



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF DIFFERENT TYPES OF PROTECTED FAT ON RUMEN  
METABOLISM, MEAT QUALITY AND METABOLOMICS IN DORPER  
CROSSBRED SHEEP***

ATIQUE AHMED BEHAN

FP 2018 82



**EFFECTS OF DIFFERENT TYPES OF PROTECTED FAT ON RUMEN  
METABOLISM, MEAT QUALITY AND METABOLOMICS IN DORPER  
CROSSBRED SHEEP**

By  
**ATIQUE AHMED BEHAN**

Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

February 2018

## COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## **DEDICATION**

This thesis is dedicated to

**MY LATE FATHER**

**AND**

**MY MOTHER WITH LOVE**

who always supported and encouraged me to do the best



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment  
of the requirement for the degree of Doctor of Philosophy

**EFFECTS OF DIFFERENT TYPES OF PROTECTED FAT ON RUMEN  
METABOLISM, MEAT QUALITY AND METABOLOMICS IN DORPER  
CROSSBRED SHEEP**

By

**ATIQUE AHMED BEHAN**

February 2018

**Chairman : Associate Professor Anjas Asmara @ Ab. Hadi Samsudin, PhD**  
**Faculty : Agriculture**

The prilled fat, lecithinized prilled fat and calcium soap (calcium salts of palm fatty acids) are the extensively used rumen protected fats (RPF). These are used in ruminant diets to protect dietary lipids from rumen biohydrogenation and to prevent detrimental effects of high fats on rumen fermentation. Supplementation of the protected fats could improve *in vitro* and *in vivo* rumen metabolism, nutrient intake and digestibility, meat quality and metabolomics; and modify meat fatty acid profile. There is very limited number of studies in the literature discusses the use of RPF and their impact on performance, meat quality characteristics and fatty acid profile in sheep. However, the influences of RPF supplementation on rumen metabolism have been highly variable and inconsistent and their impacts on meat quality remain obscure. Therefore there is a need for specific studies to permit personalized decisions and informed choices in the utilization of protected fats. Thus, the present study was conducted to examine the effects of different types of protected fats on *in vitro* and *in vivo* rumen metabolism, nutrient intake and digestibility, serum biochemistry; meat quality and fatty acid profile and meat metabolomics in Dorper crossbred sheep.

*In vitro* experiment was conducted using ruminal fluid from fistulated Dorper sheep. Treatment consisted of basal diet (70:30 concentrate to rice straw) with no added RPF (T1), basal diet plus prilled fat (T2), basal diet plus prilled fat with lecithin (T3) and basal diet plus calcium soap of palm fatty acids (T4). Completely randomized design (CRD) was followed. *In vitro* gas production, fermentation kinetics, *in vitro* dry matter digestibility (IVDMD), *in vitro* organic matter digestibility (IVOMD), rumen fermentation characteristics and apparent biohydrogenation of fatty acids were determined. The cumulative gas production and gas production kinetics were not affected by RPF. Prilled fat with lecithin increased IVDMD and IVOMD significantly.

Metabolizable energy was not affected by addition of RPF. The RPF did not influence significantly pH, ammonia nitrogen, methane, VFA and molar proportion of VFA. However, the diet containing prilled fat with lecithin (T3) reduced acetate to propionate ratio, decreased methane numerically and increased biohydrogenation of C18:2n-6 and C18 UFA without disrupting rumen fermentation.

For the *in vivo* experiment, 36 male Dorper crossbred sheep about 18 months of age were used to evaluate the effects of different protected fats on nutrient intake, nutrient digestibility, serum biochemistry, meat quality and fatty acids. The animals were fed with the four experimental diets for 90 days (including last 10 days for digestibility trial) and were slaughtered. The diets did not affect body weight (BW), feed conversion ratio (FCR) and feed efficiency. There was no significant difference ( $P>0.05$ ) seen in the intake and digestibility of all nutrients except ether extract (EE) and crude fibre (CF).

The rumen fermentation characteristics including pH, methane ( $\text{CH}_4$ ), VFA and molar proportions of VFA, acetate to propionate ratio differ significantly ( $P<0.05$ ). The maximum  $\text{CH}_4$  reduction was observed in the diet T3 while T2 showed the least reduction. Concentration of ammonia nitrogen was not different significantly ( $P>0.05$ ) among the treatments. Numerically the lowest total VFA concentration was seen in the diet prilled fat with lecithin (T3). Fatty acid profile of rumen digesta was significantly different for  $\sum\text{SFA}$ ,  $\sum\text{MUFA}$ ,  $\sum\text{PUFA}$ ,  $\sum\text{n-3}$  and  $\sum\text{n-6}$ . Neither the diet nor the sampling time influenced serum cholesterol (total, HDL, LDL and VLDL), triglycerides, glucose and fatty acids. Serum fatty acids including  $\sum\text{SFA}$ ,  $\sum\text{UFA}$ ,  $\sum\text{MUFA}$ ,  $\sum\text{PUFA}$  did not significantly differ ( $P>0.05$ ). Sheep fed with diet containing RPF had higher ( $P<0.05$ ) n-3 PUFA as compared to control. The diets did not influence ( $P>0.05$ ) the levels of ALT, ALP and AST but the concentrations of ALP and AST were affected by sampling day.

There was no difference ( $P>0.05$ ) in slaughter weight, hot and cold carcass weights, dressing percentage, chilling loss, rib eye area, non-carcass components, non-carcass fats, and primal cuts. However, back fat thickness was significantly affected among the treatments. Chemical composition of *longissimus dorsi* (LD) and *semitendinosus* (ST) muscles, meat cholesterol, meat pH, drip loss, cooking loss and shear force were not significantly affected.

The muscle and liver metabolomics was conducted using  $^1\text{H}$  NMR spectroscopy. Six metabolites were identified from the muscle tissues including choline, creatine, glycerophosphocholine, inosine, isoleucine and lactate. The concentration of choline, creatine, glycerophosphocholine, inosine and lactate were significantly different but there was no significant difference observed in the concentration of isoleucine.

The supplementation of prilled fat with lecithin (T3) decreased SFA and increased MUFA and PUFA in the meat. Also, it increased the concentrations of C18:1n-9, CLA *Cis*-9 *Trans*-11, CLA *Trans*-10 *Cis*-12, C18-2n-6, C18-3n-3 and reduced n-6:n-3 which is beneficial to human health making the meat from Dorper crossbred sheep, free from negative effect.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**KESAN JENIS LEMAK BERBEZA TERHADAP METABOLISME RUMEN,  
KUALITI DAGING DAN METABOLOMIK PADA KAMBING BIRI-BIRI  
DORPER**

Oleh

**ATIQUE AHMED BEHAN**

Februari 2018

Pengerusi : Profesor Madya Anjas Asmara @ Ab. Hadi Samsudin, PhD  
Fakulti : Pertanian

Lemak terlindung, lemak terurai dan tepu kalsium (garam kalsium daripada asid lemak sawit) adalah lemak rumen terlindung yang digunakan secara meluas (RPF). Ia digunakan dalam diet ruminan untuk melindungi lipid pemakanan daripada biohidrogenasi rumen dan untuk mencegah kesan-sesan sampingan lemak tinggi pada penapaian rumen. Tambahan lemak terlindung dapat meningkatkan rumen metabolisme *in vitro* dan dalam *in vivo*, pengambilan nutrien dan pencernaan, kualiti daging dan metabolik; dan mengubah suai profil asid lemak daging. Terdapat sangat banyak kajian dalam kajian lepas dalam membincangkan penggunaan RPF dan kesannya terhadap prestasi, ciri-ciri kualiti daging dan profil asid lemak pada kambing. Walau bagaimanapun, pengaruh suplemen RPF terhadap metabolisme rumen sangat jauh berubah dan tidak konsisten dan impak mereka terhadap kualiti daging tetap tidak jelas. Oleh itu, terdapat keperluan untuk penyelidikan khusus untuk membenarkan keputusan dan pilihan peribadi yang dimaklumkan dalam penggunaan lemak terlindung. Oleh itu, kajian ini dijalankan untuk mengkaji kesan pelbagai jenis lemak terlindung pada *in vitro* dan dalam metabolisme *in vivo*, pengambilan nutrien dan pencernaan, biokimia serum; kualiti daging dan profil asid lemak dan metabolism daging pada kambing biri-biri Dorper.

Eksperimen *in vitro* dijalankan menggunakan cecair ruminal dari kambing biri-biri Dorper yang difistula. Rawatan terdiri daripada diet asas (70:30 menumpukan konsentrat kepada jerami padi) tanpa tambahan RPF (T1), diet basal ditambah lemak terlindung (T2), diet basal ditambah lemak terlindung dengan lesitin (T3) dan diet asas serta kalsium asid lemak sawit (T4). Reka bentuk rawak sepenuhnya (CRD) diikuti. Pengeluaran gas *in vitro*, kinetik penapaian, penghadaman bahan kering dalam *in vitro* (IVDMD), *in vitro* bahan organik pencernaan (IVOMD), ciri penapaian rumen dan

biohidrogenasi asid lemak ditentukan. Pengeluaran gas kumulatif dan kinetik pengeluaran gas tidak terjejas oleh RPF. Lemak yang terlindung bersama dengan lesitin meningkat IVDMD dan IVOMD dengan ketara. Tenaga metabolizable tidak terjejas oleh penambahan RPF. RPF tidak mempengaruhi pH, amonia nitrogen, metana, VFA dan bahagian molar VFA yang ketara. Walau bagaimanapun, diet yang mengandungi lemak terlindung dengan lesitin (T3) mengurangkan asetat kepada nisbah propionat, menurunkan kadar metana dan peningkatan biohydrogenasi C18: 2n-6 dan C18 UFA tanpa mengganggu penapaian rumen.

Untuk eksperimen *in vivo*, 36 ekor kambing biri-biri Dorper jantan berusia kira-kira 18 bulan digunakan untuk menilai kesan-kesan lemak yang berlainan pada pengambilan nutrien, kecernaan nutrien, biokimia serum, kualiti daging dan asid lemak. Haiwan ini diberi makan dengan empat diet percubaan selama 90 hari (termasuk 10 hari terakhir untuk percubaan pencernaan) dan disembelih. Diet tidak mempengaruhi berat badan (BW), nisbah penukaran makanan (FCR) dan kecekapan suapan. Tidak terdapat perbezaan yang signifikan ( $P > 0.05$ ) yang dilihat dalam pengambilan dan penghadaman semua nutrien kecuali keputusan ekstrak ether (EE) dan serat mentah (CF).

Ciri-ciri penapaian rumen termasuk pH, metana (CH<sub>4</sub>), VFA dan bahagian molar VFA, asetat kepada nisbah propionat berbeza dengan ketara ( $P < 0.05$ ). Pengurangan CH<sub>4</sub> adalah maksimum diperhatikan dalam diet T3 manakala T2 menunjukkan pengurangan yang paling rendah. Kepekatan nitrogen ammonia tidak banyak berbeza ( $P > 0.05$ ) di antara semua rawatan. Secara numerik jumlah kepekatan VFA yang paling rendah dilihat dalam diet lemak terlindung dengan lesitin (T3). Profil asid lemak rumen yang dicernakan adalah berbeza dengan ketara untuk  $\Sigma$ SFA,  $\Sigma$ MUFA,  $\Sigma$ PUFA,  $\Sigma$ n-3 dan  $\Sigma$ n-6. Diet atau masa pensampelan tidak mempengaruhi kolesterol serum (jumlah, HDL, LDL dan VLDL), triglicerida, glukosa dan asid lemak. Asid lemak serum termasuk  $\Sigma$ SFA,  $\Sigma$ UFA,  $\Sigma$ MUFA,  $\Sigma$ PUFA adalah tidak berbeza jauh ( $P > 0.05$ ). Kambing yang diberi makanan yang mengandungi RPF lebih tinggi n-3 PUFA secara ketara ( $P < 0.05$ ) berbanding dengan kawalan. Diet tidak mempengaruhi ( $P > 0.05$ ) tahap ALT, ALP dan AST tetapi kepekatan ALP dan AST dipengaruhi oleh hari sampling.

Tidak ada perbezaan yang ketara ( $P > 0.05$ ) dalam berat penyembelihan, berat badan panas dan sejuk, peratusan bersama kulit, kehilangan kerengsaan, kawasan mata rusuk, komponen bukan karkas, lemak bukan karkas, dan potongan awal. Walau bagaimanapun, ketebalan lemak belakang telah terjejas dengan ketara di kalangan rawatan. Komposisi kimia longissimus dorsi (LD) dan semitendinosus (ST) otot, kolesterol daging, pH daging, kehilangan titisan, daya masakan dan daya ricih tidak terjejas dengan ketara.

Metabolisma otot dan hati telah dijalankan menggunakan spektroskopi  $^1\text{H}$  NMR. Enam metabolit dikenal pasti dari tisu otot termasuk choline, creatine, glycerophosphocholine, inosine, isoleucine dan lactate. Kepekatan choline, creatine, glycerophosphocholine, inosine dan lactate sangat berbeza tetapi tiada perbezaan yang signifikan dalam kepekatan isoleucine.

Suplemen lemak terlindung dengan lesitin (T3) menurunkan SFA dan meningkatkan MUFA dan PUFA dalam daging. Selain itu, ia meningkatkan kepekatan C18: 1n-9, CLA Cis-9 Trans-11, CLA Trans-10 Cis-12, C18-2n-6, C18-3n-3 dan mengurangkan n-6: n-3 dimana ia bermanfaat untuk kesihatan manusia yang menjadikan daging dari kambing biri-biri Dorper bebas dari kesan negatif.

## **ACKNOWLEDGEMENTS**

First and foremost, I am grateful to Almighty Allah for the strength, wellbeing and patience to complete this journey.

I would like to express my gratitude to the chairman of my supervisory committee, Associate Professor Dr. Anjas Asmara @ Ab. Hadi Bin Samsudin for his support, patience, willingness to help, encouragement and guidance throughout my candidature. I would like to extend my thanks to members of my supervisory committee, Professor Dr. Loh Teck Chwen and Associate Professor Dr. Datin Sharida Fakurazi for their encouragement, constructive criticism and valuable suggestions.

I am very grateful to Sindh Agriculture University, Tandojam, Pakistan for providing me scholarship to pursue my PhD at UPM. I would like to extend my thanks and appreciations to all staff members of the Department of Animal Science, Faculty of Agriculture and Ruminant Unit Farm 2 for their kind cooperation and help in conducting my experiments.

Special thanks are due to all friends, Malaysian and international for their fine cooperation and moral support during the hard times, among them, Dr. Tanbir Ahmed, Dr. Abdul Kareem, Dr. Osama Alsaeed, Dahiru Soli, Humam Ali Merrza, Abubakar, Muideen Ahmed Adewale, Jurhamid C. Imlan, Dr. Adeyeme Kazeem Dauda, Hasfar, Dr. Candyrine and others not mentioned here but their help is fully appreciated.

Thanks are extended to Dr. Mahdi Ebrahimi, Dr. Ubedullah Kaka, Dr. Asmatullah Kaka, Dr. M. Umar Chhalgari, Dr. Tanweer Fatah Abro, Dr. Abdul Raheem Channa, Dr. Abdul Razaque Chhachhar, Dr. Shafeeqe Ahmed Memon, Dr. Khaleeq ur Rehman Bhutto, Dr. Pasand Ali Khoso, Dr. Ghulam Mujtaba Khushk, Dr. Agha Mushtaque, Dr. Muhammad Zuber, Dr. Saifullah Bullo, Dr. Muhammad Tayyab Akhtar, Dr. Waseem Mumtaz, Zulfiqar Ahmed Maher, Noor Ahmed Brohi, Mansoor Ali Khuhro, Mazhar Iqbal, Sadaf Shakoor, Sidra Rana and Farhana Haque for their continuous help and support throughout my stay at UPM.

All Pakistani students in UPM were involved in this work in one way or another and their contribution is highly appreciated.

I extend my thanks to my well-wishers Syed Allah Bachayo Shah, Mushtaque Ahmed Memon, Zulfiqar Ali Behan, Abdul Razaque Behan, Abdul Hayue Behan, Dr. Abdul Qayoom Khanzada, Dr. Ghulam Nabi Dahri, Dr. Muhammad Haroon Baloch, Dr. Irshad Ali Korejo, Dr. Ali Hassan Buriro, Dr. Shahid Laghari, Dr. Razique Hussain Laghari, Dr. Majeed Hakeem Dhamrah, Dr. Bashir Ahmed Dahri, Dr. G. Murtaza

Laghari, Dr. Faheem KK, Dr. G. Ali Jalalani, Dr. Abdul Salam Chandio, Abdul Jabbar Dahri, Dr. Akeel Ahmed Memon, Dr. Barkatullah Qureshi, Dr. Muhammad Yaqoob Koondhar, Jam Muhammad Zaman Vako, Qari Muhammad Shareef, Aijaz ur Rehman Behan for their prayers and moral support.

Many thanks to Muhammad Yaqoob Behan, Osama Behan, Shuned Behan, Mansoor Ali, Noor Ahmed, Mashooque Ali, Saifullah, Faqeer Muhammad Pathan, Khan Muhammad Pathan, Abdul Razaque Khyber who took care of my family in my absence.

