

UNIVERSITI PUTRA MALAYSIA

IDENTIFICATION OF SEQUENCE VARIANTS IN KEY VITAMIN E GENES FROM Elaeis guineensis Jacq. GERMPLASM FOR DEVELOPMENT OF DNA-BASED MARKERS

BABURA SULAIMAN RUFA'I

FP 2018 75



IDENTIFICATION OF SEQUENCE VARIANTS IN KEY VITAMIN E GENES FROM Elaeis guineensis Jacq. GERMPLASM FOR DEVELOPMENT OF DNA-BASED MARKERS

By

BABURA SULAIMAN RUFA'I

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy



COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



This thesis is dedicated to my loves ones, especially to my parents, wife, children (Aisha and Asma'u) and all my family members who have given me enormous support and prayers since the beginning of my studies.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

IDENTIFICATION OF SEQUENCE VARIANTS IN KEY VITAMIN E GENES FROM *Elaeis guineensis* Jacq. GERMPLASM FOR DEVELOPMENT OF DNA-BASED MARKERS

By

BABURA SULAIMAN RUFA'I

February 2018

Chairman: Professor Datin Siti Nor Akmar Abdullah

Faculty: Agriculture

Vitamin E possesses important nutritional attributes that play various roles in human disease protection. The most well-known function of this noble compound is that of chain breaking antioxidant activity that scavenge free radical ions and reduce lipid peroxidation in membrane systems. Homogentisate geranylgeranyl transferase (HGGT) and homogentisate phytyl transferase (HPT) that catalyse the first committed step of tocotrienol and tocopherol biosynthesis, respectively are important in determining plant vitamin E composition. In *Elaeis guineensis*, there is high variability in the level of vitamin E among the germplasm materials from Angolan and Tanzanian origins. Therefore, the first objective of this study was to determine important sequence variants in these key vitamin E genes from E. guineensis germplasm materials that can be used for the development of DNA-based markers. The second objective was to analyse the effects of the sequence variants on vitamin E content and composition by overexpression of the HGGT gene and its mutant derivatives in Arabidopsis thaliana. Sequence analysis reveals no important variants in HPT gene that could be associated with low and high vitamin E content. However, the analysis reveals four SNPs at positions 193, 2225, 2429 and 6932 in the coding region of the *HGGT* gene that are associated with the vitamin E content. SNPs at 193 and 2429 positions lead to non-conservative amino acid changes in the sequence from Proline (CCT) in low vitamin E to Serine (TCT) in high vitamin E and from Methionine (ATG) in low to Isoleucine (ATA) in high vitamin E palms, respectively. SNP markers 193F/413R and 2225F/2429R were developed at these SNP locations for selection of high and low vitamin E germplasm materials in E. guineensis. Fourty one germplasm materials with different vitamin E level were screened to validate these two functional SNP markers using designed PCR-based mismatch primers. The results showed 100% success of the SNP-based markers in differentiating low and high vitamin E accessions. Furthermore, single nucleotide mutagenesis was successfully carried out to generate three cDNA sequence variants (193SNPHGGT, 2429SNPHGGT and HighSNPHGGT) with one or both SNP variants incorporated into the sequence of the commercial D×P genotype (LowSNPHGGT). The variant HGGT cDNA sequences together with the unmodified cDNA were successfully transformed into Arabidopsis thaliana. The relative expression levels of HGGT in T_3 homozygous lines having the four different constructs separately showed significant ($P \le 0.005$) up-regulated expression compared with untransformed wild type Arabidopsis. However, there was no significant difference observed in the expression among transgenic Arabidopsis plants harbouring the different HGGT constructs. This demonstrated that the different variants of the E. $guineensis\ HGGT$ gene was expressed at about the same levels in the transgenic Arabidopsis. HPLC analysis indicates significant increase ($p \le 0.05$) in the total tocotrienol content between wild type and all the four transgenic lines (1.50 - 1.82-fold increase). Similarly, significant difference ($p \le 0.05$) in total tocotrienol was also recorded within the transgenic lines specifically between the two lines that harboured the two SNPs changes (HighSNPHGGT) and the one harboring the unmodified gene (LowSNPHGGT), which showed 1.22-fold increase. According to these results the two SNP variants introduced into the HGGT sequence of low vitamin E commercial variety affect the tocotrienol content and composition when analysed by functional characterization in $Arabidopsis\ thaliana$.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENGENALAN VARIAN JUJUKAN DALAM GEN UTAMA VITAMIN E DARIPADA BAHAN GERMPLASMA *Elaeis guineensis* Jacq. UNTUK PEMBANGUNAN PENANDA BERASASKAN DNA

Oleh

BABURA SULAIMAN RUFA'I

Februari 2018

Pengerusi: Profesor Datin Siti Nor Akmar Abdullah, PhD

Fakulti: Pertanian

Vitamin E mempunyai ciri nutrisi penting yang memainkan pelbagai peranan dalam ketahanan penyakit manusia. Fungsi yang paling dikenali bagi sebatian adi ini adalah aktiviti antioksidan pemutus rantai yang mengaut ion radikal bebas dan mengurangkan pemperoksidaan lipid dalam sistem membran. Homogentisat geranilgeranil transferase dan homogentisat fitil transferase yang memangkin langkah berkomitmen pertama daripada biosintesis tokotrienol dan tokoferol adalah penting dalam menentukan komposisi vitamin E tumbuhan. Dalam Elaeis guineensis, terdapat variabiliti yang tinggi dalam paras vitamin E di antara bahan germplasma yang berasal dari Angola dan Tanzania. Dengan itu, objektif pertama kajian adalah untuk menentukan yarian jujukan penting dalam gen vitamin E utama dari bahan germplasma E. guineensis yang boleh digunakan untuk pembangunan penanda berasasakan DNA. Objektif kedua adalah untuk menganalisis kesan varian jujukan keatas kandungan dan komposisi vitamin E melalui pengekspresan melampau gen HGGT dan mutan yang terhasil darinya dalam Arabidopsis thaliana. Analysis jujukan mendedahkan tiada varian dengan fungsi yang penting dalam gen HPT yang boleh dikaitkan dengan kandungan vitamin E yang rendah dan tinggi. Walau bagaimanapun, analisis mendedahkan empat SNPs pada kedudukan 193, 2225, 2429 dan 6932 dalam kawasan pengekodan gen *HGGT* yang dikaitkan dengan kandungan vitamin E. SNPs pada kedudukan 193 dan 2429, masing-masing membawa kepada perubahan non-konservatif asid amino dalam jujukan dari Prolin (CCT) dalam vitamin E yang rendah kepada Serine (TCT) dalam vitamin E yang tinggi dan dari Methionein (ATG) dari yang rendah kepada Isoleusin (ATA) dalam palma yang bervitamin E tinggi. Penanda SNPs 193F/413R dan 2225F/2429R telah dibangunkan pada lokasi SNPs untuk pemilihan bahan germplasma vitamin E yang tinggi dan rendah dalam E. guineensis. Empat puluh satu bahan germplasma dengan paras vitamin E yang berbeza telah disaring untuk mengesahkan dua penanda SNP fungsional menggunakan primer berasasakan PCR yang tidak sepadan. Keputusan menunjukkan 100% kejayaan penanda SNP tersebut dalam membezakan aksesi vitamin E yang rendah dan tinggi. Tambahan pula, kaedah mutagenesis nukleotida tunggal telah berjaya dilakukan untuk menjana tiga varian jujukan cDNA (193SNPHGGT, 2429SNPHGGT dan HighSNPHGGT) dengan satu atau dua varian SNP dimasukkan ke dalam jujukan cDNA genotip D×P komersial (LowSNPHGGT). Varian jujukan cDNA bersama dengan cDNA yang tidak diubah suai telah bejaya ditransformkan kedalam Arabidopsis thaliana. Paras ekspresi relatif HGGT dalam titisan homozigot T₃ yang mempunyai empat konstruk berbeza yang berasingan menunjukkan ekspresi menaik yang ketara (P≤0.05) berbanding Arabidopsis jenis liar yang tidak diubahsuai. Walau bagaimanapun, tiada perbezaan ketara yang diperhatikan dalam ekspresi antara tumbuhan Arabidopsis transgenik yang memiliki konstruk HGGT yang berbeza. Ini menunjukkan varian gen HGGT E. guineensis yang berbeza telah diekspres pada paras yang sama dalam Arabidopsis transgenik. Analisis HPLC menunjukkan peningkatan yang ketara (P≤0.05) dalam kandungan tokotrienol keseluruhan antara ienis liar dan empat titisan transgenik (1.5-1.82 kali ganda meningkat). Begitu juga, perbezaan ketara (P≤0.05) dalam keseluruhan tokotrienol juga direkodkan diantara titisan transgenik terutama di antara dua titisan yang mempunyai dua perubahan SNPs (HighSNPHGGT) dan yang mempunyai gen tidak diubahsuai (LowSNPHGGT), yang mana menunjukkan peningkatan 1.22 kali ganda. Berdasarkan penemuan ini, kedua varian SNPs yang dimasukkan kedalam HGGT varieti komersial dengan vitamin E rendah memberi kesan terhadap kandungan dan komposisi vitamin E apabila dianalisis melalui pencirian kefungsian dalam Arabidopsis thaliana.

ACKNOWLEDGEMENTS

Alhamdulillah, I would like to express my utmost appreciation to my supervisor, Professor Datin Dr. Siti Nor Akmar Abdullah, for her great support, excellence guidance, constructive comments and motivation not only in making the completion of this thesis a reality, but for also making me a better researcher and scholar.

I would like to thank my employee, Bayero University, Kano Nigeria for nominating me to receive scholarship award from Tertiary Education Trust Fund (Tetfund).

I would also like to express my deepest thanks to my beloved wife, for her endless kindness, support and love, infinite patience and understanding, and my dearest children Aisha (Yaya Mama) and Asma'u (Husna).

I will like to express my appreciation to my lab mates, both staff and students, Mrs Rozila Abdulrahman, Miss Siti Radziah, Mr Mehdi, Usama bin Zaid, Mei Yee, Azzreena, Hannan, Shafika, Nazri, Rezique, Hazwan, Enkgo, Abbas and Isyaku who made my life easy in the laboratory.

Finally, very special thanks to my father, mother, Brothers and Sisters for their endless prayer, love and support, and sincere thanks to all of my family members and friends.

