



UNIVERSITI PUTRA MALAYSIA

***PROPERTIES OF ASIATICOSIDE IN SUPPRESSING
HYPERPERMEABILITY IN HUMAN UMBILICAL VEIN ENDOTHELIAL
CELLS INDUCED BY INTERFERON-GAMMA***

NURNABILA HUDA BINTI KHAIRUDIN

FPSK(M) 2017 75



**PROPERTIES OF ASIATICOSIDE IN SUPPRESSING
HYPERPERMEABILITY IN HUMAN UMBILICAL VEIN ENDOTHELIAL
CELLS INDUCED BY INTERFERON-GAMMA**

By

NURNABILA HUDA BINTI KHAIRUDIN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

NOVEMBER 2016

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**PROPERTIES OF ASIATICOSIDE IN SUPPRESSING
HYPERPERMEABILITY IN HUMAN UMBILICAL VEIN ENDOTHELIAL
CELLS INDUCED BY INTERFERON-GAMMA**

By

NURNABILA HUDA BINTI KHAIRUDIN

NOVEMBER 2016

Chair: Enoch Kumar Perimal, PhD

Faculty: Medicine and Health Sciences

Impairment of endothelial barrier function by vasoactive agents or inflammatory mediators promotes endothelial hyperpermeability causing the development of pathological conditions such as tissue oedema. The changes of barrier structure caused gaps formation thus promote vascular leakage of plasma fluid and macromolecules. Interferon-gamma is a well-known pro-inflammatory mediator which disturbed barrier integrity thus promotes hyperpermeability on endothelial cell. Several studies have reported that asiaticoside; a triterpenoid derived from *Centella asiatica* (local - pegaga) exhibits good anti-oxidant properties, wound healing, immunomodulatory, as well as anti-inflammatory effects on several inflammatory models. In order to elucidate the potential properties of asiaticoside on endothelial barrier preservation, its actions on Human Umbilical Vein Endothelial Cell (HUVEC) hyperpermeability induced by interferon-gamma was assessed. It has been found that interferon-gamma caused an increased in endothelial permeability, determined by the flux of fluorescein isothiocyanate-dextran through *in vitro* permeability assay. Pre-treatment of asiaticoside to interferon-gamma-activated HUVEC significantly ($p < 0.05$) attenuated endothelial hyperpermeability. Moreover, it was clearly demonstrated that asiaticoside helps in restoration of nitric oxide; a mediator which regulates barrier integrity and permeability regulator, back to its basal state after impairment by interferon-gamma. p38 Mitogen Activated Protein Kinase signalling pathway is believed to be involved in nitric oxide production. Thus, it was clearly clarified that p38 Mitogen Activated Protein Kinase phosphorylation, conducted via western blot analysis was deactivated after treatment with asiaticoside. For further clarification, cyclooxygenase-2 level as well as prostaglandin E₂ release in interferon-gamma-activated HUVEC was also studied. Interferon-gamma has increased cyclooxygenase-2 level and prostaglandin E₂ release but also stabilized by asiaticoside. These results suggest that asiaticoside regulated endothelial barrier preservation via p38 Mitogen Activated Protein Kinase phosphorylation and nitric oxide restoration. Our results suggest that asiaticoside may become an important remedy in endothelial barrier preservation induced by interferon-gamma.

Keywords: Hyperpermeability, interferon-gamma, asiaticoside, prostaglandin E₂, cyclooxygenase 2, p38 mitogen activated protein kinase.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

POTENSI ASIATICOSIDE DI DALAM SUPRESI HIPERKEBOLEHTELAPAN PADA SEL ENDOTHELIAL VENA UMBILIKAL MANUSIA YANG DIARUH OLEH INTERFERON-GAMMA

Oleh

NURNABILA HUDA BINTI KHAIRUDIN

NOVEMBER 2016

Pengerusi: Enoch Kumar Perimal, PhD
Fakulti: Perubatan dan Sains Kesihatan

Kemerosotan fungsi halangan endothelial oleh ejen vasoaktif atau pengantara keradangan menggalakkan hiperkebolehtelapan sel endothelial yang menyebabkan pembangunan keadaan patologi seperti rangkaian edema. Perubahan struktur halangan menggalakkan kebocoran vaskular cecair plasma dan makromolekul. Interferon-gamma terkenal sebagai mediator pro-radang yang mengganggu integriti halangan lalu menggalakkan peningkatan ketelapan pada sel-sel endothelial. Beberapa kajian telah melaporkan bahawa asiaticoside; salah satu daripada triterpenoids yang diperolehi daripada *Centella asiatica* (tempatan - pegaga) mempamerkan ciri-ciri anti-oksida yang baik, penyembuhan luka, peningkatan system imun, serta kesan-kesan anti-radang pada beberapa model radang. Dalam usaha untuk menjelaskan potensi asiaticoside pada pemeliharaan sistem halangan endothelial, tindakannya terhadap peningkatan ketelapan HUVEC yang diaruh oleh interferon-gamma telah dinilai. Didapati bahawa interferon-gamma menyebabkan peningkatan ketelapan endothelial yang ditentukan berdasarkan pengaliran fluorescein isothiocyanate-dextran melalui kaedah 'in vitro' assay kebolehtelapan. Pra-rawatan asiaticoside untuk interferon-gamma-aktifkan HUVEC mempunyai kesan signifikansi ($p < 0.05$) dimana peningkatan ketelapan endothelial dapat dilemahkan. Selain itu, ia jelas menunjukkan bahawa asiaticoside membantu dalam pemulihan oksida nitrik; mediator yang mengawal integriti halangan dan pengawalatur ketelapan, kembali kepada keadaan basal selepas terjejas oleh interferon-gamma. Laluan isyarat p38 Mitogen Diaktifkan Protein kinase dipercayai terlibat dalam pengeluaran nitrik oksida. Oleh itu, ia jelas menunjukkan bahawa pemfosforilan p38 Mitogen Diaktifkan Protein Kinase telah dinyahaktifkan selepas rawatan dengan asiaticoside melalui kaedah blot western. Untuk penjelasan lanjut, tahap cyclooxygenase-2 serta prostaglandin E₂ diterbitkan oleh interferon gamma-aktifkan HUVEC juga telah dikaji. Interferon-gamma telah meningkatkan aras cyclooxygenase-2 dan penghasilan prostaglandin E₂ tetapi juga mampu distabilkan oleh asiaticoside. Keputusan ini menunjukkan bahawa asiaticoside memelihara sistem halangan endothelial melalui pemfosforilan p38 Mitogen Diaktifkan Protein kinase dan pemulihan oksida nitrik. Keputusan kami mencadangkan bahawa asiaticoside boleh menjadi satu ubatan yang penting dalam pemeliharaan sistem halangan endothelial yang diaruh oleh interferon-gamma.

Kata kunci: Peningkatan ketelapan, interferon-gamma, asiaticoside, prostaglandin E₂, cyclooxygenase-2, p38 Mitogen Diaktifkan Protein Kinase



ACKNOWLEDGEMENT

In the name of Allah the most gracious and the most merciful, Alhamdulillah thanks to Allah s.w.t whom with His willing of giving me the opportunity to complete this research project with flying colours.

