

UNIVERSITI PUTRA MALAYSIA

ISOLATION AND CHARACTERISATION OF TRANSCRIPTS FOR Leafy, Globosa AND Drooping Leaf TRANSCRIPTION FACTORS FROM NORMAL AND MANTLED OIL PALM INFLORESCENCE

SHARMILAH VETARYAN

FBSB 2018 46



ISOLATION AND CHARACTERISATION OF TRANSCRIPTS FOR Leafy, Globosa AND Drooping Leaf TRANSCRIPTION FACTORS FROM NORMAL AND MANTLED OIL PALM INFLORESCENCE

By

SHARMILAH VETARYAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

March 2018

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

ISOLATION AND CHARACTERISATION OF TRANSCRIPTS FOR Leafy, Globosa AND Drooping Leaf TRANSCRIPTION FACTORS FROM NORMAL AND MANTLED OIL PALM INFLORESCENCE

By

SHARMILAH VETARYAN

March 2018

Chair : Parameswari Namasivayam, PhD Faculty : Biotechnology and Biomolecular Sciences

Flowering is a crucial process in plants to ensure the continuity of species. It is a highly regulated process of which transcription factors play a major role from initiation of the reproductive development until production of fruits. In oil palm clonal planting materials, somaclonal variation in flower development was observed, notably mantling abnormality in which feminization of male flower structures occur. Consequently, production of fruit is affected and oil yield is jeopardized. The objective of this study was to investigate the possible function and also association of LEAFY(LFY), GLOBOSA (GLO) and DROOPING LEAF(DL) transcription factor genes with the mantling phenomena. Transcripts and transcript variants of these genes were isolated and their expression patterns were characterised in oil palm organs, developing female inflorescence, as well as normal and mantled inflorescence via quantitative PCR. Furthermore, selected transcripts were analysed with RNA in situ hybridisation. Two transcripts each of EgLFY, EgGLO and EgDL were isolated. Additionally, a splice variant was isolated for both EgLFY and EgGLO respectively. Both splice variants, EgLFY2v and EgGLO1v, have an intron and a premature stop codon. Tissue-specific expression analysis showed that EgLFY, EgGLO and EgDL were expressed in apical meristem tissue, whereas EgLFY and EgGLO are expressed in both male and female inflorescences at stage 2 of development. Further analysis on expression trend of transcripts at stage 1 to stage 4 of developing female inflorescence series revealed accumulation of EgLFY transcripts are at its highest at stage 1, whereas EgDL expression begins at stage 3 and increases steeply at stage 4. Meanwhile, the expression of EgGLO was constant throughout the developmental stages. Comparative expression analysis between normal and mantled inflorescence showed that EgLFY1 transcript abundance observed in mantled inflorescence was only 75% of that of in normal inflorescence at stage 1 of development whereas, the expression of EgDL1 and EgDL2 was slightly higher than 2-fold in mantled inflorescence at stage 4 of development. However only differential expression of EgLFY1 was statistically significant. Furthermore, RNA in situ hybridisation of EgDL1 revealed expression of the transcript in the ectopic supplementary carpels of mantled inflorescence. Duplication of genes might have arisen through segmental duplication which is widespread in oil palm species. Expression of EgLFY, EgGLO and EgDL transcripts in apical meristem indicates that



these transcripts may play an additional role in vegetative growth in oil palm. All three TFs also play crucial role during initiation and development of oil palm inflorescence. Based on the qPCR and RNA in situ hybridisation expression results, it was postulated that EgLFY1 and EgDL1 are associated to the mantling abnormality in clonal palms. The findings enabled the identification of transcripts which has the potential to be developed as markers for mantling floral abnormality.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

ISOLASI DAN PENCIRIAN TRANSKRIP UNTUK FAKTOR TRANSKRIPSI Leafy, Globosa DAN Drooping Leaf DARIPADA POKOK KELAPA SAWIT NORMAL DAN MANTLE

Oleh

SHARMILAH VETARYAN

Mac 2018

Pengerusi : Parameswari Namasivayam, PhD Fakulti : Bioteknologi dan Sains Biomolekul