I certify that a Thesis Examination Committee has met on 6 February 2018 to conduct the final examination of Babura Sulaiman Rufa'i on his thesis entitled "Identification of Sequence Variants in Key Vitamin E Genes from *Elaeis guineensis* Jacq. Germplasm for Development of DNA-Based Markers" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Wong Mui Yun, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Ho Chai Ling, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

Mohd Rafii bin Yusop, PhD

Professor Institute of Tropical Agriculture and Food Security Universiti Putra Malaysia (Internal Examiner)

Zoe A Wilson, PhD

Professor University of Nottingham United Kingdom (External Examiner)

RUSLI HAJI ABDULLAH, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 30 July 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of requirement for degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Siti Nor Akmar Abdullah, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Halimi B Muhammad Saud, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

Noor Azmi Shaharuddin, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:	_

Name and Matric No.: Babura Sulaiman Rufa'i, (GS41318)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision,
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	
Name of Chairman of	
Supervisory	
Committee:	Professor Dr. Siti Nor Akmar Abdullah
Signature:	
Name of Member of	
Supervisory	
Committee:	Associate Professor Dr. Halimi B Muhammad Saud
Signature:	
Name of Member of	
Supervisory	
Committee:	Associate Professor Dr. Noor Azmi Shaharuddin

TABLE OF CONTENTS

AB AC AF DF LI LI	PPROV ECLAI ST OF ST OF ST OF	K WLEDGEMENTS		Page i iii v vi viii xiii xiv xxiv
CF	IAPTI	CR		
1		RODUCTION		1
2	LITI	CRATURE REVIEW		3
	2.1	Vitamin E		3
	2.2	Tocochromanols Biosyntl	nesis Pathways	5
	2.3		eranyl Transferase (HGGT) Gene	5
	2.4		r Tocotrienols Production and	6
		Enhancement		
	2.5	Molecular Markers for Ge	enotyping	9
			Sequencing (TGS)	10
			le Polymorphism (SNPs)	11
	2.6	Oil Palm Revolution in M		12
			Elaeis Guineensis	13
	2.7	Arabidops <mark>is thaliana</mark> as N	Model Plant	14
3	VITA GER DEV	AMIN E <mark>GENES I</mark> MPLASM MAT <mark>ERIAL</mark>	N ELAEIS GUINEENSIS JA	
	3.1	Introduction		16
	3.1	Materials and Methods		16
	3.2		on and Preparation	16
		3.2.2 Extraction of Ge		17
			tion and Purification	18
		_	or Long Range Amplification	18
		9	equence and Primers Attachment	19
		3.2.6 Gradient PCR Re		19
		3.2.7 Gel Purification		
			or Amplicon Confirmation	19 20
		3.2.8 Nested Primers f3.2.9 Sequencing	or Ampricon Commination	20
		1 0	3.0	
		1 .	r Design for PCR-Based Markers	21 21
			t Mismatch Primer	21
			Ps Markers and Haplotype Analysis	21
			or Homozygosity Identification	22
	3.3	RESULTS	7 Homozygosity identification	22
	٠.٠	TUDULID		44

	3.4	3.3.1 3.3.2 3.3.3 3.3.4 3.3.5 3.3.6 3.3.7 3.3.8 3.3.9 Discuss	Isolation and Amplification of Target Genes Amplified Fragments Confirmation Sequence Analysis Identification of Sequence Variation DNA-based Marker Development and Validation Selection of Best Mismatch Primers for Screening Validation of SNPs Markers Sequencing and Haplotype Identification Analysis of HGGT Amino Acid sion	22 26 27 27 31 33 34 38 43 45
4	TH	E PRODU	DIRECTED MUTAGENESIS OF <i>HGGT</i> cDNA FOR UCTION OF DIFFERENT EXPRESSION VECTOR TS BY PCR OVERLAP EXTENSION AND ITS	47
			AL ANALYSIS IN TRANSGENIC ARABIDOPSIS	
		ALIANA		
	4.1	Introdu	ction	47
	4.2		als and Methods	48
	7.2	4.2.1	Plant Materials	48
		4.2.2	Total RNA Extraction	48
		4.2.3	Reverse Transcriptase (RT-PCR) for cDNA Synthesis	48
		4.2.4	Isolation of <i>Homogentisate geranylgeranyl transferase</i>	49
		1.2.1	cDNA	17
		4.2.5	Site-directed Mutagenesis Primer Design	49
		1.2.3	4.2.5.1 Primary (1st) PCR	49
			4.2.5.2 Secondary (2nd) PCR and Purification	50
			4.2.5.3 Confirmation of Mutant Nucleotide	50
			Substitution	50
		4.2.6	Gateway Cloning	50
		7.2.0	4.2.6.1 BP Reaction to Create Entry Clone	51
			4.2.6.2 Transformation of BP Recombinant Plasmid	51
			4.2.6.3 BP Recombinant Plasmid Purification	52
			4.2.6.4 LR Reaction to Create Expression Clone	52
			4.2.6.5 Colony PCR	53
		4.2.7	Seed Sterilization of <i>Arabidopsis thaliana</i>	54
		4.2.8	Planting <i>Arabidopsis thaliana</i> Seeds onto Soil Mixture	54
		4.2.9	Agrobacterium Transformation Using Freeze-Thaw	54
		7.2.7	Method	54
		4.2.10	Agrobacterium-Mediated Transformation of Arabidopsis Using Floral Dip Method	55
		4.2.11	Screening for Putative Transformants using Basta	56
		4.2.12	Selection of T ₂ and T ₃ Homozygous Transformant	57
		4.2.13	DNA extraction and PCR Analysis for Putative	57
		1.2.13	Transformants Confirmation	5,
		4.2.14	Analysis of Transgenic <i>Arabidopsis thaliana</i> using Real- Time qPCR	57
		4.2.15	Tocotrienol Extraction and HPLC Analysis	58
		4.2.16	Statistical Analysis	59
	4.3	RESUL	· · · · · · · · · · · · · · · · · · ·	59
	1.5	4.3.1	Isolation and Sequence analysis of EgHGGT cDNA	59
		4.3.2	Site-directed Mutagenesis	61
				01

		4.3.3	Gateway Cloning	66
		4.3.4	Analysis of Transgenic <i>Arabidops</i> is Plants in T ₁	70
			generation	
		4.3.5	PCR Verification of T ₁ Transgenic Arabidopsis Plants	72
		4.3.6	Screening of T ₃ Transgenic Plants	73
		4.3.7	Real-time PCR Optimization	73
		4.3.8	Expression Profile of different <i>HGGTs</i> in Transgenic <i>A. thaliana</i>	74
		4.3.9	Quantitative HPLC Analysis of Tocotrienols in the	77
			Transgenic Arabidopsis thaliana	
	4.4	Discussi	on	78
5	SUM	IMARY, (CONCLUSION AND RECOMMENDATIONS FOR	81
	FUT	URE RES	SEARCH	
	5.1	Summar	у	81
	5.2	Conclusi	ion	82
	5.3	Future P	rospective	83
RE	FERI	ENCES		84
AP	PENI	DICES		95
		A OF ST	UDENT	111
		PUBLIC		112

LIST OF TABLES

Гable		Page
2.1	Biological activities of tocotrienols relative to human cellular health (Benjamin, V.T, 2007).	4
2.2	Contents of tocotrienols obtained from various sources. Each contents is expressed as mg/100g.	5
3.1	Elaeis guineensis accessions HPLC data and their vitamin E composition (ppm) collected from MPOB.	17
3.2	HGGT SNPs in the codon associated with vitamin E content.	29
3.3	Purity and Concentration of Genomic DNA from <i>Elaeis</i> guineensis Germplasm Materials with Variable Vitamin E Content.	32
3.4	Mismatch primers sequences for screening the oil palm germplasm materials. Highlighted in yellow are the SNP locations and highlighted in red are the mismatch nucleotide introduced.	33
3.5	Haplotype nucleotide at the four SNPs used as the source of identification in the 41 accessions.	37
4.1	List of primers used in site-directed mutagenesis.	49
4.2	Primers for Gateway cloning.	50
4.3	List of primers specific for oil palm <i>HGGT</i> and <i>Arabidopsis Actin8</i> , <i>Tubulin4</i> for the quantitative PCR assays.	58
4.4	HPLC analyses of tocotrienol isomers in the leaf of T ₃ homozygous <i>Arabidopsis</i> lines harbouring <i>EgHGGT</i> and its mutant derivatives compared to wild type plants.	77

LIST OF FIGURES

Figure		Page
2.1	Plant tocotrienols biosynthetic pathway. The condensation of geranylgeranyldiphosphate (GGDP) and homogentisic acid catalysed by HGGT produces 2-methyl-6-geranylgeranylbenzoquinol. This serves as an intermediate that undergoes a series of methylation, cyclization and methylation reactions to form α - and γ - tocotrienols. δ - Tocotrienols are produced when the two methylation steps are not incorporated. β - Tocotrienols are produced when only the first methylation is bypassed. Enzymes written in italics (Cahoon et al., 2003; Yang et al., 2011).	7
2.2	Chemical structures of four forms of tocotrienols (Sylvester and Theriault., 2003).	8
2.3	Schematic view of a genome-wide association study (GWAS) and whole-genome prediction using genotype and phenotype data in diverse crop varieties (Huang and Han, 2014).	10
3.1	Schematic representation of Amplified fragment showing the location of primers and barcode sequences attachment.	19
3.2	Genomic DNA extracted from six accessions with different vitamin E contents from <i>Elaeis guineensis</i> germplasm materials. Lanes L: GeneRuler DNA ladder 1kb (Thermo Scientific); Lanes 1, 2, 3, 4, 5 and 6 represent DNA from Angola (high vit. E/0.311/35), Angola (high vit. E/0.311/166), Angola (low vit. E/0.311/271), Tanzania (high vit. E/0.256/96), Tanzania (low vit. E/0.256/20) and Commercial Variety (D×P), respectively	22
3.3	Schematic representation of isolation and amplification of 8.8 kb <i>HGGT</i> gene including some part of promoter sequence using two pair of primers that produced overlapping regions in the middle of the gene. The first primer set amplified 4.28 kb including the promoter and the second primer set amplified 4.53 kb including the 3'-UTR.	23
3.4	Schematic representation of isolation and amplification of 12.8 kb <i>HPT</i> gene including some part of promoter sequence using three pairs of primers producing three overlapping PCR products. The first primer pair amplified 3.48 kb (including the promoter), while the second amplified 5.34 kb in the center of the gene and the third produced 4.05 kb (including the 3'-UTR).	23
3.5	Agarose gel electrophoresis analysis of PCR products amplified from <i>HGGT</i> gene of six different accessions of oil palm from <i>Elaeis guineensis</i> germplasm materials. Two pairs of primers were used that produced overlapping region in the	24

middle of the gene. The accessions used are representative of the materials having high or low vitamin E content from the Angolan and Tanzanian populations as indicated (A) First amplified fragment of *HGGT* including promoter of 1164 bp (4200bp). (B) Second amplified fragment of *HGGT* including 3'-UTR (4500bp). Lane: L, Gene Ruler DNA ladder 1kb (Thermo Scientific); Lanes 1, 2, 3, 4, 5 and 6 represents amplified PCR product from Angola (high vit. E/0.311/35), Angola (high vit. E/0.311/166), Angola (low vit. E/0.311/271), Tanzania (high vit. E/0.256/96), Tanzania (low vit. E/0.256/20) and commercial variety (Low).

