



UNIVERSITI PUTRA MALAYSIA

***DIFFERENTIAL POTASSIUM UPTAKE AND UTILIZATION EFFICIENCY
OF OIL PALM (*Elaeis guineensis* Jacq.) COMMERCIAL CULTIVARS***

SIM CHOON CHEAK

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By

SIM CHOON CHEAK

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy**

October 2017

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Abstract of thesis submitted to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Doctor of Philosophy.

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SIM CHOON CHEAK

October 2017

Chairman : Professor Zaharah Abdul Rahman, PhD
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Conventional approaches of using higher fertilizer inputs to sustain profitable yields in oil palm plantations can be uneconomical and produce inconsistent results. In addition, the biological potential of attaining much higher oil yield is often limited by marginal environments. Nutrient efficient genotypes could potentially lead to higher productivity when grown on marginal land and eventually improve the sustainable use of resources and production of palm oil. Studies on interaction effects between planting material and nutrient inputs show differential uptake and utilization efficiency between the commercially available oil palm planting materials. The differences in leaf nutrient contents between genotypes and yield response to K fertilizer inputs demonstrated the presence of more efficient uptake characteristics. If such potassium efficient cultivars could be widely adopted, the industry would not only be capable of saving resources but also to increase productivity as well. Potassium use in palm oil production ranges from approximately 13 to 21 kg of palm oil per kg of potassium with varying degree of efficiency depending on planting varieties. The potassium use efficiency could potentially increase by 50 % in the most potassium efficient cultivar. The objectives of this study were (1.) to evaluate the growth response of selected oil palm crosses under K deficient environment and (2.) to estimate potassium use efficiency of different oil palm genotypes as part of the effort to elucidate the physiological mechanism potassium-efficient oil palms. Phenotypic responses of 5 oil palm genotypes with genetic origin from Deli and Nigerian Dura interbred with AVROS, Nigerian and Yangambi Pisiferas grown under deficient and adequate potassium supplies were evaluated. Potassium-efficient genotypes were differentiated in this experiment, where the potassium-efficient genotypes produced higher biomass (by 37.3 %) and had higher potassium uptake activity (by 41.7 %). The efficient genotypes were capable of extracting higher amount of soil potassium (by 95 %) under deficient potassium supplies. The K-efficient genotype was capable of sustain growth and to adapt to potassium-deficient environments. Alterations in rooting behaviour (increasing fine root proliferation) and maintenance of shoot growth (frond production rate) are the primary physical traits of adaptation to

potassium-deficient environment. The ability to remobilize the limiting nutrients from sink tissues to source tissues i.e. from the bole and rachis to the pinnae (the photosynthetically active tissues) and roots (to search for more nutrients allows the plant to further acquire more resources to ensure continuous growth) is also a key trait. Comparative analysis of transcriptomic differences between the efficient and in-efficient genotypes showed significant upregulation of potassium transporters (KUP3, KUP8 and KUP11) in the roots of the K-efficient genotype and genes which confer tolerance to stress, minimizes cellular damage, stress regulation and potassium homeostasis. Traits for potassium efficiency are conferred by the interaction of multiple complex mechanisms, governed by pool of genes controlling the physiological processes of stress regulation, cellular development and metabolite homeostasis. Stress detection and regulating cellular processes to mitigate the effect of stress could be the key in first tolerating and reducing damages to cellular and consequently enhancing the genotype's ability to adapt, absorb and utilize nutrients more effectively.

Abstrak tesis yang telah dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah.

**PERBEZAAN DALAM PENGAMBILAN DAN PENGGUNAAN KALIUM
OLEH KULTIVAR KOMERSIAL KELAPA SAWIT (*Elaeis guineensis* Jacq.)**

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Pendekatan konvensional menggunakan kadar pembajaan yang tinggi untuk mengekalkan penghasilan yang menguntungkan oleh industri kelapa sawit sering mencapai keputusan yang tidak konsisten dan tidak ekonomik. Di samping itu, potensi untuk mendapatkan hasil minyak sawit yang tinggi sering dijejaskan oleh keadaan persekitaran yang tidak memberangsangkan. Genotip yang cekap di dalam penggunaan nutrien mampu meningkatkan potensi produktiviti dan mengurangkan penggunaan sumber. Kajian terhadap interaksi di antara genotip dan nutrien menunjukkan perbezaan dalam pengambilan dan penggunaan nutrien antara kultivar komersial. Penanaman kultivar yang cekap dalam penggunaan kalium memerlukan baja kalium yang rendah dan secara tidak langsung mampu meningkatkan hasil sawit nasional serta menjimatkan penggunaan sumber baja. Ketika ini, industri kelapa sawit menghasilkan antara 13 hingga 21 kg minyak sawit untuk setiap kilogram baja kalium diggunakan. Penggunaan kultivar yang cekap dalam kalium mampu meningkatkan hasil sebanyak 50 %. Objektif kajian ini adalah untuk menilai pengaruh kalium dalam tanah terhadap pertumbuhan genotip sawit terpilih dan kecekapan penggunaan kalium oleh baka tanaman sawit ini. Di samping itu, mekanisme fisiologi dalam penggunaan kalium akan dikaji. Tindak balas fenotip baka tanaman yang berlainan asal-usulnya iaitu AVROS, Nigeria dan Yangambi dibawah pengaruh kalium telah dikaji. Kadar pengambilan kalium dan pertumbuhan pokok diukur di dalam rumah kaca dengan menggunakan teknik radioisotop. Tindak balas molekular telah diprofilkan dan dikira. Genotip yang ber-kecekapan tinggi berupaya menghasilkan biojisim yang lebih tinggi (> 37.3 %), mempunyai kadar pengambilan kalium yang lebih tinggi (> 41.7 %) dan berupaya mengekstrak kandungan kalium yang lebih tinggi (> 95 %) daripada tanah yang mempunyai kandungan kalium yang rendah. Genotip yang sedemikian menyesuaikan diri kepada persekitaran kalium rendah dengan mengubah tingkah laku perakaran, meningkatkan percambahan akar dan mengekalkan nisbah akar dan daun. Tiada variasi atau perubahan morfologi di antara genotip. Analisis perbandingan perbezaan transkrip antara genotip cekap dan cekap memperlihatkan peningkatan besar pengangkut potasium (KUP3, KUP8 dan KUP11) dalam akar genotip dan gen yang efisien yang memberikan toleransi terhadap stres, mengurangkan kerosakan selular, peraturan tekanan dan kalium homeostasis. Ciri-ciri kecekapan kalium diberikan oleh interaksi pelbagai mekanisme kompleks, yang

dikendalikan oleh gen gen yang mengawal proses fisiologi pengawalan tekanan, pembangunan selular dan homeostasis metabolit. Pengesanan tekanan dan mengawal selia proses selular untuk mengurangkan kesan tekanan boleh menjadi kunci dalam menoleransi pertama dan mengurangkan kerosakan kepada selular dan seterusnya meningkatkan keupayaan genotip untuk menyesuaikan, menyerap dan menggunakan nutrien dengan lebih berkesan.



