



**UNIVERSITI PUTRA MALAYSIA**

***PREPRODUCTIVE PATHOPHYSIOLOGICAL CHANGES IN NON-PREGNANT BOER DOES INOCULATED WITH  
Corynebacterium pseudotuberculosis VIA INTRADERMAL,  
INTRANASAL AND ORAL ROUTES***

**AISHATU OTHMAN MOHAMMED**

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**By**

**AISHATU OTHMAN MOHAMMED**

**This is submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Veterinary Science.**

**December 2014**

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## DEDICATION

This work is dedicated to my loving and caring husband, my wonderful parents, siblings and relatives.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the required for the Degree of Master of Veterinary Science

**PRE PRODUCTIVE PATHOPHYSIOLOGICAL CHANGES IN NON-PREGNANT BOER DOES INOCULATED WITH *Corynebacterium pseudotuberculosis* VIA VARIOUS ROUTES.**

By

**AISHATU OTHMAN MOHAMMED**

**December 2014**

**Chairman: Faez Firdaus Jesse Abdullah, PhD.**

**Faculty of Veterinary Medicine**

An experimental study was conducted in non-pregnant Boer does inoculated with *C.pseudotuberculosis* via various routes with the objectives of determining the changes in hematology, biochemical, serum electrolyte, progesterone, estrogen, interleukin-1 $\beta$ , and interleukin 6 as well as pathological changes in the reproductive organs and their associated lymph nodes. A total of twenty (20) healthy non-pregnant Boer does were divided into 4 groups (1, 2, 3 and 4) of 5 does per group. Group 1 was kept unexposed. Group 2, 3 and 4 were inoculated with  $10^7$  cfu/ml of live *Corynebacterium pseudotuberculosis* through intradermal, intranasal and oral routes respectively.

A significant decrease in red blood cell (RBC) count ( $p < 0.05$ ) in the intradermal infected with mean value of  $11.3 \times 10^9/L$ , while the packed cell volume (PCV), hemoglobin concentration (HbC) mean corpuscular volume (MCV), and mean cellular hemoglobin concentration (MCHC) remained unaltered. Significant increase ( $p < 0.05$ ) in white blood cells (WBC) were observed in all the inoculated groups-2, 3 and 4 with mean value of  $13.68 \times 10^9/L$ ,  $9.68 \times 10^9/L$ ,  $8.67 \times 10^9/L$  respectively. Significant increase ( $p < 0.05$ ) in monocyte count was observed in intranasal route with a mean of  $0.754 \times 10^9/L$ . A significant reduction ( $p < 0.05$ ) in lymphocytic count was observed for the intranasal inoculated group with a mean value of  $3.37 \times 10^9/L$  ( $p < 0.05$ ). Significant increase ( $p < 0.05$ ) in neutrophil was observed in intranasal and intradermal route with a mean of  $8.80 \times 10^9/L$ ,  $6.95 \times 10^9/L$ , respectively. Creatinine levels increased ( $p < 0.05$ ) in the intranasal group with a mean of  $91.33 \mu\text{mol/L}$  while increased in GGT levels were observed ( $p < 0.05$ ) intradermal, intranasal and oral groups with a mean differences of  $46.52 \text{U/L}$ ,  $48.00 \text{U/L}$ ,  $36.62 \text{U/L}$  respectively. Lowered Calcium ( $\text{Ca}^{2+}$ ) concentrations were observed in the intradermal group with a mean concentration of  $2.22 \text{mmol/L}$  and  $2.23 \text{mmol/L}$  for the intranasal group. Albumin levels were decrease ( $p < 0.05$ ) in the

intranasal group with a mean of 28.74g/L. Plasma concentrations of progesterone were significantly ( $p<0.05$ ) elevated in intranasal, oral and intradermal with a mean difference of 6.67pg/ml, 3.81pg/ml and 2.4pg/ml respectively. The plasma concentrations of estrogen were elevated significantly ( $p<0.05$ ) in the oral route with a mean of 33ng/ml. However, slight rises in estrogen levels were observed in intradermal and intranasal routes in comparison with the control group.

Interleukin-1 $\beta$  showed a significant increase ( $p<0.05$ ) through the intranasal, intradermal and oral routes compared to the control group with a mean statistical difference of 99pg/ml, 67pg/ml, 42pg/ml respectively while interleukin-6 levels were significantly increased in the intranasal group 85pg/ml ( $p<0.05$ ). There were no significant differences observed in intradermal and oral groups when compared to the control group.

Histologically, inflammatory cells were observed as well as cellular necrosis, congestion in the ovary, uterine horn, cervix and vagina in intranasal and oral routes. Congestion, necrosis were observed in the ovary and uterine horn while necrosis and inflammatory cells were observed in the cervix and vagina of does inoculated through intradermal route. Lesions observed in uterus for the three groups are congestion, edema, necrosis and inflammatory cells.

The present study, therefore, highlight the effects of *C.pseudotuberculosis* on blood, progesterone, estrogen, IL-1 $\beta$ , IL-6 and cellular changes in the reproductive organs and associated lymph nodes which is significant in the understanding the pathophysiology of reproductive system in does infected with *C.pseudotuberculosis*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains Veterinar

**PRA PERUBAHAN PATOFISIOLOGICAL PEMBIAKAN DALAM KAMBING BETINA YANG TIDAK BUNTING DIJANGKITI DENGAN *Corynebacterium pseudotuberculosis* MELALUI PEL BAGAI LALUAN.**

Oleh

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Satu uji kaji telah dijalankan pada kambing betina yang tidak bunting dengan menjangkiti kambing-kambing tersebut dengan *C.pseudotuberculosis* melalui suntikan intradermal, intranasal dan laluan oral dengan objektif untuk menentukan hematologi, biokimia, elektrolit serum, profil hormon pembiakan (progesteron dan estrogen), interleukin-1 $\beta$ , interleukin 6 dan perubahan patologi dalam organ-organ pembiakan dan nodus limfa yang berkaitan. Sebanyak dua puluh (20) kambing betina yang tidak bunting dan sihat telah dibahagikan kepada 4 kumpulan (1,2,3 dan 4), 5-ekor dalam setiap kumpulan. Kambing dalam kumpulan 1 tidak dijangkiti. Kumpulan 2, 3 dan 4 telah dijangkiti dengan  $10^7$  cfu/ml *Corynebacterium pseudotuberculosis* melalui suntikan intradermal, intranasal dan laluan oral.

