

UNIVERSITI PUTRA MALAYSIA

In Vivo EFFECTS OF COPPER ON FISH TOXICITY AND CHOLINESTERASE OF Oreochromis mossambicus (W. K. H. PETERS, 1852) (BLACK TILAPIA)

AIN AQILAH BINTI BASIRUN

FBSB 2018 17



In Vivo EFFECTS OF COPPER ON FISH TOXICITY AND CHOLINESTERASE OF Oreochromis mossambicus (W. K. H. PETERS, 1852) (BLACK TILAPIA)



AIN AQILAH BINTI BASIRUN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

December 2017

COPYRIGHT

All materials contained within the thesis including without limitation text, logos, icons, photographs and all other artworks are copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from copyright holder. Commercial use of materials may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to my beloved family.



 \bigcirc

Abstract of thesis presented to Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Master of Science

In Vivo EFFECTS OF COPPER ON FISH TOXICITY AND CHOLINESTERASE OF Oreochromis mossambicus (W. K. H. PETERS, 1852) (BLACK TILAPIA)

By

AIN AQILAH BINTI BASIRUN

December 2017

Chairman: Siti Aqlima Binti Ahmad, PhD Faculty : Biotechnology and Biomolecular Sciences

Heavy metals including copper (Cu) has recently become an overwhelming pollutant towards the environment especially aquatic system. Many current researchers are focusing on cholinesterase (ChE) for biomarker and biosensor development as preliminary screening to prove the existence of xenobiotic in the aquatic system. In this study, an inhibitive assay for Cu was developed using the partially purified fraction of ChE from Oreochromis mossambicus. In addition, the biochemical, morphology and histopathology changes of O. mossambicus were observed as the biomarker for Cu exposure in vivo method. Five selected organs namely brain, blood, gills, liver and muscle depicted the alterations upon 96 h sub-acute exposure of CuSO₄. Common anomalies observed include the karyohexis and keryolysis in brain cell strutures, gills hyperplasia, melano macrophage centre (MMC) and hemosiderin formation in liver, blood cell alterations, massive formation of macrophagic cell in blood system and degeneration of muscle bundle in muscle. CuSO₄ has also inhibited ChE in in vivo analysis. ChE from five selected organs was inhibited starting from the concentration of 5 mg/L CuSO₄. Liver ChE showed the fluctuation of ChE inhibition. In vitro analysis took place where ChE of untreated O. mossambicus was partially purified through affinity chromatography using Procainamide-Sephacryl 6B as ligand. The folds of purification of ChE from brain, blood, gill, liver and muscle ChE were 5.8, 4.9, 3.6, 7.2, and 3.5, respectively. The optimisation of all ChEs were studied. Substrate specificity has specified ChE extracted from all five organs, which are brain (ATC 3.0 mM), blood (PTC 3.0 mM), gill (ATC 2.0 mM), liver (BTC 2.0 mM) and muscle (PTC 2.5 mM). The optimum pH and temperature studies of those organs recorded brain (pH 9, 20°C), blood (pH 9, 40°C), gill (pH 8, 30°C), liver (pH 9, 30°C), and muscle (pH 9, 30°C). In this case, the optimisation of ChEs from five O. mossambicus organs was not much different with each organ. Half maximal inhibitory concentration (IC_{50}) was studied to determine the Cu potency toward inhibiting O. mossambicus ChEs. IC₅₀ of Cu on brain, blood, gill, liver and muscle were 2.65, 0.297, 0.935, 7.66 and 10.58 mg/L. IC₅₀ categorised the blood PrChE of O. mossambicus as very sensitive and suitable to be used as biosensor for Cu pollution monitoring. Those alterations as well as cholinesterase inhibition of fish upon heavy metal exposure can contribute to

potential biomarker for aquatic pollution monitoring. Development of biomarker will work as preliminary screening of toxicants prior to second validation through high technology instruments followed by water rehabilitation.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

KESAN In Vivo LOGAM KUPRUM KE ATAS KETOKSIKAN IKAN DAN KOLINESTERASE DARI Oreochromis mossambicus (W. K. H. PETERS, 1852) (TILAPIA HITAM)

Oleh

AIN AQILAH BINTI BASIRUN

Disember 2017

Pengerusi : Siti Aqlima Binti Ahmad, PhD Fakulti : Bioteknologi dan Sains Biomolekul

Logam berat seperti kuprum (Cu) pada masa kini telah menjadi bahan toksik yang mencemarkan alam sekitar terutama sistem akuatik. Kajian terkini lebih tertumpu kepada kolinesterase (ChE) untuk perkembangan biopenanda dan biosensor sebagai penyaringan awal untuk mengesan kewujudan bendasing dalam sistem akuatik. Dalam kajian ini, ujian perencatan untuk Cu telah dijalankan dengan menggunakan sample hasil daripada separa penulenan ChE dari *Oreochromis mossambicus*. Di samping itu, perubahan biokimia, morfologi, dan histopatologi pada Oreochromis mossambicus dikaji untuk dijadikan sebagai biopenanda untuk pencemaran Cu dalam kaedah in vivo. Lima organ yang dipilih ialah otak, darah, insang, hati, dan otot ikan menunjukkan perubahan pada pendedahan separa-akut CuSO₄ selama 96 jam. Beberapa perubahan yang terdapat di dalamnya termasuk karyorrhexis dan karyolisis dalam struktur sel otak, hiperplasia, melano macrophage centre (MMC) dan pembentukan hemosiderin dalam sel hati, perubahan sel darah, pembentukan sel makrofaj secara besar-besaran dalam sistem darah dan kerosakan dalam tisu otot. CuSO4 juga merencat ChE dalam analisis in vivo. ChE daripada lima organ yang dipilih terencat bermula dari kepekatan 5mg/L CuSO₄. ChE daripada hati menunjukkan turun naik perencatan ChE. Analisis in vitro berlaku di mana ChE daripada O. mossambicus yang tidak terdedah kepada Cu diasing dan disepara tulenkan melalui kromatografi afinasi menggunakan Procainamide-Sephacryl 6B sebagai ligan. Faktor penulenan ChE dari otak, darah, insang, hati, dan otot ChE masing-masing adalah 5.8, 4.9, 3.6, 7.2 dan 3.5. Pengoptimuman semua ChE telah dikaji dan substrat yang spesifik telah menunjukkan ChE yang diekstrak dari semua organ termasuk otak (ATC 3.0 mM), darah (PTC 3.0 mM), insang (ATC 2.0 mM), hati (BTC 2.0 mM), dan otot (PTC 2.5 mM). Kajian pH dan suhu optimum organ-organ tersebut mencatatkan nilai; otak (pH 9, 20°C), darah (pH 9, 40°C), insang (pH 8, 30°C), hati (pH 9, 30°C) dan (pH 9, 30°C). Dalam kes ini, pengoptimuman ChEs daripada organ O. mossambicus tidak banyak berbeza di antara setiap organ. Kepekatan separa perencatan (IC₅₀) dikaji untuk menentukan potensi Cu ke arah merencat O. mossambicus ChEs. IC₅₀ pada otak, darah, insang, hati, dan otot masing-masing adalah 2.65, 0.297, 0.935, 7.66, 10.58 mg/L, IC₅₀ mengkategorikan darah PrChE of O. mossambicus sangat sensitif sebagai biosensor untuk pemantauan pencemaran Cu. Perubahan serta perencatan terhadap kolinesteres ikan apabila

didedahkan logam berat menyumbang kepada biopenanda yang berpotensi untuk pemantauan pencemaran akuatik. Perkembangan biopenanda akan berfungsi sebagai pemeriksaan awal toksik sebelum pengesahan kedua melalui instrumen teknologi tinggi dan diikuti dengan pemulihan air.



G

ACKNOWLEDGEMENTS

Bismillahi Rahmanir Rahim.

First and foremost, I am very grateful to the Almighty Allah for His guidance and mercy for me to finish my Master Degree.

I would like to express my deepest gratitude to my supervisor, Dr. Siti Aqlima Ahmad for her intellectual vigour and generous support and patience in guiding me along this wonderful research journey. I would also like to thank to all my co-supervisors, Assoc. Dr. Mohd Yunus Abd Shukor, Dr. Nur Adeela Yasid, Assoc. Prof. Dr. Hassan Daud and Dr. Mohd Khalizan Sabullah. It has been a greatly enriching experience to work under their authorative guidance.

My warmest thanks goes to my supporting pills, which are my friends, Shakirah Abdul Wahab Sha'arani, Siti Nadzirah Padrilah, Nur Muhammad Syahir Abdul Habib, Motharasan Manogaran, and all members of Bioremediation Lab especially Abubakar Aisami, Ibrahim Allamin, Umar Abubakar, Hafeez Yakasai, Fadhil Rahman for their support and advices throughout this research journey. Special thanks to academics staff and laboratory assistances at the Department of Biochemistry, Institute Biosience and Faculty of Veterinary Medicine for providing a good environment and facilities.

It is my radiant sentiment to wish million appreciations to my beloved parent, Basirun Razak and Norleha Bakar and my siblings, Amirah Asyikin, Anis Hidayah and Muhamad Adib who never desist to inspire and encourage me with their massive words and blessings.

