

UNIVERSITI PUTRA MALAYSIA

CHEMICAL CONSTITUENTS OF Syzygium aquem (MYRTACEAE) AND Dysoxylum acuntangulum (MELIACEAE)

SITI NOOR KAMILAH BINTI HAJI MOHAMAD

FS 2014 93



CHEMICAL CONSTITUENTS OF Syzygium aquem (MYRTACEAE) AND Dysoxylum acuntangulum (MELIACEAE)



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science.

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of requirements for the Master of Science

CHEMICAL CONSTITUENTS OF Syzygium aquem (MYRTACEAE) AND Dysoxylum acuntangulum (MELIACEAE)

By

SITI NOOR KAMILAH BINTI HAJI MOHAMAD

February 2014

Chairman: Professor Mawardi Rahmani, PhD

Faculty : Science

Two plant species, *Syzygium aquem* and *Dysoxylum acutangulum* were phytochemically studied. The chemical studies on the leaves and twigs of *Syzygium aquem* involves extraction using three organic solvents of different polarity and isolation of compounds by using several chromatographic techniques including gravity column chromatography. The structures of the compounds were elucidated by various spectroscopic techniques including UV, IR, EIMS and NMR.

Detail study on the chloroform crude extract twig and leaves of *Syzygium aquem* afforded three compounds where one of the compounds was identified as a chalcone, 2',4'-dihyroxy-3',5'-dimethyl-6'-methoxychalcone (1), obtained as orange needle-shaped crystals. The other two compounds are urs-12-en-3 β -ol (30) and lup-20(29)-en-3 β -ol (31).

Chemical investigation on the hexane crude extract leaves and twigs of the plant have resulted a flavanone, 5-hydroxy-7-methoxy-6,8-dimethylflavanone (26), friedelin (28), and two phytosterols, stigmasterol (30) and β -sitosterol (32). A chrotacumine C (33) were isolated from root bark of *Dysoxylum acutangulum*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KANDUNGAN KIMIA DARIPADA Syzygium aquem DAN Dysoxylum acutangulum

Oleh

SITI NOOR KAMILAH BINTI HAJI MOHAMAD

Februari 2014

Pengerusi: Profesor Mawardi Rahmani, PhD

Fakulti: Sains

Dua jenis spesis tumbuhan, Syzygium aquem dan Dysoxylum acutangulum telah dikaji secara fitokimia. Kajian kimia ke atas daun dan ranting Syzygium aquem melibatkan pengestrakan yang menggunakan tiga pelarut organik yang berbeza kekutubannya dan pemencilan sebatian dengan menggunakan beberapa teknik kromatografi termasuklah kromatografi turus graviti. Struktur-struktur sebatian ini telah dikenalpasti dengan menggunakan pelbagai kaedah spektroskopi termasuk UV, IR, MS dan NMR.

Kerja pemencilan terhadap ekstrak bahan mentah kloroform daun dan rating tumbuhan itu telah menghasilkan tiga sebatian sebatian dimana salah satunya dikenalpasti sebagai kalkon, 2',4'-dihidrosi-3',5'-dimetil-6'-metoksikalkon (1), didapati dalam bentuk kristal oren. Dua lagi sebatian tersebut ialah urs-12-en-3 β -ol (30) dan lup-20(29)-en-3 β -ol (31). Kajian terperinci ke atas ekstrak bahan mentah hexane daun dan ranting telah menghasilkan 5-hidroksi-7-metoksi-6,8-dimetilflavanon (26), friedelin (28), dan dua fitosterol, stigmasterol (27) dan β -sitosterol (29). Satu krotakumine C (32) telah dipencilkan daripada kulit akar *Dysoxylum acutangulum*.

ACKNOWLEDGEMENTS

All praises to Allah, Lord of the Universe. Only with His grace and mercy that this thesis can be completed.

I wish to express my sincere thanks to my supervisor Prof. Dr. Mawardi Rahmani for his invaluable guidance, support and continuous encouragement throughout the course of this project.

My gratitude also goes to my supervisory committee Dr Intan Safinar Ismail and Prof. Dr. Amin Ismail for their support and comments. Financial support from Malaysian Government under (Mini Bajet) and UPM fundamental research are gratefully acknowledged.

Special thanks also go to my colleagues, Dr. Najihah Hashim, Ms. Maizatulakmal, Mrs. Winda Oktima, Mrs. Kartinee, Mr. Aizat, Mrs. Nadiah, Mr. Nazil and Ms. Faiqah for sharing stress, laughter, and useful advice during this project. Special thanks are extended to staff of the Chemistry Department of UPM especially Mr. Johadi, Ms.Shareena, Mr. Fadzli, Mr. Zainal Abidin Kassim, and Mrs. Rusnani Aminuddin for their professional assistance in obtaining mass, IR, and NMR spectra of compounds.