Penurunan ketara dalam kiraan sel-sel darah merah (RBC) ( $p < 0.05$ ) dalam kambing yang dijangkiti melalui suntikan intradermal dapat dilihat, dengan nilai purata  $11.3 \times 10^9/L$ , manakala isi padu sel padat (PCV), kepekatan hemoglobin (HBC), min isi padu korpusel (MCV), dan purata kepekatan hemoglobin sel (MCHC) kekal tidak berubah. Peningkatan ketara ( $p < 0.05$ ) dalam sel-sel darah putih (WBC) diperhatikan dalam semua kumpulan yang dijangkiti-2, 3 dan 4 dengan nilai min  $13.68 \times 10^9/L$ ,  $9.68 \times 10^9/L$ ,  $8.67 \times 10^9/L$ . Sedikit penurunan ( $p < 0.05$ ) dalam kiraan limfosit diperhatikan untuk kumpulan dijangkiti secara intranasal dengan nilai min  $3.37 \times 10^9/L$  ( $p < 0.05$ ). Tahap kreatinin meningkat ( $p < 0.05$ ) dalam kumpulan intranasal dengan min  $91.33 \mu\text{mol/L}$  manakala peningkatan dalam tahap GGT diperhatikan ( $p < 0.05$ ) dalam semua kumpulan dijangkiti sama ada secara suntikan intradermal, intranasal dan oral dengan perbezaan min  $46.52U/L$ ,  $48.00U/L$ ,  $36.62U/L$ . Tidak ada ( $p < 0.05$ ) perubahan ketara dalam AST, T.Protein, APT dan elektrolit serum dalam semua kumpulan tetapi penurunan kepekatan kalsium ( $\text{Ca}^{2+}$ ) diperhatikan dalam kumpulan suntikan intradermal dengan kepekatan min  $2.22\text{mmol/L}$  dan  $2.23\text{mmol/L}$  bagi kumpulan intranasal. Tahap albumin meningkat ( $p < 0.05$ ) dalam kumpulan

intranasal dengan min 28.74g/ L manakala tiada perubahan yang diperhatikan dalam tahap kepekatan kalium (K +) dan natrium (Na +) bagi ketiga-tiga kumpulan. Kepekatan plasma progesteron meningkat secara ketara ( $p < 0.05$ ) pada kumpulan intranasal, oral dan suntikan intradermal dengan perbezaan min 6.67pg / ml, 3.81pg/ml dan 2.4pg/ml. Kepekatan plasma estrogen telah meningkat dengan ketara ( $p < 0.05$ ) dalam laluan oral dengan min 33ng/ml. Walau bagaimanapun, sedikit peningkatan dalam tahap estrogen diperhatikan dalam suntikan intradermal dan intranasal berbanding dengan kumpulan kawalan.

Interleukin-1 $\beta$  menunjukkan peningkatan yang signifikan ( $p < 0.05$ ) melalui intranasal, suntikan intradermal dan laluan oral berbanding dengan kumpulan kawalan dengan perbezaan statistik min 98pg/ml, 67pg/ ml, 42pg/ml manakala peningkatan ketara pada tahap interleukin-6 diperhatikan dalam kumpulan intranasal 85pg/ ml ( $p < 0.05$ ). Tiada perbezaan yang ketara diperhatikan dalam kumpulan suntikan intradermal dan oral berbanding kumpulan kawalan.

Melalui histologi, penyusupan sel polimorfonukleus dapat diperhatikan selain nekrosis sel, kesesakan sel darah di ovari, tanduk rahim, pangkal rahim dan faraj dalam kumpulan intranasal dan oral. Kesesakan sel darah, nekrosis dalam tanduk ovari dan rahim manakala nekrosis dan penyusupan sel neutrofil dalam serviks dan faraj pada kumpulan dijangkiti melalui suntikan intradermal. Uterus dalam ketiga-tiga kumpulan menunjukkan kesesakan sel darah, edema, nekrosis dan penyusupan neutrofil.

Oleh itu, kajian ini menyerlahkan kesan *C.pseudotuberculosis* pada progesteron, estrogen, IL-1 $\beta$ , IL-6, darah dan perubahan sel dalam organ-organ pembiakan berserta nodus limfa yang berkaitan, yang mana penting dalam memahami patofisiologi sistem pembiakan dalam kambing yang dijangkiti *C.pseudotuberculosis*.



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I certify that a Thesis examination committee has met on August 2014 to conduct the final examination of Aishatu Othman Mohammed on her thesis entitled Pre productive Pathophysiological Changes in non-pregnant Boer does inoculated with *Corynebacterium pseudotuberculosis* via various routes. In accordance with the Universities and University College Act 1971 and the constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Veterinary Science.

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## LIST OF ABBREVIATIONS

%	Percentage
µmol/L	Micromoles per liter
Alb	Albumin
AST	Aspartate transaminase
ALP	Alkaline phosphatase
AGPT	Agar gel precipitation test
Ca	Calcium
Cl	Chloride
CK	Creatinine Kinase
CLA	Caseous lymphadenitis
CFU	Colony forming unit
<i>C.pseudotuberculosis</i>	<i>Corynebacterium pseudotuberculosis</i>
DPX	Distyrene Plasticizer Xylene
E3	Estrogen
EDTA	Ethylene diaminetetraacetic acid
ELISA	Enzyme link immunosorbent assay
Fig	Figure
GGT	Gamma-glutamyl transpeptidase
g/L	Gram per liter
Hb	Haemoglobin
K	Potassium
IL	Interleukin
MCHC	Mean corpuscular hemoglobin concentration
MCV	Mean corpuscular volume
ML	Milliliters
Mmol/L	Millimolar per liter
N	Number of animals
ng	Nanogram
Na	Sodium
OD	Optical density
PBS	Phosphate buffered saline
PCV	Pack cell volume
PCR	Polymerase chain reaction
P4	Progesterone
PLD	Phospholipase D
Pg	Pictogram
RBC	Red blood count
RIA	Radioimmunoassay
TPU	Taman pertanian Universiti
T.bil	Total bilirubin
UPM	Universiti Putra Malaysia
U/L	Units per liter
WBC	White blood cells

# CHAPTER ONE

## INTRODUCTION

### 1.1 GENERAL BACKGROUND

*Corynebacterium pseudotuberculosis* is an intracellular gram-positive, facultative anaerobic small curved rod bacterium (Jesse *et al.*, 2011). *C. pseudotuberculosis* is generally regarded as an important animal pathogen and the etiological agent of the disease that is commonly called caseous lymphadenitis (CLA) in sheep and goats, and can also infect several other hosts, including human (Dorella, *et al.*, 2006; Baird, *et al.*, 2007; Guimaraes *et al.*, 2011a). The disease is distributed worldwide and has a major economic impact in most sheep and goats raising areas (Baird and Fontaine, 2007; Seyffert, *et al.*, 2010; Guimaraes, *et al.*, 2011a). The disease is also of zoonotic importance as it may on rare occasions cause regional lymphadenitis in humans, particularly in farm workers and meat inspectors (Peel, *et al.*, 1997).

