



UNIVERSITI PUTRA MALAYSIA

***FORMULATION AND LABORATORY EFFICACY OF ISARIA
FUMOSOROSEA AGAINST BAGWORMS (METISA PLANA WALKER
AND
PTEROMA PENDULA JOANNIS) (LEPIDOPTERA: PSYCHIDAE)***

MUHAMMAD NURUL YAQIN BIN SYARIF

FP 2018 66



**FORMULATION AND LABORATORY EFFICACY OF *ISARIA*
FUMOSOROSEA AGAINST BAGWORMS (*METISA PLANA* WALKER AND
PTEROMA PENDULA JOANNIS) (LEPIDOPTERA: PSYCHIDAE)**

By

MUHAMMAD NURUL YAQIN BIN SYARIF

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

February 2017

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**FORMULATION AND LABORATORY EFFICACY OF *ISARIA*
FUMOSOROSEA AGAINST BAGWORMS (*METISA PLANA* WALKER AND
PTEROMA PENDULA JOANNIS) (LEPIDOPTERA: PSYCHIDAE)**

By

MUHAMMAD NURUL YAQIN BIN SYARIF

February 2017

Chair : Dzolkhifli bin Omar, PhD
Faculty : Agriculture

Relying solely on chemical insecticides for bagworm control in oil palm plantation could lead to many consequences to the ecosystem, environment and human. Thus, developing mycoinsecticide that able to control this pest could be the alternative to reduce chemical insecticides dependence. Entomopathogenic fungi might fulfil their potential with suitable choice of formulation and stable throughout the application and storage period either in biological, chemical, or physical aspect. Wettable powder is the formulation to be developed due to its easy and cheap production procedure. In this study, three fungi isolated from the infected *Pteroma pendula* collected from United Malacca Berhad (UMB) plantations in Pahang and Negeri Sembilan were found to be *Isaria fumosorosea* based on the morphological observations. Those three isolates were identified using molecular identification procedure along with six isolates obtained from Forest Entomology Laboratory, Faculty of Forestry Universiti Putra Malaysia (UPM). Six of the isolates were *I.fumosorosea* where isolate BSB01 isolated of infected bagworms from UMB Plantation in Kemayan, Pahang was the most virulent isolates against *P.pendula*. This isolate was then mass produced and formulated with surfactants and filler. The compatibility of dispersants and wetting agents with this isolate was determined earlier before being used in wettable powder formulation. Screening the surfactants against *I.fumosorosea* BSB01 conidia viability and fungal growth were conducted on the isolate, aerial conidia and submerged propagules. Surfactants found to be qualified were TERSPERSE® 2105, TERSPERSE® 2700, ELTESOL® SC, ELTESOL® SX, sodium naphthalenesulfonate, sodium lignosulfonate, sodium polyacrylates, and SIPERNAT® 22S. Among those qualified surfactants, two of them are wetting

agents (ELTESOL® SC and ELTESOL® SX). Concentration of surfactant affects the conidia viability at 10% or higher. A surfactant system consisted of two dispersants and a wetting agent were prepared using pseudoternary phase diagram. Stable regions in each surfactant system were combined to find the mutual stable region. The single point was selected from the mutual stable region with the limit of wetting agents fixed at 15%. The stable region showed satisfying physical stability against kaolin. The best point was found to be at the ratio of 52:33:15 (Dispersant 1:Dispersant 2:Wetting agents). The surfactant ratio was then characterised with the presence of fungal aerial conidia and fungal submerged propagules. Submerged fungal propagules formulations showed better physical stability compared to the aerial conidia formulation. Although all formulations showed a steep decline of active colony counting from 9.0×10^8 to 1.0×10^8 cfu mL⁻¹ within the first three days storage, these formulations maintained this viability until 90 days. Biological efficacy of the formulated submerged propagules were determined where all formulations showed no significant difference of LT₅₀ values when compared with unformulated against *Metisa plana* and *P.pendula*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Sarjana Sains

**FORMULASI DAN KEBERKESANAN MAKMAL *ISARIA FUMOSOROSEA*
TERHADAP ULAT BUNGKUS (*METISA PLANA WALKER* DAN *PTEROMA*
PENDULA JOANNIS) (LEPIDOPTERA: PSYCHIDAE)**

Oleh

MUHAMMAD NURUL YAQIN BIN SYARIF

Februari 2017

Pengerusi : Prof Dzolkhifli b. Omar, PhD
Fakulti : Pertanian

Kebergantungan kepada racun serangga kimia untuk kawalan ulat bungkus di ladang kelapa sawit boleh membawa kepada keburukan kepada ekosistem, alam sekitar dan juga manusia. Maka, membina mikoinsektisida yang boleh mengawal perosak ini boleh menjadi alternatif untuk mengurangkan kebergantungan kepada racun serangga kimia. Kulat entomopatogenik mampu memenuhi potensi dengan pilihan formulasi yang sesuai dan stabil sepanjang penggunaan dan tempoh penyimpanan samaada dalam aspek biologi, kimia atau fizikal. Serbuk boleh basah adalah formulasi yang akan dibangunkan kerana prosedur penghasilan yang mudah dan murah. Dalam kajian ini. Tiga kulat yang diasingkan dari *Pteroma pendula* terjangkit yang dikumpulkan dari ladang-ladang United Malacca Berhad (UMB) di Pahang dan Negeri Sembilan dijumpai adalah *Isaria fumosorosea* berdasarkan pemerhatian secara morfologi. Tiga isolat tersebut telah dikenal pasti s molekul bersama dengan enam isolat yang diperolehi daripada Makmal Entomologi Hutan, Fakulti Perhutanan, Universiti Putra Malaysia (UPM). Enam isolat adalah *I.fumosorosea* di mana isolat BSB01 diasingkan dari ulat bungkus terjangkit daripada ladang UMB di Kemayan, Pahang adalah isolat paling viral terhadap *P.pendula*. Isolat ini seterusnya dihasil jisim dan diformulasi dengan surfaktant dan pemenuh. Keserasian penyerak dan agen pembasah dengan isolat ini telah ditentukan lebih awal sebelum digunakan di dalam serbuk boleh basah. Penyaringan surfaktant terhadap percambahan konidia dan perkembangan kulat *I.fumosorosea* BSB01 telah dijalankan ke atas konidia arial dan propagul tenggelam. Surfaktant didapati terlibat adalah TERSPERSE® 2105, TERSPERSE® 2700, ELTESOL® SC, ELTESOL® SX, sodium NAPHTHALENESULPHONATE sodium lignosulfonate, sodium polyacrylates, and SIPERNAT® 22S. Di antara surfactant-surfaktant yang layak, dua daripadanya adalah agen pembasah (ELTESOL® SC and ELTESOL® SX).

Kepekatan surfaktant mempengaruhi percambahan konidia pada kepekatan 10% atau lebih. Satu sistem surfaktant terdiri daripada dua penyerak dan satu agen pembasah disediakan menggunakan Rajah Fasa Pseudoternary. Rantau stabil dalam setiap sistem surfaktant digabungkan untuk mencari rantau stabil bersama. Satu titik dipilih daripada rantau stabil bersama dengan agen pembasah dihadkan kepada 15%. Rantau stabil menunjukkan ujian kestabilan fizikal yang memuaskan terhadap kaolin. Titik terbaik dijumpai pada nisbah 52:33:15 (Penyerak 1:Penyerak 2:Agen Pembasah) dan digunakan dalam penyediaan formulasi sebuk boleh basah isolat BSB01. Nisbah surfaktant ini kemudiannya dikarakterkan dengan kehadiran bahan aktif, konidia kulat udara dan propagul kulat tenggelam. Formulasi propagul kulat tenggelam menunjukkan kestabilan fizikal lebih baik berbanding dengan formulasi konidia. Walaupun semua formulasi menunjukkan kejatuhan mendadak dalam kiraan koloni aktif daripada 9.0×10^8 ke 1.0×10^8 cfu mL⁻¹ pada tiga hari pertama penyimpanan, formulasi-formulasi ini mengekalkan percambahan ini sehingga 90 hari. Keberkesanan biologi formulasi propagul kulat tenggelam telah ditentukan di mana semua formulasi menunjukkan tiada perbezaan signifikan dalam LT₅₀ dibandingkan dengan propagul tidak diformulasi terhadap *Metisa plana* dan *P.pendula*.

ACKNOWLEDGEMENTS

First of all, I would like to express my utmost gratitude to Allah s.w.t for their bless and guidance, I am able to complete this challenging journeys. There are numerous people I wish to thank for their contributions to my development as a scientist and as a person. I wish to thank my beloved parents, my father Syarif bin Umar and my mother Norsilla binti Abdul Rahman, for bringing me into this world, for providing me with loves, cares, motivational supports since I was in the womb up until now. You pour me your extraordinary love as you were able to keep your patience while I went through my teenage years and young adulthood. I also would like to thank my sisters and brothers for their encouragements and supports.

I was fortunate to have had outstanding, motivating, and inspiring supervisors. In particular, I wish to thank Prof. Dr. Dzolkhifli Omar as the chairman of my supervisory committee and Prof. Dr. Ahmad Said Sajap as I learned that four years are not enough to keep even 1% of their vast knowledges and experiences. I could not have done the work described in this disciplinary thesis without the solid foundation and guidance they provided me with.

