



UNIVERSITI PUTRA MALAYSIA

***SELENIUM-ENRICHED BACTERIAL PROTEIN AS A SOURCE OF
ORGANIC SELENIUM IN BROILER CHICKENS***

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By

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

April 2018

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DEDICATION

This work is dedicated to those who motivated and helped me throughout my study

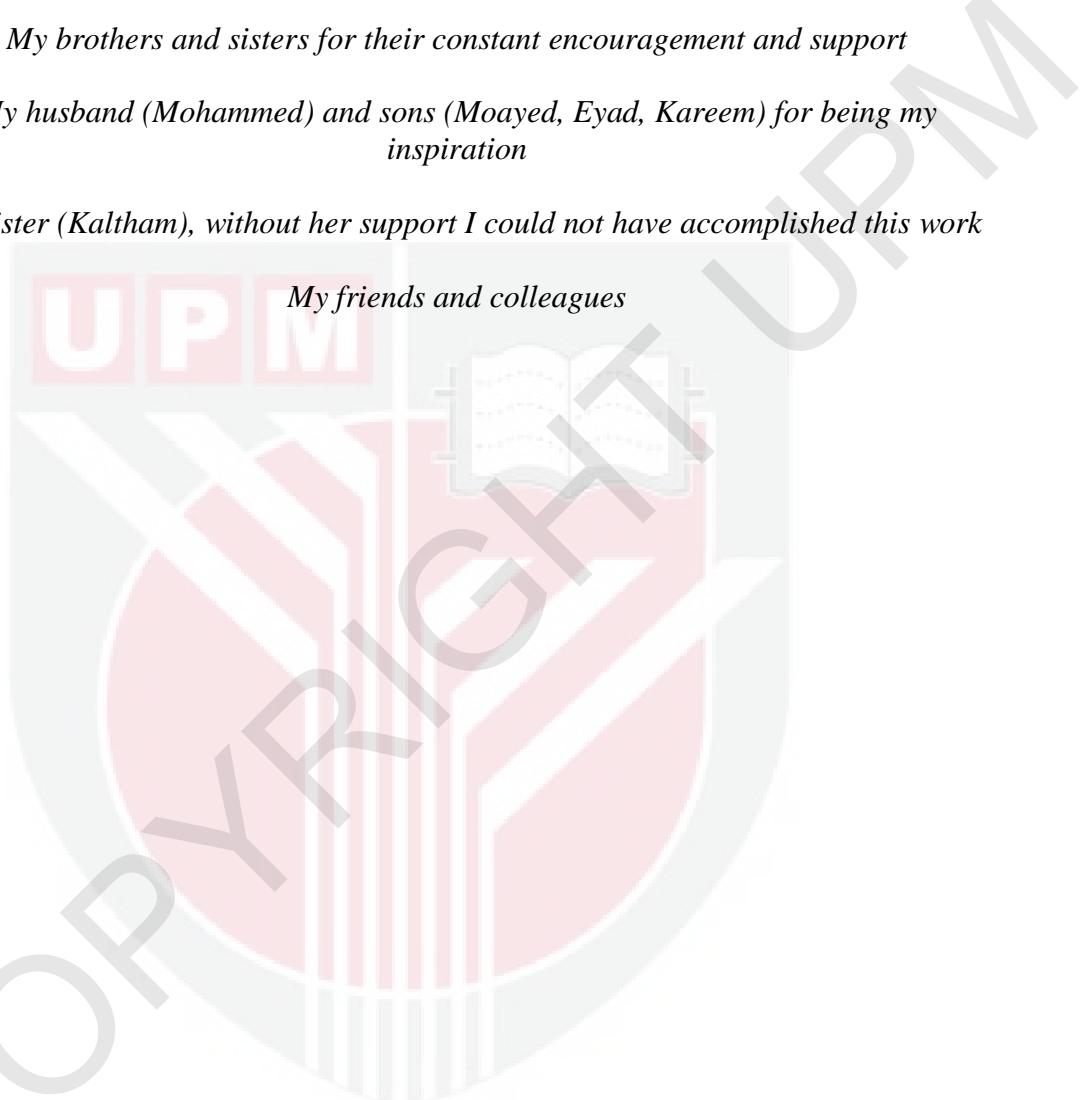
My father and mother who always prays for my success

My brothers and sisters for their constant encouragement and support

My husband (Mohammed) and sons (Moayed, Eyad, Kareem) for being my inspiration

My sister (Kaltham), without her support I could not have accomplished this work

My friends and colleagues



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of the requirement for the degree of Doctor of Philosophy

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April 2018

Chairman : Associate Professor Anjas Asmara @ Ab. Hadi Samsudin, PhD
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Selenium (Se) is an essential dietary trace mineral associated with several important roles in the biological processes. Livestock are routinely supplemented with various Se sources and many studies have indicated that organic Se is more efficient and bioavailable than inorganic sodium selenite (SS). Some bacterial strains have the ability to produce organic selenium biologically through selenite microbial reduction and accumulate it in their cells as Se-containing proteins. The possibility of using bacterial organic Se as a feed supplement may provide an interesting source of organic Se for animals and also humans through the food chain. Hence, this study aimed to identify a potential new product of bacterial organic Se and show the possibility of using it in animals' feed to improve animals' Se status and antioxidant status.

In the first experiment of this study, four isolates, identified as *Enterobacter cloacae* (ADS1, ADS7, ADS11), and *Klebsiella pneumoniae* (ADS2) from rumen fluid origin, and one isolate from hot spring water (*Stenotrophomonas maltophilia* (ADS18), were able to transform 47.17- 62.52% of the absorbed inorganic Se into organic form (Se-containing proteins). Organic Se-containing proteins in all the selected strains showed *in vitro* antioxidant properties, therefore, these strains may offer a potential source of organic Se due to their Se-tolerant nature and higher biomass organic to inorganic Se ratio.

The second experiment examined the influence of bacterial organic Se on growth performance, meat quality and meat antioxidant, immunity and selenoproteins gene expression in broiler chickens. A total of 180, day-old, broiler chicks were randomly assigned to 5 groups: T1, basal diet; T2; basal diet + 0.3 mg/kg SS; T3, basal diet + 0.3 mg/kg ADS1-Se; T4, basal diet + 0.3 mg/kg ADS2-Se; T5; basal diet + 0.3 mg/kg

ADS18-Se. Different sources of bacterial Se or SS, increased the birds' survival rate, serum and tissues antioxidants and Se accumulation, improved meat's water holding capacity, and up-regulated some selenoproteins mRNA. Birds fed bacterial Se-protein showed high intestinal villus height, better Se retention with higher tissues Se deposition, better anti-oxidative capacity and meat tenderness with lower serum and tissues TBARS level more than SS. Selenium of ADS18 had a superior action in improving Se retention, antioxidant system and expression of selenoproteins compared to ADS1 and ADS2 bacterial Se.

The third experiment examined the effects of the combination of Vit E with bacterial organic Se and inorganic Se on growth performance, meat quality, immunity and antioxidant status of broiler chickens. A total of 216, day-old chicks were randomly assigned to six dietary treatments: T1, basal diet; T2, basal diet + 100 mg/kg α -tocopherol acetate; T3, Basal diet + 0.3 mg/Kg SS; T4, Basal diet + 0.3 mg /kg ADS18-Se; T5, Basal diet + 0.3 mg /kg SS+ 100 mg/kg α -tocopherol acetate; T6, 0.3 mg/kg ADS18-Se + 100 mg/kg α -tocopherol acetate. Supplementation of ADS18-Se in broiler chickens improved the performance, antioxidant system, meat quality, caecum microbiota, and immunity response more than SS. Moreover, adding of 100 mg/kg of Vit E to the diet resulted in better performance, meat quality, and meat fatty acid modulation, with no effect on caecum microbial population. The synergistic effect of Se and Vit E appear clearly in the performance, antioxidant system, and immunity response. The supplementation of Vit E alone had a better effect on most of the antioxidant and immunity parameters than the combination with Se, while the combination with bacterial organic Se had better effect than the combination with SS. Moreover, the inclusion of 100 mg/kg Vit E with 0.3 mg/kg ADS18-Se, effectively could support performance, antioxidant system, and immunity more than using ADS18-Se alone, while no difference between using SS alone or combined with Vit E.

In conclusion, sodium selenite and bacterial organic Se of ADS1, ADS2, and ADS18 could enhance broiler biological activities, however, bacterial organic sources were more efficient, and can be used to produce Se-enriched meat. Moreover, inclusion of Vit E with bacterial organic Se could improve it's efficiency.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PROTEIN BAKTERIA YANG DIPERKAYAKAN DENGAN SELENIUM SEBAGAI SUMBER ORGANIK SELENIUM UNTUK AYAM PEDAGING

Oleh

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Selenium (Se) merupakan mineral surih pemakanan penting yang berkaitan dengan beberapa peranan penting dalam proses biologi. Secara rutin, ternakan diberi suplemen daripada pelbagai sumber Se, dan banyak kajian menunjukkan bahawa Se organik lebih efisien dan tersedia daripada segi biologi berbanding sodium selenite (SS) yang tidak organik. Sesetengah strain bakteria mempunyai keupayaan untuk menghasilkan selenium organik secara biologi melalui pengurangan mikroba selenit dan mengumpulnya dalam sel mereka sebagai protein yang mengandungi Se. Kebarangkalian menggunakan Se organik bakteria sebagai suplemen makanan dapat memberikan sumber organik Se yang menarik untuk haiwan dan manusia melalui rantai makanan. Maka, kajian ini bertujuan untuk mengenal pasti potensi satu produk baru Se organik bakteria dan menunjukkan kemungkinan untuk menggunakan其nya dalam makanan haiwan untuk menambah baik status Se haiwan.

Dalam eksperimen pertama kajian ini, empat asingan, yang dikenali sebagai *Enterobacter cloacae* (ADS1, ADS7, ADS11), dan *Klebsiella pneumoniae* (ADS2) daripada bendalir rumen, dan satu asingan daripada air mata air panas (*Stenotrophomonas maltophilia* (ADS18) untuk mengubah 47.17- 62.52% daripada Se tidak organik yang diserap, kepada organik (protein yang mengandungi Se). Protein organik yang mengandungi Se dalam semua strain terpilih menunjukkan sifat antioksidan *in vitro*. Maka, strain tersebut berpotensi menjadi sumber Se organik kerana mampu menahan Se, dan mempunyai nisbah biojisim Se organik yang tinggi berbanding Se tidak organik.

Eksperimen kedua meneliti pengaruh bakteria organik Se pada prestasi pertumbuhan, kualiti dan antioksidan daging, imuniti, dan selenoprotein ekspresi gen dalam ayam pedaging. Sebanyak 180 anak ayam pedaging berusia 1 hari, dibahagikan kepada 5 kumpulan secara rawak: T1, diet asas; T2; diet asas + 0.3 mg / kg SS; T3, diet asas + 0.3 mg / kg ADS1-Se; T4, diet asas + 0.3 mg / kg ADS2-Se; T5; diet asas + 0.3 mg / kg ADS18-Se. Sumber yang berbeza bagi Se dan SS bakteria meningkatkan kadar kelangsungan hidup burung, serum dan tisu antioksidan, dan pengumpulan Se meningkatkan kapasiti pengekalan air daging tersebut, serta “upregulated” sesetengah selenoprotein mRNA. Burung yang diberi makan bakteria organik Se menunjukkan vilus usus yang lebih tinggi, pengekalan Se yang lebih baik dengan tisu yang lebih tinggi, pengendapan Se, kapasiti anti-oksidatif yang lebih baik dan kelembutan daging dengan serum yang lebih rendah dan tahap tisu TBARS yang tinggi berbanding SS. Selenium ADS18 mempunyai hasil yang lebih baik dalam meningkatkan pengekalan Se, sistem antioksidan dan ekspresi selenoprotein, berbanding dengan ADS1 dan ADS2 Se bakteria.

Eksperimen ketiga mengkaji kesan-kesan gabungan vitamin E dengan bakteria Se organik dan Se tidak organik dalam prestasi pertumbuhan, kualiti daging, imuniti dan status antioksidan ayam pedaging. Sejumlah 216 anak ayam berusia 1 hari dibahagikan secara rawak kepada 6 rawatan pemakanan: T1, diet asas; T2, diet asas + 100 mg / kg α -tokoferol asetat; T3, diet asas + 0.3 mg / kg SS; T4, diet asas + 0.3 mg / kg ADS18-Se; T5, diet asas + 0.3 mg / kg SS + 100 mg / kg α -tocopherol asetat; T6, 0.3 mg / kg ADS18-Se + 100 mg / kg α -tokoferol asetat. Suplemen ADS18-Se bagi ayam pedaging meningkatkan prestasi, sistem antioksidan, kualiti daging, mikrobiota ceacum, dan tindak balas imuniti lebih daripada SS. Selain itu, penambahan 100 mg / kg vit E ke atas diet meningkatkan prestasi, kualiti daging dan modulasi asid lemak daging tanpa sebarang kesan pada populasi mikrob ceacum. Kesan bersinergi Se dan vit E jelas kelihatan dalam prestasi, sistem antioksidan, dan tindak balas imuniti. Suplemen vit E dengan sendiri mempunyai kesan yang lebih baik pada kebanyakan parameter antioksidan dan imuniti berbanding dengan gabungan bersama Se, sementara kombinasi dengan Se bakteria mempunyai kesan yang lebih baik berbanding kombinasi dengan SS. Selain itu, kemasukan 100 mg / kg vit E dengan 0.3 mg / kg ADS18-Se, dapat menyokong prestasi, sistem antioksidan, dan imuniti lebih daripada menggunakan ADS18-Se semata-mata, sementara SS sahaja atau gabungan dengan vit E, tidak membawa apa-apa perbezaaan.

