



UNIVERSITI PUTRA MALAYSIA

ANTI-INVASIVE EFFECTS OF 2,6-BIS-(4-HYDROXYL-3METHOXYBENZYLIDINE) CYCLOHEXANONE AND ITS MOLECULAR TARGETS ASSOCIATED WITH INVADOPODIA FORMATION IN MDA-MB-231 HUMAN BREAST CANCER CELLS

SITI NOR AINI BINTI HARUN

FPSK(M) 2018 21



ANTI-INVASIVE EFFECTS OF 2,6-BIS-(4-HYDROXYL-3METHOXYBENZYLIDINE) CYCLOHEXANONE AND ITS MOLECULAR TARGETS ASSOCIATED WITH INVADOPIA FORMATION IN MDA-MB-231 HUMAN BREAST CANCER CELLS

By

SITI NOR AINI BINTI HARUN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

May 2018

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

ANTI-INVASIVE EFFECTS OF 2,6-BIS-(4-HYDROXYL-3METHOXYBENZYLIDINE) CYCLOHEXANONE AND ITS MOLECULAR TARGETS ASSOCIATED WITH INVADOPODIA FORMATION IN MDA-MB-231 HUMAN BREAST CANCER CELLS

By

SITI NOR AINI BINTI HARUN

May 2018

Chair : Nur Fariesha binti Md Hashim, PhD
Faculty : Medicine and Health Sciences

Cancer metastasis is a pathological event occurred in cancer patients where it can colonize the distant organs. In order to metastasize, the tumor cells need to migrate and invade the surrounding tissues. Invadopodia are thought to be specialized actin-rich protrusions formed by the highly invasive cells to degrade the dense meshwork of the extracellular matrix (ECM). This is the initial step employed to drive cancer invasion. Cancer metastasis is deadliest and has affected the survival rate of cancer patients. Current cancer treatment has also produced side effects. Chemotherapy has given rise to the invasion and metastasis while radiotherapy has caused the recurrence of cancer. Identification of compound(s) capable to disrupt the metastasis of cancer especially for hindering the invadopodia formation is important so as to provide anti-metastasis targeted therapy. Curcumin has been demonstrated to produce significant effect as an anti-cancer compound. However, due to its poor bioavailability, some analogues have been formulated. A curcuminoid analogue known as 2,6-bis-(4-hydroxyl-3methoxybenzylidene) cyclohexanone or BHMC has shown good potential in inhibiting inflammation and hyperalgesia. It also possesses anti-tumor effects on 4T1 murine breast cancer cells *in vivo*. However, there is still lack of empirical evidence on how BHMC works in preventing human breast cancer invasion. In this study, we pursued to investigate the role of BHMC on MDA-MB-231 breast cancer cells its underlying mechanism of action to prevent breast cancer invasion especially on the formation of invadopodia.

Analysis revealed that treatment of BHMC at 12.5 μM and below did not interfere with the proliferation of MDA-MB-231 cells. By using scratch migration assay, transwell migration and invasion assays, we found that BHMC at 12.5 μM reduces the percentage of the migration and invasion of MDA-MB-231 cells. The gelatin degradation assay showed that BHMC reduces the number of cells forming invadopodia. Analysis of the proteins involved in invasion showed that there is significant reduction in the expression of Rho guanine nucleotide exchange factor 7 (β -PIX), matrix metalloproteinase-9 (MMP-9) and membrane type 1-MMP (MT1-MMP) in the present of BHMC treatment at 12.5 μM . It can be postulated that BHMC at 12.5 μM is the optimal concentration to prevent the invasiveness of breast cancer cells.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Master Sains

**KESAN ANTI PENYERANGAN 2,6-BIS-(4-HYDROXYL-
3METHOXYBENZYLIDINE) CYCLOHEXANONE DAN SASARAN MOLEKUL
BERKAITAN PEMBENTUKAN INVADOPODIA TERHADAP SEL KANSER
PAYUDARA MDA-MB-231**

Oleh

SITI NOR AINI BINTI HARUN

Mei 2018

Chair : Nur Fariesha binti Md Hashim, PhD
Faculty : Perubatan dan Sains Kesihatan

Metastasis (Perebakan) kanser merupakan satu keadaan patologi yang berlaku kepada pesakit kanser yang mana penyakit tersebut mengkoloni organ-organ yang lain. Sel-sel kanser perlu bermigrasi dan menyerang tisu-tisu sekeliling untuk merebak. Invadopodia merupakan benjolan yang kaya dengan actin dibentuk oleh sel-sel kanser yang tinggi daya serang ini untuk mendegradasi matriks luar sel. Pembentukan ini merupakan langkah pertama berlakunya penyerangan kanser. Perebakan kanser boleh menyebabkan kematian dan telah memberi kesan terhadap jangka hayat pesakit kanser. Rawatan kanser terkini juga menyebabkan kesan-kesan sampingan. Kemoterapi telah menyebabkan berlakunya perebakan kanser manakala radioterapi telah menyebabkan pertumbuhan semula kanser. Proses mengenal pasti suatu kompoun yang boleh bertindak menghalang perebakan sel-sel kanser terutama untuk membantutkan pembentukan invadopodia sangat penting untuk menghasilkan terapi anti-metastasis. Kurkumin telah dilaporkan mempunyai kesan yang sangat memberansangkan sebagai ubat anti-kanser. Walau bagaimanapun, disebabkan oleh tahap ketersediaan bio kurkumin yang rendah, beberapa analog kurkumin telah diformulasi. Satu analog kurkumin yang dikenali sebagai 2,6-bis-(4-hydroxyl-3-methoxybenzylidene) cyclohexanone (BHMC) telah menunjukkan potensi yang baik untuk menghalang sitokine keradangan dan hyperalgesia. Ia juga mempunyai kesan anti-kanser terhadap sel kanser payudara murin 4T1 *in vivo*. Walaupun begitu, bukti empirikal mengenai tindak balas BHMC untuk menghalang penyerangan kanser payudara masih lagi kurang. Justeru itu, kami mengkaji kesan BHMC terhadap sel kanser payudara MDA-MB-231 dan mekanisme asas BHMC untuk menghalang penyerangan sel kanser terutama terhadap pembentukan invadopodia. Analisis daripada kajian ini menunjukkan

perawatan BHMC pada kepekatan 12.5 μM dan ke bawah tidak memberikan kesan terhadap tahap pertumbuhan sel MDA-MB-231. Walaupun begitu, eksperimen pergerakan migrasi dan esei migrasi dan invasi transwell, BHMC pada kepekatan 12.5 μM mengurangkan migrasi dan invasi sel MDA-MB-231. Esei degradasi gelatin pula menunjukkan BHMC mengurangkan bilangan sel yang membentuk invadopodia. Analisis kesan BHMC terhadap protein yang berkaitan penyerangan kanser pula menunjukkan BHMC mengurangkan ekspresi Rho guanine nucleotide exchange factor 7 (β -PIX), matrix metalloproteinase-9 (MMP-9) dan membrane type 1-MMP (MT1-MMP) pada kadar kepekatan 12.5 μM dan menyimpulkan bahawa BHMC pada kepekatan 12.5 μM merupakan kepekatan optimum untuk menghalang penyerangan sel kanser payudara.