My study period in faculty of Agriculture, UPM was really wonderful apart of the struggle to complete the study. I would like to express my utmost gratitude towards Dr Hayu for motivating me throughout my study and supporting me especially towards the end of my study. Not to forget, also all Toxicology Research Group members - Yasir, Najib, Alan, Fariq, Syuna, Moon, Kak Mo, Kak Wani, Afie, Mira and others for making my time as a student so pleasant. I would like to thank Mr Jarkasi, Mr Zaki for your cooperative effort while I conducted my study. Their contributions to my student life will always be remembered.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Dzolkhifli bin Omar, PhD

Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Ahmad Said Sajap, PhD

Professor
Faculty of Forestry
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman
of Supervisory
Committee: Dzolkhifli bin Omar

Signature: _____
Name of Member of
Supervisory
Committee: Ahmad Said Sajap

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xv
 CHAPTER	
1 GENERAL INTRODUCTION	1
2 LITERATURES REVIEW	3
2.1 Entomopathogenic Fungi	3
2.1.1 <i>Isaria fumosorosea</i> Wize	4
2.2 Mycoinsecticides Formulations	5
2.3 Wettable Powder Formulations	6
2.3.1 Fungal Propagules Mass Production	6
2.3.1.1 Conidia Diphasic Fermentation	8
2.3.1.2 Submerged Propagules Liquid Fermentation	9
2.3.2 Formulation Components	10
2.3.3 Dispersants and Polymeric Dispersants	12
2.3.4 Pseudoternary Phase Diagram	13
2.4 Formulating Process	13
2.5 Physical Characteristics	14
2.5.1 Wettability	14
2.5.2 Suspensibility	15
2.5.3 Dispersability	16
2.5.4 Storage Stability	16
2.6 Bagworms in Malaysia	17
2.6.1 <i>Metisa plana</i> Walker	18
2.6.2 <i>Pteroma pendula</i> Joannis	19
3 COLLECTION, ISOLATION, IDENTIFICATION AND SCREENING OF ISARIA FUMOSOROSEA ISOLATES	20
3.1 Introduction	20
3.2 Material and Methods	20
3.2.1 Fungal collection and isolation	20
3.2.2 <i>Pteroma pendula</i> rearing	22

3.2.3	Molecular Identification	22
3.2.3.1	DNA Extraction	22
3.2.3.2	DNA Amplification	22
3.2.3.3	DNA Sequencing	23
3.2.4	Screening for virulence of <i>I.fumoso</i> rosea isolates	23
3.2.4.1	Preparation of fungal suspensions	23
3.2.4.2	Bioassay screening for <i>I. fumoso</i> rosea isolates	24
3.3	Results	24
3.3.1	Isolation and identification of <i>I.fumoso</i> rosea	24
3.3.2	Screening of <i>I.fumoso</i> rosea virulence against <i>P.pendula</i>	27
3.4	Discussion	28
3.5	Conclusion	28
4	PREPARATION AND CHARECTERIZATION OF WETTABLE POWDER (WP) FORMULATION OF ISARIA FUMOSOROSEA WIZE	30
4.1	Introduction	31
4.2	Material and Methods	31
4.2.1	Mass Production of Fungal Propagules	31
4.2.1.1	<i>I.fumoso</i> rosea Mother Culture	31
4.2.1.2	Conidia mass production	31
4.2.1.3	Submerged propagules mass production	31
4.2.2	Surfactants	32
4.2.2.1	Screening of Surfactant effect on the viability of fungal conidia	34
4.2.2.2	Screening of surfactants effect on the fungal growth	34
4.2.3	Wettable Powder Formulation of Entomopathogenic	35
4.2.3.1	Constructing Dispersants-Wetting Agent System	35
4.2.3.2	Mixing Formulation	35
4.2.4	Physical test of powder formulation	35
4.2.4.1	Wetting time test for wettability	36
4.2.4.2	Suspensibility	36
4.2.4.3	Foam and Sedimentation	37
4.2.4.4	Dispersibility	38
4.2.4.5	Calculating physical test evaluation	40
4.2.5	Storage stability	41
4.3	Results	41

4.3.1 Mass Production of Fungal Propagules	41
4.3.2 Screening Additives against Entomopathogenic Fungi	43
4.3.2.1 Screening additives against fungal conidia germination	43
4.3.2.2 Screening additives against fungal growth	44
4.3.3 Wettable Powder Formulation	45
4.3.4 Physical Test of Wettable Powder Formulations	49
4.3.5 Storage Stability	51
4.4 Discussion	51
4.5 Conclusion	56
5 LABORATORY BIOEFFICACY EVALUATION OF THE WETTABLE POWDER FORMULATIONS	57
5.1 Introduction	57
5.2 Material and Methods	57
5.2.1 Bagworm Rearing	57
5.2.2 Preparation of treatment solutions	57
5.2.3 Bioassay	58
5.3 Results	59
5.4 Discussion	62
5.5 Conclusion	63
SUMMARY OF THE STUDY	64
FUTURE RECOMMENDATIONS	65
REFERENCES	66
BIODATA OF STUDENT	83

LIST OF TABLES

Table	Page
2.1 Previous study recommended percentage on every wettable powder formulation components	12
3.1 List of ITS primers and sequences (White <i>et al</i> , 1990) used in fungi identification	23
3.2 Cycling conditions for PCR program ITS	23
3.3 Fungal isolates name and molecular result after BLAST	26
4.1 Surfactants used in the experiment	33
4.2 Criteria evaluation of dispersion state. Point I determined through observation	39
4.3 Criteria for evaluation of dispersion state time factor (f) in the evaluation of the dispersion state is that, in the case of practical products, the dispersion state after the lapse of time is more important than that immediately after dispersion.	40
4.4 Component percentage after 12% water loss from submerged propagules based wettable powder formulation	47
4.5 Physical test evaluation measurements for submerged propagules formulations	49
4.6 Physical test evaluation measurements for conidia formulations	50
5.1 Concentration equality in Wettable powder formulation after air drying process	58
5.2 Bioefficacy test of three formulations against <i>Pteroma pendula</i>	61
3.2 Bioefficacy test of three formulations against <i>Metisa plana</i>	60

LIST OF FIGURES

Figure	Page
3.1	24
3.2	25
3.3	27
4.1	42
4.2	42
4.3	43
4.4	44
4.5	45
4.6	46
4.7	48
4.8	50
4.9	51
5.1	59
5.2	60

LIST OF ABBREVIATIONS

ANOVA	Universiti Putra Malaysia
BLAST	Basic Local Alignment Search Tool
bp/kbp	Base pair/kilo base pair
cfu	Colony forming unit
cfu mL ⁻¹	Colony forming unit per millilitre
CTAB	Cetyl trimethylammonium bromide
DNA	Deoxyribonucleic acid
g	Gram
IPM	Integrated Pests Management
ITS	Internal Transcribe Spacer
LT ₅₀	Lethal time 50%
mm	millimetre
mmHg	Millimetre Hargentum
MT	Method Technical
PCR	Polymerase Chain Reaction
PCV	Pressure control valve
PDA	Potato dextrose agar
ppm	Parts per million
rpm	Revolution per minute
SAS	Statistical Analysis System
sec	Second
TAE	Tris Acetate EDTA
TE	Tris EDTA
UMB	United Malacca Berhad
UPM	Universiti Putra Malaysia
ULV	Ultra-low volume
UV	Ultra Violet
WHO	World Health organization
μL	Microlitre
μg L ⁻¹	Microgram per litre
®	Reserved
°C	Degree Celsius
%	Percent
∞	Infinity

CHAPTER 1

GENERAL INTRODUCTION

Environmental concerns remain eternal issue involving chemical pesticides and more eco-friendly approaches have been integrated to prevent economic losses caused by pests and diseases. Biological control approaches have always been highlighted on their ability to play major roles in pest and diseases management system. Recent studies on biological control have shown promising results as an alternative to chemical control. The ability to manipulate living things including microbes as biological agent has highlighted the importance of biological pesticide usage in agriculture industries today. Biopesticides are types of pesticide derived from such natural materials as animals, plants, bacteria, and certain minerals. Integrated Pest Management (IPM) program to maintain pest population below economic threshold could minimize the use of chemical pesticide. Microorganisms could give infection naturally with nature's assistance but need to continuously present to keep the pest population below economic threshold.

Entomopathogenic fungi are one of the biological agents used as biopesticides due to their epizootic nature to insects. The term used for a biopesticides utilizing entomopathogenic fungi to control insect pest is called mycoinsecticides. The mycoinsecticides highlight the concept of mass production entomopathogenic fungus formulating as active ingredient in insecticides and applying as conventional pesticides to the target insect pests. In order to successfully delivering the fungus to the target hosts, physically stable formulation has to be developed without causing much effect to the its viability and growth that might reduce the infection. A stable formulation at optimum dosage would ensure the fungus to spread at optimum dosage throughout the target host's population in the treated area.