- 3.6 Agarose gel electrophoresis analysis of PCR products amplified from HPT gene of six different accessions of oil palm from Elaeis guineensis germplasm materials. Three pairs of primers were used that produced overlapping region in the middle of the gene. The accessions used are representative of the materials having high or low vitamin E content from the Angolan and Tanzanian populations as indicated (A) First amplified fragment of HPT including the promoter of 934 bp (3000bp). (B) Second amplified fragment of *HPT* in the middle of the gene that overlapped with the first and third fragment (5300bp) and (C) Third fragment of HPT including 3'-UTR (4000bp) Lane: L, Gene Ruler DNA ladder 1kb (Thermo Scientific); Lanes 1, 2, 3, 4, 5 and 6 represents amplified PCR product from Angola (high vit. E/0.311/35), Angola (high vit. E/0.311/166), Angola (low vit. E/0.311/271), Tanzania (high vit. E/0.256/96), Tanzania (low vit. E/0.256/20)
- Confirmation of specific amplification of oil palm *HGGT* and *HPT* genes using nested PCR primers. The different PCR products (2 from HGGT and 3 from HPPT) were reamplified using gene-specific nested primers for gene identity confirmation prior to sequencing. The nested PCR amplicons were analysed by gel electrophoresis. The nested PCR products from one accession (high vit. E/0.311/35) is shown to represent the result. Gel A, B, C, D and E represents the nested PCR products of upstream HGGT amplicon, downstream HGGT amplicon, upstream HPT amplicon, middle HPT amplicon and downstream HPT amplicon, respectively. Gene Ruler DNA ladder 1kb (Thermo Scientific).

26

commercial variety (Low).

- 3.8 Schematic image of the oil palm *HGGT* gene showing the position of two fragments and their overlap for consensus sequence development.
- 3.9 Schematic image of the oil palm *HPT* gene showing the position of three fragments and their overlap for consensus sequence development.
- 3.10 Schematic image of *HGGT* genes showing the positions of the SNPs/InDels associated with vitamin E content in the coding

region. Yellow horizontal arrows represent exons location. The nucleotide sequences of HGGT AH1, HGGT AH2, HGGT TH, HGGT TL, HGGT AL and HGGT DP are from Angola (high vit. E/0.311/35), Angola (high vit. E/0.311/166), Tanzania (high vit. E/0.256/96), Tanzania (low vit. E/0.256/20), Angola (low vit. E/0.311/271) and commercial variety (Low) accessions respectively.

28

29

30

31

- 3.11 Schematic image showing the position of the SNP associated with vitamin E content in the promoters of six different accessions of *HGGT*. Arrow show the position of the SNP. The nucleotide sequences of HGGT AH1, HGGT AH2, HGGT TH, HGGT TL, HGGT AL and HGGT DP are from Angola (high vit. E/0.311/35), Angola (high vit. E/0.311/166), Tanzania (high vit. E/0.256/96), Tanzania (low vit. E/0.256/20), Angola (low vit. E/0.311/271) and commercial variety (Low) accessions respectively.
- Phylogenetic relationships between the amplified *HGGT* genes from six different accessions of *Elaeis guineensis*. The tree was generated using maximum likelihood algorithm with 1000 replicate bootstrap of Geneious software. The bar indicates the scale for branch length associated with sequence differences. The nucleotide sequences of HGGT AH1, HGGT AH2, HGGT TH, HGGT TL, HGGT AL and HGGT DP are from Angola (high vit. E/0.311/35), Angola (high vit. E/0.311/166), Tanzania (high vit. E/0.256/96), Tanzania (low vit. E/0.256/20), Angola (low vit. E/0.311/271) and commercial variety (Low) accessions respectively.
- 3.13 Distribution of SNPs/InDels in oil palm HGGT gene. (A) Number of SNPs/InDels throughout the gene. (B) SNPs/InDels likely associated with vitamin E content.
- 3.14 Schematic image of *HPT* genes showing the position of SNPs/ InDels associated with vitamin E content in the whole gene. Yellow horizontal arrow represent exon locations, Black arrows shows the SNPs/InDels location. The nucleotide sequences of HPT AH1, HPT AH2, HPT TH, HPT AL, HPT TL and HPT DP are from Angola (high vit. E/0.311/35), Angola (high vit. E/0.311/166), Tanzania (high vit. E/0.256/96), Angola (low vit. E/0.311/271), Tanzania (low vit. E/0.256/20) and commercial variety (Low) accessions respectively.
- 3.15 Schematic image showing the 100% identical sequences of *HPT* promoters from six different oil palm accessions of *E. guineensis* germplasm materials. The nucleotide sequences of HPT AH1, HPT AH2, HPT AL, HPT DP, HPT TL and HPT TH are from Angola (high vit. E/0.311/35), Angola (high vit. E/0.311/166), Angola (low vit. E/0.311/271), commercial variety (Low), Tanzania (low vit. E/0.256/20), Tanzania (high

vit. E/0.256/96), accessions respectively.

3.16 Agarose gel electrophoresis analysis of PCR products of mismatch primers tested on representative *Elaeis guineensis* accessions with high and low vitamin E content. (A) Primer set HG193F/HG413R used in PCR using template DNA from: Tanzania (0.256/43) of low vitamin E accession (Lanes 1-5), Angola (0.311/86) of high vitamin E accession (Lanes 6-10) (B) Primer set HG2225F/HG2429R used in PCR reaction, Angola (0.311/86) of high vitamin E accession (Lanes 1-5), Tanzania (0.256/43) of low vitamin E accession (Lanes 6-10). Lane: L, Gene Ruler DNA ladder 1kb (ThermoScientific)

34

35

36

- 3.17 Agarose gel electrophoresis analysis of PCR products of mismatch primer 193F/413R tested on fourty one *Elaeis guineensis* palms with low (below 1000ppm) and high (above 1000 ppm) vitamin E content. (A) Lanes 1-8 and 9-14 represent Angola accessions with low and high vitamin E content, respectively. (B) Lanes 15-22 and 23-27 represent Angola accessions with low and high vitamin E content, respectively. (C) Lanes 1-10 and 11-14 represent Tanzania accessions with low and high vitamin E content, respectively.
- 3.18 Agarose gel electrophoresis analysis of PCR products of mismatch primer 2225F/2429R tested on fourty one *Elaeis guineensis* with low (below 1000 ppm) and high (above 1000 ppm) vitamin E content. (A) Lanes 1-5 and 6-13 Angola accessions with high and low vitamin E content, respectively. (B) Lanes 14-19 and 20-27 Angola accessions with high vitamin E and low vitamin E content, respectively. (C) Lanes 1-4 and 5-14 Tanzania accessions with high and low vitamin E content, respectively.
- 3.19 Sequence alignment of six (300 bp) fragments of *E. guineensis* individuals showing the SNPs at 193 and 413 positions associated with vitamin E content. Accessions used were Tanzania high (0.256/96), Angola high (0.311/166), Angola high (0.311/35), Angola low (0.311/271), Tanzania low (0.256/20) and Commercial D×P low. Asterisk (*) under the aligned sequences indicate the conserved region. Highlighted in yellow are the two SNPs.
- 3.20 Sequence alignment of six (300 bp) fragments of *E. guineensis* individuals showing the SNP location at 2225 and 2429 positions associated with vitamin E content. Accessions used are Tanzania high (0.256/96), Angola high (0.311/166), Angola high (0.311/35), Angola low (0.311/271), Tanzania low (0.256/20) and Commercial D x P low. Asterisk (*) under shows the conserved region, highlighted in Yellow are the SNPs locations.
- 3.21 Sequence chromatogram results of selected fragment of *E.* 41 *guineensis* individual accessions. Arrows showing the SNP

	location at 193 bp position associated with high and low vitamin E content.	
3.22	Sequence chromatogram results of selected fragment of <i>E. guineensis</i> individual accession. Arrows showing the SNP location at 2429 bp position associated with high and low vitamin E content.	42
3.23	Schematic image of SignalP v4.1 program analysis showing the first 35 amino acid in <i>EgHGGT</i> served as plastid transit peptide with molecular weight of 3943.63 Da. Black arrows shows the SNP location that change the amino acid from Proline in low to Serine in high vitamin E accessions	43
3.24	Multiple sequence alignment of the 462 HGGT deduced amino acid sequences from six <i>Elaeis guineensis</i> accessions. Highlighted in red is the position where the SNP lead to the change in amino acid from Proline in low and to Serine in high vitamin E accessions at 193 position and from Methionine in low and to Isoleucine in high vitamin E accessions at 2429 position. Highlighted in yellow is where the SNP does not change the amino acid as it remains Tyrosine at 2225 and 6932 positions.	45
4.1	Schematic diagram of <i>pDONR/zeo</i> and its vector elements (Invitrogen, CA, USA)	51
4.2	Vector map of pB7WG2D.1 destination vector. This is a Gateway-compatible plant expression vector for functional analysis of genes in planta. Genes of interest are driven by constitutive dual T35S promoter (Karimi and Depicker, 2002).	53
4.3	Transformation of pB7WG2D,1 Destination Vector Plasmid Harboring different HGGT cDNA. A-E represents 193SNPHGGT, 2429SNPHGGT, HighSNPHGGT, LowSNPHGGT, and empty vector plasmids respectively. Bar = 1 cm	55
4.4	Agrobacterium-Mediated Transformation of Arabidopsis Using Floral Dip Method. A, Arabidopsis thaliana seedlings (10-15 cm) ready for transformation. B, C, D and E show dipping of the seedlings into Agrobacterium sucrose suspensions for 5-10 seconds with gentle agitation. F, show pot covered with transparent plastic sleeves with openings of the wrappers clipped to maintain the humidity for at least 24 hours. Bar = 1 cm	56
4.5	Integrity analysis of total RNA extracted from oil palm mesocarp tissues at 19 waa by agarose gel electrophoresis. (A) Total RNA samples before DNAse I treatment, with visible genomic DNA (Lanes 1, 2, 3, 4) and (B) after DNAse I	60

treatment and RNA Cleanup (Lanes 1, 2, 3, 4) without genomic DNA. The 28S and 18S rRNA bands indicate intact RNA.