ACKNOWLEDGEMENTS

Deepest gratitude and praise to the Almighty Allah S.W.T. for His permission I am able to work on my research and write this thesis completely. It would not be possible to write this thesis without the help and support of the people around me.

In particular, I would like to express my sincere appreciation to my supervisor, Dr. Nur Fariesha Md Hashim. Her encouragement, patience, guidance and support have enabled me to develop the understanding of the subject and overcome the hardship to complete my research and thesis successfully. The good advice and intellectual inputs of my co-supervisors, Prof. Dr. Daud Ahmad Israf Ali, Dr. Tham Chau Ling and Dr. Manraj Singh Cheema, have been invaluable.

Special thanks also to Mr. Zulkhairi Zainol and Mrs. Nora Asyikin Mohd Salim for their technical assistance in immunofluorescence microscopy and cell culture technique.

I would like to acknowledge the Ministry of Education, Malaysia that provided the necessary financial support under Fundamental Research Grant Scheme (FRGS) and also to University Putra Malaysia (UPM) for the Graduate Research Fellowship (GRF) award.

To my colleagues especially to Hui Min, Farizatul, Hafizan, Wafda, Aida, Audrey, Athirah, Nabilah and Mr. Kelvin Lee whom are very assuring in their encouragement and support. Also for the knowledge shared and the precious value of friendship make me heartily thankful. Exceptional thank you also to my two undergraduate brothers Izwan and Lih Sern whom are very cooperative and supportive to keep me going through the project until the end. Your assistance and support will be kept in my heart endlessly.

A million thank you to my parents, Fatimah Awi and Harun Yusof who have raised me with love and for my family for supporting me. I found my sanctuary from their utmost support.

Lastly, I offer my regards and blessings to all of those who supported me in any respect during the completion of the project. Allah bless all of you.

I certify that a Thesis Examination Committee has met on 14 May 2018 to conduct the final examination of Siti Nor Aini binti Harun on her thesis entitled "Anti-Invasive Effects of 2,6-Bis-(4-Hydroxyl-3methoxybenzylidene) Cyclohexanone and Its Molecular Targets Associated with Invadopodia Formation in MDA-MB-231 Human Breast Cancer Cells" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Suhaili binti Abu Bakar @ Jamaludin, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Yong Yoke Keong, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Radiyah Abdul Ghani, PhD

Assistant Professor
International Islamic University Malaysia
Malaysia
(External Examiner)



RUSLI HAJI ABDULLAH, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 30 August 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Nur Fariesha binti Md Hashim, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Daud Ahmad bin Israf Ali, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Tham Chau Ling, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

Manraj Singh Cheema, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Siti Nor Aini binti Harun (GS42279)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Nur Fariesha Md Hashim

Signature: _____
Name of Member of
Supervisory
Committee: Professor Daud Ahmad Israf Ali

Signature: _____
Name of Member of
Supervisory
Committee: Tham Chau Ling

Signature: _____
Name of Member of
Supervisory
Committee: Manraj Singh Cheema

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLEDGEMENTS		v
APPROVAL		vi
DECLARATION		viii
LIST OF TABLES		xii
LIST OF FIGURES		xiii
LIST OF ABBREVIATIONS		xiv
CHAPTER		
1	INTRODUCTION	1
	1.1 Background of Study	1
	1.2 Problem Statement	1
	1.3 Objectives	2
	1.4 Hypothesis	2
2	LITERATURE REVIEW	3
	2.1 Metastatic Breast Cancer	3
	2.2 Metastatic Cascade	3
	2.3 Migration and Invasion	5
	2.4 Factors That Drive Metastasis	7
	2.4.1 Growth Factors	7
	2.4.2 Tumor Hypoxia	7
	2.5 Invadopodia	8
	2.6 Invadopodia and Their Components	10
	2.6.1 Protein for Actin Regulations	10
	2.6.2 Signaling Proteins	11
	2.6.3 Proteases	12
	2.7 Curcumin	12
	2.8 BHMC	14
3	MATERIALS AND METHODOLOGY	16
	3.1 Materials	16
	3.1.1 General Reagents	16
	3.1.2 Antibodies	17
	3.1.3 General Buffers and Solution	18
	3.1.4 Cell Line	18
	3.1.5 BHMC	18
	3.2 Methodology	19
	3.2.1 Cell Culture and Maintenance	19
	3.2.2 Cryopreservation and Cell Thawing	19
	3.2.3 MTT Assay	19
	3.2.4 Proliferation Assay	20
	3.2.5 Scratch Migration Assay	20

	3.2.6	Transwell Migration and Invasion Assays	21
	3.2.7	Invadopodia Assay	21
	3.2.8	Cell Lysis and Protein Quantification	22
	3.2.9	Non-disease and Drug Control Group	23
	3.2.10	Immunoblotting	23
	3.2.11	Statistical Analysis	24
4		RESULTS AND DISCUSSION	25
	4.1	Cytotoxicity and Anti-proliferative Effects of BHMC on MDA-MB-231 Cells	25
	4.1.1	Results	25
	4.1.2	Discussion	29
	4.2	Inhibition of BHMC on Migration and Invasion of MDA-MB-231 Cells	31
	4.2.1	Results	31
	4.2.2	Discussion	35
	4.3	BHMC Has an Effect on Number of Cells Forming Invadopodia	37
	4.3.1	Results	37
	4.3.2	Discussion	40
	4.4	Inhibition of β -PIX, MMP-9 and MT1-MMP Protein Expression on MDA-MB-231 Cells upon BHMC Treatment	42
	4.4.1	Results	42
	4.4.2	Discussion	46
5		SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	49
	5.1	Summary and Conclusion	49
	5.2	Recommendations for Future Direction	50
		REFERENCES	52
		APPENDICES	62
		BIODATA OF STUDENT	63
		LIST OF PUBLICATIONS	64

LIST OF TABLES

Table		Page
3.1	General Reagents	16
3.2	Primary Antibodies	17
3.3	Secondary Antibodies	17
3.4	General Buffers and Solutions	18