Pembungaan adalah proses penting dalam tumbuhan untuk memastikan kemandirian spesies. Ia adalah proses yang terkawal ketat di mana faktor transkripsi memainkan peranan utama daripada permulaan proses pembiakan sehingga penghasilan buah .Di antara bahan tanaman klon kelapa sawit, variasi somaklonal dalam perkembangan bunga telah diperhatikan, terutamanya abnormaliti mantle di mana feminisasi organ reproduktif jantan berlaku. Akibatnya, penghasilan buah dan hasil minyak terjejas.Objektif kajian ini adalah untuk menyiasat fungsi dan juga penglibatan gen faktor transkripsi LEAFY (LFY), GLOBOSA (GLO,) dan DROOPING LEAF (DL) dengan fenomena mantle. Transkrip daripada gen-gen ini telah dipencilkan dan corak ekspresinya telah dicirikan dalam infloresen normal dan mantle melalui PCR kuantitatif. Seterusnya, transkrip yang dipilih telah dianalisis dengan hibridisasi in situ RNA. Dua salinan transkip berbeza daripada EgLFY, EgGLO dan EgDL telah diasingkan dan satu transkrip varian sambatan telah diasingkan daripada EgLFY dan EgGLO. Kedua-dua transkrip varian sambatan, EgLFY2v dan EgGLO1v, mempunyai intron dan kodon penamat prematur. Analisis ekspresi pada tisu pokok kelapa sawit menunjukkan ekspresi EgLFY, EgGLO dan EgDL dalam tisu meristem apikal dan juga ekspresi EgLFY dan EgGLO dalam infloresen jantan dan betina pada tahap 2 perkembangan. Analisis selanjutnya ke atas corak ekspresi transkrip pada tahap 1 hingga tahap 4 dalam perkembangan siri infloresen betina menunjukkan bahawa ekspresi transkripsi EgLFY adalah tertinggi pada tahap 1, manakala ekspresi EgDL bermula pada tahap 3 dan meningkat pada tahap 4. Sementara itu, ekspresi transkrip EgGLO berada pada tahap yang lebih kurang sama sepanjang peringkat perkembangan.Perbandingan analisis ekspresi antara infloresen normal dan mantle menunjukkan bahawa kuantiti transkrip EgLFY1 di dalam infloresen mantle adalah hanya 75% daripada kuantiti transkrip di dalam infloresen normal pada tahap 1 perkembangan, manakala ekspresi transkrip EgDL1 dan EgDL2 adalah lebih sedikit daripada 2 kali ganda di dalam infloresen mantle pada tahap 4 perkembangan. Walau bagaimanapun, hanya perbezaan ekspresi EgLFY1 adalah signifikan secara statistik. Tambahan pula, melalui analisis hibridasi in situ RNA, ekspresi transkrip EgDL1 diperhatikan dalam karpel ektopik tambahan pada infloresen mantle. Dua salinan gen daripada setiap faktor transkripsi mungkin terhasil melalui proses duplikasi segmen

yang berlaku secara meluas dalam spesies kelapa sawit. Ekspresi transkripsi *EgLFY*, *EgGLO* dan *EgDL* dalam meristem apikal menunjukkan bahawa transkrip-transkrip ini memainkan peranan tambahan dalam pertumbuhan vegetatif pokok kelapa sawit. Ketiga-tiga faktor transkripsi tersebut juga memainkan peranan penting semasa permulaan dan perkembangkan infloresen kelapa sawit. Berdasarkan hasil ekspresi qPCR dan *in situ* hibridisasi RNA, terdapat kemungkinan bahawa *EgLFY1* dan *EgDL1* terlibat dengan variasi *mantle* pada pokok kelapa sawit klon. Penemuan ini membolehkan pengenalpastian transkrip yang berpotensi untuk dijadikan sebagai penanda untuk variasi somaklonal *mantle*.



ACKNOWLEDGEMENTS

Firstly, I would like to thank my main supervisor, Dr. Parameswari Namasivayam for her encouragement, assistance and guidance for completion of my Masters work. I would like to also thank my co-supervisor, Dr. Ho Chai Ling, for providing helpful comments and constructive criticisms. A special thanks and sincere appreciation to my external supervisor and mentor, Dr. Kwan Yen Yen for the invaluable guidance, input and consistent motivation provided throughout the study.

My deepest appreciation to FELDA Agricultural Services Sendirian Berhad (FASSB) for funding my research project as well as for generously providing the oil palm samples for the completion of my study.

My sincere thanks to Dr. Ooi Siew Eng and Pn Rosna (MPOB) for sharing their knowledge and technical expertise on *in situ* hybridisation technique with me.

Special thanks to my close friend, Qistina, for always being there for me. Not forgetting my other friends, Suran, Puva, Reena, Bhanisha and Revathi for sharing their experiences and being supportive, thank you all.

Finally, I wish to thank my pillar of strength, my family. The words of support, patience and understanding of my wonderful parents and siblings have motivated me to complete this research. Last but not least, I wish to thank my beloved husband, Rueban for the kind understanding and encouragement throughout this journey.

v

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Parameswari Namasivayam, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Ho Chai Ling, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Kwan Yen Yen, PhD

Lead Researcher FELDA Global Ventures Innovation Centre FELDA Global Ventures Research & Development (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No.:	

Declaration by Members of Supervisory Committee

This is to confirm that:

0

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:
Signature: Name of Member of Supervisory Committee:
Signature: Name of Member of Supervisory Committee:

TABLE OF CONTENTS

ABSTRACT

ACKNOWLEDGEMENTS

ABSTRAK

Page

i

iii

V

Al Di Li Li	PPRO ECLA ST OI ST OI	VAL RATION F TABLES F FIGURES F ABBREVIATIONS	v viii xiii xiv xvi
CI	HAPT	ER	
1	INTI	RODUCTION	1
2	LITI	ERATURE REVIEW	3
	2.1	General introduction of oil palm	3
	2.2	Economic importance of oil palm and challenges in planting material improvement	3
		2.2.1 Tissue culture and somaclonal variation	4
	2.3	Oil palm flower development	4
		2.3.1 Mantled flower phenotype	5
	2.4	2.3.2 Past findings related to mantling	6
	2.4	Transcription factors and gene regulation2.4.1 Role of transcription factors in reproductive development	7 8
		of plants	0
		2.4.2 <i>LEAFY</i> transcription factor	10
		2.4.3 MADS box transcription factor genes	10
		2.4.4 Non-MADS box floral homeotic transcription factor genes	12
	2.5	Gene expression regulation of transcription factors	12
		2.5.1 Spatial and temporal expression pattern	13
		2.5.2 Alternative splicing	13
3	MAT	FERIALS AND METHODS	16
	3.1	Plant materials source	16
	3.2	Sample collection and pre-processing	16
	3.3	RNA extraction from inflorescence and vegetative tissues	18
		3.3.1 RNA quantification, purity and integrity determination	18
	3.4	Amplification of partial coding sequence of OpLFY1	19
		3.4.1 First-strand cDNA synthesis	19
		3.4.2 Determination of optimum annealing temperature	19
		3.4.3 Polymerase chain reaction (PCR) of partial CDS of <i>OpLFY1</i>	19
		3.4.4 Purification of PCR products	20
		3.4.5 Preparation of competent cell	20
		3.4.6 Cloning and transformation of PCR products	21

