



**UNIVERSITI PUTRA MALAYSIA**

***DEVELOPMENT OF ALLELOPATHIC RICE GENOTYPE WITH HIGH  
GRAIN ZINC CONTENT THROUGH MARKER-ASSISTED BACKCROSS***

**MUHAMMAD SIRAJUL MAZID**

**FP 2018 83**



**DEVELOPMENT OF ALLELOPATHIC RICE GENOTYPE WITH HIGH  
GRAIN ZINC CONTENT THROUGH MARKER-ASSISTED BACKCROSS**

By

**MUHAMMAD SIRAJUL MAZID**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirement for the Degree of Doctor of Philosophy**

**February 2018**

## **COPYRIGHT**

All material contained within the thesis including, without limitation, text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express prior written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## **DEDICATION**

**THIS THESIS IS SPECIALLY DEDICATED**

**TO**

**MY BELOVED PARENTS, BROTHERS, FAMILY AND THE PEOPLE WHO  
SUFFER DUE TO ZINC DEFICIENCY PROBLEMS**



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

## **DEVELOPMENT OF ALLELOPATHIC RICE GENOTYPE WITH HIGH GRAIN ZINC CONTENT THROUGH MARKER-ASSISTED BACKCROSS**

By

**MUHAMMAD SIRAJUL MAZID**

**February 2018**

**Chairman : Professor Abdul Shukor Juraimi, PhD**  
**Faculty : Agriculture**

Rice cultivation is hindered by biotic and abiotic stresses. Among the biotic stresses, weeds like barnyard grass can cause significant reduction of rice yield. The efficacious approach to control barnyard grass is by using allelopathic rice genotype integration management with minimum chemical control method. Zinc (Zn) deficiency is one of the critical problems of global malnutrition for humans. The problem affected approximately half of the world's population. The genetic biofortification together with agronomic biofortification of staple food crop is considered the most effective approach to prevent Zn deficiency. Marker-assisted selection (MAS) can potentially hasten breeding programs for varietal development. Marker-assisted backcrossing (MABC) is one of the most-useful ways of transferring specific gene(s) or QTL(s) into a popular variety. In this study, high grain Zn QTLs were transferred from an *indica* rice donor BRRI dhan62, using MABC method, into an allelopathic elite *indica* rice variety, MR73 to develop one superior genotype which has both allelopathic trait along with high grain Zn. Twenty eight microsatellite markers linked to the Zn QTLs or grain Zn content were selected and used in this study to determine potential association with Zn QTLs. These polymorphic foreground markers were used for confirmation of Zn QTLs or presence of grain Zn content gene in the F<sub>1</sub> population. Subsequently, only three microsatellite markers, RM152, RM3331 and RM3909 that indicated Zn QTLs located on chromosomes 8, 12 and 9 respectively, were used on BC<sub>1</sub>F<sub>1</sub>, BC<sub>2</sub>F<sub>1</sub> and BC<sub>2</sub>F<sub>2</sub> populations. Out of 405 microsatellite markers, 75 were found to have polymorphism between parental varieties and were used for recurrent parent genome recovery (RPG) analysis in each generation. Chi-square analyses of foreground markers showed an expected segregation ratio of 1:1 in BC<sub>1</sub>F<sub>1</sub> and BC<sub>2</sub>F<sub>1</sub> generations indicating inheritance in simple Mendelian fashion. This result confirmed that grain Zn content trait in BRRI dhan62 is governed by single dominant gene. The genotypic segregation of BC<sub>2</sub>F<sub>2</sub> population applying RM152, RM3331 and RM3909 exhibited 1:2:1 ratio. Results confirmed that single dominant gene control high grain Zn in BRRI

dhan62 located on chromosomes 8, 12 and 9 are linked to RM152, RM3331 and RM3909 markers respectively. The RPG analysis of the improved genotypes indicated recurrent parent genome recovery ranging from 70.20-91.40% in BC<sub>1</sub>F<sub>1</sub> generation, 80.50-93.00% in BC<sub>2</sub>F<sub>1</sub> and 93.10-95.40% in BC<sub>2</sub>F<sub>2</sub>. The average RPG in BC<sub>2</sub>F<sub>2</sub> selected improved genotypes were 93.86%, explaining the very close similarity at phenotypic level with the recipient parental variety MR73. Ten homozygous dominant Zn QTLs plants which are phenotypically closely similar for agro-morphological traits to MR73 were selected as improved high grain Zn breeding genotypes. The agronomic traits showed no significant difference between MR73 parent and Zn QTLs containing MR73 improved genotypes except the trait days to 50% flowering. Moreover, average Zn content in brown rice of improved genotypes was 40 mg kg<sup>-1</sup> which was higher than the recurrent parent MR73 (27.99 mg kg<sup>-1</sup>). Furthermore, the ability of newly improved rice genotypes to suppress the growth of barnyard grass which is expressed through average percent inhibition was 39.98%. The improved rice genotypes including these research findings could be used as valuable source for further high grain Zn rice breeding programmes in future.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia untuk memenuhi keperluan untuk Ijazah Doktor Falsafah

## **PEMBANGUNAN GENOTIP PADI ALELOPATIK DENGAN KANDUNGAN ZINK BIJIAN YANG TINGGI MELALUI KACUK BALIK BANTUAN PENANDA**

Oleh

**MUHAMMAD SIRAJUL MAZID**

**Februari 2018**

**Pengerusi : Profesor Abdul Shukor Juraimi, PhD**  
**Fakulti : Pertanian**

Penanaman padi menghadapi kekangan oleh tekanan biotik dan abiotik. Di antara tekanan biotik, rumput seperti rumput sambau boleh mengakibatkan pengurangan hasil padi yang ketara. Pendekatan yang berkesan untuk mengawal rumput sambau ini adalah melalui penggunaan genotip padi alelopatik serta pengurusan bersepadu dengan penggunaan kaedah pengawalan kimia yang minima. Kekurangan unsur zink (Zn) adalah salah satu masalah malnutrisi yang kritikal di peringkat global. Masalah ini melibatkan kira-kira separuh penduduk dunia. Peningkatan nutrien tanaman makanan ruji secara genetik dan agronomik dianggap sebagai cara yang paling berkesan untuk menangani masalah kekurangan unsur Zn. Pemilihan dengan bantuan penanda (MAS) berpotensi untuk mempercepatkan program pembiakbakaan dalam pembangunan sesuatu varieti. Kacuk balik dengan bantuan penanda (MABC) adalah salah satu kaedah yang paling berjaya untuk memindahkan gen tertentu atau QTL ke dalam sesuatu varieti popular. Dalam kajian ini, QTLs untuk unsur Zn bijian yang tinggi telah dipindahkan dari padi *indica* induk BRRI dhan62 kepada satu varieti elit padi *indica* beralelopatik, MR73 dengan kaedah MABC, untuk menghasilkan satu genotip baharu yang mempunyai ciri unsur Zn bijian yang tinggi berserta beralelopatik. Dua puluh lapan penanda mikrosatelit yang berkaitan dengan QTL Zn atau kandungan Zn bijian dipilih dan digunakan dalam kajian ini untuk mengenalpasti kaitannya dengan QTL-Zn. Penanda polimorfik berkait rapat dengan Zn ini digunakan untuk pengesanan QTL-Zn atau kewujudan gen kandungan Zn bijian dalam populasi  $F_1$ . Selepas itu, hanya tiga penanda mikrosatelit iaitu RM152, RM3331 dan RM3909 yang menunjukkan QTL Zn yang masing-masing terletak di kromosom 8, 12 dan 9 telah digunakan ke atas populasi  $BC_1F_1$ ,  $BC_2F_1$  dan  $BC_2F_2$ . Dari 405 penanda mikrosatelit, 75 penanda telah didapati memberikan polimorfik di antara varieti induk dan ianya telah digunakan untuk analisis pemuliharaan genom induk penerima (RPG) untuk setiap generasi kacuk balik. Analisa khi-kuasadua ke atas penanda berkait rapat dengan Zn

telah menunjukkan nisbah segregasi 1:1 seperti nisbah Mendelian dijangka iaitu mengikut pewarisan gen tunggal. Keputusan ini menunjukkan ciri kandungan zink bijian varieti BRRI dhan62 adalah dikawal oleh gen dominan tunggal. Nisbah segregasi genotip dalam populasi  $BC_2F_2$  yang dikesan oleh penanda RM152, RM3331 and RM3909 memberikan nisbah 1:2:1. Keputusan ini juga mengesahkan bahawa gen tunggal dominan mengawal kandungan zink bijian dalam varieti BRRI dhan62 yang terletak di kromosom 8, 12 dan 9 yang berkaitan masing-masing kepada penanda-penanda RM152, RM3331 dan RM3909. Analisis RPG ke atas hasil kacukan menunjukkan pemulihan genom induk penerima adalah dari 70.20 - 91.40% dalam generasi  $BC_1F_1$ , 80.50-93.00% dalam  $BC_2F_1$  dan 93.10-95.30% dalam  $BC_2F_2$ . Purata RPG untuk genotip terpilih  $BC_2F_2$  adalah 93.85%, dan ianya adalah hampir menyamai fenotip dengan varieti induk penerima, MR73. Sepuluh pokok dominan homozigot QTL zink yang fenotip agro-moforlogi adalah hampir sama dengan MR73 telah dipilih sebagai genotip maju padi yang tinggi kandungan bijian Zn. Analisis ciri-ciri agronomik menunjukkan tiada perbezaan antara induk MR73 dan genotip maju terpilih MR73 yang mengandungi QTL Zn yang tinggi kecuali bilangan hari 50% berbunga. Tambahan pula, purata kandungan Zn dalam beras perang genotip maju terpilih adalah 40 mg kg<sup>-1</sup>, iaitu lebih tinggi daripada varieti induk penerima MR73 (27.99 mg kg<sup>-1</sup>). Seterusnya, keupayaan genotip maju terpilih padi baharu ini untuk mengawal pertumbuhan rumput sambau yang ditunjukkan melalui purata peratus perencatan adalah 39.98%. Genotip maju padi ini serta dapatan kajian ini adalah sumber yang bernilai yang boleh digunakan dalam program peningkatan kandungan Zn bijian seterusnya dalam program pembiakbakaan di masa hadapan.



## ACKNOWLEDGEMENTS

Beginning in the name of Allah. Firstly, I would like to reveal my utmost sincere gratitude to Almighty Allah who has bestowed knowledge, energy, and patience upon me to complete this research work as well as thesis. I would like to thank my research supervisor Professor Dr. Abdul Shukor Juraimi, not only for accepting me as his student, good advices, suggestions, continuous supports on the research activities in every steps, on manuscript and his guidance but also for providing the motivation and encouragement from the beginning to the end of this project. Without his prompt support during the thesis writing period, it would not be possible to bring the thesis to the last stage. His commendable supervision facilitated to complete my research work timely.

I would like to give special thanks to my committee members Professor Dr. Mohd Raffi Yusop, Professor Dr. Mohamed Hanafi Musa, Dr. Mohamad Abdul Hakim and Dr. Jannatul Ferdous for their untiring and constructive suggestions, remarks and help in the previous years.

I also wish to give thanks to Dr. Mahbod Shahebi, Dr. Perisha, Dr. Sima Taheri and Dr. Gous Miah who provided timely support.