LIST OF FIGURES

Figure		Page
2.1	The Main Steps in Cancer Metastasis	5
2.2	Invadopodia Formation Formed by Invasive Cancer Cells that Extend Into Matrix Substratum in 2D Gelatin Degradation Assay	10
2.3	Chemical Structure of BHMC and Curcumin	14
4.1	Cell Viability Assay	26
4.2	Representative Images of PCNA Antibody Staining on MDA-MB-231 Cells	27
4.3	Graph Plotted to Represent Presence of PCNA in 150 MDA-MB-231 Cells	28
4.4	Effects of BHMC on Migration of MDA-MB-231 Cells in Scratch Migration Assay	32
4.5	Effects of BHMC on MDA-MB-231 Cell Migration in Transwell Migration Assay	33
4.6	Effects of BHMC on MDA-MB-231 Cell Invasion Assay	34
4.7	Representative Images of Invadopodia Formation in MDA-MB-231 Cells	38
4.8	Effects of BHMC on Gelatin Degradation Assay	39
4.9	Effects of BHMC on β -PIX Expression Level	43
4.10	Effects of BHMC on MMP-9 Expression Level	44
4.11	Effects of BHMC on MT1-MMP Expression Level	45
5.1	Candidate Molecules Affected by BHMC Treatment	51

LIST OF ABBREVIATIONS

2D	Two-dimensional
3D	Three-dimensional
β-arr	β-arrestin
ADAM	A disintegrin and metalloproteinase
ANGPTL	Angiotensin-like
ANOVA	Analysis of variance
AP 1	Activator protein 1
Arf 6	ADP-ribosylation factor 6
Arp	Actin-related protein
ATP	Adenosine triphosphate
b-FGF	Basic fibroblast growth factor
BCA	Bicinchoninic acid
BHMC	2,6-bis-(4-hydroxy-3-methoxybenzylidene) cyclohexanone
BRCA1	Breast cancer gene 1
BRCA2	Breast cancer gene 2
BSA	Bovine serum albumin
BSC	Bio safety cabinet
CD63	CD63 antigen
CD9	CD9 antigen
Cdc42	Cell division cycle 42
CIP	Cdc42-interacting protein
CLP	Caecal-ligation puncture
COPD	Chronic Obstructive Pulmonary Disease
CRM	Chromosome region maintenance
CSF	Colony stimulating factor
DMEM	Dulbecco's modified eagle's medium
DMOG	Dimethylxaloylglycine
DMSO	Dimethylsulfoxide
DNA	Deoxyribonucleic acid
DNA	Deoxyribonucleic acid
ECL	Enhanced chemiluminescent
ECM	Extracellular matrix
EDTA	Ethylenediaminetetraacetic acid
EGF	Epidermal growth factor
EGFR	Epidermal growth factor receptor
EMT	Epithelial mesenchymal transition
EOC	Epithelial ovarian cancer
ER/PR	Estrogen/progesterone receptor
ERRB2	Epidermal growth factor receptor 2
ET	Endothelin
EV	Extracellular vesicles
FA	Focal adhesion
FAK	Focal adhesion kinase
F-BAR	Fer/CIP4 homology-Bin/Amphiphysin/Rvs
FBS	Fetal bovine serum
GAPDH	Glyceraldehyde 3-phosphate dehydrogenase
GDP	Guanosine diphosphate

GLUT	Glucose transporter
GTP	Guanosine triphosphate
HB	Heparin binding
HER2	Human epidermal growth factor receptor 2
HGF	Hepatocyte growth factor
HIF	Hypoxia inducible factor
HNSCC	Head and neck squamous cell carcinoma
HPLC	High performance liquid chromatography
HR	Homology region
HRP	Horseradish peroxidase
IGF	Insulin-like growth factor
IL	Interleukin
kDa	Kilo Dalton
LPS	Lipopolysaccharide
MAPK	p-38 and mitogen-activated protein kinase
MCP	Monocyte chemotactic protein
MLCK	Myosin like-chain kinase
MMP	Matrix metalloproteinase
MT1-MMP	Membrane type-1 matrix metalloproteinase
mTOR	Mammalian target of rapamycin
MTT	3-(4,5-Dimethylthiazol-2-yl)-2,5 diaphenyltetrazolium bromide
NF- κ B	Nuclear factor κ B
NHE	Sodium/hydrogen exchanger
NIC	Notch 1 fragment
N-WASP	Neural-Wiskott Aldrich Syndrome
PBS	Phosphate buffer saline
PCNA	Proliferating cell nuclear antigen
PDGF	Platelet-derived growth factor
PDZ-RhoGEF	postsynaptic density protein 95/disc-large/zonula occludens-RhoGEF
PI3K	Phosphoinositide 3-kinase
PIX	PAK-interacting exchange factor
PKC	Protein kinase C
PTEN	Phosphatase and tensin
PVDF	Polyvinylidene difluoride
RIPA	Radioimmunoprecipitation assay
ROCK	Rho-associated coiled-forming kinase
RSV	Rous sarcoma virus
SDF	Stromal cell derived factor
SDS	Sodium dodecyl sulphate
SDS-PAGE	SDS-polyacrylamide gel electrophoresis
SEM	Standard error of mean
SH	Sulfhydryl
TGF- α	Transforming growth factor α
TGF- β	Transforming growth factor β
TIMP	Tissue inhibitor of metalloproteinase
TLR	Toll-like receptor
TNBC	Triple negative breast cancer
TNF	Tumor necrosis factor
TNF	Tumor necrosis factor

TOM1L	Target of MYB1-like protein
uPAR	Urokinase plasminogen activator receptor
US FDA	United State Food and Drug Administration
VCA	Verprolin-cofilin-acidic
VEGF	Vascular endothelial growth factor
VHL	von Hippel-Lindau
WAVE	WASP family verprolin-homologous protein
WIP	WASP-interacting protein



© COPYRIGHT UPM

CHAPTER 1

INTRODUCTION

1.1 Background of Study

Breast cancer is a form of disorder in which the cancer cells can grow either at the lobular, medullary, ductal or sebaceous of the breast tissue. The causes of breast cancer include the mutations of the genes such as epidermal growth factor receptor 2 (ERBB2) (also known as human epidermal growth factor receptor 2 (HER2)), breast cancer 1 (BRCA1) and breast cancer 2 (BRCA2) (Lukong, 2017). There are six hallmarks of cancer that have been proposed which encompass of the ability of the cancer cells to sustain the proliferative signaling, resist cell death, induce angiogenesis, evade the growth suppressors, enable the replicative immortality and activate invasive metastasis (Hanahan & Weinberg, 2011). Most of the patients whom endure breast cancer are likely to face mortality once the cancer cells have metastasized to other parts of body. Triple negative breast cancer patients, whom are clinically having less or absent of HER2 receptor (-HER2) and estrogen/progesterone receptor (-ER/PR) are the highly invasive cancer incidence among the patients (Bianchini et al., 2016). These types of patients are having increased likelihood of distant recurrence and believed to be more aggressive compared to other types of breast cancer (Bianchini et al., 2016).

