



UNIVERSITI PUTRA MALAYSIA

**METAGENETIC ANALYSIS OF GUT MICROBIAL COMMUNITY OF
MALAYSIAN MAHSEER *Tor tambroides* (BLEEKER, 1854)
(CYPRINIDAE)
AND ITS PROBIOTICS POTENTIAL**

TAN CHUN KEAT

FP 2018 53



METAGENETIC ANALYSIS OF GUT MICROBIAL COMMUNITY OF
MALAYSIAN MAHSEER *Tor tambroides* (BLEEKER, 1854) (CYPRINIDAE)
AND ITS PROBIOTICS POTENTIAL

By

TAN CHUN KEAT



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science

December 2017

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of
the requirement for the degree of Master of Science

**METAGENETIC ANALYSIS OF GUT MICROBIAL COMMUNITY OF
MALAYSIAN MAHSEER *Tor tambroides* (BLEEKER, 1854) (CYPRINIDAE)
AND ITS PROBIOTICS POTENTIAL**

By

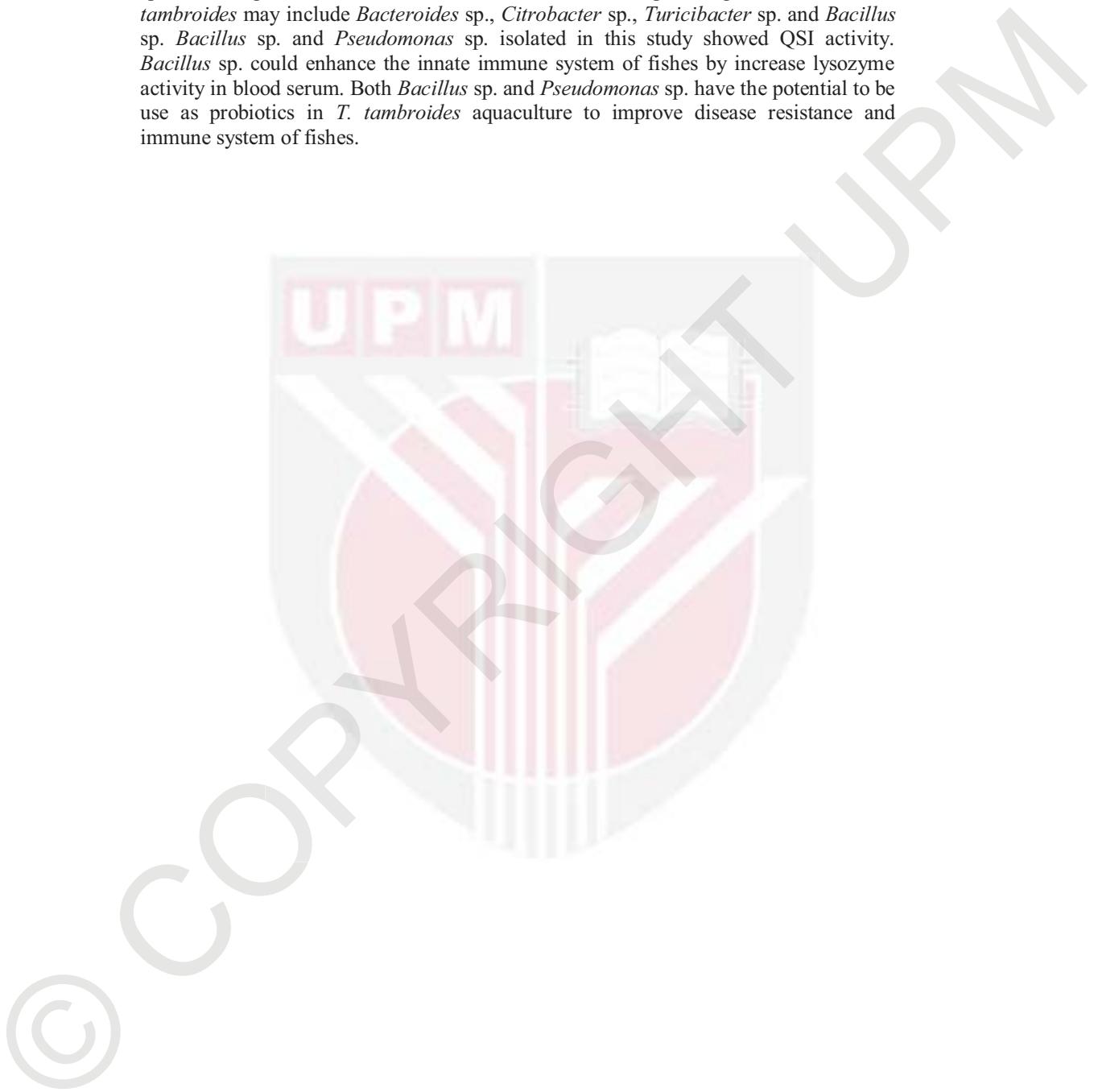
TAN CHUN KEAT

December 2017

**Chair: Natrah Fatin Mohd Ikhsan, PhD
Faculty: Agriculture**

Gut microbiota in vertebrate is complex and contains abundant of diverse beneficial microorganisms important for a dynamic host-microbe interaction. Some of these bacteria may have probiotics properties. In this study, the gut microbiota in wild and captive *T. tambroides* was identified through metagenetic sequencing of 16S rDNA V3-V4 hypervariable regions using Illumina MiSeq. The sequencing data were analyzed using Quantitative Insights into Microbial Ecology (QIIME). Bacteria were isolated from gut of *T. tambroides* to evaluate its safety and examined for its probiotics properties. These selected potential probiotics were fed to *T. tambroides* followed by challenge test to evaluate its effects on growth and disease resistance against *Aeromonas hydrophila*. The metagenetic analyses showed that the gut microbiota in *T. tambroides* was dominated by Firmicutes, Proteobacteria, Fusobacteria and Bacteroidetes. Wild *T. tambroides* gut contained *Cetobacterium* sp. (24.9%), unknown genus from Peptostreptococcaceae family (11.0%), *Bacteroides* sp. (10.1%), *PSB-M-3* from Erysipelotrichaceae family (7.89%) and *Vibrio* sp. (5.4%). Captive *T. tambroides* gut contained *Cetobacterium* sp. (27.9%), *Citrobacter* sp. (10.0%), unknown genus from Peptostreptococcaceae family (8.2%), unknown genus from Aeromonadaceae family (8.2%) and *Turicibacter* sp. (7.0%). The results showed that *Cetobacterium* sp. is the core microbiota in *T. tambroides* gut. Function of this bacterium in *T. tambroides* gut needed to be determined. Three *Aeromonas* sp., two *Bacillus* sp., two *Lysinibacillus* spp., and one *Pseudomonas* sp. were successfully isolated and identified from the wild *T. tambroides* gut sample. Both *Bacillus* sp. and *Pseudomonas* sp. showed quorum sensing inhibition activities while only *Pseudomonas* sp. showed mild antimicrobial activity against *A. hydrophila*. These two bacteria were selected for probiotics feeding experiment. Nevertheless, there was no significant difference in growth of *T. tambroides* fed with these probiotics. The *T. tambroides* juveniles were then challenged with *A. hydrophila* by intra-peritoneal injection after the probiotics feeding experiment. Both *Bacillus* sp. and *Pseudomonas* sp. appeared to be able to improve disease resistance of *T. tambroides* juveniles against *A. hydrophila* infection. Lower mortality was observed in fishes treated with *Bacillus* sp. and *Pseudomonas* sp. as compared to

positive control. Lysozyme activities in *T. tambroides* juveniles fed with *Bacillus* sp. were significantly higher ($P<0.05$) than other treatments. In conclusion, *Cetobacterium* sp. is a core gut microbiota in *T. tambroides*. The other important gut microbiota in *T. tambroides* may include *Bacteroides* sp., *Citrobacter* sp., *Turicibacter* sp. and *Bacillus* sp. *Bacillus* sp. and *Pseudomonas* sp. isolated in this study showed QSI activity. *Bacillus* sp. could enhance the innate immune system of fishes by increase lysozyme activity in blood serum. Both *Bacillus* sp. and *Pseudomonas* sp. have the potential to be used as probiotics in *T. tambroides* aquaculture to improve disease resistance and immune system of fishes.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai
memenuhi keperluan untuk Ijazah Master Sains

**ANALISIS METAGENETIK UNTUK KOMUNITI MIKROBIOTA USUS IKAN
KELAH *Tor tambroides* (BLEEKER, 1854) (CYPRINIDAE) DAN POTENSI
PROBIOTIK**

Oleh

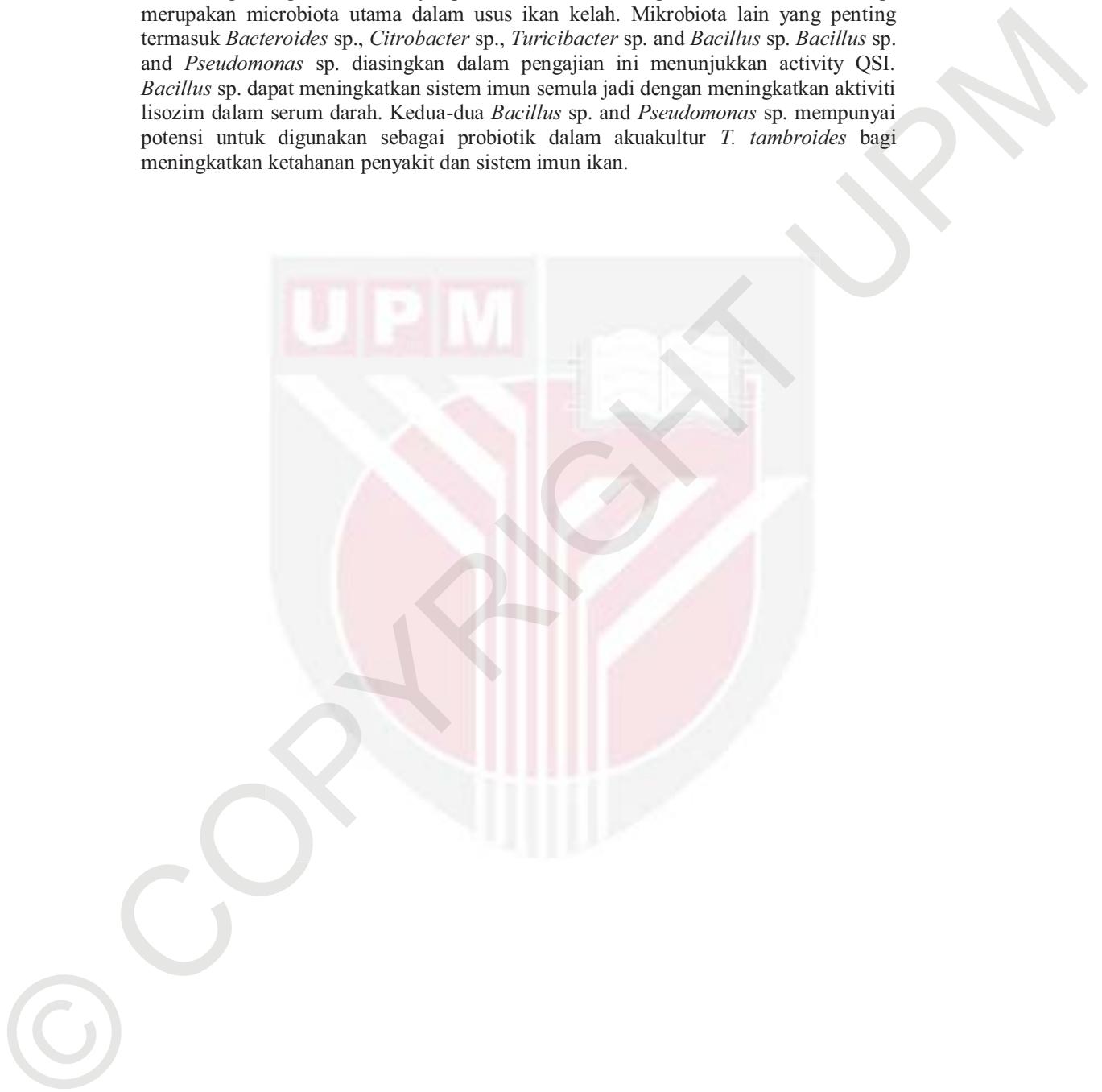
TAN CHUN KEAT

Pengerusi: Natrah Fatin Mohd Ikhsan, PhD

Fakulti: Pertanian

Mikrobiota usus dalam vertebrata adalah kompleks dan mengandungi pelbagai mikroorganisma berfaedah yang penting untuk interaksi dinamik antara mikroorganisma dengan perumahnya. Sesetengah bakteria ini mungkin mempunyai ciri-ciri probiotik. Dalam kajian ini, mikrobiota usus dalam Ikan Kelah (*Tor tambroides*) dikenal pasti melalui penjajaran metagenetik untuk bahagian V3-V4 dalam 16S rDNA menggunakan Illumina MiSeq. Data penjajaran DNA dianalisis menggunakan Quantitative Insights into Microbial Ecology (QIIME). Bakteria diasingkan dari usus *T. tambroides* dikaji untuk keselamatan dan sifat-sifat probiotik. Probiotik berpotensi yang terpilih dibagi kepada *T. tambroides* melalui pemakanan diikuti dengan ujian cabaran untuk menilai kesan-kesan penambahbaikan pertumbuhan dan ketahanan penyakit. Analisis metagenomik menunjukkan bahawa mikrobiota perut di *T. tambroides* didominasi oleh Firmicutes, Proteobacteria, Fusobacteria dan Bacteroidetes. Usus *T. tambroides* liar mengandungi *Cetobacterium* sp. (24.9%), genus yang tidak diketahui dari keluarga Peptostreptococcaceae (11.0%), *Bacteroides* sp. (10.1%), PSB-M-3 dari keluarga Erysipelotrichaceae (7.89%) dan *Vibrio* sp. (5.4%). Usus *T. tambroides* peliharaan mengandungi *Cetobacterium* sp. (27.9%), *Citrobacter* sp. (10.0%), genus yang tidak diketahui dari keluarga Peptostreptococcaceae (8.2%), genus yang tidak diketahui dari keluarga Aeromonadaceae (8.2%) dan *Turicibacter* sp. (7.0%). Ini telah menunjukkan bahawa *Cetobacterium* spp. adalah mikrobiota utama dalam usus *T. tambroides*. Fungsi bakteria ini dalam usus *T. tambroides* perlu ditentukan. Beberapa bakteria berjaya diasingkan dan dikenal pasti dari sampel usus ikan kelah liar seperti tiga *Aeromonas* sp., dua *Bacillus* sp., dua *Lysinibacillus* sp., and satu *Pseudomonas* sp. Ujian QSI menunjukkan bahawa *Bacillus* sp. dan *Pseudomonas* sp. menunjukkan aktiviti perencutan penginderaan kuorum manakala hanya *Pseudomonas* sp. menunjukkan sifat antimikrobal yang sederhana terhadap *A. hydrophila*. Kedua-dua bakteria ini dipilih untuk eksperimen pemakanan berprobiotik. Walaubagaimanapun, tiada perbezaan yang ketara dalam pertumbuhan *T. tambroides* yang diberi makan dengan dua probiotik ini. Juvana *T. tambroides* telah dicabar dengan *A. hydrophila* melalui suntikan *intra-peritoneal* selepas eksperimen pemakanan berprobiotik. Kedua-dua *Bacillus* sp. dan *Pseudomonas* sp. dapat meningkatkan ketahanan penyakit juvana *T. tambroides* terhadap jangkitan *A. hydrophila*. Kadar kematian yang lebih rendah dikesan dalam ikan yang menerima rawatan *Bacillus* sp.

dan *Pseudomonas* sp. berbanding dengan rawatan kontrol positif. Aktiviti lisozim dalam juvana *T. tambroides* menerima rawatan *Bacillus* sp. adalah lebih tinggi ($P<0.05$) berbanding dengan rawatan yang lain. Secara kesimpulan, *Cetobacterium* sp. merupakan mikrobiota utama dalam usus ikan kelah. Mikrobiota lain yang penting termasuk *Bacteroides* sp., *Citrobacter* sp., *Turicibacter* sp. and *Bacillus* sp. *Bacillus* sp. and *Pseudomonas* sp. diasingkan dalam pengajian ini menunjukkan activity QSI. *Bacillus* sp. dapat meningkatkan sistem imun semula jadi dengan meningkatkan aktiviti lisozim dalam serum darah. Kedua-dua *Bacillus* sp. and *Pseudomonas* sp. mempunyai potensi untuk digunakan sebagai probiotik dalam akuakultur *T. tambroides* bagi meningkatkan ketahanan penyakit dan sistem imun ikan.



