



UNIVERSITI PUTRA MALAYSIA

***RHEOLOGICAL, SPECTRAL AND THERMAL ANALYSES OF GELLAN/
DEXTRAN BLENDS AS GELATIN SUBSTITUTES***

NURUL HAWA AHMAD

IPPH 2014 9



**RHEOLOGICAL, SPECTRAL AND THERMAL ANALYSES OF GELLAN/
DEXTRAN BLENDS AS GELATIN SUBSTITUTES**

By

NURUL HAWA AHMAD

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirement for the Degree of Master of Science**

November 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from copyright holder. Commercial use of material only be made with express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**RHEOLOGICAL, SPECTRAL AND THERMAL ANALYSES OF GELLAN/
DEXTRAN BLENDS AS GELATIN SUBSTITUTES**

By

NURUL HAWA AHMAD

November 2014

Chair: Professor Shuhaimi Mustafa, PhD

Institute: Halal Products Research Institute

Microbial polysaccharides can serve as renewable sources of hydrocolloids that have great function as viscosifying, stabilizing, emulsifying or gelling agents which are important to improve texture and stability of food and pharmaceutical products. Similar functions are currently being offered by gelatin however the use of gelatin would give rise to health and religious concerns for particular group of consumers. Plant based polysaccharides have been utilized as gelatin substitute but plant requires longer harvest maturity and susceptible to geographical and seasonal changes. Viscosity and gel rigidity are important parameter to determine adequate food texture for mouth feel properties. Phase separation also could affect consumer perception on food products. The aims of this study are to investigate rheological properties that measure flow behaviour, viscosity and gel rigidity to be compared with gelatin as well as to predict molecular interaction between gellan (G) and dextran (D) in their binary blends (1:1, 1:2, 1:3 ratios) in varying concentration range of 0.5%, 1.5% and 3% (w/v) using spectral and thermal characteristics. For rheological measurements, all samples were subjected to steady shear and dynamic shear viscosity tests using rheometer. Spectral analysis and thermal analysis were performed using Fourier Transform Infrared (FTIR) and Differential Scanning Calorimetry (DSC) respectively. Rheological analyses revealed that during blending, dextran plays a significant role and transforms the solid-like characteristics to liquid-like behaviour of gellan with an increase in dextran concentration of G/D blends. The steady flow behaviour was well characterized by the Herschel-Bulkley model (standard error lower than 10) and G/D blends exhibited similar flow behaviour (shear thinning) as bovine and porcine gelatin at 1.5% and 3% concentration as compared to dilute concentration, 0.5% . For dynamic viscoelastic, G/D blends exhibit similar gel characteristic with both porcine and bovine gelatin at 3% as opposed to 0.5% and 1.5%. From spectral characteristic of the blend, a shift in the major

gellan band of carboxylic group at 1414 cm^{-1} and disappearance of two gellan bands at 1072 cm^{-1} and 1042 cm^{-1} as well as dextran band at 1080 cm^{-1} in glycosidic linkage were observed. DSC thermograms showed a single exothermic peak accompanied by a shifting to higher melting temperature for all blends in comparison of individual polysaccharide. These changes suggest a possible molecular interaction event between gellan and dextran. In conclusion, gellan and dextran blends exhibit desired viscosity for mouth feel properties and favorable molecular interaction depended on the amount of added dextran. This interaction is important to design food and pharmaceutical products with desired textural and stability attributes.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**ANALISIS REOLOGI, SPEKTRA DAN TERMA BAGI CAMPURAN GELLAN/
DEXTRAN SEBAGAI GANTIAN GELATIN**

Oleh

NURUL HAWA AHMAD

November 2014

Pengerusi: Profesor Shuhaimi Mustafa, PhD

Institut: Institut Penyelidikan Produk Halal

Polisakarida mikrob adalah sumber hidrokoloid yang berfungsi sebagai agen pemekat, penstabil, mengemulsi dan pelekat yang sangat penting untuk menambah baik tekstur dan kestabilan produk makanan dan farmaseutikal. Gelatin dari sumber haiwan mempunyai fungsi yang sama tetapi terdapat isu yang kritikal yang berkait rapat dengan isu keagamaan dan kesihatan pengguna. Hidrokoloid dari sumber tumbuhan digunakan untuk sebagai bahan ganti bagi gelatin tetapi tumbuhan memerlukan tempoh masa yang lama untuk mencapai tahap tuai matang dan terdedah kepada risiko perubahan geografi dan cuaca. Kelikatan dan ketegaran gel merupakan parameter yang penting untuk menentukan struktur makanan yang sesuai bagi perangsangan deria mulut. Fasa pengasingan di dalam makanan boleh persepsi pengguna kepada sesuatu produk makanan. Objektif penyelidikan ini adalah untuk mengkaji ciri-ciri reologi yang menentu ukur sifat aliran, kelikatan dan ketegaran gel untuk dibandingkan kepada ciri-ciri reologi yang ada pada gelatin serta mengkaji interaksi molekul antara gellan dan dextran pada nisbah 1:1, 1:2, 1:3 pada kepekatan 0.5%, 1.5% dan 3%. Analisa reologi iaitu *shear* stabil and dinamik dijalankan dengan menggunakan AR-G2 Rheometer. Analisa spektra dijalankan menggunakan inframerah peralihan-Fourier (FTIR) dan analisa terma menggunakan kalorimetri imbasan pembeza (DSC). Analisa reologi membuktikan bahawa dextran bertindak untuk mengubah sifat pepejal gellan kepada sifat cecair dengan peningkatan dextran di dalam campuran gellan dan dextran. Kestabilan sifat aliran juga diterangkan secara mendalam melalui model *Herschel-Bulkley* (ralat piawai di bawah 10) dan sifat aliran (*shear thinning*) campuran gellan dan dextran menunjukkan ciri yang sama pada gelatin pada kepekatan 1.5% dan 3% berbanding 0.5%. Bagi viskoelastik dinamik, campuran gellan dan dextran menunjukkan