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Forever indebted.

Sim Choon Cheak
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LIST OF ABBREVIATIONS

^{86}Rb	Radioactive isotope of Rubidium
Ca	Calcium
CEC	Cation exchange capacity
CPM	Count per minute
CRD	Complete Randomized Design
DPM	Disintegrations per minute
ICP-OES	Inductively Coupled Plasma - Optical Emission Spectrometry
K	Potassium
KdF	Potassium derived from fertilizer
KUE	Potassium use efficiency
KUpE	Potassium uptake efficiency
KUtE	Potassium utilization efficiency
Mg	Magnesium
MgUpE	Magnesium uptake efficiency
MgUtE	Magnesium utilization efficiency
N	Nitrogen
NE	Nutrient efficient
NitUpE	Nitrogen uptake efficiency
NitUtE	Nitrogen utilization efficiency
P	Phosphorus
PUpE	Phosphorus uptake efficiency
PUtE	Phosphorus utilization efficiency
RCBD	Randomized Complete Block Design
RNA	Ribonucleic acid



CHAPTER 1

INTRODUCTION

Occupying merely 4.8 % of the total oil crop area planted worldwide, the oil palm contributed approximately 34.7 % of the global vegetable oil production, ranking it the highest oil producing crop known to-date. At a global production average of 3.8 tonnes of crude palm oil (CPO) per hectare, the palm oil productivity has very much stagnated over the last two decades especially in Malaysia. The stagnation could be attributed to the expansion of oil palm plantation on marginal land where the soil and terrain is least favorable for oil palm cultivation (Jalani *et al.* 2002). The efficient dissemination of selected planting material to end users, particularly to small holders is also a key factor for productivity improvement in the oil palm.

The projected biological potential of oil palm via experimental data indicates that the best progenies could be capable of yielding 11.5 tonnes of CPO per hectare. The highest plantation yield reported to-date achieved an average of about 6.5 tonnes of CPO, which is equivalent to about half of its biological yield potential. The vast gap between this biological yield potential and actual yield attained indicates an immense opportunity for yield improvement. The productivity of CPO increased from 1.3 to 6.5 tonnes per hectare through breeding advances, introduction of pollinating weevils and improved management whilst fertilization alone accounted for 29 % of the increase (Davison, 1993; Corley and Tinker, 2003). Combination of both adequate fertilization regime and improved materials that could thrive under poor and marginal growing conditions could provide viable alternatives in ensuring and improving sustainable production of oil palms grown on those marginal areas.

Amongst all oil crops, oil palm consumed the highest amount of potash fertilizer i.e. about 7 % of total global potash consumption, whilst soybean and other oil crops collectively consumed 12 % (Heffer, 2013). On the contrary, consumption of nitrogenous and phosphorus fertilizer is relatively low with potash constituting almost 60 % of total fertilizer usage for oil palm cultivation. The demand of potassium inputs for growth and oil production increases in parallel to the yield and it is further elevated by losses caused by the degraded land in the tropics. On per hectare basis, the oil palm requires up to 250 kg of potassium annually (Goh *et al.* 1994) and its planting on tropical soils that are inherently low in potassium fertility and prone to potassium leaching (Amberger, 2006) further contributes to high potassium requirement.

Yield responses to potassium fertilization are mostly soil-dependent and are affected by the soils inherent potassium fertility; higher yield response is recorded on soils with low soil potassium reserve and vice versa. Such yield response variations often translate into varying degree of optimal potash fertilization rate which is generally site specific. Generally, less than 1.3 kg K palm⁻¹ yr⁻¹ on fertile coastal soils to as high as 6.0 kg K

$\text{palm}^{-1} \text{ yr}^{-1}$ are needed for maximum yield on sandy textured soils. On most inland sedentary soils, many of the long-term fertilization trial showed an optimum range of 1.8 to 2.3 kg of potassium $\text{palm}^{-1} \text{ yr}^{-1}$.

Realizing such limitations, gradual effort is now being concentrated into improving the oil palm productivity via adopting nutrient efficient genotypes which could potentially lead to higher productivity on marginal land and eventually to a more sustainable use of resources and to the sustainable production of palm oil. The main objectives of this study are (1) to assess the variability of oil palm genotypes in potassium use efficiency and (2) to identify the physiological mechanism underlying the adaptation to potassium-deficient soil and the increase of potassium use efficiency.



REFERENCES

- Ahn S.J., Shin R., Schachtman D.P. (2004) Expression of KT/KUP genes in Arabidopsis and the role of root hairs in K⁺ uptake. *Plant Physiology* 134: 1135-1145.
- Ali L., Rahmatullah, Aziz T., Ashraf M., Maqsood M.A., Kanwal S. (2010) Differential potassium influx influences growth of two cotton varieties in hydroponics. *Pakistan J. Bot.* 42.2: 943-953.
- Amberger A. (2006) Soil fertility and plant nutrition in the tropics and subtropics. International Fertilizer Industry Association. IFA, Paris, France. ISBN 2-9523139-0-3.
- An S.H., Sohn K.H., Choi H.W., Hwang I.S., Lee S.C., Hwang B.K. (2008) Pepper pectin methylesterase inhibitor protein CaPMEI1 is required for anti-fungal activity, basal disease resistance and abiotic stress tolerance. *Planta*. 228: 61–78.
- Andrews M., Sprent J.I., Raven J.A., Eady P.E. (1999) Relationships between shoot to root ratio, growth and leaf soluble protein concentration of *Pisum sativum*, *Phaseolus vulgaris* and *Triticum aestivum* under different nutrient deficiencies. *Plant, Cell and Environment*. 22: 949-958.
- Argen G. and Franklin O. (2003) Root : Shoot ratios, optimization and nitrogen productivity. *Annals of Botany*. 92.6: 795-800.
- Armengaud P., Breitling R., Amtmann A. (2004) The potassium dependent transcriptome of arabidopsis reveals a prominent role of jasmonic acid in nutrient signaling. *Plant Physiology*. 136: 2556-2576.
- Armengaud P., Breitling R., Amtmann A. (2010) Coronatine-Insensitive 1 (COI1) mediates transcriptional responses of *Arabidopsis thaliana* to external potassium supply. *Molecular Plant*. 3.2: 390-405.
- Ashley M.K., Grant M., Grabov A. (2006) Plant responses to potassium deficiencies: a role for potassium transport proteins. *Journal of Experimental Botany*. 57: 425–436.
- Bailey T.L., Boden M., Buske F.A., Frith M., Grant C.E., Clementi L., Ren J.Y., Li W.W., Noble W.S. (2009) MEME Suite: Tools for motif discovery and searching. *Nucleic Acids Research*. 37: 202-208.
- Baligar V.C., Fageria N.K., He Z.L. (2001) Nutrient use efficiency in plants. *Communications in Soil Science and Plant Analysis*, 32: 7-8, 921-950.
- Bange G. (1979) Comparison of compartmental analysis for Rb⁺ and Na⁺ in low-salt and high-salt barley roots. *Physiol Plant* 46:179 – 183.
- Belknap W.R. and Garbarino J.E. (1996) The role of ubiquitin in plant senescence and stress response. *Trends in Plant Science*. 1.10: 331-335.

