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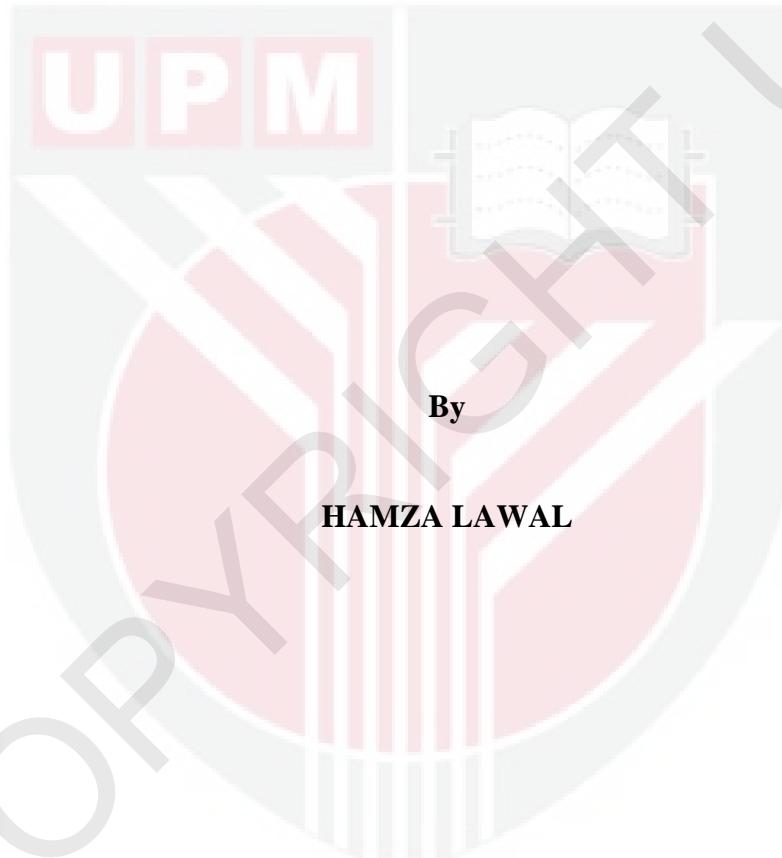
***IMMUNOMODULATORY ACTIVITY OF *Moringa oleifera L.* LEAF
ETHANOL EXTRACT ON NORMAL LYMPHOCYTES AND
LEUKAEMIC CELL LINES***

HAMZA LAWAL

FPSK(M) 2018 31



**IMMUNOMODULATORY ACTIVITY OF *Moringa oleifera* L. LEAF
ETHANOL EXTRACT ON NORMAL LYMPHOCYTES AND
LEUKAEMIC CELL LINES**



**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillments of the Requirement for the Degree of
Master of Science**

April 2018

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DEDICATIONS

This is for you, Mama.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirements for the degree of Master of Science

**IMMUNOMODULATORY ACTIVITY OF *Moringa oleifera* L. LEAF
ETHANOL EXTRACT ON NORMAL LYMPHOCYTES AND
LEUKAEMIC CELL LINES**

By

HAMZA LAWAL

April 2018

Chairman : Associate Professor Rajesh Ramasamy, PhD
Faculty : Medicine and Health Sciences

Moringa oleifera (*M. oleifera*), a member of the family Moringaceae, is a small-medium sized tree, 10-15m high, widely cultivated in East and Southeast Asia, West Indies and Polynesia. Indians have been using leaves, fruits and flowers of *M. oleifera* as part of their routine diet as this 'wonder' herb also was used in ancient Ayurveda and Siddha medicine for nearly 2000 years. Phytochemical and animal studies have shown that the therapeutic activities of *M. oleifera* have largely depended on its main polyphenols such as quercetin glucosides, kaempferol glycosides, rutin, and chlorogenic acid. To date, *M. oleifera* has been studied for their anti-oxidative, antidiabetic, anti-inflammatory and anticancer activity, yet their role in modulating the immune system is still elusive. The present study, therefore, aimed to investigate the in vitro immunomodulatory effect of *M. oleifera* leaves' ethanol extracts (MOETE) on healthy peripheral blood mononuclear cells (PBMCs) and leukaemic cell lines. Fresh and healthy leaves of *M. oleifera* were collected from an herbal farm located at Kampar, Selangor. Extracts of *M. oleifera* leaves were obtained using the mixture of ethanol and water at ratios of 100:0 (100% ethanol), 70:30 (70% ethanol), 50:50 (50% ethanol) and 0:100 (aqueous) as extraction solvents. Healthy donors were used as a source of primary lymphocytes while Jurkat and BV173 cells were utilised as transformed cell lines of T cells and B cells, respectively. The cytotoxicity of aqueous and ethanolic extracts of *M. oleifera* leaves on the cells was determined using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay while the marker compounds (quercetin, and kaempferol 3-O-glucoside) in *M. oleifera* extract were identified and quantified using High Performance Liquid Chromatography (HPLC). The immunomodulatory effect was evaluated through cell proliferation assays, cell cycle analysis and apoptosis assays. The antitumour potential of the extract on Jurkat and BV173 cells was further explored via global secretome and apoptotic proteins proteome arrays.

From the cytotoxicity analysis, 70% ethanol *M. oleifera* leaves extract exerted a dose-dependent stimulatory effect on PBMCs with an EC₅₀ of 28±3 g/mL as well as cytotoxic effects on BV173 (IC₅₀ = 125±6 g/mL) and Jurkat cells (IC₅₀ = 262±3 g/mL). Also, from the HPLC analysis, kaempferol 3-O-glucoside (standard Rt = 32.689 vs sample mean Rt = 32.671), and quercetin (standard Rt = 42.020 vs sample mean Rt = 41.981) were identified. The extract enhanced the viability and proliferation of PBMCs by committing the cells into the cell cycle and reducing apoptosis while exerting anti-proliferative effects, cell cycle arrest and apoptosis in tumour cell lines. Also, the extract induced overexpression of pro-apoptotic cytokines and proteins but suppressed the expression of angiogenic factors and pro-survival proteins in tumour cell lines. Pathway enrichment analysis revealed that MOETE induced apoptosis in BV173 and Jurkat cells mainly through activation of the mitochondrial apoptotic pathway by upregulation of mitochondrial pro-apoptotic B cell lymphoma 2 (BCL2) family of proteins like the BCL-2-associated X protein (BAX) and the BCL-2 homologous antagonist killer (BAK) while downregulation of the anti-apoptotic protein, BCL-2. *M. oleifera* ethanol extract has immunostimulatory properties on normal lymphocytes and antitumour activity on leukemic cell lines. These abilities can be exploited in developing herbal supplements that strengthen the immune system to support aged and immunocompromised individuals as well as serve as adjuvants in therapies against blood cancers.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk ijazah Master Sains

**AKTIVITI IMMUNOMODULATORY DAUN KACANG KELOR YANG
DIETANOL-EKSTRAK TERHADAP LIMFOSIT NORMAL DAN
SEL-SEL KANSER LEUKEMIA**

Oleh

HAMZA LAWAL

April 2018

Pengerusi : Profesor Madya Rajesh Ramasamy, PhD
Fakulti : Perubatan dan Sains Kesihatan

Moringa oleifera (*M. oleifera*), dari keluarga Moringaceae, adalah pokok bersaiz sederhana kecil, berketinggian 10-15m yang ditanam secara meluas di Timur dan Asia Tenggara, Hindia Barat serta Polynesia. Di Malaysia, kaum India telah menggunakan daun, buah-buahan dan bunga *M. oleifera* sebagai sebahagian daripada makanan rutin memandangkan ianya juga digunakan dalam perubatan Ayurveda dan Siddha selama hampir 2000 tahun. Kajian fitokimia dan haiwan telah menunjukkan bahawa aktiviti terapi *M. oleifera* sebahagian besarnya bergantung kepada polifenol utama seperti quercetin glukosida, kaempferol glukosida, rutin, dan asid klorogenik. Sehingga kini, *M. oleifera* telah dikaji kerana aktiviti anti-oksidatif, anti-diabetik, anti-radang dan anti-kansernya. Walaubagaimanapun, peranannya dalam modulasi sistem imun masih sukar untuk difahami. Oleh itu, kajian ini dijalankan bertujuan untuk mengkaji kesan imunomodulator ekstrak etanol daun *M. oleifera* (MOETE) pada sel mononuklear darah periferal (PBMCs) dan sel leukaemik secara *in vitro*. Daun *M. oleifera* dikumpulkan dari ladang herba yang terletak di Kampar, Selangor. Ekstrak daun *M. oleifera* diperoleh menggunakan campuran etanol dan air pada nisbah 100: 0 (100% etanol), 70:30 (70% etanol), 50:50 (50% etanol) dan 0:100 (air) sebagai pelarut ekstraksi. Penderma sihat digunakan sebagai sumber sel darah putih utama manakala sel Jurkat dan BV173 digunakan sebagai sel-sel transformasi T dan B. Sitotoksisiti ekstrak air dan etanol daun *M. oleifera* pada sel telah ditentukan dengan menggunakan ujian 3- (4,5-dimetilthiazol-2-yl) -2,5-diphenyltetrazolium bromida (MTT) manakala sebatian penanda (quercetin, dan kaempferol 3-O-glukosida) dalam ekstrak *M. oleifera* telah dikenalpasti dan dikira menggunakan Kromatografi Liquid Performance High (HPLC). Kesan imunomodulator telah dinilai melalui ujian proliferasi sel, analisis kitaran sel dan pemeriksaan apoptosis. Potensi anti-tumor ekstrak pada sel-sel Jurkat dan BV173 terus diterokai menerusi ‘global secretome’ dan apoptosis protein ‘proteome arrays’.

Dari analisis sitotoksiti, ekstrak 70% etanol daun *M. oleifera* memberikan kesan stimulasi yang kebergantungan-dos pada PBMC dengan EC₅₀ kadar 28 ± 3 g/mL serta kesan sitotoksik pada BV173 (IC₅₀ = 125 ± 6 g / mL) dan sel Jurkat (IC₅₀ = 262 ± 3 g/mL). Selain itu, dari analisa HPLC, kaempferol 3-O-glukosida (standard Rt = 32.689 berbanding purata sampel Rt = 32.671), dan quercetin (standard Rt = 42.020 berbanding purata sampel Rt = 41.981) telah dikenalpasti. Ekstrak itu meningkatkan viabiliti dan proliferasi PBMC melalui modulasi kitaran hidup sel dan mengurangkan kadar apoptosis, sambil memberi kesan-kesan anti-proliferatif, perhentian kitaran sel dan apoptosis dalam sel-sel tumor. Juga, ekstrak dapat menjana tinggi expressi sitokin dan protein pro-apoptosis sementara dapat menyekat expresi faktor-faktor angiogenik dan ‘pro-survival’ protein dalam sel-sel sel tumor. Analisa ‘pathway enrichment’ mendedahkan bahawa MOETE dapat menyebabkan apoptosis dalam sel BV173 dan sel Jurkat terutamanya melalui pengaktifan laluan apoptosis mitokondria dengan menaik-expresi protein pro-apoptosis B lymphoma 2 (BCL2) mitokondria seperti protein X BCL-2 yang berkaitan (BAX), dan pe-nyah antagonis homologous BCL-2 (BAK) sementara mengurangkan protein anti-apoptosis, BCL-2. Ekstrak etanol *M. oleifera* mempunyai sifat immunostimulatori pada limfosit normal namun aktiviti anti-tumor pada sel-sel leukemia. Keupayaan ini dapat dieksloitasi dalam mengembangkan industry suplemen herba bagi menguatkan sistem kekebalan tubuh bagi individu yang berumur dan juga imunokompromi serta berperanan sebagai pembantu dalam terapi terhadap kanser darah.