karakter gel yang sama pada gelatin pada kepekatan 0.3% berbanding 0.5% dan 1.5%. Bagi analisa spektra, hasil eksperimen mendapati terdapat gellan mengalami perubahan pada kedudukan spektrum kumpulan karbosilik pada 1414 cm^{-1} dan kehilangan dua spektrum pada 1072 cm^{-1} dan 1042 cm^{-1} di dalam interaksi glikosida. Perubahan pada interaksi glikosida dextran dilihat pada 1080 cm^{-1} . Lingkaran DSC menunjukkan satu puncak *exothermic* diiringi dengan peningkatan suhu takat cair untuk kesemua campuran gellan/dextran berbanding gellan dan dextran secara tersendiri. Perubahan ini membawa kepada potensi interaksi molekular yang mungkin berlaku antara gellan dan dextran. Kesimpulannya, campuran gellan dan dextran mempunyai kelikatan yang sesuai bagi perangsangan deria mulut dan interaksi molekular yang padan bergantung kepada jumlah dextran. Interaksi ini dapat memberi maklumat yang berguna sebelum menghasilkan sesuatu produk makanan dan farmaseutikal dengan tekstur yang dikehendaki dan lebih stabil.

ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and the Most Merciful Allah Almighty, all praises to Allah for the strength, perseverance and good health to complete this thesis. My first and foremost thanks must go to my beloved husband, Muhammad Amsyar Jafri as well as my parents, Ahmad Wardi and Rosnani Hamim, for their endless encouragement and support. I would like to express my deepest gratitude to my supervisor, Professor Shuhaimi Mustafa for his incessant guidance for this research work. I bestow my earnest appreciation to Professor Madya Dzulkifly Mat Hashim for his endless assistance, precious remark, undying efforts and constructive opinion on this project. My special thanks must go to Professor Dato' Dr. Mohd Yazid Abd Manap, Dr. Ally El Shekar, Dr. Jasim Ahmed for their constant assistances throughout the course of my study. I would like to convey my remarkable recognition to my late supervisor, Professor Dato' Dr. Yaakob Che Man for being a great mentor during his presence. His full dedication and devotion to uplift research expertise in halal products, without doubt, is a significant contribution for the nation. To my dearest Nurrulhidayah, Nina Naquiah, Noorashikin, Hajaratul Najwa, Yanty, Aisyah, Dalila, Saliza and Atiqah thank you for being there through my joy and pain. I also would like to acknowledge Amran, Zhaffan, Rosmawati and Faizal for their technical assistances for this project. This scientific research project could not be achieved without the cooperation of all aforementioned people as well as numerous individuals who have directly or indirectly affected my research experience at UPM.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Shuhaimi Mustafa, PhD

Professor
Halal Products Research Institute
Universiti Putra Malaysia
(Chairman)

Dzulkifly Mat Hashim, Msc

Professor Madya
Halal Products Research Institute
Universiti Putra Malaysia
(Member)

Mohd Yazid Abd. Manap, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work
- quotations, illustrations and citations have been duly referenced
- the thesis has not been submitted previously or concurrently for any other degree at any institutions
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be owned from supervisor and deputy vice –chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____

Date: _____

Name and Matric no.: Nurul Hawa Ahmad (GS 28829)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of
Chairman of
Supervisory
Committee: _____

Signature: _____
Name of
Member of
Supervisory
Committee: _____

Signature: _____
Name of
Member of
Supervisory
Committee: _____

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF SYMBOLS AND ABBREVIATIONS	xvi
 CHAPTER	
 1 GENERAL INTRODUCTION	
1.1 Background of the study	1
1.2 Objective of study	2
 2 LITERATURE REVIEW	
2.1 Food hydrocolloids	3
2.1.1 Functionality of food hydrocolloids	3
2.1.2 Sources of food hydrocolloids	4
2.1.2.1 Plant hydrocolloids	4
2.1.2.2 Seaweed hydrocolloids	4
2.1.2.3 Animal hydrocolloids	5
2.2 Microbial hydrocolloids	6
2.2.1 Gellan	8
2.2.2 Dextran	9
2.3 Mixed biopolymer food gels	10
2.3.1 Interaction of gellan with other biopolymer	11
2.4 Rheological measurement of polysaccharides	12
2.4.1 Concept of rheology and its application	12
2.4.2 Steady shear versus dynamic shear	14
2.5 Fourier Transform Infrared (FTIR)	15
2.6 Differential Scanning Calorimetry (DSC)	16
 3 RHEOLOGICAL STUDY OF GELLAN/DEXTRAN BLENDS	
3.1 Introduction	18
3.2 Materials and methods	18
3.2.1 Materials	18
3.2.1.1 Moisture content	18

3.2.1.2	Ash content	19
3.2.1.3	Crude protein content	19
3.2.1.4	Crude fat content	19
3.2.2	Preparation of gellan/dextran blends	19
3.2.3	Preparation of bovine and porcine solution	19
3.2.4	Rheological measurements	20
3.2.4.1	Steady shear measurement	20
3.2.4.2	Dynamic viscosity measurement	20
3.2.5	Statistical analysis	20
3.3	Results and discussion	21
3.3.1	Proximate analyses	21
3.3.2	Rheological properties	22
3.3.2.1	Steady shear viscosity	22
3.3.2.2	Dynamic viscosity	27
3.3.3	Applicability of Cox-Merz rule	38
3.4	Conclusion	50
4	SPECTRAL AND THERMAL STUDY OF GELLAN/DEXTRAN BLENDS	
4.1	Introduction	51
4.2	Materials and methods	51
4.2.1	Material	51
4.2.2	Sample preparation	51
4.2.2	FTIR measurement	52
4.2.3	Thermal measurement	52
4.3	Results and discussion	53
4.3.1	Spectral analysis	53
4.3.2	Thermal characteristics	61
4.4	Conclusion	63
5	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	
5.1	Summary and general conclusion	64
5.2	Recommendations	65
	REFERENCES	66
	APPENDICES	76
	BIODATA OF STUDENT	78
	LIST OF PUBLICATIONS	79