Transmission of CLA can be through oral, intradermal and the peritoneal routes (Goldberger *et al.*, 1981; Stoops *et al.*, 1984; Collet *et al.*, 1994; AdzaRina *et al.*, 2013). The ability of the organism to survive for a long time in the environment coupled with its resistance to commonly available antibiotics are key features contributing to its increase in transmission within a herd (Williamson, 2001; Baird and Fontaine, 2007; Seyffert *et al.*, 2010; Abdullah *et al.*, 2013). Another challenge is the difficulty in identifying sub-clinically infected animals (AL-Gaabary *et al.*, 2009). CLA in sheep and goats is usually associated with the development of large external abscesses in the subcutaneous tissues, lymph nodes and visceral organs such as liver, spleen and kidneys (Aloso *et al.*, 1993; Arsenault *et al.*, 2003).

Treatment of CLA with antibiotic is inefficient/ ineffective because its thick capsule (abscess) prevents adequate penetration to reach a minimum effective dose within the lesion (Piontkowski and Shievers, 1998; Standfort *et al.*, 1998; Williamson, 2001). The best way for preventing and controlling this disease is by vaccination, culling of infected animals respectively (Luis *et al.*, 2007).

Disturbances in blood parameters and serum electrolytes have been observed in mice challenged with *C. pseudotuberculosis* and its exotoxin phospholipase D (PLD) (Abdinasiret *et al.*, 2012). Progesterone and estrogen concentration were significantly higher in mice inoculated with *C. pseudotuberculosis* and its exotoxin phospholipase D (PLD) (Zaid *et al.*, 2013). A significant reduction in progesterone level and progesterone/estrogen ratio in cows and buffalo infected with *Brucella melitensis* compared to the control group have been reported by Jabbaret *et al.*, (2012). Progesterone production by granulosa cells is increased *in vitro* by IL-1 $\beta$  in cattle and inhibits estradiol-17 $\beta$  production (Barak *et al.*, 1992; Baratta *et al.*, 1996).

The FSH- stimulation of estrogen and progesterone released may be associated with upregulation of IL-6(Salmassi, *et al.*,2001). Experimentally challenged female mice with *C.pseudotuberculosis* and its exotoxin (PLD) via intraperitoneal route showed histopathological changes such as severe congestion, profound thrombus formation and necrosis of ovaries and uterus, while testis and epididymis showed similar lesions (Zaid *et al.*, 2012).

There has been reports that the disease causes decrease in reproduction, wasting, poor wool growth, decreased milk and meat production, premature culling, carcass condemnation and rarely death in sheep, goats and other species (Williamson 2001; Paton *et al.*, 2003; Arsenault *et al.*, 2003; Peterhanset *al.*, 2004). The disease is globally distributed and sustained by latent infections(Patoet *al.*, 1994; Arsenault *et al.*, 2003; Ivanovićet *al.*, 2007). currently, the disease has been reported in many parts of Asia (OIE 2009), including Malaysia (Komalaet *al.*, 2008).Ruminant production in Malaysia ischanging steadily from subsistence to intensive operations (Jesse *et al.*, 2013a). The ruminant's population in Malaysia is predominantly made up of cattle, buffaloes, goats and sheep. Therefore, the ruminant industry is the foremost in the production of food such as milk, meat and the by- products such as hide for the leather industry in Malaysia.

Infertility caused by the organism (*C.pseudotuberculosis*) in non-pregnant Boer does is the main constraint in farm production for economic and public health reasons, as it results in wastage from culling and meat condemnation while infected animals are capable of rapidly transmitting the disease within the flock. Hence, infertility due to *C.pseudotuberculosis* is the main focus of this study in a bid to highlight the mechanismsinvolved in the disruption of the reproductive system via known different routes of infection. Reproductive hormonal (progesterone and estrogen) changes investigated in this study would be very useful as an indicative marker of infertility response to CLA.Understanding of alterations in biochemical and haematological parameters consequent to *C.pseudotuberculosis* inoculation/infection is vital to the prompt diagnosis and scoring the health status of the animal.

There is currently dearth of information on the effects of *C.pseudotuberculosis* on pre reproductive pathophysiological changes through the various routes of infection. Therefore, this study would obtain additional data on the effects of CLA onreproduction in non-pregnant Boer does.

The objectives of this study are:

1. To study the changes in blood parameters (haematological, biochemical and serum electrolyte) of non-pregnant Boer does following infection with *C.pseudotuberculosis* via different routes of inoculation.



2. To determine the plasma progesterone and estrogen concentration in non-pregnant Boer does infected with *C.pseudotuberculosis* via different routes of inoculation.
3. To investigate the inflammatory cytokines (IL-1 $\beta$  and IL-6) responses in the serum of non-pregnant does infected with *C.pseudotuberculosis* via different routes of inoculation.
4. To observe the histopathological alterations in the reproductive organs and associated lymph nodes of the affected animals experimentally infected with *C.pseudotuberculosis* via different routes of inoculation.

The hypotheses of this study are outlined below:

1. There might be changes in the blood parameters in non-pregnant Boer does inoculated via different routes with *C.pseudotuberculosis*.
2. Infection of non-pregnant Boer does with *Corynebacterium pseudotuberculosis* via different routes of inoculation may alter the concentration of progesterone and estrogen concentration.
3. There might be changes in the concentration of inflammatory cytokine associated with *C.pseudotuberculosis* infections through different routes of inoculation in non-pregnant Boer does.
4. There will be cellular changes in the reproductive organs and associated lymph nodes of non-pregnant Boer does inoculated with *C.pseudotuberculosis* via different route of inoculation.

Therefore, this study will provide a better understanding on the pathogenesis of infertility during CLA infection in non-pregnant Boer does.





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## BIBLIOGRAPHY

- AdzaRina, M.N., Zamri-Saad, M., Jesse, F.F.A., Saharee, A.A., Haron, A.W., Shahirudin, S. (2013). Clinical and pathological changes in goats inoculated *Corynebacterium pseudotuberculosis* by intradermal, intranasal and oral routes. *Journal of Veterinary Research* © Volume 17 (2):73-81.
- Abdullah, F.F.J., Osman, A.Y., Adamu, L., Azri, N.A., Haron, A.W., Saad, M.Z., Omar, A.R., Saharee, A.A. (2013). Caseous Lymphadenitis in a Goat: Case report. *South Asian Journal of Life Sciences*, 1 (1):19-20.
- Abdinasir, O.Y., Jesse, F.F.A., Saharee, A.A., Haron, A. W., Sabri, J. and Abdullah, R. (2012). Haematological and biochemical Alteration in mice following Experimental infection with whole cell and exotoxin (PLD) Extracted from *C. Pseudotuberculosis*. *Journal of Animal and Veterinary Advances*, 11 (24): 4660-4667.
- Alcama, I.E. (1998). Theory and problems of microbiology. Schum's outline series, McGraw-Hill Companies 137:147-153.
- Al-Gaabary, M.H., Osman, S. A., Ahmed, M.S., and Oreiby, A.F. (2010). Abattoir survey on caseous lymphadenitis in sheep and goats in Tanta, Egypt. *Small Ruminant Research*, 94(1): 117-124.
- Al-Gaabary, M. H., Osman, S.A., & Oreiby, A.F. (2009). Caseous lymphadenitis in sheep and goats: Clinical, epidemiological and preventive studies. *Small Ruminant Research*, 87(1): 116-121.
- Alloui, M. N., Kaba, J., and Alloui, N. (2011). Prevalence and risk factors of caseous lymphadenitis in sheep and goats of Batna area (Algeria). *Research opinions in Animal and Veterinary sciences*, 1(3): 162-164.
- Al Rawashdel, O.F. and Al Qudah, K.M. (2000). Effect of shearing on the incidence of caseous lymphadenitis in Awassi sheep in Jordan. *Journal of Veterinary Medicine Series B-Infectious Disease and Veterinary Public Health*, 47, 287-293.
- Alonso, J.L., Simon, M.C., Girones, O., Muzquiz, J.L., Ortega, C., Garcia, J. (1993). The effect of experimental infection with *Corynebacterium pseudotuberculosis* on reproduction in adult ewes. *Research In Veterinary Science*, 52: 267-272.
- Arsenault, J., Girard, C., Dubreuil, P. (2003). Prevalence of and carcass condemnation from maedi-visna, paratuberculosis and caseous lymphadenitis in culled sheep from Quebec, Canada. *Preventive Veterinary Medicine*, 59: 67-81.
- Baird, G.J., and Fontaine, M.C. (2007). *Corynebacterium pseudotuberculosis* and its role in ovine caseous lymphadenitis. *Journal of Comparative Pathology*, 137: 179-210.

