



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF POLYSACCHARIDE-BASED
COATING CONJUGATED WITH SILVER
NANOPARTICLES ON SHELF-LIFE EXTENSION
OF PAPAYA (*Carica papaya* L. cv. *Eksotika*)***

HANISAH BINTI MUSTAFFA @ HAMZAH

FSTM 2014 31



**EFFECTS OF POLYSACCHARIDE-BASED
COATING CONJUGATED WITH SILVER
NANOPARTICLES ON SHELF-LIFE EXTENSION
OF PAPAYA (*Carica papaya* L. cv. Eksotika)**

HANISAH BINTI MUSTAFFA @ HAMZAH

**MASTER OF SCIENCE
UNIVERSITI PUTRA MALAYSIA**

2014



**EFFECTS OF POLYSACCHARIDE-BASED COATING CONJUGATED
WITH SILVER NANOPARTICLES ON SHELF-LIFE EXTENSION OF
PAPAYA (*Carica papaya* L. cv. Eksotika)**

By

HANISAH BINTI MUSTAFFA @ HAMZAH

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Master of Science**

September 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

EFFECTS OF POLYSACCHARIDE-BASED COATING CONJUGATED WITH SILVER NANOPARTICLES ON SHELF-LIFE EXTENSION OF PAPAYA (*Carica papaya* L. cv. Eksotika)

By

HANISAH BINTI MUSTAFFA @ HAMZAH

September 2014

Chair : Azizah Osman, PhD
Faculty : Food Science and Technology

Papaya is a tropical fruit that is well accepted worldwide for its good taste and nutritional content. However, like many other tropical fruits, papaya also has postharvest problems due to several reasons including short shelf life caused by disease mainly fungal infection. This problem also affects *Carica papaya* L. cv. Eksotika, which is one of the popular papaya cultivars in Malaysia. In the present study, polysaccharide-based coatings incorporated with silver nanoparticles were used to prolong the shelf life of Eksotika papaya.

In this study, a two-factor central composite design (CCD) was used to optimize two different edible coating formulations for coating of Eksotika papaya. The effect of two different coating formulations, namely, chitosan-based [chitosan (0.5 – 2.5%) and glycerol (0 – 2%)] and carrageenan-based [carrageenan (0.2 – 0.8%) and glycerol (0 – 1%)] on texture and colour components (L, a and b values) of coated papaya were evaluated. From the multiple response optimization analysis, combination of 2.32% chitosan with 0.53% glycerol, and 0.78% carrageenan with 0.85% glycerol for chitosan-based and carrageenan-based coating respectively were predicted to give desirable effect to the response variables tested, with no significant ($p > 0.05$) difference between the experimental and predicted values. However, during shelf life monitoring of the coated samples, papaya coated with the optimized chitosan-based formulation failed to ripen. In order to overcome this, visual observation of papaya coated with different concentrations of chitosan (0.5, 1.0, 1.5 and 2.32%) were conducted. 1% chitosan was found to successfully coat papaya without retarding the ripening process.

Silver nanoparticles at the range of 0-80 ppm were incorporated into the chitosan- and carrageenan-based coatings to evaluate the antimicrobial properties of the formed polysaccharide-silver nanocomposites. Chitosan alone without synergistic interaction with silver nanoparticles was sufficient in giving bactericidal and antifungal effects. Carrageenan on the other hand requires addition of 2.5 and 30 ppm of silver nanoparticles to give antifungal and bactericidal effects respectively. Application of chitosan, carrageenan-silver nanocomposite and carrageenan-based coating on Eksotika papaya were able to prolong the shelf life up to 13, 11 and 9

days at ambient condition ($26\pm 2^{\circ}\text{C}$; $70\pm 10\%$ RH), respectively. During low temperature ($15\pm 1^{\circ}\text{C}$; $60\pm 10\%$ RH) storage, the shelf life was extended up to 21 days for chitosan and carrageenan-silver nanocomposite based coating and also 17 days for carrageenan-based coating. The maximum shelf life of uncoated papaya was 7 and 13 days at ambient and low temperature, respectively. Shelf life extension of coated papaya was judged based on no visible fungal decay.

Physiological and physico-chemical evaluations during storage showed that there was no significant ($p > 0.05$) difference between coated samples and control except for firmness at ambient condition and also total soluble solids. The microbial counts between coated and uncoated papaya were also found to be insignificant ($p > 0.05$).

Determination of silver concentration in papaya fruit coated with carrageenan-silver nanocomposite revealed that, silver migrated through the peel and into the innermost pulp layer of papaya during storage. Consumer acceptance test showed that scores of all tested attributes for coated samples were above the minimum acceptable value (>5). This indicated that the coating did not give negative visual or physical perception towards the consumer. Hence, coating gave positive contribution on shelf life extension of papaya especially for marketing purposes, but incorporation of silver nanoparticles needs to be reconsidered since silver migrated into the pulp due to the thin peel layer of papaya fruit.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan Ijazah Master Sains

**KESAN SALUTAN BERASASKAN POLISAKARIDA BERKONJUGASI
DENGAN ZARAH PERAK NANO TERHADAP PERLANJUTAN JANGKA
HAYAT BUAH BETIK (*Carica papaya* L. cv. Eksotika)**

Oleh

HANISAH BINTI MUSTAFFA @ HAMZAH

September 2014

Pengerusi : Azizah Osman, PhD
Fakulti : Sains dan Teknologi Makanan

Betik ialah sejenis buah tropika yang diterima baik di seluruh dunia disebabkan oleh kandungan nutrisi dan rasanya yang sedap. Walau bagaimanapun, seperti buah-buahan tropika yang lain, betik juga mempunyai masalah lepas tuai yang berpunca daripada beberapa sebab termasuk jangka hayat penyimpanan yang pendek disebabkan oleh penyakit yang berpunca daripada jangkitan kulat. Masalah ini juga memberi kesan kepada *Carica papaya* L. cv. Eksotika yang merupakan salah satu daripada kultivar betik yang popular di Malaysia. Dalam kajian ini, salutan berasaskan polisakarida yang dikongjugasikan dengan zarah nano perak telah digunakan untuk memanjangkan jangka hayat simpanan betik Eksotika.

Dalam kajian ini, reka bentuk komposit pusat dua faktorial telah digunakan untuk mendapatkan formulasi optimum bagi dua salutan boleh makan yang berbeza untuk menyalut betik Eksotika. Kesan dua formulasi berbeza, iaitu yang berasaskan kitosan [kitosan (0.5-2.5%) dan glicerol (0-2%)] dan juga yang berasaskan karagenan [karagenan (0.2-0.8%) dan glicerol (0-1%)] terhadap tekstur dan komponen warna (nilai L, a dan b) buah betik yang disalut telah diuji. Daripada analisis respon optimum pelbagai, kombinasi 2.32% kitosan dengan 0.53% glicerol, dan kombinasi 0.78% karagenan dengan 0.85% glicerol, masing-masing bagi salutan berasaskan kitosan dan karagenan diramalkan member kesan yang diinginkan terhadap pembolehubah bergerak balas yang dikaji, dengan tiada perbezaan yang signifikan ($p > 0.05$) antara nilai eksperimen dengan nilai yang diramal. Walau bagaimanapun, semasa tempoh pemerhatian jangka hayat simpanan betik yang telah disalut, didapati bahawa betik yang disalut dengan formulasi kitosan optimum gagal meranum. Bagi mengatasi masalah ini, pemerhatian secara visual bagi betik yang disalut dengan kepekatan kitosan yang berbeza (0.5, 1.0, 1.5 dan 2.32%) telah dijalankan. Didapati kitosan pada kepekatan 1% telah berjaya menyalut betik tanpa membantutkan proses peranakan.

Zarah perak nano pada julat 0-80 ppm telah dimasukkan ke dalam salutan berasaskan kitosan dan karagenan bagi menguji sifat antimikrobial nanokomposit polisakarida dan perak yang terbentuk. Kitosan semata-mata tanpa interaksi sinergi dengan zarah perak nano adalah mencukupi untuk member kesan anti bakteria dan

anti kulat. Walau bagaimanapun, karagenan memerlukan penambahan 2.5 dan 30 ppm zarah perak nano masing-masing bagi member kesan anti kulat dan bakteriasid.

Aplikasi salutan berasaskan kitosan, nanokomposit karagenan dengan perak dan karagenan pada betik Eksotika masing-masing mampu memanjangkan jangka hayat penyimpanan buah sehingga 13, 11 dan 9 hari pada keadaan ambien ($26\pm 2^{\circ}\text{C}$; $70\pm 10\%$ RH). Semasa penyimpanan pada suhu rendah ($15\pm 1^{\circ}\text{C}$; $60\pm 10\%$ RH), jangka hayat simpanan dapat dipanjangkan sehingga 21 hari bagi salutan berasaskan kitosan dan nanokomposit karagenan dengan perak. Manakala bagi betik disalut karagenan, jangka hayat dapat dipanjangkan sehingga 17 hari. Jangka hayat simpanan maksimum bagi betik yang tidak disalut ialah 7 dan 13 hari, masing-masing bagi penyimpanan pada keadaan ambien dan suhu rendah. Peningkatan jangka hayat simpanan betik yang disalut adalah berdasarkan tiadanya kesan kerosakan akibat kulat.

Penilaian fisiologi dan fiziko-kimia semasa tempoh penyimpanan menunjukkan tiada perbezaan signifikan ($p>0.05$) antara sampel bersalut dan tidak bersalut kecuali untuk tekstur dan kandungan kesuluruhan pepejal larut. Pertumbuhan bakteria, yis dan kulat antara betik bersalut dan tidak bersalut juga didapati tidak signifikan ($p>0.05$).

Penentuan kepekatan zarah perak di dalam betik bersalut nanokomposit karagenan dan perak semasa tempoh penyimpanann menunjukkan bahawa zarah perak berjaya menembusi kulit betik dan terus ke lapisan isi betik yang paling dalam. Ujian penerimaan pengguna menunjukkan bahawa skor bagi setiap ciri betik bersalut yang dinilai adalah lebih tinggi daripada nilai minimum yang diterima (>5). Ini menunjukkan bahawa dari segi visual dan fizikal salutan yang digunakan tidak memberi persepsi negatif kepada pengguna. Justeru, penggunaan salutan dapat memberi kesan positif kepada peningkatan jangka hayat simpanan betik terutamanya bagi tujuan pemasaran. Walau bagaimanapun, penambahan zarah perak nano perlu dipertimbangkan, memandangkan terdapat penghijrahan zarah perak ke dalam isi buah betik disebabkan oleh kulitnya yang nipis.

ACKNOWLEDGEMENTS

In the name of Allah, the Most Beneficent, the Most Merciful. First and foremost, all praise to Allah for providing me the opportunity and guidance, with which had given me the strength to complete this research.

Greatest appreciation goes to my supervisor Prof. Dr. Azizah Osman for her guidance and moral support throughout the research. Her countless contributions had made this research possible. I would also like to extend my deepest gratitude to my supervisory committee members, Prof. Dr. Tan Chin Ping and Assoc. Prof. Dr. Farinazleen Mohamad Ghazali for their valuable advice, constant support and financial aid.

Very special thanks to Prof. Dr. Hasanah Mohd. Ghazali and Assoc. Prof. Dr Yaya Rukayadi for their valuable suggestions. I am also thankful to the supporting staffs of the Department of Food Science (Mrs. Norlinawati Abd. Halim, Mrs. Aziatum Omar, Mr. Zulkefli Nordin, Mrs Rosmawati Othman, Ms. Nurul Asikin Mohd Yusof, Mrs Norhafiza Abd. Razak, Ms. Nurfatihah Noor Musa, Mrs. Noratina Mohd Darus, Mr. Azman Asmat, Mrs Kamariah Jaafar, Mr. Zolkifli Johari and Mr. Azman Abu Yamin), Department of Food Technology (Mr. Mahamed Kidin, Mr. Amran Suratman, Mrs. Asmawati Mantali, Mrs. Jamaliah Ahmad, Mrs Faridah Muda and Mr. Mohamad Soib Yusof) and staffs of Research Laboratory (Mr. Kamarul Arifin Hadithon, Mr. Muhammad Raznishafiq Razak, Mrs. Noor Hesniza Muhamad Nordin, Mrs. Suraya Saad, Mrs Norliza Othman and Mrs Fatimah Mokhtar) for their cooperation in providing me a convenient working environment.