I owe to Dr. Asfaliza Ramli, Dr. Kauser Hosain, Dr. Parvez Anwar, Dr. Anwar, Dr. Moniruzzaman, scientists from BINA, my friends and lab mates Oladosu Yusuff, Osman Magaji, Ibrahim Wasi Arlou, Rehana, Masitah, Azua and Tan (from ITA FooS and Crop Science), staffs (from Crop Science, ITA FooS and Land Management) and many more who always helped me whenever it required. Special gratitude goes to the officers of School of Graduate Studies (Universiti Putra Malaysia), who selected me for International Graduate Research Fellowship (for 11 months) during my study period that encouraged me to bear the massive study load.

I would like to remember my previous teachers Prof. Dr. Kamal Uddin Ahamed, Prof. Dr. Md. Shahidur Rashid Bhuiyan, Prof. Dr. Naheed Zeba, Prof. Dr. Md. Sarowar Hossain Prof. Md. Zulfikar Ahmed Reza, Prof. Ruhul Amin of Sher-e-Bangla Agricultural University for their encouragements and inspiration during my study of B.Sc. in agriculture.

I would like to express my profound gratitude to my parents, relatives and family members specially my younger brother Muhammad Lutful Kabir, my wife, my son and daughter for their love, patience and faith.

This thesis was submitted to the senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the Doctor of Philosophy. The members of the supervisory committee were as follows:

**Abdul Shukor Juraimi**

Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Chairman)

**Mohd Rafii Yusop, PhD**

Professor  
Faculty of Agriculture  
Universiti Putra Malaysia  
(Member)

**Mohamed Hanafi Musa, PhD**

Professor  
Institute of Tropical Agriculture and Food Security  
Universiti Putra Malaysia  
(Member)

**Mohamad Abdul Hakim, PhD**

Research Fellow  
Institute of Tropical Agriculture and Food Security  
Universiti Putra Malaysia  
(Member)

**Jannatul Ferdous, PhD**

Senior Scientific officer  
Biotechnology Division  
Bangladesh Rice Research Institute  
(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_

Date: \_\_\_\_\_

Name and Matric No.: Muhammad Sirajul Mazid, GS36640

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_

Name of  
Chairman of  
Supervisory  
Committee:

\_\_\_\_\_  
Professor Abdul Shukor Juraimi

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee:

\_\_\_\_\_  
Professor Dr. Mohd Raffi Yusop

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee:

\_\_\_\_\_  
Professor Dr. Mohamed Hanafi Musa

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee:

\_\_\_\_\_  
Dr. Mohamad Abdul Hakim

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee:

\_\_\_\_\_  
Dr. Jannatul Ferdous

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	xviii
<b>LIST OF TABLES</b>	xiv
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xvi
 <b>CHAPTER</b>	
 <b>1 INTRODUCTION</b>	 1
1.1 General introduction	1
1.2 Problem statement	1
1.3 Objective of the study	3
 <b>2 LITERATURE REVIEW</b>	 4
2.1 Rice ( <i>Oryza sativa</i> L.)	4
2.1.1 Taxonomy of Rice	5
2.1.2 Status of rice production in Malaysia	5
2.1.3 Genetic resources of rice in Malaysia	5
2.1.4 MR73 high-yielding allelopathic rice variety	6
2.1.5 BRRI Dhan 62 high-yielding zinc enriched rice variety	6
2.1.6 Major weeds of rice in Malaysia	6
2.2 Barnyard grass as a major rice weed	7
2.2.1 Species of Barnyard grass in Malaysia	7
2.2.2 Management of Barnyard grass in rice field	7
2.3 Zinc deficiency problem in world population	9
2.3.1 Causes of zinc deficiency	9
2.3.2 Zinc deficiency symptoms and consequences in human body	9
2.3.3 Strategies to control zinc deficiency in human body	10
2.3.4 Physiological basis of grain zinc in rice plant	10
2.3.5 Zinc in grain of rice plant and genetic basis	12
2.3.6 Zinc in grain of rice plant and molecular basis	12
2.3.7 Molecular markers for grain zinc concentration in rice plant	14
2.3.8 Breeding approaches to enhance zinc in grain of rice plant	14
2.3.9 Microsatellites in rice breeding programme	15

	2.3.9.1 Advantages and disadvantages of microsatellites markers	15
	2.3.9.2 Applications of microsatellites markers	16
2.4	Conventional breeding and marker-assisted backcrossing	16
2.4.1	Advantages and disadvantages of marker-aided backcrossing	18
2.4.2	Pre-requisite of a successful backcross breeding program	19
2.4.3	Recurrent parent genome recovery using marker-aided backcrossing	19
2.5	Marker-aided selection and rice variety development	20
2.5.1	Marker-aided foreground selection	21
2.5.2	Marker-aided recombinant selection	22
2.5.3	Marker-aided background selection	22
<b>3</b>	<b>VALIDATION AND INHERITANCE PATTERN OF MICROSATELLITE MARKERS LINKED TO GRAIN ZINC QTLs (PUTATIVE) IN MR73 AND BRRI dhan62 CROSSES</b>	<b>23</b>
3.1	Introduction	23
3.2	Materials and Methods	23
3.2.1	Plant materials and crossing scheme	23
3.2.2	Microsatellite marker analysis	26
3.2.3	Extraction and dilution of DNA from leaf samples	28
3.2.4	Pre-conditions of polymerase chain reaction	28
3.2.5	Genotyping of marker segregation	29
3.2.6	Statistical analysis	29
3.3	Results and Discussion	29
3.3.1	Quantity and quality of DNA	29
3.3.2	Polymorphic SSR markers and markers linked to high grain zinc	30
3.3.3	Segregation analysis of markers in BC <sub>2</sub> F <sub>1</sub> population	33
3.4	Conclusion	34
<b>4</b>	<b>ESTIMATION OF RECURRENT PARENT GENOME RECOVERY IN MARKER AIDED BACKCROSS POPULATIONS DERIVED FROM CROSS BETWEEN MR73 AND BRRI dhan62</b>	<b>36</b>
4.1	Introduction	36
4.2	Materials and Methods	37
4.2.1	Plant materials and strategy of breeding	37
4.2.2	Leaf samples collection	37
4.2.3	Analysis of molecular markers	37
4.2.3.1	Marker-aided foreground selection	37
4.2.3.2	Marker-aided background selection	38
4.2.4	Extraction of DNA and pre-conditions of PCR and electrophoresis	38

4.2.5	Analysis of data	39
4.3	Results and Discussion	39
4.3.1	Polymorphism of markers in the parental lines	39
4.3.2	Genotypic survey in BC <sub>1</sub> F <sub>1</sub> population	39
4.3.2.1	Foreground selection of high grain zinc QTLs	39
4.3.2.2	Background selection for recovery of recurrent parent	42
4.3.3	Genotypic survey in BC <sub>2</sub> F <sub>1</sub> population	47
4.3.3.1	Foreground selection of high grain zinc QTLs	47
4.3.3.2	Background selection for recovery of recurrent parent	48
4.3.4	Evaluation of agro-morphological performance in BC <sub>2</sub> F <sub>1</sub> population	53
4.4	Conclusion	58
<b>5</b>	<b>PYRAMIDING OF HIGH GRAIN ZINC QTLs (PUTATIVE) INTO ELITE RICE VARIETY MR73 THROUGH MARKER-AIDED BACKCROSS BREEDING</b>	<b>59</b>
5.1	Introduction	59
5.2	Materials and Methods	60
5.2.1	Raising parental lines	60
5.2.2	Development of improved lines	60
5.2.3	Analysis of molecular markers	61
5.2.4	Extraction of DNA from leaf samples and analysis of PCR	62
5.2.5	Evaluation of improved lines for agro-morphological traits	62
5.2.6	Screening for allelopathic trait in improved rice lines	63
5.2.7	Screening for high zinc content in the grain of improved rice lines	64
5.2.8	Statistical analysis	64
5.3	Results and Discussion	65
5.3.1	Marker-assisted foreground selection for high grain zinc QTLs	65
5.3.2	Amount of recurrent parent genome in improved lines	67
5.3.3	Comparison between the parent MR73 and improved lines for agro-morphological Traits	72
5.3.4	Evaluation of allelopathy in improved lines	74
5.3.5	Evaluation of zinc content in the grains of improved lines	74
5.4	Conclusion	78

<b>6</b>	<b>GENERAL DISCUSSION, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	<b>80</b>
6.1	General discussion	80
6.2	Conclusions	82
6.3	Recommendations for future research	83
	<b>REFERENCES</b>	<b>85</b>
	<b>APPENDICES</b>	<b>107</b>
	<b>BIODATA OF STUDENT</b>	<b>114</b>
	<b>LIST OF PUBLICATIONS</b>	<b>115</b>





## LIST OF TABLES

Table	Page
2.1 Different weed control methods and their advantages and disadvantages	8
2.2 Some high grain zinc QTLs and putative candidate genes identified in rice from different studies	13
2.3 Applications of MAS and conventional breeding approaches in some breeding programmes	18
2.4 Comparison between conventional and marker-assisted backcross breeding for recovery of recurrent parent genome	20
3.1 Information on the microsatellite markers linked to high grain zinc QTLs and candidate genes used in foreground analysis in this study	27
3.2 Marker segregation analysis in BC <sub>2</sub> F <sub>1</sub> lines derived from the cross between MR73 and BRRI dhan62 rice varieties	34
4.1 Proportion of grain Zn QTLs having genotypes in BC <sub>1</sub> F <sub>1</sub> generations	41
4.2 Background and introgressed segment analysis in selected lines of BC <sub>1</sub> F <sub>1</sub> population	44
4.3 Proportion of grain Zn QTLs having genotypes in BC <sub>2</sub> F <sub>1</sub> generations	49
4.4 Background and introgressed segment analysis in selected lines of BC <sub>2</sub> F <sub>1</sub> population	51
4.5 Agro-morphological trait performance of improved lines in comparison with recurrent parent MR73	54
5.1 Descriptions of the agronomic traits measured in parents and BC <sub>2</sub> F <sub>2</sub> populations	63
5.2 Allele sizes of the foreground markers linked to high grain zinc QTLs in zinc enriched and zinc poor parents	65
5.3 Foreground markers analysis in BC <sub>2</sub> F <sub>2</sub> population	67

5.4	Background and introgressed segment analysis in ten selected improved lines	68
5.5	Major agronomic traits of promising BC <sub>2</sub> F <sub>2</sub> breeding lines carrying high grain zinc QTLs	72
5.6	Evaluation of allelopathy and grain zinc content in BC <sub>2</sub> F <sub>2</sub> lines carrying high grain zinc QTLs	74
5.7	Amount of zinc content in the improved lines based on Zn QTLs	74