My deepest gratitude to my parent and siblings for their prayers and moral support and to my husband, my deepest love for your patience and understanding.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mawardi Rahmani, PhD

Professor Faculty of Science Universiti Putra Malaysia (Chairman)

Intan Safinar Ismail, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Member)

Amin Ismail, PhD

Professor
Faculty of Medical and Health Science
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

Declaration by graduate students

I hereby confirm that:

- this thesis is my original work;
- quotation, illustrations and citation have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic) form including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Puta Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:		Date:	
Name and Matric No:			_

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	
Name of	
Chairman of	
Supervisory	
Committee:	
Signature:	
Name of	
Member of	
Supervisory	
Committee:	
Signature:	
Name of	
Member of	
Supervisory	
Committee:	
Signature:	
Name of	
Member of	
Supervisory	
Committee:	

TABLE OF CONTENTS

ABS	TRACT	Page ii
ABS	TRAK	iii
ACK	KNOWLEDGEMENTS	iv
APP	PROVAL	v
DEC	CLARATION	vii
	Γ OF TABLES	ix
	T OF FIGURES	X
LIST	T OF ABBREVIATIONS	xii
CHA	APTER	
I.	INTRODUCTION	1
	Objectives of Study	6
II.	LITERATURE REVIEW	
	Botany of Plants Studied	
	The Genus of Syzygium	7
	The Species of Syzygium aquem	7
	Phytochemical Literature of Syzygium species	8
	Bioassay of Isolated compounds of Syzygium	10
	The Family of Meliaceae	10
	The Genus of Dysoxylum	10
	The Species of Dysoxylum acutangulum	11
	Phytochemical Literature of Dysoxylum species	12
	Bioassay of Isolated compounds of Dysoxylum	15
III.	MATERIALS AND METHOD	
	General Experimental Methods	
	Plant materials and extraction	16
	Instruments	16
	Chromatography	
	Extraction and Isolation of Compounds from Syzygium aqueum	
	And Dysoxylum acutangulum	
	Syzygium aqueum	• 0
	Isolation of 5-Hydroxy-7-methoxy-6,8-dimethylflavanone (26)	20
	Isolation of Stigmasterol (27)	20
	Isolation of Friedelin (28)	21
	Isolation of β –Sitosterol (29)	21
	Isolation of 2',4'-Dihydroxy-3',5'-dimethyl-6'-	24
	methoxychalcone (1)	24
	Isolation of Urs-12-β-ol (30) Isolation of Lup-20(29)-en-3β-ol (31)	24 25
	Dysoxylum acutangulum	23
	Isolation of Chrotacumine C (33)	26
		∠0

IV.	RESULTS AND DISCUSSION	
	Characterization and isolated compounds from the Leaves and	27
	Twigs of Syzygium aquem	
	5-Hydroxy-7-methoxy-6,8-dimethylflavanone (26)	29
	Stigmasterol (27)	40
	Friedelin (28)	45
	Urs-12-en-3β-ol (30)	50
	β-Sitosterol (29)	54
	2',4'-Dihydroxy-3',5'-dimethyl-6'-methoxychalcone (1)	59
	Lup-20(29)-en-3β-ol (31)	68
	Characterization and isolated compounds from <i>Dysoxylum acutangulum</i>	78
	Chrotacumine C (32)	79
V.	CONCLUSIONS	89
BIBL	LIOGRAPHY	
BIOI	DATA OF STUDENT	

LIST OF TABLES

Table		Page
4.1	¹ H NMR (500 MHz, CDCl ₃) and ¹³ C NMR (100MHz, CDCl ₃) spectral data of 5-hydroxy-7-methoxy- 6, 8-dimethylflavanone	30
4.2	(26) ¹ H NMR (500 MHz, CDCl ₃) and ¹³ C NMR (100 MHz, CDCl ₃) spectral data of stigmasterol (27)	41
4.3	¹ H NMR (500 MHz, CDCl ₃) and ¹³ C NMR (100 MHz, CDCl ₃) spectral data of friedelin (28)	46
4.4	¹ H NMR (400 MHz, CDCl ₃) and ¹³ C NMR (100 MHz, CDCl ₃) spectral data of β-sitosterol (29)	55
4.5	¹ H NMR (400 MHz, CDCl ₃) and ¹³ C NMR (100 MHz, CDCl ₃) spectral data of 2',4'-dihydroxy-3',5'-dimethyl-6'-methoxychalcone (1)	61
4.6	¹ H NMR (400 MHz, CDCl ₃) and ¹³ C NMR (100 MHz, CDCl ₃) spectral data of lup-20(29)-en-3β-ol (31)	69
4.7	¹ H NMR (500 MHz, CDCl ₃) and ¹³ C NMR (100 MHz, CDCl ₃) spectral data of chrotacumine-C (32)	80
4.8	¹ H NMR (500 MHz, CDCl ₃) and ¹³ C NMR (100 MHz, CDCl ₃) spectral data of chrotacumine-C (32), COSY and HMBC	81