LIST OF TABLES

Table		Page
1.	Microbial Polysaccharides and Their Commercial Use	7
2.	Chemical Composition of Gellan and Dextran	21
3.	Herschel-Bulkley Model Parameters for Gellan, Dextran, G/D Blends (1:1, 1:2, 1:3), Bovine and Porcine Gelatin at 25°C	23
4.	Magnitude of Slopes (n' , n'') and Intercept (K' , K'') of G' and G'' for Gellan, Dextran, G/D Blends, Bovine and Porcine Gelatin at 25 °C.	32
5.	FTIR Spectral Characteristic for Gellan, Dextran and G/D Binary Blends	60
6.	Thermal Characteristic by DSC for G/D Blends	62

LIST OF FIGURES

Figure	Page
1. Amino Acid Composition of Gelatin	5
2. Structure of Deacylated Gellan	8
3. Structure of Dextran	10
4. Geometry Plates of Rheometer	13
5a. Apparent Viscosity-Shear Rate Plot for Gellan, Dextran, G/D Blends (1:1, 1:2, 1:3), Bovine and Porcine Gelatin at Concentration of 0.5% (w/v) at 25 °C	25
5b. Apparent Viscosity-Shear Rate Plot for Gellan, Dextran, G/D Blends (1:1, 1:2, 1:3), Bovine and Porcine gelatin at Concentration of 1.5% (w/v) at 25 °C	26
5c. Apparent Viscosity-Shear Rate Plot for Gellan, Dextran, G/D Blends (1:1, 1:2, 1:3),bovine and porcine gelatin at concentration of 3% (w/v) at 25 °C	26
6a. Mechanical Spectra of 0.5% (w/v) Gellan, Dextran, G/D Blends (1:1, 1:2, 1:3), Bovine and Porcine Gelatin at 25°C	28
6b. Mechanical Spectra of 1.5% (w/v) Gellan, Dextran, G/D Blends (1:1, 1:2, 1:3), Bovine and Porcine Gelatin at 25°C	29
6c. Mechanical Spectra of 3% (w/v) Gellan, Dextran, G/D Blends (1:1, 1:2, 1:3), Bovine and Porcine Gelatin at 25°C	30
7. Comparison of Tan Delta (at 1 Hz) of Gellan, Dextran, G/D Blends (1:1, 1:2, 1:3), Bovine and Porcine Gelatin at Different Concentrations at 25 °C	34
8a. Frequency Dependence of Complex Viscosity for Gellan, Dextran, G/D Blends (1:1 G/D, 1:2 G/D, 1:3 G/D), Bovine and Porcine Gelatin at 0.5 % (w/v) Concentration at 25°C	36
8b. Frequency Dependence of Complex Viscosity for Gellan, Dextran, G/D Blends (1:1 G/D, 1:2 G/D, 1:3 G/D), Bovine and Porcine Gelatin at 1.5 % (w/v) Concentration at 25°C	36
8c. Frequency Dependence of Complex Viscosity for Gellan, Dextran, G/D Blends (1:1 G/D, 1:2 G/D, 1:3 G/D), Bovine and Porcine Gelatin at 3 % (w/v) Concentration at 25°C	37

9a.	Applicability of Cox-Merz Rule for 0.5% (w/v) Gellan at 25 °C	39
9b.	Applicability of Cox-Merz Rule for 0.5% (w/v) Dextran at 25 °C	39
9c.	Applicability of Cox-Merz Rule for 0.5% (w/v) 1:1 G/D at 25 °C	40
9d.	Applicability of Cox-Merz Rule for 0.5% (w/v) 1:2 G/D at 25 °C	40
9e.	Applicability of Cox-Merz Rule for 0.5% (w/v) 1:3 G/D at 25 °C	41
9f.	Applicability of Cox-Merz Rule for 0.5% (w/v) Bovine at 25 °C	41
9g.	Applicability of Cox-Merz Rule for 0.5% (w/v) Porcine at 25 °C	42
10a.	Applicability of Cox-Merz Rule for 1.5% (w/v) Gellan at 25 °C	43
10b.	Applicability of Cox-Merz Rule for 1.5% (w/v) Dextran at 25 °C	43
10c.	Applicability of Cox-Merz Rule for 1.5% (w/v) 1:1 G/D at 25 °C	44
10d.	Applicability of Cox-Merz Rule for 1.5% (w/v) 1:2 G/D at 25 °C	44
10e.	Applicability of Cox-Merz Rule for 1.5% (w/v) 1:3 G/D at 25 °C	45
10f.	Applicability of Cox-Merz Rule for 1.5% (w/v) Bovine at 25 °C	45
10g.	Applicability of Cox-Merz Rule for 1.5% (w/v) Porcine at 25 °C	46
11a.	Applicability of Cox-Merz Rule for 3% (w/v) Dextran at 25 °C	47
11b.	Applicability of Cox-Merz rule for 3% (w/v) 1:1 G/D at 25 °C	47
11c.	Applicability of Cox-Merz rule for 3% (w/v) 1:2 G/D at 25 °C	48
11d.	Applicability of Cox-Merz Rule for 3% (w/v) 1:3 G/D at 25 °C	48
11e.	Applicability of Cox-Merz Rule for 3% (w/v) Bovine at 25 °C	49
11f.	Applicability of Cox-Merz Rule for 3% (w/v) Porcine at 25 °C	49
12.	FTIR (KBr) Spectra of Gellan and Dextran	54
13a.	FTIR Spectra of Gellan at Different Concentration at 25 °C	56
13b.	FTIR Spectra of Dextran at Different Concentrations at 25 °C	56
14a.	FTIR Spectra for 0.5% (w/v) Gellan, Dextran and G/D Blends at 25 °C	58

14b.	FTIR Spectra for 1.5% (w/v) Gellan, Dextran and G/D Blends at 25 °C	58
14c.	FTIR spectra for 3% (w/v) Gellan, Dextran and G/D Blends at 25 °C	59
15.	DSC thermograms showing changes in exothermic peaks for (A) gellan, (B) 1:1 GD, (C) 1:2 GD, (D) 1:3 GD and (E) dextran.	62



LIST OF ABBREVIATION AND SYMBOLS

AOAC	Association of analytical chemist
ATR	Attenuated total reflectance
C-H	Methyl group
COO ⁻	Carboxylic acid
C=O	Carboxyl group
DSC	Differential scanning calorimetry
FTIR	Fourier Transform Infrared
G'	Elastic modulus
G''	Viscous modulus
G*	Complex modulus
K	Consistency index
LVR	Linear viscoelastic region
OH	Hydroxyl group
R ²	Correlation coefficient
SE	Standard error
Tan δ	Tangent delta
T _c	Completion temperature
T _o	Onset temperature
T _m	Melting temperature
η^*	Complex viscosity
η_a	Apparent viscosity
ω	Frequency

