



UNIVERSITI PUTRA MALAYSIA

**CHARACTERIZATION OF *Colletotrichum truncatum* CP2 AND ITS  
INTERACTION WITH CHILLIES (*Capsicum annuum L.*) DURING  
PATHOGENESIS**

NURUL ATIKA BINTI MOHAMAD REMLI

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**NURUL ATIKA BINTI MOHAMAD REMLI**



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment  
of the requirement for the degree of Doctor of Philosophy

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**January 2018**

**Chairman : Associate Professor Umi Kalsom Md Shah, PhD**  
**Faculty : Biotechnology and Biomolecular Sciences**

Anthracnose caused by *Colletotrichum* species is the most destructive disease of chilli worldwide. It is responsible for worldwide yield losses and could be even more severe without a successful control that still relies on the use of fungicides. Due to the growing concern about environmental and health damages caused by this control, an understanding of the mechanisms leading to the fungal pathogenicity in a particular host is essential for the implementation of effective disease control. This study aimed to investigate the mechanism leading to pathogenesis of *Colletotrichum* species in chilli fruit as little is known about the pathogenicity factor involved in this interaction. Thirty five fungal isolates were isolated from chilli lesions of anthracnose from different geographic locations in Malaysia. The ability of fungal isolates to produce cell wall-degrading enzymes was screened and the best cell wall-degrading enzymes producer was selected for further study. Based on its morphological, biochemical and molecular identification, fungal isolate CP2 was identified as *Colletotrichum truncatum*. Successful inoculation of the *C. truncatum* CP2 on detached chilli fruits proved its pathogenicity and was confirmed to be a primary pathogen of chilli when it successfully infected the chilli fruits. In order to illustrate the infection strategy adopted by *C. truncatum* CP2, the infection process of this fungus in the chilli fruit was characterized using light, scanning and transmission microscope. *C. truncatum* CP2 exhibited a prolonged biotrophic phase of about 48 hour, before switched to necrotrophic phase at approximately 72 hour after inoculation. The first phase of necrotrophy in *C. truncatum* CP2 was characterized by formation of germ tube, appresorium and infectious hyphae. The destructive necrotrophic phase was characterized by formation of sunken lesions and production of numerous acervuli. The role of cell wall-degrading enzymes in facilitating the *C. truncatum* CP2 to colonize the host cell was investigated taking into consideration changes in the morphological and chemical compositions of the chilli fruits. The results of enzymatic activity experiment indicated that polygalacturonase (PG) was

the first cell wall-degrading enzymes detected and the activities obtained were higher ( $0.24\pm0.10$  U/mL) than other enzymes, which appeared later and in lower amount. Significant changes in the pectin (total uronide content increased up to 50.33% - 71.85%) and cellulose contents (decreased to 11.45% - 12.32%) in chilli treated with PG and combination of PG and cellulases showed the main role of these enzymes in facilitating the *C. truncatum* CP2 during pathogenesis in chilli fruits. According to Fourier transform infrared analysis, there were remarkable changes in the vibration side of cellulose (3290 cm<sup>-1</sup> and 2924 cm<sup>-1</sup>) and ring and vibration side of pectin (1581, 1337 and 1029 cm<sup>-1</sup>) in the cell wall of chilli treated with PG and mixture of both enzymes. In order to understand the exact role of PG enzymes in pathogenesis, PG enzymes from *C. truncatum* CP2 was purified using aqueous two phase system. The optimum purification condition of PG was achieved using 22% (w/w) polyethylene glycol and 15% (w/w) sodium citrate comprising crude load of 16% (w/w) at pH 7.0 with addition of 1.0% (w/w) sodium chloride. The necrotizing activity of the crude and purified PG from *C. truncatum* CP2 was then tested on detached chilli fruits. The faster lesion formation on the chilli treated with purified PG had confirmed the involvement of this enzyme in anthracnose of chilli. In conclusion, *C. truncatum* CP2 possess all the features to be termed as a serious anthracnose pathogen with the presence of pathogenicity factors such as PG enzymes. The results from this study provide a better insight into the interaction of *C. truncatum* CP2 and chilli fruits and these findings may be used in the development of efficient disease management strategies in Malaysia.

Abstrak tesis yang dikemukakan kepada Senat of Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENCIRIAN *Colletotrichum truncatum* CP2 DAN INTERAKSINYA DENGAN BUAH CILI (*Capsicum annuum* L.) SEMASA PATOGENESIS**

Oleh

**NURUL ATIKA BINTI MOHAMAD REMLI**

**Januari 2018**

**Pengerusi : Professor Madya Umi Kalsom Md Shah, PhD  
Fakulti : Bioteknologi dan Sains Biomolekul**

Penyakit antraknos yang berpunca daripada spesis *Colletotrichum* merupakan penyebab utama kepada kerosakan cili di seluruh dunia. Ia telah menyebabkan kerugian besar kepada pengusaha cili di seluruh dunia dan mungkin memberi kesan lebih teruk tanpa kawalan yang baik yang kini masih bergantung kepada penggunaan racun kulat. Peningkatan kesedaran terhadap kesan penggunaan racun kulat terhadap kerosakan alam sekitar dan kesihatan telah membawa kepada pentingnya memahami mekanisme terjadinya jangkitan penyakit ini bagi membolehkan strategi kawalan yang lebih berkesan dilaksanakan. Kajian ini dijalankan bagi menyelidik mekanisma yang membawa kepada patogenesiti spesis *Colletotrichum* terhadap buah cili di mana pengetahuan mengenai faktor yang menyumbang kepada patogenesiti ini belum diketahui. Tiga puluh lima pencilan kulat telah dipencarkan daripada lesi antraknos dari lokasi geografi yang berlainan di Malaysia. Keupayaan kulat tersebut untuk menghasilkan enzim yang terbaik telah disaring dan pengeluar enzim yang terbaik dipilih untuk kajian selanjutnya. Pencilan kulat CP2 diperiksa berdasarkan ciri-ciri morfologi, molekul dan patogenesiti. Berdasarkan keputusan yang diperoleh, pencilan kulat CP2 dikenalpasti sebagai *Colletotrichum truncatum*. Ujian patogenesiti menunjukkan bahawa *C. truncatum* CP2 adalah patogenik terhadap perumah asal dan dikenalpasti sebagai penyebab utama kepada antraknos cili. Bagi mengetahui strategi jangkitan yang diaplikasikan oleh *C. truncatum* CP2, proses jangkitan kulat ini pada cili dikaji menggunakan mikroskop cahaya, mikroskop elektron pengimbasan dan mikroskop elektron penghantaran. *C. truncatum* CP2 mempamerkan fasa biotropik selama 48 jam, sebelum beralih kepada fasa nekrotropik bermula 72 jam selepas inokulasi. Fasa biotrofi dalam *C. truncatum* CP2 dapat dikenalpasti melalui pembentukan tiub germa, appresorium dan hyphae. Fasa nekrotropi pula dapat dikenalpasti melalui pembentukan lesi dan penghasilan acervuli yang banyak pada buah cili. Peranan enzim pengurai dinding sel di dalam membantu *C. truncatum* CP2 untuk koloniasi perumah asal dikenalpasti dengan mengambil kira perubahan dalam komposisi morfologi dan komposisi kimia cili.

Keputusan eksperimen aktiviti enzimatik menunjukkan bahawa polygalakturonase (PG) merupakan enzim pengurai dinding sel pertama yang dikesan dan aktiviti yang diperoleh lebih tinggi ( $0.24 \pm 0.10$  U / mL) daripada enzim lain, yang kemudiannya muncul dan dalam jumlah yang lebih rendah. Perubahan penting dalam pektin (jumlah kandungan uronida meningkat dari 50.33% - 71.85%) dan kandungan selulosa (menurun dari 11.45% - 12.32%) dalam cili yang dirawat dengan PG dan gabungan PG dan selulase menunjukkan peranan utama enzim ini dalam membantu *C. truncatum* CP2 semasa patogenesis pada buah cili. Menurut analisis inframerah transformasi Fourier, terdapat perubahan yang luar biasa pada getaran selulosa (3290 cm<sup>-1</sup> dan 2924 cm<sup>-1</sup>) dan getaran pektin (1581, 1337 dan 1029 cm<sup>-1</sup>) di dinding sel cili dirawat dengan PG dan campuran kedua-dua enzim. Untuk memahami peranan sebenar enzim PG dalam patogenesis, enzim PG dari *C. truncatum* CP2 ditulenkan menggunakan sistem penulenan dwi-fasa. Keadaan penulenan optimum PG dicapai melalui penggunaan 22% (w/w) polietilen glikol dan 15% (w/w) sodium sitrat yang mengandungi enzim sebanyak 16% (w/w) pada pH 7.0 dengan tambahan 1.0% (w/w) natrium klorida. Kebolehan enzim PG yang telah ditulenkan untuk menguraikan buah cili kemudiannya diuji. Pembentukan lesi lebih cepat pada cili yang dirawat dengan PG yang ditulenkan telah mengesahkan penglibatan enzim ini dalam antraknos cili. Sebagai kesimpulan, *C. truncatum* CP2 mempunyai ciri-ciri yang boleh dianggap sebagai patogen antraknos yang serius dengan kehadiran faktor-faktor virulensi seperti enzim PG. Hasil daripada kajian ini memberi gambaran yang lebih baik mengenai interaksi *C. truncatum* CP2 dan buah cili dan dapatan kajian ini boleh digunakan dalam pembangunan strategi pengurusan penyakit yang lebih baik di Malaysia.

