10^7 CFU/mouse using Reed and Muench method and oral administration AMPPC at 6.38mg/mouse was nontoxic to male ICR mice, with no change in physical state and body weight as compared to the control group, PBS only. Moreover, liver function enzymes, ALT and AST were not significantly different when compared to the control group mice given PBS only. In addition, kidney function indicator, creatinine and urea levels revealed no significant difference. Histological examination of liver and kidney manifested normal histological architectures. The efficacy test of AMPPC showed that mice injected with 2×10^7 CFU/mouse of *P. aeruginosa* ATCC 10145 in 0.2 ml of AMPPC, CE and PBS solutions and evaluated after 48 hrs, indicated there were significant decreased ($P < 0.05$) in mortality rate and bacteria count in organs and blood samples, liver enzymes (AST and ALT), renal functions (creatinine and urea levels) and pro-inflammatory cytokines TNF- α and IL-6 in treatment groups by bioactive crude mucous extract, as compared to non-treatment and antibiotic treatment groups.

In conclusion, the epidermal mucous of the climbing perch are natural sources of antimicrobial proteins and peptides. Similarly, the proteomic analysis of bioactive crude extracts derived from the epidermal mucous may serve as an opening to further investigations to determine their potential medical application as antibacterial agent for preventing and elimination of *P. aeruginosa* sepsis.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN PEPTID DAN PROTEIN DARI MUKUS EPIDERMAL IKAN
PUYU, *Anabas testudineus* YANG BERSIFAT ANTIBAKTERIA DAN
IMUNOMODULATORI**

Oleh

AGHARID ALI HUSSEIN

April 2018

Pengerusi : Profesor Madya Hassan Hj Mohd Daud, PhD
Fakulti : Perubatan Veterinar

Kulit ikan mempunyai permukaan lapisan mukosa yang luas mengandungi pelbagai sebatian biologiikal aktif serta peptid antimikrob yang terungkap berurutan untuk memberi perlindungan terhadap mikrob berpotensi patogenik. Ikan puyu (*Anabas testudineus*) adalah ikan obligat sedutan udara dan tahan keadaan air tidak sesuai yang ekstrem. Badannya diseliputi oleh lapisan tebal mukus terdiri daripada mukopolisakarida, lipid dan protein yang menjadikan ikan tersebut calon yang sesuai untuk mendapatkan sebatian bioaktif seperti protein antibakteria dan peptid. Berdasarkan latarbelakang ini penilaian terhadap kesan antimikrob dan imunomodulatori bioaktif protein dan peptid dalam ekstrak kasar terbit dari epidermis ikan puyu adalah satu langkah yang penting. Olehitu tujuan kajian ini adalah untuk menentukan kesan antimikrob dan imunomodulatori ekstrak bioaktif mukus mentah dari epidermis ikan puyu untuk menguji keberkesanan dalam *in vivo* dan *in vitro*.

Ekstrak bioaktif mukus kasar telah disediakan dalam bentuk akuas dan terasid dengan memanaskan secara sederhana, diikuti masing-masing dengan penilaian komponen imun dan penilaian aktiviti antibakteria dan menentukan beberapa enzim berkaitan dengan keimunan menggunakan ujian resapan gel agar-agar. Analisis proteomik juga dijalankan untuk menganggarkan kepekatan protein seperti SDS-PAGE, aktiviti hemolitik ekstrak berasid mentah dan juga analisis histopatologiikal. Ujian kepekatan perencat minimal (MIC) dan kepekatan bakterisidal minimal (MBC) dan ujian tolerans NaCl₂ juga dijalankan. Ujian antibakteria menunjukkan aktiviti maksima terhadap *Pseudomonas aeruginosa* ATCC10145 dengan statistik yang bererti ($p < 0.05$). Walaubagaimana pun tiada aktiviti hemolisis untuk ekstrak terasid kasar terhadap sel darah merah kuda. Kepekatan protein total terlarut melalui

asei OPA dan profil SDS-PAGE menunjukkan 13 jalur (11-245 kDa) berbeza menggunakan NuPAGE 4-12% gel Bis-Tris (Novex Invitrogen). Nilai MIC dan MBC ekstrak mukus kasar sebagaimana ditentukan dengan asej larutan mikroliter berdasar resazurin menunjukkan nilai yang serupa (2.5 mg/ml), terhadap kerintangan multidrug (MDR) *P. aeruginosa* ATCC10145, tambahan kepada toleransi terhadap garam dalam larutan fisiologi. Penilaian enzim berkait imun menunjukkan nilai-nilai bererti dalam protease, lisozim dan esterase yang mana menandakan peranan mereka dalam mekanisme ketahanan penyakit. Dalam keadaan yang sama protein dan peptid antimikrob yang dipencil dengan sePak C18, pengultraempuran 30kDa dan SuperdexTMpeptide menunjukkan fraksi dalam puncak pertama merujuk pada sifat antibakteria terhadap *P. aeruginosa* ATCC 10145. Tambahan lagi, fraksi bioaktif diuji melalui Q-TOFLC/MS dan diikuti oleh pencernaan dengan tripsin bagi mengenalpasti protein dan peptid antimikrob. Ini adalah kali pertama langkah mengenalpasti ApolipoproteinA-1 yang novel (protein antimikrob), β -haemoglobin subunit (fragmen), β -actin (fragmen), faktor pemanjangan 1- α (fragmen) dan juga sitokrom-C oksidase (fragmen) dalam ekstrak kasar mukus dari kikisan epidermal ikan puyu. Peptid *de novo* AAGPKGPLGPR telah dipilih dari peptid *de novo* yang lain kerana ianya diramal sebagai peptid antibakteria mengikut cas bersih (+2), residu rendah (11), nisbah hidrofobik bernilai 27% dan kaya dengan prolina dan glisina. Selepas analisis proteomik ekstrak bioaktif kasar dan anggaran aktiviti enzim berkait keimunan, keberkesanan protein antimikrob dan potensi bioaktif peptid (AMPPC) dalam ekstrak kasar terhadap *P. aeruginosa* ATCC 10145 dijalankan dalam model haiwan. Saiz inokulum untuk MDR *P. aeruginosa* ATCC 10145 ditentukan pada 2×10^7 CFU/mencit menggunakan kaedah Reed dan Muench. Pemberian oral AMPPC pada 6.38mg/mencit adalah tidak toksid terhadap mencit ICR jantan, dengan tiada perubahan keadaan fizikal dan berat badan sebagaimana dibandingkan dengan kumpulan kawalan iaitu PBS sahaja. Juga enzim fungsi hepar, ALT dan AST tidak menunjukkan perubahan bererti apabila dibandingkan dengan kumpulan mencit kawalan yang diberi PBS sahaja. Tambahan lagi penunjuk fungsi ginjal iaitu paras kreatinina dan urea menunjukkan tiada berbezaan bererti. n Pemeriksaan histologi hepar dan ginjal mempamerkan arkitektur yang normal. Ujian keberkesanan AMPPC menunjukkan mencit yang disuntik dengan 2×10^7 CFU/mencit *P. aeruginosa* ATCC 10145 dalam 0.2 ml of larutan AMPPC, CE dan PBS dan dinilai selepas 48 jam, menunjukkan terdapat penurunan bererti ($P < 0.05$) dalam kadar mortaliti dan bilangan bakteria dalam sampel organ dan darah, enzim hepar (AST dan ALT), fungsi ginjal (paras kreatinina dan urea) dan pro-inflamatori sitokin TNF- α dan IL-6 dalam kumpulan dirawat dengan ekstrak kasar bioaktif mukus apabila dibanding dengan kumpulan tanpa rawatan dan dirawat dengan antibiotik.

Akhir kata, mukus dari epidermal ikan puyu adalah sumber asli protein dan peptid antimikrob. Secara yang serupa analisis proteomik ekstrak bioaktif kasar dari mukus epidermal boleh bertindak sebagai ruang untuk sisatan lanjut untuk menentukan kegunaan perubatan sebagai agen antibakteria bagi mengelak dan menyingkir sepsis *P. aeruginosa*.

ACKNOWLEDGEMENTS

All thanks and praise go to Allah (swt) for which he has made easy, supported and has sustained.

There is no measure of gratitude that can express my thanks to all who have helped throughout my academic journey. Foremost, I would like to sincerely thank my esteemed my supervisor Assoc. Professor Dr. Hassan B HJ Mohd Daud for all of his assistance, advice, guidance, and support to refine my Thesis and I pray and ask Allah (swt) to be pleased with him.

I also offer my thanks to the supervisor, committee: Assoc. Professor Dr. Siti Khairani Bejo Dr. Farina Mustafa Kamal who spent their time to review my thesis and provided invaluable suggestions and feedback.

Additional thanks go to my former professors and supervisors in Iraq who have propelled me throughout my academic career during both my Bachelor's and Master's degrees at the University of Baghdad, and instilled the love of for this field and research into my heart.

I would like to express my sincere thanks to Ministry of Higher Education, Iraq for the grant of scholarship for this study

My thanks go to Professor Dr. Karim Al-Jashaami for his actively helped to evaluate the histological slides. My Special thanks for my colleague Dr. Kareem, Dr. Bashiro and Dr. Amaal and all my colleagues for their invaluable help and for giving hand in solving problems

My thanks to Miss Farahayu and Miss Azira at Malaysia Genome Institute, Miss Tee at LCMS Laboratory, Monash University Malaysia, Miss. Umaira at Microbial Culture Collection Unit, IBS for their help.

I would also like to highlight my deep appreciation and recognition for all the anonymous and unknown contributors on the World Wide Web who have made available a countless wealth of knowledge which enriched my thesis research.

Last but not least, sincere thanks goes to my husband, my children, my brothers and sisters, my extended family who have provided me with all kinds of wonderful support, valuable aid, who have looked after my family during my four-year doctoral journey.



I certify that a Thesis Examination Committee has met on 20 April 2018 to conduct the final examination of Agharid Ali Hussein on her thesis entitled "Characterization of Peptides and Proteins from Epidermal Mucus of Climbing Perch *Anabas testudineus* (Bloch, 1792) with Antibacterial and Immunomodulatory Properties" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Abdul Aziz bin Saharee, PhD
Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Annie Christianus, PhD
Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Abdul Rahman bin Omar, PhD
Professor
Institute of Bioscience
Universiti Putra Malaysia
(Internal Examiner)

K. Pani Prasad, PhD
Professor
Central Institute of Fisheries Education
India
(External Examiner)



RUSLI HAJI ABDULLAH, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 30 July 2018

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Hassan B Hj Mohd Daud, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Siti Khairani Bejo, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Farina Mustaffa Kamal, PhD

Senior Lecturer
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No: Agharid Ali Hussein, GS38697

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of Chairman
of Supervisory Associate Professor
Committee: Dr. Hassan B Hj Mohd Daud

Signature: _____
Name of Member
of Supervisory Associate Professor
Committee: Dr. Siti Khairani Bejo

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Farina Mustaffa Kamal

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xvi
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xxii
 CHAPTER	
1 INTRODUCTION	1
 2 LITERATURE REVIEW	 4
2.1 Classification, distribution and economic importance of Climbing perch, <i>Anabas testudineus</i>	4
2.2 Fish immunology	5
2.2.1 Fish mucosal immunity	6
2.2.2 The mucosal layer	7
2.2.3 Cells produce mucus	7
2.2.4 Mucus	8
2.3 The antimicrobial property of epidermal mucus	9
2.4 Antimicrobial peptides and proteins (AMPPs)	9
2.5 Types of Fish AMPPs	10
2.5.1 Cationic antimicrobial peptides and proteins (AMPPs)	11
2.5.1.1 Cationic proteins :Apolipoprotein	11
2.5.1.2 Cationic antimicrobial peptides AMPs	12
2.5.1.2.a Piscidins	12
2.5.1.2.b β -Defencins	12
2.5.1.2.c Hepcidins	12
2.5.1.2.d Cathelicidins	12
2.5.1.3 Cationic peptides derived from larger proteins	13
2.5.1.3.a Histone-derived AMPs	13
2.5.1.3.b Hemoglobin-related AMPs	13
2.5.1.4 Anionic antimicrobial peptides and proteins AAMPs	14
2.5.2 Mechanism of antibacterial activity of Antimicrobial peptide	14
2.5.3 Toxicity of Antimicrobial peptides	15
2.5.4 Efficacy of Antimicrobial peptides in-vivo	16
2.5.4.1 Immunomodulatory activity of AMPs	17

	2.5.4.2	AMPs neutralization of Lipopolysaccharide <i>in vivo</i>	18
2.6		Immune-related hydrolytic enzymes	19
	2.6.1	Lysozyme	19
	2.6.2	Proteases	19
	2.6.3	Alkaline phosphatase	20
	2.6.4	Esterase	20
2.7		<i>Pseudomonas aeruginosa</i> and its Pathogenicity	20
	2.7.1	Virulence factors	21
	2.7.2	<i>Pseudomonas aeruginosa</i> bloodstream infections	22
	2.7.3	Antibiotic resistant style of <i>Pseudomonas aeruginosa</i>	23
		2.7.3.a Outer membrane permeability	23
		2.7.3.b Efflux systems	23
		2.7.3.c Production of antibiotic-inactivating enzymes	23
	2.7.4	Treatment of <i>Pseudomonas aeruginosa</i> sepsis by natural antibacterial agents	24
3		CRUDE EXTRACTION OF EPIDERMAL MUCUS OF CLIMBING PERCH <i>ANABAS TESTUDINEUS</i> AND ITS ANTIBACTERIAL AND HAEMOLYTIC ACTIVITIES	25
	3.1	Introduction	25
	3.2	Materials and methods	26
		3.2.1 Place and Duration of Experiment	26
		3.2.2 Epidermal mucus layer extraction	26
		3.2.3 Crude extraction	27
		3.2.3.1 Aqueous Extract	27
		3.2.3.2 Acidic Extract	27
		3.2.4 Detection of antimicrobial activity of both extract	27
		3.2.4.1 Bacteria and cells Preparation	27
		3.2.4.2 Preliminary screening for antimicrobial activity of both acidic and aqueous crude extracts	28
		3.2.5 Protein composition analysis of bioactive extract	28
		3.2.5.1 Protein estimation	28
		3.2.5.2 Protein profile	29
		3.2.6 Haemolytic activity of bioactive crude extract	29
		3.2.7 Statistical analysis.	30
	3.3	Results	30
		3.3.1 Antibacterial activity of both extracts extract against testing bacteria	30
		3.3.2 Protein composition analysis of bioactive extract	32
		3.3.3 Haemolytic activity	34
	3.4	Discussion	35
	3.5	Conclusion	38

4	MINIMUM INHIBITION AND MINIMUM BACTERICIDAL CONCENTRATIONS OF BIOACTIVE CRUDE AGAINST <i>P. aeruginosa</i> ATCC 10145	39
4.1	Introduction	39
4.2	Materials and methods	39
4.2.1	Preparation of bioactive crude	39
4.2.2	Criteria for selection <i>Pseudomonas aeruginosa</i> ATCC 10145	40
4.2.3	Resazurin microtiter assay for bacteriostatic and bactericidal	40
4.2.3.1	Preparation of Standardised Inoculum	40
4.2.3.2	Resazurin solution	40
4.2.3.3	Preparation of the plates	40
4.2.4	The antibiotics susceptibility test of <i>P. aeruginosa</i> ATCC 10145	40
4.2.5	Sensitivity testing of <i>P. aeruginosa</i> ATCC 10145 to ciprofloxacin and meropenem	41
4.2.6	Determination of Multiple Antibiotic Resistance Index (MAR Index)	41
4.2.7	NaCl sensitivity of bioactive crude	41
4.3	Results	42
4.3.1	Antibiotics susceptibility of <i>Pseudomonas aeruginosa</i> ATCC 10145	42
4.3.2	Minimum Inhibition(MIC) and Minimum Bactericidal Concentrations (MBC)of bioactive crude	43
4.3.3	NaCl sensitivity of bioactive crude	45
4.4	Discussion	46
4.4.1	Bacteriostatic and Bactericidal activity of bioactive crude and antibiotics profile of <i>Pseudomonas aeruginosa</i>	46
4.4.2	NaCl sensitivity of bioactive crude	47
4.5	Conclusion	47
5	ASSESSMENT OF SOME IMMUNE COMPONENTS FROM THE BIOACTIVE CRUDE DERIVED FROM EPIDERMAL MUCUS LAYER OF CLIMBING PERCH, <i>Aanabs. testudineus</i>	48
5.1	Introduction	48
5.2	Materials and methods	49
5.2.1	Estimation of immune related enzymes activities in bioactive crude	49
5.2.1.1	Protease	49
5.2.1.2	Lysozyme(LYZ)	49
5.2.1.3	Alkaline phosphatase(ALP)	50
5.2.1.4	Esterase	50
5.2.2	Isolation of Antimicrobial proteins and peptides by Size exclusion chromatography (SEC)	50
5.2.3	Proteins and Peptides estimation	51

5.2.4	Proteins and peptides profile	51
5.2.5	Identification of Antimicrobial proteins and peptides by Q-TOF LC/MS	51
5.2.5.1	Digestion by trypsin	51
5.2.5.2	Identification by Q-TOF LC/MS	52
5.3	Results	52
5.3.1	Immune-related enzymatic activities of bioactive crude	52
5.3.2	Isolation of antibacterial proteins and peptides by size-exclusion chromatography	54
5.3.3	Q-TOF LC/MS Data analysis	57
5.4	Discussion	59
5.4.1	Activities of immune related enzymes in bioactive crude	59
5.4.2	Identification of antimicrobial proteins and peptides	61
5.4.3	The Q-TOF LC/MS data analyses	61
5.5	Conclusion	64
6	<i>In vivo</i> ASSESSMENT OF AMPPC TOXICITY AND MORTALITY RATE IN ICR MICE INFECTED WITH <i>P. aeruginosa</i> ATCC 10145	65
6.1	Introduction	65
6.2	Materials and methods	66
6.2.1	Laboratory animals and management	66
6.2.2	Ethical consideration	66
6.2.3	Passage of <i>Pseudomonas aeruginosa</i> ATCC 10145 in ICR mice	66
6.2.4	Determination the Mortality Rate of <i>Pseudomonas aeruginosa</i> ATCC 10145	67
6.2.5	Acute toxicity of AMPPC	67
6.2.6	Blood collection	67
6.2.7	Serum biochemistry marker	68
6.2.8	Proinflammatory cytokines	68
6.2.8.1	Detection of IL-6 by ELISA test	68
6.2.8.2	Detection of TNF- α by ELISA test	68
6.2.8.3	Calculation of results	69
6.2.9	Histopathology	69
6.2.10	Statistical Analysis	69
6.3	Results	70
6.3.1	Mortality rate of ICR mice infected by <i>P.aeruginosa</i> ATCC 10145	70
6.3.2	Acute toxicity of AMPPC	71
6.4	Discussion	75
6.4.1	Passage of <i>Pseudomonas aeruginosa</i> ATCC 10145 and LD50	75
6.4.2	Acute toxicity of AMPPC	75
6.5	Conclusion	76

