

UNIVERSITI PUTRA MALAYSIA

POSTHARVEST QUALITIES AND METABOLIC RESPONSES OF HYDRO-COOLED ROCKMELON (Cucumis melo L. reticulatus cv Glamour) AFTER DIFFERENTIAL STORAGE DURATIONS

BOKHARY BIN ZAINAL

FP 2018 77



POSTHARVEST QUALITIES AND METABOLIC RESPONSES OF HYDRO-COOLED ROCKMELON (*Cucumis melo* L. *reticulatus* cv Glamour) AFTER DIFFERENTIAL STORAGE DURATIONS

By

BOKHARY BIN ZAINAL

Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Required for the Degree of Doctor of Philosophy

May 2018

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

POSTHARVEST QUALITIES AND METABOLIC RESPONSES OF HYDRO-COOLED ROCKMELON (*Cucumis melo* L. *reticulatus* cv Glamour) AFTER DIFFERENTIAL STORAGE DURATIONS

By

BOKHARY BIN ZAINAL

May 2018

Chairman Faculty

: Associate Professor Phebe Ding, PhD : Agriculture

Harvested rockmelon often accumulates a considerable amount of heat from the high field temperature which accelerates the respiration, metabolic changes and thus shortens the market life of the fruit. The beneficial effects of hydro-cooling to rapidly reduce the core temperature in order to maintain fruit quality during postharvest storage has been reported on various fruits. Therefore, the main objective of this study was to evaluate the cooling time (CT) which characterize the performance of hydro-cooled rockmelon during cold storage. Three experiments were conducted to determine (i) quality and postharvest performances of hydro-cooled rockmelon (ii) antioxidant enzymes activity and histological structure of hydro-cooled rockmelon and (iii) phytochemical content, in vitro antioxidant activities and metabolite composition of hydro-cooled rockmelon. To achieve the best cooling time, rockmelon fruit (25.5 °C initial temperature) was hydro-cooled to 14.8 and 9.4 °C to attain 1/2 and 3/4 CT, respectively prior to storage at 13 °C cold room of 85% relative humidity along with non-hydro-cooled fruit (0 CT) as control. Results showed that 1/2 CT was effective in slowing down weight loss and increment of soluble solids concentration, preserving higher chroma intensity of pulp colour and turgor pressure of rind and pulp firmness, and maintaining the citric acid content as compared to the others treatment during rockmelon storage. The significant weight loss in the control and 3/4 CT hydro-cooled rockmelon had influenced cell rigidity as dehydration caused the impact in a loss of cell turgor. During storage, the ascorbate peroxidase, peroxidase and superoxide dismutase enzymes activity were higher for 1/2 CT rockmelon. The significant induction of these antioxidative enzymes activity corresponded to lower lipid peroxidation as measured by malondialdehyde content. The less induction of antioxidative enzyme activity in 3/4 CT and control rockmelon pulp exposed the fruits to deleterious effect of free radicals. The severity of cell membranes disorder in structural characteristics was in line with higher malondialdehyde content due to less tolerance to oxidative stress in 3/4 CT and control rockmelon. To better understand the metabolic changes, untargeted metabolite properties were studied through Nuclear Magnetic Resonance (NMR)-based metabolomics approach to visualize variations in metabolic profile of hydro-cooled rockmelon during cold storage. Overall, sugars and amino acids were appreciably affected by hydro-cooling application. The high intensity of phenylalanine, tryptophan and tyrosine appear to be associated with biosynthesis of secondary metabolites and cold tolerance since their present were altered in 1/2 CT rockmelon, relative to the control and 3/4 CT hydro-cooled rockmelon. Noticeably, less accumulation of sucrose during advanced cold storage in control and 1/2 CT hydro-cooled rockmelon rind might reduce the cell metabolism and fruit resistibility in response to cold storage since it influences the cell structural support and protection against reactive oxygen species (ROS). In short, the 1/2 CT of hydro-cooling application could be considered as the optimum CT level for rockmelon fruit in preserving physico-chemical quality and thus, extending the storage life of rockmelon fruit.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

KUALITI LEPAS TUAI DAN TINDAK BALAS METABOLIK RENDAMAN AIR SEJUK ROCKMELON (Cucumis melo L. reticulatus cv Glamor) SELEPAS **TEMPOH PENYIMPANAN BERBEZA**

Oleh

BOKHARY BIN ZAINAL

Mei 2018

: Profesor Madya Phebe Ding, PhD

: Pertanian

Pengerusi Fakulti

Rockmelon yang dituai kerap mengumpul banyak haba dari suhu persekitaran yang tinggi di mana meningkatkan respirasi, perubahan metabolik dan seterusnya memendekkan tempoh pasaran buah. Kebaikan kesan rendaman air sejuk dengan cepat dapat merendahkan suhu teras supaya mengekalkan kualiti buah semasa penyimpanan lepas tuai telah dilaporkan pada pelbagai buahbuahan. Oleh itu, objektif kajian ini adalah untuk menilai masa penyejukan (CT) yang menunjukkan prestasi rockmelon rendaman air sejuk semasa penyimpanan sejuk. Tiga eksperimen telah dijalankan untuk menentukan (i) kualiti dan prestasi lepas tuai rockmelon rendaman air sejuk (ii) aktiviti enzim antioksidan dan struktur histologi rockmelon rendaman air sejuk dan (iii) kandungan fitokimia, aktiviti antioksidan in vitro dan komposisi metabolit rockmelon rendaman air sejuk. Untuk mencapai masa penyejukan terbaik, buah rockmelon (suhu awal 25.5 °C) telah direndam air sejuk hingga ke suhu 14.8 dan 9.4 °C untuk mencapai 1/2 dan 3/4 CT, sebelum penyimpanan pada 13 °C bilik sejuk 85% kelembapan relatif bersama dengan buah tidak direndam air sejuk (0 CT) sebagai kawalan. Keputusan menunjukkan bahawa 1/2 CT adalah berkesan dalam memperlahankan penurunan berat dan peningkatan kepekatan pepejal larut, mengekalkan ketinggian keamatan kroma bagi warna isi dan tekanan turgor kulit dan isi yang kuat, dan mengekalkan kandungan asid sitrik berbanding dengan rawatan yang lain semasa penyimpanan rockmelon. Kehilangan berat yang signifikan pada buah kawalan dan 3/4 CT rockmelon rendaman air sejuk telah mempengaruhi kesegahan sel apabila dehidrasi menyebabkan impak dalam kehilangan sel turgor. Semasa penyimpanan, aktiviti enzim ascorbate peroxidase, peroxidase dan superoxide dismutase adalah tinggi bagi 1/2 CT rockmelon. Peningkatan signifikan aktiviti enzim antioksidan ini sepadan dengan peroksidasi lipid yang rendah seperti yang diukur oleh kandungan malondialdehid. Pengurangan peningkatan aktiviti enzim antioksidan dalam isi iii

3/4 CT dan rockmelon kawalan mendedahkan buah-buahan kepada kesan buruk terhadap radikal bebas. Kerosakan membran sel dalam pencirian struktur adalah selari dengan kandungan malondialdehid yang tinggi kerana kurang toleransi dengan tekanan oksidatif pada 3/4 CT dan rockmelon kawalan. Untuk lebih memahami perubahan metabolik, kandungan metabolit yang tidak disasarkan dikaji melalui pendekatan metabolomik berasaskan Magnetik Nuklear Resonans (NMR) untuk menggambarkan variasi dalam profil metabolik rockmelon yang telah direndam air sejuk semasa penyimpanan sejuk. Secara keseluruhannya, gula dan asid amino dipengaruhi oleh aplikasi rendaman air sejuk. Intensiti yang tinggi bagi fenilalanina, triptofan dan tirosina dikaitkan dengan biosinteis metabolit sekunder dan toleransi sejuk di mana kehadirannya telah berubah dalam 1/2 CT rockmelon berbanding dengan yang rockmelon kawalan dan direndam air sejuk pada 3/4 CT. Sukrosa kurang terkumpul semasa lanjutan penyimpanan sejuk dalam kulit rockmelon kawalan dan 1/2 CT rendaman air sejuk mungkin mengurangkan metabolism sel dan daya tahan buah sebagai tindak balas terhadap penyimpanan sejuk kerana ia mempengaruhi sruktur sokongan sel dan perlindungan terhadap spesies oksigen reaktif (ROS). Ringkasnya, aplikasi 1/2 CT rendaman air sejuk boleh dianggap sebagai tahap CT optimum untuk buah rockmelon bagi mengekalkan kualiti fiziko-kimia dan sekaligus memanjangkan tempoh penyimpanan buah rockmelon.

ACKNOWLEDGEMENTS

In the name of Allah SWT, The most Beneficent, The most Merciful. Praise be to ALLAH SWT and peace and blessings be upon Prophet Muhammad SAW, his family, his companions and upon all other Prophets and Messengers.

I would like to extend my sincere gratitude to Associate Professor Dr. Phebe Ding the chairman of my supervisor committee; Professor Dr. Nazamid Saari and Associate Professor Dr. Intan Safinar Ismail, members of my supervisory committee, for their invaluable advice, guidance, immense knowledge, encouragement and patience throughout the research and the preparation of this thesis.

I am grateful to my mother, Hajah Salmah Mueit and siblings, Bakhtiar Zainal and Ain Zubaidah Zainal who have provided me through moral and emotional support in my life. I am also grateful to my other family members (brother and sister in laws) who have supported me along the way.

With a special mention to my friends, Mohd Nazif Saifuddin, Muhammad Zulhusni Munir, Khairul Azree Rosli, Alicia Jack, Salumiah Mijin, Anuar Yahaya, Dr. Shahrizim Zulkifly, Sapari Mat and Kak Nurul for their genuine support and constant encouragement throughout my research work.

And finally, last but by no means least, also everyone in the Postharvest Laboratory..it was great sharing laboratory with all of you during last five years.

Thanks for all your encouragement!

I certify that a Thesis Examination Committee has met on 2018 to conduct the final examination of Bokhary bin Zainal on his thesis entitled "Postharvest Qualities and Metabolism Responses of Hydro-cooled Rockmelon (*Cucumis melo* L. *reticulatus* cv Glamour) After Differential Storage Durations" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

(External Examiner)



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree Doctor of Philosophy. The members of Supervisory Committee were as follows:

Phebe Ding, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Nazamid Saari, PhD Professor

Faculty of Food Science and Technology Universiti Putra Malaysia (Member)

Intan Safinar Ismail, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD Professor and Dean

School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:

Name and Matric No.: Bokhary bin Zainal GS33958

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of the thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:	

TABLE OF CONTENTS

					Page
ABSTRACT	Г				i
ABSTRAK					iii
ACKNOWL	EDGE	EMENTS			V
APPROVA	_				vi
DECLARA	ΓΙΟΝ				viii
LIST OF TA		S			xiv
LIST OF FI	GURE	S			xvi
LIST OF A	BBRE	VIATION	S		xxiii
CHAPTER					
1	INTF	RODUCTI	ON		1
2			REVIEW		4
	2.1 2.2	Pre-cool		mis melo L.)	4 5
	2.2	2.2.1		ng concepts and	5
		2.2.1	cooling ra		'
		2.2.2		ng methods	9
			2.2.2.1	Hydro-cooling	9
			2.2.2.2	Room cooling	10
			2.2.2.3	Forced-air cooling	10
			2.2.2.4	Vacuum cooling	12
			2.2.2.5	Contact or packing icing	13
	2.3	Storage	of fruits		13
3			EMICAL A		
				CHARACTERISTICS	
				ulatus cv. Glamour)	
		ING STO			
	3.1	Introduc			18
	3.2	Material	s and meth	nods	19
		3.2.1	Fruit mate	erials	19
		3.2.2		oling and storage	19
		3.2.3		ss determination	20
		3.2.4		pulp colour	20
		2 2 5	determina Bind and		04
		3.2.3			21
		3.2.5	Rind and determina	pulp firmness ation	21

G

	3.2.6	Soluble solids concentration determination	21
	3.2.7	Titratable acidity determination	22
	3.2.8	pH determination	22
	3.2.9	Ascorbic acid determination	22
	3.2.10	Histological and structural	23
		characteristics	
		3.2.10.1 Tissue preparation for	23
		light microscopy	
		study	
		3.2.10.2 Tissue preparation for	24
		scanning electron	
	0.0.44	microscopy study	
	3.2.11	Experimental design and	24
3.3	Deputte	statistical analysis	24
3.3	Results 3.3.1	Waight loss	24 24
	3.3.2	Weight loss Rind and pulp colour	24
	3.3.3	Rind and pulp firmness	28
	3.3.4	Soluble solids concentration	30
	3.3.5	Titratable acidity	30
	3.3.6	pH	31
	3.3.7	Ascorbic acid	32
	3.3.8	Histological and structural	33
		studies	
3. <mark>4</mark>	Discuss		40
	3.4.1	Postharvest quality of rockmelon	40
	242	fruit after hydro-cooling	40
	3.4.2	Microstructural changes of	46
		quality related characteristics of hydro-cooled rockmelon fruit	
3.5	Conclus		47
0.0	Conolas		1
LIPI	D PEROX	(IDATION, HISTOLOGICAL	
		N AND ANTIOXIDANT ENZYMES	
ACT	IVITY OF	HYDRO-COOLED	
		N (Cucumis melo L. reticulatus	
		IN RELATION TO OXIDATIVE	
		RING STORAGE	
4.1	Introdu		49
4.2		ils and methods	50
	4.2.1	Fruit materials	50
	4.2.2	Hydro-cooling, storage and	50
	4.2.3	sampling Measurement of membrane lipid	50
	4.2.3	peroxidation	50
	4.2.4	Histological evaluation	51
	4.2.5	Enzyme extraction	51
		,	