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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Umi Kalsom Md Shah, PhD**

Associate Professor

Faculty Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

**Arbakariya B. Ariff, PhD**

Professor

Faculty Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

**Mohd Termizi B. Yusof, PhD**

Senior Lecturer

Faculty Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

**Jeffrey Lim Seng Heng, PhD**

Strategic Resources Research Centre

Malaysian Agricultural Research and Development Institute (MARDI)

Malaysia

(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

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Signature: \_\_\_\_\_  
Name of  
Chairman of  
Supervisory  
Committee: Associate Professor Dr. Umi Kalsom Md Shah

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: Professor Dr. Arbakariya B. Ariff

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: Dr. Mohd Termizi B. Yusof

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: Jeffrey Lim Seng Heng

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## LIST OF ABBREVIATIONS

ADF	Acid detergent fiber
ADL	Acid detergent lignin
ANOVA	Analysis of Variance
Ap	appresorium
ATPS	Aqueous two phase system
ATR	Attenuate total reflectance
C	conidium
Cu	cuticle
CBM	Carbohydrate binding module
CMC	Carboxy methyl cellulose
DNA	Deoxyribonucleic acid
DNS	Dinitrosalicylic acid
FAO	Food and agriculture organization
FAOSTAT	The statistics division of food and agriculture organization
FTIR	Fourier Transform Infrared
Hai	hour after inoculation
g	gram
mg	milligram
$\mu\text{g}$	microgram
g/g	Gram per gram substrate
g/L	Gram per liter
GF	gel filtration
$\text{H}_2\text{SO}_4$	Sulphuric acid
HCl	Hydrochloric acid
IU	International unit
Ha	acre
HAI	hour after inoculation
Hg/ha	Hectogram per acre
IEX	Ion exchange
ITS	Internal transcribed spacer
KCl	Potassium chloride

$\text{KH}_2\text{PO}_4$	Potassium dihydrogen phosphate
$\text{K}_2\text{HPO}_4$	Potassium hydrogen phosphate
kV	kiloVolt
$\text{MgCl}_2$	Magnesium chloride
$\text{MgSO}_4$	Magnesium sulphate
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	Magnesium sulphate heptahydrate
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	Magnesium sulphate hydrate
mm	milimetre
MOA	Ministry of agriculture
$\text{NaNO}_3$	Sodium nitrate
$\text{NaOCl}$	Sodium hypochlorite
$\text{NaOH}$	Sodium hydroxide
NCBI	National center for biotechnology information
NDF	Neutral detergent fiber
PCR	Polymerase chain reaction
PDA	Potato dextrose agar
PEG	Polyethylene glycol
PG	polygalacturonase
PL	pectin lyase
PNL	pectate lyase
PME	pectin methylesterases
pp	penetration peg
rRNA	Ribosomal ribonucleic acid
SEM	Scanning electron microscope
$\text{T ha}^{-1}$	Tons per acre
TEM	Transmission electron microscope
UF	Ultrafiltration
v/v	volume per volume
UV	Ultraviolet
w/v	weight per volume
%	percent
$^{\circ}\text{C}$	Degree celcius

# CHAPTER 1

## INTRODUCTION

### 1.1 General background

*Capsicum annuum* L. is commonly known as chilli, or chilli pepper has been a part of Malaysian diet. Chilli fruits often used as a spice added to various dishes due to their hot flavour. However, the production of chilli in Malaysia is often hampered by fungal diseases (Ali *et al.*, 2014; Heng *et al.*, 2011; Yun *et al.*, 2009). Anthracnose disease caused by *Colletotrichum* species is one of the main causes for post-harvest fruit rot of chilli and resulted in high yield losses. It can be developed on the field, during long distant transport and cold storage and these attributed to the lower fruit quality and marketability. According to FAO report (2016), area harvested with chilli in Malaysia in the year 2013 was estimated to be around 4,104 ha with estimated yield of 145,651 Hg/Ha (FAOSTAT, 2014). However, this number decreased to 3,582 ha in the year 2014 with the total estimated yield of 113,124 Hg/Ha. The statistical data has showed a reduction of approximately 22% of the yield and hence indicating the severity of anthracnose disease in chilli. Higher yield loss due to anthracnose disease will be faced by farmers if the disease is not well managed (Stanley Freeman, 2008). Such losses would increase the market price of chilli and ultimately affect both growers and consumers.

In conventional agriculture, control of chilli anthracnose at post-harvest stage is usually done by spraying the whole plant including the fruits with synthetic fungicides. The synthetic fungicides such as manganese ethylenebisdithiocarbamate (Maneb) and carbendazim are generally recommended for controlling anthracnose disease. However in recent years, the consumers' concern regarding the environmental and health damage associated with the use synthetic fungicides have increased. Furthermore, the demand for safer storage methods and vegetables free from pesticide residues by worldwide consumers is growing day by day. As a result, the pesticide legislation is becoming stricter and only limited number of fungicides are allowed for post-harvest usages. Moreover, the development of fungicide resistance pathogen due to frequent use of fungicides causes difficulty to control the anthracnose disease. Therefore, there is a need to search for another alternative control method that is considered safe for human health and environment.

In order to develop effective disease management strategies, an understanding of the mechanism leading to fungal pathogenicity on a particular host is essential. In general, plant pathogenic fungi form infection structures that accomplish crucial stages of pathogenesis including adhesion, penetration, proliferation and nutrient. *Colletotrichum* species use different strategies to attack host tissues, either through an intracellular hemibiotrophic strategy, a subcuticular intramural strategy or a combination of both strategies (Kubo *et al.*, 2016). Most members of the genus

*Colletotrichum* infected the host cells through intracellular hemibiotroph (Wharton *et al.*, 2001). Through this method, they initially colonized the host biotrophically, where the fungi obtained nutrients from living plant cells, and then they turned into necrotroph, where the fungi cause an extensive degradation of host cells.

The infection of host plants by fungal pathogens is usually mediated by numerous cell wall-degrading enzymes. The secretion of cell wall-degrading enzymes by fungal pathogens is very important as it contributes to the pathogenicity of fungal pathogens during interaction with the host (Fernando *et al.*, 2001; Kikot *et al.*, 2009). Among the various cell wall-degrading enzymes released by fungal pathogens, most research has been focused on pectinases enzyme. Earlier studies have reported that pectinases are the only enzymes that have the ability to cause tissue maceration and cell death. However, some of other cell wall-degrading enzymes such as cellulases, chitinases and xylanases have also been reported as the pathogenic determinants in disease development (Kubicek *et al.*, 2014).

Even though some *Colletotrichum* species have been identified as causal agents of chilli anthracnose disease worldwide, the information on the mechanism of infection and disease development of *Colletotrichum* species on chilli is still lacking. The extracellular enzymes profile which may have potential roles in pathogenesis of *Colletotrichum* during anthracnose infection is also unknown. A view on the strategy of infection used by pathogens along with the secretion of cell wall-degrading enzymes occur in the chilli fruits during infection would provide valuable information needed to develop a new control strategy for chilli anthracnose.

## 1.2 Objectives

The general objective of this study was to investigate the mechanism leading to pathogenesis of *Colletotrichum* species during its interaction with chilli fruit.

The specific objectives of the present investigation were as follow:

1. To isolate and identify *Colletotrichum* species isolated from lesions of chilli anthracnose using morphological, biochemical and molecular characterization.
2. To characterize the pre- and penetration process of *C. truncatum* CP2 on chilli fruit during pathogenesis.
3. To evaluate the cell wall-degrading enzymes produced by *C. truncatum* CP2 during pathogenesis on chilli.
4. To characterize the properties of cell wall-degrading enzymes produced by *C. truncatum* CP2.

## REFERENCES

- A'fifah, A. R., Ismail, M. R., Puteri, E. M. W., Abdullah, S. N. A., Berahim, Z., Bakhtiar, R., & Kausar, H. (2015). Optimum fertigation requirement and crop coefficients of chilli (*Capsicum annuum*) grown in soilless medium in the tropic climate. *International Journal of Agriculture and Biology*, 17(1), 80–88.
- Abbasiliasi, S., Tan, J. S., Ibrahim, T. A. T., Kadkhodaei, S., Ng, H. S., Vakhshiteh, F., Ariff, A. B. (2014). Primary recovery of a bacteriocin-like inhibitory substance derived from *Pediococcus acidilactici* Kp10 by an aqueous two-phase system. *Food Chemistry*, 151, 93–100.
- Aboaba, S. (2009). The role of pectinase enzyme in the development of soft rot caused by *Pseudomonas fluorescens* in the purple variety of onions (*Allium cepa*). *African Journal of Microbiology Research*, 3(4), 163–167.
- Acosta-Rodríguez, I., Piñón-Escobedo, C., Zavala-Páramo, M. G., López-Romero, E., & Cano-Camacho, H. (2005). Degradation of cellulose by the bean-pathogenic fungus *Colletotrichum lindemuthianum*. Production of extracellular cellulolytic enzymes by cellulose induction. *Antonie van Leeuwenhoek, International Journal of General and Molecular Microbiology*, 87(4), 301–310.
- Admasu,W., Sahile, S., and Kibret,M. (2014). Assessment of potential antagonists for anthracnose (*Colletotrichum gloeosporioides*) disease of mango (*Mangifera indica L.*) in NorthWestern Ethiopia (Pawe). *Arch. Phytopathol.* 47, 2176–2186.
- Agrios, G. N. (2005) Plant Pathology. St. Louis, MO: Academic Press.
- Ahmad, Y., Hameed, A., & Ghaffar, A. (2006). Enzymatic activity of fungal pathogens in corn. *Pakistan Journal of Botany*, 38(4), 1305–1316.
- Ahsol, H., Wiwin, S., & Sutarya, R. (2014). Screening for resistance to Anthracnose caused by *Colletotrichum acutatum* in chili pepper (*Capsicum annuum L.*) in Kediri, East Java. *Advances in Agriculture & Botanics- International Journal of the Bioflux Society*, 6(2), 104–118.
- Ajith, P. S., Lakshmesha, K. K., Murthy, S. M., & Lakshmidhi, N. (2012). Botanicals for control of anthracnose of bell peppers. *The Journal of Plant Protection Sciences*, 4(1), 13-19.
- Aked, J., & Jongen, W. (2002). Maintaining the postharvest quality of fruits and vegetables. *Fruit and vegetable processing: improving quality*, 119-149.
- Alberto, R.T., Tiedeman, A.V., Wolf, G. & Danzinger, H.L. (2003). Light microscopy study on the pathological histology of *Colletotrichum gloeosporioides* (Penzig) Penzig and Sacc. in onion. *Journal of Tropical Plant Pathology* 38, 47–51.
- Al-Isawi, R. H. K., Almuktar, S. A. A. A. N., & Scholz, M. (2016). Monitoring and

