

UNIVERSITI PUTRA MALAYSIA

CULTURE OF COPEPOD Oithona simplex (Farran, 1913) AND ITS POTENTIAL AS LIVE FOOD FOR FLOWER CRAB, Portunus pelagicus (Linnaeus, 1758) LARVAE

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NUR SYUHADA BINTI MAT NOOR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

May 2017



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Abstract of the thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

CULTURE OF COPEPOD Oithona simplex (Farran, 1913) AND ITS POTENTIAL AS LIVE FOOD FOR FLOWER CRAB, Portunus pelagicus (Linnaeus, 1758) LARVAE

By

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May 2017

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Research on *Oithona simplex* was carried out to further assess its potential as live food candidate for hatchery crab larval rearing. Although several species copepods from genus *Oithona* was proven to be good for fish larval rearing, no trial had ever been carried out using *O. simplex* during crab larval rearing and its life cycle development of this *O. simplex* has not been studied. All experiments were done at the International Institute of Aquaculture and Aquatic Sciences, Port Dickson and specimens were obtained from nearby coastal waters. The larval development of *O. simplex* was studied under laboratory condition of 30 °C and with 12 hours light and 12 hours dark photoperiod. The developmental stages were divided into 6 naupliar stages (NI, NII, NIII, NIV, NV, NVI), 5 copepodite stages (CI, CII, CIII, CIV, CV) and an adult (CVI) stage. The different larval stages of this species were described. The life cycle period lasted for 5 to 6 days.

Effect of some environmental parameters on the reproduction and development of *O. simplex* were also studied. This is to gather the optimum environmental condition for the culture of the species. Four different salinity levels viz. 20, 25, 30 and 35 ppt, three different temperatures of 25, 30 and 35 °C, three different photoperiods representing 12h light (L): 12h dark (D), 1h (L):23h (D), 24h (L):0h (D) and three light intensities of 5, 20, 35 µmol m⁻² s⁻¹ were employed in this study. *Oithona simplex* can tolerate a wide range of salinity of 20-35 ppt. The optimum temperature required for the maximum reproduction of *O. simplex* was 30 °C. The overall reproductive process was highest under the lowest light intensity (5 µmol m⁻² s⁻¹) compared to other light intensities. In addition, the overall reproduction and fastest development time were achieved by copepod reared under photoperiod 12h light (L): 12h dark (D) condition. The result of this study showed that *O. simplex* had a high population growth, short generation time, high reproductive potential, and a wide range of tolerance to environmental conditions.

Survival and specific growth rates of flower crab larvae from zoea 1 to megalopa stage were studied by feeding them with three different live food options that comprised of *O. simplex* alone, *Artemia* nauplii in combination with *O. simplex* and only *Artemia* nauplii. The highest survival was achieved by crab larvae fed with the combination of *O. simplex* and *Artemia* nauplii (3.16%), followed by those fed with *O. simplex* (1.31%) and *Artemia* nauplii (0.95%). Meanwhile, the highest specific growth rates were achieved by larvae fed with the combination of both live foods (28.4%) followed by *Artemia* nauplii only (27.2%) and only *O. simplex* (25.6%). No significant difference was found on the final length of crab larvae in those three different treatments. The result from this study showed that *O. simplex* meets the several requirements needed as live feed for crab larvae which indicated that it can be used to replace partially imported *Artemia* cyst during crab larval rearing.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENTERNAKAN KOPEPOD Oithona simplex (Farran, 1913) DAN POTENSINYA SEBAGAI MAKANAN HIDUP KEPADA LARVA KETAM BUNGA, Portunus pelagicus (Linnaeus, 1758)

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Penyelidikan ke atas *Oithona simplex* dijalankan untuk mengenal pasti potensinya sebagai calon makanan hidup untuk penternakan larva ketam. Walaupun beberapa spesis kopepod dari genus *Oithona* telah terbukti bagus untuk penternakan larva ikan, tiada percubaan yang pernah dijalankan menggunakan *O. simplex* untuk penternakan larva ketam dan perkembangan kitaran hayat spesis *O. simplex* ini belum pernah dikaji. Semua eksperimen telah dilakukan di Institut Antrabangsa Akuakultur dan Sains Akuatik, Port Dickson dan specimen diambil dari perairan pantai yang berdekatan. Perkembangan larva bagi *O. simplex* dikaji di bawah keadaan makmal pada suhu 30 °C dan 12 jam cahaya:12 jam gelap. Tahap perkembangan telah dibahagikan kepada 6 nauplius (NI, NII, NIII, NIV, NV, NVI), 5 peringkat kopepodit (CI, CII, CIII, CIV, CV) dan dewasa (CIV). Peringkat perbezaan larva bagi spesies ini diterangkan. Tempoh kitaran hidup mengambil masa selama 5 hingga 6 hari.

Kesan beberapa parameter persekitaran ke atas pembiakan dan perkembangan *O. simplex* juga turut dikaji. Ini adalah untuk mendapatkan faktor persekitaran yang optimum bagi penternakan untuk spesies ini. Empat tahap kemasinan berbeza iaitu 20, 25, 30 and 35 ppt, tiga suhu berbeza 25, 30 and 35 °C, tiga tahap jangka masa cahaya mewakili 12 jam cahaya:12 jam gelap, 1 jam cahaya:23 jam gelap, 24 jam cahaya:0 jam gelap dan tiga tahap keamatan cahaya 5, 20, 35 µmol m⁻² s⁻¹ telah digunakan dalam kajian ini. *Oithona simplex* boleh hidup dalam julat saliniti yang lebar iaitu 20-35 ppt. Suhu optimum yang diperlukan untuk pembiakan maksimum adalah 30 °C. Pembiakan keseluruhan adalah paling cepat di bawah keamatan cahaya yang rendah (5 µmol m⁻² s⁻¹) berbanding keamatan cahaya ujikaji yang lain. Sementara itu, masa keseluruhan pembiakan dan perkembangan terpantas telah dicapai oleh kopepod yang diternak dibawah jangka masa cahaya 12 jam cahaya: 12 jam gelap. Hasil kajian ini menunjukkan bahawa *O. simplex* mempunyai tahap pembesaran yang tinggi, selang masa generasi yang singkat, potensi pembiakkan yang tinggi dan ketahanan yang tinggi terhadap faktor persekitaran.