1.2 Problem Statement

Metastatic breast cancer is the major problem in the population as it can affect the survival of breast cancer patients (Lukong, 2017). The ability of the tumor cells to disseminate to other parts of the body such as lungs, brain and bones and only can be diagnosed at an advanced stage has been troublesome as the tumor cells cannot be eradicated totally from the body thus causing the recurrence of cancer (Scully et al., 2012). Once the invasive cancer cells have metastasized, the treatment to reduce the growth of cancer would be challenging as many cancer genes have been up-regulated and also reduce the sensitivity of cancer cells to the treatment (Massagué et al., 2017). There are several treatments used to treat breast cancer including the application of surgery, radio- and chemotherapy. radio- and chemotherapy (Lukong, 2017). However, besides killing the tumor cells, especially radio- and chemotherapy, both radio- and chemotherapy have caused side effects to the patients such as thrombocytopenia, diarrhea, infertility and others (Lukong, 2017).

Curcumin, a diferuloylmethane is a potential compound to target and modulate many proteins linked to cancer (Hasima & Aggarwal, 2012). Curcumin has been reported to have entered the clinical trials but the poor bioavailability

problem that curcumin had has been questioned (Anand et al., 2007). Having promising abilities to target multiple proteins related to cancer has urged the researchers to modify the structure of curcumin but still preserving the methyl and hydroxyl group which are responsible for the anti-proliferation and anti-oxidant properties of curcumin (Nagahama et al., 2016). One of the analogues of curcumin which is 2,6-bis-(4-hydroxyl-3methoxybenzylidene) cyclohexanone or BHMC has been studied in inflammation and hyperalgesic pain (Ming-Tatt et al., 2013; Tham et al., 2010). Recently, BHMC has shown to possess anti-tumor activity in 4T1 murine breast cancer (Razak et al., 2017); . However, BHMC is yet to be studied in human breast cancer. It will be interesting to investigate the effects of BHMC on the migration and invasion of MDA-MB-231 human breast cancer cells and specifically in the formation of invadopodia and their targeted proteins.

1.3 Objectives

This research was conducted to investigate the anti-invasive effects of BHMC and its molecular target(s) on MDA-MB-231 human breast cancer cells.

The specific objectives of the study include the following:

1. To determine the effect of BHMC on the proliferation of MDA-MB-231 human breast cancer cells
2. To determine the effect of BHMC on the migration and invasion of MDA-MB-231 human breast cancer cells
3. To determine the effect of BHMC on the formation of invadopodia in MDA-MB-231 human breast cancer cells
4. To identify the effect of BHMC on expressions of the proteins involved in invasion and invadopodia such as MMP-9, MT1-MMP and β -PIX

1.4 Hypothesis

It is hypothesized that BHMC at non-cytotoxic concentrations will reduce the migration and invasion of MDA-MB-231 human breast cancer cells. In addition, it is expected that BHMC would reduce the number of cells forming invadopodia via down-regulation of MMP-9, MT1-MMP and β -PIX expressions.

REFERENCES

- Ahn, S. J., Chung, K. W., Lee, R. A., Park, I. A., Lee, S. H., Park, D. E., & Noh, D. Y. (2003). Overexpression of β Pix-a in human breast cancer tissues. *Cancer Letters*, 193(1): 99–107.
- An, T., Qin, S., Xu, Y., Tang, Y., Huang, Y., Situ, B., Inai, J. M., & Zheng, L. (2015). Exosomes serve as tumor markers for personalized diagnostics owing to their important role in cancer metastasis. *Journal of Extracellular Vesicles*, 4(1): 1-16.
- Anand, M., Van Meter, T. E., & Fillmore, H. L. (2011). Epidermal growth factor induces matrix metalloproteinase-1 (MMP-1) expression and invasion in glioma cell lines via the MAPK pathway. *Journal of Neuro-Oncology*, 104(3): 679–687.
- Anand, P., Kunnumakkara, A. B., Newman, R. A., & Aggarwal, B. B. (2007). Bioavailability of curcumin: Problems and promises. *Molecular Pharmaceutics*, 4(6): 807-818.
- Artym, V. V., Zhang, Y., Seillier-Moisewitsch, F., Yamada, K. M., & Mueller, S. C. (2006). Dynamic interactions of cortactin and membrane type 1 matrix metalloproteinase at invadopodia: Defining the stages of invadopodia formation and function. *Cancer Research*, 66(6): 3034–3043.
- Azizah, A. M., Ibrahim, N. S., & Abdullah, N. H. (2015). *Malaysian National Cancer Registry Report 2007-2011*. Ministry of Health Malaysia.
- Baldassarre, M., Pompeo, A., Beznoussenko, G., Castaldi, C., Cortellino, S., McNiven, M. A., Luini, R., & Buccione, R. (2003). Dynamin Participates in Focal Extracellular Matrix Degradation by Invasive Cells. *Molecular Biology of the Cell*, 14(3): 1074–1084.
- Bianchini, G., Balko, J. M., Mayer, I. A., Sanders, M. E., & Gianni, L. (2016). Triple-negative breast cancer: Challenges and opportunities of a heterogeneous disease. *Nature Reviews Clinical Oncology*, 13(11): 674-690.
- Bravo-Cordero, J. J., Hodgson, L., & Condeelis, J. (2012). Directed cell invasion and migration during metastasis. *Current Opinion in Cell Biology*, 24(2): 277-283.
- Castro-Castro, A., Marchesin, V., Monteiro, P., Lodillinsky, C., Rossé, C., & Chavrier, P. (2016). Cellular and Molecular Mechanisms of MT1-MMP-Dependent Cancer Cell Invasion. *Annual Review of Cell and Developmental Biology*, 32: 555-576.
- Chen, B., Zhang, Y., Wang, Y., Rao, J., Jiang, X., & Xu, Z. (2014). Curcumin inhibits proliferation of breast cancer cells through Nrf2-mediated down-regulation of Fen1 expression. *Journal of Steroid Biochemistry and Molecular Biology*, 143: 11–18.