My deepest gratitude goes to my parents and siblings, whom I am forever indebted to. Their understanding, contributions and continuous moral supports had made the journey of completing this research possible. I would also like to express my deepest appreciation to all the lab members (Umi, Nima, Liana, Mahani and Hoda) and also a list of my close friends for their support and assistance. Last but not least, my special thanks to Parveen Devi for her willingness to edit this thesis.

I certify that a Thesis Examination Committee has met on 5 September 2014 to conduct the final examination of Hanisah binti Mustaffa @ Hamzah on her thesis entitled "Effects of Polysaccharide-Based Coating Conjugated with Silver Nanoparticles on Shelf-Life Extension of Papaya (*Carica papaya* L. cv. Eksotika)" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Roselina binti Karim, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Son Radu, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Internal Examiner)

Azizah binti Abdul Hamid, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Internal Examiner)

Norrakiah Abdullah Sani, PhD

Senior Lecturer
Universiti Kebangsaan Malaysia
Malaysia
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 9 December 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment for the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Azizah binti Osman, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Chairman)

Tan Chin Ping, PhD

Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

Farinazleen binti Mohamad Ghazali, PhD

Associate Professor
Faculty of Food Science and Technology
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice –Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____

Date: _____

Name and Matric No.: Hanisah Binti Mustaffa @ Hamzah (GS25692)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature : _____
Name of
Chairman of
Supervisory
Committee : Prof. Dr. Azizah Osman

Signature : _____
Name of
Member of
Supervisory
Committee : Prof. Dr. Tan Chin Ping

Signature : _____
Name of
Member of
Supervisory
Committee : Assoc. Prof. Dr. Farinazleen Mohamad Ghazali

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xvi
LIST OF APPENDICES	xviii
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	
2.1 Papaya	4
2.1.1 Production and geographical distribution	4
2.1.2 Characteristics and uses	4
2.1.3 Postharvest changes	8
2.1.4 Postharvest problems	10
2.1.5 Postharvest treatments	12
2.2 Edible coating	15
2.2.1 Properties and classification	15
2.2.2 Application of edible coating	18
2.3 Silver nanoparticles	20
2.3.1 Properties of silver nanoparticles	20
2.3.1 Application of silver nanoparticles	21
2.3.3 Health and safety aspects	22
2.4 Response surface methodology (RSM)	22
3 OPTIMIZATION OF POLYSACCHARIDE-BASED COATING FORMULATION FOR COATING OF PAPAYA (<i>Carica papaya</i> L. cv. Eksotika)	
3.1 Introduction	24
3.2 Materials and Methods	25
3.2.1 Materials	25
3.2.2 Preparation of edible coating formulations	25
3.2.3 Coating of papaya	26
3.2.4 Analysis of responses	26
3.2.5 Microstructure evaluation of papaya peel	27
3.2.6 Experimental design and statistical analysis	27
3.2.7 Visual observation of Eksotika papaya coated with different concentrations of chitosan	30

3.3	Results and Discussion	30
3.3.1	Response surface analysis	30
3.3.2	Firmness	32
3.3.3	Colour components (L, a and b)	33
3.3.4	Optimization and validation procedures	35
3.5	Conclusion	40
4	OPTIMIZATION OF SILVER NANOPARTICLES CONCENTRATION INCORPORATED INTO POLYSACCHARIDE-BASED COATING	
4.1	Introduction	41
4.2	Materials and Methods	42
4.2.1	Materials	42
4.2.2	Preparation of edible coating incorporated with silver nanoparticles	42
4.2.3	Characterization of silver nanoparticles	42
4.2.4	Evaluation of antibacterial activity	42
4.2.5	Evaluation of antifungal activity	43
4.2.6	Statistical analysis	43
4.3	Results and Discussion	43
4.3.1	Characterization of silver nanoparticles	43
4.3.2	Antimicrobial activity of silver nanoparticles	46
4.4	Conclusion	51
5	PHYSIOLOGICAL, PHYSICO-CHEMICAL AND MICROBIOLOGICAL CHANGES OF COATED AND UNCOATED PAPAYA (<i>Carica papaya</i> L. cv. Eksotika) DURING STORAGE	
5.1	Introduction	52
5.2	Materials and Methods	53
5.2.1	Materials	53
5.2.2	Preparation of edible coating formulations with and without silver nanopartilces	53
5.2.3	Coating and storage of papaya	53
5.2.4	Physiological analysis	54
5.2.5	Physico-chemical analysis	54
5.2.6	Microbiological analysis	55
5.2.7	Statistical analysis	56
5.3	Results and Discussion	56
5.3.1	Physiological analysis	56
5.3.2	Physico-chemical analysis	59
5.3.3	Microbiological analysis	74
5.4	Conclusion	77

6	CONCENTRATION OF SILVER NANOPARTICLES IN PEEL AND PULP OF PAPAYA AND CONSUMER ACCEPTANCE OF THE COATED AND UNCOATED PAPAYA (<i>Carica papaya</i> L. cv. Eksotika)	
	6.1 Introduction	78
	6.2 Materials and Methods	79
	6.2.1 Materials	79
	6.2.2 Preparation of edible coating formulations	79
	6.2.3 Coating and storage of papaya fruit	79
	6.2.4 Preparations of peel and pulp of papaya fruit	79
	6.2.5 Determination of silver nanoparticles in peel and pulp of papaya fruit	80
	6.2.6 Visual inspection of coated and uncoated papaya fruit	80
	6.2.7 Statistical analysis	81
	6.3 Results and Discussion	81
	6.3.1 Concentration of silver nanoparticles in peel and pulp of papaya fruit	81
	6.3.2 Consumer acceptance of coated and uncoated whole papaya fruit	83
	6.4 Conclusion	85
7	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	86
	REFERENCES	88
	APPENDICES	110
	BIODATA OF STUDENT	128
	LIST OF PUBLICATIONS	129

LIST OF TABLES

Table	Page	
2.1	Papaya export in Malaysia (2008-2011)	4
2.2	Top world papaya exporters in 2011	5
2.3.	Nutrient composition in edible portion (100g) of Eksotika papaya	7
2.4	Nutrient composition in edible portion (100g) of papaya	7
3.1.	Levels of independent variables established according to the central composite design (CCD) for carrageenan based edible coating	28
3.2	Levels of independent variables established according to the central composite design (CCD) for chitosan-based coating	28
3.3.	Central composite design and experimental data obtained for carrageenan based edible coating	28
3.4.	Central composite design and experimental data obtained for chitosan based edible coating	29
3.5.	Regression coefficients, R^2 , adjusted R^2 , probability values and lack of fit for the final reduced models of carrageenan-based coating	31
3.6	Regression coefficients, R^2 , adjusted R^2 , probability values and lack of fit for the final reduced models of chitosan-based coating	31
3.7.	Significance probability (p-values) of the independent variables for carrageenan-based coating	32
3.8.	Significance probability (p-values) of the independent variables for chitosan -based coating	32
3.9.	Predicted and experimental values of responses at optimum concentration of carrageenan and glycerol	35
3.10.	Predicted and experimental values of responses at optimum concentration of chitosan and glycerol	36
4.1.	MIC (ppm) and MBC (ppm) of chitosan- and carrageenan-silver nanoparticles nanocomposites against tested bacteria	47

4.2	Mycelial growth inhibition of <i>C. gloeosporioides</i> in PDA media with carrageenan-silver nanoparticles nanocomposite	49
4.3	Mycelial growth inhibition of <i>C. gloeosporioides</i> in PDA media with chitosan-silver nanoparticles nanocomposite	50
5.1	Changes in percent weight loss of coated and uncoated papaya fruit samples stored at ambient temperature	61
5.2	Changes in percent weight loss of coated and uncoated papaya fruit samples stored at low temperature	61
5.3	Changes in L value of coated and uncoated papaya fruit samples stored at ambient temperature	62
5.4	Changes in L value of coated and uncoated papaya fruit samples stored at low temperature	62
5.5	Changes in a value of coated and uncoated papaya fruit samples stored at ambient temperature	63
5.6	Changes in a value of coated and uncoated papaya fruit samples stored at low temperature	63
5.7	Changes in b value of coated and uncoated papaya fruit samples stored at ambient temperature	64
5.8	Changes in b value of coated and uncoated papaya fruit samples stored at low temperature	64
5.9	Changes in total soluble solids content of coated and uncoated papaya fruit samples stored at ambient temperature	68
5.10	Changes in total soluble solids content of coated and uncoated papaya fruit samples stored at low temperature	68
5.11	Changes in titratable acidity of coated and uncoated papaya fruit samples stored at ambient temperature	70
5.12	Changes in titratable acidity of coated and uncoated papaya fruit samples stored at low temperature	70
5.13	Changes in pH value of coated and uncoated papaya fruit samples stored at ambient temperature	71
5.14	Changes in pH value of coated and uncoated papaya fruit samples stored at low temperature	71
5.15	Changes in ascorbic acid content of coated and uncoated papaya fruit samples stored at ambient temperature	73

5.16	Changes in ascorbic acid content of coated and uncoated papaya fruit samples stored at low temperature	73
5.17	Changes in total plate count of coated and uncoated papaya fruit samples stored at ambient temperature	75
5.18	Changes in total plate count of coated and uncoated papaya fruit samples stored at low temperature	75
5.19	Changes in yeast and mould count of coated and uncoated papaya fruit samples stored at ambient temperature	76
5.20	Changes in yeast and mould count of coated and uncoated papaya fruit samples stored at low temperature	76
6.1.	Concentration of silver (mg/kg) in peel and pulp of papaya fruit for sample stored at ambient temperature	81
6.2.	Concentration of silver (mg/kg) in peel and pulp of papaya fruit for sample stored at low temperature	82
6.3.	Sensory scores of papaya fruit samples at day 7 of storage in ambient temperature	84
6.4.	Sensory scores of papaya fruit samples at day 13 of storage in low temperature	84

LIST OF FIGURES

Figure		Page
2.1	Visual comparison between (a) papaya of family Caricaceae and (b) 'pawpaw' of family Annonaceae	5
2.2	Chemical structure of chitosan	16
2.3	Disaccharide repeat units of κ -, ι - and λ -carrageenan	17
3.1	Response surface plots for significant ($p < 0.1$) interaction effects for (A) L value, (B) a value and (C) b value	34
3.2	Visual comparison of Eksotika papaya coated with different concentrations of chitosan	37
3.3	Visual comparison of (a) uncoated (b) optimized carrageenan-based and (c) 1.0% chitosan-based coated Eksotika papaya at 5 th day of storage	39
3.4	SEM micrographs of the surface of Eksotika papaya fruit at 5 th day of storage at ambient condition. (a) Control (b) Carrageenan (c) 1% chitosan and (d) 2.32% chitosan coated	40
4.1.	UV-visible spectra of (a) carrageenan-silver nanoparticles and (b) chitosan-silver nanoparticles nanocomposites	44
4.2.	TEM micrographs showing spherical silver nanoparticles (a) uncoated particles (b) carrageenan-silver nanoparticles and (c) chitosan-silver nanoparticles nanocomposites.	45
5.1.	Changes in respiration rate of coated and uncoated papaya fruit samples stored at (a) ambient and (b) low temperatures.	57
5.2.	Changes in ethylene production rate of coated and uncoated papaya fruit samples stored at (a) ambient and (b) low temperatures.	59
5.3	Changes in firmness of control and coated papaya samples stored at (a) ambient and (b) low temperatures.	66
5.4	SEM images of papaya mesocarp cells at 5 th day of storage at ambient condition. (a) Control (b) Carrageenan (c) Carrageenan-silver nanocomposite and (d) Chitosan coated	67