- Bennet P.C., Choi W.J., Rogera J.R. (1998) Microbial destruction of feldspars. *Mineral Management*. 8.62: 149-150.
- Benton J.J.Jr. (2001) *Laboratory guide for conducting soil tests and plant analysis*. New York: CRC Press LLC.
- Benton J.J.Jr. (2003). *Agronomic handbook. Management of crops, soils, and their fertility*. CRC Press, New York, USA.
- Bourgis F., Kilaru A., Cao X., Ngando-Ebongue G., Drira N., Ohlrogge J.B., Arondel V. (2011) Comparative transcriptome and metabolite analysis of oil palm and date palm mesocarp that differ dramatically in carbon partitioning. In: *Proceedings of the National Academy of Sciences of the United States of America*. 108.44:18186.
- Callis J. (2014) The ubiquitination machinery of the ubiquitin system. In: *The Arabidopsis Book*. 12: e0174. 1-35. doi: 10.1199/tab.0174.
- Chan P.L., Rose R.J., Abdul M.A.M., Zainal Z., Low E.T.L., Ooi C.L.L., Ooi S.E., Yahya S., Singh R. (2014) Evaluation of reference genes for quantitative real-time PCR in oil palm elite planting materials propagated by tissue culture. *PLOS One*. 9.6: e99774.
- Chen J. and Gabelman W.H. (1995) Isolation of tomato strains varying in potassium acquisition using a sand-zeolite culture system. *Plant Soil* 176 p65-70.
- Chen L. and Hellmann H. (2013) Plant E3 ligases: Flexible enzymes in a sessile world. *Molecular Plant* 6.5: 1388-1404.
- Cheng Y., Qi Y., Zhu Q., Chen X., Wang N., Zhao X., Chen H., Cui X., Xu L., Zhang W. (2009) New changes in the plasma membrane associated proteome of rice roots under salt stress. *Proteomics*. 9: 3100-3114.
- Claasen N. and Steingrobe B. (1999) Mechanistic simulation models for a better understanding of nutrient uptake from soil. In: Rengel Z (ed) *Mineral Nutrition of Crops: Fundamental Mechanisms and Implications*. Food Products Press, Binghamton, NY, 327 – 367.
- Claassen N., Syring K.M., Jungk A. (1986) Verification of a mathematical model by simulating potassium uptake from soil. *Plant Soil*. 95: 209 – 220.
- Conde A., Chaves M.M., Geros H. (2011) Membrane transport, sensing and signaling in plant adaptation to environmental stress. *Plant Cell Physiology*. 52.9: 1583-1602.
- Coolen S., Proietti S., Hickman R., Olivas N.H., Huang P., Van Verk M.C. (2016) Transcriptome dynamics of *Arabidopsis* during sequential biotic and abiotic stresses. *Plant Journal*. 86: 249-267.
- Corley R. H. V. and Tinker P. B. (2003) *The Oil Palm*, 4th edition. Blackwell Science. ISBN: 978-0-470-75036-0

- Cui F., Liu L.J., Zhao Q.Z., Zhang Z.H., Li Q.L., Lin B.Y., Wu Y.R., Tang S.Y., Xie, Q. (2012). Arabidopsis ubiquitin conjugase UBC32 is an ERAD component that functions in brassinosteroid-mediated salt stress tolerance. *Plant Cell*. 24: 233-244.
- Cuin T.A., Pottosin I.I., Shabala S.N. (2008) Mechanisms of potassium uptake and transport in higher plants. In: *Plant Membrane and Vacuolar Transporters* (ed. Jaiwal P.K., Singh R.P. and Dhanker O.P.) CABI, UK.
- Davidson L. (1993) Management for efficient cost-effective and productive plantations. In: *Proceedings of 1991 PORIM International Palm Oil Conference*, [Yusof B. et. al. (eds.)]. 153-167.
- Davies C., Shin R., Liu W., Thomas M.R., Schachtman D.P. (2006). Transporters expressed during grape berry (*Vitis vinifera* L.) development are associated with an increase in berry size and berry potassium accumulation. *Journal of Experimental Botany*. 57: 3209-3216.
- Day I.S., Reddy V.S., Ali G.S., Reddy A.S.N. (2002) Analysis of EF-hand-containing proteins in Arabidopsis. *Genome Biology*. 3.10: 56.1 - 56.24.
- Dereeper A., Guignon V., Blanc G., Audic S., Buffet S., Chevenet F., Dufayard J.F., Guindon S., Lefort V., Lescot M., Claverie J.M., Gascuel O. (2008) Phylogeny.Fr: Robust phylogenetic analysis for the non-specialist. *Nucleic Acids Research*. 36: 465-469.
- Dong B., Rengel Z., Graham R.D. (1995) Root morphology of wheat genotypes differing in zinc efficiency. *Journal of Plant Nutrition*. 18.12: 2761-2773.
- Dyson B.C., Allwood J.W., Feil R., Xu Y., Miller M., Bowsher C.G., Goodacre R., Lunn J.E., Johnson G.N. (2015) Acclimation of metabolism to light in Arabidopsis thaliana: the glucose 6-phosphate/phosphate translocator GPT2 directs metabolic acclimation. *Plant, Cell and Environment*. 38: 1404-1417.
- Dyson B.C., Webster R.E., Johnson G.N. (2014) GPT2: a glucose 6-phosphate/phosphate translocator with a novel role in the regulation of sugar signalling during seedling development. *Annals of Botany*. 113: 643-652.
- Epstein, E. and Bloom, A.J. (2004) *Mineral Nutrition of Plants: Principles and perspectives*. 2nd Edition, Sinauer Associates, Inc., Sunderland.
- Epstein, E., D.W. Rains, E. Elzam (1963) Resolution of dual mechanisms of potassium absorption by barley roots. *Proc. Nat. Acad. Sci. USA*. 49: 684-692.
- Fageria N.K. and Baligar V.C. (1994) Screening crop genotypes for mineral stresses. In: *Adaptation of plants to soil stress*, University Nebraska, Lincoln (Ed: Maranville et al.). 152-159.
- Fageria N.K., Baligar V.C., Li Y.C. (2008). The role of nutrient efficient plants in improving crop yields in the twenty first century. *Journal of Plant Nutrition*. 31:1121-1157.