- Chen, Q. Y., Jiao, D. M., Yao, Q. H., Yan, J., Song, J., Chen, F. Y., Lu, G. H. & Zhou, J. Y. (2012). Expression analysis of Cdc42 in lung cancer and modulation of its expression by curcumin in lung cancer cell lines. *International Journal of Oncology*, 40(5), 1561–1568.
- Chen, Q., Zheng, Y., Jiao, D., Chen, F., Hu, H., Wu, Y., Song, J., Yan, J, Wu, L. J. & Lv, G. (2014). Curcumin inhibits lung cancer cell migration and invasion through Rac1-dependent signaling pathway. *The Journal of Nutritional Biochemistry*, 25(2): 177–185.
- Chen, W. -T. (1989). Proteolytic activity of specialized surface protrusions formed at rosette contact sites of transformed cells. *Journal of Experimental Zoology*, 251(2): 167–185.
- Chen, W. T. (1996). Proteases associated with invadopodia, and their role in degradation of extracellular matrix. *Enzyme and Protein*, 49(1–3): 59–71.
- Chen, Y. (2013). Scratch Wound Healing Assay. *Journal of Chemical Information and Modeling*, 53(1): 1689–1699.
- Cheung, K. J., & Ewald, A. J. (2016). A collective route to metastasis: Seeding by tumor cell clusters. *Science*, 352(6282): 167-169.
- Chevalier, C., Collin, G., Descamps, S., Touaitahuata, H., Simon, V., Reymond, N., Fernandez, L., Milhiet, P. E., Georget, V., Urbach, S., Lasorsa, L., Orsetti, B., Boissiere-Michot, F., Lopez-Crapez, E., Theillet, C., Roche, S. & Benistant, C. (2016). TOM1L1 drives membrane delivery of MT1-MMP to promote ERBB2-induced breast cancer cell invasion. *Nature Communications*, 7: 1-16.
- Chhabra, E. S., & Higgs, H. N. (2007). The many faces of actin: Matching assembly factors with cellular structures. *Nature Cell Biology*, 9(10): 1110-1121.
- Chiu, T.-L., & Chin-Cheng, S. (2009). Curcumin inhibits proliferation and migration by increasing the Bax to Bcl-2 ratio and decreasing NF-kBp65 expression in breast cancer MDA-MB-231 cells. *International Journal of Molecular Medicine*, 23: 469–475.
- Choi, H., Chun, Y. S., Kim, S. W., Kim, M. S., & Park, J. W. (2006). Curcumin inhibits hypoxia-inducible factor-1 by degrading aryl hydrocarbon receptor nuclear translocator: a mechanism of tumor growth inhibition. *Molecular Pharmacology*, 70(5): 1664–1671.
- Chou, J., Wang, B., Zheng, T., Li, X., Zheng, L., Hu, J., Zhang, Y., Xing, Y. & Xi, T. (2016). MALAT1 induced migration and invasion of human breast cancer cells by competitively binding MIR-1 with cdc42. *Biochemical and Biophysical Research Communications*, 472(1): 262–269.
- Clark, A. G., & Vignjevic, D. M. (2015). Modes of cancer cell invasion and the role of the microenvironment. *Current Opinion in Cell Biology*, 36: 13-22.

- Clark, E. S., Whigham, A. S., Yarbrough, W. G., & Weaver, A. M. (2007). Cortactin is an essential regulator of matrix metalloproteinase secretion and extracellular matrix degradation in invadopodia. *Cancer Research*, 67(9): 4227–4235.
- Cowden Dahl, K. D., Symowicz, J., Ning, Y., Gutierrez, E., Fishman, D. A., Adley, B. P., Stack, M. S., & Hudson, L. G. (2008). Matrix metalloproteinase 9 is a mediator of epidermal growth factor-dependent E-cadherin loss in ovarian carcinoma cells. *Cancer Research*, 68(12): 4606–4613.
- David-Pfeuty, T., & Singer, S. J. (1980). Altered distributions of the cytoskeletal proteins vinculin and alpha-actinin in cultured fibroblasts transformed by Rous sarcoma virus. *Proceedings of the National Academy of Sciences of the United States of America*, 77(11): 6687–6691.
- Di, D., Chen, L. E. I., Guo, Y., Wang, L., Wang, H., & Ju, J. (2018). Association of BCSC-1 and MMP-14 with human breast cancer. *Oncology Letters*, 15(4): 5020–5026.
- Díaz, B., Yuen, A., Iizuka, S., Higashiyama, S., & Courtneidge, S. A. (2013). Notch increases the shedding of HB-EGF by ADAM12 to potentiate invadopodia formation in hypoxia. *Journal of Cell Biology*, 201(2): 279–292.
- Doyle, A. D., Petrie, R. J., Kutys, M. L., & Yamada, K. M. (2013). Dimensions in cell migration. *Current Opinion in Cell Biology*, 25(5): 642-649.
- Diaz, B. (2013). Invadopodia Detection and Gelatin Degradation Assay. *Bio-Protocol*, 3(24): 1-8.
- Eddy, R. J., Weidmann, M. D., Sharma, V. P., & Condeelis, J. S. (2017). Tumor Cell Invadopodia: Invasive Protrusions that Orchestrate Metastasis. *Trends in Cell Biology*, 27(8): 595-607.
- Feng, Q., Baird, D., Peng, X., Wang, J., Ly, T., Guan, J. L., & Cerione, R. A. (2006). Cool-1 functions as an essential regulatory node for EGF receptor- and Src-mediated cell growth. *Nature Cell Biology*, 8(9): 945–956.
- Ferlay, J., Soerjomataram, I., Ervik, M., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D. M., Forman, D. & Bray, F. (2012). World Cancer Research Fund International Worldwide Data. *World Cancer Research Fund*. Retrieved 1 August, 2017 from <https://www.wcrf.org/int/cancer-facts-figures/worldwide-data>.
- Fidler, I. J. (2002). The organ microenvironment and cancer metastasis. *Differentiation*, 70(9-10): 498-505.
- Fu, H., Wu, R., Li, Y., Zhang, L., Tang, X., Tu, J., Zhou, W., Wang, J., & Shou, Q. (2016). Safflower Yellow Prevents Pulmonary Metastasis of Breast Cancer by Inhibiting Tumor Cell Invadopodia. *The American Journal of*

Chinese Medicine, 44(7): 1491–1506.

Gialeli, C., Theocharis, A. D., & Karamanos, N. K. (2011). Roles of matrix metalloproteinases in cancer progression and their pharmacological targeting. *FEBS Journal*, 278(1): 16-27.

Guan, F., Ding, Y., Zhang, Y., Zhou, Y., Li, M., & Wang, C. (2016). Curcumin suppresses proliferation and migration of MDA-MB-231 breast cancer cells through autophagy-dependent Akt degradation. *Plos One*, 11(1): 1-12.