I am sincerely grateful to my family members including my mother, my wife my sons and my daughter, my sisters and my brother Muhammad Issa Behan for their love, endless support, encouragement, understanding, and reassurance during my study.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Anjas Asmara @ Ab. Hadi Bin Samsudin, PhD**

Associate Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Chairman)

**Loh Teck Chwen, PhD**

Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Member)

**Sharida Fakurazi, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Atique Ahmed Behan, GS33285

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature:

Name of Chairman  
of Supervisory  
Committee:

\_\_\_\_\_  
Associate Professor  
Dr. Anjas Asmara @ Ab. Hadi Bin Samsudin

Signature:

Name of Member  
of Supervisory  
Committee:

\_\_\_\_\_  
Professor  
Dr. Loh Teck Chwen

Signature:

Name of Member  
of Supervisory  
Committee:

\_\_\_\_\_  
Associate Professor  
Dr. Sharida Fakurazi

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iv
<b>ACKNOWLEDGEMENTS</b>	vii
<b>APPROVAL</b>	ix
<b>DECLARATION</b>	xi
<b>LIST OF TABLES</b>	xviii
<b>LIST OF FIGURES</b>	xx
<b>LIST OF APPENDICES</b>	xxi
<b>LIST OF ABBREVIATIONS</b>	xxii
 <b>CHAPTER</b>	
 <b>1 INTRODUCTION</b>	1
 <b>2 LITERATURE REVIEW</b>	5
2.1 Global contribution of livestock	5
2.2 The impact of livestock products on human health	5
2.3 Livestock industry in Malaysia	6
2.4 Sheep production in Malaysia	6
2.5 Ruminant feeds in Malaysia	7
2.6 Global meat production and consumption	8
2.7 Sheep meat production and consumption in Malaysia	8
2.8 Rumen microbial ecosystem	9
2.9 Fat supplementation in ruminant diet	10
2.10 Level of fat supplementation in the ruminant diet	10
2.11 Fat metabolism in the rumen	11
2.12 Rumen protected fats (RPF)	12
2.13 Significance of RPF supplementation	12
2.14 General properties of RPF	13
2.15 Types of RPF	13
2.15.1 Natural RPF	13
2.15.2 Chemically prepared RPF	13
2.16 Methods of RPF preparation	13
2.16.1 Crystalline or prilled fats	14
2.16.2 Calcium soaps	14
2.16.3 Formaldehyde-treated fats	15
2.16.4 Fatty acyl amides	15
2.17 RPF and <i>in vitro</i> fermentation	15
2.18 RPF and growth performance	16
2.19 RPF and nutrient intake	16
2.20 RPF and nutrient digestibility	16

2.21	RPF and rumen fermentation	17
2.22	RPF and carcass characteristics	17
2.23	The Dorper sheep	17
2.24	Carcass characteristics of Dorper sheep	18
2.25	Meat quality attributes	19
2.25.1	Meat pH	19
2.25.2	Water holding capacity (WHC)	20
2.25.2.1	Drip loss	20
2.25.2.2	Cooking loss	21
2.25.2.3	Meat colour	21
2.25.2.4	Meat tenderness	21
2.26	Metabolomics	22
2.27	Analytical techniques for metabolomics	23
2.28	Nuclear Magnetic Resonance (NMR) spectroscopy	23
2.29	Livestock metabolomics	23
2.30	Summary	24

### **3 EFFECTS OF DIFFERENT TYPES OF PROTECTED FAT ON *IN VITRO* RUMEN FERMENTATION AND APPARENT BIOHYDROGENATION OF FATTY ACIDS**

3.1	Introduction	25
3.2	Materials and Methods	26
3.2.1	Donor Animals	26
3.2.2	Treatments and experimental design	27
3.2.3	Chemical analysis	29
3.2.4	Preparation of <i>in vitro</i> samples for analysis	30
3.2.5	Collection of rumen liquor	30
3.2.6	Preparation of buffered solutions	30
3.2.7	Preparation of media	30
3.2.8	<i>In vitro</i> rumen fermentation of samples	31
3.2.9	Determination of pH	31
3.2.10	<i>In vitro</i> gas production and fermentation kinetics	31
3.2.11	<i>In vitro</i> dry matter digestibility (IVDMD)	32
3.2.12	<i>In vitro</i> organic matter digestibility (IVOMD)	32
3.2.13	Determination of volatile fatty acids (VFA)	32
3.2.14	Determination of ammonia nitrogen (NH <sub>3</sub> -N)	32
3.2.15	Determination of methane (CH <sub>4</sub> )	33
3.2.16	Determination of metabolizable energy (ME)	33
3.2.17	Fatty acid analysis	33
3.2.18	Rate of biohydrogenation	34
3.2.19	Statistical analysis	34
3.3	Results	35
3.3.1	<i>In vitro</i> cumulative gas production	35
3.3.2	<i>In vitro</i> gas production kinetics	36
3.3.3	<i>In vitro</i> rumen fermentation characteristics	37
3.3.4	Fatty acid composition of rumen liquor after 72 hours of incubation	37

3.3.5	Apparent biohydrogenation of fatty acids after incubation	72 h	38
3.4	Discussion		39
3.4.1	<i>In vitro</i> cumulative gas production		39
3.4.2	<i>In vitro</i> gas production kinetics		40
3.4.3	<i>In vitro</i> rumen fermentation characteristics		41
3.4.4	Fatty acid profile and apparent biohydrogenation of fatty acids		44
3.5	Conclusion		45
<b>4</b>	<b>EFFECTS OF DIFFERENT TYPES OF PROTECTED FAT ON NUTRIENT INTAKE, APPARENT DIGESTIBILITY, BODY WEIGHT CHANGES AND SERUM BIOCHEMISTRY IN DORPER CROSSBRED SHEEP</b>		<b>46</b>
4.1	Introduction		46
4.2	Materials and Methods		47
4.2.1	Experimental site		47
4.2.2	Animals and housing management		48
4.2.3	Treatments and experimental design		48
4.2.4	Live body weight changes and feed intake		48
4.2.5	Apparent digestibility		49
4.2.6	Chemical analysis		49
4.2.7	Blood sampling		49
4.2.8	Serum biochemistry		49
4.2.9	Slaughtering		50
4.2.10	Collection of rumen liquor		50
4.2.11	Determination of rumen liquor pH		50
4.2.12	Rumen fermentation parameters		50
4.2.13	Fatty acid analysis		50
4.2.14	DNA extraction of rumen microbes		51
4.2.15	Quantitative Real-Time PCR		51
4.2.16	Statistical analysis		51
4.3	Results		52
4.3.1	Chemical composition of experimental diets		52
4.3.2	Body weight changes and feed efficiency		52
4.3.3	Nutrient intake in Dorper sheep		53
4.3.4	Apparent nutrient digestibility in sheep		54
4.3.5	Rumen fermentation characteristics of sheep		55
4.3.6	Rumen microbial population		56
4.3.7	Fatty acid profile of ruminal digesta in Dorper sheep		57
4.3.8	Blood serum biochemistry		58
4.3.9	Serum enzyme activity		59
4.3.10	Fatty acid profile of blood serum		60
4.3.11	Sums and ratios of serum fatty acids		62
4.4	Discussion		63
4.4.1	Body weight changes and feed efficiency		63
4.4.2	Nutrient intake		65

4.4.3	Apparent nutrient digestibility	66
4.4.4	Rumen fermentation characteristics	68
4.4.5	Rumen microbial population	69
4.4.6	Rumen fatty acid profile	71
4.4.7	Blood serum biochemistry	72
4.4.8	Blood serum enzyme activity	73
4.4.9	Fatty acid profile of blood serum	74
4.5	Conclusion	75

## **5 EFFECTS OF DIFFERENT TYPES OF PROTECTED FAT ON CARCASS CHARACTERISTICS, MEAT QUALITY AND TISSUE FATTY ACID PROFILES IN DORPER CROSSBRED SHEEP**

5.1	Introduction	76
5.2	Materials and Methods	77
5.2.1	Slaughter and carcass sampling	77
5.2.2	Ageing of meat	79
5.2.3	Chemical analysis of muscles	80
5.2.4	Determination of muscle pH	80
5.2.5	Determination of meat colour coordinates	80
5.2.6	Determination of Water Holding Capacity (WHC)	81
5.2.6.1	Drip Loss	81
5.2.6.2	Cooking loss	81
5.2.7	Texture analysis	82
5.2.8	Determination of fatty acid composition	82
5.2.9	Determination of cholesterol	82
5.2.10	Statistical analysis	83
5.3	Results	83
5.3.1	Carcass composition in Dorper sheep	83
5.3.2	Non-carcass components	84
5.3.3	Non-carcass fats	84
5.3.4	Primal cuts	85
5.3.5	Chemical composition of meat	86
5.3.6	pH, water holding capacity and shear force of <i>longissimus dorsi</i> (LD) muscle in Dorper sheep	86
5.3.7	pH, water holding capacity and shear force of <i>semitendinosus</i> (ST) muscle in Dorper sheep	87
5.3.8	Colour coordinates of <i>longissimus dorsi</i> (LD) muscle in Dorper sheep	89
5.3.9	Colour coordinates of <i>semitendinosus</i> (ST) muscle in Dorper sheep	90
5.3.10	Fatty acid composition of <i>longissimus dorsi</i> (LD) muscle in sheep	91
5.3.11	Fatty acid composition of <i>semitendinosus</i> (ST) muscle in sheep	92
5.3.12	Fatty acid composition of the liver in Dorper sheep	94
5.4	Discussion	96

5.4.1	Carcass composition in Dorper sheep	96
5.4.2	Non-carcass components	97
5.4.3	Non-carcass fats	97
5.4.4	Primal cuts	98
5.4.5	Chemical composition of muscles	99
5.4.6	Cholesterol level in LD and ST muscles	99
5.4.7	pH values of LD and ST muscles of sheep	100
5.4.8	Drip loss and cooking loss in LD and ST muscles of sheep	101
5.4.9	Shear force in LD and ST muscles of sheep	102
5.4.10	Colour coordinates of LD and ST muscles of sheep	103
5.4.11	Fatty acid profile of different tissues in Dorper sheep	104
5.5	Conclusion	107
<b>6</b>	<b>EFFECTS OF DIFFERENT TYPES OF PROTECTED FAT ON MUSCLE AND LIVER METABOLOME OF DORPER SHEEP USING <math>^1\text{H}</math> NMR SPECTROSCOPY</b>	108
6.1	Introduction	108
6.2	Materials and Methods	109
6.2.1	Sample collection	109
6.2.2	Extraction of tissue metabolites	109
6.2.3	Preparation of extracts for NMR Spectroscopy	110
6.2.4	NMR measurement and data processing	110
6.2.5	Statistical analysis	110
6.3	Results	111
6.3.1	Muscle tissue	111
6.3.2	Liver Tissue	114
6.4	Discussion	117
6.5	Conclusion	118
<b>7</b>	<b>GENERAL DISCUSSION</b>	119
<b>8</b>	<b>SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH</b>	123
8.1	Summary	123
8.2	Conclusion	124
8.3	Recommendation for future research	124
<b>REFERENCES</b>		125
<b>APPENDICES</b>		157
<b>BIODATA OF STUDENT</b>		169
<b>LIST OF PUBLICATIONS</b>		170

## LIST OF TABLES

Table	Page
2.1 Production, consumption and self-sufficiency of goat and sheep meat (2004-2013) in Malaysia	9
3.1 Ingredients, chemical and fatty acid composition of experimental diets	28
3.2 Fatty acid composition of rumen protected fats	29
3.3 <i>In vitro</i> gas production, gas production kinetics and digestibility with different rumen protected fats	36
3.4 <i>In vitro</i> rumen fermentation characteristics of substrates with different rumen protected fats	37
3.5 Fatty acid composition of rumen liquor at 72 hours of incubation of substrates with different rumen protected fats	38
3.6 Apparent biohydrogenation of fatty acids at 72 hours of incubation of substrates with different rumen protected fats	39
4.1 Effect of rumen protected fat on body weight changes, average daily gain and feed conversion ratio in Dorper sheep	53
4.2 Effect of different rumen protected fats on daily average nutrient intake in Dorper sheep	54
4.3 Effect of different rumen protected fats on apparent nutrient digestibility in sheep	55
4.4 Effect of different rumen protected fats on rumen fermentation characteristics in Dorper sheep	56
4.5 Effect of different rumen protected fats on microbial population (copies/ml) in rumen of Dorper sheep	57
4.6 Effect of different rumen protected fats on fatty acid composition of rumen digesta in Dorper sheep	58
4.7 Serum biochemical parameters in sheep as influenced by diet and sampling time	59
4.8 Effect of rumen protected fats on serum enzyme activity in Dorper sheep as influenced by diet and sampling time	60

4.9	Saturated and monounsaturated fatty acids (% of total fatty acids) in serum of Dorper sheep as influenced by diet and sampling time	61
4.10	Polyunsaturated fatty acids (% of total fatty acids) in serum of Dorper sheep as influenced by diet and sampling time	62
4.11	Sums and ratios of total fatty acids in serum of Dorper sheep as influenced by diet and sampling time	63
5.1	Effect of rumen protected fat on carcass composition in sheep	83
5.2	Effect of rumen protected fat on non-carcass components in sheep	84
5.3	Effect of rumen protected fat on non-carcass fats in Dorper sheep	85
5.4	Effect of rumen protected fat on primal cuts in Dorper sheep	85
5.5	Effect of rumen protected fat on chemical composition of <i>longissimus dorsi</i> and <i>semitendinosus</i> muscles in Dorper sheep	86
5.6	Influence of rumen protected fat on pH, water holding capacity and shear force of <i>longissimus dorsi</i> (LD) muscle in Dorper sheep	87
5.7	Influence of rumen protected fat on pH, water holding capacity and shear force of <i>semitendinosus</i> (ST) muscle in Dorper sheep	88
5.8	Effect of rumen protected fat on colour coordinates of <i>longissimus dorsi</i> (LD) muscle in Dorper sheep at different ageing days	89
5.9	Effect of rumen protected fat on colour coordinates of <i>semitendinosus</i> (ST) muscle in Dorper sheep at different ageing days	90
5.10	Effect of rumen protected fat on fatty acid composition of <i>longissimus dorsi</i> (LD) muscle in Dorper sheep	92
5.11	Effect of rumen protected fat on fatty acid composition of <i>semitendinosus</i> (ST) muscle in Dorper sheep	93
5.12	Effect of rumen protected fat on fatty acid composition of liver in Dorper sheep	95
6.1	Characteristic $^1\text{H}$ NMR chemical shifts and multiplicity of indentified metabolites in muscle of Dorper sheep	111
6.2	Relative quantification of metabolites found in Dorper sheep <i>longissimus dorsi</i> muscle tissues	114

## LIST OF FIGURES

Figure	Page
2.1 Population of sheep in Malaysia from 1996 to 2015	7
3.1 Rumen protected fats (RPF)	29
3.2 <i>In vitro</i> cumulative gas production profile	35
5.1 Location of <i>Longissimus dorsi</i> (LD) and <i>Semitendinosus</i> (ST) muscles	78
5.2 Primal cuts of carcass	79
5.3 The location for measuring the rib eye area and back fat thickness	80
6.1 PCA score plot of Dorper sheep <i>longissimus dorsi</i> muscle	112
6.2 PCA Loading scatter plot of Dorper sheep <i>longissimus dorsi</i> muscle	112
6.3 $^1\text{H}$ NMR spectrum of Dorper sheep <i>longissimus dorsi</i> muscle showing identified metabolites acquired at 700 MHz	113
6.4 PCA score plot of Dorper sheep liver	115
6.5 PLS score plot of Dorper sheep liver	115
6.6 Validation of PLS-DA model with permutation test (100 permutation)	116
6.7 J-resolved spectrum of Dorper sheep <i>longissimus dorsi</i> muscle	117

## LIST OF APPENDICES

Appendix		Page
A	Names of fatty acids	157
B	Chemical analysis	158
C	Preparation of solution for <i>in vitro</i> gas production	162
D	DNA extraction protocol	163
E	Primers used for quantitative real-time polymerase chain reaction (q-PCR)	164
F	Protocol for extraction of tissue metabolites	165
G	Experimental Animals	166
H	Dressed carcasses in the chiller during <i>postmortem</i> ageing	167
I	A chromatogram of fatty acids in rumen liquor	168
J	A 700 MHz nuclear magnetic resonance (NMR) instrument	168

## LIST OF ABBREVIATIONS

ADF	Acid detergent fibre
ADG	Average daily gain
ADL	Acid detergent lignin
ALP	Alkaline phosphatase
ALT	Alanine aminotransferase
ANOVA	Analysis of variance
AST	Aspartate aminotransferase
BH	Biohydrogenation
BW	Body weight
CCW	Cold carcass weight
CF	Crude fibre
CH <sub>4</sub>	Methane
CLA	Conjugated linoleic acid
cm	Centimetre
cm <sup>2</sup>	Centimetre square
CO <sub>2</sub>	Carbon dioxide
CP	Crude protein
d	Day
DM	Dry matter
DMD	Dry matter digestibility
DMI	Dry matter intake
EBW	Empty body weight
EE	Ether extract
FA	Fatty acid

FAME	Fatty acid methyl esters
FCR	Feed conversion ratio
g	Gram
GLM	Generalized linear model
h	Hour
HCW	Hot carcass weight
HDL	High density lipoprotein
IU/L	International units per litre
IVDMD	<i>In vitro</i> dry matter digestibility
IVOMD	<i>In vitro</i> organic matter digestibility
kg	Kilogram
KPH	Kidney, pelvic and heart fat
L	Litre
LD	<i>longissimus dorsi</i>
LDL	Low density lipoprotein
ME	Metabolizable energy
mg	Milligram
MJ	Mega joule
mL	Millilitre
mm	Millimetre
mM	Millimole
mmol/L	Millimole per litre
MUFA	Mono unsaturated fatty acid
N	Nitrogen
NDF	Neutral detergent fibre
NGP	Net gas production

NH <sub>3</sub> -N	Ammonia-nitrogen
NRC	National Research Council
OM	Organic matter
OMD	Organic matter digestibility
PUFA	Poly unsaturated fatty acid
RPF	Rumen protected fat
SEM	Standard error of means
SFA	Saturated fatty acid
ST	<i>Semitendinosus</i>
TVFA	Total volatile fatty acid
UFA	Unsaturated fatty acid
VFA	Volatile fatty acid
VLDL	Very low density lipoprotein
WG	Weight gain
WHC	Water holding capacity
WHO	World Health Organization

## CHAPTER 1

### INTRODUCTION

Feed is considered a major proportion of the cost of raising ruminants. The raw ingredients for animal feed such as cereal grains, vegetable and animal proteins, mineral sources, micro-ingredients and other additives are not produced in Malaysia but are imported from other countries (Loh, 2004). So as to reduce the cost incurred on the import of these raw ingredients there is a need of utilizing available low-cost indigenous resources for animal feeds to fulfil their energy needs by producing more with spending less. In this connection, there has been a recent emphasis to utilize by-products of oil palm industry as animal feed (Alimon and Wan Zahari, 2012).