## LIST OF FIGURES

Figure		Page
2.1	The shapes and relative sizes of the <i>Oryza sativa</i> chromosomes	4
3.1	High yielding allelopathic rice variety (MR73) and high grain zinc content rice variety (BRRI dhan62)	24
3.2	Crossing and selection scheme to produce grain zinc enriched BC <sub>2</sub> F <sub>1</sub> and BC <sub>2</sub> F <sub>2</sub> genotypes	25
3.3	Steps of the seed production through crossing	26
3.4	Nanodrop technologies used for determination of DNA quantity and quality	30
3.5	Genotyping with markers (a) RM152, (b) RM3331 and (c) RM3909 linked to high grain zinc QTLs in F <sub>1</sub> population of rice derived from MR73 (PM) X BRRI dhan 62(PB).	32
3.6	Genotyping with markers (a) RM152, (b) RM3331 and (c) RM3909 linked to high grain zinc QTLs in BC <sub>2</sub> F <sub>1</sub> population of rice derived from MR73 (PM) X BRRI dhan 62(PB).	33
4.1	Screening of high grain zinc QTLs having plant using (a) RM152 (b)RM3331 and (c) RM3909 markers in BC <sub>1</sub> F <sub>1</sub> population	41
4.2	Banding pattern of background markers in BC <sub>1</sub> F <sub>1</sub> generation	43
4.3	Frequency distribution of the recurrent parent genome recovery in BC <sub>1</sub> F <sub>1</sub> population	43
4.4	Chromosome-wise the highest recurrent parent genome recovery of the plant no. 16-3 in BC <sub>1</sub> F <sub>1</sub> generation	45
4.5	Chromosome-wise recurrent genome recovery of the four selected plants in BC <sub>1</sub> F <sub>1</sub> generation.	46
4.6	Screening of high grain zinc QTLs having plant using (a) RM152 (b) RM3331 and (c) RM3909 markers in BC <sub>2</sub> F <sub>1</sub> population	48
4.7	Banding pattern of background markers in BC <sub>2</sub> F <sub>1</sub> generation	50
4.8	Frequency distribution of the recurrent parent genome recovery	51

in BC<sub>2</sub>F<sub>1</sub> population

4.9	Chromosome-wise recurrent genome recovery of the five selected plants in BC <sub>2</sub> F <sub>1</sub> generation.	52
4.10	Chromosome-wise the highest recurrent parent genome recovery of the plant no. 16-3-38 in BC <sub>2</sub> F <sub>1</sub> generation	53
4.11	The dendrogram depicting relationship between parental lines and improved MR73 line (IMR73 or BC <sub>2</sub> F <sub>1</sub> ) rice genotypes ( <i>Oryza sativa</i> ) using 13 morphological traits	54
5.1	Schematic diagram for developing of improved lines	61
5.2	Screening of high zinc content improved homozygous lines applying linked markers	67
5.3	Graphical genotypes of the selected introgression lines in MR73 background developed in this study	69
5.4	Graphical genotype represents the lowest recovery plant among the selected lines	70
5.5	Graphical genotype represents the highest recovery plant among the selected lines	71
5.6	Morphometric characteristics of (a) plant height, (b) panicle length and (c) grain length and shape of parental lines and high grain zinc content improved lines	73

## LIST OF ABBREVIATIONS

bp	Base pair
CTAB	Cetyltrimethylammonium bromide
$\chi^2$	Chi-square
df	Degree of Freedom
DNA	Deoxyribo Nucleic Acid
EDTA	Ethylene Diamine Tetra Acetate Acid
GA	Expected Genetic Advance
GCV	Genotypic Coefficient of Variance
MABC	Marker Assisted Backcrossing
MAS	Marker Assisted Selection
MABB	marker-assisted backcross breeding
ABL	Advance breeding line
MS	Mean Square
MSE	Mean Square of Error
MSG	Mean Square of Genotypes
PCA	Principal component analysis
PCR	Polymerase Chain Reaction
PCV	Phenotypic Coefficient of Variance
PVP	Polyvinyl Pyrrolidone
QTL	Quantitative Trait Loci
SSR	Simple Sequence Repeat
RAPD	Randomly Polymorphic DNA
RCBD	Randomized Complete Block Design
RH	Relative Humidity
RFLP	Restriction Fragment Length Polymorphism
RNA	Ribonucleic Acid
TE	Tris/ EDTA
UV	Ultra Violet
ml	Milliliter

mm	Millimeter
Min	Minute
M	Molar
n	Nano
PCR	Polymerase chain reaction
%	Percentage
RPG	Recurrent parent genome
μ	Micro
RNA	Ribo Nucleic Acid
°C	Degree celsius
cm	Centimeter
cM	Centi Morgan
g	Gram
rpm	Rounds per minute
s	Second
V	Volt
v/v	Volume/volume
β-BE	B-mercaptoethanol
ddH <sub>2</sub> O	Double distilled water
mg	Milligram
kg	Killogram
ha	Hectare
GA	Genetic Advance
ZIP	Zrt-,Irt-like protein
SNP	Single-nucleotide polymorphism

## CHAPTER 1

### INTRODUCTION

#### 1.1 General introduction

More than last five decades, availability and growth of calorically enriched staple crops production were the main targets of agricultural research in developing countries. Rice development programmes have been conducted towards enhancing yield, resistance to biotic factors, tolerance to abiotic stress, and curtailing growth period for the last four decades. Amount of micronutrients present in the rice grain is one of the vital characteristics. We should investigate the prospect of development of high yielding rice varieties with high micronutrients content in order to combat micronutrients deficiency. Among the micronutrients deficiency, zinc (Zn) scarcity is one of the important micronutrients deficiency problem for humans.

More than half of the global humans consume rice as a staple food. Rice grain innately contain lower amount of micronutrients like Zn. Insufficiency of Zn is a significant issue not only for the health of world population but also nutrition. One third of the global people are under threat owing to poor nutritional eating of Zn (Myers et al., 2014). Most of the people who are under risk widely dependent on cereals; particularly rice (*Oryza sativa* L.). Malaysian people also consume rice as their staple food. Therefore, people of Malaysia could be potential victims of Zn deficiency in near future.

Weed causes significant problems including yield loss in rice crop. Although many controlling measures are adopted involving popular chemical control but it significantly affect environment (Kabir et al., 2010), human health and is responsible for the emergence of herbicide resistant weed variety (Heap, 2014). Therefore, development of weed resistant rice variety or use of rice allelopathy has been suggested. In this perspective, advantageous/desired trait development in the background of allelopathic rice variety could be one of the logical approaches for weed control combined with other controlling methods.

#### 1.2 Problem statement

Deficiency of Zn in diet is considered a significant public health and dietary problem in the current world. In the globe, half of the soils are fundamentally poor in terms of Zn. Inadequate of Zn in soils is considered as extensive nutritional crisis all over the rice producing nations. Scarcity of Zn in soil is generally related to high fixation of Zn,

poor phyto-availability (Rehman et al., 2012) and cultivation of high yielding rice varieties which eliminate huge amount of soil Zn at each reaps (De Steur et al., 2014).

Zinc deficiency in humans can lead to many unexpected consequences such as dwarfism, hypogonadism, diarrhoea, pneumonia and many more (Prasad et al., 1961; Gibson et al., 2008). Among the different approaches, Zn biofortification is highly suggested to combat Zn deficiency problem.

Many efforts have been taken more than last one decade to biofortify food crops particularly for Zn. Consequently, information is available with regards to genetic, molecular and physiological basis of high Zn accumulation, influence of environmental features and agronomic practices on Zn uptake, translocation and loading into rice grain. Generally, biofortification efforts have been carried out through agronomic, transgenic and breeding approach. Agronomic biofortification is a remedy for short time and legal systems of many countries do not support products from transgenic approach. Therefore, genetic Zn biofortification through breeding is highly recommended. Rice variety should be developed in such a way so that it can include high yielding characteristics with high Zn content in grain.

Conventional breeding for the development of high Zn rice cultivar has been going for the last few years in different countries. But the efforts became less effective due to complex genetic basis of grain Zn and environmental effects (Swamy et al., 2016). Moreover, it took long time through this method and the developed line had moderate yield potential as well as modest increase in Zn concentration. However, selection of correct breeding methods, crossing programmes, selection of individual plant and processes of crop field evaluation play crucial role for fruitful development of high Zn rice as suggested by Swamy et al. (2016). Conventional breeding method is a lingering process in which normally six to eight backcrosses required to recover complete recurrent parent genome. Moreover the method includes arduous evaluation of field performance which is some time unreliable due to genotype  $\times$  environment (G $\times$ E) interactions and expensive, plant breeders require wide experience etc. The application of molecular marker-aided selection (MAS) extensively removes these impediments. Molecular markers allow the perfect tool for developing the efficacy of selection for expected traits in new rice lines. These molecular markers provide vast scope for increasing the efficacy of conventional breeding by executing selection not straightforwardly on the desirable feature but the molecular markers related to that trait. Furthermore, application of molecular marker decreases the time required for the recovery of recurrent parent genome compared to conventional method. Thus, molecular marker-aided backcrossing (MAB) facilitates to select the desired plant and to reduce the overall time requirement for completing the breeding procedure.

If high Zn trait could be introgressed into Malaysian modern allelopathic rice variety MR73 keeping its other traits unaltered, a significant amount of Zn enriched rice could



be produced and simultaneously application of herbicide could be decreased for the betterment of environment.

The current study was designed to improve allelopathic MR73 rice variety into high Zn content rice through marker-aided backcrossing (MABC). The study consisted of foreground and background selection for MABC.

The improved lines required to be tested for their Zn content in grain and phenotypic adaptabilities. Moreover, grain yield and yield-contributing characteristics were also compared.

### **1.3 Objective of the study**

1. To analyse microsatellite (SSR) markers linked to high grain Zn QTLs in MR73 and BRRI dhan62 crosses.
2. To analyse the recipient parent genome reappearance in marker-aided backcross breeding programmes.
3. To pyramid high grain Zn QTLs in MR73-rice variety through marker-aided backcross breeding.

## REFERENCES

- Abdullah, A., Arshad, F., Radam, A., Ismail, M. and Mohamed, Z. (2010). Studies on the impacts of subsidized fertilizer scheme and subsidized paddy price scheme provided by the federal government. Ministry of Agriculture and Agro-based Industry Malaysia, Putrajaya, Malaysia.
- Abutalebi, S. and Fotokian, M.-H. (2015). Haplotype diversity of microsatellite markers linked to QTLs controlling zinc content in rice grains. *Journal of Biodiversity and Environmental Sciences*, 6(4): 445-453.
- Ahloowalia, B., Maluszynski, M. and Nichterlein, K. (2004). Global impact of mutation-derived varieties. *Euphytica*, 135(2): 187-204.
- Ahmed, H.U., Henry, A., Mauleon, R., Dixit, S., Vikram, P., Tilatto, R., Verulkar, S.B., Perraju, P., Mandal, N.P. and Variar, M. (2013). Genetic, physiological, and gene expression analyses reveal that multiple QTL enhance yield of rice mega-variety IR64 under drought. *PLoS One*, 8(5): e62795.
- Akinbile, C., El-Latif, K.A., Abdullah, R. and Yusoff, M. (2011). Rice production and water use efficiency for self-sufficiency in Malaysia: a review. *Trends in Applied Sciences Research*, 6(10): 1127.
- Aliyu, R., Adamu, A., Muazu, S., Alonge, S. and Gregorio, G. (2011). Tagging and validation of SSR markers to salinity tolerance QTLs in rice (*Oryza spp*). *2010 International Conference on Biology, Environment and Chemistry*, IPCBEE, IACSIT press, Singapore.
- Allard, R.W. (1999). *Principles of Plant Breeding*, John Wiley & Sons.
- Allard, W. (1960). *Principles of Plant Breeding (John Willey and Sons. Inc. London)*.
- Ampong-Nyarko, K. and De Datta, S.K. (1991). *A handbook for weed control in rice*, International Rice Research Institute.
- Andreini, C., Banci, L., Bertini, I. and Rosato, A. (2006). Zinc through the three domains of life. *Journal of Proteome Research*, 5(11): 3173-3178.
- Antralina, M., Istina, I.N. and Simarmata, T. (2015). Effect of difference weeds control methods to yield of lowland rice in the SOBARI. *Procedia Food Science*, 3:323-329.
- Anuradha, K., Agarwal, S., Rao, Y.V., Rao, K., Viraktamath, B. and Sarla, N. (2012). Mapping QTLs and candidate genes for iron and zinc concentrations in unpolished rice of Madhukar× Swarna RILs. *Gene*, 508(2): 233-240.

