



UNIVERSITI PUTRA MALAYSIA

***ANTIGENIC ANALYSIS OF OUTER MEMBRANE PROTEIN OF *Vibrio*
SPECIES AND DEVELOPMENT OF VERSATILE RECOMBINANT *vhDnaJ*
VACCINE AGAINST VIBRIOSIS***

FATHIN AMIRAH MURSID

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By

FATHIN AMIRAH MURSIDI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Master of Science**

February 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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February 2018

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Faculty : Agriculture

Vibriosis is one of the most catastrophic bacterial disease caused by infection of *Vibrio* spp. which frequently attacks marine cultures in all life stages. Recently, vibriosis was reported to cause vast mortality of tiger grouper cultured in deep sea cages in Langkawi, Malaysia. This disease frequently occurs during dry months when the water temperature is elevated. Although vibriosis is controlled through vaccination, the existence of different strains and antigenic diversities of *Vibrio* species and their serotypes have led to slow progress of vaccine development. Therefore, the development of a versatile vaccine that can fight against multiple *Vibrio* by eliciting protection against homologous and heterologous strains is urgently needed both for hindering vibriosis infections and to avoid the exploitation of antibiotics in aquaculture industry. Hence, the aim of this research was to search for the most antigenic OMPs protein among *Vibrio* sp. by analysing the protein's ability to elicit homologous and cross antigenicities and to develop a potentially versatile recombinant *Vibrio* vaccine. The safety of the developed vaccine was also assessed in gnotobiotic *Artemia* species. OMPs of *Vibrio harveyi* strain VH1, *V. alginolyticus* strain VA2, *V. parahaemolyticus* strain VPK1 and *Photobacterium damsela* strain PDS1 isolated from diseased groupers were extracted and characterized by SDS-PAGE and detection of immunogenic proteins were done by Western immunoblotting. The polyclonal antiserum of *V. harveyi* strain VH1 raised from rabbit induced strong antigenic responses on homologous OMPs antigens of *V. harveyi* strain VH1 and cross reacted against heterologous OMPs antigens of *V. parahaemolyticus* strain VPK1, *V. alginolyticus* strain VA2 and *P. damsela* strain PDS1 at molecular weight of 32 kDa. Therefore, further studies were conducted on the antigenically heterologous 32 kDa OMP of *V. harveyi* strain VH1 as a potential vaccine candidate. The 32 kDa protein band was a molecular chaperone DnaJ, designated as vhDnaJ after submitted for N-terminal amino acid sequencing analysis.

The vhDnaJ gene was amplified and cloned in pET-32 Ek/LIC vector and expressed in host BL21 (DE3) *Escherichia coli*. Bioinformatics analysis indicated that the target gene was highly conserved among *Vibrio* sp. and highly antigenic by comprising 40 antigenic sites. Successful recombinant vhDnaJ protein expression expressed under 30°C for 10 hour was detected at 49 kDa band by SDS-PAGE and Western immunoblotting by using Anti-HisTag monoclonal antibodies. The bioencapsulation of the inactivated recombinant vhDnaJ cells vaccine into *Artemia* sp. demonstrated that the species could survive up to $\pm 83.3\%$ after 36 h post-encapsulation, signifying the vaccine was safe and might be beneficial to the host. In conclusion, the cumulative evidences of the 32 kDa OMP of *V. harveyi* strain VH1 by being the most antigenic against homologous and heterologous isolates and highly conserved among the tested *Vibrio* strains in *in-vitro* antigenicity and bioinformatics study could be a promising versatile vaccine antigen against multiple *Vibrio* sp. in grouper culture. Moreover, the successful expression of the protein of interest and verified safety of the developed recombinant vhDnaJ vaccine in *Artemia* sp. open the way for future preparation of crude recombinant vaccine as well as to assess its efficacy in marine fish.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**ANALISIS KEANTIGENAN PROTIN MEMBRANE LUAR DARI *Vibrio*
SPESIES DAN PEMBANGUNAN REKOMBINAN VAKSIN vhDnaJ YANG
VERSATIL MELAWAN VIBRIOSIS**

Oleh

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Vibriosis adalah salah satu penyakit yang amat merbahaya berpunca daripada jangkitan bakteria *Vibrio* spp. yang kebiasaannya menyerang spesies marin. Baru-baru ini, vibriosis yang menyebabkan kematian sejumlah besar ikan kerapu yang dibiakkan di sangkar laut dalam telah dilaporkan di Langkawi, Malaysia. Wabak vibriosis ini kebiasaannya berlaku apabila suhu air meningkat. Walaupun vibriosis telah dikawal melalui pemvaksinan, kewujudan strain yang berbeza dan kepelbagaian antigenik dalam *Vibrio* sp. dan serotip telah menyebabkan pembangunan vaksin semakin perlahan. Justeru itu, pembangunan vaksin versatil yang boleh melawan pelbagai patogen dengan menghasilkan perlindungan antigenik menentang homolog dan heterologus strain diperlukan dengan segera untuk menghalang jangkitan vibriosis dan mengelakkan penyalahgunaan antibiotik dalam industri akuakultur. Oleh itu, tujuan penyelidikan ini adalah untuk menyelidik protin yang paling antigenik dengan menganalisis kemampuan protin membran luar (OMP) daripada *Vibrio* sp. yang berbeza untuk menghasilkan reaksi homologi dan silang dan membangunkan rekombinan vaksin yang mempunyai potensi serbaguna. Keselamatan vaksin tersebut juga telah dinilai dengan menggunakan gnotobiotik *Artemia* model. OMP daripada *Vibrio harveyi* strain strain VH1, *V. alginolyticus* strain VA2, *V. parahaemolyticus* strain VPK1 dan *Photobacterium damsela* strain PDS1 yang telah dipisahkan daripada kerapu berpenyakit telah diekstrak dan dicirikan menggunakan kaedah sodium dodecyl sulfat-gel elektroforesis poliakrilamide (SDS-PAGE) dan pengesanan protin imunogenik melalui kaedah Pemblotan Western. Antiserum poliklonal daripada *V. harveyi* strain VH1 yang dibangunkan daripada arnab telah merangsang gerak balas yang kuat terhadap antigen protin *V. harveyi* strain VH1 dan reaksi silang terhadap protin antigen *V. parahaemolyticus* strain VPK1, *V. alginolyticus* strain VA2 dan *P. damsela* strain PDS1 di kedudukan jalur 32 kDa. Oleh itu, kajian seterusnya telah dijalankan

dengan memilih jalur protein 32 kDa *V. harveyi* strain VH1 sebagai calon vaksin. Jaluran protein tersebut telah diproses melalui penjurukan amino asid Terminal-N dan keputusan menunjukkan protein tersebut adalah “molekul pengiring DnaJ”. Gen tersebut telah diamplifikasikan dan diklonkan kedalam vector pET-32 EK/LIC sebelum dimasukkan kedalam perumah ekspresi BL21 (DE3) *Escherichia coli*. Bioinformatik analisis menunjukkan bahawa gen tersebut sangat terpelihara di kalangan *Vibrio* sp. dan sangat antigenik dengan memiliki 40 tapak antigenik. Protein yang telah berjaya diekspresikan dibawah suhu 30°C selama 10 jam dianalisa melalui “SDS-PAGE” dan Pemblotan Western menggunakan monoclonal antibodi Anti-HisTag telah menunjukkan jalur protein gabungan yang sangat menonjol pada saiz 49 kDa, yang mengandungi 32 kDa protein vhDnaJ dan protein Tag bersaiz 17 kDa. Rekombinan vhDnaJ vaksin yang tidak aktif dimasukkan ke dalam *Artemia* sp. secara bio menunjukkan spesies tersebut mampu hidup sebanyak $\pm 83.3\%$ selepas 36 jam, menunjukkan bahawa vaksin tersebut selamat dan mungkin memberi kesan yang baik kepada *Artemia*. Kesimpulannya, bukti-bukti yang terkumpul menunjukkan bahawa OMP *V. harveyi* strain VH1 bersaiz 32 kDa adalah paling antigenik dan paling terpelihara di antara *Vibrio* strain berdasarkan ujikaji antigenik dan bioinformatik *in-vitro* dan boleh menjadi antigen vaksin yang bagus serta berpotensi untuk melawan pelbagai *Vibrio* dalam ternakan kerapu. Tambahan pula, pada masa hadapan, ekspresi protein pilihan dan rekombinan vhDnaJ vaksin telah terbukti selamat dan telah membuka jalan untuk penghasilan vaksin rekombinan mentah dan untuk menilai keberkesanan vaksin tersebut dalam ternakan ikan marin.

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LIST OF ABBREVIATIONS/NOTATIONS/GLOSSARY OF TERMS

%	Percentage
µg	microgram
µl	microliter
µM	micromolar
ASW	Artificial sterile water
BHI	Brain-heart infusion
BLAST	Basic Local Alignment Search Tool
Blastp	protein-protein BLAST
bp	base pair
BSA	bovine serum albumin
CFU	colony forming units
DAB	3,3'-Diaminobenzidine
dATP	deoxyadenosine triphosphate
dNTP	deoxynucleotide triphosphate
DNA	deoxyribonucleic acid
<i>E. coli</i>	<i>Escherichia coli</i>
<i>E. fuscoguttatus</i>	<i>Epinephelus fuscoguttatus</i>
<i>E. lanceolatus</i>	<i>Epinephelus lanceolatus</i>
FKC	formalin-killed cells
H ₂ SO ₄	sulphuric acid
Ig	immunoglobulin
IHN	infectious haematopoietic necrosis
IP	intraperitoneal
IPTG	isopropyl β-D-1-thiogalactopyranoside
kb	kilobase pair
kDa	kilodalton
LB	Luria Bertani
LIC	ligation-independent cloning

M	molar
MCS	multiple cloning site
mg	milligram
MgCl ₂	magnesium chloride
MgSO ₄	magnesium sulfate
mM	millimolar
NaCl	sodium chloride
Na ₂ S ₂ O ₃	sodium thiosulfate
NaOCl	Sodium hypochlorite
ng	nanogram
nm	nanometer
OD	Optical density
OMP	Outer membrane protein
PBS	phosphate buffer saline
PCR	polymerase chain reaction
<i>P. damsela</i>	<i>Photobacterium damsela</i>
pmol	picomole
RE	restriction enzyme
rpm	revolutions per minute
RPS	relative percentage survival
SDS-PAGE	Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis
SOC	Super Optimal broth with Catabolite repression
<i>sp.</i>	species
TBE	tris-boric EDTA
TBS	tris-buffer saline
TCBS	thiosulfate-citrate-bile salts-sucrose
TMB	3,3',5,5'-tetramethylbenzidine
TSA	tryptic soy agar
TSB	tryptic soy broth

U	unit
V	voltage
<i>V. alginolyticus</i>	<i>Vibrio alginolyticus</i>
<i>V. harveyi</i>	<i>Vibrio harveyi</i>
<i>V. parahaemolyticus</i>	<i>Vibrio parahaemolyticus</i>
v/v	volume per volume
w/v	weight per volume
$x g$	multiples of gravitational acceleration



CHAPTER 1

INTRODUCTION

For many decades, aquaculture industry has blooming boundlessly throughout the world primarily in Asia. Marine grouper culture which belong to family Serranidae, subfamily Epinephelinae has recognized to be among the top species in aquaculture production in most Asian countries (Harikrishnan et al., 2011). The high interest towards grouper aquacultures has encouraged Malaysian government to increase hatcheries number and has provided infrastructures for aquacultures development (Pomeroy, 2002).