In order to fulfil the energy needs, dietary lipids are being used in ruminant nutrition. However, feeding lipids in high concentrations could adversely influence rumen microbial metabolism, affecting nutrient digestibility and animal performance (Hartati *et al.*, 2012; Szumacher-Strabel *et al.*, 2009; Naik *et al.*, 2007a). The adverse effects of lipid supplementation are because of extensive biohydrogenation (BH) of fatty acids (FA), especially BH of unsaturated fatty acids (UFA) to saturated fatty acids (SFA) by rumen microbes. Thus, the negative impact of lipid supplementation can be easily overwhelmed by feeding rumen protected fats (RPF) or rumen inert or rumen bypass fats to ruminants.

Rumen protected fats are generally, a by-product of palm oil industry, considered as insoluble fats due to their protection from microbial fermentation and biohydrogenation. They remain insoluble at normal rumen pH range of 6 to 7 and escape rumen fermentation. They are then utilized as a source of energy when absorbed through the small intestine (Warner *et al.*, 2015). The use of RPF enhance fibre digestibility in high fat supplemented diets by forming insoluble soaps (Palmquist and Jenkins 1980). Moreover, supplementation of RPF improves energy efficiency as a result of reduced production of methane from the rumen and direct use of long-chain fatty acids (Park *et al.*, 2010). Owing to their inert nature, RPF can be successfully used in relatively large amounts without compromising rumen function (Ayasan and Karakozak, 2011) and reducing feed intake (Gooden, 1977).

The common available RPF include calcium soaps (calcium salts of palm fatty acids), hydrogenated fats from palm oil, fractionated palm oil fatty acids (C16), straight fats (tallow) and dry fat premixes (blends of vegetable and/or animal fats). The common methods of RPF preparation include microencapsulation with a water-insoluble lipid coating, formaldehyde treatment of a lipid-protein matrix, the formation of calcium salts of fatty acids and preparation of fatty acyl amides (Bauman *et al.*, 2003; Putnam *et al.*, 2003).

The calcium soap of palm fatty acids and the prilled fat are the most extensively used protected fats, both of which are highly digestible. Calcium soap of palm fatty acids, prilled fat and lecithinized prilled fat are the highly concentrated source of energy supplement fats, specially produced from 100 percent fully refined palm oil fraction which are non-hydrogenated and free from trans fatty acids (TFA). The high palmitic acid contents in these fats can bypass rumen and become a direct energy source for the ruminants. Moreover, lecithinized prilled fats improve the emulsifying properties and thus increase the digestibility of animals.

The calcium salts of palm fatty acids are produced by reacting palm fatty acids distillate with calcium hydroxide to form calcium soaps. The calcium soaps would not be influenced in the rumen (pH 6.5 to 6.8), and finally are fully opened making the fatty acids accessible for absorption in abomasum and duodenum (pH 3.5) (Mierlita, 2018). It was reported that the RPF in the form of calcium soap allows normal rumen fermentation and digestibility of nutrients (Schauff and Clark, 1989; Jenkins and Palmquist, 1984). Prilled fats are made by liquefying a mixture of fatty acids high in saturated fatty acid content and spraying the mixture under pressure into a cooled atmosphere in order to form a dried prilled fatty acid supplement that is inert in the rumen and does not alter rumen fermentation (Grummer, 1988; Chalupa, 1986). Furthermore, the prilled fat with lecithin act as an emulsifier by dispersing fatty acids and enhancing fatty acid absorption (Wettstein *et al.*, 2011).

Bhatt *et al.* (2015); (2013b) reported that supplementation of protected fat of industrial grade of improved nutrient digestibility, body condition and carcass characteristics in sheep. There was no adverse effect of supplementation of RPF on the rumen fermentation (Naik *et al.*, 2013). Alexander *et al.* (2002) reported that calcium soap prepared from sunflower acid oil at 10 % of DM can be fed to sheep without affecting fibre digestibility in sheep. However, there was a significant reduction in *in vitro* DM degradability (IVDMD) with an increase in the level of bypass fat (Tangendjaja *et al.*, 1993). Manso *et al.* (2006) reported that supplementation of calcium soaps of palm fatty acids improved EE digestibility and FCR in growing lambs. Therefore protected fat supplementation could improve digestibility and FCR in adult sheep as well so there is a need to evaluate the effects of protected fats in adult sheep.

Red meat is one of the major dietary sources of protein and essential nutrients such as vitamins and minerals that play vital role in human health. However, consumption of red meat may increase the risk of cardiovascular disease (CVD) and cancer in the colon (McAfee *et al.*, 2010) because of high SFA contents in it. Thus, reducing SFA content and the n-6/n-3 ratio is of major importance in meat research (Mierlita, 2018). Therefore, modifying the FA composition of ruminant meat is of prime significance (Mapiye *et al.*, 2015; Scollan *et al.*, 2014).

The supplementation of RPF has been one of the methods to reduce undesirable SFA and increase beneficial UFA in the meat (Warner *et al.*, 2015). In this regard, several studies have been conducted on effects of dietary lipids and their FA composition on various aspects of meat quality (Wood *et al.*, 2008; Schmid *et al.*, 2006; Wood *et al.*, 2004; Palmquist and Jenkins, 1980). However, most of the published studies have been inconclusive in proving whether the use of protected fat increases the content of essential fatty acids in animal meat (Lima *et al.*, 2017). Therefore, there is a need to conduct such experiment in order to evaluate the effects of protected fats.

In previous studies effects of protected fats on live weight, carcass and meat characteristics and fatty acid composition of muscle have been evaluated. However, no characterization of the muscle and liver metabolomes of sheep has ever been evaluated. Therefore the present study was planned to characterize the metabolome of the muscle and liver of sheep, and study the effect of protected fat in these tissues, which are important from the productive and metabolic perspectives. The NMR-metabolomics based approach, which, was for the first time applied to Dorper sheep for dietary fat supplementation.

Although RPF has been extensively evaluated in dairy animals (Gowda *et al.*, 2013; Naik, 2013; Shelke *et al.*, 2012; Wadhwa *et al.*, 2012) and to some extent beef cattle (Mangrum *et al.*, 2016; Long *et al.*, 2014; Hightshoe *et al.*, 1991) yet there is a need to evaluate RPF especially in sheep for meat purpose where there is a very limited number of studies done. The studies conducted on RPF in sheep are in young lambs (Bhatt *et al.*, 2015; Bhatt *et al.*, 2013b) none of those has been conducted in adult sheep, therefore adult animals (about one and half a year of age) were selected to be used in the present study especially with intention to focus on fat deposition. It is well established that fat deposition in the body is age-dependent and subcutaneous fat is accumulated at the later stage of growth (Bhatt *et al.*, 2013a). Leon *et al.* (1999) reported higher body fat reserves at 12 and 16 months of age as compared to 8 month old lambs.

It is hypothesized that supplementation of protected fat in the form of calcium soap, prilled fat and lecithinized prilled fat will improve *in vitro* and *in vivo* rumen metabolism; nutrient intake and digestibility; meat quality, meat metabolomics and will increase the circulating unsaturated fatty acids in blood and consequently depositing them into the intramuscular meat. However, producing the meat that is low in saturated fatty acids, high in monounsaturated fatty acids (MUFA) and polyunsaturated fatty acids (PUFA), concurrently prevents the development of cardiovascular disease would be an effective approach. Therefore, different types of protected fats were selected to evaluate with the following general objective:

- To determine the effects and efficacy of RPF supplementation in improving the quality of meat production in small ruminants.

The specific objectives of the study include:

1. To evaluate the effects of RPF on *in vitro* gas production, rumen fermentation, and apparent biohydrogenation of fatty acids.
2. To determine the effects of RPF on nutrient intake, digestibility, rumen fermentation, serum biochemistry and serum fatty acid profile in Dorper sheep.
3. To determine the influences of RPF on carcass characteristics, meat quality and meat fatty acid profile.
4. To determine the effects of RPF on muscle and liver metabolites using NMR-based metabolomics.

### **Presentation of the thesis**

The thesis is divided into eight chapters. The first two chapters discuss the framework of the experimental research. Chapter 1 provides the rationale for the focus of the research. Chapter 2 presents the review of literature covering the livestock industry, sheep meat production and consumption in Malaysia, rumen microbial ecosystem, fat supplementation to ruminants and its metabolism in rumen, rumen protected fats (RFP) and significance of RFP supplementation, effects of RFP on body weight changes, nutrient intake and digestibility, rumen fermentation, carcass traits, meat quality and livestock metabolomics. From Chapter 3 to 6 present the experimental works for this study. Chapter 7 describes the major findings and highlights the practical importance. Chapter 8 presents the summary, conclusions and recommendations for future research.

## REFERENCES

- Abarghuei, M. J., Rouzbehani, Y., Salem, A. Z. M., & Zamiri, M. J. (2014). Nitrogen balance, blood metabolites and milk fatty acid composition of dairy cows fed pomegranate-peel extract. *Livestock Science*, 164, 72-80.
- Aberle, E. D., Forrest, J. C., Gerrard, D. E., & Mills, E. W. (2001). Structure and composition of animal tissues. In: *Principles of Meat Science*, 116 pp.
- AbuGhazaleh, A., Riley, M., Thies, E., & Jenkins, T. (2005). Dilution rate and pH effects on the conversion of oleic acid to trans C 18: 1 positional isomers in continuous culture. *Journal of Dairy Science*, 88, 4334-4341.
- Adeyemi, K. D. (2015). Rumen metabolism, carcass traits and meat quality in goats fed blend of canola oil and palm oil. PhD thesis, Universiti Putra Malaysia. Retrieved on 1 June 2016 from <http://www.lib.upm.edu.my/>.
- Adeyemi, K. D., & Sazili, A. Q. (2014). Efficacy of carcass electrical stimulation in meat quality enhancement: A review. *Asian-Australasian Journal of Animal Sciences*, 27, 447-456.
- Adeyemi, K. D., Ebrahimi, M., Samsudin, A. A., Alimon, A. R., Karim, R., Karsani, S. A., & Sazili, A. Q. (2015a). Influence of Carotino oil on *in vitro* rumen fermentation, metabolism and apparent biohydrogenation of fatty acids. *Animal Science Journal*, 86(3), 270-278.
- Adeyemi, K. D., Ebrahimi, M., Samsudin, A. A., Sabow, A. B., & Sazili, A. Q. (2015b). Carcass traits, meat yield and fatty acid composition of adipose tissues and Supraspinatus muscle in goats fed blend of canola oil and palm oil. *Journal of Animal Science and Technology*, 57(1), 42.
- Adeyemi, K. D., Sabow, A. B., Aghwan, Z. A., Ebrahimi, M., Samsudin, A. A., Alimon, A. R., & Sazili, A. Q. (2016). Serum fatty acids, biochemical indices and antioxidant status in goats fed canola oil and palm oil blend. *Journal of Animal Science and Technology*, 58(1), 6.
- Adeyosoye, O. I., Adesokan, I. A., Afolabi, K. D., & Ekeocha, A. H. (2010). Estimation of proximate composition and biogas production from *in vitro* gas fermentation of sweet potato (*Ipomea batatas*) and wild cocoyam (*Colocasia esculenta*) peels. *African Journal of Environmental Science and Technology*, 4(6).
- Adzitey, F. (2011). Effect of pre-slaughter animal handling on carcass and meat quality. *International Food Research Journal*, 18(2).
- Aferri, G., Leme, P. R., Silva, S. D. L., Putrino, S. M., & Pereira, A. S. C. (2005). Performance and carcass characteristics of steers fed different fat sources. *Revista Brasileira de Zootecnia*, 34(5), 1651-1658.

- Alemseged, Y., & Hacker, R. B. (2014). Introduction of Dorper sheep into Australian rangelands: implications for production and natural resource management. *The Rangeland Journal*, 36(1), 85-90.
- Alexander, G., Rao, Z. P., & Prasad, J. R. (2002). Effect of supplementing sheep with sunflower acid oil or its calcium soap on nutrient utilization. *Asian Australian Journal of Animal Sciences*, 15(9), 1288-1293.
- Alimon, A. R., & Wan Zahari, W. M. (2012). Recent advances in the utilization of oil palm by-products as animal feed. In *International Conference on Livestock Production and Veterinary Technology (ICARD)*.
- Allen, L. H. (2003). Interventions for micronutrient deficiency control in developing countries: past, present and future. *The Journal of Nutrition*, 133(11), 3875S-3878S.
- Allen, M. S. (2000). Effects of diet on short-term regulation of feed intake by lactating dairy cattle. *Journal of Dairy Science*, 83(7), 1598-1624.
- Álvarez-Sánchez, B., Priego-Capote, F., & de Castro, M. L. (2010). Metabolomics analysis I. Selection of biological samples and practical aspects preceding sample preparation. *Trends in Analytical Chemistry*, 29(2), 111-119.
- Andrade, E. N., Neto, A. P., Roça, R. O., Faria, M. H., Resende, F. D., Siqueira, G. R., & Pinheiro, R. S. B. (2014). Beef quality of young Angus x Nellore cattle supplemented with rumen-protected lipids during rearing and fattening periods. *Meat Science*, 98 (4), 591-598.
- Antosik, K., Koćwin-Podsiadła, M., & Goławski, A. (2011). Effect of different CO<sub>2</sub> concentrations on the stunning effect of pigs and selected quality traits of their meat-A Short Report. *Polish Journal of Food and Nutrition Sciences*, 61(1), 69-72.
- AOAC. (2007). Official methods of analysis of the Association of Official Analytical Chemists (18<sup>th</sup> Ed). Association of Official Analytical Chemists, Washington D.C., USA
- Ariff, O. M., Sharifah, N. Y., & Hafidz, A. W. (2015). Status of beef industry of Malaysia. *Malaysian Journal of Animal Science*, 18(2), 1-21.
- Awawdeh, M. S., Obeidat, B. S., Abdullah, A. Y., & Hananeh, W. M. (2009). Effects of yellow grease or soybean oil on performance, nutrient digestibility and carcass characteristics of finishing Awassi lambs. *Animal Feed Science and Technology*, 153(3), 216-227.
- Ayaşan, T., & Karakozak, E. (2011). Use of protected fats in animal nutrition. *Atatürk Üniversitesi Veteriner Bilimleri Dergisi*, 6(1), 85-94.
- Baldwin, R. L., & Connor, E. E. (2017). Rumen function and development. *The Veterinary Clinics of North America. Food Animal Practice*.

- Barendse, W. (2014). Should animal fats be back on the table? A critical review of the human health effects of animal fat. *Animal Production Science*, 54(7), 831-855.
- Bartle, S. J., Preston, R. L., & Miller, M. F. (1994). Dietary energy source and density: effects of roughage source, roughage equivalent, tallow level, and steer type on feedlot performance and carcass characteristics. *Journal of Animal Science*, 72(8), 1943-1953.
- Bauman, D. E., Harvatine, K. J., & Lock, A. L. (2011). Nutrigenomics, rumen-derived bioactive fatty acids, and the regulation of milk fat synthesis. *Annual Review of Nutrition*, 31, 299-319.
- Bauman, D., Perfield, J., De Veth, M., & Lock, A. (2003). New perspectives on lipid digestion and metabolism in ruminants. In *Proceedings of Cornell Nutrition Conference* (Vol. 65, pp. 175-189).
- Beckonert, O., Keun, H. C., Ebbels, T. M., Bundy, J., Holmes, E., Lindon, J. C., & Nicholson, J. K. (2007). Metabolic profiling, metabolomic and metabonomic procedures for NMR spectroscopy of urine, plasma, serum and tissue extracts. *Nature Protocols*, 2(11), 2692-2703.
- Bekhit, A. E. D., Geesink, G. H., Ilian, M. A., Morton, J. D., & Bickerstaffe, R. (2003). The effects of natural antioxidants on oxidative processes and metmyoglobin reducing activity in beef patties. *Food Chemistry*, 81(2), 175-187.
- Bertram, H. C., & ERSEN, H. J. (2004). Applications of NMR in meat science. *Annual Reports on NMR Spectroscopy*, 53, 157-202.
- Bhatt, R. S., & Sahoo, A. (2017). Effect of feeding complete feed block containing rumen protected protein, non-protein nitrogen and rumen protected fat on improving body condition and carcass traits of cull ewes. *Journal of Animal Physiology and Animal Nutrition*.
- Bhatt, R. S., Karim, S. A., Sahoo, A., & Shinde, A. K. (2013a). Growth performance of lambs fed diet supplemented with rice bran oil as such or as calcium soap. *Asian-Australasian Journal of Animal Sciences*, 26(6), 812-819.
- Bhatt, R. S., Sahoo, A., Karim, S. A., & Gadekar, Y. P. (2016). Effects of *Saccharomyces cerevisiae* and rumen bypass-fat supplementation on growth, nutrient utilisation, rumen fermentation and carcass traits of lambs. *Animal Production Science*.
- Bhatt, R. S., Sahoo, A., Shinde, A. K., & Karim, S. A. (2013b). Change in body condition and carcass characteristics of cull ewes fed diets supplemented with rumen bypass fat. *Livestock Science*, 157(1), 132-140.
- Bhatt, R. S., Sahoo, A., Shinde, A. K., & Karim, S. A. (2015). Effect of calcium salt of fatty acids supplementation on performance of Malpura lambs. *Animal Production Science*, 55(9), 1123-1130.