All in all, each and every hard work applied, none of it was from us without the blessing from Almighty Allah. Alhamdulillah!

Ain Aqilah binti Basirun, 2017.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Siti Aqlima Ahmad, PhD

Senior Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairperson)

Mohd Yunus Shukor, Ph.D

Associate Professor Faculty Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Nur Adeela Yasid, PhD

Senior Lecturer Faculty of Biotechnology Universiti Putra Malaysia (Member)

Mohd Khalizan Sabullah, PhD

Senior Lecturer Faculty of Science and Natural Resources Universiti Malaysia Sabah (Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date

Declaration by graduate student

I hereby confirm that:

- this thesis is my origical work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writing, seminar paper, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:

Name and Matric No.: Ain Aqilah binti Basirun, GS43678

Declaration by Members of Supervisory committee

This is to certify that:

- the research conducted and the writing of the thesis was under our supervision
- supervision of responsibilities slated in rule 41 in rules 2003 (revision 2012 2013) were adhered to.

Signature: ______ Dr. Siti Aqlima Ahmad ______ Signature: ______ Name of Member of Supervisory

> Associate Professor Mohd Yunus Abd Shukor

Signature: Name of Member of Supervisory Committee:

Committee:

Dr. Nur Adeela Yasid

Signature: Name of Member of Supervisory Committee:

Dr. Mohd Khalizan Sabullah

TABLE OF CONTENTS

Al Al Al Dl Ll Ll Ll	BSTR BSTR CKN(PPRO ECLA ST O ST O ST O HAPT	ACT AK OWLEDG OVAL ARATION F TABLE F FIGUR F ABBRE	EMENTS S ES EVIATIONS			Page i iii v vi viii xiii xv xix
1	INT	RODUCT	TION			1
2		ERATUR	E REVIEW			
	2.1	Heavy m	ietals.			4
		2.1.1	Heavy met	als contamination	status in Malaysian	5
	22	Conner	aquatic syste	·m.		6
	2.2	Toxicity	of Cu towards	aquatic system.		8
	2.4	Fish as b	iomarker for C	u and other recalcitra	ant in aquatic system.	9
		2.4.1	Histopatholo	gical study of fis	s <mark>h org</mark> ans upon Cu	10
		2.4.2	exposure.			10
	25	2.4.2 Cholines	Biochemical	alteration of fish upo	on Cu exposure.	13
	2.5	2.5.1	Cholinestera	se as biomarker and l	biosensor	14
	2.6	Oreochr	omis mossambi	<i>icus</i> (black tilapia) as	s a model of study.	19
3	MA	TERIALS	S AND METH	ODS		21
	3.1	Material	Chamical			21
		3.1.1	Equipment			21
		3.1.2	Species of fr	eshwater as a subject		21
	3.2	Method	1	5		
		3.2.1	Fish acclima	tisation and stabilisat	tion.	22
		3.2.2	Acute toxicit	ty exposure of copper	r sulphate.	23
		3.2.3	Histology stu	idy of O. mossambic	us exposed by Cu.	23
			3.2.3.1	Histology study	by light inverted	23
			3232	Histology study	v by transmission	24
			5.2.5.2	electron microsco	pe.	21
			3.2.3.3	Histology study	by scanning electron	24
				microscope.	-	
		3.2.4	Cholinestera	se extraction from O.	mossambicus.	24
		3.2.5	Enzyme acti	vity determination.		25
		3.2.6	Protein conte	of <i>Q</i> magazitic	us ChE hy officity	26
		5.2.1	chromatogra	phy.	us Chill by affility	20

6

		3.2.7.1	Preparation of procainamide sephacryl-	26
		3777	oB resin. ChE purification using Proceinamida	26
		3.2.1.2	Senhacryl 6B as resin	20
	328	SDS-PAGE	Sephaeryr ob as resni.	27
	329	ChE optimis	ation study	28
	5.2.7	3 2 9 1	Substrate specificity	28
		3292	nH profile	20
		3293	Temperature profile	29
	3 2 10	Half maxima	al inhibitory concentration (IC $_{50}$) study of	29
	5.2.10	Cii on <i>O</i> mo	ssambicus	25
	3.2.11	Statisticl anal	lysis	29
	0.2.11		- , 0.15	
4 RES	SULTS AN	D DISCUSSI	ONS	
4.1	Behaviou	ral and physic	ological changes of O. mossambicus upon	30
	Cu expos	ure.		
	4.1.1	Behavioural	and physiological study.	30
	4.1.2	Fish surviva	1 and LC ₅₀ study upon exposure of CuSO ₄ .	33
4.2	Histologi	cal changes of	O. mossambicus upon Cu exposure.	
	4.2.1	Organ sectio	onal image observation under LM.	34
		4.2.1.1	Brain.	34
		4.2.1.2	Gills.	36
		4.2.1.3	Liver.	38
		4.2.1.4	Muscle.	40
	4.2.2	Organ section	onal image observation under SEM.	
		4.2.2.1	Brain.	42
		4.2.2.2	Blood.	44
		4.2.2.3	Gills.	46
		4.2.2.4	Liver.	48
		4.2.2.5	Muscle.	50
	4.2.3	Organ section	onal image observation under TEM.	
		4.2.3.1	Brain.	52
		4.2.3.2	Blood.	54
		4.2.3.3	Gills.	56
		4.2.3.4	Liver.	58
		4.2.3.5	Muscle.	60
4.3	Enzyme a	activity determ	ination.	
	4.3.1	<i>In vivo</i> inhib	biton study.	62
		4.3.1.1	Brain.	62
		4.3.1.2	Blood.	64
		4.3.1.3	Gills.	66
		4.3.1.4	Liver.	69
		4.3.1.5	Muscle.	71
	4.3.2	In vitro anal	ysis	73
	4.3.2.1	Purification	of ChE from untreated O. mossambicus	73
		through a	affinity chromatography by using	
		Procanamide	e- Sepnacryl 6B as resin	72
		4.3.2.1.1	Brain.	15
		4.3.2.1.2	В100 0 .	/0
		4.3.2.1.3		17 07
		4.3.2.1.4	Liver.	ð2

		4.3.2.1.5 Muscle.	85
	4.3.2.2	ChE optimisation study.	88
	4.3.2.2.1	Substrate specificity study	88
	4.3.2.2.2	pH profile.	96
	4.3.2.2.3	Temperature.	101
	4.3.2.2.4	Half maximal inhibitory concentration (IC ₅₀) study of	106
		Cu on O. mossambicus	
4.4	Overall su	mmary	112

5 CONCLUSION

 (\mathcal{G})

REFERENCES APPENDICES BIODATA OF STUDENT LIST OF PUBLICATION 116 117

133 139

140

LIST OF TABLES

Table		Page
2.1	Guideline of water usage	5
2.2	Estimated categorizes of metals ion toxicity in a descending order	9
2.3	Bioaccumulation of heavy metal in several aquatic organism species caught at Juru River, Penang Malaysia	11
2.4	Effects on heavy metals exposure on gill, liver, and kidney from various fish species	12
2.5	List of ChE from various sources as biomarker candidate for Ecotoxicology monitoring	19
2.6	Taxonomy of <i>Oreochromis mossambicus</i>	20
3.1	Solution mixtures for stacking and resolving gels of SDS- PAGE	27
4.1	Behavioural changes and morphological deformities of <i>O.</i> <i>mossambicus</i> upon exposure of different concentration of CuSO ₄	31
4.2	Comparison between extraction and purification method of <i>O</i> . <i>mossambicus</i> ChE brain.	74
4.3	Comparison between extraction and purification method of <i>O</i> . <i>mossambicus</i> ChE blood.	77
4.4	Comparison between extraction and purification method of <i>O</i> . <i>mossambicus</i> ChE gills.	80
4.5	Comparison between extraction and purification method of <i>O</i> . <i>mossambicus</i> ChE liver.	83
4.6	Comparison between extraction and purification method of <i>O</i> . <i>mossambicus</i> ChE muscle.	86
4.7	Maximal velocity, V_{max} and Michaelis-Menten constant, K_m values of three synthetic substrates on partially purified ChE from brain extract of <i>O. mossambicus</i> to study its substrate specificity properties.	90
4.8	Maximal velocity, V_{max} and Michaelis-Menten constant, K_m values of three synthetic substrates on partially purified ChE from blood extract of <i>O. mossambicus</i> to study its substrate specificity properties.	91
4.9	Maximal velocity, V_{max} and Michaelis-Menten constant, K_m values of three synthetic substrates on partially purified ChE from gills extract of <i>O. mossambicus</i> to study its substrate specificity properties.	93
4.10	Maximal velocity, V_{max} and Michaelis-Menten constant, K_{m} values of three synthetic substrates on partially purified ChE	94

from liver extract of *O. mossambicus* to study its substrate specificity properties.