- Ashkani, S., Rafii, M., Sariah, M., Siti, N.A.A., Rusli, I., Harun, A. and Latif, M. (2011). Analysis of simple sequence repeat markers linked with blast disease resistance genes in a segregating population of rice (*Oryza sativa*). *Genetics and Molecular Research*, 10(3): 1345-1355.
- Ashraf, M. (2009). Biotechnological approach of improving plant salt tolerance using antioxidants as markers. *Biotechnology Advances*, 27(1): 84-93.
- Ashraf, M., Akram, N. and Foolad, M. (2012). Marker-assisted selection in plant breeding for salinity tolerance. *Plant Salt Tolerance: Methods and Protocols*, 305-333.
- Ashraf, M. and Akram, N.A. (2009). Improving salinity tolerance of plants through conventional breeding and genetic engineering: an analytical comparison. *Biotechnology Advances*, 27(6): 744-752.
- Ashraf, M. and Foolad, M.R. (2013). Crop breeding for salt tolerance in the era of molecular markers and marker-assisted selection. *Plant Breeding*, 132(1): 10-20.
- Azlan, S., Alias, I., Saad, A. and Habibuddin, H. (2003). Performance of potential mutant lines of MR 180. *Modern rice farming. Proceedings of the international rice conference* Serdang, Malaysia: MARDI.
- Azmi, M. (1992). Competitive ability of barnyard grass in direct-seeded rice. *Teknologi Padi*, 8:19-25.
- Azmi, M. (1995). Weed succession from transplanting to direct-seeding method in Kemubu rice area, Malaysia. *Journal of Biosciences*, 6:143-154.
- Azmi, M. (1998). Biological and ecology of *Echinochloa crus-galli* (L.) Beauv. in direct seeded rice (*Oryza sativa* L.). *Proceeding of rice integrated pest management conference*.
- Babu, N.N., Krishnan, S.G., Vinod, K., Krishnamurthy, S., Singh, V.K., Singh, M.P., Singh, R., Ellur, R.K., Rai, V. and Bollinedi, H. (2017). Marker aided incorporation of Saltol, a major QTL associated with seedling stage salt tolerance, into *Oryza sativa* 'Pusa Basmati 1121'. *Frontiers in Plant Science*, 8:41.
- Bajwa, A.A., Jabran, K., Shahid, M., Ali, H.H. and Chauhan, B.S. (2015). Eco-biology and management of *Echinochloa crus-galli*. *Crop Protection*, 75:151-162.
- Banik, M. (1999). Cold injury problems in Boro rice. Workshop on modern rice cultivation in Bangladesh. Bangladesh Rice Research Institute Joydebpur, Gazipur, Bangladesh.
- Basavaraj, S., Singh, V.K., Singh, A., Singh, A., Singh, A., Anand, D., Yadav, S., Ellur, R.K., Singh, D. and Krishnan, S.G. (2010). Marker-assisted improvement

- of bacterial blight resistance in parental lines of Pusa RH10, a superfine grain aromatic rice hybrid. *Molecular Breeding*, 26(2): 293-305.
- Bashir, K., Ishimaru, Y. and Nishizawa, N.K. (2012). Molecular mechanisms of zinc uptake and translocation in rice. *Plant and Soil*, 361(1-2): 189-201.
- Beckie, H.J. (2006). Herbicide-Resistant Weeds: Management Tactics and Practices1. *Weed Technology*, 20(3): 793-814.
- Begum, M., Juraimi, A., Azmi, M., Rajan, A. and Omar, S.S. (2005). Weed vegetation of direct seeded rice fields in muda rice granary areas of Peninsular Malaysia. *Pakistan Journal of Biological Sciences*, 8(4): 537-541.
- Bekele, B.D., Naveen, G., Rakhi, S. and Shashidhar, H. (2013). Genetic evaluation of recombinant inbred lines of rice (*Oryza sativa* L.) for grain zinc concentrations, yield related traits and identification of associated SSR markers. *Pakistan Journal of Biological Sciences*, 16(23): 1714-1721.
- Bhatia, D., Sharma, R., Vikal, Y., Mangat, G., Mahajan, R., Sharma, N., Lore, J.S., Singh, N., Bharaj, T.S. and Singh, K. (2011). Marker-assisted development of bacterial blight resistant, dwarf, and high yielding versions of two traditional basmati rice cultivars. *Crop Science*, 51(2): 759-770.
- Biradar, H., Bhargavi, M., Sasalwad, R., Parama, R. and Hittalmani, S. (2007). Identification of QTL associated with silicon and zinc content in rice (*Oryza sativa* L.) and their role in blast disease resistance. *The Indian Journal of Genetics and Plant Breeding*, 67(2): 105-109.
- Black, R.E., Allen, L.H., Bhutta, Z.A., Caulfield, L.E., De Onis, M., Ezzati, M., Mathers, C., Rivera, J., Maternal and Group, C.U.S. (2008). Maternal and child undernutrition: global and regional exposures and health consequences. *The Lancet*, 371(9608): 243-260.
- Bonman, J. (1992). Durable resistance to rice blast disease—environmental influences. *Breeding for Disease Resistance*, Springer: 115-123.
- Bonnett, D., Rebetzke, G. and Spielmeyer, W. (2005). Strategies for efficient implementation of molecular markers in wheat breeding. *Molecular Breeding*, 15(1): 75-85.
- Bormans, C.A. (2002). Development of genetic markers for blast resistance genes in rice (*Oryza sativa* L.) for use in marker assisted selection.
- Bouis, H. (2014). Biofortification Progress Briefs August 2014. Washington DC.: Harvest Plus; 2014 August 2014. 82 p.

- Bouis, H.E. (2003). Micronutrient fortification of plants through plant breeding: can it improve nutrition in man at low cost? *Proceedings of the Nutrition Society*, 62(2): 403-411.
- Brara, B., Jaina, R.K. and Jain, S. (2015). Correlation of molecular marker allele size with physio-morphological and micronutrient (Zn, Fe) traits among rice genotypes. *International Journal of Current Science*, 15:42-50.
- BRKB. (2017). Bangladesh Rice Knowledge Bank. [www.knowledgebank-brri.org](http://www.knowledgebank-brri.org) (accessed on 12-05-2017).
- Brown, K.H., Hambidge, K.M. and Ranum, P. (2010). Zinc fortification of cereal flours: current recommendations and research needs. *Food and Nutrition Bulletin*, 31(1\_suppl1): S62-S74.
- Brown, K.H., Rivera, J.A., Bhutta, Z., Gibson, R.S., King, J.C., Lönnerdal, B., Ruel, M.T., Sandtröm, B., Wasantwisut, E. and Hotz, C. (2004). International Zinc Nutrition Consultative Group (IZiNCG) technical document# 1. Assessment of the risk of zinc deficiency in populations and options for its control. *Food and Nutrition Bulletin*, 25(1 Suppl 2).
- BRRI. (2015). Modern Rice Cultivation, 18th Edition.
- Cakmak, I. (2008). Enrichment of cereal grains with zinc: agronomic or genetic biofortification? *Plant and Soil*, 302(1-2):1-17.
- Chandel, G., Banerjee, S., See, S., Meena, R., Sharma, D. and Verulkar, S. (2010). Effects of different nitrogen fertilizer levels and native soil properties on rice grain Fe, Zn and protein contents. *Rice Science*, 17(3): 213-227.
- Chandel, G., Samuel, P., Dubey, M. and Meena, R. (2011). In silico expression analysis of QTL specific candidate genes for grain micronutrient (Fe/Zn) content using ESTs and MPSS signature analysis in rice (*Oryza sativa* L.). *Journal of Plant Genetics and Transgenics*, 2(1): pp 11-22.
- Chauhan, B.S. (2012). *Weed management in direct-seeded rice systems*, International Rice Research Institute.
- Chen, H., Lin, Y.a. and Zhang, Q. (2010). Rice. *Biotechnology in Agriculture and Forestry*, 64:423.
- Cherati, F.E., Bahrami, H. and Asakereh, A. (2011). Evaluation of traditional, mechanical and chemical weed control methods in rice fields. *Australian Journal of Crop Science*, 5(8): 1007.
- Collard, B., C.Y. and Mackill, D.J. (2007). Marker-assisted selection: An approach for precision plant breeding in the twenty-first century. *Philosophical Transactions of the Royal Society of London B: Biological Sciences* 17:1-16.

- Collard, B.C. and Mackill, D.J. (2008). Marker-assisted selection: an approach for precision plant breeding in the twenty-first century. *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, 363(1491): 557-572.
- Collard, B.C., Vera Cruz, C.M., McNally, K.L., Virk, P.S. and Mackill, D.J. (2008). Rice molecular breeding laboratories in the genomics era: current status and future considerations. *International Journal of Plant Genomics*.
- Cuc, L.M., Huyen, L.T., Hien, P.T., Hang, V.T., Dam, N.Q., Mui, P.T., Quang, V.D., Ismail, A.M. and Ham, L.H. (2012). Application of marker assisted backcrossing to introgress the submergence tolerance QTL *SUB1* into the Vietnam elite rice variety-AS996. *American Journal of Plant Sciences*, 3(04): 528.
- Dalrymple, D.G. (1986). *Development and spread of high-yielding rice varieties in developing countries*, International Rice Research Institute.
- De Steur, H., Mogendi, J.B., Blancquaert, D., Lambert, W., Van Der Straeten, D. and Gellynck, X. (2014). Genetically Modified Rice with Health Benefits as a Means to Reduce Micronutrient Malnutrition: Global Status, Consumer Preferences, and Potential Health Impacts of Rice Biofortification. *Wheat and Rice in Disease Prevention and Health*, Elsevier: 283-299.
- Decoux, G. and Hospital, F. (2002). Popmin: a program for the numerical optimization of population sizes in marker-assisted backcross programs. *Journal of Heredity*, 93(5): 383-384.
- Divya, B., Robin, S., Rabindran, R., Senthil, S., Raveendran, M. and Joel, A.J. (2014). Marker assisted backcross breeding approach to improve blast resistance in Indian rice (*Oryza sativa*) variety ADT43. *Euphytica*, 200(1): 61-77.
- Doyle, J.J. and Doyle, J.L. (1990). Isolation of plant DNA from fresh tissue. *Focus*, 12:13-15.
- Duke, J.A. (1983). Handbook of energy crops.
- Dutta, R., Baset, M. and Khanam, S. (2002). Plant architecture and growth characteristics of fine grain and aromatic rices and their relation with grain yield. *International Rice Commission Newsletter (FAO) Bulletin de la Commission Internationale du Riz (FAO) Noticiario de la Comision Internacional del Arroz (FAO)*.
- Dwivedi, S.L., Crouch, J.H., Mackill, D.J., Xu, Y., Blair, M.W., Ragot, M., Upadhyaya, H.D. and Ortiz, R. (2007). The molecularization of public sector crop breeding: progress, problems, and prospects. *Advances in Agronomy*, 95:163-318.
- Fazaa, M., Anis, G., Mazal, T.M., Sabagh, A.E., Barutçular, C. and Islam, M. (2016a). Genetic Identification of Quantitative Trait Loci (QTL) for High Nutritional



