



UNIVERSITI PUTRA MALAYSIA

***MOLECULAR AND PHYSIOLOGICAL RESPONSES OF RECALCITRANT
INDICA RICE TO LIGNOSULFONATES DURING CALLUS
REGENERATION***

LOW LEE YOON

FBSB 2018 31



**MOLECULAR AND PHYSIOLOGICAL RESPONSES OF RECALCITRANT
INDICA RICE TO LIGNOSULFONATES DURING CALLUS
REGENERATION**

By

LOW LEE YOON

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

April 2018

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

MOLECULAR AND PHYSIOLOGICAL RESPONSES OF RECALCITRANT INDICA RICE TO LIGNOSULFONATES DURING CALLUS REGENERATION

By

LOW LEE YOON

April 2018

Chair : Lai Kok Song, PhD
Faculty : Biotechnology and Biomolecular Sciences

Lignosulfonate (LS) is commonly used as an enhancer to promote plant growth. The recalcitrant *Oryza sativa indica* cv. MR219 rice is an important local high yield rice cultivar that is widely cultivated in Malaysia. However, low callus regeneration rate of MR219 hinders further exploitation in cultivar improvement. Hence, LS was introduced in the culture medium in effort to enhance in vitro cultivation of MR219. To date, the effects of LS on regeneration of MR219 has not been reported. Therefore, this study was undertaken to evaluate the effects of LS on callus proliferation, shoot induction and shoot growth of MR219. The MR219 calli were proliferated on MS supplemented with different types (aNaLS and aCaLS) and concentrations (50, 100, 150, 200 mg/L) of LS. The optimum callus proliferation rate (88%) was obtained in week 3 on MS supplemented with 100 mg/L aCaLS in the presence of plant hormone. However, both LSs did not enhance the shoot induction efficiency whereby 50% of the shoot induced was albino in MS fortified with 100 mg/L CaLS. In shoot growth study, shoot apices were cultured in MS supplemented with different types (aNaLS and aCaLS) and concentrations (100, 200, 300 and 400 mg/L) of LS. The optimum shoot growth was observed in MS supplemented with 300 mg/L aNaLS that is taller by 26% of control height. To understand the growth promoting effects of LS, aCaLS treated callus was used as a study model. Results showed that aCaLS increased callus proliferation rate by 67% and adventitious root formation by 62% in MS without hormone. Hence, it was shown that the LS effect was found to be independent of hormone. Under scanning electron microscopy, adventitious roots were seen protruding out from aCaLS-treated calli. Further expression analysis of adventitious root-related genes (*OsWOX11*, *OsAUX1* and *OsIAA23*) on treated calli, *OsWOX11* expression recorded 1.7-fold expression increment, implying a positive role of aCaLS in adventitious root development. In addition, aCaLS-treated calli recorded 1.2-fold higher endogenous indole-3-acetic acid (IAA) content and increment of nutrient ions (Na, K, Ca, Mg, Fe, Mn, Zn and Cu) uptake. Consistently, expression analysis of auxin-related genes (*OsASA1*, *OsTAA1* and *OsYUC1*) and nutrient uptake-related genes (*OsAKT1*, *OsHAK5*, *OsCBL*, *OsCIPK23* and *OsCamk1*) also showed a similar increment trend. The Ca

increment was observed throughout four weeks but the major increment of K was only detected starting from week two. The observed rise of Ca following the enhancement of endogenous K content suggested the possible cross-talk between these ions uptake. The LC-MS/MS analysis suggested that there was an increased in carbon and nitrogen metabolisms in aCaLS treated callus. Taken together, the presence of aCaLS improved MR219 callus proliferation, up-regulated endogenous auxin synthesis, nutrients uptake and carbon-nitrogen metabolisms that ultimately contributed to calli growth enhancement. The findings of this study would be useful on improving the *in vitro* cultivation of the recalcitrant rice cultivars.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**PERUBAHAN MOLEKUL AND FISILOGI DALAM BERAS INDICA
REKALSITRAN TERHADAP LIGNOSULFONATES SEMASA REGENERASI
KALUS**

Oleh

LOW LEE YOON

April 2018

Pengerusi : Lai Kok Song, PhD
Fakulti : Bioteknologi and Sains Biomolekul

Lignosulfonate (LS) pada kebiasaannya digunakan sebagai perangsang bagi menggalakkan tumbesaran tumbuhan. Rekalsitran *Oryza sativa* indika cv. MR219 merupakan kultivar beras hasil tinggi tempatan yang banyak ditanam di Malaysia. Walau bagaimanapun, kadar regenerasi kalus yang rendah MR219 telah menghalang eksploitasi dalam pembaikan kultivar tersebut. Oleh itu, LS diperkenalkan dalam medium untuk memperbaiki pertumbuhan MR219 *in vitro*. Sehingga kini, kesan LS pada pertumbuhan semula beras indica rekalsitran belum dilaporkan. Oleh itu, kajian ini dijalankan untuk menilai kesan LS pada proliferasi kalus, induksi pertumbuhan pucuk MR219. Kalus MR219 dikultur di dalam MS yang ditambah dengan LS (aNaLS and aCaLS) pada kepekatan (50, 100, 150, 200 mg/L) yang berbeza-beza. Menurut pemerhatian kami, MS yang ditambah dengan 100 mg/L aCaLS menunjukkan kadar proliferasi yang paling optimum (88%) dalam medium yang mengandungi hormon. Walau bagaimanapun, kedua-dua LS tidak merangsang kadar pengaruh pucuk dimana 50% tunas yang diinduksi dalam MS yang ditambah dengan 100 mg/L aCaLS adalah albino. Dalam kajian pertumbuhan pucuk, pucuk apeks dikultur dalam MS yang ditambah dengan LS (aNaLS dan aCaLS) pada kepekatan (100, 200, 300 dan 400 mg/L) yang berbeza-beza. Pertumbuhan pucuk optimum diperhatikan pada MS yang ditambah dengan 300 mg/L aNaLS, yang mana pertumbuhan pucuk dicatatkan 26% lebih tinggi berbanding dengan kawalan. Untuk memahami kesan LS terhadap kadar pertumbuhan, kalus yang dirawat dengan aCaLS digunakan sebagai model kajian. Hasil kajian menunjukkan bahawa aCaLS meningkatkan kadar proliferasi kalus sebanyak 67% dan pembentukan akar adventif sebanyak 62% dalam media bebas hormon. LS didapati dapat memberi kesan dalam situasi bebas hormon. Di bawah pemeriksaan mikroskopi elektron, akar-akar adventif dapat dilihat menonjol keluar dari kalus yang dirawat dengan aCaLS. Kajian lebih lanjut melalui analisis gen ekspresi yang berkaitan dengan akar adventif (*OsWOX11*, *OsAUX1* dan *OsIAA23*) dijalankan pada kalus yang dirawat dengan aCaLS. Menurut analisis gen ekspresi, gen *OsWOX11* mencatatkan kenaikan 1.7 kali ganda, mengimplikasikan peranan positif aCaLS sebagai perangsang pertumbuhan akar

adventif. Tambahan pula, kalus yang dirawat aCaLS mencatatkan kandungan sebanyak 1.2 kali ganda auksin dalaman yang lebih tinggi dan peningkatan pengambilan ion nutrient telah direkodkan (Na, K, Ca, Mg, Fe, Mn, Zn dan Cu). Analisis ekspresi gen yang berkaitan dengan auksin (*OsASAI*, *OsTAA1* dan *OsYUC1*) dan gen yang berkaitan dengan pengambilan nutrien (*OsAKTI*, *OsHAK5*, *OsCBL*, *OsCIPK23* dan *OsCamk1*) juga turut menunjukkan corak peningkatan yang sama. Peningkatan Ca^{2+} telah dikesan sepanjang empat minggu tetapi kenaikan K^+ hanya dapat dikesan bermula dari minggu kedua. Peningkatan Ca^{2+} yang diikuti dengan peningkatan kandungan K^+ mencadangkan kemungkinan pengambilan ion-ion ini berhubung antara satu sama lain. Analisis LC-MS/MS menunjukkan peningkatan metabolisme karbon dan nitrogen dalam kalus yang dirawat aCaLS. Secara keseluruhannya, aCaLS telah berjaya meningkatkan proliferasi kalus MR219, penghasilan auksin dalaman, pengambilan nutrien dan metabolisme karbon-nitrogen yang akhirnya menyumbang pada penambahbaikan pertumbuhan kalus. Penemuan kajian ini berguna untuk memperbaiki pertumbuhan kultivar beras rekalsitran *in vitro*.

ACKNOWLEDGEMENTS

I would like to express my deepest gratitude to everyone who has helped me throughout the project. The completion of this study could not have been possible without the expertise and guidance of the supervisory committees: Dr. Lai, Dr. Janna and Dr. Wee. I would like to thank my family, friends and labmates for their help and moral support.

Last but not least, I would also like to acknowledge the UPM Graduate Research Fellowship and UPM Putra Grant (GP-IPS/2016/9506100) for supporting and funding the research.

Thank you from the bottom of my humble heart.