	4.2.6	Ascorbate peroxidase (APX)	51
	4.2.7	enzyme assay Peroxidase (POD) enzyme	51
	4.2.7	assay	51
	4.2.8	Superoxide dismutase (SOD)	52
		enzyme assay	
	4.2.9	Experimental design and	53
		statistical analysis	
4.3	Results		53
	4.3.1	Malondialdehyde (MDA) content	53
	4.3.2	Histological study	55
	4.3.3	Ascorbate peroxidase (APX)	58
	4.3.4	enzyme activity	50
	4.3.4	Peroxidase (POD) enzyme activity	59
	4.3.5	Superoxide dismutase (SOD)	60
	4.5.5	enzyme activity	00
4.4	Discuss		61
	4.4.1	Lipid peroxidation and structural	61
		changes in relation to oxidative	• •
		stress	
	4.4.2	The role of antioxidative	63
		enzymes as protective agents	
		against oxidative stress	
4.5	Conclus	sions	64
UNVE OF H (<i>CUC</i> GLA	EILS THE YDRO-C UMIS MI MOUR) D	ABOLOMIC PROFILING COMPOSITIONAL CHANGES OOLED ROCKMELON ELO L. RETICULATUS CV. DURING STORAGE RELATED ANTIOXIDANT ACTIVITY	
5.1			65
5.2		Is and methods	66
	5.2.1	Fruit materials	66
	5.2.2	Hydro-cooling and storage	66
	5.2.3	Beta-carotene and lycopene content determination	66
	5.2.4	Preparation of methanolic	67
	0.2.4	extracts	07
	5.2.5	Total phenolic content	67
	0.2.0	determination	0.
	5.2.6	Total flavonoids content	68
		determination	
	5.2.7	2,2-Diphenyl-1-picrylhydrazyl	69
		(DPPH) radical scavenging	
		assay	
	5.2.8	Nitric oxide (NO) scavenging assay	69

5

Ĉ

xii

		5.2.9	¹ H NMR preparati	sampling and sample	69
		5.2.10		ion of ¹ H NMR	70
		5.2.11		spectroscopy analysis	70
		5.2.12		te databases and	70
		5.2.13	Experime	ental design and ate data analysis	71
	5.3	Results			71
		5.3.1	Beta-caro content	otene and lycopene	71
		5.3.2		enolic content	73
		5.3.3		onoids content	76
		5.3.4		ant activity	78
			5.3.4.1	The 2,2-Diphenyl-1-	78
			A.A.A.	picrylhydrazyl (DPPH) radical	
				scavenging activity	
			5.3.4.2	Nitric oxide (NO)	80
				scavenging activity	
		5.3.5	Metabolit	te identification	82
			5.3.5.1	Non-hydro-cooled	82
				rockmelon	
			5.3.5.2	Hydro-cooled	90
				rockmelon of 1/2	
				cooling time	
				rockmelon	
			5.3.5.3	Hydro-cooled	96
			0.0.0.0	rockmelon of 3/4	00
				cooling time	
	5.4	Discuss	ion	cooling time	102
	0.4	5.4.1		emical and in vitro	102
		5.4.1		ant evaluation of hydro-	102
				ockmelon fruit	
		5.4.2		se of sugar and amino	105
		J.4.Z		hydro-cooling during	105
			cold stor		
	5.5	Conclus		lage	109
	5.5	Conclus	0015		109
5	SIIM		ONCLUS		110
5	RECO			FOR FUTURE	110
REFERENC	ES				112
BIODATA O					140
LIST OF PU	BLIC	ATIONS			141

LIST OF TABLES

Table		Page
3.1	Main and interaction effects of cooling time of hydro- cooling treatment and week of storage on weight loss, rind and pulp (L^* , C^* and h°) colour of rockmelon fruits.	26
3.2	Main and interaction effects of cooling time of hydro- cooling treatment and week of storage on rind and pulp firmness, soluble solids concentration (SSC) and titratable acidity (TA) of rockmelPoon fruits.	29
3.3	Correlation coefficients (r) between weight loss, rind and pulp firmness of rockmelon fruits.	30
3.4	Main and interaction effects of cooling time of hydro- cooling treatment and week of storage on pH and ascorbic acid of rockmelon pulp extracts.	32
4.1	Main and interaction effects of cooling time of hydro- cooling treatment and week of storage on malondialdehyde (MDA) content of rockmelon pulp extracts.	54
4.2	Main and interaction effects of cooling time of hydro- cooling treatment and week of storage on enzyme activity of ascorbate peroxidase (APX), peroxidase (POD) and superoxide dismutase (SOD) of rockmelon pulp extracts.	58
5.1	Main and interaction effects of cooling time of hydro- cooling treatment and week of storage on beta- carotene content and lycopene content of rockmelon pulp extracts.	72
5.2	Main and interaction effects of cooling time of hydro- cooling treatment and week of storage on total phenolic content and total flavonoids content of rockmelon rind and pulp extracts.	74
5.3	Main and interaction effects of cooling time of hydro- cooling treatment and week of storage on DPPH scavenging activity and nitric oxide scavenging activity of rockmelon rind and pulp extracts.	79

 \bigcirc

5.4 Assignments for metabolites and chemical shifts identified in the ¹H NMR spectra of rockmelon rind extracts at different week of storage with corresponding multiplicity based on Chenomx database.

.



LIST OF FIGURES

Figure

- 2.1 A cooling curve showing the drop in temperature from the initial temperature of 20 °C through one (1/2 cool), two (3/4 cool) and three (7/8 cool) half-cooling times. The end of the third half-cooling cycle represents the seven-eight cooling time. The actual rate of temperature drops and time required to achieve seven-eight cooling varies with product and cooling conditions (Chakraverty and Singh, 2014).
- 3.1 Weight loss of rockmelon fruits treated with different hydrocooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05.
- 3.2 Rind firmness of rockmelon fruits treated with different 28 hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at $P \le 0.05$.
- 3.3 Percentage citric acid of rockmelon fruits treated with different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at $P \le 0.05$.
- 3.4 Ascorbic acid content of rockmelon fruits treated with different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at $P \le 0.05$.
- 3.5 Transverse section of rockmelon fruit tissue. (A) Scanning 34 electron micrograph showing exocarp and mesocarp region of non-hydro-cooled rockmelon fruit at week of storage 0 (35x). (B) Light microscopy image showing exocarp and mesocarp region of non-hydro-cooled rockmelon fruit at week of storage 0 (10x). (C) Scanning electron micrograph of non-hydro-cooled rockmelon fruit mesocarp at week of storage 0 (70x). (D) Scanning

Page

8

electron micrograph of non-hydro-cooled rockmelon fruit mesocarp at week of storage 1 (70x). (E) Scanning electron micrograph of non-hydro-cooled rockmelon fruit mesocarp at week of storage 2 (70x). EX = exocarp, M = mesocarp, EP = epidermis, P = peridermal tissue, IS = intercellular spaces, VB = vascular bundle.

- 3.6 Transverse section of non-hydro-cooled rockmelon fruit at week of storage 3. (A) Light microscopy image showing exocarp and mesocarp region of rockmelon fruit (4x); (B) scanning electron micrograph of the mesocarp of rockmelon fruit (100x); (C) scanning electron micrograph of the mesocarp of rockmelon fruit (70x). EX = exocarp, M = mesocarp, P = peridermal tissue, IS = intercellular spaces.
- 3.7 Transverse section of scanning electron micrographs of 1/2 cooling time hydro-cooled rockmelon mesocarp. (A) Mesocarp of rockmelon fruit at week of storage 0 (100x); (B) mesocarp of rockmelon fruit at week of storage 1 (100x); (C) mesocarp of rockmelon fruit at week of storage 2 (100x) and (D) mesocarp of rockmelon fruit at week of storage 3 (100x). IS = intercellular spaces, VB = vascular bundle.
- 3.8 Transverse section of 3/4 cooling time rockmelon fruit at week of storage 0 showing transverse section of 3/4 cooling time hydro-cooled rockmelon fruit at week of storage 0. (A) Scanning electron micrograph of the mesocarp region of rockmelon fruit (35x); (B) scanning electron micrograph of rockmelon fruit with angular polyhedral interlocking profile with small intercellular spaces (150x) and (C) light microscopy image showing exocarp and mesocarp region of rockmelon fruit (20x). EX = exocarp, EP = epidermis M = mesocarp, IS = intercellular spaces.
- 3.9 Scanning electron micrographs of 3/4 cooling time hydro-38 cooled rockmelon fruit at week of storage 1. (A) Transverse section of mesocarp region of rockmelon fruit (70x); (B and C) transverse section rockmelon fruit indicated the sign dehydration (700x). M = mesocarp, IS = intercellular spaces.
- 3.10 Scanning electron micrographs of 3/4 cooling time hydro-38 cooled rockmelon fruit at week of storage 2. (A) Transverse section of mesocarp region of rockmelon fruit (70x) and (B) cells of rockmelon fruit showing prominent intercellular spaces (70x).

35

36

- 3.11 Transverse section of 3/4 cooling time rockmelon fruit at week of storage 3. (A and B) Mesocarp of 3/4 cooling time hydro-cooled rockmelon at week of storage 3 viewed under scanning electron micrograph showing aspect of dehydration, tissue leakage and collapse cell walls (70x); (C) Light microscopy image showing exocarp and mesocarp region of 3/4 cooling time hydro-cooled rockmelon at week of storage 3 (10x); (D) Light microscopy image showing clump of cells of 3/4 cooling time hydro-cooled rockmelon at week of storage 3 (40x). EX = exocarp, M = mesocarp, C = collapsed cell wall.
- 3.12 Rockmelon fruits at four different weeks of storage (WS) 41 treated with different cooling time (CT) of 4 °C cold water.
- 4.1 Bovine erythrocyte SOD (Cu/Zn) standard calibration 52 curve with linear equation and coefficient of determination (R²).
- 4.2 Changes of malondialdehyde (MDA) content of rockmelon fruits treated with different hydro-cooling treatment before cold storage for three weeks. Means (n=18) of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05. Vertical bars indicate SE of means.
- 4.3 Transverse section of rockmelon fruit at week of storage
 0. (A) Light micrograph of non-hydro-cooled rockmelon fruit (10x). (B) Light micrograph of 1/2 cooling time hydro-cooled rockmelon fruit (10x). (C) Light micrograph of 3/4 cooling time hydro-cooled rockmelon fruit (10x). The transverse section revealed the EX = exocarp, EP = epidermis, M = mesocarp and VB = vascular bundle with its elements of X = xylem and PH = phloem in the rockmelon tissue.
- 4.4 Transverse section of rockmelon fruit at week of storage 3. (A) Light micrograph of non-hydro-cooled rockmelon fruit (10x). (B) Light micrograph of 1/2 cooling time hydro-cooled rockmelon fruit (10x). (C) Light micrograph of 3/4 cooling time hydro-cooled rockmelon fruit (10x). The transverse section revealed the EX = exocarp, M = mesocarp and formation of IS = intercellular space found in the rockmelon tissue.
- 4.5 Changes of ascorbate peroxidase (APX) activities of 59 rockmelon fruits treated with different hydro-cooling treatment before cold storage for three weeks. Means

39

xviii

(n=18) of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05. Vertical bars indicate SE of means.

- 4.6 Changes of peroxidase (POD) activities of rockmelon fruits treated with different hydro-cooling treatment before cold storage for three weeks. Means (n=18) of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05. Vertical bars indicate SE of means.
- 4.7 Changes of superoxide dismutase (SOD) activities of 61 rockmelon fruits treated with different hydro-cooling treatment before cold storage for three weeks. Means (n=18) of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05. Vertical bars indicate SE of means.
- 5.1 Gallic acid standard calibration curve with linear equation 67 and coefficient of determination (R²).
- 5.2 Quercetin standard calibration curves for rockmelon rind 68 (A) and pulp (B) extracts with linear equation and coefficient of determination (R²).
- 5.3 Beta-carotene content of rockmelon pulp treated with 72 different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05.
- Lycopene content of rockmelon pulp treated with different 5.4 73 hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05.
- 5.5 75 Total phenolic content of rockmelon rind treated with different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05.

- 5.6 Total phenolic content of rockmelon pulp treated with different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05.
- 5.7 Total flavonoids content of rockmelon rind treated with different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at P≤0.05.
- 5.8 Total flavonoids content of rockmelon pulp treated with 77 different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at $P \le 0.05$.
- 5.9 DPPH scavenging activity of rockmelon rind treated with different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at $P \le 0.05$.
- 5.10 Nitric oxide scavenging activity of rockmelon rind treated 81 with different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at $P \le 0.05$.
- 5.11 Nitric oxide scavenging activity of rockmelon pulp treated 81 with different hydro-cooling treatment before cold storage for three weeks. Means of different cooling time within the same week of storage followed by the different letters are significantly different by Duncan's multiple range tests at $P \le 0.05$.
- 5.12 ¹H NMR spectra (CD₃OD + 0.2% TMS) of methanolic 83 extract of 0 cooling time (non-hydro-cooled) rockmelon rind at different week of storage (WS).
- 5.13 Two-dimensional score plot of the PCA (PC1 and PC2) 85 representing the methanolic extracts of 0 cooling time

76

ΧХ

(non-hydro-cooled) rockmelon rind shows the discrimination in week of storage.