Nevertheless, intensive farming of groupers cultivated in net cages in the brackish water plus in limited area have made them to become very susceptible to disease infection, especially by bacterial *Vibrio* species causing strenuous mortality that totally disrupting both economic and social development as well as food safety worldwide (Chatterjee and Haldar, 2012; Li et al., 2015). Vibriosis, caused by infection of *Vibrio* sp. has been extensively reported as a major threatening and most catastrophic diseases to grouper culture involving grouper fry, fingerlings, juveniles, adults and brood stocks (Sarjito et al., 2009; Ilmiah et al., 2013; Novriadi and Haw, 2014). *Vibrio* species like *Vibrio harveyi*, *V. alginolyticus*, *V. parahaemolyticus*, *V. vulnificus*, *V. anguillarum*, and *Photobacterium damsela* have long been recognized as the main pathogens in *Epinephelus* sp. and other marine species causing severe gastroenteritis syndrome and haemorrhagic septicaemia (Ningqiu et al., 2008; Li et al., 2010; Hu et al., 2012; Huang et al., 2012; Peng et al., 2016; Pang et al., 2016).

A case study in Malaysia reported only 30-40% cage-cultured groupers could survive due to various diseases including vibriosis (Liao and Leano, 2008). Besides that, Abdullah et al., (2017) reported cage-cultured groupers and snappers in fish farm in Langkawi Island in Kedah, Malaysia were majorly infected with vibriosis along with viruses for up to 77.78% and 76.05%, respectively. Abdullah et al. (2015) also reported three main *Vibrio* sp. (*V. vulnificus*, *V. alginolyticus*, and *P. damsela*) were isolated from diseased tiger grouper after vibriosis outbreak in deep sea cage in Langkawi with prevalence rates from 50% to 90%. Another case of vibriosis outbreaks reported in Malaysia discovered *V. harveyi* as the main culprit followed with *V. parahaemolyticus*, *V. alginolyticus*, *V. ponticus*, *V. fluvialis* and many other *Vibrio* sp. infecting Asian seabass (*Lates calcarifer*), orange-spotted grouper (*Epinephelus coioides*), red snapper (*Lutjanus* sp.), brown marble grouper (*Epinephelus fuscoguttatus*), and hybrid grouper (*E. fuscoguttatus* x *E. lanceolatus*) (Albert and Ransangan, 2013). This disease also commonly reported infecting grouper (*Epinephelus* sp.) (Huang et al., 2012), European sea bass (*Dicentrarchus labrax*) (Bellos et al., 2015), red sea bream (*Pagrus major*) (El-Galil and Mohamed, 2012), and catfish (*Siluriformes*) (Geng et al., 2014). Despite of this problem, government of Malaysia has boost the research and development (R&D) on the frequently isolated *Vibrio* sp. to further study on their virulence properties and

immunogenic potential for preventative measures against infection in mariculture species including grouper to maintain healthy broodstock and disease-free seed and fingerling (Pomeroy, 2002).

Preventions of vibriosis are mainly dependent on antibiotics which are still effective, however the usage are strictly not recommended since the extensive use of the antimicrobial drugs have led to the emergence of drug resistance strains and pose risk to horizontally transfer the resistance gene to fish and human's pathogen (Ningqiu et al., 2008, Maiti et al., 2012; Li et al., 2014). Moreover, application of antibiotics also caused problems of food safety and environmental pollution as well (Evelyn, 1996). Bacterial resistance to common antibiotics has reached frightening levels in multiple countries around the world making the antibiotics treatment to common diseases is no longer effective (WHO, 2014). Many studies have reported some *Vibrio* sp. such as *V. harveyi*, *V. vulnificus* and *V. parahaemolyticus* exhibit multiple antibiotic resistance in aquaculture productions (Elmahdi et al., 2016). Thus, new strategies to avoid the massive misuse of antibiotics to control infection in aquaculture are urgently needed. Vaccination is an alternative prophylactic measure with a safer strategy to control diseases by increasing antibody titre, boost immunological memory, and enhance rate of survival of infected fish (Defoirdt et al., 2007). Fish vaccination is now approved as a standard protocol in modern aquaculture (Huang et al., 2012).

Although vibriosis has been administered by vaccination, however, the existence of different strains and serotypes of *Vibrio* sp. have contributed major challenges in vaccine development. Moreover, the existence of antigenic diversity of *Vibrio* strains and their serotypes have made the vaccines unable to elicit protection against multiple vibrios resulting in slow progress of vaccine development against vibriosis (Li et al., 2014). Furthermore, according to Li et al. (2010), there are no commercial vibrio vaccines that are versatile available at the moment. Until this time, studies on vaccines that could provide cross-protections have been widely reported, however most investigations were mainly focused on cross protection against homologous strains or serotype (Mutharia et al., 1993; Sabri et al., 2000; Sun et al., 2012). The studies on cross protective ability against heterogenous serotypes and species on *Vibrio* sp. are still scarce. Therefore, more efforts are needed to develop a powerful and versatile subunit vaccine that could provide cross protection against multiple *Vibrio* strains and serotypes to combat vibriosis in fish.

Outer membrane protein (OMP), a unique composition of Gram-negative bacteria is ideally located on the cell surface of bacteria and highly immunogenic. These characteristics have made this protein to be extensively used in all range of studies of antimicrobial drugs and vaccination. OMP has been revealed to provide homologous and heterologous protection and could act as polyvalent immunogens against diverse Gram-negative bacteria (Xu et al., 2005; Li et al., 2009). For this reason, more researches have been focused on determination of immunogenic characteristics of OMPs. Moreover, interests have been growing to develop a recombinant vaccine expressing immunogenic outer membrane protein (OMPs) that elicit homologous

and cross antigenicity since this type of vaccine has been proven to induce protective efficacy against multiple bacterial species and serotypes. Recombinant cells allow production of protein in large amount which further increase antibody reaction, thus intensify protection. Moreover, in recent years, the development of versatile recombinant vaccines were concentrated on conserved OMPs antigen that exist in *Vibrio* pathogens and their serotypes since such protein could provoke highly effective immune response and able to defend against different pathogens (Li et al., 2010; Lun et al., 2014; Zha et al., 2016). Li et al. (2010) had reported recombinant ompK was a versatile vaccine candidate by providing cross protection to heterogeneous virulent *V. harveyi*, *V. alginolyticus*, and *V. parahaemolyticus* in orange spotted grouper (*Epinephelus coioides*). Another versatile vibriosis vaccine was reported by Lun et al. (2014) where immunization of zebrafish (*Danio reiro*) using recombinant LamB, a family of OMPs that was conserved antigen among various *Vibrio* sp. showed significant protection against vibriosis which was found in *V. parahaemolyticus* RIMD2210633. Therefore, this study was aimed to develop a versatile recombinant vaccine against vibriosis through the search of the most antigenic protein from different strains of *Vibrio* sp. isolated from diseased marine fish by analysing the ability of their OMPs in eliciting homologous and cross antigenicity and to develop versatile recombinant *Vibrio* vaccine as well as to assess the safety of the developed vaccine in gnotobiotic *Artemia* sp. before it can be applied into final host (fish). The objectives of this study were:

1. To characterize the outer membrane protein (OMPs) profiles of *Vibrio harveyi* strain VH1, *V. parahaemolyticus* strain VPK1, *V. alginolyticus* strain VA2, and *Photobacterium damsela* strain PDS1 isolated from diseased grouper (*Epinephelus fuscoguttatus*) and to determine homologous and cross antigenicity of the OMPs against homologous and heterologous antisera.
2. To sequence the most antigenic protein by N-terminal amino acid sequencing and to construct a recombinant cell vaccine containing the antigenic outer membrane protein vhDnaJ gene of *V. harveyi* strain VH1 in prokaryotic expression system for production of potential recombinant vaccine.
3. To evaluate the safety of formalin-killed recombinant vhDnaJ vaccine based on survivability of gnotobiotic *Artemia* model.

Hypothesis:

H₀: Outer membrane protein of *V. harveyi* strain VH1 could not elicit homologous and heterologous antigenicity against polyclonal rabbit antisera of different *Vibrio* strains and formalin-killed recombinant vhDnaJ cells vaccine could not increase the survivability of gnotobiotic *Artemia* nauplii.

H_A: Outer membrane protein of *V. harveyi* strain VH1 could elicit homologous and heterologous antigenicity against polyclonal rabbit antisera of different *Vibrio* strains and formalin-killed recombinant vhDnaJ cells vaccine could increase the survivability of gnotobiotic *Artemia* nauplii.