- 4.11 Maximal velocity, V_{max} and Michaelis-Menten constant, K_m
 96 values of three synthetic substrates on partially purified ChE from muscle extract of *O. mossambicus* to study its substrate specificity properties.
- 4.12 Summary of behavioural and physiological changes of *O*. 112 *mossambicus* affected by CuSO₄.
- 4.13 Summary of histopathological alterations and ChE of *O*. 113 *mossambicus* brain exposed by CuSO₄.
- 4.14 Summary of histopathological alterations and ChE of *O*. 113 *mossambicus* blood exposed by CuSO₄.
- 4.15 Summary of histopathological alterations and ChE of *O*. 114 *mossambicus* gills exposed by CuSO₄.
- 4.16 Summary of histopathological and ChE alterations of *O*. 114 *mossambicus* liver exposed by CuSO₄.
- 4.17 Summary of histopathological and ChE alterations of *O*. 115 *mossambicus* muscle exposed by CuSO₄.
- 4.18 Summary of *in vitro* study of ChE extracted from *O*. 115 *mossambicus*.

LIST OF FIGURES

Figure		Page
2.1	The entrance of Cu via ingestion or from secretory fluid transported into the liver, followed by excretion via urinary or faeces.	7
2.2	The ultrastructure of hepatocyte of Puntius javanicus.	13
2.3	The schematic illustration of AChE active site	15
2.4	Cholinergic synapse	16
2.5	Schematic diagram of biomarker implementation.	17
3.1	O. mossambicus as a model of the study.	22
4.1	The effect of CuSO ₄ concentration on the survival percentage (%) of <i>O.mossambicus</i> after 96-hour of sub-acute toxicity.	33
4.2	The representative section images of <i>O. mossambicus</i> brain control and exposed to CuSO ₄ at different concentrations under H&E staining	35
4.3	The representative section images of <i>O. mossambicus</i> gills control and exposed to CuSO ₄ at different concentrations under H&E staining.	37
4.4	The representative section images of <i>O. mossambicus</i> liver control and exposed by CuSO ₄ with different concentration under H&E staining.	39
4.5	The representative section images of <i>O. mossambicus</i> muscle control and exposed by CuSO ₄ with different concentration under H&E staining.	41
4.6	The representative section images of the surface of O . mossambicus mid brain (mesencephalon) control and exposed by CuSO ₄ with different concentration under visualisation SEM.	43
4.7	The representative section images of the normal <i>O</i> . <i>mossambicus</i> blood and exposed by CuSO ₄ with different concentration under visualisation SEM.	45
4.8	The representative section images of <i>O. mossambicus</i> gills control and exposed to $CuSO_4$ at different concentrations under visualisation SEM.	47
4.9	The representative section images of <i>O. mossambicus</i> liver control and exposed by CuSO ₄ with different concentration under visualisation SEM.	49
4.10	The representative section images of O . mossambicus muscle control and exposed by CuSO ₄ with different concentration under visualisation SEM.	51

4.11	Ultrastructure of <i>O. mossambicus</i> brain control and exposed by CuSO ₄ with different concentration under TEM visualisation.	53
4.12	Ultrastructure of normal O . mossambicus blood cell and exposed by CuSO ₄ with different concentration under TEM visualisation.	55
4.13	Ultrastructure of O . mossambicus gill tissue control and exposed to CuSO ₄ at different concentrations under TEM visualisation.	57
4.14	Ultrastructure of O . mossambicus liver tissue control and exposed by CuSO ₄ with different concentration under TEM visualisation.	59
4.15	Ultrastructure of <i>O. mossambicus</i> muscle tissue control and exposed by CuSO ₄ with different concentration under TEM visualisation.	60
4.16	Enzyme activity of untreated (control) brain of <i>O. mossambicus</i> hydrolysed three substrates, ATC, BTC, and PTC.	63
4.17	ChE inhibition study of brain of O. mossambicus treated by CuSO ₄ with concentration of 2.5, 5.0, 10.0 and 20.0 mg/L.	64
4.18	Enzyme activity of untreated (control) blood of <i>O. mossambicus</i> hydrolysed three substrates, ATC, BTC, and PTC.	65
4.19	ChE inhibition study of blood of <i>O. mossambicus</i> treated by CuSO ₄ with concentration of 2.5, 5.0, 10.0 and 20.0 mg/L.	66
4.20	Enzyme activity of untreated (control) gills of <i>O. mossambicus</i> hydrolysed three substrates, ATC, BTC, and PTC.	67
4.21	ChE inhibition study of gills of <i>O. mossambicus</i> treated by $CuSO_4$ with concentration of 2.5, 5.0, 10.0 and 20.0 mg/L.	68
4.22	Enzyme activity of untreated (control) liver of <i>O. mossambicus</i> hydrolysed three substrates, ATC, BTC, and PTC.	69
4.23	ChE inhibition study of liver of <i>O. mossambicus</i> treated by CuSO ₄ with concentration of 2.5, 5.0, 10.0 and 20.0 mg/L.	70
4.24	Enzyme activity of untreated (control) muscle of <i>O. mossambicus</i> hydrolysed three substrates, ATC, BTC, and PTC.	71
4.25	ChE inhibition study of muscle of <i>O. mossambicus</i> treated by CuSO ₄ with concentration of 2.5, 5.0, 10.0 and 20.0 mg/L.	72
4.26	Profile of purified ChE from brain extract of <i>O. mossambicus</i> on Procainamide–Sephacryl 6B affinity column.	74
4.27	SDS-PAGE of partially purified ChE from the brain of <i>O</i> . <i>mossambicus</i> in a 12% polyacrylamide gel.	75
4.28	Purified ChE from <i>O. mossambicus</i> brain was detected based on broad protein range standard curve at 41.36 kDa.	76

G

4.29	Profile of purified ChE from blood extract of <i>O. mossambicus</i> on Procainamide–Sephacryl 6B affinity column.	77
4.30	SDS-PAGE of partially purified ChE from the blood of <i>O</i> . <i>mossambicus</i> in a 12% polyacrylamide gel.	78
4.31	Partially purified ChE from <i>O. mossambicus</i> blood were detected based on broad protein range standard curve at 147.44 kDA, 114.14 kDa, and 62.46 kDa.	79
4.32	Profile of purified ChE from gills extract of <i>O. mossambicus</i> on Procainamide–Sephacryl 6B affinity column.	80
4.33	SDS-PAGE of partially purified ChE from the gills of <i>O</i> . <i>mossambicus</i> in a 12% polyacrylamide gel.	81
4.34	Partially purified ChE from <i>O. mossambicus</i> gills were detected based on broad protein range standard curve at 79.55 kDa and 62.46 kDa.	82
4.35	Profile of purified ChE from liver extract of <i>O. mossambicus</i> on Procainamide–Sephacryl 6B affinity column.	83
4.36	SDS-PAGE of partially purified ChE from the liver of <i>O</i> . <i>mossambicus</i> in a 12% polyacrylamide gel.	84
4.37	Partially purified ChE from <i>O. mossambicus</i> liver were detected based on broad protein range standard curve at 140.59 kDa and 93.57 kDa.	85
4.38	Profile of purified ChE from muscle extract of <i>O. mossambicus</i> on Procainamide–Sephacryl 6B affinity column.	86
4.39	SDS-PAGE of partially purified ChE from the muscle of <i>O</i> . <i>mossambicus</i> in a 12% polyacrylamide gel.	87
4.40	Partially purified ChE from <i>O. mossambicus</i> muscle were detected based on broad protein range standard curve at 34.111 kDa, 22.87 kDa and 13.45 kDa.	88
4.41	Michaelis-Menten plot of <i>O. mossamibicus</i> ChE brain incubated with different synthetic substrates.	89
4.42	Michaelis-Menten plot of <i>O. mossamibicus</i> ChE blood incubated with different synthetic substrates.	91
4.43	Michaelis-Menten plot of <i>O. mossamibicus</i> ChE gills incubated with different synthetic substrates.	92
4.44	Michaelis-Menten plot of <i>O. mossamibicus</i> ChE liver incubated with different synthetic substrates.	93
4.45	Michaelis-Menten plot of <i>O. mossamibicus</i> ChE muscle incubated with different synthetic substrates.	94
4.46	Optimisation studies of pH for the ChE from <i>O. mossambicus</i> brain using three different buffers.	95

G

4.47	Optimisation studies of pH for the ChE from <i>O. mossambicus</i> blood using three different buffers.	98
4.48	Optimisation studies of pH for the ChE from <i>O. mossambicus</i> gills using three different buffers.	99
4.49	Optimisation studies of pH for the ChE from <i>O. mossambicus</i> liver using three different buffers.	100
4.50	Optimisation studies of pH for the ChE from <i>O. mossambicus</i> muscle using three different buffers.	101
4.51	Optimisation of temperature for ChE from <i>O. mossambicus</i> brain.	102
4.52	Optimisation of temperature for ChE from <i>O. mossambicus</i> blood.	103
4.53	Optimisation of temperature for ChE from <i>O. mossambicus</i> gills.	104
4.54	Optimization of temperature for ChE from <i>O. mossambicus</i> liver.	105
4.55	Optimization of temperature for ChE from <i>O. mossambicus</i> muscle.	106
4.56	Percentage inhibition of brain ChE by Cu with series of concentrations (1 to 10 mg/L).	107
4.57	Percentage inhibition of blood ChE by Cu with series of concentrations (1 to 10 mg/L).	108
4.58	Percentage inhibition of gill ChE by Cu with series of concentrations (1 to 10 mg/L).	109
4.59	Percentage inhibition of liver ChE by Cu with series of concentrations (1 to 10 mg/L).	110
4.60	Percentage inhibition of muscle ChE by Cu with series of concentrations (1 to 10 mg/L)	111