Kesimpulannya, natrium selenit dan bakteria Se dari ADS1, ADS2 dan ADS18 boleh meningkatkan aktiviti biologi pedaging, namun, sumber bakteria organik jauh lebih cekap, dan boleh digunakan untuk menghasilkan daging diperkaya. Selain itu, kemasukan vit E dengan bakteria Se boleh juga meningkatkan kecekapan bakteria Se.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

µg	Microgram
µL	Microliter
µM	Micromole
a*	Yelloness
ADG	Average daily gain
ADS1	<i>Enterobacter cloacae</i> bacterial strain
ADS2	<i>Klebsiella pneumonia</i> bacterial strain
ADS18	<i>Stenotrophomonas maltophilia</i> bacterial strain
A/G:	Albumin/ globulin ratio
AL	Albumin
ALT	Alanine aminotransferase
AST	Aspartate aminotransferase
b*	Redness
BUN	Blood urea nitrogen
BW	Body weight
BWG	Body weight gain
CAT	Catalase
cDNA	Complementary-DNA
CFU	Colony forming units
Cm	Centimeter
CP	Crude protein
Cr	Creatinine

CRD	Complete randomized design
DIO1	Iodothyronine deiodinase 1
DIO2	Iodothyronine deiodinase 2
DNA	Deoxyribonucleic acid
EDTA	Ethylenediaminetetraacetic acid
FA	Fatty acids
FCR	Feed conversion ratio
FI	Feed intake
G	Gram
G	Globulin
GAPDH	Glyceraldehyde-3-phosphate Dehydrogenase
GLM	General linear model
GSH-Px	Glutathione peroxidase
HB	Hemoglobin
HDL-C	High density lipoprotein cholesterol
HNO ₃	Nitric acid
H ₂ O ₂	Hydrogen peroxide
H.S	Hot spring water
HSe	Selenide
ICP.MS	Inductively coupled plasma mass spectrometer
IFN- γ	Interferon gamma
IgA	Immunoglobulin A
IgG	Immunoglobulin G

IgM	Immunoglobulin M
IL	Interleukin
kg	Kilogram
L*	Lightness
L	Liter
LDH	Lactate dehydrogenase
LDL-C	Low-density lipoprotein cholesterol
MDA	Malondialdehyde
mL	Milliliter
Mm	Millimeter
mM	Millimolar
MUSFA	Mono-unsaturated fatty acids
mRNA	Messenger ribonucleic acid
NADPH	Nicotinamide phosphate adenine dinucleotide
OD	Optical density
PBS	Phosphate buffer saline
PCV	Packed cell volume
PUSFA	Poly-unsaturated fatty acids
R.B	Rumen bacteria
RBCs	Red blood cells
RNA	Ribonucleic acid
SAS	Statistical analysis system
Se	Selenium
Se-Cys	Selenocysteine

SELW1	Selenoproteins w
Se-Met	Selenomethionine
SFA	Saturated fatty acids
SOD	Superoxide dismutase
TAC	Total antioxidant
TBARS	Thiobarbituric acid reactive substances
TBS	Tris-buffered saline
TCOL	Total cholesterol
TG	Triglyceride
TNF- α	Tumor necrosis factor alpha
TP	Total protein
TCHOL	Total cholesterol
TXNDR1	Thioredoxin reductase
USFA	Un-saturated fatty acids
VLDL-C	Very density lipoprotein cholesterol
WBCs	White blood cells
WG	Weight gain

CHAPTER 1

GENERAL INTRODUCTION

Selenium (Se) is an essential trace mineral, which is a natural constituent of the earth's crust and available in living organisms in small quantities. In 1817, Se was recognised by Jacob Berzelius Jöns as a toxic compound that causes major health problems in humans and animals. However, the outcome of researches over the years demonstrated that Se can be absorbed by the intestinal tract of animals and humans, and then play a vital role in their body functions and metabolic pathways. Selenium has several important roles: For example, it is a significant structural part of at least 25 proteins, such as glutathione peroxidase (GSH-Px) and thioredoxin reductase (Zhang, 2009). These catalysts have important roles in the body, and the discovery of their structural Se inclusion illustrated the biochemical role of this mineral. Selenium is involved in the body for reproduction and fertility, thyroid hormone metabolism, redox control of enzymes and proteins, immune system function, DNA synthesis, and in the reduction of the aging process (Suttle, 2010; Méplan, 2011). Moreover, dietary Se affects the oxidative status in the host by altering seleno-proteins expression (Ferguson & Karunasinghe, 2011). Therefore, an adequate supply of Se element in the diet of animals is essential for maintenance of their health, growth, and biochemical-physiological functions. The Se status must be controlled in humans and animals and the daily recommended intake level should be adequate to perform its functions (Mehdi *et al.*, 2013).

To ensure optimal level of Se in livestock, inorganic Se is usually added to the diet in the form of sodium selenite or sodium selenate. However, most of the researches showed that inorganic Se sources are poorly utilised and poorly absorbed by livestock (Suttle, 2010), while organic Se sources such as seleno-methionine (SeMet) and seleno-cysteine (SeCys) resulted in high bioavailability of Se in the body compared to inorganic sources.

Concerning the improvement in Se status of livestock, most recent studies have tended to investigate the effective organic sources of Se to be used in animal diets. The results of these previous studies showed different sources of organic Se, but the most popular and commercially available one is Se-yeast, which, showed good results in improving performance, production, fertility and physiological aspect in poultry (Surai *et al.*, 2010). This is because yeast has the ability to produce SeMet in the presence of Se. The SeMet can be used by the host immediately for protein synthesis, or be deposited in the tissues and animal products, and in turn benefit the human population via the food chain (Kuricová *et al.*, 2003).

Many studies showed that organic Se also can be produced by bacterial species during respiration or detoxification processes (Stolz *et al.*, 2002). Several bacterial strains isolated from different environmental sources were found to be capable of converting inorganic Se to organic forms. However, limited data are available on using bacterial

Se in animal feed as a source of Se, but some studies indicated that various biological compounds can be produced by microorganisms, such as Se-enriched exopolysaccharide, which is produced by *Enterobacter cloacae* Z0206 (Zeqing *et al.*, 2013). The product of bacterial strain *Enterobacter cloacae* Z0206 showed significant improvement in the innate and humoral immunity in mice (Xu *et al.*, 2009). Moreover supplementation of Se-enriched exopolysaccharides product to broiler chickens improved the antioxidant system and enhanced the immunity response (Zeqing *et al.*, 2013).

Poultry production is associated with various stresses, and Se as an important trace mineral in poultry nutrition is the main structural component in antioxidant enzymes to overcome oxidative stresses (Fairweather-Tait *et al.*, 2010). All recent studies demonstrated that the organic Se is more bioavailable than inorganic forms in poultry nutrition (Stępińska *et al.*, 2012; Chen *et al.*, 2014), which may be due to the differences in the absorption and metabolism of organic and inorganic Se. Based on different poultry studies, it appears that excess Se supplementation of different sources could increase the concentration of the Se mineral in plasma (Kuricová *et al.*, 2003), and higher Se concentration in serum, kidney, liver, and breast muscle were observed (Wang *et al.*, 2011b). Some studies indicated that organic Se in broiler and layer nutrition affects the antioxidant status and the total antioxidant capacity in the blood and tissues of the birds; the inhibition of hydroxyl radical was also elevated when Se-yeast was used rather than sodium selenite (Chen *et al.*, 2014).

With regard to poultry performance and production, some researchers demonstrated that there is no relation between Se sources and poultry performance (Chantiratikul *et al.*, 2008a; Maysa *et al.*, 2009; and Chen *et al.*, 2014). However, some studies revealed that in comparison with inorganic Se, organic sources significantly increased live body weight in poultry (Maysa *et al.*, 2009), increased the birth rate and feed consumption and decreased mortality in the progeny of the layer (Wang *et al.*, 2011b). Although most of the reviewed studies reported better effects for organic Se on poultry performance in comparison with the inorganic Se, there are still some conflicting results between the studies.

In recent years, there has been interest in dietary incorporation of some minerals and vitamins as an economical way to enhance the body antioxidant status and increase the disease resistance in animals, as well as, enhance the delivery of essential nutrients to humans via animal products. The relation between dietary Se and Vit E had been well illustrated. Researchers suggested that supplementation of Se or Vit E has a beneficial effect on the antioxidant system and immune response (Guo *et al.*, 2003), however, their dietary combination may be more effective in improving growth performance, serum and meat antioxidant status and enhance innate and humoral immunity (Habibian *et al.*, 2015).

The possibility of delivering Se in a highly bioavailable form using Se-enriched bacterial protein has not been deeply investigated. However, according to recently data, there are some strains of bacteria capable of up-taking inorganic Se and accumulate it in their cells as Se-containing proteins. Thus, the current study was initiated to examine such effects with the following hypothesis and objectives.

Hypothesis statements

1. Se- enriched bacteria could be isolated from rumen fluid and hotspring water, and their Se- protein extract would identify as organic Se source.
2. Se-enriched bacterial protein would alter growth performance, Se- status meat quality, antioxidant status and immunity response in broiler chickens.
3. Combination of Vit E with Se-enriched bacterial protein would alter growth performance, antioxidant status, caecum microbial profile and cytokines gene expression in broiler chickens.

The general objective

To identify a potential source of bacterial organic Se from rumen fluid and hotspring water and examine the possibility of using it as a feed supplement to improve animals' Se status.

The specific objectives:

1. To isolate and identify potential selenium-enriched bacteria of high Se-containing protein, and extraction of their biomass Se-protein as a source of organic selenium.
2. To examine the effects of dietary supplementation of inorganic and bacterial organic selenium on growth performance, meat quality, selenium status, immunity, and hepatic selenoproteins gene expression in broiler chickens.
3. To investigate the effects of dietary supplementation of inorganic and bacterial organic Se sources and their Vit E combination on growth performance, antioxidant status, immunity response, cytokine gene expression in broiler chickens.

REFERENCES

- AAFCO. (2003). *Official publication. association of american feed control officials incorporated.* Olympia, WA.
- Abdel-Raheem, S. M., Abd-Allah, S. M., & Hassanein, K. M. (2012). The effects of prebiotic, probiotic and synbiotic supplementation on intestinal microbial ecology and histomorphology of broiler chickens. *International Journal for Agro Veterinary and Medical Sciences*, 6, 277–289.
- Adejumo, D. O. (2004). Performance, organ development and hematological indices of rats fed sole diets of graded levels of cassava flour and soybean flour (soygar) as substitutes for energy and protein concentrates. *Tropical Journal of Animal Science*, 7, 57–63.
- Aguilar, F., Autrup, H., Barlow, S., Castle, L., Crebelli, R., Dekant, W., ... Toldrá, F. (2008). Selenium-enriched yeast as source for selenium added for nutritional purposes in foods for particular nutritional uses and foods (including food supplements) for the general population. *The EFSA Journal*, 766, 1–42.
- Ahmadipour B, Hassanpour, H., Rafiei, F., & Khajali, F. (2015). Antioxidative, antihyperlipidemic, and growth-promoting effects of kelussia odoratissima in meat-type chickens. *Poultry Science Journal*, 1(1), 37–46.
- Ahmed, Z., Malhi, M., Soomro, S. A., Gandahi, J. A., Arijo, A., Bhutto, B., & Qureshi, T. A. (2016). Dietary selenium yeast supplementation improved some villi morphological characteristics in duodenum and jejunum of young goats. *The Journal of Animal & Plant Sciences*, 26(2), 382–387.
- Albuquerque, D. M. N., Lopes, J. B., Ferraz, M. S., Ribeiro, M. N., Silva, S. R. G., Costa, E. M. S., ... Lopes, J. C. O. (2017). Vitamin E and organic selenium for broilers from 22 to 42 days old: Performance and carcass traits. *Anais Da Academia Brasileira de Ciencias*, 89(2), 1259–1268.
- Alzate, A., Fernández-Fernández, A., Pérez-Conde, M. C., Gutiérrez, A. M., & Cámarra, C. (2008). Comparison of biotransformation of inorganic selenium by Lactobacillus and Saccharomyces in lactic fermentation process of yogurt and kefir. *Journal of Agricultural and Food Chemistry*, 56(18), 8728–8736.
- AMSA. (2012). *Meat color measurement guidelines (2nd ed.)* (201 East S). USA: American Meat Science Association.
- Andreoni, V., Luischi, M. M., Cavalca, L., Erba, D., & Ciappellano, S. (2000). Selenite tolerance and accumulation in the Lactobacillus species. *Annals of Microbiology*, 50, 77–88.

- Animashahun, R. A., Omoikhoje, S. O., & Bamgbose, A. M. (2006). Haematological and biochemical indices of weaner rabbits fed concentrates and *Syndrella nodiflora* forage supplement. In *11th Annual Conference of Animal Science Association*.
- Antonioli, P., Lampis, S., Chesini, I., Vallini, G., Rinalducci, S., Zolla, L., & Righetti, P. G. (2007). *Stenotrophomonas maltophilia* SeITE02, a new bacterial strain suitable for bioremediation of selenite-contaminated environmental matrices. *Applied and Environmental Microbiology*, 73(21), 6854–6863.
- Anugu, S., Petersson-Wolfe, C. S., Combs, G. F., & Petersson, K. H. (2013). Effect of vitamin E on the immune system of ewes during late pregnancy and lactation. *Small Ruminant Research*, 111(1–3), 83–89.
- AOAC. (1990). *Official methods of analysis*. (K. Herlick & V. A. Arlington, Eds.) (15th ed). USA: Association of official analytical chemist.
- Araúz, I. L. C., Afton, S., Wrobel, K., Caruso, J. A., Corona, J. F. G., & Wrobel, K. (2008). Study on the protective role of selenium against cadmium toxicity in lactic acid bacteria: An advanced application of ICP-MS. *Journal of Hazardous Materials*, 153(3), 1157–1164.
- Arbogast, S., & Ferreiro, A. (2010). Selenoproteins and protection against oxidative stress: selenoprotein N as a novel player at the crossroads of redox signaling and calcium homeostasis. *Antioxidants & Redox Signaling*, 12(7), 893–904.
- Arpášová, H., Haš, P., Ka, M., & Gálik, B. (2012). The effect of probiotic preparation enriched with selenium on performance parameters of laying hens. *Animal Science and Biotechnologies*, 45(1), 17–23.
- Arthur, J. R., Mckenzie, R. C., & Beckett, G. J. (2003). Selenium in the immune system. *The Journal of Nutrition*, 133(5), 1457–1459.
- Avoscan, L., Collins, R., Carriere, M., Gouget, B., & Covès, J. (2006). Seleno-L-methionine is the predominant organic form of selenium in *Cupriavidus metallidurans* CH34 exposed to selenite or selenate. *Applied and Environmental Microbiology*, 72(9), 6414–6416.
- Bartoň, L., Marounek, M., Kudrna, V., Bureš, D., & Zahrádková, R. (2007). Growth performance and fatty acid profiles of intramuscular and subcutaneous fat from Limousin and Charolais heifers fed extruded linseed. *Meat Science*, 76(3), 517–523.
- Bartosch, S., Fite, A., Macfarlane, G. T., & Mcmurdo, M. E. T. (2004). Characterization of bacterial communities in feces from healthy elderly volunteers and hospitalized elderly patients by using real-time PCR and effects of antibiotic treatment on the fecal microbiota. *Applied and Environmental Microbiology*, 70(6), 3575–3581.