7	THE EFFICACY OF ANTIMICROBIAL PROTEINS AND THEIR POTENTIAL BIOACTIVE PEPTIDES AGAINST <i>Pseudomonas aeruginosa</i> ATCC 10145 INFECTION IN MICE MODEL	77
7.1	Introduction	77
7.2	Materials and methods	78
7.2.1	Laboratory animals and management	78
7.2.2	Ethical consideration	78
7.2.3	Blood collection	78
7.2.4	Determination of bacterial quantities	78
7.2.5	Serum biochemistry marker	79
7.2.6	Proinflammatory cytokines.	79
7.2.7	Histopathology	79
7.2.8	Experimental design of AMPPC efficacy against <i>Pseudomonas aeruginosa</i> sepsis	79
7.2.9	Evaluation of treatment	81
7.2.10	Statistical Analysis	81
7.3	Results	81
7.3.1	Evaluation of treatment with AMPPC	81
7.3.1.1	Mortality rate and bacteria burden in organs and blood	81
7.3.1.2	Liver and kidney function biomarkers	84
7.3.1.3	Proinflammatory cytokines TNF- α and IL-6	86
7.3.1.4	Histopathology examination of internal organs	88
7.4	Discussion	93
7.4.1	Evaluation of treatment by AMPPC and Ciprofloxacin	93
7.4.1.1	Mortality rate	93
7.4.1.2	<i>P. aeruginosa</i> ATCC 1045 burden in organs and blood	94
7.4.1.3	Liver and kidney function biomarkers	95
7.4.1.4	Proinflammatory cytokines TNF- α and IL-6 levels	96
7.4.1.5	Histopathology changes	97
7.5	Conclusion	98
8	GENERAL DISCUSSION, LIMITATION AND RECOMMENDATIONS	100
	REFERENCES	104
	APPENDICES	143
	BIODATA OF STUDENT	153
	LIST OF PUBLICATIONS	154

LIST OF TABLES

Table		Page
2.1	Antimicrobial activity of some of freshwater fish AMPs against gram negative bacteria	10
2.2	Antimicrobial activity of some of freshwater fish AMPs against Some species of gram positive bacteria	11
4.1	Antibiotic resistance patterns of <i>P. aeruginosa</i> ATCC 10145 against different group of antibiotics	42
4.2	Zone diameter of breakpoint interpretation criterion based on CLSI	43
5.1	Innate immune parameters of enzyme hydrolysate evaluated i bioactive crude, data represented the mean values of three replicates of enzyme activity by U/mg \pm Standard S.D	52
5.2	Q-TOF LC/MS analysis details, accession number, and physical parameters of identified proteins in the bioactive fraction of Superdex™ peptide 10/300 GL.	57
5.3	Sequences and physical parameters of <i>de novo</i> peptides in the biological active fraction of Superdex™ peptide 10/300 GL chromatography	59
6.1	Cumulative number of dead and surviving mice after injected with different doses of <i>P. aeruginosa</i> ATCC 11045	70
7.1	Pathological inflammation scoring (PIS) system	79
7.2	Bacterial counts in the internal organs (lung, liver, spleen) and blood from mice for groups of non-treated and treated with AMPPC and CE via oral administration. Each value represents the mean value from three determinations \pm standard derivation (SD)	82

LIST OF FIGURES

Figure		Page
2.1	Climbing perch, <i>Anabas testudineus</i>	4
2.2	Schematic representation of fish skin showing the general structure and cellular components	7
2.3	The epidermis and mucous goblet cells of rainbow trout skin (indicated by arrowheads). The green portion in the dermis indicates the collagen-rich connective tissue	8
2.4	The proposed models of membrane-lytic mechanisms by AMPs	15
2.5	Various manners of impact of antimicrobial peptides	17
3.1	Climbing perch was acclimatized for seven days in glass aquarium at the Aquatic Animal Health Unit, Faculty of Veterinary Medicine, UPM	26
3.2	Mueller- Hinton agar plates showing well diffusion (zone of inhibition measured in mm) of ACA (E) (1.2 mg/ml), ciprofloxacin(CE) (5µg/ml) as positive control against clinical and reference isolates, plates incubated for 24 h at 37°C	31
3.3	Mean of Inhibition Zone Diameter (IZD) measured in mm of antibacterial activity of ACA against isolate strains and reference strains. <i>Pseudomonas aeruginosa</i> ATCC 10145 had the highest while, Methicillin Resistant <i>S. aureus</i> ATCC 43300 (MRSA) was the lowest value. The bacteria with different letter were significantly different ($P < 0.05$)	32
3.4	Calibration curve of BSA (0–200 µg/mL) versus absorbance at 340 nm estimated using O-Phthaldialdehyde (OPA) assay with linear regression $R^2 = 0.9965$; unknown concentration of proteins in BCE sample was calculated from graph through absorbance	33
3.5	SDS-PAGE (NuPAGE 4–12% Bis-Tris Novex Invitrogen) of ACA sample Line (a) represents calibrated marker proteins (BioRad) and line (b) represents the protein profile of ACA; arrows indicate the bands of ACA proteins	33

3.6	Calibration curve of log Mwt of marker proteins (11 - 245) kDa versus relative mobility (Rf) during electrophoresis showed a linear regression ($R^2=0.9836$), which allows the molecular weight of the unknown proteins in ACA to be read from the graph	34
3.7	96-wells plate showing haemolytic activities of acidic crude extract sample toward horse RBCs. There was no lysis showed by two-fold serial dilutions of ACA (1.2mg/ml) and negative control C (-) in the PBS saline. Positive control C (+) in the 0.2% TritonX-100 showed complete lysis of RBCs	35
4.1	Mueller-Hinton agar plate showing well diffusion assay (zone of inhibition measured in mm) of ciprofloxacin (CE) (5 μ g/well) and meropenem (ME) (10 μ g/well) for <i>P. aeruginosa</i> ATCC 10145	43
4.2	96-well plate after 24 h of incubation in modified resazurin assay used in the microdilution experiment. MIC of bioactive crude was 10 mg/mL against <i>P. aeruginosa</i> ATCC 10145. After adding the resazurin to the wells, the pink colour indicates growth and blue or colourless media indicates inhibition of growth. In the rows A, B, and C, the wells from 1 to 10 were serially diluted crude samples in broth, indicator, and bacteria, while wells 11 were negative controls containing bacteria, broth, and indicator. Wells 12 were positive controls containing ciprofloxacin (5 μ g/mL), broth, indicator, and bacteria	44
4.3	Mueller-Hinton agar plate showing inhibition growth of <i>P. aeruginosa</i> ATCC 10145 inoculated from the first eight wells of modified resazurin assay plate, MBC (minimum bactericidal concentration) value of bioactive crude was 2.5mg/ml, plates incubated for 24 h at 37°C	44
4.4	Killig rate of <i>P. aeruginosa</i> ATCC treated with bioactive crude (10mg/ml) in variable concentration of NaCl (170,150,130,110, 90) mM, through three preiod of times (10,20,30) min	45
5.1	Calibration curve of lysozyme from chicken egg white (0.1–0.006) μ g/mL (linear regression $R^2 = 0.9876$) against lysozyme activity estimated by turbidity assay using <i>M. lysodeikticus</i> . Concentration of lysozyme (μ g/mL) of bioactive crude sample was calculated from curve	53
5.2	Calibration curve with linear regression (R^2) in different concentration of p-nitrophenol concentration (μ mole p-nitrophenol with <i>O.D</i> 410	53

5.3	Size exclusion chromatography of antibacterial proteins and peptides using Superdex™ peptide 10/300 GL on AKTA Purifier 10 FPLC. Four chromatogram peaks are shown: Peak 1 (A12 – B12), Peak 2(C3–C6), Peak 3 (C8–C10), and Peak 4 (D2–D6) detected by absorbance at 280 nm(mAU)	55
5.4	Mueller-Hinton agar plate showing well diffusion assay (zone of inhibition measured in mm) against <i>P. aeruginosa</i> ATCC 10145 for (a) fraction for SepPak C18 and ultrafiltration 30 kDa and fractions at Peak 1 (b) B2–B4, (c) B6–B8, and (d) B10–B12 for Superdex™ peptide 10/300 GL column	56
5.5	SDS-PAGE (15%) analysis of elution fraction form SepPak C18 and ultrafiltration 30 kDa Line 1 is calibrated high protein marker (BioRad) and line 2 represents the protein profile of fraction.; arrows indicate the bands of fraction proteins (10-25)kDa	56
5.6	Aligned amino acid sequences (blue) coverage of apolipoprotein A-1 of bioactive fraction, matched with identified apolipoprotein A-1 from <i>Channa striata</i> (accession number A0A077H3P6)	58
5.7	Aligned peptide sequence (blue) coverage of haemoglobin subunit beta (fragment) of bioactive fractions matched with peptide sequence of identified haemoglobin subunit beta (fragment) from <i>Channa argus</i> and <i>Channa maculata</i> (accession number A0A0U2K0I0)	58
5.8	MS/MS spectrum of <i>de novo</i> peptide AAGPKGPLGPR showing the fragmentation pattern of peptide with a parent mass of 1020.25 and doubly charged	59
6.1	Dose lethality curve of <i>P. aeruginosa</i> ATCC 10145 in ICR mice (<i>linear</i> regression, $R^2= 0.9263$) using the methods of Reed and Munch (1938)	71
6.2	Microphotographs of ICR mice kidney for control group G1 (a) showing normal glomeruli and tubules, and G (AMPPC) (b) showing Bowman’s capsule (black arrow) and urinary space (blue arrow) (H& E ×20 magnification)	73
6.3	Microphotographs of ICR mice liver for (a) normal group G1; hepatocytes exhibit normal central veins, liver cell cords, and hepatic sinusoids, (b) group (AMPPC) showing normal histological architectures (H& E ×20 magnification)	74

7.1	Flowchart of the experimental animal model of AMPCC efficacy in ICR mice model. Two groups (G3 and G5) were treated with AMPPC, one group (G4) was treated with CE and group G2 was not treated. *Oral administration of AMPPC and CE by oral gavage **Mice injected with 2×10^7 CFU/mL of (MDR) <i>P. aeruginosa</i> ATCC 10145	80
7.2	Yellow colonies of (MDR) <i>P. aeruginosa</i> ATCC 10145 (non-lactose fermenter) on MAC agar after incubation of 24 h at 37 °C. Bacteria was counted and cultured from harvested organs and blood sample of nontreated group with AMPPC. (a) lung, (b) spleen, (c) liver, and (d) blood	83
7.3	Yellow colonies of (MDR) <i>P.aeruginosa</i> ATCC10145 colonies (Non-lactose fermenter) on MAC agar after incubation of 24 hr at 37°C. Bacteria was counted and culturing from harvested organs and blood sample of treated group by AMPPC	83
7.4	The mean serum AST and ALT concentration of ICR mice after 24 h of last treatment. A significant difference ($p < 0.05$) between control and other groups were indicated by different letters (error bar = S.D)	85
7.5	The mean of serum creatinine and urea concentration of ICR mice after 24 h of last treatment. A significant difference ($p < 0.05$) between control and other groups were indicated by different letters (error bar = S.D)	86
7.6	Serum levels of TNF- α at 12 and 24 h during the treatment period. Different letters indicate significant differences ($p < 0.05$) between groups (error bar = S.D)	87
7.7	Serum levels of IL-6 at 12 and 24 h during the treatment period. Different letters indicate significant differences ($p < 0.05$) between groups (error bar = S.D)	88
7.8	Microphotographs of ICR mice lungs: G1 (a) shows normal histological properties, G2 (b) shows thickening of alveolar wall and pulmonary haemorrhage. G3 (c), G4 (d), and G5 (e) display gradual decrease in the degree of congestion and haemorrhage (H&E X20 magnification)	89
7.9	Microphotographs of ICR mice spleens: G1 (a) shows normal histological architecture; G2 (b) shows loss of the typical structures of the germinal centers with congested blood vessels. G3 (c), G5 (d), and G4 (e) display the red and white pulps and germinal centres with moderate histological changes (H&E X20 magnification)	90

- 7.10 Microphotographs of ICR mice livers G1 (a) shows normal histological properties; G2 (b) shows a severe massive infiltration of inflammatory cells, severe haemorrhage, and hepatocellular necrosis. G3 (c), G5 (d), and G4 (e) show a slightly reduced inflammation and congestion (H&E X20 magnification) 92



LIST OF ABBREVIATIONS

ALT	Alanine Aminotransferase
ALP	Alkaline phosphatase
AMPs	Antimicrobial peptides and proteins
AMPP	Antimicrobial peptides and proteins
AMPPC	Antimicrobial proteins and bioactive potential peptide in the crude
APOL1	Apolipoprotein L1
AST	Aspartate Aminotransferase
ACE	acidic crude extract
BC	bioactive crude
BCE	bioactive crude extract
ACN	Acetonitrile
BHI	Heart Infusion Broth
BSA	Bovine Serum Albumin
BSI	bloodstream infections
CE	Ciprofloxacin
°C	Degree Celsius
CF	Cystic fibrosis
CFU	Colony Form Unit
CLSI	Clinical and Laboratory Standards Institute
DMSO	Dimethyl sulfoxide
D.W	Distilled Water
EML	Epidermal mucus and Epidermis Layer
HAc	Acetic acid
H&E	Harris'haematoxylin
HDAPs	Histone-derived antimicrobial peptides
h	hour
IACUC	Institutional Animal Care and Use Committee recommendations

IL-6	Interleukin-6
IL-8	Interleukin-8
IL-1b	Interleukin-1b
I	Intermediate sensitive
IP	Intraperitoneal
IZD	Inhibition Zone Diameter
IgA	Immunoglobulin A
IgG	Immunoglobulin G
kDa	Kilo Dalton
LC/MS	Liquid Chromatography–Mass Spectrometry
LPS	lipopolysaccharides
LYZ	Lysozyme
MAR	Multiple antibiotic resistance index
MBC	Minimal Bactericidal Concentration
MDR	Multidrug-resistant
Me	Meropenem
MHA	Muller–Hinton agar
MIC	Minimal Inhibitory Concentration
ml	Milliliter
μl	Microliter
mg	milligram
μg	Microgram
mm	Millimeter
mM	Millmole
μmole	Micromole
MRSA	Methicillin Resistant <i>S. aureus</i>
Mwt	Molecular weight
PBS	Phosphate Buffered Saline
PAMP	pathogen-associated molecular pattern
PA	<i>Pseudomonas aeruginosa</i>

pg.	Pictogram
PG-1	Protegrin-1
PIS	Pathological Inflammation scoring
p-NP	p-nitrophenol
p-NNP	p-nitrophenylphosphate
O.D	Optical Density
OPA	O-Phthaldialdehyde
RBCs	Red Blood Cells
Rf	Relative mobility
R	Resistant
R2	linear regression
S	Sensitive
S.D	Standard deviation
SEC	Size exclusion chromatography
SDS–PAGE	Sodium dodecyl sulfate-polyacrylamide gel electrophoresis
TFA	Trifluoroacetic acid
TNF- α	tumor necrosis factor- α
T2SS	Type II Secretion System
T3SS	Type III Secretion System

CHAPTER 1

INTRODUCTION

Bioprospecting is an organized search to find beneficial products obtained from bioresources including plants, microorganisms and animals, that use in creating and investigating new biological compounds with medicinal and commercial values for health purposes (Oyemitan,2017).The evolution and adaptation by animals that allow them to survive under extreme environmental conditions are the result of developed functions that may have resulted from acquiring traits and features containing essential bioactive components. Whenever these interesting bioactivities are found and the bioactive compounds can be isolated and identified, valuable new biological products can be developed (Chalamaiah *et al.*, 2012; Harnedy and FitzGerald 2012). Common antibiotics and other chemical-based medicines are the most widely used ways to control bacterial infection in humans and animals (Salger *et al.*, 2016). The widespread application of antibiotics has resulted in the emergence of resistant pathogens that are capable of withstanding the effect of commonly used antibiotics. Therefore, the world needs novel active biomolecules to combat the emerging and re-emerging pathogen that affect human health and aquatic lives (Manikantan *et al.*, 2016). Active biomolecules are the components of the innate immune system (Salger *et al.*, 2016). Antimicrobial peptides (AMPs) and other protein materials are produced by a large number of organisms and are essential components of the innate immune system (Costa *et al.*, 2017; Salger *et al.*, 2016 Zhang and Gallo, 2016). They are readily produced when the organisms are subjected to adverse environmental conditions (Costa *et al.*, 2017; Zhang and Gallo, 2016). These small molecules have been shown to exhibit a broad spectrum of antimicrobial activities against many organisms including bacteria, yeasts, fungi, and viruses. They also possess significant anti-inflammatory and immunomodulatory activities as well as cytotoxic activity on cancer cells (Kang *et al.*, 2017). Since the first AMP was described in humans, over 2700 kinds have been discovered in all life forms ranging from bacteria to human (Valero *et al.*, 2013). The first AMP was discovered in 1922 and this has attracted researchers' attention since the 1980s (Wang, 2017).

The aquatic habitat is home to an array of organisms that have developed features and adaptations that permit them to live remarkably in these enclaves. Notable among the inhabitants of the aquatic ecosystem are fish. They have evolved ways to survive including possessing many biological components as well as bioactive defence system that serves to protect them from the load of bacteria present in water (Harnedy and FitzGerald, 2012). The protection is conferred by both specific and nonspecific immune components. One of the major components of the nonspecific defence mechanism in fish is the antimicrobial peptides and proteins. They are the major components of the fish's innate immune response to a wide range of opportunistic pathogens (Nsrelden *et al.*, 2017). Moreover, the immunological activity of AMPs, they also possess inflammatory and modulatory roles as part of their innate and adaptive immune responses, as well as act as chemokines to recruit other

effector cells (Cuesta *et al.*, 20011; Chertov *et al.*, 1996; Oppenheim *et al.*, 2003; Kang *et al.*, 2017).