ACKNOWLEDGEMENT

This thesis becomes a reality with the kind supports and helps of many individuals. I would like to extend my sincere thanks to all of them

Foremost, I would like to express a million of sincere gratitude and appreciation to my supervisor Dr. Natrah Fatin Mohd Ikhsan for the motivation, encouragement, supervision and guidance in this study. I am also deeply grateful to my co-supervisors, Dr. Anjas Asmara Samsudin and Dr. Iswan Budy Suyub for their supervision, help and problem solving. As a postgraduate student, I am truly fortunate to have them as my supervisors and mentors during my Master study. Without their encouragement and support, this thesis would not have been completed with quality.

In addition, many thanks go to my colleagues, Mrs. Nazrien and Mrs. Marilyn in Agro-Biotechnology Institute (ABI) for their assistance during the feeding experiment and data collection. I thanked Agro-Biotechnology Institute (ABI) for allowing me to use the equipments and hatchery facilities for feeding experiment and challenge test. I truthfully appreciate the funding for this study from Ministry of Science, Technology and Innovation (MOSTI) under the flagship project of “Development of Sustainable Malaysian Mahseer/ Kelah/ Empurau Aquaculture”.

I am gratefully thanking Malaysia Genome Institute (MGI) for allowing me to use their laboratory for library preparation and MiSeq sequencing. I thanked Mr. Faizal and Mr. Mohd Noor Mat Isa for allowing me to use the computer and server facilities for bioinformatics data analysis. A special thanks to Ms. Yufi from Malaysia Genome Institute (MGI) for guiding me in the using of QIIME for bioinformatics data analysis. In my daily work, I have been blessed with friendly lab buddies Shariza Azizan, Nur Ain Yahya, Wee Wen Chen and Firdaus Asnawi for their guidances, assistances and comments.

I would specially like to thank my wife Ong Chee Huey for her selfless support, love, encouragement and continuous advice. She is always there to share the up and down in my life. I am gratefully thanked my wife for delivered our precious daughter, Tan Yu En during my Master study. I would like to thank my wife for her constant support with me when I was reading and writing my thesis.

Lastly, I am grateful to my beloved parents, grandparents and my siblings for their encouragement as well as understanding for supporting me spiritually throughout my life. Greatest gratitude was expressed to my parents for their help in postpartum period of my wife during my Master study.

I wish every one of you a great health with a lot of happiness.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Natrah Fatin Mohd Ikhsan, PhD

Senior lecturer

Faculty of Agriculture

Universiti Putra Malaysia

(Chairman)

Anjas Asmara @ Ab Hadi Bin Samsudin, PhD

Associate Professor

Faculty of Agriculture

Universiti Putra Malaysia

(Member)

Iswan Budy Suyub, PhD

Senior Lecturer

Faculty of Agriculture

Universiti Putra Malaysia

(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of Chairman of
Supervisory
Committee:

Natrah Fatin Mohd Ikhsan

Signature: _____

Name of Member of
Supervisory
Committee:

Anjas Asmara @ Ab Hadi Bin Samsudin

Signature: _____

Name of Chairman of
Supervisory
Committee:

Iswan Budy Suyub

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xv
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xix
 CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 <i>Tor spp.</i> and <i>Tor tambroides</i>	4
2.1.1 Nomenclature, taxonomy and distribution	4
2.1.2 General morphology of <i>T. tambroides</i>	5
2.1.3 Reproduction and life cycle of <i>T. tambroides</i>	7
2.1.4 Nutritional requirement of <i>T. tambroides</i>	7
2.2 Problems in <i>T. tambroides</i> farming	8
2.2.1 Depletion of wild stocks and slow growth rate	8
2.2.2 Diseases	9
2.2.3 Breeding	10
2.3 Gut microbiota in aquatic animals	10
2.4 Probiotics and their mechanisms of action	11
2.4.1 Enhancement of the epithelial barrier, increase adhesion to intestinal mucosa and competitive exclusion for nutrient and adhesion site	12
2.4.2 Production of anti-microorganism compounds	13
2.4.3 Modulation of immune system	13
2.4.4 Interference with quorum sensing signalling	14
2.4.4.1 Quorum sensing (QS) and other sensing mechanism	14
2.4.4.2 Quorum quenching (QQ) or quorum sensing inhibition	15
2.5 Benefits and effects of probiotics in aquaculture	15
2.5.1 Immune systems and lysozyme activity	19
2.5.2 Superoxide dismutase (SOD) activity	19
2.6 Deoxyribonucleic acid (DNA), 16S ribosomal RNA gene and Next Generation Sequencing	19
2.6.1 Quantitative Insights into Microbial Ecology (QIIME) Pipeline	22

3	16S rRNA METAGENETIC ANALYSIS OF GUT MICROBIAL COMMUNITY OF WILD AND CAPTIVE MALAYSIAN MAHSEER <i>Tor tambroides</i>	23
3.1	Introduction	23
3.2	Materials and Methods	23
3.2.1	Fish sampling and species verification using cytochrome b gene	23
3.2.1.1	Fish sampling at Kenyir Lake and hatchery	24
3.2.1.2	DNA extraction of <i>T. tambroides</i>	25
3.2.1.3	Polymerase chain reaction (PCR) amplification of cytochrome gene and gel electrophoresis	25
3.2.1.4	PCR purification of <i>T. tambroides</i> cytochrome b gene sequences	27
3.2.1.5	DNA sequencing of <i>T. tambroides</i> cytochrome b gene sequences	27
3.2.1.6	Fish species identification using NCBI Basic Local Alignment Search Tool (BLAST)	27
3.2.2	Comparison of gut microbial community in wild <i>T. tambroides</i> from Kenyir Lake and captive <i>T. tambroides</i> from hatchery	27
3.2.2.1	Fish dissection and gut removal	27
3.2.2.2	Metagenome DNA extraction from wild and captive <i>T. tambroides</i> gut samples	28
3.2.2.3	PCR amplification of V3-V4 region of 16S rDNA of gut microbiota	29
3.2.2.4	Size verification of the V3-V4 sequences of 16S rDNA	29
3.2.2.5	PCR clean up of V3-V4 sequences of 16S rDNA	30
3.2.2.6	Measurement of concentration of PCR product	31
3.2.2.7	Index PCR to label the V3-V4 sequences of 16S rDNA with unique indexes pairs	31
3.2.2.8	PCR clean up of the indexed library	32
3.2.2.9	Library concentration determination	32
3.2.2.10	Library validation	32
3.2.2.11	Library Quantification, Normalization and Pooling	32
3.2.2.12	Library denaturation and Illumina MiSeq Sequencing	33
3.2.2.13	Sequences processing and bioinformatics workflow using Quantitative Insights into Microbial Ecology (QIIME) pipeline	34

	3.2.2.14	Phylogenetic Investigation of Communities by Reconstruction of Unobserved States (PICRUSt) analyses for predictive functions	37
3.3	Results		37
3.3.1	Wild and captive <i>T. tambroides</i> measurements and identification		37
3.3.2	Metagenetic sequencing and bioinformatics data analysis using QIIME		38
3.3.3	MiSeq sequencing and data generation		39
3.3.4	Alpha rarefactions analysis of wild and captive <i>T. tambroides</i> gut microbiome		40
3.3.5	Gut taxonomy of wild and captive <i>T. tambroides</i>		42
3.3.6	Beta diversity analysis of wild and captive <i>T. tambroides</i> gut microbiome		44
3.3.7	PICRUSt analyses for predictive functions of gut microbiome		46
3.4	Discussion		51
3.4.1	Species identification using cytochrome b gene		51
3.4.2	Environmental factors and feed effect on wild and captive <i>T. tambroides</i> gut microbiota		51
3.4.3	Microbiome taxonomy of wild and captive <i>T. tambroides</i>		52
3.4.4	Alpha rarefactions analysis of wild and captive <i>T. tambroides</i> gut microbiome		52
3.4.5	Gut microbiota in wild and captive <i>T. tambroides</i> guts		53
3.4.6	Beta diversity analysis of wild and captive <i>T. tambroides</i> gut microbiome		54
3.4.7	Analysis of wild and captive <i>T. tambroides</i> gut microbiome for species abundance		55
3.4.8	PICRUSt function prediction of gut microbiome in wild and captive <i>T. tambroides</i>		57
3.5	Conclusion		58
4	SCREENING OF POTENTIAL PROBIOTICS ISOLATED FROM MALAYSIAN MAHSEER <i>Tor tambroides</i> GUT		59
4.1	Introduction		59
4.2	Materials and Methods		59
4.2.1	Isolation and growth of culturable gut microbiota		59
4.2.2	Deoxyribonucleic acid (DNA) extraction of isolated bacteria		60
4.2.3	16S rDNA-PCR amplification of bacterial isolates		60
4.2.4	Polymerase chain reaction (PCR) purification and sequencing		61
4.2.5	Species Identification using EzBioCloud		62
4.2.6	Antimicrobial assay		62

4.2.7	Quorum Sensing Inhibition (QSI) assay	62
4.2.8	Quantification of biofilm produced by potential probiotics	62
4.2.9	Phylogenetic analysis of the selected potential probiotics	63
4.2.10	In vitro safety evaluation of the selected potential probiotics by hemolysis assay	63
4.2.11	In vivo safety evaluation of selected potential probiotics	63
4.3	Results	63
4.3.1	Species identification of the isolated bacteria from <i>T. tambroides</i> gut	63
4.3.2	Antimicrobial activity of isolated bacteria from <i>T. tambroides</i> guts	66
4.3.3	Quorum Sensing Inhibition (QSI) assay of isolated bacteria from <i>T. tambroides</i> guts	68
4.3.4	Biofilm production by isolated bacteria from <i>T. tambroides</i> guts	70
4.3.5	Phylogenetic analysis of the selected potential probiotics	71
4.3.6	In vitro safety evaluation of selected potential probiotics by hemolysis assay	73
4.3.7	In vivo safety evaluation of potential probiotics	73
4.4	Discussion	74
4.4.1	Antimicrobial activity of selected probiotics against <i>A. hydrophila</i>	74
4.4.2	Quorum sensing inhibition (QSI) activity of selected probiotics tested using <i>C. violaceum</i> CV026	74
4.4.3	Biofilm formation ability of the selected probiotics	74
4.4.4	Safety evaluation of selected potential probiotics	75
4.5	Conclusion	75
5	EVALUATION OF SELECTED POTENTIAL PROBIOTICS EFFECTS ON GROWTH AND DISEASE RESISTANCE OF MALAYSIAN MAHSEER <i>Tor tambroides</i>	76
5.1	Introduction	76
5.2	Materials and Methods	76
5.2.1	Feeding experiment with probiotics added feeds	76
5.2.2	Challenge test with <i>Aeromonas hydrophila</i> strain AH1N	78
5.2.3	Superoxide Dismutase (SOD) analysis	78
5.2.4	Lysozyme activity analysis	79
5.3	Results	80
5.3.1	Probiotics feeding experiment and growth performance of <i>T. tambroides</i>	80
5.3.2	Challenge test of probiotics fed <i>T. tambroides</i> with pathogenic <i>A. hydrophila</i> AH1N	81

5.3.3	Lysozyme activity in blood serum of <i>T. tambroides</i> after probiotics feeding experiment	84
5.3.4	Superoxide dismutase (SOD) activity in blood serum of <i>T. tambroides</i> after probiotics feeding experiment	85
5.4	Discussion	86
5.4.1	Probiotics feeding and challenge test with <i>A. hydrophila</i> AH1N	86
5.4.2	Lysozyme activity after probiotics feeding	87
5.4.3	Superoxide dismutase activity after probiotics feeding	87
5.5	Conclusion	87
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS	88
REFERENCES		91
APPENDICES		120
BIODATA OF STUDENT		121

LIST OF TABLES

Table		Page
2.1	List of species in Genus <i>Tor</i>	4
2.2	Summary of probiotics used in aquaculture and their results	16
3.1	Primer pair used in PCR amplification of cytochrome b gene	26
3.2	PCR reaction composition for amplification of cytochrome b gene	26
3.3	PCR protocol for amplification of cytochrome b gene	26
3.4	Primers used in PCR amplification of V3-V4 region of 16S rDNA	29
3.5	PCR reaction composition for amplification of V3-V4 region of 16S rDNA	29
3.6	PCR protocol for amplification of V3-V4 region of 16S rDNA	29
3.7	Indexes used for each sample (Nextera XT Index Kit, Illumina Inc.)	31
3.8	PCR reaction composition for Index PCR	31
3.9	PCR Protocol for Index PCR	32
3.10	Quantitative Polymerase Chain Reaction (qPCR) reaction composition	33
3.11	Quantitative Polymerase Chain Reaction (qPCR) protocol	33
3.12	List of software or script used in bioinformatics data analysis	35
3.13	Measurement data and gut digesta colour for wild and captive <i>T. tambroides</i>	37
3.14	BLAST results of six <i>T. tambroides</i> cytochrome b gene sequence	38
3.15	Summary of gut microbiome in wild and captive <i>T. tambroides</i>	39
3.16	Summary of metagenetic statistics of wild and captive <i>T. tambroides</i> gut microbiome	42
3.17	Mann-Whitney U Test for 15 most abundant OTUs at genus level for wild and captive samples respectively	46
4.1	Primers used in PCR	60
4.2	PCR reaction composition	61
4.3	PCR protocol	61
4.4	Identification results of 16S rDNA from bacteria isolated from <i>T. tambroides</i> guts	65
4.5	Assay for antimicrobial activity	66
4.6	Inhibition zone of different concentration of potential probiotics on <i>A. hydrophila</i> AH1N	67
4.7	Assay for quorum sensing inhibition activity	68
4.8	Inhibition zone diameter for each treatment concentration of potential probiotics on purple pigment production by <i>Chromobacterium violaceum</i> (CV026)	69

4.9	Indirect biofilm production measurement using OD600	70
5.1	Experimental design for feeding experiment	77
5.2	Growth of <i>T. tambroides</i> fed with different treatment in 12 weeks	80



LIST OF FIGURES

Figure		Page
2.1	Malaysian Red Mahseer, <i>Tor tambroides</i>	6
2.2	Median lobe on the lower lips	6
3.1	Map of Peninsular Malaysia which showed the location of Agro-Biotechnology Institute (ABI) at Selangor and Kenyir Lake at Terengganu	24
3.2	Gel electrophoresis image of 6 samples of <i>T. tambroides</i> cytochrome b gene	38
3.3	Venn diagram showing the unique and shared gut microbiota of wild and captive <i>T. tambroides</i>	40
3.4	Alpha rarefaction curves of wild and captive <i>T. tambroides</i> gut microbiome	41
3.5	Relative abundance of phyla found in wild <i>T. tambroides</i> gut microbiome	42
3.6	Relative abundance of phyla found in captive <i>T. tambroides</i> gut microbiome	43
3.7	Comparison of top 15 most abundant observed bacteria in wild and captive <i>T. tambroides</i> guts	44
3.8	Principal Coordinates Analysis (PCoA) plots of beta diversity analysis based on weighted UniFrac distance metric	45
3.9	Top 10 most abundant predictive functions of gut microbiome in wild and captive <i>T. tambroides</i>	47
3.10	Comparison of bile secretion between wild and captive <i>T. tambroides</i> gut microbiome	48
3.11	Comparison of lysine biosynthesis between wild and captive <i>T. tambroides</i> gut microbiome	48
3.12	Comparison of glycine, serine and threonine metabolism between wild and captive <i>T. tambroides</i> gut microbiome	49
3.13	Comparison of starch and sucrose metabolism between wild and captive <i>T. tambroides</i> gut microbiome	49
3.14	Comparison of porphyrin and chlorophyll metabolism between wild and captive <i>T. tambroides</i> gut microbiome	50
3.15	Comparison of carbohydrate metabolism between wild and captive <i>T. tambroides</i> gut microbiome	50
4.1	Gel electrophoresis results of the 16S rDNA PCR product from 30 isolates	64
4.2	Antimicrobial activity assay using <i>A. hydrophila</i> and chloramphenicol (2ng/ μ l) as positive control	67
4.3	QSI assay using <i>Chromobacterium violaceum</i> (CV026) with 0.5 ppm HHL	69
4.4	Biofilm stained by crystal violet	70
4.5	Phylogenetic analysis of potential probiotic <i>Bacillus</i> sp.	71
4.6	Phylogenetic analysis of potential probiotic <i>Pseudomonas</i> sp.	72