- Fageria, N. K. and Baligar V.C. (1994) Screening crop genotypes for mineral stresses. In: *Adaptation of Plants to Soil Stress* (Ed: J.W. Maranville et al.) University of Nebraska, Lincoln, NE.
- Fatland B.L., Ke J., Anderson M.D., Mentzen W.I., Cui L.W., Allred C.C., Johnston J.L., Nikolau B.J., Wurtele E.S. (2002) Molecular characterization of a heteromeric ATP-citrate lyase that generates cytosolic acetyl-coenzyme A in Arabidopsis. *Plant Physiology*. 130: 740-756.
- Fatland B.L., Nikolau B.J., Wurtele E.S. (2005) Reverse genetic characterization of cytosolic Acetyl-CoA generation by ATP-Citrate Lyase in Arabidopsis. *The Plant Cell*. 17: 182-203.
- Foster H.L. (2002) Assessment of oil palm fertiliser requirements. In: *The oil palm – management for large and sustainable yields* (Ed. by T.H. Fairhurst & R. Hardter), Potash & Phosphate Institute of Canada (ESEAP), Singapore.
- Foster H.L. and Goh H.S. (1976) Yield response of oil palm to fertilizers in West Malaysia II: Influence of soil and climatic factors. *MARDI Research Bulletin*. 5.1: 6-22.
- Foster H.L. and Prabowo N.E. 2006. Partition and transfer of nutrients in the reserve tissues and leaves of oil palm. In: *Workshop on Nutrient Needs in Oil Palm. A Dialogue Among Experts*. October 17-18, 2006, Singapore 1-17.
- Foster H.L., Chang K.C., Tayeb D.M., Tarmizi A.M., Zin Z.Z.(1985) Oil Palm yield responses to N and K fertilizers in different environments in Peninsular Malaysia. *PORIM* 16:23.
- Foster H.L., Tayeb D.M., Gurmit S. (1988) The effect of fertilisers on oil palm bunch components in Peninsular Malaysia. In: *Proceedings of the 1987 International Oil Palm Conference*, PORIM and Inc. Soc. of Planters, Kuala Lumpur: 294-304.
- Freschet G.T., Swart E.M., Cornelissen J.H.C. (2015) Integrated plant phenotypic responses to contrasting above- and below- ground resources: key roles of specific leaf area and root mass fraction. *New Phytologist*. 206: 1247-1260.
- Gan S.T. (2014) The development and application of molecular markers for linkage mapping and quantitative trait loci analysis of important agronomic traits in oil palm (*Elaeis guineensis* Jacq.). PhD thesis. University of Nottingham, Malaysia.
- Gaydou E.M. and Arrivets J. (1983) Effects of phosphorus, potassium, dolomite, and nitrogen fertilization on the quality of soybean-Yields, proteins, and lipids. *Journal of Agricultural and Food Chemistry*, 31, 765-769.
- Gierth M. and Maser P. (2007) Potassium transporters in plants: Involvement in K⁺ acquisition, redistribution and homeostasis. *FEBS Letters*. 581: 2348-2356.
- Gierth M., Maser P., Schroeder J.I. (2005) The potassium transporter AtHAK5 functions in K⁺ deprivation-induced high affinity K⁺ uptake and AKT1 K⁺ channel

contribution to K⁺ uptake kinetics in Arabidopsis roots. *Plant Physiology*.137:1105-1114.

- Gleeson S.K. (1992) Optimization of tissue nitrogen and root-shoot allocation. *Annals of Botany*. 71: 23-31.
- Goh K.J., Chew P.S., Teo C.B. (1994a) Maximising and maintaining oil palm yields on commercial scale in Malaysia: In: ISP Planters Conference on Managing Oil Palms for Enhanced Profitability (Chee, K.H., ed), ISP, Kuala Lumpur: 121-141.
- Goh K.J., Chew P.S., Teoh K.C. (1994b) K nutrition for mature oil palms in Malaysia. *IPI Research Topics* 17: 36.
- Gomez A.K. and Gomez A.A. (1984) Statistical procedures for agricultural research. John Wiley & Sons, New York.
- Gomez-Herreros F., Miguel-Jimenez L., Morillo-Huesca M., Delgado-Ramos L., Munoz-Centeno M., Chavez S. (2012) TFIIS is required for the balanced expression of the genes encoding ribosomal components under transcriptional stress. *Nucleic Acids Research*. 40.14: 6508-6519.
- Grabov A. (2007) Plant KT/KUP/HAK potassium transporter: single family – multiple functions. *Annals of Botany*. 99.6: 1035-1041.
- Guglielmi B., Soutourina J., Esnault C., Werner M. (2007) TFIIS elongation factor and mediator act in conjunction during transcription initiation in vivo. In: In: Proceedings of the National Academy of Sciences of the United State of America, PNAS. 104.41: 16062-16067
- Guo Y., Xiong L., Ishitani M., Zhu J.K. (2002) An Arabidopsis mutation in translation elongation factor 2 causes superinduction of CBF/DREB1 transcription factor genes but blocks the induction of their downstream targets under low temperatures. In: Proceedings of the National Academy of Sciences of the United State of America, PNAS. 99.11: 7786-7791.
- Gupta M., Qiu X., Wang L., Xie W., Zhang C., Xiong L., Lian X., Zhang O. (2008) KT/HAK/KUP potassium transporters gene family and their whole-life cycle expression profile in rice (*Oryza sativa*). *Molecular Genetic Genomics*. 280: 437-452.
- Hashim M.A.T. (1999) Potassium requirement of mature oil palm on coastal soils. Phd Thesis. Universiti Putra Malaysia, Serdang, Malaysia.
- Haq M.U. and Mallarino A.P. (2005) Response of soybean grain oil and protein concentration to foliar and soil fertilization. *Journal of Agronomy*. 97: 910–918.
- Heffer P. (2009). Assessment of fertilizer use by crop at the global level (2006/07 – 2007/08). International Fertilizer Industry Association (IFA), France.
- Heffer P. (2013). Assessment of fertilizer use by crop at the global level, 2010-2011. AgCom/13/39-13/111. IFA Paris, France.

