

UNIVERSITI PUTRA MALAYSIA

AUTOCLAVE-CHEMICAL HYDROLYSIS OF CHICKEN FEATHER FOR PROTEIN HYDROLYSATE PRODUCTION

CHEONG CHOOI WEI

FBSB 2018 24



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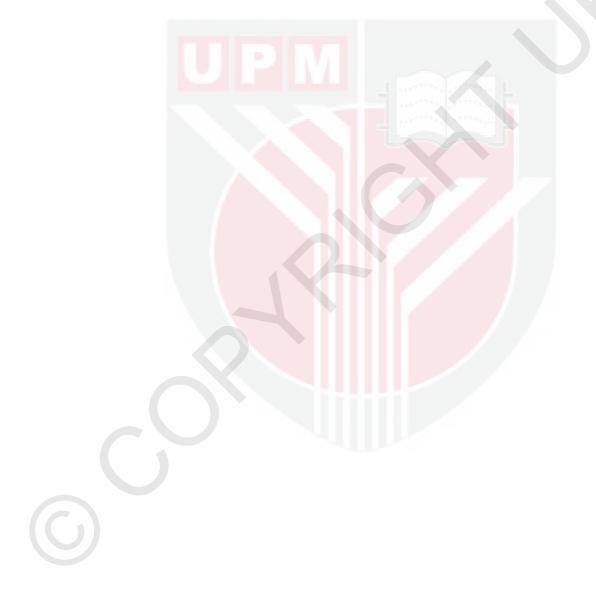
Thesis Submitted to School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

April 2018

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

AUTOCLAVE-CHEMICAL HYDROLYSIS OF CHICKEN FEATHER FOR PROTEIN HYDROLYSATE PRODUCTION

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CHEONG CHOOI WEI

April 2018

Chairman: Phang Lai Yee, PhD Faculty: Biotechnology and Biomolecular Sciences

Millions tonnes of feather waste are generated every year. Various treatments on feather waste have been developed in order to add value to them. However, these methods have their pros and cons. Autoclave-chemical treatment could hydrolyse feather within a short period but harsh conditions could lead to excessive destruction on protein and amino acids. Biological treatment is an eco-friendly treatment which could hydrolyse feather with minimum protein and amino acid destruction but it required a long reaction time. Therefore, treatment modification is necessary in order to enhance the feather hydrolysis and produce feather hydrolysate consists high amount of protein and amino acids. The objectives of this study were to investigate the effect of autoclave-chemical treatment on chicken feather hydrolysis and to examine the effect of autoclave-alkaline as pretreatment on enzymatic hydrolysis of chicken feather. Sodium hydroxide (NaOH) was selected based on the screening result and the temperature was fixed at 105°C. The NaOH concentration and autoclave holding time ranged from 0.01 M to 0.10 M and 1 min to 10 min, respectively were used in the subsequent experiments. The effect of autoclave-alkaline treatment (AAT) on feather was investigated using response surface methodology (RSM). NaOH concentration and holding time were the significant parameters in AAT that affect the feather hydrolysis and protein production (p < 0.05). The ideal conditions for AAT were 0.08 M of NaOH, 7 min holding time at 105°C in which 85.59% of feather were solubilized and 0.75 g/g of soluble protein with 633.50 mg/g of free amino acid could be recovered from chicken feather. Autoclave-alkaline method was also proposed to be used as pretreatment in this study in order to enhance the performance of the subsequent biological treatment. The effect of autoclave-alkaline as pretreatment on the enzymatic hydrolysis of chicken feather with Savinase[®] Ultra 16L by RSM was investigated. NaOH concentration was significant parameter that affected the subsequent enzymatic reaction on feather hydrolysis and protein production (p < 0.05). The pretreated chicken feather with optimized autoclave-alkaline (0.07 M of NaOH, 2 min holding time at 105°C) could improve Savinase hydrolysis up to 14 times with 80.81% of feather hydrolysis and recovered 0.69 g/g of soluble protein as well as 673.80 mg/g of free amino acid. Therefore, AAT is a potential method for feathers solubilization, as well as, recovery of soluble protein and free amino acid from chicken feather waste. Autoclave-alkaline pretreatment also could enhance enzyme reaction to degrade chicken feather to a great extent and generate feather hydrolysates rich in amino acids.



Abstrak tesis yang dikemukakan kepada Senat of Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

HIDROLISIS AUTOKLAF-KIMIA BULU AYAM UNTUK PENGHASILAN PROTEIN HIDROLISAT

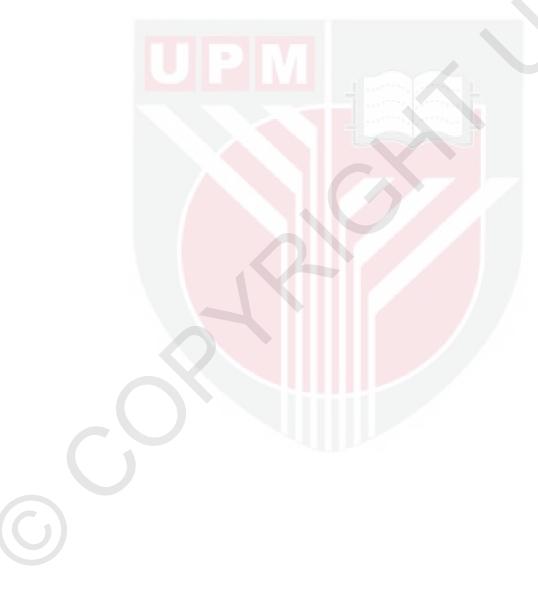
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April 2018

Pengerusi: Phang Lai Yee, PhD Fakulti: Bioteknologi dan Sains Biomolekul

Berjuta-juta tan bulu ayam telah dihasilkan setiap tahun sebagai sisa buangan. Pelbagai rawatan terhadap sisa bulu telah ditemui untuk menambah nilai sisa bulu tersebut. Walau bagaimanapun, rawatan-rawatan tersebut mempunyai kebaikan dan keburukan. Rawatan kimia-autoklaf mampu menghidrolisis bulu ayam dalam tempoh yang singkat tetapi keadaan rawatan yang tidak bersesuaian boleh mengakibatkan pemusnahan berlebihan terhadap protein dan asid amino. Rawatan biologi adalah rawatan mesra alam yang boleh menghidrolisis bulu dengan mengurangkan pemusnahan protein dan asid amino tetapi memerlukan masa reaksi yang panjang. Oleh itu, pengubahsuaian rawatan diperlukan untuk mempertingkatkan hidrolisis bulu dan menghasilkan hidrolisat bulu yang mengandungi jumlah protein dan asid amino yang tinggi. Objektif kajian ini adalah untuk mengkaji kesan rawatan kimia-autoklaf terhadap hidrolisis bulu ayam dan juga untuk mengkaji kesan pra-rawatan autoklaf-alkali terhadap hidrolisis enzimatik bulu ayam. Berdasarkan keputusan penyaringan, natrium hidroksida (NaOH) telah dipilih dan suhu telah ditetapkan pada 105°C. Kepekatan NaOH dan masa pegangan dalam autoklaf masing-masing adalah antara 0.01 M hingga 0.10 M dan 1 minit hingga 10 minit. Kesan rawatan autoklaf-alkali (AAT) terhadap bulu telah dikajikan dengan menggunakan kaedah gerak balas permukaan (RSM). Kepekatan NaOH dan masa pegangan adalah parameter penting dalam AAT yang mempengaruhi hidrolisis bulu dan penghasilan protein (p <0.05). Keadaan optima untuk AAT adalah 0.08 M NaOH, dengan masa pegangan selama 7 minit dan suhu 105°C, di mana 85.59% bulu telah dilarutkan dan 0.75 g/g hasil protein dengan 633.50 mg/g asid amino bebas boleh dipulih daripada bulu ayam. Kaedah autoklaf-alkali juga dicadangkan untuk digunakan sebagai pra-rawatan untuk meningkatkan prestasi rawatan biologi berikutnya. Kesan pra-rawatan autoklaf-alkali terhadap rawatan enzim yang berikutnya telah dikajikan dengan menggunakan Savinase® Ultra 16 L dan kaedah RSM. Kepekatan NaOH adalah parameter penting yang mempengaruhi tindak balas enzimatik berikutnya pada hidrolisis bulu dan pengeluaran protein (p <0.05). Bulu ayam yang telah dirawat dengan pra-rawatan autoklaf-alkali dalam keadaan optima (0.07 M NaOH, 2 minit masa pegangan pada 105°C) mampu meningkatkan hidrolisis Savinase sehingga 14 kali ganda dengan 80.81% hidrolisis bulu dan 0.69 g/g protein serta 673.80 mg/g asid amino bebas dipulih. Oleh itu, AAT adalah kaedah yang berpotensi untuk melarutkan bulu ayam dan memulihkan protein dengan asid amino bebas daripada bulu ayam. Pra-rawatan autoklaf-alkali juga boleh mempertingkatkan reaksi enzim untuk menguraikan bulu ayam dan menghasilkan hidrolisat bulu yang kaya dengan asid amino.