86

- 5.14 Loading column plot of component 1 (p[1]) of the ¹H NMR data representing different week of storage in 0 cooling time (non-hydro-cooled) rockmelon rind extracts (R2Y = 0.88, Q2 = 0.55). Each loading column represent metabolite and numbers in the significant loading column plot are assigned according to Table 5.4. 1 = Valine, 2 = Alanine, 3 = Citrulline, 4 = Glutamate, 5 = Aspartate, 6 = Phenylalanine, 7 = Tyrosine, 8 = Tryptophan, 9 = Glucose, 10 = Sucrose.
- 5.15 Loading column plot of component 2 (p[2]) of the ¹H NMR data representing different week of storage in 0 cooling time (non-hydro-cooled) rockmelon rind extracts (R2Y = 0.88, Q2 = 0.55). Each loading column represent metabolite and numbers in the significant loading column plot are assigned according to Table 5.4. 1 = Valine, 2 = Alanine, 6 = Phenylalanine, 7 = Tyrosine, 8 = Tryptophan, 9 = Glucose, 10 = Sucrose.
- 5.16 The biplot obtained from PCA describing the correlation 89 between identified metabolites and week of storage of the methanolic extracts of 0 cooling time (non-hydro-cooled) rockmelon rind.
- 5.17 ¹H NMR spectra (CD₃OD + 0.2% TMS) of methanolic 91 extract of 1/2 cooling time hydro-cooled rockmelon rind at different week of storage (WS).
- 5.18 Two-dimensional score plot of the PCA (PC1 and PC2) 92 representing the methanolic extracts of 1/2 cooling time hydro-cooled rockmelon rind shows the discrimination in week of storage.
- 5.19 Loading column plot of component 1 (p[1]) of the ¹H NMR 93 data representing different week of storage in 1/2 cooling time hydro-cooled rockmelon rind extracts (R2Y = 0.80, Q2 = 0.54). Each loading column represent metabolite and numbers in the significant loading column plot are assigned according to Table 5.4. 1 = Valine, 2 = Alanine, 3 = Citrulline, 4 = Glutamate, 5 = Aspartate, 6 = Phenylalanine, 7 = Tyrosine, 8 = Tryptophan, 9 = Glucose, 10 = Sucrose.
- 5.20 Loading column plot of component 2 (p[2]) of the ¹H NMR 94 data representing different week of storage in 1/2 cooling time hydro-cooled rockmelon rind extracts (R2Y = 0.80,

Q2 = 0.54). Each loading column represent metabolite and numbers in the significant loading column plot are assigned according to Table 5.4. 1 = Valine, 2 = Alanine, 3 = Citrulline, 4 = Glutamate, 5 = Aspartate, 7 = Tyrosine.

- 5.21 The biplot obtained from PCA describing the correlation between identified metabolites and week of storage of the methanolic extracts of 1/2 cooling time rockmelon rind.
- 5.22 ¹H NMR spectra (CD₃OD + 0.2% TMS) of methanolic extract of 3/4 cooling time hydro-cooled rockmelon rind at different week of storage (WS).
- 5.23 Two-dimensional score plot of the PCA (PC1 and PC2) 98 representing the methanolic extracts of 3/4 cooling time hydro-cooled rockmelon rind shows the discrimination in week of storage.
- 5.24 Loading column plot of component 1 (p[1]) of the ¹H NMR 99 data representing different week of storage in 3/4 cooling time hydro-cooled rockmelon rind extracts (R2Y = 0.90, Q2 = 0.63). Each loading column represent metabolite and numbers in the significant loading column plot are assigned according to Table 5.4. 2 = Alanine, 3 = Citrulline, 4 = Glutamate, 5 = Aspartate, 6 = Phenylalanine, 8 = Tryptophan, 9 = Glucose, 10 = Sucrose.
- 5.25 Loading column plot of component 2 (p[2]) of the ¹H NMR data representing different week of storage in 3/4 cooling time hydro-cooled rockmelon rind extracts (R2Y = 0.90, Q2 = 0.63). Each loading column represent metabolite and numbers in the significant loading column plot are assigned according to Table 5.4. 2 = Alanine, 3 = Citrulline, 5 = Aspartate, 9 = Glucose, 10 = Sucrose.
- 5.26 The biplot obtained from PCA describing the correlation 101 between identified metabolites and week of storage of the methanolic extracts of 3/4 cooling time rockmelon rind.
- 5.27 108 Pathways showing the relationship of sucrose, aromatic amino acids (tryptophan, phenylalanine, tyrosine) and glutamate in regulating cell wall integrity, secondary metabolites and y-aminobutyric acid biosynthesis in rockmelon rind, respectively. The remaining metabolites shown in the pathways were not determined, however there are included for completeness (Luengwilai et al., 2012).

97

95

LIST OF ABBREVIATIONS

	¹ H NMR	Proton nuclear magnetic resonance
	AAA	Aromatic amino acid
	ANOVA	Analysis of variance
	APX	Ascorbic peroxidase
	CD ₃ OD	Methanol-d4
	СТ	Cooling time
	CV-ANOVA	Cross validated residuals of variance analysis
	D ₂ O	Deuterium oxide
	DDPH	2,2-diphenyl-1-picrylhydrazyl
	DMRT	Duncan's multiple range test
	EDTA	Ethylenediamine tetraacetic acid
	GABA	γ-aminobutyric acid
	GAE	Gallic acid equivalents
	MVDA	Multivariate data analysis
	NO	Nitric oxide
	PC	Principal component
	PCA	Principal component analysis
	POD	Peroxidase
	PVP	Polyvinylpyrrolidone
	QE	Quercetin equivalents
	RH	Relative humidity
	SOD	Superoxide dismutase
	ТВА	Thiobarbituric acid

TCA	Trichloroacetic acid

TFC	Total flavonoids content
TPC	Total phenolic content
VIP	Variables influence on projection





CHAPTER 1

INTRODUCTION

Melon is one of the most important fruits in the world. It is the fourth world's major production after orange, banana and grape (Aguayo et al., 2004). The most common melon variety consumed as fresh fruit is cantaloupe or also known as muskmelon or rockmelon (*Cucumis melo* L. *reticulatus*). Rockmelon production in all tropical and subtropical regions of the world at present is mainly as a source of food or commodity produce. The rockmelon fruits are usually consumed directly as a fresh cut fruit, salad or as a dessert with ice cream or custard. Presently, China is the largest rockmelon-producing country with estimated production of over 31 million tonnes (Yahia, 2011). Turkey, Iran, the United States, Spain, India, Egypt, Morocco, Mexico and Italy are the next largest rockmelon-producing countries in the world (FAO, 2014).

In Malaysia, rockmelon is one of the fast growing popular fruits and has gained very high demand. The production of this crop has been tremendously increased in between 2011 to 2015 from 311 to 5,118 metric tons, respectively (Department of Agriculture, 2015). The popular variety of rockmelon type cultivated in Malaysia is 'Glamour' or locally known as 'Golden Langkawi'. Rockmelon fruits are preferably consumed during hot weather and widely accepted among consumers because of their sweet pulp, lusciously rich in flavor and pleasant aroma (Villanueva et al., 2004). In addition to their high appreciation by consumers, rockmelon fruits are extremely nutritious fruit choice as they are rich in ascorbic acid, β -carotene, flavonoids, folic acid, and potassium (Lester and Crosby, 2002; Koubala et al., 2016) as well as a number of other human health-bioactive compounds (Lester, 1997).

However, besides being highly appreciated by consumers, rockmelon fruits have a drawback of lower shelf life due to its high perishability. The substantial loss of its marketability commonly occurs within two weeks during recommended storage conditions at 7 °C with 95% RH (Zheng and Wolff, 2000; Shellie and Lester, 2004). The major factor limiting rockmelon keeping quality and short storage life is desiccation (Mayberry and Hartz, 1992). Desiccation during storage is mainly due to water loss and the amount of water loss may result in a significant loss in fresh weight. This limitation severely limits the wide commercial acceptance of rockmelon fruit as water loss can cause a subtle quality changes in colour, texture, firmness and nutritional value, then limits the possibility of extending the storage life even under current commercial handling techniques (Mayberry and Hartz, 1992).

The relative postharvest life and quality of horticultural crops is significantly affected by temperature. Quality loss after harvest and throughout storage occurs as a result of physiological and biological processes as the rates are influenced primarily by product temperature. For example, crop with high product temperature utilizes the reserves carbohydrates faster due to increase in respiration rate and becoming more perishable than those with lower respiration rates (Fonseca et al., 2002). During respiration, harvested produce gives off vital heat couple with a considerable amount of field heat which is reflected as increase in respiration rate and metabolic activities associated with its decomposition (Aroucha et al., 2016). Indeed, Nunes et al. (1995a) concluded that within six hours of harvest at 30 °C of ambient temperature, 'Chandler', 'Oso Grande' and 'Sweet Charlie' strawberries had greater water loss, low tissue firmness value and prone to browning during postharvest storage.

For the maintenance of market quality, it is necessary not only to cool the product but must be rapidly cooled after harvest to the optimal storage temperature in order to arrest the deteriorative and senescence process to maintain a high level of quality that ensures customer satisfaction (Senthilkumar et al., 2015). In practice, this means that direct cold storage is not recommended as it is slow cooling and delays the core temperature to decrease over hours to cause high metabolic activity and shorten the expected storage life in most perishable crops. Therefore, prompt pre-cooling is critical for produce to remove field heat from freshly harvested crops rapidly so it can minimize postharvest losses and meet consumer demands for high quality fresh produce. Practically, cooling time (CT) is required to establish the relationship between temperature and amount of time produce is exposed to cooling medium (Chakraverty and Singh, 2014).

There are a variety of pre-cooling techniques available for use in the horticultural industry, among them are air-cooling, hydro-cooling, cryogenic cooling and vacuum cooling (Garrido et al., 2015). These pre-cooling methods differed from each other according to the medium used as a heat transfer in lowering the commodity temperature, such as cold water, refrigerated air or ice. Among these, hydro-cooling is a practically popular pre-cooling method because of its simplicity, low cost and effectiveness to lower the temperature. Water has high thermal conductivity (Liang et al., 2013) and utilization of water as cooling medium offers faster and uniform contact with the surface of produce causing the temperature to lower quickly. Unlike air-pre-cooling, hydro-cooling does not dehydrate the produce but it may even revive slightly fruits and vegetables (Henry and Bennett, 1973). Some workers found that hydro-cooling was an effective pre-cooling treatment in preserving fruit quality of apple (Brackman and Balz, 2000), delaying senescence and prolonging postharvest life during distribution of sweet cherry (Manganaris et al., 2007), rambutan (Nampan et al., 2006) and dates (Elansari, 2008).

Therefore, the general objective of this work was to evaluate the CT which characterize the performance of hydro-cooled rockmelon. The performance of hydro-cooled product requires extensive evaluation concerning quality and

storability attributes of fruit throughout storage period. Hence, three studies were conducted with the aims:

- 1. To evaluate the effects of different CT of hydro-cooling treatment in conjunction with cold storage on the physico-chemical quality of rockmelon fruit;
- 2. To determine the activity profile of antioxidative scavenging enzymes (superoxide dismutase, ascorbate peroxidase and peroxidase), postharvest structural analysis and malondialdehyde content which is end product of lipid peroxidation in cells represent oxidative damage of the hydro-cooled 'Glamour' melon fruit during cold storage; and
- 3. To profile the phytochemical content, *in vitro* antioxidant activities and major trend in metabolite composition of hydro-cooled rockmelon fruit during storage.

REFERENCES

- Adisa, V.A. 1986. The influence of molds and some storage factors on the ascorbic acid content of orange and pineapple fruits. *Food Chemistry* 22: 139-146.
- Agar, I.T., Massantini, R., Hess-Pierce, B. and Kader, A.A. 1999. Postharvest CO₂ and ethylene production and quality maintenance of fresh-cut kiwifruit slices. *Journal of Food Science* 64: 433-440.
- Agati, G., Azzarello, E., Pollastri, S. and Tattini, M. 2012. Flavonoids as antioxidants in plants: location and functional significance. *Plant Science* 196: 67-76.
- Aghdam, M.S. and Bodbodak, S. 2013. Physiological and biochemical mechanisms regulating chilling tolerance in fruits and vegetables under postharvest salicylates and jasmonates treatments. *Scientia Horticulturae* 156: 73-85.
- Aguayo, E., Escalona, V.H. and Rtés, F.A. 2004. Metabolic behaviour and quality changes of whole and fresh processed melon. *Journal of Food Science* 69: 148-155.
- Agusti, M., Almela, V., Juan, M., Alferez, F., Tadeo, F.R. and Zacarias, L. 2001. Histological and physiological characterization of rind breakdown of 'Navelate' sweet orange. *Annals of Botany* 88: 415-422.
- Alférez, F. and Burns, J.K. 2004. Postharvest peel pitting at non-chilling temperatures in grapefruit is promoted by changes from low to high relative humidity during storage. *Postharvest Biology and Technology* 32: 79-87.
- Alique, R., Zamorano, J.P., Martinez, M.A. and Alonso, J. 2005. Effect of heat and cold treatments on respiratory metabolism and shelf-life of sweet cherry, type picota cv "Ambrunes". *Postharvest Biology and Technology* 35: 153-165.
- Álvares, V.S., Finger, F.L., de ASantos, R.C., da Silva Negreiros, J.R. and Casali, V.W. 2007. Effect of pre-cooling on the postharvest of parsley leaves. *Journal of Food Agriculture and Environment* 5: 31-34.
- Aroucha, E.M.M., Araujo, J.M.M.D., Nunes, G.H.D.S., Negreiros, M.Z.D., Paiva, C.A.D. and Souza, M.S.D. 2016. Cantaloupe melon (*Cucumis melo* L.) conservation using hydrocooling. *Revista Ceres* 63: 191-197.
- Artés, F., Conesa, M.A., Hernández, S. and Gil, M.I. 1999. Keeping quality of fresh-cut tomato. *Postharvest Biology and Technology* 17: 153-162.