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I certify that a Thesis Examination Committee has met on 17 April 2018 to conduct the final examination of Hamza Lawal on his thesis entitled "Immunomodulatory Activity of *Moringa oleifera* L. Leaf Ethanol Extract on Normal Lymphocytes and Leukaemic Cell Lines" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

Ang	Angiogenin
Apaf-1	Apoptotic protease activating factor 1
APCs	Antigen-presenting cells
ATCC	American type culture collection
BAD	Bcl-2-associated death promoter
BAX	Bcl-2-associated X protein
Bcl-2	B cell lymphoma 2
BCLx	B-cell lymphoma X
BCR	B- cell receptor
BID	Bax-like BH3 protein
c-FLIP	Cellular (FADD-like IL-1 β -converting enzyme [FLICE])-inhibitory protein
CAD	Caspase-activated Dnases
Casp	Caspase
CD	Cluster of differentiation
CDK	Cyclin-dependent kinase
CHI3L	Chitinase-3-like protein
CMI	Cell-mediated immunity
Con A	Concanavalin A
Cripto	Cryptic family protein
Cyt-c	Cytochrome c
DECs	Differentially expressed cytokines
DEPs	Differentially expressed proteins
DIABLO	Direct IAP-Binding protein with Low PI
DISC	Death inducing signaling complex
DMSO	Dimethyl sulfoxide
DNA	Deoxyribonucleic acid
DR	Death receptor
EC50	Half maximal effective concentration
FAS	First apoptotic signal
FACS	Fluorescence-activated cell sorting
FADD	Fas associated death domain
FGF	Fibroblast growth factors
FITC	Fluorescein isothiocyanate
G	Gap
GM-CSF	Granulocyte–macrophage colony-stimulating factor
GO	Gene ontology

HAVCR	Hepatitis A virus cellular receptor
HO	Heme oxygenase
HMRC	Herbal Medicine Research Centre
HPLC-DAD	High-Performance Liquid Chromatography with Diode-Array Detection
HRP	Horse radish peroxidase
HSP	Heat shock protein
IAP	Inhibitor of apoptosis protein
IC ₅₀	Half maximal inhibitory concentration
IFN	Interferon
IGFBP	Insulin-like growth factor-binding protein
IL	Interleukin
IMR	Institute for Medical Research
IUPAC	International Union of Pure and Applied Chemistry
KEGG	Kyoto encyclopaedia of genes and genomes
JAK	Janus kinase
LC-MS	Liquid chromatography mass spectrometry
LPS	Lipopolysaccharide
LT	Lymphotoxin
M	Mitosis
MHC	Major histocompatibility complex
MOETE	<i>M. oleifera</i> leaves ethanol extract
MTT	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
NCBI	National Center for Biotechnology Information
NF-κB	Nuclear factor – kappa beta
NK	Natural killer
PBMCs	Peripheral blood mononuclear cells
PBS	Phosphate-buffered Saline
PHA	Phytohemagglutinin
PI	Propidium iodide
PI3K	Phosphoinositide 3-kinase
PS	Phosphatidylserine
PWM	Pokeweed mitogen
Rb	Retinoblastoma
RNA	Ribonucleic acid
RNase	Ribonuclease
ROS	Reactive oxygen species
RPMI	Roswell park memorial institute
S	Synthesis
SD	Standard deviation
SMAC	Second Mitochondria-derived Activator of Caspases

STAT	Signal transducer and activator of transcription
TAAs	Tumour associated antigens
TCR	T-cell receptor
TGF	Transforming growth factor
TGF	Transforming growth factor
Th	T-helper
TNF	Tumour necrosis factor
TNFSF	TNF superfamily
TRAIL R	TNF-related apoptosis-inducing ligand receptor
Tregs	Regulatory T lymphocytes
uPAR	Urokinase receptor
UV	Ultraviolet rays
VEGF	Vascular endothelial growth factor
XIAP	X-linked inhibitor of apoptosis protein

CHAPTER 1

INTRODUCTION

Cell-mediated and humoral immune responses are compromised in immunodeficiency disorders and as a result, the ability of the immune system to defend the body from abnormal or invading foreign cells and disease-causing substances is impaired (Halder *et al.*, 2012). This immunocompromised condition predisposes the body not only to the nosocomial infections but also to the opportunistic infections and in addition, increase the susceptibility of developing benign and malignant tumours as a result of decreased cancer immunosurveillance (Dadi *et al.*, 2016). The modulation of immune response in the form of boosting the ability of the body to produce immune cells for the amelioration of disease and cancer susceptibility as well as positive response to conventional cancer radio- and chemotherapy can be achieved by the dietary intake and supplementation of plants materials with established immunomodulatory properties (Gupta *et al.*, 2010; Yin *et al.*, 2017). This is often referred to as “Phytoimmunotherapy” and it’s being considered as a new approach for the treatment of cancers (Efferth *et al.*, 2017; Yin *et al.*, 2017). Moreover, many plant products have been exploited for modulation of the immune system in a number of Ayurvedic formulation either alone or in groups (Anwar *et al.*, 2007).

M. oleifera, a tree plant mostly used as part of regular diet in most parts of the world especially in Southeast Asia, Polynesia, India and Africa, has been in ancient scripts, identified as having immune enhancing properties (Anwar *et al.*, 2007). Different parts of the plant (leaves, roots, fruits, flowers, resin and bark) have been reported to possess wide range of pharmacological and therapeutic properties including antitumor, antipyretic, antiepileptic, antispasmodic, anti-inflammatory, diuretic, antiulcer, hypotensive, hypolipidemic, hypoglycemic, hepatoprotective, antifungal and antibacterial activities (Bharali *et al.*, 2003; Leelavinothan *et al.*, 2007; Saini *et al.*, 2016). The polyphenolic constituents of *M. oleifera* leaves, stem, roots, and flowers have been investigated using comprehensive analytical techniques ranging from Liquid Chromatography Mass Spectrometry LC-MS (Bennett *et al.*, 2003), to High Performance Liquid Chromatography HPLC (Atawodi *et al.*, 2010; Vongsak *et al.*, 2013). These investigations have revealed several phytochemicals present in the *M. oleifera* plant. The leaves, however, have mainly flavonoids quercetin and kaempferol (mostly in their glycated form, i.e. isoquercitin and astragalin), chlorogenic acid as well as other phytochemicals like benzylamine (moringine), niazirin and niazirinin (Shanker *et al.*, 2007; Velaga *et al.*, 2017).

Previous studies have shown that pharmacological agents and naturally occurring food chemicals such as the flavonoids and other polyphenols can modify immune system as they have been shown to affect the function of T cells, B cells, NK cells, macrophages, mast cells, basophils, neutrophils, eosinophils and platelets (John *et al.*, 2011; Somerville *et al.*, 2016). Additionally, the leaves of *M. oleifera* being a

rich source of flavonoids, chlorogenic acid and many vital phytochemicals (Mishra *et al.*, 2017), are expected to exhibit immunomodulatory properties capable of enhancing the proliferation of normal lymphocytes and exerting anti-tumour activity. Although the anti-tumour activity of extracts of *M. oleifera* leaves have been explored by analysing its ability to induce apoptosis in proliferating cancer cells (Sreelatha *et al.*, 2011; Tiloke *et al.*, 2016; Tragulpakseerojn *et al.*, 2017), very few studies have investigated the immunomodulatory effect of *M. oleifera* leaves. With the exception of few animal studies, reporting increase in white blood cell or splenocyte count following treatment with extracts of *M. oleifera* leaves, (Gupta *et al.*, 2010; Mousa *et al.*, 2017; Nfambi *et al.*, 2015), close examination of the interactions between the extracts of *M. oleifera* leaves and immune cells, especially lymphocytes within controlled in vitro cultures remained unexplored. To address this research gap, the present study evaluated the effect of 70% ethanolic extract of *M. oleifera* which is mainly comprised of polyphenols on normal and malignant lymphocytes. The cytotoxicity of *M. oleifera* extract on lymphocyte-derived tumour cell lines, and healthy peripheral blood mononuclear cells were tested along with the cell cycle progression, cell proliferation, and apoptosis assays. Both negative and positive impacts of *M. oleifera* on these cells were further deciphered through the profiling of cytokines and apoptotic proteins using customised proteome-antibody arrays. Based on the inputs from detailed arrays of proteins, a panel of signalling pathways were identified which could serve as the possible mechanisms that exploit by *M. oleifera* to deliver its pharmacological effects.

1.1 Problem statement

Infectious diseases, cardiometabolic disorders as well as tumours and cancers are the most prevalent life-threatening conditions in the world today. These conditions target, destroy and/or compromise the overall immune system, leaving the body susceptible to the opportunistic infections among many others. The key to the prevention and treatment of these disease conditions is the strengthening of the immune system using most importantly, readily available, native plant materials with immune boosting properties.

1.2 Study justification

M. oleifera, a widely known herb in Malaysia and throughout South-East Asia, could be ideal for providing a panacea to the identified problem. However, with very few studies have been conducted to explore the immunomodulatory potentials of *M. oleifera* leaves, there is need to carry out an analytical assessment on its ability to exert immunomodulatory activities on normal and tumour-derived lymphocytes. The findings from this study would provide the scientific and laboratory-based evidence for the development and use of *M. oleifera* leaves' supplements in improving the immune system amongst aged and immunocompromised individuals as well as support the current trend in the immunotherapeutic approaches towards the fight against cancer and tumours.

1.3 Null hypothesis

M. oleifera leaves' ethanol extract (MOETE) has no immunomodulatory effect on normal PBMCs and leukaemic cell lines.

1.4 Objectives

1.4.1 General objective

To investigate the immunomodulatory activity of *M. oleifera* leaves' ethanol extract (MOETE) on normal and malignant lymphocytes.

1.4.2 Specific objectives

- i. To extract, analyse and identify the phytochemical constituents of MOETE.
- ii. To assess the impact of MOETE on cell viability, cell proliferation, cell cycle progression and apoptosis of normal lymphocytes (PBMCs) and leukaemic cell lines (BV173 and Jurkat cells).
- iii. To determine the effect of MOETE on cytokine and chemokine secretome of the leukaemic cell lines (BV173 and Jurkat cells).
- iv. To evaluate the effect of MOETE on the expression of apoptosis mediators in the leukaemic cell lines (BV173 and Jurkat cells).

REFERENCES

- Aalinkeel, R., Bindukumar, B., Reynolds, J. L., Sykes, D. E., Mahajan, S. D., Chadha, K. C., & Schwartz, S. A. (2008). The dietary bioflavonoid, quercetin, selectively induces apoptosis of prostate cancer cells by down-regulating the expression of heat shock protein 90. *The Prostate*, 68(16), 1773-1789.
- Abalaka, M. E., Daniyan, S. Y., Oyeleke, S. B., & Adeyemo, S. O. (2012). The antibacterial evaluation of *M. oleifera* leaf extracts on selected bacterial pathogens. *Journal of Microbiology Research*, 2(2), 1-4.
- Abukhdeir, A. M., & Park, B. H. (2008). P21 and p27: roles in carcinogenesis and drug resistance. *Expert reviews in molecular medicine*, 10.
- Adedapo, A., Mogbojuri, O., & Emikpe, B. (2009). Safety evaluations of the aqueous extract of the leaves of *M. oleifera* in rats. *Journal of Medicinal Plants Research*, 3(8), 586-591.
- Aguirre, A., Shoji, K. F., Sáez, J. C., Henríquez, M., & Quest, A. F. (2013). FasL-triggered death of Jurkat cells requires caspase 8-induced, ATP-dependent cross-talk between fas and the purinergic receptor P2X7. *Journal of cellular physiology*, 228(2), 485-493.
- Akbay, P., Basaran, A. A., Undege, U., & Basaran, N. (2003). In vitro immunomodulatory activity of flavonoid glycosides from *Urtica dioica* L. *Phytotherapy research*, 17(1), 34-37.
- Al-Asmari, A. K., Albalawi, S. M., Athar, M. T., Khan, A. Q., Al-Shahrani, H., & Islam, M. (2015). *M. oleifera* as an anti-cancer agent against breast and colorectal cancer cell lines. *PloS one*, 10(8), e0135814.
- Alamgir, A. (2017). Pharmacognostical Botany: Classification of Medicinal and Aromatic Plants (MAPs), Botanical Taxonomy, Morphology, and Anatomy of Drug Plants. In *Therapeutic Use of Medicinal Plants and Their Extracts: Volume 1* (pp. 177-293): Springer.
- Amaglo, N. K., Bennett, R. N., Curto, R. B. L., Rosa, E. A., Turco, V. L., Giuffrida, A., ... & Timpo, G. M. (2010). Profiling selected phytochemicals and nutrients in different tissues of the multipurpose tree *M. oleifera* L., grown in Ghana. *Food Chemistry*, 122(4), 1047-1054.
- Andrews, L., Clary, J. J., Terrill, J., & Bolte, H. F. (1987). Subchronic inhalation toxicity of methanol. *Journal of Toxicology and Environmental Health, Part A Current Issues*, 20(1-2), 117-124.