- Behne, D., & Kyriakopoulos, A. (2001). Mammalian selenium-containing proteins. *Annual Review of Nutrition*, 21(1), 453–473.
- Bellinger, F. P., Raman, A. V., Reeves, M. A., & Berry, M. J. (2009). Regulation and function of selenoproteins in human disease. *Biochemical Journal*, 422(1), 11–22.
- Ben-Amara, I., Soudani, N., Troudi, A., Bouaziz, H., Boudawara, T., & Zeghal, N. (2011). Antioxidant effect of vitamin E and selenium on hepatotoxicity induced by dimethoate in female adult rats. *Ecotoxicology and Environmental Safety*, 74(4), 811–819.
- Berri, C., Le Bihan-Duval, E., Debut, M., Santé-Lhoutellier, V., Baéza, E., Gigaud, V., ... Duclos, M. J. (2007). Consequence of muscle hypertrophy on characteristics of Pectoralis major muscle and breast meat quality of broiler chickens. *Journal of Animal Science*, 85, 2005–2011.
- Bianco, A. C., Salvatore, D., Gereben, B., Berry, M. J., & Larsen, P. R. (2002). Biochemistry, cellular and molecular biology, and physiological roles of the iodothyronine selenodeiodinases. *Endocrine Reviews*, 23(1), 38–89.
- Biswas, A., Ahmed, M., Bharti, V. K., & Singh, S. B. (2011a). Effect of antioxidants on physio-biochemical and hematological parameters in broiler chicken at high altitude. *Asian-Australasian Journal of Animal Sciences*, 24(2), 246–249.
- Biswas, K. C., Barton, L. L., Tsui, W. L., Shuman, K., Gillespie, J., & Eze, C. S. (2011b). A novel method for the measurement of elemental selenium produced by bacterial reduction of selenite. *Journal of Microbiological Methods*, 86(2), 140–144.
- Böck, A., Rother, M., Leibundgut, M., & Ban, N. (2006). Selenium metabolism in prokaryotes. In *Selenium* (pp. 9–28). Springer US.
- Boiago, M. M., Borba, H., Leonel, F. R., Giampietro-Ganeco, A., Ferrari, F. B., Stefani, L. M., & Souza, P. A. de. (2014). Sources and levels of selenium on breast meat quality of broilers. *Ciência Rural*, 44(9), 1692–1698.
- Bölükbaşı, Ş. C., Erhan, M. K., & Özkan, A. (2006). Effect of dietary thyme oil and vitamin E on growth, lipid oxidation, meat fatty acid composition and serum lipoproteins of broilers. *South African Journal of Animal Sciences*, 36(3), 189–196.
- Boostani, A., Sadeghi, A. A., Mousavi, S. N., Chamani, M., & Kashan, N. (2015). Effects of organic, inorganic, and nano-Se on growth performance, antioxidant capacity, cellular and humoral immune responses in broiler chickens exposed to oxidative stress. *Livestock Science*, 178, 330–336.

- Briens, M., Mercier, Y., Rouffineau, F., Mercerand, F., & Geraert, P.-A. (2014). 2-Hydroxy-4-methylselenobutanoic acid induces additional tissue selenium enrichment in broiler chickens compared with other selenium sources. *Poultry Science*, 93(1), 85–93.
- Briens, M., Mercier, Y., Rouffineau, F., Vacchina, V., & Geraert, P. A. (2013). Comparative study of a new organic selenium source v. seleno-yeast and mineral selenium sources on muscle selenium enrichment and selenium digestibility in broiler chickens. *British Journal of Nutrition*, 110(4), 617–624.
- Brown, A. J., & Jessup, W. (1999). Oxysterols and atherosclerosis. *Atherosclerosis*, 142(1), 1–28.
- Bunglavan, S. J., Garg, A. K., Dass, R. S., & Srivastava, S. (2014). Effect of supplementation of different levels of selenium as nanoparticles/sodium selenite on blood biochemical profile and humoral immunity in male Wistar rats. *Veterinary World*, 7(12), 1075–1081.
- Burk, R. F., Hill, K. E., & Motley, A. K. (2003). Selenoprotein metabolism and function: evidence for more than one function for selenoprotein P. *The Journal of Nutrition*, 133, 1517S–1520S.
- Burke, N. C., Scaglia, G., Saker, K. E., Blodgett, D. J., & Swecker, W. S. (2007). Influence of endophyte consumption and heat stress on intravaginal temperatures, plasma lipid oxidation, blood selenium, and glutathione redox of mononuclear cells in heifers grazing tall fescue. *Journal of Animal Science*, 85(11), 2932–2940.
- Burton, G. W., & Traber, M. G. (1990). Vitamin E: antioxidant activity, biokinetics, and bioavailability. *Annual Review of Nutrition*, 10(1), 357–382.
- Butler, J. A., Beilstein, M. A., & Whanger, P. D. (1989). Influence of dietary methionine on the metabolism of selenomethionine in rats. *The Journal of Nutrition*, 119(7), 1001–1009.
- Cai, S. J., Wu, C. X., Gong, L. M., Song, T., Wu, H., & Zhang, L. Y. (2012). Effects of nano-selenium on performance, meat quality, immune function, oxidation resistance, and tissue selenium content in broilers. *Poultry Science*, 91, 2532–2539.
- Calomme, M., Hu, J., Van-Den, B. K., & Berghe, D. V. (1995). Seleno-lactobacillus. *Biological Trace Element Research*, 47, 379–383.
- Cao, X., Zhang, W., & Wang, R. (2001). Effects of selenium source and level on performance, immune function and meat quality in AA broilers. In *In Alltech's 17th Annual Science & Technology in the Feed Industry Symposium*. (pp. 297–304). Nottingham: Nottingham University Press.

- Chan, K. W., Khong, N. M. H., Iqbal, S., Umar, I. M., & Ismail, M. (2012). Antioxidant property enhancement of sweet potato flour under simulated gastrointestinal pH. *International Journal of Molecular Sciences*, 13, 8987–8997.
- Chantiratikul, A., Aengwanich, W., Chinrasri, O., & Chantiratikul, P. (2008a). Plasma selenium concentration and glutathione peroxidase activity in red blood cells of laying hens fed sodium selenite or zinc-L-selenomethionine. *International Journal of Poultry Science*, 7(7), 692–695.
- Chantiratikul, A., Orawan, C., & Chantiratikul, P. (2008b). Effect of sodium selenite and zinc-L-selenomethionine on performance and selenium concentrations in eggs of laying hens. *Asian-Australasian Journal of Animal Sciences*, 21(7), 1048–1052.
- Chapman, J., Weir, E., & Regan, F. (2010). Period four metal nanoparticles on the inhibition of biofouling. *Colloids Surf B Biointerfaces*, 78, 208–216.
- Chen, G., Wu, J., & Li, C. (2014). Effect of different selenium sources on production performance and biochemical parameters of broilers. *Journal of Animal Physiology and Animal Nutrition*, 98(4), 747–754.
- Chen, J., & Berry, M. J. (2003). Selenium and selenoproteins in the brain and brain diseases. *Journal of Neurochemistry*, 86(1), 1–12.
- Chen, L., Pan, D.-D., Zhou, J., & Jiang, Y.-Z. (2005). Protective effect of selenium-enriched Lactobacillus on CCl₄-induced liver injury in mice and its possible mechanisms. *World Journal of Gastroenterology*, 11(37), 5795–5800.
- Cheng, K., Niu, Y., Zheng, X. C., Zhang, H., Chen, Y. P., Zhang, M., & Huang, X. X. (2016). A Comparison of natural (D- α -tocopherol) and synthetic (DL- α -tocopherol Acetate) vitamin E supplementation on the growth performance , meat quality and oxidative status of broilers. *Asian-Australasian Journal of Animal Sciences*, 29(5), 681–688.
- Chmielewski, J., & Tyflewska, A. (2007). Selenium in microorganisms. In M. Wierzbicka, E. Bulska, K. Pyrzyńska, I. Wysocka, & B. A. Zachara (Eds.), *Selenium. The element essential to health and fascinating for researchers*. Warsaw, Malamut.
- Choct, M., Naylor, A. J., & Reinke, N. (2004). Selenium supplementation affects broiler growth performance, meat yield and feather coverage. *British Poultry Science*, 45(5), 677–683.
- Choe, D. W., Loh, T. C., Foo, H. L., Hair-Bejo, M., & Awis, Q. S. (2012). Egg production, faecal pH and microbial population, small intestine morphology, and plasma and yolk cholesterol in laying hens given liquid metabolites produced by Lactobacillus plantarum strains. *British Poultry Science*, 53(1), 106–115.

- Combs, G., & Combs Jr, S. . (1986). *The role of selenium in nutrition*. Academic Press, Inc.
- Combs, G. F. (1988). Selenium in foods. In C. O. Chichester & B. S. Schweigert (Eds.), *Advances in Food Research* (Vol. 32, pp. 85–113). Academic Press.
- Combs, G. F. (1994). Clinical implications of selenium and vitamin E in poultry nutrition. *Veterinary Clinical Nutrition*, 1, 133–140.
- Combs, G. F., & Combs Jr, B. (1984). The nutritional biochemistry of selenium. *Annual Review of Nutrition*, 4(140), 257–80.
- Cox, A. J., Lehtinen, A. B., Xu, J., Langefeld, C. D., Freedman, B. I., Carr, J. J., & Bowden, D. W. (2013). Polymorphisms in the selenoprotein S gene and subclinical cardiovascular disease in the diabetes heart study. *Acta Diabetologica*, 50(3), 391–399.
- Cox, R., & García-Palmieri, M. (1990). Cholesterol, triglycerides, and associated lipoproteins. In H. J. Walker HK, Hall WD (Ed.), *Clinical Methods: The History, Physical, and Laboratory Examinations* (3rd ed.). Boston: Butterworths.
- Cozzi, G., Prevedello, P., Stefani, a. L., Piron, a., Contiero, B., Lante, a., ... Chevaux, E. (2011). Effect of dietary supplementation with different sources of selenium on growth response, selenium blood levels and meat quality of intensively finished Charolais young bulls. *Animal*, 5(10), 1531–1538.
- Da-Silva, I., Ribeiro, A., Canal, C., Trevizan, L., Macagnan, M., Gonçalves, T., ... Pereira, R. (2010). The impact of organic and inorganic selenium on the immune system of growing broilers submitted to immune stimulation and heat stress. *Revista Brasileira de Ciência Avícola*, 12(4), 247–254.
- Dai, M., Wu, S., Feng, M., Feng, S., Sun, C., Bai, D., ... Cao, W. (2016). Recombinant chicken interferon-alpha inhibits the replication of exogenous avian leukosis virus (ALV) in DF-1 cells. *Molecular Immunology*, 76, 62–69.
- David, L. S., Jerry, K., & Monty, S. L. (1977). The influence of dietary selenium and vitamin E on glutathione peroxidase and glutathione in the rat. *Biochimica et Biophysica Acta*, 497, 218–224.
- Debieux, C. M., Dridge, E. J., Mueller, C. M., Splatt, P., Paszkiewicz, K., Knight, I., ... Butler, C. S. (2011). A bacterial process for selenium nanosphere assembly. *Proceedings of the National Academy of Sciences of the United States of America*, 108(33), 13480–13485.
- Del-Maestro, R. (1991). Free radicals as mediators of tissue injury. In IE. Dreosti (Ed.), *Trace Elements, Micronutrients, and Free Radicals* (pp. 25–51). Springer.

- Del-Puerto, M., Cabrera, M. C., & Saadoun, A. (2017). A note on fatty acids profile of meat from broiler chickens supplemented with inorganic or organic selenium. *International Journal of Food Science*, 1–8.
- Dessi, S., & Batetta, B. (2003). Overview- intracellular cholesterol homeostasis: old and new players. In A. Dessi and S. Pani (Ed.), *Cell growth and cholesterol esters* (pp. 1–12). New York, NY, USA: Kluwer Academic/Plenum Publishers.
- Dhingra, S., & Bansal, M. P. (2006). Attenuation of LDL receptor gene expression by selenium deficiency during hypercholesterolemia. *Molecular and Cellular Biochemistry*, 282(1), 75–82.
- Dinarello, C. A. (2000). Proinflammatory cytokines. *Chest*, 118(2), 503–508.
- Djordjevic, S., Omerovic, I., Stolic, N., & Milosevic, B. (2016). Effect of organic selenium in broiler diet on slaughter traits and plasma lipids. *IOSR Journal of Agriculture and Veterinary Science*, 9(5), 71–73.
- Dlouhá, G., Ševčíková, S., Dokoupilová, A., Zita, L., Heindl, J., & Skřivan, M. (2008). Effect of dietary selenium sources on growth performance , breast muscle selenium , glutathione peroxidase activity and oxidative stability in broilers. *Czech Journal of Animal Science*, 53(6), 265–269.
- Dong, Y., Zhang, H., Hawthorn, L., Ganther, H. E., & Ip, C. (2003). Delineation of the molecular basis for selenium-induced growth arrest in human prostate cancer cells by oligonucleotide array. *Cancer Research*, 63, 52–59.
- Doyle, D. (2006). William Hewson (1739–74): the father of haematology. *British Journal of Haematology*, 133(4), 375–381.
- Droge, W. (2002). Free radicals in the physiological control of cell function. *Physiological Reviews*, 82(1), 47–95.
- Dumont, E., Vanhaecke, F., & Cornelis, R. (2006). Selenium speciation from food source to metabolites: a critical review. *Analytical and Bioanalytical Chemistry*, 385, 1304–1323.
- Dungan, R., & Frankenberger, W. (2001). Bioremoval of selenium by Enterobacter cloacae SLD1a–1: formation of dimethylselenide. *Biogeochemistry*, 55, 73–86.
- Ebeid, T. A., Zeweil, H. S., Basyony, M. M., Dosoky, W. M., & Badry, H. (2013). Fortification of rabbit diets with vitamin E or selenium affects growth performance, lipid peroxidation, oxidative status and immune response in growing rabbits. *Livestock Science*, 155(2–3), 323–331.
- El-Ramady, H., Abdalla, N., Alshaal, T., Domokos-Szabolcsy, É., Elhawat, N., Prokisch, J., ... Shams, M. (2014). Selenium in soils under climate change, the implication for human health. *Environmental Chemistry Letters*, 13, 1–19.

