



**UNIVERSITI PUTRA MALAYSIA**

***OPTIMISATION OF SIMULTANEOUS SACCHARIFICATION AND  
FERMENTATION FOR BIOBUTANOL PRODUCTION USING OIL PALM  
EMPTY FRUIT BUNCH***

**NUR ATHEERA AIZA MD RAZALI**

**FBSB 2018 40**



**OPTIMISATION OF SIMULTANEOUS SACCHARIFICATION AND  
FERMENTATION FOR BIOBUTANOL PRODUCTION USING OIL PALM  
EMPTY FRUIT BUNCH**

By

**NUR ATHEERA AIZA MD RAZALI**

**Thesis submitted to School of Graduate Studies, Universiti Putra  
Malaysia, in Fulfillment of the Requirement for the Degree of  
Master of Science**

**June 2018**

## **COPYRIGHT**

All materials contained within the thesis, including, without limitation of text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

## **OPTIMISATION OF SIMULTANEOUS SACCHARIFICATION AND FERMENTATION FOR BIOBUTANOL PRODUCTION USING OIL PALM EMPTY FRUIT BUNCH**

By

**NUR ATHEERA AIZA MD RAZALI**

**June 2018**

**Chairman :Suraini Abd Aziz, PhD**  
**Faculty :Biotechnology and Biomolecular Sciences**

Malaysia has a vast amount of tropical agricultural land that suitable for various agricultural activities. A lot of biomass generated from the agricultural activities such as rubber, paddy, fruit and oil palm biomass. Oil palm empty fruit bunch (OPEFB) has been recorded as one of the largest oil palm biomass (18–19 million tonnes/year) produced in palm oil processing mill. OPEFB has the potential as substrate for biobutanol production due to its abundance, cheap and high holocellulose content, thus provide renewability and environmentally friendly biobutanol compared to fossil fuels. Process for biobutanol production from OPEFB can be classified into two major processes: (i) separate saccharification and fermentation (SHF) and (ii) simultaneous saccharification and fermentation (SSF). SSF is a process where the enzymatic saccharification of OPEFB and ABE fermentation are carried out simultaneously in a flask. SSF process has recently gained attention and was proven to be more feasible than SHF, as it reduces the needs for additional equipments therefore lowering in capital, operational costs and time. The main disadvantage of the SSF process is the optimum temperature for fermenting *Clostridia* (37°C) does not coincide with the optimum temperature for cellulase (50°C). Thus, the objective of this study was to improve the biobutanol production through SSF using OPEFB. The optimisation study consisted of two main parts that employed; one factor at a time (OFAT) approach and using Central Composite Design (CCD) by Response Surface Methodology (RSM).

The SSF process successfully produced maximum biobutanol concentration of 1.74 g/L and biobutanol yield of 0.070 g/g at 96 h. The SSF biobutanol yield was comparable to the SHF biobutanol yield of 0.078 g/g. The SSF process was further enhanced by studying the preliminary investigation by OFAT approach. The results generated maximum biobutanol concentration of 2.91

g/L and biobutanol yield 0.12 g/g. The percentage of biobutanol increment was 40.21% and 1.71 fold. The biobutanol production was further statistically optimised using CCD. The analysis of variance (ANOVA) showed that the model was very significant ( $p < 0.0010$ ) for the biobutanol production. The optimum fermentation conditions obtained the highest biobutanol production were at temperature of 35°C, initial pH of 5.5, cellulase loading of 15 FPU/g of substrate and 5% (w/v) substrate concentration. From the validation study, the statistical optimisation resulted in a significant increment of biobutanol production of 3.97 g/L with biobutanol yield of 0.16 g/g with 55.95% increment (2.14 fold). The model and optimisation design obtained in this study helps to improve the biobutanol production in which was comparable to other studies of SSF processes using *Clostridia* species.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk Ijazah Master Sains

**PENGOTIMUMAN PROSES PENSAKARIDAAN DAN FERMENTASI  
SERENTAK BAGI PENGHASILAN BIOBUTANOL MENGGUNAKAN  
TANDAN KOSONG KELAPA SAWIT**

Oleh

**NUR ATHEERA AIZA MD RAZALI**

**Jun 2018**

**Pengerusi : Suraini Abd-Aziz, PhD**  
**Fakulti : Bioteknologi dan Sains Biomolekul**

Malaysia mempunyai sejumlah besar tanah pertanian tropika yang sesuai untuk dijadikan pelbagai aktiviti pertanian. Biojisim yang dihasilkan daripada aktiviti pertanian ini amat banyak antaranya biojisim getah, padi, buah-buahan dan kelapa sawit. Tandan kosong kelapa sawit (TKKS) telah direkodkan sebagai salah satu daripada biojisim kelapa sawit yang terbesar (18-19 juta tan/tahun) dihasilkan di kilang pemprosesan minyak kelapa sawit. TKKS merupakan substrat berpotensi untuk penghasilan biobutanol kerana kuantitinya yang banyak, murah dan kandungan holoselulosa yang tinggi malah menghasilkan biobutanol bersifat lebih mesra alam berbanding dengan bahan api fosil. Proses penghasilan biobutanol dari TKKS boleh diklasifikasikan kepada dua proses utama: (i) pensakaridaan dan penapaian berasingan (PPB) dan (ii) pensakaridaan dan penapaian serentak (PPS). PPS menggabungkan proses pensakaridaan TKKS dan penapaian ABE serentak di dalam bekas yang sama. Proses PPS telah mendapat perhatian dan terbukti lebih baik berbanding daripada proses PPB. Ini kerana proses PPS ini dapat mengurangkan keperluan untuk peralatan tambahan serta bermanfaat dalam menurunkan modal, kos operasi dan masa. Kelemahan utama proses PPS ini ialah suhu optimum untuk penapaian *Clostridia* (37°C) tidak setara dengan suhu optimum pensakaridaan selulase (50°C). Oleh yang demikian, objektif utama kajian ini adalah untuk meningkatkan pengeluaran biobutanol menerusi proses PPS menggunakan TKKS. Kajian pengoptimuman proses PPS terdiri daripada dua bahagian utama iaitu melalui satu faktor satu masa (SFSM) dan rekaan komposit pusat (RKP) yang dijalankan melalui kaedah permukaan tindakbalas (KPT).

Proses PPS telah berjaya menghasilkan pengeluaran maksimum biobutanol pada 96 jam dengan kepekatan 1.74 g/L hasil 0.07 g/g. Hasil biobutanol dalam

PPS adalah setara dengan hasil biobutanol PPB sebanyak 0.078 g/g. Proses PPS ini boleh dipertingkatkan dengan mengkaji pencirian awal yang dinilai oleh pendekatan SFMS. Maximum kepekatan biobutanol yang dihasilkan adalah 2.91 g/L dan 0.12 g/g hasil biobutanol. Peratusan kenaikan biobutanol adalah kira-kira 40.21% dan 1.71 kali ganda. Pengeluaran biobutanol boleh dilanjutkan pengoptimumannya menggunakan RKP. Menerusi analisis varians (ANOVA) model yang dihasilkan adalah sangat signifikan ( $p < 0.0010$ ) untuk pengeluaran biobutanol. Keadaan optimum yang diperolehi untuk pengeluaran biobutanol yang tertinggi adalah pada tahap suhu 35°C, pH awal 5.5, pemuatan selulase sebanyak 15 FPU/g substrat dan 5% (w/v) kepekatan substrat. Pengoptimuman statistik telah merekodkan kenaikan ketara pengeluaran biobutanol kepada 3.97 g/L dengan hasil 0.15 g/g dengan 55.95% kadar kenaikan (2.14 kali ganda). Reka bentuk model dan pengoptimuman yang diperolehi daripada kajian ini telah dapat membantu meningkatkan pengeluaran biobutanol setanding dengan kerja-kerja pengkajian PSS yang menggunakan spesies *Clostridia* yang lain.