4.7	Hemolysis assay of selected potential probiotics using blood agar	73
5.1	Timeline of feeding experiment and challenge test	78
5.2	Weight gain of <i>T. tambroides</i> fed with different probiotics in 12 weeks	81
5.3	Survival (%) of <i>T. tambroides</i> after challenged with <i>A. hydrophila</i> AH1N	82
5.4	Cummulative mortality of <i>T. tambroides</i> in 14 days after challenged with <i>A. hydrophila</i> AH1N	82
5.5	<i>T. tambroides</i> fed with probiotics and <i>T. tambroides</i> fed without probiotics after challenged with <i>A. hydrophila</i>	84
5.6	Lysozyme activity in blood serum of <i>T. tambroides</i> after probiotics feeding experiment	85
5.7	Superoxide dismutase (SOD) activities in blood serum of <i>T. tambroides</i> after probiotics feeding experiment	86

LIST OF ABBREVIATIONS

ABI	Agro-Biotechnology Institute
ACP	Alternative Complement Pathway
AHLs	N-acyl Homoserine Lactones
AI-2	Autoinducer-2
ANOVA	Analysis Of Variance
ATP	Adenosine Triphosphate
BLAST	Basic Local Alignment Search Tool
bp	Base pair
CFU	Colony Forming Unit
CI	Chloroform:isoamyl alcohol
COI	Cytochrome C Oxidase subunit I
CytB	Cytochrome b
D-loop	Displacement Loop
DAH	Days After Hatching
DCs	Dendritic Cells
DNA	Deoxyribonucleic Acid
dNTPs	Deoxynucleotide Triphosphates
DO	Dissolve Oxygen
DOF	Department Of Fisheries Malaysia
DS	Diffusion Sensing
dsDNA	Double Stranded Deoxyribonucleic Acid
EB	Elution Buffer
EFA	Essential Fatty Acids
ES	Efficiency Sensing
FAO	Food and Agricultural Organization of the United Nations
FCR	Feed Conversion Ratio
Gb	Giga Base Pair
GPS	Global Positioning System
HHL	N-Hexanoyl-L-Homoserine Lactone
IACUC	Institutional Animal Care and Use Committee
IFRPC	Indigenous Fish Research and Production Centre, Sarawak
Ig	Immunoglobulin
IgA	Immunoglobulin A
ITIS	Integrated Taxonomic Information System
IUCN	International Union for Conservation of Nature
LAB	Lactic Acid Bacteria
LB	Luria-Bertani Broth
LBA	Luria-Bertani Agar
MGI	Malaysia Genome Institute
MPO	Myeloperoxidase
MSR	MiSeq Reporter
n-3	Omega 3
n-6	Omega 6
n.d.	No Date
NA	Not Available
NCBI	National Center for Biotechnology Information
NGS	Next Generation Sequencing

NRC	National Research Council
OD	Optical Density
OTUs	Operational Taxonomic Units
PBS	Phosphate Buffered Saline
PC	Principal Coordinate
PCI	Phenol:chloroform:isoamyl alcohol
PCoA	Principal Coordinates Analysis
PCR	Polymerase Chain Reaction
PEAR	Paired-End reAd mergeR
PGM	Personal Genome Machine
PICRUSt	Phylogenetic Investigation of Communities by Reconstruction of Unobserved States
ppm	Parts Per Million
QIIME	Quantitative Insights Into Microbial Ecology
qPCR	Quantitative Polymerase Chain Reaction
QS	Quorum Sensing
QSI	Quorum Sensing Inhibition
QQ	Quorum Quenching
RAS	Recirculating Aquaculture System
RNA	Ribonucleic Acid
ROS	Reactive Oxygen Species
rpm	Revolutions per minute
rRNA	Ribosomal Ribonucleic Acid
SBs	Sequence By Synthesis
SD	Standard Deviation
SGR	Specific Growth Rate
SMRT	Single Molecule Real-Time Sequencing
SOD	Superoxide Dismutase
SOLiD	Sequencing by Oligonucleotide Ligation and Detection
spp.	Species
SRA	Sequence Read Archive
TAE	Tris-acetate-EDTA
TE	Tris-EDTA
tRNAs	Transfer Ribonucleic Acids
rRNAs	Ribosoma Ribonucleic Acids
TSA	Tryptic Soy Agar
TSB	Tryptic Soy Broth
UPM	Universiti Putra Malaysia
UV	Ultraviolet

CHAPTER 1

INTRODUCTION

Aquaculture is considered as one of the fastest-growing sector in food producing industry (FAO, 2014). Besides its importance in food and nutrition security, it also provides employment to millions of people worldwide and supports their livelihood. Aquaculture involves the farming of aquatic organisms in inland and coastal areas that intervene in the rearing process to enhance production (FAO, 2015) as oppose to capture fishery which involves harvesting of naturally occurring living resources in both marine and freshwater environments. While capture fishery production remained relatively static since the late 1980s, aquaculture has been responsible for the impressive growth in the supply of fish for human consumption. Capture fisheries production achieved 93.4 million tonnes in 2014 as compared to 73.8 million tonnes in aquaculture which equals to 44.1% of total world production (FAO, 2016). Aquaculture production is expected to be increasing in the following years and may achieve over 50% of total world production. China has played a major role in this growth as it represents more than 60 percent of the world aquaculture production (FAO, 2016).

According to Department of Fisheries Malaysia statistics (DOF, 2016) in year 2015, total landing of capture fisheries in Malaysia was 1,486,051 tonnes as compared to 1,458,128 tonnes on year 2014 (DOF, 2015). Value of capture fisheries in year 2015 was RM 9.32 million which showed an increment of 6.1% from year 2014. Aquaculture production was 506,465.25 tonnes in year 2015 which equals to 25.4% of total production in Malaysia. Total fishes production in aquaculture of Malaysia in year 2015 were dominated by freshwater catfish (45.2%) followed by Red Tilapia (27.1%), River Catfish (12.4%), Black Tilapia (4.5%) and other species (10.8%).

Tor tambroides is one of the most expensive freshwater fish in Malaysia. The market price of this fish in Malaysia ranges from RM 370-700 per kg (Azuadi et al., 2013). However, *T. tambroides* production was only 24.7 tonnes in year 2015 which was 0.02% of total aquaculture production in year 2015 (DOF, 2016). This showed huge potential for development of *T. tambroides* aquaculture. However, there are some obstacles that needed to be overcome to enable success aquaculture for *T. tambroides*. IUCN Red List of Threatened Species did not classify *T. tambroides* as endangered species due to data deficiency but the global population of this species has been observed to be decreasing due to overfishing, logging, deforestation, agriculture activities and anthropogenic activities causing river morphology modification and water flow interruption (Kottelat, 2012). The growth of mahseers reported to be slow (Lee et al., 2014; Chatta et al., 2015) which may discourage its aquaculture production.

As aquaculture activities intensify and expand, there is higher chance of disease and problems caused by parasites, bacteria, viruses, fungi and other undiagnosed and emerging pathogens. This is because maintenance of large numbers of fish crowded together in a small area provides a very conducive environment for the development

and spread of infectious diseases. Disease is a major constraint to the aquaculture industry, hampered both economic and social development in many countries (Bondad-Reantaso et al., 2005). The gills of Malaysian mahseer, *T. tambroides* obtained from Tasik Kenyir Reservoir Malaysia were frequently infected by a *Myxobolus* species (Székely et al., 2012). Golden mahseer, *Tor putitora* was susceptible to *Aeromonas hydrophila* infection (Kumar et al., 2016a). Eye lesions in golden mahseer, *T. putitora* was caused by *Pseudomonas koreensis* (Shahi and Mallik, 2014). *Tor tambera* from aquaculture fishponds were infested by Asian fish tapeworm (Muchlisin et al., 2015). Kesarcodi-Watson et al. (2008) reported the fastest treatment to diseases is the use of antimicrobial drugs but misuse of this drugs will eventually contributed to the emergence of antibiotic resistant bacteria. A feasible alternative to solve this problem is the use of probiotics or beneficial bacteria (Balcázar et al., 2006). Probiotics, prebiotics, immunostimulants, plant products and oral vaccines are some of the solutions in fish nutrition aiming to improve fish growth, feed efficiency, health, stress tolerance and resistance to diseases (Olivia-Teles, 2012; Newaj-Fyzul & Austin, 2014).

The term probiotic was defined as “a mono- or mixed culture of live microorganisms that when applied to animals or man, affect beneficially the host by improving the properties of the indigenous microflora” (Moriarty et al., 2005). Moriarty (1998) extended the definition for aquaculture to include the addition of natural bacteria to water environment in which the aquatic animals live. Probiotic strains have abilities to inhibit pathogenic bacteria both *in vitro* and *in vivo* through different mechanisms. These include enhancement of the epithelial barrier, increase adhesion to intestinal mucosa, competitive exclusion, production of anti-microorganism compounds, immune response enhancement, increase antiviral effects (Balcázar et al., 2006) and quorum sensing inhibition (Defoirdt et al., 2004; Kaufmann et al., 2008; Kalia, 2013). Addition of probiotics in feed may improve disease resistance and immune response of Malaysian mahseer. Besides, probiotics such as *Bacillus* spp. can produce digestive enzyme (Kesarcodi-Watson et al., 2008) to facilitate digestion of feed in the stomach and intestines of mahseer thus improve nutrient adsorption and growth rate. Lara-Flores et al. (2003) concluded that use of probiotic can reduce the amount of feed required for animal growth thus reduced the production cost.

There are more than 99% of prokaryotes in the environment that are uncultivable in laboratory and this limits our understanding of microbial physiology, genetics and community ecology (Schloss & Handelsman, 2005). The development of Next Generation Sequencing (NGS) technology allows the recognition of discrete populations (culturable and uncultivable) based on DNA sequences in the environmental samples (Konstantinidis & Rosselló-Móra, 2015). Gut microbiota of grass carp influenced by amount of food in the gut, diet and environment (Ni et al., 2014a). Fecal bacterial communities of *Epinephelus fuscoguttatus*, *Epinephelus sexfasciatus* and *Atule mate* collected from different geographical location showed same classes of bacteria as core microbiomes but the proportions of these bacteria were strongly varied (Hennersdorf et al., 2016). There were differences in gut microbial composition of wild and aquaculture origin *Seriola lalandi* (Ramírez & Romero, 2017). It is anticipated that the gut microbial community of wild and captive *T. tambroides* are different.

Since the wild stock of *T. tambroides* is decreasing and the growth of this species is remarkably slow, more research should be focus on the nutritional requirements, breeding and diseases control to develop a sustainable aquaculture production for this species. There has not been any report on the comparison of gut microbiome of wild and captive Malaysia mahseer. This study aims to identify and differentiate the gut microbiota in wild and captive Malaysian mahseer subsequently lead to the selection of bacteria with potential probiotics properties that can improve growth and disease resistance of Malaysian mahseer.

Objectives

1. To identify and compare gut microbiota of wild and captive *T. tambroides* by metagenetic sequencing
2. To examine the probiotics properties of bacteria isolated from gut of *T. tambroides*
3. To evaluate the effects of selected probiotics on growth and disease resistance of *T. tambroides* against *Aeromonas hydrophila*

REFERENCES

- Acharya, A. D., Behera, B. K., Paria, P., Bhowmick, S., Parida, P. K. & Das, B. K. (2018). Isolation, identification and characterization of *Klebsiella pneumoniae* from infected farmed Indian Major Carp *Labeo rohita* (Hamilton 1822) in West Bengal, India. Aquaculture 482: 111-116.
- Adeoye, A. A., Yomla, R., Jaramillo-Torres, A., Rodiles, A., Merrifield, D. L., Davies, S. J. (2016). Combined effects of exogenous enzymes and probiotic on Nile tilapia (*Oreochromis niloticus*) growth, intestinal morphology and microbiome. Aquaculture 463:61-70.
- Akhter, N., Wu, B., Memon, A. M. & Mohsin, M. (2015). Probiotics and prebiotics associated with aquaculture: A review. Fish & Shellfish Immunology 45(2): 733-741.
- Albert, M. J., Mathan, V. I. & Baker, S. J. (1980). Vitamin B12 synthesis by human small intestinal bacteria. Nature 283: 781-782.
- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K. & Walter, P. (2002). Innate Immunity. In *Molecular Biology of the Cell. 4th edition*. Garland Science, New York. Retrieved 16 July 2017 from <https://www.ncbi.nlm.nih.gov/books/NBK26846/>.
- Almada, C. N. D., Almada. C. N., Martinez, R. C. R. & Sant'Ana, A. S. (2016). Paraprobiotics: Evidences on their ability to modify biological responses, inactivation methods and perspectives on their application in foods. Trends in Food Science & Technology 58: 96-114.
- Altinok, I., Kayis, S. & Capkin, E. (2006). *Pseudomonas putida* infection in rainbow trout. Aquaculture 261(3): 850-855.
- Altinok, I., Balta, F., Capkin, E. & Kayis, S. (2007). Disease of rainbow trout caused by *Pseudomonas luteola*. Aquaculture 273(4): 393-397.
- Aly, S. M., Ahmed, Y. A. G., Ghareeb, A. A. A., Mohamed, M. F. (2008). Studies on *Bacillus subtilis* and *Lactobacillus acidophilus*, as potential probiotics, on the immune response and resistance of Tilapia nilotica (*Orechromis niloticus*) to challenge infections. Fish & Shellfish Immunology 25: 128-136.
- Ambak, M. A., Ashraf, A. H., Budin, S. (2007). Conservation of the Malaysian mahseer in Nenggiri basin through community action. Mahseer: The Biology, Culture and Conservation. Malaysian Fisheries Society Occasional Publication No. 14, Kuala Lumpur, Malaysia, pp. 217–228.
- Aoudia, N., Rieu, A., Briandet, R., Deschamps, J., Chluba, J., Jego, G., Garrido, C. & Guzzo, J. (2016). Biofilms of *Lactobacillus plantarum* and *Lactobacillus fermentum*: Effect on stress responses, antagonistic effects on pathogen growth and immunomodulatory properties. Food Microbiology 53(A): 51-59.

- Araki, H., Cooper, B. & Blouin, M. S. (2009). Carry-over effect of captive breeding reduces reproductive fitness of wild-born descendants in the wild. *Biology Letters* 5: 621-624.
- Araújo, C., Muñoz-Atienza, E., Nahuelquín, Y., Poeta, P., Igrejas, G., Hernández, P. E., Herranz, C. & Cintas, L. M. (2015). Inhibition of fish pathogens by the microbiota from rainbow trout (*Oncorhynchus mykiss*, Walbaum) and rearing environment. *Anaerobe* 32: 7-14.
- Asaduzzaman, M., Sofia, E., Shakil, A., Haque, N. F., Khan, M. N. A., Ikeda, D., Kinoshita, S. & Abol-Munafi, A. B. (2018a). Host gut-derived probiotic bacteria promote hypertrophic muscle progression and upregulate growth-related gene expression of slow-growing Malaysian Mahseer *Tor tambroides*. *Aquaculture Reports* 9: 37-45.
- Asaduzzaman, M., Iehata, S., Akter, S., Kader, M. A., Ghosh, S. K., Khan, M. N. A. & Abol-Munafi, A. B. (2018b). Effects of host gut-derived probiotic bacteria on gut morphology, microbiota composition and volatile short chain fatty acids production of Malaysian Mahseer *Tor tambroides*. *Aquaculture Reports* 9: 53-61.
- Austin, B. & Austin, D. A. (2012). *Bacterial Fish Pathogens: Disease of Farmed and Wild Animals*. Springer, Dordrecht, Netherland.
- Azuadi, N.M., S.S. Siraj, S.K. Daud, A. Christianus, S.A. Harmin, S. Sungan and R. Britin. (2011). Enhancing ovulation of Malaysian mahseer (*Tor tambroides*) in captivity by removal of dopaminergic inhibition. *Journal of Fisheries and Aquatic Science* 6: 740-750.
- Azuadi, N. M., Siraj, S. S., Daud, S. K., Christianus, A., Harmin, S. A., Sungan, S. & Britin, R. (2013). Induced Ovulation, Embryonic and Larval Development of Malaysian Mahseer, *Tor tambroides* (Bleeker, 1854) in Captivity. *Asian Journal of Animal and Veterinary Advances* 8: 761-774.
- Bai, F., Han, Y., Chen, J. & Zhang, X. H. (2008). Disruption of quorum sensing in *Vibrio harveyi* by the AiiA protein of *Bacillus thuringiensis*. *Aquaculture* 274: 36-40.
- Balcázar, J.L., Blas, I. D., Ruiz-Zarzuela, I., Cunningham, D., Vendrell, D., Múzquiz, J. L. (2006). The role of probiotics in aquaculture. *Veterinary Microbiology* 114: 173-186.
- Beck, B. R., Kim, D., Jeon, J., Lee, S. M., Kim, H. K., Kim, O. J., Lee, J. I., Suh, B. S., Do, H. K., Lee, K. H., Holzapfel, W. H., Hwang, J. Y., Kwon, M. G. & Song, S. K. (2015). The effects of combined dietary probiotics *Lactococcus lactis* BFE920 and *Lactobacillus plantarum* FGL0001 on innate immunity and disease resistance in olive flounder (*Paralichthys olivaceus*). *Fish & Shellfish Immunology* 42(1): 177-183.