Entomopathogenic fungi such as *Metarhizium anisopliae*, *Beauveria bassiana*, and *Isaria fumosorosea* are ubiquitous microbes known for their epizootic to insect pests and possess desirable traits favourable for their development as microbial control agents (Lacey *et al.*, 2015). Wettable powder formulation has been associated with the early introduction of mycoinsecticides. This type of formulation has always available in pesticides market despite several limitations. Fungus could not be delivered to its optimum due to the physical instability of the suspension formed. In order to form stable spraying suspension, formulation should consist of favourable dispersants and wetting agents to ensure good delivery of active ingredient to the target. According to Alves *et al.*, (1996), biological agents as active ingredient in biopesticide may suffer degradation in terms of germination and viability due to both biotic (microorganism antagonistic factors) and abiotic (temperature and humidity) factors. This is the main reason on why formulating mycoinsecticides should be

done meticulously considering all possible factors affecting the efficacy of the fungal active ingredient.

Wettable powder formulation is the most common and simplest form of formulation in pesticide industry. In recent years, formulators need to review the composition of wettable powder formulation for greater adaptation and more practical application. The introduction of improvised formulations such as suspension concentrates, water dispersible granule, capsule suspension and suspo-emulsion arguably improved the efficacy of pesticides delivery. Despite the emergence of new improvised formulation, this formulation is still getting much attention due to their long storage stability, good miscibility with water and convenient application using conventional spraying equipment (Brar, 2006). Knowles (2005) reported that wettable powder formulation has the advantage on having low production cost, low phytotoxicity, good persistence on porous substrates, and versatile. Developing a wettable powder formulation of mycoinsecticides requires the art of balancing the composition of epizootic fungus and inert ingredient to achieve physically stable formulation and fungal spore's viability during storage period and cause significant mortality to the target hosts.

Surfactant is probably the most important additives in the formulation (Knowles, 2005). Surfactants were more preferred be included inside the formulation. Tank mix surfactants are not recommended as compared to formulation build in surfactant for two reasons, simpler usage and proportion wise advantage where all the ingredients are in the right amount (Jones & Burges, 1998). There are two types of surfactants recommended in wettable powder formulation which are dispersants and wetting agents. These surfactants combination affects formulation stability performance in wettable powder suspension. Every formulation component should be at its optimum amount to make a good suspension and well-perform on infecting target insect pests. The amount of surfactant used in the formulation should be optimised to achieve suspensible formulation as the amount of surfactant could greatly affect the production cost. This type of study could give basic knowledge to the importance of achieving good formulation stability for commercialization. Thus, this study sets several objectives to be accomplished to overcome the all challenges faced during formulating process.

The objectives for this study were as follows:

- 1.To evaluate the compatibility of ingredients for development of Wettable Powder (WP) formulation of mycoinsecticides
- 2.To prepare and characterize the Wettable Powder (WP) formulation of entomopathogenic fungus
- 3.To evaluate the biological performance of the Wettable Powder formulations against bagworms

REFERENCES

- Al-Aidroos, K., & Roberts, D. W. (1978). Mutants of *Metarhizium anisopliae* with increase virulence toward mosquito larvae. Canadian Journal of Genetics and Cytology, 20(2), 211–219.
- Ali-Shtayeh M.S., Abu Ghdeib S.I. (1999) Antifungal activity of plant extracts against Dermatophytes Mycoses, 42, pp. 665–672
- Altre, J. A., & Vandenberg, J. D. (2001). Factors influencing the infectivity of isolates of *Paecilomyces fumosoroseus* against diamondback moth, *Plutella xylostella*. Journal of Invertebrate Pathology, 78(1), 31–36.
- Alves, R. T., Bateman, R. P., Gunn, J., Prior, C., & Leather, S. R. (2002). Effects of different formulations on viability and medium-term storage of *Metarhizium anisopliae* conidia. Neotropical Entomology, 31(1), 91–99.
- Alves, S. B., Pereira, R. M., Stimac, J. L., & S. Viera, A. (1996). Delayed Germination of *Beauveria bassiana* Conidia after Prolonged Storage at Low, Above-freezing Temperatures. Biocontrol Science and Technology, 6 (june 1996), 575–581.
- Amjad, Z., & Zuhl, R. W. (2005). The Impact of Thermal Stability on the Performance of Polymeric Dispersants for Boiler Water Systems.
- Anderson, T. E., Hajek, A. E., Roberts, D. W., Preisler, H. K., & Roberteson, J. L. (1989). Colorado Potato Beetle (Coleoptera: Chrysomelidae): Effects of Combinations of *Beauveria bassiana* with Insecticides. Journal of Economic Entomology, 82(1), 83–89.
- Anderson, W. G. (1986). Wettability literature survey. II: Wettability measurement. Journal of Petroleum Technology, 38(12), 1246–1262.
- Araki, J., Wada, M., & Kuga, S. (2001). Steric stabilization of a cellulose microcrystal suspension by poly (ethylene glycol) grafting. Langmuir, 17(1), 21–27.
- Asi, M. R., Bashir, M. H., Afzal, M., Ashfaq, M., Sahi, S.T., & Ramzan, M. (2010). Compatibility of Entomopathogenic Fungi, *Metarhizium anisopliae* and *Paecilomyces fumosoroseus* with Selective Insecticides. Pak. J. Bot., 42(6), 4207–4214.
- Auld, B. A., & Morin, L. (1995). Constraints in the Development of Bioherbicides. Weed Technology, 9(3), 638–652.
- Avery, P. B., Hunter, W. B., Hall, D. G., Jackson, M. A., Powell, C. A., & Rogers, M. E. (2009). *Diaphorina citri* (Hemiptera : Psyllidae) Infection and Dissemination of the Entomopathogenic Fungus *Isaria fumosorosea* (Hypocreales : Cordycipitaceae) Under Laboratory Conditions. Florida Entomologist, 92(4).

- Avery, P. B., Kumar, V., Shareef, F., Barbeau, E., Nelson, P., Landa, Z., Powell, C. A. (2013). Competitive capacity of *Isaria fumosorosea* with other fungal species present on the phylloplane of Ficus leaves for biocontrol of ficus whitefly. *Fungal Ecology*.
- Avery, P., Pick, D., Aristizábal, L., Kerrigan, J., Powell, C., Rogers, M., & Arthurs, S. (2013). Compatibility of *Isaria fumosorosea* (Hypocreales: Cordycipitaceae) Blastospores with Agricultural Chemicals Used for Management of the Asian Citrus Psyllid, *Diaphorina citri* (Hemiptera: Liviidae). *Insects*, 4(4), 694–711.
- Barbosa, P., Krischik, V., & Lance, D. (1989). Life-history traits of forest-inhabiting flightless Lepidoptera. *American Midland Naturalist*, 262–274.
- Bartlett, M. C., & Jaronski, S. T. (1988). Mass production of entomogenous fungi for biological control of insects. *Fungi in Biological Control Systems*, 61–85.
- Basri, M., & Kevan, P. G. (1995). Life history and feeding behaviour of the oil palm bagworm, *Metisa plana* Walker (Lepidoptera: Psychidae).
- Basri, M. W., Abdul Halim, H., & Zulkipli Masijan. (1988). Bagworms (Lepidoptera: Psychidae) of Oil Palms in Malaysia. *PORIM Occasional Paper*, (23), 1–23.
- Basri, M. W., Norman, K., & Hamdan, A. B. (1995). Natural enemies of the bagworm, *Metisa plana* Walker (Lepidoptera: Psychidae) and their impact on host population regulation. *Crop Protection*, 14(8), 637–645.
- Bateman, R. (1993). Simple, Standardized Methods For Recording Droplet Measurements And Estimation Of Deposits From Controlled Droplet Applications. *Crop Protection*, 12(3), 201–206.
- Bateman, R. P., & Alves, R. T. (2000). Delivery systems for Mycoinsecticides Using Oil-based Formulations. *Aspects of Applied Biology*, 57, 163–170.
- Bateman, R. P., Kooyman, C., & Lamer, C. (1998). Some observations on the dose transfer of mycoinsecticide sprays to desert locusts. *Crop Protection*, 17(2), 151–158.
- Behle, R., Compton, D., Laszlo, J., & Shapiro Ilan, D. (2006). Potential Use of Soyscreen in Formulations of *Beauveria bassiana* for UV protection. *Entomol. Soci. Amer. Ann. Meet. North Central Branch. Paper No. D-215*.
- Bhattacharyya, A., Barik, S., & Ganguly, P. (2009). New pesticide molecules, formulation technology and uses: Present status and future challenges. *Journal of Plant Protection Sciences*, 1(1), 9–15.