## ACKNOWLEDGEMENTS

Alhamdulillah. All praised to Allah S.W.T for his mercy and guidance for enabling me to complete this master project. With utmost gratitude and sincere thanks, I forwarded to my main supervisor Prof. Dr. Suraini Abd-Aziz for her invaluable guidance and supervision, constructive ideas, technical support and suggestions she has provided throughout my time as her student. I am particularly thankful to my co-supervisors Dr. Mohamad Faizal Ibrahim and Dr. Ezyana Kamal Bahrin for their brilliant advices, constructive critics, generous guidance and assistance that help motivate and musters the confidence in myself to complete the project and thesis. It's been an amazing privilege to have met with encouraging professor and doctors like them and be involved in this project.

I would like to express my gratitude to Universiti Putra Malaysia and Ministry of Education Malaysia (MOE) for the funding of my study. Besides that, my greatest appreciation goes to the Environmental Biotechnology Group (EB Group) for giving the opportunity to improve myself. I am grateful by the supports and friendship delivered by the EB members. It had been a great experience working with all.

My deepest gratitude, honor and sincere appreciation I forwarded to all my family members for their unconditional love, prayers and comprehensive supports that I will remember forever in my mind. I also wished to thank my friends for always willing to help me whenever I need them. I am indebted to who directly and indirectly help me in completing this project.

May Allah reward and bless all of you with goodness.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science.

The members of the Supervisory Committee were as follows:

**Suraini Abd. Aziz, PhD**

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

**Mohamad Faizal Ibrahim, PhD**

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

**Ezyana Kamal Bahrin, PhD**

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

---

**ROBIAH BINTI YUNUS, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

## Declaration by Graduate Student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Nur Atheera Aiza Md Razali (GS36879)

## Declaration by Members of Supervisory Committee

This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature : \_\_\_\_\_  
Name of Chairman  
of Supervisory Committee : Prof. Dr. Suraini Abd-Aziz

Signature : \_\_\_\_\_  
Name of Member  
of Supervisory Committee : Dr. Mohamad Faizal Ibrahim

Signature : \_\_\_\_\_  
Name of Member  
of Supervisory Committee : Dr. Ezyana Kamal Bahrin

## TABLE OF CONTENTS

<b>ABSTRACT</b>	<b>Page</b>
<b>ABSTRAK</b>	i
<b>ACKNOWLEDGEMENTS</b>	iii
<b>APPROVAL</b>	v
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	viii
<b>LIST OF FIGURES</b>	xiii
<b>LIST OF ABBREVIATIONS</b>	xv
	xvi

## CHAPTER

<b>1 INTRODUCTION</b>	
1.1 Research Background	1
1.2 Problem Statement	2
1.3 Research Objectives	3
1.4 Scope of Study	3
1.5 Limitation of Study	3
<b>2 LITERATURE REVIEW</b>	
2.1 Biobutanol	4
2.1.1 Characteristics of Biobutanol	5
2.1.2 Applications of Biobutanol	7
2.2 Microorganisms for Biobutanol Production	10
2.2.1 <i>Clostridium acetobutylicum</i>	11
2.3 Substrates for Biobutanol Production	15
2.3.1 Lignocellulosic Biomass	17
2.3.2 Oil Palm Biomass	18
2.3.3 Oil Palm Empty Fruit Bunch (OPEFB)	21
2.4 Process for Biobutanol Production	23
2.4.1 Separate Saccharification and Fermentation (SHF)	26
2.4.2 Simultaneous Saccharification and Fermentation (SSF)	26
2.5 Factors Affecting Biobutanol Production in SSF	28
2.5.1 Temperature	28
2.5.2 Initial pH	28
2.5.3 Cellulase Loading	29
2.5.4 Substrate Concentration	29
2.5.5 Inoculum Concentration	30
2.6 Optimisation of Biobutanol Production	31
2.6.1 One Factor at a Time (OFAT)	31
2.6.2 Statistical Optimisation	32
2.7 Concluding Remarks	33

<b>3</b>	<b>MATERIALS AND METHODS</b>	
3.1	Experimental Design	34
3.2	Pretreatment of Oil Palm Empty Fruit Bunch (OPEFB)	36
3.3	Preparation of Medium for SSF	36
3.3.1	Preparation of Inoculum	36
3.3.2	Maintenance of Culture	37
3.3.3	Preparation of Fermentation Medium	37
3.3.4	Production of Biobutanol	38
3.4	Optimisation of Biobutanol	39
3.4.1	One Factor at a Time (OFAT)	39
3.4.1.1	Effect of Temperature	40
3.4.1.2	Effect of Initial pH	40
3.4.1.3	Effect of Cellulase Loading	40
3.4.1.4	Effect of Substrate Concentration	41
3.4.1.5	Effect of Inoculum Concentration	41
3.4.2	Central Composite Design (CCD)	41
3.4.3	Validation of Optimum Condition	43
3.5	Analytical Methods	44
3.5.1	Lignocellulosic Composition of Oil Palm Empty Fruit Bunch (OPEFB)	44
3.5.2	Filter Paper Unit (FPU) Assay	44
3.5.3	Reducing Sugars Analysis	45
3.5.4	Determination of Cell Concentration	45
3.5.5	pH Determination	45
3.5.6	Determination of ABE and Organic Acids using GC	45
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	
4.1	Characteristic of Oil Palm Empty Fruit Bunch (OPEFB)	47
4.2	Biobutanol Production through SSF	48
4.3	Investigation the Effects of Environmental Factors by One Factor at a Time (OFAT)	50
4.3.1	Effect of Temperature	50
4.3.2	Effect of Initial pH	52
4.3.3	Effect of Cellulase Loading	55
4.3.4	Effect of Substrate Concentration	57
4.3.5	Effect of Inoculum Concentration	59
4.3.6	Summary of OFAT Study	61
4.4	Optimisation of Biobutanol Production by Central Composite Design (CCD)	62
4.4.1	Validation of Biobutanol Optimisation	70
4.5	Comparison Studies	75

## **5 CONCLUSIONS AND RECOMMENDATIONS**

5.1 Conclusions	77
5.2 Recommendations for Future Research	78

<b>REFERENCES</b>	79
-------------------	----

<b>APPENDICES</b>	92
-------------------	----

<b>BIODATA OF STUDENT</b>	104
---------------------------	-----

<b>LIST OF PUBLICATIONS</b>	105
-----------------------------	-----



## LIST OF TABLES

Table		Page
1	Properties of Biobutanol and Other Fuels	8
2	Overview of Biobutanol Production from <i>Clostridium acetobutylicum</i> Utilising Various Carbon Sources	14
3	Lignocellulosic Composition of Oil Palm Biomass	20
4	Overview of Biobutanol Production from Oil Palm Biomass	21
5	Composition of RCM	37
6	Composition of P2 Medium	38
7	Parameters Range Selection for SSF Process	40
8	Predicted Data Biobutanol Yield of Central Composite Design (CCD) from 30 Experimental Run	42
9	Lignocellulosic Content of Untreated and Treated OPEFB as compare with Other Lignocellulosic Biomass	48
10	Products Concentrations from Simultaneous Saccharification and Fermentation by <i>Clostridium acetobutylicum</i> ATCC 824.	49
11	Effect of Temperatures (30 – 50°C) on Biobutanol Production at Initial pH 5.5, 15 FPU/g, 5% (w/v) Substrate and 10% (v/v) Inoculum on Simultaneous Saccharification and Fermentation by <i>Clostridium acetobutylicum</i> ATCC 824	51
12	Effect of Initial pH (4.5 – 7.0) on Biobutanol Production at 35°C, 15 FPU/g Substrate, 5% (w/v) Substrate and 10% (v/v) Inoculum on Simultaneous Saccharification and Fermentation by <i>Clostridium acetobutylicum</i> ATCC 824	54
13	Effect of Cellulase Loading (5-30 FPU/g Substrate) on Biobutanol Production at 35°C, Initial pH 5.5, 5% (w/v) Substrate and 10% (v/v) Inoculum on Simultaneous Saccharification and Fermentation by <i>Clostridium acetobutylicum</i> ATCC 824	56
14	Effect of Substrate Concentration (1–7 % w/v) on Biobutanol Production at Temperature 35°C, Initial pH 5.5, 10 FPU/g Substrate and 10% (v/v) Inoculum on Simultaneous Saccharification and Fermentation by <i>Clostridium acetobutylicum</i> ATCC 824	58
15	Effect of Inoculum Concentration (5 – 20 % v/v) on Biobutanol Production at Temperature 35°C, Initial pH 5.5, 10 FPU/g Substrate and 5% (w/v) Substrate Concentration on Simultaneous Saccharification and Fermentation by <i>Clostridium acetobutylicum</i> ATCC 824	60
16	Summary of OFAT Experiment	61
17	Coded Values for Each Factor of the Central Composite Design (CCD) for Biobutanol Production in Simultaneous Saccharification and Fermentation (SSF)	62
18	Experimental Data of Central Composite Design (CCD) for Biobutanol Yield	63
19	The ANOVA for the Second Order Model of Central	64