- Bartz, J.A. and Brecht, J.K. 2003. *Postharvest Physiology and Pathology of Vegetables*. New York: Marcel Dekker, Inc.
- Batth, R., Singh, K., Kumari, S. and Mustafiz, A. 2017. Transcript profiling reveals the presence of abiotic stress and developmental stage specific ascorbate oxidase genes in plants. *Frontiers in Plant Science* 8: 1-15.
- Beaulieu, J.C., Gram, D.A. Lea, J.M. and Bett-Garber. K.L. 2004. Effect of harvest maturity on the sensory characteristics fresh-cut cantaloupe. *Journal of Food Science* 69: 250–258.
- Becker, B.R. and Fricke, B.A. 2002. Hydrocooling time estimation methods. International Communications in Heat and Mass Transfer 29: 165-174.
- Bett-Garber, K.L., Greene, J.L., Lamikanra, O., Ingram, D.A. and Watson, M.A. 2011. Effect of storage temperature variations on sensory quality of fresh-cut cantaloupe melon. *Journal of Food Quality* 34: 19-29.
- Bhujbal, S.S., Nanda, R.K., Deoda, R.S., Kumar, D., Kewatkar, S.M., More, L. S. and Patil, M.J. 2010. Structure elucidation of a flavonoid glycoside from the roots of *Clerodendrum serratum* (L.) Moon, Lamiaceae. *Brazilian. Journal of Pharmacognosy* 20: 1001-1002.
- Biale, J.B. 1964. Growth, maturation and senescence in fruits. *Science* 146: 880-888.
- Biswas, P., East, A.R., Hewett, E.W. and Heyes, J.A. 2016. Intermittent warming in alleviating chilling injury - a potential technique with commercial constraint. *Food and Bioprocess Technology* 9: 1-15.
- Bouche, N. and Fromm, H. 2004. GABA in plants: just a metabolite?. *Trends in Plant Science* 9: 110-115.
- Boulton, R. 1980. The general relationship between potassium, sodium and pH in grape juice and wine. *American Journal of Enology and Viticulture* 31: 182-186.
- Bouvier, F., Isner, J.C., Dogbo, O. and Camara, B. 2005. Oxidative tailoring of carotenoids: a prospect towards novel functions in plants. *Trends in Plant Science* 10: 187-194.
- Bown, A.W. and Shelp, B.J. 1997. The metabolism and functions of [gamma]aminobutyric acid. *Plant Physiology* 115: 1-5.
- Brackmann, A. and Balz, G. 2000. Effects of precooling and postharvest treatments on the quality of Gala apples after CA storage. *Revista Cientifica Rural* 5:12-20.

- Brosnan, T. and Sun, D.W. 2001. Precooling techniques and applications for horticultural products - a review. *International Journal of Refrigeration* 24: 154-170.
- Brummell, D.A. and Harpster, M.H. 2001. Cell wall metabolism in fruit softening and quality its manipulation in transgenic plants. *Plant Molecular Biology* 47: 311-340.
- Brummell, D.A., Howie, W.J., Ma, C. and Dunsmuir, P. 2002. Postharvest fruit quality of transgenic tomatoes suppressed in expression of a ripeningrelated expansin. *Postharvest Biology and Technology* 25: 209-220.
- Bustamante, C.A., Monti, L.L., Gabilondo, J., Scossa, F., Valentini, G., Budde, C.O., Lara, M.V., Fernie, A.R. and Drincovich, M.F. 2016. Differential metabolic rearrangements after cold storage are correlated with chilling injury resistance of peach fruits. *Frontiers in Plant Science* 7: 1-15.
- Cao, S., Yang, Z. and Zheng, Y. 2013. Sugar metabolism in relation to chilling tolerance of loquat fruit. *Food Chemistry* 136: 139-143.
- Cao, S., Zheng, Y., Wang, K., Jin, P. and Rui, H. 2009a. Methyl jasmonate reduces chilling injury and enhances antioxidant enzyme activity in postharvest loquat fruit. *Food Chemistry* 115: 1458–1463.
- Cao, S., Zheng, Y., Wang, K., Rui, H. and Tang. S. 2009b. Effects of 1– methylcyclopropene on oxidative damage, phospholipases and chilling injury in loquat fruit. *Journal of the Science of Food and Agriculture* 89: 2214-2220.
- Caprioli, I., Lafuente, M.T., Rodrigo, M.J. and Mencarelli, F. 2009. Influence of postharvest treatments on quality, carotenoids, and abscisic acid content of stored "Spring Belle" peach (*Prunus persica*) fruit. *Journal of Agricultural and Food Chemistry* 57: 7056-7063.
- Carvajal, F., Palma, F., Jamilena, M. and Garrido, D. 2015. Cell wall metabolism and chilling injury during postharvest cold storage in zucchini fruit. *Postharvest Biology and Technology* 108: 68-77.
- Castellanos, D.A., Mendoza, R., Gavara, R. and Herrera, A.O. 2017. Respiration and ethylene generation modeling of "Hass" avocado and feijoa fruits and application in modified atmosphere packaging. *International Journal of Food Properties* 20: 333-349.
- Chakraverty, A. and Singh, R.P. 2014. *Postharvest Technology and Food Process Engineering*. United States: CRC Press.
- Chanjirakul, K., Wang, S.Y., Wang, C.Y. and Siriphanich, J. 2006. Effect of natural volatile compounds on antioxidant capacity and antioxidant

enzymes in raspberries. *Postharvest Biology and Technology* 40: 106-115.

- Charles, M.T., Tano, K., Asselin, A. and Arul, J. 2009. Physiological basis of UV-C induced resistance to *Botrytis cinerea* in tomato fruit. V. Constitutive defence enzymes and inducible pathogenesis-related proteins. *Postharvest Biology and Technology* 51: 414-424.
- Cisneros-Zevallos, L. 2003. The use of controlled postharvest abiotic stresses as a tool for enhancing the nutraceutical content and adding-value of fresh fruits and vegetables. *Journal of Food Science* 68: 1560-1565.
- Clayton, M., Biasi, W.V. and Mitcham, E.J. 2001. Effect of temperature management methods on firmness uniformity of commercially ripened cannery pears. *Applied Engineering in Agriculture* 17: 201-208.
- Cordenunsi, B.R. and Lajolo, F.M. 1995. Starch breakdown during banana ripening: sucrose synthase and sucrose phosphate synthase. *Journal of Agricultural and Food Chemistry* 43: 347-351.
- Damodaran, S., Parkin, K.L. and Fennema, O.R. 2007. *Fennema's Food Chemistry*. New York: CRC press.
- Daun, H. 2005. Produce color and appearance. In *Produce Degradation: Pathways and Prevention,* ed. O. Lamikanra, and S.H. Imam, 191-221. United State: CRC Press.
- De Castro, E., Barrett, D.M., Jobling, J. and Mitcham, E.J. 2008. Biochemical factors associated with a CO₂–induced flesh browning disorder of Pink Lady apples. *Postharvest Biology and Technology* 48: 182-191.
- de Castro, L.R., Vigneault, C. and Cortez, L.A.B. 2004. Container opening design for horticultural produce cooling efficiency. *Journal of Food, Agriculture and Environment* 2: 135-140.
- de Souza, E.L., de Souza, A.L.K., Tiecher, A., Girardi, C.L., Nora, L., da Silva, J.A., Argenta, L.C. and Rombaldi, C.V. 2011. Changes in enzymatic activity, accumulation of proteins and softening of persimmon (*Diospyros kaki* Thunb.) flesh as a function of pre-cooling acclimatization. *Scientia Horticulturae* 127: 242-248.
- Defraeye, T., Cronje, P., Berry, T., Opara, U.L., East, A., Hertog, M., Verboven, P. and Nicolai, B. 2015. Towards integrated performance evaluation of future packaging for fresh produce in the cold chain. *Trends in Food Science and Technology* 44: 201-225.
- de Paiva, E., Serradilla, M.J., Ruiz-Moyano, S., Córdoba, M.G., Villalobos, M.C., Casquete, R. and Hernández, A. 2017. Combined effect of antagonistic yeast and modified atmosphere to control *Penicillium expansum*

infection in sweet cherries cv. Ambrunés. *International Journal of Food Microbiology* 241: 276-282.

- Dehghannya, J., Ngadi, M. and Vigneault, C. 2010. Mathematical modeling procedures for airflow, heat and mass transfer during forced convection cooling of produce: a review. *Food Engineering Reviews* 2: 227-243.
- Del Carlo, M., Sacchetti, G., Di Mattia, C., Compagnone, D., Mastrocola, D., Liberatore, L. and Cichelli, A. 2004. Contribution of the phenolic fraction to the antioxidant activity and oxidative stability of olive oil. *Journal of Agricultural and Food Chemistry* 52: 4072-4079.
- Delgado, A.E. and Sun, D.W. 2001. Heat and mass transfer models for predicting freezing processes–a review. *Journal of Food Engineering* 47: 157-174.
- Department of Agriculture. 2015. Fruit Crops Statistic Malaysia 2015; Department of Agriculture: Putrajaya.
- Der Agopian, R.G., Peroni-Okita, F.H.G., Soares, C.A., Mainardi, J.A., do Nascimento, J.R.O., Cordenunsi, B.R., Lajolo, F.M. and Purgatto, E. 2011. Low temperature induced changes in activity and protein levels of the enzymes associated to conversion of starch to sucrose in banana fruit. *Postharvest Biology and Technology* 62: 133-140.
- Dhara, P., Patel, N.L., Tanveer, A., Apeksha, P. and Kumar, V. 2017. Effect of pre-cooling packaging material on chemical and sensory quality of guava fruits *Psidium guajava* (Linn.) cv Allahabad Safeda. *Environment and Ecology* 35: 64-69.
- Dhindsa, R.S., Plumb-Dhindsa, P. and Thorpe, T.A. 1981. Leaf senescence: correlated with increased levels of membrane permeability and lipid peroxidation, and decreased levels of superoxide dismutase and catalase. *Journal of Experimental Botany* 32: 93-101.
- Díaz-Pérez, J.C., Muy-Rangel, M.D. and Mascorro, A.G. 2007. Fruit size and stage of ripeness affect postharvest water loss in bell pepper fruit (*Capsicum annuum* L.). *Journal of the Science of Food and Agriculture* 87: 68-73.
- Ding, P., Ahmad, S.H. and Ghazali, H.M. 2007. Changes in selected quality characteristics of minimally processed carambola (*Averrhoa carambola* L.) when treated with ascorbic acid. *Journal of the Science of Food and Agriculture* 87: 702-709.
- Duque, P., Barreiro, M.G. and Arrabaça, J.D. 1999. Respiratory metabolism during cold storage of apple fruit. I. Sucrose metabolism and glycolysis. *Physiologia Plantarum* 107: 14-23.

- Dutta, P., Bhowmick, N., Khalko, S., Ghosh, A. and Ghosh, S.K. 2017. Postharvest treatments on storage life of guava (*Psidium guajava* L.) in Himalayan Terai Region of West Bengal, India. *International Journal of Current Microbiology and Applied Sciences* 6: 1831-1842.
- Elansari, A.M. 2008. Hydrocooling rates of Barhee dates at the Khala stage. *Postharvest Biology and Technology* 48: 402-407.
- Elansari, A.M. 2009. Design aspects in the precooling process of fresh produce. *Global Science Books, Fresh Produce* 3: 49-57.
- FAO. 2014. Statistical Database of the Food and Agriculture Organization of the United Nations. http://faostat.fao.org/site/291/ default.aspx.
- FAO. 2015. Statistical Database of the Food and Agriculture Organization of the United Nations. http://faostat.fao.org/site/291/ default.aspx.
- Faragher, J.D., Borochov, A., Keren-Paz, V., Adam, Z. and Halevy, A.H. 1984. Changes in parameters of cell senescence in carnation flowers after cold storage. *Scientia Horticulturae* 22: 295-302.
- Fernández-Trujillo, J.P. and Martínez, J.A.J.A. 2012. Ultrastructure of the onset of chilling injury in cucumber fruit. *Journal of Applied Botany and Food Quality* 80: 100-110.
- Ferrua, M.J. and Singh, R.P. 2009. Modeling the forced-air cooling process of fresh strawberry packages, Part I: Numerical model. *International Journal of Refrigeration* 32: 335-348.
- Finger, F.L., Endres, L., Mosquim, P.R. and Puiatti, M. 1999. Physiological changes during postharvest senescence of broccoli. *Pesquisa Agropecuária Brasileira* 34: 1565-1569.
- Fleshman, M.K., Lester, G.E., Riedl, K.M., Kopec, R.E., Narayanasamy, S., Curley Jr, R.W., Schwartz, S.J. and Harrison, E.H. 2011. Carotene and novel apocarotenoid concentrations in orange-fleshed *Cucumis melo* melons: determinations of β-carotene bioaccessibility and bioavailability. *Journal of Agricultural and Food Chemistry* 59: 4448-4454.
- Fogelman, E., Kaplan, A., Tanami, Z. and Ginzberg, I. 2011. Antioxidative activity associated with chilling injury tolerance of muskmelon (*Cucumis melo* L.) rind. *Scientia Horticulturae* 128: 267-273.
- Fonseca, S.C., Oliveira, F.A. and Brecht, J.K. 2002. Modelling respiration rate of fresh fruits and vegetables for modified atmosphere packages: a review. *Journal of Food Engineering* 52: 99-119.

- Forde, B.G. and Lea, P.J. 2007. Glutamate in plants: metabolism, regulation, and signaling. *Journal of Experimental Botany* 58: 2339-2358.
- Forney, C.F., Jordan, M.A. and Nicholas, K.U.K.G. 2003. Effect of CO₂ on physical, chemical, and quality changes in 'Burlington' blueberries. *Acta Horticulturae* 600: 587-893.
- França, C.F., Ribeiro, W.S., Silva, F.C., Costa, L.C., Rêgo, E.R. and Finger, F.L. 2015. Hydrocooling on postharvest conservation of butter lettuce. *Horticultura Brasileira* 33: 383-387.
- Friess, Y., Koffler, M. and Kauffeld, M. 2016. Density modification of ice particles in ice slurry. *International Journal of Refrigeration* 62: 97-105.
- García, J.F., Olmo, M. and García, J.M. 2016. Decay incidence and quality of different citrus varieties after postharvest heat treatment at laboratory and industrial scale. *Postharvest Biology and Technology* 118: 96-102.
- Garrido, Y., Tudela, J.A. and Gil, M.I. 2015. Comparison of industrial precooling systems for minimally processed baby spinach. *Postharvest Biology and Technology* 102: 1-8.
- Génard, M., Bertin, N., Borel, C., Bussières, P., Gautier, H., Habib, R., Léchaudel, M., Lecomte, A., Lescourret, F., Lobit, P. and Quilot, B. 2007. Towards a virtual fruit focusing on quality: modelling features and potential uses. *Journal of Experimental Botany* 58: 917-928.
- Gerchikov, N., Keren-Keiserman, A., Perl-Treves, R. and Ginzberg, I. 2008. Wounding of melon fruits as a model system to study rind netting. *Scientia Horticulturae* 117: 115-122.
- Gillies, S.L. and Toivonen, P.M.A. 1995. Cooling method influences the postharvest quality of broccoli. *HortScience* 30: 313-315.
- Goldman, A. 2002. Melons for the Passionate Grower. Artisan: New York.
- Golob, P., Farrell, G. and Orchard, J.E. 2002. *Postharvest Science and Technology, Principles and Practices*. United States: Blackwell Science.
- Gómez, P., Ferrer, M., Fernández-Trujillo, J.P., Calderón, A., Artés, F., Egea-Cortines, M. and Weiss, J. 2009. Structural changes, chemical composition and antioxidant activity of cherry tomato fruits (cv. Micro-Tom) stored under optimal and chilling conditions. *Journal of the Science* of Food and Agriculture 89: 1543-1551.
- Gonda, I., Bar, E., Portnoy, V., Lev, S., Burger, J., Schaffer, A.A., Tadmor, Y., Gepstein, S., Giovannoni, J.J., Katzir, N. and Lewinsohn, E. 2010. Branched-chain and aromatic amino acid catabolism into aroma volatiles in *Cucumis melo* L. fruit. *Journal of Experimental Botany* 61: 1111-1123.