First and foremost, I would like to express my gratitude and appreciation to my late supervisor, Allahyarhamah Associate Professor Dr. Zuraini Ahmad for her guidance and patience throughout this research project. Without her support, advice and unceasing encouragement, this research project and thesis wouldn't have been done and written. I'm very grateful and it's my pleasure to have the chances to work under her supervision. She was understanding person that always give positive motivation in order to succeed in live. I also want to express my heartiest appreciation to my current supervisor, Dr. Enoch Kumar Perimal for his guidance and valuable advice for me to complete this research project, as well as hearties thanks to my co-supervisor, Dr Yong Yoke Keong for his guidance.

Special thanks go to my respective seniors and lab mates in Physiology Research Laboratory for their guidance, advices and kind assistance during my research experiments. I apologize for any unintended inconvenience cause. Not to forget, deepest thanks and appreciation to my lab-mates Mrs Nurfarahdilla and Mr. Shaari who had helping me a lot in completing my project as well as they're being the one who always supports me in every hard phase and moments.

I am also grateful and would like to give my appreciations to the lecturers and staff form Department of Biomedical Science and Physiology Laboratory for their willingness and assistance every time when needed especially in providing many facilities in Lab. They are also supportive and always get ready whenever I need to use the lab during my late nights experiment as well as during holidays. Not to forget, a special thanks to my dearest friends who always support and be there whenever I'm facing the hard time and provides me an endless support in many ways during my research stage.

Last but not least, I also would like to express my gratitude to the one that raise me, provides me an endless love, support me physically and mentally at every time and everywhere, encouragement and their prayers to my beloved parents; Mr Khairudin Hassan and Mrs Noranita Salamat. I don't think I could ever complete my Master Degree and fulfil my dream without them.

I certify that a Thesis Examination Committee has met on _____ to conduct the final examination of NurNabila Huda Khairudin on her thesis entitled “Potential Properties of asiaticoside in Suppression of Hyperpermeability in Human Umbilical Vein Endothelial Cell induced by Interferon-gamma” in accordance with the Universities and Universiti College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follow:

Mohamad Taufik Hidayat bin Baharuldin, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Mohamad Aris bin Moklas, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Zurina Hassan, PhD

Centre for Drug Research
Universiti Sains Malaysia
(External Examiner)

ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the Degree of Master of Science. The members of the Supervisory Committee were as follows:

Enoch Kumar Perimal, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Yong Yoke Keong, PhD

Senior Lecturer
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: _____

Signature: _____
Name of Member
of Supervisory
Committee: _____

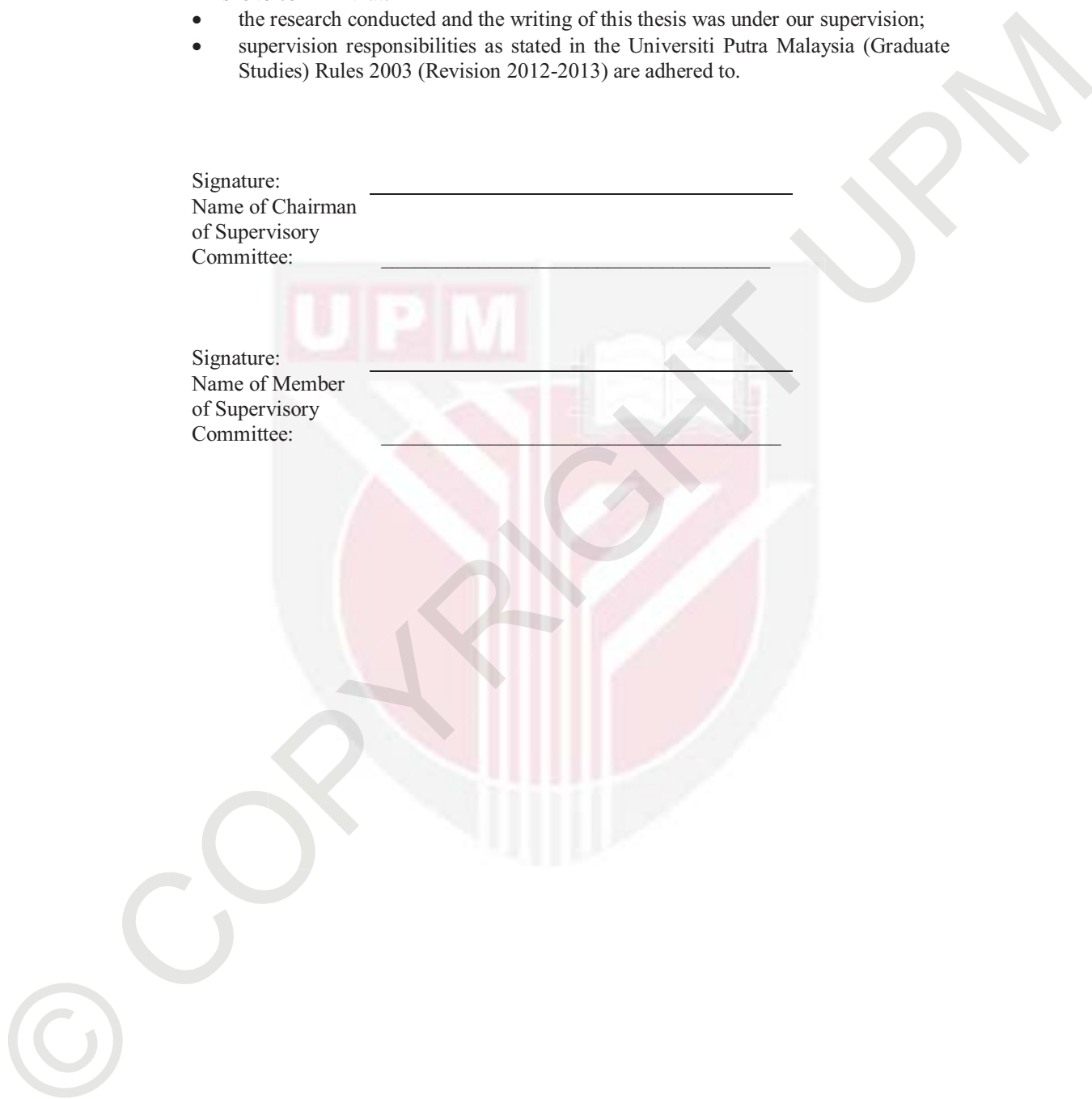


TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	xiv
CHAPTER	
1 INTRODUCTION	1
1.1 General Introduction	1
1.2 Problem statement and justification	2
1.3 Hypothesis	3
1.4 Research Objectives	3
2 LITERATURE REVIEW	4
2.1 Natural products	4
2.2 <i>Centella asiatica</i>	4
2.3 Inflammation	9
2.4 Interferon system	16
2.5 Endothelial cells	17
2.6 Endothelial permeability	19
2.7 Mechanisms of endothelial hyperpermeability	20
2.8 Anti-inflammatory medication	22
3 METHODOLOGY	24
3.1 Chemicals and reagents	24
3.2 Cell culture	25
3.3 Cell viability assay (MTT assay)	26
3.4 Mechanism involve in endothelial hyperpermeability	26
3.5 Statistical analysis	29
4 RESULTS	30
4.1 Cell viability assay (MTT assay)	30
4.2 HUVEC permeability assay	31
4.3 Attenuation of IFN- γ -induced endothelial hyperpermeability by asiaticoside	31
5 DISCUSSION	37
5.1 HUVEC viability towards asiaticoside	37
5.2 HUVEC monolayer permeability study	37
5.3 The effects of asiaticoside in endothelial hyperpermeability and mechanisms involved	38