- Bhatt, R. S., Soren, N. M., Tripathi, M. K., & Karim, S. A. (2011). Effects of different levels of coconut oil supplementation on performance, digestibility, rumen fermentation and carcass traits of Malpura lambs. *Animal Feed Science and Technology*, 164(1), 29-37.
- Bianchi, A. E., Macedo, V. P., França, R. T., Lopes, S. T., Lopes, L. S., Stefani, L. M., ... & Da Silva, A. S. (2014). Effect of adding palm oil to the diet of dairy sheep on milk production and composition, function of liver and kidney, and the concentration of cholesterol, triglycerides and progesterone in blood serum. *Small Ruminant Research*, 117(1), 78-83.
- Bindel, D. J., Drouillard, J. S., Titgemeyer, E. C., Wessels, R. H., & Löest, C. A. (2000). Effects of ruminally protected choline and dietary fat on performance and blood metabolites of finishing heifers. *Journal of Animal Science*, 78(10), 2497-2503.
- Bitman, J., Dryden, L. P., Goering, H. K., Wrenn, T. R., Yoncoskie, R. A., & Edmondson, L. F. (1973). Efficiency of transfer of polyunsaturated fats into milk. *Journal of American Oil Chemical Society*, 50, 93-98.
- Black, M. M. (2003). Micronutrient deficiencies and cognitive functioning. *The Journal of Nutrition*, 133(11), 3927S-3931S.
- Block, E., Chalupa, W., Evans, E., Jenkins, T., Moate, P., Palmquist, D., & Sniffen, C. (2005). Calcium salts are highly digestible. *Feedstuffs*, 77(30), 20-25.
- Borys, B., Borys, A., & Gasior, R. (2004). Effect of feeding rapeseed and linseed diets and their supplementation with vitamin E on health quality of lamb meat. *Archiv Fur Tierzucht*, 47(6; SPI), 189-197.
- Brooker, J., Lum, D., Miller, S., Skene, I., & O'Donovan, L. (1995). Rumen microorganisms as providers of high-quality protein. *Livestock Research for Rural Development*, 6, 1-6.
- Buccioni, A., Decandia, M., Minieri, S., Molle, G., & Cabiddu, A. (2012). Lipid metabolism in the rumen: New insights on lipolysis and biohydrogenation with an emphasis on the role of endogenous plant factors. *Animal Feed Science and Technology*, 174(1), 1-25.
- Byers, F. M. & Schelling, G. T. (1993). Lipids in ruminant nutrition. In D. C., Church (eds.) *The Ruminant Animal: Digestive Physiology and Nutrition*. Page 298-312. Waveland Press, Inc., Prospect Heights, IL.
- Calsamiglia, S., Cardozo, P. W., Ferret, A., & Bach, A. (2008). Changes in rumen microbial fermentation are due to a combined effect of type of diet and pH. *Journal of Animal Science*, 86, 702-711.
- Candyrine, S. C. L., Jahromi, M. F., Ebrahimi, M., Liang, J. B., Goh, Y. M., & Abdullah, N. (2016). In vitro rumen fermentation characteristics of goat and sheep supplemented with polyunsaturated fatty acids. *Animal Production Science*, 57(8), 1607-1612.

- Carlson, G.L. (1996). *Large Animal Internal Medicine*. In: Mosby-Year book, St. Louis, pp. 441-469.
- Carriquiry, M., Weber, W. J., Baumgard, L. H., & Crooker, B. A. (2008). *In vitro* biohydrogenation of four dietary fats. *Animal Feed Science and Technology*, 141(3), 339-355.
- Castejón, D., García-Segura, J. M., Escudero, R., Herrera, A., & Cambero, M. I. (2015). Metabolomics of meat exudate: Its potential to evaluate beef meat conservation and ageing. *Analytica Chimica Acta*, 901, 1-11.
- Castro, T., Manso, T., Mantecón, A., Guirao, J., & Jimeno, V. (2005). Fatty acid composition and carcass characteristics of growing lambs fed diets containing palm oil supplements. *Meat Science*, 69, 757-764.
- Chalupa, W., Vecchiarelli, B., Elser, A. E., Kronfeld, D. S., Sklan, D., & Palmquist, D. L. (1986). Ruminal fermentation *in vivo* as influenced by long-chain fatty acids. *Journal of Dairy Science*, 69(5), 1293-1301.
- Chang, J. H., Lunt, D. K., & Smith, S. B. (1992). Fatty acid composition and fatty acid elongase and stearoyl-CoA desaturase activities in tissues of steers fed high oleate sunflower seed. *Journal of Nutrition*, 122, 2074-2080.
- Chapinal, N., Carson, M. E., LeBlanc, S. J., Leslie, K. E., Godden, S., Capel, M., ... & Duffield, T. F. (2012). The association of serum metabolites in the transition period with milk production and early-lactation reproductive performance. *Journal of Dairy Science*, 95(3), 1301-1309.
- Chen, F., Zhu, Y., Dong, X., Liu, L., Huang, L., & Dai, X. (2010). Lignocellulose degrading bacteria and their genes encoding cellulase/hemicellulase in rumen-- A review. *Acta Microbiologica Sinica*, 50, 981-987.
- Cheng, K. J., Fay, J. P., Howarth, R. E., & Costerton, J. W. (1980). Sequence of events in the digestion of fresh legume leaves by rumen bacteria. *Applied and Environmental Microbiology*, 40(3), 613-625.
- Chiang, V. S. C., & Quek, S. Y. (2015). The relationship of red meat with cancer: Effects of thermal processing and related physiological mechanisms. *Critical Reviews in Food Science and Nutrition*, 57(6), 1153-1173.
- Chilliard, Y. (1993). Dietary fat and adipose tissue metabolism in ruminants, pigs, and rodents: A review. *Journal of Dairy Science*, 76, 3897-3931.
- Chilliard, Y., Glasser, F., Ferlay, A., Bernard, L., Rouel, J., & Doreau, M. (2007). Diet, rumen biohydrogenation and nutritional quality of cow and goat milk fat. *European Journal of Lipid Science and Technology*, 109(8), 828-855.
- Chouinard, P. Y., Girard, V., & Brisson, G. J. (1997). Lactational response of cows to different concentrations of calcium salts of canola oil fatty acids with or without bicarbonates. *Journal of Dairy Science*, 80(6), 1185-1193.

- Chouinard, P. Y., Girard, V., & Brisson, G. J. (1998). Fatty acid profile and physical properties of milk fat from cows fed calcium salts of fatty acids with varying unsaturation. *Journal of Dairy Science*, 81(2), 471-481.
- Cianzio, D. S., Topel, D. G., Whitehurst, G. B., Beitz, D. C., & Self, H. L. (1985). Adipose tissue growth and cellularity: changes in bovine adipocyte size and number. *Journal of Animal Science*, 60(4), 970-976.
- Cloete, S. W. P., Snyman, M. A., & Herselman, M. J. (2000). Productive performance of Dorper sheep. *Small Ruminant Research*, 36(2), 119-135.
- Coppock, C. E., & Wilks, D. L. (1991). Supplemental fat in high-energy rations for lactating cows: effects on intake, digestion, milk yield, and composition. *Journal of Animal Science*, 69(9), 3826-3837.
- Costa, D. P. B. D., Roça, R. D. O., Costa, Q. P. B. D., Lanna, D. P. D., Lima, E. D. S., & Barros, W. M. D. (2013). Meat characteristics of Nellore steers fed whole cottonseed. *Revista Brasileira de Zootecnia*, 42(3), 183-192.
- Čuperlović-Culf, M. (2015). Metabolomics in Animal Cell Culture. In *Animal Cell Culture* (pp. 615-646). Springer International Publishing.
- Daniel, M. (2015). Effects of different sources of fat (calcium soap) on carcass quality and fatty acid profile of muscle and adipose tissues in lambs, *Analele Universității din Oradea, Fascicula: Ecotoxicologie, Zootehnie și Tehnologii de Industrie Alimentară Vol. XIV/A*, 363–372.
- Devendra, C., & Lewis, D. (1974). The interaction between dietary lipids and fibre in the sheep 2. Digestibility studies. *Animal Science*, 19, 67-76.
- De Waal, H. O., & Combrinck, W. J. (2000). The development of the Dorper, its nutrition and a perspective of the grazing ruminant on veld. *Small Ruminant Research*, 36(2), 103-117.
- Degen, A. A., & Kam, M. (1992). Body mass loss and body fluid shifts during dehydration in Dorper sheep. *The Journal of Agricultural Science*, 119(3), 419-422.
- Demirel, G., Wachira, A. M., Sinclair, L. A., Wilkinson, R. G., Wood, J. D., & Enser, M. (2004). Effects of dietary n-3 polyunsaturated fatty acids, breed and dietary vitamin E on the fatty acids of lamb muscle, liver and adipose tissue. *British Journal of Nutrition*, 91(04), 551-565.
- Department of Health and Social Security (1984). Diet and cardiovascular disease. Report on health and social subjects, n. 28. London: HMSO (134 pp.)
- Department of Standards Malaysia (2009). MS1500: 2009 (1st revision) Halal food production, preparation, handling and storage-general guideline (pp. 1–13).

- DePeters, E. J., Getachew, G., Fadel, J. G., Zinn, R. A., Taylor, S. J., Pareas, J. W., Hinders, R. G. & Aseltine, M. S. (2003). *In vitro* gas production as a method to compare fermentation characteristics of steam-flaked corn. *Animal Feed Science and Technology*, 105(1), 109-122.
- Deutschmann, K., Phatsara, C., Sorachakula, C., Vearasilp, T., Phunphiphat, W., Cherdthong, A., Gerlach, K. & Südekum, K. H. (2017). *In vitro* gas production and *in vivo* nutrient digestibility and growth performance of Thai indigenous cattle fed fresh and conserved pangola grass. *Italian Journal of Animal Science*, 1-9.
- Devasena, B., Ravi, A., Ramana, J. V., & Prasad, J. R. (2007). Prospective Role of Protected Fat in Ruminants. *Intas Polivet*, 8(2), 300-307.
- Dinh, T. T., Thompson, L. D., Galyean, M. L., Brooks, J. C., Patterson, K. Y., & Boylan, L. M. (2011). Cholesterol content and methods for cholesterol determination in meat and poultry. *Comprehensive Reviews in Food Science and Food Safety*, 10(5), 269-289.
- Doreau, M., & Chilliard, Y. (1997). Digestion and metabolism of dietary fat in farm animals. *British Journal of Nutrition*, 78(01), S15-S35.
- Doreau, M., & Ferlay, A. (1994). Digestion and utilisation of fatty acids by ruminants. *Animal Feed Science and Technology*, 45, 379-396.
- Dror, D. K., & Allen, L. H. (2011). The importance of milk and other animal-source foods for children in low-income countries. *Food and Nutrition Bulletin*, 32(3), 227-243.
- Duarte, I. F., Marques, J., Ladeirinha, A. F., Rocha, C., Lamego, I., Calheiros, R., Silva, T. M., Marques M. P. M., Melo, J. B., Carreira, I. M. & Gil, A. M. (2009). Analytical approaches toward successful human cell metabolome studies by NMR spectroscopy. *Analytical Chemistry*, 81(12), 5023-5032.
- Dunn, W. B., & Ellis, D. I. (2005). Metabolomics: current analytical platforms and methodologies. *Trends in Analytical Chemistry*, 24(4), 285-294.
- Dutta, T. K., Agnihotri, M. K., & Rao, S. B. N. (2008). Effect of supplemental palm oil on nutrient utilization, feeding economics and carcass characteristics in post-weaned Muzaffarnagari lambs under feedlot condition. *Small Ruminant Research*, 78(1), 66-73.
- DVS. (2015). Department of Veterinary Services, Malaysia; Livestock products statistics.
- Ebrahimi, M. (2012). Production of omega-3 polyunsaturated fatty acid enriched chevon using treated oil palm (*Elaeis guineensis* jacq.) fronds. PhD thesis, Universiti Putra Malaysia. Retrieved on 12 January 2017 from <http://www.lib.upm.edu.my/>.

- Ebrahimi, M., Rajion, M. A., & Goh, Y. M. (2014). Effects of oils rich in linoleic and  $\alpha$ -linolenic acids on fatty acid profile and gene expression in goat meat. *Nutrients*, 6, 3913-3928.
- Ekeren, P. A., Smith, D. R., Lunt, D. K., & Smith, S. B. (1992). Ruminal biohydrogenation of fatty acids from high-oleate sunflower seeds. *Journal of Animal Science*, 70(8), 2574-2580.
- Elmeddah, Y., Doreau, M., & Michalet-Doreau, B. (1991). Interaction of lipid supply and carbohydrates in the diet of sheep with digestibility and ruminal digestion. *The Journal of Agricultural Science*, 116(3), 437-445.
- Enjalbert, F., Nicot, M. C., Griess, D., Vernay, M., & Moncoulon, R. (1994). Effect of different forms of polyunsaturated fatty acids on duodenal and serum fatty acid profiles in sheep. *Canadian Journal of Animal Science*, 74(4), 595-600.
- FAO. (2015b). *FAO Food Outlook 2015: Biannual Report on Global Food Markets*. Food and Agriculture Organization of the United Nations.
- FAO. (2015a). *FAO Statistical Pocketbook 2015: World Food and Agriculture*. Rome: Food and Agriculture Organization of the United Nations.
- FAO. (2017). Food and Agriculture Organization of the United Nations. Retrieved on 6 September 2017 from [http://www.fao.org/ag/againfo/themes/en/animal\\_production.html](http://www.fao.org/ag/againfo/themes/en/animal_production.html)
- FAOSTAT. (2017). Food and Agriculture Organization of the United Nations. Retrieved on 12 September 2017 from <http://www.fao.org/faostat/en/#home>
- Ferguson, J. D., Sklan, D., Chalupa, W. V., & Kronfeld, D. S. (1990). Effects of hard fats on *in vitro* and *in vivo* rumen fermentation, milk production, and reproduction in dairy cows. *Journal of Dairy Science*, 73(10), 2864-2879.
- Fay, J. P., Jakober, K. D., Cheng, K. J., & Costerton, J. W. (1990). Esterase activity of pure cultures of rumen bacteria as expressed by the hydrolysis of p-nitrophenyl palmitate. *Canadian Journal of Microbiology*, 36(8), 585-589.
- Fennema, O. R. (1990). Comparative water holding properties of various muscle foods. *Journal of Muscle Foods*, 1(4), 363-381.
- Fennema, O. R. (1996). *Food Chemistry*, 3<sup>rd</sup> Edition. Chapter 2, Pp. 17-94. USA: Marcel Dekker Publishing.
- Ferreira, E. M., Pires, A. V., Susin, I., Gentil, R. S., Parente, M. O. M., Nolli, C. P., ... & Ribeiro, C. V. D. M. (2014). Growth, feed intake, carcass characteristics, and meat fatty acid profile of lambs fed soybean oil partially replaced by fish oil blend. *Animal Feed Science and Technology*, 187, 9-18.
- Fievez, V., Vlaeminck, B., Jenkins, T., Enjalbert, F., & Doreau, M. (2007). Assessing rumen biohydrogenation and its manipulation *in vivo*, *in vitro* and *in situ*. *European Journal of Lipid Science and Technology*, 109(8), 740-756.