- Gonzalez-Aguilar, G.A., Villa-Rodriguez, J.A., Ayala-Zavala, J.F. and Yahia, E.M. 2010. Improvement of the antioxidant status of tropical fruits as a secondary response to some postharvest treatments. *Trends in Food Science and Technology* 21: 475-482.
- Grizzell, W.G. and Bennett, A.H. Hydrocooling stacked crates of celery and sweet corn. USDA Agricultural Research Report, 1966.
- Gualanduzzi, S., Baraldi, E., Braschi, I., Carnevali, F., Gessa, C.E. and De Santis, A. 2009. Respiration, hydrogen peroxide levels and antioxidant enzyme activities during cold storage of zucchini squash fruit. *Postharvest Biology and Technology* 52: 16–23.
- Gupta, A. 2016. Postharvest management and processing techniques for vegetables. *STC Agriculture and Natural Resources* 2: 28-41.
- Guy, C.L., Huber, J.L. and Huber, S.C. 1992. Sucrose phosphate synthase and sucrose accumulation at low temperature. *Plant Physiology* 100: 502-508.
- Habib, M., Bhat, M., Dar, B.N. and Wani, A.A. 2017. Sweet cherries from farm to table: A review. *Critical Reviews in Food Science and Nutrition* 57: 1638-1649.
- Hadfield, K.A., Rose, J.C.K., Yaver, D.S., Berka, R.M. and Bennett, A.B. 1998. Polygalacturonase gene expression in ripe melon fruit supports a role for polygalacturonase in ripening-associated pectin disassembly. *Plant Physiology* **117**: 363-373.
- Hamann, T. and Denness, L. 2011. Cell wall integrity maintenance in plants: lessons to be learned from yeast?. *Plant Signaling and Behavior* 6: 1706-1709.
- Han, Q., Gao, H., Chen, H., Fang, X. and Wu, W. 2017. Precooling and ozone treatments affects postharvest quality of black mulberry (*Morus nigra*) fruits. *Food Chemistry* 221: 1947-1953.
- Hardenburg, R.E., Watada, A.E. and Yang, C.Y. 1990. *The Commercial Storage* of *Fruits, Vegetables, and Florist and Nursery Stocks*. Washington: Agriculture Handbook.
- Hassan, S. 1990. *Siri Panduan Tanaman: Panduan Penanaman Tembikai Wangi*. Kuala Lumpur: Berita Publishing Sdn. Bhd.
- He, S.Y., Feng, G.P., Yang, H.S., Wu, Y. and Li, Y.F. 2004. Effects of pressure reduction rate on quality and ultrastructure of iceberg lettuce after vacuum cooling and storage. *Postharvest Biology and Technology* 33: 263-273.

- He, S.Y., Zhang, G.C., Yu, Y.Q., Li, R.G. and Yang, Q.R. 2013. Effects of vacuum cooling on the enzymatic antioxidant system of cherry and inhibition of surface-borne pathogens. *International Journal of Refrigeration* 36: 2387-2394.
- Heath, M.C. 1998. Apoptosis, programmed cell death and the hypersensitive response. *European Journal of Plant Pathology* 104: 117-124.
- Henriod, R.E. 2005. Postharvest characteristics of Navel oranges following high humidity and low temperature storage and transport. *Postharvest Biology and Technology* 42: 57–64.
- Henriod, R.E., Gibberd, M.R. and Treeby, M.T. 2005. Storage temperature effects on moisture loss and development of chilling injury in Lanes Late Navel orange. *Australian Journal of Experimental Agriculture* 45: 453-58.
- Henry, F.E. and Bennett, A.H. 1973. Hydraircooling vegetable products in unit loads. *Transactions of the American Society of Agricultural Engineers* 16: 731-733.
- Heyes, J.A. and Sealey, D.F. 1996. Textural changes during nectarine (*Prunus persica*) development and ripening. *Scientia Horticulturae* 65: 49-58.
- Higashi, K., Hosoya, K. and Ezura, H. 1999. Histological analysis of fruit development between two melon (*Cucumis melo* L. *reticulatus*) genotypes setting a different size of fruit. *Journal of Experimental Botany* 50: 1593-1597.
- Ho, L.C., Sjut, V. and Hoad, G.V. 1982. The effect of assimilate supply on fruit growth and hormone levels in tomato plants. *Plant Growth Regulation* 1: 155-171.
- Hodges, D.M. 2003. *Postharvest Oxidative Stress in Horticultural Crops*. Food Product Press: New York.
- Hodges, D.M., Lester, G.E., Munro, K.D. and Toivonen. P.M.A. 2004. Oxidative stress: importance for postharvest quality. *HortScience* 39: 924–929.
- Holland, N., Menezes, H.C. and Lafuente, M.T. 2002. Carbohydrates as related to the heat-induced chilling tolerance and respiratory rate of 'Fortune' mandarin fruit harvested at different maturity stages. *Postharvest Biology and Technology* 25: 181-191.
- Huang, R., Liu, J., Lu, Y. and Xia, R. 2008. Effect of salicylic acid on the antioxidant system in the pulp of 'Cara Cara' navel orange (*Citrus sinensis* L. Osbeck) at different storage temperatures. *Postharvest Biology and Technology* 47: 168–175.

- Huang, R., Xia, R., Hu, L., Lu, Y. and Wang, M. 2007. Antioxidant activity and oxygen–scavenging system in orange pulp during fruit ripening and maturation. *Scientia Horticulturae* 113: 166–172.
- Huang, R., Xia, R., Lu, Y., Hu, L. and Xu, Y. 2008. Effect of pre-harvest salicylic acid spray treatment on post-harvest antioxidant in the pulp and peel of 'Cara cara' navel orange (*Citrus sinenisis* L. Osbeck). *Journal of the Science of Food and Agriculture* 88: 229-236.
- Hubbard, N.L., Pharr, D.M. and Huber, S.C. 1991. Sucrose phosphate synthase and other sucrose metabolizing enzymes in fruits of various species. *Physiologia Plantarum* 82: 191-196.
- Hulme, A.C. 1971. *The Biochemistry of Fruits and Their Products*. Academic Press: London.
- Hussain, P.R., Meena, R.S., Dar, M.A. and Wani, A.M. 2012. Effect of postharvest calcium chloride dip treatment and gamma irradiation on storage quality and shelf-life extension of Red delicious apple. *Journal of Food Science and Technology* 49: 415-426.
- Imahori, Y., Takemura, M. and Bai, J. 2008. Chilling-induced oxidative stress and antioxidant responses in mume (*Prunus mume*) fruit during low temperature storage. *Postharvest Biology and Technology* 49: 54-60.
- Ismail, H.I., Chan, K.W., Mariod, A.A. and Ismail, M. 2010. Phenolic content and antioxidant activity of cantaloupe (*Cucumis melo*) methanolic extracts. *Food Chemistry* 119: 643-647.
- Janick, J. 1986. *Horticultural Science*. 4th Edition. New York: Freeman WH.
- Janisiewicz, W.J., Takeda, F., Glenn, D.M., Camp, M.J. and Jurick, W.M. 2016. Dark period following UV-C treatment enhances killing of *Botrytis cinerea* conidia and controls gray mold of strawberries. *Phytopathology* 106: 386-394.
- Jarvis, M.C., Briggs, S.P.H. and Knox, J.P. 2003. Intercellular adhesion and cell separation in plants. *Plant, Cell and Environment* 26: 977-989.
- Jayaprakasam, B., Seeram, N.P. and Nair, M.G. 2003. Anticancer and antiinflammatory activities of cucurbitacins from *Cucurbita andreana*. *Cancer Letters* 189: 11-16.
- Jeffrey, C. 1990. Systematics of the Cucurbitaceae: an overview. In *Biology and Utilization of the Cucurbitaceae*, ed. D.M. Bates, R.W. Robinson, and C. Jeffrey. pp. 3-9. Comstock Publishing Associates, Cornell University Press.

- Jeong, J., Brecht, J.K., Huber, D.J. and Sargent, S.A. 2008. Storage life and deterioration of intact cantaloupe (*Cucumis melo* L. var. *reticulatus*) fruit treated with 1-methylcyclopropene and fresh-cut cantaloupe prepared from fruit treated with 1-methylcyclopropene before processing. *HortScience* 43: 435-438.
- Jeong, J., Lee, J. and Huber, D.J. 2007. Softening and ripening of 'Athena' cantaloupe (*Cucumis melo* L. var. *reticulatus*) fruit at three harvest maturities in response to the ethylene antagonist 1-methylcyclopropene. *HortScience* 42: 1231-1236.
- Jin, C., Suo, B., Kan, J., Wang, H. and Wang, Z. 2006. Changes in cell wall polysaccharide of harvested peach fruit during storage. *Journal of Plant Physiology and Molecular Biology* 32: 657-664.
- Jum'h, I., Telfah, A., Lambert, J., Gogiashvili, M., Al-Taani, H. and Hergenröder, R. 2017. ¹³C and ¹HNMR measurements to investigate the kinetics and the mechanism of acetic acid (CH₃CO₂H) ionization as a model for organic acid dissociation dynamics for polymeric membrane water filtration. *Journal of Molecular Liquids* 227: 106-113.
- Junmatong, C., Faiyue, B., Rotarayanont, S., Uthaibutra, J., Boonyakiat, D. and Saengnil, K. 2015. Cold storage in salicylic acid increases enzymatic and non-enzymatic antioxidants of Nam Dok Mai No. 4 mango fruit. *ScienceAsia* 41: 12-21.
- Kader. A. 2002. *Postharvest Technology on Horticultural Crops, 3rd Edition.* California: University of California & Natural Resources.
- Karchi, Z. 2000. Development of melon culture and breeding in Israel. Acta Horticulturae 510: 13-18.
- Kasmire, R.F., Rappaport, L. and May, D. 1970. Effects of 2chloroethylphosphonic acid on ripening of cantaloupes. *Journal of the American Society for Horticultural Science* 95: 134-137.
- Kaur, C. and Kapoor, H.C. 2001. Antioxidants in fruits and vegetables-the millennium's health. *International Journal of Food Science and Technology* 36: 703-725.
- Kaynas, K. and Sivritepe, H.O. 1995. Effect of pre-cooling treatments on storage quality of mature green tomatoes. *Acta Horticulturae* 412: 200-209.

Kays, S.J. and Paull, R.E. 2004. *Postharvest Biology*. United States: Exon Press.

Keren-Keiserman, A., Tanami, Z., Shoseyov, O. and Ginzberg, I. 2004. Peroxidase activity associated with suberization processes of the muskmelon (*Cucumis melo*) rind. *Physiologia Plantarum* 121: 141-148.

- Khairi, A.N., Falah, M.A.F., Suyantohadi, A., Takahashi, N. and Nishina, H. 2015. Effect of storage temperatures on color of tomato fruit (*Solanum lycopersicum* Mill.) cultivated under moderate water stress treatment. *Agriculture and Agricultural Science Procedia* 3: 178-183.
- Khaliq, G., Mohamed, M.T.M., Ali, A., Ding, P. and Ghazali, H.M. 2015. Effect of gum arabic coating combined with calcium chloride on physico-chemical and qualitative properties of mango (*Mangifera indica* L.) fruit during low temperature storage. *Scientia Horticulturae* 190: 187-194.
- Kienzle, S., Sruamsiri, P., Carle, R., Sirisakulwat, S., Spreer, W. and Neidhart, S. 2011. Harvest maturity specification for mango fruit (*Mangifera indica* L. 'Chok Anan') in regard to long supply chains. *Postharvest Biology and Technology* 61: 41-55.
- Kim, B.S., Kim, D.C., Lee, S.E., Nahmgoong Choi, M.J. and Joong, M.C. 1995. Freshness prolongation of crisphead lettuce by vacuum cooling. *Agricultural Chemistry and Biotechnology* 38: 239-247.
- Kim, H.K., Choi, Y.H. and Verpoorte, R. 2010. NMR-based metabolomic analysis of plants. *Nature Protocols* 5: 536-549.
- Kim, T., Kim, J., Jin, Y. and Yun, Y. 2006. The inhibitory effect and mechanism of luteolin 7-glucoside on rat aortic vascular smooth muscle cell proliferation. *Archives of Pharmacal Research* 29: 67-72.
- Kinnersley, A.M. and Turano, F.J. 2000. Gamma aminobutyric acid (GABA) and plant responses to stress. *Critical Reviews in Plant Sciences* 19: 479-509.
- Knight, M.R. and Knight, H. 2012. Low-temperature perception leading to gene expression and cold tolerance in higher plants. *New Phytologist* 195: 737-751.
- Kitinoja, L. and Thompson, J.F. 2010. Pre-cooling systems for small-scale producers. *Stewart Postharvest Review* 6: 1-14.
- Kochhar, V. and Kumar, S. 2015. Effect of different pre-cooling methods on the quality and shelf life of broccoli. *Journal of Food Processing and Technology* 6: 424-.431.
- Koh, T.H. and Melton, L.D. 2002. Ripening-related changes in cell wall polysaccharides of strawberry cortical and pith tissues. *Postharvest Biology and Technology* 26: 23-33.
- Konarska, A. 2012. Differences in the fruit peel structures between two apple cultivars during storage. *Acta Scientiarum Polonorum-Hortorum Cultus* 11: 105-116.