LIST OF FIGURES

Figure		Page
1.1	The Fruits of Syzygium aquem	3
1.2	The Buds of Syzygium aquem	3 3 5
1.3	The Tree of Syzygium aquem	3
1.4	The Tree of <i>Dysoxylum acutangulum</i>	
1.5	The Leaves of Dysoxylum acutangulum	5
1.6	The Barks of <i>Dysoxylum acutangulum</i>	5
3.1	Compounds obtained from hexane crude extract twigs and	19
	leaves of Syzygium aquem	
3.2	Compounds obtained from chloroform crude extract twigs and	23
	leaves of Syzygium aquem	
4.1	Compounds obtained from twigs and leaves of Syzygium aquem	28
4.2	EI mass spectrum of 5-hydroxy-7-methoxy- 6,8-	31
	dimethylflavanone (26)	
4.3	IR spectrum of 5-hydroxy-7-methoxy- 6,8-dimethylflavanone	31
	(26)	
4.4	¹ H-NMR spectrum of 5-hydroxy-7-methoxy- 6,8-	32
	dimethylflavanone (26)	
4.4a	Expanded ¹ H-NMR spectrum of 5-hydroxy-7-methoxy-6,8-	33
	dimethylflavanone (26)	
4.4b	Expanded ¹ H-NMR spectrum of 5-hydroxy-7-methoxy-6,8-	34
	dimethylflavanone (26)	
4.4c	Expanded ¹ H-NMR spectrum of 5-hydroxy-7-methoxy-6,8-	35
	dimethylflavanone (26)	
4.5	¹³ C-NMR spectrum of 5-hydroxy-7-methoxy-6,8-	36
	dimethylflavanone (26)	
4.6	HMQC-NMR spectrum of 5-hydroxy-7-methoxy-6,8-	37
	dimethylflavanone (26)	
4.7	COSY-NMR spectrum of 5-hydroxy-7-methoxy-6,8-	38
	dimethylflavanone (26)	
4.8	HMBC-NMR spectrum of 5-hydroxy-7-methoxy-6,8-	39
	dimethylflavanone (26)	
4.9	EI mass spectrum of stigmasterol (27)	42
4.10	IR spectrum of stigmasterol (27)	42
4.11	¹ H-NMR spectrum of stigmasterol (27)	43
4.12	¹³ C-NMR spectrum of stigmasterol (27)	44
4.13	EI mass spectrum of friedelin (28)	47
4.14	IR spectrum of friedelin (28)	47
4.15	¹ H-NMR spectrum of friedelin (28)	48
4.16	¹³ C-NMR spectrum of friedelin (28)	49
4.17	EI mass spectrum of urs-12-ene-3β-ol (30)	51
4.18	IR spectrum of urs-12-en-3β-ol (30)	51
4.19	Mass fragmentation patterns of urs-12-en-3β-ol (30)	52
4.20	¹ H NMR spectrum of urs-12-en-3β-ol (30)	53
4.21	IR spectrum of β-sitosterol (29)	56
4.22	EI mass spectrum of β-sitosterol (29)	56
4.23	¹ H NMR spectrum of β-sitosterol (29)	57
4.24	¹³ C NMR spectrum of β-sitosterol (29)	58