$\dot{\gamma}$	Shear rate
τ	Shear stress
τ_0	Yield stress



CHAPTER 1

GENERAL INTRODUCTION

1.1 Background of the study

Hydrocolloids are polysaccharides that are dissolving in water and provide a wide range of purposes in food system, such as thickening, gelling, emulsifying, stabilizing, foaming and preventing crystal growth agent. Important sources of hydrocolloids primarily originated from plants (e.g. starch), seaweed (e.g. alginate), animal (e.g. gelatin) and microbes (xanthan). Gelatin is marked as unique and extraordinary hydrocolloid because of its multipurpose functions in a wide range of applications in various industries. For Muslim consumers, the sources of gelatin become their main religious concerns because the two largest raw materials used to manufacture gelatin worldwide are pigskin (42%) as well as cattle bones and hides (55%). In addition, global gelatin production in 2011 was dominated by Europe (40%) followed by North America (23%), inferring that there is still a great market niche for halal gelatin (GIA, 2013). Gelatins also exhibit critical health risk in which cattle infected with bovine spongiform encephalopathy (BSE) or mad cow disease is unsafe for human consumption (Karim and Bhat, 2008). Plant and seaweed hydrocolloids have now been commercialized to accommodate religious and health concerns of gelatin. However, there are still several limitations in comparison to microbial hydrocolloids wherein plant and seaweed require longer harvest maturity prior polysaccharide production and also susceptible to geographical and seasonal damage. In contrast, microbial hydrocolloids pose fast yielding process because it is only takes several days for microbes to grow. Furthermore, microbial hydrocolloids can be produced under fully controlled environment via fermentation process (Donot et al., 2012).

Among microbial hydrocolloids that have great commercial potential includes gellan and dextran. Gellan is secreted by *Pseudomonas elodea*, consists of linear repeating tetrasaccharides: β -D-glucose (DGlc), L-rhamnose (L-Rha), and D-glucuronic acid (D-GlcA) in molar ratio of 2:1:1. Gellan is rigid and fragile gel with higher thermal stability in the deacylated form (Evageliou et al., 2010; Chaudhary et al., 2013). Dextran is highly water soluble polysaccharides produced by *Leuconostoc mesenteroides*, strain B-512F and contain a backbone of consecutive α -1,6 linkages, with the remaining being branched α -1,3 linkages (Tirtaatmadja et al., 2001). Gellan has been combined with other polysaccharides such as agarose and kappa carrageenan to form mixed biopolymers but these combinations resulted phase separation. Phase separation is not desired in food products because it will affect consumer perception on food texture.

Viscosity and gel rigidity of microbial polysaccharide are important parameter to determine food texture. Rheological test such as steady state flow and dynamic viscoelastic have been utilized to characterize viscosity and gel rigidity respectively, by correlating the relationship between applied stress acting on a given sample and the resulting characteristic, either more solid (elastic) or more fluid (viscous) under defined condition. Beside rheological properties, favorable interaction of mixed polysaccharides needs to be investigated to avoid phase separation. Spectral analysis performed by Fourier Transform Infrared (FTIR) is useful to determine functional groups changes involve when two polysaccharides are mixed together. The presence of new band accompanied by disappearance of band in the polysaccharide blend could be an indicator of chemical properties alteration. Shifting of specific bands in the polysaccharides blends in comparison to pure component is another way of looking at changes of molecular interaction. Further, thermal analysis performed by Differential Scanning Calorimetry (DSC) can also be used to detect interaction occur upon blending. Thermal transition such as melting point and enthalpy changes of polysaccharides blend as opposed to single polysaccharide can be observed as associative interaction if it is thermodynamically favorable and segregative if they are less favorable.

1.2 Objective of the study

In this study, gellan/dextran blends at different concentration and ratio were prepared as this polysaccharide combination has not previously studied based on literature. Therefore, the specific objectives of this study are:

1. To characterize rheological properties including flow characteristics and gel rigidity of gellan, dextran and gellan/dextran blends in different mixing ratios as compared to porcine and bovine gelatin.
2. To investigate the molecular interaction and phase transition using spectral and thermal analysis of gellan, dextran and their blends.

REFERENCES

- Aguilera, J. M., Rademacher, B. (2004). Protein gels. In R. Y. Yada, Proteins in Food Processing. (pp. 468–482). New York: Woodhead Publishing Limited and CRC Press LLC.
- Ahmed, R. Z. Siddiqui, K., Arman, M., Ahmed, N. (2012) Characterization of high molecular weight dextran produced by *Weissella cibaria* CMGDEX3. *Carbohydrate Polymers*, 90, 441-446.
- Ahmed J., Ramaswamy H. S., Ngadi, O. (2005). Rheological characteristics of Arabic gum in combination with guar and xanthan gum using response surface methodology: effect of temperature and concentration. *International Journal of Food Properties*, 8, 179–192.
- Ahmed, J., Ramaswamy, H. S. (2006). Viscoelastic properties of sweet potato puree infant food. *Journal of Food Engineering*, 74, 376–382.
- AOAC. (2005). Official methods of analysis. Arlington: Association of Official Analytical Chemists.
- Ashtaputre, A. A., Shah, A. K. (1995). Studies on exopolysaccharide from *Sphingomonas paucimobilis* GSI: Nutritional requirement and precursors forming enzymes. *Current Microbiology*, 31, 234–238.
- Banik, R. M., Kanari, B., Upadhyay, S. N. (2000). Exopolysaccharides of gellan family: prospects and potential. *World Journal of Microbiology and Biotechnology*, 16, 407–414.
- Banik R. M., Santhiagu, A. (2006). Improvement in production and quality of gellan gum by *Sphingomonas paucimobilis* under high dissolved oxygen tension levels. *Biotechnology Letters*, 28, 1347–1350.
- Bekers, M., Urite, D., Kaminska, E., Laukevics, J., Grube, M., Vigants, A., et al. (2005). Stability of levan produced by *Zymomonas mobilis*. *Process Biochemistry*, 40, 1535–1539.
- Bhandari, P.N., Singhl, R.S., Kale, D. D. (2002). Effect of succinylation on the rheological profile of starch pastes. *Carbohydrate Polymers*, 47, 365–371.
- Bhavani, A., Nisha, J. (2010). Dextran-The polysaccharide with versatile uses. *International Journal of Pharma and Bioscience*, 1, 569–573.
- Biliaderis, C. G., Juliano, B. O. (1993). Thermal and mechanical properties of concentrated rice starch gels of varying composition. *Food Chemistry*, 48, 243–250.