- Value Traits in DHLs derived from Black Rice. *Journal of Agriculture Biotechnology*, 1(02).
- Fazaa, M., Sabagh, A.E., Anis, G., EL-Rewainy, I., Barutçular, C., Yildirim, M. and Islam, M. (2016b). Grain Quality of Doubled Haploid Lines in Rice (*Oryza sativa* L.) Produced by Anther Culture. *Journal of Agricultural Science*, 8(5): 184.
- Frisch, M. (2004). Breeding strategies: optimum design of marker-assisted backcross programs. *Molecular Marker Systems in Plant Breeding and Crop Improvement*, Springer: 319-334.
- Frisch, M., Bohn, M. and Melchinger, A.A. (1999a). Minimum sample size and optimal positioning of flanking markers in marker-assisted backcrossing for transfer of a target gene. *Crop Science*, 39(4): 967-975.
- Frisch, M., Bohn, M. and Melchinger, A.E. (1999b). Comparison of selection strategies for marker-assisted backcrossing of a gene. *Crop Science*, 39(5): 1295-1301.
- Frisch, M. and Melchinger, A.E. (2001). Marker-assisted backcrossing for simultaneous introgression of two genes. *Crop Science*, 41(6): 1716-1725.
- Frisch, M. and Melchinger, A.E. (2005). Selection theory for marker-assisted backcrossing. *Genetics*, 170(2): 909-917.
- Fu, C., Wu, T., Liu, W., Wang, F., Li, J., Zhu, X., Huang, H., Liu, Z.R., Liao, Y. and Zhu, M. (2012). Genetic improvement of resistance to blast and bacterial blight of the elite maintainer line Rongfeng B in hybrid rice (*Oryza sativa* L.) by using marker-assisted selection. *African Journal of Biotechnology*, 11(67): 13104-13114.
- Fujimaki, H. (1979). Recurrent selection by using genetic male sterility for rice improvement. *Japan Agricultural Research Quarterly*.
- Gande, N.K., Kundur, P.J., Soman, R., Ambati, R., Ashwathanarayana, R., Bekele, B.D. and Shashidhar, H. (2014). Identification of putative candidate gene markers for grain zinc content using recombinant inbred lines (RIL) population of IRRI38 X Jeerigesanna. *African Journal of Biotechnology*, 13(5).
- Gao, L.-z., Zhang, C.-H., Chang, L.-p., Jia, J.-z., Qiu, Z.-e. and Dong, Y.-s. (2005). Microsatellite diversity within *Oryza sativa* with emphasis on *indica-japonica* divergence. *Genetics Research*, 85(1): 1-14.
- García-Bañuelos, M.L., Sida-Arreola, J.P. and Sánchez, E. (2014). Biofortification-promising approach to increasing the content of iron and zinc in staple food crops. *Journal of Elementology*, 19(3).

- Garcia-Oliveira, A.L., Tan, L., Fu, Y. and Sun, C. (2009). Genetic identification of quantitative trait loci for contents of mineral nutrients in rice grain. *Journal of Integrative Plant Biology*, 51(1): 84-92.
- Garris, A.J., Tai, T.H., Coburn, J., Kresovich, S. and McCouch, S. (2005). Genetic structure and diversity in *Oryza sativa* L. *Genetics*, 169(3): 1631-1638.
- Ghislain, M., Spooner, D., Rodriguez, F., Villamón, F., Nunez, J., Vásquez, C., Waugh, R. and Bonierbale, M. (2004). Selection of highly informative and user-friendly microsatellites (SSRs) for genotyping of cultivated potato. *Theoretical and Applied Genetics*, 108(5): 881-890.
- Giannopolitis, C. and Vassiliou, G. (1989). Propanil tolerance in *Echinochloa crus-galli* (L.) Beauv. *International Journal of Pest Management*, 35(1): 6-7.
- Gibson, R. (2005). Dietary strategies to enhance micronutrient adequacy: experiences in developing countries. *Micronutrients in South and Southeast Asia*. International Centre for Integrated Mountain Development, Kathmandu, 3-7.
- Gibson, R.S. (2006). Zinc: the missing link in combating micronutrient malnutrition in developing countries. *Proceedings of the Nutrition Society*, 65(1): 51-60.
- Gibson, R.S., Hess, S.Y., Hotz, C. and Brown, K.H. (2008). Indicators of zinc status at the population level: a review of the evidence. *British Journal of Nutrition*, 99(S3): S14-S23.
- Gopalakrishnan, S., Sharma, R., Anand Rajkumar, K., Joseph, M., Singh, V., Singh, A., Bhat, K., Singh, N. and Mohapatra, T. (2008). Integrating marker assisted background analysis with foreground selection for identification of superior bacterial blight resistant recombinants in Basmati rice. *Plant Breeding*, 127(2): 131-139.
- Graham, R.D., Knez, M. and Welch, R.M. (2012). 1 How Much Nutritional Iron Deficiency in Humans Globally Is due to an Underlying Zinc Deficiency? *Advances in Agronomy*, 115(1).
- Graham, R.D., Welch, R.M., Saunders, D.A., Ortiz-Monasterio, I., Bouis, H.E., Bonierbale, M., De Haan, S., Burgos, G., Thiele, G. and Liria, R. (2007). Nutritious subsistence food systems. *Advances in Agronomy*, 92:1-74.
- Gregorio, G.B. (2002). Progress in breeding for trace minerals in staple crops. *The Journal of Nutrition*, 132(3): 500S-502S.
- Gregorio, G.B., Aliyu, R., Adamu, A., Muazu, S. and Alonge, S. (2011). Tagging and validation of SSR markers to salinity tolerance QTLs in rice (*Oryza* spp). 2010 *International Conference on Biology, Environment and Chemistry*, IPCBEE, IACSIT press, Singapore.



- Gu, Y.Q., Coleman-Derr, D., Kong, X. and Anderson, O.D. (2004). Rapid genome evolution revealed by comparative sequence analysis of orthologous regions from four *Triticeae* genomes. *Plant Physiology*, 135(1): 459-470.
- Guimaraes, E.P.a. and Correa-Victoria, F. (2000). Use of recurrent selection for develops resistance *Pyricularia grisea* Sacc. On rice, In Guimaraes EP (ed) *Advances in rice population improvement* (pp. 165-175). Embrapa Rice and Beans, Santo Antonio de Goias.
- Guo, L.-b. and Ye, G.-y. (2014). Use of major quantitative trait loci to improve grain yield of rice. *Rice Science*, 21(2): 65-82.
- Guo, S.-b., Yu, W., LI, X.-q., LIU, K.-q., HUANG, F.-k., CHEN, C.-h. and GAO, G.-q. (2013). Development and identification of introgression lines from cross of *Oryza sativa* and *Oryza minuta*. *Rice Science*, 20(2): 95-102.
- Hacisalihoglu, G. and Kochian, L.V. (2003). How do some plants tolerate low levels of soil zinc? Mechanisms of zinc efficiency in crop plants. *New Phytologist*, 159(2): 341-350.
- Hadzim, K., Ajimilah, N., Othman, O., Arasu, N., Latifah, A. and Saad, A. (1988). Mutant Mahsuri: Baka untuk beras bermutu. *Teknologi Padi*, 4:7-13.
- Haider, B.A. and Bhutta, Z.A. (2009). The effect of therapeutic zinc supplementation among young children with selected infections: a review of the evidence. *Food and Nutrition Bulletin*, 30(1\_suppl1): S41-S59.
- Hari, Y., Srinivasarao, K., Viraktamath, B.C., Prasad, H., Arremsetty, S., Laha, G.S., I Ahmed, M., Natarajkumar, P., Sujatha, K. and Srinivas Prasad, M. (2013). Marker-assisted introgression of bacterial blight and blast resistance into IR 58025B, an elite maintainer line of rice. *Plant Breeding*, 132(6): 586-594.
- Hayashi, K., Yoshida, H. and Ashikawa, I. (2006). Development of PCR-based allele-specific and InDel marker sets for nine rice blast resistance genes. *Theoretical and Applied Genetics*, 113(2): 251-260.
- Heap, I. (2014). Herbicide resistant weeds. *Integrated Pest Management*, Springer: 281-301.
- Hess, S.Y. and King, J.C. (2009). Effects of maternal zinc supplementation on pregnancy and lactation outcomes. *Food and Nutrition Bulletin*, 30(1\_suppl1): S60-S78.
- Hien, N.L., Sarhadi, W.A., Oikawa, Y. and Hirata, Y. (2007). Genetic diversity of morphological responses and the relationships among Asia aromatic rice (*Oryza sativa* L.) cultivars. *Tropics*, 16(4): 343-355.

- Hirabayashi, H., Sato, H., Nonoue, Y., Kuno-Takemoto, Y., Takeuchi, Y., Kato, H., Nemoto, H., Ogawa, T., Yano, M. and Imbe, T. (2010). Development of introgression lines derived from *Oryza rufipogon* and *O. glumaepatula* in the genetic background of japonica cultivated rice (*O. sativa* L.) and evaluation of resistance to rice blast. *Breeding Science*, 60(5): 604-612.
- Hittalmani, S., Parco, A., Mew, T., Zeigler, R. and Huang, N. (2000). Fine mapping and DNA marker-assisted pyramiding of the three major genes for blast resistance in rice. *Theoretical and Applied Genetics*, 100(7): 1121-1128.
- Holm, L.G. (1977). *The World's Worst Weeds: Distribution and Biology*, East-West Center.
- Hospital, F. (2005). Selection in backcross programmes. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 360(1459): 1503.
- Hospital, F., Chevalet, C.a. and Mulsant, P. (1992). Using markers in gene introgression breeding programs. *Genetics*, 132:1119-1120.
- Hospital, F.a. and Charcosset, A. (1997). Marker assisted introgression of quantitative trait loci. *Genetics*, 147:1469-1485.
- Hossain, M., Islam, M. and Hasanuzzaman, M. (2008). Influence of different nitrogen levels on the performance of four aromatic rice varieties. *International Journal of Agriculture and Biology*, 10(6): 693-696.
- Hotz, C. and Brown, K. (2004). *Assessment of the risk of deficiency in populations and options for its control*, International nutrition foundation: for UNU.
- Huyen, L.T., Cuc, L.M., Ismail, A.M. and Ham, L.H. (2012). Introgression the salinity tolerance QTLs Saltol into AS996, the elite rice variety of Vietnam. *American Journal of Plant Sciences*, 3(07): 981.
- Iftekharuddaula, K. (2009). Comparison of new selection strategies of marker-assisted backcrossing for a submergence tolerant gene in rice, Ph. D. dissertation.
- Iftekharuddaula, K., Newaz, M., Salam, M., Ahmed, H., Mahbub, M., Septiningsih, E., Collard, B., Sanchez, D., Pamplona, A. and Mackill, D. (2011). Rapid and high-precision marker assisted backcrossing to introgress the *SUB1* QTL into BR11, the rainfed lowland rice mega variety of Bangladesh. *Euphytica*, 178(1): 83-97.
- Iftekharuddaula, K.M., Salam, M.A., Newaz, M.A., Ahmed, H.U., Collard, B.C., Septiningsih, E.M., Sanchez, D.L., Pamplona, A.M. and Mackill, D.J. (2012). Comparison of phenotypic versus marker-assisted background selection for the *SUB1* QTL during backcrossing in rice. *Breeding Science*, 62(3): 216-222.
- Impa, S.M., Morete, M.J., Ismail, A.M., Schulin, R. and Johnson-Beebout, S.E. (2013a). Zn uptake, translocation and grain Zn loading in rice (*Oryza sativa* L.)