	Composite Design (CCD) for Biobutanol Production	
20	The ANOVA Results for Response of Biobutanol Yield	65
21	Products Concentrations from Validation of SSF in Serum Bottles and 2L Bioreactor Fermentation	75
22	Comparison Studies of Biobutanol Production through SSF Process from Various Substrates	76





## LIST OF FIGURES

Figure		Page
1	Timeline of Biobutanol Advances from 1916 to 2015	5
2	Structure of Biobutanol	5
3	Biobutanol Applications as Industrial Solvents	10
4	ABE Production Pathway	12
5	Overall Process Scheme for Biobutanol Production from Three Substrates Generation	15
6	The Upstream and Downstream Flow of Conversion of Sugar, Starch and Lignocellulosic Biomass into Biobutanol	17
7	Types of Oil Palm Biomass	19
8	(a) Oil Palm Fresh Fruit Bunch (b) Oil Palm Empty Fruit Bunch	21
9	Conversion of OPEFB into Various Product	23
10	Process Flow of Lignocellulosic Biomass into Biobutanol	25
11	General Experimental Design	35
12	A Simultaneous Saccharification and Fermentation for Biobutanol Production using Alkaline Pretreated Oil Palm Empty Fruit Bunch in 100 mL Serum Bottles	39
13	3D Surface Graphs for Model for Biobutanol Yield at Optimum Point	67
14	Simultaneous Saccharification And Fermentation (SSF) for Biobutanol using Oil Palm Empty Fruit Bunch by <i>Clostridium acetobutylicum</i> ATCC 824	71
15	Predicted Optimum Condition of Biobutanol Production through SSF using <i>Clostridium acetobutylicum</i> ATCC 824 by CCD	72

## LIST OF ABBREVIATIONS

ABE	Acetone – butanol – ethanol
ANN	Artificial neural network
ATCC	American type culture collection
CICC	China center of industrial culture collection
CCD	Central composite design
CPO	Crude palm oil
DCW	Dry cell weight
FFB	Fresh fruit bunch
FPU	Filter paper unit
GA	Genetic algorithm
GHG	Greenhouse gas
k/Pa	Kilo per pascal
KJ/kg	Kilojoule per kilogram
LCB	Lignocellulosic biomass
MJ/L	Megajoule per litre
mM	milliMolar
MON	Motor octane number
NCIMB	National collection of industrial and marine bacteria, Aberdeen, UK
OFAT	One factor at a time
OPEFB	Oil palm empty fruit bunch
OPF	Oil palm frond
OPT	Oil palm trunk
POME	Palm oil mill effluent
PKO	Palm kernel oil
RON	Research octane number
RSM	Response surface methodology
SHF	Separate hydrolysis and fermentation
SSF	Simultaneous saccharification and fermentation
SO	Saccharification only
sp	Species
VOCs	Volatile organic compounds

## CHAPTER 1

### INTRODUCTION

#### 1.1 Research Background

Gasoline derived from petrochemical route is mainly used for various mobilisations in the worldwide transport sector. However, in consideration of the probable declination of petrochemical reserves and environmental issues upon combustion of gasoline, renewable biofuels are among the accessible alternative that can partially or fully substitute the gasoline (Shao and Chen, 2015). Biofuels additional benefits are energy security, environmentally sustain as it derived from renewable substrates and mitigating climate change (Kumar and Gayen, 2012). Among other types of biofuels e.g., bioethanol, biodiesel, biohydrogen and biomethane, biobutanol possesses attractive characteristics almost similar to gasoline (Begum and Dahman, 2015). Biobutanol also has a good blending ability, which can be blend with gasoline at any ratio and used in the engine system without modification (Lee et al., 2008; Uyttebroek et al., 2013). In addition to having comparable octane number to gasoline, biobutanol also has higher energy density, lower Reid vapour pressure, lower water miscibility and is less hydroscopic than bioethanol. The demand on biobutanol is predicted to grow at a rate of 4.7% annually with the total consumption reaching about 2.9 million tonnes per year mainly in China, Europe and United States (Mascal, 2012). The biobutanol production is mainly impacted by substrates cost and improvement on the fermentation technology (Morone and Pandey, 2014).

Lignocellulosic biomass offers better selection as compared to sugar and starch substrates as it is cheaper, highly available and does not compete with food production or animal feed (Xue et al., 2013). Lignocellulosic biomass from agricultural biomass, forest residues, energy crops (switchgrass, yellow poplar, *Miscanthus* and etc.) and municipal solid wastes including food wastes, can become a potential substrate for biobutanol production (Salehi and Taherzadeh, 2015). In Malaysia, agricultural biomass is generated mainly from the palm oil industry with the current production reaching nearly 83 million tonnes and projected to increase to 85 – 110 million tonnes by 2020 (AIM, 2013). There are six types of oil palm biomass produced namely oil palm fronds (OPF), oil palm trunks (OPT), oil palm empty fruit bunch (OPEFB), kernel shells, mesocarp fibre and palm oil mill effluent (POME). Among these, OPEFB is the most abundant biomass produced at the palm oil mill with an annual production of 69,000 dry tonnes per year (Yoshizaki et al., 2013). The low cost of OPEFB is due to its unutilised capacity whereby the conventional management of OPEFB only involves dumping at the mill or mulching at the plantation (Ibrahim et al., 2013). OPEFB was also extensively studied as compost, composite materials and substrates for mushroom cultivation and fermentation (Cai et al., 2013; Sklavounos et al., 2013; Tan et al., 2010).

Harnessing OPEFB as substrate for biobutanol production has gained interest because of its high holocellulose content (Ibrahim et al., 2012; Sklavounos et al., 2013). The OPEFB high cellulose (54 – 59%) and hemicellulose (22 – 28%) content (Geng, 2013; Umikalsom et al., 1997) can be converted into fermentable sugars which comprise of hexoses and pentoses. Biobutanol producing *Clostridium* species are known to metabolised these hexoses and pentoses, therefore offer an economical strategies for biobutanol production (Jang et al., 2012).

A typical biobutanol production using OPEFB requires two major steps: (1) enzymatic saccharification of OPEFB for fermentable sugars production and (2) acetone-butanol-ethanol (ABE) fermentation for biobutanol production. These two steps are done separately and known as separate saccharification and fermentation (SHF) process. Studies have been carried out by combining those two major steps within a single reaction vessel known as simultaneous saccharification and fermentation (SSF) process (Sasaki et al., 2014). The SSF process can reduce the number of step involved in conversion of OPEFB to biobutanol as compared to SHF process (Ibrahim et al., 2015a). Besides, it can also decrease the inhibition of sugars on enzyme as the sugars released is simultaneously being consumed by the fermenting microorganisms (Su et al., 2015). The major challenges faced in SSF is the difference of optimum operating temperature for saccharification which is 40 – 50°C and ABE fermentation which occur at 30 – 37°C (Salehi and Taherzadeh, 2015). The cellulase in saccharification works optimally at higher temperature than fermenting bacteria that produce biobutanol. Besides temperature, pH and cellulase loading also are generally different (Oberoi et al., 2011b). Furthermore, the biobutanol production also required an optimum process control on various fermentation factors including initial pH, substrate concentration and inoculum concentration (Al-Shorgani et al., 2015). Due to the advantages and challenges of the biobutanol production through SSF process, further optimisation studies to improve the biobutanol production are valuable to pursue. Optimisation tools such as one factor at a time (OFAT) and response surface methodology (RSM) are among the widely implemented tools used for seeking optimum conditions for biobutanol production (Al-Shorgani et al., 2015; Razak et al., 2013).