 \bigcirc

LIST OF ABBREVIATIONS

%	Percent
<	Less than
>	Greater than
μL	Microlitre
μm	Micrometre
μΜ	Micromolar
Abs	Absorbance
Ach	Acetycholine
AChE	Acetylcholinesterase
Ag	Silver
Al	Aluminium
ALAT	Alanine transferase
ANOVA	Analysis of variance
APS	Aluminum persulphate
ARD	Acid rock drainage
As	Arsenic
ASAT	Aspirate amino transferase
ASP	Aspartate
ATC	Acetythiocholine iodide
ATPase	Adenosine Tryphosphatase
Au	Gold
Ba	Barium
BCh	Butyrylcholine
BChE	Butyrylcholinesterase
BSA	Bovine serum albumin
BTC	Butyrylthiocholine iodide
Ca	Calcium
CAT	Catalase
Cd	Cadmium
ChE	Cholinesterase
Cm	Centimetre
CNS	Central nervouse system
Co	Cobalt
Cr	Chromium
Cu	Copper
Cu ²⁺	Copper ion
CuO	Copper oxide
CuO ₂	Copper dioxide
CuSO ₄	Copper sulphate
Da	Dalton
dH ₂ O	Distilled water
DO	Dissolved oxygen
DTNB	5,5-dithio-bis-2-nitrobenzoate
EDTA	Ethylene diamine tetra acetic acid
EDX	Energy dispersive X-ray
et al	and friends
FAO	Food and Agricultural Organisation

0

Fe	Iron
g	Gram
GL	Granular layer
Glu	Glutamine
GST	Gluthathione S-tranferase
h	Hour
H&E	Hematoxylin and Eosin
Hg	Mercury
HgCl ₂	Mercury chloride
His	Histidine
IC ₅₀	Half maximal inhibitory concentration
K	Potassium
kb	Kilo base
kDa	Kilo Dalton
Kg	Kilogram
L	Litre
 LC ₅₀	Half maximal lethal concentration
Li	Lithium
IM	Light inverted microscope
LOFC	Low observed effects of concentration
M	Molar
MB	Muscle hundle
Mo	Magnesium
mg	Miligram
min	Minute
mL	Mililitre
MMC	Melano-macrophage centre
Mn	Manganese
MT	Metallothionine
MW	Molecular weight
Na	Sodium
NaOH	Sodium hydoxide
Ni	Nickel
°C	Degree Celsius
OD	Optical density
OP	Organophosphate
Pb	Lead
Phe	Phenylalanine
PL	Primary lamella
PNS	Peripheral nervous system
PrCh	Propionylcholine
PrChE	Propionylcholinesterase
Pt	Platinum
PTC	Propionylthiocholine iodide
RBC	Red blood cell
R _f	Retention factor
RNA	Ribosomal nucleic acid
ROS	Reactive oxygen species
RPM	Rotation per minute
SDS	Sodium dodecyl sulphate

SDS-PAGE	Sodium dodecyl sulphate polyacrylamide gel				
	electrophoresis				
SEM	Scanning electron microscope				
Ser	Serine				
SH-	Sulfuhydryl				
SL	Secondary lamella				
Sn	Selenium				
SOD	Superoxide dismutase				
Sr	Srontium				
TEM	Transmission electron microscope				
Trp	Tryptophan				
Tyr	Tyrosine				
U	Enzyme unit				
V	Voltage				
Vmax	Maximum velocity				
Zn	Zinc				

 \bigcirc

CHAPTER 1

INTRODUCTION

The vital environmental pollutants are those that can accumulate and are persistent due to the chemical stability and poor biodegradability (Yousafzai et al., 2017). Heavy metals effluences have become a great concern due to their multifunction, toxicity and resistance to degradation as well as their potential of bioaccumulation (Naji et al., 2014). The transformation of heavy metals into persistent metallic composites can cause bioaccumulation in an organism's body, disrupt the biological food chain and eventually give several adverse effects to the ultimate consumer, which is humankind that utilises aquatic sources to live (El-Moselhy et al., 2014; Zhou et al., 2008). Previous studies reporting several freshwater systems in Malaysia including Klang River, Langat basin, and also Mamut River in East Malaysia, Sabah, have raised a concern on their level of contamination by heavy metals such as copper, cadmium, zinc and lead (Abalaka, 2015; Ali et al., 2014; Naji et al., 2014).

The occurrence of heavy metals spillage is due to misappropriate and inconsiderate disposal of metal-rich wastes from human activities such as rapid industrial development and urbanisation, abandoned mining site and mining activities as well as agricultural and deforestation (Ibemenuga, 2016). Heavy metals accumulation in aquatic systems can affect the sustainability and productivity of aquatic organisms (Javed and Usmani, 2016; Sabullah et al., 2015a). In general, the accumulation of heavy metals in organism's tissues can enhace the production of reactive oxygen species (ROS) and eventually cause the biochemical, molecular and morphological alterations (ElGazzar et al., 2014). Heavy metals such as copper (Cu), zinc (Zn) and cadmium (Cd) are the vast metal ions those were excessively accumulate in water.

Copper is among the heavy metals that can be naturally found in several forms like copper ion (Cu^{2+}) and copper sulphate pentahydrate ($CuSO_4.5H_20$) or known as "bluestone" (Yanong, 2009). Copper plays crucial role in several integral parts of enzymes regarding respiration, collagen synthesis and to reduce free radicals (Acosta et al., 2016). Organisms require small amount of Cu to regulate body metabolism. According to Mashifane and Moyo (2014), several Cu compounds were effectively utilised in water treatment as they preserve the discolouration of water and monitor or eliminate algae development and fish parasites in freshwater and marine systems.

However, massive usage of Cu can lead to Cu pollution that would give several negative influences to the aquatic living systems. One of the causes of Cu exposure into stream systems is the abandoned copper mining site. For instance, extreme accumulation of Cu in Malaysia has become alarming especially in Mamut River near Ranau, Sabah. Copper pollution has occurred in the headwater of Mamut River due to an open pit Cu mine operated since 1975 and ceased its operation in 1999. Beyond this time range, this mine became a source of heavy metals accompanied by the increase in number of environmental problems beyond Ranau areas. The main source of heavy

metals was originated from the runoff of mine site, in addition to the floatation process used in preparing Cu concentrates (Ali et al., 2014).

Copper is tremendously toxic to aquatic life (Ezeonyejiaku et al., 2011). Cu exposure can generally disrupt the neural processes, protein function and chemosensory abilities (Dew et al., 2012). Consequences of Cu poisoning include several organ defects to organisms. As such, Cu can encourage larval mortality that will endanger the productivity of aquatic living systems, movement limitation of organisms in their habitat and cells degeneration (Gandhewar et al., 2012; Sabullah et al., 2015a). Meanwhile, exposure and accumulation of Cu toward aquatic organisms especially fish would give substantial effects. Physiological and histopathological alterations of fish upon exposure would aid in clarifying the health status of the fish as the food sources to humans. Aquatic environment makes up the major part of the human resources and is said to be interrelated to human (Ballesteros et al., 2017). Hence, fish has become a sentinel species to be included as a tool of biomonitoring for preliminary assessment of the toxicant in the aquatic systems. It is a potential biomarker for monitoring the heavy metals including Cu in aquatic living system. Fish can be a good biomarker by referring several parts such as liver, gill, muscle and brain that change due to exposure. Gills are the primary organ exposed to toxicant exposure (Sabullah et al., 2014a). Fish biomarkers are necessary to track environmentally induced alterations in accessing the effects of xenobiotics especially Cu on fish (Authman, 2015).

Biomarkers are deliberated to be one of the most capable tools for ecotoxicological applications as they could offer an early recognition of noxious waste exposure and a primary indication of possible effects at advanced levels of biological organisation such as population and ecosystem effects (Quintaneiro et al., 2015). Biomarkers have been considered as the reliable tool to screen alterations in biological responses toward environmental threat ranging from molecular through cellular and physiological response to behavioural changes, which are correlated to the exposure of xenobiotic and highly toxic compounds (Kaviraj et al., 2014; Sabullah et al., 2015a). Apart from the morphology of organism, biochemical alterations can also present as pollution biomarker. Cholinesterase (ChE), a ubiquitous enzyme was selected as the best biomarker for heavy metal detection as the response of inhibition towards vast range of inhibitors was closely accompanied by a rise in mortality and survival of aquatic organism, which was impaired due to inhibition (Nunes, 2011).

In this study, *O. mossambicus* was chosen as a test subject as the previous study mentioned about the capability of this species as an alternative biomarker for detecting of selective heavy metals known as Cu using the behavioural and histological alterations as well as inhibitive assay of ChE activity (Sabullah et al., 2015a). Therefore, this study can be perhaps referred to as another basis for future application of this species, whether in aquaculture management and production or in environmental monitoring.