6.1. Cross section of papaya; peel and pulp layers used for analysis

80



LIST OF APPENDICES

Appendix		Page
A	Maturity Index of Eksotika Papaya	110
B	Physical Appearance of Polysaccharide-Silver Nanocomposite Solutions	111
C	Appearance of 96-Well Microtitre Plate for Minimum Inhibitory Concentration (MIC) Determination	112
D	Minimum Bactericidal Concentration (MBC) Agar Plates	113
E	Mycelial Growth Inhibition Agar Plates	115
F	Formula for Gas Concentrations	117
G	Coated and Uncoated Papaya fruit Samples Stored At Ambient and Low Temperatures	118
H	Chromatograms of Ethylene Production	122
I	Chromatograms of Ascorbic Acid Content	125
J	Questionnaire for Sensory Evaluation	127

LIST OF ABBREVIATIONS

1-MCP	1- Methylcyclopropene
ANOVA	Analysis of variance
ATCC	American Type Culture Collection
CCD	Central composite design
CFU	Colony forming units
DNA	Deoxyribonucleic acid
FAOSTAT	Food and Agriculture Organization Corporate Statistical Database
HNO ₃	Nitric acid
ICP-OES	Inductively coupled plasma- optical emission spectrometry
LDPE	Low density polyethylene
MAP	Modified atmosphere packaging
MARDI	Malaysian Agricultural Research and Development Institute
MBC	Minimum bactericidal concentration
MHA	Mueller-Hinton agar
MHB	Mueller-Hinton broth
MIC	Minimum inhibitory concentration
NaOH	Natrium hydroxide/ sodium hydroxide
PCA	Plate count agar
PDA	Potato dextrose agar
PE	Polyethylene
PET	Polyethylene terephthalate
PP	Polypropylene
PVP	Polyvinylpyrrolidone
RSM	Response surface methodology

SEM	Scanning electron microscopy
TA	Titrateable acidity
TEM	Transmission electron microscopy
TPC	Total plate count
TSS	Total soluble solids
UK	United Kingdom
USA	United States of America
USDA	United States Department of Agriculture
USEPA	United States Environmental Protection Agency
UV-Vis	Ultraviolet-visible
w/v	weight/volume
YM	Yeast and mould

CHAPTER 1

INTRODUCTION

Fruits and vegetables can be classified as climacteric or non-climacteric. Climacteric fruits continue to ripen after harvest and show more dramatic changes in their physiological activity (Maftoonazad & Ramaswamy, 2005). Papaya is one of the climacteric fruit and has attained great popularity among growers due to the fact it can be intensively cultivated, its rapid returns and the increase demand for the fresh fruit and its processed products (Singh & Rao, 2011). The ripe fruit are usually eaten as a dessert but they can also be processed into jams, soft drinks, ice cream flavouring, crytallised and candied fruits, fruit rolls and as filler for sauces (Md. Yon, 1994). Papaya is an important fruit in ASEAN countries and can be divided into two groups. The first group consists of varieties which bear large, elongated fruits. The second group on the other hand bears small, pyriform or round shaped fruit with very good quality. The varieties in the first group include Subang, Sitiawan, Batu Arang, Kaegdum, Kaegnuan and Sainampeung. The common varieties in the second group are Solo and Eksotika (Chan *et al.*, 1994). Due to the small, uniform size and also its ease of handling and serving, Eksotika papaya has become very popular among local consumers (Chan, 2011).

Brazil, Mexico and Malaysia are the main papaya exporting countries mainly to the European market (Cia *et al.*, 2007) with Malaysia being the second most important exporter of Eksotika papaya. Although the export revenue from papaya is quite significant, the prospects for market expansion are rather restricted due to expensive and limited air-cargo space (Chan, 2008). Short storage life limits the export of papaya to distant markets by refrigerated sea containers (Mahmud *et al.*, 2008). The short postharvest life of papaya fruit is contributed by many factors including diseases. Postharvest losses due to diseases can reach about 97% depending upon the postharvest handling and packaging procedures (Alvarez & Nishijima, 1987). Anthracnose is an important postharvest fungal disease in most tropical and sub-tropical regions (Teixeira da Silva *et al.*, 2007). Anthracnose was found to be the main postharvest disease associated with papaya in Brazil and other countries (Cia *et al.*, 2007). In addition, inspection of papaya shipments that were delivered to the New York market revealed that anthracnose rot caused by *Colletotrichum gloeosporioides* was the major disorder (Cappellini *et al.*, 1988). It was reported that 40 to 100 % of papaya fruits were lost either due to its short postharvest life or due to lack of proper postharvest technology (Vyas *et al.*, 2014).

Low temperature storage, heat treatments and controlled/modified atmosphere packaging are some of the common techniques that have been used to prolong the shelf life of papaya and consequently reduces postharvest losses (Alvarez & Nishijima, 1987; Abd. Rahman *et al.*, 1994; Arpaia & Kader, 1997; Paull *et al.*, 1997; Gonzalez-Aguilar *et al.*, 2003; Singh & Rao, 2005). Even though these techniques were found to be successful in achieving their objectives, several limitations have made the needs for other technique necessary. Papaya fruits are sensitive to prolonged storage at too low temperature. The fruits exhibit signs of chilling injury when exposed to temperature of about 10°C or lower (Biglete *et al.*, 1994). On the other hand, exposure to heat during vapour heat treatment, hot water treatment or high ambient temperature causes hyperthermal injury in papaya (Biglete *et al.*, 1994).

These limitations have urged the needs for an alternative technique to maintain the quality of papaya. Edible coating is an alternative packaging methods for fresh products including whole fruits. It have attracted increasing attention because of environmental considerations and the trends toward the use of convenience food (Özden & Bayindirli, 2002). In addition, application of edible coatings on fresh fruit can provide an alternative to modified atmosphere storage by reducing quality changes and quantity losses through modification and control of the internal atmosphere of the individual fruits (Park, 1999). Studies have shown that semi permeable coatings are capable to improve the storage life of perishable fruits (Worrel *et al.*, 2002). Shelf life of mango was extended by coating treatments due to delayed ripening and softening, reduced decay and weight loss (Baldwin *et al.*, 1999). Edible coatings may contribute to extend the shelf life of fresh-cut fruits by reducing moisture and solute migration, gas exchange, respiration and oxidative reaction rates (Baldwin *et al.*, 1996). The application of edible coating have been expanded due to its high potential to carry active ingredients such as antibrowning agents, colourants, flavours, nutrients, spices and antimicrobial compounds that can extend product shelf life and reduce the risk of pathogenic growth on food surfaces (Pranoto *et al.*, 2005).

Range of materials including essential oils and plant extracts has been incorporated into coating formulations to improve it antimicrobial properties (Bautista-Baños *et al.*, 2003; Rojas-Graü *et al.*, 2007 & Ponce *et al.*, 2008). However, due to the high resistance of some microbial strains towards these antimicrobial agents, new ones are needed (Sondi & Salopek-Sondi, 2004). Nowadays, nanotechnologies are being used to enhance the nutritional aspects of food by means of nanoscale additives and nutrients and also nanosized delivery systems for bioactive compounds (Bouwmeester *et al.*, 2007). Metal nanoparticles with silver nanoparticles in particular are well known for their antimicrobial properties. Numerous studies have been conducted to determine the antibacterial and antifungal activity of silver nanoparticles (Sondi & Salopek-Sondi, 2004; Cho *et al.*, 2005; Kim *et al.*, 2008, 2009 & Lamsal *et al.*, 2011). Their application as antimicrobial agent in various industries including food packaging is growing rapidly. Silver nanoparticles incorporated polymer packaging materials have been used to reduce microbial load and prolong shelf life of asparagus, carrot, pear and orange juice (An *et al.*, 2008; Mohammed Fayaz *et al.*, 2009 & Emamifar *et al.*, 2010, 2011). However, studies on the application of nanoparticles incorporated into edible coating on specific fruits are rather limited. In addition, very few studies were done on the migration of these nanoparticles into the coated or packed food.

Eksotika papaya has the potential to be applied with the silver nanoparticles incorporated edible coating. This is because the fruit are being largely exported and an efficient packaging that can help in extending their shelf life and maintaining the freshness by the time they reach consumer is required. Successful shelf life extension of papaya fruit will offer a chance to exploit the export of the fruits by less expensive sea freight (Chan, 2008). Thus, the study was carried out based on the hypothesis that the application of polysaccharides-based surface coating with the addition of silver nanoparticles will help to extend the shelf life of Eksotika papaya and reduces postharvest losses due to pathological disorder, mainly anthracnose.

Therefore, the objectives of this study are:

Overall:

To extend the shelf life of Eksotika papaya using polysaccharides-silver nanocomposite-based coating,

Specific:

1. To optimize edible coating formulations (chitosan- and carrageenan-based) for coating of Eksotika papaya,
2. To determine optimum silver nanoparticles concentration that are able to give effective antimicrobial effect,
3. To evaluate the effect of edible coating incorporated with silver nanoparticles on the quality aspects of Eksotika papaya and
4. To determine silver concentration in peel and pulp of papaya fruit samples and consumer acceptance towards the coated and uncoated papaya fruits.

REFERENCES

- Abd. Rahman, A. ., Tan, S. C., & Lazan, H. (1994). Storage. In R. Md. Yon (Ed.), *Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN* (pp. 48–55). Kuala Lumpur: ASEAN Food Handling Bureau.
- Aked, J. (2002). Maintaining the post-harvest quality of fruits and vegetables. In W. Jongen (Ed.), *Fruits and Vegetables Processing: Improving Quality* (pp. 119–149). Cambridge, England: Woodhead Publishing Limited.
- Ali, A., Maqbool, M., Ramachandran, S., & Alderson, P. G. (2010). Gum arabic as a novel edible coating for enhancing shelf-life and improving postharvest quality of tomato (*Solanum lycopersicum* L.) fruit. *Postharvest Biology and Technology*, 58(1), 42–47.
- Ali, A., Muhammad, M. T. M., Sijam, K., & Siddiqui, Y. (2011). Effect of chitosan coatings on the physicochemical characteristics of Eksotika II papaya (*Carica papaya* L.) fruit during cold storage. *Food Chemistry*, 124(2), 620–626.
- Alvarenga, E. S. De. (2011). Characterization and properties of chitosan. In M. Elnashar (Ed.), *Biotechnology of Biopolymers* (pp. 91–108). Croatia: InTech.
- Alvarez, A. M., & Nishijima, W. T. (1987). Postharvest diseases of papaya. *Plant Disease*, 71(8), 681–686.
- Ames, G., & Greer, L. (2010). Pawpaw — a “ tropical ” fruit for temperate climates. *NCAT Agriculture Specialists*, 1–8.
- An, J., Zhang, M., Wang, S., & Tang, J. (2008). Physical, chemical and microbiological changes in stored green asparagus spears as affected by coating of silver nanoparticles-PVP. *LWT - Food Science and Technology*, 41(6), 1100–1107.
- An, J.-F., & Paull, R. E. (1990). Storage temperature and ethylene influence on ripening of papaya fruit. *Journal of the American Society for Horticultural Science*, 115(6), 949–953.
- Andrews, J. M. (2001). Determination of minimum inhibitory concentrations. *Journal of Antimicrobial Chemotherapy*, 48 Suppl 1, 5–16.
- Arpaia, M. L., & Kader, A. A. (1997). Papaya: Recommendations for maintaining postharvest quality. *Perishable Handling #90*. Retrieved from <http://postharvest.ucdavis.edu>
- Asharani, P. V, Lian Wu, Y., Gong, Z., & Valiyaveetil, S. (2008). Toxicity of silver nanoparticles in zebrafish models. *Nanotechnology*, 19(25), 255102.
- Avena-Bustillos, R. D. J., Krochta, J. M., & Saltveit, M. E. (1997). Water vapor resistance of red delicious apples and celery sticks coated with edible caseinate-acetylated monoglyceride films. *Journal of Food Science*, 62(2), 351–354.