		3.4.7	Purification of plasmids	21
		3.4.8	Analysis of positive clones	22
		3.4.9	Analysis of sequencing results	22
	3.5	Isolatio	n of full-length coding sequence	22
		3.5.1	Generation of 5' and 3' RACE-ready cDNAs	22
		3.5.2	Rapid amplification of cDNA ends (RACE) of OpLFY1	23
		3.5.3	PCR amplification of transcription factor's full length	23
			CDS	
		3.5.4	Analysis of full length coding sequence	24
		3.5.5	Construction of phylogenetic tree	24
	3.6		sion analysis of the isolated transcription factors'	25
		transcri	•	
		3.6.1	Quantitative PCR (QPCR) primer design	26
		3.6.2	Determination of PCR efficiency and melting curve	27
		5.0.2	analysis	2,
		3.6.3	QPCR reaction	28
		3.6.4	Statistical analysis	29
	3.7		n situ Hybridisation (RISH)	29
	5.7	3.7.1	Designing of hybridization probe	29
		3.7.2	Fixation and embedding of samples	29
		3.7.3	Tissue sectioning	30
		3.7.4	RISH - Pre-treatment, hybridization, washing and	30
		5.7.4	detection	50
			detection	
4	DESI		ND DISCUSSION	32
4	KES 4.1		and Integrity of extracted total RNA	32
	4.1		and integrity of extracted total KIVA on and characterization of $EgLFY1$, $EgLFY2$ and $EgLFY2v$	32
	4.2		sequences (CDS)	32
		4.2.1		32
			Partial and full length CDS analysis of <i>EgLFY</i> transcripts	
		4.2.2	QPCR expression analysis of <i>EgLFY</i> transcripts	38
			4.2.2.1 Organ-specific expression	38
			4.2.2.2 Expression trend in developing female	39
			inflorescence series	
			4.2.2.3 Comparative expression between normal and	40
			mantled female inflorescence	
		4.2.3	RISH of EgLFY1 transcript in normal and mantled	41
			female inflorescence	
	4.3	Isolatio	on and characterization of EgGLO1, EgGLO2 and	43
		EgGLC	D2v CDS	
		4.3.1	Full length CDS analysis of EgGLO transcripts	43
		4.3.2	QPCR expression analysis of <i>EgGLO</i> transcripts	48
			4.3.2.1 Organ specific expression	48
			4.3.2.2 Expression trend in developing female	49
			inflorescence series	
			4.3.2.3 Comparative expression between normal and	51
			mantled female inflorescence	
	4.4	Isolatio	n and characterization of EgDL1 and EgDL2 CDS	52
		4.4.1	Full length CDS analysis of EgDL transcripts	52
		4.4.2	QPCR expression analysis of $EgDL$ transcripts	55
			4.4.2.1 Organ specific expression	55

		4.4.2.2	Expression trend in developing female	56
		4.4.2.3	inflorescence series Comparative expression between normal and mantled female inflorescence	57
	4.4.3	RISH of infloresc	EgDL transcripts in normal and mantled female	58
5	SUMMARY, FUTURE RE		USION AND RECOMMENDATIONS FOR I	60

REFERENCES APPENDICES BIODATA OF STUDENT

C

62 69

85

LIST OF TABLES

	Page
EgLFY1 partial coding sequence primer sequence	20
Gene specific primers sequence for 5' and 3' RACE reaction	23
List of CDS PCR primer sequences	24
Information of the inflorescence used in studying expression trend of genes in developing normal female series	25
Information of the inflorescence used in comparative gene expression studies in normal and mantled inflorescence	26
List of qPCR primer sequences	27
List of RISH LNA probe sequences	29
	Gene specific primers sequence for 5' and 3' RACE reaction List of CDS PCR primer sequences Information of the inflorescence used in studying expression trend of genes in developing normal female series Information of the inflorescence used in comparative gene expression studies in normal and mantled inflorescence List of qPCR primer sequences