6	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	43
6.1	Summary	43
6.2	Conclusion	43
6.3	Recommendations for future research	44
	REFERENCES	45
	APPENDICES	56
	BIODATA OF STUDENT	69
	PUBLICATIONS	70



LIST OF TABLES

Table		Page
2.1	Features of acute, chronic and low grade chronic inflammation	11
2.2	Types of T helper (Th) cells and cytokines secretion	14
2.3	Properties of COX-1 and COX-2	15
3.1	Experiment design for HUVEC viability after subjected with various concentration of asiaticoside within 24 h incubation	26
3.2	Experiment design on HUVEC endothelial hyperpermeability induced by IFN- γ	27

LIST OF FIGURES

Figure		Page
2.1	The main target sites of <i>Centella asiatica</i> .	6
2.2	Chemical structure of Asiaticoside.	7
2.3	The plant of <i>Centella asiatica</i> (pegaga / pennywort).	8
2.4	Sequence of events during inflammation.	12
2.5	Endothelial cells with blue endothelial nuclei and black cell boundaries stained with silver nitrate.	18
4.1	Effects of asiaticoside on HUVEC viability. HUVEC were treated with asiaticoside at concentrations ranging from 3.125 μ M to 200 μ M for 24 h measured using MTT assay.	30
4.2	The effect of IFN- γ on HUVEC permeability after 8 h incubation time. This assay was evaluated via FITC-dextran permeability assay.	31
4.3	Effects of asiaticoside on endothelial hyperpermeability induced by 10 ng/mL of IFN- γ . This assay was evaluated via FITC-dextran permeability assay.	32
4.4	Effects of asiaticoside on IFN- γ -impaired NO bioavailability in HUVEC via DAN assays.	33
4.5	Effects of asiaticoside on phosphorylation of p38 MAP Kinase signalling in IFN- γ -activated HUVEC via western blot analysis.	34
4.6	Effects of asiaticoside on COX-2 activity after induced by 10 ng/mL of IFN- γ . HUVEC were pre-treated with asiaticoside at concentration stated.	35
4.7	Effects asiaticoside on PGE2 release in IFN- γ -activated HUVEC. HUVEC were pre-treated with asiaticoside at concentration stated.	36

LIST OF ABBREVIATIONS

AA	Arachidonic acid
BCA	Bicinchoninic acid
BSA	Bovine serum albumin
Ca ²⁺	Calcium ion
COX	Cyclooxygenase
eNOS	Constitutive nitric oxide synthase
DAN	2,3-diaminonaphthalene
DMSO	Dimethyl sulfoxide
ECL	Enhanced chemiluminescence
ECM	Endothelial cell media
EDTA	Ethylenediaminetetraacetic acid
END-D Cells	Mouse vascular endothelial cells
ELISA	Enzyme-linked Immunosorbent Assay
eNOS	Endothelial nitric oxide synthase
ERK	Extracellular signal-regulated kinases
FBS	Fetal bovine serum
FDA	Food & drug administration
FITC	Fluorescence isothiocyanate
GPCR	G protein coupled receptors
HPLC	High performance liquid chromatography
HUVEC	Human umbilical vein endothelial cell
HO	Heme oxygenase
ICAM-1	Intercellular adhesion molecule-1
IFN- α	Interferon-alpha
IFN- β	Interferon-beta
IFN- γ	Interferon-gamma
IFN- ω	Interferon-omega
IFN- τ	Interferon-tau
IL	Interleukin
iNOS	Inducible nitric oxide synthase
IP3	Inositol 1,4,5-triphosphate
JNK	C-jun N terminal kinase
LNAME	L-NG-Nitroarginine methyl ester
LPS	Lipopolysaccharide
MAF	Macrophage-activating factor
MAPK	Mitogen activated protein kinase
MHC	Major histocompatibility complex
MTT	3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
NaOH	Sodium hydroxide
nNOS	Neuronal nitric oxide synthase
NO	Nitric oxide
NK cell	Natural killer cell
NSAIDs	Non-steroidal anti-inflammatory Drugs
PAF	Platelet-activating factor
PAMP	Pathogen-associated molecular pattern
PBS	Phosphate buffered saline
PGE ₂	Prostaglandin E ₂
PKC	Protein kinase C

PKG	Protein kinase G
PVDF	Poly-vinylidenedifluoride
RIPA Buffer	Radioimmunoprecipitation assay buffer
ROS	Reactive oxygen species
SDS-PAGE	Sodium dodecyl sulfate polyacrylamide gel electrophoresis
TBS	Tris-buffered saline
Th cell	T Helper cell
TLC	Thin layer chromatography
TNF- α	Tumour Necrosis Factor- α
[Ca ²⁺] _i	Intracellular Calcium ion concentration
U/mL	Units per ml
μ M	Micromolar
μ M/mL	Micromolar per millilitre
μ g/mL	Microgram per millilitre
μ L	Microlitre
ng/mL	Nanogram per millilitre
μ m	Micrometer
mM	Millimolar
Mg	Miligram

© COPYRIGHT UPM



CHAPTER 1

INTRODUCTION

1.1 General introduction

Vascular tissue is a fundamental effector when there is body's tissues damaged due to infections and trauma. The vessels will start to release fluid due to high permeability and increases blood flow in order to stop and repair the damage. Endothelial hyperpermeability marks the occurrence of inflammatory vascular oedema. Inflammation is the reaction or response of living tissue when there is infection and injury caused by physical trauma, noxious chemicals, or microbiologic agents take place. Inflammation is a normal, protective response and one of the body's efforts to inactivate or destroy invading organisms, remove irritants and directly set the stage of tissue repair – healing. When healing process is completed, these inflammatory processes usually cease. However, if assisted repair mechanism is poorly conducted, chronic inflammatory disorder can be developed which leads to persistent tissue damage by leucocytes, lymphocytes or collagen (Nathan, 2002) and further promotion to scarring and loss of organ function (Ricciotti & FitzGerald, 2011).

Inflammation encompasses a complex cascade of reaction at the peripheral receptor terminals. It works on two ways; primary (acute) and secondary (chronic) pathways. Typically, primary pathway focuses on detoxification and repair while secondary pathway works on protection or body defence process. Inflammation can be produced with the aid of pro-inflammatory cytokines such as Interferon-gamma (IFN- γ), Tumour Necrosis Factor (TNF), Interleukin (IL)-8 as well as neuropeptide bioactive substance, Substance P (SP) which contribute to neurogenic inflammation. SP in turn may act on target cells in periphery such as mast cells, immune cells, as well as vascular smooth muscle (Richardson & Vasko, 2002; Walsh et al., 1995) thus evoking inflammatory peripheral effects such as vasodilatation, plasma extravasation (Lembeck & Holzer, 1979) and leukocyte activation (O'Connor et al., 2004). This process appears to contribute in the pathogenesis of numerous diseases including psoriasis, asthma, fibromyalgia, eczema, multiple chemical sensitivity and migraine (Richardson & Vasko, 2002). IFN- γ is pleiotropic cytokine that possesses anti-tumour (Schroder, Hertzog, Ravasi, & Hume, 2004), anti-proliferative and antiviral activity (Dinarello, 2000; Pawliczak et al., 2005). IFN- γ may also boost TNF activity and induced nitric oxide (NO) formation. Therefore, IFN- γ is considered as a pro-inflammatory cytokine (Dinarello, 2000) and since the last decades, cytokine-based therapy has brought in much attention to researchers (Ng et al., 2015).