- Binks, B. P., & Lumsdon, S. O. (2000). Influence of Particle Wettability on the Type and Stability of Surfactant-Free Emulsions. *Langmuir*, 16(23), 8622–8631.
- Bolckmans, K., Sterk, G., Eyal, J., Sels, B., & Stepman, W. (1995). PreFeRal, (*Paecilomyces fumosoroseus* strain Apopka 97), a new microbial insecticide for the biological control of whiteflies in greenhouses. *Mededelingen-Faculteit Landbouwkundige En Toegepaste Biologische Wetenschappen, Universiteit Gent*, 60(3a), 707–711.
- Bora, T., H. Ozaktan, E. Gore., and E. A. (2004). Biological Control of *Fusarium oxysporum* f. sp. melonis by Wettable Powder Formulations of the two Strains of *Pseudomonas putida*. *Journal of Phytopatology*, 475, 471–475.
- Boyetchko, S. M. (1996). Impact of soil microorganisms on weed biology and ecology. *Phytoprotection*, 77(1), 41.
- Boyette, C. D., Quimby Jr, P. C., Caesar, A. J., Birdsall, J. L., Connick Jr., W. J., Daigle, D. J., Abbas, H. K. (1996). Adjuvants, formulations, and spraying systems for improvement of mycoherbicides. *Weed Technology*, 637–644.
- Bradley, C. A., Black, W. E., Kearns, R., & Wood, P. (1992). *Frontiers in Industrial Mycology*. In G. F. Leatham (Ed.) (pp. 160–173). Boston, MA: Springer US.
- Brar, S. K., Verma, M., Tyagi, R. D., & Valéro, J. R. (2006). Recent Advances In Downstream Processing And Formulations Of *Bacillus Thuringiensis* Based Biopesticides. *Process Biochemistry*, 41(2), 323–342.
- Burges, H. D. (1998). Formulation of Mycoinsecticides. In *Formulation of Microbial Biopesticides* (pp. 131–185).
- Burges, H. D., & Hussey, N. W. (1971). *Microbial Control Of Insects and Mites*. Microbial Control of Insects and Mites.
- Butt, T. M., & Goettel, M. S. (2000). Bioassays of entomogenous fungi. In *Bioassays Of Entomopathogenic Microbes And Nematodes*. (pp. 141–195).
- Butt, T. M., Jackson, C. W., & Magan, N. (2001). *Fungi as Biocontrol Agents: Progress, Problems, and Potential*. (T. M. Butt, C. W. Jackson, & N. Magan, Eds.). CABI Publishing.
- Castillo, M.A., Moya, P., Hernandez, E., & Primo-Yufera, E. (2000). Susceptibility of *Ceratitis capitata* (Diptera: Tephritidae) to entomopathogenic fungi and their extracts. *Biological Control*, 19, 274–282.

- Cayer, C. M., Musco, V. A., & Peterson Jr, R. F. (1997, January). Oxyfluorfen dispersible granule formulation. Patent No. 5,597,777. Washington, DC: U.S. Patent and Trademark Office.: Google Patents.
- Chakraborty, R., Borglin, S. E., Dubinsky, E. A., Andersen, G. L., & Hazen, T. C. (2012). Microbial response to the MC-252 oil and Corexit 9500 in the Gulf of Mexico. *Frontiers in Microbiology*, 3, 357.
- Chander, S., & Hogg, R. (2007). Characterization of the Wetting and Dewetting Behavior of Powders. *KONA*, 25(25), 56–75.
- Charnley, A. K. (2003). Fungal pathogens of insects: Cuticle degrading enzymes and toxins. *Advances in Botanical Research*, 40, 241–321.
- Chen, C., Wang, Z., & Feng, M. (2009). Synchronous production of conidial powder of several fungal biocontrol agents in series fermentation chamber system, *African Journal of Biotechnology* 8(15), 3649–3653.
- Chengshu, W., Sibao, W., Meizhen, F., & Zengzhi, L. (1999). Relationship Between Shelf-Life And Heat Tolerance of The Conidia of *Beauveria Bassiana*. *Chinese Journal of Biological Control*, 15(4), 162–165.
- Cherry, A., Jenkins, N. E., Heviefo, G., Bateman, R. P., & Lomer, C. J. (1999). Operational and economic analysis of a west african pilotscale production plant for aerial conidia of *Metarhizium* spp. for use as a mycoinsecticide against locusts and grasshoppers. *Biocontrol Science and Technology*, 9(1), 35–51.
- Chung, G. F. (1989). Spraying and trunk injection of oil palm for pest control. *Planter*, 65(764), 500–524.
- Chung, G. F. (1998). Strategies and methods for the management of leaf-eating caterpillars of oil palm. *Planter*, 74(871), 531–558.
- Chung, G. F., & Sim, C. S. (1991). Bagworm census and control. A case study. In *PORIM* (Vol. 4, pp. 433–442). Kuala Lumpur, Malaysia: PORIM.
- Churchill, B. W. (1982). Mass Production Of Microorganisms For Biological Control. page 139-156 in *Biological Control of Weed with Plant Pathogen* R Charudattan, H. L. Walker eds, Wiley, New York
- Clayton, E. T., Elinor, J. R., & M, R. J. (1975). Cleaner Compositions. Patent No: 3,882,038 U.S. Patent Office.
- Connick Jr, W. J., Jackson, M. A., Williams, K. S., & Boyette, C. D. (1997). Stability Of Microsclerotial Inoculum of *Colletotrichum truncatum* encapsulated in wheat flour-kaolin granules. *World Journal of Microbiology and Biotechnology*, 13(5), 549–554.

- Costa, S. D., Grassano, S., & Jiancai, L. (2014). Sweet Whey Based Biopesticide Composition. Patent No: 8,623,420. Washington, DC: U.S. Patent and Trademark Office.
- Couch, T. L. (2000). Entomopathogenic Bacteria: from Laboratory to Field Application. In J.-F. Charles, A. Delécluse, & C. N.-L. Roux (Eds.) (pp. 297–316). Dordrecht: Springer Netherlands.
- Couch, T. L., & Ignoffo, C. M. (1981). Formulation of Insect Pathogens. In H. D. Burges (Ed.), In Microbial control of pest and plant diseases 1970-1980 (pp. 621–634). New York: Academic Press.
- Daoust, R. A., Ward, M. G., & Roberts, D. W. (1983). Effect of formulation on the viability of *Metarhizium anisopliae* conidia. Journal of Invertebrate Pathology, 41(2), 151–160.
- Dayakar, B. V., Narayanan, N. N., & Gnanamanickam, S. S. (2000). Cross-Compatibility and Distribution of Mating Type Alleles of the Rice Blast Fungus *Magnaporthe grisea* in India. Plant Disease, 84(6), 700–704.
- De, A., Bose, R., Kumar, A., & Mozumdar, S. (2014). Pesticide Formulations. In Targeted Delivery of Pesticides Using Biodegradable Polymeric Nanoparticles (pp. 15–23). Springer.
- Domsch, K. H., Gams, W., & Anderson, T.-H. (1980). Compendium of soil fungi. Volume 2. Academic Press (London) Ltd.
- Dunlap, C. a, Biresaw, G., & Jackson, M. a. (2005). Hydrophobic And Electrostatic Cell Surface Properties Of Blastospores of The Entomopathogenic Fungus *Paecilomyces fumosoroseus*. Colloids and Surfaces. B, Biointerfaces, 46(4), 261–6.
- Dunlap, C. a., Jackson, M. a., & Wright, M. S. (2007). A Foam Formulation of *Paecilomyces fumosoroseus*, an Entomopathogenic Biocontrol Agent. Biocontrol Science and Technology, 17(5), 513–523.
- Dziubinski, S., A. Witczal-Stawicka, & J., S. (2004). Study of suspension stability Petroleum & Coal, 47–54.
- Eilenberg, J., Hajek, A. E., & Lomer, C. (2001). Suggestions for Unifying The Terminology In Biological Control. BioControl, 46(1), 387–400.
- Elzein, A., Kroschel, J., & Müller-Stöver, D. (2004). Effects of Inoculum Type And Propagule Concentration On Shelf Life Of Pesta Formulations Containing *Fusarium oxysporum* Foxy 2, A Potential Mycoherbicide Agent for *Striga* spp. Biological Control, 30(2), 203–211.
- Er, M. K., & Gökçe, A. (2004). Effects of Selected Pesticides Used Against Glasshouse Tomato Pests on Colony Growth And Conidial Germination of *Paecilomyces fumosoroseus*. Biological Control, 31(3), 398–404.