- Antonopoulos, C., El Sanadi, C., Kaiser, W. J., Mocarski, E. S., & Dubyak, G. R. (2013). Proapoptotic chemotherapeutic drugs induce noncanonical processing and release of IL-1 β via caspase-8 in dendritic cells. *The Journal of Immunology*, 191(9), 4789-4803.
- Antonsson, B. (2001). Bax and other pro-apoptotic Bcl-2 family "killer-proteins" and their victim the mitochondrion. *Cell and tissue research*, 306(3), 347-361.
- Anwar, F., Latif, S., Ashraf, M., & Gilani, A. H. (2007). M. oleifera: a food plant with multiple medicinal uses. *Phytotherapy research*, 21(1), 17-25.
- Appleman, L. J., Berezhovskaya, A., Grass, I., & Boussiotis, V. A. (2000). CD28 costimulation mediates T cell expansion via IL-2-independent and IL-2-dependent regulation of cell cycle progression. *The Journal of Immunology*, 164(1), 144-151.
- Apte, R. N., Dotan, S., Elkabets, M., White, M. R., Reich, E., Carmi, Y., Voronov, E. (2006). The involvement of IL-1 in tumorigenesis, tumor invasiveness, metastasis and tumor-host interactions. *Cancer and Metastasis Reviews*, 25(3), 387-408.
- Arumuggam, N., Bhowmick, N. A., & Rupasinghe, H. (2015). A review: phytochemicals targeting JAK/STAT signaling and IDO expression in cancer. *Phytotherapy research*, 29(6), 805-817.
- Asare, G. A., Gyan, B., Bugyei, K., Adjei, S., Mahama, R., Addo, P., Nyarko, A. (2012). Toxicity potentials of the nutraceutical M. oleifera at supra-supplementation levels. *Journal of ethnopharmacology*, 139(1), 265-272.
- Asghar, U., Witkiewicz, A. K., Turner, N. C., & Knudsen, E. S. (2015). The history and future of targeting cyclin-dependent kinases in cancer therapy. *Nature reviews Drug discovery*, 14(2), 130-146.
- Ashraf, M. T., & Khan, R. H. (2003). Mitogenic lectins. *Medical Science Monitor*, 9(11), RA265-RA269.
- Asselin, E., Mills, G. B., & Tsang, B. K. (2001). XIAP regulates Akt activity and caspase-3-dependent cleavage during cisplatin-induced apoptosis in human ovarian epithelial cancer cells. *Cancer research*, 61(5), 1862-1868.
- Atawodi, S. E., Atawodi, J. C., Idakwo, G. A., Pfundstein, B., Haubner, R., Wurtele, G., . . . Owen, R. W. (2010). Evaluation of the polyphenol content and antioxidant properties of methanol extracts of the leaves, stem, and root barks of M. oleifera Lam. *Journal of Medicinal Food*, 13(3), 710-716.
- Attaran-Bandarabadi, F., Abhari, B. A., Neishabouri, S. H., & Davoodi, J. (2017). Integrity of XIAP is essential for effective activity recovery of apoptosome and its downstream caspases by Smac/Diablo. *International journal of biological macromolecules*, 101, 283-289.

- Awodele, O., Oreagba, I. A., Odoma, S., da Silva, J. A. T., & Osunkalu, V. O. (2012). Toxicological evaluation of the aqueous leaf extract of *M. oleifera* Lam.(Moringaceae). *Journal of ethnopharmacology*, 139(2), 330-336.
- Baig, S., Seevasant, I., Mohamad, J., Mukheem, A., Huri, H., & Kamarul, T. (2017). Potential of apoptotic pathway-targeted cancer therapeutic research: Where do we stand? *Cell death & disease*, 7(1), e2058.
- Bakre, A. G., Aderibigbe, A. O., & Ademowo, O. G. (2013). Studies on neuropharmacological profile of ethanol extract of *M. oleifera* leaves in mice. *Journal of ethnopharmacology*, 149(3), 783-789.
- Bauer, G. (2017). siRNA-based Analysis of the Abrogation of the Protective Function of Membrane-associated Catalase of Tumor Cells. *Anticancer research*, 37(2), 567-581.
- Bechtel, W., & Bauer, G. (2009). Catalase protects tumor cells from apoptosis induction by intercellular ROS signaling. *Anticancer research*, 29(11), 4541-4557.
- Bennett, R. N., Mellon, F. A., Foidl, N., Pratt, J. H., Dupont, M. S., Perkins, L., & Kroon, P. A. (2003). Profiling glucosinolates and phenolics in vegetative and reproductive tissues of the multi-purpose trees *M. oleifera* L.(horseradish tree) and *Moringa stenopetala* L. *Journal of agricultural and food chemistry*, 51(12), 3546-3553.
- Berkovich, L., Earon, G., Ron, I., Rimmon, A., Vexler, A., & Lev-Ari, S. (2013). *M. oleifera* aqueous leaf extract down-regulates nuclear factor-kappaB and increases cytotoxic effect of chemotherapy in pancreatic cancer cells. *BMC complementary and alternative medicine*, 13(1), 212.
- Bharali, R., Tabassum, J., & Azad, M. R. H. (2003). Chemomodulatory effect of *M. oleifera*, Lam, on hepatic carcinogen metabolising enzymes, antioxidant parameters and skin papillomagenesis in mice. *Asian Pacific Journal of Cancer Prevention*, 4(2), 131-140.
- Blair, G., & Cook, G. (2008). Cancer and the immune system: an overview. *Oncogene*, 27(45), 5868-5869.
- Bose, C. K. (2007). Possible role of *M. oleifera* Lam. root in epithelial ovarian cancer. *Medscape General Medicine*, 9(1), 26.
- Bournazos, S., DiLillo, D. J., & Ravetch, J. V. (2015). The role of Fc–Fc γ R interactions in IgG-mediated microbial neutralization. *Journal of Experimental Medicine*, jem. 20151267.
- Brodin, P., & Davis, M. M. (2017). Human immune system variation. *Nature Reviews Immunology*, 17(1), 21.

- Bystrom, J., Taher, T. E., Muhyaddin, M. S., Clanchy, F. I., Mangat, P., Jawad, A. S., Mageed, R. A. (2015). Harnessing the therapeutic potential of Th17 cells. *of inflammation*, 2015.
- Caltagirone, S., Rossi, C., Poggi, A., Ranelletti, F. O., Natali, P. G., Brunetti, M., ... & Piantelli, M. (2000). Flavonoids apigenin and quercetin inhibit melanoma growth and metastatic potential. *International Journal of Cancer*, 87(4), 595-600.
- Castro, N. P., Fedorova-Abrams, N. D., Merchant, A. S., Rangel, M. C., Nagaoka, T., Karasawa, H., Sharan, S. K. (2015). Cripto-1 as a novel therapeutic target for triple negative breast cancer. *Oncotarget*, 6(14), 11910.
- Castrogiovanni, C., Waterschoot, B., De Backer, O., & Dumont, P. (2017). Serine 392 phosphorylation modulates p53 mitochondrial translocation and transcription-independent apoptosis. *Cell Death and Differentiation*.
- Cavin, L. G., Wang, F., Factor, V. M., Kaur, S., Venkatraman, M., Thorgeirsson, S. S., & Arsura, M. (2005). Transforming growth factor- α inhibits the intrinsic pathway of c-myc-induced apoptosis through activation of nuclear factor- κ B in murine hepatocellular carcinomas. *Molecular cancer research*, 3(7), 403-412.
- Ceuppens, J., Baroja, M., Lorre, K., Van Damme, J., & Billiau, A. (1988). Human T cell activation with phytohemagglutinin. The function of IL-6 as an accessory signal. *The Journal of Immunology*, 141(11), 3868-3874.
- Chang, F., Lee, J., Navolanic, P., Steelman, L., Shelton, J., Blalock, W., . . . McCubrey, J. (2003). Involvement of PI3K/Akt pathway in cell cycle progression, apoptosis, and neoplastic transformation: a target for cancer chemotherapy. *Leukemia*, 17(3), 590.
- Charoensin, S. (2014). Antioxidant and anticancer activities of *M. oleifera* leaves. *Journal of Medicinal Plants Research*, 8(7), 318-325.
- Chen, D., Daniel, K. G., Chen, M. S., Kuhn, D. J., Landis-Piwowar, K. R., & Dou, Q. P. (2005). Dietary flavonoids as proteasome inhibitors and apoptosis inducers in human leukemia cells. *Biochemical pharmacology*, 69(10), 1421-1432.
- Chen, T., Zhang, Y., Wang, Z., Yang, J., Li, M., Wang, K., . . . Zhou, M. (2017). Recombinant rabies virus expressing IL-15 enhances immunogenicity through promoting the activation of dendritic cells in mice. *Virologica Sinica*, 32(4), 317-327.
- Chiang, L.-C., Ng, L. T., Chiang, W., Chang, M.-Y., & Lin, C.-C. (2003). Immunomodulatory activities of flavonoids, monoterpenoids, triterpenoids, iridoid glycosides and phenolic compounds of *Plantago* species. *Planta medica*, 69(07), 600-604.

- Chin, . Y., Jalil, J., Ng, P. Y., & Ng, S. F. (2018). Development and formulation of *M. oleifera* standardised leaf extract film dressing for wound healing application. *Journal of ethnopharmacology*, 212, 188-199.
- Chiou, W. F., Chen, C. F., & Lin, J. J. (2000). Mechanisms of suppression of inducible nitric oxide synthase (iNOS) expression in RAW 264.7 cells by andrographolide. *British Journal of Pharmacology*, 129(8), 1553-1560.
- Ciau-Uitz, A., Monteiro, R., Kirmizitas, A., & Patient, R. (2014). Developmental hematopoiesis: ontogeny, genetic programming and conservation. *Experimental hematatology*, 42(8), 669-683.
- Clark, B. D., Collins, K. L., Gandy, M. S., Webb, A. C., & Auron, P. E. (1986). Genomic sequence for human prointerleukin 1 beta: possible evolution from a reverse transcribed prointerleukin 1 alpha gene. *Nucleic acids research*, 14(20), 7897-7914.
- Coccia, M., Collignon, C., Hervé, C., Chalon, A., Welsby, I., Detienne, S., Waters, N. C. (2017). Cellular and molecular synergy in AS01-adjuvanted vaccines results in an early IFN γ response promoting vaccine immunogenicity. *npj Vaccines*, 2, 25.
- Coppin, J. P., Xu, Y., Chen, H., Pan, M.-H., Ho, C.-T., Juliani, R., Wu, Q. (2013). Determination of flavonoids by LC/MS and anti-inflammatory activity in *M. oleifera*. *Journal of Functional Foods*, 5(4), 1892-1899.
- Cory, S., Huang, D. C., & Adams, J. M. (2003). The Bcl-2 family: roles in cell survival and oncogenesis. *Oncogene*, 22(53), 8590-8607.
- Cosmi, L., Maggi, L., Santarlasci, V., Liotta, F., & Annunziato, F. (2014). T helper cells plasticity in inflammation. *Cytometry Part A*, 85(1), 36-42.
- Coulie, P. G., Hanagiri, T., & Takenoyama, M. (2001). From tumor antigens to immunotherapy. *International journal of clinical oncology*, 6(4), 163-170.
- Curiel, T. J., Coukos, G., Zou, L., Alvarez, X., Cheng, P., Mottram, P., Buow, M. (2004). Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nature medicine*, 10(9), 942-949.
- Dadi, S., Chhangawala, S., Whitlock, B. M., Franklin, R. A., Luo, C. T., Oh, S. A., Leslie, C. S. (2016). Cancer immuno surveillance by tissue-resident innate lymphoid cells and innate-like T cells. *Cell*, 164(3), 365-377.
- Dai, C., & Gu, W. (2010). p53 post-translational modification: deregulated in tumorigenesis. *Trends in molecular medicine*, 16(11), 528-536.
- Day, T. W., & Safa, A. R. (2009). RNA interference in cancer: targeting the anti-apoptotic protein c-FLIP for drug discovery. *Mini reviews in medicinal chemistry*, 9(6), 741-748.