- Azarakhsh, N., Azizah, O., Ghazali, H. M., Tan, C. P., & Mohd Adzahan, N. (2012). Optimization of alginate and gellan-based edible coating formulations for fresh-cut pineapples. *International Food Research Journal*, 19(1), 279–285.
- Azevedo, A. N., Buarque, P. R., Cruz, E. M. O., Blank, A. F., Alves, P. B., Nunes, M. L., & Santana, L. C. L. D. A. (2014). Response surface methodology for optimisation of edible chitosan coating formulations incorporating essential oil against several foodborne pathogenic bacteria. *Food Control*, 43, 1–9.
- Azevedo, I. G., Oliveira, J. G., da Silva, M. G., Pereira, T., Corrêa, S. F., Vargas, H., & Façanha, A. R. (2008). P-type H⁺-ATPases activity, membrane integrity, and apoplastic pH during papaya fruit ripening. *Postharvest Biology and Technology*, 48(2), 242–247.
- Badawy, M. E. I., & Rabea, E. I. (2011). A biopolymer chitosan and its derivatives as promising antimicrobial agents against plant pathogens and their applications in crop protection. *International Journal of Carbohydrate Chemistry*, 2011, 1–29.
- Baez-Sañudo, M., Siller-Cepeda, J., Muy-Rangel, D., & Heredia, J. B. (2009). Extending the shelf-life of bananas with 1-methylcyclopropene and a chitosan-based edible coating. *Journal of the Science of Food and Agriculture*, 89(14), 2343–2349.
- Baldwin, E. A., Burns, J. K., Kazokas, W., Brecht, J., Hagenmaier, R. D., Bender, R. J., & Pesis, E. (1999). Effect of two edible coatings with different permeability characteristics on mango (*Mangifera indica* L.) ripening during storage. *Postharvest Biology and Technology*, 17(3), 215–226.
- Baldwin, E. A., Nisperos, M. O., Chen, X., & Hagenmaier, R. D. (1996). Improving storage life of cut apple and potato with edible coating. *Postharvest Biology and Technology*, 9(2), 151–163.
- Bautista-Baños, S., Hernández-López, M., Bosquez-Molina, E., & Wilson, C. L. (2003). Effects of chitosan and plant extracts on growth of *Colletotrichum gloeosporioides*, anthracnose levels and quality of papaya fruit. *Crop Protection*, 22(9), 1087–1092.
- Becker, B. R. and Fricke, B. A. (1996). Transpiration and respiration of fruits and vegetables. In Murphy, W. E. (Ed.), *New Developments in Refrigeration for Food Safety and Quality* (pp. 110-121). International Institute of Refrigeration, Paris, France and American Society of Agricultural Engineers, St. Joseph, Michigan.
- Ben-Yehoshua, S., Burg, S. P., & Young, R. (1985). Resistance of citrus fruit to mass transport of water vapor and other gases. *Plant Physiology*, 79(4), 1048–1053.

- Bezerra, M. A., Santelli, R. E., Oliveira, E. P., Villar, L. S., & Escaleira, L. A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, *76*(5), 965–77.
- Bico, S. L. S., Raposo, M. F. J., Morais, R. M. S. C., & Morais, a. M. M. B. (2009). Combined effects of chemical dip and/or carrageenan coating and/or controlled atmosphere on quality of fresh-cut banana. *Food Control*, *20*(5), 508–514.
- Biglete, N. A., Zainon, M. A., & Lazan, H. (1994). Postharvest pathology and control of diseases. In R. Md. Yon (Ed.), *Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN* (pp. 75–82). Kuala Lumpur: ASEAN Food Handling Bureau.
- Blasco, C., & Pico, Y. (2011). Determining nanomaterials in food. *Trends in Analytical Chemistry*, *30*(1), 84–99.
- Bourtoom, T. (2008). Edible films and coatings: characteristics and properties. *International Food Research Journal*, *15*(3), 1–12.
- Bouwmeester, H., Dekkers, S., Noordam, M., Hagens, W., Bulder, A., Heer, C. De., Voorde, S. Ten., Wijnhoven, S. & Sips, A. (2007). Health impact of nanotechnologies in food production. 1-91.
- Božanić, D. K., Trandafilović, L. V., Luyt, A. S., & Djoković, V. (2010). “Green” synthesis and optical properties of silver–chitosan complexes and nanocomposites. *Reactive and Functional Polymers*, *70*(11), 869–873.
- Brady, C. J. (1984). Fruit ripening. *Annual Review of Plant Physiology*, *38*(1), 155–178.
- Brasil, I. M., Gomes, C., Puerta-gomez, A., Castell-perez, M. E., & Moreira, R. G. (2012). Polysaccharide-based multilayered antimicrobial edible coating enhances quality of fresh-cut papaya. *LWT - Food Science and Technology*, *47*(1), 39–45.
- Bron, I. U., & Jacomino, A. P. (2006). Ripening and quality of ‘ “Golden” ’ papaya fruit harvested at different maturity stages. *Brazilian Journal of Plant Physiology*, *18*(3), 389–396.
- Brownleader, M. D., Jackson, P., Mobasheri, A., Pantelides, A. T., Sumar, S., Trevan, M., & Dey, P. M. (1999). Molecular aspects of cell wall modifications during fruit ripening. *Critical Reviews in Food Science and Nutrition*, *39*(2), 149–164.
- Brummell, D. A., & Harpster, M. H. (2001). Cell wall metabolism in fruit softening and quality and its manipulation in transgenic plants. *Plant Molecular Biology*, *47*, 311–340.
- Bryan, D. S. (1974). Prepared citrus fruit halves and method of making the same. U.S. Patent 3, 851, 067.

- Burg, S. P. (2004). Horticultural commodity requirements. In *Postharvest Physiology and Hypobaric Storage of Fresh Produce* (pp. 371–439). Wallingford, UK: CABI Publishing.
- Camargo, R. J., Tadini, C. C., & Sabato, S. F. (2007). Physical–chemical analyses of irradiated papayas (*Carica papaya* L.). *Radiation Physics and Chemistry*, 76(11-12), 1866–1868.
- Campo, V. L., Kawano, D. F., Silva, D. B. Da, & Carvalho, I. (2009). Carrageenans: Biological properties, chemical modifications and structural analysis – a review. *Carbohydrate Polymers*, 77(2), 167–180.
- Cappellini, R. A., Cepaonis, M. J., & Lightner, G. W. (1988). Disorders in apricot and papaya shipments to the new york market, 1972-1985. *Plant Disease*, 72(4), 366–368.
- Chan, H. T., & Tang, C. S. (1979). The chemistry and biochemistry of papaya. In G. E. Inglett & G. Charalambous (Eds.), *Tropical Foods Vol 1* (pp. 33–54). New York: Academic Press.
- Chan, Y. K. (1989). Breeding for better eksotikas. In Y. K. Chan & R. P (Eds.), *Proceedings of the MARDI-MAPPS Seminar on Eksotika Papaya: Problems and Prospects* (pp. 8–12). Johor Bahru, Malaysia: MARDI-MAPPS.
- Chan, Y. K. (2008). Papaya (*Carica papaya*). In Y. K. Chan, S. L. Tan, & S. H. Jamaluddin (Eds.), *Breeding Horticultural Crops@ MARDI* (pp. 175–206). Kuala Lumpur: Malaysian Agricultural Research and Development Institute.
- Chan, Y. K. (2011). Kultivar. In Y. K. Chan, P. Raveendranathan, M. L. Raziah, & S. T. Choo (Eds.), *Penanaman Betik* (pp. 5–11). Kuala Lumpur, Malaysia: Institut Penyelidikan dan Kemajuan Pertanian Malaysia (MARDI).
- Chan, Y. K., Napakoonwong, U., Broto, W., Huat, K. S., & Espino, R. R. C. (1994). Commercial papaya cultivars in ASEAN. In R. Md. Yon (Ed.), *Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN* (pp. 5–17). Kuala Lumpur: ASEAN Food Handling Bureau.
- Chandrika, U. G., Jansz, E. R., Wickramasinghe, S., & Warnasuriya, N. D. (2003). Carotenoids in yellow- and red-fleshed papaya (*Carica papaya* L.). *Journal of the Science of Food and Agriculture*, 83, 1279–1282.
- Chen, X., & Schluesener, H. J. (2008). Nanosilver: a nanoproduct in medical application. *Toxicology Letters*, 176(1), 1–12.
- Cheng, G., Duan, X., Shi, J., Lu, W., Luo, Y., Jiang, W., & Jiang, Y. (2008). Effects of reactive oxygen species on cellular wall disassembly of banana fruit during ripening. *Food Chemistry*, 109(2), 319–324.

- Chien, P.-J., Sheu, F., & Yang, F.-H. (2007). Effects of edible chitosan coating on quality and shelf life of sliced mango fruit. *Journal of Food Engineering*, 78(1), 225–229.
- Chillo, S., Flores, S., Mastromatteo, M., Conte, A., Gerschenson, L., & Del Nobile, M. a. (2008). Influence of glycerol and chitosan on tapioca starch-based edible film properties. *Journal of Food Engineering*, 88(2), 159–168.
- Cho, K.-H., Park, J.-E., Osaka, T., & Park, S.-G. (2005). The study of antimicrobial activity and preservative effects of nanosilver ingredient. *Electrochimica Acta*, 51(5), 956–960.
- Cia, P., Pascholati, S. F., Benato, E. A., Camili, E. C., & Santos, C. A. (2007). Effects of gamma and UV-C irradiation on the postharvest control of papaya anthracnose. *Postharvest Biology and Technology*, 43(3), 366–373.
- Conklin, P. L. (2001). Recent advances in the role and biosynthesis of ascorbic acid in plants. *Plant Physiology and Biochemistry*, 24, 383–394.
- Damm, C., Münstedt, H., & Rösch, a. (2008). The antimicrobial efficacy of polyamide 6/silver-nano- and microcomposites. *Materials Chemistry and Physics*, 108(1), 61–66.
- Dangaran, K., Tomasula, P. G., & Qi, P. (2009). Structure and function of protein-based edible films and coatings. In M. E. Embuscado & K. C. Huber (Eds.), *Edible Films and Coatings for Food Applications* (pp. 25–56). New York: Springer Science + Business Media, LLC.
- Daniel da Silva, A. L., & Trindade, T. (2011). Biofunctional composites of polysaccharides containing inorganic nanoparticles. In A. A. Hashim (Ed.), *Advances in Nanocomposite Technology* (pp. 275–298). Croatia: InTech.
- Daniel-da-Silva, A. L., Trindade, T., Goodfellow, B. J., Costa, B. F. O., Correia, R. N., & Gil, A. M. (2007). In situ synthesis of magnetite nanoparticles in carrageenan gels. *Biomacromolecules*, 8(8), 2350–2357.
- Davey, M. W., Gilot, C., Persiau, G., Ostergaard, J., Han, Y., Bauw, G. C., & Van Montagu, M. C. (1999). Ascorbate biosynthesis in Arabidopsis cell suspension culture. *Plant Physiology*, 121, 535–43.
- Del Nobile, M. A., Cannarsi, M., Altieri, C., Sinigaglia, M., Favia, P., Iacoviello, G., & D'Agostino, R. (2004). Effect of ag-containing nano-composite active packaging system on survival of *Alicyclobacillus acidoterrestris*. *Journal of Food Science*, 69(8), E379–E383.
- Department of Agriculture. (2012). *Perangkaan Tanaman Buah-buahan Fruit Crop Statistics*. Putrajaya: Jabatan Pertanian Semenanjung Malaysia.
- Duan, X., Zhang, H., Zhang, D., Sheng, J., Lin, H., & Jiang, Y. (2011). Role of hydroxyl radical in modification of cell wall polysaccharides and aril