The teleostei taxon is made up of over 24,000 different species; hence, only small fractions of this taxon of fish species have been studied. The majority of these studies utilised them as experimental models in order to explore their commercial potentials to the aquaculture industry and about 62 different AMPs have been reported in many fish species (Smith and Fernandes, 2009; Valero *et al.*, 2013). Peptides of fish origin are reportedly active in very harsh environmental conditions, including very high salt concentrations, making them suitable candidates for development of therapeutic antimicrobial agents (Masso-Silva and Diamond, 2014).

Anabas testudineus, commonly known as climbing perch, is an obligate air breather (Sayer, 2005). The species represents an economically useful commodity and is a very good source of food in Southeast Asia especially in Thailand, Malaysia, and the Philippines (Chotipuntu *et al.*, 2011; Loh *et al.*, 2015). It is a euryhaline teleost, inhabiting both freshwater and brackish water ecosystems of Southeast Asian countries (Kohinoor *et al.*, 1991; Sarkar *et al.*, 2005) and is tolerant to extremely unfavourable water conditions and high salinity (Ip *et al.*, 2013; Khan *et al.*, 1976; Sarkar and Ponniah, 2000). The body surface of *Anabas testudineus* is covered by a thick coat of mucus which is composed of mucopolysaccharides, lipids, and proteins. These substances are important in keeping the skin moist thereby facilitating the survival of the fish in harsh situations (Agarwal *et al.*, 1980). The climbing perch has extraordinary tissue regeneration potential and this characteristic may have medical applications for skin regeneration (Srivastava *et al.*, 2013). Therefore, the skin mucus of *Anabas testudineus* is suitable for research material of bioactive compounds such as antibacterial proteins and peptides.

Natural antimicrobial proteins and peptides have the ability to kill microbes and antibiotic-resistant bacteria. Roughly all fish antimicrobial peptides and proteins (AMPPs) have antibacterial activities against several types of bacteria. Some of the fish AMPPs have the ability to act in high salt concentrations, make them perfect potential targets for development as therapeutic antimicrobials. Epidermal mucus from selected freshwater fish species is a potential source of novel antimicrobial agents against antibiotic-resistance microorganisms.

This research hypothesised that epidermal mucus of the climbing perch generates a large variety of antimicrobial proteins and peptides. It will be a new natural drug alternative to antibiotics and have immunomodulatory functions for treatment of resistant pathogenic bacteria.

The specific objectives of the present study are:

1. To isolate crude antimicrobial proteins and peptides from the epidermal mucus of the climbing perch
2. To investigate the antimicrobial activity, haemolytic activities, and NaCl tolerance of the crude antimicrobial proteins and peptides.
3. To isolate and characterise the crude antimicrobial proteins and peptides.
4. To evaluate the efficacy of the crude antimicrobial proteins and peptides against *Pseudomonas aeruginosa* infection in mice models *in vivo*.



REFERENCES

- Abdelraouf, T.K., and Tam, V. H. (2017). *Pseudomonas*. In: Mayers, D. L., Sobel, J. D., Ouellette, M., Kaye, K. S., and Marchaim, D. (Eds.), *Antimicrobial Drug Resistance: Clinical and Epidemiological Aspects* (2nd ed, Vol.2, pp 899-923). Springer Nature.
- Abraham, E., (2000). Tissue factor inhibition and clinical trial results of tissue factor pathway inhibitor in sepsis. *Critical Care Medicine*, 28(9), S31-S33.
- Agarwal, S. K., Banerjee, T. K., and Mittal, A. K. (1980). A histochemical study of the epidermis of the climbing perch, *Anabas testudineus* (Anabantidae, Pisces). *Zeitschrift Fur Mikroskopisch-Anatomische Forschung*, 94(1), 143-159.
- Ageitos, J. M., Sánchez-Pérez, A., Calo-Mata, P., and Villa, T. G. (2017). Antimicrobial peptides (AMPs): Ancient compounds that represent novel weapons in the fight against bacteria. *Biochemical Pharmacology*, 133, 117-138
- Aires, J. R., Köhler, T., Nikaido, H., and Plésiat, P. (1999). Involvement of an active efflux system in the natural resistance of *Pseudomonas aeruginosa* to aminoglycosides. *Antimicrobial Agents and Chemotherapy*, 43(11), 2624-262
- Akhila, J.S., Shyamjith, D. and Alwar, M.C. (2007). Acute toxicity studies and determination of median lethal dose. *Current Science*, 917-920.
- Alasil, S. M., Omar, R., Ismail, S., and Yusof, M. Y. (2014). Antibiofilm activity, compound characterization, and acute toxicity of extract from a novel bacterial species of *Paenibacillus*. *International journal of microbiology*, 2014.
- Alexander, J. B., and Ingram, G. A. (1992). Noncellular nonspecific defence mechanisms of fish. *Annual Review of Fish Diseases*, 2, 249-279.
- Ali, Z., Mumtaz, N., Naz, S. A., Jabeen, N., and Shafique, M. (2015). Multi-drug resistant *Pseudomonas aeruginosa*: a threat of nosocomial infections in tertiary care hospitals. *JPMA*, 65(12).
- Alikunhi, K. H. (1957). Fish culture in India. *Farm Bulletin*, (20), 1-144.
- Almaaytah, A., Mohammed, G. K., Abualhaijaa, A., and Al-Balas, Q. (2017). Development of novel ultrashort antimicrobial peptide nanoparticles with potent antimicrobial and antibiofilm activities against multidrug-resistant bacteria. *Drug Design, Development and Therapy*, 11, 3159.
- Aly, R., and Malbach, H. I. (1988). Comparative antibacterial efficacy of a 2-minute surgical scrub with chlorhexidine gluconate, povidone-iodine, and chloroxylenol sponge-brushes. *American journal of infection control*, 16(4), 173-177.

- Andrews, J.M. (2001). Determination of minimum inhibitory concentrations. *Journal of Antimicrobial Chemotherapy*, 48(suppl 1), 5-16.
- Ángeles, E. M. (2012). An overview of the immunological defenses in fish skin. *ISRN Immunology*, 2012.
- Aranishi, F. and Nakane, M. (1997). Epidermal proteases of the Japanese eel. *Fish Physiology and Biochemistry*, 16(6), 471-478.
- Aranishi, F. (2000). High sensitivity of skin cathepsins L and B of European eel (*Anguilla anguilla*) to thermal stress. *Aquaculture*, 182(3), 209-213.
- Aranishi, F. (1999). Lysis of pathogenic bacteria by epidermal cathepsins L and B in the Japanese eel. *Fish Physiology and Biochemistry*, 20, 37-41.
- Ashare, A., Monick, M. M., Powers, L. S., Yarovinsky, T., and Hunninghake, G. W. (2006). Severe bacteremia results in a loss of hepatic bacterial clearance. *American journal of respiratory and critical care medicine*, 173(6), 644-652.
- Axelrod, freshwater aquarium fishes. Neptune City, NJ: TFH Publications.
- Bansil, R., Stanley, E., and LaMont, J. T. (1995). Mucin biophysics. *Annual Review of Physiology*, 57(1), 635-657.
- Barman, D., Nen, P., Mandal, S. C., and Kumar, V. (2013). Aquaculture health management: a new approach. *Journal of Marine Science. Research and Development*, 3(4), 1.
- Battersby, A. J., Khara, J., Wright, V. J., Levy, O., and Kampmann, B. (2016). Antimicrobial proteins and peptides in early life: ontogeny and translational opportunities. *Frontiers in Immunology*, 7.
- Bauer, A. W., Kirby, W. M., Sherris, J. C., and Turck, M. (1966). Antibiotic susceptibility testing by a standardized single disk method. *American Journal Of Clinical Pathology*, 45(4), 493.
- Bergsson, G., Agerberth, B., Jörnvall, H., and Gudmundsson, G. H. (2005). Isolation and identification of antimicrobial components from the epidermal mucus of Atlantic cod (*Gadus morhua*). *The FEBS journal*, 272(19), 4960-4969
- Bezerra, R. F., Soares, M. D. C. F., Santos, A. J. G., Maciel Carvalho, E. V. M., and Coelho, L. C. B. B. (2014). Seasonality influence on biochemical and hematological indicators of stress and growth of Pirarucu (*Arapaima gigas*), an Amazonian air-breathing fish. *The Scientific World Journal*, 2014.

- Bhargava, R., Altmann, C. J., Andres-Hernando, A., Webb, R. G., Okamura, K., Yang, Y., and Faubel, S. (2013). Acute lung injury and acute kidney injury are established by four hours in experimental sepsis and are improved with pre, but not post, sepsis administration of TNF- α antibodies. *PLoS One*, 8(11), e79037.
- Birkemo, G. A., Lüders, T., Andersen, Ø., Nes, I. F., and Nissen-Meyer, J. (2003). Hippusin, a histone-derived antimicrobial peptide in Atlantic halibut (*Hippoglossus hippoglossus* L.). *Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics*, 1646(1), 207-215
- Biller-Takahashi, J. D., and Urbinati, E. C. (2014). Fish Immunology. The modification and manipulation of the innate immune system: Brazilian studies. *Anais da Academia Brasileira de Ciências*, 86(3), 1484-1506.
- Bornscheuer, U. T., and Kazlauskas, R. J. (2006). *Hydrolases in organic synthesis: regio- and stereoselective biotransformations*. John Wiley and Sons.
- Bradshaw, J. P. (2003). Cationic antimicrobial peptides. *BioDrugs*, 17(4), 233-240.
- Bragadeeswaran, S., Priyadarshini, S., Prabhu, K. and Rani, S.R.S. (2011). Antimicrobial hemolytic activity of fish epidermal mucus *Cynoglossus arel* and *Arius caelatus*. *Asian Pacific Journal of Tropical Medicine*, 4(4), 305-309.
- Breidenstein, E. B., de la Fuente-Núñez, C., and Hancock, R. E. (2011). *Pseudomonas aeruginosa*: all roads lead to resistance. *Trends in Microbiology*, 19(8), 419-426.
- Brinchmann, M. F. (2016). Immune relevant molecules identified in the skin mucus of fish using-omics technologies. *Molecular BioSystems*, 12(7), 2056-2063.
- Browne, M. J., Feng, C. Y., Booth, V., and Rise, M. L. (2011). Characterization and expression studies of Gaduscidin-1 and Gaduscidin-2; paralogous antimicrobial peptide-like transcripts from Atlantic cod (*Gadus morhua*). *Developmental and Comparative Immunology*, 35(3), 399-408.
- Bruce, T. J., and Brown, M. L. (2017). A Review of Immune System Components, Cytokines, and Immunostimulants in Cultured Finfish Species. *Journal of Animal Sciences*, 7, 267-288.
- Brunetti, J., Falciani, C., Roscia, G., Pollini, S., Bindi, S., Scali, S., Arrieta, U.C., Gómez-Vallejo, V., Quercini, L., Ibba, E. and Prato, M. (2016). In vitro and in vivo efficacy, toxicity, bio-distribution and resistance selection of a novel antibacterial drug candidate. *Scientific Reports*, 6, 26077.
- Bryant, C.E., Spring, D.R., Gangloff, M. and Gay, N.J., (2010). The molecular basis of the host response to lipopolysaccharide. *Nature Reviews Microbiology*, 8(1), 8-14.

- Bulet, P., Stöcklin, R., and Menin, L. (2004). Anti- microbial peptides: from invertebrates to vertebrates. *Immunological Reviews*, 198(1), 169-184.
- Bullen, J. J., Rogers, H. J., Spalding, P. B., and Ward, C. G. (2005). Iron and infection: the heart of the matter. *Pathogens and Disease*, 43(3), 325-330.
- Bun Ng, T., Chi Fai Cheung, R., Cheuk Wing Ng, C., Fei Fang, E., and Ho Wong, J. (2015). A review of fish lectins. *Current Protein and Peptide Science*, 16(4), 337-351.
- Buonocore, F.; Randelli, E.; Casani, D.; Picchiatti, S.; Belardinelli, M.C.; de Pascale, D.; de Santi, C.; Scapigliati, G. (2012). A piscidin-like antimicrobial peptide from the icefish *Chionodraco hamatus* (Perciformes: Channichthyidae): Molecular characterization, localization and bactericidal activity. *Fish Shellfish Immunol.* 33, 1183–1191
- Burgess, D. S. (2005). Use of pharmacokinetics and pharmacodynamics to optimize antimicrobial treatment of *Pseudomonas aeruginosa* infections. *Clinical Infectious Diseases*, 40(Supplement_2), S99-S104.
- Cahill, M. M. (1990). Bacterial flora of fishes: a review. *Microbial ecology*, 19(1), 21-41
- Cantisani, M., Finamore, E., Mignogna, E., Falanga, A., Nicoletti, G.F., Pedone, C., Morelli, G., Leone, M., Galdiero, M. and Galdiero, S. (2014). Structural insights into and activity analysis of the antimicrobial peptide myxinidin. *Antimicrobial Agents and Chemotherapy*, 58(9), 5280-5290.
- Cantisani, M., Leone, M., Mignogna, E., Kampanaraki, K., Falanga, A., Morelli, G., Galdiero, M. and Galdiero, S. (2013). Structure-activity relations of myxinidin, an antibacterial peptide derived from the epidermal mucus of hagfish. *Antimicrobial Agents and Chemotherapy*, 57(11), 5665-5673.
- Cao, H., Ke, T., Liu, R., Yu, J., Dong, C., Cheng, M., Huang, J. and Liu, S., 2015. Identification of a novel proline-rich antimicrobial peptide from *Brassica napus*. *Plo S one*, 10(9), e0137414.
- Cetin, N., Suleyman, B., Kuyrukluylidiz, U., Nalkiran, H.S., Kiran, A., Gencoglu, S., Duzgun, A., Kurtoglu, I.Z., Yarali, O., Gul, M.A. and Suleyman, H. (2016). Investigation of mucus obtained from different fish species on the acute pain induced with scalpel incision in paw of rats. *Experimental animals*, 65(1), 77-85
- Chaithany, E. R., Philip, R., Sathyan, N., and Anil Kumar, P. R. (2013). Molecular characterization and phylogenetic analysis of a histone-derived antimicrobial peptide teleostin from the marine teleost fishes, *Tachysurus jella* and *Cynoglossus semifasciatus*. *ISRN Molecular Biology*, 2013,1-7

- Chakraborty, B. K. (2010). Status and position of aquatic biodiversity of four beels and its floodplain level of northern Bangladesh with a good practice of beel nurseries and community based co-management policy. *Advances in Environmental Research*, 8, 121-164.
- Chakraborty, B. K. (2016). Sustainable Aquaculture Practice of Climbing Perch Koi, *Anabas testudineus* (Bloch, 1792) Under Semi Intensive Aquaculture System in Bangladesh. *Proceedings of the Zoological Society*, 69,1, (133-140).
- Chalamaiah, M., Hemalatha, R., and Jyothirmayi, T. (2012). Fish protein hydrolysates: proximate composition, amino acid composition, antioxidant activities and applications: a review. *Food Chemistry*, 135(4), 3020-3038.
- Chalkley, L. J., and Koornhof, H. J. (1985). Antimicrobial activity of ciprofloxacin against *Pseudomonas aeruginosa*, *Escherichia coli*, and *Staphylococcus aureus* determined by the killing curve method: antibiotic comparisons and synergistic interactions. *Antimicrobial Agents and Chemotherapy*, 28(2), 331-342.
- Chandra, G., Bhattacharjee, I., Chatterjee, S. N., and Ghosh, A. (2008). Mosquito control by larvivorous fish. *Indian Journal of Medical Research*, 127(1), 13.
- Chang, S. K., Ismail, A., Yanagita, T., Esa, N. M., and Baharuldin, M. T. H. (2015). Antioxidant peptides purified and identified from the oil palm (*Elaeis guineensis* Jacq.) kernel protein hydrolysate. *Journal of functional foods*, 14, 63-75.
- Chang, E. W. Y., Loong, A. M., Wong, W. P., Chew, S. F., Wilson, J. M., and Ip, Y. K. (2007). Changes in tissue free amino acid contents, branchial Na⁺/K⁺-ATPase activity and bimodal breathing pattern in the freshwater climbing perch, *Anabas testudineus* (Bloch), during seawater acclimation. *Journal of Experimental Zoology Part A: Ecological Genetics and Physiology*, 307(12), 708-723.
- Chang, X., Kobayashi, T., Senthilkumaran, B., Kobayashi-Kajura, H., Sudhakumari, C. C., and Nagahama, Y. (2005). Two types of aromatase with different encoding genes, tissue distribution and developmental expression in Nile tilapia (*Oreochromis niloticus*). *General and Comparative Endocrinology*, 141(2), 101-115.
- Chatterjee, M., Anju, C. P., Biswas, L., Kumar, V. A., Mohan, C. G., and Biswas, R. (2016). Antibiotic resistance in *Pseudomonas aeruginosa* and alternative therapeutic options. *International Journal of Medical Microbiology*, 306(1), 48-58.
- Chatzinikolaou, I., Abi-Said, D., Bodey, G. P., Rolston, K. V., Tarrand, J. J., and Samonis, G. (2000). Recent experience with *Pseudomonas aeruginosa* bacteremia in patients with cancer: retrospective analysis of 245 episodes. *Archives of Internal Medicine*, 160(4), 501-509.

- Chen J.Y, Lin W.J and Lin T.L (2009). A fish antimicrobial peptide, tilapia hepcidin TH2–3, shows potent antitumor activity against human fibrosarcoma cells. *Peptides*, 30, 1636–1642.
- Chen, Tinggui, Li, Y and Zhang, L. (2017) Nine Different Chemical Species and Action Mechanisms of Pancreatic Lipase Ligands Screened Out from *Forsythia suspensa* Leaves all at One Time. *Molecules*: 22:79.
- Chen, C., Mangoni, M. L., and Di, Y. P. (2017). In vivo therapeutic efficacy of frog skin-derived peptides against *Pseudomonas aeruginosa*-induced pulmonary infection. *Scientific Reports*, 7(1), 8548.
- Chen, Y., Guarnieri, M.T., Vasil, A.I., Vasil, M.L., Mant, C.T. and Hodges, R.S., 2007. Role of peptide hydrophobicity in the mechanism of action of α -helical antimicrobial peptides. *Antimicrobial Agents and Chemotherapy*, 51(4),1398-1406.
- Chertov, Oleg, Dennis F. Michiel, Luoling Xu, Ji Ming Wang, Kenji Tani, William J. Murphy, Dan L. Longo, Dennis D. Taub, and Joost J. Oppenheim (1996). Identification of defensin-1, defensin-2, and CAP37/azurocidin as T-cell chemoattractant proteins released from interleukin-8-stimulated neutrophils. *Journal of Biological Chemistry*, 271(6), 2935-2940.
- Chinchar, V. G., Bryan, L., Silphadaung, U., Noga, E., Wade, D., and Rollins-Smith, L. (2004). Inactivation of viruses infecting ectothermic animals by amphibian and piscine antimicrobial peptides. *Virology*, 323(2), 268-275.
- Cho, J.H., Park, I.Y., Kim, H.S., Lee, W.T., Kim, M.S. and Kim, S.C., (2002). Cathepsin D produces antimicrobial peptide parasin I from histone H2A in the skin mucosa of fish. *The FASEB Journal*, 16(3), 429-431.
- Chotipuntu, P., and Avakul, P. (2011). Aquaculture potential of climbing perch, *Anabas testudineus*, in brackish water. *Walailak Journal of Science and Technology (WJST)*, 7(1), 15-21.
- Chowdhury, M.I., Mahmud, A.I. and Rahman, A.F.M.A. (2014). Effects of Water Salinity on Feeding Efficiencies, Growth Performances and Survival Rate of Thai Strain Koi, *Anabas testudineus* (Bloch, 1792). *World Journal of Fish and Marine Science*, 6(5),479-486.
- Chromek, M., Slamová, Z., Bergman, P., Kovács, L., Podracká, L.U., Ehrén, I., Höckfelt, T., Gudmundsson, G.H., Gallo, R.L., Agerberth, B. and Brauner, A. (2006). The antimicrobial peptide cathelicidin protects the urinary tract against invasive bacterial infection. *Nature medicine*, 12(6), 636-641.

