

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF BRASSINOLIDE AND ANTI-TRANSPIRANTS UNDER WATER STRESS CONDITIONS ON PLANT GROWTH AND PHYSICO-CHEMICAL CHANGES OF Musa acuminata COLLA cv. BERANGAN

MD AIMAN TAKRIM BIN ZAKARIA



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MD AIMAN TAKRIM BIN ZAKARIA

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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Faculty : Agriculture

Banana (Musa acuminata) cv. Berangan relatively sensitive to dry soil condition which may influence good performance of growth, physiology and biochemical changes of banana. Global water supply crisis and uneven distribution of rainfall are some of limiting factors in agriculture sector for production of banana under Malaysia climate condition. The study was aimed to investigate the plant growth, physiology and biochemical changes on yield of banana as influenced by brassinolide (BR) and minerals solely or in combination under water stress conditions. BR represents one of the plant growth regulators essential in multiple developmental processes in plants including cell division, cell elongation and also reproductive development. One month old banana seedlings were transplanted into the 15 cm × 15 cm size of polybag and placed under rain shelter. Banana seedlings were foliar sprayed with different concentrations of BR (0, 3, 6 and 12 gL⁻¹) for every two weeks intervals. The results showed that, BR concentration gave significant effects on the growth and physiology of banana plant (Musa acuminata ev. Berangan). As the BR concentrations increased from 0 to 6 gL⁻¹, plant height, pseudo-stem diameter, total leaf numbers, total leaf area, fresh and dry weight of shoot were markedly increased from week 3 to week 8 after transplanting. The BR-induced increase in chlorophyll content which contributed the increase in photosynthesis rate. The root size and distributions were, however, not significantly affected by BR. However, exogenous application of BR at 6.88 gL⁻¹ was the best concentration for banana plant at nursery stage as it was able to increase the plant height, pseudo-stem diameter, total leaf numbers and total leaf area of the species. Additionally, water stress or synonym referring to the drought season is the major abiotic stress which affect growth, physiology and biochemical activity in plant and cause major losses to agriculture production sector. The impacts of water stress in combination with exogenous application of plant growth regulator and minerals were also studied under rain shelter and open field plot. Under rain shelter experiment, one week old banana seedlings were transplanted in the polybag and the plants were grown under optimized concentration of BR (6.88 gL⁻¹) from previous experiment. The leaves of the whole banana seedlings were foliar sprayed with three treatments: (i) BR as control, (ii) CaCO₃ + MgCO₃ (1:1, v/v) and (iii) BR + CaCO₃ + MgCO₃ (1:1:1, v/v). The solutions for BR, CaCO₃ and MgCO₃ were prepared by dissolving 6.88 g, 3.96 g and 0.23 g into 1 Litre (L) of distilled water, respectively. The plants were also subjected to water stress treatments: 50%, 75% and 100% of the FC. Water stress at 50% FC had significantly reduced major growth parameters (plant height, pseudo-stem diameter and total leaf area) but enhanced accumulation of proline and malondialdehyde content in leaves tissue. Concurrently, two months old banana seedlings were transplanted at open field plot in Field 15, Faculty of Agriculture, UPM. The similar treatments [(i) 6.88 gL⁻¹ BR as control, (ii) CaCO₃ + MgCO₃ and (iii) BR + CaCO₃ + MgCO₃] were applied at every 2 weeks interval from 2 months after transplanting until flowering stage. Banana plants have been subjected to two different water regimes: (i) rain-fed and (ii) irrigated by micro-sprinkler system as control treatment for a continuous periods about 12 months. According to the results showed that, banana plants grown under rain-fed condition significantly reduced morphological characters such as plant height, pseudo-stem, canopy diameter, but enhanced accumulation of proline and malondialdehyde content in leaves tissue. Overall, the results proved that both conditions of water stress (50% FC) and rain-fed were significantly reduced major growth parameters (plant height, pseudo-stem diameter, total leaf area and canopy diameter), but enhanced accumulation of proline and malondialdehyde content in leaves tissue, respectively. These results suggest that application of BR + CaCO₃ + MgCO₃ on banana leaves increased vapour pressure deficit but reduced stomata conductance. Foliar application of BR with combination of CaCO₃ and MgCO₃ is recommended to regulate mechanism of drought adaptation to Berangan banana plant under field condition that facing global water supply criss and uneven distribution of rainfall, without negative effect on the harvested yield.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KESAN BRASSINOLIDE DAN ANTI-TRANSPIRAN DALAM KEADAAN KETEGASAN AIR TERHADAP PERTUMBUHAN TANAMAN DAN PERUBAHAN FIZIKO-KIMIA Musa acuminata COLLA ev. BERANGAN

Oleh

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Pisang (Musa acuminata) kultivar Berangan agak sensitif terhadap keadaan tanah kering yang mungkin mempengaruhi prestasi pertumbuhan, fisiologi dan perubahan biokimia pisang. Krisis bekalan air global dan taburan hujan tidak sekata adalah beberapa faktor yang menyekat dalam sektor pertanian untuk pengeluaran pisang di bawah keadaan iklim di Malaysia. Kajian ini bertujuan untuk mengkaji pertumbuhan tanaman, fisiologi dan perubahan biokimia pada hasil pisang seperti yang dipengaruhi oleh brassinolide (BR) dan mineral semata-mata atau gabungan dalam keadaan ketegasan air. BR mewakili salah satu pengawal atur tumbuhan yang penting dalam pelbagai proses pembangunan dalam tumbuhan termasuk pembahagian sel, pemanjangan sel dan juga perkembangan pembiakan. Anak pokok berusia satu bulan ditanam ke dalam 15 cm x 15 cm saiz polibeg dan diletakkan di bawah rumah kalis hujan. Anak-anak pokok disembur dengan kepekatan BR yang berbeza (0, 3, 6 dan 12 gL⁻¹) untuk setiap selang dua minggu. Keputusan menunjukkan bahawa kepekatan BR memberi kesan yang signifikan terhadap pertumbuhan dan fisiologi pokok pisang (Musa sp.) kultivar Berangan. Peningkatan BR dari 0 hingga 6 gL⁻¹ telah meningkatkan secara ketara terhadap ketinggian pokok, diameter pseudo-batang, jumlah jumlah daun, jumlah kawasan daun, berat pucuk segar dan berat pucuk kering dari minggu 3 hingga minggu 8 selepas pemindahan. Aplikasi BR meningkatkan kandungan klorofil yang menyumbang peningkatan kadar fotosintesis. Bagaimanapun, saiz dan pembahagian akar tidak memberi perbezaan yang ketara dengan rawatan BR. Walau bagaimanapun, aplikasi BR secara luaran dengan kadar 6.88 gL⁻¹ adalah merupakan kepekatan yang terbaik untuk pokok pisang di peringkat nurseri kerana ia dapat meningkatkan ketinggian pokok, diameter pseudobatang, jumlah daun dan jumlah keluasan daun. Selain itu, ketegasan air atau sinonim yang merujuk kepada musim kemarau adalah tekanan utama abiotik yang mempengaruhi pertumbuhan, fisiologi dan aktiviti biokimia dalam tanaman dan menyebabkan kerugian besar kepada sektor pengeluaran pertanian. Kesan ketegasan air dalam gabungan dengan pengawal atur tumbuhan dan mineral juga dikaji di bawah perlindungan hujan dan plot lapangan terbuka. Eksperimen di bawah rumah kalis hujan, anak pokok berusia satu

minggu dipindahkan ke dalam polibeg dan disembur dengan kepekatan BR yang optimum (6.88 gL⁻¹) daripada eksperimen sebelumnya. Keseluruhan daun anak pokok disembur dengan tiga rawatan berbeza untuk setiap selang dua minggu; (i) BR sebagai kawalan, (ii) CaCO₃ + MgCO₃ (1: 1, v/v) dan (iii) BR + CaCO₃ + MgCO₃ (1: 1: 1, v/v). Campuran untuk BR, CaCO₃ dan MgCO₃ disediakan dengan melarutkan 6.88 g, 3.96 g dan 0.23 g ke dalam 1 liter (L) air suling. Anak-anak pokok juga diberi rawatan ketegasan air: 50%, 75% dan 100% daripada kapasiti lapangan. Ketegasan air pada 50% kapasiti lapangan telah mengurangkan parameter pertumbuhan utama dengan ketara (ketinggian pokok, diameter pseudo-batang dan jumlah keluasan daun) tetapi pengumpulan kandungan proline dan malondialdehid meningkat dalam tisu daun. Pada masa yang sama, anak pokok pisang berusia dua bulan dipindahkan di plot lapangan terbuka di Ladang 15, Fakulti Pertanian, UPM. Rawatan yang sama [(i) 6.88 gL-1 BR sebagai kawalan, (ii) CaCO₃ + MgCO₃ dan (iii) BR + CaCO₃ + MgCO₃] digunakan pada setiap selang dua minggu bermula dua bulan selepas pemindahan sehingga tahap berbunga. Anak-anak pisang telah didedahkan kepada dua rejim air yang berbeza: (i) hanya bergantung kepada hujan dan (ii) pengairan oleh sistem pemercik mikro sebagai rawatan kawalan untuk tempoh berterusan kira-kira 12 bulan. Menurut hasil kajian menunjukkan, tanaman pisang yang ditanam di bawah keadaan bergantung hujan secara ketara mengurangkan ciri morfologi seperti ketinggian tumbuhan, pseudo-batang dan diameter kanopi, tetapi meningkatkan pengumpulan kandungan proline dan malondialdehid dalam tisu daun. Secara keseluruhannya, hasil kajian membuktikan bahawa kedua-dua keadaan ketegasan air (50% kapasiti lapangan) dan bergantung kepada hujan mengurangkan pertumbuhan utama secara ketara (ketinggian tumbuhan, diameter pseudo-batang, jumlah keluasan daun dan diameter kanopi), tetapi peningkatan pengumpulan proline dan kandungan malondialdehid dalam tisu daun, masing-masing. Hasil kajian menunjukkan bahawa penggunaan BR + CaCO₃ + MgCO₃ pada daun pisang meningkat defisit tekanan wap tetapi mengurangkan konduktiviti stomata. Aplikasi foliar BR dengan gabungan CaCO₃ dan MgCO₃ disarankan untuk mengatur mekanisme adaptasi kemarau kepada tanaman pisang Berangan di bawah keadaan lapangan yang meghadapi krisis bekalan air global dan taburan hujan yang tidak rata, tanpa memberi kesan negatif terhadap hasil tuaian.