G

LIST OF FIGURES

Figure		Page	
2.1	Top five palm oil exporting countries in 2016	4	
2.2	Normal, mantle fertile and mantle parthenocarpic oil palm fruit	6	
2.3	Interaction of site specific transcription factor with general transcription machinery at transcription initiation site	8	
2.4	Schematic representation of flower structure and floral organ identity model of Arabidopsis	9	
2.5	Domain specific expression of flower development related genes and MADS-box gene interactions in floral organ specification	11	
2.6	Spatial expression of <i>DROOPING LEAF</i> gene in superwoman1 (spw1) mutant in rice	12	
2.7	Types and frequency of alternative splicing events in humans and Arabidopsis	14	
2.8	Schematic representation of non-functional heterodimer formation through peptide interference (PEPi) and mRNA degradation through nonsense-mediated decay (NMD)	15	
3.1	Harvested inflorescences from oil palm tree	17	
4.1	Schematic representation of the intron-exon boundary of the isolated <i>EgLFY1</i> , <i>EgLFY2</i> and <i>EgLFY2v</i> transcripts	33	
4.2	Amino acid sequence alignment of predicted EgLFY proteins with LFY from other species	35	
4.3	Phylogenetic tree of predicted LFY protein sequences	37	
4.4	Quantitative PCR of <i>EgLFY1</i> , <i>EgLFY2</i> and <i>EgLFY2v</i> transcripts in vegetative and reproductive organs of oil palm	38	
4.5	Quantitative PCR of <i>EgLFY1</i> , <i>EgLFY2</i> and <i>EgLFY2v</i> transcripts in developing female inflorescence series of oil palm	40	
4.6	Quantitative PCR of <i>EgLFY1</i> transcript in normal and mantled female inflorescence series at stage 1 to stage 3 of development	41	
4.7	<i>In situ</i> localization of <i>EgLFY1</i> transcript in inflorescence tissues at developmental stage 1	42	
4.8	Schematic representation of the intron-exon structure of the isolated <i>EgGLO1</i> , <i>EgGLO1v</i> and <i>EgGLO2</i> transcripts	44	
4.9	Amino acid sequence alignment of predicted EgGLO1 and EgGLO2 proteins with GLO and PI proteins from other species	46	
4.10	Phylogenetic tree of predicted oil palm GLO protein sequences with GLO and PI proteins from other species	47	

4.11	Quantitative PCR of <i>EgGLO1</i> , <i>EgGLO1v</i> and <i>EgGLO2</i> transcripts in vegetative and reproductive organs of oil palm	48
4.12	Quantitative PCR of <i>EgGLO1</i> , <i>EgGLO1v</i> and <i>EgGLO2</i> transcripts in developing female inflorescence series of oil palm	50
4.13	Quantitative PCR of <i>EgGLO1</i> , <i>EgGLO1v</i> and <i>EgGLO2</i> transcripts in normal and mantled female inflorescence	51
4.14	Schematic representation of the intron-exon structure of the isolated <i>EgDL1</i> and <i>EgDL2</i> transcripts	52
4.15	Amino acid sequence alignment of predicted EgDL1 and EgDL2 proteins with DL and CRC proteins from other species	53
4.16	Phylogenetic tree of predicted oil palm DL protein sequences with DL and CRC proteins from other species	54
4.17	Quantitative PCR of <i>EgDL1</i> and <i>EgDL2</i> transcripts in vegetative and reproductive organs of oil palm	55
4.18	Quantitative PCR of <i>EgDL1</i> and <i>EgDL2</i> transcripts in developing female inflorescence series of oil palm	56
4.19	Quantitative PCR of <i>EgDL1</i> and <i>EgDL2</i> transcript in normal and mantled female inflorescence at stage 4 of development	57
4.20	In situ localization of EgDL1 transcript in inflorescence tissues at stage 4	59

C

LIST OF ABBREVIATIONS

AP1	Apetala 1
AS	alternative splicing
ASF	accompanying staminate flower
BLAST	basic local alignment search tool
bp	base pair
cDNA	complementary DNA
CDS	coding sequences
Cq	quantification cycle
DBD	DNA-binding domain
DEPC	Diethyl pyrocarbonate
DNA	deoxyribonucleic acid
EAR	ethylene-responsive element binding factor-associated amphiphilic
	repression
GLO	GLOBOSA
GTFs	general transcription factors
LFY	LEAFY
LNA TM	Locked Nucleic Acid
MADS	MCM1 AGAMOUS DEFICIENS SRF
ORF	open reading frame
QPCR	quantitative PCR
NCBI	National Centre for Biotechnology Information
NMD	non-sense mediated decay
RISH	RNA <i>in situ</i> hybridisation
RACE	rapid amplification of cDNA ends
siPEPs	small interfering peptides
TFs	site specific transcription factors

CHAPTER 1

INTRODUCTION

Elaeis guineensis, also known as the African oil palm, is the most efficient oilbearing crop in the world. One hectare of oil palm yields about 3.7 tons/ha/year of oil and the same area will yield 0.7, 0.5 and 0.4 tons of oil if planted with rapeseed, sunflower and soy, respectively (Sumathi *et al.*, 2008). Requirement for vegetable oil worldwide is predicted to be 240 Mt in the year 2050. Due to increased demand for edible oil and its potential to be used as raw material for biofuel production in future, and also limited availability of land for expansion of oil palm planting, it is necessary to increase the oil yield productivity (Corley, 2009). Hence, high yielding hybrid *tenera* oil palm planting materials are planted to maximize the productivity of each acreage. However, advanced commercial hybrids exhibited low heritability for oil yield (Corley & Tinker, 2003). To address the yield gap in field, one of the strategies is to plant clonal planting materials that have the potential to produce genetically uniform palms that could increase up to 30% yield compared to hybrids (Mutert & Fairhurst, 1999).