In response to that, the sublethal concentration of copper sulphate (CuSO₄) acute exposure in various concentrations and the behavioural response and physiological changes of the exposed *Oreochromis mossambicus* was done in this study as the most

sensitive indication of potential toxic effects. The use of local *Oreochromis* sp. is expected to increase the potential of this fish to become a sentinel species that permits the recognition of lower contamination level of heavy metals especially Cu. Therefore, the objectives of this study are:

- 1. To determine the physical, behavioural, and morphological changes of *O*. *mossambicus* inhibited by Cu using *in vivo* method.
- 2. To purify and characterise ChE activity from different organs of untreated *O. mossambicus*.
- 3. To examine half maximal inhibitory concentration (IC₅₀) of Cu that would give inhibit ChE extracted from different organs of untreated *O. mossambicus* through *in vitro* analysis.



REFERENCES

- Abalaka, S. E. (2015). Heavy metals bioaccumulation and histopathological changes in *Auchenoglanis occidentalis* fish from Tiga dam, Nigeria. *Journal of Environmental Health Science and Engineering*, 13. doi.org/10.1186/s40201-015-0222-y.
- Abdel-Khalek, A. A., Badran, S. R., & Marie, M. A. S. (2016). Toxicity evaluation of copper oxide bulk and nanoparticles in Nile tilapia, *Oreochromis niloticus*, using hematological, bioaccumulation and histological biomarkers. *Fish Physiology and Biochemistry*, 42(4), 1225–1236.
- Abhijith, B. D., Ramesh, M., & Poopal, R. K. (2016). Responses of metabolic and antioxidant enzymatic activities in gill, liver and plasma of *Catla catla* during methyl parathion exposure. *The Journal of Basic and Applied Zoology*, 77, 31–40.
- Acheampong, M. G., Dueño, D. E., Glover, B. K., Henry, A. A., Mata, R., VanBrakle, M. L., Westblade, L. F., Sussman, J. L., & Granberry, A. L. (2012).
 Acetylcholinesterase: Substrate traffic and inhibition. *Biochemistry and Molecular Biology Education*, 40(2), 144–144. doi.org/10.1002/bmb.20604.
- Acosta, D. da S., Danielle, N. M., Altenhofen, S., Luzardo, M. D., Costa, P. G., Bianchini, A., Bonan, C. D., Da Silva, R. S., & Dafre, A. L. (2016). Copper at low levels impairs memory of adult zebrafish (*Danio rerio*) and affects swimming performance of larvae. *Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology*, 185–186, 122–130.
- Adel, M., Oliveri Conti, G., Dadar, M., Mahjoub, M., Copat, C., & Ferrante, M. (2016). Heavy metal concentrations in edible muscle of whitecheek shark, *Carcharhinus dussumieri (elasmobranchii, chondrichthyes)* from the Persian Gulf: A food safety issue. *Food and Chemical Toxicology*, 97, 135–140.
- Agius, C., & Roberts, R. J. (2003). Melano-macrophage centres and their role in fish pathology. *Journal of Fish Diseases*, 26(9), 499–509.
- Ahmad, S. A., Sabullah, M., Basirun, A. A., Yasid, N. A., Iqbal, I. M., Shamaan, N. A., Syed, M. A., & Shukor, M. (2016c). Evaluation of cholinesterase from the muscle and blood of *Anabas testudineus* as detection of metal ion. *Fresenius Environmental Bulletin*, 25(10), 4253–4260.
- Ahmad, S. A., Sabullah, M. K., Shamaan, N. A., Shukor, M. Y. A., Jirangon, H., Khalid, A., & Syed, M. A. (2016a). Evaluation of acetylcholinesterase source from fish, *Tor tambroides* for detection of carbamate. *Journal of Environmental Biology*, *37*(4), 479–484.
- Ahmad, S. A., Wong, Y. F., Shukor, M. Y., Sabullah, M. K., Yasid, N. A., Hayat, N. M., Shamaan, N. A., Khalid, A., & Syed, M. A. (2016b). An alternative bioassay using *Anabas testudineus* (Climbing perch) colinesterase for metal ions detection. *International Food Research Journal*, 23(4), 1446–1452.
- Alam, L., Mokhtar, M. B., Alam, M., Bari, M., Kathijotes, N., Ta, G. C., & Ern, L. K. (2015). Assessment of environmental and human health risk for contamination of heavy metal in tilapia fish collected from Langat Basin, Malaysia. *Asian Journal of Water, Environment and Pollution*, 12(2), 21–30.
- Al-Bairuty, G. A., Boyle, D., Henry, T. B., & Handy, R. D. (2016). Sublethal effects of copper sulphate compared to copper nanoparticles in rainbow trout

(Oncorhynchus mykiss) at low pH: physiology and metal accumulation. Aquatic Toxicology, 174, 188–198.

- Allan, D., Billah, M. M., Finean, J. B., & Michell, R. H. (1976). Release of diacylglycerol-enriched vesicles from erythrocytes with increased intracellular (Ca²⁺). *Nature*, 261, 58-60.
- Ali, B. N. M., Lin, C. Y., Cleophas, F., Abdullah, M. H., & Musta, B. (2014). Assessment of heavy metals contamination in Mamut river sediments using sediment quality guidelines and geochemical indices. *Environmental Monitoring and Assessment*, 187(1), 4190–4201
- Alina, M., Azrina, A., Mohd Yunus, A. S., Mohd Zakiuddin, S., Mohd Izuan Effendi, H., & Muhammad Rizal, R. (2012). Heavy metals (mercury, arsenic, cadmium, plumbum) in selected marine fish and shellfish along the Straits of Malacca. *International Food Research Journal*, 19(1), 135–140.
- Al-Shafi, S. M. K., Al-Azzaw, M., & Ali, M. A. (2017). The Inhibitory effect of gallic acid on human serum cholinesterase. *Iraqi Journal of Pharmaceutical Sciences*, 18(1), 33–37.
- Amanah, S. N. A., & Abidin, E. Z. (2015). Heavy metals (Pb and Cu) assessments in hair samples of goldsmiths in Kelantan, Malaysia. Asia Pacific Environmental and Occupational Health Journal, 1(1). 1–8.
- Anushia, C., Prabhu, A., & Sampath, P. (2012). Pilots study on effect of copper and cadmium toxicity in Tilapia Mossambicus. *Journal of Research in Animal Sciences*, 1, 20–27.
- Araújo, C. V. M., Shinn, C., Moreira-Santos, M., Lopes, I., Espíndola, E. L. G., & Ribeiro, R. (2014). Copper-driven avoidance and mortality in temperate and tropical tadpoles. *Aquatic Toxicology*, 146, 70–75.
- Araújo, M. C. de, Assis, C. R. D., Silva, L. C., Machado, D. C., Silva, K. C. C., Lima, A. V. A., Carvalho, L. B., Bezerra, R. S., & Oliveira, M. B. M. (2016). Brain acetylcholinesterase of jaguar cichlid (*Parachromis managuensis*): From physicochemical and kinetic properties to its potential as biomarker of pesticides and metal ions. *Aquatic Toxicology*, 177, 182–189.
- Araujo, N. de S., & Borges, J. C. S. (2016). Rodlet cells changes in Oreochromis niloticus in response to organophosphate pesticide and their relevance as stress biomarker in teleost fishes. International Journal of Aquatic Biology, 3(6), 398–408.
- Askar, K. A., Kudi, A. C., & Moody, A. J. (2011). Purification of soluble acetylcholinesterase from sheep liver by affinity chromatography. *Applied Biochemistry and Biotechnology*, *165*(1), 336–346.
- Assis, C. R. D., Bezerra, R. S., & Carvalho Jr, L. B. (2011). Fish cholinesterases as biomarkers of organophosphorus and carbamate pesticides. In *Pesticides in the Modern World-Pests Control and Pesticides Exposure and Toxicity Assessment.* InTech. Retrieved from https://www.intechopen.com/download/pdf/20784
- Assis, C. R. D., Linhares, A. G., Oliveira, V. M., França, R. C. P., Santos, J. F., Marcuschi, M., Maciel, E. V. M., Bezerra, R. S., & Carvalho, L. B. (2014). Characterization of catalytic efficiency parameters of brain cholinesterases in tropical fish. *Fish Physiology and Biochemistry*, 40(6), 1659–1668.