- Batey, R.G.(1986a).The effect of caseous lymphadenitis on body condition and weightof merino mutton carcasses.Australia Veterinary Journal, 63,268.
- Batey, R.G.(1986b).Pathogenesis of caseous lymphadenitis in sheep and goats.Australia Veterinary Journal,63,296-272.
- Benham, C.L., Seaman, A., Woodbine, M. (1962).*Corynebacterium pseudotuberculosis* and its role in diseases of animals.Veterinary Bulletin, 32: 645-657.
- Brogden, K.A., and Engen, R.L. (1990). Alteration in the phospholipid composition and Morphology of ovine erythrocytes after intravenous inoculation of *Corynebacterium pseudotuberculosis*. American Journal of Veterinary Research, 51:874-877.
- Brodgen, K.A., Cutlip, R.C. and Lehmkuhl, H.D. (1984).Experimental *Corynebacterium pseudotuberculosis* infection in lambs. American Journal of Veterinary Research, 45: 1532-1534.
- Brogden, K.A, and Engen, RL.(1990).Alterations in the phospholipid composition and morphology of ovine erthrocytes after intravenous inoculation of *Corynebacterium pseudotuberculosis*. American Journal of Veterinary Research, 51: 874-877.
- Brown, C.C., Olander, H.J. and Alves, S.F. (1987). Synergistic hemolysis-inhibition titers associated with caseous lymphadenitis in a slaughter house survey of goats and sheep in Northeastern Brazil. Canadian Journal of Veterinary Research, 51: 46-49.
- Burrell, D.H. (1978). Experimental induction of caseous lymphadenitis in sheep by intralymphatic inoculation of *Corynebacterium ovis*. Research in Veterinary Science,24: 269-276.
- Baratta, M., Basini, G., Bussolati, S., Tamanini, C (1996). Effects of interleukin-1 beta fragment (163-171) on progesterone and estradiol-17 beta release by bovine granulosa cells from different size follicles. Regulatory Peptides, 67: 187-94.
- Banerjee,M.R (1976).Responses of mammary cells to hormones.International Review of Cytology, 47: 1-97.
- Besarab, A., Caro, J.F. (1981). Increased absolute calcium binding to albumin in hypoalbuminaemia.Journal Clinical Pathology, 34: 1368-1374.
- Barak,V., Yanai, P., Trever, A.J., Roisman, I., Simon, A. &Lufer, N.(1992). Interleukin-1: local production and modulation of human granulosa luteal cells steroidogenesis. Fertility and sterility, 58, 719-725.

- Bush, R.D., R. Barnett, R., Links, I.J. and Windsor, P. A. (2012) Using abattoir surveillance and producer surveys to investigate the prevalence and current preventative management of caseous lymphadenitis in Merino flocks in Australia. *Animal Production Science* 52(7) 675-679.
- Carne, H.R. (1940). The toxin of *Corynebacterium ovis*. *Journal of Pathology and Bacteriology*, 51, 199-212.
- Collett, M.G., Bath, G.F., Cameron, C.M. (1994) *Corynebacterium pseudotuberculosis* infections In: Coetzer, J., Thomson, G.R., Tustin, R.C. (Eds), *Infectious diseases of livestock with Special Reference to Southern Africa*. (2nd ed.), Oxford University Press, Cape Town, 1387-1395.
- Carne, H.R., and Onon, E.O. (1978). Action of *Corynebacterium ovis* exotoxin on the endothelial cells of blood vessels. *Nature*, 271, 246-248.
- Çetinkaya, B., Karahan, M., Atil, E., Kalin, R., De Baere, T. and Vaneechoutte, M. (2002). Identification of *Corynebacterium pseudotuberculosis* isolates from sheep and goats by PCR. *Veterinary Microbiology*, 88: 75–83.
- Coyle, M.B., Hollis, D.G. and Groman, N.B. (1985). *Corynebacterium spp.* And other coryneform organisms. In: *Manual of Clinical Microbiology*, 4th Edith., E.H. Lennette, A. Balows, W. Hausler and H.J. Shadomy, Eds, American Society for Microbiology, Washington, 198-199.
- Carvahol, N.A.V., Mol, J.P.S., Xavier, M.N., Paixão, T.A., Lage, A.P. and Santos, R.L. (2010). Pathogenesis of bovine beucellosis. *Veterinary Journal*, 184, 146-155.
- Carolina, G., Hernández-Bello, R. and Morales-Montor, J. (2010). Regulation of Steroidogenesis in Reproductive, Adrenal and Neural Tissues by Cytokines. *Neuroendocrinology Journal*, 2010, 3, 161-169.
- Chun, S.L. (2012). Cytokine gene polymorphisms: *Kidney Research of Clinical Practice*, 31: 203-204.
- Chrousos, G.P. (1995). The hypothalamic-pituitary-adrenal axis and immune-mediated inflammation. *New England Journal of Medicine*, 332: 1351- 62.
- Chen, H.F., Shew, J.Y., Chao, K.H., Chang, L.J., Ho, H.N., Yang, Y.S. (2000). Luteinizing hormone up-regulates the expression of interleukin-1 beta mRNA in human granulosa-luteal cells. *American Journal of Reproductive Immunology*, 43: 125-33.
- Corner, G.W., Allen, W.M. (1929). Physiology of the corpus luteum. II. Production of a special uterine reaction (Progestional proliferation) by extracts of the corpus luteum. *American Journal of Physiology*, 88: 326-399.