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I certify that a Thesis Examination Committee has met on 13 August 2018 to conduct the final examination of Md Aiman Takrim bin Zakaria on his thesis entitled "Effects of Brassinolide and Anti-Transpirants under Water Stress Conditions on Plant Growth and Physico-Chemical Changes of *Musa acuminata* Colla cv. Berangan" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

% Percentage
< Less than
> Greater than

* Significantly difference at P<0.05
 ** Significantly difference at P<0.01
 *** Significantly difference at P<0.001

ANOVA Analysis of Varience

BR Brassinolide C Carbon

CEC Cation Exchange Capacity

Carbon Dioxide CO_2 Centimetre cm cv. Cultivar °C Degree Celcius Chl a Chlorophyll a Chl b Chlorophyll b $Chl_{a\; +b}$ Total Chlorophyll CaCO₃ Calcium Carbonate

DW Dry Weight
et al. And Friends
EL Electrolyte Leakage

FAOSTAT Food and Agriculture Organization Statistical Database

FC Field Capacity
FW Fresh Weight

g Gram

gL⁻¹ Gram Per Litre gcm⁻¹ Gram Per Centimetre

ha Hectare H₂O Water

IAA Indole-3-Acetic Acid

kg Kilogram kPa Kilo Pascal L Litre LA Leaf Area

LSD Least Significant Difference

LAI Leaf Area Index

m Metre

MDA Malonyldialdehyde

 $\begin{array}{lll} \mu g & Microgram \\ \mu mol & Micromole \\ mL & Millitre \\ mm & Millimetre \\ mmol & Millimole \end{array}$

MgCO₃ Magnesium carbonate

NPK Nitrogen Phosphorus Potassium

n Number of Respondence

ns Not Significant

pH Measurement of Acidity / Alkalinity

RWC Relative Water Content

RCBD Randomized Complete Block Design

second

SAS

TW

Statistical Analysis System
Turgid Weight
Universiti Putra Malaysia
United States of America
Volume per Volume UPM USA V/VVPD Vapour Pressure Deficit

Versus vs.

WAT Week After Transplanting



CHAPTER 1

INTRODUCTION

Banana (Musa acuminata) belong to the family Musaceae has a lot of potential as a sustainable crop with multi-uses product. Global water supply crisis and uneven distribution of rainfall are some of limiting factors in agriculture sector for production of banana under Malaysia climate condition. The most important source of natural water for crop growth is through rainfall. When rainfall is insufficient, thus irrigation system need be installed to guarantee a good harvest. The changes and uneven distribution of rainfall in Malaysia fluctuated heavily from -30% to +30% coupled with the rise of temperature by 0.3°C to 4.5°C and rise in sea level is expected to be about 95 cm over a hundred years (Alam et al., 2011). However, this crop is very sensitive to the climate change and may reduce banana growth performance (Ranjitkar et. al., 2016). Malaysia is one of the centers of origin for many banana varieties in the world (Siti Hawa, 1992) and cv. Berangan is the most popular one. Banana has been planted either in large or small scale for commercial in Malaysia. There were 27, 296 ha planted areas of banana in Malaysia with total production of 343, 061 metric tonnes (DOA, 2015). The largest world producers are Latin America and Carribean. Latin America and Carribean exports banana about 13.4 million tonnes in year 2013. Whereas in Asia, India is the largest producer and the Philipines is the largest exporter. In 2013, Asia region exports banana about 2.9 million tonnes (FOASTAT, 2015).

According to Lascano et al. (2007), it is expected that human population in the world will be increased 8 to 9 billion by the year 2025, meanwhile the demand or consumption of the food will be higher than present. Global issues concerning now on the climate change and food security due to human population growth and stagnating productivity of food. Other than our concent on the food productivity, there is evidence that more than 40% of the people in our planet are affected by water scarcity. By the year 2025, about 1.8 billion people will be living in the countries with absolutely lack of water resources (Lascano et al., 2007). There is also an evidence that two-third of the world's population suffered from water stress (Forouzani et al., 2012). Impact of the water stress is closely related to the fenomena of drought which is an outcome of the climatic changes that directly affected productivity of the plant, thus reducing yield in many region around the world (Riccardi et al., 2016). In Malaysia, adaptation to climate change is a great challenges to sustain agricultural productivity and attain food security (Alam et al., 2011). Forouzani et al. (2012) reported that management of water resources in a sustainable way is very important for growing population as well as the need more of food production drives the agricultural sector to manage availabl usage properly.

Banana plant requires uniformly warm and moist conditions for optimum growth and yield production (Razi et al., 2004). Tropical environments with unfavourable water stress condition may lead to low survival rate of banana plants since they are very sensitive to dry soil as well as various environmental changes. Water is one the most

important element for plant to grow well especially for transporting important nutrients through the plant. Zingaretti et al. (2011) reported that during vegetative stage growth, water is essentially required by plant to obtain maximum yield, but inadequate water uptake in this stage may reduce crop productivity.

Applications of plant hormone as well as foliar spray of mineral as anti-transpirant are possible strategies for improving adaptability of plant with climate change. Plant hormone widely used in modern agriculture at low dosages and typically applied via foliar sprays with water as a carrier that affect developmental or metabolic processes in higher plants such as banana (Rademacher, 2015). Brassinolide (BR) is one of the phytohormones essential for plant growth and development as well as important for cell division and expansion which may increase crop yield and able to alleviate various biotic and abiotic stress (Sasse, 2003; Jager et al., 2008). According to Ingram and Bartels (1996), stated that either by endogenous molecular systems or exogenous applied compounds able to protect the plant cell from severely damage and to mitigate the plant from environmental stress. Exogenous application of BR also has been reported influences growth and development processes as well as mitigates the abiotic stresses of tomato plants (Montoya et al., 2005).

Through understanding of physiological responses of plant to water stress and mechanism water use efficiency, foliar application with minerals such as magnesium carbonate (MgCO₃) and calcium carbonate (CaCO₃) as anti-transpirant on upper and lower part of the leaves able to enhance photosynthetic efficiency. Carmen et al. (2014a), noted that the application of lithovit as foliar fertilizer consist of calcium carbonate on the leaf surface of tomato plant resulted in highest photosynthesis intensity. In the modern agriculture, improving the plant growth performance and crop productivity by using anti-transpirant materials potentially reduce transpiration rate and increasing turgidity of leaves as well as stomata guard cell for increasing water stress use efficiency and water stress resistance (Davenport et al., 1972).

Therefore, the study was aimed to investigate the plant growth performance, physiology, biochemical changes and yield of Berangan banana as influenced by BR and minerals under water stress conditions. The general objectives of the study were:

- (1) To identify banana seedlings response toward usage of plant hormone and mineral of anti-transpirants with different concentrations on morphological and physiological changes.
- (2) To observe morphological and physiological response biochemical changes of banana seedlings as influenced by plant hormone and anti-transpirants under water stress conditions.
- (3) To analyse biochemical changes as affected by plant hormone, anti-transpirants and water stress conditions on yield production of banana.

REFERENCES

- Abd El Ghafar, M.S., Al-Abd M.T., Helaly A.A. & Rashwan A.M. (2016). Foliar application of lithovit and rose water as factor for increasing onion seed production. *Journal of Natural and Science*, 14(3): 53 61.
- Abd El-Kader, A.M., Saleh, M.M.S. & Ali, M.A. (2006). Effect of soil moisture levels and some antitranspirants on vegetative growth, leaf mineral content, yield and fruit quality of Williams banana plants. *Journal of Applied Sciences Research*, 2(12): 1248-1255.
- Abdel, R. (2014). Studies on yield and quality of broccoli (*Brassica oleraceae* L.) under Assiut conditions. M.Sc. thesis in Faculty of Agriculture in Assiut, Al-Azhar University.
- Abo-Sedera, F.A., Shams A.S., Mohamed M.H.M. & Hamoda A.H.M. (2016). Effect of organic fertilizer and foliar spray with some safety compounds on growth and productivity of snap bean. *Annals of Agricultural Science Moshtohor Journal*, 54 (1): 105 118.
- Abou-Shleell, M.K.K. (2017). Botanical studies on moringa plant: Ph.D. Thesis, Faculty of Agriculture, Moshtohor, Benha University, Egypt.
- Agrawal, A., Swennen, R. & Panis, B. (2004). A comparison of four methods for cryopreservation of meristems in banana (*Musa* Spp.). *CryoLetters*, 25(2):101-110.
- Akynci, S. & Losel, D.M. (2012). Plant water-Stress response mechanisms. In: Rahman, M.M., Hasegawa, H. (Eds.), Water Stress. In Tech, Croatia, pp. 15-42.
- Alam, M.M., Siwar, C., Murad, M.W. & Mohd Ekhwan, T. (2011). Impacts of Climate Change on Agriculture and Food Security Issues in Malaysia: An Empirical Study on Farm Level Assessment, *World Applied Sciences Journal* 14(3): 431-442.
- Aldrich, W.W. & Work, R.A. (1934). Evaporating power of the air and top-root ratio in relation to rate of pear fruit enlargement. *Proceeding of the American Society for Horticulture Science*, 32: 115-123.
- Ali, H.M. (2005). Effects of rice straw compost and water regimes on growth performance of tomatoes (*Lycopersicum esculentum* L.). PhD Thesis, Universiti Putra Malaysia, Serdang pp: 145.
- Alkarkhi, A.F.M., Ramli, S., Yeoh, S.Y. & Easa, A.M. (2010). Physiochemical properties of banana peel flour as influenced by variety and stage of ripeness: multivariate statistical analysis. *Asian Journal of Food Agro-Industry*, 3:349-362.