ACKNOWLEDGEMENTS

The completion of this project was made possible with the help and support from many people. First and foremost, I would like to express my sincere gratitude to my supervisor, Assoc. Prof. Dr. Phang Lai Yee for her guidance, valuable advice, supervision and support throughout my study. I would also like to express my appreciation to my co-supervisors, Dr. Siti Aqlima Ahmad and Assoc. Prof. Dr. Ooi Peck Toung for their encouragement and insightful comments.

I also indebted to the Ministry of Higher Eduction (MOHE) Malaysia and Universiti Putra Malaysia which have provided the financial aids for me to conduct my Master degree in Universiti Putra Malaysia. Besides, I also appreciate all the staffs and laboratory members in Faculty of Biotechnology and Biomolecular Sciences, especially Mrs. Norazlina, Mrs. Aluyah and Mrs. Siti Noraisyah for their assistance and technical support.

I also wish to express my appreciation to my friends and all my colleagues as well as the member of Bioproduct lab for their advice and moral support during the conduct of this research. Last but not least, I am highly grateful to my beloved family especially my parent for their love, encouragement, understanding and support that strengthen me throughout the study. I certify that a Thesis Examination Committee has met on 26 April 2018 to conduct the final examination of Cheong Chooi Wei on her thesis entitled "Autoclave-Chemical Hydrolysis of Chicken Feather for Protein Hydrolysate Production" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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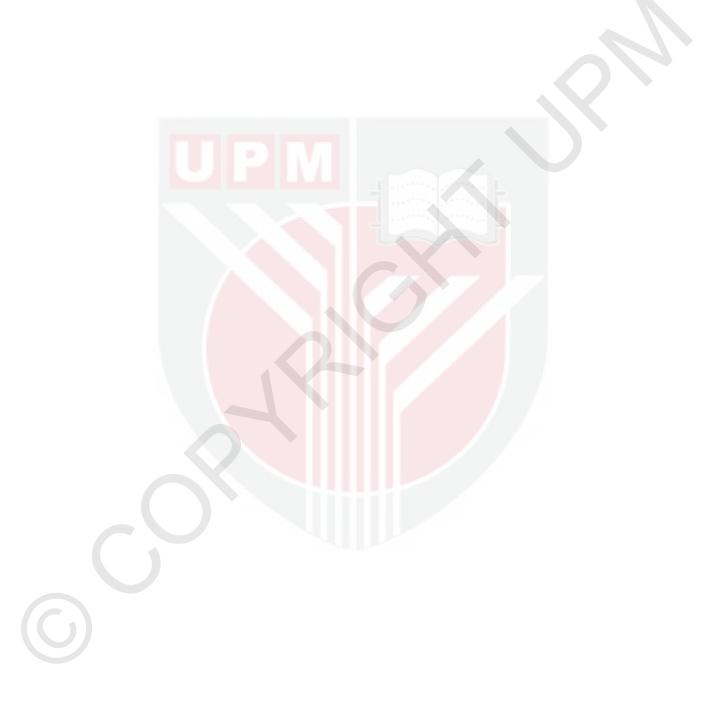
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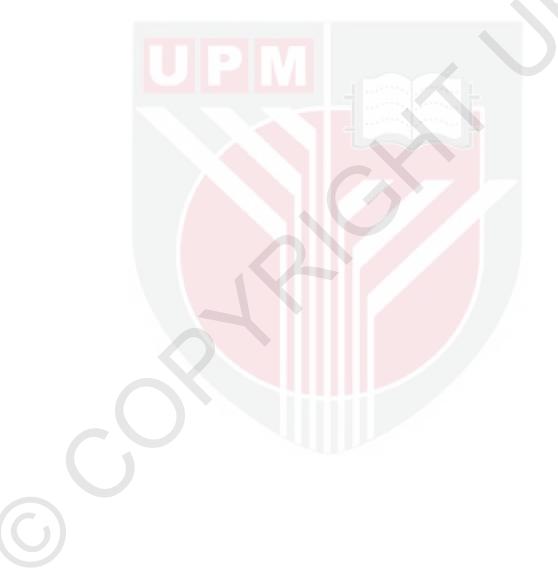
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LIST OF ABBREVIATIONS

AAS	Autoclave-alkaline Savinase combined treatment
AAT	Autoclave-alkaline treatment
°C	Degree Celsius
%	Percentage
A _{600nm}	Optical density at wavelength 600 nanometer
μL	Microliter
μm	Micrometer
CaCl ₂	Calcium chloride
g	Gram
Ľ	Litre
Μ	Molar
MT	Million tonnes
NaOH	Sodium hydroxide
HCI	Hydrochloric acid
RSM	Response surface methodology
CCD	Central composite design
FTIR	Fourier transform infrared
CHNS	Carbon, hydrogen, nitrogen and sulphur
USA	United State of America
IEC	Ion exchange chromatography
Psi	Pounds per square inch
GLN	Glutamine
GLU	Glutamic acid
ASP	Aspartic acid
ASN	Asparagine
NH4OH	Ammonium Hydroxide
NaCN	Sodium cyanide
DTT	Dithiothreitol



CHAPTER 1

INTRODUCTION

1.1 Research Background

Nowadays, the rate of consumption poultry meat is increasing year by year. In Malaysia, poultry meat consumption has reached 1632 million tonnes in 2017 (United State Department of Agriculture, 2017). This high consumption rate on poutry meat is probably due to its cheap price among the protein sources and it is suitable for all ethic and culture groups. The sharp demand on poultry meat leads to the production of noticeable amounts of by-products such as chicken feathers, from poultry processing plants. Since, chicken feathers constitute 10% of total chicken weight (Thyagarajan *et al.*, 2013), million tonnes of chicken feathers are generated annually in the worldwide poultry industries as the consumption of poultry meat increases (Ali *et al.*, 2011). Some of these feather wastes are utilized to produce dusters or decorative products as well as high-class bedding. However, most of the feathers are disposed in dumps, landfills and/or incinerators. These methods may cause contamination to our environment by generating greenhouse gases such as methane and carbon dioxide (Acda, 2010b). The application of these disposal methods were also limited due to lack of landfill spaces (Acda, 2010b) and high expenses (Tonkova *et al.*, 2009).

Feather is made up of more than 90% of protein, namely keratin (Mokrejs *et al.*, 2011) and it also contains high levels of certain amino acid which are cysteine, glycine, arginine, and phenylalanine (Kumar et al., 2012). Nowadays, essential amino acids were highly demanded in global market as it is expected to reach 10.1 million ton which is valued at USD 35.4 billion by 2022 (Radiant Insight Inc., 2015). Instead of becoming environmental pollutant, the chicken feathers can be transformed into valuable and marketable products. Up to date, they could be used to make feather meal (Kumar et al., 2012), bio-fertilizers (Saber et al., 2010) and thermoplastic (Ullah et al., 2011). Lately, feather was also suggested to be used as biomedical reagent for wound healing (Wang et al., 2017). However, feather is difficult to be hydrolyzed or degraded due to its high constitution of keratins (Mokrejs et al., 2011). Keratin consists of large amount of cysteine containing sulphur which is the building block of the disulphide bridge. Keratin is tightly packed either in α -helix or β -sheets into the keratin supercoiled polypeptides. These polypeptides are further stabilized by hydrogen bond and hydrophobic interaction (Brandelli, 2008). Also, keratin is the protein that has the high mechanical and temperature stability (Brandelli, 2008).

There are various treatments reported to be able to treat feather waste in order to add value to them (Barone and Schmidt, 2006; Chinta *et al.*, 2013; Lakshmi *et al.*, 2013; Mokrejs *et al.*, 2011b; Paul *et al.*, 2013; Staroń *et al.*, 2014; Stiborova *et al.*, 2016).