- genotypes selected for Zn deficiency tolerance and high grain Zn. *Journal of Experimental Botany*, 64(10): 2739-2751.
- Ishikawa, S., Abe, T., Kuramata, M., Yamaguchi, M., Ando, T., Yamamoto, T. and Yano, M. (2009). A major quantitative trait locus for increasing cadmium-specific concentration in rice grain is located on the short arm of chromosome 7. *Journal of Experimental Botany*, 61(3): 923-934.
- Ishimaru, Y., Bashir, K. and Nishizawa, N.K. (2011). Zn uptake and translocation in rice plants. *Rice*, 4(1): 21-27.
- Ishimaru, Y., Masuda, H., Suzuki, M., Bashir, K., Takahashi, M., Nakanishi, H., Mori, S. and Nishizawa, N.K. (2007). Overexpression of the *OsZIP4* zinc transporter confers disarrangement of zinc distribution in rice plants. *Journal of Experimental Botany*, 58(11): 2909-2915.
- Jan Stomph, T., Jiang, W. and Struik, P.C. (2009). Zinc biofortification of cereals: rice differs from wheat and barley. *Trends in Plant Science*, 14(3): 123-124.
- Jena, K. and Mackill, D. (2008). Molecular markers and their use in marker-assisted selection in rice. *Crop Science*, 48(4): 1266-1276.
- Jia, Y. (2009). Artificial introgression of a large chromosome fragment around the rice blast resistance gene *Pi-ta* in backcross progeny and several elite rice cultivars. *Heredity*, 103(4): 333.
- Jiang, G., Xu, C., Tu, J., Li, X., He, Y. and Zhang, Q. (2004). Pyramiding of insect- and disease-resistance genes into an elite *indica*, cytoplasm male sterile restorer line of rice, 'Minghui 63'. *Plant Breeding*, 123(2): 112-116.
- Jiang, S., Wu, J., Feng, Y., Yang, X. and Shi, C. (2007). Correlation analysis of mineral element contents and quality traits in milled rice (*Oryza sativa* L.). *Journal of Agricultural and Food Chemistry*, 55(23): 9608-9613.
- Jiang, W., Struik, P., Van Keulen, H., Zhao, M., Jin, L. and Stomph, T. (2008). Does increased zinc uptake enhance grain zinc mass concentration in rice? *Annals of Applied Biology*, 153(1): 135-147.
- Johnson, A.A., Kyriacou, B., Callahan, D.L., Carruthers, L., Stangoulis, J., Lombi, E. and Tester, M. (2011). Constitutive overexpression of the *OsNAS* gene family reveals single-gene strategies for effective iron- and zinc-biofortification of rice endosperm. *PLoS One*, 6(9): e24476.
- Joseph, M., Gopalakrishnan, S., Sharma, R., Singh, V., Singh, A., Singh, N. and Mohapatra, T. (2004). Combining bacterial blight resistance and Basmati quality characteristics by phenotypic and molecular marker-assisted selection in rice. *Molecular Breeding*, 13(4): 377-387.

- Kabir, A., Karim, S., Begum, M. and Juraimi, A.S. (2010). Allelopathic potential of rice varieties against spinach (*Spinacia oleracea*). *International Journal of Agriculture and Biology*, 12(6): 809-815.
- Khanh, T.D., Linh, T.H. and Xuan, T.D. (2013). Rapid and high-precision marker assisted backcrossing to introgress the *SUB1* QTL into the Vietnamese elite rice variety. *Journal of Plant Breeding and Crop Science*, 5(2): 26-33.
- Khush, G., Mackill, D. and Sidhu, G. (1989). Breeding Rice for Resistance to Bacterial Blight. *Bacterial Blight of Rice*, 207-217.
- Kiyosawa, S. (1982). Gene analysis for blast resistance. *Oryza*, 18:196-203.
- Koizumi, S. (2007). Durability of resistance to rice blast disease. *JIRCAS Working Report*, 53:1-10.
- Kong, C.H., Chen, X.H., Hu, F. and Zhang, S.Z. (2011). Breeding of commercially acceptable allelopathic rice cultivars in China. *Pest Management Science*, 67(9): 1100-1106.
- Korinsak, S., Sirithunya, P., Meakwatanakarn, P., Sarkarung, S., Vanavichit, A. and Toojinda, T. (2011). Changing allele frequencies associated with specific resistance genes to leaf blast in backcross introgression lines of Khao Dawk Mali 105 developed from a conventional selection program. *Field Crops Research*, 122(1): 32-39.
- Kumar, M., Mishra, G.P., Singh, R., Kumar, J., Naik, P.K. and Singh, S.B. (2009). Correspondence of ISSR and RAPD markers for comparative analysis of genetic diversity among different apricot genotypes from cold arid deserts of trans-Himalayas. *Physiology and Molecular Biology of Plants*, 15(3): 225-236.
- Lee, S., Persson, D.P., Hansen, T.H., Husted, S., Schjoerring, J.K., Kim, Y.S., Jeon, U.S., Kim, Y.K., Kakei, Y. and Masuda, H. (2011). Bio-available zinc in rice seeds is increased by activation tagging of nicotianamine synthase. *Plant Biotechnology Journal*, 9(8): 865-873.
- Leeson, J.Y. (2005). Prairie weed surveys of cereal, oilseed and pulse crops from the 1970s to the 2000s.
- Lewis, R.S. and Kernodle, S. (2009). A method for accelerated trait conversion in plant breeding. *Theoretical and Applied Genetics*, 118(8): 1499-1508.
- Linh, L.H., Linh, T.H., Xuan, T.D., Ham, L.H., Ismail, A.M. and Khanh, T.D. (2012). Molecular breeding to improve salt tolerance of rice (*Oryza sativa* L.) in the Red River Delta of Vietnam. *International Journal of Plant Genomics*.

- Litt, M. and Luty, J.A. (1989). A hypervariable microsatellite revealed by in vitro amplification of a dinucleotide repeat within the cardiac muscle act in gene. *American Journal of Human Genetics*, 44(3): 397.
- Liu, W.-G., Jin, S.-J., Zhu, X.-Y., Feng, W., Li, J.-H., Liu, Z.-R., Liao, Y.-L., Zhu, M.-S., Huang, H.-J. and Liu, Y.-B. (2008). Improving blast resistance of a thermo-sensitive genic male sterile rice line GD-8S by molecular marker-assisted selection. *Rice Science*, 15(3): 179-185.
- Lopez-Gerena, J. (2006). Mapping QTL controlling durable resistance to rice blast in the cultivar *Oryzica Llanos 5*, Kansas State University.
- Lu, K., Li, L., Zheng, X., Zhang, Z., Mou, T. and Hu, Z. (2008). Quantitative trait loci controlling Cu, Ca, Zn, Mn and Fe content in rice grains. *Journal of Genetics*, 87(3): 305-310.
- Mackill, D. and Ni, J. (2001). Molecular mapping and marker-assisted selection for major-gene traits in rice. *Rice Genetics IV*, 140-141.
- Mackill, D.J. (2007). Molecular markers and marker-assisted selection in rice. *Genomics Assisted Crop Improvement*, Springer: 147-168.
- Mahender, A., Anandan, A., Pradhan, S.K. and Pandit, E. (2016). Rice grain nutritional traits and their enhancement using relevant genes and QTLs through advanced approaches. *Springerplus*, 5(1): 2086.
- Man, A. and Mortimer, M. (2002). Weed species shifts in response to serial herbicide application in wet-seeded rice in Malaysia. Direct Seeding: Research Strategies and Opportunities, International Rice Research Institute, Los Banos, Philippines, 357-367.
- Marambe, B. and Amarasinghe, L. (2002). Propanil-resistant barnyardgrass [*Echinochloa crus-galli* (L.) Beauv.] in Sri Lanka: Seedling growth under different temperatures and control. *Weed Biology and Management*, 2(4): 194-199.
- MARDI. (2008). *Three Decades of Achievements in Research and Development*.
- Matsuo, T., Futsuhara, Y., Kikuchi, F.a. and Yamaguchi, H. (1997). *Science of the rice plant*. In: Matsuo T, Futsuhara Y, Kikuchi F, Yamaguchi H (eds) *Food and Agriculture Policy Research Center*, Tokyo.
- Matthews, R.B. (1995). *Modeling the impact of climate change on rice production in Asia*, International Rice Research Institute.
- Mazid, M., Rafii, M., Hanafi, M., Rahim, H., Shabanimofrad, M. and Latif, M. (2013). Agro morphological characterization and assessment of variability, heritability,



- genetic advance and divergence in bacterial blight resistant rice genotypes. *South African Journal of Botany*, 86(15-22).
- McCouch, S.R., Sweeney, M., Li, J., Jiang, H., Thomson, M., Septiningsih, E., Edwards, J., Moncada, P., Xiao, J. and Garriss, A. (2007). Through the genetic bottleneck: *O. rufipogon* as a source of trait-enhancing alleles for *O. sativa*. *Euphytica*, 154(3): 317-339.
- McCouch, S.R., Teytelman, L., Xu, Y., Lobos, K.B., Clare, K., Walton, M., Fu, B., Maghirang, R., Li, Z. and Xing, Y. (2002). Development and mapping of 2240 new SSR markers for rice (*Oryza sativa* L.). *DNA Research*, 9(6): 199-207.
- Miah, G. (2015). Development of Blast-resistant rice variety derived from crossing between MR219 and Pongsu Seribu 1 through marker-assisted selection. Doctoral dissertation. Universiti Putra Malaysia, Malaysia, 43400.
- Miah, G., Rafii, M., Ismail, M., Puteh, A., Rahim, H., Asfaliza, R. and Latif, M. (2013). Blast resistance in rice: a review of conventional breeding to molecular approaches. *Molecular Biology Reports*, 40(3): 2369-2388.
- Mohamad, O., Abdullah, M.Z.a. and Rahman, M.A. (2005). Molecular markers as a breeding tool in rice breeding: new paradigms and opportunities. In: *Proceedings of the 6th National Congress on Genetics*, Genetics Society of Malaysia, Kuala Lumpur, Malaysia.
- Mohamad, O., Mohd, N., Abdul, R., Alias, I., Azlan, S., Othman, O., Hadzim, K., Saad, A., Abdullah, M. and Habibuddin, H. (2006). Development of improved rice varieties through the use of induced mutations in Malaysia.
- Mohler, V. and Singrün, C. (2004). General considerations: marker-assisted selection. *Molecular Marker Systems in Plant Breeding and Crop Improvement*, Springer: 305-317.
- Moldenhauer, K. and Slaton, N. (2001). Rice growth and development. *Rice Production Handbook*, 7-14.
- Mondal, U., Khanom, M., Hassan, L. and Begum, S. (2014). Foreground selection through SSRs markers for the development of salt tolerant rice variety. *Journal of the Bangladesh Agricultural University*, 11(1): 67-72.
- Morgante, M., Hanafey, M. and Powell, W. (2002). Microsatellites are preferentially associated with non repetitive DNA in plant genomes. *Nature Genetics*, 30(2): 194.
- Myers, S.S., Zanobetti, A., Kloog, I., Huybers, P., Leakey, A.D., Bloom, A.J., Carlisle, E., Dietterich, L.H., Fitzgerald, G. and Hasegawa, T. (2014). Increasing CO<sub>2</sub> threatens human nutrition. *Nature*, 510(7503): 139-142.