## 1.2 Problem Statement

The SSF operating conditions were not study in depth, due to the difference in the optimum operating conditions of enzymatic saccharification and ABE fermentation if performed individually. Therefore, the SSF operating conditions needs to be optimised by investigating the factors such as temperature, initial pH, cellulase loading, substrate concentration and inoculum concentration towards biobutanol production.

### **1.3 Research Objectives**

The overall objective of this study is to improve biobutanol production through simultaneous saccharification and fermentation (SSF) using oil palm empty fruit bunch (OPEFB). The specific objectives are:

1. To investigate the effect of environmental factors using one factor at a time (OFAT) approach on biobutanol production through simultaneous saccharification and fermentation (SSF) from OPEFB.
2. To optimise the biobutanol production through simultaneous saccharification and fermentation (SSF) from OPEFB using Central Composite Design (CCD) of Response Surface Methodology (RSM).

### **1.4 Scope of Study**

This thesis is focuses on optimisation of simultaneous saccharification and fermentation for biobutanol production using oil palm empty fruit bunch. It is divided into five chapters. Chapter 1 contained a research background, problem statement, research objectives and scope of study. Chapter 2 reviewed on the literature concerning biobutanol which includes characteristics of biobutanol, microorganisms producing biobutanol, substrates used for biobutanol production and process involved with biobutanol production. Besides, factors affecting biobutanol production in SSF and optimisation of biobutanol production were also reviewed thoroughly. Chapter 3 described the materials used and methods of experimental conditions and equipment employed in this study. Results obtained in this study are presented in Chapter 4 which can be divided in five major sections: characteristics of OPEFB, biobutanol production from SSF, one factor at a time (OFAT) investigation, optimisation by Central Composite Design (CCD) and comparison studies. Chapter 5 includes the conclusion and recommendation of this study.

### **1.5 Limitation of Study**

The limitation of this study occurred on the optimisation study as it can only be conducted in 100 mL laboratory scale operation only. This is because at 2 L bioreactor laboratory scale the operating systems are different especially the temperature control, sparging unit control, stirring/agitation unit and bigger fermentation volume. These factors application would vary in the amount of materials and the operation of the bioreactor system.

## REFERENCES

- Abd-Alla, M.H., El-Enany, A.E., 2012. Production of acetone-butanol-ethanol from spoilage date palm (*Phoenix dactylifera* L.) fruits by mixed culture of *Clostridium acetobutylicum* and *Bacillus subtilis*. *Biomass and Bioenergy* 42, 172–178.
- Abdehagh, N., Tezel, F.H., Thibault, J., 2014. Separation techniques in butanol production: Challenges and developments. *Biomass and Bioenergy* 60, 222–246.
- Aditiya, H.B., Chong, W.T., Mahlia, T.M.I., Sebayang, A.H., Berawi, M.A., Nur, H., 2016. Second generation bioethanol potential from selected Malaysia's biodiversity biomasses: A review. *Waste Manag.* 47, 46–61.
- Adney, B., Baker, J., 2008. Measurement of cellulase activities. *Natl. Renew. Energy Lab. Tech. Rep.* 8.
- AIM, 2013. National Biomass Strategy 2020: New wealth creation for Malaysia's palm oil industry.
- Al-Shorgani, N.K., Shukor, H., Abdeshahian, P., Kalil, M., Yusoff, W.M.W., Hamid, A.A., 2016. Enhanced butanol production by optimization of medium parameters using *Clostridium acetobutylicum* YM1. *Saudi J. Biol. Sci.* In Press.
- Al-Shorgani, N.K., Hamid, A.A., Mohtar, W., Yusoff, W., Kalil, M.S., 2013. Pre-optimization of medium for biobutanol production by a new isolate of solvent-producing *Clostridium*. *BioResources* 8, 1420–1430.
- Al-Shorgani, N.K.N., Shukor, H., Abdeshahian, P., Nazir, M.Y.M., Kalil, M.S., Hamid, A.A., Yusoff, W.M.W., 2015. Process optimization of butanol production by *Clostridium saccharoperbutylacetonicum* N1-4 (ATCC 13564) using palm oil mill effluent in acetone-butanol-ethanol fermentation. *Biocatal. Agric. Biotechnol.* 4, 244–249.
- Ariffin, H., Hassan, M.A., Kalsom, M.S.U., Abdullah, N., Shirai, Y., 2008. Effect of physical, chemical and thermal pretreatments on the enzymatic hydrolysis of oil palm empty fruit bunch (OPEFB). *J. Trop. Agric. Food Sci.* 36, 000–000.
- Baer, S.H., Blaschek, H.P., Smith, T.L., 1987. Effect of butanol challenge and temperature on lipid composition and membrane fluidity of butanol-tolerant *Clostridium acetobutylicum*. *Appl. Environ. Microbiol.* 53, 2854–2861.
- Bahrin, E.K., Ibrahim, M.F., Abd Razak, M.N., Abd-Aziz, S., Shah, U.K.M., Alitheen, N., Salleh, M.M., 2012. Improved cellulase production by *Botryosphaeria rhodina* from OPEFB at low level moisture condition



- through statistical optimization. *Prep. Biochem. Biotechnol.* 42, 155–170.
- Begum, S., Dahman, Y., 2015. Enhanced biobutanol production using novel clostridial fusants in simultaneous saccharification and fermentation of green renewable agriculture residues. *Biofuels, Bioprod. Biorefining* 9, 529–544.
- Bezerra, M.A., Santelli, R.E., Oliveira, E.P., Villar, L.S., Escaleira, L.A., 2008. Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta* 76, 965–977.
- Boonsombuti, A., Tangmanasakul, K., Nantapipat, J., Komolpis, K., Luengnaruemitchai, A., Wongkasemjit, S., 2016. Production of biobutanol from acid pretreated corncob using *Clostridium beijerinckii* TISTR 1461: Process optimization studies. *Prep. Biochem. Biotechnol.* 46, 141–149.
- Bowles, L.K., Ellefson, W.L., 1985. Effects of butanol on *Clostridium acetobutlicum*. *Appl. Environ. Microbiol.* 50, 1165–1170.
- Buyondo, J.P., Liu, S., 2012. Processes and bioreactor designs for butanol production from lignocellulosic biomass. *J. Bioprocess Eng. Biorefinery* 1, 1–16.
- Cai, D., Zhang, T., Zheng, J., Chang, Z., Wang, Z., Qin, P., Tan, T., 2013. Biobutanol from sweet sorghum bagasse hydrolysate by a hybrid pervaporation process. *Bioresour. Technol.* 145, 97–102.
- Cardona, C.A., Sánchez, Ó.J., Gutierrez, L.F., 2010. Process synthesis for fuel ethanol production, Focus on Catalysts. CRC Press.
- Chandel, A.K., Es, C., Rudravaram, R., Narasu, M.L., Rao, V., Ravindra, P., 2007. Economics and environmental impact of bioethanol production technologies : an appraisal. *Biotechnol. Mol. Biol. Rev.* 2, 014–032.
- Chang, S.H., 2014. An overview of empty fruit bunch from oil palm as feedstock for bio-oil production. *Biomass and Bioenergy* 62, 174–181.
- Chen, W., Chen, Y., Lin, J., 2013. Evaluation of biobutanol production from non-pretreated rice straw hydrolysate under non-sterile environmental conditions. *Bioresour. Technol.* 135, 262–268.
- Chiew, Y.L., Shimada, S., 2013. Current state and environmental impact assessment for utilizing oil palm empty fruit bunches for fuel , fiber and fertilizer - A case study of Malaysia. *Biomass and Bioenergy* 51, 109–124.
- Chua, T.K., Liang, D., Qi, C., Yang, K., He, J., 2013. Characterization of a butanol-acetone-producing *Clostridium* strain and identification of its solventogenic genes. *Bioresour. Technol.* 135, 372–378.