Gupta, S. C., Patchva, S., Koh, W., & Aggarwal, B. B. (2012). Discovery of curcumin, a component of golden spice, and its miraculous biological activities. *Clinical and Experimental Pharmacology and Physiology*, 39(3): 283–299.

Hanahan, D., & Weinberg, R. A. (2011). Hallmarks of cancer: The next generation. *Cell*, 144(5): 646-674.

Harada, H. (2011). How Can We Overcome Tumor Hypoxia in Radiation Therapy? *Journal of Radiation Research*, 52(5): 545–556.

Hasima, N., & Aggarwal, B. B. (2012). Cancer-linked targets modulated by curcumin. *International Journal of Biochemistry and Molecular Biology*, 3(4): 328-351.

Hoshino, D., Branch, K. M., & Weaver, A. M. (2013). Signaling inputs to invadopodia and podosomes. *Journal of Cell Science*, 126(14): 2979–2989.

Hurst, D. R., & Welch, D. R. (2011a). Metastasis suppressor genes. At the interface between the environment and tumor cell growth. *International Review of Cell and Molecular Biology*, 286(C): 107–180.

Ichikawa, K. (2015). Synergistic effect of blocking cancer cell invasion revealed by computer simulations. *Mathematical Biosciences and Engineering: MBE*, 12(6): 1189–1202.

Justus, C. R., Leffler, N., Ruiz-Echevarria, M., & Yang, L. V. (2014). In vitro Cell Migration and Invasion Assays. *Journal of Visualized Experiments*, 88: 1-8.

Klein, T., & Bischoff, R. (2011). Physiology and pathophysiology of matrix metalloproteases. *Amino Acids*, 41(2): 271-290.

Koo, H. J., Shin, S., Choi, J. Y., Lee, K. H., Kim, B. T., & Choe, Y. S. (2015). Introduction of Methyl Groups at C2 and C6 Positions Enhances the Antiangiogenesis Activity of Curcumin. *Scientific Reports*, 5(14205): 1-12.

Krausz, A. E., Adler, B. L., Cabral, V., Navati, M., Doerner, J., Charafeddine, R. A., Chandra, D., Liang, H., Gunther, L., Clendaniel A., Harper, S, Friedman, J. M., Nosanchuk, J. D. & Friedman, A. J. (2015). Curcumin-

- encapsulated nanoparticles as innovative antimicrobial and wound healing agent. *Nanomedicine: Nanotechnology, Biology, and Medicine*, 11(1): 195–206.
- Kuo, J. C., Han, X., Hsiao, C. Te, Yates, J. R., & Waterman, C. M. (2011). Analysis of the myosin-II-responsive focal adhesion proteome reveals a role for β -Pix in negative regulation of focal adhesion maturation. *Nature Cell Biology*, 13(4): 383–395.
- Lam, K. W., Tham, C. L., Liew, C. Y., Syahida, A., Rahman, M. B. A., Israf, D. A., & Lajis, N. H. (2012). Synthesis and evaluation of DPPH and anti-inflammatory activities of 2,6-bisbenzylidenecyclohexanone and pyrazoline derivatives. *Medicinal Chemistry Research*, 21: 333-344.
- Langley, R. R., & Fidler, I. J. (2011). The seed and soil hypothesis revisited—The role of tumor-stroma interactions in metastasis to different organs. *International Journal of Cancer*, 128(11): 2527–2535.
- Lee, A. Y. L., Fan, C. C., Chen, Y. A., Cheng, C. W., Sung, Y. J., Hsu, C. P., & Kao, T. Y. (2015). Curcumin Inhibits Invasiveness and Epithelial-Mesenchymal Transition in Oral Squamous Cell Carcinoma Through Reducing Matrix Metalloproteinase 2, 9 and Modulating p53-E-Cadherin Pathway. *Integrative Cancer Therapies*, 14(5): 484–490.
- Lee, W. H., Loo, C. Y., Young, P. M., Rohanizadeh, R., & Traini, D. (2016). Curcumin Nanoparticles Attenuate Production of Pro-inflammatory Markers in Lipopolysaccharide-Induced Macrophages. *Pharmaceutical Research*, 33(2): 315–327.
- Li, Y.-Y., Zhou, C.-X., & Gao, Y. (2015). Podoplanin promotes the invasion of oral squamous cell carcinoma in coordination with MT1-MMP and Rho GTPases. *American Journal of Cancer Research*, 5(2): 514–529.
- Li, Y. Q., Yan, J. P., Xu, W. L., Wang, H., Xia, Y. J., Wang, H. J., Zhu, Y. Y., & Huang, X. J. (2013). ADAM17 mediates MMP9 expression in lung epithelial cells. *Plos One*, 8(1): 1-9.
- Liao, H., Wang, Z., Deng, Z., Ren, H., & Li, X. (2015). Curcumin inhibits lung cancer invasion and metastasis by attenuating GLUT1/MT1-MMP/MMP2 pathway. *International Journal of Clinical and Experimental Medicine*, 8(6): 8948–8957.
- Lin, S.-S., Lai, K.-C., Hsu, S.-C., Yang, J.-S., Kuo, C.-L., Lin, J.-P., Ma, Y. S., Wu, C. C. & Chung, J.-G. (2009). Curcumin inhibits the migration and invasion of human A549 lung cancer cells through the inhibition of matrix metalloproteinase-2 and -9 and Vascular Endothelial Growth Factor (VEGF). *Cancer Letters*, 285(2): 127–133.
- Linder, S., & Aepfelbacher, M. (2003). Podosomes: Adhesion hot-spots of invasive cells. *Trends in Cell Biology*, 13(7): 376-385.
- Lohmer, L. L., Kelley, L. C., Hagedorn, E. J., & Sherwood, D. R. (2014).