- Nakandalage, N., Nicolas, M., Norton, R.M., Hirotsu, N., Milham, P.J. and Seneweera, S. (2016). Improving rice zinc biofortification success rates through genetic and crop management approaches in a changing environment. *Frontiers in Plant Science*, 7.
- Namanya, P. (2011). *Towards the biofortification of banana fruit for enhanced micronutrient content*, Queensland University of Technology.
- Narayanan, N., Baisakh, N., Cruz, V., Gnanamanickam, S., Datta, K. and Datta, S. (2002). Molecular breeding for the development of blast and bacterial blight resistance in rice cv. IR50. *Crop Science*, 42(6): 2072-2079.
- Navarez, D. and Olofsdotter, M. (1996). Relay seeding technique for screening allelopathic rice (*Oryza sativa*). 2. *International Weed Control Congress*, Copenhagen (Denmark), 25-28 Jun 1996, SP.
- Neeraja, C., Maghirang-Rodriguez, R., Pamplona, A., Heuer, S., Collard, B., Septiningsih, E., Vergara, G., Sanchez, D., Xu, K. and Ismail, A. (2007). A marker-assisted backcross approach for developing submergence-tolerant rice cultivars. *Theoretical and Applied Genetics*, 115(6): 767-776.
- Nordin, M.S., Ariffin, Z., Jajuli, R., Abdullah, W.D.W. and M.G., a.D. (2007). Country report on The State of Plant Genetic Resources For Food and Agriculture in Malaysia (1997-2007), 19.
- Norton, G.J., Deacon, C.M., Xiong, L., Huang, S., Meharg, A.A. and Price, A.H. (2010). Genetic mapping of the rice ionome in leaves and grain: identification of QTLs for 17 elements including arsenic, cadmium, iron and selenium. *Plant and Soil*, 329(1-2): 139-153.
- Oliveira, E.J., Pádua, J.G., Zucchi, M.I., Vencovsky, R. and Vieira, M.L.C. (2006). Origin, evolution and genome distribution of microsatellites. *Genetics and Molecular Biology*, 29(2): 294-307.
- Olsen, L.I. and Palmgren, M.G. (2014). Many rivers to cross: the journey of zinc from soil to seed. *Frontiers in Plant Science*, 5.
- Olufowote, J.O., Xu, Y., Chen, X., Goto, M., McCouch, S.R., Park, W.D., Beachell, H.M. and Dilday, R.H. (1997). Comparative evaluation of within-cultivar variation of rice (*Oryza sativa* L.) using microsatellite and RFLP markers. *Genome*, 40(3): 370-378.
- Parameswari, Y. and Srinivas, A. (2017). Weed Management in Rice- A Review.
- Paul, S., Ali, N., Gayen, D., Datta, S.K. and Datta, K. (2012). Molecular breeding of *Oxfer2* gene to increase iron nutrition in rice grain. *GM Crops and Food*, 3(4): 310-316.

- Pfeiffer, W.H. and McClafferty, B. (2007). Biofortification: breeding micronutrient-dense crops. *Breeding Major Food Staples*, 61-91.
- Powell, W., Machray, G.C. and Provan, J. (1996). Polymorphism revealed by simple sequence repeats. *Trends in Plant Science*, 1(7): 215-222.
- Prasad, A.S. (2007). Zinc: mechanisms of host defense. *The Journal of Nutrition*, 137(5): 1345-1349.
- Prasad, A.S., Halsted, J.A. and Nadimi, M. (1961). Syndrome of iron deficiency anemia, hepatosplenomegaly, hypogonadism, dwarfism and geophagia. *The American Journal of Medicine*, 31(4): 532-546.
- Prasad, R. (2010). Zinc biofortification of food grains in relation to food security and alleviation of zinc malnutrition. *Current Science*, 1300-1304.
- Prasad, R., Shivay, Y.S. and Kumar, D. (2014). Agronomic biofortification of cereal grains with iron and zinc. *Advances in Agronomy*, 125:55-91.
- Prigge, V., Maurer, H.P., Mackill, D.J., Melchinger, A.E. and Frisch, M. (2008). Comparison of the observed with the simulated distributions of the parental genome contribution in two marker assisted backcross programs in rice. *Theoretical and Applied Genetics*, 116(5): 739-744.
- Rafii, M., Zakiah, M., Asfaliza, R., Haifaa, I., Latif, M. and Malek, M. (2014). Grain quality performance and heritability estimation in selected F1 rice genotypes. *Sains Malaysiana*, 43(1): 1-7.
- Rajamoorthy, Y. and Munusamy, S. (2015). Rice industry in Malaysia: challenges, policies and implications. *Procedia Economics and Finance*, 31:861-867.
- Rajpurohit, D., Kumar, R., Kumar, M., Paul, P., Awasthi, A., Basha, P.O., Puri, A., Jhang, T., Singh, K. and Dhaliwal, H.S. (2011). Pyramiding of two bacterial blight resistances and a semi dwarfing gene in Type 3 Basmati using marker-assisted selection. *Euphytica*, 178(1): 111-126.
- Ramesh, S.A., Shin, R., Eide, D.J. and Schachtman, D.P. (2003). Differential metal selectivity and gene expression of two zinc transporters from rice. *Plant Physiology*, 133(1): 126-134.
- Ramkumar, G., Sivaranjani, A., Pandey, M.K., Sakthivel, K., Rani, N.S., Sudarshan, I., Prasad, G., Neeraja, C., Sundaram, R. and Viraktamath, B. (2010). Development of a PCR-based SNP marker system for effective selection of kernel length and kernel elongation in rice. *Molecular Breeding*, 26(4): 735-740.
- Ranst, E.v., Verloo, M., Demeyer, A. and Pauwels, J. (1999). *Manual for the soil chemistry and fertility laboratory: Analytical methods for soils and plants equipment, and management of consumables*, Gent Universiteit.



- Rehman, H.-u., Aziz, T., Farooq, M., Wakeel, A. and Rengel, Z. (2012). Zinc nutrition in rice production systems: a review. *Plant and Soil*, 361(1-2): 203-226.
- Ren, X.-L., Liu, Q.-L., Wu, D.-X. and Shu, Q.-Y. (2006). Variations in concentration and distribution of health-related elements affected by environmental and genotypic differences in rice grains. *Rice Science*, 13(170): 178.
- Ribaut, J.-M., Jiang, C. and Hoisington, D. (2002). Simulation experiments on efficiencies of gene introgression by backcrossing. *Crop Science*, 42(2): 557-565.
- Rohlf, F. (2000). Numerical taxonomy and multivariate analysis system version 2.1. New York: Exeter Publishing.
- Rose, T.J., Impa, S.M., Rose, M.T., Pariasca-Tanaka, J., Mori, A., Heuer, S., Johnson-Beebout, S. and Wissuwa, M. (2012). Enhancing phosphorus and zinc acquisition efficiency in rice: a critical review of root traits and their potential utility in rice breeding. *Annals of Botany*, 112(2): 331-345.
- Sabu, K.K., Abdullah, M.Z., Lim, L.S. and Wickneswari, R. (2006). Development and evaluation of advanced backcross families of rice for agronomically important traits. *Communications in Biometry and Crop Science*, 1(2): 111-123.
- Salina, E., Dobrovolskaya, O., Efremova, T., Leonova, I. and Röüder, M. (2003). Microsatellite monitoring of recombination around the *Vrn-B1* locus of wheat during early backcross breeding. *Plant Breeding*, 122(2): 116-119.
- Sambrook, J. and Russell, D.W. (2001). Molecular cloning: a laboratory manual 3rd edition. Cold spring-Harbour Laboratory Press, UK.
- Samman, S. (2007). Zinc. *Nutrition and Dietetics*, 64(s4).
- Sasaki, A., Yamaji, N. and Ma, J.F. (2014). Overexpression of *OsHMA3* enhances Cd tolerance and expression of Zn transporter genes in rice. *Journal of Experimental Botany*, 65(20): 6013-6021.
- Semagn, K., Bjørnstad, Å. and Ndjiondjop, M. (2006). Progress and prospects of marker assisted backcrossing as a tool in crop breeding programs. *African Journal of Biotechnology*, 5(25).
- Septiningsih, E.M., Pamplona, A.M., Sanchez, D.L., Neeraja, C.N., Vergara, G.V., Heuer, S., Ismail, A.M. and Mackill, D.J. (2008). Development of submergence-tolerant rice cultivars: the *Sub1* locus and beyond. *Annals of Botany*, 103(2): 151-160.
- Servin, B. and Hospital, F. (2002). Optimal positioning of markers to control genetic background in marker-assisted backcrossing. *Journal of Heredity*, 93(3): 214-217.

- Shahzad, Z., Rouached, H. and Rakha, A. (2014). Combating mineral malnutrition through iron and zinc biofortification of cereals. *Comprehensive Reviews in Food Science and Food Safety*, 13(3): 329-346.
- Shamsudin, N.A.A., Swamy, B.M., Ratnam, W., Cruz, M.T.S., Raman, A. and Kumar, A. (2016). Marker assisted pyramiding of drought yield QTLs into a popular Malaysian rice cultivar, MR219. *BMC Genetics*, 17(1): 30.
- Shanti, M., Shenoy, V., Devi, G.L., Kumar, V.M., Premalatha, P., Kumar, G.N., Shashidhar, H., Zehr, U. and Freeman, W. (2010). Marker-assisted breeding for resistance to bacterial leaf blight in popular cultivar and parental lines of hybrid rice. *Journal of Plant Pathology*, 495-501.
- Shi, C., Zhang, H., Wu, J., Li, C. and Ren, Y. (2003). Genetic and genotype× environment interaction effects analysis for erucic acid content in rapeseed (*Brassica napus* L.). *Euphytica*, 130(2): 249-254.
- Shu, Q. (2009). Induced plant mutations in the genomics era. *Proceedings of an International Joint FAO/IAEA Symposium*, Vienna, Austria.
- Shu, Q., Wu, D. and Xia, Y. (1997). The most widely cultivated rice variety 'Zhefu 802' in China and its genealogy.
- Siangliw, J.L., Jongdee, B., Pantuwan, G. and Toojinda, T. (2007). Developing KDML105 backcross introgression lines using marker-assisted selection for QTLs associated with drought tolerance in rice. *Science Asia*, 33:207-214.
- Singh, A., Gopalakrishnan, S., Singh, V., Prabhu, K., Mohapatra, T., Singh, N., Sharma, T., Nagarajan, M., Vinod, K. and Singh, D. (2011). Marker assisted selection: a paradigm shift in Basmati breeding. *Indian Journal of Genetics and Plant Breeding*, 71(2): 120.
- Singh, A., Singh, V.K., Singh, S., Pandian, R., Ellur, R.K., Singh, D., Bhowmick, P.K., Gopala Krishnan, S., Nagarajan, M. and Vinod, K. (2012). Molecular breeding for the development of multiple disease resistance in Basmati rice. *AoB Plants*.
- Singh, V.K., Singh, A., Singh, S., Ellur, R.K., Choudhary, V., Sarkel, S., Singh, D., Krishnan, S.G., Nagarajan, M. and Vinod, K. (2012a). Incorporation of blast resistance into "PRR78", an elite Basmati rice restorer line, through marker assisted backcross breeding. *Field Crops Research*, 128:8-16.
- Singh, V.K., Singh, A., Singh, S., Ellur, R.K., Singh, D., Gopala Krishnan, S., Bhowmick, P., Nagarajan, M., Vinod, K. and Singh, U. (2013). Marker-assisted simultaneous but stepwise backcross breeding for pyramiding blast resistance genes *Piz5* and *Pi54* into an elite Basmati rice restorer line 'PRR78'. *Plant Breeding*, 132(5): 486-495.