I certify that a Thesis Examination Committee has met on 17 April 2018 to conduct the final examination of Low Lee Yoon on her thesis entitled "Molecular and Physiological Responses of Recalcitrant Indica Rice to Lignosulfonates During Callus Regeneration" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Parameswari a/p Namasivayam, PhD

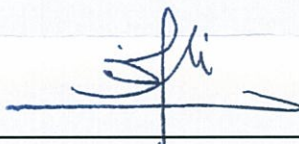
Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Noor Baity Saidi, PhD

Senior Lecturer
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal Examiner)

Tee Chong Siang, PhD

Associate Professor
Universiti Tunku Abdul Rahman
Malaysia
(External Examiner)



RUSLI HAJI ABDULLAH, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 27 September 2018

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Lai Kok Song, PhD

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Janna Ong Binti Abdullah, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

Wee Chien Yeong, PhD

Research Officer

Biotechnology and Nanotechnology Research Centre

Malaysian Agricultural Research and Development Institute (MARDI)

(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

Declaration By Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Low Lee Yoon, GS45724

Declaration By Members Of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman of
Supervisory
Committee: Dr. Lai Kok Song

Signature: _____
Name of Member of
Supervisory
Committee: Assoc. Prof. Dr. Janna Ong binti Abdullah

Signature: _____
Name of Member of
Supervisory
Committee: Dr. Wee Chien Yeong

TABLE OF CONTENTS

		Page
ABSTRACT		i
ABSTRAK		iii
ACKNOWLEDGEMENTS		v
APPROVAL		vi
DECLARATION		viii
LIST OF TABLES		xiii
LIST OF FIGURES		xiv
LIST OF ABBREVIATIONS		xvi
LIST OF APPENDICES		xvii
CHAPTER		
1	INTRODUCTION	1
2	LITERATURE REVIEW	3
	2.1 Plant tissue culture	3
	2.1.1 Callus culture and regeneration	4
	2.2 <i>Oryza sativa</i> L.	
	2.2.1 Recalcitrant <i>Oryza sativa</i> indica cv. MR219	5
	2.2.2 Indica rice tissue culture	5
	2.3 Lignosulfonate	6
	2.3.1 Application in agriculture	7
	2.3.2 Proposed LS growth promoting mechanisms	7
	2.4 Plant cell growth and development	8
	2.4.1 Auxin signaling	9
	2.4.2 Macro- and micronutrients uptake	11
	2.5 Proteomic study	12
	2.5.1 Liquid chromatography tandem mass spectrometry (LC-MS/MS)	13
3	MATERIALS AND METHODS	15
	3.1 Plant tissue culture	15
	3.1.1 Plant materials	15
	3.1.2 Lignosulfonates preparation	15
	3.1.3 Culture medium	15
	3.1.4 Seeds sterilization	15
	3.1.5 Callus induction	15
	3.1.6 Rice callus proliferation	16
	3.1.7 Shoot induction	16
	3.1.8 Shoot growth	16
	3.1.9 Plant growth data collection	17
	3.2 Scanning electron microscope (SEM)	17
	3.2.1 Sample preparation	17

3.2.2	SEM viewing	17
3.3	Chlorophyll content analysis	17
3.4	Gene expression study	18
3.4.1	Sample preparation	18
3.4.2	Real-time reverse transcription polymerase chain reaction (RT-PCR) primers design	18
3.4.3	Primers optimization and validation	18
3.4.4	Real-time reverse transcription polymerase chain reaction (RT-PCR)	19
3.4.5	RT-PCR data analysis	19
3.5	Callus nutrient ions content analysis	19
3.5.1	Sample preparation	19
3.5.2	Inductively coupled plasma mass spectrometry (ICP-MS) analysis	19
3.6	Endogenous auxin concentration analysis	20
3.6.1	Sample preparation	20
3.6.2	Liquid chromatography mass spectrometry (LC-MS) analysis	20
3.7	Proteomic analysis	21
3.7.1	Protein extraction	21
3.7.2	Liquid chromatography tandem-mass spectrometry (LC-MS/MS) analysis	21
4	RESULTS AND DISCUSSION	23
4.1	Effect of lignosulfonates treatment on <i>in vitro</i> <i>Oryza sativa</i> indica cv MR219 growth and development	23
4.1.1	Effects of different types and concentrations of lignosulfonates on callus proliferation	23
4.1.2	Effects of different types and concentrations of lignosulfonates on shoot induction	26
4.1.3	Effect of different types and concentration of lignosulfonates on shoot growth	31
4.1.4	Effect of optimized calcium lignosulfonate on callus proliferation on hormone-free medium	32
4.2	Mode of action of lignosulfonates	36
4.2.1	LS regulates endogenous auxin biosynthesis	36
4.2.2	LS regulates macro- and micronutrients uptake	39
4.3	Proteomic analysis	42
5	CONCLUSION AND FUTURE PROSPECT	51

REFERENCES	52
APPENDICES	67
BIODATA OF STUDENT	85
LIST OF PUBLICATIONS AND CONFERENCES	86



LIST OF TABLES

Table		Page
2.1	Genes involve in IAA biosynthesis in plant and their respective functions	9
2.2	Differential proteomic studies in rice plant and methods employed	12
3.1	ICP-MS operating condition.	20
4.1	Nutrient ions contents of four-week-old calli cultures on MSO (control) and MS supplemented with 100 mg/L aCaLS treated calli	40
4.2	Weekly time-point analysis of potassium and calcium ions contents in calli cultured on MSO (control) and MSO supplemented with 100 mg/L aCaLS treated calli	40
4.3	List of proteins showing significant abundance difference in aCaLS treated calli. The list is sorted according to descending fold changes of each protein and the biological process involved.	47

LIST OF FIGURES

Table	Page
4.1 FW and DW of three weeks old callus treated with different types and concentrations of LS	24
4.2 Morphology of callus in control and 50 mg/L to 200 mg/L LS treatments	25
4.3 Control callus and 100 mg/L aCaLS-treated callus with root-like structure	25
4.4 Number of callus observed with root-like structure formation in control and 100 mg/L aCaLS treatment	26
4.5 Morphological changes under aNALS and aCaLS treatments	28
4.6 Shoot induction on control and MS supplemented with LS	29
4.7 Chlorophyll contents analysis and normalized relative expression of chlorophyll biosynthesis-related genes	30
4.8 Shoot height after three weeks culture period in control and LS treatments	31
4.9 Stages involve in shoot growth from 1 cm shoot apex	32
4.10 FW and DW measurement of week 1 to week 4 calli cultured on MSO and 100 mg/L aCaLS	34
4.11 Weekly observation on the number of callus with adventitious root formation in MSO and 100 mg/L aCaLS for four weeks and normalized expression of adventitious root formation-related genes.	35
4.12 SEM analysis of calli cultured on MSO and MSO supplemented with 100 mg/L aCaLS	36
4.13 Endogenous auxin concentration in 4 week-old calli cultured in MSO and MSO supplemented with 100 mg/L aCaLS	38
4.14 The expression analysis of selected auxin biosynthesis genes in calli cultured in MSO and MSO supplemented with 100 mg/L aCaLS	38
4.15 Normalized expression of calcium signaling genes and potassium transportation genes in calli cultured in MSO and MSO supplemented with 100 mg/L aCaLS.	41

4.16	The volcano plot showing proteins significance versus fold-change difference between MSO and MSO supplemented with 100 mg/L aCaLS treatments	45
4.17	Biological processes involved in aCaLS treated calli	46
4.18	Mechanism proposed on the roles of aCaLS in callus growth.	50



LIST OF ABBREVIATIONS

2,4-D	2,4-Dichlorophenoxyacetic acid
aCaLS	Analytical grade calcium lignosulfonate
aNaLS	Analytical grade sodium lignosulfonate
ABC	Ammonium bicarbonate
ANOVA	Analysis of variance
ATP	Adenine triphosphate
BSA	Bovine serum albumin
DW	Dry weight
FW	Fresh weight
IAA	Indole-3-acetic acid
KED	Kinetic energy discrimination
LC	Liquid chromatography
LS	Lignosulfonate
NAA	1-Naphthaleneacetic acid
PMSF	Phenylmethylsulfonyl fluoride
PCR	Polymerase chain reaction
RT-PCR	Real-Time Reverse Transcription PCR
SD	Standard deviation

LIST OF APPENDICES

Appendix		Page
A	Preparation of plant tissue culture media	67
B	Sequence of primers used in RT-PCR analysis	68
C	Validation of primers used in RT-PCR analysis	69
D	Standard curves	84





© COPYRIGHT UPM

CHAPTER 1

INTRODUCTION

Rice is one of the major staple food worldwide with Asia being the top producer and consumer (Gumma *et al.*, 2011). In Malaysia, rice production was recorded at 3 million tons in 2016 (FAO, 2017). However, the current production is unable to sustain the rising local domestic consumption demands in Malaysia. Annual rice yield losses due to climate change as well as pest and disease outbreak worsens food security (Rajamoorthy and Munusamy, 2015). In order to fulfill the local rice demand, Malaysia is still largely dependent on imported rice. Thailand and Vietnam are the major countries that controlled more than 70 percent of rice imported into Malaysia. In 2016, around 675,000 tons of rice valued at 277 million US dollars was imported from these two countries (Abdul, 2017). Hence, improving rice yield and quality are seen as the most effective way in increasing the local rice production in order to reduce the high importing cost.

The *Oryza sativa indica* cv. MR219 is an important local rice cultivar that is widely cultivated in Malaysia. This cultivar is well known for its high yield, short maturation period and resistance to blast and bacterial leaf blight characteristics (FFTC, 2002). However, the recalcitrant characteristic of MR219 rice hinders genetic study on this cultivar. Low regeneration efficiency, long regeneration duration and low transformation rate of recalcitrant rice have been the major obstacles that trouble the plant biotechnologist (Raghavendra *et al.*, 2010; Sah *et al.*, 2014; Mishra and Rao, 2016). Hence, improving the *in vitro* plant cultivation medium in the critical callus regeneration stages (callus proliferation and shoot induction) is essential, before any traits enhancement modifications are performed in the recalcitrant indica rice cultivar.

Lignosulfonate (LS) is a low-cost by-product from sulfite pulping process in wooding industries that are already being commercialized as binding agent and dispersal for several industrial purposes (Yang *et al.* 2007; Almas *et al.* 2014). In agriculture, LS is widely used as an important component in fertilizer mainly because it is cost-effective and able to chelate different macro- and micronutrient (Carrasco *et al.* 2012). It is used as stimulant in plant growth, plant development and as well as in enhancing *in vitro* rooting and shoot development of ornamental plants (Telysheva *et al.*, 1992, 1997; Van der Krieken *et al.*, 2004; Rodríguez-Lucena *et al.*, 2009; Ertani *et al.*, 2011).