- Bermudez-Brito, M., Plaza-Díaz, J., Muñoz-Quezada, S., Gómez-Llorente, C. & Gil, A. (2012). Probiotic Mechanism of Action. *Annals of Nutrition and Metabolism* 61: 160-174.
- Bhatnagar, A. & Lamba, R. (2015). Antimicrobial ability and growth promoting effects of feed supplemented with probiotic bacterium isolated from gut microflora of *Cirrhinus mrigala*. *Journal of Integrative Agriculture* 14(3): 583-592.
- Bondad-Reantaso, M. G., Subasinghe, R. P., Arthur, J. R., Ogawa, K., Chinabut, S., Adlard, R., Tan, Z. & Shariff, M. (2005). Disease and health management in Asian aquaculture. *Veterinary Parasitology* 132(3-4): 249-272.
- Borsodi, A. K., Szabó, A., Krett, G., Felföldi, T., Specziár, A. & Boros, G. (2017). Gut content microbiota of introduced bigheaded carps (*Hypophthalmichthys* spp.) inhabiting the largest shallow lake in Central Europe. *Microbiological Research* 195: 40-50.
- Bromage, N. (1998). Broodstock management and the optimisation of seed supplies. *Suisan Zoshoku* 46: 395– 401.
- Calladine, C. R., Drew, H. R., Luisi, B. F. & Travers, A. A. (2004). Understanding DNA: The Molecule & How It Works, Third Edition. Elsevier Academic Press.
- Cao, J., Huang, A. L., Zhu, X. C., Li, L. & Li, J. N. (2018). Construction of *Vibrio mimicus* ghosts as a novel inactivated vaccine candidate and its protective efficacy against ascites disease in grass carps (*Ctenopharyngodon idella*). *Aquaculture* 485: 147-153.
- Caporaso, J. G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F. D., Costello, E. K., Fierer, N., Gonzalez Pena, A., Goodrich, J. K., Gordon, J. I., Huttley, G. A., Kelley, S. T., Knights, D., Koenig, J. E., Ley, R. E., Lozupone, C. A., McDonald, D., Muegge, B. D., Pirrung, M., Reeder, J., Sevinsky, J. R., Turnbaugh, P. J., Walters, W. A., Widmann, J., Yatsunenko, T., Zaneveld, J. & Knight, R. (2010a). QIIME allows analysis of high-throughput community sequencing data. *Nature Methods* 7(5): 335-336. Caporaso, J. G., Bittinger, K., Bushman, F. D., DeSantis, T. Z., Andersen, G. L., Knight, R. (2010b). PyNAST: a flexible tool for aligning sequences to a template alignment. *Bioinformatics* 26:266-267.
- Carnevali, O., Vivo, L. D., Sulpizio, R., Gioacchini, G., Olivotto, I., Silvi, S., Cresci, A. (2006). Growth improvement by probiotic in European sea bass juveniles (*Dicentrarchus labrax*, *L.*), with particular attention to IGF-1, myostatin and cortisol gene expression. *Aquaculture*;258: 430-438.
- Celebioglu, H. U., Olesen, S. V., Prehn, K., Lahtinen, S. J., Brix, S., Hachem, M. A. & Svensson, B. (2017). Mucin- and carbohydrate-stimulated adhesion and subproteome changes of the probiotic bacterium *Lactobacillus acidophilus* NCFM. *Journal of Proteomics* 163: 102-110.

- Cerezuela, R., Fumanal, M., Tapia-Paniagua, S. T., Meseguer, J., Moriñigo, M. Á. & Esteban, M. Á. (2013). Changes in intestinal morphology and microbiota caused by dietary administration of inulin and *Bacillus subtilis* in gilthead sea bream (*Sparus aurata* L.) specimens. Fish & Shellfish Immunology 34(5): 1063-1070.
- Chakravorty, S., Hleb, D., Burday, M., Connell, N. & Alland, D. (2008). A detailed analysis of 16S ribosomal RNA gene segments for the diagnosis of pathogenic bacteria. Journal of Microbiological Methods 69(2): 330-339.
- Chatta, A. M., Khan, A. M., Khan, M. N., Ayub, M. (2015). A Study on Growth Performance and Survival of Indus Golden Mahseer (*Tor macrolepis*) with Indian Major Carps in Semi-Intensive Polyculture System. The Journal of Animal & Plant Sciences 25(2): 561-566.
- Chen, F., Gao, Y., Chen, X., Yu, Z. & Li, X. (2013). Quorum Quenching Enzymes and Their Application in Degrading Signal Molecules to Block Quorum Sensing-Dependent Infection. International Journal of Molecular Sciences 14(9): 17447-17500.
- Chen, Q., Zhu, Z., Wang, J., Lopez, A. I., Li, S., Kumar, A., Yu, F., Chen, H., Cai, C. & Zhang, L. (2017). Probiotic *E. coli* Nissle 1917 biofilms on silicone substrates for bacterial interference against pathogen colonization. Acta Biomaterialia 50: 353-360.
- Cheng, Z. J., Hardy, R. W. & Usry, J. L. (2003). Effects of lysine supplementation in plant protein-based diets on the performance of rainbow trout (*Oncorhynchus mykiss*) and apparent digestibility coefficients of nutrients. Aquaculture 215(1-4): 255-265.
- Chu, L., Wang, J., Quan, F., Xing, X. H., Tang, L. & Zhang, C. (2014a). Modification of polyurethane foam carriers and application in a moving bed biofilm reactor. Process Biochemistry 49(11): 1979-1982.
- Chu, W., Zhou, S., Zhu, W. & Zhuang, X. (2014b). Quorum quenching bacteria *Bacillus* sp. QSI-1 protect zebrafish (*Danio rerio*) from *Aeromonas hydrophila* infection. Nature Scientific Reports 4: 5446.
- Cock, P. J. A., Fields, C. J., Goto, N., Heuer, M. L. & Rice, P. M. (2010). The Sanger FASTQ file format for sequences with quality scores, and the Solexa/Illumina FASTQ variants. Nucleic Acids Research 38(6): 1767-1771.
- Cockburn, D. W., Koropatkin, N. M. (2016). Polysaccharide Degradation by the Intestinal Microbiota and Its Influence on Human Health and Disease. Journal of Molecular Biology 428(16):3230-3252.
- Cordero, H., Guardiola, F. A., Tapia-Paniagua, S. T., Cuesta, A., Meseguer, J., Balebona, M. C., Moriñigo, M. C. & Estebana, M. Á. (2015). Modulation of immunity and gut microbiota after dietary administration of alginate encapsulated *Shewanella putrefaciens* Pdp11 to gilthead seabream (*Sparus aurata* L.). Fish and Shellfish Immunology 45(2): 608-618.

- Costa, P. S., Reis, M. P., Ávila, M. P., Leite, L. R., de Araújo, F. M. G., Salim, A. C. M., Oliveira, G., Barbosa, F., Chartone-Souza, E. & Nascimento, A. M. A. (2015). Metagenome of a Microbial Community Inhabiting a Metal-Rich Tropical Stream Sediment. *PLoS ONE* 10(3): e0119465. <https://doi.org/10.1371/journal.pone.0119465>.
- Craig, S. & Helfrich, L. A. (2009). Understanding Fish Nutrition, Feeds, and Feeding. Virginia Cooperative Extension Publication 420-256. Retrieved 7 June 2017 from https://www.pubs.ext.vt.edu/content/dam/pubs_ext_vt_edu/420/420-256/420-256_pdf.pdf.
- D'Argenio, V., Casaburi, G., Precone, V., Salvatore, F. (2016). Comparative Metagenomic Analysis of Human Gut Microbiome Composition Using Two Different Bioinformatic Pipelines. *BioMed Research International*: 325340. <http://dx.doi.org/10.1155/2014/325340>.
- Das, A., Nakhro, K., Chowdhury, S. & Kamilya, D. (2013). Effects of potential probiotic *Bacillus amyloliquifaciens* FPTB16 on systemic and cutaneous mucosal immune responses and disease resistance of catla (*Catla catla*). *Fish & Shellfish Immunology* 35(5): 1547-1553.
- De Silva, S. S., Ingram, B., Sungan, S., Tinggi, D., Gooley, G., Sim, S. Y. (2004). Artificial propagation of the indigenous *Tor* species, empurau (*T. tambroides*) and semah (*T. douronensis*), Sarawak, East Malaysia. *Aquaculture Asia* Volume IX No. 4 October-December 2004, pp. 15-20.
- Defoirdt, T., Boon, N., Bossier, P. & Vestraete, W. (2004). Disruption of bacterial quorum sensing: an unexplored strategy to fight infections in aquaculture. *Aquaculture* 240(1-4): 69-88.
- Defoirdt, T., Thanh, L. D., Delsen, B. V., Schryver, P. D., Sorgeloos, P., Boon, N. & Bossier, P. (2011). N-acylhomoserine lactone-degrading *Bacillus* strains isolated from aquaculture animals. *Aquaculture* 311(1-4): 258-260.
- Dehler, C. E., Secombes, C. J. & Martin, S. A. M. (2017). Environmental and physiological factors shape the gut microbiota of Atlantic salmon parr (*Salmo salar* L.). *Aquaculture* 467: 149-157.
- Denev, S., Staykov, Y., Moutafchieva, R. & Beev, G. (2009). Microbial ecology of the gastrointestinal tract of fish and the potential application of probiotics and prebiotics in finfish aquaculture. *International Aquatic Research* 1: 1-29.
- Department of Fisheries (DOF) Malaysia. (2015). *Fisheries Statistics 2014*. Retrieved 23 March 2017 from <http://www.dof.gov.my/index.php/pages/view/2600>.
- Department of Fisheries (DOF) Malaysia. (2016). *Fisheries Statistics 2015*. Retrieved 23 March 2017 from <http://www.dof.gov.my/index.php/pages/view/2614>.
- Desai, V. R. (2003). Synopsis of Biological Data on the Tor Mahseer *Tor tor* (Hamilton, 1822). FAO Fisheries Synopsis No. 158.

- DeSantis, T. Z., Hugenholtz, P., Larsen, N., Rojas, M., Brodie, E. L., Keller, K., Huber, T., Dalevi, D., Hu, P., Andersen, G. L. (2006). Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. *Applied and Environmental Microbiology* 72(7):5069-5072.
- Doan, H. V., Hoseinifar, S. H., Tapingkae, W., Tongsiri, S. & Khamtavee, P. (2016). Combined administration of low molecular weight sodium alginate boosted immunomodulatory, disease resistance and growth enhancing effects of *Lactobacillus plantarum* in Nile tilapia (*Oreochromis niloticus*). *Fish & Shellfish Immunology* 58: 678-685.
- Dong, Y. H., Wang, L. H. & Zhang, L. H. (2007). Quorum-quenching microbial infections: mechanisms and implications. *Philosophical Transactions of the Royal Society B: Biological Sciences* 362(1483): 1201- 1211.
- Dong, Y. M., Feng, L., Jiang, W. D., Liu, Y., Wu, P., Jiang, J., Kuang, S. Y., Tang, L., Tang, W. N., Zhang, Y. A. & Zhou, X. Q. (2018). Dietary threonine deficiency depressed the disease resistance, immune and physical barriers in the gills of juvenile grass carp (*Ctenopharyngodon idella*) under infection of *Flavobacterium columnare*. *Fish & Shellfish Immunology* 72: 161-173.
- Edgar, R. C. (2010). Search and clustering orders of magnitude faster than BLAST, *Bioinformatics* 26(19), 2460-2461 doi: 10.1093/bioinformatics/btq461.
- Edgar, R. C., Haas, B. J., Clemente, J. C., Quince, C. & Knight, R. (2011). UCHIME improves sensitivity and speed of chimaera detection. *Bioinformatics* btr381.
- Eichmiller, J. J., Hamilton, M. J., Staley, C., Sadowsky, M. J. & Sorensen, P. W. (2016). Environment shapes the fecal microbiome of invasive carp species. *Microbiome* 4: 44. Retrieved 3 August 2017 from <https://doi.org/10.1186/s40168-016-0190-1>.
- El-Rhman, A. M. A., Khattab, Y. A. E. & Shalaby, A. M. E. (2009). *Micrococcus luteus* and *Pseudomonas* species as probiotics for promoting the growth performance and health of Nile tilapia, *Oreochromis niloticus*. *Fish & Shellfish Immunology* 27(2): 175-180.
- Esa, Y., Siraj, S. S., Daud, S. K., Ryan, J. J. R., Rahim, K. A. A. & Tan, S. G. (2008). Molecular Systematics of Mahseers (Cyprinidae) in Malaysia Inferred from Sequencing of a Mitochondrial Cytochrome C Oxidase I (COI) Gene. *Pertanika Journal of Tropical Agricultural Science* 31(2): 263-269.
- Esposito, A. & Kirschberg, M. (2014). How many 16S-based studies should be included in a metagenomic conference? It may be a matter of etymology. *FEMS Microbiology Letters* 351: 145-146.
- Esty, W. E. (1996). The Efficiency of Good's Nonparametric Coverage Estimator. *The Annals of Statistics* 14(3): 1257-1260.

- Etyemez, M. & Balcázar, J. L. (2015). Bacterial community structure in the intestinal ecosystem of rainbow trout (*Oncorhynchus mykiss*) as revealed by pyrosequencing-based analysis of 16S rRNA genes. Research in Veterinary Science 100:8-11.
- Farzanfar A. (2006). The use of probiotics in shrimp aquaculture. EMS Immunology & Medical Microbiology 48: 149–158. doi:10.1111/j.1574-695X.2006.00116.x.
- FASTX-Toolkit. (n.d.). FASTX-Toolkit is a collection of command line tools for Short-Reads FASTA/FASTQ files preprocessing. Retrieved 4 June 2016 from http://hannonlab.cshl.edu/fastx_toolkit/index.html.
- Filippidou, S., Junier, T., Wunderlin, T., Lo, C. C., Li, P. E., Chain, P. S. & Junier, P. (2015). Under-detection of endospore-forming Firmicutes in metagenomic data. Computational and Structural Biotechnology Journal 13: 299-306.
- Food and Agriculture Organization (FAO) of the United Nations. (1980). ADCP/REP/80/11- Fish Feed Technology. United Nations Development Programme, Rome.
- Food and Agriculture Organization (FAO) of the United Nations. (1987). The Nutrition and Feeding of Farmed Fish and Shrimp – A Training Manual, 1. The Essential Nutrients. Brazil.
- Food and Agriculture Organization of United Nations (FAO). (2014). The State of World Fisheries and Aquaculture 2014: Opportunities and challenges. Rome.
- Food and Agriculture Organization of United Nations (FAO). (2015). Report of the APFIC/FAO Regional Consultation: Improving the contribution of culture-based fisheries and fishery enhancements in inland waters to Blue Growth, 25–27 May 2015, Jetwing Blue Hotel, Negombo, Sri Lanka. RAP Publication 2015/08.
- Food and Agriculture Organization of United Nations (FAO). (2016). The State of World Fisheries and Aquaculture 2016: Contributing to food security and nutrition for all. Rome.
- Franklin, R. & Gosling, R. G. (1953a). Molecular Configuration in Sodium Thymonucleate. Nature 171: 740-741.
- Franklin, R. & Gosling, R. G. (1953b). Evidence for 2-Chain Helix in Crystalline Structure of Sodium Deoxyribonucleate. Nature 172: 156-157.
- Fuqua, W. C., Winans, S. C. & Greenberg, E. P. (1994). Quorum Sensing in Bacteria: the LuxR-LuxI Family of Cell Density-Responsive Transcriptional Regulators. Journal of Bacteriology 176(2): 269-275.
- Fish Base. (n.d.). Fish Identification: Genus *Tor*. Retrieved 17 April 2017 from <http://www.fishbase.org/identification/SpeciesList.php?genus=Tor>.