- Folch, J., Lees, M., & Sloane-Stanley, G. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*, 226, 497-509.
- Friedewald, W. T., Levy, R. I., & Fredrickson, D. S. (1972). Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clinical Chemistry*, 18(6), 499-502.
- Funston, R. N. (2004). Fat supplementation and reproduction in beef females. *Journal of Animal Science*, 82(13\_suppl), E154-E161.
- Garcia, I. F. F., Almeida, A. K. D., Costa, T. I. R., Leopoldino Júnior, I., Ribeiro, J. D. S., & Souza, F. A. D. (2010). Carcass characteristics and cuts of Santa Inês lambs fed different roughage proportions and fat source. *Revista Brasileira de Zootecnia*, 39(6), 1322-1327.
- Garg, M. R. (1997). Bypass Fat Production Using Acid Oil, Its Effect on *In Vitro* Rumen Fermentation and Effect of Its Feeding on In Sacco DM Disappearance in Sheep. *Asian-Australasian Journal of Animal Sciences*, 10(6), 571-574.
- GASL. (2014). Towards Sustainable Livestock. Discussion Document. Rome: Food and Agriculture Organization of the United Nations.
- Garrett, W. N., Yang, Y. T., Dunkley, W. L., & Smith, L. M. (1976). Energy utilization, feedlot performance and fatty acid composition of beef steers fed protein encapsulated tallow or vegetable oils. *Journal of Animal Science*, 42(6), 1522-1533.
- Getachew, G., DePeters, E. J., & Robinson, P. H (2008). Gas production provides effective method for assessing ruminant feeds. *California Agriculture*, 58(1): 54–58.
- Getachew, G., DePeters, E., Robinson, P., & Taylor, S. (2001). *In vitro* rumen fermentation and gas production: Influence of yellow grease, tallow, corn oil and their potassium soaps. *Animal Feed Science and Technology*, 93, 1-15.
- Getachew, G., Robinson, P. H., DePeters, E. J. & Taylor, S. J. (2004). Relationships between chemical composition, dry matter degradation and *in vitro* gas production of several ruminant feeds. *Animal Feed Science and Technology*, 111(1-4): 57–71.
- Ghani, A. A. A., Rusli, N. D., Shahudin, M. S., Goh, Y. M., Zamri-Saad, M., Hafandi, A., & Hassim, H. A. (2017). Utilisation of oil palm fronds as ruminant feed and its effect on fatty acid metabolism. *Pertanika Journal of Tropical Agricultural Science*, 40(2).
- Gilbert, C. D., Lunt, D. K., Miller, R. K., & Smith, S. B. (2003). Carcass, sensory, and adipose tissue traits of Brangus steers fed casein-formaldehyde-protected starch and/or canola lipid. *Journal of Animal Science*, 81(10), 2457-2468.

- Goldansaz, S. A., Guo, A. C., Sajed, T., Steele, M. A., Plastow, G. S., & Wishart, D. S. (2017). Livestock metabolomics and the livestock metabolome: A systematic review. *PloS One*, 12(5), e0177675.
- Gómez, I., Mendizabal, J. A., Sarriés, M. V., Insausti, K., Albertí, P., Realini, C., Juan, M. P., Oliver, M. A., Purroy, A. & Beriain, M. J. (2015). Fatty acid composition of young Holstein bulls fed whole linseed and rumen-protected conjugated linoleic acid-enriched diets. *Livestock Science*, 180, 106-112.
- Gooden, J. M., The role of fat in ruminant nutrition (1977). *Applied Biochemistry Division, D.S.I.R., Palmerston North, N.Z*  
URL: <http://livestocklibrary.com.au/handle/1234/19333>
- Gowda, N. K. S., Manegar, A., Raghavendra, A., Verma, S., Maya, G. P. A. L. D. T., Pal, D. T. & Sampath, K. T. (2013). Effect of protected fat supplementation to high yielding dairy cows in field condition. *Animal Nutrition and Feed Technology*, 13, 125-130.
- Griinari, J., & Bauman, D. E. (1999). Biosynthesis of conjugated linoleic acid and its incorporation into meat and milk in ruminants. *Advances in Conjugated Linoleic Acid Research*, 1, 180-200.
- Grosse, S. D. (1998). Farm Animals and Children's Nutritional Status in Rural Rwanda. *Presentation at the Symposium on Human Nutrition and Livestock*, October 14, 1998, Little Rock, Arkansas: Heifer Project International.
- Grummer, R. R. (1988). Influence of prilled fat and calcium salt of palm oil fatty acids on ruminal fermentation and nutrient digestibility. *Journal of Dairy Science*, 71(1), 117-123.
- Günal, M., Pinski, B., & AbuGhazaleh, A. A. (2017). Evaluating the effects of essential oils on methane production and fermentation under *in vitro* conditions. *Italian Journal of Animal Science*, 1-7.
- Haaland, G. L., Matsushima, J. K., Johnson, D. E., & Ward, G. M. (1981). Effect of replacement of corn by protected tallow in a cattle finishing diet on animal performance and composition. *Journal of Animal Science*, 52(4), 696-702.
- Haddad, S. G., & Younis, H. M. (2004). The effect of adding ruminally protected fat in fattening diets on nutrient intake, digestibility and growth performance of Awassi lambs. *Animal Feed Science and Technology*, 113(1), 61-69.
- Hailemariam, D., Mandal, R., Saleem, F., Dunn, S. M., Wishart, D. S., & Ametaj, B. N. (2014). Identification of predictive biomarkers of disease state in transition dairy cows. *Journal of Dairy Science*, 97(5), 2680-2693.
- Hannula, T., & Puolanne, E. (2004). The effect of cooling rate on beef tenderness: The significance of pH at 7 C. *Meat Science*, 67(3), 403-408.
- Harfoot, C. G. (1978). Lipid metabolism in the rumen. *Progress in Lipid Research*, 17(1), 21-54.

- Harfoot, C. G., & Hazlewood, G. P. (1997). Lipid metabolism in the rumen. In P. N. Honson & C. S. Stewart (Ed.), *The Rumen Microbial Ecosystem* (pp. 382-426). Netherlands: Springer
- Hartati, L., Agus, A., Widyobroto, B. P., & Yusiaty, L. M. (2012). *In Vitro Digestibilities of Six Rumen Protected Fat-Protein Supplement Formulas*. *Animal Production*, 14(1).
- Healy, P. J. (1974). Serum alkaline phosphatase activity in sheep. *Australian Journal of Experimental Biology and Medical Science*, 52, 375-385.
- Henderson, C. (1971). A study of the lipase produced by *Anaerovibrio lipolytica*, a rumen bacterium. *Microbiology*, 65(1), 81-89.
- Hess, B. W., Moss, G. E., & Rule, D. C. (2008). A decade of developments in the area of fat supplementation research with beef cattle and sheep. *Journal of Animal Science*, 86(14\_suppl), E188-E204.
- Hidayah, N., Wirawan, K. G., & Suharti, S. (2014). *In vitro* rumen fermentation of ration supplemented with protected vegetable oils. *Media Peternakan*, 37(2), 129.
- Hightshoe, R. B., Cochran, R. C., Corah, L. R., Harmon, D. L., & Vanzant, E. S. (1991). Influence of source and level of ruminal-escape lipid in supplements on forage intake, digestibility, digesta flow, and fermentation characteristics in beef cattle. *Journal of Animal Science*, 69(12), 4974-4982.
- Hill, G. M., & West, J. W. (1991). Rumen protected fat in kline barley or corn diets for beef cattle: digestibility, physiological, and feedlot responses. *Journal of Animal Science*, 69(8), 3376-3388.
- HLPE. (2016). Sustainable agricultural development for food security and nutrition: What roles for livestock? A report by the High-Level Panel of Experts on Food Security and Nutrition of the Committee on World Food Security, Rome. URL <http://www.fao.org/cfs/cfs-hlpe/en/>
- Hood, R. L., & Allen, C. E. (1973). Cellularity of bovine adipose tissue. *Journal of Lipid Research*, 14(6), 605-610.
- Hristov, A. N., Ivan, M., & McAllister, T. A. (2004). *In vitro* effects of individual fatty acids on protozoal numbers and on fermentation products in ruminal fluid from cattle fed a high-concentrate, barley-based diet. *Journal of Animal Science*, 82(9), 2693-2704.
- Huff-Lonergan, E., & Lonergan, S. M. (2005). Mechanisms of water-holding capacity of meat: The role of *postmortem* biochemical and structural changes. *Meat Science*, 71, 194-204.
- Iannotti, L. L., Zavaleta, N., Huasaquiche, C., Leon, Z., & Caulfield, L. E. (2015). Early growth velocities and weight gain plasticity improve linear growth in Peruvian infants. *Maternal & Child Nutrition*, 11(1), 127-137.

- Ivan, M., Mir, P. S., Koenig, K. M., Rode, L. M., Neill, L., Entz, T., & Mir, Z. (2001). Effects of dietary sunflower seed oil on rumen protozoa population and tissue concentration of conjugated linoleic acid in sheep. *Small Ruminant Research*, 41(3), 215-227.
- Ivan, M., Petit, H., Chiquette, J., & Wright, A. (2013). Rumen fermentation and microbial population in lactating dairy cows receiving diets containing oilseeds rich in C-18 fatty acids. *British Journal of Nutrition*, 109, 1211-1218.
- Jackson, P., & Cockcroft, P. (2008). *Clinical Examination of Farm Animals*. John Wiley & Sons. Appendix 3 Laboratory Reference Values: Biochemistry, pp. 303-305.
- Jaeger, S. M. P. L., Dutra, A. R., Pereira, J. C., & Oliveira, I. D. (2004). Características da carcaça de bovinos de quatro grupos genéticos submetidos a dietas com ou sem adição de gordura protegida. *Revista Brasileira de Zootecnia*, 33(6), 1876-1887.
- Jalč, D., Čertík, M., Kundríková, K., & Kubelková, P. (2009). Effect of microbial oil and fish oil on rumen fermentation and metabolism of fatty acids in artificial rumen. *Czech Journal of Animal Science*, 54, 229-237.
- Jama, N., Muchenje, V., Chimonyo, M., Strydom, P., Dzama, K., & Raats, J. (2008). Cooking loss components of beef from Nguni, bonsmara and Angus steers. *African Journal of Agricultural Research*, 3, 416-420.
- Jamaludin, M. H., Hassan, M. H., Amin, M. R., & Zulhisyam, A. K. (2014). The future of the Malaysian beef industry. *Journal of Tropical Resources and Sustainable Science*, 2, 23-29.
- Jaworska, D., Czauderna, M., Przybylski, W., & Rozbicka-Wieczorek, A. J. (2016). Sensory quality and chemical composition of meat from lambs fed diets enriched with fish and rapeseed oils, carnosic acid and seleno-compounds. *Meat Science*, 119, 185-192.
- Jenkins, T. C. (1993). Lipid metabolism in the rumen. *Journal of Dairy Science*, 76(12), 3851-3863.
- Jenkins, T. C., & Bridges, W. C. (2007). Protection of fatty acids against ruminal biohydrogenation in cattle. *European Journal of Lipid Science and Technology*, 109(8), 778-789.
- Jenkins, T. C., & Fotouhi, N. (1990). Effects of lecithin and corn oil on site of digestion, ruminal fermentation and microbial protein synthesis in sheep. *Journal of Animal Science*, 68(2), 460-466.
- Jenkins, T. C., & Jenny, B. F. (1992). Nutrient digestion and lactation performance of dairy cows fed combinations of prilled fat and canola oil. *Journal of Dairy Science*, 75(3), 796-803.

- Jenkins, T. C., & Palmquist, D. L. (1984). Effect of fatty acids or calcium soaps on rumen and total nutrient digestibility of dairy rations. *Journal of Dairy Science*, 67(5), 978-986.
- Jenkins, T. C., Gimenez, T., & Cross, D. L. (1989). Influence of phospholipids on ruminal fermentation *in vitro* and on nutrient digestion and serum lipids in sheep. *Journal of Animal Science*, 67(2), 529-537.
- Jenkins, T. C., Wallace, R. J., Moate, P. J., & Mosley, E. E. (2008). Board-invited review: Recent advances in biohydrogenation of unsaturated fatty acids within the rumen microbial ecosystem. *Journal of Animal Science*, 86(2), 397-412.
- Jeong, D. W., Choi, Y. M., Lee, S. H., Choe, J. H., Hong, K. C., Park, H. C., & Kim, B. C. (2010). Correlations of trained panel sensory values of cooked pork with fatty acid composition, muscle fibre type, and pork quality characteristics in Berkshire pigs. *Meat Science*, 86(3), 607-615.
- Jian, G. A. O., Wang, M. Z., Jing, Y. J., Sun, X. Z., Wu, T. Y., & Shi, L. F. (2016). Impacts of the unsaturation degree of long-chain fatty acids on the volatile fatty acid profiles of rumen microbial fermentation in goats *in vitro*. *Journal of Integrative Agriculture*, 15(12), 2827-2833.
- Jouany, J. P. (1996). Effect of rumen protozoa on nitrogen utilization by ruminants. *The Journal of Nutrition*, 126(4S), 1335S.
- Juarez, M., Aldai, N., López-Campos, Ó., Dugan, M., Uttaro, B., & Aalhus, J. (2012). Beef texture and juiciness. In Y. H. Hui (Ed.), *Handbook of Meat and Meat Processing* (pp.177-206). New York: CRC Press.
- Kadim, I. T., Mahgoub, O., Al-Ajmi, D., Al-Maqbaly, R., Al-Saqri, N., & Ritchie, A. (2004). An evaluation of the growth, carcass and meat quality characteristics of Omani goat breeds. *Meat Science*, 66, 203-210.
- Kaluza, J., Åkesson, A., & Wolk, A. (2015). Long-term processed and unprocessed red meat consumption and risk of heart failure: A prospective cohort study of women. *International Journal of Cardiology*, 193, 42-46.
- Kamra, D. N. (2005). Rumen microbial ecosystem. *Current Science*, 124-135.
- Kaneko, J. J., Harvey, J. W., & Bruss, M. L. (Eds.). (2008). *Clinical biochemistry of domestic animals*. Academic Press. San Diego, London, Boston, New York, Sydney, Tokyo, Toronto. pp. 303-325.
- Kang, S., & Wanapat, M. (2013). Using plant source as a buffering agent to manipulating rumen fermentation in an *in vitro* gas production system. *Asian-Australasian Journal of Animal Sciences*, 26, 1424.
- Kang, S., Wanapat, M., & Viennasay, B. (2016a). Supplementation of banana flower powder pellet and plant oil sources on *in vitro* ruminal fermentation, digestibility, and methane production. *Tropical Animal Health and Production*, 48(8), 1673-1678.

- Kang, S., Wanapat, M., Phesatcha, K., Norrapoke, T., Foiklang, S., Ampapon, T., & Phesatcha, B. (2016b). Using krabok (*Irvingia malayana*) seed oil and *Flemingia macrophylla* leaf meal as a rumen enhancer in an *in vitro* gas production system. *Animal Production Science*, 57(2), 327-333.
- Kannan, G., Gadiyaram, K., Galipalli, S., Carmichael, A., Kouakou, B., Pringle, T., & Gelaye, S. (2006). Meat quality in goats as influenced by dietary protein and energy levels, and *postmortem* ageing. *Small Ruminant Research*, 61, 45-52.
- Karisa, B. K., Thomson, J., Wang, Z., Bruce, H. L., Plastow, G. S., & Moore, A. S. (2013). Candidate genes and biological pathways associated with carcass quality traits in beef cattle. *Canadian Journal of Animal Science*, 93(3), 295-306.
- Karisa, B. K., Thomson, J., Wang, Z., Li, C., Montanholi, Y. R., Miller, S. P., Moore, S. S. & Plastow, G. S. (2014). Plasma metabolites associated with residual feed intake and other productivity performance traits in beef cattle. *Livestock Science*, 165, 200-211.
- Khoo, S. H. G., & Al- Rubeai, M. (2007). Metabolomics. An emerging tool for understanding metabolic systems. In: Al-Rubeai M., Fussenegger M. (Eds) Systems Biology. *Cell Engineering*, vol 5. Springer, Dordrecht
- Khorasani, G. R., De Boer, G., Robinson, P. H., & Kennelly, J. J. (1992). Effect of canola fat on ruminal and total tract digestion, plasma hormones, and metabolites in lactating dairy cows. *Journal of Dairy Science*, 75(2), 492-501.
- Kilic, U. and Garipoglu, A.V. (2009). *In situ* rumen degradability, *in vitro* digestibility and *in vitro* gas production of full-fat canola seeds. *Asian Journal of Animal and Veterinary Advances* 4(4):200-208.
- Kim, S. C., Adesogan, A. T., Badingga, L., & Staples, C. R. (2007). Effects of dietary n-6: n-3 fatty acid ratio on feed intake, digestibility, and fatty acid profiles of the ruminal contents, liver, and muscle of growing lambs. *Journal of Animal Science*, 85(3), 706-716.
- Klusmeyer, I. H., Lynch, G. L., Clark, J. H., & Nelson, D. R. (1991). Effects of Calcium Salts of Fatty Acids and Protein Source on Ruminal Fermentation and Nutrient Flow to Duodenum of Cows. *Journal of Dairy Science*, 74(7), 2206-2219.
- Kodani, Y., Miyakawa, T., Komatsu, T., & Tanokura, M. (2017). NMR-based metabolomics for simultaneously evaluating multiple determinants of primary beef quality in Japanese Black cattle. *Scientific Reports*, 7.
- Koike, S., and Kobayashi, Y. (2001). Development and use of competitive PCR assays for the rumen cellulolytic bacteria: *Fibrobacter succinogenes*, *Ruminococcus albus* and *Ruminococcus flavefaciens*. *FEMS Microbiology Letters*, 204(2), 361-366.