- Fargues, J., Maniania, N. K., & Delmas, J.-C. (1994). Infectivity of Propagules of *Paecilomyces fumosoroseus* During In Vitro Development to *Spodoptera Frugiperda*. *Journal of Invertebrate Pathology*, 64(3), 173–178.
- Faria, M. R. De, & Wraight, S. P. (2007). Mycoinsecticides and Mycoacaricides: A comprehensive list with worldwide coverage and international classification of formulation types. *Biological Control*, 43(3), 237–256.
- Faria, M., & Wraight, S. P. (2001). Biological Control of *Bemisia Tabaci* With Fungi. *Crop Protection*, 20(9), 767–778.
- Feng, M. G., Poprowski, T. J., & Khachatourians, G. G. (1994). Production, Formulation and Application of the Entomopathogenic Fungus *Beauveria bassiana* for Insect Control: Current Status. *Biocontrol Science and Technology*, 4, 3–34.
- Ferron, P. (1978). Biological Control of Insect Pests by Entomogenous Fungi. *Annual Review of Entomology*, 23(1), 409–442.
- Ferron, P. (1981). Pest Control by The Fungi *Beauveria* and *Metarhizium*. In *Microbial Control of Pests and Plant Diseases* (pp. 1970–1980).
- Flexner, J. L., Lighthart, B., & Croft, B. A. (1986). The Effects of Microbial Pesticides on Non-Target, Beneficial Arthropods. *Agriculture, Ecosystems & Environment*, 16(3-4), 203–254.
- Friesen, T. J., Holloway, G., Hill, G. A., & Pugsley, T. S. (2006). Effect of Conditions and Protectants on The Survival Of *Penicillium Bilaiae* During Storage. *Biocontrol Science and Technology*, 16(1), 89–98.
- Graves, T. M., & Koundakjian, T. H. (1978, January 31). Aqueous Flowable Concentrates of Particulate Water-Insoluble Pesticides. Patent No. 4,071,617. Washington, DC: U.S. Patent and Trademark Office.
- Gregory, P. H. (1978). Distribution of Airborne Pollen and Spores And Their Long Distance Transport. *Pure and Applied Geophysics*, 116(2), 309–315.
- Grunberger, D., Banerjee, R., Eisinger, K., Oltz, E. M., Efros, L., Caldwell, M., Nakanishi, K. (1988). Preferential Cytotoxicity on Tumor Cells by Caffeic Acid Phenethyl Ester Isolated from Propolis. *Cellular and Molecular Life Sciences*, 44(3), 230–232.
- Hajek, A. E., McManus, M. L., & Junior, I. D. (2005). Catalogue of Introductions of Pathogens and Nematodes For Classical Biological Control Of Insects And Mites. FHTET, USDA Forest Service.
- Hajek, A. E., & St. Leger, R. J. (1994). Interactions Between Fungal Pathogens and Insect Hosts. *Annu. Rev. Entomol.*, 39, 293–322.

- Hall, R. A. (1981). fungus *Verticillium lecanii* as a microbial insecticide against aphids and scales. *Microbial Control of Pests and Plant Diseases*, 1970-1980.
- Hall, R. A., & Papierok, B. (1982). Fungi as Biological Control Agents of Arthropods Of Agricultural And Medical Importance. *Parasitology*, 84(04), 205–240.
- Hallsworth, J. E., & Magan, N. (1994). Effects of KCl Concentration on Accumulation Of Acyclic Sugar Alcohols And Trehalose In Conidia Of Three Entomopathogenic Fungi. *Letters in Applied Microbiology*, 18(1), 8–11.
- Hallsworth, J. E., & Magan, N. (1995). Manipulation of Intracellular Glycerol and Erythritol Enhances Germination Of Conidia At Low Water Availability. *Microbiology*, 141(5), 1109–1115.
- Hallsworth, J. E., & Magan, N. (1996). Culture Age, Temperature, And Ph Affect the Polyol and Trehalose Contents Of Fungal Propagules. *Applied and Environmental Microbiology*, 62(7), 2435–2442.
- Hassan, A. E. M., & Charnley, A. K. (1989). Ultrastructural Study Of The Penetration By *Metarhizium anisopliae* Through Dimilin-Affected Cuticle Of *Manduca sexta*. *Journal of Invertebrate Pathology*, 54(1), 117–124.
- Hawckett, B. S., Park, H. Y., Blatt, E., & Zhou, Z. (1996). Wettable Powder Formulations. Patent No: 0789510 B1 European Patent Office.
- Heath, D., Tadros, T. F., Knott, R. D., & Knowles, D. A. (1984). Stabilization of Aqueous Pesticidal Suspensions By Graft Copolymers And Their Subsequent Weak Flocculation By Addition Of Free Polymer. In H. B. Scher (Ed.), *Advances in pesticide formulation technology*, ACS Series 254 (pp. 11–28). Washington, DC: American Chemical Society.
- Heux, L., Chauve, G., & Bonini, C. (2000). Nonflocculating And Chiral-Nematic Self-Ordering of Cellulose Microcrystals Suspensions In Nonpolar Solvents. *Langmuir*, 16(21), 8210–8212.
- Holder, D. J., & Keyhani, N. O. (2005). Adhesion Of The Entomopathogenic Fungus *Beauveria* (Cordyceps) *Bassiana* To Substrata. *Applied and Environmental Microbiology*, 71(9), 5260–5266.
- Holder, D. J., Kirkland, B. H., Lewis, M. W., & Keyhani, N. O. (2007). Surface Characteristics of The Entomopathogenic Fungus *Beauveria* (Cordyceps) *Bassiana*. *Microbiology*, 153(10), 3448–3457.
- Hölker, U., Höfer, M., & Lenz, J. (2004). Biotechnological advantages of laboratory-scale solid-state fermentation with fungi. *Applied Microbiology and Biotechnology*, 64(2), 175–186.

- Hong, T. D., Gunn, J., Ellis, R. H., Jenkins, N. E., & Moore, D. (2001). The effect of storage environment on the longevity of conidia of *Beauveria bassiana*. *Mycological Research*, 105(5), 597–602.
- Hsieh, L. S., Tzean, S. S., & Wu, W. J. (1997). The genus *Akanthomyces* on spiders from Taiwan. *Mycologia*, 89(2), 319–324. <http://doi.org/Doi10.2307/3761088>
- Humphreys, A. M., Matewele, P., & Trinei, A. P. J. (1989). Effects Of Water Activity On Morphology, Growth And Blastospore Production Of *Metarhizium anisopliae*, *Beauveria bassiana* And *Paecilomyces farinosus* In Batch and Fed-Batch Culture. *Mycological Research*, 92(3), 257–264.
- Inch, J. M. M., Humphreys, A. M., Trinci, A. P. J., & Gillespie, A. T. (1986). Growth and Blastospore Formation by *Paecilomyces fumosoroseus*, A Pathogen Of Brown Planthopper (*Nilaparvata lugens*). *Transactions of the British Mycological Society*, 87(2), 215–222.
- Inglis, G. D., Goettel, M. S., Butt, T. M., & Stresser, H. (2001). Use of Hyphomycetous Fungi for Managing Insect Pests. In *Fungi as Biocontrol Agents* (pp. 23–69).
- Issaly, N., Chauveau, H., Aglevor, F., Fargues, J., & Durand, A. (2005). Influence of nutrient, pH and dissolved oxygen on the production of *Metarhizium flavoviride* Mf189 blastospores in submerged batch culture. *Process Biochemistry*, 40(3), 1425–1431.
- Ivy, C. C. H. (2011). Formulation of Mycoinsecticides as Water-Dispersible Granule for Vegetable Insect Pest Control. Universiti Putra Malaysia Unpublished.
- Jackson, M. A., Cliquet, S., & Iten, L. B. (2003). Media and Fermentation Processes for the Rapid Production of High Concentrations of Stable Blastospores of the Bioinsecticidal Fungus *Paecilomyces fumosoroseus*. *Biocontrol Science and Technology*, 13(1), 23–33.
- Jackson, M. A., Dunlap, C. A., & Jaronski, S. T. (2010). Ecological Considerations in Producing and Formulating Fungal Entomopathogens For Use In Insect Biocontrol. *BioControl*, (55), 129–145.
- Jackson, M. A., Erhan, S., & Poprawski, T. J. (2006). Influence of Formulation Additives on The Desiccation Tolerance And Storage Stability Of Blastospores Of The Entomopathogenic Fungus *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes). *Biocontrol Science and Technology*, 16(1), 61–75.
- Jackson, M. A., & Jaronski, S. T. (2009). Production of Microsclerotia Of The Fungal Entomopathogen *Metarhizium anisopliae* And Their Potential For Use As A Biocontrol Agent For Soil-Inhabiting Insects. *Mycological Research*, 113(Pt 8), 842–50.

- Jackson, M. A., McGuire, M. R., Lacey, L. A., & Wraight, S. P. (1997). Liquid Culture Production of Desiccation Tolerant Blastospores Of The Bioinsecticidal Fungus *Paecilomyces fumosoroseus*. *Mycology Research*, 101(1), 35–41.
- Jacobsen, B. J., & Backman, A. (1993). Biological And Cultural Plant Disease Controls : Alternatives And Supplements To Chemicals In IPM Systems. *Plant Disease*, 77(3), 311–315.
- Jaronski, S. T. (1997). New Paradigms in Formulating Mycoinsecticides. In *Pesticide Formulations and Application Systems: 17th Volume*. ASTM International.
- Jaronski, S. T. (2007). Mycosis Inhibits Grasshopper Necrophagy. *Entomological Society of America Miscellaneous Publications*. Paper No. 31456. Available.
- Jarrott, B. (1970). Uptake and Metabolism of Catecholamines In The Perfused Hearts Of Different Species. *Br J Pharmacol*, 38(4), 810–821.
- Jenkins, N. E., & Goettel, M. S. (1997). Methods for Mass-Production Of Microbial Control Agents Of Grasshoppers And Locusts. *Memoirs of the Entomological Society of Canada*, 129(171), 37–48.
- Jenkins, N. E., Thomasb, M. B., & Thomas, M. B. (1996). Effect of Formulation and Application Method on the Efficacy of Aerial and-Submerged Conidia of *Metarhizium flavoviride* for Locust and Grasshopper Control, 299–306.
- Jin, X., Streett, D. A., Dunlap, C. A., & Lyn, M. E. (2008). Application Of Hydrophilic – Lipophilic Balance (HLB) Number To Optimize A Compatible Non-Ionic Surfactant For Dried Aerial Conidia Of *Beauveria bassiana*, 46, 226–233.
- Jones, K. A., & Burges, H. D. (1998). Technology of formulation and application. In H. D. Burges (Ed.), *Formulation of Microbial Biopesticides* (pp. 7–30). Boston, Kluwer
- Kamarudin, N. H., Wahid, M. B., & Robinson, G. S. (1994). Common Bagworm Pests (Lepidoptera: Psychidae) Of Oil Palm in Malaysia With Notes To Related South-East Asian Species. *Malayan Nature Journal (Malaysia)*.
- Kamarudin, N., & Mohd Basri Wahid. (2010). Interactions Of The Bagworm, *Pteroma pendula* (Lepidoptera : Psychidae), And Its Natural Enemies In An Oil Palm Plantation In Perak. *Journal of Oil Palm Research*, 22(1999), 758–764.
- Kantz, B. (2007). Nature's Own Answer: Biopesticide Technology Leads The Charge Against Resistant Pests. *Florida Grower*, 100, 24.