When there is disruption or sudden disturbance of endothelial layer during pathological conditions, the injury may lead to endothelial cell damage, contraction of adjacent endothelial cell, plasma extravasation (Granger, 2010) and subsequently cause chronic inflammatory diseases (Javanmard & Dana, 2012). Endothelial gaps causing endothelial cell to become highly permeable to proteins and plasma fluids, which eventually leads to oedema and swelling (Fong et al., 2015). Endothelial cells become the main targets of the occurrence of adverse effects as they are routinely exposed to circulating immune cells and effector molecules of immunity after responding to stimuli (Javanmard & Dana, 2012). In the present study, Human Umbilical Vein

Endothelial Cell (HUVEC) was used as *in vitro* model system to study whether IFN- γ -induced endothelial hyperpermeability can be attenuated by asiaticoside which is one of the components found in *Centella asiatica*.

Present study will focused on asiaticoside in restoration of IFN- γ -induced endothelial hyperpermeability on HUVEC and the possible mechanisms involved. Traditional plant remedies can become an important source in providing new, active and potential treatment to cure diseases related to inflammation. *Centella asiatica* (local name – pegaga in Malay) has been utilized for many years in medicinal treatment while asiaticoside; a major triterpenoid saponin component of *Centella asiatica* is believed to possess various antioxidant properties (Pitella et al., 2009), anti-inflammatory activities (Gohil et al., 2010) and contribute to wound healing (Shukla et al., 1999). Apart from the valuable effects of asiaticoside stated above, the beneficial effects of asiaticoside on IFN- γ -induced endothelial hyperpermeability in HUVEC are still insufficiently declared.

This research can provide a very important finding where asiaticoside becomes a novel alternative treatment if it could be an anti-hyperpermeability induced by IFN- γ on HUVEC. Nevertheless, the aim of this study is not only to determine the anti-hyperpermeability ability of asiaticoside, but also the beneficial inhibitory effects of asiaticoside on the underlying mechanisms involved in hyperpermeability. More scientific data are required to explain the biological activities, mechanism of reaction as well as the active components in *Centella asiatica* though a large number of studies reported over the past decades.

1.2 Problem statement and justification

To date, conventional NSAIDs (non-steroidal anti-inflammatory drugs) as well as selective COX-2 inhibitor are associated with small and definite risk of several side effects. There are over 15,000 deaths annually among patients following doctor's prescriptions for NSAIDs and this toxicity is especially noticed in elderly (Odell & Sorgnard, 2008). Furthermore, the use of these drugs offers other weaknesses such as significant side effects, high cost and the route of administration. For example, at least two subcutaneous injections per week are required for an effective drug therapy (Santos, 2004). Other than that, glucocorticoid therapy is also widely used to treat inflammation. Prolonged use of glucocorticoid therapy may induce side effects involving the central nervous system, gastrointestinal tract as well as cardiovascular system (Moghadam-Kia & Werth, 2010). In this perspective, therapeutic agents which derived from natural plant may have positive contributions and benefits with regards to the treatment for inflammatory disorder as well as other diseases with inflammatory origin. Dexamethasone, a well-known glucocorticoids were used to inhibit pro-inflammatory cytokines production as well as other pro-inflammatory mediators as described by Pang & Knox (1997). It was used in the present study as the positive control to define and compare the anti-inflammatory activity of a new potential treatment from natural plant origin, Asiaticoside as well as the possible mechanisms involved.

1.3 Hypothesis

Asiaticoside has no toxic effects and will not cause cell death to HUVEC on tested concentrations. Asiaticoside will significantly attenuate IFN- γ -induced endothelial hyperpermeability in Human Umbilical Vein Endothelial Cell (HUVEC) by improvement of impaired nitric oxide (NO) levels back to its basal states. Asiaticoside also reduces cyclooxygenase-2 (COX-2) enzyme activity and prostaglandin E₂ (PGE₂) release on HUVEC. Asiaticoside aids in deactivation of p38 mitogen-activated protein kinases (MAPK) in Human Umbilical Vein Endothelial Cell (HUVEC).

1.4 Research objectives

1.4.1 General objective

To evaluate the effects of asiaticoside on IFN- γ -induced endothelial hyperpermeability and the possible mechanisms involved in cell model; Human Umbilical Vein Endothelial Cell (HUVEC).

1.4.2 Specific objectives

1. To determine the viability of HUVEC towards asiaticoside.
2. To determine the effects of asiaticoside on IFN- γ -induced endothelial hyperpermeability in HUVEC.
3. To determine the effects of asiaticoside on nitric oxide (NO) levels, cyclooxygenase-2 (COX-2) levels, and prostaglandin E₂ (PGE₂) release in activated HUVEC.
4. To determine the effects of asiaticoside on p38 mitogen-activated protein kinases (MAPK) phosphorylation in activated HUVEC.

REFERENCES

- (U.S), F. a. D. A. (2012). COX-2 Selective and Non-Selective Non-Steroidal Anti-Inflammatory Drugs (NSAIDs). from U.S. Department of Health & Human Services
- Akarasereenont, P., Techatraisak, K., Thaworn, A., & Chotewuttakorn, S. (2002). The expression of COX-2 in VEGF-treated endothelial cells is mediated through protein tyrosine kinase. *Mediators of inflammation*, 11(1), 17-22.
- Angeles Muñoz-Fernández, M., & Fresno, M. (1998). The role of tumour necrosis factor, interleukin 6, interferon- γ and inducible nitric oxide synthase in the development and pathology of the nervous system. *Progress in neurobiology*, 56(3), 307-340.
- Baluk, P. (1997). *Neurogenic inflammation in skin and airways*. Paper presented at the Journal of Investigative Dermatology Symposium Proceedings.
- Bascom, R., Meggs, W. J., Frampton, M., Hudnell, K., Killburn, K., Kobal, G., . . . Rea, W. (1997). Neurogenic inflammation: with additional discussion of central and perceptual integration of nonneurogenic inflammation. *Environmental health perspectives*, 105(Suppl 2), 531.
- Bates, D., Hillman, N., Williams, B., Neal, C., & Pocock, T. (2002). Regulation of microvascular permeability by vascular endothelial growth factors*. *Journal of anatomy*, 200(6), 581-597.
- Benbarek, H., Mouithys-Mickalad, A., Deby-Dupont, G., Deby, C., Grülke, S., Nemmar, A., . . . Serteyn, D. (1999). High concentrations of histamine stimulate equine polymorphonuclear neutrophils to produce reactive oxygen species. *Inflammation Research*, 48(11), 594-601.
- Billiau, A. (1996). Interferon-g: biology and role in pathogenesis. *Advances in immunology*, 62, 61-130.
- Billiau, A., Heremans, H., Vermeire, K., & Matthys, P. (1998). Immunomodulatory Properties of Interferon- γ : An Update. *Annals of the New York Academy of Sciences*, 856(1), 22-32.
- Bogatcheva, N. V., & Verin, A. D. (2008). The role of cytoskeleton in the regulation of vascular endothelial barrier function. *Microvascular research*, 76(3), 202-207.
- Bonte, F., Dumas, M., Chaudagne, C., & Meybeck, A. (1994). Influence of asiatic acid, madecassic acid, and asiaticoside on human collagen I synthesis. *Planta medica*, 60(02), 133-135.
- Borbiev, T., Birukova, A., Liu, F., Nurmukhambetova, S., Gerthoffer, W. T., Garcia, J. G., & Verin, A. D. (2004). p38 MAP kinase-dependent regulation of endothelial cell permeability. *American Journal of Physiology-Lung Cellular and Molecular Physiology*, 287(5), L911-L918.
- Borish, L. C., & Steinke, J. W. (2003). 2. Cytokines and chemokines. *Journal of Allergy and Clinical Immunology*, 111(2), S460-S475.