Kelansungan hidup dan kadar pertumbuhan spesifik larva ketam bunga dari zoea 1 ke peringkat megalopa dikaji dengan memberi pilihan makanan hidup yang berbeza: *O. simplex* sahaja, *Artemia* nauplius dengan gabungan *O. simplex* dan hanya *Artemia* nauplius. Kelangsungan hidup yang tinggi diperolehi oleh larva ketam yang diberi makan kombinasi *O. simplex* dan *Artemia* nauplius (3.16%), dikuti yang diberi makan *O. simplex* (1.31%) dan *Artemia* nauplius (0.95%). Sementara itu, kadar pertumbuhan spesifik dari segi berat badan yang paling tinggi telah diperolehi oleh larva ketam yang memakan gabungan kedua-dua makanan hidup (28.4%) diikuti yang memakan hanya *Artemia* nauplius (27.2%) dan hanya *O. simplex* (25.6%). Tiada perbezaan yang signifikasi didapati pada panjang akhir larva ketam dalam ketiga-tiga rawatan tersebut. Hasil daripada kajian ini menunjukkan bahawa *O. simplex* memenuhi beberapa syarat yang diperlukan sebahagian pengimportan sista *Artemia* semasa penternakan larva ketam.

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I certify that a Thesis Examination Committee has met on (2 May 2017) to conduct the final examination of Nur Syuhada binti Mat Noor on her thesis entitled "Culture of Copepod *Oithona simplex* (Farran, 1913) and its Potential as Live Food for Flower Crab, *Portunus pelagicus* (Linnaeus, 1758) Larvae" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

A1	Antennule
ARA	arachidonic acid
an	Antenna
CI - CV	copepodid I – V
CVI	adult
D	Dark
DHA	docosahexaenoic acid
EPA	eicosapentaenoic acid
FAME	fatty acid methyl esters
h	Hour
L	Light
ind	Individual
М	Megalopa
mnd	Mandible
min	Minute
MUFA	monounsaturated fatty acid
mx	Maxilla
NI - NVI	nauplius I – VI
ppm	parts per million
ppt	parts per thousand
PUFA	polyunsaturated fatty acid
SFA	saturated fatty acid
Z1 – Z4	zoea 1 – 4



CHAPTER 1

INTRODUCTION

Portunus pelagius known as a flower crab is one of the commercially important species for local consumption in Malaysia. However almost all landings are from the wild sources and there has not been much crab pond grow outs activities taking place in the country. In comparison to mud crab, flower crab aquaculture is almost nil at present. As the availability of the wild stocks is fluctuating and the practice of wild catching in the long run is unsustainable, the aquaculture development of flower crab must seriously be considered. Apart from that, flower crab culture operations also totally relying on seed collected from the wild, but now it is becoming more and dependent on hatcheries production which is a more dependable source for the future (Azra and Ikhwanuddin, 2015). Nowadays, the commercial development of crab culture, both mud crab and flower crab face some common problems mainly on live food cultures due to the difficulties in maintaining its culture, disease vector and high price of Artemia cyst (Allan and Fielder, 2004; Ouinitio et al., 2001). According to Ikhwanuddin et al. (2012a). investigation and modification of feeding during crab larvae rearing might be useful to improve survival rate of crab larvae. Therefore, research on live foods resources considerably important in establishing a better live food candidate for crab larval rearing. This is because; initial feeding at the early stages of development is one of the main components for successful larval rearing (Le Ruyet et al., 1993). In addition, most larvae would prefer feed that can be easily captured and digested, and also provides good nutrients for growth and survival (Giri et al., 2002). Thus, a great comprehension of larval morphology, behaviour, live food and artificial diet requirements, and environmental conditions are important to improve the techniques of larval rearing (Liao et al., 2001).

Most marine hatcheries are dependent on rotifers (Brachionus sp.) and brine shrimp (Artemia sp.) during the larval rearing period of fish and crustacean (Evjemo and Olsen, 1997: Hagiwara et al., 2001: Takeuchi, 2001). Artemia and rotifer are also commonly used as a live food for portunid crab larvae (Suprayudi et al., 2002; Soundarapandian et al., 2007; Baylon, 2009). Rotifers are comparatively easy to culture and it high density culture can also be reproduced rapidly (Lubzens et al., 2001). Despite the advantage, the culture of rotifer may sometime crash easily (Ananthi et al., 2011). As for brine shrimp, it is commercially available in the form of dry cysts. Recently, there has been some form of crisis in Artemia cyst availability due to insufficient supply to meet big demands by the aquaculture operators. The decreasing of harvested Artemia from the traditional productive Great Salt Lake, Utah USA, along with implementing a stricter rules of harvesting from those waters were also one of the cause of Artemia cyst availability crisis (Conceição et al., 2010). Other than that, even though Artemia is still the most preferred live food during larvicuture of fish and crustacean but this brine shrimp is not costeffective due to its high market price and the cost tends to increase from year to year following market trend (Akbary et al., 2010).

Marine copepods are the most numerous plankton that exist throughout the worlds's ocean and becoming the natural food sources for many marine fish and crustacean.