- Kassa, A. (2003). Development and testing of mycoinsecticides based on submerged spores and aerial conidia of the entomopathogenic fungi *Beauveria bassiana* and *Metarhizium anisopliae* (Deuteromycotina: Hyphomycetes) for control of locusts, grasshoppers and storage pests. Doctoral Dissertation, Georg-August-University Göttingen.
- Khachatourians, G. G. (1991). Physiology and genetics of entomopathogenic fungi. *Handbook of Applied Mycology*, 2, 613–663.
- Khoo, K. C., Ooi, P. A. C., & Tuck, H. C. (1991). Lepidoptera. In *Crop Pests and Their Management in Malaysia*. Tropical Press.
- Kim, M., Chen, Y., Liu, Y., & Peng, X. (2005). Super-Stable, High-Quality Fe₃O₄ Dendron-Nanocrystals Dispersible in Both Organic and Aqueous Solutions. *Advanced Materials*, 17(11), 1429–1432.
- Kleespies, R. G., & Zimmermann, G. (1992). Production Of Blastospores By Three Strains Of *Metarhizium anisopliae* (Metch.) Sorokin In Submerged Culture. *Biocontrol Science and Technology*, 2(2), 127–135.
- Kleinkauf, H., & Von Dohren, H. (1987). Biosynthesis of Peptide Antibiotics. *Annual Reviews in Microbiology*, 41(1), 259–289.
- Knowles, A. (2005). New Developments in Crop Protection Product Formulation. *Agrow Reports UK: T and F Informa UK Ltd*.
- Knowles, A. (2008). Recent Developments of Safer Formulations Of Agrochemicals. In *Environmentalist* (Vol. 28, pp. 35–44).
- Krogman, K. C., Druffel, T., & Sunkara, M. K. (2005). Anti-Reflective Optical Coatings Incorporating Nanoparticles. *Nanotechnology*, 16(7), S338–S343.
- Kumar S.N., Nambisan B., Sundaresan A., *et al* (2013). Isolation and Identification of Antimicrobial Secondary Metabolites From *Bacillus Cereus* Associated With A Rhabditid Entomopathogenic Nematode *Ann. Microbiol.*, 64, pp. 209–218
- Lacey, L. A., & Kirk, A. A. (1993). Foreign Exploration for Natural Enemies Of *acc Tabaci* And Implementation in Integrated Control Programs In The United States. In *Proceedings Third International Conference on Pests in Agriculture*. (pp. 351–360). Association Nationale de Protection des Plantes.
- Lane, B. S., Trinci, A. P. J., & Gillespie, A. T. (1991). Endogenous Reserves and Survival of Blastospores Of *Beauveria bassiana* Harvested From Carbon-And Nitrogen-Limited Batch Cultures. *Mycological Research*, 95(7), 821–828.

- Larsson, M., Hill, A., & Duffy, J. (2012). Suspension Stability; Why Particle Size, Zeta Potential and Rheology are Important. Annual Transactions of The Nordic Rheology Society, 20, 209–214.
- Lindstrom, J. E., & Braddock, J. F. (2002). Biodegradation of Petroleum Hydrocarbons At Low Temperature In The Presence Of The Dispersant Corexit 9500. Marine Pollution Bulletin, 44(8), 739–747.
- Liu, Z., Wei, H., Li, Y., Li, S., Zhang, L., & Chen, H. (2011). Effects of Milling and Surfactants On Suspensibility And Spore Viability In *Paenibacillus Polymyxa* Powder Formulations. Biocontrol Science and Technology, 21(9), 1103–1116.
- Lomer, C. J., Bateman, R. P., Johnson, D. L., Langewald, J., & Thomas, M. (2001). Biological Control of Locusts and Grasshoppers. Annu. Rev. Entomol., 46, 667–702.
- Loong, C. Y. (2009). Bagworm (*Pteroma pendula* Joannis) And Its Control By Entomopathogenic Fungi, (*Metarhizium anisopliae* Metchnikoff And *Paecilomyces fumosoroseus* Wize). University Putra Malaysia.
- Loong, C. Y., Sajap, A. S., Noor, H. M., Omar, D., & Abood, F. (2010). Demography of The Bagworm, *Pteroma pendula* Joannis On An Exotic Tree, *Acacia mangium* Willd In Malaysia. The Malaysian Forester, 73(1), 77–85.
- Lozano-Contreras, M. G., Elías-Santos, M., Rivas-Morales, C., Luna-Olvera, A., Galán-Wong, L. J., & Maldonado-Blanco, M. G. (2007).
- Paecilomyces fumosoroseus* Blastospore Production Using Liquid Culture in A Bioreactor. African Journal of Biotechnology, 6(18), 2095–2099.
- Lumsden, R. D., Lewis, J. A., & Fravel, D. R. (1995). Formulation and Delivery of Biocontrol Agents for Use Against Soilborne Plant Pathogens. In F. R. H. and J.W. & Barry (Eds.), Biorational Pest Control Agents (pp. 166–182). Washington, DC: American Chemical Society.
- Luo, D.-Q., Liu, Z.-Q., Liu, T., Chen, C., Li, M.-Y., Wang, Z.-Y., Wang, X. (2015). Data in Support of Fumosorinone, A Novel PTP1B Inhibitor, Activates Insulin Signaling In Insulin-Resistance HepG2 Cells And Shows Anti-Diabetic Effect In Diabetic KKAy mice. Toxicology and Pharmacology, 285, 61–70.
- Magalhães, B. P., & Boucias, D. G. (2004). Effects of Drying on The Survival Of Conidiospores of *Metarhizium anisopliae* var. *acridum* Driver & Milner. Journal of Orthoptera Research, 13(1), 155–159.
- Mahadi, N. A., Muhamad, R., & Adam, N. A. (2012). Relationship between Bagworm *Pteroma pendula* Joannis (Lepidoptera: Psychidae) Populations, Parasitoids, and Weather Parameters in Oil Palm Plantation. Journal of Agricultural Science, 4(12), 13–17.

- Mahdi, E S, Sakeena, M H F, Abdulkarim, M F, Abdullah, G Z, Sattar, M A, Noor, A M (2011) Effect of Surfactant and Surfactant Blends on Pseudoternary Phase Diagram Behavior Of Newly Synthesized Palm Kernel Oil Esters. *Drug Design, Development and Therapy*, 5, 311-323
- Mascarin, G. M., Alves, S. B., & Lopes, R. B. (2010). Culture Media Selection for Mass Production of *Isaria fumosorosea* and *Isaria farinosa*. *Brazilian Archives of Biology and Technology*, 53(4), 753–761.
- Merkl, R., Hrádková, I., Filip, V., & Šmidrkal, J. (2010). Antimicrobial and Antioxidant Properties of Phenolic Acids Alkyl Esters. *Czech J. Food Sci*, 28(4), 275–279.
- Mesquita, A. L. M., Lacey, L. A., Mercadier, G., & Leclant, F. (1996). Entomopathogenic activity of a whitefly-derived isolate of *Paecilomyces fimosoroseus* (Deuteromycotina: Hyphomycetes) against Russian wheat aphid, *Diuraphis noxia* (Hemiptera: Sternorrhyncha) with the description of an effective bioassay method. *Eur J Entomology*, 93, 69–75.
- Miles, J. W., Goette, M. B., Grimm, L., & Pearce, G. W. (1959). Factors Affecting the Suspensibility of DDT Water-Dispersable Powders. *Bull. Wld Hlth Org.*, 20, 937–945.
- Minamidate, Y., Yin, S., & Sato, T. (2010). Synthesis and characterization of plate-like ceria particles for cosmetic application. *Materials Chemistry and Physics*, 123(2-3), 516–520.
- Mitsui, T., & Takada, S. (1969). On Factors Influencing Dispersibility and Wettability of Powder in Water. *Soci. Cosmetic Chemists*, 351(20), 335–351.
- Mohd Basri Wahid. (1993). Life History, Ecology And Economic Impact Of The Bagworm, *Metisa plana* Walker (Lepidoptera: Psychidae) On The Oil Palm, *Elaeis guineensis* Jacq. (Palmae), in Malaysia. University of Guelph.
- Moore, J. (2008). Partners Against Pests. *Florida Grower*, 101, 12–13.
- Moorer, H. H., & Sandefur, C. W. (1976). Process for Making Combination Wetting-Dispersing Agent. Patent No:5,968,808 New York. U.S. Patent and Trademark Office.
- Moubasher, H. (1993). Soil Fungi in Qatar and Other Arab Countries. *Economic Botany* (Vol. 50).
- Mukherjee, K. Raghu P. K. (1997) Effect of temperature on antagonistic and biocontrol potential of *Trichoderma* sp. on *Sclerotium rolfsii* *Mycopathologia*, 139 pp. 151–155
- Mulqueen, P. (2003). Recent Advances in Agrochemical Formulation. *Advances in Colloid and Interface Science*, 106, 83–107.