- Allahmoradi P., Ghobadi M., Taherabadi S. & Taherabadi S. (2011). Physiological Aspects of Mungbean in Response to Drought Stress. IPCBEE vol.9 (2011) © (2011) IACSIT Press, Singapoore Internation Conference Food England Biotechnology.
- Aloni, B., Daie, J. & Karni, L. (1991). Water relations, photosynthesis and assimilate partitioning in leaves of pepper (*Capsicum annuum*) transplant: Effect of water stress after transplanting. *Horticulture Science*, 66: 75-85.
- Anda, M., Shamshuddin, J., Fauziah, C.I. & Syed Omar, S.R. (2010). Increasing the organic matter content of an Oxisol using rice husk compost: changes in the composition and its chemistry. *Soil Science Society of America Journal*, 74: 1167-1180.
- Anonymous, (2013). Banana cultivation guide. Retrieved from http://www.bananaplanters.com/site/banana-cultivation-guide/.
- Arias, P. (2003). The world banana economy, 1985-2002 (Vol. 1): Food and Agriculture Organization.
- Asharaf, M. & Mahmood, S. (1990). Response of four Brassica species to drought stress. *Experimental Botany*, 30: 93-100.
- Awad, M.H., Razi, M.I, Marziah, M., Mahmud, T.M. & Hassan, I. (2004). Effect of water deficit on growth and leaf gas exchange of pepper plants (*Capsicum annuum* L.) In: Environment Stress Impact on Tropical Crops, eds. Razi M.I and W.J. Davies, pp 48-63. Selangor: Jofri Publishing House.
- Ayob, K. (1986). Effect of available soil moisture on the yield of chilli (*Capsicum annuum*). *Techonlogy Sayur Sayuran*, 2: 57-59.
- Bafeel S.O. & Moftah A.E. (2008) Physiological response of eggplants grown under different irrigation regimes to antitranspirant treatments. *Saudi Journal of Biological Sciences*, 15: 259-267.
- Baher, Z.F., Mirza, M., Ghorbanli, M. & Bagher, R.M. (2002). The influence of water stress on plant height, herbal and essential oil yield and composition in Satureja hortensis L. *Flavour and Fragrance Journal*, 17(4), 275-277.
- Bao F., Shen J., Brady S.R., Muday G.K., Asami T. & Yang Z. (2004). Brassinosteroids interact with auxin to promote lateral root development in *A. thaliana. Plant Physiology*, 134: 1-8.
- Bastiah, A. (1999). Effect of rootstock on growth and water efficiency of Hevea during water stress. *Journal Rubber Research* 2(2):99-119.
- Bates, L.S., Waldren, R.P. & Teare, I.D. (1973). Rapid. Determination of free proline in water stress studies. *Plant Soil*, 39: 205-208.

- Begg, J.E. & Turner, N.C. (1976). Crop water deficits. Advance Agronomy, 28: 161-217.
- Bhatt, R.M. & Srinivasa Rao, N.K. (2005). Influence of pod load response of okra to water stress. *Indian Journal Plant Physiology*, 10: 54–59.
- Black, G.R. & Hartge, K.H. (1986). Bulk density. In: A. Klute (Ed.). Methods of Soil Analysis. Part I. Physical and Mineralogical Methods. 2^{nd.} Ed., Agronomy No. 9 (part I). ASA-SSSA. Madison, Wisconsin, USA, 363-375.
- Blokhina, O., Virolainen, E. & Fagerstedt, K.V. (2003). Antioxidants, oxidative damage and oxygen deprivation stress. *Annual Botany*, 91: 179-194.
- Blomme, G., Dita, M, Jacobsen, K.M, Vicente, L.P, Molina, A., Ocemati, W., Poussier, S., & Prior, P. (2017). Bacterial diseases of bananas and enset: current state of knowledge and integrated approaches toward sustainable management. *Frontier in Plant Science*, 8: 1290.
- Bora, K.K, & Mathur, S.R. (1998). Some plant growth regulator as antitranspirants in soybean. Ann. *Plant Physiology*, 12: 175-177.
- Boyer, J.S. (1968). Relationship of water potential to the growth of leaves. *Plant Physiology*, 43:1056-1062.
- Boyer, J.S. (1982a). Plant productivity and environment. Science, 218 (4571):443-448.
- Boyer, J.S. (1982b). Physiological adaptations to water stress. In: Adaptation of plants water stress and high temperature stress. Eds. Turner N.C., Kramer P.J., pp. 43-45
- Bray, R.H., & Kurtz, L.T. (1945). Determination of organic and available forms of phosphorus in soils. *Soil Science*, 59: 39-45.
- Briggs, L.J. & Shantz, H.L. (1912). The wilting coefficient for different plants and its indirect determination. United State Department Agriculture.230.
- Carbonell, M.V., Florez, M., Martinez, M.E., Maqueda, R. & Amaya, J.M. (2011). Study of stationary magnetic fields on initial growth of pea (*Pisum sativum L.*) seeds. *Seed Science and Technology*, 39(3): 673-679.
- Carmen, B., Radu S., Stefania, G. & Sorin, V. (2014a). Physiological Indicators Study Involved in Productivity Increasing in Tomato. *ProEnvironment*, 7: 218 224.
- Carmen, B., Sumalan, R., Gadea, S. & Vatca, S. (2014b). Physiological Indicators Study Involved in Productivity Increasing in Tomato. *ProEnvironment*, 10 (7):218 224.
- Carmi, A. & Heuer, B. (1981). The role of roots in control of bean shoot growth. *Annals of Botany*, 48: 519-527.

- Catuchi, T.A., Vitolo, H.F., Bertolli, S.S., & Souza, G.M. (2011). Tolerance to water deficiency between two soybean cultivars: transgenic *versus* conventional. *Ciencia Rural, Santa Maria*, 31: 373-378.
- Chapin, F.S. (1980). The mineral nutrition of wild plants. *Annals Ecology System*, 11: 233-260.
- Chapman, H.D. (1965). Determination of cation exchange capacity. In C.A.Black (Eds), Methods of soil analysis. *Agronomy Mongor*, 2(1): 891-900.
- Chapman, K.R. (1973). Effect of water regimes in performance of glasshouse-grown nursery apples trees. *Queensland Journal of Agriculture and Animal Science*, 30:125-135.
- Chaves, M.M. (1991). Effect of water deficits on carbon assimilation. *Journal of Experimental Botany*, 42: 1-16.
- Chaves, M.M., Pereira, J.S., Maroco, J., Rodrigues, M.L., Ricardo, C.P.P. & Osorio, M.L. (2002). How plants cope with water stress in the field: photosynthesis and growth. *Annuals of Botany*, 89: 907-916.
- Clifton-Brown, J.C. & Lewandowski, I. (2000). Water use efficiency and biomass partitioning of three different Miscanthus genotypes with limited and unlimited water supply. *Annals of Botany*, 86: 191-200.
- Clouse, S.D., & Sasse, J.M. (1998). Brassinosteroids: essential regulators of plant growth and development. *Annual Review Plant Physiology Plant Molecular Biology*, 49:427–451.
- Coombs, J., Hind, G., Leegood, R.C., Tieszen, L.L. & Vonshak, A. (1985). Analytical Techniques. In: Techniques in Bioproductivity and photosynthesis 2nd edition. (Eds.) J. Coombs, D.O. Hall, S.P. Long and J.M.O. Scurlock. Pergamon Press. pp. 219-220.
- Cornic, G. & Massacci, A. (1996). Leaf photosynthesis under drought stress. In: Baker, N.R. (ed.). Photosynthesis and the environment. Kluwer Academic Publishers, pp. 347-366.
- Cornic, G. & Briantais, J.M. (1991). Partitioning of photosynthetic electron flow between CO₂ and O₂ reduction in a C₃ leaf (*Phaseolus vulgaris* L.) at different CO₂ concentrations and during drought stress. *Planta*, 183: 178-184.
- Cornic, G. (1994). Drought stress and high light effects on leaf photosynthesis. In: Photoinhibition of photosynthesis: from molecular mechanisms to the field, eds. Baker NR, Bowyer JR, Oxford: BIOS Scientific Publishers.
- Costa, R.C.L., Lobato, A.K.S., Oliveira, Neto, C.F.., SantosFilho, B.G., Cruz, F.J.R., & Laughinghouse, IV H.D. (2008). Biochemical and physiological behavior of Vignaunguiculata (L.) Walp.under waterstress during the vegetative phase. *Asian Journal of Plant Science*, 7:44-49.

- Davenport, D.C., Martin, P.E., & Hagan, M.R. (1972). Antitranspirants for conservation of leaves water potential of transplanted citrus trees. *Horticultural science*, 7: 511-520.
- Delauney, A.J. & Verna, D.P.S. (1993). Proline biosynthesis and osmoregulation in plants. *Plant Journal*, 4: 215-223.
- Department of Agriculture, Malaysia (DOA). (2015). Booklet statistik tanaman (Subsektor Tanaman Makanan 2015. First country report. Kuala Lumpur: Department of Agriculture. pp. 49.
- Devi, M.J., Sinclair, T.R., Chen, P. & Carter, T. (2014). Evaluation of elite southern maturity soybean breeding lines for drought tolerant traits. *Agronomy Journal*, 106: 1947-1954.
- Deepak, M.K., Ritesh, G., Jun, Y.C., Jonggeun, K., Inhwan, B. & Hanhong, B. (2016). Multiple reaction monitoring mode based liquid chromatography-mass spectrometry method for simultaneous quantification of brassinolide and other plant hormones involved in abiotic stresses. *International Journal of Analytical Chemistry*, 2016:1-8.
- Dorji, K., Behboudian, M.H. & Zegbe-Dominguez, J.A. (2005). Water relations, growth, yield, and fruit quality of hot pepper under deficit irrigation and partial root zone drying. *Scientia Horticulturae*, 104: 137-149.
- Dreesmann, D.C., Harn, C. & Daie, J. (1994). Expression of genes encoding rubisco in sugarbeet Beta vulgaris L. plants subjected to gradual desiccation. *Plant Cell Physiology*, 35(4): 645-653.
- Dry, P.R. & Loveys, B.R. (1999). Grapevine shoot growth and stomatal conductance are reduced when part of the root system is dried. *Vitis*, 39: 9-12.
- Ehleringer, J. (1980). Leaf morphology and reflectance in relation to water and temperature stress. In: Adaptation of Plants to Water and High Temperature Stress, eds. N.C. Turner and P.J. Kramer, pp. 295-308.Wiley, New York.
- El-Abd, A.A.A. (1996). Studied on the effect of drainoge water and/or antitranspirants on growth and some chemical constituents of some Citrus rootstock. M.Sc.Thesis, Faculty of Agriculture Kafr El-Shikh, Tanta University, Egypt.
- Elazab, A., Molero, G., Serret, M.D. & Araus, J.L. (2012). Root traits and $\delta^{13}C$ and $\delta^{18}O$ of durum wheat under different water regimes. *Functional Plant Biology*, 39: 379–393.
- El-Kholy, M.A. & Gaballah, M.S., (2005). Productivity of wheat cultivars as affected by seeding methods and reflectant application under water stress condition. *Journal Agronomy*, 4: 23-30.