Generally, they are categorized into three groups namely, physical, chemical and biological treatments. Each of these treatments has the pros and cons in feather degradation, as well as, protein and amino acid recovering (Cheong *et al.*, 2017). Physical treatment involves fast reaction and the treatment can be easily handled. However, the feathers are usually degraded at high temperature or/and pressure which may lead to excess denaturation of certain amino acids and high amount of energy is needed for the machine operation in physical treatment. Chemical treatment can be performed easily by applying strong acids or alkaline to degrade the feathers. The use of reducing agents such as 2-mercaptoethanol, dithiothreitol (DTT), sodium mbisulphite and sodium bisulphite, also could effectively degrade chicken feathers into soluble forms (Sinkiewicz *et al.*, 2017). However, chemical treatment may also lead to loss of certain essential amino acid. For example, tryptophan, cystine, serine, and threonine were destroyed in acid hydrolysis (Haurowitz, 1955).

Biological treatment is an environmental friendly method which involves the utilization of keratinase producing microorganisms or keratinase alone to break down the rigid bonds in feathers. However, this method is time consuming due to the low reaction rate of the keratinase producing microorganisms or keratinase on feather hydrolysis (Kani *et al.*, 2012; Kim *et al.*, 2005; Matikeviciene *et al.*, 2009; Poovendran *et al.*, 2011). Considering the pros and cons of each treatment, it is necessary to develop an effective method for feathers hydrolysis in order to recover high amount of good quality protein and/or amino acid. Recently, studies of feather degradation by using combination of physical, chemical and biological treatments were done either in single step or two-steps. The combined treatments could diminish the disadvantages of the treatment methods while retaining the quality of recovered soluble protein and amino acids. For example, microwave-alkaline pretreatment (450 W, 0.05 M of NaOH for 10 min) has enhance the subsequent enzymatic hydrolysis of chicken feather to about 88% degradation with about 94% of soluble protein was recovered (Lee, 2016).

Autoclave is a thermal method that can be easily applied. Studies show that it might hydrolyse protein materials at appropriate conditions (Badadani *et al.*, 2007; Eddie *et al.*, 2016; Moore and Stem, 1963). Amino acids like glutamine and asparagine in the feather were destroyed under high-temperature and high pressure conditions (Wu, 2013). According to Taira (1973), 1.1 g/16g N of arginine and 2.0 g/16g N of lysine in soybean was lost after a treatment of 120°C for 30 min and 160°C for 10 min, respectively. However, there was no changes in amino acid when the soybean was treated under 100°C for 5 min (Taira, 1973). Hence, in order to avoid amino acids destruction, a lower temperature setting in the autoclave (105°C, 3 Psi pressure) was proposed in this research.

Strong alkaline may lead to losses of some essential amino acids. Stiborova *et al.* (2016) reported that 1% (w/w) of amino acids could be recovered from the feather being treated with 0.107 M of KOH within 24 hours at 70 °C. This indicated that at mild conditions (mild alkaline concentration and heating) feather could be hydrolyzed

to release small amount of amino acids. It could be hypothesized that autoclavechemical treatment at appropriate conditions may break down the rigid bonds in feathers and hence improve the protein and amino acid recovery. On the other hands, at certain circumstances, autoclave-chemical method may be applied as pretreatment to enhance the hydrolysis performace of biological treatment. Łaba and Szczekała (2013) reported that autoclaving in 10 mM sodium sulfite could enhance the subsequent feather hydrolysis by crude keratinase extracts of *Bacillus cereus* B5esz (86.3% degradation), as well as, the production of amino acids such as leucine, valine, glutamate, glycine, serine and cysteine.

1.2 Problem Statement

Autoclave heating is a common method used in biomass treatment. It has been classically applied for protein hydrolysis by treating the protein or peptide substances with the addition of 6 M hydrochloric acid, at 110°C for 18–24 hours. However, there is no study on the usage of autoclave-alkaline treatment on keratinous materials. Moreover, some studies also claimed that thermal-chemical process affect amino acids yield at high temperature and high concentration of chemicals (Barone and Schmidt, 2006; Guillermo *et al.*, 2016; Mokrejs *et al.*, 2011; Mehta *et al.*, 2014). Hence, research is needed to improve protein and amino acids recovery by thermal-chemical process. It is hypothesized that the overall protein hydrolysis could be improved by thermal-chemical treatment conducted at milder conditions, i.e. lower chemical concentration and temperature.

Biological treatment on chicken feather hydrolysis is an environment friendly method which can produce not only hydrolysates containing soluble proteins but also reduce loss in essential amino acids. However, biological treatment has low reaction rate and low yield, hence these drawbacks limited its application in industrial scale. Pretreatment is a common step in a fermentation process, especially if the biomass is used as feedstock due to its robust structure (Farid *et al.*, 2014; Łaba *et al.*, 2015; Wanitwattanarumlug *et al.*, 2012; Zakaria *et al.*, 2015). So, recently combined methods has been proposed in order to reduce the treatment time and to improve the yield of biological treatment. However, the reports on the usage of autoclave to pretreat feather was limited and current developed combined methods were effective in feather degradation but poor in protein recovery (Łaba and Szczekała, 2013; Lee, 2016). Research on treatment modification is needed to enhance solubilization of the feather and improve the recovery of protein and amino acids.

1.3 Research Objectives

General objective:

This study aimed to determine the feasibility of autoclave-chemical method as a treatment and/or pretreatment for chicken feather hydrolysate production.

The specific objectives of this study were:

- i. To investigate the effect of autoclave-chemical treatment on chicken feather hydrolysis;
- ii. To examine the effect of autoclave-alkaline as pretreatment on enzymatic hydrolysis of chicken feather.

1.4 Research Questions

There are some research questions were underlying with the objectives. The specific research questions of this study were:

- i. How much protein could be recovered from chicken feather by autoclavealkaline treatment?
- ii. Did autoclave-alkaline pretreatment enhance the enzymatic hydrolysis of chicken feather?

iii.

1.5 Scope of Study

This study involves three parts which are screening process (Part 1), autoclavechemical hydrolysis (Part 2) and autoclave-chemical-enzymatic hydrolysis (Part 3). In the screening process, three parameters namely, chemical types, chemical concentration, and holding time involved in autoclave-chemical hydrolysis were examined by using one-factor-at-a-time (OFAT) approach. Autoclave-alkaline as treatment and pretreatment were optimized with Response Surface Methodology (RSM). RSM is as efficient optimization tool which able to explain the effects of multiple factors and their interactions on the responses with reduced number of expriments (Bezerra et al., 2008). Center composite design (CCD) was used for the autoclave-alkaline treatment optimization. The effect of NaOH and holding time were the independent variables on total protein and percentage of solubilization were investigated. Savinase, a commercial enzyme was used in the enzymatic hydrolysis (Part 3 of the study). The solubilization of feathers in these treatments were determined based on the sample weight loss. The soluble protein production after the treatments were also analysed. Amino acid profiles of the hydrolysate produced after autoclavealkaline treatment and autoclave-alkaline-enzymatic treatment were determined. The structural and elemental changes of the treated samples were analyzed by Fourier Transform Infrared (FT-IR) Spectroscopy and Carbon, Hydrogen, Nitrogen and Sulphur (CHNS) analysis.