Copepods are common zooplankton of seawater, freshwater and brackishwater, that can represent up to 80% of the zooplankton biomass in the water column (Mauchline, 1998). The most commonly used species of free-living copepods in aquaculture belong to the three main orders which are Calanoida, Harpacticoida, and Cyclopoida (Støttrup, 2003). Copepods are abundant in the coastal waters of Malaysia and study by Rezai *et al.* (2004) stated that 177 copepod species belongs to 37 genera and 25 families are found in the Straits of Malacca. However, there is still lack of information and knowledge on these indigenous copepods especially on their potential role as a live food for fish and crustacean larvae. There are several advantages of using copepod as a live food such as improving larval survival (Shields *et al.*, 1999), increasing growth rate (Støttrup and Norsker, 1997), high contents of HUFA and has broad range of body size (Anathi *et al.*, 2011). Despite the advantage, there are still not many works regarding the application of copepod as live food for crustacean larvae. Thus, more research is needed to discover and assess the potential of copepod culture which can enhance the better growth and survivals of crustacean especially crab larvae.

Thus, the summary of the research problem in this study were stated below:

- 1) At present, the mariculture hatchery operation is completely dependent on imported and expensive *Artemia* cyst which is on the increase from year to year following market trend. Therefore, copepod has been chosen as an alternative live food to replace *Artemia* due to their unique characteristics, good nutritional content and readily available zooplankton in Malaysian coastal waters. By utilizing the naturally available live foods such as copepod to rear the crab larvae, feed costs can be significantly reduced.
- 2) Oithona simplex have been chosen in this study due to overall lack of biological information regarding this species especially on it's larval stages, life cycle reproduction and life development under environmental laboratory conditions. Lacking in information especially on their culture conditions could be one of the reason why mass culture technique is yet to be successful for hatchery copepod production. Thus, the information from this study would be vital for the mass culture production of this species.
- 3) The use of copepod as live food on the growth and survival of *P. pelagicus* larvae has not been well documented particularly for *Oithona simplex*. Thus, the finding of this study would contribute positively toward rearing of *P. pelagicus* larvae with locally abundant copepod. Higher survival rate of the larvae will pave the way for *P. pelagicus* aquaculture and this will indirectly reduce the dependent on wild catches from the sea and will also support the high growing demands of flower crab production in the country.

The overall aims of this study is to investigate in details about larval stages, culture conditions of *O. simplex* and to examine the use of *O. simplex* as alternative live food with an attempt to replace fully or partially the use of *Artemia* nauplii in crab larvae rearing. Thus, this study was established to address the following objectives:

- To investigate the different larval stages and life cycle of cyclopoid copepod O. simplex in the laboratory conditions.
- ➤ To examine the reproduction and development of *O. simplex* exposed to different sets of environmental laboratory conditions.
- > To determine the growth and percentage survival rate of crab larvae fed with copepod *O. simplex* and in combination with other live foods.



REFERENCES

- Abdullahi, B.A. (1992). Effects of the diet on growth and development of three species of cylopoids copepods. *Hydrobiologia*, 232:233-241.
- Akbary, P., Hosseini, S.A., Imanpoor, M., Sudagar, M., Makhdomi, N.M. (2010). Comparison between live food and artificial diet on survival rate, growth and body chemical composition of *Oncorhynchus mykiss* larvae. *Iranian Journal of Fisheries Sciences*, 9:19-32.
- Allan, G. and Fielder, D. (2004). Mud crab aquaculture in Australia and Southeast Asia. In Proceedings of the ACIAR Crab Aquaculture Scoping Study and Workshop, pp: 63-65, Bribbie Island.
- Alvarez-Lajonchere, L., Perez Sanchez, L., Hernandez Molenjon, O. G., Toress Gomes, E. (1996). Mass production of striped patao *Eugeress brasilianus* juvenile in Cuba. *Journal of the World Aquaculture Society*, 27:347-352.
- Anathi, P., Santhanam, P., Nandakumar, R., Ananth, S., Jothiraj, K., Dinesh kumar, S., Balaji Prasath, B., Jayalakshmi, T. (2011). Production and utilization of marine copepods as live feed for larval rearing of tiger shrimp *Penaeus monodon* with special emphasis on astaxanthin enhancement. *Indian Journal Natural Sciences*, 11:494-503.
- Andersen-Borg, C.M., Bruno, E., Kiørboe, T. (2012). The Kinematics of Swimming and Relocation Jumps in Copepod Nauplii. *Plos One*, 7(10): e47486. Doi:<u>10.1371/journal.pone.0047486</u>
- Azra, M. N., Wendy, W., Talpur, A. D., Abol-Munafi, A. B. and Ikhwanuddin, M. (2012). Effects of tank colourations on the survival, growth and development rate of flower crab, *Portunus pelagicus* (Linnaeus, 1758) larvae. *International Journal of Current Research and Review*, 4:117-123.
- Baylon, J. C. and Failaman, A. N. (1999). Larval rearing of the mud crab Scylla serrata in the Philippines. In Proceedings of Mud Crab Aquaculture and Biology Workshop, pp: 141-146, Australia.
- Baylon, J. C., Bravo, M. E. A., Maningo, N. C. (2004). Ingestion of *Brachionus* plicatilisand Artemia salina nauplii by mud crab Scylla serrate larvae. Aquaculture Research, 35:65-70.
- Baylon, J.C. (2009). Appropriate food type, feeding schedule and *Artemia* density for the zoea larvae of the mud crab, *Scylla tranquebarica* (Crustacea: Decapoda:Portunidae). *Aquaculture*, 288:190-195.
- Bell, J. G., McEvoy, L. A., Estevez, A., Shields, R.J., Sargent, J. R. (2003). Optimizing lipid nutrition in first feeding flatfish larvae. *Aquaculture*, 227: 211-220.