- Bryan, N. S., & Grisham, M. B. (2007). Methods to detect nitric oxide and its metabolites in biological samples. *Free Radical Biology and Medicine*, 43(5), 645-657.
- Calder, P. C., Ahluwalia, N., Albers, R., Bosco, N., Bourdet-Sicard, R., Haller, D., . . . Marcos, A. (2013). A consideration of biomarkers to be used for evaluation of inflammation in human nutritional studies. *British Journal of Nutrition*, 109(S1), S1-S34.
- Calder, P. C., Ahluwalia, N., Brouns, F., Buetler, T., Clement, K., Cunningham, K., . . . Lansink, M. (2011). Dietary factors and low-grade inflammation in relation to overweight and obesity. *British Journal of Nutrition*, 106(S3), S1-S78.
- ChemDiv. (2014). Anti-inflammatory Library. In C. T. C. o. Cures (Ed.).
- Chen, S., Yin, Z.-J., Jiang, C., Ma, Z.-Q., Fu, Q., Qu, R., & Ma, S.-P. (2014). Asiaticoside attenuates memory impairment induced by transient cerebral ischemia-reperfusion in mice through anti-inflammatory mechanism. *Pharmacology Biochemistry and Behavior*, 122, 7-15.
- Chen, S. C., Tsai, M. H., Wang, H. J., Yu, H. S., & Chang, L. W. (2007). Involvement of substance P and neurogenic inflammation in arsenic-induced early vascular dysfunction. *Toxicological sciences*, 95(1), 82-88.
- Claesson-Welsh, L. (2015). Vascular permeability—the essentials. *Upsala journal of medical sciences*, 120(3), 135-143.
- Conway, J. (2000). Inflammation and Repair. College of Medicine: University of Illinois.
- Créminon, C., Habib, A., Maclouf, J., Pradelles, P., Grassi, J., & Frobert, Y. (1995). Differential measurement of constitutive (COX-1) and inducible (COX-2) cyclooxygenase expression in human umbilical vein endothelial cells using specific immunometric enzyme immunoassays. *Biochimica et Biophysica Acta (BBA)-Lipids and Lipid Metabolism*, 1254(3), 341-348.
- Dias, D. A., Urban, S., & Roessner, U. (2012). A historical overview of natural products in drug discovery. *Metabolites*, 2(2), 303-336.
- Dinarello, C. A. (2000). Proinflammatory cytokines. *Chest Journal*, 118(2), 503-508.
- Donnerer, J., & Amann, R. (1993). The inhibition of neurogenic inflammation. *General Pharmacology: The Vascular System*, 24(3), 519-529.
- Farnsworth, N. R., Akerele, O., Bingel, A. S., Soejarto, D. D., & Guo, Z. (1985). Medicinal plants in therapy. *Bull World Health Organ*, 63(6), 965-981.
- Faujan, N. H., Abdullah, N., Abdullah Sani, N., & Babji, A. S. (2007). Antioxidative activities of water extracts of some Malaysian herbs. *ASEAN Food Journal*, 14(1), 61-68.
- Fish, S., Proujansky, R., & Reenstra, W. (1999). Synergistic effects of interferon γ and tumour necrosis factor α on T84 cell function. *Gut*, 45(2), 191-198.
- Fong, L. Y., Ng, C. T., Zakaria, Z. A., Baharuldin, M. T. H., Arifah, A. K., Hakim, M. N., & Zuraini, A. (2015). Asiaticoside Inhibits TNF- α -Induced Endothelial Hyperpermeability of Human Aortic Endothelial Cells. *Phytotherapy Research*.

- Förstermann, U., & Münzel, T. (2006). Endothelial nitric oxide synthase in vascular disease from marvel to menace. *Circulation*, 113(13), 1708-1714.
- Fosslien, E. (2005). Cardiovascular complications of non-steroidal anti-inflammatory drugs. *Annals of Clinical & Laboratory Science*, 35(4), 347-385.
- Funk, C. D. (2001). Prostaglandins and leukotrienes: advances in eicosanoid biology. *Science*, 294(5548), 1871-1875.
- Fürst, R., Schroeder, T., Eilken, H. M., Bubik, M. F., Kiemer, A. K., Zahler, S., & Vollmar, A. M. (2007). MAPK phosphatase-1 represents a novel anti-inflammatory target of glucocorticoids in the human endothelium. *The FASEB Journal*, 21(1), 74-80.
- Gohil, K. J., Patel, J. A., & Gajjar, A. K. (2010). Pharmacological review on *Centella asiatica*: a potential herbal cure-all. *Indian journal of pharmaceutical sciences*, 72(5), 546.
- Goldkind, L., & Laine, L. (2006). A systematic review of NSAIDs withdrawn from the market due to hepatotoxicity: lessons learned from the bromfenac experience. *Pharmacoepidemiology and drug safety*, 15(4), 213-220.
- Granger, Senchenkova E. (2010). *Inflammation and the Microcirculation*. San Rafael (CA): Morgan & Claypool Life Sciences.
- Guo, T., Fang, M., Zhang, D., & Li, X. (2013). Combination treatment with asiaticoside and rapamycin: A new hope for in-stent restenosis. *Experimental and therapeutic medicine*, 6(2), 557-561.
- Hashim, P. (2011). *Centella asiatica* in food and beverage applications and its potential antioxidant and neuroprotective effect. *International Food Research Journal*, 18(4), 1215-1222.
- Hatakeyama, T., Pappas, P. J., Hobson, R. W., Boric, M. P., Sessa, W. C., & Durán, W. N. (2006). Endothelial nitric oxide synthase regulates microvascular hyperpermeability in vivo. *The Journal of physiology*, 574(1), 275-281.
- Holzer, P. (1998). Neurogenic vasodilatation and plasma leakage in the skin. *General Pharmacology: The Vascular System*, 30(1), 5-11.
- Houle, F., & Huot, J. (2006). Dysregulation of the endothelial cellular response to oxidative stress in cancer. *Molecular carcinogenesis*, 45(6), 362-367.
- Indena. (2011). *Centella Asiatica Selected Triterpenes - A Highly Standardized Natural Remedy for The Maintenance of an Healthy Venous System*. smartshop.nazwa.pl/coffeshop/centella.pdf
- Inoue, H., Ando, K., Wakisaka, N., Matsuzaki, K.-i., Aihara, M., & Kumagai, N. (2001). Effects of nitric oxide synthase inhibitors on vascular hyperpermeability with thermal injury in mice. *Nitric Oxide*, 5(4), 334-342.
- Islam, M. A. (2015). *Centella Asiatica L.: A Concise Drug Review With Probable Clinical Uses*. *wounds*, 5, 9.
- Javanmard, S. H., & Dana, N. (2012). The effect of interferon γ on endothelial cell nitric oxide production and apoptosis. *Advanced biomedical research*, 1.