REFERENCES

- Aanifah, F. J. M., Phang, L. Y., Wasoh, H. and Abd-Aziz, S. (2014). Effect of different alkaline treatment on the release of ferulic acid from oil palm empty fruit bunch fibres. *Journal of Oil Palm Research*, 26(4): 321–331.
- Acda, M.N. (2010a). Sustainable use of waste chicken feather for durable and low cost building materials for tropical climates. In Sustainable Agriculture: Technology, Planning and Management, pp. 353–366.
- Acda, M.N. (2010b). Waste chicken feather as reinforcement in cement-bonded composites. *Philippine Journal of Science*,139(2): 161–166.
- Adigüzel, A.C., Bitlisli, B.O., Yaşa, I. and Eriksen, N.T. (2009). Sequential secretion of collagenolytic, elastolytic, and keratinolytic proteases in peptide-limited cultures of two *Bacillus cereus* strains isolated from wool. *Journal of Applied Microbiology*, 107(1): 226–234. http://doi.org/10.1111/j.1365-2672.2009.04200.x
- Al-Bahri, M.B., Al-Naimi, S.A. and Ahammed, S.H. (2009). Study the effect of hydrolysis variables on the production of soya proteins hydrolysis. *Al-Khwarizmi Engineering Journal*, 5(4): 25–38.
- Ali, T.H., Ali, N.H. and Mohamed, L. (2011). Production, purification and some properties of extracellular keratinase from feather-degradation by Aspergillus oryzae NRRL-447. Journal of Applied Sciences in Environmental Sanitation, 6(2): 123–136.
- Ameen, O.M., Usman, L.A., Muhammed, N.O., Okeola, O.F., Boluwarin, E.O. and Fadeyi, O. (2014). Effect of heat and alkaline hydrolysis on the amino acid. *African Journal of Food Agriculture, Nutrition and Development*, 14(2): 1–13.
- Anna, T. (2013). Biogas production from lignocelluloses: Pretreatment, substrate characterization, co-digestion, and economic evaluation, PhD Thesis, University of Boras.
- Ashraf, G., Sonkar, C., Masih, D. and Shams, R. (2017). Study on the effect of thermal processing on ready-to-eat poultry egg keema. *Journal of Food Processing & Technology*, 8(7): 684. http://doi.org/10.4172/2157-7110.1000684
- Asquith, R. (1977). Chemistry of natural protein fibers. Plenum Press, New York, USA.
- Ayutthaya, S.I.N., Tanpichai, S. and Wootthikanokkhan, J. (2015). Keratin extracted from chicken feather waste: Extraction, preparation, and structural characterization of the keratin and keratin/biopolymer films and electrospuns. *Journal of Polymers and the Environment*, 23(4): 506–516. http://doi.org/10.1007/s10924-015-0725-8

- Badadani, M., SureshBabu, S.V. and Shetty, K.T. (2007). Optimum conditions of autoclaving for hydrolysis of proteins and urinary peptides of prolyl and hydroxyprolyl residues and HPLC analysis. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 847(2): 267–274. http://doi.org/10.1016/j.jchromb.2006.10.021
- Barone, J.R., and Schmidt, W.F. (2006). Effect of formic acid exposure on keratin fiber derived from poultry feather biomass. *Bioresource Technology*, 97(2): 233– 242. http://doi.org/10.1016/j.biortech.2005.02.039
- Barret, G. (1985). Chemistry and Biochemistry of the amino acids.
- Barth, A. (2007). Infrared spectroscopy of proteins. *Biochimica et Biophysica Acta Bioenergetics*, 1767(9): 1073–1101. http://doi.org/10.1016/j.bbabio.2007.06.004
- Bezerra, M.A., Santelli, R.E., Oliveira, E.P., Villar, L.S. and Escaleira, L.A. (2008). Response surface methodology (RSM) as a tool for optimization in analytical chemistry. *Talanta*, 76: 965–977
- Bohacz, J. (2017). Biodegradation of feather waste keratin by a keratinolytic soil fungus of the genus *Chrysosporium* and statistical optimization of feather mass loss. *World Journal of Microbiology and Biotechnology*, 33(1): 1–16. http://doi.org/10.1007/s11274-016-2177-2
- Brandelli, A. (2008). Bacterial keratinases: Useful enzymes for bioprocessing agroindustrial wastes and beyond. *Food Bioprocess Technology*, 1(2): 105–116. http://doi.org/10.1007/s11947-007-0025-y
- Brandelli, A., Daroit, D. J. and Riffel, A. (2010). Biochemical features of microbial keratinases and their production and applications. *Applied Microbiology and Biotechnology*, 85(6): 1735–1750. http://doi.org/10.1007/s00253-009-2398-5
- Brebu, M. and Spiridon, I. (2011). Thermal degradation of keratin waste. *Journal of Analytical and Applied Pyrolysis*, 91(2): 288–295. http://doi.org/10.1016/j.jaap.2011.03.003
- Burnett, L.R., Rahmany, M.B., Richter, J.R., Aboushwareb, T.A., Eberli, D., Ward, C.L., Orlando, G., Hantgan, R.R., Van Dyke, M.E. and Van Dyke, M.E. (2013). Hemostatic properties and the role of cell receptor recognition in human hair keratin protein hydrogels. *Biomaterials*, 34(11): 2632–2640. http://doi.org/10.1016/j.biomaterials.2012.12.022
- Cai, CG. and Zheng, X. (2009). Medium optimization for keratinase production in hair substrate by a new *Bacillus subtilis* KD-N2 using response surface methodology. *Journal of Industrial Microbiology & Biotechnology*, 36(7): 875–883.
- Chen, J.Y., Ding, S.Y., Ji, Y.M., Ding, J.Y., Yang, X.Y., Zou, M.H. and Li, Z.Y. (2015). Microwave-enhanced hydrolysis of poultry feather to produce amino acid. *Chemical Engineering and Processing: Process Intensification*, 87: 104–109.

http://doi.org/https://doi.org/10.1016/j.cep.2014.11.017

- Cheong, C.W., Ahmad, S.A., Ooi, P.T., and Phang, L.Y. (2017). Treatment of chicken feather waste. *Pertanika Journal of Scholarly Research Reviews.*, 3(1): 50–59.
- Cheran, K., Rudhra, S.M. and Krithigaa, M. (2017). A performance analysis of autoclave aerated concrete. *International Journal of Science and Engineering Research (IJOSER)*, 5(3): 1112–1117.
- Chew, C.S. (2015). Effect of acid-microwave pretreatment and enzymatic treatment on chicken feather hydrolysis. Degree Thesis. Universiti Putra Malaysia.
- Chinta, S., Landage, S. and Yadav, K. (2013). Application of chicken feathers intechnical textiles. *International Journal of Innovative Research in Science, Engineering and Technology*, 2(4): 1158–1165.
- Costa, J.C., Barbosa, S.G., Alves, M.M. and Sousa, D.Z. (2012). Thermochemical preand biological co-treatments to improve hydrolysis and methane production from poultry litter. *Bioresource Technology*, 111: 141–147. http://doi.org/10.1016/j.biortech.2012.02.047
- Cotana, F., Barbanera, M., Foschini, D., Lascaro, E. and Buratti, C. (2015).
 Preliminary optimization of alkaline pretreatment for ethanol production from vineyard pruning. *Energy Procedia*, 82: 389–394. http://doi.org/10.1016/j.egypro.2015.11.814
- Darah, I., Nur-Diyana, A., Nurul-Husna, S., Jain, K. and Lim, S.H. (2013). *Microsporum fulvum* IBRL SD3: As novel isolate for chicken feathers degradation. *Applied Biochemistry and Biotechnology*, 171(7): 1900–1910. http://doi.org/10.1007/s12010-013-0496-4
- Débora, D.B., Rasiah, L., Belarmino, L.D., Pimentel, J.R., da Rocha, B.G., Galvão, A.O. and Andrade, S. M. (2012). Physical and morphological structure of chicken feathers (keratin biofiber) in natural, chemically and thermally modified forms. *Materials Sciences and Applications*, 3(12): 887–893. http://doi.org/10.4236/msa.2012.312129
- Demir, T., Hameş, E. E., Öncel, S. S. and Vardar-Sukan, F. (2015). An optimization approach to scale up keratinase production by *Streptomyces* sp. 2M21 by utilizing chicken feather. *International Biodeterioration & Biodegradation*, 103: 134–140. http://doi.org/10.1016/j.ibiod.2015.04.025
- Dhurai, B. and Saravanan, K. (2012). Exploration on amino acid content and morphological structure in chicken feather fiber. *Journal of Textile and Apparel, Technology and Management*, 7(3): 1–6.
- Dou, Y., Zhang, B., He, M., Yin, G. and Cui, Y. (2016). The structure, tensile properties and water resistance of hydrolyzed feather keratin-based bioplastics. *Chinese Journal of Chemical Engineering*, 24(3): 415–420.