- breakdown during senescence of harvested longan fruit. *Food Chemistry*, 128(1), 203–207.
- Durango, A. M., Soares, N. F., & Andrade, N. J. (2006). Microbiological evaluation of an edible antimicrobial coating on minimally processed carrots. *Food Control*, 17(5), 336–341.
- El Ghaouth, A., Ponnampalam, R., Castaigne, F., & Arul, J. (1992). Chitosan coating to extend the storage life of tomatoes. *HortScience*, 27(9), 1016–1018.
- El-Badawy, A., Feldhake, D., & Venkatapathy, R. (2010). *State of the Science Literature Review: Everything Nanosilver and More*. Washington.
- El-Ghaouth, A., Arul, J., Grenier, J., & Asselin, A. (1992). Antifungal activity of chitosan on two postharvest pathogens of strawberry fruits. *Phytopathology*, 82(4), 398–402.
- Elsabee, M. Z., & Abdou, E. S. (2013). Chitosan based edible films and coatings: a review. *Materials Science & Engineering. C, Materials for Biological Applications*, 33(4), 1819–41.
- Emamifar, A., Kadivar, M., Shahedi, M., & Soleimani-zad, S. (2010). Evaluation of nanocomposite packaging containing Ag and ZnO on shelf life of fresh orange juice. *Innovative Food Science and Emerging Technologies*, 11(4), 742–748.
- Emamifar, A., Kadivar, M., Shahedi, M., & Soleimani-zad, S. (2011). Effect of nanocomposite packaging containing Ag and ZnO on inactivation of *Lactobacillus plantarum* in orange juice. *Food Control*, 22(3-4), 408–413.
- Emeruwa, A. C. (1982). Antibacterial substance from *Carica papaya* fruit extract. *Journal of Natural Products*, 45(2), 123–127.
- Espitia, P. J. P., Avena-Bustillos, R. J., Du, W.-X., Teófilo, R. F., Soares, N. F. F., & McHugh, T. H. (2014). Optimal antimicrobial formulation and physical–mechanical properties of edible films based on açai and pectin for food preservation. *Food Packaging and Shelf Life*, 2(1), 38–49.
- Fabi, J. P., Cordenunsi, B. R., De Mattos Barreto, G. P., Mercadante, A. Z., Lajolo, F. M., & Do Nascimento, J. R. O. (2007). Papaya fruit ripening: response to ethylene and 1-methylcyclopropene. *Journal of Agricultural and Food Chemistry*, 55, 6118–6123.
- Falguera, V., Quintero, J. P., Jiménez, A., Muñoz, J. A., & Ibarz, A. (2011). Edible films and coatings: structures, active functions and trends in their use. *Trends in Food Science & Technology*, 22(6), 292–303.
- FAMA (2008). Standard FAMA: FS003-2000. *Brosur Siri Panduan Kualiti Betik Eksotika*

- Fang, M., Chen, J.-H., Xu, X.-L., Yang, P.-H., & Hildebrand, H. F. (2006). Antibacterial activities of inorganic agents on six bacteria associated with oral infections by two susceptibility tests. *International Journal of Antimicrobial Agents*, 27(6), 513–7.
- FAOSTAT. (2013). Statistical database. *Food and Agriculture Organization of the United States*. Retrieved from <http://faostat3.fao.org>
- Feng, Q. L., Wu, J., Chen, G. Q., Cui, F. Z., Kim, T. N., & Kim, J. O. (2000). A mechanistic study of the antibacterial effect of silver ions on *Escherichia coli* and *Staphylococcus aureus*. *Journal of Biomedical Materials Research*, 52(4), 662–8.
- Floros, J. D., & Matsos, K. I. (2005). Introduction to modified atmosphere packaging. In J. H. Han (Ed.), *Innovations in Food Packaging* (pp. 159–170). London: Elsevier Academic Press.
- Follett, P. A. (2009). Generic radiation quarantine treatments: the next steps. *Journal of Economic Entomology*, 102(4), 1399–1406.
- Food Act 1983 (Act 281) & Regulations (1st March 2013). Selangor, Malaysia: International Law Book Services.
- Fortunati, E., Peltzer, M., Armentano, I., Jiménez, a., & Kenny, J. M. (2013). Combined effects of cellulose nanocrystals and silver nanoparticles on the barrier and migration properties of PLA nano-biocomposites. *Journal of Food Engineering*, 118(1), 117–124.
- Garcia, E., & Barret, D. M. (2002). Preservatives treatment for fresh cut fruits and vegetables. In O. Lamikanra (Ed.), *Fresh-Cut Fruits and Vegetables* (pp. 267–304). Boca Raton, Florida: CRC Press.
- Gennadios, A. (2002). *Protein-Based Edible Films and Coatings*. Boca Raton, Florida: CRC Press.
- Ghasemzadeh, R., Karbassi, A., & Ghoddousi, H. B. (2008). Application of Edible Coating for Improvement of Quality and Shelf-life of Raisins. *World Applied Sciences Journal*, 3(1), 82–87.
- Gils, P. S., Ray, D., & Sahoo, P. K. (2010). Designing of silver nanoparticles in gum arabic based semi-IPN hydrogel. *International Journal of Biological Macromolecules*, 46(2), 237–44.
- Gleiter, H. (2000). Nanostructured materials: Basic concepts and microstructure. *Acta Materialia*, 48, 1–29.
- Gol, N. B., Patel, P. R., & Rao, T. V. R. (2013). Improvement of quality and shelf-life of strawberries with edible coatings enriched with chitosan. *Postharvest Biology and Technology*, 85, 185–195.

- Gonzalez-Aguilar, G. A., Buta, J. G., & Wang, C. Y. (2003). Methyl jasmonate and modified atmosphere packaging (MAP) reduce decay and maintain postharvest quality of papaya “ Sunrise .” *Postharvest Biology and Technology*, 28, 361–370.
- Hackenbarg, S., Scherzed, A., Kessler, M., Hummel, S., Technau, A., Froelich, K., Ginzkey, C., Koehler, C., Hagen, R. & Kleinsasser, N. (2011). Silver nanoparticles: evaluation of DNA damage, toxicity and functional impairment in human mesenchymal stem cells. *Toxicology Letters*, 201(1), 27–33.
- Hagenmaier, R. D. (2000). Evaluation of a polyethylene – candelilla coating for “Valencia” oranges. *Postharvest Biology and Technology*, 19, 147–154.
- Hagenmaier, R. D. (2002). The flavor of mandarin hybrids with different coatings. *Postharvest Biology and Technology*, 24(1), 79–87.
- Han, J., & Gennadios, A. (2005). Edible films and coatings: a review. In J. H. Han (Ed.), *Innovations in Food Packaging* (pp. 239–262). London, UK: Elsevier Academic Press.
- Hebeish, A. A., El-Rafie, M. H., Abdel-Mohdy, F. a., Abdel-Halim, E. S., & Emam, H. E. (2010). Carboxymethyl cellulose for green synthesis and stabilization of silver nanoparticles. *Carbohydrate Polymers*, 82(3), 933–941.
- Hernández, Y., Lobo, M. G., & González, M. (2006). Determination of vitamin C in tropical fruits: A comparative evaluation of methods. *Food Chemistry*, 96(4), 654–664.
- Hernández-Muñoz, P., Almenar, E., Del, V., Velez, D., & Gavara, R. (2008). Effect of chitosan coating combined with postharvest calcium treatment on strawberry (*Fragaria x ananassa*) quality during refrigerated storage. *Food Chemistry*, 110, 428–435.
- Hoagland, P. D., & Parris, N. (1996). Chitosan / pectin laminated films. *Journal of Agricultural and Food Chemistry*, 44(7), 1915–1919.
- HunterLab. (2008). Hunter L, a, b color scale. *Application Note*. 8(9). Retrieved from www.hunterlab.com
- Hussain, S. M., Hess, K. L., Gearhart, J. M., Geiss, K. T., & Schlager, J. J. (2005). In vitro toxicity of nanoparticles in BRL 3A rat liver cells. *Toxicology in Vitro*, 19(7),
- Ilag, L. L., Muid, S., Tan, S. C., Prabawati, S., & Vichitrananda, S. (1994). Postharvest pathology and control of diseases. In R. Md. Yon (Ed.), *Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN* (pp. 83–98). Kuala Lumpur: ASEAN Food Handling Bureau.

- Imeson, A. P. (2009). Carrageenan and furcellaran. In G.O. Philips & P. A. Williams (Eds.), *Handbook of hydrocolloids* (pp. 164–185). Cambridge, UK: Woodhead Publishing Limited.
- Isabelle, M., Lee, B. L., Lim, M. T., Koh, W.-P., Huang, D., & Ong, C. N. (2010). Antioxidant activity and profiles of common fruits in Singapore. *Food Chemistry*, 123(1), 77–84.
- Jacxsens, L., Devlieghere, F., & Debevere, J. (2002). Predictive modelling for packaging design: equilibrium modified atmosphere packages of fresh-cut vegetables subjected to a simulated distribution chain. *International Journal of Food Microbiology*, 73(2-3), 331–41.
- Jafarizadeh Malmiri, H. (2012). *Quality Improvement of Banana using Edible Surface Coating Conjugated with Silver Nanoparticles*. Universiti Putra Malaysia.
- Jafarizadeh Malmiri, H., Osman, A., Tan, C. P., & Abdul Rahman, R. (2011). Development of an edible coating based on chitosan-glycerol to delay “Berangan ” banana (*Musa sapientum* cv . Berangan) ripening process. *International Food Research Journal*, 18(3), 989–997.
- Jagtiani, J., Chan, H. T. J., & Sakai, W. S. (1988). *Tropical Fruit Processing*. London: Academic Press Inc. Ltd.
- Jaouen, V., Brayner, R., Lantiat, D., Steunou, N., & Coradin, T. (2010). In situ growth of gold colloids within alginate films. *Nanotechnology*, 21(18), 185605.
- Jiang, Y., & Li, Y. (2001). Effects of chitosan coating on postharvest life and quality of longan fruit. *Food Chemistry*, 73, 139–143.
- Jokar, M., & Abdul Rahman, R. (2014). Study of silver ion migration from melt-blended and layered-deposited silver polyethylene nanocomposite into food simulants and apple juice. *Food Additives & Contaminants: Part A*, 31(4), 734–42.
- Jung, W. K., Koo, H. C., Kim, K. W., Shin, S., Kim, S. H., & Park, Y. H. (2008). Antibacterial activity and mechanism of action of the silver ion in *Staphylococcus aureus* and *Escherichia coli*. *Applied and Environmental Microbiology*, 74(7), 2171–8.
- Kader, A. A., & Saltveit, M. E. (2003). Respiration and gas exchange. In J. A. Bartz & J. K. Brecht (Eds.), *Postharvest Physiology and Pathology of Vegetables* (pp. 7–29). New York: Marcel Dekker, Inc.
- Kang, H. J., Jo, C., Kwon, J. H., Kim, J. H., Chung, H. J., & Byun, M. W. (2007). Effect of a pectin-based edible coating containing green tea powder on the quality of irradiated pork patty. *Food Control*, 18(5), 430–435.