- Hiscox J.D. and Israelstam G.F. (1979) A method for the extraction of chlorophyll from leaf tissue without maceration. *Canadian Journal of Botany*. 57.12: 1332-1334.
- Hoagland D.R. and Arnon D.I. (1950) The water-culture method for growing plants without soil. *California Agricultural Experiment Station Circular*. 347: 1 – 32.
- Houston K., Tucker M.R., Chowdhury J., Shirley N., Little A. (2016) The plant cell wall: A complex and dynamic structure as revealed by the responses of genes under stress conditions. *Frontiers in Plant Science*. 7.984.: 1-18.
- Hozain M., Abdelmageed H., Lee J., Kang M., Fokar M., Allen R.D., Holaday A.S. 2012. Over-expressing AtSAP5 in cotton up-regulates putative stress-responsive genes and improves the tolerance to rapidly developing water deficit and moderate heat stress. *Journal of Plant Physiology*. 169: 1261-1270.
- Hsia M.M. and Callis J. (2010) BRIZ1 and BRIZ2 proteins form a heteromeric E3 ligase complex required for seed germination and post-germination growth in *Arabidopsis thaliana*. *The Journal of Biological Chemistry*. 285.47: 37070-37081.
- Huang B. and Eissenstat D.M. (2000) Root plasticity in exploiting water and nutrient heterogeneity. In: *Plant-Environment Interaction* (Ed: Wilkinson R.) Marcel Dekker Inc. New York.
- Inthapanya P., Sipaseuth, Sihavong P., Suhathep V., Chanphengsay M., Fukai S., Basnayake J. (2000). Genotype differences in nutrient uptake and utilization for grain yield production of rainfed lowland rice under fertilized and non-fertilized conditions. *Field Crops Research*. 65: 57-68.
- Jaafar H.Z.E. and Hafiz M.I. (2012) Photosynthesis and quantum yield of oil palm seedlings to elevated carbon dioxide. In: *Advances in Photosynthesis - Fundamental Aspects* (Ed: Najafpour M.). 16: 321-340. ISBN:978-953-307-9288.
- Jacquemard J.C., Ollivier J., Erwanda., Edyana S., Pelep P. (2010). Genetic signature in mineral nutrition in oil palm (*Elaeis guineensis* Jacq.): A new panorama for high yielding materials at low fertilizer cost. In: *International Oil Palm Conference*, 1-3 July 2010, Yogyakarta, Indonesia.
- Jacquemard J.C., Tailliez B., Dadang K., Ouvrier M., Asmady. (2002). Oil Palm (*Elaeis guineensis* Jacq.) nutrition: Planting material effect. In: *International Oil Palm Conference & Exhibition*, 8-12 July 2002, Bali, Indonesia.
- Jalani B.S., Yusuf B., Darus A., Chan K.W., Rajanaidu N. (2002) Prospects of elevating national oil palm productivity: a Malaysian perspective. *Oil Palm Industry Economic Journal*. 2.2: 1-9.
- Jia Y., Yang X., Feng Y., Jilani G. (2008) Differential response of root morphology to potassium deficient stress among rice genotypes varying in potassium efficiency. *Journal of Zhejiang University*. 9.5: 427-434.

- Jones B.Jr. (2003) Agronomic handbook: management of crops, soils, and their fertility. CRC Press, Florida.
- Jourdan C and Rey H. (1997) Architecture and development of the oil palm (*Elaeis guineensis* Jacq.) root system. *Plant and Soil*. 189: 33-48.
- Jungk A. (2002) Dynamics of nutrient movement at the soil-root interface. In: *Plant Roots: The Hidden Half*, 3rd edition. (Eds. Waisel et al.) 587-616. Marcel Dekker Inc., New York.
- Kang M., Fokar M., Abdelmageed H., Allen, R.D. (2011). Arabidopsis SAP5 functions as a positive regulator of stress responses and exhibits E3 ubiquitin ligase activity. *Plant Molecular Biology*. 75: 451-466.
- Kang M., Abdelmageed H., Lee S., Reichert A., Mysore K.S., Allen R.D. (2013). AtMBP-1, an alternative translation product of LOS2, affects abscisic acid responses and is modulated by the E3 ubiquitin ligase AtSAP5. *The Plant Journal*. 76: 481-493.
- Karim M., Shafiqur R., Mainur R. (1971) Rb86 as tracer for potassium: I. Uptake of Rb and K by rice plant in nutrient solution. *Plant and Soil*. 35.1: 179-181.
- Kim E.J., Kwak J.M., Uozumi N., Schroeder J.I. (1998) AtKUP1: an Arabidopsis gene encoding high-affinity potassium transport activity. *Plant Cell*. 10.1: 51-62.
- Kiribuchi K., Sugimori M., Takeda M., Otani T., Okada K., Onodera H., Ugaki M., Tanaka Y., Tomiyama-Akimoto C., Yamaguchi T., Nishiyama M., Nojiri H., Yamane H. (2004). RERJ1, a jasmonic acid-responsive gene from rice, encodes a basic helix-loop-helix protein. *Biochemical and Biophysical Research Communication*. 325: 857-863.
- Kosova K., Prasil I.T., Vitamvas P. (2013) Protein contribution to plant salinity response and tolerance acquisition. *International Journal of Molecular Sciences*. 14: 6757-6789.
- Krafczyk I., Trolldenier G., Beringer H. (1984) Soluble root exudates of maize: Influence of potassium supply and rhizosphere microorganisms. *Soil Biology and Biochemistry*. 16.4: 315-322.
- Kronzucker H.J., Szczerba M.W., Britto D.T. (2003) Cytosolic potassium homeostasis revisited: 42K-tracer analysis in *Hordeum vulgare* L. reveals set-point variations in [K⁺]. *Planta*, 217: 540 – 546.
- Kunz H.H., Häusler R.E., Fettke J., Herbst K., Niewiadomski P., Gierth M., Bell K., Steup M., Flügge U.I., Schneider A. (2010) The role of plastidial glucose-6-phosphate/phosphate translocators in vegetative tissues of *Arabidopsis thaliana* mutants impaired in starch biosynthesis. *Plant Biology*. 12:115-128.
- Läuchli A. and Epstein M. (1970) Transport of potassium and rubidium in plant roots. *Plant Physiology*. 45, 639 – 641.