http://doi.org/10.1016/j.cjche.2015.11.007

- Eddie, T.T.T., Ken, W.L.Y., Wong, S.H., Arcy, B.R.D., Jassim, R.A., De Voss, J.J. and Fletcher, M.T. (2016). Thermo-alkaline treatment as a practical degradation strategy to reduce indospicine contamination in camel meat. *Journal of Agricultural and Food Chemistry*, 64(44): 8447–8453. http://doi.org/10.1021/acs.jafc.6b03499
- Eslahi, N., Dadashian, F. and Nejad, N.H. (2013). An investigation on keratin extraction from wool and feather waste by enzymatic hydrolysis. *Preparative Biochemistry* and *Biotechnology*, 43(7): 624–648. http://doi.org/10.1080/10826068.2013.763826
- Fakhfakh, N., Haddar, A., Hmidet, N., Frikha, F. and Nasri, M. (2010). Application of statistical experimental design for optimization of keratinases production by *Bacillus pumilus* A1 grown on chicken feather and some biochemical properties. *Process Biochemistry*, 45(5): 617–626. http://doi.org/10.1016/j.procbio.2009.12.007
- Fakhfakh, N., Ktari, K., Haddar, A., Hamza, I., Dahmen, M. and Nasri, M. (2011). Total solubilisation of the chicken feathers by fermentation with a keratinolytic bacterium, *Bacillus pumilus* A1, and the production of protein hydrolysate with high antioxidative activity. *Process Biochemistry*, 46(9): 1731–1737. http://doi.org/https://doi.org/10.1016/j.procbio.2011.05.023
- Farid, M.A., Noor El-Deen, A.M. and Shata, H.M. (2014). Optimization of microwave pretreatment and enzymatic hydrolysis of pith bagasse with *Trichoderma* cellulase. *Indian Journal of Biotechnology*, 13(1): 98–107.
- Figueras, M. (1997). Ultra- structural aspects of hair digestion in black piedra infection. Journal of Medical and Veterinary Mycology, 35: 1–6.
- Fisher, M.L., Leeson, S., Morrison, W.D. and Summers, J. (1981). Feather growth and feather composition of broiler chickens. *Canadian Journal of Animal Science*, 61: 769–773.
- Forgács, G. (2012). Biogas production from citrus wastes and chicken feather : pretreatment and co-digestion. PhD Thesis, University of Boras.
- Fountoulakis, M. and Lahm, H.-W. (1998). Hydrolysis and amino acid composition analysis of proteins. *Journal of Chromatography A*, 826(2): 109–134. http://doi.org/10.1016/S0021-9673(98)00721-3
- Fraser, R.D.B., MacRae, T.P., Parry, D.A.D. and Suzuki, E. (1971). The structure of feather keratin. *Polymer*, 12(1): 35–56. http://doi.org/10.1016/0032-3861(71)90011-5
- Fujine, K. (2013). FlashEA 1112 elemental analyzer (CHNS): User guide. International Ocean Discovery Program.

- Galarza, B., Cavello, I., Garro, L., Gortari, C., Hours, R. and Cantera, C. (2012). Evaluation of increase at the production of keratinolytics enzymes. *Journal of Aqeic*, 63(3): 70–76.
- Gerald, G., Flory, K.R., Browne, B.A., Majid, S., Ichida, J.M. and Burtt, E.H. (2004). Bacterial degradation of black and white feathers. *The Auk*, 121(3): 656–659.
- Goodwin, W.D. (1976). Nutrient protein from keratinaceous material solubilized with N,N,-dimethylformamide. Patent *US3970614 A*, The Athlon Corporation.
- Gousterova, A., Nustorova, M., Paskaleva, D., Naydenov, M., Neshev, G. and Vasileva-Tonkova, E. (2012). Assessment of feather hydrolysate from thermophilic actinomycetes for soil amendment and biological control application. *International Journal of Environmental Research*, 6(2): 467–474.
- Grazziotin, A., Pimentel, F. A., De Jong, E. V. and Brandelli, A. (2006). Nutritional improvement of feather protein by treatment with microbial keratinase. *Animal Feed Science and Technology*, 126(1-2): 135–144. http://doi.org/10.1016/j.anifeedsci.2005.06.002
- Guillermo, C.K., Agbogbo, F.K. and Holtzapple, M. (2016a). Lime treatment of keratinous materials for the generation of highly digestible animal feed: 2. Animal hair. *Bioresource Technology*, 97(11): 1344–1352.
- Guillermo, C.K., Chang, V.S., Agbogbo, F.K. and Holtzapple, M. (2016b). Lime treatment of keratinous materials for the generation of highly digestible animal feed: 1. Chicken feathers. *Bioresource Technology*, 97(11): 1337–1343.
- Gunderson, A.R., Frame, A.M., Swaddle, J.P. and Forsyth, M. H. (2008). Resistance of melanized feathers to bacterial degradation: is it really so black and white? *Journal of Avian Biology*, 39(5): 539–545. http://doi.org/10.1111/j.2008.0908-8857.04413.x
- Guo, T., He, A.Y., Du, T.F., Zhu, D.W., Liang, D.F., Jiang, M., Wei, P. and Ouyang, P. K. (2013). Butanol production from hemicellulosic hydrolysate of corn fiber by a *Clostridium beijerinckii* mutant with high inhibitor-tolerance. *Bioresource Technology*, 135: 379–385. http://doi.org/10.1016/j.biortech.2012.08.029
- Gupta, A., Kamarudin, N., Chua, G.K. and Yunus, R.M. (2012). Extraction of keratin protein from chicken feather. *Journal of Chemistry and Chemical Engineering*, 6 (8): 732-737. ISSN 1934-7375.
- Gupta, R. and Ramnani, P. (2006). Microbial keratinases and their prospective applications: An overview. *Applied Microbiology and Biotechnology*, 70(1): 21–33. http://doi.org/10.1007/s00253-005-0239-8
- Hamad, A.J. (2014). Materials, production, properties and application of aerated lightweight concrete: Review. *International Journal of Materials Science and Engineering*, 2(2): 152–157. http://doi.org/10.12720/ijmse.2.2.152-157

- Harlander-Matauschek, A. and Feise, U. (2009). Physical characteristics of feathers play a role in feather eating behavior. *Poultry Science*, 88(9): 1800–1804. http://doi.org/10.3382/ps.2008-00566
- Haurowitz, F. (1955). *Biochemistry. An Introductory Textbook*. New York: John Wiley & Sons, Inc.
- Hegarty, M.P. and Pound, A. (1970). Indospicine, a hepatotoxic amino acid from *Indigofera spicata*: Isolation, structure, and biological studies. *Australia Journal of Biological Science*, 23: 831–842.
- Hii, K., Baroutian, S., Parthasarathy, R., Gapes, D.J. and Eshtiaghi, N. (2014). A review of wet air oxidation and thermal hydrolysis technologies in sludge treatment. *Bioresource Technology*, 155: 289–299. http://doi.org/10.1016/j.biortech.2013.12.066.
- Horvath, A.L. (2009). Solubility of structurally complicated materials: 3. Hair. *The Scientific World Journal*, 9: 255–271. http://doi.org/10.1100/tsw.2009.27
- Hossain, M.S., Balakrishnan, V., Rahman, N.N.N., Sarker, M.Z.I. and Kadir, M.O.A. (2012). Treatment of clinical solid waste using a steam autoclave as a possible alternative technology to incineration. *International Journal of Environmental Research and Public Health*, 9(3): 855–867. http://doi.org/10.3390/ijerph9030855
- Hugo, W. (1991). A brief history of heat and chemical preservation and disinfection. *Journal of Applied Bacteriology*, 71(1): 9–18.
- Ismail, S.N. and Manaf, L. (2013). The challenge of future landfill: A case study of Malaysia. *Journal of Toxicology and Environmental Health Sciences*, 5(6): 86– 96. http://doi.org/10.5897/JTEHS12.058
- Jayaraman, K., Munira, H., Chowdhury, D. and Iranmanesh, M. (2013). The preference and consumption of chicken lovers with race as a moderator An empirical study in Malaysia. *International Food Research Journal*, 20(1): 165–174.
- Jayathilakan, K., Sultana, K., Radhakrishna, K. and Bawa, A. (2012). Ultilization of byproducts and waste materials from meat, poultry and fish processing inductries: A review. *Journal of Food and Technology*, 49(3): 278–293.
- Kadri, S.H., Allen, W.M. and Pikel, J. (1979). Method for the preparation of watersoluble keratinaceous protein using saturated steam and water. Patent *US4172073 A*. Chemetron Corporation.
- Kakkar, P., Madhan, B. and Shanmugam, G. (2014). Extraction and characterization of keratin from bovine hoof: A potential material for biomedical applications. *SpringerPlus*, 3(1): 596. http://doi.org/10.1186/2193-1801-3-596