- Dayal, B., Yannamreddy, V. R., Amin, R., Lea, M. A., & Attygalle, A. B. (2013). Bioactive Compounds in *M. oleifera*: isolation, structure elucidation, and their antiproliferative properties. In *Tropical and Subtropical Fruits: Flavors, Color, and Health Benefits* (pp. 203-219): ACS Publications.
- De Almagro, M., & Vucic, D. (2012). The inhibitor of apoptosis (IAP) proteins are critical regulators of signaling pathways and targets for anti-cancer therapy. *Exp Oncol*, 34(3), 200-211.
- de Andrade Luz, L., Rossato, F. A., e Costa, R. A. P., Napoleão, T. H., Paiva, P. M. G., & Coelho, L. C. B. B. (2017). Cytotoxicity of the coagulant *M. oleifera* lectin (cMoL) to B16-F10 melanoma cells. *Toxicology in Vitro*, 44, 94-99.
- De Coninck, B., Timmermans, P., Vos, C., Cammue, B. P., & Kazan, K. (2015). What lies beneath: belowground defense strategies in plants. *Trends in plant science*, 20(2), 91-101.
- de Melo, C. M. L., Paim, B. A., Zecchin, K. G., Morari, J., Chiaratti, M. R., Correia, M. T. S., . . . Paiva, P. M. G. (2010). Cramoll 1, 4 lectin increases ROS production, calcium levels, and cytokine expression in treated spleen cells of rats. *Molecular and cellular biochemistry*, 342(1-2), 163-169.
- De Palma, M., Biziato, D., & Petrova, T. V. (2017). Microenvironmental regulation of tumour angiogenesis. *Nature Reviews Cancer*, 17(8), 457.
- Debnath, S., Biswas, D., Ray, K., & Guha, D. (2011). *M. oleifera* induced potentiation of serotonin release by 5-HT 3 receptors in experimental ulcer model. *Phytomedicine*, 18(2), 91-95.
- De-la-Peña, C., Badri, D. V., Lei, Z., Watson, B. S., Brandão, M. M., Silva-Filho, M. C., . . . Vivanco, J. M. (2010). Root secretion of defense-related proteins is development-dependent and correlated with flowering time. *Journal of Biological Chemistry*, 285(40), 30654-30665.
- Del Poeta, G., Venditti, A., Del Principe, M. I., Maurillo, L., Buccisano, F., Tamburini, A., Mazzone, C. (2003). Amount of spontaneous apoptosis detected by Bax/Bcl-2 ratio predicts outcome in acute myeloid leukemia (AML). *Blood*, 101(6), 2125-2131.
- Del Prete, G. (1998). The concept of type-1 and type-2 helper T cells and their cytokines in humans. *International reviews of immunology*, 16(3-4), 427-455.
- Devaraj, V. C., Krishna, B., & Viswanatha, G. L. (2011). Simultaneous determination of quercetin, rutin and kaempferol in the leaf extracts of *M. oleifera* Lam. and *Raphinus sativus* Linn. by liquid chromatography-tandem mass spectrometry. *Journal of Chinese Integrative Medicine*, 9(9), 1022-1030.

- Efferth, T., Saeed, M. E., Mirghani, E., Alim, A., Yassin, Z., Saeed, E., Daak, S. (2017). Integration of phytochemicals and phytotherapy into cancer precision medicine. *Oncotarget*, 8(30), 50284.
- Eggers, N. A., & de Blanco, E. J. C. (2016). Phytochemicals in Nutrition and Health. *Nutrition: An Approach to Good Health and Disease Management*, 201.
- Ehrke, M. J. (2003). Immunomodulation in cancer therapeutics. *International immunopharmacology*, 3(8), 1105-1119.
- Eigenbrod, T., Park, J.-H., Harder, J., Iwakura, Y., & Núñez, G. (2008). Cutting edge: critical role for mesothelial cells in necrosis-induced inflammation through the recognition of IL-1 α released from dying cells. *The Journal of Immunology*, 181(12), 8194-8198.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic pathology*, 35(4), 495-516.
- Emens, L. A., Ascierto, P. A., Darcy, P. K., Demaria, S., Eggermont, A. M., Redmond, W. L., Marincola, F. M. (2017). Cancer immunotherapy: Opportunities and challenges in the rapidly evolving clinical landscape. *European Journal of Cancer*, 81, 116-129.
- England, H., Summersgill, H. R., Edye, M. E., Rothwell, N. J., & Brough, D. (2014). Release of interleukin-1 α or interleukin-1 β depends on mechanism of cell death. *Journal of Biological Chemistry*, 289(23), 15942-15950.
- Escribese, M., & Barber, D. (2017). New insight into cancer immunotherapy. *Allergologia et immunopathologia*.
- Faizi, S., Siddiqui, B. S., Saleem, R., Siddiqui, S., Aftab, K., & Gilani, A.-u. H. (1994). Isolation and structure elucidation of new nitrile and mustard oil glycosides from *M. oleifera* and their effect on blood pressure. *Journal of Natural Products*, 57(9), 1256-1261.
- Fernandes, E. E., Pulwale, A. V., Patil, G. A., & Moghe, A. S. (2016). Probing regenerative potential of *M. oleifera* aqueous extracts using In vitro cellular assays. *Pharmacognosy research*, 8(4), 231.
- Fili, L., Cardilicchia, E., Maggi, E., & Parronchi, P. (2014). Perspectives in vaccine adjuvants for allergen-specific immunotherapy. *Immunology letters*, 161(2), 207-210.
- Fonsatti, E., Altomonte, M., Nicotra, M. R., Natali, P. G., & Maio, M. (2003). Endoglin (CD105): a powerful therapeutic target on tumor-associated angiogenetic blood vessels. *Oncogene*, 22(42), 6557-6563.
- Francis, S., Nair, J., Shiji, P., Mohamed, S., & Geetha, P. (2016). A Case Series of Acute Methanol Poisoning from Northern Kerala. *Emergency Med*, 6(312), 2.

- Frankel, T., Lanfranca, M. P., & Zou, W. (2017). The Role of Tumor Microenvironment in Cancer Immunotherapy. In *Tumor Immune Microenvironment in Cancer Progression and Cancer Therapy* (pp. 51-64): Springer.
- Fuglie, L. J. (1999). The miracle tree: *M. oleifera*, natural nutrition for the tropics.
- Fulda, S. (2009). Tumor resistance to apoptosis. *International journal of cancer*, 124(3), 511-515.
- Fulda, S. (2010). Evasion of apoptosis as a cellular stress response in cancer. *International journal of cell biology*, 2010.
- Gallagher, G., Dickensheets, H., Eskdale, J., Izotova, L., Mirochnitchenko, O., Peat, J., Kotenko, S. (2000). Cloning, expression and initial characterisation of interleukin-19 (IL-19), a novel homologue of human interleukin-10 (IL-10). *Genes and immunity*, 1(7), 442.
- Galun, D., Srdic-Rajic, T., Bogdanovic, A., Loncar, Z., & Zuvela, M. (2017). Targeted therapy and personalized medicine in hepatocellular carcinoma: drug resistance, mechanisms, and treatment strategies. *Journal of hepatocellular carcinoma*, 4, 93.
- Garzon, R., & Croce, C. M. (2008). MicroRNAs in normal and malignant hematopoiesis. *Current opinion in hematology*, 15(4), 352-358.
- Gaur, U., & Aggarwal, B. B. (2003). Regulation of proliferation, survival and apoptosis by members of the TNF superfamily. *Biochemical pharmacology*, 66(8), 1403-1408.
- Gill, Z. P., Perks, C. M., Newcomb, P. V., & Holly, J. M. (1997). Insulin-like growth factor-binding protein (IGFBP-3) predisposes breast cancer cells to programmed cell death in a non-IGF-dependent manner. *Journal of Biological Chemistry*, 272(41), 25602-25607.
- Gismondi, A., Canuti, L., Impei, S., Di Marco, G., Kenzo, M., Colizzi, V., & Canini, A. (2013). Antioxidant extracts of African medicinal plants induce cell cycle arrest and differentiation in B16F10 melanoma cells. *International journal of oncology*, 43(3), 956-964.
- Gondi, C. S., Kandhukuri, N., Dinh, D. H., Gujrati, M., & Rao, J. S. (2007). Down-regulation of uPAR and uPA activates caspase-mediated apoptosis and inhibits the PI3K/AKT pathway. *International journal of oncology*, 31(1), 19-27.
- Gopalakrishnan, L., Doriya, K., & Kumar, D. S. (2016). *M. oleifera*: A review on nutritive importance and its medicinal application. *Food Science and Human Wellness*, 5(2), 49-56.

- Gopalan, C., Rama Sastri, B., & Balasubramanian, S. (1980). Nutrition value of Indian foods.
- Graf, F., Mosch, B., Koehler, L., Bergmann, R., Wuest, F., & Pietzsch, J. (2010). Cyclin-dependent kinase 4/6 (cdk4/6) inhibitors: perspectives in cancer therapy and imaging. *Mini reviews in medicinal chemistry*, 10(6), 527-539.
- Granado-Serrano, A. B., Martín, M. A., Bravo, L., Goya, L., & Ramos, S. (2006). Quercetin induces apoptosis via caspase activation, regulation of Bcl-2, and inhibition of PI-3-kinase/Akt and ERK pathways in a human hepatoma cell line (HepG2). *The Journal of nutrition*, 136(11), 2715-2721.
- Green, D. R., & Llambi, F. (2015). Cell death signaling. *Cold Spring Harbor perspectives in biology*, 7(12), a006080.
- Greene, L. M., Nathwani, S. M., & Zisterer, D. M. (2016). Inhibition of γ -secretase activity synergistically enhances tumour necrosis factor-related apoptosis-inducing ligand induced apoptosis in T-cell acute lymphoblastic leukemia cells via upregulation of death receptor 5. *Oncology letters*, 12(4), 2900-2905.
- Gubin, M. M., Artyomov, M. N., Mardis, E. R., & Schreiber, R. D. (2015). Tumor neoantigens: building a framework for personalized cancer immunotherapy. *The Journal of clinical investigation*, 125(9), 3413.
- Guillerman, R. P., Voss, S. D., & Parker, B. R. (2011). Leukemia and lymphoma. *Radiologic Clinics of North America*, 49(4), 767-797.
- Guon, T. E., & Chung, H. S. (2017). *M. oleifera* fruit induce apoptosis via reactive oxygen species-dependent activation of mitogen-activated protein kinases in human melanoma A2058 cells. *Oncology letters*, 14(2), 1703-1710.
- Gupta, A., Gautam, M. K., Singh, R. K., Kumar, M. V., Rao, C. V., Goel, R., & Anupurba, S. (2010). Immunomodulatory effect of *M. oleifera* Lam. extract on cyclophosphamide induced toxicity in mice.
- Gupta, S., Jain, R., Kachhwaha, S., & Kothari, S. (2017). Nutritional and medicinal applications of *M. oleifera* Lam.—Review of current status and future possibilities. *Journal of Herbal Medicine*.
- Hajighasemi, F., & Tajic, S. (2017). Assessment of Cytotoxicity of Dimethyl Sulfoxide in Human Hematopoietic Tumor Cell Lines. *Iranian Journal of Blood and Cancer*, 9(2), 48-53.
- Halder, S., Mehta, A. K., & Mediratta, P. K. (2012). Augmented humoral immune response and decreased cell-mediated immunity by *Aloe vera* in rats. *Inflammopharmacology*, 20(6), 343-346.

- Harrington, L. E., Hatton, R. D., Mangan, P. R., Turner, H., Murphy, T. L., Murphy, K. M., & Weaver, C. T. (2005). Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nature immunology*, 6(11), 1123-1132.
- Hassan, M., Watari, H., AbuAlmaaty, A., Ohba, Y., & Sakuragi, N. (2014). Apoptosis and molecular targeting therapy in cancer. *BioMed research international*, 2014.
- Helmstetter, C., Flossdorf, M., Peine, M., Kupz, A., Zhu, J., Hegazy, A. N., Radbruch, A. (2015). Individual T helper cells have a quantitative cytokine memory. *Immunity*, 42(1), 108-122.
- Hoang, T. T., & Raines, R. T. (2017). Molecular basis for the autonomous promotion of cell proliferation by angiogenin. *Nucleic acids research*, 45(2), 818-831.
- Holcik, M., Gibson, H., & Korneluk, R. G. (2001). XIAP: apoptotic brake and promising therapeutic target. *Apoptosis*, 6(4), 253-261.
- Hostanska, K., Hajto, T., Weber, K., Fischer, J., Lentzen, H., Sütterlin, B., & Saller, R. (1996). A natural immunity-activating plant lectin, Viscum album agglutinin-I, induces apoptosis in human lymphocytes, monocytes, monocytic THP-1 cells and murine thymocytes. *Natural immunity*, 15(6), 295-311.
- Hu, Z.-Q., Toda, M., Okubo, S., Hara, Y., & Shimamura, T. (1992). Mitogenic activity of (-) epigallocatechin gallate on B-cells and investigation of its structure-function relationship. *International journal of immunopharmacology*, 14(8), 1399-1407.
- Iwasaki, A., & Medzhitov, R. (2015). Control of adaptive immunity by the innate immune system. *Nature immunology*, 16(4), 343-353.
- Jadlowiec, C. C., Taner, T., & Wiesner, R. H. (2018). Immunosuppression: The Global Picture. *Schiff's Diseases of the Liver, Twelfth Edition*, 1065-1078.
- Jafarain, A., Asghari, G., & Ghassami, E. (2014). Evaluation of cytotoxicity of *M. oleifera* Lam. callus and leaf extracts on Hela cells. *Advanced biomedical research*, 3.
- Jankovic, J., & Aguilar, L. G. (2008). Current approaches to the treatment of Parkinson's disease. *Neuropsychiatric disease and treatment*, 4(4), 743.
- Jayanthi, M., Garg, S. K., Yadav, P., Bhatia, A., & Goel, A. (2015). Some newer marker phytoconstituents in methanolic extract of *M. oleifera* leaves and evaluation of its immunomodulatory and splenocytes proliferation potential in rats. *Indian journal of pharmacology*, 47(5), 518.