- Dada, O., Kalil, M.S., Yusoff, W.M.W., 2012. Effects of inoculum and substrate concentration in anaerobic fermentation of treated rice bran to acetone, butanol, ethanol. *Bacteriol. J.* 2, 79–89.
- Dahnum, D., Tasum, S.O., Triwahyuni, E., Nurdin, M., Abimanyu, H., 2015. Comparison of SHF and SSF processes using enzyme and dry yeast for optimization of bioethanol production from empty fruit bunch. *Energy Procedia* 68, 107–116.
- Darshini, D., Dwivedi, P., Glenk, K., 2013. Capturing stakeholder's views on oil palm-based biofuel and biomass utilisation in Malaysia. *Energy Policy* 62, 1128–1137.
- Desai, K.M., Survase, S.A., Saudagar, P.S., Lele, S.S., Singhal, R.S., 2008. Comparison of artificial neural network (ANN) and response surface methodology (RSM) in fermentation media optimization: Case study of fermentative production of scleroglucan. *Biochem. Eng. J.* 41, 266–273.
- Dong, J.J., Ding, J.C., Zhang, Y., Ma, L., Xu, G.C., Han, R.Z., Ni, Y., 2016. Simultaneous saccharification and fermentation of dilute alkaline-pretreated corn stover for enhanced butanol production by *Clostridium saccharobutylicum* DSM 13864. *FEMS Microbiol. Lett.*
- Dowe, N., McMillan, J., 2008. SSF Experimental Protocols — Lignocellulosic Biomass Hydrolysis and Fermentation, National Renewable Energy Laboratory Technical Report.
- Durre, P., 2007. Biobutanol: An attractive biofuel. *Biotechnol. J.* 2, 1525–1534.
- Elbeshbishy, E., Dhar, B.R., Hafez, H., Lee, H., 2015. Acetone-butanol-ethanol production in a novel continuous flow system. *Bioresour. Technol.* 190, 315–320.
- Esfahanian, M., Nikzad, M., Najafpour, G., Ghoreyshi, A.A., 2013. Modeling and optimization of ethanol fermentation using *Saccharomyces cerevisiae*: Response surface methodology and artificial neural network. *Chem. Ind. Chem. Eng. Quartely* 19, 241–252.
- Ezeji, T.C., Qureshi, N., Blaschek, H.P., 2004. Butanol fermentation research: Upstream and downstream manipulations. *Chem. Rec.* 4, 305–314.
- Ezeji, T.C., Qureshi, N., Blaschek, H.P., 2007. Bioproduction of butanol from biomass: From genes to bioreactors. *Curr. Opin. Biotechnol.* 18, 220–227.
- Frey, D.D., Engelhardt, F., Greitzer, E.M., 2003. A role for “one-factor-at-a-time” experimentation in parameter design. *Res. Eng. Des.* 14, 65–74.
- Fujii, T., Fang, X., Inoue, H., Murakami, K., Sawayama, S., 2009. Enzymatic



- hydrolyzing performance of *Acremonium cellulolyticus* and *Trichoderma reesei* against three lignocellulosic materials. *Biotechnol. Biofuels* 2, 24.
- Gallego, L.J., Escobar, A., Peñuela, M., Peña, J.D., Rios, L.A., 2015. King Grass: A promising material for the production of second-generation butanol. *Fuel* 143, 399–403.
- García, V., Pääkilä, J., Ojamo, H., Muurinen, E., Keiski, R.L., 2011. Challenges in biobutanol production: How to improve the efficiency? *Renew. Sustain. Energy Rev.* 15, 964–980.
- Geng, A., 2013. Conversion of oil palm empty fruit bunch to biofuels, in: Fang, Z. (Ed.), *Liquid, Gaseous and Solid Biofuels - Conversion Techniques*. InTech.
- Gheshlaghi, R., Scharer, J.M., Moo-Young, M., Chou, C.P., 2009. Metabolic pathways of *Clostridia* for producing butanol. *Biotechnol. Adv.* 27, 764–781.
- Goering, H.K., Van Soest, P.J., 1970. Forage fiber analyses (Apparatus, reagents, procedures, and some applications), in: *Agriculture Handbook No 379*. US Government Printing Office, Washington, pp. 1–19.
- Goh, C.S., Tan, K.T., Lee, K.T., Bhatia, S., 2010. Bio-ethanol from lignocellulose: Status, perspectives and challenges in Malaysia. *Bioresour. Technol.* 101, 4834–4841.
- Green, E.M., 2011. Fermentative production of butanol-The industrial perspective. *Curr. Opin. Biotechnol.* 22, 337–343.
- Gu, Y., Hu, S., Chen, J., Shao, L., He, H., Yang, Y., Yang, S., Jiang, W., 2009. Ammonium acetate enhances solvent production by *Clostridium acetobutylicum* EA 2018 using cassava as a fermentation medium. *J. Ind. Microbiol. Biotechnol.* 36, 1225–1232.
- Guan, W., Shi, S., Tu, M., Lee, Y.Y., 2016. Acetone–butanol–ethanol production from Kraft paper mill sludge by simultaneous saccharification and fermentation. *Bioresour. Technol.* 200, 713–721.
- Gupta, A., Verma, J.P., 2015. Sustainable bio-ethanol production from agro-residues: A review. *Renew. Sustain. Energy Rev.* 41, 550–567.
- Han, M., Kim, Y., Kim, S.W., Choi, G.W., 2011. High efficiency bioethanol production from OPEFB using pilot pretreatment reactor. *J. Chem. Technol. Biotechnol.* 86, 1527–1534.
- Ibrahim, M.F., Abd-Aziz, S., Razak, M.N.A., Phang, L.Y., Hassan, M.A., 2012. Oil palm empty fruit bunch as alternative substrate for acetone-butanol-ethanol production by *Clostridium butylicum* EB6. *Appl. Biochem.*

Biotechnol. 166, 1615–1625.

- Ibrahim, M.F., Abd-Aziz, S., Yusoff, M.E.M., Phang, L.Y., Hassan, M.A., 2015a. Simultaneous enzymatic saccharification and ABE fermentation using pretreated oil palm empty fruit bunch as substrate to produce butanol and hydrogen as biofuel. *Renew. Energy* 77, 447–455.
- Ibrahim, M.F., Linggang, S., Jenol, M.A., Yee, P.L., Abd-Aziz, S., 2015b. Effect of buffering system on acetone-butanol-ethanol fermentation by *Clostridium acetobutylicum* ATCC 824 using pretreated oil palm empty fruit bunch. *BioResources*.
- Ibrahim, M.F., Ramli, N., Kamal Bahrin, E., Abd-Aziz, S., 2017. Cellulosic biobutanol by Clostridia: Challenges and improvements. *Renew. Sustain. Energy Rev.* 79, 1241–1254.
- Ibrahim, M.F., Razak, M.N.A., Phang, L.Y., Hassan, M.A., Abd-Aziz, S., 2013. Crude cellulase from oil palm empty fruit bunch by *Trichoderma asperellum* UPM1 and *Aspergillus fumigatus* UPM2 for fermentable sugars production. *Appl. Biochem. Biotechnol.* 170, 1320–1335.
- Inui, M., Suda, M., Kimura, S., Yasuda, K., Suzuki, H., Toda, H., Yamamoto, S., Okino, S., Suzuki, N., Yukawa, H., 2008. Expression of *Clostridium acetobutylicum* butanol synthetic genes in *Escherichia coli*. *Appl. Microbiol. Biotechnol.* 77, 1305–1316.
- Irfan, M., Asghar, U., Nadeem, M., Nelofer, R., Syed, Q., Shakir, H.A., Qazi, J.I., 2016. Statistical optimization of saccharification of alkali pretreated wheat straw for bioethanol production. *Waste and Biomass Valorization* 7, 1389–1396.
- Jang, Y., Malaviya, A., Cho, C., Lee, J., Lee, S.Y., Yup, S., 2012. Butanol production from renewable biomass by clostridia. *Bioresour. Technol.* 123, 653–663.
- Janssen, H., Wang, Y., Blaschek, H.P., 2014. *Clostridium acetobutylicum*, in: Batt, C., Batt, C.A. (Eds.), *Encyclopedia of Food Microbiology*, Volume 1. Elsevier, pp. 449–457.
- Jeon, H., Kang, K., Jeong, J., Gong, G., Choi, J., Abimanyu, H., Ahn, B.S., Suh, D., Choi, G., 2014. Production of anhydrous ethanol using oil palm empty fruit bunch in a pilot plant. *Biomass and Bioenergy* 67, 99–107.
- Jiang, W., Wen, Z., Wu, M., Li, H., Yang, J., Lin, J., Lin, Y., Yang, L., Cen, P., 2014. The effect of pH control on acetone-butanol-ethanol fermentation by *Clostridium acetobutylicum* ATCC 824 with xylose and D-glucose and D-xylose mixture. *Chinese J. Chem. Eng.* 22, 937–942.
- Jin, C., Yao, M., Liu, H., F. Lee, C., Ji, J., 2011. Progress in the production and