To date, a few hypotheses had been suggested on the enhancing effects of LS in plant growth and development. Nevertheless, the molecular mechanisms underlying these growth enhancing responses induced by LS remained largely unknown. In general applications, the incorporation of LS in planting field would allow soil amendment by reducing the soil pH and increased soil organic matters as a form of disease control strategy for soil borne disease through the enrichment of beneficial microbiota (Lazarovits, 2001; Lazarovits *et al.*, 2001; Abbasi *et al.*, 2002; Almas *et al.*, 2014). Besides, researchers also suggested that LS may play role in regulating endogenous

auxin concentration (Hausman *et al*, 1995; Gaspar *et al*, 1996); as auxin protector (Soteras, 1994); increasing tissue sensitivity to auxin (Telysheva *et al*. 1992, 1997); and as mineral-balancer where LS facilitates the transfer of macro- and micronutrients into the plant cell compartments (Yamashita and Thomas, 1996; Cieschi *et al*, 2016; Carrasco *et al*, 2012).

These interesting enhancing effects of LS have made it a potential enhancing supplement for micropropagation of various type of plant without causing any negative side-effects. To achieve a better usage of the LS components as plant media supplement, understanding the mode of activity of LS is necessary. Hence, we hypothesized that LS could improve the *in vitro* cultivation of MR219 through regulation of endogenous auxin and nutrient uptake in MR219. To address this problem, the present study was undertaken to improve the *in vitro* cultivation medium of MR219 rice through the supplementation of growth enhancer namely lignosulfonate (LS). Taken together, this study was aimed to investigate the effects of LS on callus regeneration of MR219 rice *in vitro* at three different stages which were callus proliferation, shoot induction and shoot growth. Furthermore, the present experimental study also aimed to further determine and elucidate the mode of action of LS in MR219 metabolism, in an effort of verifying potential hypothesized mechanisms that lead to plant growth enhancement phenomenon.

Therefore, the objectives of this study were:

1. To study the effects of LS on callus proliferation, shoot induction and shoot growth of recalcitrant MR219.
2. To study the molecular and physiological responses of recalcitrant MR219 to LS.

REFERENCES

- Abbasi, P.A., Al-Dahmani, J., Sahin, F., Hoitink, H.A.J. and Miller, S.A. (2002). Effect of compost amendments on disease severity and yield of tomato in conventional and organic production systems. *Plant Disease*, 86(2): 156-161.
- Abdul, G. W. (2017). GAIN REPORT: Grain and Feed Annual 2017. *USDA Foreign Agricultural Service*, MY7001: 1-8.
- Abiri, R., Maziah, M., Shaharuddin, N.A., Yusof, Z.N.B., Atabaki, N., Hanafi, M.M., Sahebi, M., Azizi, P., Kalhori, N. and Valdiani, A. (2017). Enhancing somatic embryogenesis of Malaysian rice cultivar MR219 using adjuvant materials in a high-efficiency protocol. *International Journal of Environmental Science and Technology*, 14(5): 1091-1108.
- Abu-Zaitoon, Y. M., Bennett, K., Normanly, J. and Nonhebel, H. M. (2012). A large increase in IAA during development of rice grains correlates with the expression of tryptophan aminotransferase *OsTARI* and a grain-specific *YUCCA*. *Physiologia Plantarum*, 146(4): 487-499.
- Ahmad, I., Mian, A. and Maathuis, F. J. (2016). Overexpression of the rice *AKT1* potassium channel affects potassium nutrition and rice drought tolerance. *Journal of Experimental Botany*, 67(9): 2689-2698.
- Ahmed, T., Biswas, S., Elias, S. M., Rahman, M. S., Tuteja, N. and Seraj, Z. I. (2017). In Planta transformation for conferring salt tolerance to a tissue-culture unresponsive indica rice (*Oryza sativa* L.) cultivar. *In Vitro Cellular & Developmental Biology-Plant*, pp: 1-12.
- Ahsan, N., Lee, D. G., Alam, I., Kim, P. J., Lee, J. J., Ahn, Y. O., Kwak, S. S., Lee, I. J., Bahk, J. D., Kang, K. Y. and Renaut, J. (2008). Comparative proteomic study of arsenic-induced differentially expressed proteins in rice roots reveals glutathione plays a central role during As stress. *Proteomics*, 8(17): 3561-3576.
- Alam, M. J., Imran, M., Hassan, L., Rubel, M. H. and Shamsuddoha, M. J. (2012). *In vitro* regeneration of high yielding indica rice (*Oryza sativa* L.) varieties. *Journal of Environmental science and Natural Resources*, 5: 173-177.
- Aldemita, R. R., Hodges, T. K. (1996). *Agrobacterium tumefaciens* mediated transformation of japonica and indica rice varieties. *Planta*, 199: 612-617.
- Almas, Å.R., Afanou, A.K. and Krogstad, T. (2014). Impact of lignosulfonate on solution chemistry and phospholipid fatty acid composition in soils. *Pedosphere*, 24(3): 308-321.
- Armengaud, P., Breiting, R. and Amtmann, A. (2004). The potassium-dependent transcriptome of Arabidopsis reveals a prominent role of jasmonic acid in nutrient signaling. *Plant physiology*, 136(1): 2556-2576.
- Aro, T. and Fatehi, P., 2017. Production and application of lignosulfonates and sulfonated lignin. *ChemSusChem*, 10(9): 1861–1877.
- Aryal, U. K., Xiong, Y., McBride, Z., Kihara, D., Xie, J., Hall, M. C. and Szymanski, D. B. (2014). A proteomic strategy for global analysis of plant protein complexes. *The Plant Cell*, 26(10): 3867-3882.
- Assani, A., Chabane, D., Foughi-Wehr, B. and Wenzel, G. (2006). An improved protocol for microcallus production and whole plant regeneration from recalcitrant banana protoplasts (*Musa spp.*). *Plant cell, Tissue and Organ Culture*, 85(3): 257-264.

- Bardini, M., Labra, M., Winfield, M. and Sala, F., 2003. Antibiotic-induced DNA methylation changes in calluses of *Arabidopsis thaliana*. *Plant Cell, Tissue and Organ Culture*, 72(2): 157-162.
- Barford, D. (2004). The role of cysteine residues as redox-sensitive regulatory switches. *Current Opinion In Structural Biology*, 14(6): 679-686.
- Bartel, B. and Fink, G.R. (1994). Differential regulation of an auxin-producing nitrilase gene family in *Arabidopsis thaliana*. *Proceedings of the National Academy of Sciences*, 91(14): 6649-6653.
- Bhatia, S. and Sharma, K. (2015). Microenvironmentation in micropropagation. *Modern Applications of Plant Biotechnology in Pharmaceutical Sciences*, pp: 345-360
- Biswas, A. and Mandal, A. B. (2007). Plant regeneration in different genotypes of indica rice. *Indian Journal of Biotechnology*, 6(4): 532-540.
- Bogeat-Triboulot, M. B., Brosché, M., Renaut, J., Jouve, L., Le Thiec, D. and Fayyaz, P. (2007). Gradual soil water depletion results in reversible changes of gene expression, protein profiles, ecophysiology, and growth performance in *Populus euphratica*, a poplar growing in arid regions. *Plant Physiology*, 143: 876-892.
- Britto, D.T. and Kronzucker, H.J. (2008). Cellular mechanisms of potassium transport in plants. *Physiologia Plantarum*, 133(4): 637-650.
- Carrasco, J., Kovács, K., Czech, V., Fodor, F., Lucena, J. J., Vértes, A. and Hernández-Apaolaza, L. (2012). Influence of pH, iron source, and Fe/ligand ratio on iron speciation in lignosulfonate complexes studied using Mössbauer spectroscopy. Implications on their fertilizer properties. *Journal of agricultural and food chemistry*, 60(13): 3331-3340.
- Cava-Montesinos, P., Cervera, M.L., Pastor, A. and de la Guardia, M. (2005). Room temperature acid sonication ICP-MS multielemental analysis of milk. *Analytica Chimica Acta*, 531(1): 111-123.
- Chen, G., Hu, Q., Luo, L. E., Yang, T., Zhang, S., Hu, Y., Yu, L. and Xu, G. (2015). Rice potassium transporter OsHAK1 is essential for maintaining potassium-mediated growth and functions in salt tolerance over low and high potassium concentration ranges. *Plant, Cell and Environment*, 38(12): 2747-2765.
- Chen, X. Y., Kim, S. T., Cho, W. K., Rim, Y., Kim, S., Kim, S. W., Kang, K. Y., Park, Z. Y. and Kim, J. Y. (2009). Proteomics of weakly bound cell wall proteins in rice calli. *Journal of Plant Physiology*, 166(7): 675-685.
- Chitteti, B. R., Tan, F., Mujahid, H., Magee, B.G., Bridges, S. M. and Peng, Z. (2008). Comparative analysis of proteome differential regulation during cell dedifferentiation in *Arabidopsis*. *Proteomics*, 8(20): 4303-4316.
- Chu, C. C., Wang, C. S., Sun, C. S., Hsu, V., Yin, K. C., Chu, C. Y. and Bi, F. Y. (1975). Establishment of an efficient medium for anther culture of rice through experiments on the nitrogen source. *Scientia Sinica*, 18(5): 659-668.
- Cieschi, M. T., Benedicto, A., Hernández-Apaolaza, L. and Lucena, J. J. (2016). EDTA shuttle effect vs. lignosulfonate direct effect providing Zn to Navy Bean plants (*Phaseolus vulgaris* L 'Negro Polo') in a calcareous soil. *Frontiers in plant science*, 7: 1.
- Corey, A. M., Wamsley, K. G. S., Winowiski, T. S. and Moritz, J. S. (2014). Effects of calcium lignosulfonate, mixer-added fat, and feed form on feed manufacture and broiler performance. *Journal of Applied Poultry Research*, 23(3): 418-428.
- Cui, S., Huang, F., Wang, J., Ma, X., Cheng, Y., Liu, J. (2005). A proteomic analysis of cold stress responses in rice seedlings. *Proteomics*, 5: 3162-3172.
- Dagla, H.R. (2012). Plant tissue culture. *Resonance*, 17(8):759-767.