- John, C. M., Sandrasaigaran, P., Tong, C. K., Adam, A., & Ramasamy, R. (2011). Immunomodulatory activity of polyphenols derived from Cassia auriculata flowers in aged rats. *Cellular immunology*, 271(2), 474-479.
- Kajstura, M., Halicka, H. D., Pryjma, J., & Darzynkiewicz, Z. (2007). Discontinuous fragmentation of nuclear DNA during apoptosis revealed by discrete “sub-G1” peaks on DNA content histograms. *Cytometry Part A*, 71(3), 125-131.
- Kamath, N., Swaminathan, R., & Desai, N. (2016). Antibacterial activity of Indian medicinal plant-M. oleifera against MRSA and Klebsiella Spp.(ESBL) which are commonly isolated bacteria in hospital environments. *IJAR*, 2(8), 515-517.
- Kang, M. H., & Reynolds, C. P. (2009). Bcl-2 inhibitors: targeting mitochondrial apoptotic pathways in cancer therapy. *Clinical cancer research*, 15(4), 1126-1132.
- Katoh, O., Takahashi, T., Oguri, T., Kuramoto, K., Mihara, K., Kobayashi, M., Watanabe, H. (1998). Vascular endothelial growth factor inhibits apoptotic death in hematopoietic cells after exposure to chemotherapeutic drugs by inducing MCL1 acting as an antiapoptotic factor. *Cancer research*, 58(23), 5565-5569.
- Khalafalla, M. M., Abdellatef, E., Dafalla, H. M., Nassrallah, A. A., Aboul-Enein, K. M., Lightfoot, D. A., El-Shemy, H. A. (2010). Active principle from M. oleifera Lam leaves effective against two leukemias and a hepatocarcinoma. *African Journal of Biotechnology*, 9(49), 8467-8471.
- Khanuja, S. P. S., Arya, J. S., Tiruppadiripuliyur, R. S. K., Saikia, D., Kaur, H., Singh, M., . . . Srivastava, S. K. (2005). Nitrile glycoside useful as a bioenhancer of drugs and nutrients, process of its isolation from M. oleifera. In: Google Patents.
- Khatun, S., Khan, M., Ashraduzzaman, M., Pervin, F., Bari, L., & Absar, N. (2009). Antibacterial activity and cytotoxicity of three lectins purified from drumstick (M. oleifera Lam.) leaves. *Journal of Bio-Science*, 17, 89-94.
- Kim, K., Han, J., Lee, T. R., Shin, D. W., Chang, H., Cho, A.-R., Kwon, O. (2014). Comparative Analysis of Human Epidermal and Peripheral Blood $\gamma\delta$ T Cell Cytokine Profiles. *Annals of dermatology*, 26(3), 308-313.
- Kim, K. Y., Jang, W. Y., Lee, J. Y., Jun, D. Y., Ko, J. Y., Yun, Y. H., & Kim, Y. H. (2016). Kaempferol activates G2-checkpoint of the cell cycle resulting in G2-arrest and mitochondria-dependent apoptosis in human acute leukemia Jurkat T cells. *Journal of microbiology and biotechnology*, 26(2), 287-294.
- Koff, J. L., Ramachandiran, S., & Bernal-Mizrachi, L. (2015). A time to kill: targeting apoptosis in cancer. *International journal of molecular sciences*, 16(2), 2942-2955.

- Kraiphet, S., Butryee, C., Rungsipipat, A., Budda, S., Rattanapinyopitak, K., & Tuntipopipat, S. (2017). Apoptosis induced by *M. oleifera* Lam. pod in mouse colon carcinoma model. *Comparative Clinical Pathology*, 1-10.
- Kuhn, C., Rezende, R., & Weiner, H. L. (2016). IL-6R signaling inhibits generation of Th3 cells and is a promising therapeutic target for enhancing oral tolerance induction. In: Am Assoc Immnol.
- Kumari, D. J. (2010). Hypoglycaemic effect of *M. oleifera* and *Azadirachta indica* in type 2 diabetes mellitus. *Bioscan*, 5(20), 211-214.
- Lalas, S., Athanasiadis, V., Karageorgou, I., Batra, G., Nanos, G. D., & Makris, D. P. (2017). Nutritional Characterization of Leaves and Herbal Tea of *M. oleifera* Cultivated in Greece. *Journal of Herbs, Spices & Medicinal Plants*, 1-14.
- Landskron, G., De la Fuente, M., Thuwajit, P., Thuwajit, C., & Hermoso, M. A. (2014). Chronic inflammation and cytokines in the tumor microenvironment. *Journal of immunology research*, 2014.
- Larson, J. L., Wolf, D. C., & Butterworth, B. E. (1993). Acute hepatotoxic and nephrotoxic effects of chloroform in male F-344 rats and female B6C3F1 mice. *Fundamental and applied toxicology*, 20(3), 302-315.
- Laxminarayan, R., Duse, A., Wattal, C., Zaidi, A. K., Wertheim, H. F., Sumpradit, N., . . . Goossens, H. (2013). Antibiotic resistance—the need for global solutions. *The Lancet infectious diseases*, 13(12), 1057-1098.
- Lee, D.-H., Szczepanski, M., & Lee, Y. J. (2008). Role of Bax in quercetin-induced apoptosis in human prostate cancer cells. *Biochemical pharmacology*, 75(12), 2345-2355.
- Lee, S., & Margolin, K. (2011). Cytokines in cancer immunotherapy. *Cancers*, 3(4), 3856-3893.
- Lee, T. B., Min, Y. D., Lim, S. C., Kim, K. J., Jeon, H. J., Choi, S. M., & Choi, C. H. (2002). Fas (Apo-1/CD95) and Fas ligand interaction between gastric cancer cells and immune cells. *Journal of gastroenterology and hepatology*, 17(1), 32-38.
- Leelavinothan, P., Karamać, M., Kosińska, A., Rybarczyk, A., & Amarowicz, R. (2007). Antioxidant activity of the crude extracts of drumstick tree (*M. oleifera* Lam.) and sweet broomweed (*Scoparia dulcis* L.) leaves. *Polish Journal of Food and Nutrition Sciences*, 57(2), 203-208.
- Leen, A., Tzannou, I., Bilgi, M., Liu, H., Vera, J. F., Gerdemann, U., Gee, A. P. (2015). Immunotherapy for lymphoma using T cells targeting multiple tumor associated antigens. In: Am Soc Hematology.

- Leone, A., Fiorillo, G., Criscuoli, F., Ravassenghi, S., Santagostini, L., Fico, G., Pozzi, F. (2015). Nutritional characterization and phenolic profiling of *M. oleifera* leaves grown in Chad, Sahrawi Refugee Camps, and Haiti. *International journal of molecular sciences*, 16(8), 18923-18937.
- Levine, A. J. (1997). p53, the cellular gatekeeper for growth and division. *Cell*, 88(3), 323-331.
- Li, A., Chen, B., Sumners, C., Gu, D., Jiang, G., Hu, G., Li, J. (2017). Angiotensin II type 2 receptor promotes apoptosis and inhibits angiogenesis in bladder cancer. *Journal of Experimental & Clinical Cancer Research*, 36(1), 77.
- Liao, W., Lin, J.-X., & Leonard, W. J. (2011). IL-2 family cytokines: new insights into the complex roles of IL-2 as a broad regulator of T helper cell differentiation. *Current opinion in immunology*, 23(5), 598-604.
- Liao, Y.-C., Liang, W.-G., Chen, F.-W., Hsu, J.-H., Yang, J.-J., & Chang, M.-S. (2002). IL-19 induces production of IL-6 and TNF- α and results in cell apoptosis through TNF- α . *The Journal of Immunology*, 169(8), 4288-4297.
- Libreros, S., Garcia-Areas, R., & Iragavarapu-Charyulu, V. (2013). CHI3L1 plays a role in cancer through enhanced production of pro-inflammatory/pro-tumorigenic and angiogenic factors. *Immunologic research*, 57(1-3), 99-105.
- Lipinski, C. A., Lombardo, F., Dominy, B. W., & Feeney, P. J. (1997). Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced drug delivery reviews*, 23(1-3), 3-25.
- Long, J., & Ryan, K. (2012). New frontiers in promoting tumour cell death: targeting apoptosis, necroptosis and autophagy. *Oncogene*, 31(49), 5045-5060.
- Lord, J. D., McIntosh, B. C., Greenberg, P. D., & Nelson, B. H. (2000). The IL-2 receptor promotes lymphocyte proliferation and induction of the c-myc, bcl-2, and bcl-x genes through the trans-activation domain of Stat5. *The Journal of Immunology*, 164(5), 2533-2541.
- Luo, H., Jiang, B., Li, B., Li, Z., Jiang, B. H., & Chen, Y. C. (2012). Kaempferol nanoparticles achieve strong and selective inhibition of ovarian cancer cell viability. *International journal of nanomedicine*, 7, 3951.
- Lu, Y., Wang, R., Guo, B., & Jia, Y. (2016). Quercetin inhibits angiotensin II induced apoptosis via mitochondrial pathway in human umbilical vein endothelial cells. *Eur. Rev. Med. Pharmacol. Sci*, 20, 1609-1616.
- Luheshi, N., Davies, G., Poon, E., Wiggins, K., McCourt, M., & Legg, J. (2014). Th1 cytokines are more effective than Th2 cytokines at licensing anti-tumour functions in CD40-activated human macrophages in vitro. *European journal of immunology*, 44(1), 162-172.

- Luqman, S., Srivastava, S., Kumar, R., Maurya, A. K., & Chanda, D. (2012). Experimental assessment of *M. oleifera* leaf and fruit for its antistress, antioxidant, and scavenging potential using in vitro and in vivo assays. *Evidence-Based Complementary and Alternative Medicine*, 2012.
- Ma, M., Zhao, L., Sun, G., Zhang, C., Liu, L., Du, Y., Shan, B. (2016). Mda-7/IL-24 enhances sensitivity of B cell lymphoma to chemotherapy drugs. *Oncology reports*, 35(5), 3122-3130.
- Madi, N., Dany, M., Abdoun, S., & Usta, J. (2016). *M. oleifera*'s nutritious aqueous leaf extract has anticancerous effects by compromising mitochondrial viability in an ROS-dependent manner. *Journal of the American College of Nutrition*, 35(7), 604-613.
- Madian, A. G., Wheeler, H. E., Jones, R. B., & Dolan, M. E. (2012). Relating human genetic variation to variation in drug responses. *Trends in Genetics*, 28(10), 487-495.
- Mahajan, S. G., & Mehta, A. A. (2010). Immunosuppressive activity of ethanolic extract of seeds of *M. oleifera* Lam. in experimental immune inflammation. *Journal of ethnopharmacology*, 130(1), 183-186.
- Mahdi, H. J., Khan, N. A. K., Asmawi, M. Z. B., Mahmud, R., & Murugaiyah, V. A. (2017). in vivo anti-arthritis and anti-noceptic effects of ethanol extract of *M. oleifera* leaves on complete Freund's adjuvant (CFA)-induced arthritis in rats. *Integrative Medicine Research*.
- Majchrzak, K., Kaspera, W., Szymaś, J., Bobek-Billewicz, B., Hebda, A., & Majchrzak, H. (2013). Markers of angiogenesis (CD31, CD34, rCBV) and their prognostic value in low-grade gliomas. *Neurologia i Neurochirurgia Polska*, 47(4), 325-331.
- Marrufo, T., Encarnaçāo, S., Silva, O. M. D., Duarte, A., Neto, F. F., Barbosa, F. M., & Agostinho, A. B. (2013). Chemical characterization and determination of antioxidant and antimicrobial activities of the leaves of *M. oleifera*. *International Network Environmental Management Conflicts*, 2(1), 1-15.
- Marzabadi, C. H., & Franck, R. W. (2017). Small-Molecule Carbohydrate-Based Immunostimulants. *Chemistry-A European Journal*, 23(8), 1728-1742.
- Matsui, W., Huff, C. A., Wang, Q., Malehorn, M. T., Barber, J., Tanhehco, Y., Jones, R. J. (2004). Characterization of clonogenic multiple myeloma cells. *Blood*, 103(6), 2332-2336.
- Maurya, S. K., Tewari, M., Sharma, B., & Shukla, H. S. (2013). Expression of procaspase 3 and activated caspase 3 and its relevance in hormone-responsive gallbladder carcinoma chemotherapy. *The Korean journal of internal medicine*, 28(5), 573.