- application of n-butanol as a biofuel. *Renew. Sustain. Energy Rev.* 15, 4080–4106.
- Jinn, C.M., San, H.P., Ling, C.K., Wen, C.E., Tahir, P.M., Hua, L.S., Chen, L.W., Chuah, L., Maminski, M., 2015. Empty fruit bunches in the race for energy, biochemical, and material industry, in: Hakeem, K.R., Jawaid, M., Alothman, O.Y. (Eds.), *Agricultural Biomass Based Potential Materials*. Springer International Publishing Switzerland, pp. 375–389.
- Jones, D.T., Woods, D.R., 1986. Acetone-butanol fermentation revisited. *Microbiol. Mol. Biol. Rev.* 50, 484–524.
- Karimi, K., Tabatabaei, M., Horváth, I.S., Kumar, R., 2015. Recent trends in acetone, butanol, and ethanol (ABE) production. *Biofuel Res. J.* 8, 301–308.
- Khamaiseh, E.I., Abdul Hamid, A., Abdesahian, P., Wan Yusoff, W.M., Kalil, M.S., 2014. Enhanced butanol production by *Clostridium acetobutylicum* NCIMB 13357 grown on date fruit as carbon source in P2 medium. *Sci. World J.* 2014.
- Khamaiseh, E.I., Kalil, M.S., Dada, O., El-Shawabkeh, I., Yusoff, W.M.W., 2012. Date fruit as carbon source in RCM-modified medium to produce biobutanol by *Clostridium acetobutylicum* NCIMB 13357. *J. Appl. Sci.* 12, 1160–1165.
- Khamaiseh, E.I.S., Hamid, A.A., Yusoff, W.M.W., Kalil, M.S., 2013. Effect of some environment parameters on biobutanol production by *Clostridium acetobutylicum* NCIMB 13357 in date fruit medium. *Pakistan J. Biol. Sci.* 16, 1145–1151.
- Kim, J.S., Lee, Y.Y., Kim, T.H., 2016. A review on alkaline pretreatment technology for bioconversion of lignocellulosic biomass. *Bioresour. Technol.* 199, 42–48.
- Komonkiat, I., Cheirsilp, B., 2013. Felled oil palm trunk as a renewable source for biobutanol production by *Clostridium* spp. *Bioresour. Technol.* 146, 200–207.
- Krishna, S.H., Chowdary, G. V., 2000. Optimization of simultaneous saccharification and fermentation for the production of ethanol from lignocellulosic biomass. *J. Agric. Food Chem.* 48, 1971–1976.
- Kumar, M., Gayen, K., 2011. Developments in biobutanol production: New insights. *Appl. Energy* 88, 1999–2012.
- Kumar, M., Gayen, K., 2012. Biobutanol: The future biofuel, in: Baskar, C., Baskar, S., Dhillon, R.S. (Eds.), *Biomass Conversion*. Springer-Verlag Berlin Heidelberg, pp. 221–236.

- Lan, T.Q., Lou, H., Zhu, J.Y., 2012. Enzymatic saccharification of lignocelluloses should be conducted at elevated pH 5.2-6.2. *BioEnergy Res.* 6, 476–485.
- Law, L., 2010. Production of biobutanol from white grape pomace by *Clostridium saccharobutylicum* using submerged fermentation. Auckland University of Technology.
- Lee, K.T., Ofori-boateng, C., 2013. Sustainability of biofuel production from oil palm biomass. Springer Singapore Heidelberg, New York Dordrecht London.
- Lee, S.Y., Park, J.H., Jang, S.H., Nielsen, L.K., Kim, J., Jung, K.S., 2008. Fermentative butanol production by clostridia. *Biotechnol. Bioeng.* 101, 209–228.
- Li, J., Wang, L., Chen, H., 2016. Periodic peristalsis increasing acetone-butanol-ethanol productivity during simultaneous saccharification and fermentation of steam exploded corn straw. *J. Biosci. Bioeng.* 122, 620–626.
- Lin, Y., Wang, J., Wang, X.M., Sun, X.H., 2011. Optimization of butanol production from corn straw hydrolysate by *Clostridium acetobutylicum* using response surface method. *Chinese Sci. Bull.* 56, 1422–1428.
- Linggang, S., Phang, L.Y., Wasoh, H., Abd-Aziz, S., 2013. Acetone – butanol – ethanol production by *Clostridium acetobutylicum* ATCC 824 using sago pith residues hydrolysate. *Bioenergy Resour.* 6, 321–328.
- Linggang, S., Phang, L.Y., Wasoh, M.H., Abd-Aziz, S., 2012. Sago pith residue as an alternative cheap substrate for fermentable sugars production. *Appl. Biochem. Biotechnol.* 167, 122–131.
- López-Contreras, A.M., Martens, A.A., Szijarto, N., Mooibroek, H., Claassen, P.A.M., Oost, V.D.J., de Vos, W.M., 2003. Production by *Clostridium acetobutylicum* ATCC 824 of CelG, a cellulosomal glycoside hydrolase belonging to family 9. *Appl. Environ. Microbiol.* 69, 869–877.
- Loyarkat, S., Cheirsilp, B., Umsakul, K., 2013. Decanter cake waste as a renewable substrate for biobutanol production by *Clostridium beijerinckii*. *Process Biochem.* 48, 1933–1941.
- Maddox, I.S., Steiner, E., Hirsch, S., Wessner, S., Gutierrez, N.A., Gapes, J.R., Schuster, K.C., 2000. The cause of acid crash and acidogenic fermentations during the batch acetone-butanol-ethanol (ABE) fermentation process. *J. Mol. Microbiol. Biotechnol.* 2, 95–100.
- Madiah, M.S., Ariff, A.B., Sahaid, K.M., Suraini, A.A., Karim, M.I.A., 2001. Direct fermentation of gelatinized sago starch to acetone-1-butanol-