- 4.6 Isolation of *Elaeis guineensis HGGT* cDNA from oil palm mesocarp tissues at 19 waa by PCR. Lanes 1 and 2: the amplified PCR products of the expected size. L: DNA ladder (ThermoScientific).
- 60
- Phylogenetic tree showing relationship between EgHGGT 4.7 (DxP)) cDNA and other *HGGT* cDNA. The tree was generated using maximum likelihood algorithm with 1000 replicate bootstrap of Geneious software. The bar indicates the scale for branch length. The nucleotide sequences homologues are as follows: Elaeis guineensis HGGT1 (GeneBank accession no. KP878511.1). EgHGGT2 (GeneBank accession KP878512.1), EgHGGT transcript x1 (GeneBank accession no. XM 010933929.2). EgHGGT transcript x2 (GeneBank accession no. XM 008778000.2), Musa acuminita HGGT variant x1 (GeneBank accession no. XM 18818924.1), MaHGGT variant x2 (GeneBank accession no. 18818925.1), Amborella tricopoda HPT (GeneBank accession no. XM 020670041.1), Phoenix dactylifera HGGT (GeneBank accession no. XM 008778000.2), Triticum aestivum HGGT (GeneBank accession no. AY 222861.1)

61

- 4.8 Primers for nucleotide replacement in *EgHGGT*. The *EgHGGT* cDNA with the complete coding sequence is shown. The mutation sites at 91 and 615 positions highlighted in red. The positions of the primers B₁ and C₁ used to introduce the mutation at position 91 and primers B₂ and C₂ used to introduce the mutations at position 615 highlighted in yellow. The outer bound primers A and D used for amplifying the entire coding region also highlighted in yellow. Arrows show the directions of amplification by the specific primers.
- 62

- 4.9 Schematic diagram for site-directed mutagenesis of the *HGGT* cDNA at position 91. The mutated site (indicated by the box) introduced in the oligonucleotide internal primers b₁ and c₁. Intermediate PCR products AB and CD that overlapped with each other were first produced by pairing one outer bound primer with one internal primer with an altered nucleotide. These PCR products were used as templates to produce the entire product with the substituted nucleotide by the outer bound primers a and d.
- 63

64

Agarose gel electrophoresis analysis of the PCR product for replacing first nucleotide at position 91 in *EgHGGT* by site-directed mutagenesis. The primary PCR produced cDNA segments AB (101 bp) and CD (1288 bp) with primers b₁ and c₁ carrying the substituted nucleotides of AT instead of GC. The products were used as template to generate AD products (A) First fragment AB of 101 bp (B) Second fragment CD of 1288 bp (C) Entire fragment AD of 1389 bp. Lane: L, Gene Ruler DNA Ladder (ThermoScientific).

4.11 A schematic diagram for site-directed mutagenesis of the *HGGT* cDNA at position 615. The mutated site (indicated by the box) introduced in the oligonucleotide internal primers b₂ and c₂. Intermediate PCR products AB and CD that overlapped with each other were first produced by pairing one outer bound primer with one internal primer with an altered nucleotide. These PCR products were used as templates to produce the entire product with the substituted nucleotide by the outer bound primers a and d.

65

66

67

- 4.12 Agarose gel electrophoresis analysis of the PCR product for replacing second nucleotide at position 615 in oil palm *HGGT* by site-directed mutagenesis. The primary PCR produced AB fragment (629 bp long) and CD fragment (760 bp) with primers b₂ and c₂ carrying the substituted nucleotides of T/A instead of C/G. The products were used as template to generate AD products (A) First Fragment AB of 629 bp (B) Second fragment CD of 760 bp (C) Entire fragment AD of 1389 bp. Lane: L, Gene Ruler DNA Ladder (ThermoScientific)
- 4.13 Cloning of mutated HGGT cDNAs for expression in A. thaliana using Gateway cloning strategies. AttB sites was introduced by PCR to flank 193SNPHGGT, 2429SNPHGGT, HighSNPHGGT and LowSNPHGGT. The purified attBflanked products cloned into pDONRTM/Zeo. Each confirmed plasmid was subsequently used in the LR recombination reaction with destination vector, pB7WG2D.1 to generate expression clones. PCR analysis was carried out using recombinant plasmids purified from the transformed E. coli to monitor the cloning process. (A) Entry clones plasmids (B) Expression clones plasmids. (C) Lanes 1, 2, 3 and 4 represent 193SNPHGGT. 2429SNPHGGT, attB-flanked products HighSNPHGGT and **LowSNPHGGT PCR** respectively. Lane L Gene Ruler DNA Ladder (ThermoScientific).
- 4.14 Bacterial colonies harbouring expression clones of mutated *HGGT* in pB7WG2D.1 destination vector. A, B, C and D represent *193SNPHGGT*, *2429SNPHGGT*, *HighSNPHGGT* and *LowSNPHGGT* constructs respectively. Bar = 1 cm
- 4.15 Nucleotide sequences of entry clone of *HGGT* mutated at 193 position produced by Gateway Cloning. Highlighted in pink is the successful mutated site for the construct. The *attB* sites for Gateway cloning, start and stop codons and gene-specific primers, are highlighted in cyan, red and yellow respectively.
- 4.16 Nucleotide sequences of entry clone of *HGGT* mutated at 2429 position produced by Gateway Cloning. Highlighted in pink is the successful mutated site for the construct. The *attB* sites for Gateway cloning, start and stop codons and gene-specific primers, are highlighted in cyan, red and yellow respectively.

4.17 Nucleotide sequences of entry clone of *HGGT* mutated at 193 69 and 2429 positions produced by Gateway Cloning. Highlighted in pink is the successful mutated site for the construct. The attB sites for Gateway cloning, start and stop codons and genespecific primers, are highlighted in cyan, red and yellow respectively. 4.18 Nucleotide sequences of entry clone of non-mutated HGGT 70 produced by Gateway Cloning. Highlighted in pink is the successful mutated site for the construct. The attB sites for Gateway cloning, start and stop codons and gene-specific primers, are highlighted in cyan, red and yellow respectively. 4.19 Vegetative Growth Phase of Healthy Arabidopsis thaliana 71 before Transformation. Bars = 5 cm 4.20 Selection of T₁ generation of transgenic *Arabidopsis* plants by 71 Basta spraying. A, B, C, D and E represent the surviving plants from HighSNPHGGT, 193SNPHGGT, 2429SNPHGGT, LowSNPHGGT construct, T₁ plants transformed with empty vector and wild type untransformed lines respectively. White arrows indicate the healthy transgenic plants. One plant represents one independent T₁ line after two weeks of Basta selection. Bar = 1 cm 4.21 Confirmation of putative transgenic Arabidopsis plants 72. harbouring normal and mutated *EgHGGT* gene. extracted from the leaves of independent T₁ transgenic lines harbouring normal and mutated oil palm HGGT gene and used in PCR analysis using Elaeis guineensis HGGT-specific primers (HGT-F₁ and HGT-R₁). Lane: M, Gene ruler DNA Scientific); ladder (Thermo -C₁: negative control (untransformed wild type Arabidopsis gDNA); -C₂: negative control (transformed with empty vector). Lanes labeled 1-8 of gel A, B, C and D show PCR products of eight independent transgenic lines harbouring *HighSNPHGGT*, 193SNPHGGT, 2429SNPHGGT and LowSNPHGGT, respectively. 4.22 Selection for T₃ homozygous lines of Arabidopsis plants 73 harbouring normal and mutated EgHGGT gene. The T_3 plants were screened using Basta and pots with 100% survival were chosen. A, B, C, D and E represents T₃ homozygous plants harbouring HighSNPHGGT, 193SNPHGGT, 2429SNPHGGT, LowSNPHGGT and empty vector construct, respectively. The picture shows two representative pots for two out of eight independent transgenic lines that were obtained for each construct. Bar = 5 cm 4.23 Total RNAs analyses from randomly selected samples of T₃ 74

homozygous lines by agarose gel electrophoresis. The samples were analysed on 1% agarose gel at 80 V for 60 minutes. Two micrograms of RNA were loaded with RNA loading dye. Lanes 1, 2, 3, 4 are samples before DNAse I treatment and Lanes 5,

6, 7, 8 after DNAse I treatment and RNA Cleanup. The 28S and 18S rRNA bands indicate intact RNA. Genomic DNA was not visible after DNAse I treatment.

75

75

- 4.24 Relative normalized expression of HighSNPHGGT in three independent T_3 homozygous lines (B1-B3). The target gene was normalized to two reference genes. The value of the control (untransformed Arabidopsis) was set at 1 and the relative normalized expression (fold increase) was then compared to that value. All results are means of three technical replicates \pm standard error.
- 4.25 Relative normalized expression of 193SNPHGGT in three independent T_3 homozygous lines (B1-B3). The target gene was normalized to two reference genes. The value of the control (untransformed Arabidopsis) was set at 1 and the relative normalized expression (fold increase) was then compared to that value. All results are means of three technical replicates \pm standard error.
- 4.26 Relative normalized expression of *2429SNPHGGT* in three independent T₃ homozygous lines (B1-B3). The target gene was normalized to two reference genes. The value of the control (untransformed *Arabidopsis*) was set at 1 and the relative normalized expression (fold increase) was then compared to that value. All results are means of three technical replicates ± standard error.
- 4.27 Relative normalized expression of normal *EgHGGT* in three independent T₃ homozygous lines (B1-B3). The target gene was normalized to two reference genes. The value of the control (untransformed *Arabidopsis*) was set at 1 and the relative normalized expression (fold increase) was then compared to that value. All results are mean of three technical replicates ± standard error.
- 4.28 Comparison of the normalized expression of EgHGGT 76 (lowSNPHGGT) and its mutant derivatives. The value of the control_(untransformed Arabidopsis) was calibrated at 1 and the relative normalized expression (fold increase) was then compared to that value. All results are the means \pm SEM of three homozygous transgenic lines of T_3 generation. For the control, the means were from three wildtype plants. Same letter alphabets show no significant difference at $p \le 0.05$ using DMRT.

LIST OF APPENDICES

Appendix		Page
1	APPENDIX A: REAGENTS AND BUFFERS	95
2	APPENDIX B: PRIMERS AND SEQUENCING DATA	97
3	APPENDIX C: REAL-TIME qPCR VALIDATION	102
	DATA	
4	APPENDIX D: HPLC ANALYSIS DATA	104
5	APPENDIX E: ANOVA TABLES	106



LIST OF ABBREVIATIONS

μl Micro-liter

A.thaliana Arabidopsis thaliana A.tumefaciens Agrobacterium tumefaciens

ANOVA Analysis of variance

BLAST Basic Local Alignment Search Tool

bp Base Pairs

CaMV Cauliflower Mosaic Virus cDNA Complementary DNA

CDS Coding Region

CTAB Hexacetyltrimethyl Ammonium Bromide

D x P
Dura x Pesifera
DEPC
Diethyl Pyrocarbonate
DLD-1
Colorectal anticancer agent
DNA
Deoxyribonucleic Acid
DNase
Deoxyribonuclease
dNTPs
Deoxynucleotides

EDTA Ethylenediaminetetraacetic acid

EgHGGT Elaeis guineensis HGGT
EtBr Ethidium Bromide
FFB Fresh fruit bunches

Gb Giga bite

GBS Genotyping by sequencing
GGDP Geranylgeranyl diphosphate
GWAS Genome-wide association study

HGA Homogentisic acid

HGGT Homogentisate geranylgeranyl transferase
HPLC High performance liquid chromatography
HPPD Hydroxyphenyl pyruvate dioxygenase