- Kanchana, R. and Mesta, D. (2013). Native feather degradation by a keratinophilic fungus. *International Journal of ChemTech Research*, 5(6): 2947–2954.
- Kani, T.P., Subha, K., Madhanraj, P., Senthilkumar, G. and Panneerselvam, A. (2012). Degradation of chicken feathers by *Leuconostoc* sp. and *Pseudomonas microphilus*. *European Journal of Experimental Biology*, 2(2): 358–362.
- Katharine, G. (2011). New process turns waste chicken feathers into biodegradable plastic. Retrieved May 18, 2016, from http://phys.org/news/2011-04-chicken-feathers-biodegradable-plastic.html
- Kazilek, C.J. (2009). Feather biology. Retrieved November 26, 2017, from https://askabiologist.asu.edu/explore/feather-biology
- Khandve P.V. (2016). Applications of AAC in construction industry assistant. Journal of Environmental Science, Computer Science and Engineering and Technology, 5(1): 91–97.
- Kim, J.M., Yang, M.C. and Hyung, J.S. (2005). Preparation of feather digests as fertilizer with *Bacillus pumilis* KHS-1. *Journal of Microbiol and Biotechnology*, 15(3): 472–476.
- Kim, W.K., Lorenz, E.S. and Patterson, P.H. (2002). Effect of enzymatic and chemical treatments on feather solubility and digestibility. *Poultry Science*, 81(1): 95–98.
- King'ori, A. M. (2012). Management of poultry processing by-products: Utilization of feathers. *International Journal of Livestock Research ISSN*, 2(3): 58–64.
- Kothari, V., Patadia, M. and Trivedi, N. (2011). Microwave sterilized media supports better microbial growth than autoclaved media. *Research in Biotechnology*, 2(5): 63–72.
- Kowata, K., Nakaoka, M., Nishio, K., Fukao, A., Satoh, A., Ogoshi, M., Takahashi, S., Tsudzuki, M. and Takeuchi, S. (2014). Identification of a feather β-keratin gene exclusively expressed in pennaceous barbule cells of contour feathers in chicken. *Gene*, 542(1): 23–28. http://doi.org/10.1016/j.gene.2014.03.027
- Kratchanova, M., Pavlova, E. and Panchev, I. (2004). The effect of microwave heating of fresh orange peels on the fruit tissue and quality of extracted pectin. *Carbohydrate Polymers*, 56(2): 181–185. http://doi.org/https://doi.org/10.1016/j.carbpol.2004.01.009
- Krejci, O., Mokrejs, P. and Sukop, S. (2011). Preparation and characterization of keratin hydrolysates, pp.308–311. Retrieved from http://www.wseas.us/e-library/conferences/2011/Catania/Catania-53.pdf
- Krishna, P.K., Sabiha, S.S., Rohini, K.K. and Abraham, P. (2014). Bioplastic from chicken feather waste, 27(65): 373–375.

- Kumar, D.J.M., Priya, P., Balasundari, S.N., Devi, G.S.D.N., Rebecca, A.I.N. and Kalaichelvan, P.T. (2012). Production and optimization of feather protein hydrolysate from *Bacillus* sp. MPTK6 and its antioxidant potential. *Middle-East Journal of Scientific Research*, 11(7): 900–907. Retrieved from http://www.idosi.org/mejsr/mejsr11(7)12/9.pdf
- Kumar, R., Johnsy, G., Rajamanickam, R., Lakshmana, J.H., Kathiravan, T., Nataraju, S. and Nadanasabapathi, S. (2013). Effect of gamma irradiation and retort processing on microbial, chemical and sensory quality of ready-to-eat (RTE) chicken pulav. *International Food Research Journal*, 20(4): 1579–1584.
- Kumaran, P., Gupta, A. and Sharma, S. (2017). Synthesis of wound-healing keratin hydrogels using chicken feathers proteins and its properties, 9(2): 171-178. http://doi.org/10.22159/ijpps.2017v9i2.15620
- Kunert, J. (1992). Effect of redicing agents on proteolytic and keratinolytic activity of enzymes of *Microsporium gypseum*. *Mycoses*, 35: 343–348.
- Kunert, J. (2000). Physiology of keratinophilic fungi. In Kushwaha RKS, Guarro J (eds) Biology of dermatophytes and other keratinophilic fungi, 1st edn. pp. 77– 85.
- Łaba, W. and Szczekała, K. (2013). Keratinolytic proteases in biodegradation of pretreated feathers. *Polish Journal of Environmental Studies*, 22(4): 1101–1109.
- Łaba, W., Kopec, W., Chorazyk, D., Kancelista, A., Piegza, M. and Malik, K. (2015). Biodegradation of pretreated pig bristles by *Bacillus cereus* B5esz. *International Biodeterioration and Biodegradation*, 100: 116–123. http://doi.org/10.1016/j.ibiod.2015.02.024
- Lai, L.W., Teo, C.H. and Idris, A. (2013). Different pretreatment methods to evaluate lignin content in oil palm trunk. In *International Conference on Industrial Engineering and Management Science*. pp. 156–161.
- Lai, L.W., Ibrahim, M., Md Rahim, N., Hashim, E. F., Ya'cob, M. Z., Idris, A. and Akhtar, J. (2016). Study on composition, structural and property changes of oil palm frond biomass under different pretreatments. *Cellulose Chemistry and Technology*, 50(9-10): 951-959.
- Lakshmi, T.B.V., Chowdary, Y.A., Soumya, M., Devi, P.P., Anuradha, K. and Madhuri, A. (2013). Degradation of chicken feathers by *Proteus vulgaris* and *Micrococcus luteus*. *International Journal of Pharmaceutical & Biological Archives*, 4(2): 366–370.
- Lee, Y.Z., Phang, L.Y., Ahmad, S.A. and Ooi, P. T. (2016). Microwave-alkali treatment of chicken feathers for protein hydrolysate production. *Waste and Biomass Valorization*, 7(5): 1147–1157. http://doi.org/10.1007/s12649-016-9483-7

- Lee, Y.Z. (2016). Microwave-alkaline treatment of chicken feathers for protein hydrolysate production. Msc Thesis, Universiti Putra Malysia.
- Lopes, F.C., Silva, L.A.D., Tichota, D.M., Daroit, D.J., Velho, R.V., Pereira, J.Q., Corrêa, A.P.F. and Brandelli, A. (2011). Production of proteolytic enzymes by a keratin-degrading *Aspergillus niger*. *Enzyme Research*, 2011: 1–9. http://doi.org/10.4061/2011/487093
- Loralyn, H.L. and Marshall, R.T. (2010). Microbiological spoilage of dairy products. In *Compendium of the microbiological spoilage of foods and beverages*. pp. 41-67. http://doi.org/10.1007/978-1-4419-0826-1
- Lowry, O.H., Rosebrough, N.J., Farr, A.L. and Randall, R.J. (1951). Total protein estimation by Lowry's method. *The Journal of Biological Chemistry*, 193: 265.
- Ma, B.M., Qiao, X., Hou, X.L. and Yang, Y.Q. (2016). Pure keratin membrane and fibers from chicken feather. *International Journal of Biological Macromolecules*, 89: 614–621. http://doi.org/10.1016/j.ijbiomac.2016.04.039
- Mabrouk, M.E.M. (2008). Feather degradation by a new keratinolytic *Streptomyces* sp. MS-2. *World Journal of Microbiology and Biotechnology*, 24(10): 2331–2338.
- Mackey, B.M., Miles, C., Parsons, S.E. and Seymour, D. (1991). Thermal denaturation of whole cells and cell components of *Escherichia coli* examined by differential scanning calorimetry. *Journal of General Microbiology*, 137(10): 2361–2374. http://doi.org/10.1099/00221287-137-10-2361
- Magee, T.R.A. and Neill, G. (2012). Effects of heat treatment on protein denaturation and starch gelatinisation in wheat flour. *Journal of Food Engineering*, 113: 422–426.
- Maheshi, D., Passel, S.V., Nelen, D., Tielemans, Y. and Acker, K.V. (2013). Environmental and socio-economic impacts of landfills. In *Linnaeus ECO-TECH* 2012. pp. 40–52.
- Malviya, H.K., Rajak, R.C. and Hasija, S.K. (1992). Synthesis and regulation of extracellular keratinase in three fungi isolated from the grounds of a gelatin factory, Jabalpur, India. *Mycopathologia*, 120: 1–4.
- Martinez-Hernandez, A.L., Velasco-Santos, C., De Icaza, M and Castano, V. M. (2015). Microstructural characterisation of keratin fibres from chicken feathers. *International Journal of Environment and Pollution*, 23(2): 162–178. http://doi.org/10.1504/IJEP.2005.006858
- Martinez-Hernandez, A.L. and Velasco-Santos, C. (2012). Keratin fibers from chicken feathers: Structure and advances in polymer composites. In *Keratin: Structure, Properties and Applications*. pp. 149–211. Nova Science Publishers, Inc.