- Geng, Y., Liu, D., Han, S., Zhou, Y., Wang, K. Y., Huang, X. L., Chen, D. F., Peng, X. & Lai, W. M. (2014). Outbreaks of vibriosis associated with *Vibrio mimicus* in freshwater catfish in China. *Aquaculture* 433:82-84.
- Giatsis, C., Sipkema, D., Smidt, H., Heilig, H., Benvenuti, G., Verreth, J., & Verdegem, M. (2015). The impact of rearing environment on the development of gut microbiota in tilapia larvae. *Scientific Reports* 5: 18206.
- Gihring, T. M., Green, S. J. & Schadt, C. W. (2011). Massively parallel rRNA gene sequencing exacerbates the potential for biased community diversity comparisons due to variable library sizes. *Environmental Microbiology* 14(2): 285-290.
- Giri, S. S., Sen, S. S. & Sukumaram, V. (2012). Effects of dietary supplementation of potential probiotic *Pseudomonas aeruginosa* VSG-2 on the innate immunity and disease resistance of tropical freshwater fish, *Labeo rohita*. *Fish & Shellfish Immunology* 32:1135-1140.
- Giri, S. S., Sukumaran, V. & Oviya, M. (2013). Potential probiotic *Lactobacillus plantarum* VSG3 improves the growth, immunity, and disease resistance of tropical freshwater fish, *Labeo rohita*. *Fish & Shellfish Immunology* 34: 660-666.
- Giri, S. S. Sen, S. S., Jun, J. W., Park, S. C. & Sukumaran, V. (2016). Heat-killed whole-cell products of the probiotic *Pseudomonas aeruginosa* VSG2 strain affect *in vitro* cytokine expression in head kidney macrophages of *Labeo rohita*. *Fish & Shellfish Immunology* 50: 310-316.
- Gobi, N., Malaikozhundan, B., Sekar, V., Shanthi, S., Vaseeharan, B., Jayakumar, R. & Nazar, A. K. (2016). GFP tagged *Vibrio parahaemolyticus* Dahv2 infection and the protective effects of the probiotic *Bacillus licheniformis* Dahb1 on the growth, immune and antioxidant responses in *Pangasius hypophthalmus*. *Fish & Shellfish Immunology* 52: 230-238.
- Gogineni, V. K., Morrow, L. E. & Malesker, M. A. (2013). Probiotics: Mechanisms of Action and Clinical Applications. *Journal of Probiotics & Health* 1: 101.
- Gono, R. K., Muzondiwa, J., Chihanga, I. & Manhondo, P. R. (2015). Socio-economic factors affecting the successful implementation of aquaculture projects in Zimbabwe; a case study of Chivi District. *Livestock Research for Rural Development* 27(2). Retrieved 10 July 2017 from <http://www.lrrd.org/lrrd27/2/gono27027.html>.
- Goujon, M., McWilliam, H., Li, W., Valentin, F., Squizzato, S., Paern, J. & Lopez, R. 2010). A new bioinformatics analysis tools framework at EMBL-EBI. *Nucleic Acids Research*: W695-9 doi:10.1093/nar/gkq313.
- Griffiths, A. J. F., Wessler, S. R., Lewontin, R. C., Gelbart, W. M., Suzuki, D. T. & Miller, J. H. (2004). An Introduction to Genetic Analysis, Eight Edition. W.H. Freeman and Company.

- Guardiola, F. A., Bahi, A., Bakhrouf, A. & Esteban, M. A. (2017). Effects of dietary supplementation with fenugreek seeds, alone or in combination with probiotics, on gilthead seabream (*Sparus aurata* L.) skin mucosal immunity. Fish & Shellfish Immunology 65: 169-178.
- Guerreiro, I., Serra, C. R., Enes, P., Couto, A., Salvador, A., Costas, B & Oliva-Teles, A. (2016). Effect of short chain fructooligosaccharides (scFOS) on immunological status and gut microbiota of gilthead sea bream (*Sparus aurata*) reared at two temperatures. Fish & Shellfish Immunology 49: 122-131.
- Guo, X., Chen, D. D., Peng, K. S., Cui, Z. W., Zhang, X. J., Li, S. & Zhang, Y. A. (2016). Identification and characterization of *Bacillus subtilis* from grass carp (*Ctenopharyngodon idellus*) for use as probiotic additives in aquatic feed. Fish & Shellfish Immunology 52: 74-84.
- Gupta, A., Gupta, P. & Dhawan, A. (2014). Dietary supplementation of probiotics affects growth, immune response and disease resistance of *Cyprinus carpio* fry. Fish & Shellfish Immunology 41: 113-119.
- Hai, N. V. & Fotedar, R. (2009). Comparison of the effects of the prebiotics (Bio-Mos® and β-1,3-D-glucan) and the customised probiotics (*Pseudomonas synxantha* and *P. aeruginosa*) on the culture of juvenile western king prawns (*Penaeus latisulcatus* Kishinouye, 1896). Aquaculture 289: 310-316.
- Haas, B. J., Gevers, D., Earl, A. M., Feldgarden, M., Ward, D. V., Giannoukos, G., Ciulla, D., Tabbaa, D., Highlander, S. K., Sodergren, E., Methé, B., DeSantis, T. Z., The Human Microbiome Consortium., Petrosino, J. F., Knight, R. & Birren, B. W. (2011). Chimeric 16S rRNA sequence formation and detection in Sanger and 454-pyrosequenced PCR amplicons. Genome Research 21(3): 494-504.
- Hargittai, I. (2009). The tetranucleotide hypothesis: a centennial. Structural Chemistry 20:753-756.
- Heather, J. M. & Chain, B. (2016). The sequence of sequencers: The history of sequencing DNA. Genomics 107(1): 1-8.
- Heiman, M. L. & Greenway, F. L. (2016). A healthy gastrointestinal microbiome is dependent on dietary diversity. Molecular Metabolism 5(5):317-320.
- Hennersdorf, P., Kleinertz, S., Theisen, S., Abdul-Aziz, M. A., Mrotzek, G., Palm, H. W. & Saluz, H. P. (2016). Microbial Diversity and Parasitic Load in Tropical Fish of Different Environmental Conditions. PLoS ONE 11(3): e0151594.
- Hense, B. A., Kuttler, C., Müller, J., Rothbäcker, M., Hartmann, A., Kreft J-U. 2007 Does efficiency sensing unify diffusion and quorum sensing? Nature Reviews Microbiology 5: 230–239.
- Hershey, A. D. & Chase, M. (1952). Independent Functions of Viral Protein and Nucleic Acid in Growth of Bacteriophage. Journal of General Physiology 36(1): 39-56.

- Hochart-Behra, A. C., Drobecq, H., Tourret, M., Dubreuil, L., Behra-Miellet, J. (2014). Anti-stress proteins produced by *Bacteroides thetaiotaomicron* after nutrient starvation. *Anaerobe* 28:18-23.
- Holley, R. W., Apgar, J., Everett, G. A., Madison, J. T., Marquisee, M., Merrill, S. H., Penswick, J. R. & Zamir, A. (1965). Structure of a Ribonucleic Acid. *Science* 147(3364): 1465-1467.
- Holmes, A. J., Chew, Y. V., Colakoglu, F., Cliff, J. B., Klaassens, E., Read, M. N., Solon-Biet, S. M., McMahon, A. C., Cogger, V. C., Ruohonen, K., Raubenheimer, D., Le Couteur, D. G. & Simpson, S. J. (2017). Diet-Microbiome Interactions in Health Are Controlled by Intestinal Nitrogen Source Constraints. *Cell Metabolism* 25(1): 140-151.
- Hossain, M. A. & Yoshimatsu, T. (2014). Dietary calcium requirement in fishes. *Aquaculture Nutrition* 20(1): 1-11.
- Hosseini, M., Miandare, H. K., Hoseinifar, S. H. & Yarahmadi, P. (2016). Dietary *Lactobacillus acidophilus* modulated skin mucus protein profile, immune and appetite genes expression in gold fish (*Carassius auratus gibelio*). *Fish & Shellfish Immunology* 59: 149-154.
- Hostins, B., Lara, G., Decamp, O., Cesar, D. E. & Wasielesky, W. J. (2017). Efficacy and variations in bacterial density in the gut of *Litopenaeus vannamei* reared in a BFT system and in clear water supplemented with a commercial probiotic mixture. *Aquaculture* 480: 58-64.
- Hughes, J. B. & Bohannan, B. J. M. (2004). Application of ecological diversity statistics in microbial ecology. *Molecular Microbial Ecology Manual*, 2nd Edition 7.01: 1321-1344.
- Hughes, J. B., Hellmann, J. J., Ricketts, T. H. & Bohannan, B. J. M. (2001). Counting the Uncountable: Statistical Approaches to Estimating Microbial Diversity. *Applied Environmental Microbiology* 67(10): 4399-4406.
- Hsieh, C. H., Chang, W. T., Chang, H. C., Hsieh, H. S., Chung, Y. L. & Hwang, D. F. (2010). Puffer fish-based commercial fraud identification in a segment of cytochrome b region by PCR-RFLP analysis. *Food Chemistry* 121(4): 1305-1311.
- Hsieh, Y. H., Peterson, C. M., Raggio, A., Keenan, M. J., Martin, R. J., Ravussin, E. & Marco, M. L. (2016). Impact of Different Fecal Processing Methods on Assessments of Bacterial Diversity in the Human Intestine. *Frontiers in Microbiology* 7:1643.
- Huang, J. Y., Lee, S. M., Mazmanian, S. K. (2011). The human commensal *Bacteroides fragilis* binds intestinal mucin. *Anaerobe* 17(4):137-141.

Huyben, D., Nyman, A., Vidaković, A., Passoth, V., Moccia, R., Kiessling, A., Dicksved, J. & Lundh, T. (2017). Effects of dietary inclusion of the yeasts *Saccharomyces cerevisiae* and *Wickerhamomyces anomalus* on gut microbiota of rainbow trout. *Aquaculture* 473: 528-537.

Illumina. (n.d.). 16S Metagenomic Sequencing Library Preparation. Retrieved 14 Mac 2-16 from https://support.illumina.com/content/dam/illumina-support/documents/documentation/chemistry_documentation/16s/16s-metagenomic-library-prep-guide-15044223-b.pdf.

Illumina. (n.d.). Optimizing Cluster Density on Illumina Sequencing Systems. Retrieved 6 Septembe 2016 from <https://www.illumina.com/content/dam/illumina-marketing/documents/products/other/miseq-overclustering-primer-770-2014-038.pdf>.

Ingerslev, H. C., Strube, M. L., Jørgensen, L. V. G., Dalsgaard, I., Boye, M. & Madsen, L. (2014). Diet type dictates the gut microbiota and the immune response against *Yersinia ruckeri* in rainbow trout (*Oncorhynchus mykiss*). *Fish & Shellfish Immunology* 40(2): 624-633.

Ingram, B., Sungan, S., Gooley, G., Sim, S. Y., Tinggi, D. and De Silva, S. S. (2005). Induced spawning, larval development and rearing of two indigenous Malaysian mahseer, *Tor tambroides* and *T. douronensis*. *Aquaculture Research* 36: 1001–1014.

Ingram, B., Sungan, S., Tinggi, D., Sim, S. Y., Gooley, G. J. & De Silva, S. S. (2006). Developments in the spawning of *Tor tambroides* and *T. duoronensis* in captivity. Mahseer: The Biology, Culture and Conservation. Malaysian Fisheries Society Occasional Publication No. 14, Kuala Lumpur, Malaysia, pp. 123–126.

Ingram, B., Sungan, S., Tinggi, D., Sim, S. Y. & De Silva, S. S. (2007a). Breeding performance of Malaysian mahseer, *Tor tambroides* and *T. douronensis* broodfish in captivity. *Aquaculture Research* 38(8): 809-818.

Ingram, B., Sungan, S., Tinggi, D., Sim, S. Y., Gooley, G. J., De Silva S. S. (2007b). Observations on the growth of cage- and pond-reared *Tor tambroides* and *T. douronensis* in Sarawak, Malaysia. Mahseer: The Biology, Culture and Conservation, Vol. 14, Malaysian Fisheries Society Occasional Publication, Kuala Lumpur, Malaysia; 2007.

Integrated Taxonomic Information System (ITIS). (n.d.). *Tor tambroides* Taxonomic Serial No.: 690205. Retrieved 27 March 2017 from https://www.itis.gov/servlet/SingleRpt/SingleRpt?search_topic=TSN&search_value=690205#null.

Irianto, A. & Austin, B. (2002). Probiotics in aquaculture. *Journal of Fish Diseases* 25(11): 633-642.

- Ishak, S. D., Kamarudin, M. S., Ramezani-Fard, E. & Yusof, Y. A. (2016). Effects of varying dietary carbohydrate levels on growth performance, body composition and liver histology of Malaysian mahseer fingerlings (*Tor tambroides*). Journal of Environmental Biology 37(4): 756-764.
- Ismail, M. F. S., Siraj, S. S., Daud, S. K. & Harmin, S. A. (2011). Association of annual hormonal profile with gonad maturity of mahseer (*Tor tambroides*) in captivity. General and Comparative Endocrinology 170(1): 125-130.
- Iwana, G. K., Pickering, A. D., Sumpter, J. P., Schreck, C. B. (1997). Fish Stress and Health in Aquaculture. Society for Experimental Biology Seminar Series 62.
- Izquierdo, M. S., Fernandez-Palacios, H. and Tacon, A. G. J. (2000). Effect of broodstock nutrition on reproductive performance of fish. Aquaculture 197: 25–42.
- Janda, J. M. & Abbott, S. L. (2007). 16S rRNA Gene Sequencing for Bacterial Identification in the Diagnostic Laboratory: Pluses, Perils, and Pitfalls. Journal of Clinical Microbiology 45(9): 2761-2764.
- Jeon, Y. S., Lee, K., Park, S. C., Kim, B. S., Cho, Y. J., Ha, S. M. & Chun, J. (2014). EzEditor: a versatile sequence alignment editor for both rRNA- and protein-coding genes. International Journal of Systematic and Evolutionary Microbiology 64: 689-691.
- Jha, D. K., Bhujel, R. C., Anal, A. K. (2015). Dietary supplementation of probiotics improves survival and growth of Rohu (*Labeo rohita* Ham.) hatchlings and fry in outdoor tanks. Aquaculture 435: 475-479.
- Jiang, H., Chen, T., Sun, H., Tang, Z., Yu, J., Lin, Z., Ren, P., Zhou, X., Huang, Y., Li, X. & Yu, X. (2017). Immune response induced by oral delivery of *Bacillus subtilis* spores expressing enolase of *Clonorchis sinensis* in grass carps (*Ctenopharyngodon idellus*). Fish & Shellfish Immunology 60: 318-325.
- Jin, Y., Wu, S., Zeng, Z. & Fu, Z. (2017). Effects of environmental pollutants on gut microbiota. Environmetal Pollution 222: 1-9.
- Jonsson, V., Österlund, T., Nerman, O. & Kristiansson, E. (2016). Statistical evaluation of methods for identification of differentially abundant genes in comparative metagenomics. BMC Genomics 17:78.
- Kalia, V. C. (2013). Quorum sensing inhibitors: An overview. Biotechnology Advances 31(2):224-245.
- Kanehisa, M. & Goto, S. (2000). KEGG: Kyoto Encyclopedia of Genes and Genomes. Nucleic Acids Research 28(1): 27-30.
- Kanehisa, M., Tanabe, M., Sato, Y., and Morishima, K. (2017). KEGG: new perspectives on genomes, pathways, diseases and drugs. Nucleic Acids Research 45: D353-D361.