- Jin, H., Koyama, T., Hatanaka, Y., Akiyama, S., Takayama, F., & Kawasaki, H. (2006). Histamine-induced vasodilation and vasoconstriction in the mesenteric resistance artery of the rat. *European journal of pharmacology*, 529(1), 136-144.
- Jongsung, L. (2006). Asiaticoside Induce Human Collagen I Synthesis through TGF β Receptor I Kinase (T β RI Kinase)-Independent Smad Signaling. *United States: Planta Med*, 72, 324-328.
- Kähler, C. M., Kirchmair, R., Kaufmann, G., Kähler, S. T., Reinisch, N., Fischer-Colbrie, R., . . . Wiedermann, C. J. (1997). Inhibition of proliferation and stimulation of migration of endothelial cells by secretoneurin in vitro. *Arteriosclerosis, thrombosis, and vascular biology*, 17(5), 932-939.
- Khalil, Z., & Helme, R. D. (1989). Sequence of events in substance P-mediated plasma extravasation in rat skin. *Brain research*, 500(1), 256-262.
- Kilbourn, R. G., & Belloni, P. (1990). Endothelial cell production of nitrogen oxides in response to interferon γ in combination with tumor necrosis factor, interleukin-1, or endotoxin. *Journal of the National Cancer Institute*, 82(9), 772-776.
- Komarova, Y., & Malik, A. B. (2010). Regulation of endothelial permeability via paracellular and transcellular transport pathways. *Annual review of physiology*, 72, 463-493.
- Kornfeld, R. A. (2009). Five Ways To Reduce Inflammation Naturally. Retrieved from http://www.huffingtonpost.com/dr-robert-a-kornfeld/5-ways-to-reduce-inflamma_b_271640.html
- Kumar, P., Shen, Q., Pivetti, C. D., Lee, E. S., Wu, M. H., & Yuan, S. Y. (2009). Molecular mechanisms of endothelial hyperpermeability: implications in inflammation. *Expert reviews in molecular medicine*, 11, e19.
- Larsen, G. L., & Henson, P. M. (1983). Mediators of inflammation. *Annual review of immunology*, 1(1), 335-359.
- Lee, Kim, H. L., Lee, M. H., You, K. E., Kwon, B. J., Seo, H. J., & Park, J. C. (2012). Asiaticoside enhances normal human skin cell migration, attachment and growth in vitro wound healing model. *Phytomedicine*, 19(13), 1223-1227.
- Lee, Shin, Y. J., Won, C., Lee, Y.-S., Park, C.-G., Ye, S.-K., & Chung, M.-H. (2009). Simvastatin acts as an inhibitor of interferon gamma-induced cyclooxygenase-2 expression in human THP-1 cells, but not in murine RAW264. 7 cells. *Biocell*, 33(2), 107-114.
- Lembeck, F., & Holzer, P. (1979). Substance P as neurogenic mediator of antidromic vasodilation and neurogenic plasma extravasation. *Naunyn-Schmiedeberg's archives of pharmacology*, 310(2), 175-183.
- Levine, J., Dardick, S., Basbaum, A., & Scipio, E. (1985). Reflex neurogenic inflammation. I. Contribution of the peripheral nervous system to spatially remote inflammatory responses that follow injury. *The Journal of Neuroscience*, 5(5), 1380-1386.

- Li, T., Hu, J., Du, S., Chen, Y., Wang, S., & Wu, Q. (2014). ERK1/2/COX-2/PGE2 signaling pathway mediates GPR91-dependent VEGF release in streptozotocin-induced diabetes. *Molecular vision*, 20, 1109.
- Lin, C.-H., Wu, C.-H., Thum, W.-Y., Ho, Y.-S., & Lee, H.-M. (2002). Involvement of p38 mitogen-activated protein kinase in PLL-AGE-induced cyclooxygenase-2 expression. *European journal of pharmacology*, 438(3), 143-152.
- Luo, Y., Fu, C., Wang, Z., Zhang, Z., Wang, H., & Liu, Y. (2015). Asiaticoside attenuates the effects of spinal cord injury through antioxidant and anti-inflammatory effects, and inhibition of the p38- MAPK mechanism. *Molecular medicine reports*, 12(6), 8294-8300.
- Mandal, A. (2013). What are cytokines (Medical Information Service). Retrieved 17 January 2015, from AZoM.com <http://www.news-medical.net/health/What-are-Cytokines.aspx>
- Mantyh, P. W. (2003). Neurobiology of substance P and the NK1 receptor. *The Journal of clinical psychiatry*, 63(suppl 11), 6-10.
- Martin, S., Maruta, K., Burkart, V., Gillis, S., & Kolb, H. (1988). IL-1 and IFN-gamma increase vascular permeability. *Immunology*, 64(2), 301.
- Matsuura, H., Sakaue, M., Subbaramaiah, K., Kamitani, H., Eling, T. E., Dannenberg, A. J., . . . Jetten, A. M. (1999). Regulation of Cyclooxygenase-2 by Interferon γ and Transforming Growth Factor α in Normal Human Epidermal Keratinocytes and Squamous Carcinoma Cells ROLE OF MITOGEN-ACTIVATED PROTEIN KINASES. *Journal of Biological Chemistry*, 274(41), 29138-29148.
- McGeachie, J. (1998). More About Endothelial Cells. 2014, from <http://www.lab.anhb.uwa.edu.au/mb140/moreabout/endothel.htm>
- Mehta, D., & Malik, A. B. (2006). Signaling mechanisms regulating endothelial permeability. *Physiological reviews*, 86(1), 279-367.
- Mestre, J. R., Mackrell, P. J., Rivadeneira, D. E., Stapleton, P. P., Tanabe, T., & Daly, J. M. (2001). Redundancy in the signaling pathways and promoter elements regulating cyclooxygenase-2 gene expression in endotoxin-treated macrophage/monocytic cells. *Journal of Biological Chemistry*, 276(6), 3977-3982.
- Mishra, B. B., & Tiwari, V. K. (2011). Natural products: an evolving role in future drug discovery. *European journal of medicinal chemistry*, 46(10), 4769-4807.
- Misko, T. P., Schilling, R., Salvemini, D., Moore, W., & Currie, M. (1993). A fluorometric assay for the measurement of nitrite in biological samples. *Analytical biochemistry*, 214(1), 11-16.
- Moghadam-Kia, S., & Werth, V. P. (2010). Prevention and treatment of systemic glucocorticoid side effects. *International journal of dermatology*, 49(3), 239-248.
- Molinari, G. (2009). Natural products in Drug Discovery: Present Status and Perspective. In C. A. Guzman & G. Z. Feuerstein (Eds.), *Advances Experimental Medicine and Biology* (Vol. 655). Texas, USA: LandesBioscience.