- ethanol by *Clostridium acetobutylicum*. Folia Microbiol. (Praha). 46, 197–204.
- Marchal, R., Rebeller, M., Vandecasteele, J.P., 1984. Direct bioconversion of alkali-pretreated straw using simultaneous enzymatic hydrolysis and acetone-butanol fermentation. Biotechnol. Lett. 6, 523–528.
- Mascal, M., 2012. Chemicals from biobutanol: Technologies and markets. Biofuels, Bioprod. Biorefining 6, 483–493.
- Meng Hon, L., Joseph, 2010. A case study on palm empty fruit bunch as energy feedstock. SEGi Rev. 3, 3–15.
- Miller, G.L., 1959. Use of dinitrosalicylic acid reagent for determination of reducing sugar. Anal. Chem. 31, 426–428.
- Monot, F., Martin, J.R., Petitdemange, H., Gay, R., 1982. Acetone and butanol production by *Clostridium acetobutylicum* in synthetic medium. Appl. Environ. Microbiol. 44, 1318–1324.
- Moon, H.G., Jang, Y.S., Cho, C., Lee, J., Binkley, R., Lee, S.Y., 2016. One hundred years of clostridial butanol fermentation. FEMS Microbiol. Lett. 363.
- Morone, A., Pandey, R.A., 2014. Lignocellulosic biobutanol production: Gridlocks and potential remedies. Renew. Sustain. Energy Rev. 37, 21–35.
- Nasr, N., Hafez, H., El Naggar, M.H., Nakhla, G., 2013. Application of artificial neural networks for modeling of biohydrogen production. Int. J. Hydrogen Energy 38, 3189–3195.
- Nasrah, N.S.M., Zahari, M.A.K.M., Masngut, N., Ariffin, H., 2017. Statistical optimization for biobutanol production by *Clostridium acetobutylicum* ATCC 824 from oil palm frond (OPF) juice using response surface methodology. MATEC Web Conf. 111, 1–8.
- Ndaba, B., Chiyanzu, I., Marx, S., 2015. N-Butanol derived from biochemical and chemical routes: A review. Biotechnol. Reports 8, 1–9.
- Nigam, P.S., Singh, A., 2011. Production of liquid biofuels from renewable resources. Prog. Energy Combust. Sci. 37, 52–68.
- Noomtim, P., Cheirsilp, B., 2011. Production of butanol from palm empty fruit bunches hydrolyzate by *Clostridium acetobutylicum*. Energy Procedia 9, 140–146.
- O'Brien, R.W., Morris, J.G., 1971. Oxygen and the growth and metabolism of *Clostridium acetobutylicum*. J. Gen. Microbiol. 68, 307–318.



- Oberoi, H.S., Vadlani, P. V., Nanjundaswamy, A., Bansal, S., Singh, S., Kaur, S., Babbar, N., 2011a. Enhanced ethanol production from Kinnow mandarin (*Citrus reticulata*) waste via a statistically optimised simultaneous saccharification and fermentation process. *Bioresour. Technol.* 102, 1593–1601.
- Oberoi, H.S., Vadlani, P. V., Saida, L., Bansal, S., Hughes, J.D., 2011b. Ethanol production from banana peels using statistically optimized simultaneous saccharification and fermentation process. *Waste Manag.* 31, 1576–1584.
- Olofsson, K., Bertilsson, M., Lidén, G., 2008. A short review on SSF - an interesting process option for ethanol production from lignocellulosic feedstocks. *Biotechnol. Biofuels* 1.
- Patakova, P., Linhova, M., Rychtera, M., Paulova, L., Melzoch, K., 2013. Novel and neglected issues of acetone – butanol – ethanol (ABE) fermentation by clostridia: *Clostridium* metabolic diversity, tools for process mapping and continuous fermentation systems. *Biotechnol. Adv.* 31, 58–67.
- Philippidis, G.P., Smith, T.K., Wyman, C.E., 1993. Study of the enzymatic hydrolysis of cellulose for production of fuel ethanol by the simultaneous saccharification and fermentation process. *Biotechnol. Bioeng.* 41, 846–853.
- Qdais, H.A., Hani, K.B., Shatnawi, N., 2010. Modeling and optimization of biogas production from a waste digester using artificial neural network and genetic algorithm. *Resour. , Conserv. Recycl.* 54, 359–363.
- Qi, B., Chen, X., Yi, S., Wan, Y., 2013. Inhibition of cellulase,  $\beta$ -glucosidase, and xylanase activities and enzymatic hydrolysis of dilute acid pretreated wheat straw by acetone-butanol-ethanol fermentation products. *Environ. Prog. Sustain. Energy* 1–7.
- Qureshi, N., Saha, B.C., Cotta, M.A., 2007. Butanol production from wheat straw hydrolysate using *Clostridium beijerinckii*. *Bioprocess Biosyst. Eng.* 30, 419–427.
- Qureshi, N., Saha, B.C., Cotta, M.A., 2008. Butanol production from wheat straw by simultaneous saccharification and fermentation using *Clostridium beijerinckii*: Part II—Fed-batch fermentation. *Biomass and Bioenergy* 32, 176–183.
- Qureshi, N., Singh, V., Liu, S., Ezeji, T.C., Saha, B.C., Cotta, M.A., 2014. Process integration for simultaneous saccharification, fermentation and recovery (SSFR): Production of butanol from corn stover using *Clostridium beijerinckii* P260. *Bioresour. Technol.* 154, 222–228.
- Rahnama, N., Mamat, S., Md Shah, U.K., Hooi Ling, F., Abdul Rahman, N.A.,

- Ariff, A.B., 2013. Effect of alkali pretreatment of rice straw on cellulase and xylanase production by local *Trichoderma harzianum* SNRS3 under solid state fermentation. *BioResources* 8, 2881–2896.
- Ranjan, A., Khanna, S., Moholkar, V.S., 2013a. Feasibility of rice straw as alternate substrate for biobutanol production. *Appl. Energy* 103, 32–38.
- Ranjan, A., Mayank, R., Moholkar, V.S., 2013b. Development of semi-defined rice straw-based medium for butanol production and its kinetic study. *3 Biotech* 3, 353–364.
- Razak, M.N.A., Ibrahim, M.F., Phang, L.Y., Hassan, M.A., Abd-Aziz, S., 2013. Statistical optimization of biobutanol production from oil palm decanter cake hydrolysate by *Clostridium acetobutylicum* ATCC 824. *Bioresources* 8, 1758–1770.
- Salehi, J.G., Taherzadeh, M.J., 2015. Advances in consolidated bioprocessing systems for bioethanol and butanol production from biomass: A comprehensive review. *Biofuel Res. J.* 5, 152–195.
- Sasaki, C., Kushiki, Y., Asada, C., Nakamura, Y., 2014. Acetone–butanol–ethanol production by separate hydrolysis and fermentation (SHF) and simultaneous saccharification and fermentation (SSF) methods using acorns and wood chips of *Quercus acutissima* as a carbon source. *Ind. Crops Prod.* 62, 286–292.
- Schmidt, F.R., 2005. Optimization and scale up of industrial fermentation processes. *Appl. Microbiol. Biotechnol.* 68, 425–435.
- Shah, M.M., Lee, Y.Y., 1992. Simultaneous saccharification and extractive fermentation for acetone /butanol production from pretreated hardwood. *Appl. Biochem. Biotechnol.* 34/35, 557–568.
- Shao, M., Chen, H., 2015. Feasibility of acetone-butanol-ethanol (ABE) fermentation from *Armorphophallus konjac* waste by *Clostridium acetobutylicum* ATCC 824. *Process Biochem.* 50, 1301–1307.
- Shen, J., Agblevor, F.A., 2008. Optimization of enzyme loading and hydrolytic time in the hydrolysis of mixtures of cotton gin waste and recycled paper sludge for the maximum profit rate. *Biochem. Eng. J.* 41, 241–250.
- Shirkavand, E., Baroutian, S., Gapes, D.J., Young, B.R., 2016. Combination of fungal and physicochemical processes for lignocellulosic biomass pretreatment - A review. *Renew. Sustain. Energy Rev.* 54, 217–234.
- Shukor, H., Al-Shorgani, N.K.N., Abdeslahian, P., Hamid, A.A., Anuar, N., Rahman, N.A., Kalil, M.S., 2014. Production of butanol by *Clostridium saccharoperbutylacetonicum* N1-4 from palm kernel cake in acetone-butanol-ethanol fermentation using an empirical model. *Bioresour.*

Technol. 170, 565–573.

Sklavounos, E., Iakovlev, M., Survase, S., Granström, T., Heiningen, A. V., 2013. Oil palm empty fruit bunch to biofuels and chemicals via SO<sub>2</sub>-ethanol – water fractionation and ABE fermentation. *Bioresour. Technol.* 147, 102–109.