- Kappel, K., Haase, I., Käppel, C., Sotelo, C. G. & Schröder, U. (2017). Species identification in mixed tuna samples with next-generation sequencing targeting two short cytochrome b gene fragments. *Food Chemistry* 234: 212-219.
- Kato, C. D., Kahuma, C. E., Namulawa, V. T. & Kasozi, N. (2016). Antibacterial Activity of *Lactobacillus* spp. and *Lactococcus* spp. Isolated from Various Parts of Pebbley Fish, *Alestes baremoze*. *British Microbiology Research Journal* 17(2): 1-7.
- Kaufmann, G. F., Park, J., Janda, K. D. (2008). Bacterial quorum sensing: a new target for anti-infective immunotherapy. *Expert Opinion on Biological Therapy* 8(6):719-724.
- Kesarcodi-Watson, A., Kasper, H., Lategan, M. J. & Gibson, L. (2008). Probiotics in aquaculture: The need, principles and mechanisms of action and screening processes. *Aquaculture* 274: 1-14.
- Khojasteh, S. M. B. (2012). The morphology of the post-gastric alimentary canal in teleost fishes: a brief review. *International Journal of Aquatic Science* 3(20):71-88.
- Klindworth, A., Pruesse, E., Schweer, T., Peplies, J., Quast, C., Horn, M & Glöckner, O. (2013). Evaluation of general 16S ribosomal RNA gene PCR primers for classical and next-generation sequencing-based diversity studies. *Nucleic Acids Research* 41(1):e1.
- Konstantinidis, K. T. & Rosselló-Móra, R. (2015). Classifying the uncultivated microbial majority: A place for metagenomic data in the Candidatus proposal. *Systematic and Applied Microbiology* 38(4): 223-230.
- Koo, H., Hakim, J. A., Powell, M. L., Kumar, R., Eipers, P. G., Morrow, C. D., Crowley, M., Lefkowitz, E. J., Watts, S. A. & Bej, A. K. (2017). Metagenomics approach to the study of the gut microbiome structure and function in zebrafish *Danio rerio* fed with gluten formulated diet. *Journal of Microbiological Methods* 135: 69-76.
- Kottelat, M. (2012). *Tor tambroides*. The IUCN Red List of Threatened Species 2012: e.T187939A1837406. Retrieved 27 March 2017 from <http://dx.doi.org/10.2305/IUCN.UK.2012-1.RLTS.T187939A1837406.en>.
- Krogdahl, A., Hemre, G. I. & Mommsen, T. P. (2005). Carbohydrates in fish nutrition: Digestion and absorption in postlarval stages. *Aquaculture Nutrition* 11(2): 103-122.
- Kumar, R., Pande, V., Singh, L., Sharma, L., Saxena, N., Thakuria, D., Singh, A. K. & Sahoo, P. K. (2016a). Pathological Findings of Experimental *Aeromonas hydrophila* Infection in Golden Mahseer (*Tor putitora*). *Fisheries and Aquaculture Journal* 7(1): 1000160.

- Kumar, S., Stecher, G. & Tamura, K. (2016b). MEGA7: Molecular Evolutionary Genetics Analysis version 7.0. *Molecular Biology and Evolution* 33(7):1870-1874.
- Kunlapapuk, S. & Kulabtong, S. (2011). Breeding, Nursing and Biology of Thai Mahseer (*Tor tambroides*) in Malaysia: An Overview. *Journal of Agricultural Science and Technology A* 1: 1214-1216.
- Lade, H., Paul, D. & Kweon, J. H. (2014). Quorum Quenching Mediated Approaches for Control of Membrane Biofouling. *International Journal of Biological Sciences* 10(5): 550-565.
- Lalloo, R., Moonsamy, G., Ramchuran, S., Görgens, J. & Gardiner, N. (2010). Competitive exclusion as a mode of action of a novel *Bacillus cereus* aquaculture biological agent. *Letters in Applied Microbiology* 50: 563-570.
- Lane, D.J. (1991). 16S/23S rRNA sequencing. In *Nucleic acid techniques in bacterial systematics*, ed. Stackebrandt, E., and Goodfellow, M., pp. 115-175. John Wiley and Sons, New York.
- Langille, M. G. I., Zaneveld, J., Caporaso, J. G., McDonald, D., Knights, D., Reyes, J., Clemente, J. C., Burkepile, D. E., Vega Thurber, R. L., Knight, R., Beiko, R. G. and Huttenhower, C. (2013). Predictive functional profiling of microbial communities using 16S rRNA marker gene sequences. *Nature Biotechnology* 31: 814-821.
- Lara-Flores, M., Olvera-Novoa, M. A., Guzmán-Méndez, B. E. and López-Madrid, W. (2003). Use of the bacteria *Streptococcus faecium* and *Lactobacillus acidophilus*, and the yeast *Saccharomyces cerevisiae* as growth promoters in Nile tilapia (*Oreochromis niloticus*). *Aquaculture* 216: 193-201.
- Larsen, A. M., Mohammed, H. H. & Arias, C. R. (2014). Characterization of the gut microbiota of three commercially valuable warmwater fish species. *Journal of Applied Microbiology* 116:1396-1404.
- Laskar, B. A., Bhattacharjee, M. J., Dhar, B., Mahadani, P., Kundu, S. & Ghosh, S. K. (2013). The Species Dilemma of Northeast Indian Mahseer (Actinopterygii: Cyprinidae): DNA Barcoding in Clarifying the Riddle. *PLoS ONE* 8(1): e53704.
- Leary, S., Underwood, W., Anthony, R., Cartner, S., Corey, D., Grandin, T., Greenacre, C., Gwaltney-Brant, S., McCrackin, M. A., Meyer, R., Miller, D., Shearer, J. & Yanong, R. (2013). AVMA Guidelines for the Euthanasia of Animals: 2013 Edition. American Veterinary Medical Association.
- Lee, K. S., Lihan, S., Dasthagir, F. F. G., Mikal, K. M., Collick, F., Ng, K. H. (2014). Microbiological and Physicochemical Analysis of Water from Empurau Fish (*Tor tambroides*) Farm in Kuching, Sarawak, Malaysian Borneo. *International Journal of Scientific & Technology Research* 3(6): 285-292.

- Lee, S., Katya, K., Park, Y., Won, S., Seong, M., Hamidoghi, A. & Bai, S. C. (2017). Comparative evaluation of dietary probiotics *Bacillus subtilis* WB60 and *Lactobacillus plantarum* KCTC3928 on the growth performance, immunological parameters, gut morphology and disease resistance in Japanese eel, *Anguilla japonica*. Fish & Shellfish Immunology 61: 201-210.
- Li, J., Ni, J., Li, J., Wang, C., Li, X., Wu, S., Zhang, T., Yu, Y. & Yan, Q. (2014). Comparative study on gastrointestinal microbiota of eight fish species with different feeding habits. Journal of Applied Microbiology 117(6): 1750-1760.
- Li, Y. L., Weng, J. C., Hsiao, C. C., Chou, M. T., Tseng, C. W. & Hung, J. H. (2015). PEAT: an intelligent and efficient paired-end sequencing adapter trimming algorithm. BMC Bioinformatics 16(Supplement 1): S2.
- Lin, H. L., Shiu, Y. L., Chiu, C. S., Huang, S. L. & Liu, C. H. (2017). Screening probiotic candidates for a mixture of probiotics to enhance the growth performance, immunity, and disease resistance of Asian seabass, *Lates calcarifer* (Bloch), against *Aeromonas hydrophila*. Fish & Shellfish Immunology 60: 474-482.
- Liu, H., Guo, X. W., Gooneratne, R., Lai, R. F., Zeng, C., Zhan, F. B., Wang, W. M. (2015). The gut microbiome and degradation enzyme activity of wild freshwater fishes influenced by their trophic levels. Nature Scientific Reports 6, 24340; 2015. doi:10.1038/srep24340.
- Liu, Y., White, R. H. & Whitman, W. B. (2010). Methanococci Use the Diaminopimelate Aminotransferase (DapL) Pathway for Lysine Biosynthesis. Journal of Bacteriology 192(13): 3303-3310.
- Liu, Y. M., Zhang, Q. Z., Xu, D. H., Fu, Y. W., Lin, De Jie., Zhou, S. Y. (2018). Antiparasitic efficacy of commercial curcumin against *Ichthyophthirius multifiliis* in grass carp (*Ctenopharyngodon idellus*). Aquaculture 480: 65-70.
- Lobo, C., Tapia-Paniagua, S., Moreno-Ventas, X., Alarcón, F. J., Rodríguez, C., Balebona, M. C., Moriñigo, M. A., De La Banda, I. G. (2014). Benefits of probiotic administration on growth and performance along metamorphosis and weaning of Senegalese sole (*Solea senegalensis*). Aquaculture 433: 183-195.
- Long, S. L., Gahan, C. G. M. & Joyce, S. A. (2017). Interactions between gut bacteria and bile in health and disease. Molecular Aspects of Medicine 56: 54-65.
- Low, V. L., Tan, T. K., Lim, P. E., Domingues, L. N., Tay, S. T., Lim, Y. A. L., Goh, T. G., Panchadcharam, C. & Bathmanaban, P. (2014). Use of COI, CytB and ND5 genes for intra- and inter-specific differentiation of *Haematobia irritans* and *Haematobia exigua*. Veterinary Parasitology 204(3-4): 439-442.
- Lozupone, C. & Knight, R. (2005). UniFrac: a new phylogenetic method for comparing microbial communities. Applied Environmental Microbiology 71(12): 8228-8235.

- Lozupone, C. A., Hamady, M., Kelley, S. T., Knight, R. (2007). Quantitative and qualitative β diversity measures lead to different insights into factors that structure microbial communities. *Applied and Environmental Microbiology* 73:1576–1585.
- Lü, A., Hu, X., Zheng, L., Zhu, A., Cao, C. & Jiang, J. (2011). Isolation and characterization of *Citrobacter* spp. from the intestine of grass carp *Ctenopharyngodon idellus*. *Aquaculture* 313(1-4): 156-160.
- Madsen, K. L. (2012). Enhancement of Epithelial Barrier Function by Probiotics. *Journal of Epithelial Biology and Pharmacology* 5: 55-59.
- Mahseer Trust. (n.d.). About Mahseer. Retrieved 18 April 2017 from <http://mahseertrust.org/about/about-mahseer/>.
- Malekinejad, H., Bazargani-Gilani, B., Tukmechi, A. & Ebrahimi, H. (2012). A cytotoxicity and comparative antibacterial study on the effect of *Zataria multiflora* Boiss, *Trachyspermum copticum* essential oils, and Enrofloxacin on *Aeromonas hydrophila*. *Avicenna Journal of Phytomedicine* 2(4): 188-195.
- Martin, S. A. M., Dehler, C. E. & Król, E. (2016). Transcriptomic responses in the fish intestine. *Developmental & Comparative Immunology* 64: 103-117.
- Mao, Z., Ye, J., Li, M., Xu, H. & Chen, J. (2013). Vaccination efficiency of surface antigens and killed whole cell of *Pseudomonas putida* in large yellow croaker (*Pseudosciaena crocea*). *Fish & Shellfish Immunology* 35(2): 375-381.
- Mehri, I., Turki, Y., Chair, M., Chérif, H., Hassen, A., Meyer, J. M. & Gtari, M. (2011). Genetic and functional heterogeneities among fluorescent *Pseudomonas* isolated from environmental samples. *Journal of General and Applied Microbiology* 57: 101-114.
- Meidong, R., Khotchanalekha, K., Doolgindachbapom, S., Nagasawa, T., Nakao, M., Sakai, K. & Tongpim, S. (2018). Evaluation of probiotic *Bacillus aerius* B81e isolated from healthy hybrid catfish on growth, disease resistance and innate immunity of Pla-mong *Pangasius bocourti*. *Fish & Shellfish Immunology* 73: 1-10.
- Merrifield, D. L., Dimitroglou, A., Foey, A., Davies, S. J., Baker, R. T. M., Bøgwald, J., Castex, M. & Ringø, E. (2010). The current status and future focus of probiotic and prebiotic applications for salmonids. *Aquaculture* 302: 1-18.
- Miao, J., Xu, M., Guo, H., He, L., Gao, X., DiMarco-Crook, C., Xiao, H. & Cao, Y. (2015). Optimization of culture conditions for the production of antimicrobial substances by probiotic *Lactobacillus paracasei* subsp. *Tolerans* FX-6. *Journal of Functional Foods* 18(1): 244-253.
- Miller, M. B. & Blaser, B. L. (2011). Quorum sensing in Bacteria. *Annual Review of Microbiology* 55: 165-199.

- Mishra, P., Samanta, M., Mohanty, S. & Maiti, N. K. (2010). Characterization of *Vibrio* species isolated from freshwater fishes by ribotyping. Indian Journal of Microbiology 50(1): 101-103.
- Misieng, J. D., Kamarudin, M. S. & Musa, M. (2011). Optimum dietary protein requirement of Malaysian mahseer (*Tor tambroides*) fingerling. Pakistan Journal of Biological Sciences 14(3): 232-235.
- Modanloo, M., Soltanian, S., Akhlaghi, M. & Hoseinifar, S. H. (2017). The effects of single or combined administration of galactooligosaccharide and *Pediococcus acidilactici* on cutaneous mucus immune parameters, humoral immune responses and immune related genes expression in common carp (*Cyprinus carpio*) fingerlings. Fish & Shellfish Immunology 70: 391-397.
- Mohammadian, T., Alishahi, M., Tabandeh, M. R., Ghorbanpoor, M., Gharibi, D., Tollabi, M. & Rohanizade, S. (2016). Probiotic effects of *Lactobacillus plantarum* and *L. delbrueckii* ssp. *bulganicus* on some immune-related parameters in *Barbus grypus*. Aquaculture International 24(1): 225-242.
- Modu, B. M., Zaleha, K. & Shaharom-Harrison, F. M. (2014). Water Quality Assessment using Monogenean Gill Parasites of Fish in Kenyir Lake, Malaysia. Nigerian Journal of Fisheries and Aquaculture 2(1): 37-47.
- Moriarty, D. J. W. (1998). Control of luminous *Vibrio* species in penaeid aquaculture ponds. Aquaculture 164: 351-358.
- Moriarty, J. D. W., Decamp, O. & Lavens, P. (2005). Probiotics in aquaculture. AQUA Culture Asia Pacific Magazine September/October; 2005. Retrieved 12 July 2016 from <https://www.aquaasiapac.com/document-download.php?id=75>.
- Muchlisin, Z. A., Fuadi, Z., Munazir, A. M., Fadli, N., Winaruddin, W., Defira, C. N. & Hendri, A. (2015). First Report On Asian Fish Tapeworm (*Bothriocephalus Acheilognathi*) Infection of Indigenous Mahseer (*Tor Tambra*) From Nagan Raya District, Aceh Province, Indonesia. Bulgarian Journal of Veterinary Medicine 18(4): 361-366.
- Muchlisin, Z. A., Murda, T., Yulvizar, C., Dewiyanti, I., Fadli, N., Afrido, F., Siti-Azizah, M. N., Muhammadar, A. A. (2017). Growth performance and feed utilization of keureleng fish *Tor tambra* (Cyprinidae) fed formulated diet supplemented with enhanced probiotic. F1000Research 6: 137. Retrieve 9 July 2017 from <http://doi.org/10.12688/f1000research.10693.1>.
- Munir, M. B., Hashim, R., Chai, Y. H., Marsh, T. L. Nor, S. A. M. (2016). Dietary prebiotics and probiotics influence growth performance, nutrient digestibility and the expression of immune regulatory genes in snakehead (*Channa striata*) fingerlings. Aquaculture 460: 59-68.
- Murtaza, N., Cuív, P. Ó. & Morrison, M. (2016). Diet and the Microbiome. Gastroenterology Clinics of North America 46(1): 49-60.

- Nadiatul, H. H., Daud, S. K., Siraj, S. S., Sungan, S. & Moghaddam, F. Y. (2011). Genetic diversity of Malaysian indigenous Mahseer, *Tor douronensis* in Sarawak river basins as revealed by cytochrome c oxidase I gene sequences. *Iranian Journal of Animal Biosystematics* 7(2): 119-127.
- Nakayama, K., Yamashita, R. & Kitamura, S. I. (2017). Use of common carp (*Cyprinus carpio*) and *Aeromonas salmonicida* for detection of immunomodulatory effects of chemicals on fish. *Marine Pollution Bulletin* 124(2): 710-713.
- National Center for Biotechnology Information (NCBI). (1988). Bethesda (MD): National Library of Medicine (US), National Center for Biotechnology Information. Retrieved 24 February 2017 from <https://www.ncbi.nlm.nih.gov/>.
- National Research Council (NRC). (1993). Nutrient Requirements of Fish. National Academy Press, Washington, D.C. 114 pp.
- Natrah, F. M. I. (2011). Role of bacterial quorum sensing and micro-algae in fish and crustacean larviculture. PhD thesis, Ghent University, Ghent, Belgium.
- Natrah, F. M. I., Alam, M. I., Pawar, S., Harzevili, A. S., Nevejan, N., Boon, N., Sorgeloos, P., Bossier, P. & Defoirdt, T. (2012). The impact of quorum sensing on the virulence of *Aeromonas hydrophila* and *Aeromonas salmonicida* towards burbot (*Lota lota L.*) larvae. *Veterinary Microbiology* 159(1-2): 77-82.
- NavinChandran, M., Iyapparaj, P., Moovendhan, S., Ramasubburayan, R., Prakash, S., Immanuel, G. & Palavesam, A. (2014). Influence of probiotic bacterium *Bacillus cereus* isolated from the gut of wild shrimp *Penaeus monodon* in turn as a potent growth promoter and immune enhancer in *P. monodon*. *Fish & Shellfish Immunology* 36(1): 38-45.
- Nayar, R., Shukla, I., Sultan, A. (2014). Epidemiology, Prevalence and Identification of *Citrobacter* Species in Clinical Specimens in a Tertiary Care Hospital in India. *International Journal of Scientific and Research Publication* 4(4):1-6.
- Neiffer, D. L. & Stamper, M. A. (2009). Fish Sedation, Anesthesia, Analgesia, and Euthanasia: Considerations, Methods, and Types of Drugs. *ILAR Journal* 50(4): 343-360.
- Neuman, C., Hatje, E., Zarkasi, K. Z., Smullen, R., Bowman, J. P. & Katouli, M. (2016). The effect of diet and environmental temperature on the faecal microbiota of farmed Tasmanian Atlantic Salmon (*Salmo salar L.*). *Aquaculture Research* 47(2): 660-672.
- Neuman, H. & Koren, O. (2015). The Gut Microbiome. Reference Module in Biomedical Sciences, from Encyclopedia of Cell Biology 2015;2:799-808.
- Newaj-Fyzul, A. & Austin, B. (2014). Probiotics, immunostimulants, plant products and oral vaccines, and their role as feed supplements in the control of bacterial fish diseases. *Journal of Fish Diseases* 38(11): 937-955.