4.25	IR spectrum of 2',4'-dihydroxy-3',5'-dimethyl-6'-	62
	methoxychalcone (1)	
4.26	EI mass spectrum of 2',4'-dihydroxy-3',5'-dimethyl-6'-	62
	methoxychalcone (1)	
4.27	¹ H NMR spectrum of 2',4'-dihydroxy-3',5'-dimethyl-6'-	63
	methoxychalcone (1)	
4.28	¹³ C NMR spectrum of 2',4'-dihydroxy-3',5'-dimethyl-6'-	64
	methoxychalcone (1)	
4.29	DEPT NMR spectrum of 2',4'-dihydroxy-3',5'-dimethyl-6'-	65
	methoxychalcone (1)	
4.30	HMBC NMR spectrum of 2',4'-dihydroxy-3',5'-dimethyl-6'-	66
	methoxychalcone (1)	
4.31	HSQC NMR spectrum of 2',4'-dihydroxy-3',5'-dimethyl-6'-	67
	methoxychalcone (1)	
4.32	EI mass spectrum of lup-20(29)-en-3β-ol (31)	70
4.33	IR spectrum of lup-20(29)-en-3β-ol (31)	70
4.34	H NMR spectrum of lup-20(29)-en-3β-ol (31)	71
4.34a	Extended ¹ H NMR spectrum of lup-20(29)-en-3β-ol (31)	72
4.34b	Extended ¹ H NMR spectrum of lup-20(29)-en-3β-ol (31)	73
4.35	¹³ C NMR spectrum of lup-20(29)-en-3β-ol (31)	74
4.35a	Extended ¹³ C NMR spectrum of lup-20(29)-en-3β-ol (31)	75
4.36	DEPT NMR spectrum of lup-20(29)-en-3β-ol (31)	76
4.37	HMBC NMR spectrum of lup-20(29)-en-3β-ol (31)	77
4.38	¹ H NMR spectrum of chrotacumine-C (32)	82
4.39a	Extended ¹ H NMR spectrum of chrotacumine-C (32)	83
4.39b	Extended ¹ H NMR spectrum of chrotacumine-C (32)	84
4.40	¹³ C NMR spectrum of chrotacumine-C (32)	85
4.41	¹ H- ¹ H COSY NMR spectrum of chrotacumine-C (32)	86
4.42	HMQC NMR spectrum of chrotacumine-C (32)	87
4.43	HMBC NMR spectrum of chrotacumine-C (32)	88

LIST OF ABBREVIATIONS

 $\alpha \hspace{1cm} Alpha$

 β Beta

δ Chemical shift in ppm

γ Gamma

μg Micro gram

brs Broad singlet

¹³C Carbon-13

CHCl₃ Chloroform

CDCl₃ Deuterated chloroform

CD₃OD Deuterated methanol

COSY Correlated Spectorscopy

d Doublet

dd Doublet of doublet

DEPT Distortionless Enhancement by Polarization Transfer

DMSO Dimethylsulfoxide

dt Doublet of triplet

EtOAc Ethyl acetate

EtOH Ethanol

EI-MS Electron Ionization Mass Spectroscopy

g Gram

GC Gas Chromatography

GC-MS Gas Chromatography- Mass Spectroscopy

¹H Proton

HMBC Heteronuclear Multiple Bond Connectivity by 2D Multiple

Quantum

HPLC High Performance Liquid Chromatography

HR-MS High Resolution Mass Spectroscopy

HSQC Heteronuclear Single Quantum Coherence

Hz Hertz

IR Infra Red

J Coupling constants in Hz

L Litre

m Multiplet

(CD₃)₂CO Deuterated Acetone

MeOH Methanol

m.p Melting point

MS Mass Spectrum/Spectra/Spectrometer

NMR Nuclear Magnetic Resonance

ppm Parts per million

s Singlet

t Triplet

TLC Thin Layer Chromatography

UV Ultra Violet

WHO World Health Organization

CHAPTER I

INTRODUCTION

1.1 General Introduction

Organic compounds are found in various fungi, micro-organism, plants or animals. Some of these compounds can be found in many different organisms. Sometimes these compounds can only be found in specific species only. A natural product is a study on organic compounds and considered as a major driving force in development of organic chemistry and medicinal chemistry. Plants that are used in traditional medicine provide some of the first prototype drugs used clinically in the treatment of wide variety of diseases. The need to purify natural products from complex mixtures and to determine their structures have led to the development of more sophisticated methods for separations of compounds and structural analysis by chemical, and spectroscopic procedures. Some of the organic compounds are useful for biological activity and perform various functions in nature.

According to the World Health Organization (WHO), as much as 80 % of the world's population depends on traditional medicine for their primary health care needs (Cordell, 1995). The major part of traditional therapy involves the use of plants extracts. Today, traditional medicine still remains a popular method of treatment. Plants are almost exclusive source of medicine for the majority of the world populations, especially in the developing countries. No accurate data are available to assess the value and extent of the use of plants or active principles derived from them in the health care systems throughout the world.

Over half of all deaths in the United States are caused by diseases of the heart and malignant neoplasm as stated by the Centers of Disease Control and Prevention (Minino et al., 2006). A diet which is high in vegetables and fruits suggested by epidemiological evidences are linked to a reduced incidence of heart disease, cancer, and some neurodegenerative disorders (Arts and Hollman, 2005).