- Cirioni, O., Silvestri, C., Pierpaoli, E., Barucca, A., Kamysz, W., Ghiselli, R., Scalise, A., Brescini, L., Castelli, P., Orlando, F. and Kamysz, E. (2013). IB-367 pre-treatment improves the in vivo efficacy of teicoplanin and daptomycin in an animal model of wounds infected with meticillin-resistant *Staphylococcus aureus*. *Journal of Medical Microbiology*, 62(10),1552-1558.
- Cirioni, O., Silvestri, C., Ghiselli, R., Orlando, F., Riva, A., Gabrielli, E., Mocchegiani, F., Cianforlini, N., Trombettoni, M.M.C., Saba, V. and Scalise, G., (2009). Therapeutic efficacy of buforin II and rifampin in a rat model of *Acinetobacter baumannii* sepsis. *Critical Care Medicine*, 37(4), 1403-1407.
- Coates, C. J., and Decker, H. (2017). Immunological properties of oxygen-transport proteins: hemoglobin, hemocyanin and hemerythrin. *Cellular and Molecular Life Sciences*, 74(2), 293-317.
- Cole, A.M., Weis, P. and Diamond, G. (1997). Isolation and characterization of pleurocidin, an antimicrobial peptide in the skin secretions of winter flounder. *Journal of Biological Chemistry*, 272(18), 12008-12013.
- Cole, S. J., Records, A. R., Orr, M. W., Linden, S. B., and Lee, V. T. (2014). Catheter-associated urinary tract infection by *Pseudomonas aeruginosa* is mediated by exopolysaccharide-independent biofilms. *Infection and immunity*, 82(5), 2048-2058.
- Colerangle, J. B. (2013). Preclinical development of non-oncogenic drugs (Small and Large Molecules). Retrieved from <http://dx.doi.org/10.1016/B978-0-12-387815-1.00022,30/12/2017>.
- Conceição, K., Monteiro-dos-Santos, J., Seibert, C. S., Silva, P. I., Marques, E. E., Richardson, M., and Lopes-Ferreira, M. (2012). Potamotrygon cf. henlei stingray mucus: Biochemical features of a novel antimicrobial protein. *Toxicon*, 60(5), 821-829.
- Concha, M. I., Molina, S., Oyarzún, C., Villanueva, J., and Amthauer, R. (2003). Local expression of apolipoprotein AI gene and a possible role for HDL in primary defence in the carp skin. *Fish and Shellfish Immunology*, 14(3), 259-273.
- Concha, M. I., Smith, V. J., Castro, K., Bastías, A., Romero, A., and Amthauer, R. J. (2004). Apolipoproteins A- I and A- II are potentially important effectors of innate immunity in the teleost fish *Cyprinus carpio*. *The FEBS Journal*, 271(14), 2984-2990.
- Conlon, J. M. (2007). Purification of naturally occurring peptides by reversed-phase HPLC. *Nature protocols*, 2(1), 191-197.
- Cooper, R., and Molan, P. (1999). The use of honey as an antiseptic in managing *Pseudomonas* infection. *Journal of Wound Care*, 8(4), 161-164.

- Cordero, H., Brinchmann, M. F., Cuesta, A., Meseguer, J., and Esteban, M. A. (2015). Skin mucus proteome map of European sea bass (*Dicentrarchus labrax*). *Proteomics*, 15(23-24), 4007-4020.
- Cordero, H., Cuesta, A., Meseguer, J., and Esteban, M. A. (2016a). Changes in the levels of humoral immune activities after storage of gilthead seabream (*Sparus aurata*) skin mucus. *Fish and Shellfish Immunology*, 58, 500-507.
- Cordero, H., Morcillo, P., Cuesta, A., Brinchmann, M. F., and Esteban, M. A. (2016b). Differential proteome profile of skin mucus of gilthead seabream (*Sparus aurata*) after probiotic intake and/or overcrowding stress. *Journal of Proteomics*, 132, 41-50.
- Costa, R.J., C Silva, N., Sarmiento, B., and Pintado, M. (2017). Delivery Systems for Antimicrobial Peptides and Proteins: Towards Optimization of Bioavailability and Targeting. *Current pharmaceutical biotechnology*, 18(2), 108-120.
- Cox, C. D. (1979). Passage of *Pseudomonas aeruginosa* in compromised mice. *Infection and immunity*, 26(1), 118-124
- Cuesta, A., Meseguer, J., and Esteban, M. A. (2011). Molecular and functional characterization of the gilthead seabream β -defensin demonstrate its chemotactic and antimicrobial activity. *Molecular immunology*, 48(12), 1432-1438.
- Cyriac, J. M., & James, E. (2014). Switch over from intravenous to oral therapy: A concise overview. *Journal of Pharmacology and Pharmacotherapeutics*, 5(2), 83.
- Dash, S., Samal, J., and Thatoi, H. (2014). A comparative study on innate immunity parameters in the epidermal mucus of Indian major carps. *Aquaculture international*, 22(2), 411-421.
- De Jong, H.K., Van Der Poll, T. and Wiersinga, W.J., (2010). The systemic pro-inflammatory response in sepsis. *Journal of Innate Immunity*, 2(5), 422-430.
- de Almeida Vaucher, R., Gewehr, C. D. C. V., Correa, A. P. F., Sant 'Anna, V., Ferreira, J., & Brandelli, A. (2011). Evaluation of the immunogenicity and in vivo toxicity of the antimicrobial peptide P34. *International Journal of Pharmaceutics*, 421(1), 94-98.
- Dellinger, R.P., Levy, M.M., Rhodes, A., Annane, D., Gerlach, H., Opal, S.M., Sevransky, J.E., Sprung, C.L., Douglas, I.S., Jaeschke, R. and Osborn, T.M. (2013). Surviving Sepsis Campaign: international guidelines for management of severe sepsis and septic shock, 2012. *Intensive Care Medicine*, 39(2), 165-228.
- Delvaeye, M., and Conway, E. M. (2009). Coagulation and innate immune responses: can we view them separately? *Blood*, 114(12), 2367-2374.

- Diamond, G., Zasloff, M., Eck, H., Brasseur, M., Maloy, W. L., and Bevins, C. L. (1991). Tracheal antimicrobial peptide, a cysteine-rich peptide from mammalian tracheal mucosa: peptide isolation and cloning of a cDNA. *Proceedings of the National Academy of Sciences*, 88(9), 3952-3956.
- Di Bonaventura, I., Jin, X., Visini, R., Probst, D., Javor, S., Gan, B.H., Michaud, G., Natalello, A., Doglia, S.M., Köhler, T. and van Delden, C. (2017). Chemical space guided discovery of antimicrobial bridged bicyclic peptides against *Pseudomonas aeruginosa* and its biofilms. *Chemical Science*, 8(10), 6784-6798.
- Dickerson, H. W. (2009). The biology of teleost mucosal immunity. *Fish Defenses*, 2, 1-42.
- Doi, K., Leelahavanichkul, A., Yuen, P. S., and Star, R. A. (2009). Animal models of sepsis and sepsis-induced kidney injury. *The Journal of Clinical Investigation*, 119(10), 2868.
- Domingos, M.O., Franzolin, M.R., dos Anjos, M.T., Franzolin, T.M., Albes, R.C.B., de Andrade, G.R., Lopes, R.J. and Barbaro, K.C. (2011). The influence of environmental bacteria in freshwater stingray wound-healing. *Toxicon*, 58(2), 147-153.
- Dong, W., Mao, X., Guan, Y., Kang, Y., and Shang, D. (2017). Antimicrobial and anti-inflammatory activities of three chensinin-1 peptides containing mutation of glycine and histidine residues. *Scientific Reports*, 7, 40228.
- Dongarrà, M. L., Rizzello, V., Muccio, L., Fries, W., Cascio, A., Bonaccorsi, I., and Ferlazzo, G. (2013). Mucosal immunology and probiotics. *Current Allergy and Asthma Reports*, 13(1), 19-26.
- Douglas, S.E.; Gallant, J.W.; Gong, Z.; Hew, C (2001). Cloning and developmental expression of a family of pleurocidin-like antimicrobial peptides from winter flounder, *Pleuronectes americanus* (Walbaum). *Developmental and Comparative Immunology*. 25, 137–147.
- Dubois, V., Arpin, C., Melon, M., Melon, B., Andre, C., Frigo, C., and Quentin, C. (2001). Nosocomial outbreak due to a multiresistant strain of *Pseudomonas aeruginosa* P12: efficacy of cefepime-amikacin therapy and analysis of β -lactam resistance. *Journal of Clinical Microbiology*, 39(6), 2072-2078.
- Duran-Bedolla, J., de Oca-Sandoval, M. A. M., Saldaña-Navor, V., Villalobos-Silva, J. A., Rodriguez, M. C., and Rivas-Arancibia, S. (2014). Sepsis, mitochondrial failure and multiple organ dysfunction. *Clinical and Investigative Medicine*, 37(2), 58-69.

- Easy, R. H., and Ross, N. W. (2009). Changes in Atlantic salmon (*Salmo salar*) epidermal mucus protein composition profiles following infection with sea lice (*Lepeophtheirus salmonis*). *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*, 4(3), 159-167.
- Ebbensgaard, A., Mordhorst, H., Overgaard, M.T., Nielsen, C.G., Aarestrup, F.M. and Hansen, E.B., (2015) Comparative evaluation of the antimicrobial activity of different antimicrobial peptides against a range of pathogenic bacteria. *PloS one*, 10(12), e0144611.
- Ebran, N., Julien, S., Orange, N., Saglio, P., Lemaître, C., and Molle, G. (1999). Pore-forming properties and antibacterial activity of proteins extracted from epidermal mucus of fish. *Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology*, 122(2), 181-189.
- Ellis, A.E., (1999). Immunity to bacteria in fish. *Fish and Shellfish Immunology*, 9(4), 291-308.
- El-Solh, A.A., Hattemer, A., Hauser, A.R., Alhajhusain, A. and Vora, H. (2012). Clinical outcomes of type III *Pseudomonas aeruginosa* bacteremia. *Critical Care Medicine*, 40(4), 1157.
- Esteban, M.A (2012). An overview of the immunological defenses in fish skin. *International Scholarly Research Network, Immunology*, 2012,1-29
- Esteban, M.A. and Cerezuela, R. (2015) Fish Mucosal Immunity: Skin. In: Beck and Peatman, (Eds.), *Mucosal Health in Aquaculture* (pp.67-920). Cambridge: Academic Press.
- Estévez, R.A., Mostazo, M.G.C., Rodríguez, E., Espinoza, J.C., Kuznar, J., Jónsson, Z.O., Guðmundsson, G.H. and Maier, V.H. (2018). Inducers of salmon innate immunity: An in vitro and in vivo approach. *Fish and Shellfish Immunology*, 72,247-258
- Estévez, Rosana A., Miriam G. Contreras Mostazo, Eduardo Rodríguez, Juan Carlos Espinoza, Juan Kuznar, Zophonías O. Jónsson, Guðmundur H. Guðmundsson, and Valerie H. Maier. "Inducers of salmon innate immunity: An in vitro and in vivo approach." *Fish and Shellfish Immunology* 72 (2018): 247-258.
- Faix, J. D. (2013). Biomarkers of sepsis. *Critical Reviews in Clinical Laboratory Sciences*, 50(1), 23-36.
- Fast, M. D., Sims, D. E., Burka, J. F., Mustafa, A., and Ross, N. W. (2002). Skin morphology and humoral non-specific defence parameters of mucus and plasma in rainbow trout, coho and Atlantic salmon. *Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology*, 132(3), 645-657.

- Feng, X., Sambanthamoorthy, K., Palys, T., and Parnavitana, C. (2013). The human antimicrobial peptide LL-37 and its fragments possess both antimicrobial and antibiofilm activities against multidrug-resistant *Acinetobacter baumannii*. *Peptides*, 49, 131-137.
- Ferguson H. W., Morrison, D., Ostland, V. E., Lumsden, J., and Byrne, P. (1992). Responses of mucus-producing cells in gill disease of rainbow trout (*Oncorhynchus mykiss*). *Journal of comparative pathology*, 106(3), 255-265.
- Fernandes, J. M., MOLLE, M. G., and Smith, V. J. (2002a). Anti-microbial properties of histone H2A from skin secretions of rainbow trout, *Oncorhynchus mykiss*. *Biochemical Journal*, 368(2), 611-620.
- Fernandes, J.M. and Smith, V.J.(2002b). A novel antimicrobial function for a ribosomal peptide from rainbow trout skin. *Biochemical and Biophysical Research Communications*, 296(1), 167-171
- Fernandes, J.M., Molle, G., Kemp, G.D. and Smith, V.J. (2004a). Isolation and characterisation of oncorhyncin II, a histone H1-derived antimicrobial peptide from skin secretions of rainbow trout, *Oncorhynchus mykiss*. *Developmental and Comparative Immunology*, 28(2), 127-138.
- Fernandes, J. M. O., Kemp, G. D., and Smith, V. (2004b). Two novel muramidases from skin mucosa of rainbow trout (*Oncorhynchus mykiss*). *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 138(1), 53-64.
- Fernandes, J.M., Saint, N. and Smith, V.J. (2003). Oncorhyncin III: a potent antimicrobial peptide derived from the non-histone chromosomal protein H6 of rainbow trout, *Oncorhynchus mykiss*. *Biochemical Journal*, 373(2), 621-628.
- FishBas (2011). Retrieved from <http://www.fishbase.org>
- Fletcher, T. C. (1982). Non-specific defence mechanisms of fish. *Developmental and Comparative Immunology*, 2, 123-132.
- Fox, J. L. (2013). Antimicrobial peptides stage a comeback. *Nature Biotechnology*, 31, 5, 379-82
- Frimodt-Mo, N., Ier, J. D., and Espersen, E. (1999). The Mouse Peritonitis / Sepsis Model. In: Zak, O and Merle A. Sande, A.M. (Eds.), *Handbook of Animal Models of Infection: Experimental Models in Antimicrobial Chemotherapy* (pp.127-136). Academic Press.
- Furuya, N., Hirakata, Y., Tomono, K., Matsumoto, T., Tateda, K., Kaku, M., and Yamaguchi, K. (1993). Mortality rates amongst mice with endogenous septicemia caused by *Pseudomonas aeruginosa* isolates from various clinical sources. *Journal of Medical Microbiology*, 39(2), 141-146.

- Gaglione, R., Dell'Olmo, E., Bosso, A., Chino, M., Pane, K., Ascione, F., Itri, F., Caserta, S., Amoresano, A., Lombardi, A. and Haagsman, H.P. (2017). Novel human bioactive peptides identified in Apolipoprotein B: Evaluation of their therapeutic potential. *Biochemical pharmacology*, 130, 34-50.
- Ganz, T., Gabayan, V., Liao, H. I., Liu, L., Oren, A., Graf, T., and Cole, A. M. (2003). Increased inflammation in lysozyme M-deficient mice in response to *Micrococcus luteus* and its peptidoglycan. *Blood*, 101(6), 2388-2392.
- Geleta, B., Makonnen, E., and Debella, A. (2016). Toxicological evaluations of the crude extracts and fractions of *Moringa stenopetala* leaves in liver and kidney of rats. *Journal of Cytology and Histology*, 7(383), 10-4172.
- Gellatly, S. L., and Hancock, R. E. (2013). *Pseudomonas aeruginosa*: new insights into pathogenesis and host defenses. *Pathogens and Disease*, 67(3), 159-173.
- Girlich, D., Naas, T. and Nordmann, P. (2004). Biochemical characterization of the naturally occurring oxacillinase OXA-50 of *Pseudomonas aeruginosa*. *Antimicrobial Agents and Chemotherapy*, 48(6)2043-2048.
- Gomez, D., Sunyer, J.O. and Salinas, I. (2013). The mucosal immune system of fish: the evolution of tolerating commensals while fighting pathogens. *Fish and Shellfish Immunology*, 35(6), 1729-1739.
- Gómez-Zorrilla, S., Calatayud, L., Juan, C., Cabot, G., Tubau, F., Oliver, A., Dominguez, M.A., Ariza, J. and Peña, C. (2017). Understanding the acute inflammatory response to *Pseudomonas aeruginosa* infection: differences between susceptible and multidrug-resistant strains in a mouse peritonitis model. *International journal of antimicrobial agents*, 49(2), 198-203.
- Gonnert, F.A., Recknagel, P., Seidel, M., Jbeily, N., Dahlke, K., Bockmeyer, C.L., Winning, J., Lösche, W., Claus, R.A. and Bauer, M. (2011). Characteristics of clinical sepsis reflected in a reliable and reproducible rodent sepsis model. *Journal of Surgical Research*, 170(1), e123-e134.
- Grisaru-Soen, G., Lerner-Geva, L., Keller, N., Berger, H., Passwell, J. H., and Barzilai, A. (2000). *Pseudomonas aeruginosa* bacteremia in children: analysis of trends in prevalence, antibiotic resistance and prognostic factors. *The Pediatric Infectious Disease Journal*, 19(10), 959-963.
- Guardiola, F. A., Cuartero, M., del Mar Collado-González, M., Baños, F. G. D., Cuesta, A., Moriñigo, M. Á., and Esteban, M. Á. (2017). Terminal carbohydrates abundance, immune related enzymes, bactericidal activity and physico-chemical parameters of the Senegalese sole (*Solea senegalensis*, Kaup) skin mucus. *Fish and Shellfish Immunology*, 60, 483-491.