- DalCorso, G., Farinati, S. and Furini, A. (2010). Regulatory networks of cadmium stress in plants. *Plant signaling and behavior*, 5(6): 663-667.
- Dhaliwal, G.S., Jindal, V. and Dhawan, A.K. (2010). Insect pest problems and crop losses: changing trends. *Indian Journal of Ecology*, 37(1): 1-7.
- Dharmasiri, N., Dharmasiri, S. and Estelle, M. (2005). The F-box protein TIR1 is an auxin receptor. *Nature*, 435: 441-445.
- Docquier, S., Kevers, C., Lambe, P., Gaspar, T. and Dommes, J. (2007). Beneficial use of lignosulfonates in in vitro plant cultures: stimulation of growth, of multiplication and of rooting. *Plant Cell, Tissue and Organ Culture*, 90(3): 285-291.
- Dong, M., Yang, L.L., Williams, K., Fisher, S.J., Hall, S.C., Biggin, M.D., Jin, J. and Witkowska, H.E. (2008). A “tagless” strategy for identification of stable protein complexes genome-wide by multidimensional orthogonal chromatographic separation and iTRAQ reagent tracking. *Journal of Proteome Research*, 7(5): 1836-1849.
- Duclercq, J., Sangwan-Norreel, B., Catterou, M. and Sangwan, R. S. (2011). *De novo* shoot organogenesis: from art to science. *Trends in plant science*, 16(11): 597-606.
- Durgbanshi, A., Arbona, V., Pozo, O., Miersch, O., Sancho, J. V. and Gómez-Cadenas, A. (2005). Simultaneous determination of multiple phytohormones in plant extracts by liquid chromatography-electrospray tandem mass spectrometry. *Journal of Agricultural and Food Chemistry*, 53(22): 8437-8442.
- Ertani, A., Francioso, O., Tugnoli, V., Righi, V. and Nardi, S. (2011). Effect of commercial lignosulfonate-humate on *Zea mays* L. metabolism. *Journal of agricultural and food chemistry*, 59(22): 11940-11948.
- Fan, M., Xu, C., Xu, K. and Hu, Y. (2012). LATERAL ORGAN BOUNDARIES DOMAIN transcription factors direct callus formation in Arabidopsis regeneration. *Cell research*, 22(7): 1169-1180.
- Fasahat, P., Muhammad, K., Abdullah, A. and Ratnam, W. (2012). Proximate nutritional composition and antioxidant properties of *Oryza rufipogon*, a wild rice collected from Malaysia compared to cultivated rice, MR219. *Australian Journal of Crop Science*, 6(11): 1502-1507.
- Fatehi, P. and Ni, Y. (2011). Sustainable production of fuels, chemicals, and fibers from forest biomass. *American Chemical Society Symposium Series*, pp. 409-441
- Fazeli-nasab, B., Omidi, M. and Amiritokaldani, M. (2012). Callus induction and plant regeneration of wheat mature embryos under abscisic acid treatment. *International Journal of Agriculture and Crop Sciences*, 4: 17-23.
- Fen, L. L., Ismail, M. R., Zulkarami, B., Rahman, M. S. A. and Islam, M. R. (2015). Physiological and molecular characterization of drought responses and screening of drought tolerant rice varieties. *Bioscience Journal*, 31(3).
- Food and Agriculture Organization. (2017). GIEWS - Global information and early warning system. *The United Nation*, accessed 10 November 2017, <http://www.fao.org/giews/countrybrief/country.jsp?code=MYS>
- Food and Fertilizer Technology Center. (2002). MR219, A New High-Yielding Rice Variety with Yields of More Than 10 MT/Ha. For the Asian and Pacific Region, accessed 11 December 2017, http://www.fftg.agnet.org/library.php?func=view&id=20110725142748&type_id=8
- Frank, M., Rupp, H. M., Prinsen, E., Motyka, V., Van Onckelen, H., Schmülling, T. (2000). Hormone autotrophic growth and differentiation identifies mutant lines of

- Arabidopsis with altered cytokinin and auxin content or signaling. *Plant Physiology*, 122(3): 721-729.
- Gamborg, O.L., Miller, R. and Ojima, K. (1968). Nutrient requirements of suspension cultures of soybean root cells. *Experimental cell research*, 50(1): 151-158.
- Gaspar, T., Kevers, C., Penel, C., Greppin, H., Reid, D. M. and Thorpe, T. A. (1996). Plant hormones and plant growth regulators in plant tissue culture. *In Vitro Cellular and Developmental Biology-Plant*, 32(4): 272-289.
- Ge, X., Chu, Z., Lin, Y. & Wang, S. (2006). A tissue culture system for different germplasms of indica rice. *Plant Cell Reports*, 25: 392-402.
- Gierth, M. and Mäser, P. (2007). Potassium transporters in plants involvement in K⁺ acquisition, redistribution and homeostasis. *FEBS Letters*, 581: 2348– 2356.
- Gierth, M., Mäser, P. and Schroeder, J. I. (2005). The potassium transporter AtHAK5 functions in K⁺ deprivation-induced high-affinity K⁺ uptake and AKT1 K⁺ channel contribution to K⁺ uptake kinetics in Arabidopsis roots. *Plant Physiology*, 137(3): 1105-1114.
- Goren, R., Altman, A. and Giladi, I. (1979). Role of ethylene in abscisic acid-induced callus formation in citrus bud cultures. *Plant Physiology*, 63(2): 280-2.
- Gray, W.M., Kepinski, S., Rouse, D., Leyser, O. and Estelle, M. (2001). Auxin regulates SCFTIR1-dependent degradation of AUX/IAA proteins. *Nature*, 414(6861): 271-276.
- Guilfoyle, T.J. and Hagen, G. (2007). Auxin response factors. *Current Opinion in Plant Biology*, 10(5): 453-460.
- Gumma, M. K., Nelson, A., Thenkabail, P. S. and Fingh, A.N. (2011). Mapping rice areas of South Asia using MODIS multitemporal data. *Journal of Applied Remote Sensing*, 5(1): 053547-053547.
- Guo, L., Devaiah, S.P., Narasimhan, R., Pan, X., Zhang, Y., Zhang, W. and Wang, X. (2012). Cytosolic glyceraldehyde-3-phosphate dehydrogenases interact with phospholipase D δ to transduce hydrogen peroxide signals in the Arabidopsis response to stress. *The Plant Cell*, 24(5): 2200-2212.
- Gutierrez, L., Mongelard, G., Floková, K., Păcurar, D. I., Novák, O., Staswick, P., Kowalczyk, M., Păcurar, M., Demailly, H., Geiss, G. and Bellini, C. (2012). Auxin controls Arabidopsis adventitious root initiation by regulating jasmonic acid homeostasis. *The Plant Cell*, 24(6): 2515-2527.
- Hakim, M. A., Juraimi, A. S., Begum, M., Hanafi, M. M., Ismail, M. R. and Selamat, A. (2010). Effect of salt stress on germination and early seedling growth of rice (*Oryza sativa* L.). *African Journal of Biotechnology*, 9(13): 1911-1918.
- Hausman, J.F., Kevers, C. and Gaspar, T. (1995). Auxin-polyamine interaction in the control of the rooting inductive phase of poplar shoots *in vitro*. *Plant Science*, 110(1): 63-71.
- Hayat, S., Hayat, Q., Alyemeni, M.N., Wani, A.S., Pichtel, J. and Ahmad, A. (2012). Role of proline under changing environments: a review. *Plant Signaling & Behavior*, 7(11): 1456-1466.
- He, C., Chen, X., Huang, H. and Xu, L. (2012). Reprogramming of *H3K27me3* is critical for acquisition of pluripotency from cultured Arabidopsis tissues. *PLoS Genetics*, 8(8): 1002911.
- He, C., Lin, Z., McElroy, D. and Wu, R. (2009). Identification of a rice *Actin2* gene regulatory region for high-level expression of transgenes in monocots. *Plant Biotechnology Journal*, 7(3): 227-239.
- He, Y. and Zhao, Y. (2015). A key link between jasmonic acid signaling and auxin biosynthesis. *Science China. Life sciences*, 58(3): 311.