Mammalian system naturally produced reactive oxygen species (ROS) as a result of oxidative metabolism. Reactive oxygen species damage cell membranes and DNA, cancerous mutation, and the oxidation of low-density lipoprotein are major factors in the promotion of heart disease. In chronic inflammatory disease, as well as the etiology of cancers and heart disease, inflammation plays a major factor (Hu and Willett, 2002). Endogenous antioxidants can balance oxidative damage. Nutritive and non-nutritive elements from food provided are one of the additional protections for disease chemoprevention.

According to the (Art and Hollman, 2005) in the prevention of oxidative and inflammatory disease, colourful fruits are a potentially rich source of many dietary phenolic antioxidants and are believed to play an important role. Anthocyanins pigments contain in the fruit colour plants are responsible for many of the bright fruits and flowers

colours. It acts as a strong antioxidants and anti-inflammatories, with antimutagenic and cancer chemopreventative activities (Reynertson et al., 2006).

1.2 The Family of Myrtaceae

The plant family Myrtaceae is a pan-tropical plant and widely found in South America, Southeast Asia, and Australia. The plants in the family can generate economic returns for these countries in food production, agricultural crops, and ornamentals, such as the Mediteranean genus *Myrtus* (myrtle), spices such as clove (*Syzygium aromaticum*) and bay rum (*Pimenta racemosa*). Another example of fruit plants under this family are *Psidium* (guavas), *Myrciaria*, *Eugenia*, *Syzygium*, *Plinia* and *Luma*.

Syzygium species are known for their medicinal properties. Syzygium aqueum is the watery rose apple or water apple. It is originated from south of India. It is still grown wild in India and parts of Malaysia. The fruits have an uneven shape, being wider at the apex than base. The colour varies from white to bright pink. It is, crispy and watery flesh makes a good thirst quencher. Although they have a high water content, their skins are full of fruit sugars and vitamin A. Other names include: 'jambu air' (Malaysia/Indonesia) and 'tambis' (Philippines). The samples of Syzygium aqueum for this research were collected from Bandar Baru Bangi, Selangor.

In this study, twigs and leaves of syzygium aqueum was investigated in detail. The purposes of this research are to extract and identify the chemical constituents of twigs and leaves syzygium aquem species using chromatographic and modern spectroscopic methods. Some members of this genus have been investigated or studied in detail, however, syzygium aquem have not been well studied especially twigs and leaves parts. Thus, there is a need to identify bioactive compounds from this genus for effective drug development (Chattopadhayay et al., 1998).



Figure 1.1: The Fruits of Syzygium aquem



Figure 1.2: The Buds of Syzygium aquem



Figure 1.3: The Tree of Syzygium aquem

1.3 The Family of Meliaceae

This family comprises of 51 genera and about 575 species of trees and shrubs, native to tropical and subtropical regions also known as mahogany family of flowering plants. Most members of the family have fruit which are fleshy and coloured or leathery and also have large compound leaves. The leaflets arranged in the form of a feather, and branch flower clusters.

The bark of dysoxylum acutangulum was chosen as the second plant for this research. The plant was collected in Terengganu, Malaysia. In Sumatera, Indonesia seeds of Dysoxylum acutangulum have been traditionally used as fish-poison. The active principles of this plant have been investigated by monitoring the toxicity against a species of fish, Oryzias latipes, with the isolation of a phenolic sesquiterpene as a major toxic constituent. The compound showed a significant fish- toxicity against Oryzias latipes at 5 ppm concentration and moderate antibacterial activity against gram-positive bacteria, such as Staphylococcus aureus, Candida albicans and Trichophyton mentagrophytes at 5-20 ppm (MIC), but it is ineffective against Gram-negative bacteria, such as Esherichia coli or Pseudomonas aeruginosa (Nishizawa et al., 1983).



Figure 1.4: The Tree of Dysoxylum acutangulum



Figure 1.5: The Leaves of Dysoxylum acutangulum



Figure 1.6: The Barks of Dysoxylum acutangulum

1.4 Objectives of study

- 1. To extract and isolate the chemical constituents of the twigs and leaves of *Syzygium aquem* and bark of *Dysoxylum acutangulum*.
- 2. To identify and elucidate the structure of the compounds by using spectroscopic methods.