- Neves, P. M. O. J., Hirose, E., Tchujo, P. T., & Moino Jr, A. (2001). Compatibility of Entomopathogenic Fungi with Neonicotinoid Insecticides. *Neotropical Entomology*, 30(2), 263–268.
- Ordas, E. P. (1970). Wettable Powder Pesticide Concentrate. Patent No: 3,595,423 U.S. Patent Office.
- Osborne, L. S. (1990). Biological Control of Whiteflies and Other Pests with a Fungal Pathogen. Patent No: 4,942,030 Florida U.S. Patent Office.
- Pan, X., Zeng, F., & Fu, X. (1999). Research on The Relationship Between The Humic Substances Grades And Their Ability As CWM Dispersants. *Coal Conver*, 22, 38–42.
- Payne, C. C., Hafez, M., Jones, K., Harpaz, I., Akingbohunge, A. E., Paterson, R. R. M., & Ahmed, S. M. K. H. (1988). Pathogens for the Control of Insects: Where Next? *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, 318(1189), 225–248.
- Pekrul, S., & Gula, E. A. (1979). Mode of Infection Of The Corn Earworm (*Heliothis Zea*) By *Beauveria bassiana* As Revealed By Scanning Electron Microscopy. *Journal of Invertebrate Pathology*, 34(3), 238–247.
- Perdana Rozziansha, T. A., Sudharto, A. S., Chenon, R. D. de, Prasetyo, A. E., & Susanto, A. (2011). Organisme Pengganggu Tanaman *Pteroma pendula* Joannis. Inf PT, H - 0008,
- Pereira, R. M., & Roberts, D. W. (1991). Alginate and Cornstarch Mycelial Formulations of Entomopathogenic Fungi, *Beauveria bassiana* and *Metarhizium anisopliae*. *Journal of Economic Entomology*, 84(6), 1657–1661.
- Powell, K. A., & Jutsum, A. R. (1993). Technical and commercial aspects of biocontrol products. *Pesticide Science*, 37(4), 315–321.
- Purwar, J. P., & Sachan, G. C. (2006). Synergistic Effect Of Entomogenous Fungi On Some Insecticides Against Bihar Hairy Caterpillar *Spilarctia Obliqua* (Lepidoptera: Arctiidae). *Microbiological Research*, 161(1), 38–42.
- Quintela, E. D., & McCoy, C. W. (1998). Synergistic Effect Of Imidacloprid And Two Entomopathogenic Fungi On The Behavior And Survival Of Larvae Of *Diaprepes abbreviatus* (Coleoptera: Curculionidae) In Soil. *Journal of Economic Entomology*, 91(1), 110–122.
- Ramarethinam, S., Marimuthu, S., Murugesan, N. V, Loganathan, S., & Ramarethinam, S. (2000). Evaluation Of *Paecilomyces fumosoroseus*, An Entomopathogenic Fungus For Controlling The Red Spider Mite, *Oligonychus coffeae* (Nietner)(Acarina: Tetranychidae), Infesting Tea In India. *Pestology*, 24, 1–5.

- Reddy, N., & Yang, Y. (2010). Structure And Properties Of Ultrafine Silk Fibers Produced By *Theriodopteryx ephemeraeformis*. Journal of Materials Science, 45(24), 6617–6622.
- Reed, J. S. (1995). Principles of Ceramics Processing. New York: Wiley.
- Reichelderfer, K. (1984). Factors Affecting the Economic Feasibility of the Biological Control of Weeds. In E. S. Delfosse (Ed.), Proceedings IV International Symposium Biological Control of Weeds (pp. 19–25). Vancouver, Canada: Agric. Can.
- Rhainds, M., Davis, D. R., & Price, P. W. (2009). Bionomics of Bagworms (Lepidoptera: Psychidae). Annual Review of Entomology, 54, 209–26.
- Rogers, M. E., Dewdney, M. M., & Futch, S. H. (2009). 2014 Florida Citrus Pest Management Guide: Pesticides Registered for Use on Florida Citrus.
- Rombach, M. C. (1989). Production of *Beauveria bassiana* [Deuteromycotina. Hyphomycetes] Symptoduloconidia in Submerged Culture. BioControl, 34(1), 45–52.
- Sahayaraj, K., & Namasivayam, S. K. R. (2008). Mass Production of Entomopathogenic Fungi Using Agricultural Products and By Products. African Journal of Biotechnology, 7(12), 1907–1910.
- Sajap, A. S., Rozihawati, Z., Omar, D., & Lau, W. H. (2014). *Isaria fumosorosea* And *Metarhizium anisopliae* For Controlling *Atteva sciodoxa* (Lepidoptera: Yponomeutidae), A Pest of *Eurycoma longifolia*. Journal of Tropical Forest Science (Malaysia), 26(1), 84–91.
- Salim, H., & Hamid, N. H. (2007). Evaluation of Several Chemical Control Oil Palm Plantations. The Planter, 88(1040), 785–799.
- Salim, H., & Hamid, N. H. (2012). Evaluation of Several Chemical Control Approaches Against Bagworm, *Metisa plana* Walker (Lepidoptera: Psychidae) In FELDA Oil Palm Plantations. Planter, 88(1040), 785–799.
- Samish, M., Gindin, G., Alekseev, E., & Glazer, I. (2001). Developmental Fungi To Different of Entomopathogenic Pathogenicity Sanguineus (Acari : Ixodidae) Stages of Rhipicephalus. Journal of Parasitology, 87(6), 1355–1359.
- Samson, R. A. (1974). Notes On Entomogenous Fungi From Ghana.II. The Genus *Akanthomyces*. Acta Bot Neerl, 23(1), 28–35.
- Samson, R. A. (1974). *Paecilomyces* And Some Allied Hyphomycetes. Stud. Mycol., 6, 1–119.

- Samuels, R. I., & Paterson, I. C. (1995). Cuticle Degrading Proteases from Insect Moulting Fluid And Culture Filtrates Of Entomopathogenic Fungi. *Comparative Biochemistry and Physiology. Part B, Biochemistry & Molecular Biology*, 110(4), 661–9.
- Sanchez-Murillo, R. I., Torre-Martinez, M. de la, Aguire-Linares, J., & Harrera-Estrella, A. (2004). Light-Regulated Asexual Reproduction In *Paecilomyces fumosoroseus*. *Microbiology*, 150(2004), 311–319.
- Sandoval-Coronado, C. F., Luna-Olvera, H. A., Arevalo-Nino, K., Jackson, M. A., Poprowski, T. J., & Galan-Wong, L. J. (2001). Drying and Formulation Of Blastospores Of *Paecilomyces fumosoroseus* (Hyphomycetes) Produced In Two Different Liquid Media. *World Journal of Microbiology & Biotechnology*, 17, 423–428.
- Sarwar, M. (2015). Commonly Available Commercial Insecticide Formulations and Their Applications in the Field. *International Journal of Materials Chemistry and Physics*, 1(2), 116–123.
- Serebrov, V. V., Khodyrev, V. P., Gerber, O. N., & Tsvetkova, V. P. (2005). Perspectives of Combined Use Of Entomopathogenic Fungi And Chemical Insecticides Against Colorado Beetle (*Leptinotarsa decemlineata*). *Mikologiya I Fitopatologiya*, 39(3), 89–98.
- Shimizu, S., & Yamaji, M. (2002). Pathogenicity Of Entomopathogenic Fungi To The Termite, *Reticulitermes speratus*. *Japanese Journal of Applied Entomology and Zoology (Japan)*.
- Siti Ramlah, A. A., Norman, K., W, M., Basri, A, M., Najib, Kushairi, A. (2007). Sistem Pengurusan Perosak Bersepadu bagi Kawalan Ulat Bungkus di Ladang Sawit. MPOB Press.
- Smith, P. (1993). Control Of *Bemisia tabaci* And The Potential Of *Paecilomyces fumosoroseus* As A Biopesticide. *Biocontrol News and Information*, 14(4), 71N–78N.
- Starnes, R. L., Liu, C. L., & Marrone, P. G. (1993). History, Use, and Future of Microbial Insecticides. *American Entomologist*, 39(2), 83–91.
- Sterk, G., Bolckmans, K., & Eyal, J. (1996). A New Microbial Insecticide, *Paecilomyces fumosoroseus* Strain Apopka 97, For the Control of The Greenhouse Whitefly. In Brighton Crop Protection Conference: Pests & Diseases-1996: Volume 2: Proceedings of an International Conference, Brighton, UK, 18-21 November 1996 (pp. 461–466). British Crop Protection Council.
- Sudharto. (1991). Hama Tanaman Kelapa Sawit dan Cara Pengendaliannya.: Pusat Penelitian Perkebunan Marihat Permatang Siantar, Indonesia.