- El-Shenawy, N. S., AL-Harbi, M. S., & Hamza, R. Z. (2015). Effect of vitamin E and selenium separately and in combination on biochemical, immunological and histological changes induced by sodium azide in male mice. *Experimental and Toxicologic Pathology*, 67(1), 65–76.
- Erol, H., Imik, H., Gumus, R., & Halici, M. (2017). The effects of different amount of protein and vitamin E supplementation in rations on lipid and antioxidant metabolism of broilers exposed to heat stress. *Brazilian Journal of Poultry Science*, 19(2), 289–296.
- Eszenyi, P., Sztrik, A., Babka, B., & Prokisch, J. (2011). Elemental, nano-sized (100–500 nm) selenium production by probiotic lactic acid bacteria. *International Journal of Bioscience, Biochemistry and Bioinformatics*, 1(2), 148–152.
- Fairweather-Tait, S. J., Collings, R., & Hurst, R. (2010). Selenium bioavailability: current knowledge and future research. *The American Journal of Clinical Nutrition*, 91(2), 1484S–1491S.
- Fan, Y. K., Croom, J., Christensen, V. L., Black, B. L., & Bird, A. R. (1997). Jejunal glucose uptake and oxygen consumption in turkey poult selected for rapid growth. *Poultry Science*, 76(12), 1738–1745.
- Fawzy, M. M., El-sadawi, H. A., & El-dien, M. H. (2016). Hematological and biochemical performance of poultry following zinc oxide and sodium selenite supplementation as food additives. *Annals of Clinical Pathology*, 4(4), 1076.
- Federal Register. (2000). *Food additive permitted in feed and drinking water: selenium yeast* (Vol. 65). Federal Register.
- Ferguson, L. R., & Karunasinghe, N. (2011). Nutrigenetics, nutrigenomics, and selenium. *Frontiers in Genetics*, 2, 1–10.
- Fernández-Llamosas, H., Castro, L., Blázquez, M., Díaz, E., & Carmona, M. (2016). Biosynthesis of selenium nanoparticles by Azoarcus sp. CIB. *Microbial Cell Factories*, 15, 109.
- Fesharaki, P. J., Nazari, P., Shakibaie, M., Rezaie, S., Banoei, M., Abdollahi, M., & Shahverdi, A. R. (2010). Biosynthesis of selenium nanoparticles using Klebsiella pneumoniae and their recovery by a simple sterilization process. *Brazilian Journal of Microbiology*, 41(2), 461–466.
- Finch, J., & Turner, R. . (1996). Effects of selenium and vitamin E on the immune responses of domestic animals. *Research in Veterinary Science*, 60(2), 97–106.
- Finley, J. (2006). Bioavailability of selenium from foods. *Nutrition Reviews*, 64, 146–151.

- Fletcher, D. (2002). Poultry meat quality. *World's Poultry Science Journal*, 58, 131–145.
- Flohe, L., Günzler, W. A., & Schock, H. H. (1973). Glutathione peroxidase: a selenoenzyme. *FEBS Letters*, 32(1), 132–134.
- Florian, S., Wingler, K., Schmehl, K., Jacobasch, G., Kreuzer, O. J., Meyerhof, W., & Brigelius-Flohé, R. (2001). Cellular and subcellular localization of gastrointestinal glutathione peroxidase in normal and malignant human intestinal tissue. *Free Radical Research*, 35(6), 655–663.
- Folch, J., Lees, M., & Stanley, G. H. S. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *The Journal of Biological Chemistry*. 497-509.
- Fomenko, D. E., Novoselov, S. V., Natarajan, S. K., Lee, B. C., Koc, A., Carlson, B. A., ... Gladyshev, V. N. (2009). MsrB1 (Methionine-Rsulfoxide Reductase 1) knock-out mice: roles of MsrB1 in redox regulation and identification of a novel selenoprotein form. *Journal of Biological Chemistry*, 284(9), 5986–5993.
- Fordyce, F. (2013). Selenium deficiency and toxicity in the environment. In O. Selinus, E. Brian, C. B. Davies, & C. D. Theo (Eds.), *Essentials of Medical Geology* (pp. 373–415). Netherlands: Springer.
- Fox, T., Van den Heuvel, E., Atherton, C., Dainty, J., Lewis, D., Langford, N., ... Fairweather-Tait, S. (2004). Bioavailability of selenium from fish, yeast and selenate: a comparative study in humans using stable isotopes. *European Journal of Clinical Nutrition*, 58, 343–349.
- Friedewald, W., Levy, R., & Fredrickson, D. (1972). Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clinical Chemistry*, 18, 499–502.
- Funari, J. P., Albuquerque, R. D., Murarolli, V. D. A., Raspantini, L. E. R., Cardoso, A. L. S. P., Tessari, E. N. C., & Alves, F. R. (2012). Different sources and levels of selenium on humoral immunity of broiler chickens. *Ciência Rural*, 42(1), 154–159.
- Galano, E., Mangiapane, E., Bianga, J., Palmese, A., Pessione, E., Szpunar, J., ... Amoresano, A. (2013). Privileged incorporation of selenium as selenocysteine in *Lactobacillus reuteri* proteins demonstrated by selenium-specific imaging and proteomics. *Molecular & Cellular Proteomics*, 12(8), 2196–2204.
- Gallo-Torres, D. C. (1980). Absorption, blood transport and metabolism of vitamin E. In L. J. Machlin (Ed.), *A Comprehensive Treatise* (pp. 170–267). New York: Marcel Dekker.,

- Garbisu, C., Ishii, T., Leighton, T., & Buchanan, B. (1996). Bacterial reduction of selenite to elemental selenium. *Chemical Geology*, 132, 199–204.
- Gault, N. F. S. (1985). The relationship between water-holding capacity and cooked meat tenderness in some beef muscles as influenced by acidic conditions below the ultimate pH. *Meat Science*, 15, 15–30.
- Gaweł, S., Wardas, M., Niedworok, E., & Wardas, P. (2004). Malondialdehyde (MDA) as a lipid peroxidation marker. *Wiadomosci Lekarskie (Warsaw, Poland: 1960)*, 57, 453–455.
- Gelderman, A., & Clapper, J. (2013). Effects of inorganic or organic selenium on immunoglobulins in swine. *Journal of Animal Science and Biotechnology*, 4(47), 1–9.
- Göçmen, R., Yazgan, O., & Cufadar, Y. (2016). Effect of different organic and inorganic selenium levels on performance, selenium concentrations of some tissues, glutathione peroxidase enzyme activity and meat quality in broilers. *The Journal of Animal & Plant Sciences*, 26(4), 916–923.
- Grashorn, M. A. (2007). Functionality of poultry meat. *Journal of Applied Poultry Research*, 16(1), 99–106.
- Gromer, S., Eubel, J., Lee, B., & Jacob, J. (2005). Human selenoproteins at a glance. *Cellular and Molecular Life Sciences*, 62, 2414–2437.
- Grumolato, L., Ghzili, H., Montero-Hadjadje, M., Gasman, S., Lesage, J., Tanguy, Y., Galas, L., ... Elkahloun, A. G. (2008). Selenoprotein T is a PACAP-regulated gene involved in intracellular Ca²⁺ mobilization and neuroendocrine secretion. *The FASEB Journal*, 22(6), 1756–1768.
- Gružauskas, R., Barštys, T., Racevičiute-Stupeliene, A., Kliševičiute, V., Buckiuniene, V., & Bliznikas, S. (2014). The effect of sodium selenite, selenium methionine and vitamin E on productivity, digestive processes and physiologic condition of broiler chickens. *Veterinarija Ir Zootechnika*, 65(87), 22–29.
- Guo, Y., Zhang, G., Yuan, J., & Nie, W. (2003). Effects of source and level of magnesium and vitamin E on prevention of hepatic peroxidation and oxidative deterioration of broiler meat. *Animal Feed Science and Technology*, 107(1), 143–150.
- Habibian, M., Ghazi, S., & Moeini, M. M. (2015). Effects of dietary selenium and vitamin E on growth performance, meat yield, and selenium content and lipid oxidation of breast meat of broilers reared under heat stress. *Biological Trace Element Research*, 169(1), 142–152.

- Habibian, M., Ghazi, S., Moeini, M. M., & Abdolmohammadi, A. (2014). Effects of dietary selenium and vitamin E on immune response and biological blood parameters of broilers reared under thermoneutral or heat stress conditions. *International Journal of Biometeorology*, 58(5), 741–752.
- Hadley, K. B., & Sunde, R. A. (2001). Selenium regulation of thioredoxin reductase activity and mRNA levels in rat liver. *The Journal of Nutritional Biochemistry*, 12(12), 693–702.
- Halliwell, B., & Gutteridge, J. M. (2015). *Free radicals in biology and medicine*. USA: Oxford University Press,.
- Halliwell, B., & Poulsen, H. E. (2006). *Oxidative stress. Cigarette smoke and oxidative stress*. Berlin: Springer-Verlag.
- Harsini, S. G., Habibiyan, M., Moeini, M. M., & Abdolmohammadi, A. R. (2012). Effects of dietary selenium, vitamin E, and their combination on growth, serum metabolites, and antioxidant defense system in skeletal muscle of broilers under heat stress. *Biological Trace Element Research*, 148(3), 322–330.
- Hashemi, Z., Sharifi, N., Khani, B., Aghadavod, E., & Asemi, Z. (2017). The effects of vitamin E supplementation on endometrial thickness, and gene expression of vascular endothelial growth factor and inflammatory cytokines among women with implantation failure. *The Journal of Maternal-Fetal & Neonatal Medicine*, 1–8.
- Hasty, J. L., van Heugten, E., See, M. T., & Larick, D. K. (2002). Effect of vitamin E on improving fresh pork quality in Berkshire- and Hampshire-sired pigs1. *Journal of Animal Science*, 80, 3230–3237.
- He, J., Zhang, K. Y., Chen, D. W., Ding, X. M., Feng, G. D., & Ao, X. (2013). Effects of vitamin E and selenium yeast on growth performance and immune function in ducks fed maize naturally contaminated with aflatoxin B1. *Livestock Science*, 152(2–3), 200–207.
- Henry P. R. and Ammerman.C, B. (1995). Selenium bioavailability. In C.B. Ammerman, D.H. Baker and A.J. Lewis (Ed.), *Bioavailability of nutrients for animals* (pp. 303–336). New York, USA.: Academic Press.
- Hernken, R. W., Harmon, R. J., & Tramsmel, S. (1998). Selenium of dairy cattle: A role for organic selenium. In T. P. Lyons & K. A. Jacques (Eds.), *Biotechnology in feed industry.Proceedings Alltech 14th Ann. Symp.* (pp. 797–803). Loughborough, LEC, UK: Nottingham University Press.
- Hidiroglon, M., Heaney, D. P., & Jenkins, K. J. (1968). Metabolism of inorganic selenium in rumen bacteria. *Canadian Journal of Physiology and Pharmacology*, 46, 229–232.

- Hoffmann, P. R., & Berry, M. J. (2008). The influence of selenium on immune responses. *Molecular Nutrition and Food Research*. 1273-1280.
- Hogan, G. R., & Jackson, P. D. (1986). Dichotomous effects of cadmium and selenium on erythropoiesis in mice. *Bulletin of Environmental Contamination and Toxicology*, 36(1), 674–679.
- Hosseini Mansoub, N. (2011). Influence of organic selenium source on carcass characteristics and oxidative stability of meat of male broilers. *Advances in Environmental Biology*, 5, 1832-1836.
- Hu, C. H., Li, Y. L., Xiong, L., Zhang, H. M., Song, J., & Xia, M. S. (2012). Comparative effects of nano elemental selenium and sodium selenite on selenium retention in broiler chickens. *Animal Feed Science and Technology*, 177(3–4), 204–210.
- Huang, X., Sun, B., Zhang, J., Gao, Y., & Li, G. (2017). Selenium deficiency induced injury in chicken muscular stomach by downregulating selenoproteins. *Biological Trace Element Research*, 1–7.
- Huber, R., Sacher, M., Huber, H., & Rose, D. (2000). Respiration of arsenate and selenate by hyperthermophilic archaea. *Systematic and Applied Microbiology*, 23, 305–314.
- Hucker, G. J., & Conn, H. J. (1923). *Methods of Gram staining*. Technical Bulletin of the New York State Agricultural Experimental Station, no. 93.
- Hudman, J. F., & Glenn, A. R. (1984). Selenite uptake and incorporation by *Selenomonas ruminantium*. *Arch Microbiol*, 140, 252–256.
- Hudman, J. F., & Glenn, A. R. (1985). Selenium uptake by *Butyrivibrio fibrisolvens* and *Bacteroides ruminicola*. *FEMS Microbiology Letters*, 27(2), 215–220.
- Humann-Ziehank, E., Renko, K., Mueller, A. S., Roehrig, P., Wolfsen, J., & Ganter, M. (2013). Comparing functional metabolic effects of marginal and sufficient selenium supply in sheep. *Journal of Trace Elements in Medicine and Biology*, 27(4), 380–390.
- Hunter, W. J., & Manter, D. K. (2009). Reduction of selenite to elemental red selenium by *Pseudomonas* sp. strain CA5. *Current Microbiology*, 58, 493–498.
- Hussain, A., Jinke, T., Jianjun, W., Muhammad Ammar, K., Yuanxiao, W., Lili, Z., & Tian, W. (2012). Effects of dietary sodium selenite and selenium yeast on antioxidant enzyme activities and oxidative stability of chicken breast meat. *Journal of Agricultural and Food Chemistry*, 60, 7111–7120.