- Boral, S., Saxena, A., Bohidar, H. B. (2008). Universal growth of microdomains and gelation transition in Agar hydrogels. *Journal of Physical Chemistry*, 112, 3625–3632.
- Burey, P., Bhandari, B. R., Howes, T., Gidley, M. J. (2008). Hydrocolloid Gel Particles: Formation, Characterization, and Application. *Critical Reviews in Food Science and Nutrition*, 48, 361–377.
- Caggioni, M., Spicer, P. T., Blair, D. L., Lindberg, S. E., Weitz, D. A. (2007). Rheology and microrheology of a microstructured fluid: The gellan gum case. *Journal of Rheology*, 51, 851–865.
- Carlfors, J., Edsman, K., Peterson, R., Jorning, K. (1998). Rheological evaluation of Gelrite in situ gels for ophthalmic use. *European Journal Pharmaceutical*, 6, 113–119.
- Chaudhary, V., Small, D. M., Kasapis, S. (2013). Effect of a glassy gellan / polydextrose matrix on the activity of. *Carbohydrate Polymers*, 95, 389–396.
- Choi, S. S., Regenstien, J. M. (2000). Physicochemical and sensory characteristic of fish gelatin. *Journal of Food Science*, 65, 194–199.
- Coviello, T., Matricardi, P., Marianecchi, C., Alhaique, F. (2007). Polysaccharide hydrogels for modified release formulations. *Journal of Controlled Release : Official Journal of the Controlled Release Society*, 119, 5–24.
- Cox, W. P., Merz, E. H. (1958). Correlation of dynamic and steady state flow viscosities. *Journal Polymer Science*, 28, 619–622.
- Cui, S. W. (2005). Food carbohydrates: Chemistry, Physical properties and application. USA Davidson: CRC/ Taylor and Francis.
- Dai, L., Liu, X., Liu, Y., Tong, Z. (2008). Concentration dependence of critical exponents for gelation in gellan gum aqueous solutions upon cooling. *European Polymer Journal*, 44, 4012–4019.
- Dentini, M., Coviello, T., Burchard, W., Crescenzi, V. (1988). Solution properties of exocellular microbial polysaccharides. 3. Light scattering from gellan and from the exocellular polysaccharide of *Rhizobium trifolii* (strain TA-1) in the ordered state. *Macromolecules*, 21, 3312–3320.
- Dickinson, E. (2003). Hydrocolloids at interfaces and the influence on properties of dispersed systems. *Food Hydrocolloids*, 17, 25–39.
- Dickinson, E., Lopez, G. (2001). Comparison of the emulsifying properties of fish gelatin and commercial milk proteins. *Journal of Food Science*, 66, 118–123.

- Donot, F., Fontana, A., Baccou, J. C., Schorr-Galindo, S. (2012). Microbial exopolysaccharides: Main examples of synthesis, excretion, genetics and extraction. *Carbohydrate Polymers*, 87, 951–962.
- Drevetton, E., Daniel, B., Lonen, C. (1994). Effect of mixing and mass transfer conditions on gellan production by *Auromonas elodea*. *Journal of Fermentation and Bioengineering*, 77, 642–649.
- Evageliou, V., Tseliou, G., Mandala, I., Komaitis, M. (2010). Effect of inulin on texture and clarity of gellan gels. *Journal of Food Engineering*, 101, 381–385.
- Evans, J. D., Haisman, D. R. (1979). Rheology of getinized starch suspensions. *Journal of Texture Studies*, 10, 347–370.
- Falconer, D. J., Mukerjea, R., Robyt, J. F. (2011). Biosynthesis of dextrans with different molecular weights by selecting the concentration of *Leuconostoc mesenteroides* B-512FMC dextransucrase, the sucrose concentration and the temperature. *Carbohydrate Polymers*, 364, 280–284.
- Fathi, E., Atyabi, N., Imani, M., Alinejad, Z. (2011). Physically crosslinked polyvinyl alcohol–dextran blend xerogels: Morphology and thermal behavior. *Carbohydrate Polymers*, 84, 145–152.
- Ferry, A. –L., Hort, J., Mitchell, J. R., Cook, D. J., Lagarrigue, S., Valles Pamies, B. (2006) Viscosity and flavour perception: Why is starch different from hydrocolloids? *Food Hydrocolloids*, 20, 855–862
- Fischer, P., Windhab, E. J. (2011). Rheology of food materials. *Current Opinion in Colloid & Interface Science*, 16, 36–40.
- Fonkwe, L. G., Narsimhan, G., Cha, A. S. (2003). Characterization of gelation time and texture of gelatin and gelatin–polysaccharide mixed gels. *Food Hydrocolloids*, 17, 871–883.
- Funami, T. (2011). Next target for food hydrocolloid studies: Texture design of foods using hydrocolloid technology. *Food Hydrocolloids*, 25, 1904–1914.
- Funami, T., Hiroe, M., Noda, S., Asai, I., Ikeda, S., Nishinari, K. (2007). Influence of molecular structure imaged with atomic force microscopy on the rheological behavior of carrageenan aqueous systems in the presence or absence of cations. *Food Hydrocolloids*, 21, 617–629.
- GIA (2013) Gelatin market by raw materials (pigskin, bovine hides, bones and others) for food & beverage, nutraceuticals, pharmaceuticals, photograph, cosmetics and other applications-Global Industry Analyst, size, share, growth, trends and forecast 2012-2018. <http://www.transparencymarketresearch.com/gelatin.html>. Retrieved 22 November 2014