In vitro propagation is capable of multiplying a single elite palm into tens of thousands of ramets. However, incidence of mantled somaclonal variation remains the headwind for planters to embrace clonal planting materials commercially. Mantled somaclonal variation is characterized by the feminization of male counterparts in the flower, resulting in abnormal fruits containing supplementary carpel structures. Mantled phenotype is observed in 5% of the tissue-culture derived regenerants, affecting oil yields due to bunch failure (Jaligot *et al.*, 2000). At the pre-nursery stage, an oil palm ramet is sold at approximately RM 22. Taking into consideration the price of ramet only, a 5% rate of mantling will result in RM 1.1 million loss in a company with 1 million production of ramet/year.

Recently, differences in the methylation status of *Karma* transposon located in the intron of *DEFICIENS* (*DEF*) gene have been identified as the culprit for mantled somaclonal variation. Loss of methylation of *Karma* region has introduced a new splicing site at *DEF* gene transcript that resulted in the production of a truncated transcript, *kDEF* (Ong-Abdullah *et al.*, 2015). Expression of *kDEF* is restricted only on mantled inflorescence at stage 3 onwards, coinciding with initiation of the floral reproductive organs. However, not much information is available on what triggers the hypomethylation of *Karma* element which leads to this abnormality. Therefore, further investigations are necessary in order to identify other players involved in contributing to mantled condition to obtain a better understanding of the phenomena.

In line with the objective, three candidate genes which are involved in flower structure formation, *LEAFY (LFY)*, *GLOBOSA (GLO)* and *DROOPING LEAF (DL)*

G

were selected to be analysed in the present study. LFY gene functions both as a meristem identity gene and an upstream regulator of MADS-box genes (Liljegren *et al.*, 1999). *GLO* is a member of B-class MADS-box gene, necessary for specification of reproductive organs, while *DL* is required for specification of carpel structure (Yamaguchi *et al.*, 2004). Full-length transcripts and splice variants of these genes were isolated and the gene expression were characterized in oil palm. This provided an opportunity to both investigate the possible function of these genes in oil palm flower development as well as identify association of the genes in mantling abnormality. This fundamental study on the candidate genes comprises of the following objectives.

- I. To isolate *EgLFY*, *EgGLO* and *EgDL* transcripts and associated splice variant transcripts;
- II. To profile the isolated transcripts' expression level in different oil palm organs, inflorescence developmental stages and comparative expression level in both normal and mantled inflorescence;
- III. To determine the localization of selected transcripts in normal and mantled inflorescence through *in situ* hybridization.

REFERENCES

- Adam, H., Jouannic, S., Morcillo, F., Verdeil, J.-L., Duval, Y., & Tregear, J. W. (2007). Determination of flower structure in Elaeis guineensis: do palms use the same homeotic genes as other species? *Annals of Botany*, 100(1), 1–12.
- Adam, H., Jouannic, S., Morcillo, F., Richaud, F., Duval, Y., & Tregear, J. W. (2006). MADS box genes in oil palm (Elaeis guineensis): patterns in the evolution of the SQUAMOSA, DEFICIENS, GLOBOSA, AGAMOUS, and SEPALLATA subfamilies. *Journal of Molecular Evolution*, 62(1), 15-31.
- Adam, H., Jouannic, S., Escoute, J., Duval, Y., Verdeil, J. L., & Tregear, J. W. (2005). Reproductive developmental complexity in the African oil palm (Elaeis guineensis, Arecaceae). *American Journal of Botany*, 92(11), 1836-1852.
- Ali, G. S., & Reddy, A. S. N. (2008) Regulation of alternative splicing of premRNAs by stresses. *Nuclear pre-mRNA Processing in Plants*. Springer Berlin Heidelberg, 257-275.
- Alwee, S. S., Van der Linden, C. G., Van der Schoot, J., De Folter, S., Angenent, G. C., Cheah, S. C., & Smulders, M. J. M. (2006). Characterization of oil palm MADS box genes in relation to the mantled flower abnormality. *Plant Cell*, *Tissue and Organ Culture*, 85(3), 331-344.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology*, 109(4), 289-295.
- Benlloch, R., Roque, E., Ferrándiz, C., Cosson, V., Caballero, T., Penmetsa, R.V., Beltrán, J.P., Cañas, L.A., Ratet, P. & Madueño, F. (2009). Analysis of B function in legumes: PISTILLATA proteins do not require the PI motif for floral organ development in Medicago truncatula. *The Plant Journal*, 60(1), 102-111.
- Berbel, A., Navarro, C., Ferrándiz, C., Cañas, L. A., Beltrán, J. P., & Madueño, F. (2005). Functional conservation of PISTILLATA activity in a pea homolog lacking the PI motif. *Plant physiology*, 139(1), 174-185.
- Besse, I., VERDEIL, J. L., Duval, Y., Sotta, B., Maldiney, R., & Miginiac, E. (1992). Oil palm (Elaeis guineensis Jacq.) clonal fidelity: endogenous cytokinins and indoleacetic acid in embryogenic callus cultures. *Journal of Experimental Botany*, 43(7), 983-989.
- Bomblies, K., Wang, R. L., Ambrose, B. A., Schmidt, R. J., Meeley, R. B., & Doebley, J. (2003). Duplicate FLORICAULA/LEAFY homologs zfl1 and zfl2 control inflorescence architecture and flower patterning in maize. *Development*, 130(11), 2385-2395.

Bowman, J. L., & Smyth, D. R. (1999). CRABS CLAW, a gene that regulates carpel

and nectary development in Arabidopsis, encodes a novel protein with zinc finger and helix-loop-helix domains. *Development*, 126(11), 2387-2396.