- El-Kholy, M.A., Gaballah, M.S., El-Ashry, S. & El-Bawab, A.M. (2005). Combating drought using yield stabilizing agents in barley. *International Journal Agriculture Biology*, 7(3): 369-437.
- El-Zohiri, S.S.M. & Abdelal, A.M.H., (2014). Improve the adverse impacts of water stress on growth, yield and its quality of taro plants by using glycinebetaine, MgCO₃ and defoliation under delta conditions. *Middle-East Journal Agriculture Research*, 3: 799-814.
- Epron, D. (1997). Effect of drought on photosynthesis and on the thermotolerance of photosystem II in seedling of Cedar *Cedrus atlantica* and *C. libani. Journal of Experimental Botany*, 48(315): 1835-1841.
- Esitken, A. & Turan, M. (2004). Alternating magnetic field effects on yield and plant nutrient element composition of strawberry (*Fragaria* × *Ananassa* cv. camarosa). *Acta Agriculturae Scandinavica, Section B- Plant Soil Science*, 54(3): 135-139.
- Evans, J.R. and Loreto, F. (2000). Acquisition and diffusion of CO₂ in higher plant leaves. In: Photosynthesis: Physiology and Metabolism, eds. Leegood, R.C., Sharkey, T.D., Von Caemmerer, S., pp. 321-351.
- FAO. (2017). Banana market review. Retrieved on 2 April 2018 from http://www.fao.org/3/a-i7410e.pdf.
- Farooq, M., Basra, S.M.A., Wahid, A., Cheema, Z.A., Cheema, M.A.and Khaliq, A. (2008). Physiological role of exogenously applied glycinebetaine in improving drought tolerance of fine grain aromatic rice (*Oryza sativa L.*). *Journal Agronomy of Crop Science*, 194: 325–333.
- Farquhar, G.D. (1978). Feed forward responses of stomata to humidity. *Australian Journal of Plant Physiology*, Melbourne, Vol. 5, p.787-800.
- Farrag, A.M. and El-Nagar, A.M. (2005). Response of cucumber plants to drip irrigation levels, NPK fertilization rates and salinity distribution in the soil. In: *Conference on Recent Technologies in Agriculture*, 14-16 November, Cairo University, Egypt, pp. 519-534.
- Faten, S.A., Mona, M.A.M. & Aisha, H.A. (2008). Combined effect of irrigation intervals and foliar application of some antitranspirants on eggplant growth, fruits yield and its physical and chemical properties. *Research Journal of Agriculture and Biology Science*, 4(5): 416-423.
- Fariduddin, Q., Yusuf, M., Ahmad, I., & Ahmad, A. (2014). Brassinosteroids and their role in response of plants to abiotic stresses. *Biologia Plantarum*, 58 (1): 9-17.
- Feroza, K.C., Rosa, M.R., Eduardo, B. & Ron, M. (2017). Reactive oxygen species, abiotic stress and stress combination. *The Plant Journal*, 90: 856–867.

- Fischer, G., Shah, M.M., Van Velthuizen, H.T. & Nachtergaele, F.O. (2002). Global agroecological assessment for agriculture in 21st century: Methodology and Results. In IIASA Research Report RR-02-002, Laxenburg, Austria.
- Flore, J.A. & Lakso, A.N. (1988). Environmental and physiological regulation of photosynthesis in fruit crops. *Horticulture Reviews*, 11:111-157.
- FOASTAT. (2015). Retrieved on 22 August 2016 from http://www.foastat.org.
- Forouzani, M., Karami, E., Zibaei, M., & Zamani, G.H. (2012). Agricultural Water Poverty Index for a Sustainable World. *Farming for Food and Water Security*, 127-155.
- Fujii, S. & Saka, H. (2001). Distribution of assimilates to each organ in rice plants exposed to a low temperature at the ripening state, and the effect of brassinolide on the distribution. *Plant Productive Science* 4, 136–144.
- Futoshi, S. & Naoto, I. (2013). Brassinolide induce stem growth of water spinach. Bulletin Shinshu University Alpine Field Centre, 11:1-4.
- Gale, J. and Hagan, R.M. (1966). Plant anti-transpirants. *Annual Review on Plant Physiology*, 17: 269-282.
- Geller, G.N. & Smith, W.K. (1982). Influence of leaf size, and arrangement on temperature and transpiration in three high-elevation, large-leafed herbs. *Oecologia* (Berl), 53: 227-234.
- Gerten, D., & Rost, S. (2010). Development and climate change: climate change impacts on agricultural water stress and impact mitigation potential. Potsdam Institute for Climate Impact Research (PIK), Germany, pp. 8.
- Giasson, B.I., Ischiropoulos, H., Virginia, M., Lee, Y. & Trojanowski, J.Q. (2002). The relationship between oxidative/nutritive stress and pathological inclusion in Alzhemier's and Parkinson's diseases. *Free Radical Biology and Medicine* 32: 1264-1275.
- Giorio, P., Sorrentino, G., & d'Andria, R. (1999). Stomatal behaviour leaf water status and photosynthetic response in field-grown olive trees under water deficit. *Environmental Experimental Botany*, 42: 95-104.
- Goode, J.E., Higgs, K.H. and Hyrvcz, K.J. (1979). Effect of water stress control in apple trees by misting. *Journal of Horticultural Science*, 54: 1-11.
- Gowing, D.J.G., Davies, W.J., & Jones, H.G. (1990). Appositive root-sourced signal as an indicator of soil drying in apple, *Mallus* × *domestica* Borkh. *Journal of Experimental Botany*, 41: 1535-1540.
- Grassi, G. & Magnani, F. (2005). Stomatal, mesophyll conductance and biochemical limitations to photosynthesis as affected by drought and leaf ontogeny in ash and oak trees. *Plant, Cell and Environment,* 28: 834–849.

- Grove, M., Spencer, G., Rohwedder, W., Mandava, N., Worley, J., Warthen, J., Jr., Steffens, G., Flippen-Anderson, J. & Cook, J. (1979). Brassinolide, a plant growth-promoting steroid isolated from *Brassica napus* pollen. *Nature*, 281:216-217.
- Gunes, A., Inal A., Adak M.S, Bagci E.G., Cicek N., & Eraslan F., (2008). Effect of drought stress implemented at pre- or post- anthesis stage some physiological as screening criteria in chickpea cultivars. *Russian Journal Plant Physiolohy*, 55: 59-67.
- Gusta, L.V., & Chen, T.H.H., (1987). The physiology of water and temperature stress. In: Heyne, E.G. (Ed.), Wheat and Wheat Improvement, USA, pp. 115-150.
- Hale, M.G. & Orcutt D.M. (1987). The physiology of plants under stress. Wiley-InterScience, New York. pp. 93.
- Hameed, M.A., Reid, J.B. & Rowe R.N. (1987). Root confinement and its effects on the water relations, growth and assimilate partitioning of tomato (Lycopersicum esculentum Mill). *Annals of Botany*, 59: 685-692.
- Harris, R.W. (1992). Root-shoot ratios. Journal of Arboriculture, 18(1):39-42.
- Hasan, S.A., Hayat, S., Ali, B.B. & Ahmad, A. (2008). 28- Homobrassinolide protects chickpea (*Cicer arietinum*) from cadmium toxicity by stimulating antioxidants. *Environment Pollution*, 151:60-66.
- Haubrick, L.L., Torsethaugen, G. & Assmann, S.M. (2006). Effect of brassinolide, alone and in concert with abscisic acid, on control of stomatal aperture and potassium currents of *Vicia faba* guard cell protoplasts. *Physiology Plant*, 128: 134–143.
- Heath, R.L. & Packer, L. (1968). Photoperoxidation in isolated chloroplasts. I. Kinetics and stoichiometry of fatty acid peroxidation. *Arch Biochemistry Biophysiology* 125: 189-198.
- Heslop-Harisson, J.S. & Schawarzacher, T. (2007). Domestication, genomics and the future of banana. *Annals of Botany*, 100: 1073-1084.
- Heuer, B. (1999). Osmoregulatory role of proline in plants exposed to environmental stresses. Handbook pp. 675-695.
- Hoffmann, H. P. & Turner, D.W. (1993). Soil water deficits reduce the elngation rate of emerging banana leaves but the night/ day elongation ratio remains unchanged. *Scientia Horticulture*, 54, 1-12.
- Hsiao, T.C. & Acevedo, E. (1974). Plant responses to water deficits, water-use efficiency, and drought resistance. *Agricultural Meteorology*, 14: 59-84.
- Hsiao, T.C. & Xu, L.K. (2000). Sensitivity of growth of roots versus leaves to water stress: Biophysical analysis and relation to water transport. *Journal of Experimental Botany*, 51: 1595-1616.