- assessment of treated river, rain, gully pot and grey waters for irrigation of *Capsicum annuum*. *Environmental Monitoring and Assessment*, 188(5), 287.
- Ali, A., Bordoh, P. K., Singh, A., Siddiqui, Y., & Droby, S. (2016). Post-harvest development of anthracnose in pepper (*Capsicum spp*): Etiology and management strategies. *Crop Protection*, 90, 132–141.
- Anand, G., Yadav, S., & Yadav, D. (2017). Purification and biochemical characterization of an exo-polygalacturonase from *Aspergillus flavus* MTCC 7589. *Biocatalysis and Agricultural Biotechnology*, 10(August 2016), 264–269.
- Anand, T., Bhaskaran, R., Raguchander, T., Karthikeyan, G., Rajesh, M., & Senthilraja, G. (2008). Production of cell wall degrading enzymes and toxins by *Colletotrichum capsici* and *Alternaria alternata* causing fruit rot of chillies. *Journal of Plant Protection Research*, 48(4), 437–451.
- Anand, T., Chandrasekaran, A., Kuttalam, S., Senthilraja, G., & Samiyappan, R. (2010). Integrated control of fruit rot and powdery mildew of chilli using the biocontrol agent *Pseudomonas fluorescens* and a chemical fungicide. *Biological Control*, 52(1), 1–7.
- Anderson, J.L. & Walker, J.C. (1961). Histology of watermelon anthracnose. *Phytopathology* 52, 650–654.
- Andrade Pinto, J. M., Souza, E. A., & Oliveira, D. F. (2010). Use of plant extracts in the control of common bean anthracnose. *Crop Protection*, 29(8), 838–842.
- Aneja, K.R. (2005). Production of pectinolytic enzymes. New Age International (P) Ltd., In: Exp in Microb. Plant Path. & Biotech. New Delhi, 4th Ed. pp.251–253.
- Antov, M. G., Peričin, D. M., & Pejin, S. N. (2004). Pectinases partitioning in aqueous two-phase systems: An integration of the systems poly (ethylene glycol) crude dextran and poly (ethylene glycol) ammonium sulphate. *Journal of the Serbian Chemical Society*, 69(4), 299–307.
- Antov, M. G., Peričin, D. M., & Dašić, M. G. (2006). Aqueous two-phase partitioning of xylanase produced by solid-state cultivation of *Polyporus squamosus*. *Process Biochemistry*, 41(1), 232–235.
- Aphidech Sangdee. (2011). Morphological, pathological and molecular variability of *Colletotrichum capsici* causing anthracnose of chilli in the North-east of Thailand. *African Journal of Microbiology Research*, 5(25), 4368–4372.
- Arroyo, F. T., Moreno, J., García-Herdugo, G., Santos, B. D. L., Barrau, C., Porras, M. & Romero, F. (2005). Ultrastructure of the early stages of *Colletotrichum acutatum* infection of strawberry tissues. *Canadian journal of botany*, 83(5), 491–500.
- Asenjo, J. A., & Andrews, B. A. (2011). Aqueous two-phase systems for protein separation: a perspective. *Journal of Chromatography A*, 1218(49), 8826–8835.

- Asenjo, J. A., & Andrews, B. A. (2012). Aqueous two-phase systems for protein separation: phase separation and applications. *Journal of Chromatography A*, 1238, 1-10.
- Ashwini, N., & Srividya, S. (2014). Potentiality of *Bacillus subtilis* as biocontrol agent for management of anthracnose disease of chilli caused by *Colletotrichum gloeosporioides* OGC1. *3 Biotech*, 4(2), 127–136.
- Auyong, A. S. M., Ford, R., & Taylor, P. W. J. (2012). Genetic transformation of *Colletotrichum truncatum* associated with anthracnose disease of chili by random insertional mutagenesis. *Journal of basic Microbiology*, 52(4), 372-382.
- Babalola, O. O. (2010). Pectinolytic and cellulolytic enzymes enhance *Fusarium compactum* virulence on tubercles infection of egyptian broomrape. *International Journal of Microbiology*, 2010.
- Bailey, M. J., Biely, P., & Poutanen, K. (1992). Interlaboratory testing of methods for assay of xylanase activity. *Journal of biotechnology*, 23(3), 257-270.
- Barreto, A. L. H., Vasconcelos, I. M., Grangeiro, T. B., Melo, V. M. M., Matos, T. E., Eloy, Y. R. G. & Oliveira, J. T. A. (2007). Infection process and host defense responses in compatible and incompatible interactions between cowpea (*Vigna unguiculata*) and *Colletotrichum gloeosporioides*. *International journal of plant sciences*, 168(2), 193-203.
- Baum, A., Dominiak, M., Vidal-Melgosa, S., Willats, W. G. T., Søndergaard, K. M., Hansen, P. W. & Mikkelsen, J. D. (2017). Prediction of Pectin Yield and Quality by FTIR and Carbohydrate Microarray Analysis. *Food and Bioprocess Technology*, 10(1), 143–154.
- Bellincampi, D., Cervone, F., & Lionetti, V. (2014). Plant cell wall dynamics and wall-related susceptibility in plant-pathogen interactions. *Frontiers in Plant Science*, 5, 228.
- Benavides, J., Rito-Palomares, M., & Asenjo, J. A. (2011). Aqueous Two-Phase Systems. *Comprehensive Biotechnology*, (September 2016), 697–713.
- Bhadauria, V., Banniza, S., Vandenberg, A., Selvaraj, G., & Wei, Y. (2013). Overexpression of a novel biotrophy-specific *Colletotrichum truncatum* effector, CtNUDIX, in hemibiotrophic fungal phytopathogens causes incompatibility with their host plants. *Eukaryotic Cell*, 12(1), 2–11.
- Bhat, T. M., & Kudesia, R. (2011). Evaluation of genetic diversity in five different species of family solanaceae using cytological characters and protein profiling. *Genetic Engineering and Biotechnology Journal*, 20, 20-25.
- Blais, P., Rogers, P. A., & Charest, P. M. (1992). Kinetic of the production of polygalacturonase and pectin lyase by two closely relatedFormae speciales of *Fusarium oxysporum*. *Experimental Mycology*, 16(1), 1-7.
- Blumenkrantz, N., & Asboe-Hansen, G. (1973). New method for quantitative determination of uronic acids. *Analytical Biochemistry*, 54(2), 484–489.

- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical biochemistry*, 72(1-2), 248-254.
- Brummell, D. A., & Harpster, M. H. (2001). Cell wall metabolism in fruit softening and quality and its manipulation in transgenic plants. *Plant Molecular Biology*.
- Buyer, J. S., Roberts, D. P., Millner, P., & Russek-Cohen, E. (2001). Analysis of fungal communities by sole carbon source utilization profiles. *Journal of Microbiological Methods*, 45(1), 53–60.
- Caffall, K. H., & Mohnen, D. (2009). The structure, function, and biosynthesis of plant cell wall pectic polysaccharides. *Carbohydrate Research*, 344(14), 1879–1900.
- Cai, L., Hyde, K.D., Taylor, P.W.J., Weir, B.S., Waller, J.M., Abang, M.M. (2009). A polyphasic approach for studying *Colletotrichum*. *Fungal Diversity*, 39, 183-204.
- Cannon, P.F., Buddie, A.G., & Bridge, P.D. (2008). The typification of *Colletotrichum gloeosporioides*. *Mycotaxon*, 104, 189-204.
- Cannon, P. F., Damm, U., Johnston, P. R., & Weir, B. S. (2012). *Colletotrichum* – current status and future directions. *Studies in Mycology*, 73, 181–213.
- Cerkauskas, R. (2004). Cercospora leaf spot. *AVRDC-The World Vegetable Center, Shanhua, Taiwan*. Available online at: <http://www.avrdc.org/pdf/pepper/cercospora.pdf#search=%22cercospora%22>.
- Cerkauskas, R., Mertely, J. C., Legard, D. E., Photita, W., Taylor, P. W. J., Ford, R. and Garg, R. (2009). Chilli anthracnose disease caused by *Colletotrichum* species. *Journal of Zhejiang University. Science. B*, 4(1), 764–78.
- Chen, Z., Nunes, M. a., Silva, M. C., & Jr., C. J. R. (2004). Appressorium Turgor Pressure of *Colletotrichum kahawae* Might Have a Role in Coffee Cuticle Penetration. *Mycologia*, 96(6), 1199.
- Choi, J., Kim, K.-T., Jeon, J., & Lee, Y.-H. (2013). Fungal plant cell wall-degrading enzyme database: a platform for comparative and evolutionary genomics in fungi and *Oomycetes*. *BMC Genomics*, 14 Suppl 5(Suppl 5), S7
- Choquer, M., Fournier, E., Kunz, C., Levis, C., Pradier, J. M., Simon, A., & Viaud, M. (2007). *Botrytis cinerea* virulence factors: New insights into a necrotrophic and polyphagous pathogen. *FEMS Microbiology Letters*, 277(1), 1–10.
- Chowdappa, P., Chethana, C. S., Bharghavi, R., Sandhya, H., & Pant, R. P. (2012). Morphological and molecular characterization of *Colletotrichum gloeosporioides* (Penz) Sac. isolates causing anthracnose of orchids in India. *Biotechnol. Bioinf. Bioeng.*, 2(1), 567–572.
- Collmer, A., & Keen, N. T. (1986). The role of pectic enzymes in plant

- pathogenesis. *Annual review of phytopathology*, 24(1), 383-409.
- Cosgrove, D. J. (2005). Growth of the plant cell wall. *Nature Reviews. Molecular Cell Biology*, 6(November), 850–861.
- Crouch, J.A., Clarke, B.B., White, J.F. and Hillman, B.I. (2009). Systematic analysis of falcate-spored graminicolous *Colletotrichum* and a description of six new species from warm-season grasses. *Mycologia*, 101, 717-732.
- Curry, K. J., Abril, M., Avant, J. B., & Smith, B. J. (2002). Strawberry Anthracnose: Histopathology of *Colletotrichum acutatum* and *C. fragariae*. *Phytopathology*, 92(10), 1055–1063.
- D’Ovidio, R., Mattei, B., Roberti, S., & Bellincampi, D. (2004). Polygalacturonases, polygalacturonase-inhibiting proteins and pectic oligomers in plant-pathogen interactions. *Biochimica et Biophysica Acta - Proteins and Proteomics*.
- Damm, U., Woudenberg, J. H. C., Cannon, P. F., & Crous, P. W. (2009). *Colletotrichum* species with curved conidia from herbaceous hosts. *Fungal Diversity*, 39(1802), 45–87.
- De Bary, A. (1886). Ueber einige Sclerotinien und Sclero. *Botanische Zeitung*, 44, 378.
- De Lorenzo, G., & Ferrari, S. (2002). Polygalacturonase-inhibiting proteins in defense against phytopathogenic fungi. *Current opinion in plant biology*, 5(4), 295-299.
- De Silva, D. D., Ades, P. K., Crous, P. W., & Taylor, P. W. J. (2016). *Colletotrichum* species associated with chili anthracnose in Australia. *Plant Pathology*.
- Deising, H. B., Werner, S., & Wernitz, M. (2000). The role of fungal appressoria in plant infection. *Microbes and infection*, 2(13), 1631-1641.
- Deshmukh, N., Talkal, R., Jha, K., Singh, P. G., & Prajapati, D. C. (2012). Production , Purification, Characterization and Comparison of Polygalacturonase from various strains of *Aspergillus*. *International Journal*, 1(9).
- Diao, Y., Zhang, C., Xu, J., Lin, D., Liu, L., Mtung'e, O. G., & Liu, X. (2015). Genetic differentiation and recombination among geographic populations of the fungal pathogen *Colletotrichum truncatum* from chili peppers in China. *Evolutionary applications*, 8(1), 108-118.
- Diéguez-Uribeondo, J., Förster, H., Soto-Estrada, A., & Adaskaveg, J. E. (2005). Subcuticular-Intracellular Hemibiotrophic and Intercellular Necrotrophic Development of *Colletotrichum acutatum* on Almond. *Phytopathology*, 95(7), 751–758.
- Dinh, S. Q., Chongwungse, J., Pongam, P., & Sangchote, S. (2003). Fruit infection by *Colletotrichum gloeosporioides* and anthracnose resistance of some mango cultivars in Thailand. *Australasian Plant Pathology*, 32(4), 533–538.