- Carlberg C., Molnár F. (2014) The Basal Transcriptional Machinery. In: Mechanisms of Gene Regulation. Springer, Dordrecht
- Coen, E. S., & Meyerowitz, E. M. (1991). The war of the whorls: genetic interactions controlling flower development. *Nature*, 353(6339), 31.
- Corley, R. H. V. (2009). How much palm oil do we need?. *Environmental Science* & *Policy*, 12(2), 134-139.
- Corley, R. H. V., & Tinker, B. (2003). *The oil palm 4th edition*. John Wiley and Sons, Hoboken, NJ.
- Corley, R. H. V. (1998). What is the upper limit to oil extraction ratio?. In International Conference on Oil and Kernel production in oil palm. A global perspective. September 27-28.Kuala Lumpur, Malaysia (No. L-0372). PORIM.
- Danilevskaya, O. N., Meng, X., Selinger, D. A., Deschamps, S., Hermon, P., Vansant, G., Gupta, R., Ananiev, E. V., & Muszynski, M. G. (2008). Involvement of the MADS-box gene ZMM4 in floral induction and inflorescence development in maize. *Plant physiology*, 147(4), 2054-2069.
- Fitzherbert, E. B., Struebig, M. J., Morel, A., Danielsen, F., Brühl, C. A., Donald, P. F., & Phalan, B. (2008). How will oil palm expansion affect biodiversity?. *Trends in ecology & evolution*, 23(10), 538-545.
- Goto, K., & Meyerowitz, E. M. (1994). Function and regulation of the Arabidopsis floral homeotic gene PISTILLATA. *Genes & Development*, 8(13), 1548-1560.
- Guo, S., Sun, B., Looi, L. S., Xu, Y., Gan, E. S., Huang, J., & Ito, T. (2015). Coordination of flower development through epigenetic regulation in two model species: rice and Arabidopsis. *Plant and Cell Physiology*, 56(5), 830-842.
- Hamès, C., Ptchelkine, D., Grimm, C., Thevenon, E., Moyroud, E., Gérard, F., Martiel, J., Benlloch, R., Parcy, F., Müller, C. W. (2008). Structural basis for LEAFY floral switch function and similarity with helix-turn-helix proteins. *The EMBO Journal*, 27(19), 2628–2637.
- Honma, T., & Goto, K. (2000). The Arabidopsis floral homeotic gene PISTILLATA is regulated by discrete cis-elements responsive to induction and maintenance signals. *Development*, 127(10), 2021-2030.

Huala, E., & Sussex, I. M. (1992). LEAFY interacts with floral homeotic genes to regulate Arabidopsis floral development. *The Plant Cell*, 4(8), 901-913.

- Jaligot, E., Rival, A., Beulé, T., Dussert, S., & Verdeil, J. L. (2000). Somaclonal variation in oil palm (Elaeis guineensis Jacq.): the DNA methylation hypothesis. *Plant cell reports*, 19(7), 684-690.
- Kang, H. G., Jeon, J. S., Lee, S., & An, G. (1998). Identification of class B and class C floral organ identity genes from rice plants. *Plant molecular biology*, 38(6), 1021-1029.
- Kagale, S., & Rozwadowski, K. (2011). EAR motif-mediated transcriptional repression in plants. *Epigenetics*, 6(2), 141–146.
- Kelly, A. J., Bonnlander, M. B., & Meeks-Wagner, D. R. (1995). NFL, the tobacco homolog of FLORICAULA and LEAFY, is transcriptionally expressed in both vegetative and floral meristems. *The Plant Cell*, 7(2), 225-234.
- Kubis, S. E., Castilho, A. M., Vershinin, A. V., & Heslop-Harrison, J. S. P. (2003). Retroelements, transposons and methylation status in the genome of oil palm (Elaeis guineensis) and the relationship to somaclonal variation. *Plant molecular biology*, 52(1), 69-79.Kumaran, M. K., Bowman, J. L., & Sundaresan, V. (2002). YABBY polarity genes mediate the repression of KNOX homeobox genes in Arabidopsis. *The Plant Cell*, 14(11), 2761-2770.
- Kushairi, A. Malaysian Oil Palm Industry Performance 2016 and Prospects for 2017. Presented at Palm Oil Economic Review & Outlook Seminar 2017, Kuala Lumpur.
- Kushairi, A., Tarmizi, A. H., Zamzuri, I., Ong-Abdullah, M., Samsul Kamal, R., Ooi, S. E., & Rajanaidu, N. (2010). Production, performance and advances in oil palm tissue culture. In *International Seminar On Advances In Oil Palm Tissue Culture*. Yogyakarta.
- Kwon, Y. J., Park, M. J., Kim, S. G., Baldwin, I. T., & Park, C. M. (2014). Alternative splicing and nonsense-mediated decay of circadian clock genes under environmental stress conditions in Arabidopsis. *BMC plant biology*, 14(1), 136.
- Lange, M., Orashakova, S., Lange, S., Melzer, R., Theißen, G., Smyth, D.R. and Becker, A. (2013). The seirena B class floral homeotic mutant of California Poppy (Eschscholzia californica) reveals a function of the enigmatic PI motif in the formation of specific multimeric MADS domain protein complexes. *The Plant Cell*, 25(2), 438-453.
- Liljegren, S. J., Gustafson-Brown, C., Pinyopich, a, Ditta, G. S., & Yanofsky, M. F. (1999). Interactions among APETALA1, LEAFY, and TERMINAL FLOWER1 specify meristem fate. *The Plant Cell*, 11(6), 1007–1018.
- Liu, L., White, M. J., & Macrae, T. H. (1999). Transcription factors and their genes in higher plants functional domains, evolution and regulation. *European Journal of Biochemistry*, 262(2), 247–257.