HPT Homogentisate petyltransferase

InDels Insertion Deletion

KASP Competitive allele specific PCR

kb Kilo Base-Pair Km Michaelis constant LB Luria-Bertani

LiAc Lithium Acetate
LiCl Lithium Chloride

MAS Marker-assisted selection MPOB Malaysian palm oil board

mRNA Messenger RNA
MYB Myeloblastosis gene
NaCl Sodium Chloride
NaOH Natrium Hydroxide

NCBI National Center For Biotechnology Information

ng Nanogram

NGS Next-generation sequencing

OD Optical Density

OFR Open reading frame PacBio Pacific Bioscience

PCR Polymerase Chain Reactions

PDP Phytyl diphosphate pH potential of hydrogen

PTGs Post transcriptional gene silencing

PVP Polyvinylpyrrolidone

qPCR Quantitative real-time PCR
QTLs Quantitative trait loci
RNA Ribonucleic Acid
RNase Ribonuclease
rpm Round per minute

rtITP reversibly terminated deoxyinosine triphosphate RT-PCR Reverse Transcriptase Polymerase Chain Reaction

S.O.C Super Optimal Broth
SDS Sodium Dodecyl Sulphate
SGS Second generation sequencing
SMRT Single-molecule real-time
SNP Single nucleotide polymorphism

SSRs Single nucleotide polymorphis
SSRs Simple sequence repeats

TAE Tris-acetate EDTA
TE buffer Tris-EDTA buffer

TGS Third generation sequencing

Tm Melting temperature UTR Untranslated region

UV Ultraviolent

w.a.a Week After Anthesis

WT Wild Type

193SNPHGGT *HGGT* with mutated SNP at 193 position 2429SNPHGGT *HGGT* with mutated SNP at 2429 position

HighSNPHGGT HGGT with two mutated SNPs at 193 and 2429 position

LowSNPHGGT Non Mutated HGGT



CHAPTER 1

INTRODUCTION

The term vitamin E is used to describe eight lipophilic, naturally occurring compounds that include four tocopherols designated as α -, β -, γ - δ - tocopherols and their four corresponding tocotrienols (Peh et al., 2015). Tocopherols and tocotrienols isomers are distinguished based on the number and position of the methyl groups on their chromanol ring. Tocopherols have saturated phytyl tail that differentiates them from tocotrienols. which possess unsaturated tail containing three double bonds. Tocopherols are important lipid soluble antioxidants that protect cell membrane from oxidation by reacting with lipid radicals produced in the lipid peroxidation chain reaction (Hunter and Cahoon, 2007). Like tocopherols, tocotrienols are good antioxidants that tend to guard plant cells against any biochemical stress especially those arising from the breakdown of unsaturated fatty acids is the seeds (Das et al., 2005). Compared with tocopherols, tocotrienols are sparsely studied, but the current research direction is starting to give more attention to the tocotrienols, the lesser known but more potent antioxidant in vitamin E. Tocotrienols are believed to possess greater ability than tocopherols in scavenging free radical ions and reducing peroxidation of lipids in membrane system (Shahidi et al., 2010). In addition, some studies suggested that tocotrienols have specialized role in protecting neurons from damage (neurodegradation) (Sen et al., 2006) and cholesterol reduction properties (Das et al., 2005). Oral consumption of tocotrienols protects against stroke-associated brain damage in vivo (Khanna et al., 2005). Generally, most reports on vitamin E have shown that many of the properties in tocotrienols are not present in tocopherols. Tocotrienols are commercially produced from extracts of rice and oil palm and can be purchased in different forms (Cahoon et al., 2003).

Crude red palm oil is a unique vegetable oil obtained from the fruits of oil palm tree (*Elaeis guineensis*). The only natural oil produces a mixture of different antioxidants and phytonutrients such as tocopherol/tocotreinol (vitamin E), alpha and beta-carotene (provitamin A) in high level, phytosterol complex and coenzymes. No other vegetable oil has this natural combinations of phytonutrients (Corley, 2007). Malaysian Palm Oil Board (MPOB) has most member of *Elaeis guineensis* germplasm materials in the whole world (Zaki et al., 2012). Among the *E. guineensis* germplasm materials, the Angolan and Tanzanian materials were observed to have high variability in the level of vitamin E content ranging from 300 – 1600 ppm, while the level is 500 – 1000 ppm in the commercial (D×P) variety (Wahid et al., 2005). This variation may arise from the molecular set up of the genes responsible for the production of vitamin E in the plant. Thus, it would be interesting to study functional nucleotide variants in the key vitamin E biosynthetic genes (*HGGT* and *HPT*), which catalyses the first committed step of tocotreinol and tocopherol biosynthesis, respectively.

Identification of potentially functionally important sequence variants in the form of SNPs or indels as well as functional analysis of *HGGT* responsible for tocotrienol biosynthesis

and *HPT* gene responsible for tocopherol biosynthesis in oil palm would be very valuable. It could help in identifying the sources of vitamin E content variation among the accessions, and in the production of nutritionally rich palm oil in the long term. Since such studies have not been done so far, the results obtained will serve as valuable background information for genetic improvement of the oil palm. The objectives of this study are:

- 1) To determine the sequence variations in *homogentisate geranylgeranyl* transferase (HGGT) and *homogentisate* phytyltransferase (HPT) genes that differentiates accessions producing high and low vitamin E in *Elaeis guineensis* germplasm materials.
- 2) To develop SNPs/Indels markers for high and low vitamin E germplasm materials identification based on potential functional nucleotide variants.
- 3) To produce different expression vector constructs by introducing different combination of variant nucleotides into the *HGGT* sequence from D×P variety for functional studies.
- 4) To analyse the effects of sequences variant on vitamin E composition and content by overexpression of the *HGGT* gene and its mutant derivatives in transgenic model plant (*Arabidopsis thaliana*).

REFERENCES

- Abbasi, A. R., Saur, A., Hennig, P., Tschiersch, H., Hajirezaei, M., Hofius, D., Sonnewald, U., Voll, L.M., 2009. Tocopherol deficiency in transgenic tobacco (*Nicotiana tabacum L.*) plants leads to accelerated senescence. *Plant Cell Environment*, 32: 144–57.
- Abdullah, N., Rafii Yusop M., Saleh, G., Latif M.A., 2011. Genetic variability of oil palm parental genotypes and performance of its progenies as revealed by molecular markers and quantitative traits. *Comptes Rendus Biologies*, 334: 290-299.
- Agarwal, M., Shrivastava, N., Padh, H., 2008. Advances in molecular marker techniques and their applications in plant sciences. *Plant Cell Report*. 27: 617–631.
- Aggarwal, B.B., Sundaram, C., Prasad, S., Kannappan, R., 2010. Tocotrienols, the vitamin E of the 21st century: Its potential against cancer and other chronic diseases. *Biochemical. Pharmacology*, 30: 1613-1631.
- Ahsan, H., Ahad, A., Siddiqui, W.A., 2015. A review of characterization of tocotrienols from plant oils and foods. *Journal of Chemistry and Biology*, 8: 45–59.
- Andersen, J.R., Lübberstedt, T., 2003. Functional markers in plants. *Trends in Plant Science*, 8: 554–560.
- Arabidopsis Genome Initiative, 2000. Analysis of the genome sequence of the Fowering plant *Arabidopsis thaliana*. *Nature*, 408: 796–815.
- Arif, I.A., Bakir, M.A., Khan, H.A., Al Farhan, A.H., Al Homaidan, A.A., Bahkali, A.H., Al Sadoon, M., Shobrak, M., 2010. A brief review of molecular techniques to assess plant diversity. *International. Journal of. Molecular Science*, 11: 2079–2096.
- Asensi-Fabado, M.A., Munné-Bosch, S., 2010. Vitamins in plants: occurrence, biosynthesis and antioxidant function. *Trends in Plant Science*, 15: 582–592.
- Atanassov, I.I., Etchells, J.P., Turner, S.R., 2009. A simple, flexible and efficient PCR-fusion/Gateway cloning procedure for gene fusion, site-directed mutagenesis, short sequence insertion and domain deletions and swaps. *Plants Methods*, 5:14
- Barcelos, E., Rios, S. De, A., Cunha, R.N.V., Lopes, R., Motoike, S.Y., Babiychuk, E., Skirycz, A., Kushnir, S., 2015. Oil palm natural diversity and the potential for yield improvement. *Frontiers in Plant Science*, 6: 190.
- Baud, S., Mendoza, M.S., To, A., Harscoët, E., Lepiniec, L., Dubreucq, B., 2007. WRINKLED1 specifies the regulatory action of LEAFY COTYLEDON2 towards fatty acid metabolism during seed maturation in *Arabidopsis*. *Plant Journal*, 50: 825–838.
- Berendzen, K., Searle, I., Ravenscroft, D., Koncz, C., Batschauer, A., Coupland, B., Somssich, I.E., Ulker, B., 2005. A rapid and versatile combined DNA/RNA extraction protocol and its application to the analysis of a novel DNA marker set polymorphic between *Arabidopsis thaliana* ecotypes Col-0 and *Landsberg Erecta*. *Plant Methods*, 1: 340–55.

- Berlin, K., Koren, S., Chin, C.-S., Drake, J.P., Landolin, J.M., Phillippy, A.M., 2015. Assembling large genomes with single-molecule sequencing and locality-sensitive hashing. *Nature Biotechnology*, 33: 623–630.
- Brown, S.D., Nagaraju, S., Utturkar, S., De Tissera, S., Segovia, S., Mitchell, W., Land, M.L., Dassanayake, A., Köpke, M., 2014. Comparison of single-molecule sequencing and hybrid approaches for finishing the genome of *Clostridium autoethanogenum* and analysis of CRISPR systems in industrial relevant Clostridia. *Biotechnology and Biofuels*, 7: 40.
- Bui, M., Liu, Z., 2009. Simple allele-discriminating PCR for cost-effective and rapid genotyping and mapping. *Plant Methods*, 5: 1.
- Cahoon, E.B., Dietrich, C.R., Meyer, K., Damude, H.G., Dyer, J.M., Kinney, A.J., 2006. Conjugated fatty acids accumulate to high levels in phospholipids of metabolically engineered soybean and *Arabidopsis* seeds. *Phytochemistry*, 67: 1166–1176.
- Cahoon, E.B., Hall, S.E., Ripp, K.G., Ganzke, T.S., Hitz, W.D., Coughlan, S.J., 2003. Metabolic redesign of vitamin E biosynthesis in plants for tocotrienol production and increased antioxidant content. *Nature Biotechnology*, 21: 1082–1087.
- Che, P., Zhao, Z., Glassman, K., Dolde, D., Hu, T.X., Jones, T.J., Obukosia, S., Wambugu, F., Che, P., Zhao, Z., Glassman, K., Dolde, D., Hu, T.X., Jones, T.J., Fred, D., 2016. Elevated vitamin E content improves all *trans* β-carotene accumulation and stability in biofortified sorghum. *Proceedings of the National Academy of Sciences of the United States of America*, 113: 8209–8209.
- Chen, C., Mitchell, S.E., Elshire, R.J., Buckler, E.S., El-Kassaby, Y.A., 2013. Mining conifers' mega-genome using rapid and efficient multiplexed high-throughput genotyping-by-sequencing (GBS) SNP discovery platform. *Tree Genetics and Genomes*, 9: 1537–1544.
- Chen, J., Liu, C., Shi, B., Chai, Y., Han, N., Zhu, M., Bian, H., 2017. Overexpression of *HvHGGT* Enhances Tocotrienol Levels and Antioxidant Activity in Barley. *Journal of Agricultural and Food Chemistry*, 7: 439.
- Chen, M.-H., Bergman, C.J., 2015. Vitamin E homologs and γ-oryzanol levels in rice (*Oryza sativa L.*) during seed development. *Cereal Chemistry Journal*, CCHEM-07-15-0152-R.
- Chiu, J., March, P.E., Lee, R., Tillett, D., 2004. Site-directed, Ligase-Independent Mutagenesis (SLIM): a single-tube methodology approaching 100 % efficiency in 4 h. *Nucleic Acids Research*, 32: 174-179.
- Choong, C.G., McKay, A., 2014. Sustainability in the Malaysian palm oil industry. *Journal of Clean Production*, 85: 258–264.
- Clough, S.J. and Bent, A.F., 1998. Floral dip: A simplified method for *Agrobacterium*-mediated transformation of *Arabidopsis thaliana*. *The Plant Journal*, 16: 735-743.
- Collakova, E., DellaPenna, D., 2001. Isolation and Functional Analysis of Homogentisate Phytyltransferase from *Synechocystis sp.* PCC 6803 and *Arabidopsis. Plant Physiology*, 127: 1113–1124.