- Cameron, C.M., Minnaar, J.L., Engelbrecht, M., and Purdom, M.R. (1972). Immune response of Merino sheep to inactivated *Corynebacterium pseudotuberculosis* vaccine. Onderstepoort Journal of Veterinary Research, 39, 11-24.
- Dorella, F.A., Pacheco, L.G.C., Oliveira, S.C., Miyoshi, A., & Azevedo, V. (2006). *Corynebacterium pseudotuberculosis*: microbiology, biochemical properties, pathogenesis and molecular studies of virulence. Veterinary research, 37(2), 201-218.
- Dinareello, C.A. (2010). Anti-inflammatory agents: present future. Journal of cell, 140 (6): 935-950.
- Egen, N.B., Cuevas, W.A., McNamara, P.J., Sammons, D.W., Humphreys, R., Songer, J.G. (1989). Purification of the phospholipase D of *Corynebacterium pseudotuberculosis* by recycling isoelectric focusing. American Journal of Veterinary Research, 50: 1319-1322.
- Euzeby, J.P. (2005). List of bacterial names with standing in nomenclature- genus *Actinomadura* (online). Available from <http://www.bacterio.cict.fr/a/actinomadura>.
- Ehrhart-Bornstein, M., Bornstein, S.R., Scherbaum, W.A. (1996). Sympathoadrenal system and immune system in the regulation of adrenocortical function. European Journal of Endocrinology, 135: 19-26.
- Ellis, J.A., Hawk, D.A., Holler, L.D., Mills, K.W., Pratt, D.L. (1990). Differential antibody responses to *Corynebacterium pseudotuberculosis* in sheep with naturally acquired caseous lymphadenitis. Journal of the American Veterinary Medical Association, 196: 1609-1613.
- El-Enbaawy, M.I., Saad, M.M., Selim, S.A. (2005). Humoral and cellular immune responses of a murine model against *Corynebacterium pseudotuberculosis* antigens. Egyptian Journal of Immunology, 12: 13-20.
- Fraser, G. (1961). Haemolytic activity of *Corynebacterium ovis*. Nature, 189-246.
- Fontain, M.C., Baird, G., Connor, K.M., Rudge, K., Sales, J. and Donachie, W. (2006). Vaccination confers significant protection of sheep against infection with a virulent United Kingdom strain *Corynebacterium pseudotuberculosis*. Vaccine, 24: 5986-5996.
- Fontaine, M.C., Baird, G.J. (2008). Caseous lymphadenitis. Small Ruminant Research, 76: 42-48.
- Freitas, V.J.F., Baril, G., Saumande, J. (1997). Estrus synchronization in dairy goats: use of fluorogestone acetate vaginal sponges or norgestomet ear implants. Animal Reproduction Sciences, Volume 46, Issues 3-4, Pages 237-244.

- Gameel, A.A. and Tartour.(1974). Haematological and Plasma protein changes in sheep experimentally infected with *Corynebacterium pseudotuberculosis*.Journal of Comparative Pathology, Vol.84.
- Goldberger, A.C., Lipsky, B.A., Plorde, J.J. (1981). Suppurative granulomatous lymphadenitis caused by *Corynebacterium ovis* (*pseudotuberculosis*). American Journal of Clinical Pathology, 76: 486-490.
- Gomes, W.R., and Erb, R.E.(1965).Progesterone in bovine reproduction:A review. Journal of Dairy sciences.48, 314.
- Guimaraes, A.S., Carmo, F.B., Pauletti, R.B., Seyffert, N., Ribeiro, D., Lage, A.P., Heinemann, M.B., Miyoshi, A., Azevedo, V. and Gouveia, A.M.G. (2011). Caseous lymphadenitis: epidemiology, diagnosis, and control. Institute of Integrative Omics and Applied Biotechnology Journal, 2(2): 33–43.
- Gonzalez-Hernandez, J.A., Bornstein, S.R., Ehrhart-Bornstein, M., Spath-Schwalbe, E., Jirikowski, G., Scherbaum, W.A.(1994).Interleukin-6 messenger ribonucleic acid regulation of adrenal function. Journal ofClinical Endocrinology and Metabolism,79: 1492-7.
- Gonzalez-Hernandez, J.A., Bornstein, S.R., Ehrhart-Bornstein, M., et al.(1995). IL-1 is expressed in human adrenal gland in vivo. Possible role in a local immune-adrenal axis.Journal of Clinical and Experimental Immunology, 99: 137-41.
- Gonzalez-Hernandez, J.A., Ehrhart-Bornstein, M., Spath-Schwalbe, E., Scherbaum, W.A., Bornstein, S.R.(1996). Human adrenal cells express tumor necrosis factor-alpha messenger ribonucleic acid: evidence for paracrine control of adrenal function. Journal of Clinical Endocrinology and Metabolism, 81: 807-13.266.
- George, F. W. and Wilson, J .D. (1988).Sex determination and differentiation. In Knobil E et al: The Physiology ofReproduction. New York, Raven Press, pp 3-26.
- García-Gómez, J.M.,*et al.*, (2013). Sparse Manifold Clustering and Embedding to discriminate gene expression profiles of glioblastoma and meningioma tumors. Computers in Biology and Medicine, 43(11):1863–1869.
- Guilloteau, L., Pepin, M., Pardon, P., et al. 91990). Recruitment of 99m-technetium- or 111-indium-labelled polymorphonuclear leucocytes in experimentally induced pyogranulomas in lambs. Journal ofLeukocyte Biology 48: 343–352.
- Hard, G.C. (1972).Examination by electron microscopy of the interaction between peritoneal phagocyte and *Corynebacterium ovis*. Journal of Medical Microbiology, 5: 483-491.