BIBLIOGRAPHY

- Aalbersberg, W., Y. Singh. (1991). Phytochemistry, 30, 921-926.
- Andersson Dunstan, C., Norren, Y., Serrano, G., Cox, P. A., Perera, P., & Bohlin, L. (1997). Evaluation of some Samoan and Peruvian medicinal plants by prostaglandin biosynthesis and rat ear oedema assays. *Journal of Enthnopharmacology*, 57(1), 35-36.
- Amin, I., & Tan, S. H. (2002). Antioxidant activity of selected seaweeds. *Malaysian Journal of Nutrition*, 8, 167-177.
- Amin, I., & Mukhrizah, O. (2006). Antioxidant capacity of methanolic and water extracts prepared from food-processing by products. *Journal of the Science of Food and Agriculture*, 86, 778-784.
- Arts, I. C. W., & Hollman, P. C. H. (2005). Polyphenols and disease risk in epidemiologic studies. *American Journal of Clinic Nutrition*, 81(1), 3175-3255.
- Amor, E. C., Villasenor, I. M., Ghayur, M. N., Gilani, A. H., & Choudhary, M. I. (2005). Spasmolytic flavonoids from *Syzygium samarangense* (Blume) merr. And I.M. Perry. *Zeitschrift fuer Naturforschung, C: Journal of Biosciences*, 60(1/2), 67-71.
- Amor, E. C., Villasenor, I. M., Yasin, A., & Choudhary, M. I. (2004). Prolyl endopeptidase inhibitors from *Syzygium samarangense* (Blume) merr. and I.M. Perry. *Zeitschrift fuer Naturforschung, C: Journal of Biosciences*, 59(1/2), 86-92.
- Bhatia, I. S., & Bajaj, K. L. (1975). Chemical constituents of the seeds and bark of *Syzygium cumini*. *Planta Medica*, 28(4), 346-352.
- Bhatia, I. S., Baja, K. L., & Ghangas, G. S. (1971). Tannins in black plum seeds. *Phytochemistry*, 10(1), 219-220.
- Buckingham, J., Macdonald, F.M. and Bradley, H.M. (1994). *Dictionary of Natural Product*, Vol. 4, 2587. London: Chapman & Hall.
- Cai, Y.Z., Sun, M., Jie, X., Luo, Q., & Corke, H. (2006). Structure radical scavenging activity relationships of phenolic compounds from traditional Chinese medicinal plants. *Life Sciences*. 78(25), 2872-2888.
- Chattopadhayay, E. D., Sinha, B. K., & Vaid, L. K. (1998). Antibacterial activity of syzygium species. Fitoterapia, 119(4), 365-367.
- Cordell, G.A., (1995) Changing Strategies in Natural Products Chemistry. *Phytochemistry*, 40, 1585-1612.

- Connolly, J.D. and Hill, R.A. (1991). Triterpenoids. In *Methods in plant biochemistry*, ed. P.M. Dey, and J.B. Harborne, p 353. London: Academic Press Limited.
- Diaz D., P. P., Arias, C. T. and Joseph-Nathan, P. (1987). *Phytochemistry*, 26, 809.
- Gafner, S., Wolfender, J.-L., Mavi, S., & Hostettmann, K. (1996). Antifungal and antibacterial chalcones from *Myrica serrate*. *Planta Medica*, 62(1), 67-69.
- Ghayur, m. N., Gilani, A. H., Khan, A., Amor, E. C., Villasenor, I.M. & Choudhary, M. I. (2006). Presence of calcium antagonist activity explains the use of *Syzygium samarangense* in diarrhea. *Phytotherapy Research*, 20(1), 49-52.
- Go, M. L., Wu, X., & Liu, X. L. (2005). Chalcones: An update on cytotoxic and chemoprotective properties. *Current Medicinal Chemistry*, 12(4), 483-499.
- H. Liu.; N. Qiu.; H. Ding.; & R. Yao. (2008). Polyphenols content and antioxidant capacity of 68 Chinese herbals suitable for medical or food uses. *Food Research International*, 41, 363-370.
- He, K., T. Barbara, A.J. Aladesanmi, Z. Lu, (1996). *Phytochemistry*, 35(6), 1455-1456.
- Hill, R.A., Kirk, D.N., Makin, H.L.J. and Murphy, G.M. (1991). *Dictionary of Steroids*. Great Britain: Chapman & Hall.
- Hu, F. B., & Willett, W. C. (2002). Optimal diets for prevention of coronary heart disease. *Journal of the American Medical Association*, 288(20), 2569-2578.
- Holland, H. L., Diakow, P. R. P., and Taylor, G. J. (1978). ¹³C Nuclear Magnetic Resonance Spectra of some C-19 hydroxy, C-5,6,epoxy and C-24 ethyl steroids. *Canada Journal. Chemistry*, 56, 3121-3127.
- Ismail, I.S.; Nagakura, Y.; Hirasawa, Y.; Hosoya, T.; Lazim, M.I.M; Lajis, N. H.; Morita, H.(2009). Chrotacumine A-D, Chromone Alkaloids from *Dysoxylum acutangulum. Journal Natural Product*, 72, 1879-1883.
- Jung, H.A.; Su, B.N.; Keller, W.J.; Mehta, R.G.; Kinghorn, A.D. Antioxidant xanthones from
- pericarp of Garcinia mangostana (Mangosteen). J. Agric. Food Chem. 2006, 54, 2077-2082.
- Kashiwada, Y., T. Fujioka, J.J. Chen, K. Mihashi, K.H. Lee, J. (1992). *Journal Organic Chemistry*, 57(1), 6946-6953.
- Kähkönen, M., Hopia, A., Vuorela, H., Rauha, J., Pihlaja, K., Kujala, T., & Heinonen, M. (1999). Antioxidant activity of plant extracts containing phenolic compounds. *Journal of Agricultural and Food Chemistry*, 47, 3954-3962.