- Slamet-Loedin, I.H., Johnson-Beebout, S.E., Impa, S. and Tsakirpaloglou, N. (2015). Enriching rice with Zn and Fe while minimizing Cd risk. *Frontiers in Plant Science*, 6.
- Sperotto, R.A., Boff, T., Duarte, G.L., Santos, L.S., Grusak, M.A. and Fett, J.P. (2010). Identification of putative target genes to manipulate Fe and Zn concentrations in rice grains. *Journal of Plant Physiology*, 167(17): 1500-1506.
- Sreewongchai, T., Toojinda, T., Thanintorn, N., Kosawang, C., Vanavichit, A., Tharreau, D. and Sirithunya, P. (2010). Development of elite *indica* rice lines with wide spectrum of resistance to Thai blast isolates by pyramiding multiple resistances QTLs. *Plant Breeding*, 129(2): 176-180.
- Stangoulis, J.C., Huynh, B.-L., Welch, R.M., Choi, E.-Y. and Graham, R.D. (2007). Quantitative trait loci for phytate in rice grain and their relationship with grain micronutrient content. *Euphytica*, 154(3): 289-294.
- Steele, K., Price, A., Shashidhar, H. and Witcombe, J. (2006). Marker-assisted selection to introgress rice QTLs controlling root traits into an Indian upland rice variety. *Theoretical and Applied Genetics*, 112(2): 208-221.
- Suh, J.-P., Jeung, J.-U., Noh, T.-H., Cho, Y.-C., Park, S.-H., Park, H.-S., Shin, M.-S., Kim, C.-K. and Jena, K.K. (2013). Development of breeding lines with three pyramided resistance genes that confer broad-spectrum bacterial blight resistance and their molecular analysis in rice. *Rice*, 6(1): 5.
- Suh, J.-P., Yang, S.-J., Jeung, J.-U., Pamplona, A., Kim, J.-J., Lee, J.-H., Hong, H.-C., Yang, C. I., Kim, Y.-G. and Jena, K.K. (2011). Development of elite breeding lines conferring *Bph18* gene-derived resistance to brown plant hopper (BPH) by marker-assisted selection and genome wide background analysis in japonica rice (*Oryza sativa* L.). *Field Crops Research*, 120(2): 215-222.
- Suh, J., Roh, J., Cho, Y., Han, S., Kim, Y. and Jena, K. (2009). The *Pi40* gene for durable resistance to rice blast and molecular analysis of *Pi40*-advanced backcross breeding lines. *Phytopathology*, 99(3): 243-250.
- Sundaram, R.M., Vishnupriya, M., Laha, G.S., Rani, N.S., Rao, P.S., Balachandran, S.M., Reddy, G.A., Sarma, N.P. and Sonti, R.V. (2009). Introduction of bacterial blight resistance into Triguna, a high yielding, mid-early duration rice variety. *Biotechnology Journal*, 4(3): 400-407.
- Sundaram, R.M., Vishnupriya, M.R., Biradar, S.K., Laha, G.S., Reddy, G.A., Rani, N.S., Sarma, N.P. and Sonti, R.V. (2008). Marker assisted introgression of bacterial blight resistance in Samba Mahsuri, an elite *indica* rice variety. *Euphytica*, 160(3): 411-422.

- Susanto, U. (2008). Mapping of quantitative trait loci for high iron and zinc content in polished rice (*Oryza sativa* L) grain and some agronomic traits using simple sequence repeats markers.
- Swamy, B., Kaladhar, K., Anuradha, K., Batchu, A., Longvah, T., Viraktamath, B. and Sarla, N. (2011). Enhancing iron and zinc concentration in rice grains using wild species. *ADNAT Convention and International Symposium on Genomics and Biodiversity*, CCMB, Hyderabad.
- Swamy, B.M., Rahman, M.A., Inabangan-Asilo, M.A., Amparado, A., Manito, C., Chadha Mohanty, P., Reinke, R. and Slamet-Loedin, I.H. (2016). Advances in breeding for high grain Zinc in Rice. *Rice*, 9(1): 49.
- Takeuchi, Y., Ebitani, T., Yamamoto, T., Sato, H., Ohta, H., Hirabayashi, H., Kato, H., Ando, I., Nemoto, H. and Imbe, T. (2006). Development of isogenic lines of rice cultivar Koshihikari with early and late heading by marker-assisted selection. *Breeding Science*, 56(4): 405-413.
- Tanksley, S., Young, N., Paterson, A. and Bonierbale, M. (1989). RFLP mapping in plant breeding: new tools for an old science. *Nature Biotechnology*, 7(3): 257-264.
- Tian, F., Li, D.J., Fu, Q., Zhu, Z.F., Fu, Y.C., Wang, X.K. and Sun, C.Q. (2006). Construction of introgression lines carrying wild rice (*Oryza rufipogon* Griff.) segments in cultivated rice (*Oryza sativa* L.) background and characterization of introgressed segments associated with yield related traits. *Theoretical and Applied Genetics*, 112(3): 570-580.
- Toojinda, T., Tragoonrun, S., Vanavichit, A., Siangliw, J.L., Pa-In, N., Jantaboon, J., Siangliw, M. and Fukai, S. (2005). Molecular breeding for rainfed lowland rice in the Mekong region. *Plant Production Science*, 8(3): 330-333.
- Torres, A.M. (2010). Application of molecular markers for breeding disease resistant varieties in crop plants. *Molecular Techniques in Crop Improvement*, Springer: 185-205.
- Tosiah, S., Kadir, J., Sariah, M., Juraimi, A., Lo, N. and Soetikno, S. (2009). Survey and evaluation of native fungal pathogens for biocontrol of barnyard grass (*Echinochloa crus-galli* complex). *Journal of Tropical Agriculture and Food Science*, 37(1): 119-128.
- van Berloo, R. (2008). GGT 2.0: versatile software for visualization and analysis of genetic data. *Journal of Heredity*, 99(2): 232-236.
- Visscher, P. (1996). Proportion of the variation in genetic composition in backcrossing programs explained by genetic markers. *Journal of Heredity*, 87(2): 136-138.

- Visser, P.M., Haley, C.S. and Thompson, R. (1996). Marker-assisted introgression in backcross breeding programs. *Genetics*, 144(4): 1923-1932.
- Walker, C., Fischer, Ezzati, M. and Black, R. (2009). Global and regional child mortality and burden of disease attributable to zinc deficiency. *European Journal of Clinical Nutrition*, 63(5): 591.
- Wang, L., Zhang, Z., Wei, L., Zhang, D., Teng, F., Tao, Y. and Zheng, Y. (2009). The residual background genome from a donor within an improved line selected by marker-assisted selection: impact on phenotype and combining ability. *Plant Breeding*, 128(5): 429-435.
- Waters, B.M. and Sankaran, R.P. (2011). Moving micronutrients from the soil to the seeds: genes and physiological processes from a biofortification perspective. *Plant Science*, 180(4): 562-574.
- Wei, D., Liping, C., Zhijun, M., Guangwei, W. and Ruirui, Z. (2010). Review of non-chemical weed management for green agriculture. *International Journal of Agricultural and Biological Engineering*, 3(4): 52-60.
- White, P.J. and Broadley, M.R. (2011). Physiological limits to zinc biofortification of edible crops. *Frontiers in Plant Science*, 2.
- William, H.M., Morris, M., Warburton, M. and Hoisington, D.A. (2007). Technical, economic and policy considerations on marker-assisted selection in crops: lessons from the experience at an international agricultural research centre. *Marker-Assisted Selection*, 381.
- Wu, C.-y., Lu, L.-l., Yang, X.-e., Feng, Y., Wei, Y.-y., Hao, H.-l., Stoffella, P. and He, Z.-l. (2010). Uptake, translocation, and remobilization of zinc absorbed at different growth stages by rice genotypes of different Zn densities. *Journal of Agricultural and Food Chemistry*, 58(11): 6767-6773.
- Xi, Z.-Y., He, F.-H., Zeng, R.-Z., Zhang, Z.-M., Ding, X.-H., Li, W.-T. and Zhang, G.-Q. (2006). Development of a wide population of chromosome single-segment substitution lines in the genetic background of an elite cultivar of rice (*Oryza sativa* L.). *Genome*, 49(5): 476-484.
- Xiao, J., Grandillo, S., Ahn, S.N., McCouch, S.R., Tanksley, S.D., Li, J. and Yuan, L. (1996b). Genes from wild rice improve yield. *Nature* (London), 384(6606): 223-224.
- Xiao, J., Li, J., Yuan, L., McCouch, S. and Tanksley, S. (1996a). Genetic diversity and its relationship to hybrid performance and heterosis in rice as revealed by PCR-based markers. *Theoretical and Applied Genetics*, 92(6): 637-643.



- Yamaji, N., Xia, X.J., Mitani-Ueno, N., Yokosho, K. and Ma, J.F. (2013). Preferential delivery of Zn to developing tissues in rice is mediated by a P-type ATPases, *OsHMA2*. *Plant Physiology*, pp. 113.216564.
- Ye, G. and Smith, K.F. (2008). Marker-assisted gene pyramiding for inbred line development: Basic principles and practical guidelines. *International Journal of Plant Breeding*, 2(1): 1-10.
- Ye, G. and Smith, K.F. (2010). Marker-Assisted Gene Pyramiding for Cultivar Development. *Plant Breeding Reviews*, 33:219-256.
- Yin, H., Gao, X., Stomph, T., Li, L., Zhang, F. and Zou, C. (2016). Zinc concentration in rice (*Oryza sativa* L.) grains and allocation in plants as affected by different zinc fertilization strategies. *Communications in Soil Science and Plant Analysis*, 47(6): 761-768.
- Young, N. and Tanksley, S. (1989a). RFLP analysis of the size of chromosomal segments retained around the *Tm-2* locus of tomato during backcross breeding. *Theoretical and Applied Genetics*, 77(3): 353-359.
- Young, N. and Tanksley, S. (1989b). Restriction fragment length polymorphism maps and the concept of graphical genotypes. *Theoretical and Applied Genetics*, 77(1): 95-101.
- Yu, S., Xu, W., Vijayakumar, C., Ali, J., Fu, B., Xu, J., Jiang, Y., Marghirang, R., Domingo, J. and Aquino, C. (2003). Molecular diversity and multilocus organization of the parental lines used in the International Rice Molecular Breeding Program. *Theoretical and Applied Genetics*, 108(1): 131-140.
- Zhang, J., Gong, L., Qu, Y. and Qu, H. (2005). The rice ears were seriously damaged by the fifth and sixth generation of small brown plant hopper in Changshu City in 2004. *China Plant Protection*, 25(4): 39.
- Zhang, M., Guo, B. and Peng, Z. (2004). Genetic effects on Fe, Zn, Mn and P contents in *Indica* black pericarp rice and their genetic correlations with grain characteristics. *Euphytica*, 135(3): 315-323.
- Zhang, M., Pinson, S.R., Tarpley, L., Huang, X.-Y., Lahner, B., Yakubova, E., Baxter, I., Guerinot, M.L. and Salt, D.E. (2014). Mapping and validation of quantitative trait loci associated with concentrations of 16 elements in unmilled rice grain. *Theoretical and Applied Genetics*, 127(1): 137-165.
- Zhang, X., Zhang, G., Guo, L., Wang, H., Zeng, D., Dong, G., Qian, Q. and Xue, D. (2011). Identification of quantitative trait loci for Cd and Zn concentrations of brown rice grown in Cd polluted soils. *Euphytica*, 180(2): 173-179.
- Zhou, Y.-L., Uzokwe, V.N., Zhang, C.-H., Cheng, L.-R., Wang, L., Chen, K., Gao, X.-Q., Sun, Y., Chen, J.-J. and Zhu, L.-H. (2011). Improvement of bacterial blight

resistance of hybrid rice in China using the *Xa23* gene derived from wild rice (*Oryza rufipogon*). *Crop Protection*, 30(6): 637-644.

Zou, J., Pan, X., Chen, Z., Xu, J., Lu, J., Zhai, W. and Zhu, L. (2000). Mapping quantitative trait loci controlling sheath blight resistance in two rice cultivars (*Oryza sativa* L.). *Theoretical and Applied Genetics*, 101(4): 569-573.