- Guardiola, F.A., Cuesta, A., Abellán, E., Meseguer, J. and Esteban, M.A. (2014). Comparative analysis of the humoral immunity of skin mucus from several marine teleost fish. *Fish and Shellfish Immunology*, 40(1), 24-31.
- Guo, M., Wei, J., Huang, X., Huang, Y. and Qin, Q. (2012). Antiviral effects of β -defensin derived from orange-spotted grouper (*Epinephelus coioides*). *Fish and Shellfish Immunology*, 32(5), 828-838.
- Gupta, R., Sarkar, S., and Srivastava, S. (2014). In vivo toxicity assessment of antimicrobial peptides (AMPs LR14) derived from *Lactobacillus plantarum* strain LR/14 in *Drosophila melanogaster*. *Probiotics and Antimicrobial Proteins*, 6(1), 59-67.
- Haard, N. F., and Simpson, B. K. (1994). Proteases from aquatic organisms and their uses in the seafood industry. In *Fisheries Processing* 132-154. Springer US.
- Hale, J. D., and Hancock, R. E. (2007). Alternative mechanisms of action of cationic antimicrobial peptides on bacteria. *Expert Review of Anti-infective Therapy*, 5(6), 951-959.
- Halim, A. M., Nabi, M. M., and Nahar, S. (2017). Study on optimization of stocking density of climbing perch (*Anabas testudineus*, Bloch 1792) in marginal farmer earthen ponds. *Journal of Entomology and Zoology Studies*, 5(3), 833-837.
- Hames, B. D. (Ed.). (1998). *Gel electrophoresis of proteins: a practical approach* (Vol. 197). OUP Oxford.
- Hancock, R. E., and Sahl, H. G. (2006). Antimicrobial and host-defense peptides as new anti-infective therapeutic strategies. *Nature Biotechnology*, 24(12), 1551-1557.
- Hancock, R. E., and Scott, M. G. (2000). The role of antimicrobial peptides in animal defenses. *Proceedings of The National Academy of Sciences*, 97(16), 8856-8861.
- Hancock, R. E., and Speert, D. P. (2000). Antibiotic resistance in *Pseudomonas aeruginosa*: mechanisms and impact on treatment. *Drug Resistance Updates*, 3(4), 247-255.
- Hancock, R.E., and Brinkman, F.S. (2002). Function of *Pseudomonas* porins in uptake and efflux. *Annual Reviews in Microbiology*, 56(1), 17-38.
- Haniffa, M. A., Viswanathan, S., Jancy, D., Poomari, K., & Manikandan, S. (2014). Antibacterial studies of fish mucus from two marketed air-breathing fishes—*Channa striatus* and *Heteropneustes fossilis*. *International Research Journal of Microbiology*, 5(2), 22-27.

- Harnedy, P. A., and FitzGerald, R. J. (2011). Bioactive proteins, peptides, and amino acids from macroalgae. *Journal of Phycology*, 47(2), 218-232.
- Harris, F., Dennison, S. R., and Phoenix, D. A. (2009). Anionic antimicrobial peptides from eukaryotic organisms. *Current Protein and Peptide Science*, 10(6), 585-606
- Hartley, B.S., (1960). Proteolytic enzymes. *Annual review of biochemistry*, 29(1), 45-72.
- Hauser, A. R. (2009). The type III secretion system of *Pseudomonas aeruginosa*: infection by injection. *Nature Reviews Microbiology*, 7(9), 654-665.
- Heidari, B. and Farzadfar, F. (2017). Effects of temperature and gonadal growth on the lysozyme level of immune tissues in the male and female Caspian kutum (*Rutilus frisikutum*). *Aquaculture Research*, 48(2), 377-385.
- Held, P. (2006). Quantitation of total protein using OPA. *Nature Methods Application Notes*.
- Hellio, C., Pons, A. M., Beaupoil, C., Bourgougnon, N., and Le Gal, Y. (2002). Antibacterial, antifungal and cytotoxic activities of extracts from fish epidermis and epidermal mucus. *International Journal of Antimicrobial Agents*, 20(3), 214-219.
- Hirakata, Y., Furuya, N., Tateda, K., Kaku, M., & Yamaguchi, K. (1993). In vivo production of exotoxin A and its role in endogenous *Pseudomonas aeruginosa* septicemia in mice. *Infection and Immunity*, 61(6), 2468-2473.
- Hisar, Ş. A., Hisar, O., Yılmaz, S., Çakır, F., Şahin, T., and Uyanık, M. H. (2014). In Vitro Antimicrobial and Antifungal Activities of Aqueous Skin Mucus from Rainbow Trout (*Oncorhynchus Mykiss*) on Human Pathogens. *Marine Science and Technology Bulletin*, 3(1):19-22.
- Hiwarale, D. K., Khillare, Y. K., Khillare, K., Wagh, U., Sawant, J., and Magare, M. (2016). Assessment of antibacterial properties of fish mucus. *World Journal of Pharmacy and Pharmaceutical Sciences*, 5, 5, 666-672.
- Hooper, D. C. (2001). Emerging mechanisms of fluoroquinolone resistance. *Emerging Infectious Diseases*, 7(2), 337.
- Hotchkiss, R. S., and Karl, I. E. (2003). The pathophysiology and treatment of sepsis. *New England Journal of Medicine*, 348(2), 138-150.
- Huang, P. H., Chen, J. Y., and Kuo, C. M. (2007). Three different hepcidins from tilapia, *Oreochromis mossambicus*: analysis of their expressions and biological functions. *Molecular Immunology*, 44(8), 1922-1934.
- Hudzicki, J. (2009). Kirby-Bauer disk diffusion susceptibility test protocol.

- Hussin, N.M., Shaarani, S.M., Sulaiman, M.R., Ahmad, A.H. and Vairappan, C.S. (2017). Chemical Composition and Antioxidant Activities of Catfish Epidermal Mucus. *Journal of Advanced Agricultural Technologies*, 4(1)
- Iger, Y. and Abraham, M. (1997). Rodlet cells in the epidermis of fish exposed to stressors. *Tissue and Cell*, 29(4), 431-438.
- Iger, Y., and Abraham, M. (1990). The process of skin healing in experimentally wounded carp. *Journal of Fish Biology*, 36(3), 421-437.
- Inderlied, C. B., Lancero, M. G., and Young, L. S. (1989). Bacteriostatic and bactericidal in-vitro activity of meropenem against clinical isolates, including *Mycobacterium avium* complex. *Journal of Antimicrobial Chemotherapy*, 24(suppl_A), 85-99.
- Ingram, G.A. (1980). Substances involved in the natural resistance of fish to infection—a review. *Journal of Fish Biology*, 16(1), 23-60.
- Infante, C., Asensio, E., Cañavate, J. P., and Manchado, M. (2008). Molecular characterization and expression analysis of five different elongation factor 1 alpha genes in the flatfish Senegalese sole (*Solea senegalensis* Kaup): differential gene expression and thyroid hormones dependence during metamorphosis. *BMC molecular biology*, 9(1), 19.
- Ip, K.Y., Melody M. L. Soh¹, Xiu L. Chen¹, Jasmine L. Y. Ong¹, You R. Chng¹, Biyun Ching¹, Wai P. Wong¹, Siew H. Lam^{1,3}, Shit F. Chew (2013) Molecular Characterization of Branchial aquaporin and Effects of Seawater Acclimation, Emersion or Ammonia Exposure on Its mRNA Expression in the Gills, Gut, Kidney and Skin of the Freshwater Climbing Perch, *Anabas testudineus*. *Plos One*: 8: 4: e61163
- Ivanov, V. T., Karelin, A. A., Philippova, M. M., Nazimov, I. V., and Pletnev, V. Z. (1997). Hemoglobin as a source of endogenous bioactive peptides: The concept of tissue- specific peptide pool. *Peptide Science*, 43(2), 171-188.
- Izadpanah, A., and Gallo, R. L. (2005). Antimicrobial peptides. *Journal of the American Academy of Dermatology*, 52(3), 381-390.
- Janeway Jr, C. A., and Medzhitov, R. (2002). Innate immune recognition. *Annual Review of Immunology*, 20(1), 197-216.
- Jones, S. R. (2001). The occurrence and mechanisms of innate immunity against parasites in fish. *Developmental and Comparative Immunology*, 25(8), 841-852.
- Millán, J. L. (2006). Alkaline Phosphatases: Structure, substrate specificity and functional relatedness to other members of a large superfamily of enzymes. *Purinergic Signal*, 2(2), 335-41.

- Jothi, G.E.G., Deivasigamani, B., Priyadarshini, P., Rajasekar, T., Balamurugan, S. and Manikantan, G. (2014). Haemolytic and Antimicrobial Efficacy of the Epidermal Mucus of the Striped Dwarf Catfish, *Mystus vittatus* (Bloch 1794) from Vellar Estuary, Parangipettai. *Inventi Rapid: Pharm Biotech and Microbio*, 2014, 1, 1-6.
- Jouyban, A., and Fakhree, M. A. (2012). Experimental and Computational Methods Pertaining to Drug Solubility Retrieved from <https://www.intechopen.com/books/toxicity-and-drug-testing>, 12/11/2017
- Johnston, L. D., Brown, G., Gauthier, D., Reece, K., Kator, H., & Van Veld, P. (2008). Apolipoprotein AI from striped bass (*Morone saxatilis*) demonstrates antibacterial activity in vitro. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 151(2), 167-175.
- Jung, T.S., Del Castillo, C.S., Javaregowda, P.K., Dalvi, R.S., Nho, S.W., Park, S.B., Jang, H.B., Cha, I.S., Sung, H.W., Hikima, J.I. and Aoki, T (2012). Seasonal variation and comparative analysis of non-specific humoral immune substances in the skin mucus of olive flounder (*Paralichthys olivaceus*). *Developmental and Comparative Immunology*, 38(2), 295-301.
- Jung, W.K., Mendis, E., Je, J.Y., Park, P.J., Son, B.W., Kim, H.C., Cho, Y.K. and Kim, S.K. (2006). Angiotensin I-converting enzyme inhibitory peptide from yellowfin sole (*Limanda aspera*) frame protein and its antihypertensive effect in spontaneously hypertensive rats. *Food Chemistry*, 94(1), 26-32.
- Jyot, J., Balloy, V., Jouvion, G., Verma, A., Touqui, L., Huerre, M., Chignard, M. and Ramphal, R., (2011). Type II secretion system of *Pseudomonas aeruginosa*: in vivo evidence of a significant role in death due to lung infection. *Journal of Infectious Diseases*, 203(10), 1369-1377.
- Khan, H.A; B. B. Ghosh and S. K. Mukhopadhyay. (1976). Observations on the salinity tolerance of *Anabas testudineus* (Bloch) Central. Inl. Fish. Res. Inst., Barrackpore, West-Bengal, India. *Journal of the Inland Fisheries Society of India*, 8: 111-112.
- Kalle, M., Papareddy, P., Kasetty, G., Mörgelin, M., van der Plas, M.J., Rydengård, V., Malmsten, M., Albiger, B. and Schmidtchen, A. (2012). Host defense peptides of thrombin modulate inflammation and coagulation in endotoxin-mediated shock and *Pseudomonas aeruginosa* sepsis. *PLoS One*, 7(12), e51313.
- Kamel, G.M., Edeen, N.A.E., Yousef El-Mishad, M. and Ezzat, R.F. (2011). Susceptibility pattern of *Pseudomonas aeruginosa* against antimicrobial agents and some plant extracts with focus on its prevalence in different sources. *Global Veterinaria*, 6(1), 61-72.

- Kang, H. K., Kim, C., Seo, C. H., and Park, Y. (2017). The therapeutic applications of antimicrobial peptides (AMPs): A Patent Review. *Journal of Microbiology*, 55(1), 1-12.
- Marimuthu, K., Arumugam, J., Sandragasan, D., and Jegathambigai, R. (2009). Studies on the fecundity of native fish climbing perch (*Anabas testudineus*, Bloch) in Malaysia. *American-Eurasian Journal of Sustainable Agriculture*, 3(3), 266-274.
- Kessler, E., Safrin, M., Gustin, J. K., and Ohman, D. E. (1998). Elastase and the LasA protease of *Pseudomonas aeruginosa* are secreted with their propeptides. *Journal of Biological Chemistry*, 273(46), 30225-30231.
- Khansari, A. R., Balasch, J. C., Reyes-López, F. E., and Tort, L. (2017). Stressing the Inflammatory Network: Immuno-endocrine Responses to Allostatic Load in Fish. *Journal of Marine Science Research and Technology*, 1: 002,
- Khong, H. K., Kuah, M. K., Jaya-Ram, A., and Shu-Chien, A. C. (2009). Prolactin receptor mRNA is upregulated in discus fish (*Symphysodon aequifasciata*) skin during parental phase. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 153(1), 18-28.
- Kirschnek, S., and Gulbins, E. (2006). Phospholipase A2 functions in *Pseudomonas aeruginosa*-induced apoptosis. *Infection and Immunity*, 74(2), 850-860.
- Kitts, D.D. and Weiler, K., (2003). Bioactive proteins and peptides from food sources. Applications of bioprocesses used in isolation and recovery. *Current Pharmaceutical Design*, 9(16), 1309-1323.
- Kobayashi, F. (1971). Experimental infection with *Pseudomonas aeruginosa* in mice. *Japanese Journal of Microbiology*, 15(4), 301-307.
- Kohinoor, A.H.M., M. Akhteruzzaman, M.G. Hussain, M.S. Shah (1991). Observation on the induced breeding of koi fish (*Anabas testudineus*, Bloch) in Bangladesh. *Bangladesh Journal of Fisheries*, 14(1-2): 73-77.
- Köhler, T., and van Delden, C. (2009). *Pseudomonas aeruginosa* infections: from bench to bedside. *Revue Medicale Suisse*, 5(197), 732-734.
- Kong, K.F., Jayawardena, S.R., del Puerto, A., Wiehlmann, L., Laabs, U., Tümmler, B. and Mathee, K. (2005). Characterization of *poxB*, a chromosomal-encoded *Pseudomonas aeruginosa* oxacillinase. *Gene*, 358, 82-92.
- Kowser, M.M., Hoque, M.M. and Fatema, N. (2009). Determination of MIC and MBC of selected tetracycline capsule commercially available in Bangladesh. *ORION*, 32(3).

- Kravitz, M. S., Pitashny, M., and Shoenfeld, Y. (2005). Protective molecules—C-reactive protein (CRP), serum amyloid P (SAP), pentraxin3 (PTX3), mannose-binding lectin (MBL), and apolipoprotein A1 (Apo A1), and their autoantibodies: prevalence and clinical significance in autoimmunity. *Journal of Clinical Immunology*, 25(6), 582.
- Krumperman, P. H. (1983). Multiple antibiotic resistance indexing of *Escherichia coli* to identify high-risk sources of fecal contamination of foods. *Applied and Environmental Microbiology*, 46(1), 165-170.
- Kumar, H., Kawai, T., and Akira, S. (2011). Pathogen recognition by the innate immune system. *International Reviews of Immunology*, 30(1), 16-34.
- Kuppulakshmi, C., Prakash, M., Gunasekaran, G., Manimegalai, G., and Sarojini, S. (2008). Antibacterial properties of fish mucus from. *European Review for Medical and Pharmacological Sciences*, 12, 149-153.
- Ladokhin, A.S. and White, S.H., (2001). 'Detergent-like' permeabilization of anionic lipid vesicles by melittin. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1514(2), 253-260.
- Laemmli, U. K., and Favre, M. (1973). Maturation of the head of bacteriophage T4: I. DNA packaging events. *Journal of Molecular Biology*, 80(4), 575IN1593-592IN4599.
- Lambert, P.A. (2002). Mechanisms of antibiotic resistance in *Pseudomonas aeruginosa*. *Journal of the Royal Society of Medicine*, 95 (41),22.
- Laurent, P. (1984). Gill internal morphology. In: Hoar, W.S. and Randall, D.J. (Eds.), *Fish Physiology* (pp. 73-183). Orlando: Academic Press.
- Lauth, X., Shike, H., Burns, J.C., Westerman, M.E., Ostland, V.E., Carlberg, J.M., Van Olst, J.C., Nizet, V., Taylor, S.W., Shimizu, C. and Bulet, P. (2002). Discovery and characterization of two isoforms of moronecidin, a novel antimicrobial peptide from hybrid striped bass. *Journal of Biological Chemistry*, 277(7), 5030-5039.
- Lazarovici, P., Primor, N., and Loew, L. M. (1986). Purification and pore-forming activity of two hydrophobic polypeptides from the secretion of the Red Sea Moses sole (*Pardachirus marmoratus*). *Journal of Biological Chemistry*, 261(35), 16704-1671
- Le, C. F., Fang, C. M., and Sekaran, S. D. (2017). Intracellular targeting mechanisms by antimicrobial peptides. *Antimicrobial Agents and Chemotherapy*, 61(4), e02340-16.

- Lee, M.T., Sun, T.L., Hung, W.C. and Huang, H.W. (2013). Process of inducing pores in membranes by melittin. *Proceedings of the National Academy of Sciences*, 110(35), 14243-14248.
- Lee, Y. J., Kim, C. H., Oh, H. Y., Go, H. J., and Park, N. G. (2015). Antimicrobial, Antioxidant and Hemolytic Activity of Water-soluble Extract of Mottled Anemone *Urticina crassicornis*. *Fisheries and Aquatic Sciences*, 18(4), 341-347.
- Li, C., Zhu, J., Wang, Y., Chen, Y., Song, L., Zheng, W., Li, J. and Yu, R. (2017). Antibacterial Activity of AI-Hemocidin 2, a Novel N-Terminal Peptide of Hemoglobin Purified from *Arca inflata*. *Marine Drugs*, 15(7), 205.
- Li, P., Wohland T., Ho B., Ding J. L. (2004). Perturbation of lipopolysaccharide (LPS) micelles by Sushi 3 (S3) antimicrobial peptide. The importance of an intermolecular disulfide bond in S3 dimer for binding, disruption, and neutralization of LPS. *The Journal of Biological Chemistry*. 279(48):50150–50156.
- Li, Z., Zhang, S., Gao, J., Guang, H., Tian, Y., Zhao, Z., Wang, Y. and Yu, H. (2013). Structural and functional characterization of CATH_BRALE, the defense molecule in the ancient salmonoid, *Brachymystax lenok*. *Fish and Shellfish Immunology*, 34(1), 1-7.
- Liang, Y., Guan, R., Huang, W., and Xu, T. (2011). Isolation and identification of a novel inducible antibacterial peptide from the skin mucus of Japanese eel, *Anguilla japonica*. *The Protein Journal*, 30(6), 413.
- Liepke, C., Baxmann, S., Heine, C., Breithaupt, N., Ständker, L., and Forssmann, W. G. (2003). Human hemoglobin-derived peptides exhibit antimicrobial activity: a class of host defense peptides. *Journal of Chromatography B*, 791(1),345-356.
- Livermore, D. M. (2002). Multiple mechanisms of antimicrobial resistance in *Pseudomonas aeruginosa*: our worst nightmare? *Clinical infectious diseases*, 34(5), 634-640.
- Lobo, S. M., and Lobo, F. R. M. (2007). Markers and mediators of inflammatory response in infection and sepsis. *Revista Brasileira de Terapia Intensiva*, 19(2), 210-215.
- Loganathan K, Arulprakash A, Prakash M and Senthilraja P. (2013a). Lysozyme, Protease, Alkaline phosphatase and Esterase activity of epidermal skin mucus of freshwater snake head fish *Channa striatus*. *International Journal of Research in Pharmaceutical and Biosciences*, 3(1): 17-20.