- Invadopodia and basement membrane invasion in vivo. *Cell Adhesion and Migration*, 8(3): 246-255.
- Lucien, F., Brochu-Gaudreau, K., Arsenault, D., Harper, K., & Dubois, C. M. (2011). Hypoxia-induced invadopodia formation involves activation of NHE-1 by the p90 ribosomal s6 kinase (p90RSK). *Plos One*, 6(12): 1-11.
- Lukong, K. E. (2017). Understanding breast cancer – The long and winding road. *BBA Clinical*, 7: 64-77.
- Mader, C. C., Oser, M., Magalhaes, M. A. O., Bravo-Cordero, J. J., Condeelis, J., Koleske, A. J., & Gil-Henn, H. (2011). An EGFR-Src-Arg-Cortactin Pathway Mediates Functional Maturation of Invadopodia and Breast Cancer Cell Invasion. *Cancer Research*, 71(5): 1730–1741.
- Maity, G., Choudhury, P. R., Sen, T., Ganguly, K. K., Sil, H., & Chatterjee, A. (2011). Culture of human breast cancer cell line (MDA-MB-231) on fibronectin-coated surface induces pro-matrix metalloproteinase-9 expression and activity. *Tumor Biology*, 32(1): 129–138.
- Marchesin, V., Castro-Castro, A., Lodillinsky, C., Castagnino, A., Cyrta, J., Bonsang-Kitzis, H., Fuhrmann, L., Irondelle, M., Infante, E., Montagnac, G., Reyfal F., Vincent-Salomon, A., & Chavrier, P. (2015). ARF6-JIP3/4 regulate endosomal tubules for MT1-MMP exocytosis in cancer invasion. *Journal of Cell Biology*, 211(2): 339–358.
- Massagué, J., Batlle, E., & Gomis, R. R. (2017). Understanding the molecular mechanisms driving metastasis. *Molecular Oncology*, 11(1): 3-4.
- Md Hashim, N. F. (2012). *Invadopodia formation in breast cancer*. PhD Thesis. King's College London.
- Md Hashim, N. F., Nicholas, N. S., Dart, A. E., Kiriakidis, S., Paleolog, E., & Wells, C. M. (2013). Hypoxia-induced invadopodia formation: a role for β -PIX. *Open Biology*, 3(6): 120159–120159.
- Mendonça, A. M., Na, T.-Y., & Gumbiner, B. M. (2018). E-cadherin in contact inhibition and cancer. *Oncogene*:1-12.
- Ming-Tatt, L., Khalivulla, S. I., Akhtar, M. N., Lajis, N., Perimal, E. K., Akira, A., Ali, D. A., & Sulaiman, M. R. (2013). Anti-Hyperalgesic effect of a benzilidene-cyclohexanone analogue on a mouse model of chronic constriction injury-induced neuropathic pain: Participation of the κ -Opioid receptor and K_{ATP} . *Pharmacology Biochemistry and Behavior*, 114–115: 58–63.
- Moldovan, G. L., Pfander, B., & Jentsch, S. (2007). PCNA, the Maestro of the Replication Fork. *Cell*, 129(4): 665-679.
- Murphy, D. A., & Courtneidge, S. A. (2011). The “ins” and “outs” of podosomes and invadopodia: Characteristics, formation and function. *Nature Reviews Molecular Cell Biology*, 12(7): 413-426.

- Nagahama, K., Utsumi, T., Kumano, T., Maekawa, S., Oyama, N., & Kawakami, J. (2016). Discovery of a new function of curcumin which enhances its anticancer therapeutic potency. *Scientific Reports*, 6(30962): 1-14.
- Nishida-Aoki, N., Tominaga, N., Takeshita, F., Sonoda, H., Yoshioka, Y., & Ochiya, T. (2017). Disruption of Circulating Extracellular Vesicles as a Novel Therapeutic Strategy against Cancer Metastasis. *Molecular Therapy*, 25(1): 181–191.
- Orsetti, B., Nugoli, M., Cervera, N., Lasorsa, L., Chuchana, P., Ursule, L., Nguyen, C., Redon, R., Du Manoir, S., Rodriguez, C., & Theillet, C. (2004). Genomic and expression profiling of chromosome 17 in breast cancer reveals complex patterns of alterations and novel candidate genes. *Cancer Research*, 64(18): 6453–6460.
- Paget, S. (1889). The distribution of secondary growths in cancer of the breast. *The Lancet*, 133(3421): 571–573.
- Paz, H., Pathak, N., & Yang, J. (2014). Invading one step at a time: The role of invadopodia in tumor metastasis. *Oncogene*, 33(33): 4193-4202.
- Pollard, T. D. (2016). Actin and actin-binding proteins. *Cold Spring Harbor Perspectives in Biology*, 8(8): 1-10.
- Quintavalle, M., Elia, L., Price, J. H., Heynen-Genel, S., & Courtneidge, S. A. (2011). A cell-based high-content screening assay reveals activators and inhibitors of cancer cell invasion. *Science Signaling*, 4(183): 1-22.
- Raposo, G., & Stoorvogel, W. (2013). Extracellular vesicles: Exosomes, microvesicles, and friends. *Journal of Cell Biology*, 200(4): 373-383.
- Razak, N. A., Akhtar, M. N., Abu, N., Ho, W. Y., Tan, S. W., Zareen, S., Tajuddin, S. N., Long, K., Alitheen, N. B. & Yeap, S. K. (2017). The in vivo anti-tumor effect of curcumin derivative (2E-,6E)-2-bis(4-hydroxy-3-methoxybenzylidene) cyclohexanone (BHMC) on 4T1 breast cancer cells. *RSC Advances*, 7(57): 36185–36192.
- Ridley, A. J. (2015). Rho GTPase signaling in cell migration. *Current Opinion in Cell Biology*, 36: 103-112.
- Riggi, N., Aguet, M., & Stamenkovic, I. (2018). Cancer Metastasis: A Reappraisal of Its Underlying Mechanisms and Their Relevance to Treatment. *Annual Review of Pathology: Mechanisms of Disease*, 13(1): 117–140.
- Rockwell, S., Dobrucki, I. T., Kim, E. Y., Marrison, S. T., & Vu, V. T. (2009). Hypoxia and radiation therapy: past history, ongoing research, and future promise. *Current Molecular Medicine*, 9(4): 442–458.
- Sasich, L. D., & Sukkari, S. R. (2012). The US FDA's withdrawal of the breast cancer indication for Avastin (bevacizumab). *Saudi Pharmaceutical*

Journal, 20(4): 381–385.

Schnoor, M., Stradal, T. E., & Rottner, K. (2017). Cortactin: Cell Functions of A Multifaceted Actin-Binding Protein. *Trends in Cell Biology*, 28(2): 79-98.

Scully, O. J., Bay, B.-H., Yip, G., & Yu, Y. (2012). Breast cancer metastasis. *Cancer Genomics and Proteomics*, 9(5): 311-320.

Semenza, G. L. (2016). The hypoxic tumor microenvironment: A driving force for breast cancer progression. *Biochimica et Biophysica Acta*, 1863(3): 382–391.

Shao, Z.-M., Shen, Z.-Z., Liu, C.-H., Sartippour, M. R., Go, V. L., Heber, D., & Nguyen, M. (2002). Curcumin exerts multiple suppressive effects on human breast carcinoma cells. *International Journal of Cancer*, 98(2): 234–240.