- Bengtson, D.A., Leger, P., Sorgeloos, P. (1991). Use of Artemia as a food source for aquaculture. In R. A. Browns., P. Sageloos., C.N.A. Trotman. (Eds.), Artemia Biology (pp. 255-285). Boca Raton, Florida: CRC Press Inc.
- Bligh, E. G. and Dyer, W. F. (1959). A rapid method of total lipid extraction and purification. *Canadian Journal Biochemical Physiology*, 37:911-917.
- Bottger-Schnack, R. (1988). Observations on the taxonomic composition and vertical distribution of cylopoid copepods in the central Red Sea. *Hydrobiologia*,47:311-318.
- Boxshall, G. A. and Halsey, S. H. (2003). *An introduction to copepod diversity*. UK: The Ray Society.
- Camus, T. and Zeng, C. (2008). Effects of photoperiod on egg production and hatching success, naupliar and copepodite development, adult sex ratio and life expectancy of the tropical calanoid copepod *Acartia sinjiensis*. *Aquaculture*, 280:220-226.
- Camus, T. (2012). The improvement of copepods intensive culture protocols as live feeds for aquaculture hatcheries. Ph.D. thesis. Australia: James Cook University.
- Chang, W. B. and Lei, C. H. (1993). Development and energy content of a brackish water copepod, *Apocylops royi* (Linberg) reared in laboratory. *Bulletin of the Institute of Zoology, Academia Sinica*, 32:62-81.
- Chinnery, F. E. and Williams, J. A. (2003). Photoperiod and temperature regulation of diapause egg production in *Acartia bifilosa* from Southampton Water. *Marine Ecology, Progress. Series*, 263:149-157.
- Chong, B.J. and Chua, T.E. (1973). A preliminary study on the distribution of the cyclopoid copepods of the family Oithonidae in the Malaysian waters. In proceedings of Pasific Science Association Marine Sciences Symposium, pp.32-36, Hong Kong.
- Christou, E.D. and Moraitou-Apostolopoulou, M. (1995). Metabolism and feeding of mesozooplankton of the eastern Mediterranean coast (Hellenic waters). *Marine Ecology Progress Series*, 126:39-48.
- Conceicãao, L.E.C., Yufera, M., Makridis, P., Morais, S., Dinis, M.T., (2010). Live feeds for early stages of fish rearing. *Aquaculture Research*, 41:613-640.
- Conway, D.V.P., White, R.G., Hugues-Dit-Ciles, J., Gallienne, C.P., Robin, D.B. (2003). Guide to the Coastal and Surface Zooplankton of the South-Western Indian Ocean. United Kingdom: Occasional Publication of the Marine Biology Association of the United Kingdom.
- Dahms, H.-U. (1993). Pictorial keys for the identification of crustacean nauplii from the marine meiobenthos. *Journal Crustacean Biology*, 13:609-616.

- Davis, J. A., Wille, M., Hecht, T. and Sorgeloos, P. (2005). Optimum time for weaning South AfricanScylla serrate (Forskal) larvae from rotifers to Artemia. Aquaculture International, 13:203-216.
- Devreker, D., Souissi, S., Seuront, L. (2005). Effects of chlorophyll concentration and temperature variation on the reproduction and survival of *Temora longicornis* (Copepoda: Calanoida) in the Eastern English Channel. *Journal of Experimental Marine Biology and Ecology*, 318:145-162.
- Drillet, G., Stephane, F., Mie, H.S., PerM, J., Jonas, K.H., Almagri, K.J., Benni, W.H. (2011). Status and recommendation on marine copepod cultivation for use as live feed. *Aquaculture*, 315:155-166.
- Dussart, B.H. and Defaye, D. (2001). Introduction to the Copepoda. Leiden: Backhuys.
- Edgar, G.J. (1990). Predator-prey interactions in seagrass beds. II. Distribution and diet of the blue manna crab*Portunus pelagicus* Linnaeus at Cliff Head, Western Ausralia. *Journal of Experimental Marine Biology and Ecology*, 139:23-32.
- Edward, J.C. (2005). Copepods as live prey: A review of factors that influence the feeding success of marine fish larvae. In L. Cheng-Sheng., J.O. Patricia., H.M. Nancy. (Ed.), *Copepods in Aquaculture* (pp. 133-149). UK: Blackwell Publishing.
- Evjemo, J. O. and Olsen, Y. (1997). Lipid and fatty acid content in cultivated live feed organisms compared to marine copepods. *Hydrobiologia*, 358:159-162.
- Farhadian, O. (2006). Culture of a planktonic cylopoid, Apocyclops dengizicus (Lepeshkin, 1900) and its suitability as live feed for the postlarvae of giant black tiger shrimp, Penaeus Monodon Fabricus (1798). Ph.D. thesis. Malaysia: Universiti Putra Malaysia.
- Farhadian, O, Md Yusoff, F. and Arshad, A. (2014). Effects of salinity, temperature, light intensity and light regimes on production, growth and reproduction parameters of *Apocyclops dengizicus*. *Irinian Journal of Fisheries Sciences*, 13(1):30-46.
- Folch, J., Lees, M., Sloanestanley, G. H. (1957). A simple method for the isolation and purification of total lipides from animal tissues. *Journal Biology of Chemistry*, 226 (1):497-509.
- Fujaya, Y., Trijuno, D.D., Nikhlani, A., Cahyono, I., Hasnidar (2014). The use of mulberry (Morus alba) extract in the mass production of flower crab (*Portunus pelagicus*) larvae to overcome the mortality rate due to molting syndrome. Aquatic Science and Technology, 2 (1):1-14.
- Gallienne, C. P. and Robin, D. B. (2001). Is *Oithona* the most important copepod in the world's oceans? *Journal of Plankton Research*, 23(12): 1421-1432.