- Koubala, B.B., Bassang'na, G., Yapo, B.M. and Raihanatou, R. 2016. Morphological and biochemical changes during muskmelon (*Cucumis melo* var. Tibish) fruit maturation. *Journal of Food and Nutrition Sciences* 4: 18-28.
- Kupferman, E.M. 1995. Managing cherry temperature for optimum quality. *Good Fruit Grower* 46: 39-41.
- Lacan, D. and Baccou. J.C. 1998. High levels of antioxidant enzymes correlate with delayed senescence in non-netted muskmelon fruits. *Planta* 204: 377-382.
- Lamikanra, O., Chen, J.C., Banks, D. and Hunter, P.A. 2000. Biochemical and microbial changes during the storage of minimally processed cantaloupe. *Journal of Agricultural and Food Chemistry* 48: 5955-5961.
- Lamikanra, O., Imam, S. and Ukuku, D. 2005. *Produce Degradation: Pathways and Prevention*. Florida: CRC Press.
- Lamikanra, O., Inyang, I.D. and Leong, S. 1995. Distribution and effect of grape maturity on organic acid content of Red Muscadine grapes. *Journal of Agricultural and Food Chemistry* 43: 3026-3028.
- Larrigaudiére, C., Lentheric, I., Pinto, E. and Vendrell, M. 2001. Short-term effects of air and controlled atmosphere storage on antioxidant metabolism in Conference pears. *Journal of Plant Physiology* 158: 1015-1022.
- Larrigaudiére, C., Vilaplana, R., Soria, Y. and Recasens, I. 2004. Oxidative behaviour of Blanquilla pears treated with 1–methylcyclopropene during cold storage. *Journal of the Science of Food and Agriculture* 84: 1871-1877.
- Lee, J., Rudell, D.R., Davies, P.J. and Watkins, C.B. 2012. Metabolic changes in 1-methylcyclopropene (1-MCP)-treated 'Empire' apple fruit during storage. *Metabolomics* 8: 742-753.
- Lester, G. 1990. Lipoxygenase activity of hypodermal-and middle-mesocarp tissues from netted muskmelon fruit during maturation and storage. *Journal of the American Society for Horticultural Science* 115: 612-615.
- Lester, G. 1996. Calcium alters senescence rate of postharvest muskmelon fruit disks. *Postharvest Biology and Technology* 7: 91-96.
- Lester, G.E. and Dunlap, J.R. 1985. Physiological changes during development and ripening of 'Perlita' muskmelon fruits. *Scientia Horticulturae* 26: 323-331.

- Lester, G. 1997. Melon (*Cucumis melo* L.) fruit nutritional quality and health functionality. *HortTechnology* 7: 222-227.
- Lester, G.E. 2008. Antioxidant, sugar, mineral, and phytonutrient concentrations across edible fruit tissues of orange-fleshed honeydew melon (*Cucumis melo* L.). *Journal of Agricultural and Food Chemistry* 56: 3694-3698.
- Lester, G.E. and Bruton, B.D. 1986. Relationship of netted muskmelon fruit water loss to postharvest storage life. *Journal of the American Society for Horticultural Science* 111: 727-731.
- Lester, G.E. and Crosby, K.M. 2002. Ascorbic acid, folic acid, and potassium content in postharvest green-flesh honeydew muskmelons: Influence of cultivar, fruit size, soil type, and year. *Journal of the American Society for Horticultural Science* 127: 843-847.
- Lester, G.E. and Eischen, F. 1996. Beta-carotene content of postharvest orangefleshed muskmelon fruit: effect of cultivar, growing location and fruit size. *Plant Foods for Human Nutrition* 49: 191-197.
- Lester, G.E. and Hodges, D.M. 2008. Antioxidants associated with fruit senescence and human health: Novel orange-fleshed non-netted honey dew melon genotype comparisons following different seasonal productions and cold storage durations. *Postharvest Biology and Technology* 48: 347-354.
- Lester, G.E., Jifon, J.L. and Rogers, G. 2005. Supplemental foliar potassium applications during muskmelon fruit development can improve fruit quality, ascorbic acid, and beta-carotene contents. *Journal of the American Society for Horticultural Science* 130: 649-653.
- Liang, Y.S., Wongmetha, O., Wu, P.S. and Ke, L.S. 2013. Influence of hydrocooling on browning and quality of litchi cultivar Feizixiao during storage. *International Journal of Refrigeration* 36: 1173-1179.
- Liao, Y.T., Syamaladevi, R.M., Zhang, H., Killinger, K. and Sablani, S. 2017. Inactivation of *Listeria monocytogenes* on frozen red raspberries by using UV-C Light. *Journal of Food Protection* 80: 545-550.
- Lin, T.T. and Pitt, R.E. 1986. Rheology of apple and potato tissue as affected by cell turgor pressure. *Journal of Texture Studies* 17: 291-313.
- Liu, L., Kakihara, F., and Kato, M. 2004. Characterization of six varieties of *Cucumis melo* L. based on morphological and physiological characters, including shelf-life of fruit. *Euphytica* 135: 305-313.
- López Camelo, A.F. and Gómez, P.A. 2004. Comparison of color indexes for tomato ripening. *Horticultura Brasileira* 22: 534-537.

- Luengwilai, K., Saltveit, M. and Beckles, D.M. 2012. Metabolite content of harvested Micro-Tom tomato (*Solanum lycopersicum* L.) fruit is altered by chilling and protective heat-shock treatments as shown by GC–MS metabolic profiling. *Postharvest Biology and Technology* 63: 116-122.
- Lurie, S., Shapiro, B. and Ben-Yehoshua, S. 1986. Effects of water stress and degree of ripeness on rate of senescence of harvested bell pepper fruit. *Journal of the American Society for Horticultural Science* 111: 880-885.
- Luza, J.G., Van Gorsel, R., Polito, V.S. and Kader, A.A. 1992. Chilling injury in peaches: a cytochemical and ultrastructural cell wall study. *Journal of the American Society for Horticultural Science* 117: 114-118.
- Lyons, J.M., McGlasson, W.B. and Pratt, H.K. 1962. Ethylene production, respiration, and internal gas concentrations in cantaloupe fruits at various stages of maturity. *Plant Physiology* 37: 31-36.
- Lyons, J.M. 1973. Chilling injury in plants. *Annual Review of Plant Physiology* 24: 445-466.
- Ma, W., Cao, J., Ni, Z., Tian, W., Zhao, Y. and Jiang, W. 2012. Effects of 1methylcyclopropene on storage quality and antioxidant quality of harvested "Yujinxiang" melon (*Cucumis melo* L.) fruit. *Journal of Food Biochemistry* 36: 413-420.
- MacDonald, M.J. and D'Cunha, G.B. 2007. A modern view of phenylalanine ammonia lyase. *Biochemistry and Cell Biology* 85: 273-282.
- Macheix, J.J. and Fleuriet, A. 1990. Fruit Phenolics. Floride: CRC press.
- MacRae, E., Quick, W.P., Benker, C. and Stitt, M. 1992. Carbohydrate metabolism during postharvest ripening in kiwifruit. *Planta* 188: 314-323.
- Mahajan, P.V., Oliveira, F.A.R. and Macedo, I. 2008. Effect of temperature and humidity on the transpiration rate of the whole mushrooms. *Journal of Food Engineering* 84: 281-288.
- Makwana, S.A., Polara, N.D. and Viradia, R.R. 2014. Effect of pre-cooling on post harvest life of mango (*Mangifera indica* L.) cv. Kesar. *Food Science and Technology* 2: 6-13.
- Mallek-Ayadi, S., Bahloul, N. and Kechaou, N. 2017. Characterization, phenolic compounds and functional properties of *Cucumis melo* L. peels. *Food chemistry* 221: 1691-1697.
- Manganaris, G.A., Ilias, I.F., Vasilakakis, M. and Mignani, I. 2007. The effect of hydrocooling on ripening related quality attributes and cell wall

physicochemical properties of sweet cherry fruit (*Prunus avium* L.). *International Journal of Refrigeration* 30: 1386-1392.

- Mayberry, K.S. and Hartz, T.K. 1992. Extension of muskmelon storage through the use of hot water life treatment and polyethylene wraps. *HortScience* 27: 324-326.
- Mei, X., Chen, Y., Zhang, L., Fu, X., Wei, Q., Grierson, D., Zhou, Y., Huang, Y., Dong, F. and Yang, Z. 2016. Dual mechanisms regulating glutamate decarboxylases and accumulation of gamma-aminobutyric acid in tea (*Camellia sinensis*) leaves exposed to multiple stresses. *Scientific Reports* 6: 23685-23695.
- Melgar-Lalanne, G., Hernández-Álvarez, A.J., Jiménez-Fernández, M. and Azuara, E. 2017. Oleoresins from *Capsicum* spp.: extraction methods and bioactivity. *Food and Bioprocess Technology* 10: 51-76.
- McCollum, T.G., Huber, D.J. and Cantliffe, D.J. 1989. Modification of polyuronides and hemicelluloses during muskmelon fruit softening. *Physiologia Plantarum* 76: 303-308.
- McDonald, K. and Sun, D.W. 2000. Vacuum cooling technology for the food processing industry: A review. *Journal of Food Engineering* 45: 55-65.
- Menon, S.V. and Rao, T.R. 2012. Nutritional quality of muskmelon fruit as revealed by its biochemical properties during different rates of ripening. *International Food Research Journal* 19: 1621-1628.
- Milind, P. and Kulwant, S. 2011. Muskmelon is eat-must melon. International Research Journal of Pharmacy 2: 52-57.
- Mishra, S., Jha, A.B. and Dubey, R.S. 2011. Arsenite treatment induces oxidative stress, upregulates antioxidant system, and causes phytochelatin synthesis in rice seedlings. *Protoplasma* 248: 565-577.
- Mitchell, F.G., Mayer, G., Maxie, E.C. and Coates, W.W. 1974. Cold storage effects on fresh market peaches, nectarines and plums. I. Estimating freezing points. II. Using low temperatures to delay internal breakdown. *California Agriculture* 28: 12-14.
- Mittler, R. 2002. Oxidative stress, antioxidants and stress tolerance. *Trends in Plant Science* 7: 405-410.
- Mliki, A., Staub, J.E., Zhangyong, S. and Ghorbel, A. 2001. Genetic diversity in melon (*Cucumis melo* L.): An evaluation of African germplasm. *Genetic Resources and Crop Evolution* 48: 587-597.
- Moing, A., Aharoni, A., Biais, B., Rogachev, I., Meir, S., Brodsky, L., Allwood, J.W., Erban, A., Dunn, W.B., Kay, L., de Koning, S., de Vos, R.C.H.,

Jonker, H., Mumm, R., Deborde, C., Maucourt, M., Bernillon, S., Gibon, Y., Hansen, T.H., Husted, S., Goodacre, R., Kopka, J., Schjoerring, J.K., Rolin, D. and Hall, R.D. 2011. Extensive metabolic cross-talk in melon fruit revealed by spatial and developmental combinatorial metabolomics. *New Phytologist* 190: 683-696.

- Mondal, K., Sharma, N.S., Malhotra, S.P., Dhawan, K. and Singh, R. 2004. Antioxidant systems in ripening tomato fruits. *Biologia Plantarum* 48: 49-53.
- Montes, C., Vicario, I.M., Raymundo, M., Fett, R. and Heredia, F.J. 2005. Application of tristimulus colorimetry to optimize the extraction of anthocyanins from Jaboticaba (*Myricia jaboticaba* Berg.). *Food Research International* 38: 983-988.
- Mori, K., Goto-Yamamoto, N., Kitayama, M. and Hashizume, K. 2007. Loss of anthocyanins in red-wine grape under high temperature. *Journal of Experimental Botany* 58: 1935-1945.
- Mworia, E.G., Yoshikawa, T., Salikon, N., Oda, C., Asiche, W.O., Yokotani, N., Abe, D., Ushijima, K., Nakano, R. and Kubo, Y. 2012. Low-temperaturemodulated fruit ripening is independent of ethylene in 'Sanuki Gold' kiwifruit. *Journal of Experimental Botany* 63: 963-971.
- Nagata, M. and Yamashita, I. 1992. Simple method for simultaneous determination of chlorophyll and carotenoids in tomato fruit. *Journal-Japanese Society of Food Science and Technology* 39: 925-925.
- Nakano, Y. and Asada, K. 1981. Hydrogen peroxide is scavenged by ascorbatespecific peroxidase in spinach chloroplasts. *Plant and Cell Physiology* 22: 867-880.
- Nalbandi, H., Seiiedlou, S., Ghasemzadeh, H.R. and Rangbar, F. 2016. Innovative parallel airflow system for forced-air cooling of strawberries. *Food and Bioproducts Processing* 100: 440-449.
- Nampan, K., Techavuthiporn, C. and Kanlayanarat, S. 2006. Hydrocooling improves quality and storage life of "Rong-Rein" rambutan (*Nephelium lappaceum* L.) fruit. *Acta Horticulturae* 712: 763-770.
- Niranjana, P., Gopalakrishna, K.P.R., Sudhakar, D.V.R. and Madhusudhan, B. 2009. Effect of pre-cooling and heat treatment on antioxidant enzymes profile of mango and banana. *African Journal of Food, Agriculture, Nutrition and Development* 9: 1211-1225.
- Nunes, M.C.D.N. 2008. Impact of environmental conditions on fruit and vegetable quality. *Stewart Postharvest Review* 4: 1-14.