- Lim C.K., Chong C.Y.W., Lin L.T.C., Ng P.H.C., Goh K.J., Melling L. (2016) Dynamic growth responses of various oil palm planting materials under different rate of fertilizers. In: Proceedings of the 7th International Crop Science Congress. 14th to 19th August. Beijing, China. Abstract No: E1-044.
- Lee C.T. (2004) Effects of genotypes, terrain and irrigation on oil palm yield, and leaf and rachis nutrient concentrations. PhD Thesis. Universiti Putra Malaysia, Serdang, Malaysia.
- Lee D., Redfern O., Orengo C. (2007) Predicting protein function from sequence and structure. *Molecular Cell Biology*. 8: 995-1005.
- Lee S.C., Lan W.Z., Kim B.G., Li L., Cheong Y.H., Pandey G.K., Lu G., Buchanan B.B., Luan S. (2007) A protein phosphorylation/dephosphorylation network regulates a plant potassium channel. In: Proceedings of the National Academy of Sciences of the United States of America. 104: 15959-15964.
- Le-Gall H., Philippe F., Domon J., Gillet F., Pelloux J., Rayon C. (2015) Cell wall metabolism in response to abiotic stress. *Plants*. 4:112-166
- Liao M.T., Fillery I.R.P., Palta J.A. (2004) Early vigorous growth is a major factor influencing nitrogen uptake in wheat. *Functional Plant Biology*. 31: 121-129.
- Liew V.K. (2008) Effects of empty fruit bunches application on oil palm root distribution, proliferation and nutrient uptake. PhD Thesis. Universiti Putra Malaysia, Serdang, Malaysia.
- Liu J. and Zhu J.K. (1998) A calcium sensor homolog required for plant salt tolerance. *Science*. 280: 1943 – 1945.
- Liu L.L., Ren H.M., Chen L.Q. Wang Y., Wu W.H. (2013) A protein kinase, calcineurin B-like protein-interacting protein kinase 9, interacts with calcium sensor calcineurin B-like protein 3 and regulates potassium homeostasis under low potassium stress in Arabidopsis. *Plant Physiology*. 161: 266-277.
- Livak K.J. and Schmittgen T.D. (2001) Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) method. *Methods*. 25.4: 402-408.
- Ljungman M. (2007) The transcription stress response. *Cell Cycle*. 6.18: 2252-2256.
- Lu L., Chen Y., Lu L., Lu Y., Li L. (2015) Transcriptome analysis reveals dynamic changes in the gene expression of tobacco seedlings under low potassium stress. *Journal of Genetics*. 94: 397 – 406.
- Maas E.V. and Leggett J.E. (1968) Uptake of ⁸⁶Rb and K by excised maize roots. *Plant Physiology*. 43.12: 2054-2056.
- Maathuis F.J.M., Filatov V., Herzyk P., Krijger G.C., Axelsen K.B., Chen S., Green B.J., Li Y., Madagan K.L., Sanchez-Fernandez R., Forde B.G., Palmgren M.G., Rea P.A., Williams L.E., Sanders D., Amtmann A. (2003) Transcriptome analysis of

root transporters reveals participation of multiple gene families in the response to cation stress. *The Plant Journal*. 35. 675-692.

- Mangeon A., Junqueira R.M., Sachetto-Martins G. (2010) Functional diversity of the plant glycine-rich proteins superfamily. *Plant Signalling & Behaviour*. 5.2: 99-104.
- Mao K., Dong Q., Li C., Liu C., Ma F. (2017) Genome wide identification and characterization of Apple bHLH transcription factors and expression analysis in response to drought and salt stress. *Frontiers in Plant Science*. 8: 480.
- Marschner, H. (1995). *The Mineral Nutrition of Higher Plants* (2nd Edition). Academic Press, USA.
- Marston A.L. (2015) Shugoshins: Tension-sensitive pericentromeric adaptors safeguarding chromosome segregation. *Molecular Cell Biology*. 35:634–648.
- Maser P., Thomine S., Schroeder J.I., Ward J.M., Hirschi K., Sze H., Talke I.N., Amtmann A., Maathuis F.J.M., Sanders D., Harper J.F., Tchieu J., Gribskov M., Persans M.W., Salt D.E., Kim S.A., Guerinot M.L. (2001) Phylogenetic relationships within cation transporter families of Arabidopsis. *Plant Physiology*. 126: 1646-1667.
- Melvin S.G., Lu G.Q., Zhou W.J. (2002). Genotypic variation for potassium uptake and utilization efficiency in sweet potato (*Ipomoea batatas* L.). *Field Crops Research*. 77: 7-15.
- Memon A.R., Saccomani M., Glass A.D.M. (1985) Efficiency of potassium utilization by barley *Hordeum vulgare* varieties the role of subcellular compartmentation. *Journal Experimental Botany*. 36: 1860 – 1876.
- Mi H., Dong Q., Muruganujan A., Gaudet P., Lewis S., Thomas O.D. (2010) PANTHER version 7: Improved phylogenetic trees, orthologs and collaboration with the Gene Ontology Consortium. *Nucleic Acids Research*. 38: 204-210.
- Mochida K. and Shinozaki K. (2011) Advances in omics and bioinformatics tools for system analyses of plant functions. *Plant Cell Physiology*. 52.12: 2017-2038.
- Mohidin H., Hanafi M.M., Rafii Y.M., Abdullah S.N.A., Idris A.S., Man S., Idris J., Sahebi M. (2015) Determination of optimum levels of nitrogen, phosphorus and potassium of oil palm seedlings in solution culture. *Bragantia*. 74.3: 247-254.
- Ng S.K. (2002) Nutrition and nutrient management of oil palm: New thrust for the future perspective. In: Pasricha, N.S., Bansal, S.K. (Eds.), *Potassium for Sustainable Crop Production: International Symposium on the Role of Potassium*. Potash Research Institute of India and International Potash Institute, New Delhi. 415–429.
- Nielsen N.C., Adey A., Stumpf P.K. (1979) Fat metabolism in higher plants: Further characterization of wheat germ Acetyl coenzyme A carboxylase. *Arch. Biochem. Biophys*. 192: 446-456.