- Henrique, O.C., Ney Pentead de, C.N., Lia, M.R., Ieda, M., Flavia,C., Leonardo da, S.(2014). Influence of estradiol administration on estrogen receptors of nasal mucosa: an experimental study on guinea pigs. *Brazil Journal of Otorhinolaryngology*, 80: 18-23.
- Hair-Bejo, M., Salina, S., Hafiza, H. and Julaida, S. (2000). In vivo vaccination against infectious bursal disease in broiler chickens. *Journal of Veterinary Malaysia*, 12:63-69.
- Hodgson, A.L., Bird, P. and Nisbet, I.T. (1990). Cloning, nucleotide sequence, and expression in *Escherichia coli* of the phospholipase D gene from *Corynebacterium pseudotuberculosis* phospholipase D. *Vaccine*, 17:802-808.
- Hard, G.C. (1969). Electron microscopic study of the differentiation of mouse peritoneal macrophages stimulated by *Corynebacterium ovis* infection. *Laboratory Investigation*, 21 (4): 309–315.
- Holland, M.B. and Roy, D. (1995) Estrogen-induced cell proliferation and differentiation in the mammary gland of the female Noble rat. *Carcinogenesis*, 16: 1955–1961.
- Imura, H., Fukata, J.(1994). Endocrine-paracrine interaction in communication between the immune and endocrine systems. Activation of the hypothalamic-pituitary-adrenal axis in inflammation. *European Journal of Endocrinology*, 130: 32-7.
- Ibtisam, M. A. (2008). Some clinicopathological and pathological studies of *C. ovis* infection in sheep. *Egypt Journal of comparative Pathology and Clinical Pathology*, 21 (1):327-343.
- Jalal, A., Jazayeri, A., Graeme, J., Carroll, B., Ann, B., Vernallis. (2010). Interleukin-6 subfamily cytokines and rheumatoid arthritis: Role of antagonists. *International Immunopharmacology*; 10: 1–8.
- Jesse, F.F.A., Randolph, P.S.S., Saharee, A.A., Wahid, A.H., Zamri-Saad, M., Jasni, S., Omar, A.R., Adamu, L. and Abdinasir, O.Y.(2013). Clinico-pathological response of mice following oral route infection of *C. pseudotuberculosis*. *Journal of Agriculture and Veterinary Science*, 2: 38-42.
- Jesse, F.F.A., Azlan, C.M., Saharee, A.A., Murugaiyah, M., Noordin, M.M., Jasni, S., Ragavan, K., Hassan, M.D., Haron, A.W., Siti, K.B., Hazilawati, H. and Mahmud, T. (2008). Control of Caseous Lymphadenitis (CLA) in goat at UPM Farm. *Proceedings: 20th Veterinary Association Malaysia*, 2008.
- Jesse, F.F.A., Sang, S.L., Saharee, A.A. and Shahirudin, S. (2011). Pathological Changes in the organs of mice model inoculated with *Corynebacterium pseudotuberculosis* Organism. *Pertanika Journal of tropical Agricultural Sciences*, 34 (1):145-149.

- Jesse, F.F.A., Adamu, L., Abdinasir, Y.O., Zakaria, Z. and Abdullah, R *et al.*, (2013a). Clinico-pathological responses of calves associated with infection of *Pasteurella multocida* type B and the bacterial lipopolysaccharide and outer membrane protein immunogens. *International Journal of Animal and Veterinary Advances*, 5: 190-198.
- Jones, S.A., Horiuchi, S., Topley, N., Yamamoto, N., Fuller, G.M.(2001): The soluble interleukin-6 receptor :mechanisms of production and implications in disease. *The FASEB Journal*, 1:43-58.
- Jyoti, K., Bhupendra, N.T., Rajiv, K., Ganesh, G.S., Shivendra, K.D. (2013). Rapid detection of *Corynebacterium pseudotuberculosis* in clinical samples from sheep. *Tropical Animal Health and Production*, 45, (6): 1429-1435.
- Jolly, R.D.(1965).The pathogenic action of the exotoxin of *Corynebacterium ovis*. *Journal of Comparative pathology*,75:417-431.
- Jain, N.C. (2000): *Schalm's Veterinary Hematology* 6th edition. Lea and Febiger Philadelphia, USA.
- Jabbar, A.A., AL-Sa'aidi., Mohsen, A. A., Ali, A. N.(2012). Clinical, Serological, Hormonal, Bacteriological and Molecular Detection of Brucellosis in Aborted Cows and Buffalos. *International Conference on Applied Life Sciences (ICALS2012)*.
- Kaplanski, G., Marin, V., Montero-Julian, F., Mantovani, A., Farnarier, C. (2003):IL-6: a regulator of the transition from neutrophil to monocytereruitment during inflammation. *Trends Immunology*, 24: 25-29.
- Kaba, J.,Nowicki,M.T., Frymus, D.,Nowicka, L., Witkowski,O., Szaluś-Jordanow, M., Czopowicz, M.T. (2011). Evaluation of the risk factors influencing the spread of caseous lymphadenitis in goat herds. *Polish Journal of Veterinary Sciences*, 14, (2): 231-237.
- Kuria, J.K., Mbuthia, P.G., Kang' ethe, E.K, and Wahome, R.G.(2001).Caseous lymphadenitis in goats: the pathogenesis, incubation period and serological response after experimental infection. *Veterinary Research Communications*. 25:89-97.
- Kuria, J.K.N. and Holstad, G. (1989).Serological investigation of *Corynebacterium pseudotuberculosis* infection in sheep correlation between the hemolysisinhibition test and the ELISA test.*Acta Veterinary Scandinavia*, 30:109-110.
- Komala, T.S., Ramlan, M.,Yeoh N.N., Surayani, A,R. And SharifahHamidah, S.M.A.(2008).Survey of caseous lymphadenitis in small ruminant farms from two districts in perak, Malaysia- Kinta and Hilir Perak. *Tropical Biomedicine* 25(3):196-201.



- Kishimoto, T., Tanaka, T., Yoshida, K., Akira, S., Taga, T.(1995): Cytokine signal transduction through homo- or heterodimer of gp 130. *Annals of the New York Academy of Sciences*,766: 224-234.
- Kylie, H., Van der Hoek, C., Woodhouse,M., Mats Brännström, M and Robert, J. N.(1998).Effects of interleukin (IL) 6 on Luteinizing Hormone- and IL-1 $\beta$ -Induced Ovulation and Steroidogenesis in the Rat Ovary1 .*Biology of Reproduction*, 58, 1266-1271.
- Luis, G.C., Pacheco R. R., PenaThiago, L.P., Castro,F. A., Dorella R. C., Bahia, R. C., Marcilio, N.M., Frota, S. C., Oliveira, R. M., Francisco, S.F., Alves, A. M. and Vasco A.(2007).Multiplex PCR assay for identification of *Corynebacterium pseudotuberculosis* from pure cultures and for rapid detection of this pathogen in clinical samples. *Journal of Medical Microbiology*, 56:480-486.
- Liu, T.B., Cheng, S.P., Hung, C.M., Yuh, L.Y.(2002). Serum progesterone changes in luteal cyclicity and duration of estrous cycle in Formosan Sika Deer. *Zoology Sciences*,19(9):1033-1037.
- Lan, D.T.B., Makino, S., Shirahata, T.,*et al.* (1999).Tumor necrosis factor and  $\gamma$  interferon are required for the development of protective immunity to secondary *Corynebacterium pseudotuberculosis* infection in mice. *Journal of Veterinary Medical Sciences*, 61: 1203–1208.
- Lehmann, K.B. and Neumann, R.O.(1896). In: *Atlasund Grundriss der Bakteriologieundlehrbuch der SpezillenBakteriologischenDiagnostik. Lehmann's MedizinischeHandatlanten, Bd.10* .
- Moore, R., Miyoshi, A., Pacheco, L.G.C., Seyffert, N., Azevedo, V. (2010).*Corynebacterium and Arcanobacterium* In: *Pathogenesis of bacterial infections in animals*. (4th ed.), Blackwell Publishing, Iowa.
- Mahre, M.B., Wahid, H., Rosnina, Y., Jesse, F.F.A., Azlan, C.A., Yap, K.C.(2013). Plasam progesterone changes and length of oestrous cycle in Rusa Deer (*Rusatrimorensis*). *Animal Reproductive Science*.141:148-153.
- Merhej, V., Royer-Carenzi, M., Pontarotti, P., Raoult, D. (2009). Massive comparative genomic analysis reveals convergent evolution of specialized bacterial. *Biology Direct*, 4:13-37.
- Moshage, H. (1997). Cytokines and the hepatic acute phase. *Journal of pathology*, 181:257-266.
- Muckle, C.A. and Gyles, C.L. (1986).Exotoxic activities of *Corynebacterium pseudotuberculosis*. *Current Microbiology*, 13: 57-60.
- Muckle, C.A. and Gyles, C.L. (1983).Relation of lipid content and exotoxin production to virulence of *Corynebacterium pseudotuberculosis* in mice. *American Journal of Veterinary Research*, 44: 1149-1153.