- Geiling, W. T. and Campbell, R.S. (1972). The effect of temperature on development rate of the major life stages of *Diaptomus pallidus* Herrick. *Limnology and Oceanography*, 17:304-307.
- Gilbert, J. J. and C. E. Williamson. (1983). Sexual dimorphism in zooplankton (Copepoda, Cladocera, and Rotifera). Annual Review of Ecology, Evolutionand Systematics, 14:1-33.
- Giri, S. S., Sahoo, S. K., Shu, B. B., Sahu, A. K., Mohanty, S. N., Mohanty, P. K., Ayyapan, S. (2002). Larval survival and growth in *Wallago attu* (Bloch and Schneider): Effect of light, photoperiod and feeding regimes. *Aquaculture*, 213:157-161.
- Golez, M. S. N., Takashi, T., Ishimaru, T., Ohno, A. (2004). Post-embryonic development and reproduction of *Pseudodiaptamus annandalei* (Copepoda: Calanoida). *Plankton Biology Ecology*, 51 (1):15-25.
- Gonzalez-Felix, M. L., Gatlin III, D. M., Lawrence, A. L., Perez-Velasquez, M. (2002). Effect of dietry phospholipids on essential fatty acid requirements and tissue lipid composition of *Litopenaeus vannamei* juveniles. *Aquaculture*, 207:151-167.
- Goolish, R. and Burton, R. (1989). Energetics of osmoregulation in an intertidal copepod: effects of anoxia and lipid reserves on the pattern of free amino acid accumulation. *Functional Ecology*, 3:81–89.
- Gopakumar, G., Santhosi. I., Ramamurthy, N. (2009). Breeding and larviculture of the sapphire devil damselfish *Chrysiptera cynaea*. Journal Marine Biology Association of India, 52:130-136.
- Grice, G. D. (1960). Copepods of the genus *Oithona* from the Gulf of Mexico. *Bulletin of Marine Science*, 10 (4):485-490.
- Hamilton, R. D. (1979). Sterilization.In J.R. Stein (Ed.), Handbook of Phycological Methods, Culture Methods and Growth Measurements (pp448). London: Cambridge University Press.
- Hagiwara, A., Gallardo, W. G., Assavaaree, M., Kotani, T., Araujo, A. B. (2001). Live food production in Japan: recent progress and future aspects. *Aquaculture*, 200:111-127.
- Harris, R. (2001). Copepods. In: H.S. John., A.T. Steve., K.T. Karl (Ed.). Encyclopedia of ocean sciences, Academic Press, (pp. 512-523). Amsterdam: Elsevier Science Direct.
- Hernandez Molejon, O. G. and Alvarez-Lajonchere, L. (2003). Culture experiments with *Oithona aculata* Farran, 1913 (Copepoda: Cyclopoida), and its advantages as food for marine fish larvae. *Aquaculture*, 219:471-483.

- Hicks, G. R. F. and Coull, B. C. (1983). The ecology of marine meiobenthic harpacticoid copepods. *Oceanography and Marine Biology Annual Review*, 21:67-175.
- Hirche, H. J., Meyer, U., Niehoff, B. (1997). Egg production of *Calanus* finmarchicus effect of food, temperature and season. Marine Biology, 127:609-620.
- Hopcroft, R. R. and Roff. J. C. (1996). Zooplankton growth rates: diel egg production in the copepods *Oithona, Euterpina* and *Corycaeus* from tropical waters. *Journal of Plankton Research*, 18:789-803.
- Huys, R. and Boxshall, G.A. (1991). Copepod evolution. London: The Gresham Press.
- Iguchi N. and Tsujimoto, R. (1997). Seasonal changes in the copepod assemblage as food for larval anchovy in Toyama bay, southern Japan Sea. *Bulletin Japan Sea National Fisheries Research Institute*, 47:79-94. (in Japanese with English abstract)
- James, C. M. and Al-Kars, A. M. (1986). Studies on the production of planktonic copepods for aquaculture. In proceedings of the Second International Conference on Copepoda, pp. 333-340, Ottawa, Canada.
- Johan, I., Idris, B. G., Ismail, A. and Hishamudin, O. (2002). Distribution of planktonic calanoid copepods in the Straits of Malacca. In Proceedings of Second International Conference on the Straits of Malacca, pp. 393-408, Penang, Malaysia.
- Johan, I., Wan Maznah, W. O., Mashhor, M., Abu Hena, M. K., Amin, S. M. N. (2012). Spartial distribution of copepods along the salinity gradient of Perai River Estuary, Penang, Malaysia. *Pakistan Journal of Biological Sciences*, 15(13): 647-652.
- Jones, D. A., Kamarudin, M. S., Le Vay, L. (1993). The potential for replacement of live feeds in larval culture. *Journal of the World Aquaculture Society*, 24:199-210.
- Josileen, J. and Menon, N. G. (2004). Larval stages of the blue swimmer crab, *Portunus pelagicus* (Linnaeus, 1758) (Decapoda, Brachyura). *Crustaceana*, 77(7):785-803.
- Kanazawa, A. (2003). Nutrition of marine fish larvae. *Journal Applied Aquaculture*, 13:103-143.
- Koedijk, R. M., Folkvord, A., Foss, A., Pittman, K., Stefansson, S. O., Handeland, S., Imsland, A. K. (2010). The influence of first-feeding diet on the Atlantic cod *Gadus morhua* phenotype: survival, development and long term consequences for growth. *Journal of Fish Biology*, 77:1-19.