- Hernández-Apaolaza, L., Martín-Ortiz, D. and Gárate, A. (2016). Response of wheat seedlings to Mn-lignosulfonate adhered to granular NPK. *Journal of Plant Nutrition and Soil Science*, 179(1): 113-119.
- Hiei, Y. and Komari, T. (2008). Agrobacterium-mediated transformation of rice using immature embryos or calli induced from mature seed. *Nature Protocols*, 3: 824-834.
- Htwe, N. N., Maziah, M., Ling, H. C., Zaman, F. Q. and Zain, A. M. (2011). Responses of some selected Malaysian rice genotypes to callus induction under in vitro salt stress. *African Journal of Biotechnology*, 10(3): 350-362.
- Hu, Y., Xie, Q. and Chua, N. H. (2003). The Arabidopsis auxin-inducible gene *ARGOS* controls lateral organ size. *The Plant Cell*, 15(9): 1951-1961.
- Huang, C., Ragauskas, A. J., Wu, X., Huang, Y., Zhou, X., He, J., Huang, C., Lai, C., Li, X. and Yong, Q. (2017). Co-production of bio-ethanol, xylic acid and slow-release nitrogen fertilizer from low-cost straw pulping solid residue. *Bioresource technology*, 1: 1.
- Huang, T., Harrar, Y., Lin, C., Reinhart, B., Newell, N. R., Talavera-Rauh, F., Hokin, S. A., Barton, M. K. and Kerstetter, R. A. (2014). Arabidopsis *KANADII* acts as a transcriptional repressor by interacting with a specific cis-element and regulates auxin biosynthesis, transport, and signaling in opposition to HD-ZIPIII factors. *The Plant Cell*, 26(1): 246-262.
- Hussey, P. J., Ketelaar, T. and Deeks, M. J. (2006). Control of the actin cytoskeleton in plant cell growth. *Annual Review of Plant Biology*, 57: 109-125.
- Ikeuchi, M., Sugimoto, K. and Iwase, A. (2013). Plant callus: mechanisms of induction and repression. *The Plant Cell*, 25(9): 3159-3173.
- Inukai, Y., Sakamoto, T., Ueguchi-Tanaka, M., Shibata, Y. and Gomi, K. (2005). *Crown rootless1*, which is essential for crown root formation in rice, is a target of an AUXIN RESPONSE FACTOR in auxin signaling. *Plant Cell*, 17: 1387-1396.
- Ito, J., Bath, T.S., Petzold, C. J., Redding-Johanson, A. M., Mukhopadhyay, A., Verboom, R., Meyer, E. H., Millar, A. H. and Heazlewood, J. L. (2011). Analysis of the Arabidopsis cytosolic proteome highlights subcellular partitioning of central plant metabolism. *Journal of Proteome Research*, 10(4): 1571-1582.
- Jansson, S. (1994). The light-harvesting chlorophyll a/b-binding proteins. *Biochimica et Biophysica Acta*, 1184: 1-19.
- Jansson, S. (1999). A guide to the *Lhc* genes and their relatives in *Arabidopsis*. *Trends in Plant Sciences*, 4: 236-240.
- Jiang, L., He, L. and Fountoulakis, M. (2004). Comparison of protein precipitation methods for sample preparation prior to proteomic analysis. *Journal of Chromatography A*, 1023(2): 317-320.
- Jorrín, J. V., Maldonado, A. M. and Castillejo, M. A. (2007). Plant proteome analysis: a 2006 update. *Proteomics*, 7(16): 2947-2962.
- Takei, Y., Nakamura, A., Yamamoto, M., Ishida, Y., Yamazaki, C., Sato, A., Narukawa-Nara, M., Soeno, K. and Shimada, Y. (2017). Biochemical and chemical biology study of rice *OsTAR1* revealed that tryptophan aminotransferase is involved in auxin biosynthesis: identification of a potent *OsTAR1* inhibitor, pyruvamine2031. *Plant and Cell Physiology*, 58(3): 598-606.
- Kang, S., Kang, K., Lee, K. and Back, K. (2007). Characterization of rice tryptophan decarboxylases and their direct involvement in serotonin biosynthesis in transgenic rice. *Planta*, 227(1): 263-272.
- Kepinski, S. and Leyser, O. (2004). Auxin-induced SCFTIR1–Aux/IAA interaction involves stable modification of the SCFTIR1 complex. *Proceedings of the*

- National Academy of Sciences of the United States of America*, 101(33): 12381-12386.
- Kevers, C., Soterias, G., Baccou, J.C. and Gaspar, T. (1999). Lignosulfonates: Novel promoting additives for plant tissue cultures. *In Vitro Cellular and Developmental Biology-Plant*, 35(5): 413-416.
- Khaleda, L. and Al-Forkan, M. (2006). Stimulatory effects of casein hydrolysate and proline in in vitro callus induction and plant regeneration from five deep water rice (*Oryza sativa* L.). *Biotechnology*, 5(3): 379-384.
- Khanna, H.K. and Raina, S.K. (2002). Elite indica transgenic rice plants expressing modified Cry1Ac endotoxin of *Bacillus thuringiensis* show enhanced resistance to yellow stem borer (*Scirpophaga incertulas*). *Transgenic research*, 11(4): 411-423.
- Kim, D. W., Rakwal, R., Agrawal, G. K., Jung, Y. H., Shibato, J. and Jwa, N. S. (2005). A hydroponic rice seedling culture model system for investigating proteome of salt stress in rice leaf. *Electrophoresis*, 26: 4521-4539.
- Kishor, K., Polavarapu, B., Hima Kumari, P., Sunita, M.S.L. and Sreenivasulu, N. (2015). Role of proline in cell wall synthesis and plant development and its implications in plant ontogeny. *Frontiers in Plant Science*, 6: 544.
- Komatsu, S., Kojima, K., Suzuki, K., Ozaki, K. and Higo, K. (2004). Rice Proteome Database based on two-dimensional polyacrylamide gel electrophoresis: its status in 2003. *Nucleic acids research*, 32(1): D388-D392.
- Kosová, K., Vítámvás, P., Prášil, I. T. and Renaut, J. (2011). Plant proteome changes under abiotic stress contribution of proteomics studies to understanding plant stress response. *Journal of Proteomics*, 74(8): 1301-1322.
- Krasensky, J. and Jonak, C. (2012). Drought, salt, and temperature stress-induced metabolic rearrangements and regulatory networks. *Journal of Experimental Botany*, 63(4): 1593-1608.
- Kruger, N.J. (1994). The Bradford method for protein quantitation. *Basic protein and peptide protocols*, pp: 9-15.
- Kumar, S. S. and Ajinder, K. (2013). Genotype independent tissue culture base line for high regeneration of japonica and indica rice. *Research Journal of Biotechnology*, 8: 12.
- Kunze, M., Pracharoenwattana, I., Smith, S. M. and Hartig, A. (2006). A central role for the peroxisomal membrane in glyoxylate cycle function. *Biochimica Et Biophysica Acta (BBA)-Molecular Cell Research*, 1763(12): 1441-1452.
- Lai, K.S., Yusoff, K. and Maziah, M. (2011). Extracellular matrix as the early structural marker for *Centella asiatica* embryogenic tissues. *Biologia Plantarum*, 55(3): 549-553.
- Larcher W. (2003). *Physiological Plant Ecology (4th ed.)*. Berlin, Heidelberg: Springer Verlag.
- Lazarovits, G. (2001). Management of soil-borne plant pathogens with organic soil amendments: a disease control strategy salvaged from the past. *Canadian Journal of Plant Pathology*, 23(1): 1-7.
- Lazarovits, G., Tenuta, M. and Conn, K.L. (2001). Organic amendments as a disease control strategy for soil borne diseases of high-value agricultural crops. *Australasian Plant Pathology*, 30(2): 111-117.
- Lebo, S. E., Gargulak, J. D. and McNally, T. J. (2001). Lignin, Kirk-Othmer Encyclopedia of Chemical Technology. *John Wiley & Sons, Inc. doi*, 10(1002): 0471238961.

- Lee, D. G., Ahsan, N., Lee, S. H., Kang, K. Y., Bahk, J. D., Lee, I. J. and Lee, B. H. (2007). A proteomic approach in analyzing heat-responsive proteins in rice leaves. *Proteomics*, 7(18): 3369-3383.
- Lehmann, S., Funck, D., Szabados, L. and Rentsch, D. (2010). Proline metabolism and transport in plant development. *Amino acids*, 39(4): 949-962.
- Lennicke, C., Rahn, J., Lichtenfels, R., Wessjohann, L.A. and Seliger, B. (2015). Hydrogen peroxide–production, fate and role in redox signaling of tumor cells. *Cell Communication and Signaling*, 13(1): 39.
- Lentini, Z., Reyes, P., Martínez, C. P. and Roca, W. M. (1995). Androgenesis of highly recalcitrant rice genotypes with maltose and silver nitrate. *Plant Science*, 110(1): 127-138.
- Leyser, O. (2017). Auxin signaling. *Plant Physiology*, pp. 00765.
- Li, H., Yang, Y., Duan, Y., Li, J., Cong, X., Ni, D., Song, F., Li, L., Wei, P. and Yang, J. (2013). Mapping QTLs for the tissue culture performance of rice mature embryo using indica japonica recombinant inbred lines. *Australian Journal of Crop Science*, 7: 440-445.
- Li, J., Long, Y., Qi, G. N., Xu, Z. J., Wu, W. H. and Wang, Y. (2014). The OsAKT1 channel is critical for K⁺ uptake in rice roots and is modulated by the rice CBL1-CIPK23 complex. *The Plant Cell*, 26(8): 3387-3402.
- Li, L., Hou, X., Tsuge, T., Ding, M. and Aoyama, T. (2008). The possible action mechanisms of indole-3-acetic acid methyl ester in Arabidopsis. *Plant Cell Reports*, 27: 575-584.
- Lim, Y. Y. and Lai, K. S. (2017). Generation of transgenic rice expressing cyclotide precursor oldenlandia affinis kalata B1 protein. *JAPS: Journal of Animal and Plant Sciences*, 27(2): 1.
- Lin, Y. J., Zhang, Q. (2005). Optimizing the tissue culture conditions for high efficiency transformation of indica rice. *Plant Cell Reports*, 23: 540-547.
- Linsmaier, E. M. and Skoog, F. (1965). Organic growth factor requirements of tobacco tissue cultures. *Plant Physiology*, 18: 100-127.
- Liu, H., Wang, S., Yu, X., Yu, J., He, X., Zhang, S., Shou, H. and Wu, P. (2005). ARL1, a LOB-domain protein required for adventitious root formation in rice. *The Plant Journal*, 43(1): 47-56.
- Liu, J., Sheng, L., Xu, Y., Li, J., Yang, Z., Huang, H. and Xu, L. (2014). *WOX11* and *12* are involved in the first-step cell fate transition during *de novo* root organogenesis in Arabidopsis. *The Plant Cell*, 26(3): 1081-1093.
- Livak, K.J. and Schmittgen, T.D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2⁻ΔΔCT method. *Methods*, 25(4): 402-408.
- López-Meyer, M. and Nessler, C.L. (1997). Tryptophan decarboxylase is encoded by two autonomously regulated genes in *Camptotheca acuminata* which are differentially expressed during development and stress. *The Plant Journal*, 11(6): 1167-1175.
- Lu, T. C., Meng, L. B., Yang, C. P., Liu, G. F., Liu, G. J., Ma, W. and Wang, B. C. (2008). A shotgun phosphoproteomics analysis of embryos in germinated maize seeds. *Planta*, 228(6): 1029-1041.
- Mahmood, U. H., Ahmed, Z., Munir, M., Malik, S.I., Shahzad, K. (2009) Effect of sorbitol in callus induction and plant regeneration in wheat. *African Journal of Biotechnology*, 8: 6529-6535.
- Malvi, U. R. (2011). Interaction of micronutrients with major nutrients with special reference to potassium. *Karnataka Journal of Agricultural Sciences*, 24(1).