- Lohmann, J. U., & Weigel, D. (2002). Building beauty: the genetic control of floral patterning. *Developmental Cell*, 2(2), 135–42.
- Long, J. C., & Caceres, J. F. (2009). The SR protein family of splicing factors: master regulators of gene expression. *Biochemical Journal*, 417(1), 15-27.
- Masek, T., Vopalensky, V., Suchomelova, P., & Pospisek, M. (2005). Denaturing RNA electrophoresis in TAE agarose gels. *Analytical Biochemistry*, 336(1), 46-50.
- Menzel, G., Apel, K., & Melzer, S. (1996). Identification of two MADS box genes that are expressed in the apical meristem of the long-day plant Sinapis alba in transition to flowering. *The Plant Journal*, 9(3), 399-408.
- Mitsuda, N., & Ohme-Takagi, M. (2009). Functional analysis of transcription factors in Arabidopsis. *Plant and Cell Physiology*, 50(7), 1232-1248.
- Mutert, E., & Fairhurst, T. H. (1999). Oil palm clones: Productivity enhancement for the future. *Better Crops International*, 13(1), 45.
- Nieto Moreno, N., Giono, L. E., Botto, C., Adrián, E., Muñoz, M. J., & Kornblihtt, A. R. (2015). Chromatin, DNA structure and alternative splicing. *FEBS letters*, 589(22), 3370-3378.
- Moyroud, E., Kusters, E., Monniaux, M., Koes, R., & Parcy, F. (2010). LEAFY blossoms. *Trends in Plant Science*, 15(6), 346-352.
- Malaysian Palm Oil Council, Soaring with a vision; MPOC Annual report: Kuala lumpur, 2016.
- Nagasawa, N., Miyoshi, M., Sano, Y., & Satoh, H. (2003). SUPERWOMAN1 and DROOPING LEAF genes control floral organ identity in rice. *Development*, 130(4), 705–718.
- Ohmori, Y., Toriba, T., Nakamura, H., Ichikawa, H., & Hirano, H. Y. (2011). Temporal and spatial regulation of DROOPING LEAF gene expression that promotes midrib formation in rice. *The Plant Journal*, 65(1), 77-86.
- Ooi, S. E., Ramli, Z., Alwee, S. S. R. S., Kulaveerasingam, H., & Ong-Abdullah, M. (2016). EgHOX1, a HD-Zip II gene, is highly expressed during early oil palm (Elaeis guineensis Jacq.) somatic embryogenesis. *Plant Gene*, 8, 16-25.
- Ong-Abdullah, M., Ordway, J. M., Jiang, N., Ooi, S. E., Kok, S. Y., Sarpan, N., Azimi, N., Hashim, A.T., Ishak, Z., Rosli, S.K. & Malike, F. A. (2015). Loss of Karma transposon methylation underlies the mantled somaclonal variant of oil palm. *Nature*, 525(7570), 533.
- Orashakova, S., Lange, M., Lange, S., Wege, S., & Becker, A. (2009). The CRABS CLAW ortholog from California poppy (Eschscholzia californica,

Papaveraceae), EcCRC, is involved in floral meristem termination, gynoecium differentiation and ovule initiation. *The Plant Journal*, 58(4), 682-693.

- Rotem, N., Shemesh, E., Peretz, Y., Akad, F., Edelbaum, O., Rabinowitch, H.D., Sela, I. and Kamenetsky, R. (2007). Reproductive development and phenotypic differences in garlic are associated with expression and splicing of LEAFY homologue gaLFY. *Journal of experimental botany*, 1133-1141.
- Parcy, F. (2004). Flowering: a time for integration. International Journal of Developmental Biology, 49(5-6), 585-593.
- Production of crude palm oil 2016 (n.d.) In Economics and Industry Development Division MPOB. Retrieved from http://bepi.mpob.gov.my/index.php/en/ statistics/production/168-production-2016/746-production-of-crude-oilpalm-2016.html.
- Ramachandran, V., Ong-Abdullah, M., Ho, C. L., Alwee, S. S. R., & Namasivayam, P. (2014). Modified RNA in situ hybridisation protocol for oil palm (Elaeis guineensis Jacq.) fruit and inflorescence. *Journal of Oil Palm Research*, 26(4), 300-307.
- Reddy, A. S. N., Marquez, Y., Kalyna, M., & Barta, A. (2013). Complexity of the alternative splicing landscape in plants. *The Plant Cell*, 25(10), 3657–83.
- Robles, P., & Pelaz, S. (2004). Flower and fruit development in Arabidopsis thaliana. *International Journal of Developmental Biology*, 49(5-6), 633-643.
- Rebbapragada, I., & Lykke-Andersen, J. (2009). Execution of nonsense-mediated mRNA decay: what defines a substrate? *Current opinion in cell biology*, 21(3), 394-402.
- Rival, A., Beule, T., Barré, P., Hamon, S., Duval, Y., & Noirot, M. (1997). Comparative flow cytometric estimation of nuclear DNA content in oil palm (Elaeis guineensis Jacq) tissue cultures and seed-derived plants. *Plant Cell Reports*, 16(12), 884-887.
- Sambrook, J., Fritsch, E. F., & Maniatis, T. (1989). *Molecular cloning: a laboratory manual* (No. Ed. 2). Cold spring harbor laboratory press.
- Seo, P. J., Park, M. J., & Park, C. M. (2013). Alternative splicing of transcription factors in plant responses to low temperature stress: mechanisms and functions. *Planta*, 237(6), 1415-1424.
- Singh, R., Ong-Abdullah, M., Low, E.T.L., Manaf, M.A.A., Rosli, R., Nookiah, R., Ooi, L.C.L., Ooi, S.E., Chan, K.L., Halim, M.A. and Azizi, N. (2013). Oil palm genome sequence reveals divergence of interfertile species in Old and New worlds. *Nature*, 500(7462), 335.