- Hsiao, T.C. (1973). Plant responses to water stress. *Annual review of plant physiology*, 24(1): 519-570.
- Hu, S., Wang, C., Sanchez, D.L., Lipka, A.E., Liu P., Yin Y. & Lubberstedt T. (2017). Gibberellins Promote Brassinosteroids Action and Both Increase Heterosis for Plant Height in Maize (*Zea mays* L.). *Frontiers in Plant Science* 8: 1039.
- Hu. Y. & Schmidhalter, U. (1998). Spatial distribution of inorganics ions and carbohydrates contributing to osmotic adjustment in the elongating wheat leaf under saline conditions. *Australian Journal of Plant Physiology*, 25: 591-597.
- Hurd, E. A. (1975). Phenotype and drought tolerance in wheat. In: J.F. Stone (ed.) Plant modification for more efficient water use. Elsever Pun., New York.
- Ibrahim, E.A. & Selim, E.M. (2010). Effect of irrigation intervalles and antitranspirants (Kaolin) on summer squash (Cucurbita pepo L.) growth, yield quality and economics. Journal of Soil Science and Agriculture Engineering, Mansoura University 1: 883-894.
- Ibrahim, E.A. & Selim, E.M. (2007). Effect of irrigation intervals and nitrogen fertilizer rates on summer squash (*Cucurbita pepo* L.) growth, yield, nutritional status and water use efficiency. *Journal of Agriculture Science, Mansoura University*, 32: 10333-10345.
- Ingram, J. & Bartels, D. (1996). The molecular basis of dehydration tolerance in plants. *Plant Physiology*, 47: 377-403.
- Iqbal, M.M., Shah, S.M., Mohammad, W. & Nawaz, H. (1999). Field response of potato subjected to water stress at different growth stages. In: Kirda, C., Mountonnet, P., Hera, C., Nielsen, D.R. (Eds.). Crop Yield Response to Deficit Irrigation (pp. 213-223). The Netherlands: Kluwer Academic Publishers.
- Isabelle, M., Lee, B.L., Lim, M.T., Koh, W.P., Huang, D. & Ong, C.N. (2010). Antioxidant activity and profiles of common fruits in Singapore. *Food Chemistry*, 123: 77-84.
- Ismail, M.R., Davies W.J. & Awad M.H. (2002). Leaf growth and stomatal sensitivity to ABA in droughted pepper plants. Science Horticulture, 96: 313-327.
- Iwahari S., Tominaga S., & Higuchi S. (1990). Retardation of abscission of citrus leaf and fruitlet explants by brassinolide. *Plant Growth Regulation*, 9:119–125.
- Jager, C.E., Symons, G.M., Ross, J.J. & Reid, J.B. (2008). Do brassinosteroids mediate the water stress response. *Journal Physiology Plant*, 133: 417–425.
- Jain A. & Srivaslava H.S. (1981) Effect of salicyalic acid on nitrate reductase activity in maize seedlings. *Physiologia Plantarum*, 51: 339-345.
- Jaleel, C.A., Gopi R. & Panneerselvam, R. (2008). Growth and photosynthetic pigments responses of two varieties of *Catharanthus roseus* to triadimefon treatment. *Comptes Rendus Biologies*, 331: 272–277.

- Javan, M., Tajbakhsh, M., Mandoulakani, B.A. (2013). Effect of anti-transpirants application on yield and yield components in soybean (*Glycine max L.*) under limited irrigation. *Journal Application Biology Science*, 7: 70-74.
- Jha, Y., & Subramanian, R.B. (2016). Regulation of plant physiology and antioxidant enzymes for alleviating salinity stress by potassium-mobilizing bacteria. In: Meena V., Maurya B., Verma J., Meena R. (eds) Potassium Solubilizing Microorganisms for Sustainable Agriculture. Springer, New Delhi, pp. 149-162.
- Jian, H.N., Shakeel, A.A., Ran, W., Jin, H.L., Mei, L., Ji, X.S., Ali, Z., Jun, L., San, W. & Xue, F.Z. (2016). Exogenous application of brassinolide can alter morphological and physiological traits of *Leymus chinensis* (Trin.) under room and high temperatures. *Journal of Agricultural Research*, 76(1): 27-33.
- Jones, H.G. (1985). Partitioning stomatal and non-stomatal limitations to photosynthesis. *Plant, Cell & Environment,* 8: 95-104.
- Jones, I.D. (1931). Preliminary report on relation of soil moisture and leaf area to fruit development of the Georgia Belle peach. *Proceeding of the American Society for Horticulture Science*, 28: 6-14.
- Kaiser, W.M. (1987). Effect of water deficit on photosynthetic capacity. *Physiologia Plantarum*, 71: 142-149.
- Kallarackal, J., Milburn, J.A. & Baker, D.A. (1990). Water relations of the banana. III effects of controlled water stress on water potential, transpiration, photosynthesis and leaf growth. *Australian Journal of Plant Physiology*, 17: 79-90.
- Kamel, A. & Loser, D.M. (1995). Contribution of carbohydrates and other solutes to osmotic adjustment in wheat leaves under water stress. *Journal Plant Physiology*. 145, 363–366.
- Kameli, A. & Losel, D.M. (1996). Growth and sugar accumulation in durum wheat plants under water stress. *New Phytologist*, 132(1):57-62.
- Kanazawa, K. & Sakakibara, H. (2000). High content of a dopamine, a strong antioxidant, in Cavendish banana. *Journal of Agriculture and Food Chemistry*, 48:844-848.
- Kasim, W.A., Osman, M.E., Omar, M.N., Abd El-Daim, I.A., Bejai, S., & Meijer, J., (2013). Control of drought stress in wheat using plant growth promoting bacteria. *Journal Plant Growth Regulator*, 32: 122-130.
- Kayat, F., Mohammad, A., Idris A.A., Ibrahim, M.F., Soon J.M., Ahmad Yusuf M.K., Wong K.F. & Zulariff A.L. (2016). Proceeding from Second Asia Pacific Conference on Advanced Research (APCAR): Study of the intention of banana growers in improving the production in Jeli, Kelantan, APCAR, Melbourne, February, 2016.

- Khalil, S.E. (2006) Physiological study on sesame plants grown under saline water irrigation condition. PhD thesis, Cairo University, pp. 229.
- Khalil, S.E., Abd El-Aziz, N.G. & Abou-Leila, B.H. (2010). Effect of water stress and ascorbic acid on some morphological and biochemical composition of *Ocimum basilium* plant. *Journal of American Science*, 6: 33-44.
- Khan, A. M. Farooque., Haque, M. A., Rahim M. A. & Hoque M. A. (2008). Effect of water stress at vigorous growth stages on the physio-morphological characters and yield in chilli. *Bangladesh Journal Agricultural Research*, 33(3): 353-362.
- Kharkhina, T.G., Ottosen, C.O. & Rosenqvist, E. (1999). Effects of root restriction on the growth and physiology of cucumber plants. *Physiologia Plantarum*, 105(3): 434-441.
- Kirkham, M.B. (2004). Water-Use Efficiency. Encyclopedia of Soils in the Environment. 315-322.Uchida, J., & P. Levin, & S. Miyasaka, & G. Teves, & J. Hollyer, & S. Nelson, & J. Ooka, 2008. Taro Mauka to Makai. College of Tropical Agriculture and Human Resources, University of Hawaii, Honolulu, Hawaiiwaii.
- Kleinendorst, A., (1975). An explosion of leaf growth after stress conditions. Netherlands *Journal of Agriculture Science*, 20: 203-217.
- Kohl, D.H., Kennelly, E.J., Zhu Y.X., Schubert, K.R. & Shearer, G. (1991). Proline accumulation, nitrogenase (C₂H₂ reducing) activity and activities of enzymes related to proline metabolism in drought stressed soybean nodules. *Journal of Experimental Botany*, 42:831-837.Ke L.S. (1979). Studies on the physiological characteristics of bananas in Taiwan. II. Effects of soil moisture on some physiological functions and yield of the banana plant. *Journal Agriculture Association, China*, 108: 11-23.
- Koyel, D., Arkendu, G., Bhanu P., Piyali, D., & Suddhasuchi, D. (2016). Impact of climate change and mitigation strategies on fruit production. *Advances in Life Sciences*, 5(7): 2588-2596.
- Krasensky, J. & Jonak, C. (2012). Drought, salt, and temperature stress-induced metabolic rearrangements and regulatory networks. *Journal Experimental Botany*, 63(4):1593-608.
- Krishna, P. (2003). Brassinosteroid-mediated stress responses. Journal Plant Growth Regulator, 22:289–297.
- Krizek, D.T., Carmi, A., Mirecki, R.M., Synder, F.W. & Bunce, J.A. (1985). Comparative effects of soil moisture stress and restricted root zone volume in morphogenetic and physiological response of soybean (*Glycine max L. Merr.*). *Journal of Experimental Botany*, 36(162): 25-38.
- Kuang, J.B, Turner, N.C. & Henson, I. E. (1990). Influence of xylem water potential on leaf elongation and osmotic adjustment of wheat and lupin. *Journal of Experimental Botany*, 41:217-221.

- Kulkarni, V.M, Suprasanna, P., & Bapat, V.A. (2006). Plant regeneration through multiple shoot formation and somatic embryogenesis in a commercially important and endangered Indian banana cv. Rajeli. *Current Science*, 90: 842-846.
- Kumashiro, K., & Tateishi, S. (1997). The effect of soil moisture on the tree growth, yield and fruit quality of Jonathan apples. *Journal Japanese Society Horticulturae Science*, 36: 9-20.
- Lascano, R.J., Sojka, R.E., Adamsen, F., Agronomy, A.S.O., America, C.S.S.O., & America, S.S.S.O. (2007). Irrrigation of agricultural crops: *American Society of Agronomy*, 30: 279-335.
- Leonid, V.K., Michael, A.B., Thomas, G.B., & Richard, P.P. (2015). Structure–function relationships of four stereoisomers of a brassinolide mimetic on hypocotyl and root elongation of the brassinosteroid-deficient det2-1 mutant of *Arabidopsis*. *Journal Plant Growth Regulation* 2016: 215–221.
- Leskovar, D.I. & Stoffella, P.J. (1995). Vegetable seedling root systems: morphology, development and importance. *Horticulture Science*, 30: 1153-1159.
- Levitt, J., (1980). Responses of Plants to Environmental Stress, vol. 2. Academic Press, New York, pp. 433.
- Lee, W.C. & Baharuddin, A.H. (2018) Impacts of climate change on agriculture in Malaysia. In: Omran A., Schwarz-Herion O. (Eds) The Impact of Climate Change on Our Life. Springer, Singapore, pp. 179-195.
- Liang Z., Zhang F., Shao M., & Zhang J. (2002). The relations of stomata conductance, water consumption, growth rate to leaf water potential during soil drying and rewatering cycle of wheat (*Tricitum aestivum*). *Botanical Bulletin of Academica Sinica*, 43: 187-192.
- Lisar, S.Y.S, Motafakkerazad, R., Hossain, M.M., & Rahman, I.M.W. (2012). Water stress in plants: causes, effects and responses, In Rahman, I.M.W. and Hasegawa, H. (Ed.) Water Stress. USA: Ed. In Tech, New York.
- Lockhart, J.A. (1965). An analysis of irreversible plant cell elongation. *Journal of Teoritical Biology*, 8: 264-275.
- Long, S.P., Farage, P.K. & Garcia, R.L. (1996). Measurement of leaf and canopy photosynthetic CO₂ exchange in the field. *Journal of Experimental Botany*, 47: 1629-1642.
- Loveys, B.R., Stoll, M., Dry, P.R., & McCarthy, M.G. (1998). Partial root zone drying stimulates stress responses in grapevine to improve water efficiency while maintaining crop yield and quality. *Australian Grape grower and Winemaker, Technical Issue*, 414: 108-114.
- Luo, H.H., Zhang, Y.L. & Zhang, W.F. (2016). Effects of water stress and rewatering on photosynthesis, root activity, and yield of cotton with drip irrigation under mulch. *Photosynthetica*, 54 (1): 65-73.