- Mano, Y. and Nemoto, K. (2012). The pathway of auxin biosynthesis in plants. *Journal of experimental Botany*, 63(8): 2853-2872.
- Mano, Y., Nemoto, K., Suzuki, M., Seki, H., Fujii, I. and Muranaka, T. (2009). The *AMI1* gene family: indole-3-acetamide hydrolase functions in auxin biosynthesis in plants. *Journal of Experimental Botany*, 61(1): 25-32.
- Martinez-Trujillo, M., Cabrera-Ponce, J.L. and Herrera-Estrella, L. (2003). Improvement of rice transformation using bombardment of scutellum-derived calli. *Plant Molecular Biology Reporter*, 21(4): 429-437.
- Mashiguchi, K., Tanaka, K., Sakai, T., Sugawara, S., Kawaide, H., Natsume, M., Hanada, A., Yaeno, T., Shirasu, K., Yao, H. and McSteen, P. (2011). The main auxin biosynthesis pathway in Arabidopsis. *Proceedings of the National Academy of Sciences*, 108(45): 18512-18517.
- Mattioli, R., Costantino, P. and Trovato, M. (2009). Proline accumulation in plants: not only stress. *Plant Signaling & Behavior*, 4(11): 1016-1018.
- McCann, H.C., Rikkerink, E.H., Bertels, F., Fiers, M., Lu, A., Rees-George, J., Andersen, M.T., Gleave, A.P., Haubold, B., Wohlers, M.W. and Guttman, D.S. (2013). Genomic analysis of the kiwifruit pathogen *Pseudomonas syringae* pv. actinidiae provides insight into the origins of an emergent plant disease. *PLoS Pathogens*, 9(7):1003503.
- Mikkelsen, M. D., Hansen, C. H., Wittstock, U. and Halkier, B. A. (2000). Cytochrome P450 CYP79B2 from Arabidopsis catalyzes the conversion of tryptophan to indole-3-acetaldoxime, a precursor of indole glucosinolates and indole-3-acetic acid. *Journal of Biological Chemistry*, 275(43): 33712-33717.
- Minnocci, A., Francini, A., Romeo, S., Sgrignuoli, A. D., Povero, G. and Sebastiani, L. (2018). Zn-localization and anatomical changes in leaf tissues of green beans (*Phaseolus vulgaris* L.) following foliar application of Zn-lignosulfonate and ZnEDTA. *Scientia Horticulturae*, 231: 15-21.
- Mishra, R. and Rao, G. J. N. (2016). In-vitro Androgenesis in Rice: Advantages, Constraints and Future Prospects. *Rice Science*, 23(2): 57-68.
- Mohanty, A., Kathuria, H., Ferjani, A., Sakamoto, A., Mohanty, P., Murata, N. and Tyagi, A. (2002). Transgenics of an elite indica rice variety Pusa Basmati 1 harbouring the codA gene are highly tolerant to salt stress. *Theoretical and Applied Genetics*, 106(1): 51-57.
- Morgan, J. B. and Connolly, E. L. (2013). Plant-Soil Interactions: Nutrient Uptake. *Nature Education Knowledge*, 4(8): 2.
- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bioassays with tobacco tissue culture. *Plant Physiology*, 15: 473-497.
- Naik, N., Rout, P., Umakanta, N., Verma, R. L., Katara, J. L., Sahoo, K. K., Singh, O. N. and Samantaray, S. (2017). Development of doubled haploids from an elite indica rice hybrid (BS6444G) using anther culture. *Plant Cell, Tissue and Organ Culture*, 128(3): 679-689.
- Nakamura, A., Umemura, I., Gomi, K., Hasegawa, Y., Kitano, H., Sazuka, T., Matsuoka, M. (2006). Production and characterization of auxin-insensitive rice by overexpression of a mutagenized rice IAA protein. *The Plant Journal*, 46: 297-306.
- Neu, D., Lehmann, T., Elleuche, S. and Pollmann, S. (2007). Arabidopsis amidase 1, a member of the amidase signature family. *The FEBS Journal*, 274(13): 3440-3451.
- Nishimura, T., Hayashi, K.I., Suzuki, H., Gyohda, A., Takaoka, C., Sakaguchi, Y., Matsumoto, S., Kasahara, H., Sakai, T., Kato, J.I. and Kamiya, Y. (2014). Yucasin is a potent inhibitor of YUCCA, a key enzyme in auxin biosynthesis. *The Plant Journal*, 77(3): 352-366.

- Northey, R. A. (2002). The use of lignosulfonates as water reducing agents in the manufacture of gypsum wallboard. *Chemical modification, and usage of lignin*, pp.139-50.
- Nunes-Nesi, A., Fernie, A.R. and Stitt, M. (2010). Metabolic and signaling aspects underpinning the regulation of plant carbon nitrogen interactions. *Molecular plant*, 3(6): 973-996.
- Okazaki, Y. and Saito, K. (2012). Recent advances of metabolomics in plant biotechnology. *Plant biotechnology reports*, 6(1): 1-15.
- Olatunji, D., Geelen, D. and Verstraeten, I. (2017). Control of Endogenous Auxin Levels in Plant Root Development. *International Journal of Molecular Sciences*, 18(12): 2587.
- Pacek, A. W., Ding, P., Garrett, M., Sheldrake, G. and Nienow, A. W. (2013). Catalytic conversion of sodium lignosulfonate to vanillin: engineering aspects. Part 1. Effects of processing conditions on vanillin yield and selectivity. *Industrial and Engineering Chemistry Research*, 52(25): 8361-8372.
- Pacheco-Villalobos, D., Díaz-Moreno, S. M., van der Schuren, A., Tamaki, T., Kang, Y. H., Gujas, B., Novak, O., Jaspert, N., Li, Z., Wolf, S. and Oecking, C. (2016). The effects of high steady state auxin levels on root cell elongation in *Brachypodium*. *The Plant Cell Online*, 28(5): 1009-1024.
- Park, H.Y., Kim, S.A., Korlach, J., Rhoades, E., Kwok, L. W., Zipfel, W. R., Waxham, M. N., Webb, W. W. and Pollack, L. (2008). Conformational changes of calmodulin upon Ca²⁺ binding studied with a microfluidic mixer. *Proceedings of the National Academy of Sciences*, 105(2): 542-547.
- Park, W. J., Kriechbaumer, V., Müller, A., Piotrowski, M., Meeley, R. B., Gierl, A. and Glawischnig, E. (2003). The nitrilase ZmNIT2 converts indole-3-acetonitrile to indole-3-acetic acid. *Plant physiology*, 133(2): 794-802.
- Pazuki, A., Asghari, J., Sohani, M.M., Pessaraki, M. and Aflaki, F. (2015). Effects of Some Organic Nitrogen Sources and Antibiotics on Callus Growth of Indica Rice Cultivars. *Journal of Plant Nutrition*, 38(8): 1231-1240.
- Philippar, K., Fuchs, I., Lüthen, H., Hoth, S., Bauer, C. S., Haga, K., Thiel, G., Ljung, K., Sandberg, G., Böttger, M. and Becker, D. (1999). Auxin-induced K⁺ channel expression represents an essential step in coleoptile growth and gravitropism. *Proceedings of the National Academy of Sciences*, 96(21): 12186-12191.
- Pontaroli, A.C. and Camadro, E.L. (2005). Somaclonal variation in *Asparagus officinalis* plants regenerated by organogenesis from long-term callus cultures. *Genetics and Molecular Biology*, 28(3), pp.423-430.
- Posch, A., Franz, T., Hartwig, S., Knebel, B., Al-Hasani, H., Passlack, W., Kunz, N., Hinze, Y., Li, X., Kotzka, J. and Lehr, S. (2013). 2D-ToGo workflow: increasing feasibility and reproducibility of 2-dimensional gel electrophoresis. *Archives of Physiology and Biochemistry*, 119(3): 108-113.
- Rafique, M. Z., Rashid, H., Chaudhary, M. F., Chaudhry, Z. and Cheema, N. M. (2011). Study on callogenesis and organogenesis in local cultivars of rice (*Oryza sativa* L.). *Pakistan Journal of Botany*, 43(1): 191-203.
- Raghavendra, G., Kumaraswamy, G. K., Ramya, B., Sandesh, H. S., Yogendra, K. N., Deepak, N. and Gowda, P. H. R. (2010). Direct Multiple Shoot Regeneration of indica rice (*Oryza sativa*) Var. 'Rasi'. *Asian and Australasian Journal of Plant Science and Biotechnology*, 4(1): 71-73.
- Raina, S. K., Balachandran, S. M., Virmani, S. S. and Zapata, F. J. (1989). Improved medium for efficient anther culture of some indica rice hybrids. *International Rice Research Newsletter (Philippines)*, 14: 3-4.