- Kähkönen, M., Hopia, A. I., & Heinonen, M. (2001). Berry phenolics and their antioxidant activity. *Journal of Agricultural and Food Chemistry*, 49(8), 4076-4082.
- Kirtikar, K. R., & Basu, B. D. (1988). *Indian Medicinal Plants*. Dehradun: International Book Distributors.
- Klass, J., Tinto, W. F., Mclean, S. and Reynolds, W.F. 1992. Friedelane triterpenoids from *Peritassa compta:* Complete ¹H and ¹³C assignments by 2D NMR spectrocospy. *Journal of Natural products*, 55, 1626-1630.
- Kuo, Y,-C., Yang, L.-M., & Lin, L.-C. (2004). Isolation and inmunomodulatory effects of flavonoids from *Syzygium samarangense*. *Planta Medica*, 70(1), 1237-1239.
- Lakshmi, V., K. Pandey, S.K. Agarwal. (2009). Bioactivity of the compounds in genus *Dysoxylum*. *Acta Ecologica Sinica*, 29, 30-44.
- Lee, J.-H., Jung, H. S., Giang, P. M., jin, X., Lee, S., Son, P. T., et al. (2006). Blockade of nuclear factor-κb signaling pathway and anti-inflammatory activity of cardamomin, a chalcone
- analog from *Alpinia conchigera*. *Journal of Pharmacology and Experimental Therapeutics*, 316(1), 271-278.
- Li, T. S. C.; Wang, L. C. H (1998). Physiological components and health effects of ginseng, Echinacea, and sea buckthorn. In *Functional Foods: Biochemical and Processing Aspects;* Mazza, G., Ed.; Technomic Publishing: Lancaster, PA,; pp 239-256.
- Luo, X.D., S.H. Wu, D.G. Wu Y.B. MA, S.H. Qi, (2002). Tetrahedron, 58, 6691-6695.
- Malterud KE. Anthonsen T, Lorentzen GB. (1977). Two New C-methylated flavonoids from Myrica gale. *Phytochemistry* 16: 1805-1809.
- Mahmoud, I. I., Marzouk, M. S. A., Moharram, F. A., El-Gindi, M. R., & Hassan, A. M. K. (2001). Acylated flavonol glycosides from *Eugenia jambolana* leaves. *Phytochemistry*, 58(8), 1239-1244.
- Minino, A. M., Heron, M. P., & Smith, B. L. (2006). Deaths: Preliminary data for 2004. *National Vital Statistics Reports*, 54(19), 1-49.
- Morton, J. (1987). Fruits of Warm Climates. Winterville, NC: Julia Morton.
- Naik, R.J., S.L. Kattige, S.V. Bhat, B. Alreja, N.J. de Souza, R.H. Rupp, (1988). *Tetrahedron*, 44(7), 2081-2086.
- Nair, A. G. R., Krishnan, S., Ravikrishnan, C., & Madhusudanan, K. P. (1999). New and rare flavonol glycosides from leaves of *Syzygium samarangense*. *Fitoterapia*, 70(2), 148-151.

Nishizawa, M.; Inoue, A.; Sastrapradija, S.; Hayashi, Y. *Phytochemistry* 1983, 22, 2083-2085.

Nonaka, G.-i., Aiko, Y., Aritake, K., & Nishioka, I. (1992). Tanins and related compounds. CXIX. Samarangenins A and B, novel proanthocyanidins with doubly bonded structures from *Syzygium samarangense* and *S. aqueum. Chemical and Pharmaceutical Bulletin*, 40(10), 2671-2673.

Okuda, T., Yoshida, T., Hatano, T., Yazaki, K., & Ashida, M. (1982). Ellagitannins of the Casuarinaceae, Stachyuraceae, and Myrtaceae. *Phtochemistry*, 21(12), 2871-2874.