- Loganathan, K., Prakash, M. and Senthilraja, P. (2013b). Antibacterial activity of ammonium precipitate extract of viral fish (*Channa striatus*) skin mucus. *International Journal of Innovation Research*, 1, 001-004.
- Loh, J. Y., and Ting, A. S. Y. (2015). Comparative study of analogue hormones and the embryonic, larval and juvenile development on the induced breeding of climbing perch (*Anabas testudineus*, Bloch, 1792). *Journal of Fisheries and Aquatic Science* 2, 277-278.
- Lu, X. J., Chen, J., Huang, Z. A., Shi, Y. H., and Lu, J. N. (2011). Identification and characterization of a novel cathelicidin from ayu, *Plecoglossus altivelis*. *Fish and Shellfish Immunology*, 31(1), 52-57.
- Lüders, T., Birkemo, G.A., Nissen-Meyer, J., Andersen, Ø. and Nes, I.F. (2005). Proline conformation-dependent antimicrobial activity of a proline-rich histone H1 N-terminal peptide fragment isolated from the skin mucus of Atlantic salmon. *Antimicrobial Agents and Chemotherapy*, 49(6), 2399-2406.
- Lugtenberg, B., and Van Alphen, L. (1983). Molecular architecture and functioning of the outer membrane of *Escherichia coli* and other gram-negative bacteria. *Biochimica et Biophysica Acta (BBA)-Reviews on Biomembranes*, 737(1), 51-115.
- Lundberg, J. S., Perl, T. M., Wiblin, T., Costigan, M. D., Dawson, J., Nettleman, M. D., and Wenzel, R. P. (1998). Septic shock: an analysis of outcomes for patients with onset on hospital wards versus intensive care units. *Critical Care Medicine*, 26(6), 1020-1024.
- Lyczak, J. B., Cannon, C. L., and Pier, G. B. (2000). Establishment of *Pseudomonas aeruginosa* infection: lessons from a versatile opportunist. *Microbes and Infection*, 2(9), 1051-1060.
- Mackman, N., (2006). Role of tissue factor in hemostasis and thrombosis. *Blood Cells, Molecules, and Diseases*, 36(2), 104-107.
- Magnadóttir, B. (2006). Innate immunity of fish (overview). *Fish and Shellfish Immunology*, 20(2), 137-151.
- Magnadóttir, B. (2010). Immunological control of fish diseases. *Marine Biotechnology*, 12(4), 361-379.
- Magnadóttir, B., Lange, S., Gudmundsdóttir, S., Bøgwald, J. and Dalmo, R.A. (2005). Ontogeny of humoral immune parameters in fish. *Fish and Shellfish Immunology*, 19(5), 429-439.

- Mahmood, S., Ali, M. S., and Anwar-Ul-Haque, M. O. H. A. M. M. A. D. (2004). Effect of different feed on larval/fry rearing of climbing perch, *Anabas testudineus* (Bloch), Bangladesh: II. Growth and survival. *Pakistan Journal of Zoology*, 36(1), 13-20.
- Manikantan, G., Lyla, S., Khan, S. A., Vijayanand, P., and Jothi, G. E. G. (2016). Journal of Coastal Life Medicine. *Journal of Coastal Life Medicine*, 4(7), 510-520.
- Manivasagan, P., Annamalai, N., Ashokkumar, S. and Sampathkumar, P., 2009. Studies on the proteinaceous gel secretion from the skin of the catfish, *Arius maculatus* (Thunberg, 1792). *African Journal of Biotechnology*, 8(24).
- Marcos-López, M., Ruiz, C.E., Rodger, H.D., O'Connor, I., MacCarthy, E. and Esteban, M.A., (2017). Local and systemic humoral immune response in farmed Atlantic salmon (*Salmo salar* L.) under a natural amoebic gill disease outbreak. *Fish and Shellfish Immunology*, 66,207-216.
- Marimuthu, K., Arumugam, J., Sandragasan, D., and Jegathambigai, R. (2009). Studies on the fecundity of native fish climbing perch (*Anabas testudineus*, Bloch) in Malaysia. *American-Eurasian Journal of Sustainable Agriculture*, 3(3), 266-274.
- Martin, L., van Meegern, A., Doemming, S., and Schuerholz, T. (2015). Antimicrobial peptides in human sepsis. *Frontiers in immunology*, 6, 404.
- Masso-Silva, J. A., and Diamond, G. (2014). Antimicrobial peptides from fish. *Pharmaceuticals*, 7(3), 265-310.
- McCarthy, K. L., and Paterson, D. L. (2017). Long-term mortality following *Pseudomonas aeruginosa* bloodstream infection. *Journal of Hospital Infection*, 95(3), 292-299.
- Mehta, R. L., Kellum, J. A., Shah, S. V., Molitoris, B. A., Ronco, C., Warnock, D. G., and Levin, A. (2007). Acute Kidney Injury Network: report of an initiative to improve outcomes in acute kidney injury. *Critical Care*, 11(2), R31.
- Meletis, G and Bagkeri, M. (2013). *Pseudomonas aeruginosa*: multi-drugresistance development and treatment options, infection control. In: *pseudomonas-aeruginosa-multi-drug-resistance-development and-treatment options* Retrieve dfrom: <https://www.intechopen.com/books/infection-control/12/10/2017>
- Mello, C.M. and Soares, J., (2008). Membrane Selectivity of Antimicrobial Peptides. *Microbial Surfaces ACS* 984:52–62.

- Ming, L., Xiaoling, P., Yan, L., Lili, W., Qi, W., Xiyong, Y., Boyao, W. and Ning, H. (2007). Purification of antimicrobial factors from human cervical mucus. *Human Reproduction*, 22(7), 1810-1815.
- Mookherjee, N., and Hancock, R. E. W. (2007). Cationic host defence peptides: innate immune regulatory peptides as a novel approach for treating infections. *Cellular and Molecular Life Sciences*, 64(7-8), 922.
- Moore, N.M. and Flaws, M.L. (2011a). Antimicrobial resistance mechanisms in *Pseudomonas aeruginosa*. *Clinical Laboratory Science*, 24(1), 47.
- Moore, N. M., and Flaws, M. L. (2011b). Epidemiology and pathogenesis of *Pseudomonas aeruginosa* infections. *Clinical Laboratory Science*, 24(1), 43.
- Mulero, I., Noga, E. J., Meseguer, J., García-Ayala, A., and Mulero, V. (2008). The antimicrobial peptides piscidins are stored in the granules of professional phagocytic granulocytes of fish and are delivered to the bacteria-containing phagosome upon phagocytosis. *Developmental and Comparative Immunology*, 32(12), 1531-1538.
- Murty, V. L. N., Sarosiek, J., Slomiany, A., and Slomiany, B. L. (1984). Effect of lipids and proteins on the viscosity of gastric mucus glycoprotein. *Biochemical and Biophysical Research Communications*, 121(2), 521-529.
- Mydin, H. H., Corris, P.A., Nicholson, A., Perry, J.D., Meachery, G., Marrs, E.C., Peart, S., Fagan, C., Lordan, J.L., Fisher, A.J. and Gould, F.K. (2012). Targeted antibiotic prophylaxis for lung transplantation in cystic fibrosis patients colonised with *Pseudomonas aeruginosa* using multiple combination bactericidal testing. *Journal of Transplant*, 2012, 135738.
- Nagashima, Y., Kikuchi, N., Shimakura, K., and Shiomi, K. (2003). Purification and characterization of an antibacterial protein in the skin secretion of rockfish *Sebastes schlegeli*. *Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology*, 136(1), 63-71.
- Nagashima, Y., Sendo, A., Shimakura, K., Shiomi, K., Kobayashi, T., Kimura, B., and Fujii, T. (2001). Antibacterial factors in skin mucus of rabbitfishes. *Journal of Fish Biology*, 58(6), 1761-1765.
- Nahar, F., Haque, W., Ahsan, D. A., and Mustafa, M. G. (2016). Effects of salinity changes on growth performance and survival of climbing perch, *Anabas testudineus* (Bloch, 1795). *Dhaka University Journal of Biological Sciences*, 25(1), 65-73.
- Narayana, J. L., Huang, H. N., Wu, C. J., and Chen, J. Y. (2015). Efficacy of the antimicrobial peptide TP4 against *Helicobacter pylori* infection: in vitro membrane perturbation via micellization and in vivo suppression of host immune responses in a mouse model. *Oncotarget*, 6(15), 12936

- Nateche, F., Martin, A., Baraka, S., Palomino, J.C., Khaled, S. and Portaels, F. (2006). Application of the resazurin microtitre assay for detection of multidrug resistance in *Mycobacterium tuberculosis* in Algiers. *Journal of Medical Microbiology*, 55(7), 857-860.
- Nathwani, D., Raman, G., Sulham, K., Gavaghan, M., and Menon, V. (2014). Clinical and economic consequences of hospital-acquired resistant and multidrug-resistant *Pseudomonas aeruginosa* infections: a systematic review and meta-analysis. *Antimicrobial Resistance and Infection control*, 3(1), 32.
- Neu, H. C. (1983). The role of *Pseudomonas aeruginosa* in infections. *Journal of Antimicrobial Chemotherapy*, 11(suppl_B), 1-13.
- Ng, A., Heynen, M., Luensmann, D., Subbaraman, L. N., and Jones, L. (2013). Optimization of a fluorescence-based lysozyme activity assay for contact lens studies. *Current Eye Research*, 38(2), 252-259.
- Nicas, T. I., and Hancock, R. E. (1983). *Pseudomonas aeruginosa* outer membrane permeability: isolation of a porin protein F-deficient mutant. *Journal of Bacteriology*, 153(1), 281-285.
- Nigam, A. K., Kumari, U., Mittal, S., and Mittal, A. K. (2012). Comparative analysis of innate immune parameters of the skin mucous secretions from certain freshwater teleosts, inhabiting different ecological niches. *Fish Physiology and Biochemistry*, 38(5), 1245-1256.
- Nigam, A. K., Kumari, U., Mittal, S., and Mittal, A. K. (2017). Evaluation of antibacterial activity and innate immune components in skin mucus of Indian major carp, *Cirrhinus mrigala*. *Aquaculture Research*, 48(2), 407-418.
- Nigam, A.K., Kumari, U., Mittal, S. and Mittal, A.K. (2014). Characterization of carboxylesterase in skin mucus of *Cirrhinus mrigala* and its assessment as biomarker of organophosphate exposure. *Fish Physiology and Biochemistry*, 40(3), 635-644.
- Nijnik, A., and Hancock, R. E. W. (2009). Host defence peptides: antimicrobial and immunomodulatory activity and potential applications for tackling antibiotic-resistant infections. *Emerging Health Threats Journal*, 2(1), 7078
- Niu, Su-Fang, Yuan Jin, Xin Xu, Ying Qiao, Yang Wu, Yong Mao, Yong-Quan Su, and Jun Wang. (2013). Characterization of a novel piscidin-like antimicrobial peptide from *Pseudosciaena crocea* and its immune response to *Cryptocaryon irritans*. *Fish and Shellfish Immunology*, 35(2), 513-524.
- Nizet, V., Ohtake, T., Lauth, X., Trowbridge, J., Rudisill, J., Dorschner, R.A., Pestonjamas, V., Piraino, J., Huttner, K. and Gallo, R.L. (2001). Innate antimicrobial peptide protects the skin from invasive bacterial infection. *Nature*, 414(6862), 454

- Noga, E. J., Fan, Z., and Silphaduang, U. (2002). Host site of activity and cytological effects of histone like proteins on the parasitic dinoflagellate *Amyloodinium ocellatum*. *Diseases of Aquatic Organisms*, 52(3), 207-215.
- Noga, E. J., Silphaduang, U., Park, N. G., Seo, J. K., Stephenson, J., and Kozłowicz, S. (2009). Piscidin 4, a novel member of the piscidin family of antimicrobial peptides. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 152(4), 299-305.
- Noga, E. J., Ullal, A. J., Corrales, J., and Fernandes, J. M. (2011). Application of antimicrobial polypeptide host defenses to aquaculture: Exploitation of downregulation and upregulation responses. *Comparative Biochemistry and Physiology Part D: Genomics and Proteomics*, 6(1), 44-54.
- Nollet, E., Van Craenenbroeck, E.M., Martinet, W., Rodrigus, I., De Bock, D., Berneman, Z., Pintelon, I., Ysebaert, D., Vrints, C.J., Conraads, V.M. and Van Hoof, V.O., (2015). Bone matrix vesicle-bound alkaline phosphatase for the assessment of peripheral blood admixture to human bone marrow aspirates. *Clinica Chimica Acta*, 446,253-260.
- Nonaka, M., and Miyazawa, S. (2001). Evolution of the initiating enzymes of the complement system. *Genome biology*, 3(1), reviews1001-1.
- Nsrelden, R. M., Horiuchi, H., and Furusawa, S. (2017). Expression of ayu antimicrobial peptide genes after LPS stimulation. *Journal of Veterinary Medical Science*, 16-0609.
- Oddo, A., and Hansen, P. R. (2017). Hemolytic Activity of Antimicrobial Peptides. *Antimicrobial Peptides: Methods and Protocols*, 427-435.
- OECD. In Acute oral toxicity–acute oral toxic class method. Guideline 423, (adopted23/06/1996) eleventh addendum to the OECD guidelines for testing of chemicals, organization for economic co-operation and development: Paris, 2001. [http://w.w.w.research.murdoch.edu.au/ethics/arec/oral toxicity](http://w.w.w.research.murdoch.edu.au/ethics/arec/oral%20toxicity).
- O’Fagain, C. (2004). Lyophilisation of proteins. In: Paul Cutler(Eds.), *Protein Purification Protocols* (pp.309-322). Totowa: Humana Press.
- Oppenheim, J. J., Biragyn, A., Kwak, L. W., and Yang, D. (2003). Roles of antimicrobial peptides such as defensins in innate and adaptive immunity. *Annals of The Rheumatic Diseases*, 62(suppl 2), ii17-ii21.
- Ostorhazi, E., Rozgonyi, F., Sztodola, A., Harnos, F., Kovalszky, I., Szabo, D., Knappe, D., Hoffmann, R., Cassone, M., Wade, J.D. and Bonomo, R.A., (2010). Preclinical advantages of intramuscularly administered peptide A3-APO over existing therapies in *Acinetobacter baumannii* wound infections. *Journal of Antimicrobial Chemotherapy*, 65(11), 2416-2422.

- Overbye, K. M., and Barrett, J. F. (2005). Antibiotics: where did we go wrong? *Drug Discovery Today*, 10(1), 45-52.
- Oyemitan, I. A. (2017). African Medicinal Spices of Genus Piper. In Kuete, V (Eds.), *Medicinal Spices and Vegetables from Africa* (pp. 581-597). Academic press
- Page, M. G., and Heim, J. (2009). Prospects for the next anti-Pseudomonas drug. *Current Opinion in Pharmacology*, 9(5), 558-565.
- Pai, H., Kim, J. W., Kim, J., Lee, J. H., Choe, K. W., and Gotoh, N. (2001). Carbapenem resistance mechanisms in *Pseudomonas aeruginosa* clinical isolates. *Antimicrobial Agents and Chemotherapy*, 45(2), 480-484.
- Palaksha, K. J., Shin, G. W., Kim, Y. R., and Jung, T. S. (2008). Evaluation of non-specific immune components from the skin mucus of olive flounder (*Paralichthys olivaceus*). *Fish and Shellfish Immunology*, 24(4), 479-488.
- Pan, C.Y., Chen, J.Y., Cheng, Y.S.E., Chen, C.Y., Ni, I.H., Sheen, J.F., Pan, Y.L. and Kuo, C.M (2007). Gene expression and localization of the epinecidin-1 antimicrobial peptide in the grouper (*Epinephelus coioides*), and its role in protecting fish against pathogenic infection. *DNA and cell biology*, 26(6), 403-413.
- Pan, C. Y., Chen, J. Y., Lin, T. L., and Lin, C. H. (2009). In vitro activities of three synthetic peptides derived from epinecidin-1 and an anti-lipopolysaccharide factor against *Propionibacterium acnes*, *Candida albicans*, and *Trichomonas vaginalis*. *Peptides*, 30(6), 1058-1068.
- Pan, C. Y., Chen, J. C., Sheen, J. F., Lin, T. L., and Chen, J. Y. (2014). Epinecidin-1 has immunomodulatory effects, facilitating its therapeutic use in a mouse model of *Pseudomonas aeruginosa* sepsis. *Antimicrobial Agents and Chemotherapy*, 58(8):4264-74.
- Papo, N., and Shai, Y. (2003). Exploring peptide membrane interaction using surface plasmon resonance: differentiation between pore formations versus membrane disruption by lytic peptides. *Biochemistry*, 42(2), 458-466.
- Parameswaran, N., & Patial, S. (2010). Tumor necrosis factor- α signaling in macrophages. *Critical Reviews™ in Eukaryotic Gene Expression*, 20(2).
- Park, C. B., Lee, J. H., Park, I. Y., Kim, M. S., and Kim, S. C. (1997). A novel antimicrobial peptide from the loach, *Misgurnus anguillicaudatus*. *FEBS letters*, 411(2-3), 173-178.
- Park, C. H., Valore, E. V., Waring, A. J., and Ganz, T. (2001). Hepsidin, a urinary antimicrobial peptide synthesized in the liver. *Journal of Biological Chemistry*, 276(11), 7806-7810.