Soplah, S., Abdullah, S., Shirai, Y., Kamal, E., Ali, M., 2015. Fresh oil palm frond juice as a renewable, non-food, non-cellulosic and complete medium for direct bioethanol production. *Ind. Crops Prod.* 63, 357–361.

Spindler, D.D., Wyman, C.E., Grohmann, K., Mohagheghi, A., 1989. Simultaneous saccharification and fermentation of pretreated wheat straw to ethanol with selected yeast strains and  $\beta$ -glucosidase supplementation. *Appl. Biochem. Biotechnol.* 20/21, 529–540.

Srirangan, K., Akawi, L., Moo-Young, M., Chou, C.P., 2012. Towards sustainable production of clean energy carriers from biomass resources. *Appl. Energy* 100, 172–186.

Stanbury, P.F., Whitaker, A., Hall, S.J., 2003. Principles of fermentation technology, Butterworth Heinemann. Elsevier Science.

Steen, E.J., Chan, R., Prasad, N., Myers, S., Petzold, C.J., Redding, A., Ouellet, M., Keasling, J.D., 2008. Metabolic engineering of *Saccharomyces cerevisiae* for the production of n-butanol. *Microb. Cell Fact.* 7.

Stenberg, K., Bollók, M., Réczey, K., Galbe, M., Zacchi, G., 2000. Effect of substrate and cellulase concentration of simultaneous saccharification and fermentation of steam-pretreated softwood for ethanol production. *Biotechnol. Bioeng.* 68, 204–210.

Su, H., Liu, G., He, M., Tan, F., 2015. A biorefining process: Sequential, combinational lignocellulose pretreatment procedure for improving biobutanol production from sugarcane bagasse. *Bioresour. Technol.* 187, 149–160.

Sudiyani, Y., Sembiring, K.C., Hendarsyah, H., Alawiyah, S., 2010. Alkaline pretreatment and enzymatic saccharification of oil palm empty fruit bunch fiber for ethanol production. *Menara Perkeb.* 78, 70–74.

Sukumaran, R.K., Gottumukkala, L.D., Rajasree, K., Alex, D., Pandey, A., 2011. Butanol fuel from biomass: Revisiting ABE fermentation, in: Pandey, A., Larroche, C., Ricke, S.C., Dussap, C., Gnansounou, E. (Eds.), *Biofuels: Alternative Feedstocks and Conversion Processes*. Academic Press, p. 642.

Swana, J., Yang, Y., Behnam, M., Thompson, R., 2011. An analysis of net



- energy production and feedstock availability for biobutanol and bioethanol. *Bioresour. Technol.* 102, 2112–2117.
- Tan, H.T., Lee, K.T., Mohamed, A.R., 2010. Second-generation bio-ethanol (SGB) from Malaysian palm empty fruit bunch: Energy and exergy analyses. *Bioresour. Technol.* 101, 5719–5727.
- Tashiro, Y., Sonomoto, K., 2010. Advances in butanol production by clostridia, in: Mendez-Vilas, A. (Ed.), *Current Research, Technology and Education Topics in Applied Microbiology and Microbial Biotechnology*. Formatex Research Center, pp. 1383–1394.
- Thang, V.H., Kobayashi, G., 2013. A novel process for direct production of acetone-butanol-ethanol from native starches using granular starch hydrolyzing enzyme by *Clostridium saccharoperbutylacetonicum* N1-4. *Appl. Biochem. Biotechnol.*
- Thorn, G.J., King, J.R., Jabbari, S., 2013. pH-induced gene regulation of solvent production by *Clostridium acetobutylicum* in continuous culture: parameter estimation and sporulation modelling. *Math. Biosci.* 241, 149–66.
- Triwahyuni, E., Muryanto, Sudiyani, Y., Abimanyu, H., 2015. The effect of substrate loading on simultaneous saccharification and fermentation process for bioethanol production from oil palm empty fruit bunches. *Energy Procedia* 68, 138–146.
- Umikalsom, M., Ariff, A.B., Karim, M.I., 1998. Saccharification of pretreated oil palm empty fruit bunch fiber using cellulase of *Chaetomium globosum*. *J. Agric. Food Chem.* 46, 3359–3364.
- Umikalsom, M., Ariff, A.B., Zulkifli, H.S., Tong, C.C., Hassan, M.A., 1997. The treatment of oil palm empty fruit bunch fibre for subsequent use as substrate for cellulase production by *Chaetomium globosum* kunze. *Bioresour. Technol.* 62, 1–9.
- Uyttebroek, M., Hecke, W. V., Vanbroekhoven, K., 2013. Sustainability metrics of 1-butanol. *Catal. Today* 239, 7–10.
- Väisänen, S., Havukainen, J., Uusitalo, V., Havukainen, M., Soukka, R., Luoranen, M., 2016. Carbon footprint of biobutanol by ABE fermentation from corn and sugarcane. *Renew. Energy* 89, 401–410.
- Wang, Y., Blaschek, H.P., 2011. Optimization of butanol production from tropical maize stalk juice by fermentation with *Clostridium beijerinckii* NCIMB 8052. *Bioresour. Technol.* 102, 9985–9990.
- Wang, Z., Cao, G., Jiang, C., Song, J., Zheng, J., Yang, Q., 2013. Butanol production from wheat straw by combining crude enzymatic hydrolysis

and anaerobic fermentation using *Clostridium acetobutylicum* ATCC824. Energy and Fuels 1–7.

- Wingren, A., Galbe, M., Zacchi, G., 2003. Techno-economic evaluation of producing ethanol from softwood: Comparison of SSF and SHF and identification of bottlenecks. Biotechnol. Prog. 46, 1109–1117.
- Wu, P., Wang, G., Wang, G., Børresen, B.T., Liu, H., Zhang, J., 2016. Butanol production under microaerobic conditions with a symbiotic system of *Clostridium acetobutylicum* and *Bacillus cereus*. Microb. Cell Fact. 15, 1–11.
- Wyman, C.E., 1994. Ethanol from lignocellulosic biomass: Technology, economics, and opportunities. Bioresour. Technol. 50, 3–16.
- Xue, C., Zhao, X., Liu, C., Chen, L., Bai, F., 2013. Prospective and development of butanol as an advanced biofuel. Biotechnol. Adv. 31, 1575–84.
- Yamanobe, T., Mitsuishi, Y., Takasaki, Y., 1985. Method for manufacture cellulase, United States Patent.
- Yang, X., Tu, M., Xie, R., Adhikari, S., Tong, Z., 2013. A comparison of three pH control methods for revealing effects of undissociated butyric acid on specific butanol production rate in batch fermentation of *Clostridium acetobutylicum*. AMB Express 3, 3.
- Yoshizaki, T., Shirai, Y., Hassan, M.A., Baharuddin, A.S., Raja Abdullah, N.M., Sulaiman, A., Busu, Z., 2013. Improved economic viability of integrated biogas energy and compost production for sustainable palm oil mill management. J. Clean. Prod. 44, 1–7.
- Zahari, M.A.K.M., Zakaria, M.R., Ariffin, H., Mokhtar, M.N., Salihon, J., Shirai, Y., Hassan, M.A., 2012. Renewable sugars from oil palm frond juice as an alternative novel fermentation feedstock for value-added products. Bioresour. Technol. 110, 566–71.
- Zanirun, Z., Bahrin, E.K., Lai-Yee, P., Hassan, M.A., Abd-Aziz, S., 2015. Enhancement of fermentable sugars production from oil palm empty fruit bunch by ligninolytic enzymes mediator system. Int. Biodeterior. Biodegrad. 105, 13–20.
- Zhang, W., Lin, Y., Zhang, Q., Wang, X., Wu, D., Kong, H., 2013. Optimisation of simultaneous saccharification and fermentation of wheat straw for ethanol production. Fuel 112, 331–337.
- Zhu, Y., Xin, F., Chang, Y., Zhao, Y., Weichong, W., 2015. Feasibility of reed for biobutanol production hydrolyzed by crude cellulase. Biomass and Bioenergy 76, 24–30.