- Medina, K. (2016). Overview of the immune system. *Handbook of clinical neurology*, 133, 61-76.
- Menezes, M. E., Bhatia, S., Bhoopathi, P., Das, S. K., Emdad, L., Dasgupta, S., Fisher, P. B. (2014). MDA-7/IL-24: multifunctional cancer killing cytokine. In *Anticancer Genes* (pp. 127-153): Springer.
- Meyerovich, K., Fukaya, M., Terra, L. F., Ortis, F., Eizirik, D. L., & Cardozo, A. K. (2016). The non-canonical NF- κ B pathway is induced by cytokines in pancreatic beta cells and contributes to cell death and proinflammatory responses in vitro. *Diabetologia*, 59(3), 512-521.
- Miean, K. H., & Mohamed, S. (2001). Flavonoid (myricetin, quercetin, kaempferol, luteolin, and apigenin) content of edible tropical plants. *Journal of agricultural and food chemistry*, 49(6), 3106-3112.
- Mishra, G., Singh, P., Srivastav, S., & Verma, R. K. (2017). Moringa olifera-An important medicinal plant: A Review of Its Traditional Uses, Phytochemistry and Pharmacological Properties. no. February, 2011.
- Mishra, G., Singh, P., Verma, R., Kumar, S., Srivastav, S., Jha, K., & Khosa, R. (2011). Traditional uses, phytochemistry and pharmacological properties of M. oleifera plant: An overview. *Der Pharmacia Lettre*, 3(2), 141-164.
- Mocellin, S., Wang, E., & Marincola, F. M. (2001). Cytokines and immune response in the tumor microenvironment. *Journal of immunotherapy*, 24(5), 392-407.
- Moore, M. G., Wetterau, L. A., Francis, M. J., Peehl, D. M., & Cohen, P. (2003). Novel stimulatory role for insulin-like growth factor binding protein-2 in prostate cancer cells. *International journal of cancer*, 105(1), 14-19.
- Moreno-Lorenzana, D., Avilés-Vazquez, S., Sandoval Esquivel, M. A., Alvarado-Moreno, A., Ortiz-Navarrete, V., Torres-Martínez, H., . . . Chavez-Gonzalez, A. (2016). CDKIs p18INK4c and p57Kip2 are involved in quiescence of CML leukemic stem cells after treatment with TKI. *Cell Cycle*, 15(9), 1276-1287.
- Mousa, M., Osman, A., & Hady, H. A. (2017). Performance, immunology and biochemical parameters of M. oleifera and/or Cichorium intybus addition to broiler chicken ration. *Journal of Veterinary Medicine and Animal Health*, 9(10), 255-263.
- Movafagh, A., Heydary, H., Mortazavi-Tabatabaei, S. A., & Azargashb, E. (2011). The significance application of indigenous phytohemagglutinin (PHA) mitogen on metaphase and cell culture procedure. *Iranian journal of pharmaceutical research: IJPR*, 10(4), 895.
- Muller, P. A., & Vousden, K. H. (2013). p53 mutations in cancer. *Nature cell biology*, 15(1), 2-8.

- Mun'im, A., Puteri, M. U., & Sari, S. P. (2016). Anti-Anemia Effect of Standardized Extract of *M. oleifera* Lamk. Leaves on Aniline Induced Rats. *Pharmacognosy Journal*, 8(3).
- Myers, J. S., von Lersner, A. K., Robbins, C. J., & Sang, Q.-X. A. (2015). Differentially expressed genes and signature pathways of human prostate cancer. *PloS one*, 10(12), e0145322.
- Nagata, S., & Tanaka, M. (2017). Programmed cell death and the immune system. *Nature reviews Immunology*, 17(5), 333-340.
- Nahm, D. H. (2015). Personalized immunomodulatory therapy for atopic dermatitis: an allergist's view. *Annals of dermatology*, 27(4), 355-363.
- Nair, S., & Varalakshmi, K. (2011). Anticancer, cytotoxic potential of *M. oleifera* extracts on HeLa cell line. *Journal of Natural Pharmaceuticals*, 2(3), 138-138.
- Nambiar, V. S., Guin, P., Parnami, S., & Daniel, M. (2010). Impact of antioxidants from drumstick leaves on the lipid profile of hyperlipidemics. *J Herb Med Toxicol*, 4(1), 165-172.
- Nassiri, F., Cusimano, M. D., Scheithauer, B. W., Rotondo, F., Fazio, A., Yousef, G. M., Lloyd, R. V. (2011). Endoglin (CD105): a review of its role in angiogenesis and tumor diagnosis, progression and therapy. *Anticancer research*, 31(6), 2283-2290.
- Nelson, M. H., & Paulos, C. M. (2015). Novel immunotherapies for hematologic malignancies. *Immunological reviews*, 263(1), 90-105.
- Nfambi, J., Bbosa, G. S., Sembajwe, L. F., Gakunga, J., & Kasolo, J. N. (2015). Immunomodulatory activity of methanolic leaf extract of *M. oleifera* in Wistar albino rats. *Journal of basic and clinical physiology and pharmacology*, 26(6), 603-611.
- Nicholas, C., & Lesinski, G. B. (2011). Immunomodulatory cytokines as therapeutic agents for melanoma.
- Nicholls, E. F., Madera, L., & Hancock, R. E. (2010). Immunomodulators as adjuvants for vaccines and antimicrobial therapy. *Annals of the New York Academy of Sciences*, 1213(1), 46-61.
- Nikolich-Žugich, J. (2018). The twilight of immunity: emerging concepts in aging of the immune system. *Nature immunology*, 19(1), 10.
- Normanno, N., Luca, A. D., Bianco, C., Maiello, M. R., Carriero, M. V., Rehman, A., . . . Sanicola, M. (2004). Cripto-1 overexpression leads to enhanced invasiveness and resistance to anoikis in human MCF-7 breast cancer cells. *Journal of cellular physiology*, 198(1), 31-39.

- O'connell, P. J., Kuypers, D. R., Mannon, R. B., Abecassis, M., Chadban, S. J., Gill, J. S., . . . Stock, P. G. (2017). Clinical trials for immunosuppression in transplantation: the case for reform and change in direction. *Transplantation*, 101(7), 1527-1534.
- Omodanisi, E. I., Aboua, Y. G., Chegou, N. N., & Oguntibeju, O. O. (2017). Hepatoprotective, antihyperlipidemic, and anti-inflammatory activity of *M. oleifera* in diabetic-induced damage in male wistar rats. *Pharmacognosy research*, 9(2), 182.
- Otto, T., & Sicinski, P. (2017). Cell cycle proteins as promising targets in cancer therapy. *Nature Reviews Cancer*, 17(2), 93-115.
- Oudah, K. H., Abdou, N. S., Serya, R., & Abouzid, K. (2017). An overview on the prospective CDKs inhibitors as anti-cancer drugs. *Journal of American Science*, 13(4).
- Ouyang, L., Shi, Z., Zhao, S., Wang, F. T., Zhou, T. T., Liu, B., & Bao, J. K. (2012). Programmed cell death pathways in cancer: a review of apoptosis, autophagy and programmed necrosis. *Cell proliferation*, 45(6), 487-498.
- Paiva, P. M., Santana, G. M., Souza, I. F., Albuquerque, L. P., Agra-Neto, A. C., Albuquerque, A. C., Coelho, L. C. (2011). Effect of lectins from *Opuntia ficus indica* cladodes and *M. oleifera* seeds on survival of *Nasutitermes corniger*. *International biodeterioration & biodegradation*, 65(7), 982-989.
- Pal, S. K., Mukherjee, P. K., & Saha, B. (1995). Studies on the antiulcer activity of *M. oleifera* leaf extract on gastric ulcer models in rats. *Phytotherapy research*, 9(6), 463-465.
- Paliard, X., de Waal Malefijt, R., Yssel, H., Blanchard, D., Chretien, I., Abrams, J., Spits, H. (1988). Simultaneous production of IL-2, IL-4, and IFN-gamma by activated human CD4+ and CD8+ T cell clones. *The Journal of Immunology*, 141(3), 849-855.
- Pamok, S., Vinitketkumnuen, S. S. U., & Saenphet, K. (2012). Antiproliferative effect of *M. oleifera* Lam. and *Pseuderanthemum palatiferum* (Nees) Radlk extracts on the colon cancer cells. *Journal of Medicinal Plants Research*, 6(1), 139-145.
- Pan, S.-Y., Litscher, G., Gao, S.-H., Zhou, S.-F., Yu, Z.-L., Chen, H.-Q., Ko, K.-M. (2014). Historical perspective of traditional indigenous medical practices: the current renaissance and conservation of herbal resources. *Evidence-Based Complementary and Alternative Medicine*, 2014.
- Pandey, A., Pandey, R., Tripathi, P., Gupta, P., Haider, J., Bhatt, S., & Singh, A. (2012). *M. oleifera* Lam. (*Sahjan*)-A Plant with a Plethora of Diverse Therapeutic Benefits: An Updated Retrospection. *Medicinal and Aromatic Plants*, 1(1), 1-8.

- Papapetropoulos, A., Fulton, D., Mahboubi, K., Kalb, R. G., O'Connor, D. S., Li, F., . . . Sessa, W. C. (2000). Angiopoietin-1 inhibits endothelial cell apoptosis via the Akt/survivin pathway. *Journal of Biological Chemistry*, 275(13), 9102-9105.
- Parasuraman, S., Sujithra, J., Syamittra, B., Yeng, W. Y., Ping, W. Y., Muralidharan, S., Dhanaraj, S. A. (2014). Evaluation of sub-chronic toxic effects of petroleum ether, a laboratory solvent in Sprague-Dawley rats. *Journal of basic and clinical pharmacy*, 5(4), 89.
- Park, H., Li, Z., Yang, X. O., Chang, S. H., Nurieva, R., Wang, Y.-H., Tian, Q. (2005). A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nature immunology*, 6(11), 1133-1141.
- Parkin, J., & Cohen, B. (2001). An overview of the immune system. *The Lancet*, 357(9270), 1777-1789.
- Parvathy, M., & Umamaheshwari, A. (2007). Cytotoxic effect of *M. oleifera* leaf extracts on human multiple myeloma cell lines. *Trends in Medical Research*, 2(1), 44-50.
- Patel, M. P., Masood, A., Patel, P. S., & Chanan-Khan, A. A. (2009). Targeting the Bcl-2. *Current opinion in oncology*, 21(6), 516-523.
- Pawelec, G., Borowitz, A., Krammer, P. H., & Wernet, P. (1982). Constitutive interleukin 2 production by the JURKAT human leukemic T cell line. *European journal of immunology*, 12(5), 387-392.
- Pegoraro, L., Matera, L., Ritz, J., Levis, A., Palumbo, A., & Biagini, G. (1983). Establishment of a Ph1-positive human cell line (BV173). *Journal of the National Cancer Institute*, 70(3), 447-453.
- Pereira, L. M. S., Gomes, S. T. M., Ishak, R., & Vallinoto, A. C. R. (2017). Regulatory T Cell and Forkhead Box Protein 3 as Modulators of Immune Homeostasis. *Frontiers in Immunology*, 8.
- Pingel, J. T., & Thomas, M. L. (1989). Evidence that the leukocyte-common antigen is required for antigen-induced T lymphocyte proliferation. *Cell*, 58(6), 1055-1065.
- Possenti, M., Baima, S., Raffo, A., Durazzo, A., Giusti, A. M., & Natella, F. (2017). Glucosinolates in Food. *Glucosinolates*, 87-132.
- Post, S., Weng, Y.-C., Cimprich, K., Chen, L. B., Xu, Y., & Eva, Y.-H. L. (2001). Phosphorylation of serines 635 and 645 of human Rad17 is cell cycle regulated and is required for G1/S checkpoint activation in response to DNA damage. *Proceedings of the National Academy of Sciences*, 98(23), 13102-13107.