REFERENCES

- Abdel-Aziz, M., Eissa, A. E., Hanna, M., & Okada, M. A. (2013). Identifying some pathogenic *Vibrio/Photobacterium* species during mass mortalities of cultured Gilthead seabream (*Sparus aurata*) and European seabass (*Dicentrarchus labrax*) from some Egyptian coastal provinces. *International Journal of Veterinary Science and Medicine*, 1(2): 87-95.
- Abdullah, A., Mansor, N. N., Ramli, R., Ridzuan, M. S., Saad, M. Z., & Abdullah, S. Z. (2015). *Concurrent Infections in Tiger Grouper (Epinephelus fuscoguttatus) Cultured in Deep Sea Cages in Langkawi*. Paper presented at International Fisheries Symposium (IFS 2015), The Gurney Hotel, Penang. December 2015.
- Abdullah, A., Ramli, R., Ridzuan, M. S. M., Murni, M., Hashim, S., Sudirwan, F., & Amal, M. N. A. (2017). The presence of Vibrionaceae, Betanodavirus and Iridovirus in marine cage-cultured fish: role of fish size, water physicochemical parameters and relationships among the pathogens. *Aquaculture Reports*, 7: 57-65.
- Abiodun, A. A., Yusoff, M., Sabri, M., Babatunde, D. A., Md Yasin, I. S., & Abu Hassim, H. (2016). Immunoprophylaxis: a better alternative protective measure against shrimp vibriosis—a review. *Pertanika Journal of Scholarly Research Reviews*, 2(2): 57-68.
- Aguirre- Guzmán, G., Mejia Ruíz, H., & Ascencio, F. (2004). A review of extracellular virulence product of *Vibrio* species important in diseases of cultivated shrimp. *Aquaculture Research*, 35(15): 1395-1404.
- Albert, V., & Ransangan, J. (2013). Effect of water temperature on susceptibility of culture marine fish species to vibriosis. *International Journal of Research in Pure and Applied Microbiology*, 3(3): 48-52.
- Arulvasu, C., Shobana, S., Banu, H. A. A., Chandhirasekar, D., & Prabhu, D. (2012). Bioencapsulation of *Artemia* nauplii with herbal extract for promoting growth of fish fry *Poecilia sphenops* val. *Journal of Modern Biotechnology*, 1: 37-44.
- Arunagiri, K., Jayashree, K., & Sivakumar, T. (2013). Isolation and identification of Vibrios from marine food resources. *International Journal Current Microbiology Application Science*, 2(7): 217-32.
- Asensio, L., González, I., Fernández, A., Rodríguez, M. A., Hernández, P. E., García, T., & Martín, R. (2001). PCR-SSCP: a simple method for the authentication of grouper (*Epinephelus guaza*), wreck fish (*Polyprion americanus*), and Nile perch (*Lates niloticus*) fillets. *Journal of Agricultural and Food Chemistry*, 49(4): 1720-1723.

- Aslizah, M.A., Ina-Salwany, M.Y., Zamri-Saad, M., Hassan, M.D., & Norfarrah, M.A. (2016). Molecular characterization of *Vibrio harveyi* virulence associated serine protease and outer membrane protein genes for vaccine development. *International Journal of Biosciences*, 8(3): 2222-5234
- Austin, B., & Zhang, X. H. (2006). *Vibrio harveyi*: a significant pathogen of marine vertebrates and invertebrates. *Letters in Applied Microbiology*, 43 (2): 119-124.
- Austin, B., Austin, D. A., Austin, B., & Austin, D. A. (2012). Bacterial fish pathogens, *Springer* (pp. 652). Heidelberg, Germany
- Austin, B., Austin, D., Sutherland, R., Thompson, F., & Swings, J. (2005). Pathogenicity of vibrios to rainbow trout (*Oncorhynchus mykiss*, Walbaum) and *Artemia nauplii*. *Environmental Microbiology*, 7(9): 1488-1495.
- Beamer, L. J., Carroll, S. F., & Eisenberg, D. (1998). The BPI/LBP family of proteins: a structural analysis of conserved regions. *Protein Science*, 7(4): 906-914.
- Bellos, G., Angelidis, P., & Miliou, H. (2015). Effect of temperature and seasonality principal epizootiological risk factor on vibriosis and photobacteriosis outbreaks for European sea bass in Greece (1998-2013). *Journal of Aquaculture Research and Development*, 6(5): 338.
- Benedetti, L. S. (2013). *Marine Protected Areas (MPAs) as a Fisheries Management Tool for the Nassau Grouper (Epinephelus striatus) in Belize*. (Unpublished doctoral dissertation). University of Laurentian, Ontario, Canada.
- Bondad-Reantaso, M. G., Subasinghe, R. P., Arthur, J. R., Ogawa, K., Chinabut, S., Adlard, R. & Shariff, M. (2005). Disease and health management in Asian aquaculture. *Veterinary Parasitology*, 132(3): 249-272.
- Cai, S. H., Lu, Y. S., Wu, Z. H., & Jian, J. C. (2013). Cloning, expression of *Vibrio alginolyticus* outer membrane protein- OmpU gene and its potential application as vaccine in crimson snapper, *Lutjanus erythropterus* Bloch. *Journal of Fish Diseases*, 36(8): 695-702.
- Cai, S. H., Wu, Z. H., Jian, J. C., & Lu, Y. S. (2007). Cloning and expression of the gene encoding an extracellular alkaline serine protease from *Vibrio alginolyticus* strain HY9901, the causative agent of vibriosis in *Lutjanus erythropterus* (Bloch). *Journal of Fish Diseases*, 30(8): 493-500.
- Caipang, C. M. A., Jomer, B. L., & Clara, L. M. (2014). Updates on the vaccination against bacterial diseases in tilapia, *Oreochromis spp.* and Asian seabass, *Lates calcarifer*. *AACL Bioflux*, 7(3): 184-193.
- Campbell, R., Adams, A., Tatner, M. F., Chair, M., & Sorgeloos, P. (1993). Uptake of *Vibrio anguillarum* vaccine by *Artemia salina* as a potential oral delivery system to fish fry. *Fish & Shellfish Immunology*, 3(6): 451-459.
- Casali, N., & Preston, A. (2003). *E. coli* plasmid vectors: methods and applications (Vol. 235). Springer Science & Business Media.

- Ceccarelli, D., Hasan, N. A., Huq, A., & Colwell, R. R. (2013). Distribution and dynamics of epidemic and pandemic *Vibrio parahaemolyticus* virulence factors. *Frontiers in Cellular and Infection Microbiology*, 3: 97.
- Chair, M., Gapasin, R. S. J., Dehasque, M., & Sorgeloos, P. (1994). Vaccination of European sea bass fry through bioencapsulation of *Artemia* nauplii. *Aquaculture International*, 2(4): 254-261.
- Chatterjee, S. N., & Chaudhuri, K. (2012). Gram-negative bacteria: The cell membranes. In Springer Berlin Heidelberg, *Outer Membrane Vesicles of Bacteria*, (pp. 15-34).
- Chatterjee, S., & Halder, S. (2012). *Vibrio* related diseases in aquaculture and development of rapid and accurate identification methods. *Journal of Marine Science Research and Development* S, 1.
- Chen, M., Daha, M. R., & Kallenberg, C. G. (2010). The complement system in systemic autoimmune disease. *Journal of Autoimmunity*, 34(3): 276-286.
- Chen, Q., Yan, Q., Wang, K., Zhuang, Z., & Wang, X. (2008). Portal of entry for pathogenic *Vibrio alginolyticus* into large yellow croaker *Pseudosciaena crocea*, and characteristics of bacterial adhesion to mucus. *Diseases of Aquatic Organisms*, 80(3): 181-188.
- Chen, Y. M., Shih, C. H., Liu, H. C., Wu, C. L., Lin, C. C., Wang, H. C., & Lin, J. H. Y. (2011). An oral nervous necrosis virus vaccine using *Vibrio anguillarum* as an expression host provides early protection. *Aquaculture*, 321(1): 26-33.
- Cheng, A. C., Cheng, S. A., Chen, Y. Y., & Chen, J. C. (2009). Effects of temperature change on the innate cellular and humoral immune responses of orange-spotted grouper *Epinephelus coioides* and its susceptibility to *Vibrio alginolyticus*. *Fish and Shellfish Immunology*, 26(5): 768-772.
- Chifiriuc, M. C., Pircalabioru, G., Lazăr, V., Gicirc, B., Dascălu, L., Enache, G., & Bleotu, C. (2011). Immunogenicity of different cellular fractions of *Vibrio parahaemolyticus* strains grown under sub-lethal heat and osmotic stress. *African Journal of Microbiology Research*, 5(1): 65-72.
- Chinabut, S. (2001). Health management for sustainable aquaculture. In *Responsible Aquaculture Development in Southeast Asia*. Proceedings of the Seminar-Workshop: *Aquaculture Development in Southeast Asia organized by the SEAFDEC Aquaculture Department, 12-14 October 1999, Iloilo City, Philippines*. SEAFDEC Aquaculture Department.
- Cline, J., Braman, J. C., & Hogrefe, H. H. (1996). PCR fidelity of Pfu DNA polymerase and other thermostable DNA polymerases. *Nucleic Acids Research*, 24(18): 3546-3551.

- Dan, H., Balachandran, A., & Lin, M. (2009). A pair of ligation-independent *Escherichia coli* expression vectors for rapid addition of a polyhistidine affinity tag to the N-or C-termini of recombinant proteins. *J Biomol Tech*, 20: 241-248.
- Dang, W., Zhang, M., & Sun, L. (2011). *Edwardsiella tarda* DnaJ is a virulence-associated molecular chaperone with immunoprotective potential. *Fish and Shellfish Immunology*, 31(2): 182-188.
- Davies, R. L. (1991). Outer membrane protein profiles of *Yersinia ruckeri*. *Veterinary Microbiology*, 26(1-2): 125-140.
- Defoirdt, T., Boon, N., Sorgeloos, P., Verstraete, W., & Bossier, P. (2007). Alternatives to antibiotics to control bacterial infections: luminescent vibriosis in aquaculture as an example. *Trends in Biotechnology*, 25(10): 472-479.
- Defoirdt, T., Bossier, P., Sorgeloos, P., & Verstraete, W. (2005). The impact of mutations in the quorum sensing systems of *Aeromonas hydrophila*, *Vibrio anguillarum* and *Vibrio harveyi* on their virulence towards gnotobiotically cultured *Artemia franciscana*. *Environmental Microbiology*, 7(8): 1239-1247.
- Dhont, J., & Sorgeloos, P. (2002). Applications of *Artemia*. In *Artemia: Basic and applied biology* (pp. 251-277). Springer Netherlands.
- Dhont, J., Dierckens, K., Støttrup, J., Van Stappen, G., Wille, M., & Sorgeloos, P. (2013). Rotifers, *Artemia* and copepods as live feeds for fish larvae in aquaculture. *Advances in Aquaculture Hatchery Technology*: 157-202.
- El-Galil, M. A., & Mohamed, M. H. (2012). First Isolation of *Vibrio alginolyticus* from Ornamental Bird Wrasse Fish (*Gomphosus caeruleus*) of the Red Sea in Egypt. *Journal of Fisheries and Aquatic Science*, 7(6): 461.
- Elmahdi, S., DaSilva, L. V., & Parveen, S. (2016). Antibiotic resistance of *Vibrio parahaemolyticus* and *Vibrio vulnificus* in various countries: A review. *Food Microbiology*, 57: 128-134.
- Esteve-Gassent, M. D., & Amaro, C. (2004). Immunogenic antigens of the eel pathogen *Vibrio vulnificus* serovar E. *Fish and Shellfish Immunology*, 17(3): 277-291.
- Evelyn, T. P. (1996). A historical review of fish vaccinology. *Developments In Biological Standardization*, 90: 3-12.
- Fang, H. M., Ling, K. C., Ge, R., & Sin, Y. M. (2000). Enhancement of protective immunity in blue gourami, *trichogaster trichopterus* (pallas), against *Aeromonas hydrophila* and *Vibrio anguillarum* by *A. hydrophila* major adhesin. *Journal of Fish Diseases*, 23(2): 137-145.
- FAO (Fisheries and Aquaculture Department). (2016). National Aquaculture Sector Overview. Malaysia. National Aquaculture Sector Overview Fact Sheets. *Fisheries and Aquaculture Information and Statistics Branch*.