Shen, H. L., Liu, Q. J., Yang, P. Q., & Tian, Y. (2015). Protein interactions of cortactin in relation to invadopodia formation in metastatic renal clear cell carcinoma. *Tumor Biology*, 36(5): 3417–3422.

Smid, M., Wang, Y., Klijn, J. G. M., Sieuwerts, A. M., Zhang, Y., Atkins, D., Martens, J. W. & Foekens, J. A. (2006). Genes Associated With Breast Cancer Metastatic to Bone. *Journal of Clinical Oncology*, 24(15): 2261–2267.

Steeg, P. S. (2016). Targeting metastasis. *Nature Reviews Cancer*, 16: 201-218.

Sun, K., Duan, X., Cai, H., Liu, X., Yang, Y., Li, M., Zhang, X., & Wang, J. (2016). Curcumin inhibits LPA-induced invasion by attenuating RhoA/ROCK/MMPs pathway in MCF7 breast cancer cells. *Clinical and Experimental Medicine*, 16(1): 37–47.

Sun, Y. S., Zhao, Z., & Zhu, H. P. (2015). Hispolon inhibits TPA-induced invasion by reducing MMP-9 expression through the NF- κ B signaling pathway in MDA-MB-231 human breast cancer cells. *Oncology Letters*, 10(1): 536–542.

Tarone, G., Cirillo, D., Giancotti, F. G., Comoglio, P. M., & Marchisio, P. C. (1985). Rous sarcoma virus-transformed fibroblasts adhere primarily at discrete protrusions of the ventral membrane called podosomes. *Experimental Cell Research*, 159(1): 141–157.

Tham, C. L., Harith, H. H., Lam, K. W., Chong, Y. J., Cheema, M. S., Sulaiman, M. R., Lajis, N. H., & Israf, D. A (2015). The synthetic curcuminoid BHMC restores endotoxin-stimulated HUVEC dysfunction: Specific disruption on enzymatic activity of p38 MAPK. *European Journal of Pharmacology*, 749: 1–11.

Tham, C. L., Lam, K. W., Rajajendram, R., Cheah, Y. K., Sulaiman, M. R., Lajis, N. H., Kim, M. K. & Israf, D. A. (2011). The effects of a synthetic

- curcuminoid analogue, 2,6-bis-(4-hydroxyl-3-methoxybenzylidene)cyclohexanone on proinflammatory signaling pathways and CLP-induced lethal sepsis in mice. *European Journal of Pharmacology*, 652(1-3): 136-144.
- Tham, C. L., Liew, C. Y., Lam, K. W., Mohamad, A. S., Kim, M. K., Cheah, Y. K., Zakaria, Z. A., Sulaiman, M. R., & Israf, D. A. (2010). A synthetic curcuminoid derivative inhibits nitric oxide and proinflammatory cytokine synthesis. *European Journal of Pharmacology*, 628(1-3): 247-254.
- Tolde, O., Rösel, D., Veselý, P., Folk, P., & Brábek, J. (2010). The structure of invadopodia in a complex 3D environment. *European Journal of Cell Biology*, 89(9): 674–680.
- Tsai, J.-R., Liu, P.-L., Chen, Y.-H., Chou, S.-H., Cheng, Y.-J., Hwang, J.-J., & Chong, I.-W. (2015). Curcumin Inhibits Non-Small Cell Lung Cancer Cells Metastasis through the Adiponectin/NF- κ b/MMPs Signaling Pathway. *Plos One*, 10(12): 1-12.
- Volk-Draper, L., Hall, K., Griggs, C., Rajput, S., Kohio, P., DeNardo, D., & Ran, S. (2014). Paclitaxel therapy promotes breast cancer metastasis in a TLR4-dependent manner. *Cancer Research*, 74(19): 5421–5434.
- Wang, S. C. (2014). PCNA: A silent housekeeper or a potential therapeutic target? *Trends in Pharmacological Sciences*, 35(4): 178-186.
- Wang, S., Li, E., Gao, Y., Wang, Y., Guo, Z., He, J., Zhang, J., Gao, Z. & Wang, Q. (2013). Study on Invadopodia Formation for Lung Carcinoma Invasion with a Microfluidic 3D Culture Device. *Plos One*, 8(2): 1-7.
- Wang, T., Liu, N. S., Seet, L. F., & Hong, W. (2010). The emerging role of VHS domain-containing Tom1, Tom1L1 and Tom1L2 in membrane trafficking. *Traffic*. 11(9): 1119-1128.
- Wang, Z., Liang, X., Cai, M., & Du, G. (2016). Analysis of invadopodia formation in breast cancer cells. *Methods in Molecular Biology*, 1406: 203–210.
- Ward, J. D., Ha, J. H., Jayaraman, M., & Dhanasekaran, D. N. (2015). LPA-mediated migration of ovarian cancer cells involves translocation of G α_{12} to invadopodia and association with Src and β -pix. *Cancer Letters*, 356(2): 382–391.
- Weaver, A. M. (2006). Invadopodia: Specialized cell structures for cancer invasion. *Clinical and Experimental Metastasis*, 23(2): 97-105.
- Woo, M. S., Jung, S. H., Kim, S. Y., Hyun, J. W., Ko, K. H., Kim, W. K., & Kim, H. S. (2005). Curcumin suppresses phorbol ester-induced matrix metalloproteinase-9 expression by inhibiting the PKC to MAPK signaling pathways in human astrogloma cells. *Biochemical and Biophysical Research Communications*, 335(4): 1017–1025.

- Yang, J., Wang, C., Zhang, Z., Chen, X., Jia, Y., Wang, B., & Kong, T. (2017). Curcumin inhibits the survival and metastasis of prostate cancer cells via the Notch-1 signaling pathway. *APMIS*, 125(2): 134–140.
- Yousef, E. M., Tahir, M. R., St-Pierre, Y., & Gaboury, L. A. (2014). MMP-9 expression varies according to molecular subtypes of breast cancer. *BMC Cancer*, 14(609): 1-12.
- Yu, X., & Machesky, L. M. (2012). Cells assemble invadopodia-like structures and invade into matrigel in a matrix metalloprotease dependent manner in the circular invasion assay. *Plos One*, 7(2): 1-12.
- Zambonin-Zallone, A., Teti, A., Carano, A., & Marchisio, P. C. (1988). The distribution of podosomes in osteoclasts cultured on bone laminae: Effect of retinol. *Journal of Bone and Mineral Research*, 3(5): 517–523.
- Zhen, L., Fan, D., Yi, X., Cao, X., Chen, D., & Wang, L. (2014). Curcumin inhibits oral squamous cell carcinoma proliferation and invasion via EGFR signaling pathways. *International Journal of Clinical and Experimental Pathology*, 7(10): 6438–6446.