- Kolkovski, S., Creny, S., Yackey, C., Moreu, R., Dabrowski, K. (1998). The effect of vitamins C and E in n-3 HUFA enriched *Artemia* nauplii on growth, survival and stress resistance of walleye *Stizostedion vitreum* fry. *World Aquaculture*, 303-304.
- Ikhwanuddin, M., Shabidin, M.L. and Khairulhayadi, B. (2005). The development of seed production technology of flower crab, *Portunus pelagicus* in Sarawak. Proceeding of the Tropical Fish Aquaculutre Conference, pp. 1-2, Kuala Terengganu, Malaysia.
- Ikhwanuddin, M., Nor Adila, T., Azra, M. N., Hii, Y. S., Talpur, A. D., Abol-Munafi, A. B. (2011). Determination of live prey ingestion capability of blue swimming crab, *Portunus pelagicus* (Linnaeus, 1758) larvae. World Journal of fish and Marine Sciences, 3(6):570-575.
- Ikhwanuddin, M., Azra, M.N., Yeong, Y.S., Abol-Munafi, A.B., Shabdin, M.L. (2012a). Live foods for Juveniles' Production of Blue Swimming Crab, *Portunus pelagicus* (Linnaeus, 1766). Journal of Fisheries and Aquatic Science, 7:266-278.
- Ikhwanuddin, M., Azra, M. N., Redzuari, A., Aizam, Z. A., Abol-Munafi, A. B.(2012b). Ingestion rates of *Brachionus* sp. and *Artemia* sp. nauplii by crab, *Portunus pelagicus* (Linnaeus, 1758) larvae. *Journal of Fisheries and Aquatic Science*, 7(6):402-411.
- Ikhwanuddin, M., Talpur, A. D., Azra, M. N., Mohd Azlie, B., Hii, Y. S. and Abol-Munafi, A. B. (2012c). Effects of stocking density on the survival, growth and development rate of early stages blue swimming crab, *Portunus pelagicus* (Linnaeus, 1758) larvae. World Applied Science Journal, 18:379-384.
- Kailola, P. J., Williams, M. J., Stewart, P. C., Reichelt, R. E., McNee, A., Grieve, C. (1993). Australian Fisheries Resources. Bureau of Resources and the Fisheries Research and Development Corporation, Canberra, Australia.
- Lee. C.S., O'Bryen., P.J., Marcus, N.H. (2005). *Copepods in Aquaculture*. USA: Black well publishing.
- Le Ruyet, P. J., Alexandre, J. C., Thebaud, L., Mugnier, C. (1993). Marine fish larvae feeding: formulated diets of live preys?. *Journal of the World Aquaculture Society*, 24:211-224.
- Liao, I. C., Su, H. M., Chang, E.Y. (2001). Techniques in finfish larviculture in Taiwan. Aquaculture, 200:1-31.
- Lindley, L. C., Phelps, R. P., Davis, D. A., Cummins, K. A. (2011). Salinity acclimation and free amino acid enrichment of copepod nauplii for firstfeeding of larval marine fish. *Aquaculture*, 318:402-406.

- Liu, G. and Xu, D. (2009). Effects of calanoid copepod *Schmackeria poplesia* as live food on the growth, survival and fatty acid composition of larvae and juvenile of Japanese Flounder, *Paralichtys olivaceus*. *Journal of Ocean University of China*, 8:359-365.
- Lubzens, E., Zmora, O., Barr, Y. (2001). Biotechonoly and aquaculture of rotifers. *Hydrobiologia*, 446:337-353.
- Luis, E., Conceicao, C., Yuera, M., Makridis, P., Morais, S., Dinis, T. M. (2010). Live feeds for early stages of fish rearing. *Aquaculture Research*,41:613-640.
- Maheswarudu, G., Jose, J., Nair, K. R., Arputharaj, M. R., Ramakrishna, A., Vairamani, A. and Ramamoorthy, N. (2008). Evaluation of the seed production and gow out culture of blue swimming crab *Portunus pelagicus* (Linnaeus, 1758) in India. *Indian Journal of Marine Sciences*, 37:313-321.
- Mauchline, J. (1998). Advances in Marine Biology: The Biology of Calanoid Copepods. New York: Academic Press.
- Matias-Peralta, H., Yusoff, F. M., Shariff, M., Arshad, A. (2005). Effect of some environmental parameters on the reproduction and development of tropical marine harpacticoid copepod *Nitocra affinis* f. californica Lang. *Marine Pollution Bulletin*, 51:722-728.
- Matias-Peralta, H. M., Yusoff, F. M., Shariff, M., Suhaila, M. (2011). Small-scale continuous production of a tropical marine copepod, *Nitocra affinis* californica Lang and it potential as live food for aquaculture. *African Journal of Agricultural Research*, 6:1611-1620.
- Matias-Peralta, H. M., Yusoff, F. M., Shariff, M., Mohamed, S. (2012a). Reproductive performance, growth and development time of a tropical harpacticoid copepod, *Nitocra affinis* californica Lang, 1965 fed with different microalgae diets. *Aquaculture*, 344-349.
- Matias-Peralta, H. M., Yusoff, F. M., Shariff, M., Mohamed, S. (2012b). A tropical harpacticoid copepod, *Nitocra affinis* californica Lang as an effective live feed for black tiger shrimp larvae *Penaeus monodon* Fabricius.*Pertanika Journal Tropical Agriculture Science*, 35(4):695-710.
- McEvoy, L., Naess, T., Bell., J. G., Lie, O. (1998). Lipid and fatty acid composition of normal and malpigmented Atlantic halibut (*Hippoglossus hippoglossus*) fed enriched *Artemia*: a comparison with fry fed wild copepods. *Aquaculture*, 163:235-248.
- Milio, H. (1992). Effects of light (photoperiod, spectral composition) on the population dynamics of *Tisbe holothuriae* Humes (Copepoda: Harpacticoida). *Hydrobiologia*, 232:201-209.
- Moore, H. (1972). Aspects stress in the tropical marine environment. *Advances in Marine Biology*, 10:217-264.