- FAO FishStatJ. (2013). A tool for fishery statistics analysis. FAO Fisheries and Aquaculture Department. FIPS - Statistics and information, Rome.
- Farmer Iii, J. J., & Hickman-Brenner, F. W. (2006). The genera *Vibrio* and *Photobacterium*. *The prokaryotes* (pp. 508-563). Springer New York.
- Farzanfar, A. (2006). The use of probiotics in shrimp aquaculture. *Pathogens and Disease*, 48(2): 149-158.
- Feder, M. E., & Hofmann, G. E. (1999). Heat-shock proteins, molecular chaperones, and the stress response: evolutionary and ecological physiology. *Annual Review of Physiology*, 61(1): 243-282.
- Firdaus-Nawi, M., Zamri-Saad, M., Nik-Haiha, N. Y., Zuki, M. A. B., & Effendy, A. W. M. (2013). Histological assessments of intestinal immuno-morphology of tiger grouper juvenile, *Epinephelus fuscoguttatus*. *SpringerPlus*, 2(1): 611.
- Fouz, B., & Amaro, C. (2003). Isolation of a new serovar of *Vibrio vulnificus* pathogenic for eels cultured in freshwater farms. *Aquaculture*, 217(1): 677-682.
- Frans, I., Michiels, C. W., Bossier, P., Willems, K. A., Lievens, B., & Rediers, H. (2011). *Vibrio anguillarum* as a fish pathogen: virulence factors, diagnosis and prevention. *Journal of Fish Diseases*, 34(9): 643-661.
- Galdiero, S., Falanga, A., Cantisani, M., Tarallo, R., Elena Della Pepa, M., D'Oriano, V., & Galdiero, M. (2012). Microbe-host interactions: structure and role of Gram-negative bacterial porins. *Current Protein and Peptide Science*, 13(8): 843-854.
- Gatewood, D. M., Fenwick, B. W., & Chengappa, M. M. (1994). Growth-condition dependent expression of *Pasteurella haemolytica* A1 outer membrane proteins, capsule, and leukotoxin. *Veterinary Microbiology*, 41(3): 221-233.
- Geng, Y., Liu, D., Han, S., Zhou, Y., Wang, K. Y., Huang, X. L., & Lai, W. M. (2014). Outbreaks of vibriosis associated with *Vibrio mimicus* in freshwater catfish in China. *Aquaculture*, 433: 82-84.
- Gomez-Gil, B., Herrera-Vega, M. A., Abreu-Grobois, F. A., & Roque, A. (1998). Bioencapsulation of Two Different *Vibrio* species in nauplii of the Brine Shrimp (*Artemia franciscana*). *Applied and Environmental Microbiology*, 64(6): 2318-2322.
- Gomez-Gil, B., Soto-Rodríguez, S., Lozano, R., & Betancourt-Lozano, M. (2014). Draft genome sequence of *Vibrio parahaemolyticus* strain M0605, which causes severe mortalities of shrimps in Mexico. *Genome Announcements*, 2(2): 55-14.
- Gomez-Gil, B., Thompson, C. C., Matsumura, Y., Sawabe, T., Iida, T., Christen, R., & Sawabe, T. (2014). The Family Vibrionaceae. In *The prokaryotes* (pp. 659-747). Springer Berlin Heidelberg.

- Gopal, G. J., & Kumar, A. (2013). Strategies for the production of recombinant protein in *Escherichia coli*. *The protein Journal*, 32(6): 419-425.
- Govindaraju, G. S., & Jayasankar, P. (2004). Taxonomic relationship among seven species of groupers (genus *Epinephelus*; family Serranidae) as revealed by RAPD fingerprinting. *Marine Biotechnology*, 6(3): 229-237.
- Gräslund, S., Nordlund, P., Weigelt, J., Bray, J., Gileadi, O., Knapp, S., & Park, H. W. (2008). Protein production and purification. *Nature Method*, 5(2): 135-146.
- Grisez, L., & Tan, Z. I. L. O. N. G. (2005). Proceedings of the Fifth Symposium in Asian Aquaculture Edited by: Walker P, Lester R, Bondad-Reantaso MG. Goldcoast, Australia: *Vaccine development for Asian aquaculture*.
- Gudding, R., & Van Muiswinkel, W. B. (2013). A history of fish vaccination: science-based disease prevention in aquaculture. *Fish and Shellfish Immunology*, 35(6): 1683-1688.
- Gudding, R., Lillehaug, A., & Evensen, O. (Eds.). (2014). *Fish vaccination*. John Wiley & Sons.
- Gunter, D. (1980). The interaction of *Vibrio* with *Artemia nauplii*. *The Brine Shrimp Artemia Morphology, Genetics, Radiobiology, Toxicology*, 1: 318.
- Haenen, O., Fouz, B., Amaro, C., Isern, M. M., Mikkelsen, H., Zrnčić, S., & Hellstrom, A. (2014). Vibriosis in aquaculture. 16th EAAP Conference, Tampere, Finland, 4th September 2013: workshop report. *Bulletin of the European Association of Fish Pathologists*, 34(4): 138-148.
- Haldar, S., Chatterjee, S., Sugimoto, N., Das, S., Chowdhury, N., Hinenoya, A., & Yamasaki, S. (2011). Identification of *Vibrio campbellii* isolated from diseased farm-shrimps from south India and establishment of its pathogenic potential in an *Artemia* model. *Microbiology*, 157(1): 179-188.
- Hanafī, H. H., Arshad, M. A., and Yahaya, S. Regional Study and Workshop on the Environmental Assessment and Management of Aquaculture Development (TCP/RAS/2253). Network of Aquaculture Centres in Asia-Pacific, Bangkok, Thailand. 1995.
- Harikrishnan, R., Balasundaram, C., & Heo, M. S. (2011). Fish health aspects in grouper aquaculture. *Aquaculture*, 320(1): 1-21.
- Hashem, M., & El-Barbary, M. (2013). *Vibrio harveyi* infection in Arabian Surgeon fish (*Acanthurus sohal*) of Red Sea at Hurghada, Egypt. *The Egyptian Journal of Aquatic Research*, 39(3): 199-203.
- Heemstra, P. C., & Randall, J. E. (1993). FAO species catalogue. v. 16: Groupers of the world (Family Serranidae, Subfamily Epinephelinae).
- Hengen, P. N. (1995). Purification of His-Tag fusion proteins from *Escherichia coli*. *Trends in Biochemical Sciences*, 20(7): 285-286.

- Hoel, K., Reitan, L. J., & Lillehaug, A. (1998). Immunological cross reactions between *Aeromonas salmonicida* and *Vibrio salmonicida* in Atlantic salmon (*Salmo salar* L.) and rabbit. *Fish and Shellfish Immunology*, 8(3): 171-182.
- Hu, Y. H., & Sun, L. (2011). A bivalent *Vibrio harveyi* DNA vaccine induces strong protection in Japanese flounder (*Paralichthys olivaceus*). *Vaccine*, 29(26): 4328-4333.
- Hu, Y. H., Deng, T., Sun, B. G., & Sun, L. (2012). Development and efficacy of an attenuated *Vibrio harveyi* vaccine candidate with cross protectivity against *Vibrio alginolyticus*. *Fish and shellfish immunology*, 32(6): 1155-1161.
- Huang, H. Y., Chen, Y. C., Wang, P. C., Tsai, M. A., Yeh, S. C., Liang, H. J., & Chen, S. C. (2014). Efficacy of a formalin-inactivated vaccine against *Streptococcus iniae* infection in the farmed grouper *Epinephelus coioides* by intraperitoneal immunization. *Vaccine*, 32(51): 7014-7020.
- Huang, Z., Tang, J., Li, M., Fu, Y., Dong, C., Zhong, J. F., & He, J. (2012). Immunological evaluation of *Vibrio alginolyticus*, *Vibrio harveyi*, *Vibrio vulnificus* and infectious spleen and kidney necrosis virus (ISKNV) combined-vaccine efficacy in *Epinephelus coioides*. *Veterinary Immunology and Immunopathology*, 150(1): 61-68.
- Huicab-Pech, Z. G., Landeros-Sánchez, C., Castañeda-Chávez, M. R., Lango-Reynoso, F., López-Collado, C. J., & Platas-Rosado, D. E. (2016). Current state of bacteria pathogenicity and their relationship with host and environment in tilapia (*Oreochromis niloticus*). *J Aquac Res Development*, 7(428): 2.
- Igbinsola, E. O., & Okoh, A. I. (2008). Emerging *Vibrio* species: an unending threat to public health in developing countries. *Research in Microbiology*, 159(7): 495-506.
- Ilmiah, Kamaruzaman, J., Sukenda, Widanarni, & Rustam (2013). The role of probiotic bacteria on controlling vibriosis in tiger grouper fry (*Epinephelus foscoguttatus*). *World Journal of Fish and Marine Sciences*, 5 (6): 622-627.
- Immanuel, G. (2016). Bioencapsulation of brine shrimp *Artemia* nauplii with probiotics and their resistance against *Vibrio* pathogens. *Aquat. Sci*, 11: 323-330.
- Ina-Salwany. Y., M., Yusoff, S. M., Mohd, Z. S., & Mohd, E. A. W. (2011). Efficacy of an inactivated recombinant vaccine encoding a fimbrial protein of *Pasteurella multocida* B: 2 against hemorrhagic septicemia in goats. *Tropical Animal Health and Production*, 43(1): 179-187.
- Jayasinghe, C. V. L., Ahmed, S. B. N., & Kariyawasam, M. G. I. U. (2010). The isolation and identification of *Vibrio* species in marine shrimps of Sri Lanka. *Journal of Food and Agriculture*, 1(1): 36-44.