- Dogan, N., & Tari, C. (2008). Characterization of three-phase partitioned exo-polygalacturonase from *Aspergillus sojae* with unique properties. *Biochemical Engineering Journal*, 39(1), 43–50.
- Douaiher, M. N., Nowak, E., Durand, R., Halama, P., & Reignault, P. (2007). Correlative analysis of *Mycosphaerella graminicola* pathogenicity and cell wall-degrading enzymes produced in vitro: The importance of xylanase and polygalacturonase. *Plant Pathology*, 56(1), 79–86.
- Ebringerova, A., Hromadkova, Z., & Heinze, T. (2005). Hemicellulose. *Advances in Polymer Science*, 186(August), 1–67
- Edgar, R. C. (2004). MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic acids research*, 32(5), 1792–1797.
- El-Batal, A. I., Osman, E. M., & Shaima, I. A. M. (2013). Optimization and characterization of polygalacturonase enzyme produced by gamma irradiated *Penicillium citrinum*. *Journal of Chemical and Pharmaceutical Research*, 5(1), 336–347.
- El-Zoghbi, M. (1994). Biochemical changes in some tropical fruits during ripening. *Food Chemistry*, 49, 33–37.
- Eshel, D., Miyara, I., Ailing, T., Dinoor, A., & Prusky, D. (2002). pH regulates endoglucanase expression and virulence of *Alternaria alternata* in persimmon fruit. *Molecular Plant-Microbe Interactions : MPMI*, 15(8), 774–779.
- FAOSTAT (2011). Production Data. Available online at: <http://faostat.fao.org/>
- Fawole, O., Ahmed, O., & Balogun, O. (2010). Pathogenicity and cell wall-degrading enzyme activities of some fungal isolates from cowpea (*Vigna unguiculata* [L] Walp). *Biokemistri*, 18(1), 45–51.
- Fellah, A., Anjukandi, P., Waterland, M. R., & Williams, M. A. K. (2009). Determining the degree of methylesterification of pectin by ATR/FT-IR: Methodology optimisation and comparison with theoretical calculations. *Carbohydrate Polymers*, 78(4), 847–853.
- Fernando, T. H. P. S., Jayasinghe, C. K., & Wijesundera, R. L. C. (2001). Cell wall degrading enzyme secretion by *Colletotrichum acutatum* the causative fungus of secondary leaf fall of *Hevea brasiliensis*. *Mycological Research*, 105(2001), 195–201.
- Freeman, S., Minz, D., Kolesnik, I., Barbul, O., Zveibil, A., Maymon, M., et al. (2004). Trichoderma biocontrol of *Colletotrichum acutatum* and *Botrytis cinerea* and survival in strawberry. *Eur. J. Plant Pathol.* 110, 361–370.
- Freeman, S. (2008). Management, survival strategies, and host range of *Colletotrichum acutatum* on strawberry. *HortScience*, 43(1), 66–68.
- Freeman, S., Horowitz, S., & Sharon, a. (2001). Pathogenic and Nonpathogenic Lifestyles in *Colletotrichum acutatum* from Strawberry and Other Plants.

- Phytopathology*, 91, 986–992.
- Gadre, R. V., Van Driessche, G., Van Beeumen, J., & Bhat, M. K. (2003). Purification, characterisation and mode of action of an endo-polygalacturonase from the psychrophilic fungus *Mucor flavus*. *Enzyme and Microbial Technology*, 32(2), 321–330.
- Gattas, E. (2003). Isolation and characterization of pectinolytic fungi. *Revista de Ciencias Farmaceuticas*, 24(1), 33-37.
- Gautam, A. K. (2014). Physiology & Pathology *Colletotrichum gloeosporioides*: Biology , Pathogenicity and Management in India. *Journal of Plant Physiology & Pathology*, 2(2), 2–11.
- Ge, Y., & Guest, D. I. (2011). Light and scanning electron microscopy studies on the infection process of melon leaves by *Colletotrichum lagenarium*. *Physiological and Molecular Plant Pathology*, 76(1), 67–74.
- Gilbert, H. J. (2010). The biochemistry and structural biology of plant cell wall deconstruction. *Plant Physiology*, 153(2), 444–455.
- Glyk, A., Schepel, T., & Beutel, S. (2015). PEG–salt aqueous two-phase systems: an attractive and versatile liquid–liquid extraction technology for the downstream processing of proteins and enzymes. *Applied microbiology and biotechnology*, 99(16), 6599-6616.
- Goering and Van Soest (1970)
- Grabke, A., Williamson, M., Henderson, G. W., & Schnabel, G. (2014). First report of anthracnose on peach fruit caused by *Colletotrichum truncatum* in South Carolina. *Plant Disease*, 98(8), 1154-1154.
- Gregori, R., Mari, M., Bertolini, P., Barajas, J. A. S., Tian, J. B., & Labavitch, J. M. (2008). Reduction of *Colletotrichum acutatum* infection by a polygalacturonase inhibitor protein extracted from apple. *Postharvest Biology and Technology*, 48(2), 309–313.
- Grilo, A. L., Raquel Aires-Barros, M., & Azevedo, A. M. (2016). Partitioning in Aqueous Two-Phase Systems: Fundamentals, Applications and Trends. *Separation & Purification Reviews*, 45(1), 68–80.
- Guidarelli, M., Carbone, F., Mourguès, F., Perrotta, G., Rosati, C., Bertolini, P., & Baraldi, E. (2011). *Colletotrichum acutatum* interactions with unripe and ripe strawberry fruits and differential responses at histological and transcriptional levels. *Plant Pathology*, 60(4), 685–697.
- Gummadi, S. N., & Panda, T. (2003). Purification and biochemical properties of microbial pectinases—a review. *Process Biochemistry*, 38, 987–996.
- Hamdy, H. S. (2005). Purification and characterization of the pectin lyase produced by *Rhizopus oryzae* grown on orange peels. *Annals of Microbiology*, 55(3), 205–211.
- Hankin, L., & Anagnostakis, S. L. (1975). The use of solid media for detection of enzyme production by fungi. *Mycologia*, 597-607.

- Hayman, M., & Kam, P. C. A. (2008). Capsaicin: A review of its pharmacology and clinical applications. *Current Anaesthesia & Critical Care*, 19(5–6), 338–343.
- Heng, J. L. S., Shah, U. K. M., & Hamzah, H. (2011). Isolation, characterization and identification of potential Actinobacteria with antifungal activities towards chilli anthracnose. *African Journal of Biotechnology*, 10(32), 5979–5987.
- Herbert, C., O'Connell, R., Gaulin, E., Salesses, V., Esquerre-Tugayé, M. T., & Dumas, B. (2004). Production of a cell wall-associated endopolygalacturonase by *Colletotrichum lindemuthianum* and pectin degradation during bean infection. *Fungal Genetics and Biology*, 41(2), 140–147.
- Herculano, P. N., Porto, T. S., Maciel, M. H. C., Moreira, K. A., Souza-Motta, C. M., & Porto, A. L. F. (2012). Partitioning and purification of the cellulolytic complex produced by *Aspergillus japonicus* URM5620 using PEG-citrate in an aqueous two-phase system. *Fluid Phase Equilibria*, 335, 8–13.
- Hsieh, T.F., Huang, J.W. & Shiang, T. (2001). Light and scanning electron microscopy studies on the infection of oriental lily leaves by *Botrytis elliptica*. *Eur J Plant Pathol*, 107:571.
- Horowitz, S., Yarden, O., Zveibil, A., & Freeman, S. (2004). Development of a robust screening method for pathogenicity of *Colletotrichum* spp. on strawberry seedlings enabling forward genetic studies. *Plant Disease*, 88(8), 845–851.
- Hu, G. G., Linning, R., & Bakkeren, G. (2003). Ultrastructural comparison of a compatible and incompatible interaction triggered by the presence of an avirulence gene during early infection of the smut fungus, *Ustilago hordei*, in barley. *Physiological and Molecular Plant Pathology*, 62(3), 155–166.
- Huber, D. J., Karakurt, Y., & Jeong, J. (2001). Pectin degradation in ripening and wounded fruits. *Revista Brasileira de Fisiologia Vegetal*, 13(2), 224-241.
- Hugouvieux, V., Centis, S., Lafitte, C., & Esquerre-Tugaye, M. (1997). Induction by (alpha)-L-Arabinose and (alpha)-L-Rhamnose of Endopolygalacturonase Gene Expression in *Colletotrichum lindemuthianum*. *Applied and environmental microbiology*, 63(6), 2287-2292.
- Iqbal, S., Tak, H. I., Inam, A., Inam, A., Sahay, S., & Chalkoo, S. (2015). Comparative Effect of Wastewater and Groundwater Irrigation Along with Nitrogenous Fertilizer on Growth, Photosynthesis and Productivity of Chilli (*Capsicum annuum* L.). *Journal of Plant Nutrition*, 38(7), 1006–1021.
- Jagtap, G. & Sontakke, P. L. (2009). Taxonomy and morphology of *Colletotrichum truncatum* isolates pathogenic to Soybean. *African journal of Agricultural research*, 4(12), 1483-1487.
- Jaihan, P., Sangdee, K., & Sangdee, A. (2016). Selection of entomopathogenic fungus for biological control of chili anthracnose disease caused by