- Gil, E. C., Colarte A. I., Ghzaoui, A. E., Durand, D., Delarbe, J. L., Bataille, B. (2008). A sugarcane native dextran as an innovative functional excipient for the development of pharmaceutical tablets. *European Journal of Pharmaceutical and Biopharmaceutical*, 68, 319–329.
- Gilsenan, P. M., Ross-Murphy, S. B. (2000). Rheological characterizations of gelatins from mammalian and marine sources. *Food Hydrocolloids*, 14, 191–195.
- Glickman, M. (1982). Origins and classification of hydrocolloids. In *Food Hydrocolloids* (3rd ed., pp. 3–18). Boca Raton, Florida: CRC Press.
- Gloria Hernández, H., Livings, S., Aguilera, J. M., Chiralt, A. (2011). Phase transitions of dairy proteins, dextrans and their mixtures as a function of water interactions. *Food Hydrocolloids*, 25, 1311–1318.
- Gómez-Guillén, M. C., Giménez, B., López-Caballero, M. E., Montero, M. P. (2011). Functional and bioactive properties of collagen and gelatin from alternative sources: A review. *Food Hydrocolloids*, 25, 1813–1827.
- Gómez-Ordóñez, E., Rupérez, P. (2011). FTIR-ATR spectroscopy as a tool for polysaccharide identification. *Food Hydrocolloids*, 25, 1514–1520.
- Griffiths, P. R., Haseth, J. A. (2007). Fourier-transform infrared spectrometry (pp. 171–175). Wiley Interscience.
- Gunasekaran, S., Ak, M. M. (2000). Dynamic oscillatory shear testing of foods in selected applications. *Trends in Food Science and Technology*, 11, 115–127.
- Hafidz, R.M., Yaakob, C. M., Amin, I. and Noorfaizan, A. (2011). Chemical and functional properties of bovine and porcine skin gelatin. *International Food Research Journal*, 18, 813–817.
- Hansen, P. M. T. (1993). Food hydrocolloids in the dairy industry. In E. Nishinari, K., Doi, Food hydrocolloids: structure, properties and function (pp. 211–224). New York: Plenum press.
- Hennink, W. E., De Jong, S. J., Bos, G. W., Veldhuis T. F. J., van Nostrum, C. F. (2004). Biodegradable dextran hydrogel crosslinked by stereocomplex formation for the controlled release of pharmaceutical proteins. *International Journal of Pharmaceutical*, 277, 99–104.
- Henshaw, F. O., McWatters, K. H., Akingbala, J. O., Chinnan, M. S. (2003). Thermal properties of cowpea flour: a study by differential scanning calorimetry. *Die Nahrung*, 47, 161–5.

- Hidaka, S., Liu, S. Y. (2002). Effects of gelatins on calcium phosphate precipitation: a possible application for distinguishing bovine bone gelatin from porcine skin gelatin. *Journal of Food Composition and Analysis*, 16, 477–483.
- Icoz, D. Z., Kokini, J. L. (2007). Probing the boundaries of miscibility in model carbohydrates consisting of chemically derivatized dextrans using DSC and FTIR spectroscopy. *Carbohydrate Polymers*, 68, 68–76.
- Icoz, D. Z., Moraru, C. I., Kokini, J. L. (2005). Polymer–polymer interactions in dextran systems using thermal analysis. *Carbohydrate Polymers*, 62, 120–129.
- Ikeda, S., Nitta, Y., Sook, B., Temsiripong, T. (2004). Single-phase mixed gels of xyloglucan and gellan. *Food Hydrocolloids*, 18, 669–675.
- Imenson, A. (2010). Food stabilisers, thickeners and gelling agents. Oxford, England: Blackwell Publishing Ltd.
- Kacurakova, M., Wilson, R. H. (2001). Developments in mid-infrared FT-IR spectroscopy of selected carbohydrates. *Carbohydrate Polymers*, 44, 291–303.
- Kang, K. S., Veeder, G. T., Colegrove, G. T. (1983). Deacetylated polysaccharide S-60. US Patent.
- Kang, K. S., Veeder, G. T., Mirrasoul, P. J., Kaneko, T., Cottrell, I. W. (1982). Agar-like polysaccharides produced by a *Pseudomonas* species: production and basic properties. *Applied Environmental Microbiology*, 43, 1086–1091.
- Karim, A. A., Bhat, R. (2008). Gelatin alternatives for the food industry: recent developments, challenges and prospects. *Trends in Food Science and Technology*, 19, 644–656.
- Kiani, H., Mousavi, M. E., Razavi, H., Morris, E. R. (2010). Effect of gellan, alone and in combination with high-methoxy pectin, on the structure and stability of doogh, a yogurt-based Iranian drink. *Food Hydrocolloids*, 24, 744–754.
- Kim, M. K., Lee, I. Y. (1999). Higher intercellular levels of uridine monophosphate under nitrogen limited conditions enhance the metabolic flux of curdlan synthesis in *Agrobacterium* species. *Biotechnology and Bioengineering*, 62, 317–323.
- Koocheki, A., Mortazavi, S. A., Shahidi, F., Razavi, S. M. A., Taherian, A. R. (2009). Rheological properties of mucilage extracted from *Alyssum homolocarpum* seed as a new source of thickening agent. *Journal of Food Engineering*, 91, 490–496.
- Koocheki, A., Taherian, A. R., Bostan, A. (2011). Studies on the steady shear flow behavior and functional properties of *Lepidium*. *Food Research International*, 50, 446–456.