- Augustinsson, K. B. (1961). Multiple forms of esterase in vertebrate blood plasma. Annals of the New York Academy of Sciences, 94(1), 844–860.
- Authman, M. M. (2015). Use of fish as bio-indicator of the effects of heavy metals pollution. Journal of Aquaculture Research and Development, 6(4). doi.org/10.4172/2155-9546.1000328
- Baiomy, A. A. (2016). Histopathological biomarkers and genotoxicity in gill and liver tissues of Nile tilapia, *Oreochromis niloticus* from a polluted part of the Nile River, Egypt. African Journal of Aquatic Science, 41(2), 181–191.
- Balasubramanian, J., & Kumar, A. (2015). Study of histopathology alterations in the liver of arsenic exposed *Heteropneustis fossilis* and the chelating effect of zeolite. *International Journal of Pharmacology and Biological Sciences*, 9(1), 13–19.
- Ballesteros, M. L., Rivetti, N. G., Morillo, D. O., Bertrand, L., Amé, M. V., & Bistoni, M. A. (2017). Multi-biomarker responses in fish (*Jenynsia multidentata*) to assess the impact of pollution in rivers with mixtures of environmental contaminants. *Science of The Total Environment*, 595, 711–722.
- Bandmann, O., Weiss, K. H., & Kaler, S. G. (2015). Wilson's disease and other neurological copper disorders. *The Lancet. Neurology*, 14(1), 103–113.
- Bartozek, E. C. R., Bueno, N. C., Feiden, A., & Rodrigues, L. C. (2016). Response of phytoplankton to an experimental fish culture in net cages in a subtropical reservoir. *Brazilian Journal of Biology*, 76(4), 824–833.
- Berg, J. M., Tymoczko, J. L., Stryer, L., & Stryer, L. (2007). Carbonic anhydrase and heavy metals. In *Biochemistry* (6th ed, pp. 206–228). New York: W.H. Freeman.
- Bonacci, S., Browne, M. A., Dissanayake, A., Hagger, J. A., Corsi, I., Focardi, S., & Galloway, T. S. (2004). Esterase activities in the bivalve mollusc *Adamussium* colbecki as a biomarker for pollution monitoring in the Antarctic marine environment. *Marine Pollution Bulletin*, 49(5), 445–455.
- Bose, M., Ilavazhahan, M., Tamilselvi, R., & Viswanathan, M. (2013). Effect of heavy metals on the histopathology of gills and brain of fresh water fish *Calta calta*. *Biomedical and Pharmacology Journal*, 6(1), 99–105.
- Bradbury, S., Carlson, R., Henry, T., Padilla, S., & Cowden, J. (2008). Toxic responses of the fish nervous system. In *The Toxicology of Fishes* (pp. 417–444). New York: CRC Press Taylor and Francis.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72(1-2), 248–254.
- Braithwaite, V. (2006). Hooked on a myth. *Los Angeles Times*. Retrieved from http://articles.latimes.com/print/2006/oct/08/opinion/oe-braithwaite. (Accessed on 27th July 2017)
- Buratowski, S. and Chodosh, L.A. (2001). Mobility shift dna-binding assay using gel electrophoresis. *Current Protocols in Molecular Biology*. John Wiley and Sons, Inc, United States, pp 1-11.
- Camargo, M. M. P., & Martinez, C. B. R. (2007). Histopathology of gills, kidney and liver of a Neotropical fish caged in an urban stream. *Neotropical Ichthyology*, 5(3), 327–336.

- Campos-Garcia, J., Martinez, D. S. T., Rezende, K. F. O., da Silva, J. R. M. C., Alves, O. L., & Barbieri, E. (2016). Histopathological alterations in the gills of Nile tilapia exposed to carbofuran and multiwalled carbon nanotubes. *Ecotoxicology and Environmental Safety*, 133, 481–488.
- Caropreso, S., Bondioli, L., Capannolo, D., Cerroni, L., Macchiarelli, R., & Condò, S. G. (2000). Thin sections for hard tissue histology: a new procedure. *Journal of Microscopy*, 199(3), 244–247. https://doi.org/10.1046/j.1365-2818.2000.00731.x.
- Casares, M. V., de Cabo, L. I., Seoane, R. S., Natale, O. E., Castro Ríos, M., Weigandt, C., & de Iorio, A. F. (2012). Measured copper toxicity to *Cnesterodon decemmaculatus* (Pisces: Poeciliidae) and predicted by biotic ligand model in Pilcomayo River water: A step for a cross-fish-species extrapolation. *Journal of Toxicology*. https://doi.org/10.1155/2012/849315
- Cavas, T. (2007). In vivo genotoxicity of mercury chloride and lead acetate: micronucleus test on acridine orange stained fish cells. *Food and Chemical Toxicology*, 46, 352 – 258.
- Chatonnet, A., & Lockridge, O. (1989). Comparison of butyrylcholinesterase and acetylcholinesterase. *Biochemical Journal*, 260(3), 625-634.
- Chen, H., Teng, Y., Lu, S., Wang, Y., & Wang, J. (2015). Contamination features and health risk of soil heavy metals in China. *Science of The Total Environment*, *512-513*, 143–153.
- Choi, J. S., Haulader, S., Karki, S., Jung, H. J., Kim, H. R., & Jung, H. A. (2015). Acetyl and butyrylcholinesterase inhibitory activities of the edible brown alga *Eisenia bicyclis. Archives of Pharmacal Research*, 38(8), 1477–1487.
- Chor, W. K., Lim, L. S., & Shapawi, R. (2013). Evaluation of feather meal as a dietary protein source for African catfish fry, *Clarias gariepinus*. Journal of Fisheries and Aquatic Science, 8(6), 697–705.
- Ciamarro, C. M., Pereira, B. F., Alves, R. M. da S., Valim, J. R. T., Pitol, D. L., & Caetano, F. H. (2015). Changes in muscle and collagen fibers of fish after exposure to urban pollutants and biodegradable detergents. *Microscopy Research*, 3(3), 33–40.
- Ciena, A. P., Luques, I. U., Dias, F. J., Yokomizo de Almeida, S. R., Iyomasa, M. M., & Watanabe, I. (2010). Ultrastructure of the myotendinous junction of the medial pterygoid muscle of adult and aged Wistar rats. *Micron*, 41(8), 1011–1014.
- Colovic, M. B., Krstic, D. Z., Lazarevic-Pasti, T. D., Bondzic, A. M., & Vasic, V. M. (2013). Acetylcholinesterase inhibitors: pharmacology and toxicology. *Current Neuropharmacology*, 11(3), 315–335.
- Dang, M., Nørregaard, R., Bach, L., Sonne, C., Søndergaard, J., Gustavson, K., Aasrtup, P., & Nowak, B. (2017). Metal residues, histopathology and presence of parasites in the liver and gills of fourhorn sculpin (*Myoxocephalus quadricornis*) and shorthorn sculpin (*Myoxocephalus scorpius*) near a former lead-zinc mine in East Greenland. *Environmental Research*, 153, 171–180. https://doi.org/10.1016/j.envres.2016.12.007
- David, M., & Kartheek, R. M. (2016). In vivo studies on hepato-renal impairments in freshwater fish *Cyprinus carpio* following exposure to sublethal

concentrations of sodium cyanide. *Environmental Science and Pollution Research International*, 23(1), 722–733.

- Devi, M., & Fingerman, M. (1995). Inhibition of acetylcholinesterase activity in the central nervous system of the red swamp crayfish, *Procambarus clarkii*, by mercury, cadmium, and lead. *Bulletin of Environmental Contamination and Toxicology*, 55(5), 746–750.
- Dew, W. A., Wood, C. M., & Pyle, G. G. (2012). Effects of continuous copper exposure and calcium on the olfactory response of fathead minnows. *Environmental Science and Technology*, 46(16), 9019–9026.
- Di Giulio, R. T., & Hinton, D. E. (2008). Biomarkers. In *The Toxicology of Fishes* (pp. 685–731). New York: CRC Press Taylor and Francis.
- Doaa, M. M., & Hanan, H. (2013). Histological changes in selected organs of Oreochromis niloticus exposed to doses of lead acetate. *Journal of Life Science Biomedicine*, 3(3), 256–263.
- DOE. (2011). *Malaysia Environmental quality report 2007*. Department of Environment, Ministry of Natural Resources and Environment, Malaysia.
- ElGazzar, A., Ashry, K., & ElSayed, Y. (2014). Physiological and Oxidative Stress Biomarkers in the Freshwater Nile Tilapia, *Oreochromis niloticus L.*, exposed to sublethal doses of cadmium. *Alexandria Journal of Veterinary Sciences*, 40(1), 29–43.
- Ellman, G. L., Courtney, K. D., Andres, V., & Featherstone, R. M. (1961). A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochemical Pharmacology*, 7(2), 88–95.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology*, *35*(4), 495–516.
- El-Moselhy, K. M., Othman, A. I., Abd El-Azem, H., & El-Metwally, M. E. A. (2014). Bioaccumulation of heavy metals in some tissues of fish in the Red Sea, Egypt. *Egyptian Journal of Basic and Applied Sciences*, 1(2), 97–105.
- Evans, D. H., Piermarini, P. M., & Choe, K. P. (2005). The multifunctional fish gill: dominant site of gas exchange, osmoregulation, acid-base regulation, and excretion of nitrogenous waste. *Physiological Reviews*, 85(1), 97–177.
- Ezeonyejiaku, C. D., Obiakor, M. O., & Ezenwelu, C. O. (2011). Toxicity of copper sulphate and behavioural locomotor response of Tilapia (*Oreochromis niloticus*) and Catfish (*Clarias gariepinus*) species. *Online Journal of Animal and Feed Research*, 1(4), 130–134.
- Falugi, C., & Aluigi, M. G. (2012). Early appearance and possible functions of nonneuromuscular cholinesterase activities. *Frontiers in Molecular Neuroscience*, 5. https://doi.org/10.3389/fnmol.2012.00054
- Frasco, M. F., Colletier, J. P., Weik, M., Carvalho, F., Guilhermino, L., Stojan, J., & Fournier, D. (2007). Mechanisms of cholinesterase inhibition by inorganic mercury: Cholinesterase inhibition by mercury. *FEBS Journal*, 274(7), 1849– 1861.
- Fukuto, T. R. (1990). Mechanism of action of organophosphorus and carbamate insecticides. *Environmental Health Perspectives*, 87, 245–254.
- Gandhewar, S. S., Zade, S. B., Sitre, S. R., & Satyanarayan, S. (2012). Acute toxicity bioassay of copper on the freshwater fish *Rasbora daniconius*. *Indian Journal of Environmental Protection*, *32*(2), 158–162.