- Monter, L. W. (2016, 13th October 2016). Inflammation Protection Protocol. Retrieved 24th December 2016, 2016, from http://healthyprotocols.com/2_inflammation.htm
- Moore, K. W., O'garra, A., Malefyt, R. d. W., Vieira, P., & Mosmann, T. R. (1993). Interleukin-10. *Annual review of immunology*, 11(1), 165-190.
- Morikawa, A., Koide, N., Kato, Y., Sugiyama, T., Chakravorty, D., Yoshida, T., & Yokochi, T. (2000). Augmentation of nitric oxide production by gamma interferon in a mouse vascular endothelial cell line and its modulation by tumor necrosis factor alpha and lipopolysaccharide. *Infection and immunity*, 68(11), 6209-6214.
- Morita, I. (2002). Distinct functions of COX-1 and COX-2. *Prostaglandins & other lipid mediators*, 68, 165-175.
- Nathan, C. (2002). Points of control in inflammation. *Nature*, 420(6917), 846-852.
- Ng, & Celermajer. (2004). Glucocorticoid treatment and cardiovascular disease. *Heart*, 90(8), 829-830.
- Ng, Fong, L. Y., Sulaiman, M. R., Mohd Moklas, M. A., Yong, Y. K., Hakim, M. N., & Ahmad, Z. (2015). Interferon-Gamma Increases Endothelial Permeability by Causing Activation of p38 MAP Kinase and Actin Cytoskeleton Alteration. *Journal of Interferon & Cytokine Research*.
- Norzaharaini, M., Norshazwani, W., Hasmah, A., Izani, N. N., & Rapeah, S. (2011). A preliminary study on the antimicrobial activities of asiaticoside and asiatic acid against selected gram positive and gram negative bacteria. *Health Environ J*, 2(2), 23-26.
- O'Connor, T. M., O'Connell, J., O'Brien, D. I., Goode, T., Bredin, C. P., & Shanahan, F. (2004). The role of substance P in inflammatory disease. *Journal of cellular physiology*, 201(2), 167-180.
- Oberleithner, H., Riethmüller, C., Ludwig, T., Shahin, V., Stock, C., Schwab, A., . . . Schillers, H. (2006). Differential action of steroid hormones on human endothelium. *Journal of cell science*, 119(9), 1926-1932.
- Odell, R. H., & Sorgnard, R. E. (2008). Anti-inflammatory effects of electronic signal treatment. *Pain physician*, 11(6), 891-907.
- Onat, D., Brillon, D., Colombo, P. C., & Schmidt, A. M. (2011). Human vascular endothelial cells: a model system for studying vascular inflammation in diabetes and atherosclerosis. *Current diabetes reports*, 11(3), 193-202.
- Orhan, I. E. (2012). Centella asiatica (L.) urban: from traditional medicine to modern medicine with neuroprotective potential. *Evidence-based complementary and alternative medicine*, 2012.
- Oshima, T., Laroux, F. S., Coe, L. L., Morise, Z., Kawachi, S., Bauer, P., . . . Jennings, S. (2001). Interferon- γ and interleukin-10 reciprocally regulate endothelial junction integrity and barrier function. *Microvascular research*, 61(1), 130-143.

- Pang, L., & Knox, A. J. (1997). Effect of interleukin-1 β , tumour necrosis factor- α and interferon- γ on the induction of cyclo-oxygenase-2 in cultured human airway smooth muscle cells. *British journal of pharmacology*, 121(3), 579-587.
- Paolino, D., Cosco, D., Cilurzo, F., Trapasso, E., Morittu, V. M., Celia, C., & Fresta, M. (2012). Improved in vitro and in vivo collagen biosynthesis by asiaticoside-loaded ultradeformable vesicles. *Journal of Controlled Release*, 162(1), 143-151.
- Parfenova, H., Parfenov, V. N., Shlopov, B. V., Levine, V., Falkos, S., Pourcyrus, M., & Leffler, C. W. (2001). Dynamics of nuclear localization sites for COX-2 in vascular endothelial cells. *American Journal of Physiology-Cell Physiology*, 281(1), C166-C178.
- Pawitan, J. A. (2011). Potential agents against plasma leakage. *ISRN pharmacology*, 2011.
- Pawliczak, R., Logun, C., Madara, P., Barb, J., Suffredini, A. F., Munson, P. J., . . . Shelhamer, J. H. (2005). Influence of IFN- γ on gene expression in normal human bronchial epithelial cells: modulation of IFN- γ effects by dexamethasone. *Physiological genomics*, 23(1), 28-45.
- Petrache, I., Birukova, A., Ramirez, S. I., Garcia, J. G., & Verin, A. D. (2003). The role of the microtubules in tumor necrosis factor- α -induced endothelial cell permeability. *American Journal of Respiratory Cell and Molecular Biology*, 28(5), 574-581.
- Pittella, F., Dutra, R. C., Junior, D. D., Lopes, M. T., & Barbosa, N. R. (2009). Antioxidant and cytotoxic activities of *Centella asiatica* (L) Urb. *International journal of molecular sciences*, 10(9), 3713-3721.
- Platanias, L. C. (2005). Mechanisms of type-I-and type-II-interferon-mediated signalling. *Nature Reviews Immunology*, 5(5), 375-386.
- Poher, J. S., Min, W., & Bradley, J. R. (2009). Mechanisms of endothelial dysfunction, injury, and death. *Annual Review of Pathological Mechanical Disease*, 4, 71-95.
- Potapova, I. A., Cohen, I. S., & Doronin, S. V. (2013). Caspases and p38 MAPK Regulate Endothelial Cell Adhesiveness for Mesenchymal Stem Cells. *PLoS one*, 8(9), e73929.
- Qiu, S., Liu, N., Jia, L., Yang, G., Su, W., Li, J., . . . Zhang, C. (2012). A new treatment for neurogenic inflammation caused by EV71 with CR2-targeted complement inhibitor. *Virology journal*, 9(1), 285.
- Rauch, I., Müller, M., & Decker, T. (2013). The regulation of inflammation by interferons and their STATs. *Jak-Stat*, 2(1).
- Raychaudhuri, S. K., Raychaudhuri, S. P., Weltman, H., & Farber, E. M. (2001). Effect of nerve growth factor on endothelial cell biology: proliferation and adherence molecule expression on human dermal microvascular endothelial cells. *Archives of dermatological research*, 293(6), 291-295.
- Razani, B., & Lisanti, M. P. (2001). Caveolin-deficient mice: insights into caveolar function human disease. *Journal of Clinical Investigation*, 108(11), 1553.

- Ricciotti, E., & FitzGerald, G. A. (2011). Prostaglandins and inflammation. *Arteriosclerosis, thrombosis, and vascular biology*, 31(5), 986-1000.
- Richardson, J. D., & Vasko, M. R. (2002). Cellular mechanisms of neurogenic inflammation. *Journal of Pharmacology and Experimental Therapeutics*, 302(3), 839-845.
- Roy, S. K., Hu, J., Meng, Q., Xia, Y., Shapiro, P. S., Reddy, S. P., . . . Pritchard, C. (2002). MEKK1 plays a critical role in activating the transcription factor C/EBP- β -dependent gene expression in response to IFN- γ . *Proceedings of the National Academy of Sciences*, 99(12), 7945-7950.
- Rush, W., Murray, G., & Graham, D. (1993). The comparative steady-state bioavailability of the active ingredients of Madecassol. *European journal of drug metabolism and pharmacokinetics*, 18(4), 323-326.
- Santos, A. R. (2004). Anti-inflammatory compounds of plant origin. Part II. Modulation of pro-inflammatory cytokines, chemokines and adhesion molecules. *Planta Med*, 70, 93-103.
- Schroder, K., Hertzog, P. J., Ravasi, T., & Hume, D. A. (2004). Interferon- γ : an overview of signals, mechanisms and functions. *Journal of leukocyte biology*, 75(2), 163-189.
- Sen, G. C. (2001). Viruses and interferons. *Annual Reviews in Microbiology*, 55(1), 255-281.
- Sessa, W. C. (2004). eNOS at a glance. *Journal of cell science*, 117(12), 2427-2429.
- Shukla, A., Rasik, A., Jain, G., Shankar, R., Kulshrestha, D., & Dhawan, B. (1999). In vitro and in vivo wound healing activity of asiaticoside isolated from *Centella asiatica*. *Journal of ethnopharmacology*, 65(1), 1-11.
- Simoneau, B., Houle, F., & Huot, J. (2012). Regulation of endothelial permeability and transendothelial migration of cancer cells by tropomyosin-1 phosphorylation. *Vasc Cell*, 4(1), 18-18.
- Singh, S., Gautam, A., Sharma, A., & Batra, A. (2010). *Centella asiatica* (L.): a plant with immense medicinal potential but threatened. *International Journal of Pharmaceutical Sciences Review and Research*, 4(2), 9-17.
- Somchit, M., Sulaiman, M., Zuraini, A., Samsuddin, L., Somchit, N., Israf, D., & Moin, S. (2004). Antinociceptive and antiinflammatory effects of *Centella asiatica*. *Indian Journal of Pharmacology*, 36(6), 377.
- Suzuki, H., & Kou, K. (1983). Direct and indirect effects of histamine on the smooth muscle cells of the guinea-pig main pulmonary artery. *Pflügers Archiv*, 399(1), 46-53.
- Tamura, M., Sebastian, S., Yang, S., Gurates, B., Fang, Z., & Bulun, S. E. (2002). Interleukin-1 β elevates cyclooxygenase-2 protein level and enzyme activity via increasing its mRNA stability in human endometrial stromal cells: an effect mediated by extracellularly regulated kinases 1 and 2. *The journal of clinical endocrinology & metabolism*, 87(7), 3263-3273.