- Newaj-Fyzul, A., Al-Harbi, A. H. & Austin, B. (2014). Review: Developments in the use of probiotics for disease control in aquaculture. Aquaculture Volume 431: 1-11.
- Ng, C. K. (2004). King of the rivers: mahseer in Malaysia and the region. Inter Sea Fishery (M). Sdn Bhd, Kuala Lumpur.
- Ng, W. K., Abdullah, N. and De Silva S. S. (2008). The dietary protein requirement of the Malaysian mahseer, *Tor tambroides* (Bleeker), and the lack of protein-sparing action by dietary lipid. Aquaculture 284: 201-206.
- Ng, W. K. & Andin, V. C. (2011). The Malaysian mahseer, *Tor tambroides* (Bleeker), requires low dietary lipid levels with a preference for lipid sources with high omega-6 and low omega-3 polyunsaturated fatty acids. Aquaculture 322-323: 82-90.
- Nguyen, T. M. (2015). Effects of Dietary Probiotics and Temperatures Stress on Growth and Immunity related Genes Expression in Malaysian Mahseer (*Tor tambroides*). Master Thesis, Universiti Malaysia Terengganu.
- Nguyen, N. P., Warnow, T., Pop, M. & White, B. (2016). A perspective on 16S rRNA operational taxonomic unit clustering using sequence similarity. Npj Biofilms and Microbiomes 2, 16004. Retrieved 31 July 2017 from <https://www.nature.com/articles/npjbiofilms20164.pdf>.
- Nguyen, T. L., Park, C. I. & Kim, D. H. (2017). Improved growth rate and disease resistance in olive flounder, *Paralichthys olivaceus*, by probiotic *Lactococcus lactis* WFLU12 isolated from wild marine fish. Aquaculture 471: 113-120.
- Ni, J., Yan, Q., Yu, Y. & Zhang, T. (2014a). Fish gut microecosystem: a model for detecting spatial pattern of microorganisms. Chinese Journal of Oceanology and Limnology 32(1): 54-57.
- Ni, J., Yan, Q., Yu, Y. & Zhang, T. (2014b). Factors influencing the grass carp gut microbiome and its effect on metabolism. FEMS Microbiology Ecology 87(3): 704-714.
- Nie, L., Zhou, Q. J., Qiao, Y. & Chen, J. (2017). Interplay between the gut microbiota and immune responses of ayu (*Plecoglossus altivelis*) during *Vibrio anguillarum* infection. Fish & Shellfish Immunology 68: 479-487.
- Norfatimah, M. Y., The, L. K., Salleh, M. Z., Mat Isa, M. N. & SitiAzizah, M. N. (2014). Complete mitochondrial genome of Malaysian Mahseer (*Tor tambroides*). Gene 548(2): 263-269.
- Nyman, A., Huyben, D., Lundh, T. & Dicksved, J. (2017). Effects of microbe- and mussel-based diets on the gut microbiota in Arctic charr (*Salvelinus alpinus*). Aquaculture Reports 5: 34-40.
- Nyrén, P. I. & Lundin, A. (1985). Enzymatic method for continuous monitoring of inorganic pyrophosphate synthesis. Analytical Biochemistry 509: 504-509.

- O'Hara, A. M., & Shanahan, F. (2006). The gut flora as a forgotten organ. *EMBO Reports* 7(7): 688-693.
- O'Hara, A. M., & Shanahan, F. (2007). Mechanisms of Action of Probiotics in Intestinal Diseases. *The Scientific World Journal* 7: 31-46.
- Ohland, C. L. & MacNaughton, W. K. (2010). Probiotic bacteria and intestinal epithelial barrier function. *American Journal of Physiology – Gastrointestinal and Liver Physiology* 298(6): G807-G819.
- Olivia-Teles, A. (2012). Nutrition and health of aquaculture fish. *Journal of Fish Diseases* 35(20): 83-108.
- Onifade, A. A., Obiyan, R. I., Onipede, E., Adejumo, O. A., Abu, O. A. & Babatune, G. M. (1999). Assessment of the effects of supplementing rabbit diets with a culture of *Saccharomyces cerevisiae* using growth performance, blood composition and clinical enzyme activities. *Animal Feed Science and Technology* 77: 25-32.
- Orlich, M. J., Siapco, G. & Jung, S. (2017). 24 – Vegetarian Diets and the Microbiome. In *Vegetarian and Plant-Based Diets in Health and Disease Prevention 1st Edition*, ed. Mariotti, F., pp. 429-461. Academic Press.
- Othman, A. A. (2003). Freshwater Fishes of Gelami Lemi. Freshwater Fisheries Research Center, Department of Fisheries Malaysia.
- Pajarillo, E. A., Chae, J. P., Balolong, M. P., Kim, H. B., Seo, K. S. & Kang, D. K. (2015). Characterization of the Fecal Microbial Communities of Duroc Pigs using 16S rRNA Gene Pyrosequencing. *Asian-Australian Journal of Animal Sciences* 28(4): 584-591.
- Palumbi, S., Martin, A., Romano, S., McMillan, W. O., Stice, L., Grabowski, G. (1991). The Simple Fool's Guide to PCR. Department of Zoology and Kewalo Marine Laboratory, University of Hawaii, Honolulu.
- Pande, G. S. J., Scheie, A. A., Benneche, T., Wille, M., Sorgeloos, P., Bossier, P. & Defoirdt, T. (2013). Quorum sensing-disrupting compounds protect larvae of the giant freshwater prawn *Macrobrachium rosenbergii* from *Vibrio harveyi* infection. *Aquaculture* 406-407: 121-124.
- Panigrahi, A., Kiron, V., Puangkaew, J., Kobayashi, T., Satoh, S., Sugita, H. (2005). The viability of probiotic bacteria as a factor influencing the immune response in rainbow trout *Oncorhynchus mykiss*. *Aquaculture* 243:241-254.
- Park, Y., Moniruzzaman, M., Lee, S., Hong, J., Won, S., Lee, J. M., Yun, H., Kim, K. W., Ko, D. & Bai, S. C. (2016). Comparison of the effects of dietary single and multi-probiotics on growth, non-specific immune responses and disease resistance in starry flounder, *Platichthys stellatus*. *Fish & Shellfish Immunology* 59: 351-357.

- Patel, R. M. & Denning, P. W. (2013). Therapeutic Use of Prebiotics, Probiotics, and Postbiotics to Prevent Necrotizing Enterocolitis: What is the Current Evidence? *Clinics in Perinatology* 40(1): 11-25.
- Peng, B., Ye, J., Han, Y., Zeng, L., Zhang, J. & Li, H. (2016). Identification of polyvalent protective immunogens from outer membrane proteins in *Vibrio parahaemolyticus* to protect fish against bacterial infection. *Fish & Shellfish Immunology* 54:204-210.
- Phan, T. H. & Nguyen, D. L. (2012). Species-specificity of DNA trimer densities in chromosomes and their use in the classification of closely related organisms. *Journal of Microbiological Methods* 91(1): 30-37.
- Plummer, E., Twin, J., Bulach, D. M., Garland, S. M. & Tabrizi, S. N. (2015). A Comparison of Three Bioinformatics Pipelines for the Analysis of Preterm Gut Microbiota using 16S rRNA Gene Sequencing Data. *Journal of Proteomics & Bioinformatics* 8(12): 283-291.
- Pray, L. A. (2008). Discovery of DNA structure and function: Watson and Crick. *Nature Education* 1(1):100.
- Price, S. A., Friedman, S. T. & Wainwright, P. C. (2015). How predation shaped fish: the impact of fin spines on body form evolution across teleosts. *Proceedings of the Royal Society B* 282: 20151428.
- Quail, M. A., Smith, M., Coupland, P., Otto, T. D., Harris, S. R., Connor, T. R., Bertoni, A., Swerdlow, H. P. & Gu, Y. (2012). A tale of three next generation sequencing platforms: comparison of Ion Torrent, Pacific Biosciences and Illumina MiSeq sequencers. *BMC Genomics* 13:341.
- Rada, V., Splichal, I., Rockova, S., Grmanova, M. & Vlkova, E. (2010). Susceptibility of bifidobacteria to lysozyme as a possible selection criterion for probiotic bifidobacterial strains. *Biotechnology Letters* 32(3): 451-455.
- Radovanović, T. B., Mitić, S. S. B., Perendija, B. R., Despotović, S. G., Pavlović, S. Z., Cakić, P. D. & Saičić, Z. S. (2010). Superoxide Dismutase and Catalase Activities in the Liver and Muscle of Barbel (*Barbus Barbus*) and Its Intestinal Parasite (*Pomphoryinchus Laevis*) from the Danube River, Serbia. *Archives of Biological Sciences* 62(1): 97-105.
- Ramesh, D., Vinothkanna, A., Rai, A. K. & Vignesh, V. S. (2015). Isolation of potential probiotic *Bacillus* spp. and assessment of their subcellular components to induce immune responses in *Labeo rohita* against *Aeromonas hydrophila*. *Fish & Shellfish Immunology* 45(2): 268-276.
- Ramesh, D., Souissi, S., Ahmad, S. T. (2017). Effects of the potential probiotics *Bacillus aerophilus* KADR3 in inducing immunity and disease resistance in *Labeo rohita*. *Fish & Shellfish Immunology* 70: 408-415.

- Ramezani-Fard, E. & Kamarudin, M. S. (2012). Malaysian Mahseer: New Candidate for Asian Aquaculture? Global Aquaculture Alliance March/April 2012. Retrieved 25 April 2017 from <https://pdf.gaalliance.org/pdf/GAA-Fard-Mar12.pdf>.
- Ramezani-Fard, E., Kamarudin, M. S., Saad, C. R., Harmin, S. A. & Goh, Y. M. (2012). Dietary Lipid Levels Affect Growth and Fatty Acid Profiles of Malaysian Mahseer *Tor tambroides*. North American Journal of Aquaculture 74(4): 530-536.
- Ramirez, R. F. & Dixon, B. A. (2003). Enzyme production by obligate intestinal anaerobic bacteria isolated from oscars (*Astronotus ocellatus*), angelfish (*Pterophyllum scalare*) and southern flounder (*Paralichthys lethostigma*). Aquaculture 227(1-4): 417-426.
- Ramírez, C. & Romero, J. (2017). The Microbiome of *Seriola lalandi* of Wild and Aquaculture Origin Reveals Differences in Composition and Potential Function. Frontiers in Microbiology 8: 1844 doi: 10.3389/fmicb.2017.01844.
- Rasheeda, M. K., Ramgamaran, V. R., Srinivasan, S., Ramaiah, S. K., Gunasekaran, R., Jaypal, S., Gopal, D. & Ramalingam, K. (2017). Comparative profiling of microbial community of three economically important fishes reared in sea cages under tropical offshore environment. Marine Genomics 34: 57-65.
- Ray, W. (1972). Nucleotide Sequence Analysis of DNA. Nature New Biology 236:198-200.
- Redfield, R. J. (2002). Is quorum sensing a side effect of diffusion sensing? Trends in Microbiology 10(8): 365-370.
- Rideout, J. R., He, Y., Navas-Molina, J. A., Walters, W. A., Ursell, L. K., Gibbons, S. M., Chase, J., McDonald, D., Gonzalez, A., Robbins-Pianka, A., Clemente, J. C., Gilbert, J. A., Huse, S. M., Zhou, H. W., Knight, R., Caporaso, J. G. (2014). Subsampled open-reference clustering creates consistent, comprehensive OTU definitions and scales to billions of sequences. PeerJ 2:e545. Retrieved 19 May 2016 from <https://peerj.com/articles/545/>.
- Ringø, E., Dimitroglou, A., Hoseinifar, S. H. and Davies, S. J. (2014). Prebiotics in Finfish: An Update. In *Aquaculture Nutrition: Gut Health, Probiotics and Prebiotics*, ed. Merrifield, D. and Ringø, E, John Wiley & Sons, Ltd, Chichester, UK.
- Ringø, E., Zhou, Z., Vecino, J. L. G., Wadsworth, S., Romero, J., Korgdahl, A., Olsen, R. E., Dimitroglou, A., Foey, A., Davies, S., Owen, M., Lauzon, H. L., Martinsen, L. L., De Schryver, P., Bossier, P., Sperstad, S. & Merrifield, D. L. (2016). Effect of dietary components on the gut microbiota of aquatic animals. A never-ending story? Aquaculture Nutrition 22(2): 219-282.
- Roeselers, G., Mittge, E. K., Stephens, W. Z., Parichy, D. M., Cavanaugh, C. M., Guillemin, K. & Rawls, J. F. (2011). Evidence for a core gut microbiota in the zebrafish. ISME Journal 5(10):1595-1608.

- Roeselers, G., Bouwman, J. & Levin, E. (2016). The human gut microbiome, diet, and health: “Post hoc non ergo propter hoc”. Trends in Food Science & Technology 57(B): 302-305.
- Rossi-Tamisier, M., Benamar, S., Raoult, D. & Fournier, P. E. (2015). Cautionary tale of using 16S rRNA gene sequence similarity values in identification of human-associated bacterial species. International Journal of Systematic and Evolutionary Microbiology 65(Pt 6):1929-1934.
- Rouf, A. J. M. A., Phang, S. M. & Ambak, M. A. (2010). Depth distribution and ecological preferences of periphytic algae in Kenyir Lake, the largest tropical reservoir of Malaysia. Chinese Journal of Oceanology and Limnology 28(4): 856-867.
- Ryan, J. R. J. & Esa, Y. B. (2006). Phylogenetic Analysis of *Hampala* Fishes (Subfamily Cyprininae) in Malaysia Inferred from Partial Mitochondrial Cytochrome b DNA Sequences. Zoological Science 23: 893-901.
- Safari, R., Adel, M., Lazado, C. C., Caipang, C. M. A. & Dadar, M. (2016). Host-derived probiotics *Enterococcus casseliflavus* improves resistance against *Streptococcus iniae* infection in rainbow trout (*Oncorhynchus mykiss*) via immunomodulation. Fish & Shellfish Immunology 52: 198-205.
- Safari, R., Hoseinifar, S. H., Nejadmoqaddam, S. & Khalili, M. (2017). Apple cider vinegar boosted immunomodulatory and health promoting effects of *Lactobacillus casei* in common carp (*Cyprinus carpio*). Fish & Shellfish Immunology. In Press, Accepted Manuscript. Available online 7 June 2017.
- Salas-Jara, M. J., Ilabaca, A., Vega, M. & García, A. (2016). Biofilm Forming *Lactobacillus*: New Challenges for the Development of Probiotics. Microorganisms 4(3): 35.
- Salinas, I & Magadán, S. (2017). Omics in fish mucosal immunity. Developmental & Comparative Immunology 75: 99-108.
- Sanger, F., Brownlee, G. G. & Barrell, B. G. (1965). A two-dimensional fractionation procedure for radioactive nucleotides. Journal of Molecular Biology 13(2): 373-398.
- Sanger, F., Nicklen, S & Coulson, A. R. (1977). DNA sequencing with chain-terminating inhibitors. Proceedings of the National Academy of Sciences of the United States of America 74(12): 5463-5467.
- Santhanam, R. (2015). Nutritional Freshwater Life. CRC Press, pp. 154.
- Sati, J., Sah, S., Pandey, H., Ali, S., Sahoo, P. K., Pande, V. & Barat, A. (2013). Phylogenetic relationship and molecular identification of five Indian Mahseer species using COI sequence. Journal of Environmental Biology 34: 933-939.
- Schloss, P. D. & Handelsman, J. (2005). Metagenomics for studying unculturable microorganisms: cutting the Gordian knot. Genome Biology 6(8): 229.