- Park, I. Y., Park, C. B., Kim, M. S., and Kim, S. C. (1998). Parasin I, an antimicrobial peptide derived from histone H2A in the catfish, *Parasilurus asotus*. *FEBS letters*, 437(3), 258-262.
- Parseghian, M.H. and Luhrs, K.A. (2006). Beyond the walls of the nucleus: the role of histones in cellular signaling and innate immunity. *Biochemistry and Cell Biology*. 84, 589–604.
- Pastrana, L., González, R., Estévez, N., Pereira, L., Amado, I.R., Fuciños, P., Fuciños, C., Rúa, M.L., Alonso, E. and Troncoso, R., (2016). Functional Foods. *Current Developments in Biotechnology and Bioengineering: Food and Beverages Industry*, 165.
- Pasupuleti, M., Schmidtchen, A., and Malmsten, M. (2012). Antimicrobial peptides: key components of the innate immune system. *Critical Reviews in Biotechnology*, 32(2), 143-171.
- Patel, D. M., and Brinchmann, M. F. (2017). Skin mucus proteins of lumpsucker (*Cyclopterus lumpus*). *Biochemistry and Biophysics Reports*, 9, 217-225.
- Patel, J. B., Cockerill, F. R., Alder, J., Bradford, P. A., Eliopoulos, G. M., Hardy, D and Powell, M. (2014). Performance standards for antimicrobial susceptibility testing; twenty-fourth informational supplement. *CLSI standards for antimicrobial susceptibility testing*, 34(1), 1-226.
- Paterson, D. L. (2006). The epidemiological profile of infections with multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter* species. *Clinical Infectious Diseases*, 43(Supplement_2), S43-S48.
- Patrzykat, A., Zhang, L., Mendoza, V., Iwama, G. K., and Hancock, R. E. (2001). Synergy of histone-derived peptides of coho salmon with lysozyme and flounder pleurocidin. *Antimicrobial Agents and Chemotherapy*, 45(5), 1337-1342.
- Paul, S., Bezbaruah, R. L., Roy, M. K., and Ghosh, A. C. (1997). Multiple antibiotic resistance (MAR) index and its reversion in *Pseudomonas aeruginosa*. *Letters in Applied Microbiology*, 24(3), 169-171.
- Pawlinski, R., Pedersen, B., Schabbauer, G., Tencati, M., Holscher, T., Boisvert, W., Andrade-Gordon, P., Frank, R.D. and Mackman, N., (2004). Role of tissue factor and protease-activated receptors in a mouse model of endotoxemia. *Blood*, 103(4), 1342-1347.
- Peres, C. M., Alves, M., Hernandez-Mendoza, A., Moreira, L., Silva, S., Bronze, M. R., ... and Malcata, F. X. (2014). Novel isolates of lactobacilli from fermented Portuguese olive as potential probiotics. *LWT-Food Science and Technology*, 59(1), 234-246.

- Perez-Vilar, J., and Hill, R. L. (1999). The structure and assembly of secreted mucins. *Journal of Biological Chemistry*, 274(45), 31751-31754.
- Pérez-Sánchez, J., Terova, G., Simó-Mirabet, P., Rimoldi, S., Folkedal, O., Caldich-Giner, J.A., Olsen, R.E. and Sitjà-Bobadilla, A. (2017). Skin mucus of gilthead sea bream (*Sparus aurata* L.). Protein mapping and regulation in chronically stressed fish. *Frontiers in Physiology*, 8, 34.
- Peschel, A., and Sahl, H. G. (2006). The co-evolution of host cationic antimicrobial peptides and microbial resistance. *Nature Reviews Microbiology*, 4(7), 529-536.
- Peterson, T. S. (2015). 3-Overview of mucosal structure and function in teleost fishes. *Mucosal Health Aquac.*, Academic Press, San Diego, 55-65.
- Pethiyagoda, R. (1991). *Freshwater fishes of Sri Lanka: Wildlife Heritage Trust of Sri Lanka*. Colombo. 362.
- Pickering, A. D., and Fletcher, J. M. (1987). Sacciform cells in the epidermis of the brown trout, *Salmo trutta*, and the Arctic char, *Salvelinus alpinus*. *Cell and Tissue Research*, 247(2), 259-265.
- Pires, J., Siriwardena, T.N., Stach, M., Tinguely, R., Kasraian, S., Luzzaro, F., Leib, S.L., Darbre, T., Reymond, J.L. and Endimiani, A. (2015). In vitro activity of the novel antimicrobial peptide dendrimer G3KL against multidrug-resistant *Acinetobacter baumannii* and *Pseudomonas aeruginosa*. *Antimicrobial Agents and Chemotherapy*, 59(12), 7915-7918.
- Pitt, T. L. (1986). Biology of *Pseudomonas aeruginosa* in relation to pulmonary infection in cystic fibrosis. *Journal of the Royal Society of Medicine*, 79(Suppl 12), 13.
- Ponniah, A. G., and Sarkar, U. K. (2000). *Fish biodiversity of North East India*. Lucknow, India: NBFGR – NATP Publications.
- Pridgeon, J. W., and Klesius, P. H. (2013). Apolipoprotein A1 in channel catfish: transcriptional analysis, antimicrobial activity, and efficacy as plasmid DNA immunostimulant against *Aeromonas hydrophila* infection. *Fish and Shellfish Immunology*, 35(4), 1129-1137.
- Prier, J., Bartola E., and Friedman, H. (1973) *Quality Control in Microbiology*. University Park Press, Baltimore.
- Raetz, C. R., and Whitfield, C. (2002). Lipopolysaccharide endotoxins. *Annual Review of biochemistry*, 71(1), 635-700.

- Rahal, J. J. (2006). Novel antibiotic combinations against infections with almost completely resistant *Pseudomonas aeruginosa* and *Acinetobacter* species. *Clinical infectious diseases*, 43(Supplement_2), S95-S99.
- Rahman, A. K. A. (1989). *Freshwater fishes of Bangladesh* Zoological Society of Bangladesh: Department of Zoology. University of Dhaka, 364.
- Rahman, S., Monir, M. S., and Khan, M. H. (2013). Culture potentials of climbing perch, Thai Koi, *Anabas testudineus* (Bloch) under different stocking densities in northern regions of Bangladesh. *Journal of Experimental Biology*, 1, 3.
- Rahnamaeian, M., Cytryńska, M., Zdybicka-Barabas, A., and Vilcinskas, A. (2016). The functional interaction between abaecin and pore-forming peptides indicates a general mechanism of antibacterial potentiation. *Peptides*, 78, 17-23.
- Rajamuthiah, R., Jayamani, E., Conery, A.L., Fuchs, B.B., Kim, W., Johnston, T., Vilcinskas, A., Ausubel, F.M. and Mylonakis, E. (2015). A defensin from the model beetle *Tribolium castaneum* acts synergistically with Telavancin and Daptomycin against multidrug resistant *Staphylococcus aureus*. *PLoS One*, 10(6), e0128576.
- Rajan, B., Fernandes, J. M., Caipang, C. M., Kiron, V., Rombout, J. H., and Brinchmann, M. F. (2011). Proteome reference map of the skin mucus of Atlantic cod (*Gadus morhua*) revealing immune competent molecules. *Fish and shellfish immunology*, 31(2), 224-231.
- Rajan, B., Lokesh, J., Kiron, V., and Brinchmann, M. F. (2013). Differentially expressed proteins in the skin mucus of Atlantic cod (*Gadus morhua*) upon natural infection with *Vibrio anguillarum*. *BMC veterinary research*, 9(1), 103.
- Rajanbabu, V., and Chen, J. Y. (2011). Applications of antimicrobial peptides from fish and perspectives for the future. *Peptides*, 32(2), 415-420.
- Rakers, S., Gebert, M., Uppalapati, S., Meyer, W., Maderson, P., Sell, A.F., Kruse, C. and Paus, R. (2010). 'Fish matters': the relevance of fish skin biology to investigative dermatology. *Experimental dermatology*, 19(4), 313-324.
- Rakers, S., Niklasson, L., Steinhagen, D., Kruse, C., Schaubert, J., Sundell, K., and Paus, R. (2013). Antimicrobial peptides (AMPs) from fish epidermis: perspectives for investigative dermatology. *Journal of Investigative Dermatology*, 133(5), 1140-1149.
- Rao, V., Marimuthu, K., Kupusamy, T., Rathinam, X., Arasu, M.V., Al-Dhabi, N.A. and Arockiaraj, J., (2015). Defense properties in the epidermal mucus of different freshwater fish species. *Aquaculture, Aquarium, Conservation and Legislation-International Journal of the Bioflux Society (AAFL Bioflux)*, 8(2).

- Reed, L. J., and Muench, H. (1938). A simple method of estimating fifty per cent endpoints. *American Journal of Epidemiology*, 27(3), 493-497.
- Rittirsch, D., Flierl, M.A. and Ward, P.A., (2008). Harmful molecular mechanisms in sepsis. *Nature Reviews Immunology*, 8(10), 776-787.
- Robinette, D., Wada, S., Arroll, T., Levy, M. G., Miller, W. L., and Noga, E. J. (1998). Antimicrobial activity in the skin of the channel catfish *Ictalurus punctatus*: characterization of broad-spectrum histone-like antimicrobial proteins. *Cellular and Molecular Life Sciences*, 54(5), 467-475.
- Roberts, T. R. (1989). *The freshwater fishes of western Borneo (Kalimantan Barat, Indonesia)*.
- Rombout, J. H. W. M., Huttenhuis, H. B. T., Picchiatti, S., and Scapigliati, G. (2005). Phylogeny and ontogeny of fish leucocytes. *Fish and shellfish immunology*, 19(5), 441-455.
- Rosenfeld, Y., and Shai, Y. (2006). Lipopolysaccharide (Endotoxin)-host defense antibacterial peptides interactions: role in bacterial resistance and prevention of sepsis. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1758(9), 1513-1522.
- Rosenfeld, Y., Papo, N., and Shai, Y. (2006). Endotoxin (Lipopolysaccharide) Neutralization by Innate Immunity Host-Defense Peptides peptide properties and plausible modes of action. *Journal of Biological Chemistry*, 281(3), 1636-1643.
- Ross, N. W., Firth, K. J., Wang, A., Burka, J. F., and Johnson, S. C. (2000). Changes in hydrolytic enzyme activities of naive Atlantic salmon *Salmo salar* skin mucus due to infection with the salmon louse *Lepeophtheirus salmonis* and cortisol implantation. *Diseases of Aquatic Organisms*, 41(1), 43-51.
- Roth, M. (1971). Fluorescence reaction for amino acids. *Analytical Chemistry*, 43(7), 880-882
- Runti, G., Benincasa, M., Giuffrida, G., Devescovi, G., Venturi, V., Gennaro, R., and Scocchi, M. (2017). The Mechanism of Killing by the Proline-Rich Peptide Bac7 (1–35) against Clinical Strains of *Pseudomonas aeruginosa* Differs from That against Other Gram-Negative Bacteria. *Antimicrobial Agents and Chemotherapy*, 61(4), e01660-16.
- Sadikot, R. T., Blackwell, T. S., Christman, J. W., and Prince, A. S. (2005). Pathogen–host interactions in *Pseudomonas aeruginosa* pneumonia. *American journal of Respiratory and Critical Care Medicine*, 171(11), 1209-1223.

- Saito, T., Sawazaki, R., Ujiie, K., Oda, M., and Saitoh, H. (2012). Possible factors involved in oral inactivity of meropenem, a carbapenem antibiotic. *Pharmacology and Pharmacy*, 3(02), 201.
- Sakurai, A., Sakamoto, Y., Mori, F., and Loiselle, P. V. (1993). *Aquarium fish of the world: The comprehensive guide to 650 species*. Chronicle Books.
- Sahoo, T. K., Jena, P. K., Prajapati, B., Gehlot, L., Patel, A. K., and Seshadri, S. (2017). In Vivo Assessment of Immunogenicity and Toxicity of the Bacteriocin TSU4 in BALB/c Mice. *Probiotics and Antimicrobial Proteins*, 9(3), 345-354.
- Salger, S. A., Cassady, K. R., Reading, B. J., and Noga, E. J. (2016). A Diverse Family of Host-Defense Peptides (Piscidins) Exhibit Specialized Anti-Bacterial and Anti-Protozoal Activities in Fishes. *PloS one*, 11(8), e0159423.
- Salinas, I. (2015). The mucosal immune system of teleost fish. *Biology*, 4(3), 525-53
- Sanchooli, O., Hajimoradloo, A., and Ghorbani, R. (2012). Measurement of alkaline phosphatase and lysozyme enzymes in epidermal mucus of different weights of *Cyprinus carpio*. *World Journal of Fish and Marine Sciences* 4(5), 521-524.
- Smeianov, V., Scott, K., and Reid, G. (2000). Activity of cecropin P1 and FA-LL-37 against urogenital microflora. *Microbes and Infection*, 2(7), 773-777.
- Sarkar, S., Rai, B. K., Bhutia, D., Singh, S., and Pal, J. (2015). Study on the breeding performance and developmental stages of climbing perch, *Anabas testudineus* (Bloch, 1792) in the laboratory (Siliguri, India). *International Journal of Fisheries and Aquatic Studies*, 2(6), 198-201.
- Sarkar, U. K., and Ponniah, A. G. (2000). Evaluation of North East Indian fishes for their potential as cultivable, sport and ornamental fishes along with their conservation and endemic status. *Fish Biodiversity of Northeast India*, 2, 11-30.
- Sarkar, U. K., Deepak, P. K., Kapoor, D., Negi, R. S., Paul, S. K., and Singh, S. (2005). Captive breeding of climbing perch *Anabas testudineus* (Bloch, 1792) with Wova- FH for conservation and aquaculture. *Aquaculture Research*, 36(10), 941-945.
- Sarker, S.D., Nahar, L. and Kumarasamy, Y. (2007). Microtitre plate-based antibacterial assay incorporating resazurin as an indicator of cell growth, and its application in the in vitro antibacterial screening of phytochemicals. *Methods*, 42(4), 321-324.
- Sarmaşık, A., (2002). Antimicrobial peptides: a potential therapeutic alternative for the treatment of fish diseases. *Turkish Journal of Biology*, 26(4), 201-207.

- Saurabh, S., and Sahoo, P. K. (2008). Lysozyme: an important defence molecule of fish innate immune system. *Aquaculture Research*, 39(3), 223-239.
- Sayer, M. D. (2005). Adaptations of amphibious fish for surviving life out of water. *Fish and Fisheries*, 6(3), 186-211.
- Schulte, W., Bernhagen, J., and Bucala, R. (2013). Cytokines in sepsis: potent immunoregulators and potential therapeutic targets—an updated view. *Mediators of Inflammation*, 2013.
- Scicluna, B. P., van't Veer, C., Nieuwdorp, M., Felsmann, K., Wlotzka, B., Stroes, E. S., and van der Poll, T. (2013). Role of tumor necrosis factor- α in the human systemic endotoxin-induced transcriptome. *PloS one*, 8(11), e79051.
- Secombes, J. C and T. Wang (2012) The innate and adaptive immune system of fish. In: Austin, B. (Eds.), *Infectious disease in aquaculture: prevention and control* (pp3-6). Elsevier.
- Seemann, S., Zohles, F., and Lupp, A. (2017). Comprehensive comparison of three different animal models for systemic inflammation. *Journal of Biomedical Science*, 24(1), 60.
- Semple, F., and Dorin, J. R. (2012). β -Defensins: multifunctional modulators of infection, inflammation and more? *Journal of Innate Immunity*, 4(4), 337-348.
- Senati, M., Polacco, M., Grassi, V.M., Carbone, A. and De-Giorgio, F. (2013). Child abuse followed by fatal systemic *Pseudomonas aeruginosa* infection. *Legal Medicine*, 15(1), 28-31.
- Sengupta, D., Leontiadou, H., Mark, A. E., and Marrink, S. J. (2008). Toroidal pores formed by antimicrobial peptides show significant disorder. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1778(10), 2308-2317.
- Shabir, U., Ali, S., Magray, A. R., Ganai, B. A., Firdous, P., Hassan, T., and Nazir, R. (2017). Fish antimicrobial peptides (AMP's) as essential and promising molecular therapeutic agents: A review. *Microbial Pathogenesis*.1-30
- Shai, Y. (1999). Mechanism of the binding, insertion and destabilization of phospholipid bilayer membranes by α -helical antimicrobial and cell non-selective membrane-lytic peptides. *Biochimica et Biophysica Acta (BBA)-Biomembranes*, 1462(1), 55-70.
- Shai, Y. (2002). Mode of action of membrane active antimicrobial peptides. *Peptide Science*, 66(4), 236-248.
- Shapiro, E.D. (2008). Fever without localizing signs. In: Long S, Pickering, L.K, Prober, C.G, (Eds.), *Pediatric infectious diseases* (pp. 124–6). Churchill Livingstone.

- Shenoy, V. P., Ballal, M., Shivananda, P. G., and Bairy, I. (2012). Honey as an antimicrobial agent against *Pseudomonas aeruginosa* isolated from infected wounds. *Journal of Global Infectious Diseases*, 4(2), 102.
- Shephard, K. L. (1995). Functions for fish mucus. *Oceanographic Literature Review*, 10(42), 876.
- Shephard, K.L. (1994). Functions for fish mucus. *Reviews in fish biology and fisheries*, 4(4), 401-429.
- Shephard, K.L., 1993. Mucus on the epidermis of fish and its influence on drug delivery. *Advanced Drug Delivery Reviews*, 11(3), pp.403-417.
- Shih, M. F., Chen, L. Y., Tsai, P. J., and Cherng, J. Y. (2012). In vitro and in vivo therapeutics of β -thujaplicin on LPS-induced inflammation in macrophages and septic shock in mice. *International journal of immunopathology and pharmacology*, 25(1), 39-48.
- Shoriridge, V. D., Lazdunski, A., and Vasil, M. L. (1992). Osmoprotectants and phosphate regulate expression of phospholipase C in *Pseudomonas aeruginosa*. *Molecular microbiology*, 6(7), 863-871.
- Silva, T. and Gomes, M.S. (2017). Immuno-Stimulatory Peptides as a Potential Adjunct Therapy against Intra-Macrophagic Pathogens. *Molecules*, 22(8), 1297.
- Smith, R. J. F. (1992). Alarm signals in fishes. *Reviews in Fish Biology and Fisheries*, 2(1), 33-63.
- Smith, V. J., & Fernandes, J. M. (2009). Antimicrobial peptides of the innate immune system. *Fish Defenses*, 1, 241-275.
- Sobieszczyk, M. E., Furuya, E. Y., Hay, C. M., Pancholi, P., Della-Latta, P., Hammer, S. M., and Kubin, C. J. (2004). Combination therapy with polymyxin B for the treatment of multidrug-resistant Gram-negative respiratory tract infections. *Journal of Antimicrobial Chemotherapy*, 54(2), 566-569.
- Sokheng, C., Chhea, C.K., Viravong, S., Bouakhamvongsa, K., Suntornratana, U., Yoorong, N., Tung, N.T., Bao, T.Q., Poulsen, A.F. and Jorgensen, J.V. (1999). Fish migrations and spawning habits in the Mekong mainstream: a survey using local knowledge (basin-wide). *Assessment of Mekong fisheries: Fish Migrations and spawning and the Impact of water Management Project (AMFC)*. AMFP Report, 2, 99.
- Song, R., Wei, R. B., Luo, H. Y., and Wang, D. F. (2012). Isolation and characterization of an antibacterial peptide fraction from the pepsin hydrolysate of half-fin anchovy (*Setipinna taty*). *Molecules*, 17(3), 2980-2991.