- Lutts, S., Kinet, J.M. & Bouharmont, J. (1996b). NaCl-induced senescence in leaves of rice (*Oryza sativa* L.) cultivars differing in salinity resistance. *Annual Botany*, 78: 389-398.
- Lutts, S., Kinet, J.M. & Bourharmont, J., (1996a). Effects of salt stress on growth, mineral nutrition and proline accumulation in relation to osmotic adjustment in rice (*Oryza sativa* L.) cultivars differing in salinity resistance. *Plant Growth Regulator*, 19: 2017.
- Ma, R.K., Jian, J.L. (1991) Effect of antitranspirant on bean (*Pcia faba*) grown under conventional and conservation tillage systems. *Boden Kulture*, 42 (4), 327 (*Plant Breeding Abstracts* 62, 2959)
- Maestri, M., Da Matta, F.M., Regazzi, A.J. & Barros, R.S. (1995). Accumulation of proline and quaternary ammonium compounds in mature leaves of water stressed coffee plants *Coffee Arabica* and *C. canephora. Journal of Horticultural Science*, 70(2): 229-233.
- Magdy, A.S., Hazem, M.M.H., Alia, A.M.N. & Alshaimaa, A.I. (2016). The influence of lithovit fertilizer on the chemical constituents and yield characteristics of cotton plant under drought stress. *International Journal of Chemical Technology Research*, 9(8), pp. 01-11.
- Magness, J.R. (1935). Soil moisture and irrigation investigations in eastern apple orchards. United Stated Department Agriculture. pp. 491.
- Makhmudov, S.H.A. (1983). A study on chlorophyll formation in wheat leaves under moisture stress. Baku, Azerbiajan SSR, pp. 40-44.
- Makus, D.J. (1997). Effect of an antitranspirant on cotton grown under conventional tillage systems. Proc. betwide cotton conf., New Orleans, LA, USA.January, 6(10): 642-644.
- Mandava, N.B. (1988). Plant growth-promoting brassinosteroids. *Annual Review Plant Physiology and Plant Molecular Biology*, 39: 23-52.
- Manivannan, P., Abdul Jaleel, C., Sankar, B., Kishorekumar, A., Somasundaram, R., Lakshmanan, G.M.A. & Panneerselvam, R. (2007). Growth, biochemical modifications and proline metabolism in *Helianthus annuus* L. as induced by drought stress. *Colloids and Surfaces B: Biointerfaces*, 59: 141–149.
- Manzel, C.M., Turner, D.W., Doogan, V.J., & Simpson, D.R. (1994). Root shoots interactions in passion fruit (*Passiflora* sp.) under the influence of changing root volumes and soil temperatures. *Journal of Horticultural Science*, 69(3): 553-564.
- Maswada, H.F., & Abd El-Rahman, L.A. (2014). Inducing salinity tolerance in wheat plants by hydrogen peroxide and lithovit "a nano-CaCO₃ fertilizer". *Journal Agricultural Research*. Kafr El-Sheikh University, 40 (4): 696-719.

- Mazorra, L.M., Nunez, M., Hechavarria, M., Coll, F., & Sanchez-Blanco, M.J. (2002). Influence of brassinosteroids on antioxidant enzymes activity in tomato under different temperatures. *Biology Plant*, 45:593–596.
- Mclean, E.O. (1982). Soil pH and lime requirement. In: Page, A.L., Ed., Methods of Soil Analysis. Part 2. Chemical and Microbiological Properties, American Society of Agronomy, Soil Science Society of America, Madison, 199-224.
- Meloni, D.A, Golotta, M.R., Martinez, C.A., & Oliva, M.A. (2004). The effects of salt stress on growth, nitrate reduction and proline and glycinebetaine accumulation in *Prosopis alba. Brazilian Journal of Plant Physiology*, 16 (1): 201-214.
- Metwally, M.M., Afify, M.M., Wahba, H.E., Makarem, A.M., Mohamed, A., Mohamed, A.E. & Mahfouz, S. (2002) Effect of irrigation and Vapor Guard on growth, yield and chemical composition of roselle plant. *Bulletin of the NRC*, *Egypt*, 27: 533-548.
- Moftah, A.E., & Al-Humaid, A.I. (2006). Response of vegetative and reproductive parameters of water stressed Tuberose plants to vapor grad and Kaolin Antitranspirants. *Journal King Saud University Agriculture Science*, 18(2): 127-139.
- Mohankrishna, T., Bhasjar, C.V.S., Sanjeeva Rao, P., Chandrashekar, T.R., Sethuraj, M.R., & Vijayakumar, K.R. (1991). Effect of irrigation on physiological performance of immature plants of *Heavea brasiliensis* in North Konkan. *Indian Journal of Natural Rubber Research*, 4(1):36-45.
- Mok, C.K. (1979). Water potential components, growth and physiological responses of soybeans to osmotically induced water stress. Iowa University, America. PhD Thesis.
- Montoya, T., Namura, T., Yokota, T., Farrar, K., Harrison, K., Jones, J.G.D., Kaneta, T., Kamiya, W., Szekeres, M. & Bishop, G.R. (2005). Patterns of dwarf expression and brassinosteroid accumulation in tomato reveal the importance of brassinosteroid synthesis during fruit development. *Plant Journal*, 42: 26-269.
- Munns, R. & Cramer, G.R. (1996). Is co-ordination of leaf and root growth mediated by abscisic acid? Opinion. *Plant and Soil*, 185: 33-49.
- Musick, J.T. (1994). General guidelines for deficit irrigation management. Paper Presented at Central Plains Irrigation Short Course, Garden City, KS, 7-8 February 1994.
- Naidu, B.P., Paleg, L.G., Aspinall, D., Jenning, A.G. and Jones, G.P. (1990). Rate of imposition of water stress alters the accumulation of nitrogen containing solutes by wheat seedlings. *Australian Journal Plant Physiology*, 17: 377-382.
- Naik, G.P. & Joshi, G.V. (1983). Ineffectual role of proline metabolism in salt-stressed sugarcane leaves. *Procedure Indian Academy Science*, 92: 265-272.
- Nakano, A., & Uehara, Y. (1996). The effect of kaolin clay on cuticle transpiration in tomato. *Acta Horticulturae*, 440, 233-238.

- Naomi, R. & George G.G. (1994). How emergent plants experience water regime in a Mediterranean-type wetland. *Aquatic Botany*, 49 (1994) 117-136.
- Nasraoui, B. (1993). Role of antitranspirant films in protecting plants against fungal diseases. *Annals of the National Institute of Agronomic Research of Tunisia*, 66: 125-135.
- Natali, S., Bignami, C. &Fusari, A. (1991). Water consumption, photosynthesis, transpiration and leaf water potential in *Olea europeae* L., cv. Frantoio, at different levels of available water. *Agriculture Media*, 121: 205-212.
- Nelson, S.C., Ploetz R.C. & Kepler. A.K. (2006). *Musa* species (bananas and plantains), ver. 2.2. In: C.R. Elevitch (Ed.), Species Profiles for Pacific Island Agroforestry. Permanent Agriculture Resources (PAR), Holualoa, Hawai'i.
- Nermeen, T.S. & Emad, A.S. (2011). Influence of some chemical compounds as antitranspirant agent on vase life of *Monstera deliciosa* leaves. *African Journal of Agricultural Research*, 6 (1):132-139.
- Niu, J., Anjum, S.A. and Wang, R. (2016). Exogenous application of brassinolide can alter morphological and physiological traits of *Leymus chinensis* (Trin.) Tzvelev under room and high temperatures. *Chilean Journal Agricultural Research*, 76: 27-33.
- Noble, P.S. (1980). Leaf anatomy and water use efficiency. In: Adaptation of Plants to Water and High Temperature Stress, eds. N.C. Turner and P.J. Kramer, pp. 43-55, Wiley, New York.
- Noctor, G., Veliovic-Jovanovic, S., & Foyer, C.H. (2000). Peroxide processing in photosynthesis: antioxidant coupling and redox signaling. *Philosophical transaction of the royal society* B, 355: 1465-1475.
- Oliver, S., Barber, A.S. (1966). An evaluation of the mechanisms governing the supply of Ca, Mg, K and Na to soybean roots (Glycine max L.). Soil Science Society of America Proceedings, 30: 82-86.
- Ookawa, T., Naruoka, Y., Yamazaki, T., Suga, J., Hirasawa, T. (2003). A comparison of the accumulation and partitioning of nitrogen in plants between two rice cultivars. Akenohoshi and apponbare, at the ripening stage. *Plant Production Science*, 6: 172-178.
- Ouda, S.A., El-Mesiry, T., & Gaballah, M.S. (2007). Effect of using stabilizing agents on increasing yield and water use efficiency in barley grown under water stress. *Australian Journal of Basic and Applied Science*, 1:571-577.
- Ozdemir, F., Bor, M., Demiral, T. & Turkan, I. (2004). Effect of 24-epibrassinolide on seed germination, seedling growth, lipid peroxidation, proline content and antioxidative system of rice (*Oryza sativa* L.) under salinity stress. *Plant Growth Regulation*, 42 (3): 203-211.