- Siriwardana, N. S., & Lamb, R. S. (2012). The poetry of reproduction: the role of LEAFY in Arabidopsis thaliana flower formation. *The International Journal of Developmental Biology*, *56*(4), 207–221.
- Sumathi, S., Chai, S. P., & Mohamed, A. R. (2008). Utilization of oil palm as a source of renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, 12(9), 2404-2421.
- Sun, W., Huang, W., Li, Z., Lv, H., Huang, H., & Wang, Y. (2013). Characterization of a Crabs Claw gene in basal eudicot species Epimedium sagittatum (Berberidaceae). *International journal of molecular sciences*, 14(1), 1119-1131.
- Sayou, C., Nanao, M.H., Jamin, M., Posé, D., Thévenon, E., Grégoire, L., Tichtinsky, G., Denay, G., Ott, F., Llobet, M.P. & Schmid, M. (2016). A SAM oligomerization domain shapes the genomic binding landscape of the LEAFY transcription factor. *Nature communications*, 7, 11222.
- Thomas, B. S., Kumar, S., & Arel, H. S. (2017). Sustainable concrete containing palm oil fuel ash as a supplementary cementitious material–A review. *Renewable and Sustainable Energy Reviews*, 80, 550-561.
- Thomas, M. C., & Chiang, C.-M. (2006). The General Transcription Machinery and General Cofactors. *Critical Reviews in Biochemistry and Molecular Biology*, 41(3), 105–178.
- Tröbner, W., Ramirez, L., Motte, P., Hue, I., Huijser, P., Lönnig, W. E., Heinz, S., Sommer, Hans., & Schwarz-Sommer, Z. (1992). GLOBOSA: a homeotic gene which interacts with DEFICIENS in the control of Antirrhinum floral organogenesis. *The EMBO Journal*, 11(13), 4693.
- Van der Vossen, H. A. M. (1974). Towards more efficient selection for oil yield in the oil palm (Elaeis guineensis Jacquin) (p. 108). Pudoc.
- Tsai, W. C., Lee, P. F., Chen, H. I., Hsiao, Y. Y., Wei, W. J., Pan, Z. J., ... & Chen, H. H. (2005). PeMADS6, a GLOBOSA/PISTILLATA-like gene in Phalaenopsis equestris involved in petaloid formation, and correlated with flower longevity and ovary development. *Plant and Cell Physiology*, 46(7), 1125-1139.
- Wada, M., Cao, Q. F., Kotoda, N., Soejima, J. I., & Masuda, T. (2002). Apple has two orthologues of FLORICAULA/LEAFY involved in flowering. *Plant molecular biology*, 49(6), 567-577.
- Wang, A. J., Tang, J. F., Zhao, X. Y., & Zhu, L. H. (2008). Isolation of LiLFY1 and its expression in lily (Lilium longiflorum Thunb.). *Agricultural Sciences in China*, 7(9), 1077-1083.

Weigel, D., Alvarez, J., Smyth, D. R., Yanofsky, M. F., & Meyerowitz, E. M.

(1992). LEAFY controls floral meristem identity in Arabidopsis. *Cell*, 69(5), 843–859. doi:10.1016/0092-8674(92)90295-N

- Yamaguchi, T., Nagasawa, N., Kawasaki, S., Matsuoka, M., Nagato, Y., & Hirano, H. Y. (2004). The YABBY gene DROOPING LEAF regulates carpel specification and midrib development in Oryza sativa. *The Plant Cell*, 16(2), 500-509.
- Yeap, W. C., Loo, J. M., Wong, Y. C., & Kulaveerasingam, H. (2014). Evaluation of suitable reference genes for qRT-PCR gene expression normalization in reproductive, vegetative tissues and during fruit development in oil palm. *Plant Cell, Tissue and Organ Culture*, 116(1), 55-66.
- Yuan, L., & Perry, S. E., (*Eds.*). (2011). Plant Transcription Factors: Methods and Protocols. 754. Humana Press, New York.
- Yun, J., Kim, S.-G., Hong, S., & Park, C.-M. (2008). Small interfering peptides as a novel way of transcriptional control. *Plant Signaling and Behavior*, 3(9), 615–617.
- Yu, H., & Goh, C. J. (2000). Identification and characterization of three orchid MADS-box genes of the AP1/AGL9 subfamily during floral transition. *Plant Physiology*, 123(4), 1325-1336.