- Moraito-Apostolopoulou, M., Verriopoulos, G., Dikaiakou, N. (1982). Influence of light condition on the offspring production and the sex ratio of *Tisbe holothuriae*, Humes (Copepoda: Haracticoida). Archivium fur Hydrobiologie, 96:120-127.
- Nadiah, W. N., Ikhwanuddin, M., Abol-Munafi, A. B. (2012). Remarks on the mating behaviour and success of flower crab, *Portunus pelagicus* (Linnaeus,1766) through the induction of limb autotomy technique. *Journal of Animal and Veterinary Advances*, 11(8):1149-1157.
- Nandakumar, R., Prasath, B. B., Santhanam, P., Ananth, S., Jayalakshmi, T., Kumar, S. D., Devi, A. S. (2015). Optimization of culture conditions for marine copepod Macrosetella grancilis (Dana, 1847) with emphasis on salinity and algal diets. *Indian Journal of Geo-Marine Sciences*, 44(10): 1521-1527.
- Nanton, D. A. and Castell, J. D. (1999). The effects of temperature and dietary fatty acids on the fatty acid composition of harpacticoid copepod for use as a live food for marine fish larvae. *Aquaculture*, 175:167-181.
- Neffati, N., Yahia-Kefi, O. D., Bonnet, D., Carlotti, F., Yahla, M. N. D. (2014). Reproductive traits of two calanoid copepods: *Centropages ponticus* and *Temora stylifera*, in autumn in Bizerte Channel. *Journal of Plankton Research*, 35:80-96.
- Nishida, S. (1985). Taxonomy and distribution of the family Oithonidae (Copepoda, Cyclopoida) in the Pacific and Indian Oceans. Bulletin of the Ocean Research Institute, University of Tokyo, 20:1-167.
- Omori, M and Ikeda, T. (1984). *Methods in Marine Zooplankton Ecology*. New York: Wiley.
- Otto, R. A., Jamieson, G. S., Boutillier, T., Zhuang, Z. M., Hong, S. V., Armstrong, D. A., Sekiguchi, H., Ivanov, B. G., Rodin, V., and Yeon, I. J. (2001). Commercial important crabs, shrimps and lobsters of North Pacific Ocean. *National Pacific Marine Science Organization (Pices)*, 19:1-74.
- Palanisamy, V., Latif, F. A., Resat, R.M. (1991). A Guide on the Production of Algal Culture for Use in Shrimp Hatcheries. Kuala Lumpur: Department of fisheries, Ministry of Agriculture, Malaysia.
- Peck, M. A. and Holste, L. (2006). Effects of salinity, photoperiod and adult stocking density on egg production and egg hatching success in *Acartia tonsa* (Calanoida: Copepoda): optimizing intensive cultures. *Aquaculture*, 255: 341–350.
- Perumal, P., Rajkumar, M., Santhanam, P. (2009). Biochemical composition of wild copepods, Acartia spinicauda and Oithona similis, from Parangipettai coastal waters in relation to environment parameters. Journal of Environmental Biology, 30:995-1005.

- Phelps, R. P., Sumiarsa, G. S., Lipman, E. E., Lan, H. P., Moss, K. K., Davis, A. D. (2005). Intensive and extensive production techniques to provide copepod nauplii for feeding larval Red Snapper *Lutjanus campechanus*. In C. S. Lee., P. J. O'Bryen., N. H. Marcus. (Eds.), *Copepods in Aquaculture*. (p. 151-168). Oxford: Blackwell Publishing.
- Pinto, C. S. C, Souzo-Santos, L. P., Santos, P. J. P. (2001). Development and population dynamics of *Tisbe biminiensis* (Copepoda: Harpacticoida) reared on different diets. *Aquaculture*, 198:253-267.
- Quinitio, E. T., Parado-Estepa, F. D., Millamena, O. M., Rodriguez, E., Borlongan. E. (2001). Seed production of mud crab Scylla serrate juveniles. Asian Fisheries Sciences, 14:161-174.
- Rainuzzo, J. R., Reitan, K. I., Olsen, Y. (1997). The significant of lipids at early stages of marine fish: a review. *Aquaculture*, 155:103-115.
- Rajion, M. A., McLean, J. G., Cahill, R. N. (1985). Essential fatty acids in the fetal and newborn lamb. *Australian Journal of Biological Sciences*, 38(1):33-40.
- Rajkumar, M. and Kumaraguru vasagam, K. P. (2006). Suitability of the copepod, *Acartia clausi* as a live feed for Seabass larvae (*Lates calcarifer* Bloch): Compared to traditional live-food organisms with special emphasis on the nutritional value. *Aquaculture*, 261:649–658.
- Ravi, R. and Manisseri, M. K. (2012). Survival rate and development period of the larvae of *Portunus pelagicus* (Decapoda, Brachyura, Portunidae) in relation to temperature and salinity. *Fisheries and Aquaculture Journal*, 49:1-7
- Ravi, R. and Manisseri, M. K. (2013). The effect of different pH and photoperiod regimens on the survival rate and developmental period of the larvae of *Portunus pelagicus* (Decapoda, Brachyura, Portunidae). *Iranian Journal of Fisheries Sciences*, 12:490-499.
- Redzuari, A., Azra, M. N., Abol-Munafi, A. B., Aizam, Z. A., Hii, Y. S., Ikhwanuddin, M. (2012). Effects of feeding regimes on survival, development and growth of flower crab, *Portunus pelagicus* (Linnaeus, 1758) larvae. *World Applied Sciences Journal*, 18(4):472-478.
- Rezai, H., Yusoff, F. M., Arshad, A., Kawamura, A., Nishida, S., Ross, O. (2004). Spatial and temporal distribution of copepods in the Straits of Malacca. *Zoological Studies*, 43:486-497.
- Ricker, W. E. (1979). Growth rates and models. In W.S. Hoar, D.J. Randall, J.R. Brett. (Ed.) *Fish physiology* (pp.677-743). New York: Academic Press.
- Sargent, J., McEvoy, L. A., Estevez, A., Bell, G., Bell, M., Henderson, J., Tocher, D. (1999). Lipid nutrition of marine fish during early development current status and future directions. *Aquaculture*, 179:217-229.