- Ibrahim, H. A. M., Zhu, Y., Wu, C., Lu, C., Ezekwe, M. O., Liao, S. F., & Haung, K. (2012). Selenium-enriched probiotics improves murine male fertility compromised by high fat diet. *Biological Trace Element Research*, 147(1–3), 251–260.
- Imik, H., Atasever, M. A., Urcar, S., Ozlu, H., Gumus, R., & Atasever, M. (2012). Meat quality of heat stress exposed broilers and effect of protein and vitamin E. *British Poultry Science*, 53(5), 689–698.
- Jahromi, M. F., Altaher, Y. W., Shokryazdan, P., Ebrahimi, R., Ebrahimi, M., Idrus, Z., ... Liang, J. B. (2016). Dietary supplementation of a mixture of Lactobacillus strains enhances performance of broiler chickens raised under heat stress conditions. *International Journal of Biometeorology*, 60(7), 1099–1110.
- Jamilian, M., Samimi, M., Afshar Ebrahimi, F., Aghadavod, E., Mohammadbeigi, R., Rahimi, M., & Asemi, Z. (2017). Effects of selenium supplementation on gene expression levels of inflammatory cytokines and vascular endothelial growth factor in patients with gestational diabetes. *Biological Trace Element Research*, 1–8.
- Jenkins, K. J., Hidiroglou, M., & Collins, F. W. (1993). Influence of various flavonoids and simple phenolics on development of exudative diathesis in the chick. *Journal of Agricultural and Food Chemistry*, 41, 441–445.
- Jia, X., Li, N., & Chen, J. (2005). A subchronic toxicity study of elemental Nano-Se in Sprague-Dawley rats. *Life Sciences*, 76(17), 1989–2003.
- Jiang, Z., Lin, Y., Zhou, G., Luo, L., Jiang, S., & Chen, F. (2009). Effects of dietary selenomethionine supplementation on growth performance, meat quality and antioxidant property in yellow broilers. *Journal of Agricultural and Food Chemistry*, 57(20), 9769–9772.
- Juniper, D. T., Phipps, R. H., Ramos-Morales, E., & Bertin, G. (2009). Effects of dietary supplementation with selenium enriched yeast or sodium selenite on selenium tissue distribution and meat quality in lambs. *Animal Feed Science and Technology*, 149(3–4), 228–239.
- Juszczuk-Kubiak, E., Bujko, K., Cymer, M., Wicińska, K., Gabryszuk, M., & Pierzchała, M. (2016). Effect of Inorganic Dietary Selenium Supplementation on Selenoprotein and Lipid Metabolism Gene Expression Patterns in Liver and Loin Muscle of Growing Lambs. *Biological Trace Element Research*, 172(2), 336–345.
- Kang, B. P. S., Bansal, M. P., & Mehta, U. (2000). Hyperlipidemia and type I 5'-monodeiodinase activity. *Biological Trace Element Research*, 77(3), 231–239.

- Kareem, K. Y., Loh, T. C., Foo, H. L., Asmara, S. A., Akit, H., Abdulla, N. R., & Foongooi, M. (2015). Carcass, meat and bone quality of broiler chickens fed with postbiotic and prebiotic combinations. *International Journal of Probiotics and Prebiotics*, 10(1), 23–30.
- Kasaikina, M. V., Kravtsova, M. A., Lee, B. C., Seravalli, J., Peterson, D. A., Walter, J., ... Gladyshev, V. N. (2011). Dietary selenium affects host selenoproteome expression by influencing the gut microbiota. *The FASEB Journal*, 25(7), 2492–2499.
- Kessi, J., & Hanselmann, K. (2004). Similarities between the abiotic reduction of selenite with glutathione and the dissimilatory reaction mediated by *Rhodospirillum rubrum* and *Escherichia coli*. *Journal of Biological Chemistry*, 279, 50662–50669.
- Kessi, J., Ramuz, M., Wehrli, E., & Spycher, M. (1999). Reduction of selenite and detoxification of elemental selenium by the phototrophic bacterium *Rhodospirillum rubrum*. *Applied and Environmental Microbiology*, 65(11), 4734–4740.
- Khan, A. Z., Kumbhar, S., Liu, Y., Hamid, M., Pan, C., Nido, S. A., ... Huang, K. (2017). Dietary supplementation of selenium-enriched probiotics enhances meat quality of broiler chickens (*Gallus gallus domesticus*) raised under high ambient temperature. *Biological Trace Element Research*.
- Khan, M. Z. I., Akter, S. H., Islam, M. N., Karim, M. R., Islam, M. R., & Kon, Y. (2008). The effect of selenium and vitamin e on the lymphocytes and immunoglobulin-containing plasma cells in the lymphoid organ and mucosa-associated lymphatic tissues of broiler chickens. *Journal of Veterinary Medicine Series C: Anatomia Histologia Embryologia*, 37(1), 52–59.
- Kheradmand, E., Rafii, F., Yazdi, M. H., Sepahi, A. A., Shahverdi, A. R., & Oveis, M. R. (2014). The antimicrobial effects of selenium nanoparticle-enriched probiotics and their fermented broth against *Candida albicans*. *Daru : Journal of Pharmaceutical Sciences*, 22(1), 48.
- Kieliszek, M., & Błazejak, S. (2013). Selenium: Significance, and outlook for supplementation. *Nutrition*, 29(5), 713–718.
- Kim, Y. J., Park, W. Y., & Choi, I. H. (2010). Effects of dietary alpha-tocopherol, selenium, and their different combinations on growth performance and meat quality of broiler chickens. *Poultry Science*, 89(3), 603–608.
- King, N., & Whyte, R. (2006). Does it look cooked? A review of factors that influence cooked meat color. *Journal of Food Science*, 71(4), R31–R40.

- Kobayashi, Y., Ogra, Y., Ishiwata, K., Takayama, H., Aimi, N., & Suzuki, K. T. (2002). Selenosugars are key and urinary metabolites for selenium excretion within the required to low-toxic range. In *Proceedings of the National Academy of Sciences* (Vol. 99, pp. 15932–15936).
- Köhrle, J., Brigelius-Flohé, R., Böck, A., Gärtner, R., Meyer, O., & Flohé, L. (2000). Selenium in biology: facts and medical perspectives. *Biological Chemistry*, 381(9–10), 849–864.
- Koreleski, J., & Swiatkiewicz, S. (2007). Dietary supplementation with plant extracts, xanthophylls and synthetic antioxidants: Effect on fatty acid profile and oxidative stability of frozen stored chicken breast meat. *Journal of Animal and Feed Sciences*, 16(3), 463–471.
- Kralík, Z., Kralík, G., Biažík, E., Straková, E., & Suchý, P. (2013). Effects of organic selenium in broiler feed on the content of selenium and fatty acid profile in lipids of thigh muscle tissue. *Acta Veterinaria Brno*, 82, 277–282.
- Krittaphol, W., Wescombe, P. A., Thomson, C. D., McDowell, A., Tagg, J. R., & Fawcett, J. P. (2011). Metabolism of L-selenomethionine and selenite by probiotic bacteria: In vitro and in vivo studies. *Biological Trace Element Research*, 144(1–3), 1358–1369.
- Kryukov, G. V., Castellano, S., Novoselov, S. V., Lobanov, A. V., Zehtab, O., Guigó, R., & Gladyshev, V. N. (2003). Characterization of mammalian selenoproteomes. *Science*, 300(5624), 1439–1443.
- Kuricová, S., Boldiárová, K., Áková, G. R. E., Bobáek, R., & Levkut, M. (2003). Chicken selenium status when fed a diet supplemented with Se-Yeast. *Acta Veterinaria Brno*, 72, 339–346.
- Lamberti, C., Mangiapane, E., Pessione, A., Mazzoli, R., Giunta, C., & Pessione, E. (2011). Proteomic characterization of a selenium-metabolizing probiotic *Lactobacillus reuteri* Lb2 BM for nutraceutical applications. *Proteomics*, 11, 2212–2221.
- Lampis, S., Zonaro, E., Bertolini, C., Cecconi, D., Monti, F., Micaroni, M., ... Vallini, G. (2017). Selenite biotransformation and detoxification by *Stenotrophomonas maltophilia* SeITE02: Novel clues on the route to bacterial biogenesis of selenium nanoparticles. *Journal of Hazardous Materials*, 324, 3–14.
- Lanari, M. C., Hewavitharana, A. K., Becu, C., & De Jong, S. (2004). Effect of dietary tocopherols and tocotrienols on the antioxidant status and lipid stability of chicken. *Meat Science*, 68(2), 155–162.

- Larsen, E. H., Hansen, M., Fan, T., & Vahl, M. (2001). Speciation of selenoamino acids, selenonium ions and inorganic selenium by ion exchange HPLC with mass spectrometric detection and its application to yeast and algae. *Journal of Analytical Atomic Spectrometry*, 16, 1403–1408.
- Leeson, S., Namkung, H., Caston, L., Durosoy, S., & Schlegel, P. (2008). Comparison of selenium levels and sources and dietary fat quality in diets for broiler breeders and layer hens. *Poultry Science*, 87, 2605–2612.
- Leshchinsky, T. V., & Klasing, K. C. (2003). Profile of chicken cytokines induced by lipopolysaccharide is modulated by dietary alpha-tocopheryl acetate. *Poultry Science*, 82(8), 1266–1273.
- Li, J.-L., & Sunde, R. (2016). Selenoprotein transcript level and enzyme activity as biomarkers for selenium status and selenium requirements of chickens (*Gallus gallus*). *PloS One*, 11(4), e0152392.
- Li, J. L., Zhang, L., Yang, Z. Y., Zhang, Z. Y., Jiang, Y., Gao, F., & Zhou, H. G. (2017). Effects of different selenium sources on growth performance , antioxidant capacity and meat quality of local chinese subei chickens. *Biological Trace Element Research*.
- Liao, X., Lu, L., Li, S., Liu, S., Zhang, L., Wang, G., ... Luo, X. (2012). Effects of selenium source and level on growth performance, tissue selenium concentrations, antioxidation, and immune functions of heat-stressed broilers. *Biological Trace Element Research*, 150(1–3), 158–165.
- Liesegang, A., Staub, T., Wichert, B., Wanner, M., & Kreuzer, M. (2008). Effect of vitamin e supplementation of sheep and goats fed diets supplemented with polyunsaturated fatty acids and low in Se. *Journal of Animal Physiology and Animal Nutrition*, 92(3), 292–302.
- Lin, S. L., Wang, C. W., Tan, S. R., Liang, Y., Yao, H. D., Zhang, Z. W., & Xu, S. W. (2014). Selenium deficiency inhibits the conversion of thyroidal thyroxine (T4) to triiodothyronine (T3) in chicken thyroids. *Biological Trace Element Research*, 161(3), 263–271.
- Lisiak, D., Janiszewski, P., Blicharski, T., Borzuta, K., Grześkowiak, E., Lisiak, B., ... Hammermeister, A. (2014). Effect of selenium supplementation in pig feed on slaughter value and physicochemical and sensory characteristics of meat. *Annals of Animal Science*, 14(1), 213–222.
- Liu, C. P., Fu, J., Lin, S. L., Wang, X. S., & Li, S. (2014). Effects of dietary selenium deficiency on mRNA levels of twenty-one selenoprotein genes in the liver of layer chicken. *Biological Trace Element Research*, 159(1–3), 192–198.

- Liu, Q., Lanari, M. C., & Schaefer, D. M. (1995). A review of dietary vitamin E supplementation for improvement of beef quality. *Journal of Animal Science*, 73(10), 3131–3140.
- Losi, M., & Frankenberger Jr., W. T. (1997). Reduction of selenium oxyanions by enterobacter cloacae strain SLD1a-1: reduction of selenate to selenite. *Environmental Toxicology and Chemistry*, 16(9), 1851–1858.
- Lu, Z., Jin, M., Huang, M., Wang, Y., & Wang, Y. (2013). Bioactivity of selenium-enriched exopolysaccharides produced by Enterobacter cloacae Z0206 in broilers. *Carbohydrate Polymers*, 96(1), 131–136.
- Luan, Y., Zhao, J., Yao, H., Zhao, X., Fan, R., Zhao, W., ... Xu, S. (2016). Selenium deficiency influences the mRNA expression of selenoproteins and cytokines in chicken erythrocytes. *Biological Trace Element Research*, 171(2), 427–436.
- Łukaszewicz, E., Kowalczyk, A., & Jerysz, A. (2016). Effect of dietary selenium and vitamin E on chemical and fatty acid composition of goose meat and liver. *Animal Science Papers and Reports*, 34(2), 181–194.
- Lv, C. H., Wang, T., Regmi, N., Chen, X., Huang, K., & Liao, S. F. (2015). Effects of dietary supplementation of selenium-enriched probiotics on production performance and intestinal microbiota of weanling piglets raised under high ambient temperature. *Journal of Animal Physiology and Animal Nutrition*, 99(6), 1161–1171.
- Lyons, M. P., Papazyan, T. T., & Surai, P. F. (2007). Selenium in food chain and animal nutrition: Lessons from nature - Review. *Asian-Australasian Journal of Animal Sciences*, 20(7), 1135–1155.
- Macit, M., Aksakal, V., Emsen, E., Aksu, M. I., Karaoglu, M., & Esenbuga, N. (2003). Effects of vitamin E supplementation on performance and meat quality traits of Morkaraman male lambs. *Meat Science*, 63(1), 51–55.
- Macpherson, A. (1994). Selenium, vitamin E and biological oxidation. In D.J. Cole and P.J. Garnsworthy (Ed.), *Recent Advances in Animal Nutrition* (pp. 3–30). Oxford: Butterworth-Heinemann.,
- Mahan, D. C., Cline, T. R., & Richert, B. (1999). Effects of dietary levels of selenium-enriched yeast and sodium selenite as selenium sources fed to growing-finishing pigs on performance, tissue selenium, serum glutathione peroxidase activity, carcass characteristics, and loin quality. *Journal of Animal Science*, 77, 2172–2179.
- Mahan, D. C., & Parrett, N. A. (1996). Evaluating the efficacy of selenium-enriched yeast and sodium selenite on tissue selenium retention and serum glutathione peroxidase activity in grower and finisher swine. *Journal of Animal Science*, 74(12), 2967–2974.