- Karbowiak, T., Debeaufort, F., & Voilley, A. (2007). Influence of thermal process on structure and functional properties of emulsion-based edible films. *Food Hydrocolloids*, 21, 879–888.
- Kemp, M. M., Kumar, A., Clement, D., Ajayan, P., Mousa, S., & Linhardt, R. J. (2009). Hyaluronan- and heparin-reduced silver nanoparticles with antimicrobial properties. *Nanomedicine (Lond)*, 4(4), 421–429.
- Kilinceker, O., Dogan, İ. S., & Kucukoner, E. (2009). Effect of edible coatings on the quality of frozen fish fillets. *LWT - Food Science and Technology*, 42(4), 868–873.
- Kim, K. W., Min, B. J., Kim, Y.-T., Kimmel, R. M., Cooksey, K., & Park, S. I. (2011). Antimicrobial activity against foodborne pathogens of chitosan biopolymer films of different molecular weights. *LWT - Food Science and Technology*, 44(2), 565–569.
- Kim, K.-J., Sung, W. S., Moon, S.-K., Choi, J.-S., Kim, J. G., & Lee, D. G. (2008). Antifungal effect of silver nanoparticles on dermatophytes. *Journal of Microbiology and Biotechnology*, 18(8), 1482–4.
- Kim, K.-J., Sung, W. S., Suh, B. K., Moon, S.-K., Choi, J.-S., Kim, J. G., & Lee, D. G. (2009). Antifungal activity and mode of action of silver nano-particles on *Candida albicans*. *Biometals*, 22(2), 235–42.
- Kittur, F. S., Saroja, N., Habibunnisa, & Tharanathan, R. N. (2001). Polysaccharide-based composite coating formulations for shelf-life extension of fresh banana and mango. *European Food Research and Technology*, 213, 306–311.
- Krishna, K. L., Paridhavi, M., & Patel, J. A. (2008). Review on nutritional , medicinal and pharmacological properties of Papaya (*Carica papaya Linn.*). *Natural Product Radiance*, 7(4), 364–373.
- Krochta, J. M., Saltveit, M., & Cisneros-zevallos, L. (1996). Method of preserving natural color on fresh and minimally processed fruits and vegetables. U.S. Patent 5,547,693.
- Kuo, P.-L., & Chen, W.-F. (2003). Formation of silver nanoparticles under structured amino groups in pseudo-dendritic poly(allylamine) derivatives. *The Journal of Physical Chemistry B*, 107(41), 11267–11272.
- Lacroix, M., & Tien, C. Le. (2005). Edible films and coatings from non- starch polysaccharides. In J. . Han (Ed.), *Innova* (pp. 338–361). London, UK: Elsevier Academic Press.
- Lagarón, J. M., Cabedo, L., Cava, D., Feijoo, J. L., Gavara, R., & Gimenez, E. (2005). Improving packaged food quality and safety. Part 2: nanocomposites. *Food Additives and Contaminants*, 22(10), 994–998.

- Lamsal, K., Kim, S. W., Jung, J. H., Kim, Y. S., Kim, K. S., & Lee, Y. S. (2011). Application of silver nanoparticles for the control of *Colletotrichum* species in vitro and pepper anthracnose disease in field. *Mycobiology*, 39(3), 194–9.
- Larmond, E. (1977). *Laboratory Methods for Sensory Evaluation of Food*. Canada: Communication Branch, Agriculture.
- Laurila, E., & Ahvenainen, R. (2002). Minimal processing of fresh fruits and vegetables. In W. Jongen (Ed.), *Fruits and Vegetable Processing: Improving Quality* (pp. 288–309). Cambridge, England: Woodhead Publishing Limited.
- Layne, D. R. (1996). The pawpaw [*Asimina triloba* (L.) Dunal]: a new fruit crop for Kentucky and the United States. *HortScience*, 3(5), 777–784.
- Lazan, H., Ali, Z. M., Liang, K. S., & Yee, K. L. (1989). Polygalacturonase activity and variation in ripening of papaya fruit with tissue depth and heat treatment. *Physiologia Plantarum*, 77(1), 93–98.
- Lee, J. Y., Park, H. J., Lee, C. Y., & Choi, W. Y. (2003). Extending shelf-life of minimally processed apples with edible coatings and antibrowning agents. *LWT - Food Science and Technology*, 36(3), 323–329.
- Li, H., Li, F., Wang, L., Sheng, J., Xin, Z., Zhao, L., Xiao, H., Zheng, Y. & Hu, Q. (2009). Effect of nano-packing on preservation quality of Chinese jujube (*Ziziphus jujuba* Mill. var. *inermis* (Bunge) Rehd). *Food Chemistry*, 114(2), 547–552.
- Li, P., Li, J., Wu, C., Wu, Q., & Li, J. (2005). Synergistic antibacterial effects of β -lactam antibiotic combined with silver nanoparticles. *Nanotechnology*, 16, 1912–1917.
- Li, X.-F., Feng, X.-Q., Yang, S., Wang, T.-P., & Su, Z.-X. (2008). Effects of molecular weight and concentration of chitosan on antifungal activity against *Aspergillus niger*. *Iranian Polymer Journal*, 17(11), 843–852.
- Lin, Q.-B., Li, B., Song, H., & Wu, H.-J. (2011). Determination of silver in nano-plastic food packaging by microwave digestion coupled with inductively coupled plasma atomic emission spectrometry or inductively coupled plasma mass spectrometry. *Food Additives and Contaminants: Part A*, 28, 1123–1128.
- Liu, N., Chen, X.-G., Park, H.-J., Liu, C.-G., Liu, C.-S., Meng, X.-H., & Yu, L.-J. (2006). Effect of MW and concentration of chitosan on antibacterial activity of *Escherichia coli*. *Carbohydrate Polymers*, 64(1), 60–65.
- Liu, X. F., Guan, Y. L., Yang, D. Z., Li, Z., & Yao, K. De. (2000). Antibacterial action of chitosan and carboxymethylated chitosan. *Journal of Applied Polymer Science*, 79, 1324–1335.

- Lloret, E., Picouet, P., & Fernández, A. (2012). Matrix effects on the antimicrobial capacity of silver based nanocomposite absorbing materials. *LWT - Food Science and Technology*, 49(2), 333–338.
- Lurie, S. (1998). Review: postharvest heat treatments. *Postharvest Biology and Technology*, 14(3), 257–269.
- Madrigal, L. S., Ortiz, A. N., Cooke, R. D., & Fernandez, R. H. (1980). The dependence of crude papain yields on different collection (“Tapping”) procedures for papaya latex. *Journal of the Science of Food and Agriculture*, 31, 279–285.
- Maftoonazad, N., & Ramaswamy, H. S. (2005). Postharvest shelf-life extension of avocados using methyl cellulose-based coating. *LWT - Food Science and Technology*, 38(6), 617–624.
- Mahmud, T. M. M., Al Eryani-Raqeeb, A., Syed Omar, S. R., Mohamed Zaki, A. R., & Al Eryani, A.-R. (2008). Effects of different concentrations and applications of calcium on storage life and physicochemical characteristics of papaya (*Carica Papaya L.*). *American Journal of Agricultural and Biological Sciences*, 3(3), 526–533.
- Manenoi, A., & Paull, R. E. (2007). Papaya fruit softening, endoxylanase gene expression, protein and activity. *Physiologia Plantarum*, 131(3), 470–80.
- Manoto, E. C., & Zulkifly, S. (1994). The oriental fruitfly and its disinfection. In R. Md. Yon (Ed.), *Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN* (pp. 99–104). Kuala Lumpur: ASEAN Food Handling Bureau.
- Manrique, G. D., & Lajolo, F. M. (2004). Cell-wall polysaccharide modifications during postharvest ripening of papaya fruit (*Carica papaya*). *Postharvest Biology and Technology*, 33(1), 11–26.
- Manshardt, R. M. (1992). Papaya. In F. A. Hammerschlag & R. E. Litz (Eds.), *Biotechnology in Agriculture No. 8. Biotechnology of Perennial Fruit Crops* (pp. 489–511). Wallingford, UK: CAB International.
- Mantilla, N., Castell-Perez, M. E., Gomes, C., & Moreira, R. G. (2012). Multilayered antimicrobial edible coating and its effect on quality and shelf-life of fresh-cut pineapple (*Ananas comosus*). *LWT - Food Science and Technology*, 51(1), 37–43.
- Maqbool, M., Ali, A., Ramachandran, S., Smith, D. R., & Alderson, P. G. (2010). Control of postharvest anthracnose of banana using a new edible composite coating. *Crop Protection*, 29(10), 1136–1141.

- Martinez-Abad, A., Lagaron, J. M., & Ocio, M. J. (2012). Development and characterization of silver-based antimicrobial ethylene–vinyl alcohol copolymer (EVOH) films for food-packaging applications. *Journal of Agricultural and Food Chemistry*, *60*, 5350–5359.
- Martín-Belloso, O., Rojas-Graü, M. A. & Soliva-Fortuny, R. (2009). Delivery of flavor and active ingredients using edible films and coatings. In M.E. Embuscado and K.C. Huber (Eds.), *Edible Films and Coatings for Food Applications* (pp. 295-313). New York: Springer Science + Business Media, LLC.
- Md. Yon, R. (1994). Introduction. In R. Md. Yon (Ed.), *Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN* (pp. 1–4). Kuala Lumpur: ASEAN Food Handling Bureau.
- Mirhosseini, H., Tan, C. P., Hamid, N. S. A., & Yusof, S. (2008). Effect of Arabic gum, xanthan gum and orange oil on flavor release from diluted orange beverage emulsion. *Food Chemistry*, *107*, 1161–1172.
- Mirhosseini, H., Tan, C. P., Hamid, N. S. A., Yusof, S., & Chern, B. H. (2009). Characterization of the influence of main emulsion components on the physicochemical properties of orange beverage emulsion using response surface methodology. *Food Hydrocolloids*, *23*(2), 271–280.
- Mohammed Fayaz, A., Balaji, K., Girilal, M., Kalaichelvan, P. T., & Venkatesan, R. (2009). Mycobased synthesis of silver nanoparticles and their incorporation into sodium alginate films for vegetable and fruit preservation. *Journal of Agricultural and Food Chemistry*, *57*(14), 6246–52.
- Mohd. Ali, Z., Lizada, M. C. C., & Lazan, H. (1994). Ripening and senescence. In R. Md. Yon (Ed.), *Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN1* (pp. 56–74). Kuala Lumpur: ASEAN Food Handling Bureau.
- Montgomery, D. C. (2009). *Design and Analysis of Experiments*. Hoboken, New Jersey: John Wiley and Sons, Inc.
- Morillon, V., Debeaufort, F., Blond, G., Capelle, M., & Voilley, A. (2002). Factors affecting the moisture permeability of lipid- based edible films: a review. *Critical Reviews in Food Science and Nutrition*, *42*(1), 67–89.
- Morones, J. R., Elechiguerra, J. L., Camacho, A., Holt, K., Kouri, J. B., Ramírez, J. T., & Yacaman, M. J. (2005). The bactericidal effect of silver nanoparticles. *Nanotechnology*, *16*(10), 2346–53.
- Morton, J. F. (1987). Papaya. In J. F. Morton (Ed.), *Fruits of Warm Climates* (pp. 336–346). Miami, Fl: Creative Resource Systems, Inc.
- Nakasone, H. Y., & Paull, R. E. (1998). *Tropical Fruits (Crop Production Science in Horticulture No. 7)*. Wallingford, UK: CAB International.