Matikeviciene, V., Masiliuniene, D. and Grigiskis, S. (2009). Degradation of keratin

containing wastes by bacteria with keratinolytic activity. In proceedings of the 7th International Scientific and Practical Conference 1: 284–289.

- Mazotto, A.M, Ascheri, J.L.R., de Oliveira Godoy, R.L., Damaso, M.C.T., Couri, S. and Vermelho, A.B. (2017). Production of feather protein hydrolyzed by *B. subtilis* AMR and its application in a blend with cornmeal by extrusion. *Food Science and Technology*, 84: 701–709. http://doi.org/10.1016/j.lwt.2017.05.077
- Mehta, R.S., Jholapara, R.J. and Sawant, C.S. (2014). Isolation of a novel featherdegrading bacterium and optimization of its cultural conditions for enzyme production. *International Journal of Pharmacy and Pharmaceutical Sciences*, 6(1): 194–201.
- Meyers, M.D., Chen, P.Y., Lin, Y.M. and Seki, Y. (2008). Biological materials: Structure and mechanical properties. *Progress in Materials Science*, 53:1–206
- Mokrejs, P., Svoboda, P., Hrncirik, J., Janacova, D. and Vasek, V. (2011). Processing poultry feathers into keratin hydrolysate through alkaline-enzymatic hydrolysis. Waste Management & Research : The Journal of the International Solid Wastes and Public Cleansing Association, ISWA, 29(3): 260–267. http://doi.org/10.1177/0734242X10370378
- Moore, S. and Stem, W.H. (1963). Chromatographic determination of amino acids by the use of automatic recording equipment. *Methods in Enzymology.*, 6: 819–831.
- Mousavi, S., Salouti, M., Shapoury, R. and Heidari, Z. (2013). Optimizatin of keratinase production for feather degradation by *Bacillus subtilis*. *Journal of Microbiology*, 6(8): e7160. http://doi.org/10.5812/jjm.7160
- Mukherjee, A.K., Rai, S.K. and Bordoloi, N.K. (2011). Biodegradation of waste chicken-feathers by an alkaline β-keratinase (Mukartinase) purified from a mutant *Brevibacillus* sp. strain AS-S10-II. *International Biodeterioration & Biodegradation*, 65(8): 1229–1237. http://doi.org/10.1016/j.ibiod.2011.09.007
- Munira, H. (2011). The factors influencing chicken lovers in Malaysia, race as the moderating variable. MSc Thesis Universiti Sains Malaysia.
- Nagal, S. and Jain, P. (2010). Feather degradation by strains of *Bacillus* isolated from decomposing feathers. *Brazilian Journal of Microbiology*, 41(1): 196–200. http://doi.org/10.1590/S1517-83822010000100028
- Nasir, I. and Tinia, M.G. (2015). Pretreatment of lignocellulosic biomass from animal manure as a means of enhancing biogas production. *Engineering in Life Sciences Journal*, 15(7): 733–742.
- Nayaka, S. and Vidyasagar, G.M. (2013). Development of eco-friendly bio-fertilizer using feather compost. *Annals of Plant Sciences*, 2(7): 238 244.

- Nikhilesh, B., Sachin, Z.A., Vishal, T. and Dipesh, J. (2013). A review: Steam sterilization a method of sterilization. *Journal of Biological & Scientific Opinion*, 1(2): 138–141. http://doi.org/10.7897/2321
- Oladele, I.O., Omotoyimbo, J.A. and Ayemidejor, S.H. (2014). Mechanical properties of chicken feather and cow hair fibre reinforced high density polyethylene composites. *International Journal of Science and Technology*, 3(1): 66-72.
- Onifade, A.A., Al-Sane, N.A., Al-Musallam, A.A. and Al-Zarban, S. (1998). A review: Potentials for biotechnological applications of keratin-degrading microorganisms and their enzymes for nutritional improvement of feathers and other keratins as livestock feed resources. *Bioresource Technology*, 66(1): 1–11. http://doi.org/10.1016/S0960-8524(98)00033-9
- Papadopolous, M.C, El-Boushy, A.R., Roodbeen, A.E. and Ketelaars, E. (1986). Effects of processing time and moisture content on amino acids composition and nitrogen characteristics of feather meal. *Animal Feed Science and Technology*, 14: 279–290.
- Pariatamby, A. and Tanaka, M. (2015). *Municipal solid waste management in Asia* and the Pacific Islands: Challenges and strategic solutions. Springer-Verlag Singapore 2014. http://doi.org/10.1177/139156140901000108
- Paul, T., Halder, S.K., Das, A., Bera, S., Maity, C., Mandal, A., Das, P.S., Mohapatra, P.K.D., Pati, B.R. and Mondal, K.C. (2013). Exploitation of chicken feather waste as a plant growth promoting agent using keratinase producing novel isolate *Paenibacillus woosongensis* TKB2. *Biocatalysis and Agricultural Biotechnology*, 2(1): 50–57. http://doi.org/10.1016/j.bcab.2012.10.001
- Pavani, P. and Rajeswari, T. (2014). Impact of plastics on environmental pollution. Journal of Chemical and Pharmaceutical Sciences, 3: 87–93.
- Poovendran, P., Kalaigandhi, V., Kanan, VK., Jamunarani, E. and Poongunran, E. (2011). A study of feather keratin degradation by *Bacillus licheniformis* and quantification of keratinase enzyme produced. *Journal of Microbiology and Biotechnology Research*, 1(3): 120–126.
- Priorclaves. (2011). Operation & Maintenance Manual.pp.1-51. London.
- Radiant Insight Inc. (2015). *Amino acids market size & research report*, 2022. Retrieved from https://www.radiantinsights.com/research/amino-acids-market
- Rahayu, S., Syah, D. and Suhartono, M.T. (2012). Degradation of keratin by keratinase and disulfide reductase from *Bacillus* sp. MTS of Indonesian origin. *Biocatalysis and Agricultural Biotechnology*, 1(2): 152–158. http://doi.org/10.1016/j.bcab.2012.02.001
- Rahmany, M.B., Hantgan, R.R. and Dyke, M.V. (2013). A mechanistic investigation of the effect of keratin-based hemostatic agents on coagulation. *Biomaterials*,

34(10): 2492–2500. http://doi.org/10.1016/j.biomaterials.2012.12.008

- Ramesh, S., Muthuvelayudham, R., Kannan, R.R. and Viruthagiri, T. (2013). Enhanced production of cellulase from tapioca stem using response surface methodology. *Innovative Romanian Food Biotechnology*, 12: 40–51. http://doi.org/doi:10.1155/2013/514676.
- Rathi, S.O. and Khandve, P.V. (2015). AAC block A new eco-friendly material for construction. *International Journal of Advance Engineering and Research Development*, 2(4): 410–414.
- Reddy, N. and Yang, Y. (2007). Structure and properties of chicken feather barbs as natural protein fibers. *Journal of Polymers and the Environment*, 15(2): 81–87. http://doi.org/10.1007/s10924-007-0054-7
- Riffel, A., and Brandelli, A. (2002). Isolation and characterization of a featherdegrading bacterium from the poultry processing industry. *Journal of Industrial Microbiology and Biotechnology*. 29:255–258.
- Ruangmee, A. and Sangwichien, C. (2013). Response surface optimization of enzymatic hydrolysis of narrow-leaf cattail for bioethanol production. *Energy Conversion* and Management, 73: 381–388. http://doi.org/10.1016/j.enconman.2013.05.035
- Rutala, W.A. and Weber, D.J. (2013). Disinfection and sterilization: An overview. *American Journal of Infection Control*, 41: S2–S5. http://doi.org/10.1016/j.ajic.2012.11.005
- Saber, W.I.A., El-Metwally, M.M. and El-Hersh, M. S. (2010). Keratinase production and biodegradation of some keratinous waste by *Alternaria tenuissima* and *Aspergillus nidulans*. *Research Journal of Microbiology*, 5(1): 21–35.
- Samsuddin, N.S., Sharaai, A.H. and Ismail, M.M. (2015). Advances in environmental biology sustainability of chicken meat production in achieving food security in Malaysia. *Advances in Environmental Biology*, 9(23): 1–6.
- Sangali, S. and Brandelli, A. (2000). Feather keratin hydrolysis by a *Vibrio* sp. strain KR2. *Journal of Applied Microbiology*, 89(5): 735–743. http://doi.org/10.1046/j.1365-2672.2000.01173.x
- Schmidt, W.F. (1998). Innovative feather utilization strategies. *Proceedings of National Poultry Waste Management conference*, pp276-282.
- Schor, R. and Krimm, S. (1961). Studies on the structure of feather keratin: II. A β-helix model for the structure of feather keratin. *Biophysical Journal*, 1(6): 489–515. http://doi.org/10.1016/S0006-3495(61)86904-X
- Sinkiewicz, I, Sliwinska, A, Staroszczyk, H. and Kolodziejska, I. (2017). Alternative methods of preparation of soluble keratin from chicken feathers. *Waste and*