- Jeyasekaran, G., Raj, K. T., Shakila, R. J., Thangarani, A. J., & Sukumar, D. (2011). Multiplex polymerase chain reaction-based assay for the specific detection of toxin-producing *Vibrio cholerae* in fish and fishery products. *Applied Microbiology and Biotechnology*, 90(3): 1111-1118.
- Jia, B., & Jeon, C. O. (2016). High-throughput recombinant protein expression in *Escherichia coli*: current status and future perspectives. *Open Biology*, 6(8): 160-196.
- Johnston, B., & Yeeting, B. (2006). Proceedings of a workshop: *Economics and marketing of the live reef fish trade in Asia-Pacific*. Noumea, New Caledonia Secretariat of the Pacific Community.
- Joosten, P. H., Aviles-Trigueros, M., Sorgeloos, P., & Rombout, J. H. W. M. (1995). Oral vaccination of juvenile carp (*Cyprinus carpio*) and gilthead seabream (*Sparus aurata*) with bioencapsulated *Vibrio anguillarum* bacterin. *Fish and Shellfish Immunology*, 5(4): 289-299.
- Joseph, B. C., Pichaimuthu, S., Srimeenakshi, S., Murthy, M., Selvakumar, K., Ganesan, M., & Manjunath, S. R. (2015). An overview of the parameters for recombinant protein expression in *Escherichia coli*. *Journal of Cell Science and Therapy*, 6(5): 1.
- Jun, L., & Woo, N. Y. (2003). Pathogenicity of Vibrios in fish: an overview. *Journal of Ocean University of Qingdao*, 2(2): 117-128.
- Jung, C., Park, M., & Heo, M. (2005). Immunization with major outer membrane protein of *Vibrio vulnificus* elicits protective antibodies in a murine model. *Journal of Microbiology-Seoul*, 43(5): 437.
- Kaufmann, S. H. (1990). Heat shock proteins and the immune response. *Immunology Today*, 11: 129-136.
- Kawai, K., Liu, Y., Ohnishi, K., & Oshima, S. I. (2004). A conserved 37 kDa outer membrane protein of *Edwardsiella tarda* is an effective vaccine candidate. *Vaccine*, 22(25): 3411-3418.
- Khan, M. N., Bansal, A., Shukla, D., Paliwal, P., Sarada, S. K. S., Mustoori, S. R., & Banerjee, P. K. (2006). Immunogenicity and protective efficacy of DnaJ (hsp40) of *Streptococcus pneumoniae* against lethal infection in mice. *Vaccine*, 24(37): 6225-6231.
- Khushiramani, R. M., Maiti, B., Shekar, M., Girisha, S. K., Akash, N., Deepanjali, A., & Karunasagar, I. (2012). Recombinant *Aeromonas hydrophila* outer membrane protein 48 (Omp48) induces a protective immune response against *Aeromonas hydrophila* and *Edwardsiella tarda*. *Research in Microbiology*, 163(4): 286-291.
- Khushiramani, R., Girisha, S. K., Karunasagar, I., & Karunasagar, I. (2007). Cloning and expression of an outer membrane protein ompTS of *Aeromonas hydrophila* and study of immunogenicity in fish. *Protein Expression and Purification*, 51(2): 303-307.

- Lee, K. K., Liu, P. C., & Chuang, W. H. (2002). Pathogenesis of gastroenteritis caused by *Vibrio carchariae* in cultured marine fish. *Marine Biotechnology*, 4(3): 267-277
- Li, C., Ye, Z., Wen, L., Chen, R., Tian, L., Zhao, F., & Pan, J. (2014). Identification of a novel vaccine candidate by immunogenic screening of *Vibrio parahaemolyticus* outer membrane proteins. *Vaccine*, 32(46): 6115-6121.
- Li, H., Xiong, X. P., Peng, B., Xu, C. X., Ye, M. Z., Yang, T. C., & Peng, X. X. (2009). Identification of broad cross-protective immunogens using heterogeneous antiserum-based immunoproteomic approach. *Journal of Proteome Research*, 8(9): 4342-4349.
- Li, H., Ye, M. Z., Peng, B., Wu, H. K., Xu, C. X., Xiong, X. P., & Peng, X. X. (2010). Immunoproteomic identification of polyvalent vaccine candidates from *Vibrio parahaemolyticus* outer membrane proteins. *Journal of Proteome Research*, 9(5): 2573-2583.
- Li, J., Ma, S., & Woo, N. (2015). Vaccination of Silver Sea Bream (*Sparus sarba*) against *Vibrio alginolyticus*: protective evaluation of different vaccinating modalities. *International Journal of Molecular Sciences*, 17(1): 40.
- Li, J., Zhou, L., & Woo, N. Y. (2003). Invasion route and pathogenic mechanisms of *Vibrio alginolyticus* to silver sea bream *Sparus sarba*. *Journal of Aquatic Animal Health*, 15(4): 302-313.
- Liao, I.C. and Leaño, E.M. (eds.) (2008). The Aquaculture of Groupers. Asian Fisheries Society.
- Lin, C. C., Lin, J. H. Y., Chen, M. S., & Yang, H. L. (2007). An oral nervous necrosis virus vaccine that induces protective immunity in larvae of grouper (*Epinephelus coioides*). *Aquaculture*, 268(1): 265-273.
- Lin, C. Y., Huang, Z., Wen, W., Wu, A., Wang, C., & Niu, L. (2015). Enhancing protein expression in HEK-293 cells by lowering culture temperature. *PloS One*, 10(4): 123-152.
- Lin, J., Huang, S., & Zhang, Q. (2002). Outer membrane proteins: key players for bacterial adaptation in host niches. *Microbes and Infection*, 4(3): 325-331.
- Liu, L., Ge, M., Zheng, X., Tao, Z., Zhou, S., & Wang, G. (2016). Investigation of *Vibrio alginolyticus*, *V. harveyi*, and *V. parahaemolyticus* in large yellow croaker, *Pseudosciaena crocea* (Richardson) reared in Xiangshan Bay, China. *Aquaculture Reports*, 3: 220-224.
- Liu, Y., Wang, H., Zhang, S., Zeng, L., Xu, X., Wu, K., & Yin, Y. (2014). Mucosal immunization with recombinant fusion protein DnaJ-ΔA146Ply enhances cross-protective immunity against *Streptococcus pneumoniae* infection in mice via interleukin 17A. *Infection and Immunity*, 82(4): 1666-1675.
- Lo, J. H., Baker, T. A., & Sauer, R. T. (2001). Characterization of the N- terminal repeat domain of *Escherichia coli* ClpA—A class I Clp/HSP100 ATPase. *Protein Science*, 10(3): 551-559.

- Lun, J., Xia, C., Yuan, C., Zhang, Y., Zhong, M., Huang, T., & Hu, Z. (2014). The outer membrane protein, LamB (maltoporin), is a versatile vaccine candidate among the *Vibrio* species. *Vaccine*, 32(7): 809-815.
- Luo, Z., Fu, J., Li, N., Liu, Z., Qin, T., Zhang, X., & Nie, P. (2016). Immunogenic proteins and their vaccine development potential evaluation in outer membrane proteins (OMPs) of *Flavobacterium columnare*. *Aquaculture and Fisheries*, 1: 1-8.
- Maiti, B., Raghunath, P., & Karunasagar, I. (2009). Cloning and expression of an outer membrane protein OmpW of *Aeromonas hydrophila* and study of its distribution in *Aeromonas* spp. *Journal of Applied Microbiology*, 107(4): 1157-1167.
- Maiti, B., Shetty, M., Shekar, M., Karunasagar, I., & Karunasagar, I. (2012). Evaluation of two outer membrane proteins, Aha1 and OmpW of *Aeromonas hydrophila* as vaccine candidate for common carp. *Veterinary Immunology and Immunopathology*, 149(3): 298-301.
- Malik, A., Alsenaidy, A. M., Elrobh, M., Khan, W., Alanazi, M. S., & Bazzi, M. D. (2016). Optimization of expression and purification of HSPA6 protein from *Camelus dromedarius* in *E. coli*. *Saudi Journal of Biological Sciences*, 23(3): 410-419.
- Mao, Z., Yu, L., You, Z., Wei, Y., & Liu, Y. (2007). Cloning, expression and immunogenicity analysis of five outer membrane proteins of *Vibrio parahaemolyticus* zj2003. *Fish and Shellfish Immunology*, 23(3): 567-575.
- Marandi, M. V., Dubreuil, J. D., & Mittal, K. R. (1996). The 32 kDa major outer-membrane protein of *Pasteurella multocida* capsular serotype D. *Microbiology*, 142(1): 199-206.
- Marco-Noales, E., Milán, M., Fouz, B., Sanjuán, E., & Amaro, C. (2001). Transmission to eels, portals of entry, and putative reservoirs of *Vibrio vulnificus* serovar E (biotype 2). *Applied and Environmental Microbiology*, 67(10): 4717-4725.
- Marques, A., Dhont, J., Sorgeloos, P., & Bossier, P. (2004). Evaluation of different yeast cell wall mutants and microalgae strains as feed for gnotobiotically grown brine shrimp *Artemia franciscana*. *Journal of Experimental Marine Biology and Ecology*, 312(1): 115-136.
- Marques, A., Dinh, T., Ioakeimidis, C., Huys, G., Swings, J., Verstraete, W. & Bossier, P. (2005). Effects of bacteria on *Artemia franciscana* cultured in different gnotobiotic environments. *Applied and Environmental Microbiology*, 71(8): 4307-4317.
- Marques, A., Thanh, T. H., Verstraete, W., Dhont, J., Sorgeloos, P., & Bossier, P. (2006). Use of selected bacteria and yeast to protect gnotobiotic *Artemia* against different pathogens. *Journal of Experimental Marine Biology and Ecology*, 334(1): 20-30.