- Pan X.W., Li W.B., Zhang Q.Y., Li Y.H., Liu M.S. (2008). Assessment on phosphorus efficiency characteristics of soybean genotypes in phosphorus-deficient soils. *Agricultural Sciences in China*. 7: 958-969.
- Paramanathan S, (2000). *Soils of Malaysia: Their characteristics and identification*, Volume 1. Academy of Sciences Malaysia, Kuala Lumpur, Malaysia.
- Park S.J., Kwak K.J., Oh T.R., Kim Y.O., Kang H. (2009) Cold shock domain proteins affect seed germination and growth of *Arabidopsis thaliana* under abiotic stress conditions. *Plant Cell Physiology*. 50: 869-878.
- Pelechano V., Jimeno-Gonzalez S., Rodriguez-Gil A., Garcia-Martinez J., Perez-Ortin J.E., Chavez S. (2009) Regulon-specific control of transcription elongation across the yeast genome. *PLoS Genetic*. 5: e1000614.
- Peter B.B., Jonathan R.H., Janina J., Rafael L.B., Saroj P., Caroline E.S., Malcolm J.H. (2010). Nitrogen efficiency of wheat: Genotypic and environmental variation and prospects for improvement. *European Journal of Agronomy*. 33: 1-11.
- Pettersson S. and Jensen P. (1983) Variation among species and varieties in uptake and utilization of potassium. *Plant and Soil*. 72.2: 231-237.
- Pettigrew W.T. (2008) Potassium influences on yield and quality production for maize, wheat, soybean and cotton. *Physiologia Plantarum*. 133: 670-681.
- Pilot G., Gaymard F., Mouline K., Cherel I., Sentenac H. (2003) Regulated expression of *Arabidopsis* Shaker K⁺ channel genes involved in K⁺ uptake and distribution in the plant. *Plant Molecular Biology*. 51: 773-787.
- Polley L.D. and Johns W.H. (1979) Rubidium (Potassium) uptake by *Arabidopsis*. *Plant Physiology*. 64: 374-378.
- Prabowo N.E. and Foster H.L. (1998) Variation in oil and kernel extraction rates of oil palms in North Sumatra due to nutritional and climatic factors. In: *International Oil Palm Conference*. Nusa Dua-Bali, Indonesia. September 23-25, 1998.
- Prabowo N.E., Foster H.L., Silalahi A.J. (2006) Recycling oil palm bunch nutrients. In: *International Oil Palm Conference*. June 19 – 23, 2006, Nusa Dua, Bali, Indonesia.
- Qian H., Lu H., Ding H., Lavoie M., Li Y., Liu W., Fu Z. (2015) Analyzing *Arabidopsis thaliana* root proteome provides insights into the molecular bases of enantioselective imazethapyr toxicity. *Scientific Report*. 5: 11975.
- Rankine I.R. and Fairhurst T.H. (1999) *Field Handbook: Oil Palm Series Volume 1ñ Nursery*. (Oil Palm Series), Potash & Phosphate Institute (PPI), Potash & Phosphate Institute of Canada (PPIC) and 4T Consultants (4T), Singapore, 135 p.
- Rengel Z. (2013) *Improving Water and Nutrient-Use Efficiency in Food Production Systems*. John-Wiley and Son. ISBN: 978-0-8138-1989-1.

- Rengel Z. and Damon P.M. (2008) Crops and genotypes differ in efficiency of potassium uptake and use. *Physiologia Plantarum*. 133.4: 624-636.
- Ringli C., Keller B., Ryser U. (2001) Glycine-rich proteins as structural components of plant cell walls. *Cellular and Molecular Life Sciences*. 58: 1430-1441.
- Rivera Y.D.M., Chacon L.M., Bayona C.J., Romero H.M. (2012) Physiological response of oil palm interspecific hybrids (*Elaeis oleifera* H.B.K. Cortes versus *Elaeis guineensis* Jacq.) to water deficit. *Brazilian Journal of Plant Physiology*. 24.4: 273-280.
- Römer W. and Schenk H. (1998). Influence of genotype on phosphate uptake and utilization efficiencies in spring barley. *European Journal of Agronomy*. 8: 215-224.
- Ruan L., Zhang J., Xin X., Zhang C., Ma D., Chen L., Zhao B. (2015) Comparative analysis of potassium deficiency-responsive transcriptomes in low potassium susceptible and tolerant wheat (*Triticum aestivum* L.) *Scientific Reports*. 5: 10090.
- Sachetto-Martins G., Franco L.O., Oliveira D. (2000) Plant glycine-rich proteins: a family or just proteins with a common motif. *Biochimica et Biophysica Acta*. 1942. 1 - 14.
- Samal D., Sadana U.S., Gill A.A.S. (2003) Mechanistic approach to study manganese influx and its depletion in the rhizosphere of wheat and raya. *Communications in Soil Science and Plant Analysis*. 34: 3033-3044.
- Sassi A., Mieulet D., Khan I., Moreau B., Gaillard I., Sentenac H., Very A. (2012). The rice monovalent cation transporter OsHKT2;4: revisited ionic selectivity. *Plant Physiology*. 160: 498–510.
- Schmittgen T.D. and Livak K.J. (2008) Analyzing real-time PCR data by the Comparative CT method. *Nature Protocols*. 3: 1101-1108.
- Seguin P. and Zheng W. (2006) Potassium, phosphorus, sulfur, and boron fertilization effects on soybean isoflavone content and other seed characteristics. *Journal of Plant Nutrition*. 29: 681–698.
- Sievers F., Wilm A., Dineen D., Gibson T.J., Karplus K., Li W.Z., Lopez R., McWilliam H., Remmert M., Soding J., Thompson J.D., Higgins D.G. (2011) Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Molecular Systems Biology*. 7: 539.
- Simkins C.A. and Overdahl C.J. (1982) Fertilizing sunflowers on sandy lands. In: *Better Crops*, Volume LXVI, Potash and Phosphate Institute, USA.
- Singh P. and Blanke M.M. (2000) Deficiency of potassium but not phosphorus enhances root respiration. *Plant Growth Regulation*. 32:77-81.