- Mahmoud, K. Z., & Edens, F. W. (2003). Influence of selenium sources on age-related and mild heat stress-related changes of blood and liver glutathione redox cycle in broiler chickens (*Gallus domesticus*). *Comparative Biochemistry and Physiology - B Biochemistry and Molecular Biology*, 136(4), 921–934.
- Maini, S., Rastogi, S. K., Korde, J. P., Madan, A. K., & Shukla, S. K. (2007). Evaluation of oxidative stress and its amelioration through certain antioxidants in broilers during summer. *The Journal of Poultry Science*, 44, 339–347.
- Marsh, J. A., Dietert, R. R., & Combs Jr, G. F. (1981). Influence of dietary selenium and vitamin E on the humoral immune response of the chick. *Proceedings of the Society for Experimental Biology and Medicine*, 166(2), 228–236.
- Maysa, M., El-Sheikh, A., & Abdalla, E. (2009). The effect of organic selenium supplementation on productive and physiological performance in a local strain of chicken. 1-the effect of organic selenium (Sel-PlexTM). *Egyptian Poultry Science*, 29(4), 1061–1084.
- Mcsheehy, S., Yang, L., Sturgeon, R., & Mester, Z. (2005). Determination of methionine and selenomethionine in selenium-enriched yeast by species-specific isotope dilution with liquid chromatography-mass spectrometry and inductively coupled plasma mass spectrometry detection. *Analytical Chemistry*, 77(1), 344–349.
- Mego, M., Májek, J., Končeková, R., Ebringer, L., Čierniková, S., Rauko, P., ... Zajac, V. (2005). Intramucosal bacteria in colon cancer and their elimination by probiotic strain *Enterococcus faecium* M-74 with organic selenium. *Folia Microbiologica*, 50(5), 443–447.
- Mehdi, Y., Hornick, J. L., Istasse, L., & Dufrasne, I. (2013). Selenium in the environment, metabolism and involvement in body functions. *Molecules*, 18(3), 3292–3311.
- Méplan, C. (2011). Trace elements and ageing, a genomic perspective using selenium as an example. *Journal of Trace Elements in Medicine and Biology*, 25, 11–16.
- Meschy, F. (2010). *Nutrition minérale des ruminants*. Versaille, France,: Editions Quae.
- Miezeliene, A., Alencikiene, G., Gruzauskas, R., & Barstys, T. (2011). The effect of dietary selenium supplementation on meat quality of broiler chickens. *Biotechnology, Agronomy, Society and Environment*, 15, 61–69.
- Mitruka, B., & Rawlsley, H. (1981). *Clinical biochemical and hematological reference values in normal experimental animals* (2nd ed). Masson Publishing U.S.A.

- Mogna, L., Nicola, S., Pane, M., Lorenzini, P., Strozzi, G., & Mogna, G. (2012). Selenium and zinc internalized by lactobacillus buchneri Lb26 (DSM 16341) and bifidobacterium lactis Bb1 (DSM 17850). *Journal of Clinical Gastroenterology*, 46, S41–S45.
- Mohapatra, P., Swain, R., Mishra, S., Behera, T., Swain, P., Mishra, S., ... Jayasankar, P. (2014). Effects of dietary nano-selenium on tissue selenium deposition, antioxidant status and immune functions in layer chicks. *International Journal of Pharmacology*, 10(3), 160–167.
- Mohiti-Asli, M., Shariatmadari, F., & Lotfollahian, H. (2010). The influence of dietary vitamin E and selenium on egg production parameters, serum and yolk cholesterol and antibody. *Arch. Geflügelk.*, 74(1), 43–50.
- Molan, A. L. (2013). Antioxidant and prebiotic activities of selenium-containing green tea. *Nutrition*, 29(2), 476–477.
- Molan, A. L., Flanagan, J., Wei, W., & Moughan, P. J. (2009). Selenium-containing green tea has higher antioxidant and prebiotic activities than regular green tea. *Food Chemistry*, 114(3), 829–835.
- Monsen, E. R. (2000). Dietary reference intakes for the antioxidant nutrients: vitamin C, vitamin E, selenium, and carotenoids. *Journal of the American Dietetic Association*. 637–640.
- Montgomery, J. B., Wichtel, J. J., Wichtel, M. G., McNiven, M. A., McClure, J. T., Markham, F., & Horohov, D. W. (2012). Effects of selenium source on measures of selenium status and immune function in horses. *Canadian Journal of Veterinary Research*, 76(4), 281–291.
- Moore, M. D., & Kaplan, S. (1992). Identification of intrinsic high-level resistance to rare-earth oxides and oxyanions in members of the class Proteobacteria: characterization of tellurite, selenite and rhodium sesquioxide reduction in Rhodobacter sphaeroides. *Journal of Bacteriology*, 174, 1505–1514.
- Mostert, V. (2000). Selenoprotein P: properties, functions, and regulation. *Archives of Biochemistry and Biophysics*, 376(2), 433–438.
- Mykkanen, H. ., & Wasserman, R. . (1989). Uptake of 75Se-selenite by brush border membrane vesicles from chick duodenum stimulated by vitamin D. *The Journal of Nutrition*, 119(2), 242–247.
- Nam, H.-M., Srinivasan, V., Gillespie, B. E., Murinda, S. E., & Oliver, S. P. (2005). Application of SYBR green real-time PCR assay for specific detection of *Salmonella* spp. in dairy farm environmental samples. *International Journal of Food Microbiology*, 102(2), 161–171.

- Nanchariah, Y. V., & Lens, P. N. L. (2015). The ecology and biotechnology of selenium-respiring bacteria. *Microbiology and Molecular Biology Reviews*, 79, 61–80.
- Navidshad, B., Liang, J. B., & Jahromi, M. F. (2012). Correlation coefficients between different methods of expressing bacterial quantification using real time PCR. *International Journal of Molecular Sciences*, 13(2), 2119–2132.
- NRC (National Research Council). (1994). *Nutrient requirements of poultry* (9th ed.). Washington, DC, USA.: National Academy Press.
- Okunlola, D., AkandeT.O, & Nuga, H. (2015). Haematological and serum characteristics of broiler birds fed diets supplemented with varying levels of selenium powder. *Journal of Biology, Agriculture and Healthcare*, 5(1), 107–111.
- Oliveira, T. F. B., Rivera, D. F. R., Mesquita, F. R., Braga, H., Ramos, E. M., & Bertechini, A. G. (2014). Effect of different sources and levels of selenium on performance, meat quality, and tissue characteristics of broilers. *Journal of Applied Poultry Research*, 23(1), 15–22.
- Oremland, R. S., Herbel, M. J., Blum, J. S., Langley, S., Beveridge, T. J., Ajayan, P. M., ... Curran, S. (2004). Structural and spectral features of selenium nanospheres produced by se-respiring bacteria. *Applied and Environmental Microbiology*, 70(1), 52–60.
- Oshima, S., Osame, S., Ichijo, S., (1990). Changes in serum selenium levels and blood glutathione peroxidase activities in pregnant ewes injected with organic and inorganic selenium. *Journal of the Japanese Veterinary Medical Association*, 43, 784–788.
- Ozkan, A., Fiskin, K., & Ayhan, A. (2007a). Effect of vitamin E and selenium on antioxidant enzymes in brain, kidney and liver of cigarette smoke-exposed mice. *Biologia*, 62(3), 360–364.
- Ozkan, S., Malayoğlu, H. B., Yalçın, S., Karadas, F., Koçtürk, S., Cabuk, M., ... Ergül, M. (2007b). Dietary vitamin E (alpha-tocopherol acetate) and selenium supplementation from different sources: performance, ascites-related variables and antioxidant status in broilers reared at low and optimum temperatures. *British Poultry Science*, 48(5), 580–593.
- Palomo, M., Gutiérrez, A. M., Pérez-Conde, M. C., Cámaras, C., & Madrid, Y. (2014). Se metallomics during lactic fermentation of Se-enriched yogurt. *Food Chemistry*, 164, 371–379.
- Papp, L., Lu, J., Holmgren, A., & Khanna, K. (2007). From selenium to selenoproteins: synthesis, identity, and their role in human health. *Antioxidants & Redox Signaling*, 9, 775–806.

- Pappas, A. C., Zoidis, E., Surai, P. F., & Zervas, G. (2008). selenoproteins and maternal nutrition. *Comparative Biochemistry and Physiology, Part B*, 151(4), 361–372.
- Payne, R. L., Lavergne, T. K., & Southern, L. L. (2005). Effect of inorganic versus organic selenium on hen production and egg selenium concentration. *Poultry Science*, 84(2), 232–7.
- Payne, R. L., & Southern, L. L. (2005). Comparison of inorganic and organic selenium sources for broilers. *Poultry Science*, 84(6), 898–902.
- Pereira, A. S. C., Santos, M. V. dos, Aferri, G., Corte, R. R. P. da S., Silva, S. da L. e., De Freitas Júnior, J. E., ... Rennó, F. P. (2012). Lipid and selenium sources on fatty acid composition of intramuscular fat and muscle selenium concentration of Nellore steers. *Revista Brasileira de Zootecnia*, 41(11), 2357–2363.
- Perić, L., Milošević, N., Žikić, D., Kanački, Z., Džinić, N., Nollet, L., & Spring, P. (2009). Effect of selenium sources on performance and meat characteristics of broiler chickens. *Journal of Applied Poultry Research*, 18(3), 403–409.
- Pesti, G. M., & Combs Jr, G. F. (1976). Studies on the enteric absorption of selenium in the chick using localized coccidial infections. *Poultry Science*, 55(6), 2265–2274.
- Pfaffl, M. W. (2001). A new mathematical model for relative quantification in real-time RT-PCR. *Nucleic Acids Research*, 29(9), e45.
- Pieniz, S., Andreazza, R., Pereira, J. Q., De Oliveira Camargo, F. A., & Brandelli, A. (2013). Production of selenium-enriched biomass by *Enterococcus durans*. *Biological Trace Element Research*, 155(3), 447–454.
- Połatajko, A., Śliwka-Kaszyńska, M., Dernovics, M., Ruzik, R., Ruiz Encinar, J., & Szpunar, J. (2004). A systematic approach to selenium speciation in selenized yeast. *Journal of Analytical Atomic Spectrometry*, 19(1), 114.
- Ponce-De Leon, C., Bayon, M., Paquin, C., & Caruso, J. (2002). Selenium incorporation into *Saccharomyces cerevisiae* cells: A study of different incorporation methods. *Journal of Applied Microbiology*, 92, 602–610.
- Pronina, N. A., Kovshova, Y. I., Popova, V. V., Lapin, A. B., Alekseeva, S. G., Baum, R. F., ... Tsoglin, L. N. (2002). The effect of selenite ions on growth and selenium accumulation in *Spirulina platensis*. *Russian Journal of Plant Physiology*, 49(2), 235–241.
- Quinteiro-Filho, W. M., Ribeiro, A., Ferraz-De-Paula, V., Pinheiro, M. L., Sakai, M., Sá, L. R. M., ... Palermo-Neto, J. (2010). Heat stress impairs performance parameters, induces intestinal injury, and decreases macrophage activity in broiler chickens. *Poultry Science*, 89(9), 1905–1914.

- Rajion, M. A., Mclean, J. G., & Cahill, R. N. P. (1985). Essential Fatty Acids in the Fetal and Newborn Lamb. *Australian Journal of Biological Sciences*, 57, 33–40.
- Rama Rao, S. V., Prakash, B., Raju, M. V. L. N., Panda, A. K., Poonam, S., & Murthy, O. K. (2013). Effect of supplementing organic selenium on performance, carcass traits, oxidative parameters and immune responses in commercial broiler chickens. *Asian-Australasian Journal of Animal Sciences*, 26(2), 247–252.
- Ranjard, L., Prigent-combaret, C., Nazaret, S., & Cournoyer, B. (2002). Methylation of Inorganic and Organic Selenium by the bacterial thiopurine methyltransferase. *Journal of Bacteriology*, 184(11), 3146–3149.
- Rasmussen, A., & Andersson, M. (1996). New method for determination of drip loss in pork muscles. In *Proceedings of the 42nd International Congress of Meat Science and Technology (ICoMST '96)* (pp. 286–287). Lillehammer, Norway.
- Rasmussen, M., & Thomas, C. (1997). Rumen Bacteria That Reduce Selenium. In *American Society of Microbiologists Abstracts*.
- Rasoli, M., Yeap, S. K., Tan, S. W., Moeini, H., Ideris, A., Bejo, M. H., ... Omar, A. R. (2014). Alteration in lymphocyte responses, cytokine and chemokine profiles in chickens infected with genotype VII and VIII velogenic Newcastle disease virus. *Comparative Immunology, Microbiology and Infectious Diseases*, 37(1), 11–21.
- Rayman, M. P. (2000). The importance of selenium to human health. *The Lancet*, 356(9225), 233–241.
- Rayman, M. P. (2004). The use of high-selenium yeast to raise selenium status: how does it measure up? *The British Journal of Nutrition*, 92(4), 557–573.
- Rayman, M. P. (2012). Selenium and human health. *The Lancet*, 379(9822), 1256–1268.
- Read-Snyder, J., Edens, F. W., Cantor, A. H., Pescatore, A. J., & Pierce, J. L. (2009). Effect of dietary selenium on small intestine villus integrity in reovirus-challenged broilers. *International Journal of Poultry Science*, 8(9), 829–835.
- Reeves, M. A., & Hoffmann, P. R. (2009). The human selenoproteome: recent insights into functions and regulation. *Cellular and Molecular Life Sciences*, 66(15), 2457–2478.
- Ren, Z., Zhao, Z., Wang, Y., & Huang, K. (2011). Preparation of selenium/zinc-enriched probiotics and their effect on blood selenium and zinc concentrations, antioxidant capacities, and intestinal microflora in canine. *Biological Trace Element Research*, 141(1–3), 170–183.