- Membrebe, J. D., Yoon, N. K., Hong, M., Lee, J., Lee, H., Park, K., & Ahn, J. (2016). Protective efficacy of *Streptococcus iniae* derived enolase against Streptococcal infection in a zebrafish model. *Veterinary Immunology and Immunopathology*, 170: 25-29.
- Morimoto, R. I. (1998). Regulation of the heat shock transcriptional response: cross talk between a family of heat shock factors, molecular chaperones, and negative regulators. *Genes and Development*, 12(24): 3788-3796.
- Morimoto, R. I. (1998). Regulation of the heat shock transcriptional response: cross talk between a family of heat shock factors, molecular chaperones, and negative regulators. *Genes & Development*, 12(24): 3788-3796.
- Muktar, Y., Tesfaye, S., & Tesfaye, B. (2016). Present status and future prospects of fish vaccination: a review. *Journal Veterinar Science Technology*, 2: 299.
- Muktar, Y., Tesfaye, S., & Tesfaye, B. (2016). Present status and future prospects of fish vaccination: A review. *J. Veterinar. Sci. Technol.*, 7: 2.
- Mutharia, L. W., Raymond, B. T., Dekievit, T. R., & Stevenson, R. M. (1993). Antibody specificities of polyclonal rabbit and rainbow trout antisera against *Vibrio ordalii* and serotype 0: 2 strains of *Vibrio anguillarum*. *Canadian Journal of Microbiology*, 39(5): 492-499.
- Naceur, H. B., Jenhani, A. B. R., & Romdhane, M. S. (2009). New distribution record of the brine shrimp *Artemia* (Crustacea, Branchiopoda, Anostraca) in Tunisia. *Check List*, 5(2): 281-288.
- Nagasawa, K., & Cruz-Lacierda, E. R. (2004). *Diseases of cultured groupers*. Aquaculture Department, Southeast Asian Fisheries Development Center.
- Nakaguchi, Y. (2013). Contamination by *Vibrio parahaemolyticus* and its virulent strains in seafood marketed in Thailand, Vietnam, Malaysia, and Indonesia. *Tropical Medicine and Health*, 41(3): 95-102.
- Nandi, B., Nandy, R. K., Sarkar, A., & Ghose, A. C. (2005). Structural features, properties and regulation of the outer-membrane protein W (OmpW) of *Vibrio cholerae*. *Microbiology*, 151(9): 2975-2986.
- Nascimento, I. P., & Leite, L. C. C. (2012). Recombinant vaccines and the development of new vaccine strategies. *Brazilian Journal of Medical and Biological Research*, 45(12): 1102-1111.
- Nazifah, N. M. (2013). *Development and evaluation of recombinant vector cells carrying cell wall surface anchor family proteins as a vaccine against streptococcosis in red hybrid tilapia (Oreochromis spp.)*. (Unpublished doctoral dissertation, Universiti Putra Malaysia, Malaysia).
- Nehlah, R., Firdaus-Nawi, M., Nik-Haiha, N. Y., Karim, M., Zamri-Saad, M., & Ina-Salwany, M. Y. (2017). Recombinant vaccine protects juvenile hybrid grouper, *Epinephelus fuscoguttatus* × *Epinephelus lanceolatus*, against infection by *Vibrio alginolyticus*. *Aquaculture International*, 25(6): 2047-2059.

- Nehlah, R., Ina-Salwany, M. Y., & Zulperi, Z. (2016). Antigenicity Analysis and Molecular Characterization of Two Outer Membrane Proteins of *Vibrio alginolyticus* Strain VA2 as Vaccine Candidates in Tiger Grouper Culture. *Journal of Biological Sciences*, 16(1): 1.
- Ng, B. (2009). Current status and future prospects for the aquaculture industry in Malaysia. *World aquaculture*.
- Ningqiu, L., Junjie, B., Shuqin, W., Xiaozhe, F., Haihua, L., Xing, Y., & Cunbin, S. (2008). An outer membrane protein, OmpK, is an effective vaccine candidate for *Vibrio harveyi* in Orange-spotted grouper (*Epinephelus coioides*). *Fish & shellfish immunology*, 25(6): 829-833.
- Noor, R., Islam, Z., Munshi, S. H., & Rahman, F. (2013). Influence of temperature on *Escherichia coli* growth in different culture media. *Journal of Pure Applied Microbiology*, 7(2): 899-904.
- Noorlis, A., Mohamad Ghazali, F., Cheah, Y. K., Chilek, T., Zainazor, T., Ponniah, J., & Radu, S. (2011). Prevalence and quantification of *Vibrio* species and *Vibrio parahaemolyticus* in freshwater fish at hypermarket level. *International Food Research Journal*, 18(2): 689-695
- Novriadi, R., & Haw, K. B. (2014). Immunostimulation effects of herbal bio conditioners on tiger grouper (*Epinephelus fuscoguttatus*) against *V. parahaemolyticus* infection. *International Journal of Fisheries and Aquatic Studies*, 1(3): 73-78
- Nurfarizatul, Z. Z. (2015). *Characterization of three Vibrio species by polymerase chain reaction (PCR) and study of their pathogenic potential in a gnotobiotic Artemia model*. (Unpublished degree dissertation), Universiti Putra Malaysia, Malaysia.
- Olivares- Fuster, O., Terhune, J. S., Shoemaker, C. A., & Arias, C. R. (2010). Cloning, expression, and immunogenicity of *Flavobacterium columnare* heat shock protein DnaJ. *Journal of Aquatic Animal Health*, 22(2): 78-86.
- Orozco-Medina, C., Maeda-Martínez, A. M., & López-Cortés, A. (2002). Effect of aerobic Gram-positive heterotrophic bacteria associated with *Artemia franciscana* cysts on the survival and development of its larvae. *Aquaculture*, 213(1): 15-29.
- Ortiz-Carrillo, I., Estrella-Gómez, N. E., Zamudio-Maya, M., & Rojas-Herrera, R. (2015). Diversity of *Vibrio* spp in Karstic Coastal Marshes in the Yucatan Peninsula. *PloS One*, 10(8): 134953.
- Othman, M. F. (1998). Challenges ahead in meeting aquaculture production in Malaysia under the Third National Agricultural Policy, Nap3 (1998-2010). *Brackish Water Aquaculture Research Center (BARC)*. Ministry of Agricultural and Agro-Based Industry, Department of Fisheries Malaysia.

- Pan, J., Li, C., & Ye, Z. (2016). Immunoproteomic approach for screening vaccine candidates from bacterial outer membrane proteins. *Vaccines for Veterinary Diseases*, 2: 519-528.
- Pang, H., Chen, L., Hoare, R., Huang, Y., & Jian, J. (2016). Identification of DLD, by immunoproteomic analysis and evaluation as a potential vaccine antigen against three *Vibrio* species in *Epinephelus coioides*. *Vaccine*, 34(9): 1225-1231.
- Papaneophytou, C. P., Rinotas, V., Douni, E., & Kontopidis, G. (2013). A statistical approach for optimization of RANKL overexpression in *Escherichia coli*: Purification and characterization of the protein. *Protein Expression and Purification*, 90(1): 9-19.
- Pasnik, D. J., & Smith, S. A. (2005). Immunogenic and protective effects of a DNA vaccine for *Mycobacterium marinum* in fish. *Veterinary Immunology and Immunopathology*, 103(3): 195-206.
- Pati, U. S., Srivastava, S. K., Roy, S. C., & More, T. (1996). Immunogenicity of outer membrane protein of *Pasteurella multocida* in buffalo calves. *Veterinary Microbiology*, 52(3): 301-311.
- Patra, S. K., & Mohamed, K. S. (2003). Enrichment of *Artemia nauplii* with the probiotic yeast *Saccharomyces boulardii* and its resistance against a pathogenic *Vibrio*. *Aquaculture international*, 11(5): 505-514.
- Pellequer, J. L., Westhof, E., & Van Regenmortel, M. H. (1993). Correlation between the location of antigenic sites and the prediction of turns in proteins. *Immunology letters*, 36(1): 83-99.
- Peng, B., Ye, J. Z., Han, Y., Zeng, L., Zhang, J. Y., & Li, H. (2016). Identification of polyvalent protective immunogens from outer membrane proteins in *Vibrio parahaemolyticus* to protect fish against bacterial infection. *Fish and Shellfish Immunology*, 54: 204-210.
- Phillips, R., Williams, J. N., Tan, W. M., Bielecka, M. K., Thompson, H., Hung, M. C. & Christodoulides, M. (2013). Immunization with recombinant Chaperonin60 (Chp60) outer membrane protein induces a bactericidal antibody response against *Neisseria meningitidis*. *Vaccine*, 31(22): 2584-2590.
- Plant, K. P., & LaPatra, S. E. (2011). Advances in fish vaccine delivery. *Developmental and Comparative Immunology*, 35(12): 1256-1262.
- Pomeroy, R. S. (2002). The status of grouper culture in Southeast Asia. *SPC Live Reef Fish Information Bulletin*, 10: 22-26.
- Poornima, M., Vijaya Kumar, R., Santiago, T.C2. & Arasu, A.R.T1. (2013). Recombinant outer membrane protein, omp26la of *Vibrio anguillarum* is an effective vaccine candidate. *Journal of Aquatic Biology and Fisheries*, 2: 436-440.