- Schloss, P. D. & Westcott, S. L. (2011). Assessing and Improving Methods Used in Operational Taxonomic Unit-Based Approaches for 16S rRNA Gene Sequence Analysis. *Applied and Environmental Microbiology* 77(10): 3219-3226.
- Shahi, N. & Mallik, S. K. (2014). Recovery of *Pseudomonas koreensis* from eye lesions in golden mahseer, *Tor putitora* (Hamilton, 1822) in Uttarakhand, India. *Journal of Fish Diseases* 37(5): 497-500.
- Shahi, N., Sharma, P., Pandey, J., Bisht, I. & Malik, S. K. (2018). Characterization and pathogenicity study of *Chryseobacterium scophthalmum* recovered from gill lesions of diseased golden mahseer, *Tor putitora* (Hamilton, 1822) in India. *Aquaculture* 485: 81-92.
- Shiau, S. Y. (1997). Utilization of carbohydrates in warmwater fish- with particular reference to tilapia, *Oreochromis niloticus* X *O. aureus*. *Aquaculture* 151(1-4): 79-96.
- Shobharani, P., Padmaja, R. J., Halami, P. M. (2015). Diversity in the antibacterial potential of probiotic cultures *Bacillus licheniformis* MCC2514 and *Bacillus licheniformis* MCC2512. *Research in Microbiology* 166(6): 546-554.
- Sievers, F., Wilm, A., Dineen, D. G., Gibson, T. J., Karplus, K., Li, W., Lopez, R., McWilliam, H., Remmert, M., Söding, J., Thompson, J. D. & Higgins, D. (2011). Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Molecular Systems Biology* 7: 539 doi:10.1038/msb.2011.75.
- Simbolo, M., Gottardi, M., Corbo, V., Fassan, M., Maffioli, A., Malpeli, G., Lawlor, R. T. & Scarpa, A. (2013). DNA Qualification Workflow for Next Generation Sequencing of Histopathological Samples. *PLoS ONE* 8(6): e62692. <https://doi.org/10.1371/journal.pone.0062692>.
- Singh, V., Chaudhary, D. K. & Mani, I. (2012). Molecular Characterization and Modeling of Secondary Structure of 16S rRNA from *Aeromonas veronii*. *International Journal of Applied Biology and Pharmaceutical Technology* 3(1): 253-260.
- Song, W., Li, L., Huang, H., Jiang, K., Zhang, F., Chen, X., Zhao, M. & Ma, L. (2016). The Gut Microbial Community of Antarctic Fish Detected by 16S rRNA Gene Sequence Analysis. *BioMed Research International* 2016: ID3241529.
- Spanggaard, B., Huber, I., Nielsen, J., Sick, E. B., Pipper, C. B., Martinussen, T., Slierendrech, W. J. & Gram, L. (2001). The probiotic potential against vibriosis of the indigenous microflora of rainbow trout. *Environmental Microbiology* 3(12): 755-765.
- Spellerberg, I. F. & Fedor, P. J. (2003). A tribute to Claude Shannon (1916–2001) and a plea for more rigorous use of species richness, species diversity and the ‘Shannon–Wiener’ Index. *Global Ecology and Biogeography* 12(3): 177-179.

- Srinivasan, R., Karaoz, U., Volegova, M., MacKichan, J., Kato-Maeda, M., Miller, S., Nadarajan, R., Brodie, E. L. & Lynch, S. V. (2015). Use of 16S rRNA Gene for Identification of a Broad Range of Clinically Relevant Bacterial Pathogens. *PLoS ONE* 10(2): e0117617.
- Srisapoome, P. & Areechon, N. (2017). Efficacy of viable *Bacillus pumilus* isolated from farmed fish on immune responses and increased disease resistance in Nile tilapia (*Oreochromis niloticus*): Laboratory and on-farm trials. *Fish & Shellfish Immunology* 67: 199-210.
- Sugita, H., Miyajima, C. & Deguchi, Y. (1999). The vitamin B₁₂-producing ability of the intestinal microflora of freshwater fish. *Aquaculture* 92: 267-276.
- Sullam, K. E., Essinger, S. D., Lozupone, C. A., O'Connor, M. P., Rosen, G. L., Knight, R., Kilham, S. S. & Russel, J. A. (2012). Environmental and ecological factors that shape the gut bacterial communities of fish: a meta-analysis. *Molecular Ecology* 21(13): 10.1111/j.1365-294X.2012.05552.x.
- Sun, Y. Z., Yang, H. L., Ma, R. L. & Lin, W. Y. (2010). Probiotic applications of two dominant gut *Bacillus* strains with antagonistic activity improved the growth performance and immune responses of grouper *Epinephelus coioides*. *Fish & Shellfish Immunology* 29: 803-809.
- Suzer, C., Çoban, D., Kamaci, H. O., Saka, S., Firat, K., Otgucuoğlu, Ö., Küçüksarı, H. (2008). *Lactobacillus spp.* bacteria as probiotics in gilthead sea bream (*Sparus aurata*, L.) larvae: effect on growth performance and digestive enzyme activities. *Aquaculture* 280:140-145.
- Székely, C., Shaharom, F., Cech, G., Mohamed, K., Zin, N. A., Borkhanuddin, M. H., Ostoros, G. & Molnár, K. (2012). Myxozoan infection of the Malaysian mahseer, *Tor tambroides*, of Tasik Kenyir Reservoir, Malaysia: description of a new species *Myxobolus tambroides* sp.n. *Parasitology Research* 111(4): 1749-1756.
- Tan, E. S. P. (1980). Some Aspects of the Biology of Malaysian Riverine Cyprinids. *Aquaculture* 20: 281-289.
- Tan, W. S., Abdullah, J. O., Sieo, C. C., Shafee, N., Mustafa, S., Leow, T. C. A., Jahanshiri, F., Saad, W. Z., Tong, C. C., Wahab, M. N. A. & Abdullah, N. (Eds.). (2008). A Laboratory Manual on Molecular Techniques for Identification of Bacteria. Department of Microbiology, Faculty of Biotechnology and Biomolecular Sciences, Universiti Putra Malaysia.
- Thériault, V., Moyer, G. R., Jackson, L. S., Blouin, M. S. & Banks, M. A. (2011). Reduced reproductive success of hatchery coho salmon in the wild: insights into most likely mechanisms. *Molecular Ecology* 20: 1860-1869.
- Thomas, T., Gilbert, J. & Meyer, F. (2012). Metagenomics - a guide from sampling to data analysis. *Microbial Informatics and Experimentation* 2:3.

- Thy, H. T. T., Tri, N. N., Quy, O. M., Fotedar, R., Kannika, K., Unajak, S. & Areechon, N. (2017). Effects of the dietary supplementation of mixed probiotic spores of *Bacillus amyloliquefaciens* 54A, and *Bacillus pumilus* 47B on growth, innate immunity and stress responses of striped catfish (*Pangasianodon hypophthalmus*). Fish & Shellfish Immunology 60: 391-399.
- Tobe, S. S., Kitchener, A. C. & Linacre, A. (2011). Assigning confidence to sequence comparisons for species identification: A detailed comparison of the cytochrome *b* and cytochrome oxidase subunit I mitochondrial genes. Forensic Science International: Genetics Supplement Series 3(1): e246-e247.
- Tocher, D. R. (2010). Fatty acid requirements in ontogeny of marine and freshwater fish. Aquaculture Research 41(5): 717-732.
- Travers, A. & Muskhelishvili, G. (2015). DNA Structure and Function. The FEBS Journal 282: 2279-2295.
- Tsuchiya, C., Sakata, T. & Sugita, H. (2007). Novel ecological niche of *Cetobacterium somerae*, an anaerobic bacterium in the intestinal tracts of freshwater fish. The Society for Applied Microbiology, Letter in Applied Microbiology 46:43-48.
- Turcatti, G., Romieu, A., Fedurco, M. & Tairi, A. P. (2008). A new class of cleavable fluorescent nucleotides: synthesis and optimization as reversible terminators for DNA sequencing by synthesis. Nucleic Acids Research 36(4): e25.
- Turner, S., Pryer, K. M., Miao, V. P. W., & Palmer, J. D. (1999). Investigating deep phylogenetic relationships among cyanobacteria and plastids by small subunit rRNA sequence analysis. Journal of Eukaryotic Microbiology 46: 327–338.
- Udayangani, R. M. C., Dananjaya, S. H. S., Nikapitiya, C., Heo, G. J., Lee, J. & Zoysa, M. D. (2017). Metagenomics analysis of gut microbiota and immune modulation in zebrafish (*Danio rerio*) fed chitosan silver nanocomposites. Fish & Shellfish Immunology 66: 173-184.
- Van Kessel, M. A. H. J., Dutilh, B. E., Neveling, K., Kwint, M. P., Veltman, J. A., Flik, G., Jetten, M. S. M., Klaren, P. H. M., Camp, J. M. O. D. (2011). Pyrosequencing of 16S rRNA gene amplicons to study the microbiota in the gastrointestinal tract of carp (*Cyprinus carpio L.*). AMBI Express 1: 41.
- Varela, J. L., Ruiz-Jarabo, I., Vargas-Chacoff, L., Arijo, S., León-Rubio, J. M., García-Millán, I., Río, M. P. M., Moriñigo, M. A. & Mancera, J. M (2010). Dietary administration of probiotic Pdp11 promotes growth and improves stress tolerance to high stocking density in gilthead seabream *Sparus auratus*. Aquaculture 309: 265-271.
- Vazquez-Baeza, Y., Pirrung, M., Gonzalez, A. & Knight, R. (2013). Emperor: A tool for visualizing high-throughput microbial community data. Gigascience 2(1):16.

- Vinoj, G., Jayakumar, R., Chen, J. C., Withyachumnarnkul, B., Shanthi, S. & Vaseeharan, B. (2015). N-hexanoyl-L-homoserine lactone-degrading *Pseudomonas aeruginosa* PsDAHP1 protects zebrafish against *Vibrio parahaemolyticus* infection. Fish & Shellfish Immunology 42(1): 204-212.
- Voelkerding, K. V., Dames, S. A. & Durtschi, J. D. (2009). Next-generation sequencing: from basic research to diagnostics. Clinical Chemistry 55: 641-658.
- Wang, Y. B. & Xu, Z. R. (2006). Effect of probiotics for common carp (*Cyprinus carpio*) based on growth performance and digestive enzyme activities. Animal Feed Science and Technology 127: 283-292.
- Wang, Y. B., Tian, Z. Q., Yao, J. T. & Li, W. F. (2008). Effect of probiotics, *Enteroccus faecium*, on tilapia (*Oreochromis niloticus*) growth performance and immune response. Aquaculture 277: 203-207.
- Watson, J. D. & Crick, F. H. C. (1953). Molecular Structure of Nucleic Acids. Nature 171(4356): 737-738.
- West, S. A., Winzer, K., Gardner, A. & Diggle, S. P. (2012). Quorum sensing and the confusion about diffusion. Trends in Microbiology 20(12): 586-594.
- Wesseling, W., Wittka, S., Kroll, S., Soltmann, C., Kegler, P., Kunzmann, A., Riss, H. W. & Lohmeyer, M. (2015). Functionalised ceramic spawning tiles with probiotic *Pseudoalteromonas* biofilms designed for clownfish aquaculture. Aquaculture 446: 57-66.
- Whittaker, R. H. (1972). Evolution and measurement of species diversity. Taxon 21:213-251.
- Wilkins, M. H. F., Stokes, A. R. & Wilson, H. R. (1953). Molecular Structure of Deoxypentose Nucleic Acids. Nature 171: 738-740.
- Wilson, R. P. (2002). Amino Acids and Proteins. In *Fish Nutrition 3rd Edition*, ed. Halver, J. E. and Hardy, R. W., pp. 143-179. Academic Press.
- Woese, C. R. & Fox, G. E. (1977). Phylogenetic structure of the prokaryotic domain: The primary kingdoms. Proceedings of the National Academy of Sciences of the United States of America 74(11): 5088-5090.
- Woo, P. C. Y., Lau, S. K. P., Teng, J. L. L., Tse, H. & Yuen, K. Y. (2008). Then and now: use of 16S rDNA gene sequencing for bacterial identification and discovery of novel bacteria in clinical microbiology laboratories. Clinical Microbiology and Infection 14(10): 908-934.
- Wu, S., Wang, G., Angert, E. R., Wang, W., Li, W. & Zou, H. (2012). Composition, Diversity, and Origin of the Bacterial Community in Grass Carp Intestine. PLoS ONE 7(2) e30440.

- Wu, Z. Q., Jiang, C., Ling, F. & Wang, G. X. (2015). Effects of dietary supplementation of intestinal autochthonous bacteria on the innate immunity and disease resistance of grass carp (*Ctenopharyngodon idellus*). Aquaculture 438: 105-114.
- Xia, J. H., Lin, G., Fu, G. H., Wan, Z. Y., Lee, M., Wang, L., Liu, X. J., Yue, G. H. (2014). The intestinal microbiome of fish under starvation. BMC Genomics 15:266. doi: 10.1186/1471-2164-15-266.
- Xie, D., Yang, L., Yu, R., Chen, F., Lu, R., Qin, C. & Nie, G. (2017). Effects of dietary carbohydrate and lipid levels on growth and hepatic lipid deposition of juvenile tilapia, *Oreochromis niloticus*. Aquaculture 479: 696-703.
- Xing, M., Hou, Z., Yuan, J., Liu, Y., Qu, Y. & Liu, B. (2013). Taxonomic and functional metagenomic profiling of gastrointestinal tract microbiome of the farmed adult turbot (*Scophthalmus maximus*). FEMS Microbiology Ecology 86(3): 432-443.
- Xu, T. & Zhang, X. H. (2014). *Edwardsiella tarda*: an intriguing problem in aquaculture. Aquaculture 431: 129-135.
- Xu, J., Zeng, X., Jiang, N., Zhou, Y. & Zeng, L. (2015). *Pseudomonas alcaligenes* infection and mortality in cultured Chinese sturgeon, *Acipenser sinensis*. Aquaculture 446: 37-41.
- Yan, Q., Li, J., Yu, Y., Wang, J., He, Z., Nostrand, J. D. V., Kemphher, M. L., Wu, L., Wang, Y., Liao, L., Li, X., Wu, S., Ni, J., Wang, C. & Zhou, J. (2016). Environmental filtering decreases with fish development for the assembly of gut microbiota. Environmental Microbiology DOI: 10.1111/1462-2920.13365.
- Yoon, S. H., Ha, S. M., Kwon, S., Lim, J., Kim, Y., Seo, H. & Chun, J. (2017). Introducing EzBioCloud: A taxonomically united database of 16S rRNA and whole genome assemblies. International Journal of Systematic and Evolutionary Microbiology 67:1613-1617.
- Zhang, J., Kobert, K., Flouri, T. & Stamatakis, A. (2014). PEAR: a fast and accurate Illumina Paired-End reAd merger. Bioinformatics 30(5): 614-620.
- Zhang, M., Sun, Y., Liu, Y., Qiao, F., Chen, L., Liu, W. T., Du, Z. & Li, E. (2016). Response of gut microbiota to salinity change in two euryhaline aquatic animals with reverse salinity preference. Aquaculture 454: 72-80.
- Zhao, Y., Yuan, L., Wan, J., Sun, Z., Wang, Y. & Sun, H. (2016). Effects of potential probiotic *Bacillus cereus* EN25 on growth, immunity and disease resistance of juvenile sea cucumber *Apostichopus japonicas*. Fish & Shellfish Immunology 49: 237-242.
- Zhou, X., Tian, Z., Wang, Y. & Li, W. (2010). Effect of treatment with probiotics as water additives on tilapia (*Oreochromis niloticus*) growth performance and immune response. Fish Physiology and Biochemistry 36(3): 501-509.

Zorriehzahra, M. J., Delshad, S. T., Adel, M., Tiwari, R., Karthik, K., Dhama, K. & Lazado, C. (2016). Probiotics as beneficial microbes in aquaculture: an update on their multiple modes of action: a review. Veterinary Quarterly 36(4): 228-241.

Zupa, R., Rodríguez, C., Mylonas, C. C., Rosenfeld, H., Fakriadis, I., Papadaki, M., Pérez, J. A., Pousis, C., Basilone, G & Corriero, A. (2017). Comparative Study of Reproductive Development in Wild and Captive-Reared Greater Amberjack *Seriola dumerili* (Risso, 1810). PLoS ONE. Retrieved 6 May 2017 from <https://doi.org/10.1371/journal.pone.0169645>.