- Garfin, D.E. (2003). Gel electrophoresis of protein. Essential Cell Biology. Oxford University Press, United Kingdom, pp 1-34.
- Gauthier, P. T., Norwood, W. P., Prepas, E. E., & Pyle, G. G. (2016). Behavioural alterations from exposure to Cu, phenanthrene, and Cu-phenanthrene mixtures: linking behaviour to acute toxic mechanisms in the aquatic amphipod, *Hyalella azteca*. Aquatic Toxicology, 170, 377–383.
- Geethanjali, S. (2013). Isolation, purification, characterization and immobilization of protease from *Labeo rohita* viscera. http://hdl.handle.net/10603/13311. Accessed on 17 July 2017
- Ghazala, Mahboob, S., Ahmad, L., Sultana, S., Alghanim, K., Al-Misned, F., & Ahmad, Z. (2014). Fish cholinesterases as biomarkers of sublethal effects of organophosphorus and carbamates in tissues of *Labeo rohita*. *Journal of Biochemical and Molecular Toxicology*, 28(3), 137–142.
- Ghazala, Mahboob, S., Alghanim, K., Al-Misned, F., Ahmed, L., & Ahmed, Z. (2016). Effect of Triazophos on esterase activity and protein contents of liver, kidney, brain, blood and muscles of *Catla catla, Labeo rohita* and *Cirrhinus mrigala*. *Pakistan Journal of Zoology*, 48(2), 513–518.
- Ghazali, A. R., Abdul Razak, N. E., Othman, M. S., Othman, H., Ishak, I., Lubis, S. H.,
 ... Abdullah, R. (2012). Study of Heavy Metal Levels among Farmers of Muda Agricultural Development Authority, Malaysia. *Journal of Environmental and Public Health*, 4. doi: 10.1155/2012/758349
- Giannuzzi, L. A., & Stevie, F. A. (1999). A review of focused ion beam milling techniques for TEM specimen preparation. *Micron*, 30(3), 197–204. https://doi.org/10.1016/S0968-4328(99)00005-0.
- Glusker, J., Katz, A., & Bock, C. (1999). Metal Ions in Biological Systems, Volume 33. *The Rigaku Journal*, 16(2), 8–19.
- Gomes, I. D. L., Lemos, M. F. L., Soares, A. M. V. M., Barata, C., & Faria, M. (2014). The use of cholinesterase as potential biomarker: In vitro characterization in the polychaete *Capitella teleta*. *Marine Pollution Bulletin*, 85(1), 179–185.
- Gray, D. P., & Harding, J. S. (2012). Acid mine drainage index (AMDI): a benthic invertebrate biotic index for assessing coal mining impacts in New Zealand streams. New Zealand Journal of Marine and Freshwater Research, 46(3), 335–352.
- Gregory, M., Deane, M., & Mars, M. (2003). Ultrastructural changes in untraumatised rabbit skeletal muscle treated with deep transverse friction. *Physiotherapy*, 89(7), 408–416.
- Gunasekaran, B. (2011). Purification, characterisation and inhibition studies of protease from *Coriandrum sativum*. (PhD Thesis). Universiti Putra Malaysia.
- Haluzová, I., Modrá, H., Blahová, J., Havelková, M., Široká, Z., & Svobodová, Z. (2011). Biochemical markers of contamination in fish toxicity tests. *Interdisciplinary Toxicology*, 4(2). 85–89.
- Hayat, N. M., Shamaan, N. A., Sabullah, M. K., Shukor, M. Y., Syed, M. A., Khalid, A., Dahalan, F. A., & Ahmad, S. A. (2016). The use of *Lates calcarifer* as a biomarker for heavy metals detection. *Rendiconti Lincei*, 27(3), 463–472.
- Hayat, N. M., Shamaan, N. A., Shukor, M. Y., Sabullah, M. K., Syed, M. A., Khalid, A., Dahalan, F. A., & Ahmad, S. A. (2015). Cholinesterase-based biosensor

using *Lates calcarifer* (Asian Seabass) brain for detection of heavy metals. *Journal of Chemical and Pharmaceutical Sciences*, 8(2), 376–381.

- Hayat, N., Ahmad, S., Shamaan, N., Sabullah, M., Syed, M., Khalid, A., Khalil, K., & Dahalan, F. (2017). Characterisation of cholinesterase from kidney tissue of Asian seabass *Lates calcarifer* and its inhibition in presence of metal ions. *Journal of Environmental Biology*, 38, 381–388.
- Hege, D., & Matsuda, R. (2015). Affinity chromatography: Methods and protocols, methods in molecular biology. Springer Science Business Media, 1286, 1–19.
- Hernández-Moreno, D., de, la C.-R. I., Maria, F. J., González-Gómez, M. J., María, N. C., Soler, F., & Pérez-López, M. (2014). Different enzymatic activities in Carp (*Cyprinus carpio L.*) as potential biomarkers of exposure to the pesticide methomyl. Archives of Industrial Hygiene and Toxicology, 65(3), 311–318.
- Hidouri, S., Ensibi, C., Landoulsi, A., & Daly-Yahia, M. N. (2017). Effects of chronic exposure to silver nanoparticles on *Ruditapes decussatus* gills using biochemical markers. *Water, Air, and Soil Pollution, 228*(2). https://doi.org/10.1007/s11270-017-3265-0
- Hinton, D. E., Segner, H., Au, D., Kullman, S. W., & Hardman, R. C. (2008). Liver toxicity. In *The Toxicology of Fishes* (pp. 327–368). New York: CRC Press Taylor and Francis.
- Hoseini, S. M., Hedayati, A., Taheri Mirghaed, A., & Ghelichpour, M. (2016). Toxic effects of copper sulfate and copper nanoparticles on minerals, enzymes, thyroid hormones and protein fractions of plasma and histopathology in common carp *Cyprinus carpio. Experimental and Toxicologic Pathology*, 68(9), 493–503.
- Ibemenuga, K. N. (2016). Bioaccumulation and toxic effects of some heavy metals in freshwater fishes. *Animal Research International*, 10(3), 1792–1798.
- Idriss, A. A., & Ahmad, A. K. (2015). Heavy metal concentrations in fishes from Juru River, estimation of the health risk. *Bulletin of Environmental Contamination and Toxicology*, 94(2), 204–208.
- Ismail, I., & Mat Saleh, I. (2012). Analysis of heavy metals in water and fish (Tilapia sp.) samples from Tasik Mutiara, Puchong. The Malaysia Journal of Analytical Sciences, 16(3), 346–352.
- Ivanov, I., Matafonov, A., Sun, M., Cheng, Q., Dickeson, S. K., Verhamme, I. M., Emsley, J., & Gailani, D. (2017). Proteolytic properties of single-chain factor XII: a mechanism for triggering contact activation. *Blood*,. https://doi.org/10.1182/blood-2016-10-744110
- Jaffal, A., Betoulle, S., Biagianti-Risbourg, S., Terreau, A., Sanchez, W., & Paris-Palacios, S. (2015). Heavy metal contamination and hepatic toxicological responses in brown trout (*Salmo trutta*) from the Kerguelen Islands. *Polar Research*, 34(22784).

https://doi.org/http://dx.doi.org/10.3402/polar.v34.22784

- Jagadeshwarlu, R., Reddy, P., & Sunitha, D. (2015). Toxicity of Copper sulphate (CuSO₄, 5H₂O) to *Oreochromis mossambicus* (Tilapia). *Biolife*, 3(3), 657–661.
- Jalaludeen, M., Arunachalam, M., Raja, M., Nandagopal, S., Showket, A., Sundar, S., & Palanimuthu, D. (2012). Histopathology of the gill, liver and kidney tissues

of the freshwater fish *Tilapia mossambica* exposed to cadmium sulphate. *International Journal of Advanced Biological Research*, 24(4), 572–578.