- Nazeeb, M., & Broughton, W. J. (1978). Storage conditions and ripening of papaya "Bentong" and "Taiping." *Scientia Horticulturae*, 9, 265–277.
- Necas, J., & Bartosikova, L. (2013). Carrageenan : a review. *Veterinarni Medicina*, 58(4), 187–205.
- Nickols-Richardson, S. M. (2007). Nanotechnology: implications for food and nutrition professionals. *Journal of the American Dietetic Association*, 107(9), 1494–1497.
- Nishijima, K., Couey, H. M., & Alvarez, A. M. (1987). Internal yellowing, a bacterial disease of papaya fruits caused by *Enterobacter cloacae*. *Plant Disease*, 71(11), 1029–1034.
- Nishijima, K. ., Miura, C. K., Armstrong, J. W., Brown, S. A., & Hu, B. K. S. (1992). Effect of forced, hot-air treatment of papaya fruit on fruit quality and incidence of postharvest diseases. *Plant Disease*, 76, 723–727.
- Nisperos-carriedo, M. O., Baldwin, E. A., & Shaw, P. E. (1991). Development of and edible coating for extending postharvest life of selected fruits and vegetables. *Proceedings of the Florida State Horticultural Society*, 104, 122–125.
- No, H. K., Meyers, S. P., Prinyawiwatkul, W., & Xu, Z. (2007). Applications of chitosan for improvement of quality and shelf life of foods : a review. *Journal of Food Science*, 72, 87–100.
- Nunes, M. C. N., Emond, J. P., & Brecht, J. K. (2006). Brief deviations from set point temperatures during normal airport handling operations negatively affect the quality of papaya (*Carica papaya*) fruit. *Postharvest Biology and Technology*, 41(3), 328–340.
- Olivas, G. I., & Barbosa-Cánovas, G. (2009). Edible films and coatings for fruits and vegetables. In M. E. Embuscado & K. C. Huber (Eds.), *Edible Films and Coatings for Food Applications* (pp. 211–244). New York: Springer Science + Business Media, LLC.
- Özden, Ç., & Bayindirli, L. (2002). Effects of combinational use of controlled atmosphere, cold storage and edible coating applications on shelf life and quality attributes of green peppers. *European Food Research and Technology*, 214(4), 320–326.
- Pal, S., Tak, Y. K., & Song, J. M. (2007). Does the antibacterial activity of silver nanoparticles depend on the shape of the nanoparticle? A study of the Gram-negative bacterium *Escherichia coli*. *Applied and Environmental Microbiology*, 73(6), 1712–20.
- Paliyath, G., & Murr, D. P. (2008). Biochemistry of fruits. In G. Paliyath, D. P. Murr, A. K. Handa, & S. Lurie (Eds.), *Postharvest Biology and Technology of Fruits, Vegetables, and Flowers* (pp. 21–25). Maiden MA, USA: Wiley-Blackwell Publishing.

- Panáček, A., Kolár, M., Vecerová, R., Pucek, R., Soukupová, J., Krystof, V., Hamal, P., Zboril, R. & Kvítek, L. (2009). Antifungal activity of silver nanoparticles against *Candida* spp. *Biomaterials*, 30(31), 6333–40.
- Panacek, A., Kvítek, L., Pucek, R., Kolar, M., Vecerova, R., Pizúrova, N., Sharma, V. K., Navecna, T. & Zboril, R. (2006). Silver colloid nanoparticles: synthesis, characterization, and their antibacterial activity. *The Journal of Physical Chemistry. B*, 110(33), 16248–53
- Pantastico, E. B. (1975). *Postharvest Physiology, Handling and Utilization of Tropical and Subtropical Fruits and Vegetables*. Westport, USA: Avi Publishing Company, Inc.
- Park, H. J. (1999). Development of advanced edible coatings for fruits. *Trends in Food Science & Technology*, 10, 254–260.
- Paull, R. E. (1996). Ripening behavior of papaya (*Carica papaya* L.) exposed to gamma irradiation. *Postharvest Biology and Technology*, 7, 359–370.
- Paull, R. E., & Chen, N. J. (1983). Postharvest variation in cell wall-degrading enzymes of papaya (*Carica papaya* L.) during fruit ripening. *Plant Physiology*, 72, 382–385.
- Paull, R. E., & Chen, N. J. (1990). Heat shock response in field-grown , ripening papaya fruit. *Journal of the American Society for Horticultural Science*, 115(4), 623–631.
- Paull, R. E., & Chen, N. J. (2000). Heat treatment and fruit ripening. *Postharvest Biology and Technology*, 21(1), 21–37.
- Paull, R. E., & Duarte, O. (2011). *Tropical Fruits Volume 1*. Wallingford, UK: CAB International.
- Paull, R. E., Gross, K., & Qiu, Y. (1999). Changes in papaya cell walls during fruit ripening. *Postharvest Biology and Technology*, 16(1), 79–89.
- Paull, R. E., Irikura, B., Wu, P., Turano, H., Chen, N. J., Blas, A., Fellman, J. K., Gschwend, A. R., Wai, C. M., Yu, Q., Presting, G. Alam, M. & Ming, R. (2008). Fruit development, ripening and quality related genes in the papaya genome. *Tropical Plant Biology*, 1, 246–277.
- Paull, R. E., Nishijima, W., Reyes, M., & Cavaletto, C. (1997). Postharvest handling and losses during marketing of papaya (*Carica papaya* L.). *Postharvest Biology and Technology*, 11, 165–179.
- Pavlath, A. E., & Orts, W. (2009). Edible films and coatings: why, what and how? In M. E. Embuscado & K. C. Huber (Eds.), *Edible Films and Coatings for Food Applications* (pp. 1–23). New York: Springer Science + Business Media, LLC.

- Pereira, T., de Almeida, P. S. G., de Azevedo, I. G., da Cunha, M., de Oliveira, J. G., da Silva, M. G., & Vargas, H. (2009). Gas diffusion in “Golden” papaya fruit at different maturity stages. *Postharvest Biology and Technology*, 54(3), 123–130.
- Peretto, G., Du, W.-X., Avena-Bustillos, R. J., Berrios, J. D. J., Sambo, P., & McHugh, T. H. (2014). Optimization of antimicrobial and physical properties of alginate coatings containing carvacrol and methyl cinnamate for strawberry application. *Journal of Agricultural and Food Chemistry*, 62, 984–990.
- Perez-carrillo, E., & Yahia, E. M. (2004). Effect of postharvest hot air and fungicide treatments on the quality of “Maradol” papaya (*Carica Papaya* L.). *Journal of Food Quality*, 27(2004), 127–139.
- Persley, D. M., & Ploetz, R. C. (2003). Diseases of papaya. In R. C. Ploetz (Ed.), *Diseases of Tropical Fruit Crops* (pp. 373–412). Wallingford, UK: CAB International.
- Pinto, R. J. B., Almeida, A., Fernandes, S. C. M., Freire, C. S. R., Silvestre, A. J. D., Neto, C. P., & Trindade, T. (2013). Antifungal activity of transparent nanocomposite thin films of pullulan and silver against *Aspergillus niger*. *Colloids and Surfaces B: Biointerfaces*, 103, 143–8.
- Plotto, A., Narciso, J. a, Rattanapanone, N., & Baldwin, E. a. (2010). Surface treatments and coatings to maintain fresh-cut mango quality in storage. *Journal of the Science of Food and Agriculture*, 90(13), 2333–41.
- Ponce, A. G., Roura, S. I., Valle, C. E. D., & Moreira, M. R. (2008). Antimicrobial and antioxidant activities of edible coatings enriched with natural plant extracts: In vitro and in vivo studies. *Postharvest Biology and Technology*, 49, 294–300.
- Potara, M., Jakab, E., Damert, A., Popescu, O., Canpean, V., & Astilean, S. (2011). Synergistic antibacterial activity of chitosan-silver nanocomposites on *Staphylococcus aureus*. *Nanotechnology*, 22, 135101.
- Pranoto, Y., Salokhe, V. M., & Rakshit, S. K. (2005). Physical and antibacterial properties of alginate-based edible film incorporated with garlic oil. *Food Research International*, 38(3), 267–272.
- Prasanna, V., Prabha, T. N., & Tharanathan, R. N. (2007). Fruit ripening phenomena-an overview. *Critical Reviews in Food Science and Nutrition*, 47(1), 1–19.
- Preuss, H. G., Echard, B., Enig, M., Brook, I., & Elliott, T. B. (2005). Minimum inhibitory concentrations of herbal essential oils and monolaurin for Gram-positive and Gram-negative bacteria. *Molecular and Cellular Biochemistry*, (272), 29–34.
- Prucek, R., Tuček, J., Kilianová, M., Panáček, A., Kvítek, L., Filip, J., Kolár, M., Tománková, K. & Zbořil, R. (2011). The targeted antibacterial and antifungal properties of magnetic nanocomposite of iron oxide and silver nanoparticles. *Biomaterials*, 32(21), 4704–13.

- Qiu, Y., Nishina, M. S., & Paull, R. E. (1995). Papaya fruit Growth , Calcium Uptake , and Fruit Ripening. *Journal of the American Society for Horticultural Science*, 120(2), 246–253.
- Quezada-Gallo, J-A. (2009). Delivery of food additives and antimicrobials using edible films and coatings. In M.E. Embuscado and K.C. Huber (Eds.), *Edible Films and Coatings for Food Applications* (pp. 315-333). New York: Springer Science + Business Media, LLC.
- QWI-OF/17-41 (2011). Determination of metal and mineral in food and traditional medicine. In-House Method. ALS TECHNICHEM (M) Sdn. Bhd.
- Rabea, E. I., Badawy, M. E.-T., Stevens, C. V, Smagghe, G., & Steurbaut, W. (2003). Chitosan as antimicrobial agent: applications and mode of action. *Biomacromolecules*, 4(6), 1457–65.
- Radheshkumar, C., & Münstedt, H. (2006). Antimicrobial polymers from polypropylene/silver composites—Ag⁺ release measured by anode stripping voltammetry. *Reactive and Functional Polymers*, 66(7), 780–788.
- Radi, Z., Roperos, N. I., Nanthachai, S., Broto, W., Alim, J. H., & Huat, K. S. (1994). Status of the papaya industry. In R. Md. Yon (Ed.), *Papaya: Fruit Development, Postharvest Physiology, Handling and Marketing in ASEAN* (pp. 18–31). Kuala Lumpur: ASEAN Food Handling Bureau.
- Raffi, M., Hussain, F., Bhatti, T. M., Akhter, J. I., Hameed, A., & Hasan, M. M. (2008). Antibacterial Characterization of Silver Nanoparticles against *E. Coli* ATCC-15224. *Journal of Materials Science & Technology*, 24(2), 192–196.
- Rahman, M. A., Mahmud, T. M. M., Abdul Rahman, R., Kadir, J., & Begum, M. M. (2012). Potential co-application of *Burkholderia cepacia* , calcium and chitosan on enhancement of storage life and quality of papaya fruits. *Pertanika Journal of Tropical Agricultural Science*, 35(3), 439–458.
- Rahman, M. A., Mahmud, T. M. M., Kadir, J., Abdul Rahman, R., & Begum, M. M. (2008a). Antimicrobial activities of chitosan and calcium chloride on in vitro growth of *Colletotrichum gloeosporioides* from papaya. *Pertanika Journal of Tropical Agricultural Science*, 31(2), 223–232.
- Rahman, M. A., Mahmud, T. M. M., Kadir, J., Abdul Rahman, R., & Begum, M. M. (2008b). Major postharvest fungal diseases of *Papaya* cv . “ Sekaki ” in Selangor, Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 31(1), 27–34.
- Rai, M., Yadav, A., & Gade, A. (2009). Silver nanoparticles as a new generation of antimicrobials. *Biotechnology Advances*, 27(1), 76–83.
- Ranggana, S. (1977). *Manual of Analysis of Fruit and Vegetable Products*. New Delhi: McGraw-Hill.