- Mikuli, M.(1995).Effect of interleukin-2 and interleukin-6 on ovary in the ovulatory period- establishment of the new ovarian perfusion system and influence of interleukins on ovulation rate and steroid secretion. Hokkaido IgakuZasshi, 70:561-572.
- Middleton, M.J., Epstein, V.M. and Gregory, G.G.(1991).Caseous lymphadenitis on Flanders Island: prevalence and management surveys. Australian Veterinary Journal, 68:311-312.
- Maki, L.R., Shen, S.H., Bergstrom, R.C. and Stetzenbach, L.D.(1985). Diagnosis of *Corynebacterium pseudotuberculosis* infections in sheep, using an enzyme-linked immunosorbent assay.American Journal of Veterinary Research,46:212-214.
- Martin, L., Finn, C.A. and Trinder,G. (1973). Hypertrophy and hyperplasiain the mouse uterus after estrogen treatment: an autoradiographic study.Journal of . Endocrinology, 56: 133–141.
- Musa, M. (1998). “Hemolytic interactions of dermatophiluscongolesis. “ Zbi.Veterinarmed (B) 39(2):139-142.
- Nocard, E. (1896). Annales de l’Institut Pasteur, 10: 609.
- Sood, N.K., Sandhu,B.S.,Gupta, K.,Narang, D., Vasudeva, K., Singh, N.D.(2012). Mesenteric caseous lymphadenitis in a cow calf caused by *Corynebacterium pseudotuberculosis* a case report: VeterinarniMedicina, 57, (7): 371–375.
- Salmassi, A., Lü, S., Hedderich, J., Oettinghaus, C., Jonat, W.(2001).Interaction of interleukin-6 on human granulosa cell steroid secretion.Journal of endocrinology,17, 471-478.
- Nagy, G. (1976). Caseous lymphadenitis in sheep.Methods of infection. Journal of the South Africa Veterinary Association, 47:197-199.
- Nagel-Alne, G.E., Sølverød, L.S., Valle, P.S., Simon-Nilsen, M. (2009).Prevalence and handling of infection with CLA in sheep flocks in Malangenpeninsula in Troms County. In: Proceedings of the 7th International Sheep Veterinary Congress, Stavanger, 95–96.
- O’Reilly, K.M., Green, L.E., Malone, F.E., et al., (2008).Parameter estimation and simulations of a mathematical model of *Corynebacterium pseudotuberculosis* transmission in sheep. Preventive Veterinary Medicine 83: 242–259.
- OIE World Organization For Animal Health (2009).
- Paton,M.W., Walker, S.B., Rose, I.R. and Watt, G.F. (2003). Prevalence of caseous lymphadenitis and usage of caseous lymphadenitis vaccines in sheep flocks. Australian Veterinary Journal, 81: 91–95.

- Pépin, M., Seow, H.F., Corner, L., Rothel, J.S., Hodgson, A.L., Wood, P.R. (1997). Cytokine gene expression in sheep following experimental infection with various strains of *Corynebacterium pseudotuberculosis* differing in virulence. *Veterinary Research*, 28(2):149-163.
- Pépin, M., Pardon, P., Marly, J. et al. (1988). *Corynebacterium pseudotuberculosis* infection in adult ewes by inoculation in the external ear. *American Journal of Veterinary Research*, 49: 459–463.
- Piontkowski, M.D. and Shivvers, D.W. (1998). Evaluation of a commercially available vaccine against *Corynebacterium pseudotuberculosis* for use in sheep. *Journal of American Veterinary Medicine Association*, 212:1765-1768.
- Palmieri, C., Schiavi, E., Della Salda, L. (2011). Congenital and acquired pathology of ovary and tubular genital organs in ewes: *Journal of theriogenology*, 75:393-410.
- Paule, B.J.A., Azevedo, V., Regis, L.F., Carminati, R., Bahia, C.R., Vale, V.L.C., Moura-Costa, L.F., Freire, S.M., Nascimento, I., Goes, A.M., Meyer, R. (2003). Experimental *Corynebacterium pseudotuberculosis* primary infection in goats: Kinetics of IgG and interferon-gamma production, IgG avidity and antigen recognition by western blotting. *Veterinary Immunology and Immunopathology*, 96:129-139.
- Pépin, M., Paton, M. and Hodgson, A.L. (1994a). Pathogenesis and epidemiology of *Corynebacterium pseudotuberculosis* infection in sheep. *Current Topics in Veterinary Research*, 1: 63-82.
- Peterhans, E., Greenland, T., and Badiola, J. (2004). Routes of transmission and consequences of small ruminant lentiviruses (SRLV'S) infection and eradication schemes. *Veterinary Research*, 35: 257-274.
- Papaioannou, N., Zavlaris, M., Giadinis N., Petridou, E., & Psychas, V. (2010). A case of kidney infection by *Corynebacterium pseudotuberculosis* in sheep. *Journal of the Hellenic Veterinary Medical Society*, 61(1):29-35.
- Peel, M.M., Palmer, G.G., Stacpoole, A.M., and Kerr, T.G. (1997). Human lymphadenitis due to *Corynebacterium pseudotuberculosis*: report of ten cases from Australia and review. *Clinical Infectious Disease*, 24:185–191.
- Pugh, D.G. (2004) *Clínica de ovinos e caprinos*. 1st edn Roca, São Paulo.
- Preisz, H. and Guinard, L. (1891). *Journal Medicine Veterinaire*, 16: 563.
- Quinn, P.J., Carter M.E., Markey B. and Carter G.R. (1994). *Corynebacterium species* and *Rhodococcusequi*. In: *Clinical Veterinary microbiology and microbiology*, Wolfe Publishing Company, London, 137-147.
- Quarmby, V.E., and Korach, K.S. (1984) Influence of 17 $\beta$ -estradiol on the pattern of cell division in the uterus. *Endocrinology*, 114: 694–702.