- Colletotrichum* spp. *European Journal of Plant Pathology*, 146(3), 551–564.
- Jain, N., Dhawan, K., Malhotra, S. P., Siddiqui, S., & Singh, R. (2001). Compositional and enzymatic changes in guava (*Psidium guajava* L.) fruits during ripening. *Acta Physiologiae Plantarum*, 23(3), 357–362.
- Jayani, R. S., Saxena, S., & Gupta, R. (2005). Microbial pectinolytic enzymes: A review. *Process Biochemistry*, 40(9), 2931–2944.
- Jayasinghe, C. K., Wijayaratne, S. C. P., & Fernando, T. H. P. S. (2004). Characterization of cell wall degrading enzymes of *Thanatephorus cucumeris*. *Mycopathologia*, 157(1), 73-79.
- Jia, Y. J., Feng, B. Z., Sun, W. X., & Zhang, X. G. (2009). Polygalacturonase, pectate lyase and pectin methylesterase activity in pathogenic strains of *phytophthora capsici* incubated under different conditions. *Journal of Phytopathology*, 157(10), 585–591.
- Juge, N. (2006). Plant protein inhibitors of cell wall degrading enzymes. *Trends in Plant Science*, 11(7), 359–367.
- Jurick, W. M., Vico, I., McEvoy, J. L., Whitaker, B. D., Janisiewicz, W., & Conway, W. S. (2009). Isolation, Purification, and Characterization of a Polygalacturonase Produced in *Penicillium solitum*- Decayed “Golden Delicious” Apple Fruit. *Postharvest Pathology and Mycotoxins Isolation*, 636–641.
- Juwon, A. D., Akinyosoye, F. A., & Kayode, O. A. (2012). Purification, characterization and application of polygalacturonase from *Aspergillus niger* CSTRF. *Malaysian Journal of Microbiology*, 8(3), 175–183.
- Kacurakowa, M., Capek, P., Sasinkova, V., Wellner, N., & Ebringerova, A. (2000). FT-IR study of plant cell wall model compounds : pectic polysaccharides and hemicelluloses. *Carbohydrate Polymers*, 43, 195–203.
- Kanchana-udomkan C, Taylor PWJ, Mongkolporn O, 2004. Development of a bioassay to study anthracnose infection of *Capsicum chinense* Jacq. fruit caused by *Colletotrichum capsici*. Thai Journal of Agricultural Science 37, 293–7
- Kant, S., Vohra, A., & Gupta, R. (2013). Purification and physicochemical properties of polygalacturonase from *Aspergillus niger* MTCC 3323. *Protein Expression and Purification*, 87(1), 11–16.
- Kasso, M., & Bekele, A. (2016). Post-harvest loss and quality deterioration of horticultural crops in Dire Dawa Region, Ethiopia. *Journal of the Saudi Society of Agricultural Sciences*.
- Kaur, G., Kumar, S., & Satyanarayana, T. (2004). Production, characterization and application of a thermostable polygalacturonase of a thermophilic mould *Sporotrichum thermophile Apinis*. *Bioresource Technology*, 94(3), 239–243.
- Kaur, M., Sharma, O. P., and Sharma, P. (2006). In vitro effect of *Trichoderma*

- species on *Colletotrichum capsici* causing fruit rot of chilli (*Capsicum annuum* L.). *Indian Phytopathol.* 59, 243–245.
- Kharwar, R. N., Upadhyay, R. S., Dubey, N. K., & Raghuwanshi, R. (2014). Microbial diversity and biotechnology in food security. *Microbial Diversity and Biotechnology in Food Security*, 1–610.
- Khokhar, I., Haider, M. S., Mushtaq, S., & Mukhtar, I. (2012). Isolation and screening of highly cellulolytic filamentous fungi. *Journal of Applied Sciences and Environmental Management*, 16(3).
- Khor, B. S. (2013). *Morphological characterization, molecular identification and pathotyping of Colletotrichum Species in Peninsular Malaysia/Khor Bee Sym* (Doctoral dissertation, University of Malaya).
- Kikot, G. E., Hours, R. A., & Alconada, T. M. (2009). Contribution of cell wall degrading enzymes to pathogenesis of *Fusarium graminearum*: a review. *Journal of basic microbiology*, 49(3), 231-241.
- Kim, K.-H., Yoon, J.-B., Park, H.-G., Park, E. W., & Kim, Y. H. (2004). Structural Modifications and Programmed Cell Death of Chili Pepper Fruit Related to Resistance Responses to *Colletotrichum gloeosporioides* Infection. *Phytopathology*, 94(12), 1295–1304.
- Kim, Y. S., Min, J. Y., Kang, B. K., Van Bach, N., Choi, W. B., Park, E. W., & Kim, H. T. (2007). Analyses of the less benzimidazole-sensitivity of the Isolates of *Colletotrichum spp.* causing the anthracnose in pepper and strawberry. *Plant Pathology Journal*, 23(3), 187–192.
- King, B. C., Waxman, K. D., Nenni, N. V., Walker, L. P., Bergstrom, G. C., & Gibson, D. M. (2011). Arsenal of plant cell wall degrading enzymes reflects host preference among plant pathogenic fungi. *Biotechnology for Biofuels*, 4(1), 4.
- Koche, M. D., Kothikar, R. B., Konde, S. A., & Patil, C. U. (2012). Morphological and cultural variation among isolates of *Colletotrichum dematium*. *Karnataka Journal of Agricultural Sciences*, 24(4).
- Kubicek, C. P., Starr, T. L., & Glass, N. L. (2014). Plant Cell Wall-Degrading Enzymes and Their Secretion in Plant-Pathogenic Fungi. *Annual Review of Phytopathology*, (June), 1–25.
- Kubo, Y. (2013). Function of peroxisomes in plant-pathogen interactions. In *Peroxisomes and their Key Role in Cellular Signaling and Metabolism* (pp. 329-345). Springer, Dordrecht.
- Kubo, Y., Harata, K., Kodama, S., & Fukada, F. (2016). Development of the infection strategy of the hemibiotrophic plant pathogen, *Colletotrichum orbiculare*, and plant immunity. *Physiological and Molecular Plant Pathology*, 95, 32–36.
- Kumar, V., Gupta, V. P., Babu, A. M., Mishra, R. K., Thiagarajan, V., & Datta, R. K. (2001). Surface ultrastructural studies on penetration and infection process

- of *Colletotrichum gloeosporioides* on mulberry leaf causing black spot disease. *Journal of Phytopathology*, 149(11–12), 629–633.
- Laine, M. J., Haapalainen, M., Wahlroos, T., Kankare, K., Nissinen, R., Kassuwi, S., & Metzler, M. C. (2000). The cellulase encoded by the native plasmid of *Clavibacter michiganensis* ssp. *sepedonicus* plays a role in virulence and contains an expansin-like domain. *Physiological and Molecular Plant Pathology*, 57(5), 221–233.
- Lakshmesha, K. K., Lakshmidevi, N., & Mallikarjuna Aradhya, S. (2005). Inhibition of cell wall degrading cellulase enzyme - The incitant of anthracnose disease caused by *Colletotrichum capsici* on capsicum fruit. *Archives of Phytopathology and Plant Protection*, 38(4), 295–302.
- Lakshmesha, K. K., Lakshmidevi, N., & Mallikarjuna, S. A. (2005). Changes in pectinase and cellulase activity of *Colletotrichum capsici* mutants and their effect on Anthracnose disease on capsicum fruit. *Archives of Phytopathology and Plant protection*, 38(4), 267-279.
- Latif, Z., & Sohail, M. (2012). Molecular characterization of polygalacturonase producing *Klebsiella* and *Staphylococcus* species by 16S rRNA sequencing collected from rotten fruits and vegetables. *African Journal of Microbiology Research*, 6(46), 7319–7323.
- Latunde-Dada, A. O., & Lucas, J. A. (2007). Localized hemibiotrophy in *Colletotrichum*: Cytological and molecular taxonomic similarities among *C. destructivum*, *C. linicola* and *C. truncatum*. *Plant Pathology*, 56(3), 437–447.
- Latunde-Dada, A. O., O'Connell, R. J., Nash, C., & Lucas, J. A. (1999). Stomatal penetration of cowpea (*Vigna unguiculata*) leaves by a *Colletotrichum* species causing latent anthracnose. *Plant Pathology*, 48(6), 777–785.
- Leandro, L. F. S., Gleason, M. L., Wegulo, S. N., & Nutter, F. W. (2002). Survival and sporulation of *Colletotrichum acutatum* on symptomless strawberry leaves. In *Acta Horticulturae* (Vol. 567, pp. 627–629)
- Lee, K., Kamala-Kannan, S., Sub, H., Seong, C., & Lee, G. (2008). Biological control of Phytophthora blight in red pepper (*Capsicum annuum* L.) using *Bacillus subtilis*. *World Journal of Microbiology and Biotechnology*, 24(7), 1139–1145.
- Leone, G. (1992). Significance of polygalacturonase production by *Botrytis cinerea* in pathogenesis. *Recent Advances in Botrytis Research*, 63–68.
- Lerouxel, O., Cavalier, D. M., Liepman, A. H., & Keegstra, K. (2006). Biosynthesis of plant cell wall polysaccharides - a complex process. *Current Opinion in Plant Biology*.
- Lima, Á. S., Alegre, R. M., & Meirelles, A. J. (2002). Partitioning of pectinolytic enzymes in polyethylene glycol/potassium phosphate aqueous two-phase systems. *Carbohydrate Polymers*, 50(1), 63-68.
- Lin Q, Kanchana-udomkan C, Jaunet T, Mongkolporn O, 2002. Genetic analysis of

- resistance to pepper anthracnose caused by *Colletotrichum capsici*. *Thai Journal of Agricultural Science* 35, 259–64.
- Liu, Y., Yu, Y. L., Chen, M. Z., & Xiao, X. (2011). Advances in Aqueous Two-Phase Systems and Applications in Protein Separation and Purification. *Canadian Journal on Chemical Engineering Technology*, 2(2), 1–7.
- Loliam, B., Morinaga, T., & Chaiyanan, S. (2012). Biocontrol of *Phytophthora infestans*, fungal pathogen of seedling damping off disease in economic plant nursery. *Psyche: A Journal of Entomology*, 2012.
- Ma, Y., Sun, S., Hao, H., & Xu, C. (2016). Production, purification and characterization of an exo-polygalacturonase from penicillium janthinellum sw09. *Anais Da Academia Brasileira de Ciencias*, 88, 479–487.
- Maciel, M. de H. C., Ottoni, C. A., Herculano, P. N., Porto, T. S., Porto, A. L. F., Santos, C. & Souza-Motta, C. (2014). Purification of polygalacturonases produced by *Aspergillus niger* using an aqueous two-phase system. *Fluid Phase Equilibria*, 371(August), 125–130.
- Malinovsky, F. G., Fangel, J. U., & Willats, W. G. (2014). The role of the cell wall in plant immunity. *Frontiers in plant science*, 5, 178.
- Manteau, S., Abouna, S., Lambert, B., & Legendre, L. (2003). Differential regulation by ambient pH of putative virulence factor secretion by the phytopathogenic fungus *Botrytis cinerea*. *FEMS Microbiology Ecology*, 43(3), 359–366.
- Martins, E. D. S., Leite, R. S. R., Da Silva, R., & Gomes, E. (2013). Purification and properties of polygalacturonase produced by thermophilic fungus *thermoascus aurantiacus cbmai-756* on solid-state fermentation. *Enzyme Research*, 2013.
- Martins, E. S., Silva, D., Leite, R. S. R., & Gomes, E. (2007). Purification and characterization of polygalacturonase produced by thermophilic *Thermoascus aurantiacus* CBMAI-756 in submerged fermentation. *Antonie van Leeuwenhoek, International Journal of General and Molecular Microbiology*, 91(3), 291–299.
- Martos, M. A., Zubreski, E. R., Garro, O. A., & Hours, R. A. (2013). Production of Pectinolytic enzymes by the yeast *Wickerhamomyces anomalus* isolated from citrus fruits peels. *Biotechnology research international*, 2013.
- McKay, S. F., Freeman, S., Minz, D., Maymon, M., Sedgley, M., Collins, G. C., & Scott, E. S. (2009). Morphological, genetic, and pathogenic characterization of *Colletotrichum acutatum*, the cause of anthracnose of almond in Australia. *Phytopathology*, 99(8), 985–995.
- Md Sidek, N. L., Tan, J. S., Abbasilasi, S., Wong, F. W. F., Mustafa, S., & Ariff, A. B. (2016). Aqueous two-phase flotation for primary recovery of bacteriocin-like inhibitory substance (BLIS) from *Pediococcus acidilactici* Kp10. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 1027, 81–87.