- Ravishankar, R. V., & Jamuna, B. A. (2011). Nanoparticles and their potential application ad antimicrobials. In *Science Against Microbial Pathogens: Communicating Current Research and Technological Advances* (pp. 197–209). Spain: FORMATEX.
- Resende, E. C. O., Martins, P. F., Azevedo, R. A. De, Jacomino, A. P., & Bron, I. U. (2012). Oxidative processes during “ Golden ” papaya fruit ripening. *Brazilian Journal of Plant Physiology*, 24(2), 85–94.
- Ribeiro, C., Vicente, A. A., Teixeira, J. A., & Miranda, C. (2007). Optimization of edible coating composition to retard strawberry fruit senescence. *Postharvest Biology and Technology*, 44(1), 63–70.
- Rinaudo, M. (2006). Chitin and chitosan : Properties and applications. *Progress in Polymer Science*, 31, 603–632.
- Rojas-Graü, M. a., Raybaudi-Massilia, R. M., Soliva-Fortuny, R. C., Avena-Bustillos, R. J., McHugh, T. H., & Martín-Belloso, O. (2007). Apple puree-alginate edible coating as carrier of antimicrobial agents to prolong shelf-life of fresh-cut apples. *Postharvest Biology and Technology*, 45(2), 254–264.
- Rojas-Graü, M. A., Soliva-Fortuny, R., & Martín-Belloso, O. (2009). Edible coatings to incorporate active ingredients to fresh-cut fruits: a review. *Trends in Food Science & Technology*, 20(10), 438–447.
- Rojas-Graü, M. A., Tapia, M. S., & Martín-Belloso, O. (2008). Using polysaccharide-based edible coatings to maintain quality of fresh-cut Fuji apples. *LWT - Food Science and Technology*, 41(1), 139–147.
- Rojas-Graü, M. a., Tapia, M. S., Rodríguez, F. J., Carmona, a. J., & Martin-Belloso, O. (2007). Alginate and gellan-based edible coatings as carriers of antibrowning agents applied on fresh-cut Fuji apples. *Food Hydrocolloids*, 21(1), 118–127.
- Sancho, L. E. G., Yahia, E. M., Martínez-téllez, M. A., & González-aguilar, G. A. (2010). Effect of maturity stage of papaya maradol on physiological and biochemical parameters. *American Journal of Agricultural and Biological Sciences*, 5(2), 194–203.
- Sandhya. (2010). Modified atmosphere packaging of fresh produce: Current status and future needs. *LWT - Food Science and Technology*, 43(3), 381–392.
- Sankat, C. K., & Maharaj, R. (1997). Papaya. In S. K. Mitra (Ed.), *Postharvest Physiology and Storage of Tropical and Subtropical Fruits* (pp. 167–189). Wallingford, UK: CAB International.
- Saxena, A., Tripathi, R. M., Zafar, F., & Singh, P. (2012). Green synthesis of silver nanoparticles using aqueous solution of *Ficus benghalensis* leaf extract and characterization of their antibacterial activity. *Materials Letters*, 67(1), 91–94.

- Sebti, I., Martial-Gros, A., Carnet-Pantiez, A., Grelier, S., & Coma, V. (2005). Chitosan polymer as bioactive coating and film against *Aspergillus niger* contamination. *Journal of Food Science*, 70(2), 100–104.
- Seehanam, P., & Boonyakiat, D. (2010). Physiological and physicochemical responses of “Sai Nam Phueng” tangerine to commercial coatings. *HortScience*, 45(4), 605–609.
- Serry, N. K. . (2011). Postharvest handling of solo papaya fruits harvested at different maturity stages. *American-Eurasian Journal of Agriculture and Environmental Science*, 2, 205–210.
- Shackel, K. A., Greve, C., Labavitch, J. M., & Ahmadi, H. (1991). Cell turgor changes associated with ripening in tomato pericarp tissue. *Plant Physiology*, 97(2), 814–816.
- Shameli, K., Ahmad, M. Bin, Yunus, W. M. Z. W., Rustaiyan, A., Ibrahim, N. A., Zargar, M., & Abdollahi, Y. (2010). Green synthesis of silver/montmorillonite/chitosan bionanocomposites using the UV irradiation method and evaluation of antibacterial activity. *International Journal of Nanomedicine*, 5, 875–87.
- Shin, S. Y., Johari, S., Maheswary, V., & Kalsom, A. B. U. (2011). Isolation of fruit ripening genes from *Carica papaya* var . Eksotika 1 cDNA libraries. *Journal of Tropical Agriculture and Food Science*, 39(2), 1–9.
- Shit, S. C., & Shah, P. M. (2014). Edible polymers : challenges and opportunities. *Journal of Polymers*, 1–13.
- Shrivastava, S., Bera, T., Roy, A., Singh, G., Ramachandrarao, P., & Dash, D. (2007). Characterization of enhanced antibacterial effects of novel silver nanoparticles. *Nanotechnology*, 18(22), 225103.
- Siegrist, M., Stampfli, N., Kastenholz, H., & Keller, C. (2008). Perceived risks and perceived benefits of different nanotechnology foods and nanotechnology food packaging. *Appetite*, 51(2), 283–290.
- Singh, S. P., & Rao, D. V. S. (2005). Effect of modified atmosphere packaging (MAP) on the alleviation of chilling injury and dietary antioxidants levels in “Solo” papaya during low temperature storage. *European Journal of Horticultural Science*, 70(5), 246–252.
- Singh, S. P., & Rao, D. V. S. (2011). Papaya. In E. M. Yahia (Ed.), *Posrharvest Biology and Technology of Tropical and Subtropical Fruits Volume 4: Mangosteen to White Sapote* (pp. 86–124). Cambridge, UK: Woodhead Publishing Limited.
- Sondi, I., & Salopek-Sondi, B. (2004). Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram-negative bacteria. *Journal of Colloid and Interface Science*, 275(1), 177–182.

- Sothornvit, R., & Krochta, J. M. (2005). Plasticizers in edible films and coatings. In J. H. Han (Ed.), *Innovations in Food Packaging* (pp. 403–433). London, UK: Elsevier Academic Press.
- Sujiprihati, S., & Suketi, K. (2010). *Tanaman Betik*. Kuala Lumpur: Synergy Media.
- Tapia, M. S., Rojas-Graü, M. a., Carmona, A., Rodríguez, F. J., Soliva-Fortuny, R., & Martín-Belloso, O. (2008). Use of alginate- and gellan-based coatings for improving barrier, texture and nutritional properties of fresh-cut papaya. *Food Hydrocolloids*, 22(8), 1493–1503.
- Tecante, A., & Santiago, M. del C. N. (2012). Solution properties of κ -carrageenan and its interaction with other polysaccharides in aqueous media. In J. De Vicente (Ed.), *Rheology* (pp. 241–264). Croatia: InTech.
- Tee, E. S., Noor, M. I., Azudin, M. N., & Idris, K. (1997). *Nutrient Composition of Malaysian Foods*. Kuala Lumpur: Institute for Medical Research.
- Teixeira da Silva, J. A., Rashid, Z., Nhut, D. T., Sivakumar, D., Gera, A., Souza Jr, M. T., & Tennant, P. F. (2007). Papaya (*Carica papaya* L.) biology and biotechnology. *Tree and Forestry Science and Biotechnology*, 1(1), 47–73.
- Tortora, G. J., Funke, B. R., & Case, C. L. (2004). *Microbiology an Introduction*. San Francisco: Pearson/Benjamin-Cummings Publishing Company.
- Travan, A., Pelillo, C., Donati, I., Marsich, E., Benincasa, M., Scarpa, T., Paoletti, S. (2009). Non-cytotoxic silver nanoparticle-polysaccharide nanocomposites with antimicrobial activity. *Biomacromolecules*, 10, 1429–1435.
- Tucker, G. A. (1993). Introduction. In G. B. Seymour, J. E. Taylor, & G. A. Tucker (Eds.), *Biochemistry of Fruit Ripening* (pp. 1–51). London: Chapman & Hall.
- USDA National Nutrient Database for Standard Ref, Release 26. (2013). *United State Department of Agriculture (USDA), Agricultural Research Service. Nutrient Data Laboratory Home Page*. Retrieved from <http://www.ars.usda.gov>
- USEPA. (2012). Drinking Water Standards and Health Advisories. *United State Environmental Protection Agency (USEPA)*. Retrieved from <http://www.epa.gov>
- Vaclavik, V. A., & Christian, E. W. (2014). *Essentials of Food Science* (pp. 83–113). New York, NY: Springer New York.
- Valero, D., Díaz-Mula, H. M., Zapata, P. J., Guillén, F., Martínez-Romero, D., Castillo, S., & Serrano, M. (2013). Effects of alginate edible coating on preserving fruit quality in four plum cultivars during postharvest storage. *Postharvest Biology and Technology*, 77, 1–6.

- Vargas, M., Pastor, C., Chiralt, A., McClements, D. J., & González-Martínez, C. (2008). Recent advances in edible coatings for fresh and minimally processed fruits. *Critical Reviews in Food Science and Nutrition*, 48(6), 496–511.
- Vasilev, K., Sah, V. R., Goreham, R. V, Ndi, C., Short, R. D., & Griesser, H. J. (2010). Antibacterial surfaces by adsorptive binding of polyvinyl-sulphonate-stabilized silver nanoparticles. *Nanotechnology*, 21, 215102.
- Vyas, P. B., Gol, N. B., & Rao, T. V. R. (2014). Postharvest quality maintenance of papaya fruit using polysaccharide-based edible coatings. *International Journal of Fruit Science*, 14(1), 81–94.
- Wall, M. M. (2006). Ascorbic acid, vitamin A, and mineral composition of banana (*Musa sp.*) and papaya (*Carica papaya*) cultivars grown in Hawaii. *Journal of Food Composition and Analysis*, 19(5), 434–445.
- Wang, C. Y. (2010). Alleviation of chilling injury in tropical and subtropical fruits. *Acta Horticulturae*, 864, 267–274.
- Wang, D., Zhang, H., Wu, F., Li, T., Liang, Y., & Duan, X. (2013). Modification of pectin and hemicellulose polysaccharides in relation to aril breakdown of harvested longan fruit. *International Journal of Molecular Sciences*, 14, 23356–68.
- Wang, J., Wang, B., Jiang, W., & Zhao, Y. (2007). Quality and shelf life of mango (*Mangifera Indica* L. cv. 'Tainong') coated by using chitosan and polyphenols. *Food Science and Technology International*, 13(4), 317–322.
- Watada, A. E., Herner, R. C., Kader, A. A., Roman, R. J., & Staby, G. L. (1984). Terminology for the description of developmental stages of horticultural crops. *HortScience*, 19(1), 20–21.
- Wei, D., Sun, W., Qian, W., Ye, Y., & Ma, X. (2009). The synthesis of chitosan-based silver nanoparticles and their antibacterial activity. *Carbohydrate Research*, 344(17), 2375–2382.
- Wills, R. B. H., McGlasson, W. B., Graham, D., & Joyce, D. C. (2007). *Postharvest: An Introduction to the Physiology and Handling of Fruit, Vegetables and Ornamentals 5th Edition*. Wallingford, UK: CAB International.
- Woolf, A. B., & Lay-Yee, M. (1997). Pretreatments at 38°C of “Hass” avocado confer thermotolerance to 50°C hot water treatments. *HortScience*, 32(4), 705–708.
- Worrell, D. B., Sean Carrington, C. M., & Huber, D. J. (2002). The use of low temperature and coatings to maintain storage quality of breadfruit, *Artocarpus altilis* (Parks.) Fosb. *Postharvest Biology and Technology*, 25(1), 33–40.

- Yang, G., Yue, J., Gong, X., Qian, B., Wang, H., Deng, Y., & Zhao, Y. (2014). Blueberry leaf extracts incorporated chitosan coatings for preserving postharvest quality of fresh blueberries. *Postharvest Biology and Technology*, 92, 46–53.
- Yoksan, R., & Chirachanchai, S. (2010). Silver nanoparticle-loaded chitosan–starch based films: fabrication and evaluation of tensile, barrier and antimicrobial properties. *Materials Science and Engineering: C*, 30(6), 891–897.
- Younes, I., Sellimi, S., Rinaudo, M., Jellouli, K., & Nasri, M. (2014). Influence of acetylation degree and molecular weight of homogeneous chitosans on antibacterial and antifungal activities. *International Journal of Food Microbiology*, 185, 57–63.
- Yousef, A. E., & Carlsrom, C. (2003). *Food Microbiology: A Laboratory Manual*. New Jersey: John Wiley and Sons, Inc.
- Zhao, M., Moy, J., & Paul, R. E. (1996). Effect of gamma-irradiation on ripening papaya pectin. *Postharvest Biology and Technology*, 8, 209–222.
- Zhong, K., Lin, W., Wang, Q., & Zhou, S. (2012). Extraction and radicals scavenging activity of polysaccharides with microwave extraction from mung bean hulls. *International Journal of Biological Macromolecules*, 51(4), 612–617.
- Zuhair, R. A., Aminah, A., Sahilah, A. M., & Eqbal, D. (2013). Antioxidant activity and physicochemical properties changes of papaya (*Carica papaya* L . cv . Hongkong) during different ripening stage. *International Food Research Journal*, 20(4), 1653–1659.