- Kongmun, P., Wanapat, M., Pakdee, P., & Navanukraw, C. (2010). Effect of coconut oil and garlic powder on *in vitro* fermentation using gas production technique. *Livestock Science*, 127, 38-44.
- Kott, R. W., Hatfield, P. G., Bergman, J. W., Flynn, C. R., Van Wagoner, H., & Boles, J. A. (2003). Feedlot performance, carcass composition, and muscle and fat CLA concentrations of lambs fed diets supplemented with safflower seeds. *Small Ruminant Research*, 49(1), 11-17.
- Lau, C. H. (2014). Exploring novel aspects of choline phospholipid metabolism in cancer using metabolomics (Doctoral dissertation, Imperial College London).
- Lahucky, R., Krska, P., Kuchenmeister, U., Nurnberg, K., Liptaj, T., Nurnberg, G., Bahelka, I., Demo, P., Kuhn, G. & Ender, K. (2000). Effect of Vitamin E on changes in Phosphorus Compounds assessed by P NMR Spectroscopy and ATPase from *postmortem* Muscle samples and Meat quality of Pigs. *Archiv Für Tierzucht, Dummerstorf*, 43(5), 487-497.
- Lane, D. J. (1991). 16S/23S rRNA sequencing. In 'Nucleic acid techniques in bacterial systematics'. (Eds E Stackebrandt, M Goodfellow) pp. 115–175.
- Lawrie, R. A. (1998). *Lawrie's meat science*, 6th edition. Headington Hill Hall, Oxford, England: Pergamon Press, 336 pp.
- Lawrie, R., & Ledward, D. (2006). *Lawrie's Meat Science*. Cambridge: Woodhead Publishing Ltd.
- Lee, Y. (2013). Effect of pH on conjugated linoleic acid (CLA) formation of linolenic acid biohydrogenation by ruminal microorganisms. *Journal of Microbiology*, 51(4), 471-476.
- Lee, Y. J., & Jenkins, T. C. (2011). Biohydrogenation of linolenic acid to stearic acid by the rumen microbial population yields multiple intermediate conjugated diene isomers. *The Journal of Nutrition*, 141(8), 1445-1450.
- Leo'n, A' .E., Olmos, M.C., Cruz, E., Garc'ia, R. (1999). Accumulation of body fat in Cuban Pelibuey lambs according to age and feeding level. *Archivos de Zootecnia*, 48, 219–222.
- Li, X. Z., Yan, C. G., Lee, H. G., Choi, C. W., & Song, M. K. (2012). Influence of dietary plant oils on mammary lipogenic enzymes and the conjugated linoleic acid content of plasma and milk fat of lactating goats. *Animal Feed Science and Technology*, 174(1), 26-35.
- Lima, E. S., Morais, J. P. G., Roça, R. O., Costa, Q. P. B., Andrade, E. N., Vaz, V. P., ... & Costa, D. P. B. (2015a). Meat characteristics of Nellore cattle fed different levels of lipid-based diets. *Journal of Agricultural Science*, 7(7), 174.

- Lima, E. S., Morais, J. P. G., Roça, R. O., Andrade, E. N., Valente, T. N. P., Costa, Q. P. B., & Deminicis, B. B. (2015b). Effect of different sources of lipids in diet on the qualitative characteristics of Longissimus thoracis muscle of cattle finished in feedlots. *African Journal of Agricultural Research*, 10(29), 2835-2840.
- Lima, E. S., Valente, T. N. P., Roca, R. O., Cezario, A. S., dos Santos, W. B. R., Deminicis, B. B., & Ribeiro, J. C. (2017). Effect of whole cottonseed or protected fat dietary additives on carcass characteristics and meat quality of beef cattle: A review. *Journal of Agricultural Science*, 9(5), 175.
- Lindon, J. C., Beckonert, O. P., Holmes, E., & Nicholson, J. K. (2009). High-resolution magic angle spinning NMR spectroscopy: Application to biomedical studies. *Progress in Nuclear Magnetic Resonance Spectroscopy*, 55(2), 79-100.
- Liu, S., Bu, D., Wang, J., Liu, L., Liang, S., Wei, H., & Loor, J. (2012). Effect of incremental levels of fish oil supplementation on specific bacterial populations in bovine ruminal fluid. *Journal of Animal Physiology and Animal Nutrition*, 96, 9-16.
- Loh, T. C. (2004). Livestock production and the feed industry in Malaysia. In *Animal Production and Health Proceedings (FAO)*.
- Long, N. M., Burns, T. A., Duckett, S. K., & Schafer, D. W. (2014). Reproductive performance and serum fatty acid profiles of underdeveloped beef heifers supplemented with saturated or unsaturated rumen bypass fat compared to an isocaloric control. *The Professional Animal Scientist*, 30(5), 502-509.
- Lough, D. S., Solomon, M. B., Rumsey, T. S., Kahl, S., & Slyter, L. L. (1993). Effects of high-forage diets with added palm oil on performance, plasma lipids, and carcass characteristics of ram and ewe lambs. *Journal of Animal Science*, 71(5), 1171-1176.
- Lourenço, M., Ramos-Morales, E., & Wallace, R. J. (2010). The role of microbes in rumen lipolysis and biohydrogenation and their manipulation. *Animal*, 4(07), 1008-1023.
- Maccarana, L., Cattani, M., Tagliapietra, F., Bailoni, L., & Schiavon, S. (2016). Influence of main dietary chemical constituents on the *in vitro* gas and methane production in diets for dairy cows. *Journal of Animal Science and Biotechnology*, 7(1), 54.
- Machmüller, A. (2006). Medium-chain fatty acids and their potential to reduce methanogenesis in domestic ruminants. *Agriculture, Ecosystems & Environment*, 112(2), 107-114.
- Machmüller, A., Ossowski, D., & Kreuzer, M. (2000). Comparative evaluation of the effects of coconut oil, oilseeds and crystalline fat on methane release, digestion and energy balance in lambs. *Animal Feed Science and Technology*, 85, 41-60.

- Machmüller, A., Soliva, C. R., & Kreuzer, M. (2003). Methane-suppressing effect of myristic acid in sheep as affected by dietary calcium and forage proportion. *The British Journal of Nutrition*, 90(3), 529.
- Maheri-Sis, N.M, Chamani Sadeghi, A.A., Mirza- Aghazadeh, A.G. and Abolfazl, (2008). Nutritional evaluation of kabuli and desi type chickpeas (*cicer arietinum* L.) for ruminants using *in vitro* gas production technique *African Journal of Biotechnology* 7(16): 2946-2951.
- Maia, M. R., Chaudhary, L. C., Bestwick, C. S., Richardson, A. J., McKain, N., Larson, T. R., Graham I, A. & Wallace, R. J. (2010). Toxicity of unsaturated fatty acids to the biohydrogenating ruminal bacterium, *Butyrivibrio fibrisolvens*. *BMC Microbiology*, 10(1), 52.
- Males, J. R., & Purser, D. B. (1970). Relationship between rumen ammonia levels and the microbial population and volatile fatty acid proportions in faunated and defaunated sheep. *Applied Microbiology*, 19(3), 485-490.
- Maltin, C., Balcerzak, D., Tilley, R., & Delday, M. (2003). Determinants of meat quality: tenderness. *Proceedings of the Nutrition Society*, 62(2), 337-347.
- Mancini, R. A., & Hunt, M. (2005). Current research in meat colour. *Meat Science*, 71(1), 100-121.
- Mangrum, K. S., Tuttle, G., Duckett, S. K., Sell, G. S., Krehbiel, C. R., & Long, N. M. (2016). The effect of supplementing rumen undegradable unsaturated fatty acids on marbling in early-weaned steers. *Journal of Animal Science*, 94(2), 833-844.
- Manso, T., Castro, T., Mantecón, A. R., & Jimeno, V. (2006). Effects of palm oil and calcium soaps of palm oil fatty acids in fattening diets on digestibility, performance and chemical body composition of lambs. *Animal Feed Science and Technology*, 127(3), 175-186.
- Mansor, R. (2012). *Proteomic and metabolomic studies on milk during bovine mastitis* (Doctoral dissertation, University of Glasgow).
- Mapiye, C., Vahmani, P., Mlambo, V., Muchenje, V., Dzama, K., Hoffman, L., & Dugan, M. (2015). The trans-octadecenoic fatty acid profile of beef: Implications for global food and nutrition security. *Food Research International*, 76, 992-1000.
- Marais, P.G., Schoeman, A. (1990). Geographic distribution of Dorper sheep in the republic. *Dorper News* 46, 4±5. Retrieved on 10 September, 2017 from <http://gadi.agric.za/articles/Agric/geographic.php>
- Mattos, R., Staples, C. R., & Thatcher, W. W. (2000). Effects of dietary fatty acids on reproduction in ruminants. *Reviews of Reproduction*, 5(1), 38-45.

- McAfee, A. J., McSorley, E. M., Cuskelly, G. J., Moss, B. W., Wallace, J. M., Bonham, M. P., & Fearon, A. M. (2010). Red meat consumption: An overview of the risks and benefits. *Meat Science*, 84(1), 1-13.
- McCarthy, R. D., Klusmeyer, T. H., Vicini, J. L., Clark, J. H., & Nelson, D. R. (1989). Effects of source of protein and carbohydrate on ruminal fermentation and passage of nutrients to the small intestine of lactating cows. *Journal of Dairy Science*, 72(8), 2002-2016.
- McCartor, M. M., & Smith, G. C. (1978). Effect of protected lipids on feedlot performance and carcass characteristics of short-fed steers. *Journal of Animal Science*, 47(1), 270-275.
- McDonald, P. R., Edward, A., Greenhalgh, J. F. D., Morgan, C. A., Sinclair L. A. and Wilkinson R. G. (2010). *Animal Nutrition* 7th (Ed) Longmans Scientific and Technological, John Wiley and Sons. Inc. New York.
- McPhee, M. J., Oltjen, J. W., Fadel, J. G., Perry, D., & Sainz, R. D. (2008). Development and evaluation of empirical equations to interconvert between twelfth-rib fat and kidney, pelvic, and heart fat respective fat weights and to predict initial conditions of fat deposition models for beef cattle. *Journal of Animal Science*, 86(8), 1984-1995.
- Melzer, N., Wittenburg, D., Hartwig, S., Jakubowski, S., Kesting, U., Willmitzer, L., Lisec, J., Reinsch, N. & Repsilber, D. (2013). Investigating associations between milk metabolite profiles and milk traits of Holstein cows. *Journal of Dairy Science*, 96(3), 1521-1534.
- Menke, K. H., & Steingass, H. (1988). Estimation of the energetic feed value obtained from chemical analysis and *in vitro* gas production using rumen fluid. *Animal Research and Development*, 28(1), 7-55.
- Mierlita, D. (2018). Effect of diets containing essential fatty acids-rich oil calcium soaps on functional lipid components of lamb tissues. *Romanian Biotechnological Letters*, (23)1, 13205-13213.
- Miller, M. F., Carr, M. A., Ramsey, C. B., Crockett, K. L., & Hoover, L. C. (2001). Consumer thresholds for establishing the value of beef tenderness. *Journal of Animal Science*, 79(12), 3062-3068.
- Milne, C. (2000). The history of the Dorper sheep. *Small Ruminant Research*, 36(2), 99-102.
- Mobeen, A., Riaz, M., & Yaqoob, M. U. (2017). Effect of by-pass fat supplementation on the performance of Sahiwal dairy cows. *International Journal of Agriculture & Biology*, 19(3).
- Modzelewska-Kapituła, M., Dąbrowska, E., Jankowska, B., Kwiatkowska, A., & Cierach, M. (2012). The effect of muscle, cooking method and final internal temperature on quality parameters of beef roast. *Meat Science*, 91(2), 195-202.

- Mohamed, Z.A. (2007). The Livestock Industry. In S. A. Idid Fatimah, M. A., Raja Abdullah, N. M., Kaur, B., Abdullah, A. M. (Eds.), *50 Years of Malaysian Agriculture: Transformation Issues, Challenges and Direction* (553-584). Selangor, Malaysia Universiti Putra Malaysia Press.
- Moss, A. R., Jouany, J. P., & Newbold, J. (2000, May). Methane production by ruminants: its contribution to global warming. In *Annales de Zootechnie* (Vol. 49, No. 3, pp. 231-253). EDP Sciences.
- Muchenje, V., Dzama, K., Chimonyo, M., Strydom, P. E., Hugo, A., & Raats, J. G. (2009). Some biochemical aspects pertaining to beef eating quality and consumer health: A review. *Food Chemistry*, 112(2), 279-289.
- Mudgal, V., Baghel, R. P. S., Ganie, A., & Srivastava, S. (2012). Effect of feeding bypass fat on intake and production performance of lactating crossbred cows. *Indian Journal of Animal Research*, 46(1).
- Nagaraja, T. G., Newbold, C. J., Van Nevel, C. J., & Demeyer, D. I. (1997). Manipulation of ruminal fermentation. In *The Rumen Microbial Ecosystem* (pp. 523-632). Springer Netherlands.
- Naik, P. K. (2013). Bypass fat in dairy ration-a review. *Animal Nutrition and Feed Technology*, 13(1), 147-63.
- Naik, P. K., Saijapaul, S., & Rani, N. (2007a). Evaluation of rumen protected fat prepared by fusion method. *Animal Nutrition and Feed Technology*, 7, 95-101.
- Naik, P. K., Saijapaul, S., & Rani, N. (2007b). Preparation of rumen protected fat and its effect on nutrient utilization in buffaloes. *Indian Journal of Animal Nutrition*, 24, 212-215.
- Naik, P. K., Saijapaul, S., & Rani, N. (2009). Effect of ruminally protected fat on *in vitro* fermentation and apparent nutrient digestibility in buffaloes (*Bubalus bubalis*). *Animal Feed Science and Technology*, 153(1), 68-76.
- Nangia, O. P., & Shrivastava, P. N. (1989). Effect of presence or absence of protozoa on rumen digestive functions in buffaloes on straw-based diets. *Asian-Australasian Journal of Animal Sciences*, 2, 493-496.
- Nestel, P. J., Havenstein, N., Whyte, H. M., Scott, T. W., & Cook, L. J. (1973). Lowering of plasma cholesterol and enhanced sterol excretion with the consumption of polyunsaturated ruminant fats. *New England Journal of Medicine*, 288, 279-282.
- Nestel, P. J., Poyser, A., Hood, R. L., Mills, S. C., Willis, M. R., Cook, L. J., & Scott, T. W. (1978). The effect of dietary fat supplements on cholesterol metabolism in ruminants. *Journal of Lipid Research*, 19(7), 899-909.

- Ngidi, M. E., Loerch, S. C., Fluharty, F. L., & Palmquist, D. L. (1990). Effects of calcium soaps of long-chain fatty acids on feedlot performance, carcass characteristics and ruminal metabolism of steers. *Journal of Animal Science*, 68(8), 2555-2565.
- Nicholson, J. K., Lindon, J. C., & Holmes, E. (1999). 'Metabonomics': understanding the metabolic responses of living systems to pathophysiological stimuli via multivariate statistical analysis of biological NMR spectroscopic data. *Xenobiotica*, 29(11), 1181-1189.
- Nor, N. A. A. M., & Rosali, M. H. (2015). The Development and Future Direction of Malaysia's Livestock Industry. Retrieved on 6 September 2017 from <http://ap.fftca.net/index.php>
- Nowak, B., Mueffling, T. V., & Hartung, J. (2007). Effect of different carbon dioxide concentrations and exposure times in stunning of slaughter pigs: Impact on animal welfare and meat quality. *Meat Science*, 75(2), 290-298.
- NRC, (2007). National Research Council. Nutrient requirements of small ruminants (6th ed.). Washington, D. C., USA: National Academy Press. Pp. 384.
- Ocak, S., Ogun, S., & Yilmaz, O. (2016). Dorper sheep utilizing feed resources efficiently: a Mediterranean case study. *Revista Brasileira de Zootecnia*, 45(8), 489-498.
- OECD/FAO. (2015). *OECD-FAO Agricultural Outlook 2015*. Paris: Organisation for Economic Co-operation and Development. URL [http://dx.doi.org/10.1787/agr\\_outlook-2015-en](http://dx.doi.org/10.1787/agr_outlook-2015-en)
- OECDSTAT. (2017) . OECD-FAO Agricultural Outlook 2017-2026. Retrieved on 6 September 2017 from <http://stats.oecd.org/viewhtml.aspx>
- Offer, G., & Knight, P. (1988). The structural basis of water-holding in meat part 2: Drip losses. In: Lawrie, R., (Ed), (Vol. 4): *Developments in Meat Science*. London: Elsevier Applied Science, pp.172-243.
- Offer, G., & Trinick, J. (1983). On the mechanism of water holding in meat: the swelling and shrinking of myofibrils. *Meat Science*, 8(4), 245-281.
- Ohajuruka, O. A., Wu, Z., & Palmquist, D. L. (1991). Ruminal Metabolism, Fiber, and Protein Digestion by Lactating Cows Fed Calcium Soap or Animal-Vegetable Fat1. *Journal of Dairy Science*, 74(8), 2601-2609.
- Olivecrona, N. (2015). *Investigation of the Confinement Odour Problem in Exported Lamb using NMR-based Metabolomics: A thesis presented in partial fulfilment of the requirements for the degree of Master of Science in Chemistry at Massey University, Manawatū, New Zealand* (Doctoral dissertation, Massey University) retrieved on 23 November 2017.