- Nunes, M.C.N., Brecht, J.K., Morais, A.M.M.B. and Sargent, S.A. 1995a. Physical and chemical quality characteristics of strawberries after storage are reduced by a short delay to cooling. *Postharvest Biology and Technology* 6: 17-28.
- Nunes, M.C.N., Brecht, J.K., Sargent, S.A. and Morais, A.M.M.B. 1995b. Effects of delays to cooling and wrapping on strawberry quality (cv. Sweet Charlie). *Food Control* 6: 323-328.
- Nuñez-Palenius, H.G., Gomez-Lim, M., Ochoa-Alejo, N., Grumet, R., Lester, G. and Cantliffe, D.J. 2008. Melon fruits: genetic diversity, physiology, and biotechnology features. *Critical Reviews in Biotechnology* 28: 13-55.
- O'Sullivan, J.L., Ferrua, M.J., Love, R., Verboven, P., Nicolaï, B. and East, A. 2016. Modelling the forced-air cooling mechanisms and performance of polylined horticultural produce. *Postharvest Biology and Technology* 120: 23-35.
- O'Sullivan, J.L., Ferrua, M.J., Love, R., Verboven, P., Nicolaï, B. and East, A. 2017. Forced-air cooling of polylined horticultural produce: Optimal cooling conditions and package design. *Postharvest Biology and Technology* 126: 67-75.
- Padayachee, A., Netzel, G., Netzel, M., Day, L., Zabaras, D., Mikkelsen, D. and Gidley, M.J. 2012. Binding of polyphenols to plant cell wall analogues– Part 1: Anthocyanins. *Food Chemistry* 134: 155-161.
- Paniagua, A.C., East, A.R., Hindmarsh, J.P. and Heyes, J. 2013. Moisture loss is the major cause of firmness change during postharvest storage of blueberry. *Postharvest Biology and Technology* 79: 13-19.
- Parsons, R. 1998. American Society of Heating, Refrigerating and Air-Conditioning Engineers Handbook Refrigeration. United States: American Society of Heating, Refrigerating and Air-Conditioning Engineers.
- Pathare, P.B., Opara, U.L., Vigneault, C., Delele, M.A. and Al-Said, F.A.J. 2012. Design of packaging vents for cooling fresh horticultural produce. *Food and Bioprocess Technology* 5: 2031-2045.
- Paull, R. 1999. Effect of temperature and relative humidity on fresh commodity quality. *Postharvest Biology and Technology* 15: 263-277.
- Pedreschi, R., Franck, C., Lammertyn, J., Erban, A., Kopka, J., Hertog, M., Verlinden, B. and Nicolaï, B. 2009. Metabolic profiling of 'Conference' pears under low oxygen stress. *Postharvest Biology and Technology* 51: 123-130.

- Pelletier, W., Brecht, J.K., do Nascimento Nunes, M.C. and Émond, J.P. 2011. Quality of strawberries shipped by truck from California to Florida as influenced by postharvest temperature management practices. *HortTechnology* 21: 482-493.
- Perkins-Veazie, P., Beaulieu, J.C. and Siddiq, M. 2012. Watermelon, cantaloupe and honeydew. In *Tropical and Subtropical Fruits: Postharvest Physiology, Processing and Packaging*, ed Siddiq, M, Ahmed, J., Lobo, M.G. and Ozadali, F, 549-568. New Delhi: John Wiley & Sons, Inc,
- Phan, A.D.T., Flanagan, B.M., D'Arcy, B.R. and Gidley, M.J. 2017. Binding selectivity of dietary polyphenols to different plant cell wall components: Quantification and mechanism. *Food Chemistry* 233: 216-227.
- Phebe, D., Chew, M.K., Suraini, A.A., Lai, O.M. and Janna, O.A. 2009. Redfleshed pitaya (*Hylocereus polyrhizus*) fruit colour and betacyanin content depend on maturity. *International Food Research Journal* 16: 233-242.
- Purvis, A.C. 1984. Importance of water loss in the chilling injury of grapefruit stored at low temperature. *Scientia Horticulturae* 23: 261-267.
- Purvis, A.C. and Grierson, W. 1982. Accumulation of reducing sugar and resistance of grapefruit peel to chilling injury as related to winter temperatures. *Journal of American Society for Horticultural Science* 107: 139-142.
- Purvis, A.C. and Shewfelt, R.L. 1993. Does the alternative pathway ameliorate chilling injury in sensitive plant tissues? *Physiologia Plantarum* 88: 712-718.
- Quettier-Deleu, C., Gressier, B., Vasseur, J., Dine, T., Brunet, C., Luyckx, M., Cazin, M., Cazin, J., Bailleul, F. and Trotin, F. 2000. Phenolic compounds and antioxidant activities of buckwheat (*Fagopyrum esculentum* Moench) hulls and flour. *Journal of Ethnopharmacology* 72: 35-42.
- Rab, A., Rehman, H., Haq, I., Sajid, M., Nawab, K. and Ali, K. 2013. Harvest stages and pre-cooling influence the quality and storage life of tomato fruit. *Journal of Animal and Plant Sciences* 23: 1347-1352.
- Ranganna, S. 1977. Manual of analysis of fruit and vegetable products. New Delhi: Tata McGraw Hill Publishing Co. Ltd.
- Rice-Evans, C., Miller, N. and Paganga, G. 1997. Antioxidant properties of phenolic compounds. *Trends in Plant Science* 2: 152-159.
- Robles-Sánchez, R.M., Islas-Osuna, M.A., Astiazarán-García, H., Vázquez-Ortiz, F.A., Martín-Belloso, O., Gorinstein, S. and González-Aguilar,

G.A. 2009. Quality index, consumer acceptability, bioactive compounds, and antioxidant activity of fresh-cut "Ataulfo" mangoes (*Mangifera indica* L.) as affected by low-temperature storage. *Journal of Food Science* 74: 126-134.

- Rodoni, L.M., Hasperué, J.H., Ortiz, C.M., Lemoine, M.L., Concellón, A. and Vicente, A.R. 2016. Combined use of mild heat treatment and refrigeration to extend the postharvest life of organic pepper sticks, as affected by fruit maturity stage. *Postharvest Biology and Technology* 117: 168-176.
- Rodríguez-Pérez, C., Quirantes-Piné, R., Fernández-Gutiérrez, A. and Segura-Carretero, A. 2013. Comparative characterization of phenolic and other polar compounds in Spanish melon cultivars by using high-performance liquid chromatography coupled to electrospray ionization quadrupoletime of flight mass spectrometry. *Food Research International* 54: 1519-1527.
- Rojas, A.M., Castro, M.A., Alzamora, S.M. and Gerschenson, L.N. 2001. Turgor pressure effects on textural behavior of honeydew melon. *Journal of Food Science* 66: 111-117.
- Rolim, P.M., Fidelis, G.P., Padilha, C.E.A., Santos, E.S., Rocha, H.A.O. and Macedo, G.R. 2018. Phenolic profile and antioxidant activity from peels and seeds of melon (*Cucumis melo* L. var. *reticulatus*) and their antiproliferative effect in cancer cells. *Brazilian Journal of Medical and Biological Research* doi: 10.1590/1414-431X20176069.
- Rose, J.K., Catalá, C., Gonzalez-Carranza, Z.H. and Roberts, J.A. 2003. Cell wall disassembly. *Annual Plant Reviews* 8: 264-324.
- Rose, J.K.C., Hadfield, K.A., Labavitch, J.M. and Bennett, A.B. 1998. Temporal sequence of cell wall disassembly in rapidly ripening melon fruit. *Plant Physiology* 117: 345-361.
- Rudnucki, R.M., Nowak, J. and Goszczynska, D.M. 1991. Cold storage and transportation conditions for cut flowers cuttings and potted plants. *Acta Horticulturae* 298: 225-320.
- Ruelland, E., Vaultier, M.N., Zachowski, A. and Hurry, V. 2009. Cold signaling and cold acclimation in plants. *Advances in Botanical Research* 49: 35-150.
- Ruiz-Garcia, Y., Smith, P.A. and Bindon, K.A. 2014. Selective extraction of polysaccharide affects the adsorption of proanthocyanidin by grape cell walls. *Carbohydrate Polymers* 114: 102-114.
- Rux, G., Schlüter, O., Geyer, M. and Herppich, W.B. 2017. Characterization of high hydrostatic pressure effects on fresh produce cell turgor using

pressure probe analyses. *Postharvest Biology and Technology* 132: 188-194.

- Saba, M.K. and Moradi, S. 2017. Sodium nitroprusside (SNP) spray to maintain fruit quality and alleviate postharvest chilling injury of peach fruit. *Scientia Horticulturae* 216: 193-199.
- Sajid, M., Mukhtiar, M., Rab, A., Shah, S.T. and Jan, I. 2014. Influence of calcium chloride (CaCl₂) on fruit quality of pear (*Pyrus communis*) cv. Le Conte during storage. *Pakistan Journal of Agricultural Sciences* 51: 113-121.
- Sala, J.M. 1998. Involvement of oxidative stress in chilling injury in cold-stored mandarin fruits. *Postharvest Biology and Technology* 13: 255-261.
- Sala, J.M. and Lafuente, M.T. 2000. Catalase enzyme activity is related to tolerance of mandarin fruits to chilling. *Postharvest Biology and Technology* 20: 81-89.
- Sala, J.M. and Lafuente, M.T. 2004. Antioxidant enzymes activities and rind staining in 'Navelina' oranges as affected by storage relative humidity and ethylene conditioning. *Postharvest Biology and Technology* 31: 277-285.
- Saladié, M., Matas, A.J., Isaacson, T., Jenks, M.A., Goodwin, S.M., Niklas, K.J., Xiaolin, R., Labavitch, J.M., Shackel, K.A., Fernie, A.R., Lytovchenko, A., O'Neill, M.A., Watkins, C.B. and Rose, J.K.C. 2007. A reevaluation of the key factors that influence tomato fruit softening and integrity. *Plant Physiology* 144: 1012-1028.
- Sargent, S.A., Talbot, M.T. and Brecht, J.K. 1988. Evaluating precooling methods for vegetable packinghouse operations. *Proceedings of the Florida State Horticultural Society* 101: 175-182.
- Scott, G., Rupar, M., Fletcher, A.G.D., Dickinson, M. and Shama, G. 2017. A comparison of low intensity UV-C and high intensity pulsed polychromatic sources as elicitors of hormesis in tomato fruit. *Postharvest Biology and Technology* 125: 52-58.
- Sekse, L., Wermund, U., Vidrih, R., Simcic, M. and Vangdal, E. 2011. Fruit firmness as related to quality attributes in two plum cultivars (*Prunus domestica* L.) of different maturity. *The European Journal of Plant Science and Biotechnology* 5: 93-97.
- Senapati, A.K., Raj, D., Jain, R. and Patel, N.L. 2016. Advances in packaging and storage of flowers. In *Commercial Horticulture*, ed. N.L. Patel, S.L. Chawla and T.R. Ahlawat, 473-488. New Delhi: New India Publishing Agency.

- Senthilkumar, S., Vijayakumar, R.M. and Kumar, S. 2015. Advances in precooling techniques and their implications in horticulture sector: A review. *International Journal of Environmental and Agricultural Research* 1: 24-30.
- Serradilla, M.J., Bernalte-Garcia, M.J. and Lopez-Corrales, M. Eds. 2013. Preceedings from VII International Cherry Symposium 1161: Precooling application before cold storage delayed ripening and maintained high antioxidant activity of 'Sonata' sweet cherry. Díaz-Mula, H.M., Valverde, J.M., Martínez-Romero, D., Zapata, P.J., Castillo, S. and Serrano, M: Acta Horticulturae 1161: 561-568.
- Sevillano, L., Sanchez-Ballesta, M.T., Romojaro, F. and Flores, F.B. 2009. Physiological, hormonal and molecular mechanisms regulating chilling injury in horticultural species. Postharvest technologies applied to reduce its impact. *Journal of the Science of Food and Agriculture* 89: 555-573.
- Seymour, G.B. and McGlasson. W.B. 1993. Melons. In *Biochemistry of Fruit Ripening*, ed. G. Seymour, J. Taylor, and G. Tucker, pp. 273-290. United Kingdom: Chapman and Hall, Cambridge University Press.
- Seymour, G.B., Tucker, G. and Leach, L.A. 2004. Cell adhesion molecules in plants and animals. *Biotechnology and Genetic Engineering Reviews* 21: 123-132.
- Shahid, M., Salim, J., Noor, M.R.M., Hamid, A.H.A., Manas, M.A. and Ahmad, S.A. 2009. *Manual Teknologi Fertigasi Penanaman Cili, Rockmelon dan Tomato*. Kuala Lumpur: Institut Penyelidikan dan Kemajuan Pertanian Malaysia (MARDI).
- Shellie, K.C. and Lester, G. 2004. Netted melons. In: K.C., Gross, C.Y., Wang. and M. Saltveit (Ed), *The commercial storage of fruits, vegetables, and florist and nursery stocks* (pp. 423-426). Washington: United States Department of Agriculture.
- Shewfelt, R.L. and Del Rosario, B.A. 2000. The role of lipid peroxidation in storage disorders of fresh fruits and vegetables. *HortScience* 35: 575-579.
- Simandjuntak, V., Barrett, D.M. and Wrolstad, R.E. 1996. Cultivar and frozen storage effects on muskmelon (*Cucumis melo*) colour, texture and cell wall polysaccharide composition. *Journal of the Science of Food and Agriculture* 71: 291-296.
- Singh, R. and Dwivedi, U.N. 2008. Effect of ethrel and 1–methylcyclopropene (1–MCP) on antioxidants in mango (*Mangifera indica* var. Dashehari) during fruit ripening. *Food Chemistry* 111: 951-956.