- Mehrnoosh, A., Sarker, M. Z. I., Mustafa, S., & Yazid, A. M. M. (2011). Direct Purification of Pectinase from Mango (*Mangifera Indica Cv. Chokanan*) Peel Using a PEG/Salt-Based Aqueous Two Phase System. *Molecules*, 16(12), 8419–8427.
- Mehta, A., & Mehta, P. (1985). Production of Pectolytic and Cellulolytic Enzymes by *Fusarium oxysporum* and *F. moniliforme* under Different Cultivation Conditions. *Folia Microbiologica*, 30, 42–50.
- Melton, L. D., & Smith, B. G. (2001). Determination of the Uronic Acid Content of Plant Cell Walls Using a Colorimetric Assay. In *Current Protocols in Food analytical Chemistry* (p. E3.3.1-4).
- Mendgen, K., & Hahn, M. (2002). Plant infection and the establishment of fungal biotrophy. *Trends in Plant Science*, 7(8), 352–356.
- Miller, G. L. (1959). Modified DNS method for reducing sugars. *Anal. Chem*, 31(3), 426-428.
- Mitchell, J. I., Roberts, P. J., & Moss, S. T. (1995). Sequence or structure?: a short review on the application of nucleic acid sequence information to fungal taxonomy. *Mycologist*, 9(2), 67-75.
- Mishra, R. (2015). *Studies on Anthraenose of chilli (Capsicum Annum L.) Caus Ed by Colletoticum Capsici (Sudow) Butler and Bis by* (Doctoral dissertation, Rvskvv, Gwalior (MP)).
- Mochizuki, M., Yamamoto, S., Aoki, Y., & Suzuki, S. (2012). Isolation and characterisation of *Bacillus amyloliquefaciens* S13-3 as a biological control agent for anthracnose caused by *Colletotrichum gloeosporioides*. *Biocontrol Science and Technology*, 22(August 2016), 697–709.
- MOA (2015). Statistik Utama Pemasaran FAMA, Kementerian Pemasaran Pertanian Persekutuan.
- Mohamed, S. A., Farid, N. M., Hossiny, E. N., & Bassuiny, R. I. (2006). Biochemical characterization of an extracellular polygalacturonase from *Trichoderma harzianum*. *Journal of Biotechnology*, 127(1), 54–64.
- Mokhtarani, B., Karimzadeh, R., Amini, M. H., & Manesh, S. D. (2008). Partitioning of Ciprofloxacin in aqueous two-phase system of poly(ethylene glycol) and sodium sulphate. *Biochemical Engineering Journal*, 38(2), 241–247.
- Montri, P., Taylor, P. W. J., & Mongkolporn, O. (2009). Pathotypes of *Colletotrichum capsici*, the Causal Agent of Chili Anthracnose, in Thailand. *Plant Disease*, 93(1), 17–20.
- Moraes, S. R. G., Tanaka, F. A. O., & Massola Jr, N. S. (2013). Histopathology of *Colletotrichum gloeosporioides* on guava fruits (*Psidium guajava*). *Revista Brasileira de Fruticultura*, 35(2), 657–664.
- Moreira, F. G., Dos Reis, S., Ferreira Costa, M. A., Marques De Souza, C. G., &

- Peralta, R. M. (2005). Production of hydrolytic enzymes by the plant pathogenic fungus *Myrothecium verrucaria* in submerged cultures. *Brazilian Journal of Microbiology*, 36(1), 7–11.
- Mueller, M., Hobiger, S., & Jungbauer, A. (2010). Anti-inflammatory activity of extracts from fruits, herbs and spices. *Food Chemistry*, 122(4), 987–996.
- Münch, S., Lingner, U., Floss, D. S., Ludwig, N., Sauer, N., & Deising, H. B. (2008). The hemibiotrophic lifestyle of *Colletotrichum* species. *Journal of Plant Physiology*, 165(1), 41–51.
- Nadar, S. S., Pawar, R. G., & Rathod, V. K. (2017). Recent advances in enzyme extraction strategies: A comprehensive review. *International Journal of Biological Macromolecules*, 101, 931–957.
- Naganagouda, K., & Mulimani, V. H. (2008). Aqueous two-phase extraction (ATPE): an attractive and economically viable technology for downstream processing of *Aspergillus oryzae*  $\alpha$ -galactosidase. *Process Biochemistry*, 43(11), 1293–1299.
- Nagy, Z., Daood, H., Ambrózy, Z., & Helyes, L. (2015). Determination of Polyphenols, Capsaicinoids, and Vitamin C in New Hybrids of Chili Peppers. *Journal of Analytical Methods in Chemistry*, 2015.
- Nelson, N. (1944). A photometric adaptation of the Somogyi method for the determination of glucose. *J. biol. Chem*, 153(2), 375–380.
- Nguyen, P. T. H., Pettersson, O. V., Olsson, P., & Liljeroth, E. (2010). Identification of *Colletotrichum* species associated with anthracnose disease of coffee in Vietnam. *European Journal of Plant Pathology*, 127(1), 73–87.
- Niture, S. K. (2008). Comparative biochemical and structural characterizations of fungal polygalacturonases. *Biologia*, 63(1), 1–19.
- Nwanekezi, E. C., Alawuba, O. C. G., & Mkpolulu, C. C. M. (1994). Characterization of pectic substances from selected tropical fruits. *Journal of Food Science And Technology-Mysore*, 31(2), 159–161.
- O'Connell, R., Herbert, C., Sreenivasaprasad, S., Khatib, M., Esquerre-Tugayé, M. T., & Dumas, B. (2004). A novel *Arabidopsis-Colletotrichum* pathosystem for the molecular dissection of plant-fungal interactions. *Molecular Plant-Microbe Interactions*, 17(3), 272–282.
- Oeser, B., Heidrich, P. M., Müller, U., Tudzynski, P., & Tenberge, K. B. (2002). Polygalacturonase is a pathogenicity factor in the *Claviceps purpurea*/rye interaction. *Fungal Genetics and Biology*, 36(3), 176–186.
- Olutiola, P. O., & Akintunde, O. A. (1979). Pectin lyase and pectin methylesterase production by *Penicillium citrinum*. *Transactions of the British Mycological Society*, 72(1), 49–55.
- Oo, M. M., & Oh, S. (2016). Chilli anthracnose ( *Colletotrichum* spp .) disease and its management approach. *Korean Journal of Agricultural Science*, 43(2),

- Ooi, C. W., Tey, B. T., Hii, S. L., Ariff, A., Wu, H. S., Lan, J. C. W. & Ling, T. C. (2009). Direct purification of *Burkholderia Pseudomallei* lipase from fermentation broth using aqueous two-phase systems. *Biotechnology and Bioprocess Engineering*, 14(6), 811–818.
- Orobiyi, a, Dansi, a, Assogba, P., Loko, L. Y., Dansi, M., Vodouhè, R., ... Sanni, a. (2013). Chili (*Capsicum annuum* L.) in southern Benin: production constraints, varietal diversity, preference criteria and participatory evaluation. *International Research Journal of Agricultural Science and Soil Scienc*, 3(April), 107–120.
- Oyagbemi, a a, Saba, a B., & Azeez, O. I. (2015). Capsaicin: a novel chemopreventive molecule and its underlying molecular mechanisms of action. *Indian Journal of Cancer*, 47, 53–58.
- Padder, B. A., and Sharma, P. N. (2011). In vitro and in vivo antagonism of biocontrol agents against *Colletotrichum lindemuthianum* causing bean anthracnose. *Arch. Phytopathol.* 44, 961–969.
- Pakdeevaraporn, P., Wasee, S., Taylor, P. W. J., & Mongkolporn, O. (2005). Inheritance of resistance to anthracnose caused by *Colletotrichum capsici* in *Capsicum*. *Plant Breeding*, 124(2), 206–208.
- Pandey, A., Pandey, B. K., Muthukumar, M., Yadava, L. P., & Chauhan, U. K. (2012). Histopathological Study of Infection Process of *Colletotrichum gloeosporioides* Penz and Sacc. on *Mangifera indica* L. *Plant Pathology Journal*.
- Pathak, N., Mishra, S., & Sanwal, G. G. (2000). Purification and characterization of polygalacturonase from banana fruit. *Phytochemistry*, 54(2), 147–152.
- Paulert, R., Talamini, V., Cassolato, J. E. F., Duarte, M. E. R., Noseda, M. D., Smania, A., & Stadnik, M. J. (2009). Effects of sulfated polysaccharide and alcoholic extracts from green seaweed *Ulva fasciata* on anthracnose severity and growth of common bean (*Phaseolus vulgaris* L.). *Journal of Plant Diseases and Protection*, 116(6), 263-270.
- Papaspyridi, L. M., Katapodis, P., Gonou- Zagou, Z., Kapsanaki- Gotsi, E., & Christakopoulos, P. (2011). Growth and biomass production with enhanced β- glucan and dietary fibre contents of *Ganoderma australe* ATHUM 4345 in a batch- stirred tank bioreactor. *Engineering in Life Sciences*, 11(1), 65-74.
- Pedrolli, D. B., & Carmona, E. C. (2014). Purification and characterization of a unique pectin lyase from *Aspergillus giganteus* able to release unsaturated monogalacturonate during pectin degradation. *Enzyme research*, 2014.
- Peeran, M. F., Kuppusami, P. & Thiruvengadam, R. (2014). Pathogenesis of *Colletotrichum lindemuthianum* the incitant of anthracnose disease in beans mediated by macerating enzymes. *The Bioscan*, 9(1), 295-300.