- Ozgur, T. & Mithat, N.G., (2008). Influence of water stress on proline accumulation, lipid peroxidation and water content of wheat. *Asian Journal of Plant Sciences*, 7: 409-412.
- Pandey, R.K., Herrera, W.A.T., Villegas, A.W., & Penletion, J.W. (1984). Drought response of grain legumes under irrigation gradient. III. Plant growth. *Agronomy Journal*, 76: 557-560.
- Passioura, J.B. (1992). The physical chemistry of the primary cell wall: implication for the control of expansion rate. *Journal of Experimental Botany*, 45: 1675-1682.
- Pennazio, S., Roggero, P. (1984). Effect of ABA and DNP on stomatal resistance in detached tobacco leaves. *Giornale Botanico Italian*, 118, 132-140.
- Peterson, T. A., Reinsel, M.D. & Krizek, D.T. (1991). Tomato (*Lycopersicon esculentum* cv. 'Better Bush') plant response to root restriction. II. Root respiration and ethylene generation. *Journal of Experimental Botany*, 42: 1241-1249.
- Petropoulos, S.A., Daferera, D., Polissiou, M.G. & Passam, H.C. (2008). The effect of water deficit stress on the growth, yield and composition of essential oils of parsley. *Science Horticulture*, 115: 393-397.
- Pistacco, A., Lain, O. & Giulivo, C. (1990). Gas exchange and architecture of sweet pepper as affected by water stress. *Congress of the European Society of Agronomy*, 1: 32-38.
- Ploetz, R.C. (2006). Panama disease: An old nemesis rears its ugly head. Part2. The Cavendish era and beyond. Plant Health Progress. St. Paul USA: Plant Management Network.
- Poljakoff-Mayber, A., & Gale, J. (1975) Plants in saline environments. In: *Ecological Studies* (Vol 15), Springer Verlag, Berlin, pp 61-69.
- Poltez, R.C. Kelper, A.K., Daniells, J., and Nelson, C. (2007) Banana and plantain-an overview on pacific island cultivars, Species profiles for pacific island agroforestry. Retrived on 28 March 2018 from www.traditionaltree.org.
- Pompelli, M.F., Barata-Luis, R., Vitorino, H., Gonclaves, E., Rolim, E., Santos, M., Almeida-Cortez, J., & Endrez, L. (2010). Photosynthesis, photoprotection and antioxidant activity of purging nut under drought deficit and recovery. *Biomass Bioenergy*, 34:1207-1215.
- Prior, S.A., Rogers H.H, Runion, G.B & Mauney, J.R. (1994): Effects of free- air CO₂ enrichment on cotton root growth. *Agriculture and Forestry Meteorology*, 70: 69-86.
- Proebsting, E.L., Jerie, P.H. & Irvine, J. (1989). Water deficit and rooting volume modify peach tree growth and water relations. *Journal of American Society of Horticulture Science*, 114: 368-372.

- Prusakova, L.D, Chizhova, S.I., Tretyakov, N.N., Ageeva, L.F., Golantseva, E.N. and Yakovlev, A.F. (1999). Ecost and epibrassinolide antistress functions on spring wheat under the conditions of the Central Non-Chernozem zone. *Agrarian Russia*: 39-41.
- Quan, R., Shang M., Zhang H., Zhao Y. & Zhang J. (2004). Engineering of enhanced glycine betaine synthesis improves drought tolerance in maize. *Plant Biotechnology Journal*, 2(6): 477-486.
- Quick, W.P., Chaves, M.M., & Wendler, R. (1992). The effect of water stress on photosynthetic carbon metabolism in four species grown under field conditions. *Plant Cell Environment*, 15: 25-35.
- Quisenberry, J.E. (1982). Breeding for drought resistance and plant water use efficiency. In: M.N. Christaninson and C.F. Lewis (Eds.). Breeding for less favourable environments. Wiley-InterSci., New York.
- Rademacher, W. (2015). Plant growth regulators: backgrounds and uses in plant production. *Journal of Plant Growth Regulation*, 34(4): 845-872.
- Radhakrishnan, R. & Kumari, B.D.R. (2012). Pulsed magnetic field: A contemporary approach offers to enhance plant growth and yield of soybean. *Plant Physiology and Biochemical*, 51: 139-144.
- Rafiei, Shirvan, M. & Asgharipur, M.R. (2009). Yield reaction and morphological characteristics of some mung bean genotypes to drought stress. *Journal Modern Agriculture Knowledge*, 5(15): 67-76.
- Ranjitkar, S., Sujakhu, N.M., Merz, J., Kindt, R., Xu, J. & Matin, M.A. (2016). Suitability analysis and projected climate change impact on banana and coffee production zones in Nepal. *PLoS ONE*, 11(9): 1-18.
- Ranney, T.G., Bassuk, N.L., & Whitlow, T.H. (1989). Effect of transplanting practice on growth and water relation of "colt" cherry trees during reestblishment. Department of Horticulture, Aalabama Agricultural Experimental Station, Auburn University, Auburn, AL 36849, USA.
- Razi, M.I & Awad, M.H. (2000). Survival of young mangosteen in the field and responses to shading and water stress. *Transaction of the Malaysian Society of Plant Physiology*, pp. 79-81.
- Razi, M.I. & Davies, W.J. (2000). Leaf growth and stomatal sensitivity after water stress relief and its relation to xylem sap absicis acid. *Pertanika Journal Tropical Agriculture Science* 23(2): 67-73.
- Razi, M.I. & Davies, W.J. (2004). Environment stress impact on tropical crops. Malaysia: Jofri Publishing House, Serdang. 1-8 pp.
- Razi, M.I., Burrage, S.W. Tarmizi, H. & Aziz, M.A. (1994). Growth, plant water relation, photosynthesis rate and accumulation of proline in young carambola plants in relation to water stress. *Scientia Horticulturae*, 60: 101-114.

- Razi, M.I., Kamil, Y. & Marziah, M. (2004). Growth, water relations, stomata conductance and proline concentration in water stressed banana (*Musa* spp.) plant. *Asian Journal Plant Science*, 3(6): 709-713.
- Reddy, M.P. & Vora, A.B. (1986). Drought induced changes in pigment composition and chlorophyllase activity of wheat. Indian. *Journal Plant Physiology*, 29:331-334.
- Riazi, A. Matsuda, K., & Arslan, A. (1985). Water stress induced changes in concentration of proline and other solutes in growing regions of young barley leaves. *Journal of Experimental Botany*, 36: 1716-1724.
- Riccardi, M., Pulvento, C., Albrizio, R. & Barbieri, G. (2016). Drought stress response in long-storage tomatoes: physiological and biochemical traits. *Science Horticulture*, 200:25-35.
- Richard, W.H. (1992). Root-Shoot Ratio. Journal of Arboriculture 18(1): 39-42.
- Richards, L.A. (1947). Pressure-membrane apparatus-construction and use. *Agricultural Engineering*, 28: 451-454.
- Robinson, J.C. & Nel, D.J. (1985). Comparative morphology, phenology, and production potential of banana cultivars "Dwarf Cavendish' and 'Williams' in the Eastern Transvaal Lowveld. *Scientia Horticulturae*. 25:149-161.
- Robinson, J.C. (1996). Bananas and Plantain. CAB International, Cambridge.
- Roghayyeh, S., Sedghi M., Shishevan M.T. & Sharifi R.S. (2010). Effects of nano-iron oxide particles on agronomic traits of soybean. *Not Science Biology*, 2 (2): 112-113.
- Sairam, R.K. (1994). Effects of homobrassinolide application on plant metabolism and grain yield under irrigated and moisture-stress conditions of two wheat varieties. *Plant Growth Regulation*, 14:173–181.
- Sakurai, A. & Fujioka, S. (1993). The current status of physiology and biochemistry of brassinosteroids: A review. *Journal Plant Growth Regulation*, 13: 147-159.
- Samir, A.Z.M. (1988) .Water use efficiency, growth and yield of corn as influenced by antitranspirant materials. MSc thesis, Ain Shams University, Cairo, Egypt.
- Sasse, J.M. (2003). Physiological actions of brassinosteroids: an update. *Journal Plant Growth Regulation*. 22: 276–288.
- Schulze, E.D., Lange, O.L., & Buschbom, U. (1972). Stomatal response to changes in humidity in plants growing in the desert. Planta, Berlin, v. 108, n. 2, p. 259-270.
- Sengupta, K., Banik, N.C., Bhui, S. & Mitra, S. (2011). Effect of brassinolide on growth and yield of summer green gram crop. *Journal of Crop and Weed*, 7(2): 152-154.

- Shamshuddin, J. & Fauziah, C. I. (2010). Weathered tropical soils: The Ultisol and Oxisol. UPM Press, Serdang, Malaysia.
- Shanan, N.T., & Shalaby, E.A., (2011). Influence of some chemical compounds as antitranspirant agents on vase life of *Monstera deliciosa* leaves. *Journal of Agricultural Research*, 6: 132-139.
- Shao, H.B, Chu, L.Y., Jaleel, C.A. & Zhao, C.X. (2008). Water-deficit stress-induced anatomical changes in higher plants. *Comptes Rendus Biologies*, 331: 215-225.
- Shimber, G.T., Razi, M.I., Kausar, H., Marziah, M., Ramlan M.F. (2013). Plant water relations, crop yield and quality of arabica coffee (*Coffea arabica*) as affected by supplemental deficit irrigation. *Australian Journal of Crop Science*, 7(9): 1361.
- Siddique, M., Hamid, R.B. & Islam, M.A. (1999). Drought stress effect on photosynthetic rate and leaf gas exchange of wheat. *Botanical Bulletin of Academia Sinica* 40: 141-145.
- Siddiqui, M.H., Al Whaibi, M.H., Firoz M. & Al Khaishany, M.Y. (2015): Role of nanoparticles in plants. *Nanotechnology and Plant Science*, 2: 19 35.
- Simons, R.K. (1986). Comparative anatomy of leaves and shoots of Golden Delicious and Jonared apples trees grown with high and low moisture supply. *Procedure American Society Horticulture Science*, 68: 20-26.
- Sinamo, V., Hanafi, N. D., & Wahyuni, T. H. (2018). Legume Plant Growth at Various Levels of Drought Stress Treatment. *Indonesian Journal of Agricultural Research*, 1(1): 9-19.
- Sinclair, T.R, Salado-Navarro, L.R., Salas, G., & Purcell, L.C. (2007). Soybean yields and soil water status in Argentina: simulation analysis, *Agriculture System*, 94:471–477.
- Singer, S.M., Helmy, Y.I., Karas, A.N. & Abou-Hadid, A.F. (2003). Influences of different water-stress treatments on growth, development and production of snap bean (*Phaseolus vulgaris L.*). *Acta Horticulture*, 614: 605-611.
- Siram, R.K. and Saxena, D.C. (2000). Oxidative stress and antioxidative in wheat genotypes: Possible mechanism of water stress tolerance. *Journal Agronomy Crop Science*, 184(1): 55-61.
- Siti Zaharah, S., Razi, I.M. (2009). Growth, stomata aperture, biochemical changes and branch anatomy in mango (Mangifera indica) cv. Chokanan in response to root restriction and water stress. *Scientia horticulturae*, 123(1): 58-67.
- Siti Hawa, J. & Novak, F. J. (1992). Somatic embryogenesis and plant regeneration of banana cultivars, *Musa* cv. Mas (AA) and *Musa* cv. Rastali (AAB). In: Valmayer, R.V., Hwang S.C., Ploetz, R., Lee, S.W. and Roa, V.N. (Eds.), International Symposium on Recent Developments in Banana Cultivation