- Pore, D., Chowdhury, P., Mahata, N., Pal, A., Yamasaki, S., Mahalanabis, D., & Chakrabarti, M. K. (2009). Purification and characterization of an immunogenic outer membrane protein of *Shigella flexneri* 2a. *Vaccine*, 27(42): 5855-5864.
- Pridgeon, J. W., & Klesius, P. H. (2013). Major bacterial diseases in aquaculture and their vaccine development. *Anim. Sci. Rev*, 2012: 141.
- Puente, M. E., Vega- Villasante, F., Holguin, G., & Bashan, Y. (1992). Susceptibility of the brine shrimp *Artemia* and its pathogen *Vibrio parahaemolyticus* to chlorine dioxide in contaminated sea- water. *Journal of Applied Bacteriology*, 73(6): 465-471.
- Qian, R. H., Xiao, Z. H., Zhang, C. W., Chu, W. Y., Wang, L. S., Zhou, H. H., & Yu, L. (2008). A conserved outer membrane protein as an effective vaccine candidate from *Vibrio alginolyticus*. *Aquaculture*, 278(1): 5-9.
- Qian, R., Chu, W., Mao, Z., Zhang, C., Wei, Y., & Yu, L. (2007). Expression, characterization and immunogenicity of a major outer membrane protein from *Vibrio alginolyticus*. *Acta biochimica biophysica Sinica*, 39(3): 194-200.
- Qin, Y. X., & Yan, Q. P. (2010). International Conference 2010: *The antigenicity of the flagellin, outer membrane proteins and lipopolysaccharide of Vibrio harveyi*. In Bioinformatics and Biomedical Technology (ICBBT).
- Rabbi, F., Sultana, N., Rahman, T., Al-Emran, H. M., Uddin, M. N., Hossain, M., & Ahsan, C. R. (2008). Analysis of immune responses and serological cross reactivities among *Vibrio cholerae* O1, *Shigella flexneri* 2a and *Haemophilus influenzae* b. *Cellular and Molecular Immunology*, 5(5): 393
- Ramesh Kumar, P., Kalidas, C., Tamilmani, G., Sakthivel, M., Nazar, A. A., Maharshi, V. A., & Gopakumar, G. (2014). Microbiological and histopathological investigations of *Vibrio alginolyticus* infection in cobia *Rachycentron canadum* (Linnaeus, 1766) cultured in sea cage. *Indian Journal of Fisheries*, 61(1): 124-127.
- Ransangan, J., & Mustafa, S. (2009). Identification of *Vibrio harveyi* isolated from diseased Asian Seabass (*Lates calcarifer*) by use of 16S ribosomal DNA sequencing. *Journal of Aquatic Animal Health*, 21(3): 150-155.
- Ransangan, J., Lal, T. M., & Al-Harbi, A. H. (2012). Characterization and experimental infection of *Vibrio harveyi* isolated from diseased Asian seabass (*Lates calcarifer*). *Malaysian Journal of Microbiology*, 8(2): 104-115.
- Rico-Mora, R., & Voltolina, D. (1995). Effects of bacterial isolates from *Skeletonema costatum* cultures on the survival of *Artemia franciscana* nauplii. *Journal of Invertebrate Pathology*, 66(2): 203-204.

- Rimmer, M. A., Thampisamraj, Y. C., Jayagopal, P., Thineshsanthar, D., Damodar, P. N., & Toledo, J. D. (2013). Spawning of tiger grouper *Epinephelus fuscoguttatus* and squaretail coral grouper *Plectropomus areolatus* in sea cages and onshore tanks in Andaman and Nicobar Islands, India. *Aquaculture*, 410: 197-202.
- Roiha, I. S., Otterlei, E., & Samuelsen, O. B. (2010). Bioencapsulation of florfenicol in brine shrimp, *Artemia franciscana* nauplii. *Journal of Bioanalysis & Biomedicine*, 1(2): 1-5.
- Rosano, G. L., & Ceccarelli, E. A. (2014). Recombinant protein expression in *Escherichia coli*: advances and challenges. *Recombinant protein expression in microbial systems*, 7.
- Ruwandeeepika, D., Arachchige, H., Sanjeewa Prasad Jayaweera, T., Paban Bhowmick, P., Karunasagar, I., Bossier, P., & Defoirdt, T. (2012). Pathogenesis, virulence factors and virulence regulation of *Vibrios* belonging to the Harveyi clade. *Reviews in Aquaculture*, 4(2): 59-74.
- Sabri, M. Y., Zamri-Saad, M., Mutalib, A. R., Israf, D. A., & Muniandy, N. (2000). Efficacy of an outer membrane protein of *Pasteurella haemolytica* A2, A7 or A9-enriched vaccine against intratracheal challenge exposure in sheep. *Veterinary Microbiology*, 73(1): 13-23.
- Sagi, S. S., Paliwal, P., Bansal, A., Mishra, C., Khan, N., Mustoori, S. R. & Banerjee, P. K. (2006). Studies on immunogenicity and protective efficacy of DnaJ of *Salmonella Typhi* against lethal infection by *Salmonella Typhimurium* in mice. *Vaccine*, 24(49): 7135-7141.
- Saleema, M. (2015). *Molecular Characterization of Aeromonas hydrophila and Development of Recombinant Cells Vaccine Expressing Outer Membrane Proteins Against It in African Catfish (Clarias gariepinus Burchell, 1822)*. (Master of Science dissertation), Universiti Putra Malaysia, Malaysia.
- Sankar, G., Saravanan, J., Krishnamurthy, P., Chandrakala, N., & Rajendran, K. (2012). Isolation and identification of *Vibrio* spp. in diseased *Channa punctatus* from aquaculture fish farm. *Journal of Molecular Sciences*, 41(2): 159-163.
- Sapkota, A., Sapkota, A. R., Kucharski, M., Burke, J., McKenzie, S., Walker, P., & Lawrence, R. (2008). Aquaculture practices and potential human health risks: current knowledge and future priorities. *Environment International*, 34(8): 1215-1226.
- Sardar, P., Joardar, S. N., Ganesan, P., & Abraham, T. J. (2004). Seroreactivity of somatic soluble proteins of *Vibrio alginolyticus*. *Indian Journal of Fisheries*, 51(2): 239-244.
- Sarjito, S., Radjasa, O. K., Sabdono, A., Prayitno, S. B., & Hutabarat, S. (2009). Phylogenetic diversity of the causative agents of vibriosis associated with groupers fish from Karimunjawa Islands, Indonesia. *Current Research in Bacteriology*, 2(1): 14-21.

- Sawabe, T., Ogura, Y., Matsumura, Y., Feng, G., Amin, A. R., Mino, S., & Satomi, M. (2013). Updating the *Vibrio* clades defined by multilocus sequence phylogeny: proposal of eight new clades, and the description of *Vibrio tritonius* sp. *Frontiers in Microbiology*, 4.
- Schein, C. H., & Noteborn, M. H. (1988). Formation of soluble recombinant proteins in *Escherichia coli* is favored by lower growth temperature. *Nature Biotechnology*, 6(3): 291-294.
- Schwab, H. (1993). Principles of genetic engineering for *Escherichia coli*. *Biotechnology Set, Second Edition*, 372-425.
- Senderovich, Y., Izhaki, I., & Halpern, M. (2010). Fish as reservoirs and vectors of *Vibrio cholerae*. *PLoS One*, 5(1): 8607.
- Sharma, K., Ramachandra, S., Rathore, G., Verma, D. K., Sadhu, N., & Philipose, K. K. (2012). *Vibrio alginolyticus* infection in Asian seabass (*Lates calcarifer*, Bloch) reared in open sea floating cages in India. *Aquaculture Research*, 44(1): 86-92.
- Shikongo-Nambabi, M. N. N., & Schneider, M. B. (2012). The role, isolation and identification of *Vibrio* species on the quality and safety of seafood. *Biotechnology and Molecular Biology Reviews*, 7(2): 16-30.
- Singh, V., Somvanshi, P., Rathore, G., Kapoor, D., & Mishra, B. N. (2009). Gene cloning, expression and homology modeling of hemolysin gene from *Aeromonas hydrophila*. *Protein expression and Purification*, 65(1): 1-7.
- Sivashanmugam, A., Murray, V., Cui, C., Zhang, Y., Wang, J., & Li, Q. (2009). Practical protocols for production of very high yields of recombinant proteins using *Escherichia coli*. *Protein Science*, 18(5): 936-948.
- Soltanian, S. (2007). *Protection of gnotobiotic Artemia against Vibrio campbellii using baker's yeast strains and extracts* (Unpublished doctoral dissertation). Ghent University.
- Sommerset, I., Krossøy, B., Biering, E., & Frost, P. (2005). Vaccines for fish in aquaculture. *Expert Review of Vaccines*, 4(1): 89-101.
- Sonia, A.S. G., & Lipton, A.P. (2012). Pathogenicity and antibiotic susceptibility of *Vibrio* species isolated from the captive-reared tropical marine ornamental blue damselfish, *Pomacentrus caeruleus* (Quoy and Gaimard, 1825). *Indian Journal of Geo-Marine Sciences*, 41(4): 348-354.
- Sørensen, H. P., & Mortensen, K. K. (2005). Soluble expression of recombinant proteins in the cytoplasm of *Escherichia coli*. *Microbial Cell Factories*, 4(1): 1.
- Stabili, L., Miglietta, A. M., & Belmonte, G. (1999). Lysozyme-like and trypsin-like activities in the cysts of *Artemia franciscana* Kellogg, 1906: is there a passive immunity in a resting stage. *Journal of Experimental Marine Biology and Ecology*, 237(2): 291-303.