- Pereira, M., Wu, Y.-T., Venâncio, A., & Teixeira, J. (2003). Aqueous two-phase extraction using thermoseparating polymer: a new system for the separation of endo-polygalacturonase. *Biochemical Engineering Journal*, 15(2), 131–138.
- Peres, N. A. R., Kuramae, E. E., Dias, M. S. C., & De Souza, N. L. (2002). Identification and characterization of *Colletotrichum* spp. affecting fruit after harvest in Brazil. *Journal of Phytopathology*, 150(3), 128–134..
- Peres, N. a., Timmer, L. W., Adaskaveg, J. E., & Correll, J. C. (2005). Lifestyles of *Colletotrichum acutatum*. *Plant Disease*, 89(8), 784–796.
- Perfect, S. E., & Green, J. R. (2001). Infection structures of biotrophic and hemibiotrophic fungal plant pathogens. *Molecular Plant Pathology*, 2(2), 101-108.
- Perfect, S. E., Green, J. R., & O'Connell, R. J. (2001). Surface characteristics of necrotrophic secondary hyphae produced by the bean anthracnose fungus, *Colletotrichum lindemuthianum*. *European Journal of Plant Pathology*, 107(8), 813–819.
- Perfect, S. E., Hughes, H. B., O'Connell, R. J., & Green, J. R. (1999). *Colletotrichum*: A model genus for studies on pathology and fungal-plant interactions. *Fungal Genetics and Biology : FG & B*, 27(2–3), 186–98.
- Photita, W., Taylor, P. W. J., Ford, R., Hyde, K. D., & Lumyong, S. (2005). Morphological and molecular characterization of *Colletotrichum* species from herbaceous plants in Thailand. *Fungal Diversity*, 18, 117–133.
- Phoulivong, S. (2011). *Colletotrichum* , naming , control , resistance , biocontrol of weeds and current challenges. *Current Research in Environmental & Applied Mycology*, 1, 53–73.
- Po, L. G. (2011). Chili, Peppers, and Paprika. *Handbook of Vegetables and Vegetable Processing*, 581-603.
- Prihastuti, H., Cai, L., Chen, H., McKenzie, E. H. C., & Hyde, K. D. (2009). Characterization of *Colletotrichum* species associated with coffee berries in northern Thailand. *Fungal Diversity*, 39(1), 89-109.
- Prumputtha, I., Jeewon, R., Lumyong, S., McKenzie, E.H.C. and Hyde, K.D. (2005). Ribosomal DNA fingerprinting in the identification of non-sporulating endophytes from *Magnolia liliifera* (*Magnoliaceae*). *Fungal Diversity*, 20, 167–86.
- Protsenko, M. A., Bulantseva, E. A., & Korobleva, N. P. (2010). Polygalacturonase-inhibiting proteins in plant fleshy fruits during their ripening and infections. *Russian Journal of Plant Physiology*, 57(3), 356–362.
- Prusky, D., McEvoy, J. L., Leverentz, B., & Conway, W. S. (2001). Local modulation of host pH by *Colletotrichum* species as a mechanism to increase virulence. *Molecular Plant-Microbe Interactions*, 14(9), 1105-1113.

- Prusky, D., & Yakoby, N. (2003). Pathogenic fungi: leading or led by ambient pH? *Molecular Plant Pathology*, 4(6), 509-516.
- Quiroga, E. N., Sgariglia, M. A., Molina, C. F., Sampietro, D. A., Soberón, J. R., & Vattuone, M. A. (2009). Purification and characterization of an exo-polygalacturonase from *Pycnoporus sanguineus*. *Mycological Research*, 113(12), 1404–1410.
- Rahimpour, F., & Baharvand, A. R. (2009). Phase Equilibrium in Aqueous Two-phase Systems Containing Poly ( propylene glycol ) and Sodium Citrate at Different pH. *Engineering and Technology*, 150–153.
- Rahman, A. M., Rahman, M. M., Azad, A. K., & Alam, M. F. (2011). Inhibitory effect of different plant extracts and antifungal metabolites of *Trichoderma* strains on the conidial germination and germ tube growth of *Colletotrichum capsici* causing chili anthracnose. *International journal of agronomy and Agricultural Research*, 1(1), 20-28.
- Raja, S., Murty, V. R., Thivaharan, V., Rajasekar, V., & Ramesh, V. (2012). Aqueous Two Phase Systems for the Recovery of Biomolecules – A Review. *Science and Technology*, 1(1), 7–16.
- Ramdial, H., & Rampersad, S. N. (2014). Characterization of *Colletotrichum* spp. causing anthracnose of bell pepper (*Capsicum annuum* L.) in Trinidad. *Phytoparasitica*, 43(1), 37–49.
- Ranathunge, N. P., Mongkolporn, O., Ford, R., & Taylor, P. W. J. (2012). *Colletotrichum truncatum* Pathosystem on *Capsicum* spp: Infection, colonization and defence mechanisms. *Australasian Plant Pathology*, 41(5), 463–473.
- Rao, S., & Nandineni, M. R. (2017). Genome sequencing and comparative genomics reveal a repertoire of putative pathogenicity genes in chilli anthracnose fungus *Colletotrichum truncatum*. *PloS one*, 12(8), e0183567.
- Rashid, M. M., Kabir, M. H., Hossain, M. M., Bhuiyan, M. R., & Khan, M. A. I. (2015). Eco-friendly management of chilli anthracnose (*Colletotrichum capsici*). *International Journal of Plant Pathology*, 6(1), 1–11.
- Ratanapongleka, K. (2010). Recovery of Biological Products in Aqueous Two Phase Systems. *International Journal of Chemical Engineering and Applications*, 1(2), 191–198.
- Rawlings, S. L., O'Connell, R. J., & Green, J. R. (2007). The spore coat of the bean anthracnose fungus *Colletotrichum lindemuthianum* is required for adhesion, appressorium development and pathogenicity. *Physiological and Molecular Plant Pathology*, 70(4-6), 110-119.
- Reiter, W. D. (2002). Biosynthesis and properties of the plant cell wall. *Current opinion in plant biology*, 5(6), 536-542.
- Rogers, L. M., Kim, Y. K., Guo, W., González-Candelas, L., Li, D., & Kolattukudy, P. E. (2000). Requirement for either a host- or pectin-induced pectate lyase

- for infection of *Pisum sativum* by *Nectria hematococca*. *Proceedings of the National Academy of Sciences of the United States of America*, 97(17), 9813–8.
- Rojo-Báez, I., García-Estrada, R. S., León-Félix, J., Sañudo-Barajas, A., & Allende-Molar, R. (2016). Histopatología del proceso de infección de *Colletotrichum truncatum* en hojas de papaya y chícharo. *Revista Mexicana de Fitopatología, Mexican Journal of Phytopathology*, 9–18.
- Rose, J. K. C. (2003). The plant cell wall. Annual plant reviews, vol. 8. *Garsington: Blackwell Publishing*.
- Rosidah, S., Syukur, M., & Widodo, W. (2014). Pendugaan Parameter Genetik Ketahanan Tanaman Cabai terhadap Penyakit Antraknosa. *Jurnal Fitopatologi Indonesia*, 10, 202–209.
- Ryder, L. S., & Talbot, N. J. (2015). Regulation of appressorium development in pathogenic fungi. *Current opinion in plant biology*, 26, 8-13.
- Saad, N., Briand, M., Gardarin, C., Briand, Y., & Michaud, P. (2007). Production, purification and characterization of an endopolygalacturonase from *Mucor rouxii* NRRL 1894. *Enzyme and Microbial Technology*, 41(6–7), 800–805.
- Saadoun, M., & Allagui, M. B. (2013). Management of chili pepper root rot and wilt (caused by *Phytophthora nicotiana*) by grafting onto resistant rootstock. *Phytopathologia Mediterranea*, 52(1), 141–147.
- Sangdee, A., Sachan, S., & Khankhum, S. (2011). Morphological, pathological and molecular variability of *Colletotrichum capsici* causing anthracnose of chilli in the North-east of Thailand. *African Journal of Microbiology Research*, 5(25), 4368-4372.
- Sarah, E.P., Green, J. R., & O'connell, R. J. (2001). Surface characteristics of necrotrophic secondary hyphae produced by the bean anthracnose fungus, *Colletotrichum lindemuthianum*. *European Journal of Plant Pathology*, 107(8), 813-819.
- Sathiyanraj, G., Srinivasan, S., Kim, H. Bin, Subramaniyam, S., Lee, O. R., Kim, Y. J., & Yang, D. C. (2011). Screening and optimization of pectin lyase and polygalacturonase activity from ginseng pathogen *cylindrocarpon destructans*. *Brazilian Journal of Microbiology*, 42(2), 794–806.
- Saxena, A., Raghuwanshi, R., Gupta, V. K., & Singh, H. B. (2016). Chilli Anthracnose: The Epidemiology and Management. *Frontiers in Microbiology*, 7, 1527.
- Sazci, A., Erenler, K., & Radford, A. (1986). Detection of cellulolytic fungi by using Congo red as an indicator: a comparative study with the dinitrosalicylic acid reagent method. *Journal of Applied Microbiology*, 61(6), 559-562.
- Scheller, H. V., Jensen, J. K., Sørensen, S. O., Harholt, J., & Geshi, N. (2007). Biosynthesis of pectin. *Physiologia Plantarum*, 129(2), 283-295.

- Scheller, H. V., & Ulvskov, P. (2010). Hemicelluloses. *Annual Review of Plant Biology*, 61(1), 263–289.
- Schumacher, J., & Tudzynski, P. (2012). Morphogenesis and Pathogenicity in Fungi. *Topics in Current Genetics*, 22, 243–264.
- Sella, L., Castiglioni, C., Roberti, S., D'Ovidio, R., & Favaron, F. (2004). An endo-polygalacturonase (PG) of *Fusarium moniliforme* escaping inhibition by plant polygalacturonase-inhibiting proteins (PGIPs) provides new insights into the PG-PGIP interaction. *FEMS Microbiology Letters*, 240(1), 117–124.
- Selvarajan, E., & Veena, R. (2017). Recent advances and future perspectives of thermostable xylanase. *Biomedical and Pharmacology Journal*, 10(1), 261–279.
- Shaligram, N. S., & Singhal, R. S. (2010). Surfactin -a review on biosynthesis, fermentation, purification and applications. *Food Technology and Biotechnology*, 48(2), 119–134.
- Sharma, P. N., Kaur, M., Sharma, O. P., Sharma, P., & Pathania, A. (2005). Morphological, pathological and molecular variability in *Colletotrichum capsici*, the cause of fruit rot of chillies in the subtropical region of north-western India. *Journal of Phytopathology*, 153(4), 232–237.
- Sharma, S. K., Vij, A. S., & Sharma, M. (2013). Mechanisms and clinical uses of capsaicin. *European journal of pharmacology*, 720(1-3), 55-62.
- Shen, S., Goodwin, P., & Hsiang, T. (2001). Hemibiotrophic infection and identity of the fungus, *Colletotrichum destructivum*, causing anthracnose of tobacco. *Mycological Research*, 105(11), 1340–1347.
- Shenoy, B. D., Jeewon, R., Lam, W. H., Bhat, D. J., Than, P. P., Taylor, P. W. J., & Hyde, K. D. (2007). Morpho-molecular characterisation and epitypification of *Colletotrichum capsici* (*Glomerellaceae, Sordariomycetes*), the causative agent of anthracnose in chilli. *Fungal Diversity*, 27(1), 197–211.
- Shovan, L. R., Bhuiyan, M. K. A., Begum, J. A., & Pervez, Z. (2008). In vitro control of *Colletotrichum dematium* causing anthracnose of soybean by fungicides, plant extracts and *Trichoderma harzianum*. *Int. J. Sustain. Crop Prod*, 3(3), 10-17.
- Siddiqui, M. A., Pande, V., & Arif, M. (2012). Production, purification, and characterization of polygalacturonase from *Rhizomucor pusillus* isolated from decomposting orange peels. *Enzyme Research*, 2012.
- Silva, C. F. B., & Michereff, S. J. (2014). Biology of *Colletotrichum* spp. and epidemiology of the anthracnose in tropical fruit trees. *Revista Caatinga*, 26, 130–138.
- Silva, S. A. M., Rodrigues, R., Gonçalves, L. S. A., Sudré, C. P., Bento, C. S., Carmo, M. G. F., & Medeiros, A. M. (2014). Resistance in *Capsicum* spp. to anthracnose affected by different stages of fruit development during pre- and postharvest. *Tropical Plant Pathology*, 39(4), 335–341.