- Post, S. M., Tomkinson, A. E., & Lee, E. Y. H. (2003). The human checkpoint Rad protein Rad17 is chromatin-associated throughout the cell cycle, localizes to DNA replication sites, and interacts with DNA polymerase ϵ . *Nucleic acids research*, 31(19), 5568-5575.
- Pradhan, A. K., Talukdar, S., Bhoopathi, P., Shen, X.-N., Emdad, L., Das, S. K., . . . Fisher, P. B. (2017). mda-7/IL-24 Mediates Cancer Cell-Specific Death via Regulation of miR-221 and the Beclin-1 Axis. *Cancer research*, 77(4), 949-959.
- Puri, V., Gupta, A. D., Chaudhry, N., & Saran, R. K. (2015). Reversible cerebral and brain stem dysfunction in n: Hexane neuropathy. *Annals of Indian Academy of Neurology*, 18(4), 464.
- Rabinovich, G. A., & Croci, D. O. (2012). Regulatory circuits mediated by lectin-glycan interactions in autoimmunity and cancer. *Immunity*, 36(3), 322-335.
- Rachmawati, I., & Rifa'i, M. (2014). In Vitro Immunomodulatory Activity of Aqueous Extract of *M. oleifera* Lam. Leaf to the CD4+, CD8+ and B220+ Cells in *Mus musculus*. *The Journal of Experimental Life Science*, 4(1), 15-20.
- Radulescu, R. T. (2017). Retinoblastoma Tumor Suppressor Protein (RB). *Encyclopedia of Signaling Molecules*, 1-4.
- Rahman, M. M., Sheikh, M. M. I., Sharmin, S. A., Islam, M. S., Rahman, M. A., Rahman, M. M., & Alam, M. F. (2009). Antibacterial activity of leaf juice and extracts of *M. oleifera* Lam. against some human pathogenic bacteria. *CMU J Nat Sci*, 8(2), 219.
- Rao, A. V., Devi, P. U., & Kamath, R. (2001). In vivo radioprotective effect of *M. oleifera* leaves.
- Rao, D., & Said, J. (2017). Pathology and Molecular Pathology of Hematologic Malignancies. In *Pathology and Epidemiology of Cancer* (pp. 571-590): Springer.
- Reddy, D. B., & Reddanna, P. (2009). Chebulagic acid (CA) attenuates LPS-induced inflammation by suppressing NF- κ B and MAPK activation in RAW 264.7 macrophages. *Biochemical and Biophysical Research Communications*, 381(1), 112-117.
- Reichert, T. E., Nagashima, S., Kashii, Y., Stanson, J., Gao, G., Dou, Q. P., & Whiteside, T. L. (2000). Interleukin-2 expression in human carcinoma cell lines and its role in cell cycle progression. *Oncogene*, 19(4), 514.
- Ren, B., Cam, H., Takahashi, Y., Volkert, T., Terragni, J., Young, R. A., & Dynlacht, B. D. (2002). E2F integrates cell cycle progression with DNA repair, replication, and G2/M checkpoints. *Genes & development*, 16(2), 245-256.

- Restifo, N. P., Dudley, M. E., & Rosenberg, S. A. (2012). Adoptive immunotherapy for cancer: harnessing the T cell response. *Nature Reviews Immunology*, 12(4), 269.
- Rodríguez-Pérez, C., Quirantes-Piné, R., Fernández-Gutiérrez, A., & Segura-Carretero, A. (2015). Optimization of extraction method to obtain a phenolic compounds-rich extract from *M. oleifera* Lam leaves. *Industrial Crops and Products*, 66, 246-254.
- Rosenberg, S. A. (2008). Overcoming obstacles to the effective immunotherapy of human cancer. *Proceedings of the National Academy of Sciences*, 105(35), 12643-12644.
- Rosenberg, S. A., Yang, J. C., & Restifo, N. P. (2004). Cancer immunotherapy: moving beyond current vaccines. *Nature medicine*, 10(9), 909-915.
- Russo, M., Nigro, P., Rosiello, R., D'arienzo, R., & Russo, G. (2007). Quercetin enhances CD95-and TRAIL-induced apoptosis in leukemia cell lines. *Leukemia*, 21(5), 1130-1130.
- Sadelain, M., Brentjens, R., & Rivière, I. (2013). The basic principles of chimeric antigen receptor design. *Cancer discovery*, 3(4), 388-398.
- Saini, R. K., Sivanesan, I., & Keum, Y.-S. (2016). Phytochemicals of *M. oleifera*: a review of their nutritional, therapeutic and industrial significance. *3 Biotech*, 6(2), 203.
- Santos, A. F., Luz, L. A., Argolo, A. C., Teixeira, J. A., Paiva, P. M., & Coelho, L. C. (2009). Isolation of a seed coagulant *M. oleifera* lectin. *Process biochemistry*, 44(4), 504-508.
- Sasaki, H., Sheng, Y., Kotsuji, F., & Tsang, B. K. (2000). Down-regulation of X-linked inhibitor of apoptosis protein induces apoptosis in chemoresistant human ovarian cancer cells. *Cancer research*, 60(20), 5659-5666.
- Sathyananarayanan, V., & Neelapu, S. S. (2015). Cancer immunotherapy: Strategies for personalization and combinatorial approaches. *Molecular oncology*, 9(10), 2043-2053.
- Schijns, V., Tartour, E., Michalek, J., Stathopoulos, A., Dobrovolskienė, N. T., & Strioga, M. M. (2014). Immune adjuvants as critical guides directing immunity triggered by therapeutic cancer vaccines. *Cytotherapy*, 16(4), 427-439.
- Schlee, M., & Hartmann, G. (2016). Discriminating self from non-self in nucleic acid sensing. *Nature reviews Immunology*, 16(9), 566-580.
- Schmaltz, C., Alpdogan, O., Kappel, B. J., Muriglan, S. J., Rotolo, J. A., Ongchin, J., Crawford, J. M. (2002). T cells require TRAIL for optimal graft-versus-tumor activity. *Nature medicine*, 8(12).

- Schmitt, N., & Ueno, H. (2015). Regulation of human helper T cell subset differentiation by cytokines. *Current opinion in immunology*, 34, 130-136.
- Schneider, U., Schwenk, H. U., & Bornkamm, G. (1977). Characterization of EBV-genome negative "null" and "T" cell lines derived from children with acute lymphoblastic leukemia and leukemic transformed non-Hodgkin lymphoma. *Int J Cancer*, 19(5), 621-626.
- Schroeder, J. T. (2014). Diagnostic Components: T Helper Cell Cytokines (IL-4, IL-5, IL-9, IL-10, IL-13, IL-17). In *Encyclopedia of Medical Immunology* (pp. 221-226): Springer.
- Seki, N., Hayakawa, Y., Brooks, A. D., Wine, J., Wiltz, R. H., Yagita, H., . . . Sayers, T. J. (2003). Tumor necrosis factor-related apoptosis-inducing ligand-mediated apoptosis is an important endogenous mechanism for resistance to liver metastases in murine renal cancer. *Cancer research*, 63(1), 207-213.
- Sena, L. A., Li, S., Jairaman, A., Prakriya, M., Ezponda, T., Hildeman, D. A., . . . Bryce, P. J. (2013). Mitochondria are required for antigen-specific T cell activation through reactive oxygen species signaling. *Immunity*, 38(2), 225-236.
- ShaheenáSiddiqui, B. (1994). Novel hypotensive agents, niazimin A, niazimin B, niazicin A and niazicin B from *M. oleifera*: Isolation of first naturally occurring carbamates. *Journal of the Chemical Society, Perkin Transactions I*(20), 3035-3040.
- Shankaran, V., Ikeda, H., Bruce, A. T., White, J. M., Swanson, P. E., Old, L. J., & Schreiber, R. D. (2001). IFN γ and lymphocytes prevent primary tumour development and shape tumour immunogenicity. *Nature*, 410(6832), 1107-1111.
- Shanker, K., Gupta, M. M., Srivastava, S. K., Bawankule, D. U., Pal, A., & Khanuja, S. P. (2007). Determination of bioactive nitrile glycoside (s) in drumstick (*M. oleifera*) by reverse phase HPLC. *Food chemistry*, 105(1), 376-382.
- Sharifudin, S. A., Fakurazi, S., Hidayat, M. T., Hairuszah, I., Aris Mohd Moklas, M., & Arulselvan, P. (2013). Therapeutic potential of *M. oleifera* extracts against acetaminophen-induced hepatotoxicity in rats. *Pharmaceutical Biology*, 51(3), 279-288.
- Sharon, N. (2007). Lectins: carbohydrate-specific reagents and biological recognition molecules. *Journal of Biological Chemistry*, 282(5), 2753-2764.
- Shatrova, A., Mityushova, E., Aksnov, N., & Marakhova, I. (2015). CD25 expression on the surface of Jurkat cells. *Cell and Tissue Biology*, 9(5), 364-370.

- Shen, H., & Goldstein, D. R. (2009). IL-6 and TNF- α synergistically inhibit allograft acceptance. *Journal of the American Society of Nephrology*, 20(5), 1032-1040.
- Shen, X., Xi, G., Maile, L. A., Wai, C., Rosen, C. J., & Clemmons, D. R. (2012). Insulin-like growth factor (IGF) binding protein 2 functions coordinately with receptor protein tyrosine phosphatase β and the IGF-I receptor to regulate IGF-I-stimulated signaling. *Molecular and cellular biology*, 32(20), 4116-4130.
- Sherr, C. J., & Sicinski, P. (2018). The D-Type Cyclins: A Historical Perspective. In *D-type Cyclins and Cancer* (pp. 1-26): Springer.
- Singh, J. K., Simões, B. M., Howell, S. J., Farnie, G., & Clarke, R. B. (2013). Recent advances reveal IL-8 signaling as a potential key to targeting breast cancer stem cells. *Breast Cancer Research*, 15(4), 210.
- Singh, V., Ram, M., Kumar, R., Prasad, R., Roy, B. K., & Singh, K. K. (2017). Phosphorylation: Implications in Cancer. *The protein journal*, 36(1), 1-6.
- Sinha, M., Das, D. K., Bhattacharjee, S., Majumdar, S., & Dey, S. (2011). Leaf extract of *M. oleifera* prevents ionizing radiation-induced oxidative stress in mice. *Journal of Medicinal Food*, 14(10), 1167-1172.
- Sinha, M., Das, D. K., Datta, S., Ghosh, S., & Dey, S. (2012). Amelioration of ionizing radiation induced lipid peroxidation in mouse liver by *M. oleifera* Lam. leaf extract.
- Škerget, M., Kotnik, P., Hadolin, M., Hraš, A. R., Simonič, M., & Knez, Ž. (2005). Phenols, proanthocyanidins, flavones and flavonols in some plant materials and their antioxidant activities. *Food chemistry*, 89(2), 191-198.
- Somerville, V. S., Braakhuis, A. J., & Hopkins, W. G. (2016). Effect of flavonoids on upper respiratory tract infections and immune function: A systematic review and meta-analysis. *Advances in Nutrition: An International Review Journal*, 7(3), 488-497.
- Son, Y. O., Lee, K. Y., Kook, S. H., Lee, J. C., Kim, J. G., Jeon, Y. M., & Jang, Y. S. (2004). Selective effects of quercetin on the cell growth and antioxidant defense system in normal versus transformed mouse hepatic cell lines. *European journal of pharmacology*, 502(3), 195-204.
- Song, S., Jacobson, K. N., McDermott, K. M., Reddy, S. P., Cress, A. E., Tang, H., ... Makino, A. (2015). ATP promotes cell survival via regulation of cytosolic [Ca $^{2+}$] and Bcl-2/Bax ratio in lung cancer cells. *American Journal of Physiology-Cell Physiology*, 310(2), C99-C114.
- Sreelatha, S., Jeyachitra, A., & Padma, P. (2011). Antiproliferation and induction of apoptosis by *M. oleifera* leaf extract on human cancer cells. *Food and Chemical Toxicology*, 49(6), 1270-1275.