Patra, A., Chadury, S.K. and Rubgger, H. 1990. Complete ¹³C and ¹H spectral assignments of friedelin by INADEQUATE and Heteronuclear (¹³C-¹H) correlation experiments. *J. Indian Chem. Soc.* 67, 394-397.

Rahman, I., & Adcock, I. M. (2006). Oxidative stress and redox regulation of lung inflammation in COPD. *European Respiratory Journal*, 28(1), 219-242.

Reynertson, K.A., Basile, M.J., & Kennelly, E. J. (2005). Antioxidant potential f seven Myrtaceous fruits. *Enthnobotany Research & Applications*, 3, 25-35.

Reynertson, K. A., Wallace, A.M., Adachi, S., Gil, R. R., Yang, H., Basile, M. J., D'Armiento, J., Weinstein, I. B., & Kenelley, E. J. (2006). Bioactive depsides and anthocyanins from jaboticaba (*Myrciaria cauliflora*). *Journal of Natural Products*, 69(8), 1228-1230.

Resurreccion-Magno, M. H. C., Villasenor, I. M., Harada, N., & Monde, K. (2005). Antihyperglycaemic flavonoids from *Syzygium samarangense* (Blume) merr. and perry. *Phytotherapy Research*, 19(3), 246-253.

Schwartz, J.J. and All, M.E. (1955). Isolation of the sterols of the white potato. *J. Am. Chem. Soc*, 77, 5442-5443.

Seeram, N.P., Lee, R., scheuller, H.S., & Heber, D. (2006). Identification of phenolic compounds in strawberries by liquid chromatography electrospray ionization mass spectroscopy. *Food Chemistry*, 97, 1-11.

Simirgiotis M.J., et al., (2008). Cytotoxic chalcones and antioxidants from the fruits of *Syzygium samarangense* (Wax Jambu). *Food Chemistry*, 107, 813-819.

Singh, S., H.S. Garg, N.M. Khanna, (1976). *Phytochemistry*, 15(12), 2001-2002.

Singhakumara, B.M.P., Harshi K. Gamage, Mark S. Ashton, (2003). Forest Ecology and Management, 174, 511-520.

- Srivastava, R., Shaw, A. K., & Kulshreshtha, D.K. (1995). Triterpenoids and chalcone from *Syzygium samarangense*. *Phytochemistry*, 38(3), 687-689.
- Tatsuzaki, J., Bastow, K. F., Nakagawa-Goto, K., Nakamura, S., Itokawa, H., & Lee, K.-H. (2006). Dehydrozingerone, chalcone, and isoeugenol analogues as in vitro anticancer agents. *Journal of Natural Products*, 69(10), 1445-1449.
- Trakoontivakorn, K., Nakahara, H., Shinmoto, M., Takenaka, M., Onishi-Kameyama, H., Ono, M., et al. (2001). Structural analysis of a novel antimutagenic compound, 4-hydroxypanduratin a, and the antimutagenic activity of flavonoids in a Thai spice, fingerroot (*Boesenbergi pandurata* Schult.) against mutagenic heterocylic amines. *Journal of Agricultural and Food Chemistry*, 49(6), 3046-3050.
- Velioglu, Y. S., Mazza, G., Gao, L., & Oomah, B. D. (1998). Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. *Journal of Agricultural and Food Chemistry*, 46, 4113-4117.
- Wang, H., Nair, M.G., Strasburg, G. M., Chang, Y. C., Booren, A. M., Gray, J.I., & Dewitt, D. L. (1999). Antioxidant and anti-inflammatory activities of anthocyanins and their aglycon cyaniding from tart cherries. *Journal of Natural products*, 62(2), 294-296.
- Wong, K.C., & Lai, F.Y. (1996). Volatile constituents from the fruits of four *Syzygium* species grown in Malaysia. *Flavour and Fragrance Journal*, 11, 61-66.
- Zanatta, C. F., Cuevas, E., Bobbio, F. O., Winterhalter, P., & Mercadante, A. (2005). Determination of anthocyanins from camu-camu *Myrciaria dubia* by HPLC-PDA, HPLC-MS and NMR. *Journal of Agricultural and Food Chemistry*, 53, 9531-9535.
- Zheng, W., & Wang, S. Y. (2003). Oxygen radical absorbing capacity of phenolics in blueberries, cranberries, chokeberries and lingoberries. *Journal of Agricultural and Food Chemistry*, 51(2), 502-509.
- Zhishen, J., Mengcheng, T., & Jianming, W. (1999). The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chemistry*, 64(4), 555-559.