- Singh, M. P. (2009). Application of Biolog FF MicroPlate for substrate utilization and metabolite profiling of closely related fungi. *Journal of microbiological methods*, 77(1), 102-108.
- Smith, B. J., & Black, L. L. (1990). Main content area Morphological, cultural, and pathogenic variation among *Colletotrichum* species isolated from strawberry. *Plant Disease*, 74(1), 69-76.
- Soliman, S., El-Zawahry, Y., & El-Moughith, A. (2013). Fungal Biodegradation of Agro-Industrial Waste. *Cellulose - Biomass Conversion*, 1–28.
- Somogyi, M. (1952). Notes on sugar determination. *Journal of biological chemistry*, 195, 19-23.
- Soytong, K., Kanokmedhakul, S., Rattanacherdchai, K., & Charoenporn, C. (2014). Microbial elicitors to induce immunity for plant disease control in chilli and tomato. In *Basic and Applied Aspects of Biopesticides* (pp. 99-125). Springer, New Delhi.
- Srinivas, N. D. (2000). *Aqueous Two-Phase Extraction for the Downstream Processing of Enzymes* (Doctoral dissertation, University of Mysore).
- Stefanowicz, A. (2006). The Biolog Plates Technique as a Tool in Ecological Studies of Microbial Communities. *Polish Journal of Environmental Studies*, 15(5).
- Suhaimi, M. Y., Mohamad, A. M., & Hani, M. N. F. (2014). Potential and Viability Analysis for Ginger Cultivation using Fertigation Technology in Malaysia, 9(1), 421–427.
- Sun, R., Sun, X. F., & Tomkinson, J. (2003). Hemicelluloses and Their Derivatives. In *Hemicelluloses: Science and Technology* (pp. 2–22).
- Sutton, B. C. (1992). The genus *Glomerella* and its anamorph *Colletotrichum*. *The genus Glomerella and its anamorph Colletotrichum*, 1-26
- Synytsya, A., Čopíková, J., Matějka, P., & Machovič, V. (2003). Fourier transform Raman and infrared spectroscopy of pectins. *Carbohydrate Polymers*, 54(1), 97-106.
- Talbot, N. J. (2003). On the Trail of a Cereal Killer: Exploring the Biology of *Magnaporthe grisea*. *Annual Review of Microbiology*, 57(1), 177–202.
- Tamura, K., Nei, M., & Kumar, S. (2004). Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proceedings of the National Academy of Sciences of the United States of America*, 101(30), 11030-11035.
- Tewari, R., Tewari, R., & Hoondal, G. (2005). Microbial pectinases. *Methods in Biotechnology*, Vol. 17: *Microbial Enzymes and Biotransformations*, 17(2000), 191–208.
- Thakur, B. R., Singh, R. K., Handa, A. K., & Rao, M. A. (1997). Chemistry and uses of pectin—a review. *Critical Reviews in Food Science & Nutrition*, 37(1), 47-73.

- Thakur, A., Pahwa, R., Singh, S., & Gupta, R. (2010). Production, Purification, and Characterization of Polygalacturonase from *Mucor circinelloides* ITCC 6025. *Enzyme Research*, 2010, 170549.
- Than, P. P., Jeewon, R., Hyde, K. D., Pongsupasamit, S., Mongkolporn, O., & Taylor, P. W. J. (2008). Characterization and pathogenicity of *Colletotrichum* species associated with anthracnose on chilli (*Capsicum spp.*) in Thailand. *Plant Pathology*, 57(3), 562–572.
- Than, P. P., Prihastuti, H., Phoulivong, S., Taylor, P. W. J., & Hyde, K. D. (2008). Chilli anthracnose disease caused by *Colletotrichum* species. *Journal of Zhejiang University. Science. B*, 9(10), 764–78.
- Tozze Jr, H. J., Massola Jr, N. M., Camara, M. P. S., Gioria, R., Suzuki, O., Brunelli, K. R., & Kobori, R. F. (2009). First report of *Colletotrichum boninense* causing anthracnose on pepper in Brazil. *Plant Disease*, 93(1), 106-106.
- USDA. (2015). United States Department of Agriculture Agricultural Research Service, USDA Food composition database. Retrieved August 8, 2017 from <https://www.ars.usda.gov/northeast-area/beltsville-md/beltsville-human-nutrition-research-center/nutrient-data-laboratory/>
- Vazquez, C., Reyes, F., & Martinez, M. J. (1993). Comparative study of pectic activities from different formae speciales of *Fusarium oxysporum*. *Letters in applied microbiology*, 16(4), 210-213.
- Valette-Collet, O., Cimerman, A., Reignault, P., Levis, C., & Boccardo, M. (2003). Disruption of *Botrytis cinerea* pectin methylesterase gene *Bcpme1* reduces virulence on several host plants. *Molecular Plant-Microbe Interactions : MPMI*, 16(4), 360–367.
- Venard, C., & Vaillancourt, L. (2007). Penetration and colonization of unwounded maize tissues by the maize anthracnose pathogen *Colletotrichum graminicola* and the related nonpathogen *C. sublineolum*. *Mycologia*, 99(3), 368–77.
- Veneault-Fourrey, C., Laugé, R., & Langin, T. (2005). Nonpathogenic strains of *Colletotrichum lindemuthianum* trigger progressive bean defense responses during appressorium-mediated penetration. *Applied and Environmental Microbiology*, 71(8), 4761–4770.
- Viera, R. G., Rodrigues Filho, G., de Assunção, R. M., Meireles, C. D. S., Vieira, J. G., & de Oliveira, G. S. (2007). Synthesis and characterization of methylcellulose from sugar cane bagasse cellulose. *Carbohydrate Polymers*, 67(2), 182-189
- Waller, J. M., Bridge, P. D., Black, R., & Hakiza, G. (1993). Characterization of the coffee berry disease pathogen, *Colletotrichum kahawae* sp. nov. *Mycological Research*, 97(8), 989-994.
- Wang, Y. S., Chen, Y. J., & Zhang, Y. (2012). Isolation, Identification and Carbon Metabolic Fingerprinting Analysis of Four Pathogens Isolated from Postharvest Plum Fruit. *Food Science*, 13, 051.

- Wanjiru, W. M., Zhensheng, K., & Buchenauer, H. (2002). Importance of cell wall degrading enzymes produced by *Fusarium graminearum* during infection of wheat heads. *European Journal of Plant Pathology*, 108(8), 803–810.
- Watanabe, K., Ikeda, H., Sakashita, T., & Sato, T. (2016). Anthracnose of genus *Mandevilla* caused by *Colletotrichum truncatum* and *C. siamense* in Japan. *Journal of general plant pathology*, 82(1), 33-37.
- Weir, B. S., Johnston, P. R., & Damm, U. (2012). The *Colletotrichum gloeosporioides* species complex. *Studies in mycology*, 73, 115-180.
- Wharton, Phillip S, J. D.-U. (2004). The biology of *Colletotrichum acutatum* by. *Anales Del Jardin Botanico de Madrid*, 61(1), 3–22.
- Wharton, P. S., Julian, a M., & O'Connell, R. J. (2001). Ultrastructure of the Infection of Sorghum bicolor by *Colletotrichum sublineolum*. *Phytopathology*, 91(2), 149–158.
- Wharton, P. S., & Schilder, A. C. (2008). Novel infection strategies of *Colletotrichum acutatum* on ripe blueberry fruit. *Plant Pathology*, 57(1), 122-134.
- Wu, Y. T., Pereira, M., Venâncio, A., & Teixeira, J. (2001). Separation of endo-polygalacturonase using aqueous two-phase partitioning. *Journal of Chromatography A*, 929(1–2), 23–29.
- Xu, Y., Souza, M. A., Pontes, M. Z., Vitolo, M., & Pessoa Júnior, A. (2003). Liquid-liquid extraction of enzymes by affinity aqueous two-phase systems. *Brazilian archives of biology and technology*, 46(4), 741-750.
- Yakoby, N., Beno-Moualem, D., Keen, N. T., Dinoor, A., Pines, O., & Prusky, D. (2001). *Colletotrichum gloeosporioides pelB* is an important virulence factor in avocado fruit-fungus interaction. *Molecular Plant-Microbe Interactions*, 14(8), 988–995.
- Yang, H., Goja, A. M., Cui, M., & Li, C. (2013). Aqueous Two-Phase Extraction Advances for Bioseparation. *Journal of Bioprocessing and Biotechniques*, 44172(10), 2155–9821.
- Zhuang, Y., Chen, L., Sun, L., & Cao, J. (2012). Bioactive characteristics and antioxidant activities of nine peppers. *Journal of Functional Foods*, 4(1), 331–338.
- Zimmer, A. R., Leonardi, B., Miron, D., Schapoval, E., Oliveira, J. R. De, & Gosmann, G. (2012). Antioxidant and anti-inflammatory properties of *Capsicum baccatum*: From traditional use to scientific approach. *Journal of Ethnopharmacology*, 139(1), 228–233.
- Živković, S., Stojanović, S., Ivanović, Ž., Trkulja, N., Dolovac, N., Aleksic, G., & Balaz, J. (2010). Morphological and Molecular Identification of *Colletotrichum acutatum* from Tomato Fruit. *Pestic. Phytomed.*, 25(3), 231–239.