- Susanto, A., Prasetyo, A. E., Simanjuntak, D., Priwiratama, H., Sudharto, Chenon, R. D. de, Purba, R. Y. (2012). EWS - Ulat Api Ulat Kantung Ulat Bulu. Indonesian Oil Palm Research Institute.
- Thomas, M. B. (2000). 18 Development of a Mycoinsecticide for Biological Control of Locusts in Southern Africa. In R. A. Cheke, L. J. Rosenberg, & M. E. Kieser (Eds.), Workshop on Research Priorities for Migrant Pests of Agriculture in Southern Africa, Plant Protection Research Institute, Pretoria, South Africa, 24–26 March 1999. (pp. 173–182). Chatham, UK: Natural Resources Institute.
- Thomas, M. B., Wood, S. N., Langewald, J., & Lomer, C. J. (1997). Persistence of *Metarhizium flavoviride* and consequences for biological control of grasshoppers and locusts. *Pesticide Science*, 49(1), 47–55.
- Tigano-Milani, M. S., Honeycutt, R. J., Lacey, L. A., Assis, R., McClelland, M., & Sobral, B. W. (1995). Genetic Variability of *Paecilomyces fumosoroseus* Isolates Revealed by Molecular Markers. *Journal of Invertebrate Pathology*, 65(3), 274–82.
- Tseng, W. J., & Chen, C. (2003). Effect of Polymeric Dispersant on Rheological Behaviour Of Nickel Á Terpeneol Suspensions. *Materials Science and Engineering*, A347, 145–153.
- Tuck, H. C. (2002). Ecological Studies on *Pteroma pendula* Joannis and *Metisa plana* Walker (Lepidoptera: Psychidae) towards Improved Integrated Management of Infestation in Oil Palm. Universiti Putra Malaysia PhD Thesis.
- Vandenberg, J. D., Jackson, M. a, & Lacey, L. a. (1998). Relative Efficacy of Blastospores and Aerial Conidia of *Paecilomyces fumosoroseus* against the Russian Wheat Aphid. *Journal of Invertebrate Pathology*, 72, 181–183.
- Vandenberg, J. D., Sandvot, L. E., Jaronski, S. T., Jackson, M. A., & Halberts, S. E. (2001). Efficacy of Fungi for Control of Russian Wheat Aphid (Homoptera: Aphididae) In Irrigated Wheat. *Southwestern Entomologist*, 26(1), 73–85.
- Vanderberg, J. D., Ramos, M., & Altre, J. A. (1998). Dose-Response and Age- and Temperature-Related Susceptibility of the Diamondback Moth (Lepidoptera: Plutellidae) to Two Isolates of *Beauveria bassiana* (Hyphomycetes: Moniliaceae). *Environmental Entomology*, 27(4), 1017–1021.
- Vänninen, I., & Hokkanen, H. (1988). Effect of Pesticides on Four Species Of Entomopathogenic Fungi In Vitro. *Annales Agriculturae Fenniae*, 27(4), 345–353.
- Vanninen, I., Hokkanen, H., & Tyni-Juslin, J. (1999). Attempts to Control Cabbage Root Flies *Delia Radicum* L. And *Delia Floralis* (Fall.) (Dipt.,

- Anthomyiidae) With Entomopathogenic Fungi: Laboratory and Greenhouse Tests. *Journal of Applied Entomology*, 123(2), 107–113.
- Vega, F., Dowd, P., McGuire, M., Jackson, M., & Nelsen, T. (1997). In Vitro Effects of Secondary Plant Compounds on Germination of Blastospores of the Entomopathogenic Fungus *Paecilomyces fumosoroseus* (Deuteromycotina: Hyphomycetes). *Journal of Invertebrate Pathology*, 70(3), 209–13.
- Vega, F. E., Jackson, M. A., & McGuire, M. R. (1999a). Germination of conidia and blastospores of *Paecilomyces fumosoroseus* on the cuticle of the silverleaf whitefly, *Bemisia argentifolii*. *Mycopathologia*, 147, 33–35.
- Vega, F. E., Jackson, M. A., & McGuire, M. R. (1999b). Germination Of Conidia And Blastospores Of *Paecilomyces fumosoroseus* On The Cuticle Of The Silverleaf Whitefly, *Bemisia argentifolii*, 33–35.
- Vega, F. E., Posada, F., Catherine Aime, M., Pava-Ripoll, M., Infante, F., & Rehner, S. A. (2008). Entomopathogenic Fungal Endophytes. *Biological Control*, 46(1), 72–82.
- Vidal, C., Fargues, J., Lacey, L. A., & Jackson, M. A. (1998). Effect of Various Liquid Culture Media On Morphology, Growth, Propagule Production, And Pathogenic Activity To *Bemisia argentifolii* Of The Entomopathogenic Hyphomycete, *Paecilomyces fumosoroseus*. *Mycopathologia*, 143(1), 33–46.
- Villani, M. G., Krueger, S. R., & Nyrop, J. P. (1992). A Case Study of The Impact Of The Soil Environment On Insect/Pathogen Interactions: Scarabs In Turfgrass. In Anne R. Leslie (Ed.), *Handbook of Integrated Pest Management for Turf and Ornamentals*. Lewis Publisher.
- White, T., Bruns, T., Lee, S., & Taylor, J. (1990). Amplification and Direct Sequencing Of Fungal Ribosomal RNA Genes For Phylogenetics. In: *PCR Protocols: a guide to methods and applications*. Academic Press, 64, 315–322.
- Wood, B. J., Corley, R. H. V., & Goh, K. H. (1973). Studies on the effect of pest damage on oil palm yield. Communication, Chemara Research Station. Incorporated Society of Planters.
- Woodward, F. E., & Stefcik, A. (1967). United States Patent, (2), 6–10.
- Wraight, S., & Bradley, C. A. (1996). Production, Formulation, And Application Technologies for Use of Entomopathogenic Fungi To Control Field Crop Pests. Fifth Simp Sio de Controle Bi Logico, Anais: Confer Ncias Palestras. Embrapa-CNPSo, Foz Do Igua-Cu, Brazil, 177.
- Wraight, S. P., & Carruthers, R. I. (1999). Production, Delivery, And Use of Mycoinsecticides For Control of Insect Pests On Field Crops. In F. R. Hall & J. J. Menn (Eds.), *Methods in Biotechnology Vol 5 Biopesticides:*

Use and Delivery (pp. 233–269). Totowa. New Jersey: Humana Press Inc.

Wraight, S. P., Carruthers, R. I., Bradley, C. A., Jaronski, S. T., Lacey, L. A., Wood, P., & Galaini-Wraight, S. (1998). Pathogenicity Of The Entomopathogenic Fungi *Paecilomyces* spp. And *Beauveria Bassiana* Against the Silverleaf Whitefly, *Bemisia argentifolii*. *Journal of Invertebrate Pathology*, 71(3), 217–26.

Wright, M. S., William J., Connick, J. R., & Jackson, M. A. (2003). Use of *Paecilomyces* spp. as Pathogenic Agents Against Subterranean Termites. Patent No: 6,660,291 U.S. Patent Office.

Yang, D., Qiu, X., Zhou, M., & Lou, H. (2007). Properties of Sodium Lignosulfonate as Dispersant Of Coal Water Slurry. *Energy Conversion and Management*, 48(9), 2433–2438.

Yin, F. M. (1983). Storage of Conidial Powder of *Beauveria bassiana*. *Guangdong Forestry Newsletter* (in Chinese), 3/4, 13–16.

Yokoyama, T., Fujikata, M., & Fujiae, A. (1993). Improvement of Infectivity of *Metarhizium anisopliae* (Metchnikoff) to *Anomala cuprea* Hope (Coleoptera: Scarabaeidae) by Ultraviolet Irradiation of Protoplasts. *Applied Entomology and Zoology*, 28(4), 451–461.

Yu, W., France, D. M., Routbort, J. L., & Choi, S. U. S. (2008). Review and Comparison of Nanofluid Thermal Conductivity and Heat Transfer Enhancements. *Heat Transfer Engineering*, 29(5), 432–460.

Zimmermann, G. (2008). The Entomopathogenic Fungi *Isaria farinosa* (formerly *Paecilomyces farinosus*) and the *Isaria fumosorosea* species complex (formerly *Paecilomyces fumosoroseus*): Biology, Ecology and Use in Biological Control. *Biocontrol Science and Technology*, 18(9), 865–901.