- Oliveira, E., Sampaio, A. A. M., Henrique, W., Pivaro, T., Rosa, B., Fernandes, A., & Andrade, A. (2012). Quality traits and lipid composition of meat from Nellore young bulls fed with different oils either protected or unprotected from rumen degradation. *Meat Science*, 90, 28-35.
- Oliver, S. G., Winson, M. K., Kell, D. B., & Baganz, F. (1998). Systematic functional analysis of the yeast genome. *Trends in Biotechnology*, 16(9), 373-378.
- Oltjen, R. R., & Dinius, D. A. (1975). Production practices that alter the composition of foods of animal origin. *Journal of Animal Science*, 41(2), 703-722.
- Ono, Y., Sorimachi, H., & Suzuki, K. (1998). Structure and physiology of calpain, an enigmatic protease. *Biochemical and Biophysical Research Communications*, 245(2), 289-294.
- Ørskov, E. R. (Ed.). (2012). *Energy Nutrition in Ruminants*. Springer Science & Business Media.
- Ørskov, E. R., & McDonald, I. (1979). The estimation of protein degradability in the rumen from incubation measurements weighted according to rate of passage. *The Journal of Agricultural Science*, 92(02), 499-503.
- Ørskov, E. R., & Ryle, M. (1990). *Energy Nutrition in Ruminants*. Essex: Elsevier Science Publishers Ltd.
- Pal, K., Patra, A. K., & Sahoo, A. (2015). Evaluation of feeds from tropical origin for *in vitro* methane production potential and rumen fermentation *in vitro*. *Spanish Journal of Agricultural Research*, 13(3), 0608.
- Palma, M., Scanlon, T., Kilminster, T., Milton, J., Oldham, C., Greeff, J., Matzapetakis, M. & Almeida, A. M. (2016). The hepatic and skeletal muscle ovine metabolomes as affected by weight loss: a study in three sheep breeds using NMR-metabolomics. *Scientific Reports*, 6.
- Palmquist, D. L. (1994). The role of dietary fats in efficiency of ruminants. *The Journal of Nutrition*, 124(8 Suppl), 1377S-1382S.
- Palmquist, D. L., & Jenkins, T. C. (1980). Fat in Lactation Rations: Review. *Journal of Dairy Science*, 63(1), 1-14.
- Palmquist, D. L., Beaulieu, A. D., & Barbano, D. M. (1993). Feed and animal factors influencing milk fat composition. *Journal of Dairy Science*, 76(6), 1753-1771.
- Palmquist, D. L., Lock, A. L., Shingfield, K. J., & Bauman, D. E. (2005). Biosynthesis of conjugated linoleic acid in ruminants and humans. *Advances in Food and Nutrition Research*, 50, 179-217.
- Pardun, H. (1988). *Die pflanzenlecithine: Gewinnung, eigenschaften, verarbeitung und anwendung planzlicher phosphatidpräparate*. Verlag für chemische industrie H. Ziolkowsky KG.

- Paredi, G., Raboni, S., Bendixen, E., de Almeida, A. M., & Mozzarelli, A. (2012). “Muscle to meat” molecular events and technological transformations: The proteomics insight. *Journal of Proteomics*, 75(14), 4275-4289.
- Park, B. K., Choi, N. J., Kim, H. C., Kim, T.I., Cho, Y. M., Oh, Y. K., Im, S. K., Kim, Y. J., Jang, J. S., Hwang, I. H., Jang, H. Y., Kim, J. B. & Kwon, E. G. (2010). Effects of amino acid-enriched ruminally protected fatty acids on plasma metabolites, growth performance and carcass characteristics of Hanwoo steers. *Asian-Australasian Journal of Animal Sciences*, 23 (8), 1013–1021
- Parvar, R., Ghoorchi, T., & Shargh, M. S. (2017). Influence of dietary oils on performance, blood metabolites, purine derivatives, cellulase activity and muscle fatty acid composition in fattening lambs. *Small Ruminant Research*, 150, 22-29.
- Pearce, K. L., Rosenvold, K., Andersen, H. J., & Hopkins, D. L. (2011). Water distribution and mobility in meat during the conversion of muscle to meat and ageing and the impacts on fresh meat quality attributes—A review. *Meat Science*, 89, 111-124.
- Pethick, D. W., Harper, G. S., & Oddy, V. H. (2004). Growth, development and nutritional manipulation of marbling in cattle: A review. *Animal Production Science*, 44, 705-715.
- Pethick, D., Barendse, W., Hocquette, J., Thompson, J., & Wang, Y. (2007). Regulation of marbling and body composition-growth and development, gene markers and nutritional biochemistry. *Publication-European Association for Animal Production*, 124, 75.
- Phillipson A T. 1982. Ruminant digestion. *Dukes Physiology of Domestic Animals*. 9th Edition, pp. 250. (Ed.) Swenson M J. Cornell Univ. Press, London.
- Pinto, A. P. P., Furusho-Garcia, I. F., Leopoldino Júnior, I., Olalquiaga Pérez, J. R., Alves, N. G., & Pereira, I. G. (2011). Performance and carcass characteristics of lambs fed diets with fat and vitamin E. *Revista Brasileira de Zootecnia*, 40(12), 2911-2921.
- Plascencia, A., Estrada, M., & Zinn, R. A. (1999). Influence of free fatty acid content on the feeding value of yellow grease in finishing diets for feedlot cattle. *Journal of Animal Science*, 77(10), 2603-2609.
- Prevolnik, M., Čandek-Potokar, M., & Škorjanc, D. (2010). Predicting pork water-holding capacity with NIR spectroscopy in relation to different reference methods. *Journal of Food Engineering*, 98(3), 347-352.
- Ponnampalam, E. N., Sinclair, A. J., Egan, A. R., Blakeley, S. J., Li, D., & Leury, B. J. (2001). Effect of dietary modification of muscle long-chain n-3 fatty acid on plasma insulin and lipid metabolites, carcass traits, and fat deposition in lambs. *Journal of Animal Science*, 79(4), 895-903.

- Pontoizeau, C. (2012). *High-Field NMR Metabolomics: Phenotyping the metabolic complexity from humans to cells* (Doctoral dissertation, Ecole Normale Supérieure de Lyon-ENS LYON).
- Prezioso, G., Russo, C., Casarosa, L., Campodon, G., Piloni, S., & Cianci, D. (1999). Effect of diet energy source on weight gain and carcass characteristics of lambs. *Small Ruminant Research*, 33(1), 9-15.
- Purushothaman, S., Kumar, A., & Tiwari, D. P. (2008). Effect of feeding calcium salts of palm oil fatty acids on performance of lactating crossbred cows. *Asian Australasian Journal of Animal Sciences*, 21(3), 376.
- Putnam, D., Garrett, J., & Kung, L. (2003). Evaluation key to use of rumen-stable encapsulates. *Feedstuffs*, 75, 10-12.
- Quistorff, B., Moesgaard, B., Larsen, I. E., Therkelsen, I., Christensen, V. G., & Jørgensen, P. F. (1993). Effect of dietary magnesium on *postmortem* phosphocreatine utilization in skeletal muscle of swine: a non-invasive study using <sup>31</sup>P-NMR spectroscopy. *Acta Veterinaria Scandinavica (Print Edition)*, 34(4), 397-404.
- Rahmani, M. G., Kamalyan, R. G., Dehghan-Banadaky, M. J., & Marmaryan, G. Y. (2012). The effect of oral administration of choline on some liver function characterized blood plasma enzymes of early lactating dairy cows. *Biological Journal of Armenia*, 64(3), 83-86.
- Rajion, M., McLean, J., & Cahill, R. N. (1985). Essential fatty acids in the fetal and newborn lamb. *Australian Journal of Biological Sciences*, 38, 33-40.
- Reddy, Y. R., Krishna, N., Rao, E. R., & Reddy, T. J. (2003). Influence of dietary protected lipids on intake and digestibility of straw-based diets in Deccani sheep. *Animal Feed Science and Technology*, 106(1), 29-38.
- Ribeiro Pereira, Luiz Gustavo, Machado, Fernanda S, Campos, Mariana M, Guimaraes Júnior, Roberto, Tomich, Thierry R, Reis, Larissa G, & Coombs, Cassius. (2015). Enteric methane mitigation strategies in ruminants: a review. *Revista Colombiana de Ciencias Pecuarias*, 28(2), 124-143.
- Ríos-Rincón, F. G., Estrada-Angulo, A., Plascencia, A., López-Soto, M. A., Castro-Pérez, B. I., Portillo-Loera, J. J., ... & Dávila-Ramos, H. (2014). Influence of protein and energy level in finishing diets for feedlot hair lambs: growth performance, dietary energetics and carcass characteristics. *Asian-Australasian Journal of Animal Sciences*, 27(1), 55.
- Rizzi, L., Simioli, M., Sardi, L., & Monetti, P. G. (2002). Carcass quality, meat chemical and fatty acid composition of lambs fed diets containing extruded soybeans and sunflower seeds. *Animal Feed Science and Technology*, 97(1), 103-114.

- Rogers, J. A., & Davis, C. L. (1982). Effects of intraruminal infusions of mineral salts on volatile fatty acid production in steers fed high-grain and high-roughage diets. *Journal of Dairy Science*, 65(6), 953-962.
- Rowe, A., Macedo, F. A. F., Visentainer, J. V., Souza, N. E., & Matsushita, M. (1999). Muscle composition and fatty acid profile in lambs fattened in dry lot or pasture. *Meat Science*, 51(4), 283-288.
- Roy, A., Mandal, G. P., & Patra, A. K. (2013). Evaluating the performance, carcass traits and conjugated linoleic acid content in muscle and adipose tissues of Black Bengal goats fed soybean oil and sunflower oil. *Animal Feed Science and Technology*, 185(1), 43-52.
- Roy, A., Mandal, G. P., & Patra, A. K. (2017). Effects of different vegetable oils on rumen fermentation and conjugated linoleic acid concentration *in vitro*. *Veterinary World*, 10(1), 11.
- Rozanski, S., Vivian, D. R., Kowalski, L. H., Prado, O. R., Fernandes, S. R., de Souza, J. C., & de Freitas, J. A. (2017). Carcass and meat traits and non-carcass components of lambs fed ration containing increasing levels of urea. *Semina: Ciências Agrárias*, 38(3), 1577-1594.
- Ruckebusch, Y., & Thivend, P. (1980). Digestive physiology and metabolism in ruminants. In *International Symposium on Ruminant Physiology. (USA)*. 1980..
- Rudel, L., & Morris, M. (1973). Determination of cholesterol using ophthalmaldehyde. *Journal of Lipid Research*, 14, 364-366.
- Russell, J. B., & Strobel, H. J. (2005). Microbial energetics. In J., Dijkstra, J. M., Forbes & J., France (Eds.). *Quantitative Aspects of Ruminant Digestion and Metabolism*. CABI Publishing. Wallingford, Oxfordshire, United Kingdom.
- Rymer, C., Huntington, J. A., Williams, B. A., & Givens, D. I. (2005). *In vitro* cumulative gas production techniques: History, methodological considerations and challenges. *Animal Feed Science and Technology*, 123, 9-30.
- Sabow, A. B., Sazili, A. Q., Zulkifli, I., Goh, Y. M., Kadir, M. Z. A. A., & Adeyemi, K. D. (2015). Physico-chemical characteristics of Longissimus lumborum muscle in goats subjected to halal slaughter and anaesthesia (halothane) pre-slaughter. *Animal Science Journal*, 86(12), 981-991.
- Sadler, K., & Catley, A. (2009). *Milk Matters: the role and value of milk in the diets of Somali pastoralist children in Liben and Shinile, Ethiopia*. Addis Ababa, Feinstein International Center.
- Saijpaul, S., Naik, P. K., & Rani, N. (2010). Effects of rumen protected fat on *in vitro* dry matter degradability of dairy rations. *Indian Journal of Animal Sciences*, 80(10), 993.

- Saleem, F., Ametaj, B. N., Bouatra, S., Mandal, R., Zebeli, Q., Dunn, S. M., & Wishart, D. S. (2012). A metabolomics approach to uncover the effects of grain diets on rumen health in dairy cows. *Journal of Dairy Science*, 95(11), 6606-6623.
- Saleem, F., Bouatra, S., Guo, A. C., Psychogios, N., Mandal, R., Dunn, S. M., Ametaj, B. N. & Wishart, D. S. (2013). The bovine ruminal fluid metabolome. *Metabolomics*, 9(2), 360-378.
- Salinas, J., Ramirez, R. G., Domínguez, M. M., Reyes-Bernal, N., Trinidad-Larraga, N., & Montano, M. F. (2006). Effect of calcium soaps of tallow on growth performance and carcass characteristics of Pelibuey lambs. *Small Ruminant Research*, 66(1), 135-139.
- Sallam, S. M. A . (2005). Nutritive value assessment of the alternative feed resources by gas production and rumen fermentation *in vitro*. *Research Journal of Agriculture and Biological Science*, 1(2): 200–209.
- Salter, A. M. (2013). Dietary fatty acids and cardiovascular disease. *Animal*, 7(s1), 163-171.
- Salvatori, G., Pantaleo, L., Di Cesare, C., Maiorano, G., Filetti, F., & Oriani, G. (2004). Fatty acid composition and cholesterol content of muscles as related to genotype and vitamin E treatment in crossbred lambs. *Meat Science*, 67(1), 45-55.
- Santos-Silva, J., Mendes, I. A., Portugal, P. V., & Bessa, R. J. B. (2004). Effect of particle size and soybean oil supplementation on growth performance, carcass and meat quality and fatty acid composition of intramuscular lipids of lambs. *Livestock Production Science*, 90(2), 79-88.
- Santra, A., Chaturvedi, O., Tripathi, M., Kumar, R., & Karim, S. (2003). Effect of dietary sodium bicarbonate supplementation on fermentation characteristics and ciliate protozoal population in rumen of lambs. *Small Ruminant Research*, 47, 203-212.
- SAS (2013). Statistical Analysis System package (SAS) Version 9.4 software. SAS Institute Inc., Cary, NC, USA
- Sazili, A. Q., Abdul Rahim, A. H., & Hilmi, M. (2008). A study on the effects of conditioning on shear force values and water holding capacity of different skeletal muscles in Malaysian indigenous (Malin) sheep. In *54th International Congress of Meat Science & Technology, Cape Town, South Africa*.
- Scarpino, F. B. O., Ezequiel, J. M. B., Silva, D. A. V., & van Cleef, E. H. C. B. (2014). Soybean oil and residual soybean oil in diets for feedlot sheep: blood parameters. *Archivos de Zootecnia*, 63(241), 207-210.
- Schauff, D. J., & Clark, J. H. (1989). Effects of Prilled Fatty Acids and Calcium Salts of Fatty Acids on Rumen Fermentation, Nutrient Digestibilities, Milk Production, and Milk Composition1. *Journal of Dairy Science*, 72(4), 917-927.

- Schauff, D. J., & Clark, J. H. (1992). Effects of Feeding Diets Containing Calcium Salts of Long-Chain Fatty Acids to Lactating Dairy Cows. *Journal of Dairy Science*, 75(11), 2990-3002.
- Schauff, D. J., Elliott, J. P., Clark, J. H., & Drackley, J. K. (1992). Effects of feeding lactating dairy cows diets containing whole soybeans and tallow. *Journal of Dairy Science*, 75(7), 1923-1935.
- Scheffler, T. L., Scheffler, J. M., Kasten, S. C., Sosnicki, A. A., & Gerrard, D. E. (2013). High glycolytic potential does not predict low ultimate pH in pork. *Meat Science*, 95(1), 85-91.
- Schmid, A., Collomb, M., Sieber, R., & Bee, G. (2006). Conjugated linoleic acid in meat and meat products: A review. *Meat Science*, 73(1), 29-41.
- Schmidely, P., Ghazal, S., & Berthelot, V. (2017). Effect of rumen-protected conjugated linoleic acid on ruminal biohydrogenation and transfer of fatty acids to milk in dairy goats. *Livestock Science*, 199, 7-13.
- Schoeman, S. J. (2000). A comparative assessment of Dorper sheep in different production environments and systems. *Small Ruminant Research*, 36(2), 137-146.
- Scollan, N. D., Choi, N. J., Kurt, E., Fisher, A. V., Enser, M., & Wood, J. D. (2001). Manipulating the fatty acid composition of muscle and adipose tissue in beef cattle. *British Journal of Nutrition*, 85(1), 115-124.
- Scollan, N. D., Dannenberger, D., Nuernberg, K., Richardson, I., MacKintosh, S., Hocquette, J., & Moloney, A. P. (2014). Enhancing the nutritional and health value of beef lipids and their relationship with meat quality. *Meat Science*, 97, 384-394.
- Scollan, N. D., Enser, M., Gulati, S. K., Richardson, I., & Wood, J. D. (2003). Effects of including a ruminally protected lipid supplement in the diet on the fatty acid composition of beef muscle. *British Journal of Nutrition*, 90(3), 709-716.
- Sebsibe, A. (2008). Sheep and goat meat characteristics and quality. *Sheep and Goat Production Handbook for Ethiopia. Ethiopian Sheep and Goats Productivity Improvement Program (ESGPIP)*, Addis Ababa, Ethiopia. pp323-328.
- Shelke, S. K., Thakur, S. S., & Shete, S. M. (2012). Productive and reproductive performance of Murrah buffaloes (*Bubalus bubalis*) supplemented with rumen protected fat and protein. *Indian Journal of Animal Nutrition*, 29(4), 317-323.
- Sigman, M., McDonald, M. A., Neumann, C., & Bwibo, N. (1991). Prediction of cognitive competence in Kenyan children from toddler nutrition, family characteristics and abilities. *Journal of Child Psychology and Psychiatry*, 32(2), 307-320.