- Cordoba, E., Salmi, M., León, P., 2009. Unravelling the regulatory mechanisms that modulate the MEP pathway in higher plants. *Journal of Experimental Botany*, 60: 2933–43.
- Corley, P., and Re, .H.V., 2007. The Products of the Oil Palm and Their Extraction, in: The Oil Palm. *Blackwell Science* Ltd, Oxford, UK, 445–466.
- Corrêa, T.R., Motoike, S.Y., Coser, S.M., da Silveira, G., de Resende, M.D.V., Chia, G.S., 2015. Estimation of genetic parameters for *invitro* oil palm characteristics (*Elaeis guineensis Jacq.*) and selection of genotypes for cloning capacity and oil yield. *Industrial Crops and Production*, 77: 1033–1038.
- Das, S., Powell, S.R., Wang, P., Divald, A., Nesaretnam, K., Tosaki, A., Cordis, G. A., Maulik, N., Das, D.K., 2005. Cardioprotection with palm tocotrienol: antioxidant activity of tocotrienol is linked with its ability to stabilize proteasomes. *American Journal of Physiology Heart Circulation and Physiology*, 289: 361-367.
- Dellapenna D.A., 2005. A decade of progress in understanding vitamin E synthesis in plants *Journal of Plant Physiology*, 162: 729-737
- Derrington, I.M., Butler, T.Z., Collins, M.D., Manrao, E., Pavlenok, M., Niederweis, M., Gundlach, J.H., 2010. Nanopore DNA sequencing with MspA. *Proceedings of the National Academy of Sciences of the United States of America*, 107: 16060-16065.
- Doyle, J.J and Doyle, L.J. 1990. Isolation of plant DNA from fresh tissue. *Focus*, 12: 13-15.
- Drive, B., Louis, S., 2006. Gateway-compatible vectors for plant functional genomics and proteomics. *The Plant Journal*, 45: 616–629.
- Duran, C., Edwards, D., Batley, J., 2009. Molecular Marker Discovery and Genetic Map Visualisation, in: *Bioinformatics*. Springer New York, 165–189.
- Ebrahimi, M., Nor, S., Abdullah, A., Aziz, M.A., Namasivayam, P., 2015. A novel CBF that regulates abiotic stress response and the ripening process in oil palm (*Elaeis guineensis*) fruits. *Tree Genetics & Genomes*, 11: 56
- Eid, J., Fehr, A., Gray, J., Luong, K., Lyle, J., Otto, G., Peluso, P., Rank, D., Baybayan, P., Bettman, B., Bibillo, A., Bjornson, K., Chaudhuri, B., Christians, F., Cicero, R., Clark, S., Dalal, R., deWinter, A., Dixon, J., Foquet, M., Gaertner, A., Hardenbol, P., Heiner, C., Hester, K., Holden, D., Kearns, G., Kong, X., Kuse, R., Lacroix, Y., Lin, S., Lundquist, P., Ma, C., Marks, P., Maxham, M., Murphy, D., Park, I., Pham, T., Phillips, M., Roy, J., Sebra, R., Shen, G., Sorenson, J., Tomaney, A., Travers, K., Trulson, M., Vieceli, J., Wegener, J., Wu, D., Yang, A., Zaccarin, D., Zhao, P., Zhong, F., Korlach, J., Turner, S., 2009. Real-Time DNA Sequencing from Single Polymerase Molecules. *Science*, 80: 323.
- Foster, J.T., Allan, G.J., Chan, A.P., Rabinowicz, P.D., Ravel, J., Jackson, P.J., Keim, P., 2010. Single nucleotide polymorphisms for assessing genetic diversity in castor bean (*Ricinus communis*). *BMC Plant Biology*, 10: 13.
- Ghosh, S., Mandi, S. Sen, 2015. SNP in Chalcone Synthase gene is associated with variation of 6-gingerol content in contrasting landraces of *Zingiber officinale.Roscoe*. *Gene*, 566: 184-188.

- Giraud, T., Refrégier, G., Le Gac, M., De Vienne, D.M., Hood, M.E., 2008. Speciation in fungi. *Fungal Genetic and Biology*, 45: 791-802.
- Haller, G., Alvarado, D., Mccall, K., Mitra, R.D., Dobbs, M.B., Gurnett, C.A., 2016. Massively parallel single- nucleotide mutagenesis using reversibly terminated inosine. *Nature Methods*, 13: 923–925.
- Han, M., May, Y., Ngan, A., Hock, C., Ali, M., 2004. Separation of Vitamin E (tocopherol, tocotrienol, and tocomonoenol) in Palm Oil. *Lipids*, 39: 1031–1035.
- Hassanien, M.M.M., Abdel-Razek, A.G., Rudzińska, M., Siger, A., Ratusz, K., Przybylski, R., 2014. Phytochemical contents and oxidative stability of oils from non-traditional sources. *European Journal of Lipid Science and Technology*, 116: 1563–1571.
- Hayashi, K., Hashimoto, N., Daigen, M., Ashikawa, I., 2004. Development of PCR-based SNP markers for rice blast resistance genes at the Piz locus. *Theoretical and Applied Genetics*, 108: 1212–1220.
- Hayward, A.C., Tollenaere, R., Dalton-Morgan, J., Batley, J., 2015. Molecular Marker Applications in Plants. *Methods in Molecular Biology*, 1245: 13–27.
- Heckman, K.L., Pease, L.R., 2007. Gene splicing and mutagenesis by PCR-driven overlap extension. *Nature Protocol*, 2: 924–932.
- Horvath, G., Wessjohann, L., Bigirimana, J., Jansen, M., Guisez, Y., Caubergs, R., Horemans, N., 2006. Differential distribution of tocopherols and tocotrienols in photosynthetic and non-photosynthetic tissues. *Phytochemistry*, 67: 1185–1195.
- Huang, X., Han, B., 2014. Natural Variations and Genome-Wide Association Studies in Crop Plants. *Annual Review of Plant Biology*, 65: 531–551.
- Hunter, S.C., Cahoon, E.B., 2007. Enhancing vitamin E in oilseeds: unraveling tocopherol and tocotrienol biosynthesis. *Lipids*, 42: 97–108.
- Huq, A., Akter, S., Nou, S., Kim, H.T., Jung, Y.J., Kang, K.K., 2016. Identification of functional SNPs in genes and their effects on plant phenotypes. *Journal of Plant Biotechnology*, 43: 1–11.
- Husain, K., Centeno, B.A., Chen, D.-T., Hingorani, S.R., Sebti, S.M., Malafa, M.P., 2013. Vitamin E δ-tocotrienol prolongs survival in the LSL-KrasG12D/+; LSL-Trp53R172H/+ Pdx-1-Cre (KPC) transgenic mouse model of pancreatic cancer. *Cancer Prevention Research (Philadelphia)*, 6: 1074–1083.
- Idrees, M., Irshad, M., 2015. Molecular Markers in Plants for Analysis of Genetic Diversity: A Review. *European Academic Research*, 2: 1513–1540.
- Joshi, V., Joung, J.G., Fei, Z., Jander, G., 2010. Interdependence of threonine, methionine and isoleucine metabolism in plants: Accumulation and transcriptional regulation under abiotic stress. *Amino Acids*, 39: 933–947.
- Juan, G., Liu, G., Shuangyan, C., Aly, A.A., 2009. Vitamin E Metabolic Modulation in Plants, in: Herbal Drugs: *Ethnomedicine to Modern Medicine*, Springer Berlin, Heidelberg, Germany. 333–352.