- Ribeiro, M.G., Dias Junior, J.G., Paes, A.C., *et al.* (2001). Punção aspirativa com agulha fina no diagnóstico de *Corynebacterium pseudotuberculosis* na linfadenite caseosa caprina. *Arq Inst Biol São Paulo*, 68: 23-28.
- Rebouças, M.F., Portela, R.W., Lima, D.D., Loureiro, D., Bastos, B.L., Moura-Costa, L.F., Meyer, R. (2011). *Corynebacterium pseudotuberculosis* secreted antigen-induced specific gamma-interferon production by peripheral blood leukocytes: potential diagnostic marker for caseous lymphadenitis in sheep and goats. *Journal of Veterinary Diagnostic Investigation*, 23(2): 213-220.
- Roberta, S.D., Bruna, F.R.F., Sebastião, L.A.G., Maria, L.R.R., Euloir, P., Adriana, C.P.S. (2012). Influence of combined oral contraceptives on the periodontal condition. *Journal of Applied Oral Sciences*, 20: 253-259.
- Radostits, O.M., Gay, C.C., Blood, D.C. (2002). *Veterinary Medicine. A text book of the disease of cattle, sheep, pig, goats and horses*. 13th edition. Billiere Tindall. USA. 545-591.
- Reichlin, S. (1993). Neuroendocrine-immune interactions. *New England Journal of Medicine*, 329: 1246-53.
- Seyffert, N., Guimaraes, A.S., Pacheco, L.G., Portela, R.W., *et al.* (2010). High seroprevalence of caseous lymphadenitis in Brazilian goat herds revealed by *Corynebacterium pseudotuberculosis* secreted proteins-based ELISA. *Research Veterinary Sciences*, 88: 50-55.
- Seddon, H.R., Belschner, H.G. and Rose, A.L. (1992). Further observations on the method of infection in caseous lymphadenitis in sheep. *Australian Veterinary Journal*, 5: 139-149.
- Songer, J.G., Beckenbach, K., Marshall, M.M., Olson, G.B. and Kelley, L. (1988). Biochemical and genetic characterization of *Corynebacterium pseudotuberculosis*. *American Journal of Veterinary Research*, 49: 221-226.
- Stefanska, I., Rzewuska, M., Binek, M. (2008) Evaluation of three methods for DNA fingerprinting of *Corynebacterium pseudotuberculosis* strain isolated from goats in Poland. *Polish Journal of Microbiology*, 57: 105-112.
- Stoops, S.G., Renshaw, H.W. and Thilsted, J.P. (1984). Ovine caseous lymphadenitis: disease prevalence, lesion distribution, and thoracic manifestations in a population of mature culled sheep from western United States. *American Journal of Veterinary Research*, 45: 557-561.
- Sutherland, S.S., Ellis, T.M., Mercy, A.R., Paton, M. and Middleton, H. (1987). Evaluation of an enzyme-linked immunosorbent assay for the detection of *Corynebacterium pseudotuberculosis* infection in sheep. *Australian Veterinary Journal*, 64: 263-266.

- Standford, K., Brogden, K., McClelland, L.A., Kozub, G.C., and Audibert, F.(1998).The incidence of caseous lymphadenitis in Alberta sheep and assessment of impact by vaccination with commercial and experimental vaccines. *Canadian Journal of Veterinary Research*,62:38-43.
- Silver,M. (1953).A quantitative analysis of the role of oestrogen in mammary development in the rat. *Journal of Endocrinology*, 10:17–34.
- Shigidi, M.T.(1979).A comparison of five serological tests for the diagnosis of experimental *Corynebacterium Ovis* infection in sheep.*British Veterinary Journal*, 135:172-177.
- Simmons, C.P., Dunstan, S.J., Tachedjian, M., Krywult, J., Hodgson, A.L., and Strugnell, R.A.(1998). Vaccine potential of attenuated mutants of *Corynebacterium pseudotuberculosis* in sheep. *Infection and Immunity*,66:474-479.
- Sting, R.,Steng, G., and Spengler, D. (1998).Serological studies on *Corynebacterium pseudotuberculosis* infections in goats using enzyme-linked immunosorbent assay. *Journal of Veterinary Medicine Series B*, 45:209-216.
- Siitiri,P.K., and Seron-Ferren, M.(1981).Some new though ts on the fetoplacenta unit and parturition in primate: In Novy MJ and Resko JA (eds) fetal endocrinology. Academic press, New York.1-34.
- Tausk, M., De Visser, J.(1971).Effects of progesterone on the structure and functions of the Fallopian tubes.In: Tausk, M., ed. *Pharmacology of the endocrine system and related drugs: progesterone, progestational drugs and antifertility agents*. Vol. 1. New York, Pergamon Press, (International Encyclopedia of Pharmacology and Therapeutics, Section 48).251-263.
- The Writing Group for the PEPI Trial (1996) *Journal of American Medical Association*, 276:1389–1396.
- The Writing Group for the PEPI Trial (1995) *J Am Med Assoc* 273:199–208.
- Tashjian, J.J. and Campbell, S.G. (1983). Interaction between caprine macrophages and *Corynebacterium pseudotuberculosis*: an electron microscopic study. *American Journal of Veterinary Research*, 44:690-693.
- Verhoeven, G., Cailleau, J., Van Damme, J., Billiau, A.(1988). Interleukin-1 stimulates steroidogenesis in cultured rat Leydig cells. *Molecular Cell Endocrinology*, 57: 51-60.
- Valli, V.E.O. And Parry, B.W.(1993).Caseous lymphadenitis, In:*Pathology of Domestic Animals*, Vol.3 4<sup>th</sup> Edit., K.V.F. Jubb,P.C. Kennedy AND n.Palmer,,Eds,Academic Press, San Diego, 238-240.



Windsor P. A. (2011). Control of caseous lymphadenitis. The Veterinary Clinics of North America. Food Animal Practice, 27: 193–202.

Williamson, L.H.(2001).Clinical Small Ruminants.Veterinary Clinical North American,17 :359–371.

Welschen, R., Osman, P., Dullaart, W.J.DeGreef, J.TH.,Ulenbroek, J. and De Jong, F.H.(1975). Levels of follicles-stimulating hormone, luteinizing hormone,oestradiol-17 $\beta$  and progesterone, and follicular growth in the pseudopregnancy rat.Journal of Endocrinology, 1, 64:37-47.

Yozwiak, M.L., and Songer, J.G. (1993). Effect of *Corynebacterium pseudotuberculosis* phospholipase D on viability and chemotactic responses of ovine neutrophils. American Journal of Veterinary Research, 54: 39-397.

Zaid, K.,Abdulnasir,Y.O.,Jesse, F. F. A.,Haron, A. W., Saharee, A.A., Sabri, J.,Yusoff, R. and Abdullahi,R. (2012). Sex hormone profiles and cellular changes of reproductive organs of mice experimentally infected with *C.Pseudotuberculosis* and its exotoxin phospholipase D (PLD). Journal of Agriculture and Veterinary Sciences, I: 24-29.

Zychlinsky, A.B.,Fitting, J.M., Vaillon, C.andSansoneetti, P.J. (1994).Interleukin-1 is released by macrophages during apoptosis induced by Shigella flexneri. Journal of Clinical Investigation,94:1328-1332.