- Shija, D. S., Mtenga, L. A., Kimambo, A. E., Laswai, G. H., Mushi, D. E., Mgheni, D. M., Mwilawa, A. J., Shirima E. J. & Safari, J. G. (2013). Preliminary Evaluation of Slaughter Value and Carcass Composition of Indigenous Sheep and Goats from Traditional Production System in Tanzania. *Asian-Australasian Journal of Animal Sciences*, 26(1), 143.
- Shingfield, K. J., Kairenus, P., Ärölä, A., Paillard, D., Muetzel, S., Ahvenjärvi, S., Vanhatalo, A., Huhtanen, P., Toivonen, V., Grinari, J.M. & Wallace, R. J. (2012). Dietary fish oil supplements modify ruminal biohydrogenation, alter the flow of fatty acids at the omasum, and induce changes in the ruminal Butyrivibrio population in lactating cows. *The Journal of Nutrition*, 142(8), 1437-1448.
- Sirohi, S. K., Walli, T. K., & Mohanta, R. K. (2010). Supplementation effect of bypass fat on production performance of lactating crossbred cows. *Indian Journal of Animal Sciences*, 80(8), 733.
- Slyter, L.L. (1976). Influence of acidosis on rumen function. *Journal of Animal Science* 43(4): 910-929.
- Snyder, T. J., Rogers, J. A., & Muller, L. D. (1983). Effects of 1.2% sodium bicarbonate with two ratios of corn silage: grain on milk production, rumen fermentation, and nutrient digestion by lactating dairy cows. *Journal of Dairy Science*, 66(6), 1290-1297.
- Soares, S. B., Furusho-Garcia, I. F., Pereira, I. G., Alves, D. D. O., Silva, G. R. D., Almeida, A. K. D., ... & Sena, J. A. B. (2012). Performance, carcass characteristics and non-carcass components of Texel× Santa Inês lambs fed fat sources and monensin. *Revista Brasileira de Zootecnia*, 41(2), 421-431.
- Solomon, M. B., Lynch, G. P., & Lough, D. S. (1992). Influence of dietary palm oil supplementation on serum lipid metabolites, carcass characteristics, and lipid composition of carcass tissues of growing ram and ewe lambs. *Journal of Animal Science*, 70(9), 2746-2751.
- Solomon, M. B., Lynch, G. P., Ono, K., & Paroczay, E. (1990). Lipid composition of muscle and adipose tissue from crossbred ram, wether and cryptorchid lambs. *Journal of Animal Science*, 68(1), 137-142.
- Solorzano, L. (1969). Determination of ammonia in natural waters by the phenolhypochlorite method. *Limnology and Oceanography*, 799–801
- Somarny, W. W., Erin, A. R., Suhaimi, A. H. M. S., Nurulhuda, M. O., & Hifzan, R. M. (2013). A study of major prolificacy genes in Malin and Dorper sheep in Malaysia. *Journal of Tropical Agriculture and Food Science*, 41(2), 265-272.
- Sontakke, U. B., Kaur, H., Tyagi, A., Kumar, M., & Saikh, A. H. (2014). Effect of feeding rice bran lysophospholipids and rumen protected fat on feed intake, nutrient utilization and milk yield in crossbred cows. *Indian J. Anim. Sci*, 84, 998-1003.

- St John, L. C., Young, C. R., Knabe, A. D., & Smith, S. B. (1987). FA profiles and sensory and carcass traits of tissues from steers and swine fed an elevated monosaturated fat diet. *Journal of Animal Science*, 64(5), 1441-1447.
- Stanstrup, J. (2014). *Metabolomics investigation of whey intake: Discovery of markers and biological effects supported by a computer-assisted compound identification pipeline* (Doctoral dissertation, Department of Nutrition, Exercise and Sports, Faculty of Science, University of Copenhagen).
- Staples, C. R., Burke, J. M., & Thatcher, W. W. (1998). Influence of Supplemental Fats on Reproductive Tissues and Performance of Lactating Cows. *Journal of Dairy Science*, 81(3), 856-871.
- Streeter, M. N., Hill, G. M., Wagner, D. G., Hibberd, C. A., & Owens, F. N. (1993). Chemical and physical properties and *in vitro* dry matter and starch digestion of eight sorghum grain hybrids and maize. *Animal Feed Science and Technology*, 44(1-2), 45-58.
- Suen, G., Stevenson, D. M., Bruce, D. C., Chertkov, O., Copeland, A., Cheng, J. F., Detter, C., Detter, J. C., Goodwin, L. A., Han, C. S., Hauser, L. J., Ivanova, N. N., Kyrides, N. C., Land, M. L., Lapidus, A., Lucas, S., Pitluck, S., Tapia, R., Woyke, T., Boyum, J., Mead, D. & Hauser, L. J. (2011). Complete genome of the cellulolytic ruminal bacterium *Ruminococcus albus* 7. *Journal of Bacteriology*, 193(19), 5574-5575.
- Sundekilde, U. K., Poulsen, N. A., Larsen, L. B., & Bertram, H. C. (2013). Nuclear magnetic resonance metabonomics reveals strong association between milk metabolites and somatic cell count in bovine milk. *Journal of Dairy Science*, 96(1), 290-299.
- Sutter, F., Casutt, M. M., Ossowski, D. A., Scheeder, M. R. L., & Kreuzer, M. (2000). Comparative evaluation of rumen-protected fat, coconut oil and various oilseeds supplemented to fattening bulls: 1. Effects on growth, carcass and meat quality. *Archives of Animal Nutrition*, 53(1), 1-23.
- Sylvester, J. T., Karnati, S. K., Yu, Z., Morrison, M., & Firkins, J. L. (2004). Development of an assay to quantify rumen ciliate protozoal biomass in cows using real-time PCR. *The Journal of Nutrition*, 134(12), 3378-3384.
- Szumacher-Strabel, M., Cieślak, A., & Nowakowska, A. (2009). Effect of oils rich in linoleic acid on *in vitro* rumen fermentation parameters of sheep, goats and dairy cows. *Journal of Animal and Feed Sciences*, 18, 440-452.
- Tangendjaja B., Santoso B. and Wina E. (1993). Protected fat: preparation and digestibility. *Proceedings of Workshop on Advances in Small Ruminant Research in Indonesia*, August 3-4, 1993, Ciawi, Indonesia, pp. 165-178.

- Terlouw, E. M. C., Arnould, C., Auperin, B., Berri, C., Le Bihan-Duval, E., Deiss, V., Lefevre, F., Leninski, B. J. & Mounier, L. (2008). Pre-slaughter conditions, animal stress and welfare: current status and possible future research. *Animal*, 2(10), 1501-1517.
- Thakur, S. S., & Shelke, S. K. (2010). Effect of supplementing bypass fat prepared from soybean acid oil on milk yield and nutrient utilization in Murrah buffaloes. *Indian Journal of Animal Sciences*, 80(4), 354-357.
- Top Farmer. (2017). *How to get value from the Dorper sheep*. Agriculture News. Top Farmer. Retrieved on 10 September 2017 from [www.topfarmer.co.ke](http://www.topfarmer.co.ke)
- Tshabalala, P. A., Strydom, P. E., Webb, E. C., & De Kock, H. L. (2003). Meat quality of designated South African indigenous goat and sheep breeds. *Meat Science*, 65(1), 563-570.
- Tyagi, N., Thakur, S. S., & Shelke, S. K. (2009). Effect of feeding bypass fat supplement on milk yield, its composition and nutrient utilization in crossbred cows. *Indian Journal of Animal Nutrition*, 26(1), 1-8.
- Tyagi, N., Thakur, S. S., & Shelke, S. K. (2010). Effect of bypass fat supplementation on productive and reproductive performance in crossbred cows. *Tropical Animal Health and Production*, 42(8), 1749-1755.
- Ugwuowo, L. C., Ezekwe, A. G., Ani, A. O., Eze, S. I., Anyanwu, C. N., & Ofomatah, A. (2015). The efficacy of palm oil sludge in reducing ruminal methanogenesis using rumen simulation technique. *African Journal of Biotechnology*, 14(42), 2937-2942.
- Valente, T. N. P., Sampaio, C. B., Lima, E. S., Deminicis, B. B., Cezário, A. S., & Santos, W. B. R. (2017). Aspects of acidosis in ruminants with a focus on nutrition: A review. *Journal of Agricultural Science*, 9(3), 90.
- Valente, T. N. P., Lima E. S., Santos, W. B. R., Cesario, A. S., Tavares, C. J., Fernandes, I. L. & Freitas, M. A. M. (2016). Ruminal microorganism consideration and protein used in the metabolism of the ruminants: A Review. *African Journal of Microbiology Research*, 10(14), 456-464.
- Vandoni, S. L., Dell'Orto, V., & Sgoifo Rossi, C. A. (2010). Effects of administration of three different by-pass lipids on growth performance, rumen activity and feeding behaviour of beef cattle. *Italian Journal of Animal Science*, 9(2), e44.
- Van Soest, P. V., Robertson, J. B., & Lewis, B. A. (1991). Methods for dietary fibre, neutral detergent fibre, and nonstarch polysaccharides in relation to animal nutrition. *Journal of Dairy Science*, 74(10), 3583-3597.
- Van Nevel, C., & Demeyer, D. I. (1995). Lipolysis and biohydrogenation of soybean oil in the rumen *in vitro*: inhibition by antimicrobials. *Journal of Dairy Science*, 78(12), 2797-2806.

- Van Nevel, C. J., & Demeyer, D. I. (1996). Influence of pH on lipolysis and biohydrogenation of soybean oil by rumen contents *in vitro*. *Reproduction Nutrition Development*, 36(1), 53-63.
- Vargas, J. E., Andres, S., Snelling, T. J., Lopez-Ferreras, L., Yanez-Ruiz, D. R., Garcia-Estrada, C., & Lopez, S. (2017). Effect of sunflower and marine oils on ruminal microbiota, *in vitro* fermentation and digesta fatty acid profile. *Frontiers in Microbiology*, 8, 1124.
- Varnam, A. H., & Sutherland, J. P. (1995). *Meat and Meat Products: Technology, Chemistry and Microbiology*. London: Chapman and Hall.
- Venturini, R. S., Carvalho, S., Pacheco, P. S., Pellegrin, A. C. R. S. D., Martins, A. A., Lopes, J. F., Moro, A. B. & Simões, R. R. (2017). Characteristics of carcass and of non-carcass components of lambs and hoggets fed high-concentrate corn or sorghum diets. *Revista Brasileira de Zootecnia*, 46(3), 257-263.
- Viant, M. R. (2007). Revealing the metabolome of animal tissues using <sup>1</sup>H nuclear magnetic resonance spectroscopy. *Metabolomics: Methods and Protocols*, 229-246.
- Vieira, C., Fernández-Diez, A., Mateo, J., Bodas, R., Soto, S., & Manso, T. (2012). Effects of addition of different vegetable oils to lactating dairy ewes' diet on meat quality characteristics of suckling lambs reared on the ewes' milk. *Meat Science*, 91(3), 277–283.
- Vogel, H. J., Lundberg, P., Fabiansson, S., Rudérus, H., & Tornberg, E. (1985). Postmortem energy metabolism in bovine muscles studied by non-invasive phosphorus-31 nuclear magnetic resonance. *Meat Science*, 13(1), 1-18.
- Voigt, J., Kuhla, S., Gaafar, K., Derno, M., & Hagemeister, H. (2006). Digestibility of rumen protected fat in cattle. *Slovak Journal of Animal Science*, 39, 16-19.
- Wadhwa, M., Grewal, R. S., Bakshi, M. P. S., & Brar, P. S. (2012). Effect of supplementing bypass fat on the performance of high yielding crossbred cows. *Indian Journal of Animal Sciences*, 82(2), 200.
- Wang, C., Liu, Q., Guo, G., Huo, W. J., Ma, L., Zhang, Y. L., Pei, C. X., Zhang, S. L. & Wang, H. (2016). Effects of rumen-protected folic acid on ruminal fermentation, microbial enzyme activity, cellulolytic bacteria and urinary excretion of purine derivatives in growing beef steers. *Animal Feed Science and Technology*, 221, 185-194.
- Ward, P. F. V., Scott, T. W., & Dawson, R. M. C. (1964). The hydrogenation of unsaturated fatty acids in the ovine digestive tract. *Biochemical Journal*, 92(1), 60.

- Warner, C. M., Hahm, S. W., Archibeque, S. L., Wagner, J. J., Engle, T. E., Roman-Muniz, I. N., Woerner, D., Sponsler, M. & Han, H. (2015). A comparison of supplemental calcium soap of palm fatty acids versus tallow in a corn-based finishing diet for feedlot steers. *Journal of Animal Science and Technology*, 57(1), 25.
- Warriss, P. D. (2010). *Meat Science: An Introductory Text*. Cambridge: CABI Publishing, Wallingford, Oxfordshire, United Kingdom.
- Watkins, S. M., & German, J. B. (1998). Omega Fatty Acids. In C. C. Akoh & D. B. Min (Eds.), *Food Lipids: Chemistry, Nutrition and Biotechnology* (pp. 463-493). New York, NY: Marcel Dekker Inc.
- Weikard, R., Altmaier, E., Suhre, K., Weinberger, K. M., Hammon, H. M., Albrecht, E., Setoguchi, K., Takasuga, A. & Kühn, C. (2010). Metabolomic profiles indicate distinct physiological pathways affected by two loci with major divergent effect on Bos taurus growth and lipid deposition. *Physiological Genomics*, 42(2), 79-88.
- Wettstein, H. R., Quarella Forni, M. G., Kreuzer, M., & Sutter, F. (2000). Influence of plant lecithin partly replacing rumen-protected fat on digestion, metabolic traits and performance of dairy cows. *Journal of Animal Physiology and Animal Nutrition*, 84(5), 165-177.
- Wettstein, H. R., Scheeder, M. R., Sutter, F., & Kreuzer, M. (2001). Effect of lecithins partly replacing rumen-protected fat on fatty acid digestion and composition of cow milk. *European Journal of Lipid Science and Technology*, 103(1), 12-22.
- Widiawati, Y. & Thalib, A (2009). Comparison of fermentation kinetics (*in vitro*) of grass and shrub legume leaves: The pattern of VFA concentration, estimated CH<sub>4</sub> and microbial biomass production. *Indonesian Journal of Agriculture* 2(1): 21-27.
- Williams, G. L., & Stanko, R. L. (2000). Dietary fats as reproductive nutraceuticals in beef cattle. *Journal of Animal Science*, 77(E-Suppl), 1-12.
- Wilson, J.R. and Mertens, D.R. (1995). Cell wall accessibility and cell structure limitations to microbial digestion of forage. *Crop Science* 35(1): 251-259.
- Wishart, D. S. (2009). Computational strategies for metabolite identification in metabolomics. *Bioanalysis*. 2009; 1: 1579-1596. PMID: 21083105.
- Wood, J. D., Enser, M., Fisher, A. V., Nute, G. R., Sheard, P. R., Richardson, R. I., Hughes, S. I. & Whittington, F. M. (2008). Fat deposition, fatty acid composition and meat quality: A review. *Meat Science*, 78(4), 343-358.
- Wood, J. D., Richardson, R. I., Nute, G. R., Fisher, A. V., Campo, M. M., Kasapidou, E., Sheard, P. R., & Enser, M. (2004). Effects of fatty acids on meat quality: a review. *Meat Science*, 66(1), 21-32.

- World Health Organization. (2015). Carcinogenicity of consumption of red and processed meat. Report of the International Agency for Research on cancer. Press Release no 240, Lyon France, October 26, 2015.
- Wu, Z., Ohajuruka, O. A., & Palmquist, D. L. (1991). Ruminal Synthesis, Biohydrogenation, and Digestibility of Fatty Acids by Dairy Cows1. *Journal of Dairy Science*, 74(9), 3025-3034.
- Yap, C. Y., & Aw, T. C. (2010). Liver function tests (LFts). *Proceedings of Singapore Healthcare*, 19(1), 80-82.
- Yousuf, M. B., Adeloye, A. A., Okukpe, K. M., Adeyemi, K. D., & Ogundun, N. J. (2014). Influence of dietary sunflower (*tithonia diversifolia*) leaf extracts on performance characteristics of goats fed cassava peeling wastes-based diet. *Journal of Agricultural Technology*, 10, 59-65.
- Zahari, M. W., & Wong, H. K. (2009). Research and development on animal feed in Malaysia. *WARTAZOA. Indonesian Bulletin of Animal and Veterinary Sciences*, 19(4), 172-179.
- Zezza, A., Pica-Ciamarra, U., Mugera, H. K., Mwisomba, T., & Okello, P. (2016). Measuring the Role of Livestock in the Household Economy. *The Living Standards Measurement Study (LSMS) Guidebook*. URL <http://www.fao.org/3/a-i6739e.pdf>
- Zhou, M. I., & Hernandez-Sanabria, E. (2009). Assessment of the microbial ecology of ruminal methanogens in cattle with different feed efficiencies. *Applied and Environmental Microbiology*, 75(20), 6524-6533.
- Zeisel, S. H. & Blusztajn, J. K. (1994). Choline and human nutrition. *Annual Review of Nutrition 1994*; 14: 269-271.
- Zimmerman, H. J., Dujovne, C. A., & Levy, R. (1968). The correlation of serum levels of two transaminases with tissue levels in six vertebrate species. *Comparative Biochemistry and Physiology*, 25(3), 1081-1089.
- Zinn, R. A. (1989). Influence of level and source of dietary fat on its comparative feeding value in finishing diets for steers: feedlot cattle growth and performance. *Journal of Animal Science*, 67(4), 1029-1037.
- Zinn, R. A., & Plascencia, A. (1996). Effects of forage level on the comparative feeding value of supplemental fat in growing-finishing diets for feedlot cattle. *Journal of Animal Science*, 74(6), 1194-1201.
- Zinn, R. A., Gulati, S. K., Plascencia, A., & Salinas, J. (2000). Influence of ruminal biohydrogenation on the feeding value of fat in finishing diets for feedlot cattle. *Journal of Animal Science*, 78(7), 1738-1746.