- Rajamoorthy, Y. and Munusamy, S. (2015). Rice industry in Malaysia: challenges, policies and implications. *Procedia Economics and Finance*, 31: 861-867.
- Rakwal, R. and Agrawal, G. K. (2003). Rice proteomics: current status and future perspectives. *Electrophoresis*, 24(19): 3378-3389.
- Rashid, H., Yokoi, S., Torijama, K., Hinata, K. (1996) Transgenic plant production mediated by *Agrobacterium* in indica rice. *Plant Cell Reports*, 15: 727-730.
- Reid, R. and Hayes, J. (2003). Mechanisms and control of nutrient uptake in plants. *International review of cytology*, 229: 73-114.
- Rodríguez López, C. M., Wetten, A. C. and Wilkinson, M. J. (2010). Progressive erosion of genetic and epigenetic variation in callus-derived cocoa (*Theobroma cacao*) plants. *New Phytologist*, 186(4): 856-868.
- Rodríguez-Lucena, P., Tomasi, N., Pinton, R., Hernández-Apaolaza, L., Lucena, J. J. and Cesco, S. (2009). Evaluation of 59Fe-lignosulfonates complexes as Fe-resources for plants. *Plant and soil*, 325(1-2): 53.
- Rodríguez-Navarro, A. and Rubio, F. (2006). High-affinity potassium and sodium transport systems in plants. *Journal of Experimental Botany*, 57(5): 1149-1160.
- Sah, S. K., Kaur, A., Kaur, G., Cheema, G. S. (2014). Genetic Transformation of Rice: Problems, Progress and Prospects. *Rice Research: Open Access*, 3(1): 132.
- Sahoo, K.K., Tripathi, A.K., Pareek, A., Sopory, S.K. and Singla-Pareek, S.L. (2011). An improved protocol for efficient transformation and regeneration of diverse indica rice cultivars. *Plant Methods*, 7(1): 49.
- Schaller, G. E., Bishopp, A. and Kieber, J. J. (2015). The yin-yang of hormones: cytokinin and auxin interactions in plant development. *The Plant Cell*, 27(1): 44-63.
- Sekimoto, H., Seo, M., Dohmae, N., Takio, K., Kamiya, Y. and Koshiba, T. (1997). Cloning and molecular characterization of plant aldehyde oxidase. *Journal of Biological Chemistry*, 272(24): 15280-15285.
- Sekimoto, H., Seo, M., Kawakami, N., Komano, T., Desloire, S., Liotenberg, S., Marion-Poll, A., Caboche, M., Kamiya, Y. and Koshiba, T. (1998). Molecular cloning and characterization of aldehyde oxidases in *Arabidopsis thaliana*. *Plant and Cell Physiology*, 39(4): 433-442.
- Sharma, C., Kaur, M., Kaur, A. and Gosal, S. S. (2012). In vitro plant regeneration studies in three indica rice varieties. *International Journal of Agriculture and Environmental Biotechnology*, 5(4): 309-313.
- Silva, T.D. (2010). Indica rice anther culture: can the impasse be surpassed?. *Plant Cell, Tissue and Organ Culture*, 100(1): 1-11.
- Shen, S., Sharma, A. and Komatsu, S. (2003). Characterization of proteins responsive to gibberellin in the leaf-sheath of rice (*Oryza sativa* L.) seedling using proteome analysis. *Biological and Pharmaceutical Bulletin*, 26(2): 129-136.
- Skoog, F. and Miller, C. O. (1957). Chemical regulation of growth and organ formation in plant tissues cultured in vitro. *Symposia of the Society for Experimental Biology*, 11:118-30.
- Soltani, N., Conn, K.L., Abbasi, P.A. and Lazarovits, G. (2002). Reduction of potato scab and verticillium wilt with ammonium lignosulfonate soil amendment in four Ontario potato fields. *Canadian Journal of Plant Pathology*, 24(3): 332-339.
- Soteras, G. (1994). *Mode d'action des lignosulfonates de fer chez les végétaux. Isolement de la molécule active* (Doctoral dissertation). University de Montpellier II, pp.176.
- Spartz, A. K., Lor, V. S., Ren, H., Olszewski, N. E., Miller, N. D., Wu, G., Spalding, E. P. and Gray, W. M. (2016). Constitutive expression of the auxin-related

- AtSAUR19 protein confers auxin-independent hypocotyl elongation. *Plant Physiology*, pp. 01514.
- Spollen, W.G., Tao, W., Valliyodan, B., Chen, K., Hejlek, L.G., Kim, J.J., LeNoble, M.E., Zhu, J., Bohnert, H.J., Henderson, D. and Schachtman, D.P. (2008). Spatial distribution of transcript changes in the maize primary root elongation zone at low water potential. *BMC Plant Biology*, 8(1): 32.
- Steffens, B. and Rasmussen, A. (2016). The physiology of adventitious roots. *Plant Physiology*, 170(2): 603-617.
- Stepanova, A. N., Robertson-Hoyt, J., Yun, J., Benavente, L. M., Xie, D. Y., Doležal, K., Schlereth, A., Jürgens, G. and Alonso, J. M. (2008). TAA1-mediated auxin biosynthesis is essential for hormone crosstalk and plant development. *Cell*, 133(1): 177-191.
- Su, N., Hu, M. L., Wu, D. X., Wu, F. Q., Fei, G. L., Lan, Y., Chen, X. L., Shu, X. L., Zhang, X., Guo, X. P. and Cheng, Z. J. (2012). Disruption of a rice pentatricopeptide repeat protein causes a seedling-specific albino phenotype and its utilization to enhance seed purity in hybrid rice production. *Plant physiology*, 159(1): 227-238.
- Sugimoto, K., Jiao, Y. and Meyerowitz, E.M. (2010). Arabidopsis regeneration from multiple tissues occurs via a root development pathway. *Developmental cell*, 18(3): 463-471.
- Sunkar, R., Kapoor, A. and Zhu, J.K. (2006). Posttranscriptional induction of two Cu/Zn superoxide dismutase genes in Arabidopsis is mediated by downregulation of miR398 and important for oxidative stress tolerance. *The Plant Cell*, 18(8): 2051-2065.
- Tan, L. W., Rahman, Z. A., Goh, H. H., Hwang, D. J., Ismail, I. and Zainal, Z. (2017). Production of transgenic rice (indica cv. MR219) overexpressing *Abp57* gene through agrobacterium-mediated transformation. *Sains Malays*, 46(5).
- Tan, X., Calderon-Villalobos, L. I. A., Sharon, M., Zheng, C., Robinson, C. V., Estelle, M. and Zheng, N. (2007). Mechanism of auxin perception by the TIR1 ubiquitin ligase. *Nature*, 446(7136): 640-645.
- Tao, Y., Ferrer, J. L., Ljung, K., Pojer, F., Hong, F., Long, J. A., Li, L., Moreno, J. E., Bowman, M. E., Ivans, L. J. and Cheng, Y. (2008). Rapid synthesis of auxin via a new tryptophan-dependent pathway is required for shade avoidance in plants. *Cell*, 133(1): 164-176.
- Teale, W. D., Ditengou, F. A., Dovzhenko, A. D., Li, X., Molendijk, A. M., Ruperti, B., Paponov, I. and Palme, K. (2008). Auxin as a model for the integration of hormonal signal processing and transduction. *Molecular Plant*, 1(2): 229-237.
- Teale, W. D., Paponov, I. A. and Palme, K. (2006). Auxin in action: signalling, transport and the control of plant growth and development. *Nature Reviews Molecular Cell Biology*, 7(11): 847-859.
- Telysheva, G., Lebedeva, G., Dizhbite, T., Zaimenko, N., Grivinya, D. and Virzina, O. (1997). Novel ligno-silicon products promoting root system development. *In Biology of Root Formation and Development*, pp. 92-93.
- Telysheva, G., Lebedeva, G., Zaimenko, N. and Viesturs, U. (1992). New lignosilicon fertilizers and their action on soil biota. *In International symposium, Soil Decontamination Using Biological Processes. Karlsruhe, Deutschland*, pp. 525-530.
- Thoday-Kennedy, E. L., Jacobs, A. K. and Roy, S. J. (2015). The role of the CBL–CIPK calcium signalling network in regulating ion transport in response to abiotic stress. *Plant Growth Regulation*, 76(1): 3-12.

- Titouh, K., Boufis, N. and Khelifi, L. (2017). Microcalli Induction in Protoplasts Isolated from Embryogenic Callus of Date Palm. *In Date Palm Biotechnology Protocols*, 1: 227-237.
- Trovato, M., Maras, B., Linhares, F. and Costantino, P. (2001). The plant oncogene rolD encodes a functional ornithine cyclodeaminase. *Proceedings of the National Academy of Sciences*, 98(23): 13449-13453.
- Tuteja, N. and Mahajan, S. (2007). Calcium signaling network in plants: an overview. *Plant signaling & behavior*, 2(2): 79-85.
- Ueno, M., Shibata, H., Kihara, J., Honda, Y. and Arase, S. (2003). Increased tryptophan decarboxylase and monoamine oxidase activities induce Sekiguchi lesion formation in rice infected with *Magnaporthe grisea*. *The Plant Journal*, 36(2): 215-228.
- Urban, M.C. (2015). Accelerating extinction risk from climate change. *Science*, 348(6234): 571-573.
- Van der Krieken, W., Kok, C. and Stevens, L. (2004). Compositions comprising lignosulfonates for crop protection and crop improvement. U.S. Patent Application 10/543,702.
- Vanderschuren, H., Lentz, E., Zainuddin, I. and Gruissem, W. (2013). Proteomics of model and crop plant species: status, current limitations and strategic advances for crop improvement. *Journal of Proteomics*, 93: 5-19.
- Vanneste, S. and Friml, J. (2009). Auxin: a trigger for change in plant development. *Cell*, 136(6): 1005-1016.
- Vaughan, D.A., Morishima, H. and Kadowaki, K. (2003). Diversity in the *Oryza* genus. *Current Opinion in Plant Biology*, 6(2): 139-146.
- Vaughan, D.A., Lu, B.R. and Tomooka, N. (2008). The evolving story of rice evolution. *Plant science*, 174(4): 394-408.
- Véry, A.A. and Sentenac, H. (2003). Molecular mechanisms and regulation of K⁺ transport in higher plants. *Annual Review of Plant Biology*, 54(1): 575-603.
- Visarada, K. B. R. S., Sailaja, M., Sarma, N. P. (2002). Effect of callus induction media on morphology of embryogenic calli in rice genotypes. *Biologia Plantarum*, 45: 495-502.
- Visarada, K. B. R. S. and Sarma, N. P. (2004). Transformation of indica rice through particle-bombardment: factors influencing transient expression and selection. *Biologia Plantarum*, 48: 25-31.
- Vogeser, M. and Parhofer, K. G. (2007). Liquid chromatography tandem-mass spectrometry (LC-MS/MS)-technique and applications in endocrinology. *Experimental and clinical endocrinology & diabetes*, 115(09): 559-570.
- Walck, J.L., Hidayati, S.N., Dixon, K.W., Thompson, K.E.N. and Poschlod, P. (2011). Climate change and plant regeneration from seed. *Global Change Biology*, 17(6): 2145-2161.
- Wang, F., Bai, M. Y., Deng, Z., Osés-Prieto, J. A., Burlingame, A. L., Lu, T., Chong, K. and Wang, Z. Y. (2010). Proteomic study identifies proteins involved in brassinosteroid regulation of rice growth. *Journal of integrative plant biology*, 52(12): 1075-1085.
- Wang, M., Shen, Q., Xu, G. and Guo, S. (2014). New insight into the strategy for nitrogen metabolism in plant cells. *International Review of Cell and Molecular Biology*, 310: 1-37.