- Technology. Taiwan Banana Research Institute, Pingtung, Taiwan, 14-18 December, 1992. Pp. 201-212.
- Sivadjian, M.J. (1967). Chemical inhibitors of plant transpirations. Phenyl Mercuric Acetate. *Bulletine Society Botany France*, 114 (1-2): 1-4.
- Sivan, P. (1995). Drought tolerance and the effect of potassium supply on growth of taro (*Colocasia esculenta* (L.) Schott) and tannia (*Xanthosoma sagittifolium* (L.) Schott). Ph.D Thesis. University of Queensland, Australia.
- Soha, E.K. & Atef, A.S. (2011). The Influence of soil moisture stress on growth, water relation and fruit quality of *Hibiscus sabdariffa* L. grown within different soil types. *Nature and science*, 9:62-74.
- Specht, J.E., Chase, K., Macrander, M., Graef, G.L, Chung, J., Markwell, J.P., Germann, M., Orf, J.H. & Lark, K.G. (2001). Soybean response to water. A QTL analysis of drought tolerance. *Crop Science*, 41: 493–509.
- Srivalli, B., Chinnusamy, V., Chopra, R.K. (2003). Antioxidant defense in response to abiotic stresses in plants. *Journal Plant Biology* 30: 121-139.
- Srivastava, N.K., & Srivastava, A.K. (2010). Influence of some heavy metals on growth, alkaloid content and composition in *Catharanthus roseus* L. Indian *Journal of Pharmaceutical Science*, 72: 775-778.
- Stang, E.J., Feree, D.C. Hall, F.R. & Spotts, R.A. (1978). Over tree misting for bloom delay in Golden Delicious apple. *Journal American Society Horticultural Science* 103: 82-87.
- Steudle, E. & Peterson, C.A. (1998). How does water get through roots? *Journal of Experimental Botany*, 49:775-788.
- Suriyan, C., Thapanee, S. & Chalermpol, K. (2013). Glycinebetaine alleviates water deficit stress in indica rice using proline accumulation, photosynthetic efficiencies, growth performances and yield attributes. *Australian Journal of Crop Science* 7(2):213-218.
- Swarowsky, A., Tate, K.W., Hopmans, J.W. & O'Geen, A.T. (2011). Catchment-scale soil water dynamics in a Mediterranean-type oak woodland. *Vadose Zone Journal*, 10, 800-815.
- Taiz, L. & Zeiger, E., (2003). Plant Physiology. Panima Publishing Corporation, New Delhi/Bangalore, pp. 690.
- Tatar, O. & Gevrek, M.N. (2008). Influence of water stress on proline accumulation, lipid peroxidation and water content of wheat. *Asian Journal Plant Science*, 7(4): 409-412.
- Tester, M. & Davenport, R.J. (2003). Na tolerance in higher plants. *Annual of Botany* 91:503-527.

- The, C.B.S. & Jamal, T. (2006). Soil Physics Analysis. Serdang: UPM press.
- Thomas, D.S. & Turner, D.W. (1998). Leaf exchange of droughted and irrigated banana cv. Williams (*Musa* spp.) growing in hot, arid conditions. *Journal of Horticultural Science and Biotechnology*, 73:419-429.
- Torrecillas, A., Guillaume, C. Alarcon, J.J., & Ruiz-Sanchez, M.C. (1995). Water relation of two tomato species under water stress and recovery. *Plant Science* 105: 169-176.
- Turner, D.W. & Lahav, E. (1983). The growth of banana plants in relation to temperature.

 Australian Journal of Plant Physiology 10:43-54
- Turner, D.W. & Rosales, F.E. (2005). Banana Root System: towards a better understanding for its productive management. International Network for the Improvement of Banana and Plantain, Montpellier, France.
- Turner, N.C. (1979). Drought resistance and adaptation to water deficits in crop plants. In: Stress Physiology in Crop Plants (Eds.) H. Mussell and R.C. Staples, pp. 343-372. New York: John Wiley and Sons.
- Valentovic, P., Luxova, M., Kolarovic, L. & Gasparikova, O. (2006). Effect of osmotic stress on compatible solutes content, membrane stability and water relations in two maize cultivars. *Plant Soil Environment*, 4: 186-191.
- Van Asten, P.J.A., Fermont, A.M. & Taulya, G. (2010). Drought is a major yield loss factor for rainfed East African highland banana. *Agriculture Water Management*, 98: 541–552.
- Veihmeyer, F.J. & Hendrickson, A.H. (1936). Essential of irrigation and cultivation of orchards. California Agricultural Experiment Station. *Extension Service Circular*. 50: 1-24 (Revised).
- Vendruscolo, A.C.G., Schuster, L., Pileggi, M., Scapim, C.A., Molinari, H.B.C, Marur, C.J., & Vieira, L.G.C. (2007). Stress-induced synthesis of proline confers tolerance to water deficit in transgenic wheat. *Journal of Plant Physiology* 164(10):1367-1376.
- Vijayakumar, K.R., Dey, S.K., Chandrasekhar, T.K., Devakumar, A.S., Mohankrishma, T., Sanjeeva Rao, P., & Sethuraj, M.R. (1998). Irrigation requirement of rubber trees (*Hevea brasiliensis*) in the subhumid tropics. *Agriculture Water Management*, 35(3): 245-259.
- Wang, T.W., Cosgrove, D.J. & Arteca, R.N. (1993). Brassinosteroid stimulation of hypocotyl elongation and wall relaxation in pakchoi (*Brassica chinensis* cv Lei-Choi). *Plant Physiology* 101: 965-968.
- Weatherly, P.E. (1950). Studies in water relations of the cotton plant. *New Phytologist* 49 (1).81-97.

- Weinert, M. & Simpson, M. (2016). Sub-tropical banana nutrition: matching nutrition requirements to growth demands. Queensland government, Australia.
- Wieble, J., Chacko, E.K. and Downton, W.J.S. (1992). Mangosteen (Garcinia mangostana L.)- a potential crop for tropical northern Australia. *Acta Horticulturae*, 321: 132-137.
- Wilfried, C. (2004). Proline as a measure of stress in tomato plants. *Plant Science* 108: 145-152.
- Wilson, J.R. (1982). Environmental and nutritional factors affecting herbage quality. Pp 111-131. In: J.B. Hacker (Eds.) Nutritional limits to animal production from pastures. Commenwelth Agricultural Bureaux, Farnham Royal, U.K.
- Wu, Q.S., Xia, R.X. & Zou, Y.N. (2008). Improved soil structure and citrus growth after inoculation with three arbuscular mycorrhizal fungi under drought stress. *European Journal Soil Biology*, 44: 122–128.
- Wullschleger, S.D., Yin, T.M., DiFazio, S.P., Tschaplinski, T.J., Gunter, L.E., Davis, M.F., & Tuskan, G.A. (2005). Phenotypic variation in growth and biomass distribution for two advanced-generation pedigrees of hybrid poplar. *Canada Journal Forest Research*. 35:1779-1789.
- Wei, H., Lian, Z.W., Wei, W.T., Yan, Y., Zehong, D., Juhua, L., Meiying, L., Ming, P., Biyu, X. & Zhiqiang, J. (2016). Genome-wide analyses of the bZIP family reveal their involvement in the development, ripening and abiotic stress response in banana. *Scientific reports*, 6: 1-15.
- Yousef, R.M.M., Khalil, S.E. & El-Said, N.A.M. (2013). Response of Echinacea purpurea L. to irrigation water regime and bio-fertilization in sandy soils. *World Applied Science Journal*, 26: 771-782.
- Yueqing, H., Shitou, X., Yi, S., Huiqun, W., Weigui, L., Shengging, S. & Langtao, X. (2016). Brassinolide increase potato root growth in vitro in a dose-dependent way and alleviates salinity stress. *Biomed Research Internationa*, *l* 2016: 1-11.
- Yuriko, O., Keishi, O., Kazuo, S. & Lam-Son, P.T. (2014). Response of plants to water stress. *Frontiers in Plant Science*, 5(86):1-8.
- Zainudin, M. (2000). Root restriction for growth control and precocity in starfruit (*Averrhoa carambola* L.). PhD Thesis, Universiti Putra Malaysia.
- Zeovita, G.M.B.H. (2007). Internal report of Zeovita: Lithovit results.
- Zhang, H., Mallik, A., & Zeng, R.S. (2013). Control of Panama disease of banana by rotating and intercropping with Chinese chive (*Allium tuberosum*): role of plant volatiles. *Journal of chemical ecology*, 39(2): 243-252.
- Zhen, G., Xiao-Gui, L., Li, Z., Shan, L., Xue, Z., Li-Li, Z., Si, S. & Shun-Li, Z. (2017). Spraying exogenous 6-benzyladenine and brassinolide at tasseling increases

- maize yield by enhancing source and sink capacity. *Field Crops Research*, 211: 1–9.
- Zhu, J., Ingram, P. A., Benfey, P.N. & Elich, T. (2011). From lab to field, new approaches to phenotyping root system architecture. *Plant Biology* 14, 310–317.
- Zingaretti, S.M., Rodrigues, F.A., Da Graça, J.P., De Matos Pereira, L. & Lourenço, M.V. (2011). Sugarcane responses at water deficit conditions. *Plant Stress, Intech, Croatia*, 255-276.
- Zlatev, Z. & Stoyanov, Z. (2005). Effects of water stress on leaf water relations of young bean plants. *Journal Central Europe Agriculture*, 6(1):5-14.
- Zurek, D.M., Rayle, D.L., McMorris, T.C. & Clouse, S.D. (1994). Investigation of gene expression, growth kinetics, and wall extensibility during brassinosteroid-regulated stem elongation. *Plant Physiology*, 104: 505-513.
- Zwiazek, T.J. & Blake, T.J. (1990). Effects of pre-conditioning on electrolyte leakage and lipid composition in black spruce (*Picea mariana*) stresses with polyethylene glycol. *Plant Physiology*, 79: 71–77.