- Singh R., Ong-Abdullah M., Low E.T.L., Manaf M.A.A., Rosli R., Nookiah R., *et. al.* (2013). Oil palm genome sequence reveals divergence of interfertile species in Old and New worlds. *Nature*. 500:7462, 335-339.
- Song F., Han X., Zhu X., Herbert S.J. (2012) Response to water stress of soil enzymes and root exudates from drought and non-drought tolerant corn hybrids at different growth stages. *Canadian Journal of Soil Science*. 92.3: 501-507.
- Song S.K. and Huang P.M. (1988) Dynamics of potassium release from potassium bearing minerals as influenced by oxalic and citric acids. *Soil Science Society of America Journal*. 52: 383-390.
- Song Z.Z., Ma R.J., Yu M.L. (2015) Genome-wide analysis and identification of KT/HAK/KUP potassium transporter gene family in peach (*Prunus persica*). *Genetics and Molecular Research*. 14.1: 774-787.
- Sparks D.L. (2000) Bioavailability of soil potassium. In: *Handbook of Soil Science*, Section D1.4, pg D-38 to D-53. CRC Press, USA.
- Steingrobe B. (2005) A sensitive analysis for assessing the relevance of fine-root turnover for P and K uptake. *Journal of Plant Nutrition and Soil Science*. 168: 496-502.
- Szczerba M.W., Britto D.T., Kronzucker H.J. (2009) K⁺ transport in plants: Physiology and molecular biology. *Journal of Plant Physiology*. 166.5: 447-466.
- Taiz L. and Zeiger E. (2002) *Plant Physiology* (3rd Edition). Sinauer Associates, Inc. USA.
- Tan C.C. (2008) Nursery practices for production of superior oil palm planting materials. In: *Proc. Seminar on agronomic principles and practices of oil palm cultivation*. Agricultural Crop Trust Preprint. Also published in Goh K.J., Chiu S.B. and Paramanathan S. (eds). 2011. *Agronomic Principles and Practices of Oil Palm Cultivation*. Agricultural Crop Trust: 145-169.
- Terry J.R., Juan P.T., Michael T.R., Fukuta Y., Wissuwa M. (2010). Genotypic variation in grain phosphorus concentration and opportunities to improve P-use efficiency in rice. *Field Crops Research*. 119: 154-160.
- Tirol-Padre A., Ladha J.K., Singh U., Laureles E., Punzalan G., Akita S. (1996) Grain yield performance of rice genotypes at suboptimal levels of soil N as affected by N uptake and utilization efficiency. *Field Crops Research*. 46: 127-143.
- Trehan S.P. and Sharma R.C. (2002) Potassium uptake efficiency of young plants of three potato cultivars as related to root and shoot parameters. *Communications in Soil Science and Plant Analysis*. 33: 1813 - 1823.
- Trehan S.P., Dessougi H.E., Claassen N. (2005) Potassium efficiency of 10 potato cultivars as related to their capability to use non-exchangeable soil potassium by chemical mobilization. *Communications in Soil Science and Plant Analysis*. 36: 1809 - 1822.

- Tu S.X., Guo Z.F., Sun J.H. (2007) Effect of oxalic acid on potassium release from typical chinese soils and minerals. *Pedosphere*. 17.4: 457-466.
- Wang H., Inukai Y., Yamauchi A. (2006) Root development and nutrient uptake. *Critical Reviews in Plant Sciences*. 25: 279-301.
- Wang X., Vignjevic M., Jiang D., Jacobsen S., Wollenweber B. (2014) Improved tolerance to drought stress after anthesis due to priming before anthesis in wheat (*Triticum aestivum* L.) var. Vinjett. *Journal of Experimental Botany*. 65.22: 6441-6456.
- Wang H.Y., Shen Q.H., Zhou J.M., Wang J., Du C.W., Chen X.Q. (2011) Plants use alternative strategies to utilize nonexchangeable potassium in minerals. *Plant Soil*. 343: 209-220.
- Waters S., Gilliam M., Hrmova M. (2013) Plant High-Afinity Potassium (HKT) Transporters involved in salinity tolerance: Structural insight probe difference in ion selectivity. *International Journal of Molecular Sciences*. 14: 7660-7680.
- Weber E.J. (1985) Role of potassium in oil metabolism. In: *Potassium in Agriculture* (Munson R.D., ed). ASA, CSSA and SSSA, Madison, WI, pp 425-442.
- Wellburn A.R. (1994) The spectral determination of chlorophylls a and b, as well as total carotenoids, using various solvents with spectrophotometers of different resolution. *Journal of Plant Physiology*. 144: 307-313.
- White A.C., Rogers A., Rees M., Osborne C.P. (2016) How can we make plants grow faster? A source-sink perspective on growth rate. *Journal of Experimental Botany*. 67.1: 31-45.
- Wu R., Grissom J.E., McKeand S.E., O'Malley D.M. (2004) Phenotypic plasticity of fine root growth increases plant productivity in pine seedlings. *BMC Ecology*. 4: 14.
- Yan X.L., Wu P., Ling H.Q., Xu G.H., Xu F.S., Zhang Q.F. (2006) Plant Nutriomics in China: An Overview. *Annals of Botany* 98: 473-482.
- Yang T.Z., Lu L.M., Xia W., Fan J.H. (2007) Characteristics of potassium-enriched, flue-cured tobacco genotype in potassium absorption, accumulation and inward potassium currents of root cortex. *Agricultural Sciences in China*. 6.12: 1479-1486.
- Yin X. and Vyn T.J. (2003) Potassium placement effects on yield and seed composition of no-till soybean seed in alternate row widths. *Journal of Agronomy*. 95: 126-132.
- Zaharah A.R., Sharifuddin H.A.H., Sahali A.M., Hussein M.M.S. (1989) Fertilizer placement studies in mature oil palm using isotope technique. *Planter*. 65: 384-388.
- Zaharah, A.R., H.A.H. Sharifuddin, A.M. Sahali. (1991) The use of ^{86}Rb as a tracer to quantify potassium uptake by oil palm. *Proceedings of the PORIM International*

Palm Oil Conference: Progress, Prospects and Challenges Towards the 21st Century, September 9-14, 1991, Kuala Lumpur, Malaysia, pp: 499-501.

Zapata, F. and Axmann H. (1995) ^{32}P isotopic techniques for evaluating the agronomic effectiveness of rock phosphate materials. *Fertilizer Research*. 41: 189-195.

Zhao Y., Wang T., Zhang W., Li Xia (2011) SOS3 mediates lateral root development under low salts stress through regulation of auxin redistribution and maxima in *Arabidopsis*. *New Phytologist*. 189: 1122 - 1134

Zhiponova M.K., Morohashi K., Vanhoutte I., Machermer-Noonan K., Revalska M., Montagu M.V., Grotewold E., Russinova E. (2014) Helix-loop-helix/basic helix-loop-helix transcription factor network represses cell elongation in *Arabidopsis* through an apparent incoherent feed-forward loop. In: *Proceedings of the National Academy of Sciences of the United State of America, PNAS*. 111.7: 2824-2829.