- Singh, R.P. and Chakraverty, A. 2001. *Postharvest technology: cereals, pulses, fruits and vegetables*. Madison: Science Publishers.
- Sohail, M., Ayub, M., Khalil, S.A., Zeb, A., Ullah, F., Afridi, S.R. and Ullah, R. 2015. Effect of calcium chloride treatment on post harvest quality of peach fruit during cold storage. *International Food Research Journal* 22: 2225-2229.
- Srivastava, S. and Dubey, R.S. 2011. Manganese-excess induces oxidative stress, lowers the pool of antioxidants and elevates activities of key antioxidative enzymes in rice seedlings. *Plant Growth Regulation 64*: 1-16.
- Stepansky, A. and Kovalski, I. 1999. Intraspecific classification of melons (*Cucumis melo* L.) in view of their phenotypic and molecular variation. *Plant Systematics Evolution* 217: 313-332.
- Stincone, A., Prigione, A., Cramer, T., Wamelink, M., Campbell, K., Cheung, E., Olin-Sandoval, V., Grüning, N.M., Krüger, A., Alam, M.T., Keller, M.A., Breitenbach, M., Brindle, K.M., Rabinowitz, J.D. and Ralser, M. 2015. The return of metabolism: biochemistry and physiology of the pentose phosphate pathway. *Biological Reviews* 90: 927-963.
- Storey, R. and Treeby, M.T. 1994. The morphology of epicuticular wax and albedo cells of orange fruit in relation to albedo breakdown. *Journal of Horticultural Science* 69: 329-338.
- Sun, D.W. and Zheng, L. 2006. Vacuum cooling technology for the agri-food industry: Past, present and future. *Journal of Food Engineering* 77: 203-214.
- Supapvanich, S. and Tucker, G.A. 2013. Cell wall hydrolysis in netted melon fruit (*Cucumis melo var. reticulatus* L. Naud) during storage. *Chiang Mai Journal of Science* 40: 447-458.
- Takahama, U. and Oniki, T. 1997. A peroxidase/phenolics/ascorbate system can scavenge hydrogen peroxide in plant cells. *Physiologia Plantarum* 101: 845-852.
- Tamagnone, L., Merida, A., Stacey, N., Plaskitt, K., Parr, A., Chang, C.F., Lynn, D., Dow, J.M., Roberts, K. and Martin, C. 1998. Inhibition of phenolic acid metabolism results in precocious cell death and altered cell morphology in leaves of transgenic tobacco plants. *The Plant Cell* 10: 1801-1816.
- Tang, M., Bie, Z.L., Wu, M.Z., Yi, H.P. and Feng, J.X. 2010. Changes in organic acids and acid metabolism enzymes in melon fruit during development. *Scientia Horticulturae* 123: 360-365.

- Tao, F., Zhang, M. and Yu, H.Q. 2007. Effect of vacuum cooling on physiological changes in the antioxidant system of mushroom under different storage conditions. *Journal of Food Engineering* 79: 1302-1309.
- Thomas, P. and Joshi, M.R. 1988. Reduction of chilling injury in ripe Alphonso mango fruit in cold storage by temperature conditioning. *International Journal of Food Science and Technology* 23: 447-455.
- Thomashow, M.F. 2010. Molecular basis of plant cold acclimation: insights gained from studying the CBF cold response pathway. *Plant Physiology* 154: 571-577.
- Thompson, A.K. 2003. *Fruits and Vegetables: Harvesting, Handling and Storage*. Oxford: Blackwell publishing.
- Todd, J.F., Paliyath, G. and Thompson, J.E. 1990. Characteristics of a membrane-associated lipoxygenase in tomato fruit. *Plant Physiology* 94: 1225-1232.
- Tohge, T., Watanabe, M., Hoefgen, R. and Fernie, A.R. 2013. Shikimate and phenylalanine biosynthesis in the green lineage. *Frontiers in Plant Science* 4: 1-13.
- Toivonen, P.M. 1997. The effects of storage temperature, storage duration, hydro-cooling, and micro-perforated wrap on shelf life of broccoli (*Brassica oleracea* L., Italica Group). *Postharvest Biology and Technology* 10: 59-65.
- Toivonen, P.M.A. 2004. Postharvest storage procedures and oxidative stress. *HortScience* 39: 938-942.
- Toivonen, P.M.A. and Brummell, D.A. 2008. Biochemical bases of appearance and texture changes in fresh-cut fruit and vegetables. *Postharvest Biology and Technology* 48: 1-14.
- Tripathy, B.C. and Oelmüller, R. 2012. Reactive oxygen species generation and signaling in plants. *Plant Signaling and Behavior* 7: 1621-1633.
- Turmanidze, T., Gulua, L., Jgenti, M. and Wicker, L. 2017. Potential antioxidant retention and quality maintenance in raspberries and strawberries treated with calcium chloride and stored under refrigeration. *Brazilian Journal of Food Technology* doi: 10.1590/1981-6723.8916.
- Tutar, M., Erdogdu, F. and Toka, B. 2009. Computational modeling of airflow patterns and heat transfer prediction through stacked layers' products in a vented box during cooling. *International Journal of Refrigeration* 32: 295-306.

- Tzin, V. and Galili, G. 2010. The biosynthetic pathways for shikimate and aromatic amino acids in *Arabidopsis thaliana*. *The Arabidopsis Book* 8: 1-18.
- Van Den Bogaart, G., Hermans, N., Krasnikov, V., de Vries, A.H. and Poolman, B. 2007. On the decrease in lateral mobility of phospholipids by sugars. *Biophysical Journal* 92: 1598-1605.
- Van den Ende, W. and Valluru, R. 2008. Sucrose, sucrosyl oligosaccharides, and oxidative stress: scavenging and salvaging?. *Journal of Experimental Botany* 60: 9-18.
- Vallone, S., Sivertsen, H., Anthon, G.E., Barrett, D.M., Mitcham, E.J., Ebeler, S.E. and Zakharov, F. 2013. An integrated approach for flavour quality evaluation in muskmelon (*Cucumis melo* L. *reticulatus* group) during ripening. *Food Chemistry* 139: 171-183.
- Velioglu, Y.S., Mazza, G., Gao, L. and Oomah, B.D. 1998. Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of Agricultural and Food Chemistry* 46: 4113-4117.
- Vigneault, C. and Goyette, B. 2001. Loss of ice through container openings during liquid-ice cooling of horticultural crops. *Canadian Biosystems Engineering* 43: 3-45.
- Vigneault, C., Thompson, J. and Wu, S. 2009. Designing container for handling fresh horticultural produce. *Postharvest Technologies for Horticultural Crops* 2: 25-47.
- Vilaplana, R., Valentines, M.C., Toivonen, P.M.A. and Larrigaudiére. C. 2006. Antioxidant potential and peroxidative state of 'Golden Smoothee' apples treated with 1–methylcyclopropene. *Journal of the American Society for Horticultural Science* 131: 104-109.
- Villano, D., Fernández-Pachón, M.S., Moyá, M.L., Troncoso, A.M. and García-Parrilla, M.C. 2007. Radical scavenging ability of polyphenolic compounds towards DPPH free radical. *Talanta* 71: 230-235.
- Villanueva, M.J., Tenorio, M.D., Esteban, M.A. and Mendoza, M.C. 2004. Compositional changes during ripening of two cultivars of muskmelon fruits. *Food Chemistry* 87: 179-185.
- Vouldoukis, I., Lacan, D., Kamate, C., Coste, P., Calenda, A., Mazier, D., Conti, M. and Dugas, B. 2004. Antioxidant and anti-inflammatory properties of a *Cucumis melo* LC. extract rich in superoxide dismutase activity. *Journal of Ethnopharmacology* 94: 67-75.
- Wakabayashi, K. 2000. Changes in cell wall polysaccharides during fruit ripening. *Journal of Plant Research* 113: 231-237.

- Wallace, G. and Fry, S.C. 1994. Phenolic components of the plant cell wall. *International Review of Cytology* 151: 229-267.
- Wang, B., Wang, J., Feng, X., Lin, L., Zhao, Y. and Jiang, Y. 2009. Effects of 1– MCP and exogenous ethylene on fruit ripening and antioxidants in stored mango. *Plant Growth Regulation* 57: 185-192.
- Wang, K., Shao, X., Gong, Y., Zhu, Y., Wang, H., Zhang, X., Yu, D., Yu, F., Qiu, Z. and Lu, H. 2013. The metabolism of soluble carbohydrates related to chilling injury in peach fruit exposed to cold stress. *Postharvest Biology and Technology* 86: 53-61.
- Wang, L., Chen, S., Kong, W., Li, S. and Archbold, D.D. 2006. Salicylic acid pretreatment alleviates chilling injury and affects the antioxidant system and heat shock proteins of peaches during cold storage. *Postharvest Biology and Technology* 41: 244-251.
- Wang, Y.S., Tian, S.P. and Xu, Y. 2005. Effects of high oxygen concentration on pro- and anti-oxidant enzymes in peach fruits during postharvest periods. *Food Chemistry* 91: 99-104.
- Wang, Y.S., Tian, S.P., Xu, Y., Qin, G.Z. and Yao, H. 2004. Changes in the activities of pro- and anti-oxidant enzymes in peach fruit inoculated with *Cryptococcus laurentii* or *Penicillium expansum* at 0 or 20°C. *Postharvest Biology and Technology* 34: 21-28.
- Wills, R. and Golding, J. 2016. Postharvest: An Introduction to the Physiology and Handling of Fruit and Vegetables. Sydney: New South Wales University Press.
- Wills, R.B.H., McGlasson, W.B., Graham, D. and Joyce, D. 1998. Postharvest-An Introduction to the Physiology and Handling of Fruit, Vegetables and Ornamental. 4th ed. Sydney: New South Wales University Press.
- Wisniewski, M., Spadoni, A., Guidarelli, M., Phillips, J. and Mari, M. 2015. Transcriptional profiling of apple fruits in response to heat treatment: involvement of a defense response during *P. expansum* infection. *Postharvest Biology and Technology* 101: 32-48.
- Woolf, A.B., Cox, K.A., White, A. and Ferguson, I.B., 2003. Low temperature conditioning treatments reduce external chilling injury of 'Hass' avocados. *Postharvest Biology and Technology* 28: 113-122.
- Xanthopoulos, G.T., Templalexis, C.G., Aleiferis, N.P. and Lentzou, D.I. 2017. The contribution of transpiration and respiration in water loss of perishable agricultural products: The case of pears. *Biosystems Engineering* 158: 76-85.

- Yahia, E.M. 2011. Postharvest Biology and Technology of Tropical and Subtropical Fruits: Fundamental Issues. Philadelphia: Woodhead Publishing Limited.
- Yahia, E.M. and Rivera, M. 1992. Modified atmosphere packaging of muskmelon. *Lebensmittel-Wissenschaft und-Technologie* 25: 38-42.
- Yan, J., Song, Y., Li, J. and Jiang, W. 2017. Forced-air precooling treatment enhanced antioxidant capacities of apricots. *Journal of Food Processing* and Preservation. https://doi.org/ 10.1111/jfpp.13320
- Yan, W., Bai, L., Zhang, L., Chen, G., Fan, J., Gu, X., Cui, W. and Guo. Z. 2010. Comparative study for cold acclimation physiological indicators of *Forythia mandshurica* Uyeki and *Forsythia viridissima* Indl. *Middle East Journal of Scientific Research* 6: 556-562.
- Yang, Q., Rao, J., Yi, S., Meng, K., Wu, J. and Hou, Y. 2012. Antioxidant enzyme activity and chilling injury during low-temperature storage of Kiwifruit cv. Hongyang exposed to gradual postharvest cooling. *Horticulture, Environment, and Biotechnology* 53: 505-512.
- Yang, T., Zhang, L., Zhang, T., Zhang, H., Xu, S. and An, L. 2005. Transcriptional regulation network of cold-responsive genes in higher plant. *Plant Science* 169: 987-995.
- Yoo, H.J., Park, W.J., Lee, G.M., Oh, C.S., Yeam, I., Won, D.C., Kim, C.K. and Lee, J.M. 2017. Inferring the genetic determinants of fruit colors in tomato by carotenoid profiling. *Molecules* 22: 764-777.
- Zhang, L., Cleland, A.C. and Cleland, D.J. 1996. A simple method for prediction of chilling times for objects of two-dimensional irregular shape. *International Journal of Refrigeration* 19: 95-106.
- Zhang, Z., Nakano, K. and Maezawa, S. 2009. Comparison of the antioxidant enzymes of broccoli after cold or heat shock treatment at different storage temperatures. *Postharvest Biology and Technology* 54: 101-105.
- Zhang, Z., Zhu, Q., Hu, M., Gao, Z., An, F., Li, M. and Jiang, Y. 2017. Lowtemperature conditioning induces chilling tolerance in stored mango fruit. *Food Chemistry* 219: 76-84.
- Zhao, Z., Cao, J., Jiang, W., Gu, Y. and Zhao, Y. 2009. Maturity–related chilling tolerance in mango fruit and the antioxidant capacity involved. *Journal of the Science of Food and Agriculture* 89: 304-309.
- Zhao, Z., Jiang, W., Cao, J., Zhao, Y. and Gu, Y. 2006. Effect of cold–shock treatment on chilling injury in mango (*Mangifera indica* L. cv. 'Wacheng') fruit. *Journal of the Science of Food and Agriculture* 86: 2458-2462.

- Zheng, L.Y. and Sun, D.W. 2004. Vacuum cooling for the food industry-a review of recent research advances. *Trends in Food Science and Technology* 15: 555-568.
- Zheng, X., Tian, S., Meng, X. and Li, B. 2007. Physiological and biochemical responses in peach fruit to oxalic acid treatment during storage at room temperature. *Food Chemistry* 104: 156-162.
- Zheng, X.Y. Wolff, D.W. 2000. Ethylene production, shelf-life and evidence of RFLP polymorphisms linked to ethylene genes in melon (*Cucumis melo* L.). *Theoretical and Applied Genetics* 101: 613-624.
- Zhou, R., Wang, X., Hu, Y., Zhang, G., Yang, P. and Huang, B. 2015. Reduction in Hami melon (*Cucumis melo var. saccharinus*) softening caused by transport vibration by using hot water and shellac coating. *Postharvest Biology and Technology* 110: 214-223.
- Zulkarami, B., Ashrafuzzaman, M., Husni, M.O. and Ismail, M.R. 2011. Effect of pyroligneous acid on growth, yield and quality improvement of rockmelon in soilless culture. *Australian Journal of Crop Science* 5: 1508-1514.