- Staudt, L. M. (2003). Molecular diagnosis of the hematologic cancers. *New England Journal of Medicine*, 348(18), 1777-1785.
- Stohs, S. J., & Hartman, M. J. (2015). Review of the safety and efficacy of *M. oleifera*. *Phytotherapy research*, 29(6), 796-804.
- Suchal, K., Malik, S., Gamad, N., Malhotra, R. K., Goyal, S. N., Bhatia, J., & Arya, D. S. (2016). Kampeferol protects against oxidative stress and apoptotic damage in experimental model of isoproterenol-induced cardiac toxicity in rats. *Phytomedicine*, 23(12), 1401-1408.
- Sudha, P., Asdaq, S. M. B., Dhamingi, S. S., & Chandrakala, G. K. (2010). Immunomodulatory activity of methanolic leaf extract of *M. oleifera* in animals.
- Sujatha, B., & Patel, P. (2017). *M. oleifera*—Nature's Gold. *Imperial Journal of Interdisciplinary Research*, 3(5).
- Sun, J. C., Ugolini, S., & Vivier, E. (2014). Immunological memory within the innate immune system. *The EMBO journal*, 33(12), 1295-1303.
- Sun, Z.-Y., Chen, P.-G., Liu, Y.-F., Shi, L., Zhang, B.-D., Wu, J.-J., . . . Li, Y.-M. (2017). Self-assembled Nano-immunostimulant for Synergistic Immune Activation. *ChemBioChem*.
- Sundrarajan, M., Jegatheeswaran, S., Selvam, S., Sanjeevi, N., & Balaji, M. (2015). The ionic liquid assisted green synthesis of hydroxyapatite nanoplates by *M. oleifera* flower extract: a biomimetic approach. *Materials & Design*, 88, 1183-1190.
- Sunila, E., & Kuttan, G. (2004). Immunomodulatory and antitumor activity of *Piper longum* Linn. and piperine. *Journal of ethnopharmacology*, 90(2), 339-346.
- Swain, S. L. (1995). T-Cell Subsets: Who does the polarizing? *Current Biology*, 5(8), 849-851.
- Tahiliani, P., & Kar, A. (2000). Role of *M. oleifera* leaf extract in the regulation of thyroid hormone status in adult male and female rats. *Pharmacological research*, 41(3), 319-323.
- Tan, W. S., Arulselvan, P., Karthivashan, G., & Fakurazi, S. (2015). *M. oleifera* flower extract suppresses the activation of inflammatory mediators in lipopolysaccharide-stimulated RAW 264.7 macrophages via NF-κB pathway. *Mediators of inflammation*, 2015.
- Teixeira, E. M. B., Carvalho, M. R. B., Neves, V. A., Silva, M. A., & Arantes-Pereira, L. (2014). Chemical characteristics and fractionation of proteins from *M. oleifera* Lam. leaves. *Food chemistry*, 147, 51-54.

- Tesmer, L. A., Lundy, S. K., Sarkar, S., & Fox, D. A. (2008). Th17 cells in human disease. *Immunological reviews*, 223(1), 87-113.
- Tiloke, C., & Chuturgoon, A. A. (2017). The Antiproliferative and Antibacterial Effect of M. oleifera-Mediated Gold Nanoparticles: A Review. In *Metabolic Engineering for Bioactive Compounds* (pp. 269-291): Springer.
- Tiloke, C., Phulukdaree, A., Anand, K., Gengan, R. M., & Chuturgoon, A. A. (2016). M. oleifera Gold Nanoparticles Modulate Oncogenes, Tumor Suppressor Genes, and Caspase-9 Splice Variants in A549 Cells. *Journal of cellular biochemistry*, 117(10), 2302-2314.
- Tiloke, C., Phulukdaree, A., & Chuturgoon, A. A. (2013). The antiproliferative effect of M. oleifera crude aqueous leaf extract on cancerous human alveolar epithelial cells. *BMC complementary and alternative medicine*, 13(1), 226.
- Tormo, M., Marugán, I., & Calabuig, M. (2010). Myelodysplastic syndromes: an update on molecular pathology. *Clinical and Translational Oncology*, 12(10), 652-661.
- Tragulpakseerojn, J., Yamaguchi, N., Pamonsinlapatham, P., Wetwitayaklung, P., Yoneyama, T., Ishikawa, N., Apirakaramwong, A. (2017). Anti-proliferative effect of M. oleifera Lam (Moringaceae) leaf extract on human colon cancer HCT116 cell line. *Tropical Journal of Pharmaceutical Research*, 16(2), 371-378.
- Tran, E., Turcotte, S., Gros, A., Robbins, P. F., Lu, Y.-C., Dudley, M. E., Hinrichs, C. S. (2014). Cancer immunotherapy based on mutation-specific CD4+ T cells in a patient with epithelial cancer. *Science*, 344(6184), 641-645.
- Trapani, J. A., & Smyth, M. J. (2002). Functional significance of the perforin/granzyme cell death pathway. *Nature reviews Immunology*, 2(10), 735-747.
- Treutter, D. (2006). Significance of flavonoids in plant resistance: a review. *Environmental Chemistry Letters*, 4(3), 147.
- Tsareva, E., Kulakova, O., Boyko, A., & Favorova, O. (2016). Pharmacogenetics of multiple sclerosis: personalized therapy with immunomodulatory drugs. *Pharmacogenetics and genomics*, 26(3), 103-115.
- Tsareva, E., Kulakova, O., Makarycheva, O., Boiko, A., Shchur, S., Lashch, N., L'vov, D. (2011). Pharmacogenomics of multiple sclerosis: association of immune response genes polymorphism with copaxone treatment efficacy. *Molekuliarnaia biologija*, 45(6), 963-972.
- Tsiantoulas, D., Diehl, C. J., Witztum, J. L., & Binder, C. J. (2014). B cells and humoral immunity in atherosclerosis. *Circulation research*, 114(11), 1743-1756.

- Tsoporis, J., Izhar, S., Salpeas, V., Rizos, I., & Parker, T. (2016). VEGF-induced myocardial cell anti-apoptotic events requires S100A6 signaling. *Canadian Journal of Cardiology*, 32(10), S142.
- Valigra, L. (1994). Engineering the future of antibiotics. *New Scientist*, 142(1923), 25-27.
- Van Dam, N. M., Tytgat, T. O., & Kirkegaard, J. A. (2009). Root and shoot glucosinolates: a comparison of their diversity, function and interactions in natural and managed ecosystems. *Phytochemistry Reviews*, 8(1), 171-186.
- Vasanth, K., Ilango, K., MohanKumar, R., Agrawal, A., & Dubey, G. P. (2014). Anticancer activity of *M. oleifera* mediated silver nanoparticles on human cervical carcinoma cells by apoptosis induction. *Colloids and surfaces B: Biointerfaces*, 117, 354-359.
- Vázquez-León, L., Páramo-Calderón, D., Robles-Olvera, V., Valdés-Rodríguez, O., Pérez-Vázquez, A., García-Alvarado, M., & Rodríguez-Jimenes, G. (2017). Variation in bioactive compounds and antiradical activity of *M. oleifera* leaves: influence of climatic factors, tree age, and soil parameters. *European Food Research and Technology*, 243(9), 1593-1608.
- Velaga, v. S. A. R., Suryadevara, N., Chee, I., & Ismail, N. E. (2017). Phytochemical analysis and immuno-modulatory effect of *M. oleifera* flowers.
- Venkatalakshmi, P., Vadivel, V., & Brindha, P. (2016). Role of phytochemicals as immunomodulatory agents: A review. *International Journal of Green Pharmacy* (Medknow Publications & Media Pvt. Ltd.), 10(1).
- Vergara-Jimenez, M., Almatrafi, M. M., & Fernandez, M. L. (2017). Bioactive components in *M. oleifera* leaves protect against chronic disease. *Antioxidants*, 6(4), 91.
- Verma, S., Singh, A., & Mishra, A. (2015). Complex disruption effect of natural polyphenols on Bcl-2-Bax: molecular dynamics simulation and essential dynamics study. *Journal of Biomolecular Structure and Dynamics*, 33(5), 1094-1106.
- Vernon-Wilson, E. F., Auradé, F., & Brown, S. B. (2006). CD31 promotes β 1 integrin-dependent engulfment of apoptotic Jurkat T lymphocytes opsonized for phagocytosis by fibronectin. *Journal of leukocyte biology*, 79(6), 1260-1267.
- Vignon, C., Debeissat, C., Georget, M.-T., Bouscary, D., Gyan, E., Rosset, P., & Herault, O. (2013). Flow cytometric quantification of all phases of the cell cycle and apoptosis in a two-color fluorescence plot. *PloS one*, 8(7), e68425.
- Vogelstein, B., & Kinzler, K. W. (2004). Cancer genes and the pathways they control. *Nature medicine*, 10(8), 789-799.

- Vongsak, B., Sithisarn, P., Mangmool, S., Thongpraditchote, S., Wongkrajang, Y., & Gritsanapan, W. (2013a). Maximizing total phenolics, total flavonoids contents and antioxidant activity of *M. oleifera* leaf extract by the appropriate extraction method. *Industrial Crops and Products*, 44, 566-571.
- Vongsak, B., Sithisarn, P., & Gritsanapan, W. (2013). Simultaneous HPLC quantitative analysis of active compounds in leaves of *M. oleifera* Lam. *Journal of chromatographic science*, 52(7), 641-645.
- Voskoboinik, I., Whisstock, J. C., & Trapani, J. A. (2015). Perforin and granzymes: function, dysfunction and human pathology. *Nature reviews Immunology*, 15(6), 388-400.
- Waldmann, T. A. (2017). Cytokines in Cancer Immunotherapy. *Cold Spring Harbor perspectives in biology*, a028472.
- Walker, E. H., Pacold, M. E., Perisic, O., Stephens, L., Hawkins, P. T., Wymann, M. P., & Williams, R. L. (2000). Structural determinants of phosphoinositide 3-kinase inhibition by wortmannin, LY294002, quercetin, myricetin, and staurosporine. *Molecular cell*, 6(4), 909-919.
- Wang, Q., Zhang, L., Yuan, X., Ou, Y., Zhu, X., Cheng, Z., Zhang, L. (2016). The relationship between the Bcl-2/Bax proteins and the mitochondria-mediated apoptosis pathway in the differentiation of adipose-derived stromal cells into neurons. *PloS one*, 11(10), e0163327.
- Wang, S., & El-Deiry, W. S. (2003). TRAIL and apoptosis induction by TNF-family death receptors. *Oncogene*, 22(53), 8628-8633.
- Wang, Y., Xu, P., Qiu, L., Zhang, M., Huang, Y., & Zheng, J. (2016). CXCR7 Participates in CXCL12-mediated Cell Cycle and Proliferation Regulation in Mouse Neural Progenitor Cells. *Current molecular medicine*, 16(8), 738-746.
- Wei, M. C., Zong, W.-X., Cheng, E. H.-Y., Lindsten, T., Panoutsakopoulou, V., Ross, A. J., . . . Korsmeyer, S. J. (2001). Proapoptotic BAX and BAK: a requisite gateway to mitochondrial dysfunction and death. *Science*, 292(5517), 727-730.
- Wiman, K., & Zhivotovsky, B. (2017). Understanding cell cycle and cell death regulation provides novel weapons against human diseases. *Journal of internal medicine*, 281(5), 483-495.
- Wojdyło, A., Oszmiański, J., & Czemerys, R. (2007). Antioxidant activity and phenolic compounds in 32 selected herbs. *Food chemistry*, 105(3), 940-949.
- Wolf, A. M. (2016). Aging and the Immune System. *Encyclopedia of Immunotoxicology*, 11-14.

- Wynn, T. A. (2015). Type 2 cytokines: mechanisms and therapeutic strategies. *Nature reviews Immunology*, 15(5), 271-282.
- Yan, Y., Mahotka, C., Heikaus, S., Shibata, T., Wethkamp, N., Liebmann, J., Gerharz, C. (2004). Disturbed balance of expression between XIAP and Smac/DIABLO during tumour progression in renal cell carcinomas. *British journal of cancer*, 91(7), 1349-1357.
- Yin, S.-Y., Yang, N.-S., & Lin, T.-J. (2017). Phytochemicals approach for developing cancer immunotherapeutics. *Frontiers in pharmacology*, 8, 386.
- Zhivotovsky, B., & Orrenius, S. (2001). Current concepts in cell death. *Current Protocols in Cell Biology*, 18.11. 11-18.11. 18.
- Zhu, Z., Cuss, S. M., Singh, V., Gurusamy, D., Shoe, J. L., Leighty, R., Hurwitz, A. A. (2015). CD4+ T cell help selectively enhances high-avidity tumor antigen-specific CD8+ T cells. *The Journal of Immunology*, 195(7), 3482-3489.
- Zoller, V., Funcke, J.-B., Keuper, M., El Hay, M. A., Debatin, K.-M., Wabitsch, M., & Fischer-Posovszky, P. (2016). TRAIL (TNF-related apoptosis-inducing ligand) inhibits human adipocyte differentiation via caspase-mediated downregulation of adipogenic transcription factors. *Cell death & disease*, 7(10), e2412.