- Kannappan, R., Gupta, S.C., Kim, J.H., Aggarwal, B.B., 2012. Tocotrienols fight cancer by targeting multiple cell signaling pathways. *Genes and Nutrition*, 7: 43-57.
- Karimi, M., Inzé, D., Depicker, A., 2002. Gateway vectors for *Agrobacterium*-mediated plant transformation. *Trends in Plant Science*, 7: 193–195.
- Karunanandaa, B., Qi, Q., Hao, M., Baszis, S.R., Jensen, P.K., Wong, Y.-H.H., Jiang, J., Venkatramesh, M., Gruys, K.J., Moshiri, F., Post-Beittenmiller, D., Weiss, J.D., Valentin, H.E., 2005. Metabolically engineered oilseed crops with enhanced seed tocopherol. *Metabolic Engineering*, 7: 384–400.
- Kaur, J., Sharma, R., 2006. Directed Evolution: An Approach to Engineer Enzymes. *Critical Review in Biotechnology*, 26: 165–199.
- Khanna, S., Patel, V., Rink, C., Roy, S., Sen, C., 2005. Delivery of orally supplemented α-tocotrienol to vital organs of rats and tocopherol-transport protein deficient mice. *Radical Biology and Medicine*, 39: 1310-1319.
- Kim, Y.H., Lee, Y.Y., Kim, Y.H., Choi, M.S., Jeong, K.H., Lee, S.K., Seo, M.J., Yun, H.T., Lee, C.K., Kim, W.H., Lee, S.C., Park, S.K., Park, H.M., 2011. Antioxidant Activity and Inhibition of Lipid Peroxidation in Germinating Seeds of Transgenic Soybean Expressing OsHGGT. Journal of Agricultural and Food Chemistry, 59: 584–591.
- Kiryu, I., Nishioka, T., Yuasa, K., Kurita, J., Shimahara, Y., Ototake, M., Ikegami, N., Oseko, N., 2014. Rapid and Simple Detection Method of *Candidatus Xenohaliotis californiensis* Using Fecal PCR in Abalone *Haliotis discus discus* and *H. gigantea*. *Fish Pathology*, 49: 41–48.
- Kong, S.L., Abdullah, S.N.A., Ho, C.L., Amiruddin, M.D., 2016. Molecular cloning, gene expression profiling and in silico sequence analysis of vitamin E biosynthetic genes from the oil palm. *Plant Gene*, 5: 100–108.
- Koornneef, M., Meinke, D., Linne, C., West, R., 2010. The development of *Arabidopsis* as a model plant. *The Plant Journal*, 61: 909–921.
- Krivanek, O.L., Chisholm, M.F., Nicolosi, V., Pennycook, T.J., Corbin, G.J., Dellby, N., Murfitt, M.F., Own, C.S., Szilagyi, Z.S., Oxley, M.P., Pantelides, S.T., Pennycook, S.J., 2010. Atom-by-atom structural and chemical analysis by annular dark-field electron microscopy. *Nature*, 464: 571–574.
- Kumar, B., Abdel-ghani, A.H., Pace, J., Reyes-matamoros, J., Hochholdinger, F., Lübberstedt, T., 2014. Plant Science Association analysis of single nucleotide polymorphisms in candidate genes with root traits in maize (*Zea mays L.*) seedlings. *Plant Science*, 224: 9–19.
- Kushairi, A, Tarmizi, A. H., Zamzuri, I., R, S.K., Ooi, S.E., Palm, M., Board, O., Institusi, N.P., Bangi, B.B., 2010. Production, Performance and Advances in Oil Palm Tissue Culture. *Internatinal Seminar Advance in Oil Palm Tissue Culture*, 1–23.
- Lee, C.P., Taylor, N.L., Millar, A.H., 2013. Recent advances in the composition and heterogeneity of the *Arabidopsis* mitochondrial proteome. *Fronteirs in Plant Science*, 4: 4.

- Leonelli, S., 2007. *Arabidopsis*, the botanical *Drosophila*: from mouse cress to model organism. *Endeavour*, 31: 34–38.
- Liu, J., Huang, S., Sun, M., Liu, S., Liu, Y., Wang, W., Zhang, X., Wang, H., Hua, W., 2012. An improved allele-specific PCR primer design method for SNP marker analysis and its application. *Plant Methods*, 8: 34.
- Luan, B., Peng, H., Polonsky, S., Rossnagel, S., Stolovitzky, G., Martyna, G., 2010. Base-By-Base Ratcheting of Single Stranded DNA through a Solid-State Nanopore. *Physical Review Letters*, 104: 238103.
- Ma, Y., Coyne, C.J., Grusak, M.A., Mazourek, M., Cheng, P., Main, D., McGee, R.J., 2017. Genome-wide SNP identification, linkage map construction and QTL mapping for seed mineral concentrations and contents in pea (*Pisum sativum L.*). *BMC Plant Biology*, 17: 43.
- Maarasyid, C., Muhamad, I.I., Supriyanto, E., 2014. Potential Source and Extraction of Vitamin E From Palm Based Oils: A Review. *Journal Teknologi*, 4: 43–50.
- May, C.Y., Nesaretnam, K., 2014. Highlight Article Research advancements in palm oil nutrition. *European Journal of Lipid Science and Technology*, 116: 1301–1315.
- McCullum, E.O., Williams, B.A.R., Zhang, J., Chaput, J.C., 2010. Random mutagenesis by error-prone PCR. *Methods in Molecular Biology*, 634: 103–109.
- Meinke, D.W., 1998. *Arabidopsis thaliana*: A Model Plant for Genome Analysis. *Science*, 80: 662–682.
- Munusamy, U., Abdullah, S.N.A., Aziz, M.A., Khaza`Ai, H., 2015. Metabolic Engineering Of A-Tocotrienolthrough PGTS Mechanisms Andisoprenoid/Non-Mevalonate Pathways in perennial Crops. *Plant Cell Biotechnology and Molecular Biology*, 16: 119–129.
- Nesaretnam, K., Meganathan, P., Veerasenan, S.D., Selvaduray, K.R., 2012. Tocotrienols and breast cancer: The evidence to date, in: *Genes and Nutrition*, 7: 3–9.
- Paez-Garcia, A., Motes, C., Scheible, W.-R., Chen, R., Blancaflor, E., Monteros, M., 2015. Root Traits and Phenotyping Strategies for Plant Improvement. *Plants*, 4: 334–355.
- Pavlov, M.Y., Watts, R.E., Tan, Z., Cornish, V.W., Ehrenberg, M., Forster, A.C., 2009. Slow peptide bond formation by proline and other N-alkylamino acids in translation. *Proceedings of the National Academy of Sciences*, 106: 50–54.
- Peh, H.Y., Ho, W.E., Cheng, C., Chan, T.K., Seow, A.C.G., Lim, A.Y.H., Fong, C.W., Seng, K.Y., Ong, C.N., Wong, W.S.F., 2015. Vitamin E Isoform γ-Tocotrienol Downregulates House Dust Mite–Induced Asthma. *The Journal of Immunology*, 195: 2.
- Pfaffl, M. W., Horgan, G. W. and Dempfle, L. 2002. Relative expression software tool (REST) for group- wise comparison and statistical analysis of relative expression results in real-time PCR. *Nucleic Acids Research*, 30: 36

- Pootakham, W., Jomchai, N., Ruang-areerate, P., Shearman, J.R., Sonthirod, C., Sangsrakru, D., Tragoonrung, S., Tangphatsornruang, S., 2015. Genome-wide SNP discovery and identification of QTL associated with agronomic traits in oil palm using genotyping-by-sequencing (GBS). *Genomics*, 105: 288–295.
- Prescott, A. and Martin, C., 1987. A rapid method for the quantitative assessment of levels of specific mRNAs in plants. *Plant Molecular Biology Report*, 4: 219-224.
- Provart, N.J., Alonso, J., Assmann, S.M., Bergmann, D., Brady, S.M., Brkljacic, J., Browse, J., Chapple, C., Colot, V., Cutler, S., Dangl, J., Ehrhardt, D., Friesner, J.D., Frommer, W.B., Grotewold, E., Meyerowitz, E., Nemhauser, J., Nordborg, M., Pikaard, C., Shanklin, J., Somerville, C., Stitt, M., Torii, K.U., Waese, J., Wagner, D., Mccourt, P., 2016. 50 years of *Arabidopsis* research: Highlights and future directions. *New Phytologist*, 209: 921–944.
- Ouail, M., Smith, M.E., Coupland, P., Otto, T.D., Harris, S.R., Connor, T.R., Bertoni, A., Swerdlow, H.P., Gu, Y., Rothberg, J., Hinz, W., Rearick, T., Schultz, J., Mileski, W., Davey, M., Leamon, J., Johnson, K., Milgrew, M., Edwards, M., Eid, J., Fehr, A., Gray, J., Luong, K., Lyle, J., Otto, G., Peluso, P., Rank, D., Baybayan, P., Bettman, B., Bentley, D., Balasubramanian, S., Swerdlow, H., Smith, G., Milton, J., Brown, C., Hall, K., Evers, D., Barnes, C., Bignell, H., Kozarewa, I., Ning, Z., Quail, M., Sanders, M., Berriman, M., Turner, D., Quail, M., Otto, T., Gu, Y., Harris, S., Skelly, T., McQuillan, J., Swerdlow, H., Oyola, S., Syed, F., Grunenwald, H., Caruccio, N., Lam, H., Clark, M., Chen, R., Chen, R., Natsoulis, G., O'Huallachain, M., Dewey, F., Habegger, L., Carver, T., Harris, S., Berriman, M., Parkhill, J., McQuillan, J., Ponsting, N., Ning, Z., Otto, T., Sanders, M., Berriman, M., Newbold, C., Nakamura, K., Oshima, T., Morimoto, T., Ikeda, S., Yoshikawa, H., Shiwa, Y., Ishikawa, S., Linak, M., Hirai, A., Takahashi, H., Diep, B., Gill, S., Chang, R., Phan, T., Chen, J., Davidson, M., Lin, F., Lin, J., Carleton, H., Mongodin, E., Achidi, E., Gardner, M., Hall, N., Fung, E., White, O., Berriman, M., Hyman, R., Carlton, J., Pain, A., Nelson, K., Bowman, S., Choi, M., Scholl, U., Ji, W., Liu, T., Tikhonova, I., Zumbo, P., Nayir, A., Bakkaloglu, A., Ozen, S., Sanjad, S., Down, T., Rakyan, V., Turner, D., Flicek, P., Li, H., Kulesha, E., Graf, S., Johnson, N., Herrero, J., Tomazou, E., Giresi, P., Kim, J., McDaniell, R., Iyer, V., Lieb, J., Johnson, D., Mortazavi, A., Myers, R., Wold, B., Langridge, G., Phan, M., Turner, D., Perkins, T., Parts, L., Haase, J., Charles, I., Maskell, D., Peters, S., Dougan, G., Licatalosi, D., Mele, A., Fak, J., Ule, J., Kayikci, M., Chi, S., Clark, T., Schweitzer, A., Blume, J., Wang, X., Mamanova, L., Andrews, R., James, K., Sheridan, E., Ellis, P., Langford, C., Ost, T., Collins, J., Turner, D., Myllykangas, S., Buenrostro, J., Natsoulis, G., Bell, J., Ji, H., Shao, N., Hu, H., Yan, Z., Xu, Y., Hu, H., Menzel, C., Li, N., Chen, W., Khaitovich, P., Wang, Z., Gerstein, M., Snyder, M., Gnerre, S., Maccallum, I., Przybylski, D., Ribeiro, F., Burton, J., Walker, B., Sharpe, T., Hall, G., Shea, T., Sykes, S., Levin, J., Yassour, M., Adiconis, X., Nusbaum, C., Thompson, D., Friedman, N., Gnirke, A., Regev, A., Adey, A., Asan, Xun, X., Kitzman, J., Turner, E., Stackhouse, B., MacKenzie, A., Caruccio, N., Zhang, X., Flusberg, B., Webster, D., Lee, J., Travers, K., Olivares, E., Clark, T., Korlach, J., Turner, S., Holden, T., Lindsay, J., Corton, C., Quail, M., Cockfield, J., Pathak, S., Batra, R., Parkhill, J., Bentley, S., Edgeworth, J., Li, H., Durbin, R., Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., Durbin, R., Angiuoli, S., Salzberg, S., 2012. A tale of three next generation