- Spencer, R. C. (1996). Predominant pathogens found in the European prevalence of infection in intensive care study. *European Journal of Clinical Microbiology and Infectious Diseases*, 15(4), 281-285.
- Spitzer, R. H., and Koch, E. A. (1998). Hagfish skin and slime glands. In *The biology of hagfishes* (pp. 109-132). Springer, Dordrecht.
- Srinivasan, L., Harris, M. C., & Kilpatrick, L. E. (2017). Cytokines and Inflammatory Response in the Fetus and Neonate. In: Polin, A.R., Fox, W.W., Abman, H.S(Eds.), *Fetal and Neonatal Physiology* (5th, Vol 2, pp. 1241-1254).
- Srivastava, P. P., Chowdhury, R and Raizada, D (2013) Tissue Regeneration Potential of Climbing Perch, *Anabas testudines* (Bloch.). *Asian Journal of Experimental Biological Sciences*4 (3) 487-490
- Sterba, G. (1973). *Freshwater fishes of the world*. Tropical Fish Hobbyist Publications, Inc., Neptune City, NJ.
- Strateva, T., and Yordanov, D. (2009). *Pseudomonas aeruginosa*—a phenomenon of bacterial resistance. *Journal of Medical Microbiology*, 58(9), 1133-1148.
- Strous, G. J., and Dekker, J. (1992). Mucin-type glycoproteins. *Critical reviews in Biochemistry and Molecular Biology*, 27(1-2), 57-92.
- Stuart, K.R and M Pollack, M. (1982). *Pseudomonas aeruginosa* exotoxin A inhibits proliferation of human bone marrow progenitor cells in vitro. *Infect Immun*, 38(1): 206–211.
- Subhashini, S., Lavanya, J., Jain, S., and Agihotri, T. (2013). Screening of antibacterial and cytotoxic activity of extracts from epidermis and epidermal mucus of *Barbonymus schwanenfeldii* (Tinfoil barb fish). *International Journal of Engineering Research and Technology*, 2(4), 492-7.
- Subramanian, S., MacKinnon, S. L., and Ross, N. W. (2007). A comparative study on innate immune parameters in the epidermal mucus of various fish species. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 148(3), 256-263.
- Subramanian, S., Ross, N. W., and MacKinnon, S. L. (2008a). Comparison of antimicrobial activity in the epidermal mucus extracts of fish. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 150(1), 85-92.
- Subramanian, S., Ross, N. W., and MacKinnon, S. L. (2008b). Comparison of the biochemical composition of normal epidermal mucus and extruded slime of hagfish (*Myxine glutinosa* L.). *Fish and shellfish immunology*, 25(5), 625-632.

- Subramanian, S., Ross, N. W., and MacKinnon, S. L. (2009). Myxinidin, a novel antimicrobial peptide from the epidermal mucus of hagfish, *Myxine glutinosa* L. *Marine biotechnology*, 11(6), 748.
- Suh, S. J., Silo-Suh, L., Woods, D. E., Hassett, D. J., West, S. E., and Ohman, D. E. (1999). Effect of *rpoS* mutation on the stress response and expression of virulence factors in *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 181(13), 3890-3897.
- Sun, B. J., Xie, H. X., Song, Y., and Nie, P. (2007). Gene structure of an antimicrobial peptide from mandarin fish, *Siniperca chuatsi* (Basilewsky), suggests that moronecidins and pleurocidins belong in one family: the piscidins. *Journal of Fish Diseases*, 30(6), 335-343.
- Sun, Y., and Shang, D. (2015). Inhibitory effects of antimicrobial peptides on lipopolysaccharide-induced inflammation. *Mediators of inflammation*, 2015, 1-8
- Tagai, C., Morita, S., Shiraishi, T., Miyaji, K., and Iwamuro, S. (2011). Antimicrobial properties of arginine-and lysine-rich histones and involvement of bacterial outer membrane protease T in their differential mode of actions. *Peptides*, 32(10), 2003-2009.
- Takahashi, D., Shukla, S. K., Prakash, O., and Zhang, G. (2010). Structural determinants of host defense peptides for antimicrobial activity and target cell selectivity. *Biochimie*, 92(9), 1236-1241.
- Tamang, D. G., and Saier Jr, M. H. (2006). The cecropin superfamily of toxic peptides. *Journal of Molecular Microbiology and Biotechnology*, 11(1-2), 94-10
- Tamber, S., Ochs, M. M., and Hancock, R. E. (2006). Role of the novel OprD family of porins in nutrient uptake in *Pseudomonas aeruginosa*. *Journal of Bacteriology*, 188(1), 45-54.
- Tan, L.Z.W., Hong, Z., Yam, J.K.H., Salido, M.M.S., Woo, B.Y., Li, S.F.Y., Yang, L., Givskov, M. and Chng, S.S. (2017). Auranofin inhibits virulence in *Pseudomonas aeruginosa*. *bioRxiv*, 198820.
- Tessera, V., Guida, F., Juretić, D., and Tossi, A. (2012). Identification of antimicrobial peptides from teleosts and anurans in expressed sequence tag databases using conserved signal sequences. *The FEBS journal*, 279(5), 724-736.
- Thompson, S. A., Tachibana, K., Nakanishi, K., and Kubota, I. (1986). Melittin-like peptides from the shark-repelling defense secretion of the sole *Pardachirus pavoninus*. *Science*, 233, 341-344.

- Toufekoula, C., Papadakis, V., Tsaganos, T., Routsis, C., Orfanos, S.E., Kotanidou, A., Carrer, D.P., Raftogiannis, M., Baziaka, F. and Giamarellos-Bourboulis, E.J. (2013). Compartmentalization of lipid peroxidation in sepsis by multidrug-resistant gram-negative bacteria: experimental and clinical evidence. *Critical Care*, 17(1), R6.
- Trautmann, M., Heinemann, M., Zick, R., Möricke, A., Seidelmann, M., and Berger, D. (1998). Antibacterial activity of meropenem against *Pseudomonas aeruginosa*, including antibiotic-induced morphological changes and endotoxin-liberating effects. *European Journal of Clinical Microbiology and Infectious Diseases*, 17(11), 754-760.
- Traeger, T., Mikulcak, M., Eipel, C., Abshagen, K., Diedrich, S., Heidecke, C.D., Maier, S. and Vollmar, B., (2010). Kupffer cell depletion reduces hepatic inflammation and apoptosis but decreases survival in abdominal sepsis. *European Journal of Gastroenterology and Hepatology*, 22(9), 1039-1049.
- Ulevitch, R. J., and Tobias, P. S. (1999). Recognition of gram-negative bacteria and endotoxin by the innate immune system. *Current Opinion in Immunology*, 11(1), 19-22.
- Ullal, A. J., and Noga, E. J. (2010). Antiparasitic activity of the antimicrobial peptide Hb β P- 1, a member of the β - haemoglobin peptide family. *Journal of Fish Diseases*, 33(8), 657-664.
- Ullal, A. J., Litaker, R. W., and Noga, E. J. (2008). Antimicrobial peptides derived from hemoglobin are expressed in epithelium of channel catfish (*Ictalurus punctatus*, Rafinesque). *Developmental and Comparative Immunology*, 32(11), 1301-1312.
- Uribe, C., Folch, H., Enriquez, R., and Moran, G. (2011). Innate and adaptive immunity in teleost fish: a review. *Veterinarni Medicina*, 56(10), 486-503.
- Uthayakumar, V., Ramasubramanian, V., Senthilkumar, D., Priyadarisini, V. B., and Harikrishnan, R. (2012). Biochemical characterization, antimicrobial and hemolytic studies on skin mucus of fresh water spiny eel *Mastacembelus armatus*. *Asian Pacific Journal of Tropical Biomedicine*, 2(2), S863-S869.
- Uzzell, T., Stolzenberg, E. D., Shinnar, A. E., and Zasloff, M. (2003). Hagfish intestinal antimicrobial peptides are ancient cathelicidins. *Peptides*, 24(11), 1655-1667.
- Valero, Y., Chaves-Pozo, E., Meseguer, J., Esteban, M. A., and Cuesta, A. (2013). *Appleton. Biological role of fish antimicrobial peptides*. In: Seong and Hak, *Antimicrobial peptides* (pp 31-60) Nova Science Publishers, Inc.

- Van Delden, C. (2007). *Pseudomonas aeruginosa* bloodstream infections: how should we treat them? *International Journal of Antimicrobial Agents*, 30, 71-75
- van der Marel, M., Caspari, N., Neuhaus, H., Meyer, W., Enss, M. L., and Steinhagen, D. (2010). Changes in skin mucus of common carp, *Cyprinus carpio* L., after exposure to water with a high bacterial load. *Journal of Fish Diseases*, 33(5), 431-439.
- Vatsos, I. N., Kotzamanis, Y., Henry, M., Angelidis, P., and Alexis, M. N. (2010). Monitoring stress in fish by applying image analysis to their skin mucous cells. *European Journal of Histochemistry: EJH*, 54(2).
- Vaure, C., and Liu, Y. (2014). A comparative review of toll-like receptor 4 expression and functionality in different animal species. *Frontiers in immunology*, 5, 316.
- Veeruraj, A., Arumugam, M., Ajithkumar, T., and Balasubramanian, T. (2008). Isolation and biological properties of neurotoxin from sea anemone (*Stichodactyla mertensii*, *S. haddoni*). *Internet J Toxicol*, 5(2).
- Veesenmeyer, J. L., Hauser, A. R., Lisboa, T., and Rello, J. (2009). *Pseudomonas aeruginosa* virulence and therapy: evolving translational strategies. *Critical care medicine*, 37(5), 1777.
- Verdugo, P. (1990). Goblet cells secretion and mucogenesis. *Annual review of physiology*, 52(1), 157-176.
- Vieira, F.A., Gregório, S.F., Ferrareso, S., Thorne, M.A., Costa, R., Milan, M., Bargelloni, L., Clark, M.S., Canario, A.V. and Power, D.M. (2011). Skin healing and scale regeneration in fed and unfed sea bream, *Sparus auratus*. *BMC genomics*, 12(1), 490.
- Villarroel, F., Bastías, A., Casado, A., Amthauer, R., and Concha, M. I. (2007). Apolipoprotein AI, an antimicrobial protein in *Oncorhynchus mykiss*: evaluation of its expression in primary defence barriers and plasma levels in sick and healthy fish. *Fish and shellfish immunology*, 23(1), 197-209.
- Walters, M. C., Roe, F., Bugnicourt, A., Franklin, M. J., and Stewart, P. S. (2003). Contributions of antibiotic penetration, oxygen limitation, and low metabolic activity to tolerance of *Pseudomonas aeruginosa* biofilms to ciprofloxacin and tobramycin. *Antimicrobial Agents and Chemotherapy*, 47(1), 317-323.
- Wang, D., Yin, Y., and Yao, Y. (2014). Advances in sepsis-associated liver dysfunction. *Burns and Trauma*, 2(3), 97.
- Wang, G. (Ed.). (2017). *Antimicrobial peptides: discovery, design and novel therapeutic strategies*. Cabi.

- Wang, G., Li, X., and Zasloff, M. (2010). A database view of naturally occurring antimicrobial peptides: nomenclature, classification and amino acid sequence analysis. *Antimicrobial peptides: discovery, design and novel therapeutic strategies*, 1-21.
- Wang, S., Zeng, X., Yang, Q., and Qiao, S. (2016). Antimicrobial peptides as potential alternatives to antibiotics in food animal industry. *International Journal of Molecular Sciences*, 17(5), 603.
- Wang, Y., Liu, X., Ma, L., Yu, Y., Yu, H., Mohammed, S., Chu, G., Mu, L. and Zhang, Q. b (2012). Identification and characterization of a hepcidin from half-smooth tongue sole *Cynoglossus semilaevis*. *Fish and Shellfish Immunology*, 33(2), 213-219.
- Waters, V., and Smyth, A. (2015). Cystic fibrosis microbiology: advances in antimicrobial therapy. *Journal of Cystic Fibrosis*, 14(5), 551-560.
- Wei, O. Y., Xavier, R., and Marimuthu, K. (2010). Screening of antibacterial activity of mucus extract of snakehead fish, *Channa striatus* (Bloch). *European Review for Medical and Pharmacological Sciences*, 14(8), 675-681.
- Whitear, M. (1981). Secretion in the epidermis of polypteriform fish. *Zeitschrift fur Mikroskopisch-anatomische Forschung*, 95(4), 531-543.
- Whitear, M. (1986). The skin of fishes including cyclostomes: epidermis. In: Bereiter-Hahn, J., Matoltsy, A.G and Richards, K.S (Eds.), *Biology of the integument*, 2 Vertebrates (Vol. 2, pp. 9-64). Verlag Berlin Heidelberg: Springer.
- Williamson, K. S., Richards, L. A., Perez-Osorio, A. C., Pitts, B., McInnerney, K., Stewart, P. S., and Franklin, M. J. (2012). Heterogeneity in *Pseudomonas aeruginosa* biofilms includes expression of ribosome hibernation factors in the antibiotic-tolerant subpopulation and hypoxia-induced stress response in the metabolically active population. *Journal of Bacteriology*, 194(8), 2062-2073.
- Wisplinghoff, H., Bischoff, T., Tallent, S. M., Seifert, H., Wenzel, R. P., and Edmond, M. B. (2004). Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from a prospective nationwide surveillance study. *Clinical Infectious Diseases*, 39(3), 309-317.
- Wu, D., Zhou, S., Hu, S., and Liu, B. (2017). Inflammatory responses and histopathological changes in a mouse model of *Staphylococcus aureus*-induced bloodstream infections. *The Journal of Infection in Developing Countries*, 11(04), 294-305.

- Xu, Z., Parra, D., Gómez, D., Salinas, I., Zhang, Y.A., von Gersdorff Jørgensen, L., Heinecke, R.D., Buchmann, K., LaPatra, S. and Sunyer, J.O. (2013). Teleost skin, an ancient mucosal surface that elicits gut-like immune responses. *Proceedings of the National Academy of Sciences*, 110(32), 13097-13102.
- Yagel, S. K., Barrett, J. F., Amaratunga, D. J., and Frosco, M. B. (1996). In vivo oral efficacy of levofloxacin for treatment of systemic *Pseudomonas aeruginosa* infections in a murine model of septicemia. *Antimicrobial Agents and Chemotherapy*, 40(12), 2894-2897.
- Yan, J., Li, S. and Li, S., (2014). The role of the liver in sepsis. *International Reviews of Immunology*, 33(6), 498-510.
- Yang, L., Harroun, T. A., Weiss, T. M., Ding, L., and Huang, H. W. (2001). Barrel-stave model or toroidal model? A case study on melittin pores. *Biophysical Journal*, 81(3), 1475-1485.
- Yano, T. (1996). The nonspecific immune system In: William, S., Hoar, S.W., David J. Randall, J.R., Iwama, G and Nakanishi, N (Eds.), *Fish Physiology series*(Vol 15, pp. 105-157). Academic Press.
- Yenugu, S., Hamil, K. G., Birse, C. E., Ruben, S. M., & French, F. S. (2003). Antibacterial properties of the sperm-binding proteins and peptides of human epididymis 2 (HE2) family; salt sensitivity, structural dependence and their interaction with outer and cytoplasmic membranes of *Escherichia coli*. *Biochemical Journal*, 372(2), 473-483.
- Yayan, J., Ghebremedhin, B., and Rasche, K. (2015). Antibiotic resistance of *pseudomonas aeruginosa* in pneumonia at a single university hospital center in germany over a 10-year period. *Plos one*, 10(10), e0139836.
- Yeaman, M. R., and Yount, N. Y. (2003). Mechanisms of antimicrobial peptide action and resistance. *Pharmacological Reviews*, 55(1), 27-55.
- Yi, Y., You, X., Bian, C., Chen, S., Lv, Z., Qiu, L., and Shi, Q. (2017). High-Throughput Identification of Antimicrobial Peptides from Amphibious Mudskippers. *Marine drugs*, 15(11), 364.
- Zhang, L. J., and Gallo, R. L. (2016). Antimicrobial peptides. *Current Biology*, 26(1), R14-R19.
- Zhao, J.G., Zhou, L., Jin, J.Y., Zhao, Z., Lan, J., Zhang, Y.B., Zhang, Q.Y. and Gui, J.F. (2009). Antimicrobial activity-specific to Gram-negative bacteria and immune modulation-mediated NF- κ B and Sp1 of a medaka β -defensin. *Developmental and Comparative Immunology*, 33(4),624-637.

Zhao, X., Findly, R. C., and Dickerson, H. W. (2008). Cutaneous antibody-secreting cells and B cells in a teleost fish. *Developmental and Comparative Immunology*, 32(5), 500-508.

Zhao, H., Yang, H., Sun, J., Chen, Y., Liu, L., Li, G., and Liu, L. (2016). The complete mitochondrial genome of the *Anabas testudineus* (Perciformes, Anabantidae). *Mitochondrial DNA Part A*, 27(2), 1005-1007.

Zilberstein, D., Schuldiner, S., and Padan, E. (1979). Proton electrochemical gradient in *Escherichia coli* cells and its relation to active transport of lactose. *Biochemistry*, 18(4), 669-673.

Zou, J., Mercier, C., Koussounadis, A., and Secombes, C. (2007). Discovery of multiple beta-defensin like homologues in teleost fish. *Molecular Immunology*, 44(4), 638-647.