- Sampey, A., McKinnon, A. D., Meekan, M. G., McCormick, M. I. (2007). Glimpse into guts: overview of the feeding of larvae of tropical shore fishes. *Marine Ecology Progress Series*, 339:243-257.
- Santhanam, P., Perumal, P., Rajkumar, M. (2004). Effect of feeding Artemia on growth and survival of *P. monodon* larvae. Journal Applied Fisheries Aquaculture, 4(2):42-46.
- Santhanam, P. and Perumal, P. (2012). Evalution of the marine copepod Oihona rigida Giesbrecht as live feed for larviculture of Asian seabass Lates calcarifer Bloch with special reference to nutritional value. Indian Journal Fisheries, 59(2):127-134.
- Schipp, G. R., Bosmans, J.M.P., Marshall, A.J. (1999). A method for hatchery culture of tropical calanoid copepods, *Acartia* spp. *Aquaculture*, 174 (1999):81-88.
- Sergestrale, S.G. (1970). Light control of the reproductive cycle of *Pontoporeia affinis* Lindstrom (Crusteceana: Amphipoda). *Journal of Experimental Marine Biology and Ecology*, 5:272-275.
- Shansudin, L., Yusof, M., Azis, A., Shukri, Y. (1997). The potential of certain indigenous copepod species as live food for commercial fish larval rearing. *Aquaculture*, 151:351-357.
- Shields, R. J., Bell, J. G., Luizi, F. S., Gara, B., Bromage., N. R., Sargent, J. R. (1999). Naturals copepods are superior to enriched *Artemia* nauplii as feed for halibut larvae (*Hippoglosssus hippoglosssus*) in terms of survival, pigmentation and retinal morphology: relation to dietry essential fatty acids. *The Journal of Nutrition*, 129:1186-1194.
- Soundarapandian, P., Thamizhazhagan, E., Samuel, N.J. (2007). Seed production of commercially important flower crab *Portunus pelagicus* (Linnaeus). *Journal Fish Aquatic Science*, 2:302-309.
- Stephenson, W. (1972). An annotated check list and key to the Indo-West-Pacific swimming crabs (Crustacea:Decapoda:Portunidae). Bulletin of the Royal Society of New Zealand, 10:1-64.
- Støttrup, J. G. and Norsker, N. H. (1997). Production and use of copepods in marine fish larviculture. *Aquaculture*, 155:231-247.
- Støttrup, J. G. (2000). The elusive copepods: their production and suitability in marine aquaculture. *Aquaculture Research*, 31:703-711.
- Støttrup, J.G. (2003). Production and nutritional value of copepods. In J.G. Støttrup, J.G. and L. A. McEvoy. (Eds.), *Live Feeds in Marine Aquaculture* (p. 145-195). United Kingdom: Blackwell Science Ltd

- Stottrup, J. G. (2006). A Review on the status and progress in rearing copepods for marine larviculture. Advantages and disadvantages among Calanoid, Harpacticoid and Cyclopoid copepods. In: L.C.S. Elizabeth, Denis, D.R. Marie, M. T. Salazar, G. Martha, N. Lopez, D. A.V. Cavazos, A. C. Cruz, A.G. Ortega. (Eds.). Advances en NutricionAcuicola VIII (pp 62-83). Mexico: Universidad Autonoma de Nuevo Leon, Monterrey, Nuevo Leon.
- Su, H. M., Su, M. S. and Liao, C. I. (1997). Collection and culture of live foods for aquaculture in Taiwan. *Hydrobiologia*, 358:37-40.
- Sumpton, W. D., Potter, M. A., Smith, G. S. (1994). Reproduction and growth of the commercial sand crab, *Portunus pelagicus* (L.) in Moreton Bay, Queensland. *Asian Fisheries Science*, 7:103-113.
- Suprayudi, M. A., Takeuchi, T., Hamasaki, K., Hirikawa, J. (2002). Effects of *Artemia* feeding schedule and density on the survival and development of larval mud crab (Scylla serrate). *Fish Science*, 68: 1295-1303.
- Takashi, T. and Uchiyama, I. (2007). Morphology of the naupliar stages of some Oithona species (Copepoda:Cyclopoida) occurring in Toyama Bay, southern Japan Sea. Plankton Benthos Research, 2(1):12-27. Takeuchi, T., 2001. A review of feed development or early life stages of marine finfish in Japan. Aquaculture, 200:203-222.
- Talpur, A. D. and Ikhwanuddin, M. (2012). Effects of stress test on larvae of blue swimming crab, *Portunus pelagicus* (Linnaeus, 1758). Advances in Environmental Biology, 6:1909-1915
- Toledo, J. D., Golez, M. S., Doi, M., Ohno, A. (1999). Use of copepod nauplii during early feeding stage of grouper *Epinephelus coioides*. *Fish Science*, 65:390-397.
- Thach, N.C. (2009). Seed production and grow-out of mud crab (*Scylla paramamosain*) in Vietnam. Philippines: Southeast Asian Fisheries Development Center.
- Tompkins, J. (1995). Culture collection of algae and protozoa.In: M.M., Day and Turner, M.F. (Ed.), *Catalogue of Strains: Natural Evironment Research Council* (pp.144-173). United Kingdom: Titus Wilson and Sons Ltd, Kendal.
- Trijuno, D. D., Fujaya, Y., Agviranti., Marhama, S. (2015). Quality of Blue Swimming Crab *Portunus pelagicus* Larvae from Domesticated Broodstock. Aquaculture Indonesiana, 16(1):22-28.
- Walter, T. C. and Boxshall, G. (2014). World of Copepods database. Accessed at <u>http://www.marinespecies.org/aphia.php?p=taxdetails&id=106657</u> on 2016-05-04
- Walter, T. C. and Boxshall, G. (2016). World of Copepods database. Accessed at <u>http://www.marinespecies.org/copepoda</u> on 2016-05-03

- Watanabe, T., Kitajima, C., Fujita, S. (1983). Nutritional values of live organism used in Japan for mass propagation of fish: a review. *Aquaculture*, 34:115-143.
- Wetzel, R. G. (2001). *Limnology: Lake and River Ecosystems (3rd ed.)*. San Diego: Academic Press.
- Williams, M.J. (1982). Natural food and feeding in the commercial san crab *Portunus* pelagicus Linneaus, 1766 (Crustacea:Decapoda:Portunidae) in Moreton Bay, Quensland. Journal of Experimental Marine Biology and Ecology, 59:165-176.
- Yoshida, T., Matias-Peralta, H., Yusoff, F. M. D., Toda, T., Othman, B. H. R. (2012). Zooplankton research in Malaysia: Current status and future prospects. *Coastal Marine Science*, 35(1):208-213.
- Zaleha, K. and Busra, I. (2012). Culture of Harpacticoid Copepods: Understanding the Reproduction and Effect of Environmental Factors. In Z. Muchlisin (Ed.), *Aquaculture* (p 343-360). Croatia: InTech.