- Tousoulis, D., Kampoli, A.-M., Tentolouris Nikolaos Papageorgiou, C., & Stefanadis, C. (2012). The role of nitric oxide on endothelial function. *Current vascular pharmacology*, 10(1), 4-18.
- Valledor, A. F., Sánchez-Tilló, E., Arpa, L., Park, J. M., Caelles, C., Lloberas, J., & Celada, A. (2008). Selective roles of MAPKs during the macrophage response to IFN- γ . *The Journal of Immunology*, 180(7), 4523-4529.
- Van Furth, R., Cohn, Z., Hirsch, J., Humphrey, J., Spector, W., & Langevoort, H. (1972). The mononuclear phagocyte system: a new classification of macrophages, monocytes, and their precursor cells. *Bulletin of the World Health Organization*, 46(6), 845.
- van Hinsbergh, V. W. (1997). Endothelial permeability for macromolecules Mechanistic aspects of pathophysiological modulation. *Arteriosclerosis, thrombosis, and vascular biology*, 17(6), 1018-1023.
- van Hinsbergh, V. W., & van Nieuw Amerongen, G. P. (2002). Intracellular signalling involved in modulating human endothelial barrier function*. *Journal of anatomy*, 200(6), 549-560.
- van Nieuw Amerongen, G. P., Draijer, R., Vermeer, M. A., & van Hinsbergh, V. W. (1998). Transient and Prolonged Increase in Endothelial permeability induced by histamine and thrombin role of protein kinases, calcium, and RhoA. *Circulation research*, 83(11), 1115-1123.
- van Nieuw Amerongen, G. P., & van Hinsbergh, V. W. (2002). Targets for pharmacological intervention of endothelial hyperpermeability and barrier function. *Vascular pharmacology*, 39(4), 257-272.
- Vega-Ostertag, M., Casper, K., Swerlick, R., Ferrara, D., Harris, E. N., & Pierangeli, S. S. (2005). Involvement of p38 MAPK in the up-regulation of tissue factor on endothelial cells by antiphospholipid antibodies. *Arthritis & Rheumatism*, 52(5), 1545-1554.
- Villablanca, A. C., & Reid, T. W. (1997). Substance P stimulates vascular endothelial cellular reducing capacity in the presence of insulin and human plasma factors. *Journal of cellular biochemistry*, 66(4), 471-481.
- Visweswari, G., Prasad, K. S., Chetan, P. S., Lokanatha, V., & Rajendra, W. (2010). Evaluation of the anticonvulsant effect of *Centella asiatica* (gotu kola) in pentylenetetrazol-induced seizures with respect to cholinergic neurotransmission. *Epilepsy & Behavior*, 17(3), 332-335.
- Walsh, D. T., Weg, V. B., Williams, T. J., & Nourshargh, S. (1995). Substance P-induced inflammatory responses in guinea-pig skin: The effect of specific NK1 receptor antagonists and the role of endogenous mediators. *British journal of pharmacology*, 114(7), 1343-1350.
- Wan, J., Gong, X., Jiang, R., Zhang, Z., & Zhang, L. (2013). Antipyretic and Anti-inflammatory Effects of Asiaticoside in Lipopolysaccharide-treated Rat through Up-regulation of Heme Oxygenase-1. *Phytotherapy Research*, 27(8), 1136-1142.
- Weiming, X., Li Zhi, L., Loizidou, M., Ahmed, M., & Charles, I. G. (2002). The role of nitric oxide in cancer. *Cell research*, 12(5), 311-320.

- Weis, S. M. (2008). Vascular permeability in cardiovascular disease and cancer. *Current opinion in hematology*, 15(3), 243-249.
- White, M. V. (1990). The role of histamine in allergic diseases. *Journal of Allergy and Clinical Immunology*, 86(4), 599-605.
- WHO. (2002). WHO Pharmaceuticals Newsletter No. 3, 2002. In W. H. Organization (Ed.), (Vol. 3, pp. 18).
- Wojciak-Stothard, B., & Ridley, A. J. (2002). Rho GTPases and the regulation of endothelial permeability. *Vascular pharmacology*, 39(4), 187-199.
- Yong, Y. K., Sulaiman, N., Hakim, M. N., Lian, G. E. C., Zakaria, Z. A., Othman, F., & Ahmad, Z. (2013). Suppressions of Serotonin-Induced Increased Vascular Permeability and Leukocyte Infiltration by *Bixa orellana* Leaf Extract. *BioMed research international*, 2013.
- Yuan, & Sarah. (2002). Protein kinase signaling in the modulation of microvascular permeability. *Vascular pharmacology*, 39(4), 213-223.
- Yuan, R. R. (2010). *Chapter 5 Signaling Mechanisms in the Regulation of Endothelial* San Rafael (CA): Morgan & Claypool Life Sciences.
- Zainol, N., Voo, S., MR, S., & RA, A. (2008). Profiling of *Centella asiatica* (L.) Urban extract. *The Malaysian Journal of Analytical Sciences*, 12(2), 322-327.
- Zhang. (2007). Yin and yang interplay of IFN- γ in inflammation and autoimmune disease. *Journal of Clinical Investigation*, 117(4), 871.
- Zhang, Li, H.-z., Gong, X., Luo, F.-l., Wang, B., Hu, N., . . . Wan, J.-y. (2010). Protective effects of Asiaticoside on acute liver injury induced by lipopolysaccharide/D-galactosamine in mice. *Phytomedicine*, 17(10), 811-819.
- Zhang, Marconi, A., Xu, L. m., Yang, C. x., Sun, G. w., Feng, X. l., . . . d'Alessio, P. (2006). Tripterine inhibits the expression of adhesion molecules in activated endothelial cells. *Journal of leukocyte biology*, 80(2), 309-319.
- Zingarelli, B., Southan, G. J., Gilad, E., O'Connor, M., Salzman, A. L., & Szabö, C. (1997). The inhibitory effects of mercaptoalkylguanidines on cyclo-oxygenase activity. *British journal of pharmacology*, 120(3), 357-366.