- Ridnour, L. A., Isenberg, J. S., Espey, M. G., Thomas, D. D., Roberts, D. D., & Wink, D. A. (2005). Nitric oxide regulates angiogenesis through a functional switch involving thrombospondin-1. *Proceedings of the National Academy of Sciences of the United States of America*, 102(37), 13147–13152.
- Rooke, J. A., Robinson, J. J., & Arthur, J. R. (2004). Effects of vitamin E and selenium on the performance and immune status of ewes and lambs. *The Journal of Agricultural Science*, 142(3), 253–262.
- Rother, M. (2012). Selenium metabolism in prokaryotes. In D.L. Hatfield, M.J. Berry and V.N. Gladyshev (Ed.), *Selenium* (pp. 457–470). New York: Springer.
- Rotruck, J. T., Pope, A. L., Ganther, H. E., Swanson, A. B., Hafeman, D. G., & Hoekstra, W. (1973). Selenium : Biochemical role as a component of glutathione peroxidase. *Science*, 179(4073), 588–590.
- Rovenský, J., Švík, K., Stančíková, M., Ištok, R., Ebringer, L., & Ferenčík, M. (2002). Treatment of experimental adjuvant arthritis with the combination of methotrexate and lyophilized Enterococcusfaecium enriched with organic selenium. *Folia Microbiol*, 47(5), 573–578.
- Roy, M., Kiremidjian-Schumacher, L., Wishe, H. I., Cohen, M. W., & Stotzky, G. (1992). Effect of selenium on the expression of high affinity interleukin 2 receptors. In *Proceedings of the Society for Experimental Biology and Medicine* (Vol. 200, pp. 36–43). SAGE Publications.
- Rutkowski, M., & Grzegorczyk, K. (2007). Modifications of spectrophotometric methods for antioxidative vitamins determination convenient in analytic practice. *ACTA Scientiarum Polonorum Technologia Alimentaria*, 6(13), 17–28.
- Safarizadeh, A., & Zakeri, A. (2013). The effect of vitamin A and complex of vitamin E and selenium on growth factors and Humoral immunity in broiler chickens. *European Journal of Experimental Biology*, 3(4), 99–102.
- Sahin, K., Sahin, N., Onderci, M., Yaralioglu, S., & Kucuk, O. (2001). Protective role of supplemental vitamin e on lipid peroxidation, vitamins E, a and some mineral concentrations of broilers reared under heat stress. *Veterinarni Medicina*, 46(5), 140–144.
- Santini, F., Pinchera, A., Ceccarini, G., Castagna, M., Rosellini, V., Mammoli, C., ... Chiovato, L. (2001). Evidence for a role of the type III-iodothyronine deiodinase in the regulation of 3, 5, 3'-triiodothyronine content in the human central nervous system. *European Journal of Endocrinology*, 144(6), 577–583.
- Sareen, S., & Gropper, J. L. S. (2012). *Advanced nutrition and human metabolism* (6th). Cengage Learning.

- Sarret, G., Avoscan, L., Marie, C., Richard, C., Nicolas, G., Francine, C., ... Barbara, G. (2005). Chemical Forms of Selenium in the Metal-Resistant Bacterium. *Applied and Environmental Microbiology*, 71(5), 2331–2337.
- SAS. (2005). SAS software, version 9.2, SAS Institute Cary, NC.
- Sas, B. (1993). Contribution to the pathobiochemistry of furazolidone-induced oxidative toxicity in chickens. *Acta Veterinaria Hungarica*, 41(1–2), 103–121.
- Sazili, A. Q., Parr, T., Sensky, P. L., Jones, S. W., Bardsley, R. G., & Buttery, P. J. (2005). The relationship between slow and fast myosin heavy chain content, calpastatin and meat tenderness in different ovine skeletal muscles. *Meat Science*, 69(1), 17–25.
- Schäfer, K., Kyriakopoulos, A., Gessner, H., Grune, T., & Behne, D. (2004). Effects of selenium deficiency on fatty acid metabolism in rats fed fish oil-enriched diets. *Journal of Trace Elements in Medicine and Biology*, 18(1), 89–97.
- Schalm, O., Jain, N., & Carol, E. (1975). *Veterinary haematology* (3rd Editio). Lea and Febiger Philadelphia.
- Schrauzer, G. (2001). Nutritional selenium supplements: product types, quality, and safety. *The Journal of the American College of Nutrition*, 20, 1–4.
- Schrauzer, G. N. (2000). Selenomethionine: a review of its nutritional significance, metabolism and toxicity. *The Journal of Nutrition*, 130(7), 1653–6.
- Schrauzer, G. N. (2003). The nutritional significance, metabolism and toxicology of selenomethionine. *Advances in Food and Nutrition Research*, 47(3), 73–112.
- Schrauzer, G. N., & Surai, P. F. (2009). Selenium in human and animal nutrition: resolved and unresolved issues. A partly historical treatise in commemoration of the fiftieth anniversary of the discovery of the biological essentiality of selenium, dedicated to the memory of Klaus Schwarz (1914-1978) on the occasion of the thirtieth anniversary of his death. *Critical Reviews in Biotechnology*, 29(1), 2–9.
- Schwarz, K., & Foltz, C. M. (1957). Selenium as an integral part of factor 3 against dietary necrotic liver degeneration. *Journal of the American Chemical Society*, 79(12), 3292–3293.
- Seiler, A., Schneider, M., Förster, H., Roth, S., Wirth, E. K., Culmsee, C., ... Wurst, W. and Bornkamm, G. W. (2008). Glutathione peroxidase 4 senses and translates oxidative stress into 12/15-lipoxygenase dependent-and AIF-mediated cell death. *Cell Metabolism*, 8(3), 237–248.
- Sen, C. K., & Packer, L. (1996). Antioxidant and redox regulation of gene transcription. *The FASEB Journal*, 10(7), 709–20.

- Sevcikova, S., Skřivan, M., Dlouhá, G., & Koucký, M. (2006). The effect of selenium source on the performance and meat quality of broiler chickens. *Czech Journal of Animal Science*, 51(10), 449–457.
- Shah, M. K., Khan, A., Rizvi, F., & Siddique, M. (2007). Effect of cypermethrin on clinico-haematological parameters in rabbits. *Pakistan Veterinary Journal*, 27(4), 171–175.
- Shahverdi, A. R., Shakibaie, M., & Nazari, P. (2011). Basic and practical procedures for microbial synthesis of nanoparticles. In R. Mahendra and N. Duran (Ed.), *Metal nanoparticles in microbiology* (pp. 177–195). Springer, Berlin, Heidelberg.
- Shimaa, A. E., Ryad, K., Emad, A. H., & Abdelhakeem, E. (2014). Protective effects of selenium and alpha-tocopherol against lead-induced hepatic and renal toxicity in oreochromis niloticus. *Journal of Aquaculture Research & Development*, 6(1), 1–5.
- Simmons, D. B., & Wallschläger, D. (2005). A critical review of the biogeochemistry and ecotoxicology of selenium in lotic and lentic environments. *Environmental Toxicology and Chemistry*, 24, 1331–1343.
- Singh, H., Sodhi, S., & Kaur, R. (2006). Effects of dietary supplements of selenium, vitamin E or combinations of the two on antibody responses of broilers. *British Poultry Science*, 47(6), 714–719.
- Singh, V. P., Sahu, D. S., Singh, M. K., & Manoj, J. (2016). Effect of supplementation of Selenium and Ashwagandha (Withania somnifera) on some haematological and immunological parameters of broiler chickens. *Scholars Journal of Agriculture and Veterinary Sciences*, 3(6), 406–410.
- Skřivan, M., Marounek, M., Dlouhá, G., & Ševčíková, S. (2008). Dietary selenium increases vitamin E contents of egg yolk and chicken meat. *British Poultry Science*, 49(4), 482–486.
- Skřivan, M., Marounek, M., Englmaierová, M., & Skřivanová, E. (2012). Influence of dietary vitamin C and selenium, alone and in combination, on the composition and oxidative stability of meat of broilers. *Food Chemistry*, 130(3), 660–664.
- Steinbrenner, H., & Sies, H. (2009). Protection against reactive oxygen species by selenoproteins. *Biochimica et Biophysica Acta*, 1790, 1478–1485.
- Stępińska, M., Emilia, M., & Jan, J. (2012). The effect of dietary selenium source on embryonic development in turkeys. *Folia Biologica*, 60(3), 235–241.
- Stolz, J. F., Basu, P., & Oremland, R. S. (2002). Microbial transformation of elements: The case of arsenic and selenium. *International Microbiology*, 5(4), 201–207.

- Suchý, P., Straková, E., & Herzig, I. (2014). Selenium in poultry nutrition: A review. *Czech Journal of Animal Science*, 59(11), 495–503.
- Suhajda, A., Hegoczki, J., Janzso, B., Pais, I., & Vereczkey, G. (2000). Preparation of selenium yeasts I. Preparation of selenium-enriched *Saccharomyces cerevisiae*. *Journal of Trace Elements in Medicine and Biology*, 14, 43–47.
- Sunde, R. A., Li, J., & Taylor, R. M. (2016). Insights for setting of nutrient requirements , gleaned by comparison of selenium status biomarkers in turkeys and chickens versus rats .. *Advances in Nutrition: An International Review Journal*, 7(6), 1129–1138.
- Sunde, R. A., & Raines, A. M. (2011). Selenium regulation of the selenoprotein and non-selenoprotein transcriptomes in rodents. *Advances in Nutrition: An International Review Journal*, 2(2), 138–150.
- Surai, P. F. (2006). *Selenium in nutrition and health*. Nottingham, UK.: Nottingham University Press.
- Surai, P. F. (2002). *Natural antioxidants in avian nutrition and reproduction*. Nottingham University Press: Nottingham, UK.
- Surai, P. F. (2007). Natural antioxidants in poultry nutrition: New developments. *16th European Symposium on Poultry Nutrition*, (1998), 669–676.
- Surai, P. F., & Dvorska, J. E. (2002). Effect of selenium and vitamin E content of the diet on lipid peroxidation in breast muscle tissue of broiler breeder hens during storage. In *Proceeding Australian Poultry Science Symposium* (p. 14).
- Surai, P. F., & Fisinin, V. I. (2014). Selenium in poultry breeder nutrition: An update. *Animal Feed Science and Technology*, 191, 1–15.
- Surai, P. F., Pappas, A., Karadas, F., Papazyan, T., & Fisinin, V. (2010). Selenium enigma: Health implications of an inadequate supply. In F. De Meester, S. Zibadi and D. Ross-Watson (Ed.), *Modern Dietary Fat Intakes in Disease Promotion* (pp. 379–403). Springer.
- Suttle, N. (2010). *Mineral nutrition of livestock* (4th Ed). London, UK.: MPG Books Group.
- Suzuki, K. T. (2005). Metabolomics of Selenium: Se Metabolites Based on Speciation Studies. *Journal of Health Science*, 51(2), 107–114.
- Suzuki, K. T., Itoh, M., & Ohmichi, M. (1995). Selenium distribution and metabolic profile in relation to nutritional selenium status in rats. *Toxicology*, 103, 157–165.

- Suzuki, K. T., Doi, C., & Suzuki, N. (2006). Metabolic pathway of 76 Semethylselenocysteine compared with that of 77Se-selenomethionine and 82Se selenite. *Toxicology and Applied Pharmacology*, 217, 185–195.
- Suzuki, K. T., Tsuji, Y., Ohta, Y., & Suzuki, N. (2008). Preferential organ distribution of methylselenol source Se-methylselenocysteine relative to methylseleninic acid. *Toxicology and Applied Pharmacology*, 227, 76–83.
- Suzuki, Y., Hashiura, Y., Matsumura, K., Matsukawa, T., Shinohara, A., & Furuta, N. (2010). Dynamic pathways of selenium metabolism and excretion in mice under different selenium nutritional statuses. *Metalomics*, 2(2), 126–132.
- Svoboda, M., Fajt, Z., Baňoch, T., Drábek, J., & Saláková, A. (2010). The use of selenium enriched Enterococcus faecium as an alternative selenium source for growing-finishing pigs. *Acta Veterinaria Brno*, 79(4), 511–517.
- Swain, B. K., Johri, T. S., & Majumdar, S. (2000). Effect of supplementation of vitamin E, selenium and their different combinations on the performance and immune response of broilers. *British Poultry Science*, 41(3), 287–292.
- Teo, A., Hon, S.-M., Se, C.-Y., & Ian, H. (2005). Production of organic and inorganic selenium compounds by lactic acid bacteria. U.S.Patent Application No 11/243,391.
- Thibodeau, A., Letellier, A., Yergeau, É., Larrivière-Gauthier, G., & Fraval, P. (2017). Lack of evidence that selenium-yeast improves chicken health and modulates the caecal microbiota in the context of colonization by *Campylobacter jejuni*. *Frontiers in Microbiology*, 8, 1–9.
- Thiry, C., Ruttens, A., De Temmerman, L., Schneider, Y.-J., & Pussemier, L. (2012). Current knowledge in species-related bioavailability of selenium in food. *Food Chemistry*, 130(4), 767–784.
- Thomas, P., Sekhar, A. C., Uperti, R., Mujawar, M. M., & Pasha, S. S. (2015). Optimization of single plate-serial dilution spotting (SP-SDS) with sample anchoring as an assured method for bacterial and yeast cfu enumeration and single colony isolation from diverse samples. *Biotechnology Reports*, 8, 45–55.
- Thomson, C. (2004). Assessment of requirements for selenium and adequacy of selenium status: a review. *European Journal of Clinical Nutrition*, 58, 391–402.
- Touat-Hamici, Z., Legrain, Y., Bulteau, A. L., & Chavatte, L. (2014). Selective up-regulation of human selenoproteins in response to oxidative stress. *Journal of Biological Chemistry*, 289(21), 14750–14761.