- Kumar, A. S., Mody, K., Jha, B. (2007). Bacterial exopolysaccharides—a perception. *Journal of Basic Microbiology*, 47, 103–117.
- Kwon, B. D., Foss, P. A., Rha, C. (1987). Rheological characteristics of high viscosity polysaccharides. In M. Yalpani, Industrial polysaccharides: Genetic engineering structure–property relations and applications. (pp. 253–266). Amsterdam: Elsevier Science Publisher.
- Leathers, T. D. (2003). Biotechnological production and applications of pullulan. *Applied Microbiology and Biotechnology*, 62, 468–473.
- Leefflang, B. R., Faber, E. J., Erbel P., Vliegenthart, J.F.G. (2000). Structure elucidation of glycoprotein glycans and of polysaccharides by NMR spectroscopy. *Journal of Biotechnology*, 77, 115–122.
- León, P. G., Lamanna, M. E., Gerschenson, L. N., Rojas, A. M. (2008). Influence of composition of edible films based on gellan polymers on l-(+)-ascorbic acid stability. *Food Research International*, 41, 667–675.
- Liu, C., Lin, Q., Gao, Y., Ye, L., Xing, Y., Xi, T. (2007). Characterization and antitumor activity of polysaccharide from *Strongylocentrotus nudus* eggs. *Carbohydrate Polymers*, 67, 1048–1054.
- Majumder, A., Goyal, A. (2009). Rheological and gelling properties of a novel glucan from *Leuconostoc dextranicum* NRRL B-1146. *Food Research International*, 12, 525–528.
- Marcotte, M., Taherian Hoshahili, A.R., Ramaswamy, H. S. (2001). Rheological properties of selected hydrocolloids as a function of concentration and temperature. *Food Research International*, 34, 695–703.
- Mathur, V., Mathur, N. K. (2006). Microbial polysaccharides based food hydrocolloid additives. *Science Tech Entrepreneur*, 1–10.
- McCurdy, R. D., Goff, H. D., Stanley, D. W. (1994). Properties of dextran as a cryoprotectant in ice cream. *Food Hydrocolloids*, 8, 625–633.
- McKenna, B. M., Lyng, J. G. (2003). Introduction to food rheology and its measurement. In McKenna, B. M., Texture in food: Vol 1: Semi solid foods (pp. 130–160). Washington: CRC Press.
- Meena, R., Prasad, K., Siddhanta, a. K. (2009). Development of a stable hydrogel network based on agar–kappa-carrageenan blend cross-linked with genipin. *Food Hydrocolloids*, 23, 497–509.
- Milani, J., Maleki, G. (2012). Hydrocolloids in Food Industry. In B. Valdez, Food Industrial Process- Methods and Equipment (pp. 17–38). Rijeka, Croatia: InTech.

- Miyoshi, E., Takaya, T., Nishinari, K. (1995). Gel-sol transition in gellan gum aqueous solutions. *Makromolecules Symposium*, 99, 83–91.
- Miyoshi, E., Takaya, T., Nishinari, K. (1996). Gel-sol transition on gellan gum aqueous solutions. I. Rheological studies on the effects of salts. *Carbohydrate Polymers*, 36, 109–120.
- Morris, E. R. (1990). Shear thinning of “random coil” polysaccharides: characterization by two parameters from a simple linear plot. *Carbohydrate Polymers*, 13, 85–86.
- Morris, E. R., Nishinari, K., Rinaudo, M. (2012). Gelation of gellan – A review. *Food Hydrocolloids*, 28, 373–411.
- Morris, V. J. (2006). Bacterial polysaccharides. In P. A. A. M. Stephen, G. O. Phillips and Williams (Eds.), *Food polysaccharides and their applications* (2nd ed., pp. 413–454). Boca Raton, FL, USA: CRC Press, Taylor and Francis Group.
- Morris, V. J. (1986). Multicomponent gel. In P. A. Phillips, G.O., Wedlock, D.J. and Williams, Ed *Gums and Stabilisers for the Food Industry*.) (pp. 87–99). Elsevier Applied Science Publishers.
- Naessens, M., Cerdobbel, A., Soetaert, W., Vandamme, E.J. (2005). Leuconostoc dextranase and dextran: production, properties and applications. *Journal of Chemical Technology Biotechnology*, 80, 845–860.
- Nampoothiri, K. M., Singhanian, R. R. (2003). Fermentative production of gellan using *Sphingomonas paucimobilis*. *Process Biochemistry*, 38, 1513–1519.
- Nishinari, K., Watase, M., Rinaudo, M., Milas, M. (1996). Characterization and properties of gellan- κ -carrageenan mixed gels. *Food Hydrocolloids*, 10, 277–283.
- Nishinari, K., Takaya, T., Watase, M. (1994). Rheology and DSC of gellan-agarose. In E. Nishinari, K., Doi, *Food hydrocolloids: structure, properties and function* (pp. 473–476). New York: Plenum press.
- Nitta, K., Shin, Y., Hashiguchi, H., Tanimoto, S., Terano, M. (2005). Morphology and mechanical properties in the binary blends of isotactic polypropylene and novel propylene-co-olefin random copolymers with isotactic propylene sequence 1. Ethylene-propylene copolymer. *Polymer*, 46, 965–975.
- Noda, S., Funami, T., Nakauma, M., Asai, I., Takahashi, R., Al-Assaf, S., et al. (2008). Molecular structures of gellan gum imaged with atomic force microscopy in relation to the rheological behaviour in aqueous systems. 1. Gellan gum with various acyl contents in the presence and absence of potassium. *Food Hydrocolloids*, 22, 1148–1159.