- sequencing platforms: comparison of Ion torrent, pacific biosciences and illumina MiSeq sequencers. *BMC Genomics*, 13: 341.
- Qureshi, A.A., Sami, S.A., Salser, W.A., Khan, F.A., 2002. Dose-dependent suppression of serum cholesterol by tocotrienol-rich fraction (TRF25) of rice bran in hypercholesterolemic humans. *Atherosclerosis*, 161: 199–207.
- Rajanaidu, N., kushairi H., 2004. Ps8: high vitamin e breeding population. MPOB Information Series, *Science Technology*, 229.
- Rajanaidu, N.; Johari, O.; Ahmad, S. F. Veriappan, A. 2008. Proceedings of the 3rd Seminar on performance of MPOB PS1 and PS2 materials and elite ge: MPOB Head Office, Bandar Baru Bangi, Selangor. 91-116
- Ramli, A., 2015. Strengthening Agricultural Sector Superior Commodities -Based Against the Economic Growth in South Sulawesi, Indonesia. *International Journal of Advanced Research*, 3: 753–760.
- Reid, J.G., Carroll, A., Veeraraghavan, N., Dahdouli, M., Sundquist, A., English, A., Bainbridge, M., White, S., Salerno, W., Buhay, C., Yu, F., Muzny, D., Daly, R., Duyk, G., Gibbs, R.A., Boerwinkle, E., 2014. Launching genomics into the cloud: deployment of Mercury, a next generation sequence analysis pipeline. *BMC Bioinformatics*, 15: 30.
- Rhoads, A., Au, K.F., 2015. PacBio Sequencing and Its Applications. *Genomics, Proteomics and Bioinformatics*, 13, 278–289.
- Riju, A., Arunachalam, V., 2009. Interspecific differences in single nucleotide polymorphisms (SNPs) and indels in expressed sequence tag libraries of oil palm *Elaeis guineensis* and *E. oleifera*. *Nature Precedings*, 3593: 1.
- Rival Alain and Parveez Ghulam, 2014. Biotechnology of Fruit and Nut Crops Richard E Litz (Ed.), CABI Publishing, Massachusset USA, 113–136.
- Roslinda, Sajari., NH Abd Razak., Farida Yusof., S.A., 2014. Improved Efficiency of Tocotrienol Extraction from Fresh and Processed Latex. *Journal of Rubber Research*, 17: 245–260.
- Schadt, E.E., Turner, S., Kasarskis, A., 2010. A window into third-generation sequencing. *Human Molecular Genetics*, 19: 227–240.
- Sekine, D., Murata, K., Kimura, T., Nakagawa, K., Miyazawa, T., 2016. Identification of a Genetic Factor Required for High γ-Isoform Concentration in Rice Vitamin E. *Journal of Agricultural and Food Chemistry*, 64: 9368–9373.
- Semagn, K., Babu, R., Hearne, S., Olsen, M., 2014. Single nucleotide polymorphism genotyping using Kompetitive Allele Specific PCR (KASP): Overview of the technology and its application in crop improvement. *Molecular Breeding*, 33: 1–14.
- Sen, C.K., Khanna, S., Roy, S., 2006. Tocotrienols: Vitamin E beyond tocopherols. *Life Science*, 78: 2088–2098.
- Shahidi, F., De Camargo, A., 2016. Tocopherols and Tocotrienols in Common and Emerging Dietary Sources: Occurrence, Applications, and Health Benefits. *Intertional Journal of Molecular Sciences*, 17: 1745

- Shahidi, F., Zhong, Y., Song, C., Ahn, C.B., Shin, T.S., Cha, Y.J., Shahidi, F., Jeon, Y.J., 2010. Lipid oxidation and improving the oxidative stability. *Chemical Society Review*, 39: 4067.
- Shapiro, E., Biezuner, T., Linnarsson, S., 2013. Single-cell sequencing-based technologies will revolutionize whole-organism science. *Nature Review Genetics*, 14: 618–630.
- Sheppard, A.J., Pennington, J.A.T., Weihrauch, J.L., 1993. Analysis and distribution of vitamin E in vegetable oils and foods. *Vitamin E in Health and Disease*, 9: 31.
- Shi, Z., Liu, S., Noe, J., Arelli, P., Meksem, K., Li, Z., 2015. SNP identification and marker assay development for high-throughput selection of soybean cyst nematode resistance. *BMC Genomics*, 16: 314.
- Singh, R., Ong-Abdullah, M., Low, E.-T.L., Manaf, M.A.A., Rosli, R., Nookiah, R., Ooi, L.C.-L., Ooi, S.-E., Chan, K.-L., Halim, M.A., Azizi, N., Nagappan, J., Bacher, B., Lakey, N., Smith, S.W., He, D., Hogan, M., Budiman, M.A., Lee, E.K., DeSalle, R., Kudrna, D., Goicoechea, J.L., Wing, R.A., Wilson, R.K., Fulton, R.S., Ordway, J.M., Martienssen, R.A., Sambanthamurthi, R., 2013. Oil palm genome sequence reveals divergence of interfertile species in Old and New worlds. *Nature*, 500: 335–339.
- Sylvester, P.W and Theriault, A., 2003. Role of Tocotrienols in the Prevention of Cardiovascular Disease and Breast Cancer. *Current Topics in Nutraceutical Research*, 1: 121-136.
- Stacey, G.S. and M.G., 2013. Metabolic engineering of plants for increased homogentisate and tocochromanol production. *Patent US* 9624500 B2.
- Suzuki, Y., Tsuchiya, M., Wassall, S., 1993. Structural and dynamic membrane properties of. Alpha.-tocopherol and. Alpha-tocotrienol: implication to the molecular mechanism of their antioxidant potency. *Biochemistry*, 32: 10692–10699.
- Tavva, V.S., Kim, Y.H., Kagan, I.A., Dinkins, R.D., Kim, K.H., Collins, G.B., 2007. Increased α-tocopherol content in soybean seed overexpressing the *Perilla frutescens* γ-tocopherol methyltransferase gene. *Plant Cell Reports*, 26, 61–70.
- Teoh C.H., 2002. The palm oil industry in malaysia from seed to frying pan prepared for the palm oil industry in malaysia: from seed to frying pan. *Plantation Agriculture*, WWF Malaysia, 49, Jalan SS 23/15. Taman SEA, 47400 Petaling Jaya, Selangor, Malaysia
- Theriault, A., Chao, J.-T., Wang, Q., Gapor, A., Adeli, K., 1999. Tocotrienol: A review of its therapeutic potential. *Clinical Biochemistry*, 32: 309–319.
- Urban, A., 1997. A rapid and efficient method for site-directed mutagenesis using one-step overlap extension PCR. *Nucleic Acids Research*, 25: 2227–2228.
- Valentin, H.E., Qi, Q., 2005. Biotechnological production and application of vitamin E: Current state and prospects. *Applied Microbiology and Biotechnology*, 68: 436–444.
- Vangelatos, I., Vlachakis, D., Sophianopoulou, V., Diallinas, G, 2009. Modelling and mutational evidence identify the substrate binding site and functional elements in APC amino acid transporters. *Molecular Membrane Biology*, 26: 356–370.

- Verhage, L., Angenent, G.C., Immink, R.G.H., 2014. Research on floral timing by ambient temperature comes into blossom. *Trends in Plant Science*, 19: 583–591.
- Wahid, M.B., Abdullah, S.N.A., Henson, I.E., 2005. Oil Palm-Achievements and Potential. *Plant Production Science*, 8: 288–297.
- Wang, C., Husain, K., Zhang, A., Centeno, B.A., Chen, D., Tong, Z., Sebti, S.M., Malafa, M.P., 2015. ScienceDirect EGR-1 / Bax pathway plays a role in vitamin E δ tocotrienol-induced apoptosis in pancreatic cancer cells. *Journal of Nutritional Biochemistry*, 26: 797–807.
- Wang, J., Sun, N., Deng, T., Zhang, L., Zuo, K., 2014. Genome-wide cloning, identification, classification and functional analysis of cotton heat shock transcription factors in cotton (*Gossypium hirsutum*). *BMC Genomics*, 15: 961.
- Wangkumhang, P., Chaichoompu, K., Ngamphiw, C., Ruangrit, U., Chanprasert, J., Assawamakin, A., Tongsima, S., 2007. WASP: a Web-based Allele-Specific PCR assay-designing tool for detecting SNPs and mutations. *BMC Genomics*, 8: 275.
- Weigel, D., and Glazebrook, J. 2002. *Arabidopsis. Cold Spring Harbor Laboratory Press.* Cold Spring Harbor: Retrieved from http://tocs.ulb.tudarmstadt.de/108129926.pdf
- Whiteford, N., Skelly, T., Curtis, C., Ritchie, M.E., Löhr, A., Zaranek, A.W., Abnizova, I., Brown, C., 2009. Swift: Primary data analysis for the Illumina Solexa sequencing platform. *Bioinformatics*, 25: 2194–2199.
- Winifred, M.C., Thelma, S.V., Edward, H.W., Beverly, D.W., 1983. Vitamin E content of feedstuffs determined by high performance liquid chromatographic fluorescence. *Journal of Agricultural and Food Chemistry*, 31: 1330-1333.
- Woittiez, L. S., van Wijk, M. T., Slingerland, M., van Noordwijk, M., Giller, K.E., 2017. Yield gaps in oil palm: A quantitative review of contributing factors. *European Journal of Agronomy*, 83: 57–77.
- Wong, R.S.Y., Radhakrishnan, A.K., 2012. Tocotrienol research: past into present. *Nutrition Reviews*, 70: 483–90.
- Yang, W., Cahoon, R.E., Hunter, S.C., Zhang, C., Han, J., Borgschulte, T., Cahoon, E.B., 2011. Vitamin E biosynthesis: functional characterization of the monocot homogentisate geranylgeranyl transferase. The Plant Journal, 65: 206–217.
- Zaki, N.M., Singh, R., Rosli, R., Ismail, I., 2012. Elaeis oleifera Genomic-SSR Markers: Exploitation in Oil Palm Germplasm Diversity and Cross-Amplification in Arecaceae. *International Journal of Molecular Science*, 13: 4069–4088.
- Zeng, F., Zhang, Y., Zhang, Z., Malik, A.A., Lin, Y., Zeng, F., 2017. Multiple-site fragment deletion, insertion and substitution mutagenesis by modified overlap extension PCR 2818. *Biotechnology & Biotechnological Equipment*, 31: 339–348
- Zhang, C., Cahoon, R.E., Hunter, S.C., Chen, M., Han, J., Cahoon, E.B., 2013. Genetic and biochemical basis for alternative routes of tocotrienol biosynthesis for enhanced vitamin e antioxidant production. *The Plant Journal*, 73: 628–639.

Zhang, G.Y., Liu, R.-R., Xu, G., Zhang, P., Li, Y., Tang, K. X., Liang, G. H., Liu, Q. Q., 2013. Increased α-tocotrienol content in seeds of transgenic rice overexpressing *Arabidopsis* γ-tocopherol methyltransferase. *Transgenic Research*, 22, 89–99.