- Wang, P., Xu, D.Y., Wang, G. and Ji, J. (2007). Comparison study on tissue culture and regeneration ability of mature embryo in indica-japonica rice two subspecies. *Seed*, 26(10): 66-67.
- Wang, X. D., Nolan, K. E., Irwanto, R. R., Sheahan, M. B. and Rose, R. J. (2011). Ontogeny of embryogenic callus in *Medicago truncatula*: the fate of the pluripotent and totipotent stem cells. *Annals of Botany*, 107(4): 599-609.
- Wang, Y. and Wu, W.H. (2013). Potassium transport and signaling in higher plants. *Annual Review of Plant Biology*, 64: 451-476.
- Wang, Y. Q., Liang, C. Y. (2006). Efficient plant regeneration from in vitro culture of Induca rice Zhong A (*Oryza sativa* L.). *Acta Scientiarum Naturalium Universitatis Sunyaatseni*, 45: 81-84.
- Wani, S. H., Sanghera, G. S. and Gosal, S. S. (2011). An efficient and reproducible method for regeneration of whole plants from mature seeds of a high yielding indica rice (*Oryza sativa* L.) Variety PAU 201. *New Biotechnology*, 28: 418-422.
- Wawer, I., Bucholc, M., Astier, J., Anielska-Mazur, A., Dahan, J., Kulik, A., Wystouch-Cieszynska, A., Zaręba-Kozioł, M., Krzywinska, E., Dadlez, M. and Dobrowolska, G. (2010). Regulation of *Nicotiana tabacum* osmotic stress-activated protein kinase and its cellular partner GAPDH by nitric oxide in response to salinity. *Biochemical Journal*, 429(1): 73-83.
- Wei, S., Hu, W., Deng, X., Zhang, Y., Liu, X., Zhao, X., Luo, Q., Jin, Z., Li, Y., Zhou, S. and Sun, T. (2014). A rice calcium-dependent protein kinase *OsCPK9* positively regulates drought stress tolerance and spikelet fertility. *BMC Plant Biology*, 14(1): 133.
- Wirtz, M. and Hell, R. (2006). Functional analysis of the cysteine synthase protein complex from plants: structural, biochemical and regulatory properties. *Journal of Plant Physiology*, 163(3): 273-286.
- Won, C., Shen, X., Mashiguchi, K., Zheng, Z., Dai, X., Cheng, Y., Kasahara, H., Kamiya, Y., Chory, J. and Zhao, Y. (2011). Conversion of tryptophan to indole-3-acetic acid by TRYPTOPHAN AMINOTRANSFERASES OF ARABIDOPSIS and YUCCAs in Arabidopsis. *Proceedings of the National Academy of Sciences*, 108(45): 18518-18523.
- Wong, C. K. F., Lai, K. S., Yun, W. M. and Mahmood, M. (2015). Efficient regeneration and Agrobacterium-mediated transformation protocol for recalcitrant indica rice (*Oryza sativa* L.). *Emirates Journal of Food and Agriculture*, 27(11): 837.
- Woodward, A. W. and Bartel, B. (2005). Auxin: regulation, action, and interaction. *Annals Botany*, 95: 707-735.
- Xu, H., Zhang, W., Gao, Y., Zhao, Y., Guo, L. and Wang, J. (2012). Proteomic analysis of embryo development in rice (*Oryza sativa*). *Planta*, 235(4): 687-701.
- Xu, J., Li, H. D., Chen, L. Q., Wang, Y., Liu, L. L., He, L. and Wu, W. H. (2006). A protein kinase, interacting with two calcineurin B-like proteins, regulates K⁺ transporter AKT1 in Arabidopsis. *Cell*, 125(7): 1347-1360.
- Yamada, M., Greenham, K., Prigge, M.J., Jensen, P.J. and Estelle, M. (2009). The *TRANSPORT INHIBITOR RESPONSE2* gene is required for auxin synthesis and diverse aspects of plant development. *Plant Physiology*, 151(1): 168-179.
- Yamamoto, Y., Kamiya, N., Morinaka, Y., Matsuoka, M. and Sazuka, T. (2007). Auxin biosynthesis by the YUCCA genes in rice. *Plant Physiology*, 143(3): 362-371.
- Yamashita, T.T. and Thomas T. (1996). *Method and composition for promoting and controlling growth of plants*. U.S. Patent 5,549,729.

- Yan, L. N., Li, X., Cai, Q. S. (2009). Research in the induction and regeneration of mature embryo-derived calli from rice Wugeng 3. *Jiangsu Agriculture Sciences*, 6: 67-69.
- Yan, L. N., Xia, L. I. and Dan, W. U. (2010). The comparison in tissue culture ability of mature embryo in different cultivars of rice. *Agricultural Sciences in China*, 9(6): 840-846.
- Yang, D., Qiu, X., Zhou, M. and Lou, H. (2007). Properties of sodium lignosulfonate as dispersant of coal water slurry. *Energy Conversion and Management*, 48(9): 2433-2438.
- Yang, T. and Poovaiah, B.W. (2003). Calcium/calmodulin-mediated signal network in plants. *Trends in plant science*, 8(10): 505-512.
- Yang, X., Nian, J., Xie, Q., Feng, J., Zhang, F., Jing, H., Zhang, J., Dong, G., Liang, Y., Peng, J. and Wang, G. (2016). Rice Ferredoxin-Dependent glutamate synthase regulates nitrogen-carbon metabolomes and its genetically differentiated between japonica and indica subspecies. *Molecular Plant*, 9(11): 1520-1534.
- Yang, Y., Xu, R., Ma, C. J., Vlot, A. C., Klessig, D. F. and Pichersky, E. (2008). Inactive methyl indole-3-acetic acid ester can be hydrolyzed and activated by several esterases belonging to the AtMES esterase family of Arabidopsis. *Plant Physiology*, 147: 1034-1045.
- Yang, Z., Gao, Q., Sun, C., Li, W., Gu, S. and Xu, C. (2009). Molecular evolution and functional divergence of HAK potassium transporter gene family in rice (*Oryza sativa* L.). *Journal of Genetics and Genomics*, 36: 161-172.
- Yap, W. S. and Lai, K. S. (2017). Biochemical Properties of Twelve Malaysia Rice Cultivars in Relation to Yield Potential. *Asian Journal of Agricultural Research*, ISSN: 1819-1894.
- Yin, L., Tao, Y., Zhao, K., Shao, J., Li, X., Liu, G., Liu, S. and Zhu, L. (2007a). Proteomic and transcriptomic analysis of rice mature seed-derived callus differentiation. *Proteomics*, 7(5): 755-768.
- Yin, X. M., Xu, Q. G. and Li, H. L. (2007b). Establishment of plant regeneration system of rice callus from mature embryos of Xian-type hybrid rice parents. *Progress in Modern Biomedicine*, 7: 347-350.
- Yu, J., Liu, W., Liu, J., Qin, P. and Xu, L. (2017). Auxin Control of Root Organogenesis from Callus in Tissue Culture. *Frontiers in Plant Science*, 8: 1385.
- Zeeman, S. C., Kossmann, J. and Smith, A. M. (2010). Starch: its metabolism, evolution, and biotechnological modification in plants. *Annual Review of Plant Biology*, 61: 209-234.
- Zhang, X. H., Rao, X. L., Shi, H. T., Li, R. J. and Lu, Y. T. (2011). Overexpression of a cytosolic glyceraldehyde-3-phosphate dehydrogenase gene OsGAPC3 confers salt tolerance in rice. *Plant Cell, Tissue and Organ Culture*, 107(1): 1.
- Zhang, Z., Zhang, J., Chen, Y., Li, R., Wang, H. and Wei, J. (2012). Genome-wide analysis and identification of HAK potassium transporter gene family in maize (*Zea mays* L.). *Molecular Biology Reports*, 39: 8465-8473.
- Zhang, Z., Zhao, H., Tang, J., Li, Z., Li, Z., Chen, D. and Lin, W. (2014). A proteomic study on molecular mechanism of poor grain-filling of rice (*Oryza sativa* L.) inferior spikelets. *PloS one*, 9(2): 89140.
- Zhao, Q., Zhang, H., Wang, T., Chen, S. and Dai, S. (2013). Proteomics-based investigation of salt-responsive mechanisms in plant roots. *Journal of Proteomics*, 82: 230-253.
- Zhao, Y. (2010). Auxin biosynthesis and its role in plant development. *Annual review of plant biology*, 61: 49-64.

- Zhao, Y., Christensen, S.K., Fankhauser, C., Cashman, J. R., Cohen, J. D., Weigel, D. and Chory, J. (2001). A role for flavin monooxygenase-like enzymes in auxin biosynthesis. *Science*, 291(5502): 306-309.
- Zhao, Y., Hu, Y., Dai, M., Huang, L. and Zhou, D.X. (2009). The *WUSCHEL*-related *homeobox* gene *WOX11* is required to activate shoot-borne crown root development in rice. *The Plant Cell Online*, 21(3): 736-748.
- Zunica-Soto, E., Mullins, E. and Dedicova, B. (2015). Ensifer-mediated transformation: an efficient non-Agrobacterium protocol for the genetic modification of rice. *SpringerPlus*, 4(1): 600.
- Zuraida, A. R., Zulkifli, A. S., Habibuddin, H. and Naziah, B. (2012). Regeneration of Malaysian Rice variety MR219 via somatic embryogenesis. *Journal of Tropical Agriculture and Food Science*, 39(2): 167-177.