- Noor, I. S. M., Majid, S. R., Arof, a. K., Djurado, D., Claro Neto, S., Pawlicka, A. (2012). Characteristics of gellan gum–LiCF₃SO₃ polymer electrolytes. *Solid State Ionics*, 225 649–653.
- Papageorgiou, M., Gothard, M. G., Willoughby, L. E., Kasapis, S., Richardson, R. K., Morris, E. R. (1994). Rheology and structure of gellan-alginate co-gels. In P. A. Phillips, G.O., Wedlock, D.J. and Williams. *Gums and Stabilisers for the Food Industry-7* (pp. 345–356). Oxford: Oxford Univ. Press.
- Papageorgiou, M., Kasapsi, S., Richardson, R. K. (1994). Steric exclusion phenomena in gellan/gelatin system I. Physical properties of single and binary gels. *Food Hydrocolloids*, 8, 97–112.
- Patel, R. M., Patel, V. P. (2011). Microbial polysaccharides: Current innovations and future trends in medical science. *Current Pharmaceutical Research*, 1, 204–209.
- PB Gelatins (2014) Amino acid composition of gelatin. <http://www.pbgelatins.com/about-gelatin/physical-and-chemical-properties/amino-acidcomposition/> Retrieved on 19 November 2014
- Pérez-Campos, S. J., Chavarría-Hernández, N., Tecante, A., Ramírez-Gilly, M., Rodríguez-Hernández, A. I. (2012). Gelation and microstructure of dilute gellan solutions with calcium ions. *Food Hydrocolloids*, 28, 291–300.
- Prajapati, V. D., Jani, G. K., Zala, B. S. et al. (2013). An insight into the emerging exopolysaccharide gellan gum as a novel polymer. *Carbohydrate Polymers*, 93, 670–678.
- Pranoto, Y., Lee, C. M., Park, H. J. (2007). Characterizations of fish gelatin films added with gellan and κ-carrageenan. *LWT - Food Science and Technology*, 40, 766–774.
- Purama, R. K., Goswami, P., Khan, A. T., Goyal, A. (2009). Structural analysis and properties of dextran produced by *Leuconostoc mesenteroides* NRRL B-640. *Carbohydrate Polymers*, 76, 30–35.
- Rao, M. (1999). Rheology of fluids and semisolid foods. Maryland: Aspen Publishers, Inc.
- Richardson, R. K., Ross-Murphy, S. B. (1987). Nonlinear viscoelasticity polysaccharide solutions. 2: Xanthan polysaccharide solutions. *International Journal of Biology Macromolecules*, 9, 257–264.
- Roller, S., Dea, I. C. M. (1992). Biotechnology in the production and modification of biopolymers for foods. *Critical Review in Biotechnonology*, 12, 261–277.

- Ross-Murphy, S. B. (1984). Rheological methods. London, UK: SCI. In H. W.-S. Chan, Biophysical methods in food research. Critical reports on applied chemistry (pp. 195–290). London, UK: SCI.
- Sahin, H., Ozdemir, F. (2004). Effect of some hydrocolloids on the rheological properties of different formulated ketchups. *Food Hydrocolloids*, 18, 1015–1022.
- Sahin, S., Sumnu, S. G. (2006). Physical properties of foods. New York, USA: Springer
- Sanderson, G.R. (1982). The interaction of xanthan gum in food system. *Progress in Food Nutrition Science*, 6, 77–87.
- Santacruz, S., Ruales, J., Eliasson, A. C. (2003). Three under-utilized sources of starch from the Andean region in Ecuador. Part III. Rheological characterization. *Carbohydrate Polymers*, 51, 85–92.
- Schrieber, R., Gareis, H. (2007). Gelatine handbook theory and industrial practice. Weinheim: Wiley-VCH Verlag GmbH & Co.KGaA
- Schurks, N., Wingender, J., Flemming, H. C., Mayer, C. (2002). Monomer composition and sequence of alginates from *Pseudomonas aeruginosa*. *International Journal Biology and Macromolecules*, 30, 105–111.
- Shigel, K. I. (2002). Determination of structural peculiarities of dextran, pullulan and gamma irradiated pullulan by Fourier-transform IR spectroscopy. *Carbohydrate Research*, 337, 2649–2701.
- Shoraku, A., Takigawa, T., Masuda, T. (2002). Effects of alkaline metal salts on viscosity of gellan aqueous solutions. *Journal of Society of Rheology*, 30, 13–17.
- Simi, C. K., Abraham, T. E. (2010). Transparent xyloglucan–chitosan complex hydrogels for different applications. *Food Hydrocolloids*, 24, 72–80.
- Sudhamani, S. R., Prasad, M. S., Udaya Sankar, K. (2003). DSC and FTIR studies on Gellan and Polyvinyl alcohol (PVA) blend films. *Food Hydrocolloids*, 17, 245–250.
- Tabilo-Munizaga, G., Barbosa-Cánovas, G. V. (2005). Rheology for the food industry. *Journal of Food Engineering*, 67, 147–156.
- Tester, R. F., Karkalas, J. (2002). Starch. In S. D. B. A. Steinbüchel, E. J. Vandamme, Biopolymers Vol. 6, Polysaccharides II: Polysaccharides from Eukaryotes (pp. 381–438). Weinheim: Wiley-VCH.
- Tirtaatmadja, V., Dunstan, D. E., Boger, D. (2001). Rheology of dextran solutions. *Journal of Non-Newton Fluid Mechanic*, 97, 295–301.

- Varum, K. M., Smidsrod, O. (2006). Chitosans. In P. A. W. A.M. Stephen, G.O. Phillips, Food polysaccharides and their applications (pp. 497–520). CRC Press, Taylor Francis Group.
- Wanchoo, R. K., Sharma, P. K. (2003). Viscometric study on the compatibility of some water-soluble polymerepolymer mixtures. *European Polymer Journal*, 39, 1481–1490.
- Wang, Q., Cui, S. (2005). Understanding the physical properties of food polysaccharides. In S.W.Cui, Food carbohydrates: chemistry, physical properties, and applications (pp. 162–214). Boca Raton, Florida: Taylor and Francis.
- Wasswa, J., Tang, J., Gu, X. (2007). Utilization of fish processing by- products in the gelatin industry. *Food Reviews International*, 23, 159–174.
- Williams, P. A., Phillips, G. O. (2000). Introduction to food hydrocolloids. In P. A. Phillips, G.O., Williams, Handbook of hydrocolloids (pp. 1–19). Cambridge: Woodhead Publishing Ltd.
- Wu, X., Li, O., Chen, Y., Zhu, L., Qian, C., Teng, Y., Tao, X. (2011). A carotenoid-free mutant strain of *Sphingomonas paucimobilis* ATCC 31461 for the commercial production of gellan. *Carbohydrate Polymers*, 84, 1201–1207.
- Xu, X., Li, B., Kennedy, J. F., Xie, B. J., Huang, M. (2007). Characterization of konjac glucomannan–gellan gum blend films and their suitability for release of nisin incorporated therein. *Carbohydrate Polymers*, 70, 192–197.
- Yang, L., Zhang, L. M. (2009). Chemical structural and chain conformation characterization of some bioactive polysaccharides isolated from natural sources. *Carbohydrate Polymers*, 76, 349–361.
- Zhu, G., Sheng, L., Tong, Q. (2013). Preparation and characterization of carboxymethyl gellan and pullulan blend films. *Food Hydrocolloids*, 35, 1–7.