- Stevens, R. C. (2000). Design of high-throughput methods of protein production for structural biology. *Structure*, 8(9): 177-185.
- Subasinghe, R., & Shariff, M. (1993). Recent advances in aquaculture health management. *Diseases in Aquaculture: The Current Issues*
- Subhadra, B., Hurwitz, I., Fieck, A., Rao, D. V. S., Subba Rao, G., & Durvasula, R. (2010). Development of paratransgenic *Artemia* as a platform for control of infectious diseases in shrimp mariculture. *Journal of Applied Microbiology*, 108(3): 831-840.
- Sun, B. G., Dang, W., Sun, L., & Hu, Y. H. (2014). *Vibrio harveyi* Hsp70: Immunogenicity and application in the development of an experimental vaccine against *V. harveyi* and *Streptococcus iniae*. *Aquaculture*, 418: 144-147.
- Sun, K., Zhang, W. W., Hou, J. H., & Sun, L. (2009). Immunoprotective analysis of VhhP2, a *Vibrio harveyi* vaccine candidate. *Vaccine*, 27(21): 2733-2740.
- Sun, Y., Hu, Y. H., Liu, C. S., & Sun, L. (2012). Construction and comparative study of monovalent and multivalent DNA vaccines against *Streptococcus iniae*. *Fish and Shellfish Immunology*, 33(6): 1303-1310.
- Sung, Y. Y., Ashame, M. F., Chen, S., MacRae, T. H., Sorgeloos, P., & Bossier, P. (2009b). Feeding *Artemia franciscana* (Kellogg) larvae with bacterial heat shock protein, protects from *Vibrio campbellii* infection. *Journal of Fish Diseases*, 32(8): 675-685.
- Sung, Y. Y., Dhaene, T., Defoirdt, T., Boon, N., MacRae, T. H., Sorgeloos, P., & Bossier, P. (2009a). Ingestion of bacteria overproducing DnaK attenuates *Vibrio* infection of *Artemia franciscana* larvae. *Cell Stress and Chaperones*, 14(6): 603.
- Swain, P., Behera, T., Mohapatra, D., Nanda, P. K., Nayak, S. K., Meher, P. K., & Das, B. K. (2010). Derivation of rough attenuated variants from smooth virulent *Aeromonas hydrophila* and their immunogenicity in fish. *Vaccine*, 28(29): 4626-4631.
- Swain, P., Nayak, S. K., Sahu, A., Meher, P. K., & Mishra, B. K. (2003). High antigenic cross-reaction among the bacterial species responsible for diseases of cultured freshwater fishes and strategies to overcome it for specific serodiagnosis. *Comparative Immunology, Microbiology and Infectious Diseases*, 26(3): 199-211.
- Tanguay, J. A., Reyes, R. C., & Clegg, J. S. (2004). Habitat diversity and adaptation to environmental stress in encysted embryos of the crustacean *Artemia*. *Journal of Biosciences*, 29(4): 489-501.
- Thompson, B., & Subasinghe, R. (2011). Aquaculture's role in improving food and nutrition security. *Combating micronutrient deficiencies: Food-based approaches*, 150-162.

- Thompson, F. L., Iida, T., & Swings, J. (2004). Biodiversity of *Vibrios*. *Microbiology and Molecular Biology Reviews*, 68(3): 403-431.
- Tomassen, J. (2010). Assembly of outer-membrane proteins in bacteria and mitochondria. *Microbiology*, 156(9): 2587-2596.
- Toranzo, A. E., Magariños, B., & Romalde, J. L. (2005). A review of the main bacterial fish diseases in mariculture systems. *Aquaculture*, 246(1): 37-61.
- Touraki, M., Mourelatos, S., Karamanlidou, G., Kalaitzopoulou, S., & Kastritsis, C. (1996). Bioencapsulation of chemotherapeutics in *Artemia* as a means of prevention and treatment of infectious diseases of marine fish fry. *Aquacultural Engineering*, 15(2): 133-147.
- Towbin, H., Staehelin, T., & Gordon, J. (1979). Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. *Proceedings of the National Academy of Sciences*, 76(9): 4350-4354.
- Tupper, M., & Sheriff, N. (2008). Capture-based aquaculture of groupers. *Capture-based Aquaculture. Global Overview*, 217-253.
- Uchiyama, H. (2000). Distribution of *Vibrio* species isolated from aquatic environments with TCBS agar. *Environmental Health and Preventive Medicine*, 4(4): 199-204.
- Urakawa, H., & Rivera, I. N. G. (2006). The biology of *Vibrios*. Washington DC, 20036-2904.
- Van Muiswinkel, W. B. (2008). A history of fish immunology and vaccination I. The early days. *Fish and Shellfish Immunology*, 25(4): 397-408.
- Van Muiswinkel, W. B., & Nakao, M. (2014). A short history of research on immunity to infectious diseases in fish. *Developmental and Comparative Immunology*, 43(2): 130-150.
- Van Stappen, G. (1996). Introduction, biology and ecology of *Artemia*. *Manual on the production and use of live food for aquaculture*, 361: 79-106.
- Van Stappen, G. (2002). In *Artemia: Basic and Applied Biology. Zoogeography*. (pp. 171-224). Springer, Dordrecht.
- Vanmalae S, Defoirdt T, Cleenwerck I, De Vos P, Bossier P. (2015). Characterization of the virulence of *Harveyi* clade *Vibrios* isolated from a shrimp hatchery in vitro and in vivo, in a brine shrimp (*Artemia franciscana*) model system. *Aquaculture*, 435: 28-32.
- Verschuere, L., Heang, H., Criel, G., Sorgeloos, P., Verstraete, W. (2000). Selected bacterial strains protect *Artemia* spp. from the pathogenic effects of *Vibrio proteolyticus* CW8T2. *Appl. Environ. Microbiol.*, 66: 1139– 1146.
- Walia, A., & Balkhi, M. U. H. (2016). Fish vaccination and therapeutics. *International Journal of Multidisciplinary Research and Development*, 3(4): 55-60

- Wang, Q., Chen, J., Liu, R., & Jia, J. (2011). Identification and evaluation of an outer membrane protein OmpU from a pathogenic *Vibrio harveyi* isolate as vaccine candidate in turbot (*Scophthalmus maximus*). *Letters in Applied Microbiology*, 53(1): 22-29.
- Wang, Y., Wu, W., Negre, N. N., White, K. P., Li, C., & Shah, P. K. (2011). Determinants of antigenicity and specificity in immune response for protein sequences. *BMC Bioinformatics*, 12(1):251.
- WHO, 2014. Antimicrobial Resistance Global Report on Surveillance. World Health Organization [Online] Available at: <http://www.who.int/drugresistance/documents/surveillancereport/en/>.
- Wongtavatchai, J., López-Dóriga, M. V., & Francis, M. J. (2010). Effect of AquaVac TM VibromaxTM on size and health of post larva stage of Pacific White shrimp *Litopenaeus vannamei* and Black Tiger shrimp *Penaeus monodon*. *Aquaculture*, 308(3): 75-81.
- Xie, Z. Y., Hu, C. Q., Chen, C., Zhang, L. P., & Ren, C. H. (2005). Investigation of seven *Vibrio* virulence genes among *Vibrio alginolyticus* and *Vibrio parahaemolyticus* strains from the coastal mariculture systems in Guangdong, China. *Letters in Applied Microbiology*, 41(2): 202-207.
- Xiong, X. P., Zhang, B. W., Yang, M. J., Ye, M. Z., Peng, X. X., & Li, H. (2010). Identification of vaccine candidates from differentially expressed outer membrane proteins of *Vibrio alginolyticus* in response to NaCl and iron limitation. *Fish and Shellfish Immunology*, 29(5): 810-816.
- Xu, C., Wang, S., Zhaoxia, Z., & Peng, X. (2005). Immunogenic cross-reaction among outer membrane proteins of Gram-negative bacteria. *International immunopharmacology*, 5(7): 1151-1163.
- Yanuhar, U. (2010). The Role of immunogenic adhesin *Vibrio alginolyticus* 49 kDa to molecule expression of major histocompatibility complex on receptors of humpback grouper *Cromileptes altivelis*. *Proceeding World Academy of Science, Engineering and Technology*, 67: 968-973.
- Young, C. L., Britton, Z. T., & Robinson, A. S. (2012). Recombinant protein expression and purification: a comprehensive review of affinity tags and microbial applications. *Biotechnology Journal*, 7(5): 620-634.
- Yu, L. P., Hu, Y. H., Sun, B. G., & Sun, L. (2013). Immunological study of the outer membrane proteins of *Vibrio harveyi*: insights that link immunoprotectivity to interference with bacterial infection. *Fish and Shellfish Immunology*, 35(4): 1293-1300.
- Yusoff, A. (2015). Proceedings of the International Workshop on Resource Enhancement and Sustainable Aquaculture Practices in Southeast Asia 2014 (RESA): *Status of resource management and aquaculture in Malaysia*. Aquaculture Department, Southeast Asian Fisheries Development Center.

- Yusoff, M., & Sabri, M. (1999). *Immunology of the Outer Membrane Proteins of Pasteurella Haemolytica A2, A7 and A9 in Sheep*. (Unpublished doctoral dissertation). Universiti Putra Malaysia, Malaysia.
- Zha, Z., Li, C., Li, W., Ye, Z., & Pan, J. (2016). LptD is a promising vaccine antigen and potential immunotherapeutic target for protection against *Vibrio* species infection. *Scientific Reports*, 6.
- Zhang X, Robertson P, Austin B., Xu, H. (1997). Comparison of outer membrane protein profiles of *Vibrio* sp. *Acta Microbiologica Sinica*, 37(6): 449-454.
- Zhang, C., Yu, L., & Qian, R. (2007). Characterization of OmpK, GAPDH and their fusion OmpK–GAPDH derived from *Vibrio harveyi* outer membrane proteins: their immunoprotective ability against vibriosis in large yellow croaker (*Pseudosciaena crocea*). *Journal of Applied Microbiology*, 103(5): 1587-1599.
- Zhang, W. W., Sun, K., Cheng, S., & Sun, L. (2008). Characterization of DegQVh, a serine protease and a protective immunogen from a pathogenic *Vibrio harveyi* strain. *Applied and Environmental Microbiology*, 74(20): 6254-6262.
- Zhang, X. H., & Austin, B. (2000). Pathogenicity of *Vibrio harveyi* to salmonids. *Journal of Fish Diseases*, 23(2): 93-102.
- Zhang, X. H., & Austin, B. (2005). Haemolysins in *Vibrio* species. *Journal of Applied Microbiology*, 98(5): 1011-1019.
- Zhang, X., Robertson, P., Austin, B., & Xu, H. (1997). Comparison of outer membrane protein profiles of *Vibrio* sp. *Acta Microbiologica Sinica*, 37(6): 449-454.
- Zhou, L., Liu, H. M., & Zhan, W. B. (2003). Isolation and characteristics of major outer membrane proteins of aquatic pathogens *Vibrio anguillarum* and *Vibrio alginolyticus*. *Journal of Fishery Sciences of China*, 10(1): 31-35.
- Zorrilla, I., Arijo, S., Chabrillon, M., Diaz, P., Martinez- Manzanares, E., Balebona, M. C., & Morinigo, M. A. (2003a). *Vibrio* species isolated from diseased farmed sole, *Solea senegalensis* (Kaup), and evaluation of the potential virulence role of their extracellular products. *Journal of fish diseases*, 26(2): 103-108.
- Zorrilla, I., Morinigo, M. A., Castro, D., Balebona, M. C., & Borrego, J. J. (2003b). Intraspecific characterization of *Vibrio alginolyticus* isolates recovered from cultured fish in Spain. *Journal of Applied Microbiology*, 95(5): 1106-1116.