- Touchette, K. J., Carroll, J. A., Allee, G. L., Matteri, R. L., Dyer, C. J., Beausang, L. A., & Zannelli, M. E. (2002). Effect of spray-dried plasma and lipopolysaccharide exposure on weaned pigs: I. Effects on the immune axis of weaned pigs. *Journal of Animal Science*, 80(2), 494–501.
- Tres, A., Bou, R., Codony, R., & Guardiola, F. (2010). Moderately oxidized oils and dietary zinc and α -tocopheryl acetate supplementation: effects on the oxidative stability of rabbit plasma, liver, and meat. *Journal of Agricultural and Food Chemistry*, 58(16), 9112–9119.
- Tsuji, P. A., Carlson, B. A., Anderson, C. B., Seifried, H. E., Hatfield, D. L., & Howard, M. T. (2015). Dietary selenium levels affect selenoprotein expression and support the interferon- γ and IL-6 immune response pathways in mice. *Nutrients*, 7(8), 6529–6549.
- Underwood, J., & Suttle, F. (1999). The detection and correction of mineral imbalances. In *The mineral nutrition of livestock* (Vol. 3, p. 47).
- Upton, J. R., Edens, F. W., & Ferket, P. R. (2009). The effects of dietary oxidized fat and selenium source on performance, glutathione peroxidase, and glutathione reductase activity in broiler chickens. *Journal of Applied Poultry Research*, 18(2), 193–202.
- Vaishnav, R. A., Getchell, M. L., Huang, L., Hersh, M. A., Stromberg, A. J., & Getchell, T. V. (2008). Cellular and molecular characterization of oxidative stress in olfactory epithelium of Harlequin mutant mouse. *Journal of Neuroscience Research*, 68(1), 165–182.
- Wahlen, R., Evans, L., Turner, J., & Hearn, R. (2005). The use of collision/reaction cell ICP-MS for the determination of elements in blood and serum samples. *Spectroscopy*, 20(12), 1–8.
- Waldenstedt, L. (2006). Nutritional factors of importance for optimal leg health in broilers: A review. *Animal Feed Science and Technology*, 126(3–4), 291–307.
- Wang, C., Wang, H., Luo, J., Hu, Y., Wei, L., Duan, M., & He, H. (2009). Selenium deficiency impairs host innate immune response and induces susceptibility to *Listeria monocytogenes* infection. *BMC Immunology*, 10(1), 55.
- Wang, W. X., & Fisher, N. S. (1999). Assimilation efficiencies of chemical contaminants in aquatic invertebrates: A synthesis. *Environmental Toxicology and Chemistry*, 18(9), 2034–2045.
- Wang, Y. (2009). Differential effects of sodium selenite and nano-se on growth performance, tissue se distribution, and glutathione peroxidase activity of avian broiler. *Biological Trace Element Research*, 128(2), 184–190.

- Wang, Y. B., & Xu, B. H. (2008). Effect of different selenium source (sodium selenite and selenium yeast) on broiler chickens. *Animal Feed Science and Technology*, 144(3–4), 306–314.
- Wang, Y., Zhan, X., Zhang, X., Wu, R., & Yuan, D. (2011a). Comparison of different forms of dietary selenium supplementation on growth performance, meat quality, selenium deposition, and antioxidant property in broilers. *Biological Trace Element Research*, 143(1), 261–273.
- Wang, Y., Zhan, X., Yuan, D., Zhang, X., & Wu, R. (2011b). Influence of dietary selenomethionine supplementation on performance and selenium status of broiler breeders and their subsequent progeny. *Biological Trace Element Research*, 143(3), 1497–1507.
- Wang, Y. X., Zhan, X., Yuan, D., Zhang, X. W., & Wu, R. J. (2011c). Effects of selenomethionine and sodium selenite supplementation on meat quality, selenium distribution and antioxidant status in broilers. *Czech Journal of Animal Science*, 56(7), 305–313.
- Wang, Z. G., Pan, X. J., Zhang, W. Q., Peng, Z. Q., Zhao, R. Q., & Zhou, G. H. (2010). Methionine and selenium yeast supplementation of the maternal diets affects antioxidant activity of breeding eggs. *Poultry Science*, 89(5), 931–937.
- Wolfram, S., Berger, B., Grenacher, B., & E., S. (1989). Transport of seleno amino acids and their sulphur analogues across the intestinal brush border membrane. *Journal of Nutrition*, 119, 706–712.
- Wright, P., & Bell, M. (1966). Comparative metabolism of selenium and tellurium in sheep and swine. *American Journal of Physiology Legacy Content*, 211(1), 6–10.
- Wu, R., Zhan, X., Wang, Y., Zhang, X., Wang, M., & Yuan, D. (2011). Effect of different selemethionine forms and levels on performance of breeder hens and se distribution of tissue and egg inclusion. *Biological Trace Element Research*, 143(2), 923–931.
- Xia, S. K., Chen, L., & Liang, J. Q. (2007). Enriched selenium and its effects on growth and biochemical composition in *Lactobacillus bulgaricus*. *Journal of Agricultural and Food Chemistry*, 55, 2413–2417.
- Xu, C. L., Wang, Y. Z., Jin, M. L., & Yang, X. Q. (2009). Preparation, characterization and immunomodulatory activity of selenium-enriched exopolysaccharide produced by bacterium *Enterobacter cloacae* Z0206. *Bioresource Technology*, 100(6), 2095–2097.
- Yang, J., Huang, K., Qin, S., Wu, X., Zhao, Z., & Chen, F. (2009). Antibacterial action of selenium-enriched probiotics against pathogenic *Escherichia coli*. *Digestive Diseases and Sciences*, 54(2), 246–254.

- Yang, J., Liu, L., Sheikhahmadi, A., Wang, Y., Li, C., Jiao, H., ... Song, Z. (2015). Effects of corticosterone and dietary energy on immune function of broiler chickens. *PLoS One*, 10(3), 1–14.
- Yang, Y. R., Meng, F. C., Wang, P., Jiang, Y. B., Yin, Q. Q., Chang, J., ... Liu, J. X. (2012). Effect of organic and inorganic selenium supplementation on growth performance, meat quality and antioxidant property of broilers. *African Journal of Biotechnology*, 11(12), 3031–3036.
- Yazdi, M. H., Mahdavi, M., Kheradmand, E., & Shahverdi, A. R. (2012). The preventive oral supplementation of a selenium nanoparticle-enriched probiotic increases the immune response and lifespan of 4T1 breast cancer bearing mice. *Arzneimittel-Forschung/Drug Research*, 62(11), 525–531.
- Yazdi, M. H., Mahdavi, M., Setayesh, N., Esfandyar, M., & Shahverdi, A. R. (2013). Selenium nanoparticle-enriched *Lactobacillus brevis* causes more efficient immune responses in vivo and reduces the liver metastasis in metastatic form of mouse breast cancer. *Daru : Journal of Faculty of Pharmacy, Tehran University of Medical Sciences*, 21(1), 33.
- Yoon, I., Werner, T. M., & Butler, J. M. (2007). Effect of source and concentration of selenium on growth performance and selenium retention in broiler chickens. *Poultry Science*, 86, 727–730.
- Youcef, M., Jean-Luc, H., Louis, I., & Isabelle, D. (2013). Selenium in the environment, metabolism and involvement in body functions. *Molecules*, 18, 3292–3311.
- Yuan, D., Zhan, X. A., & Wang, Y. X. (2011). Effects of selenium sources and levels on reproductive performance and selenium retention in broiler breeder, egg, developing embryo, and 1-day-old chick. *Biological Trace Element Research*, 144(1–3), 705–714.
- Yuan, D., Zhan, X. A., & Wang, Y. X. (2012). Effect of selenium sources on the expression of cellular glutathione peroxidase and cytoplasmic thioredoxin reductase in the liver and kidney of broiler breeders and their offspring. *Poultry Science*, 91(4), 936–942.
- Zakharyan, R. A., Tsaprailis, G., Chowdhury, U. K., Hernandez, A., & Aposhian, H. V. (2005). Interactions of sodium selenite, glutathione, arsenic species, and omega class human glutathione transferase. *Chemical Research in Toxicology*, 18(8), 1287–1295.
- Zamani Moghaddam, A. K., Mehraei Hamzekolaei, M. H., Khajali, F., & Hassanpour, H. (2017). Role of selenium from different sources in prevention of pulmonary arterial hypertension syndrome in broiler chickens. *Biological Trace Element Research*, 1–7.

- Zanini, S. F., Torres, C. A. A., Bragagnolo, N., Turatti, J. M., Silva, M. G., & Zanini, M. S. (2004). Effect of oil sources and vitamin E levels in the diet on the composition of fatty acids in rooster thigh and chest meat. *Journal of the Science of Food and Agriculture*, 48, 672–682.
- Zduńczyk, Z., Drazbo, A., Jankowski, J., Juśkiewicz, J., Czech, A., & Antoszkiewicz, Z. (2013). The effect of different dietary levels of vitamin E and selenium on antioxidant status and immunological markers in serum of laying hens. *Polish Journal of Veterinary Sciences*, 16(2), 333–339.
- Zduńczyk, Z., Gruzauskas, R., Semaskaite, A., Juskiewicz, J., & Raceviciute-Stupeliene, A Wroblewska, M. (2011). Fatty acid profile of breast muscle of broiler chickens fed diets with different levels of selenium and vitamin E. *Archiv Für Geflügelkunde*, 75(4), 264–267.
- Zeqing, L., Mingliang, J., Ming, H., Youming, W., & Yizhen, W. (2013). Bioactivity of selenium-enriched exopolysaccharides produced by Enterobacter cloacae Z0206 in broilers. *Carbohydrate Polymers*, 96(1), 131–136.
- Zhan, X. A., Wang, H. F., Yuan, D., Wang, Y., & Zhu, F. (2014). Comparison of different forms of dietary selenium supplementation on gene expression of cytoplasmic thioredoxin reductase, selenoprotein P, and selenoprotein W in broilers. *Czech Journal of Animal Science*, 59(12), 571–578.
- Zhang, W., Lee, B. D., Lee, S. K., Lee, K. W., An, G. H., Song, K. B., & Lee, C. H. (2005a). Effects of yeast (*Saccharomyces cerevisiae*) cell components on growth performance, meat quality, and ileal mucosa development of broiler chicks. *Poultry Science*, 84(7), 1015–1021.
- Zhang, B., Zhou, K., Zhang, J., Chen, Q., Liu, G., Shang, N., ... Lin, F. (2009). Accumulation and species distribution of selenium in Se-enriched bacterial cells of the *Bifidobacterium animalis* 01. *Food Chemistry*, 115(2), 727–734.
- Zhang, J. (2009). Evaluation of nanotoxicity of foods and drugs: biological properties of red elemental selenium at nano size (Nano-Se) In Vitro and In Vivo. In S. Sahu and D. Casciano (Ed.), *Nanotoxicity, from in vivo and in vitro models to health risk* (pp. 97-114). John Wiley & Sons, Inc.
- Zhang, J., Wang, H., Yan, X., & Zhang, L. (2005b). Comparison of short-term toxicity between Nano-Se and selenite in mice. *Life Sciences*, 76(10), 1099–1109.
- Zhang, J., Xu, B., Huang, X., Gao, Y., Chen, Y., & Shan, A. (2016). Selenium deficiency affects the mRNA expression of inflammatory factors and selenoprotein genes in the kidneys of broiler chicks. *Biological Trace Element Research*, 171(1), 201–207.

- Zhang, L., Wang, Y. X., Zhou, Y., Zheng, L., Zhan, X. a, & Pu, Q. H. (2014a). Different sources of maternal selenium affect selenium retention, antioxidant status, and meat quality of 56-day-old offspring of broiler breeders. *Poultry Science*, 93(9), 2210–2219.
- Zhang, W., Zhang, R., Wang, T., Jiang, H., Guo, M., Zhou, E., ... Zhang, N. (2014b). Selenium inhibits LPS-induced pro-inflammatory gene expression by modulating MAPK and NF-κB signaling pathways in mouse mammary epithelial cells in primary culture. *Inflammation*, 37(2), 478–485.
- Zhang, Y., Shan, A., Jiang, W., Bi, C., & Li, Z. (2013). The effect of vitamin E on growth performance and meat quality in broilers given diets containing distillers' dried grain with solubles (DDGS). *British Poultry Science*, 54(1), 138–143.
- Zhang, Z. W., Wang, Q. H., Zhang, J. L., Li, S., Wang, X. L., & Xu, S. W. (2012). Effects of oxidative stress on immunosuppression induced by selenium deficiency in chickens. *Biological Trace Element Research*, 149(3), 352–361.
- Zhou, J. C., Zhao, H., Li, J. G., Xia, X. J., Wang, K. N., Zhang, Y. J., ... Lei, X. G. (2009). Selenoprotein gene expression in thyroid and pituitary of young pigs is not affected by dietary selenium deficiency or excess. *Journal of Nutrition*, 139(6), 1061–1066.
- Zhou, X., & Wang, Y. (2011). Influence of dietary nano elemental selenium on growth performance, tissue selenium distribution, meat quality, and glutathione peroxidase activity in Guangxi Yellow chicken. *Poultry Science*, 90(3), 680–686.
- Ziaeい, N., Moradi, N., & Pour, E. E. (2013). The effects of different levels of vitamin-E and organic selenium on performance and immune response of laying hens. *African Journal of Biotechnology*, 12(24), 3884–3890.
- Zingg, J. M. (2007). Vitamin E: an overview of major research directions. *Molecular Aspects of Medicine*, 28(5), 400–422.