Biomass Valorization, 8: 1043–1048. http://doi.org/10.1007/s12649-016-9678-y

- Sivakumar, T., Shankar, T., Vijayabaskar, P. and Ramasubramanian, V. (2012). Optimization for keratinase enzyme production using *Bacillus thuringiensis* TS2. *Academic Journal of Plant Sciences*, 5(3): 102–109. http://doi.org/10.5829/idosi.ajps.2012.5.3.6279
- Staroń, P., Banach, M., Kowalski, Z., Staroń, A., Materiałów, H., Pochodzących, K., and Drobiarskiego, Z.P. (2014). Hydrolysis of keratin materials derived from poultry industry. *Proceedings of ECOpole*, 8(2): 443–448. http://doi.org/10.2429/proc.2014.8(2)050
- Stiborova, H., Branska, B., Vesela, T., Lovecka, P., Stranska, M., Hajslova, J., Jiru, M., Patakova, P. and Demnerova, K. (2016). Transformation of raw feather waste into digestible peptides and amino acids. *Journal of Chemical Technology and Biotechnology*, 91(6): 1629–1637. http://doi.org/10.1002/jctb.4912
- Sun, P., Liu, Z.T. and Liu, Z.W. (2009). Particles from bird feather: A novel application of an ionic liquid and waste resource. *Journal of Hazardous Materials*, 170(2-3): 786–790. http://doi.org/10.1016/j.jhazmat.2009.05.034
- Taira, H. (1973). Heat destruction of amino acids in soybean products used for the fermented soybean foods such amino acid composition of soy- bean products. *Japan Agricultural Research Quarterly (JARQ)*, 7(4): 267–273.
- Tesfaye, T., Sithole, B., Ramjugernath, D. and Chunilall, V. (2017a). Valorisation of chicken feathers: Characterisation of chemical properties. *Waste Management*, 68: 626-635 http://doi.org/10.1016/j.jclepro.2017.02.112
- Tesfaye, T., Sithole, B., Ramjugernath, D. and Chunilall, V. (2017b). Valorisation of chicken feathers: Characterisation of physical properties and morphological structure. *Journal of Cleaner Production*, 149: 349–365. http://doi.org/10.1016/j.jclepro.2017.02.112
- Thyagarajan, D., Barathi, M. and Sakthivadivu, R. (2013). Scope of poultry waste utilization. *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)*, 6(5): 29–35.
- Tiwary, E. and Gupta, R. (2012). Rapid conversion of chicken feather to feather meal using dimeric keratinase from *Bacillus licheniformis* ER-15. *Journal of Bioprocessing & Biotechniques*, 2(4): 1–4. http://doi.org/10.4172/2155-9821.1000123
- Tonkova, E.V., Gousterova, A. and Neshev, G. (2009). Ecologically safe method for improved feather wastes biodegradation. *International Biodeterioration & Biodegradation*, 61: 1008–1012.
- Tork, S., Aly, M.M. and Nawar, L. (2010). Biochemical and molecular characterization of a new local keratinase producing *Pseudomomanas* sp., MS21.

Asian Journal of Biotechnology, 2(1): 1–13. http://doi.org/10.3923/ajbkr.2010.1.13

- Tuna, A., Okumuş, Y., Çelebi, H and Seyhan, A.T. (2015). Thermochemical conversion of poultry chicken feather fibers of different colors into microporous fibers. *Journal of Analytical and Applied Pyrolysis*, 115: 112–124. http://doi.org/10.1016/j.jaap.2015.07.008
- Ullah, A., Vasanthan, T., Bressler, D., Elias, A.L. and Wu, J. (2011). Bioplastics from feather quill. *Biomacromolecules*, 12(10): 3826–3832. http://doi.org/10.1021/bm201112n
- Umar, A., Abdullahi, I. and Aliyu, A. (2015). Development and performance evaluation of chicken feather - plastic composite particle board. In *International Engineering Conference*. pp. 445–451. Retrieved from http://http//dspace.futminna.edu.ng/jspui/handle/1/4429
- United States Department of Agriculture (2017). Malaysia broiler meat (poultry) domestic consumption by year. Retrieved September 26, 2017, from http://www.indexmundi.com/agriculture/?country=my&commodity=broilermea t&graph=domestic-consumption
- Voet, D. and Voet, J. (1995). Proteins: Three-dimensional structure. In *Biochemistry*, 2nd ed. pp. 132–134.
- Wahab, A.G. and Rittgers, C. (2014). Broiler Meat Sector, Malaysia. USDA Foreign Agriculture Service, 4. Retrieved September 13, 2017, from http://www.thefarmsite.com/reports/contents/MalaysiaPoultry14March2014.pdf
- Wang, X. and Parson, C. (1997). Effect of processing systems on protein quality of feather meal and hog hair meals. *Poultry Science*, 76: 491–496.
- Wang, J., Hao, S.L., Luo, T.T., Cheng, Z.J., Li, W.F., Gao, F.Y., Guo, T.W., Gong, Y.H. and Wang, B.C. (2017). Feather keratin hydrogel for wound repair: Preparation, healing effect and biocompatibility evaluation. *Colloids and Surfaces B: Biointerfaces*, 149: 341–350. http://doi.org/10.1016/j.colsurfb.2016.10.038
- Wang, J., Hao, S.L, Luo, T.T., Yang, Q. and Wang, B.C. (2016). Development of feather keratin nanoparticles and investigation of their hemostatic efficacy. *Materials Science and Engineering: C*, 68: 768–773. http://doi.org/10.1016/j.msec.2016.07.035
- Wanitwattanarumlug, B., Luengnaruemitchai, A. and Wongkasemjit, S. (2012). Characterization of corn cobs from microwave and potassium hydroxide pretreatment. *International Scholarly and Scientific Research & Innovation*, 6(4): 327-331.

Weidele, T. (2009). Methods for using biomass in biogas process. Patent, US7781194

B2.

- Winandy, J.E., Muehl, J.H., Glaeser, J.A. and Schmidt, W. (2007). Chicken feather fiber as an additive in MDF composites. *Journal of Natural Fibers*, 4(1): 35–48. http://doi.org/10.1300/J395v04n01
- Wu, G. (2013). Amino acid: Biochemistry and Nutrition. CRC Press.
- Yahaya, N.S., Lim, C.S., Taha, M.R. and Jacqueline, J. (2016). Exposure of municipal solid waste disposal sites to climate related geohazards: Case study of Selangor. *Bulletin of the Geological Society of Malaysia*, 62: 57–63.
- Yusuf, I. (2015). Biodegradation of highly recalcitrant chicken feather wastes by locally isolated multi heavy metal tolerant bacterium-*Alcalegenes* sp. strain AQ05-001. PhD Thesis. Universiti Putra Malaysia.
- Zakaria, M.R., Hirata, S. and Hassan, M.A. (2015). Hydrothermal pretreatment enhanced enzymatic hydrolysis and glucose production from oil palm biomass. *Bioresource Technology*, 176: 142–148. http://doi.org/10.1016/j.biortech.2014.11.027
- Zhao, W., Yang, R.J., Zhanga, Y.Q. and Wua, L. (2012). Sustainable and practical utilization of feather keratin by an innovative physicochemical pretreatment: high density steam flash-explosion. *Green Chemistry*, 14: 3352–3360. http://doi.org/10.1039/C2GC36243K