- Jasim, M. A., Sofian-Azirun, M., Yusoff, I., & Rahman, M. M. (2016). Bioaccumulation and histopathological changes induced by toxicity of mercury (HgCl₂) to Tilapia fish *Oreochromis niloticus*. Sains Malaysiana, 45(1), 119–127.
- Javed, M., & Usmani, N. (2016). Accumulation of heavy metals and human health risk assessment via the consumption of freshwater fish Mastacembelus armatus inhabiting, thermal power plant effluent loaded canal. *SpringerPlus*, 5(1). https://doi.org/10.1186/s40064-016-2471-3
- Jayarama, N., Vanita, S., Abhilah, V., & Basavaraj, M. (2015). Influence of flaxseed on the body weight, biochemical constituents and histology of muscle and liver tissues of common carp, *Cyprinus carpio*: A comparative study. *International Journal of Fisheries and Aquatic Studies*, 2(6), 170–174.
- Jebali, J., Khedher, S. B., Sabbagh, M., Kamel, N., Banni, M., & Boussetta, H. (2013). Cholinesterase activity as biomarker of neurotoxicity: utility in the assessment of aquatic environment contamination. *Revista de Gestão Costeira Integrada*, *13*(4), 425–437.
- Jindal, R., & Kaur, M. (2014). Acetylcholinesterase inhibition and assessment of its recovery response in some organs of *Ctenopharyngodon idellus* induced by chlorpyrifos. *International Journal of Science, Environment and Technology*, 3(2), 473–480.
- Kamran, A., Bibi, Z. and Kamal, M. (2014). Purification and molecular weight estimation of protease from a thermophilic Bacillus species. *Pakistan Journal* of Biochemistry and Molecular Biology, 47(3-4), 155-157.
- Kang, D., Gho, Y.S., Suh, M. and Kang, C. (2002). Highly sensitive and fast protein detection with coomassie brilliant blue in sodium dodecyl sulfatepolyacrylamide gel electrophoresis. *Bulletin Korean Chemistry and Society*, 23(11), 1511-1512.
- Kaviraj, A., Unlu, E., Gupta, A., & Nemr, A. El. (2014). Biomarkers of environmental pollutants. *BioMed Research International*, 2014, e806598. https://doi.org/10.1155/2014/806598
- Khajah, M. A., & Luqmani, Y. . (2016). Involvement of membrane blebbing in immunological disorder and cancer. *Medical Principle and Practice*, 25(2), 18–27.
- Khantaphant, S. and Benjakul, S. (2010). Purification and characterization of trypsin from the pyloric caeca of brownstripe red snapper (*Lutjanus vitta*). Food *Chemistry*, 120, 658-664.
- Khati, W., Ouali, K., Mouneyrac, C., & Banaoui, A. (2012). Metallothioneins in aquatic invertebrates: Their role in metal detoxification and their use in biomonitoring. *Energy Procedia*, 18, 784–794.
- Kim, W. K., Lee, S. K., & Jung, J. (2010). Integrated assessment of biomarker responses in common carp (*Cyprinus carpio*) exposed to perfluorinated organic compounds. *Journal of Hazardous Materials*, 180(1–3), 395–400.
- Kipigroch, K., Janosz-Rajczyk, M., & Skowron-Grabowska, B. (2016). The use of algae in the removal of Cd and Cu in the process of wastewater recovery. *Desalination and Water Treatment*, *57*(3), 1508–1514.

- Kleinow, K. M., Nichols, J. W., Hayton, W. L., McKim, J. M., & Barron, M. G. (2008). Toxicokinetics in Fishes. In *The Toxicology of Fishes* (pp. 55–152). New York: CRC Press Taylor and Francis.
- Koenig, S., & Solé, M. (2014). Muscular cholinesterase and lactate dehydrogenase activities in deep-sea fish from the NW Mediterranean. *Marine Environmental Research*, 94, 16–23.
- Krisna, C., Duong-Ly, and Gabelli, S.B., (2014). Chapter Eight- Using ion exchange chromatography to purify recombinantly expressed protein. *Method in Enzymology* 541, 95-103.
- Kröner, F. and Hubbuch, J. (2013). Systematic generation of buffer systems for pH gradient ion exchange chromatography and their application. *Journal of Chromatography, A 1285*, 78-87.
- Kumar, M., Kumar, D., & Kumar, R. (2017). Sublethal effects of cadmium and copper on the blood characteristics of catfish *Clarias batrachus* (Linn.). *International Journal of Advanced Research in Biological Sciences*, 4(1), 123–128.
- Kumar, N., Krishnani, K. K., Gupta, S. K., & Singh, N. P. (2017). Cellular stress and histopathological tools used as biomarkers in *Oreochromis mossambicus* for assessing metal contamination. *Environmental Toxicology and Pharmacology*, 49, 137–147.
- Laith, A., Ambak, M. A., Hassan, M., Sheriff, S. M., Nadirah, M., Draman, A. S., Wahab, W., Ibrahim, W. N. W., Aznan, A., Jabar, A., & Najiah, M. (2017). Molecular identification and histopathological study of natural Streptococcus agalactiae infection in hybrid tilapia (*Oreochromis niloticus*). Veterinary World, 10(1), 101–111. https://doi.org/10.14202/vetworld.2017.101-111
- Langley, A., & Dameron, C. T. (2013). Copper and Anesthesia: Clinical Relevance and Management of Copper Related Disorders. *Anesthesiology Research and Practice*, 2013. doi.org/10.1155/2013/750901
- Lari, E., & Pyle, G. G. (2017). Rainbow trout (*Oncorhynchus mykiss*) detection, avoidance, and chemosensory effects of oil sands process-affected water. *Environmental Pollution*, 225, 40–46.
- Ledy, K., Giamberini, L., & Pihan, J. C. (2003). Mucous cell responses in gill and skin of brown trout *Salmo trutta fario* in acidic, aluminium containing stream water. *Disease of Aquatic Organisms*, 56(3), 235–240.
- Leo, N., & Mat, N. (2017). Mozambique tilapia (*Oreochromis mossambicus*) Fact sheet [Non Indigenous Aquatic Species Database]. Retrieved 6 September 2017, from https://nas.er.usgs.gov/queries/factsheet.aspx?SpeciesID=466
- Lionetto, M. G., Caricato, R., Calisi, A., Giordano, M. E., & Schettino, T. (2013). Acetylcholinesterase as a biomarker in environmental and occupational medicine: New insights and future perspectives. *BioMed Research International*, 2013, 321–213.
- Liu, Y., Gu, P., Jia, L., & Zhang, G. (2016). An investigation into the use of cuprous chloride for the removal of radioactive iodide from aqueous solutions. *Journal of Hazardous Materials*, 302, 82–89. https://doi.org/10.1016/j.jhazmat.2015.09.045.
- Lu, Y., Wu, Z., Song, Z., Xiao, P., Liu, Y., Zhang, P., & You, F. (2016). Insight into the heat resistance of fish via blood: Effects of heat stress on metabolism, oxidative stress and antioxidant response of olive flounder *Paralichthys*

olivaceus and turbot Scophthalmus maximus. Fish & Shellfish Immunology, 58, 125–135.

- Mackay, D., & Milford, L. (2008). Exposure assessment and modelling in the aquatic environment (pp. 645–658). New York: CRC Press Taylor and Francis.
- Maharajan, A., Kitto, M. R., Paruruckumani, P. S., & Ganapiriya, V. (2016). Histopathology biomarker responses in Asian sea bass, *Lates calcarifer* (Bloch) exposed to copper. *The Journal of Basic & Applied Zoology*, 77, 21– 30.
- Mashifane, T. B., & Moyo, N. A. G. (2014). Acute toxicity of selected heavy metals to Oreochromis mossambicus fry and fingerlings. African Journal of Aquatic Science, 39(3), 279–285.
- Meng, F., Li, M., Tao, Z., Yuan, L., Song, M., Ren, Q., Xin, X., Meng, Q., & Wang, R. (2016). Effect of high dietary copper on growth, antioxidant and lipid metabolism enzymes of juvenile larger yellow croaker *Larimichthys croceus*. *Aquaculture Reports*, 3, 131 – 135.
- Monserrat, J. M., Bianchini, A., & Bainy, A. C. D. (2002). Kinetic and toxicological characteristics of acetylcholinesterase from the gills of oysters (*Crassostrea rhizophorae*) and other aquatic species. *Marine Environmental Research*, 54(3–5), 781–785.
- Muraoka, S., & Miura, T. (2003). Inactivation of cholinesterase induced by chlorpromazine cation radicals. *Pharmacology & Toxicology*, 92(2), 100–104.
- Muthukumaravel, K., Murthy, A., Kumarasamy, P., & Amsath, A. (2008). Light and scanning electron microscopic evaluation of effects of copper sulphate on the gill architecture of fresh water fish *Oreochromis mossambicus*. *Pollution Research*, 27(4), 715–719.
- Naji, A., Ismail, A., Kamrani, E., & Sohrabi, T. (2014). Correlation of MT levels in livers and gills with heavy metals in wild tilapia (*Oreochromis mossambicus*) from the Klang river, Malaysia. Bulletin of Environmental Contamination Toxicology, 92, 674–679.
- Nave, K. A., & Werner, H. B. (2014). Myelination of the nervous system: mechanisms and functions. *Annual Review of Cell and Developmental Biology*, 30, 503– 533. https://doi.org/10.1146/annurev-cellbio-100913-013101
- Nimet, J., Guimarães, A. T. B., & Delariva, R. L. (2017). Use of muscular cholinesterase of *Astyanax bifasciatus* (Teleostei, Characidae) as a biomarker in biomonitoring of rural streams. *Bulletin of Environmental Contamination and Toxicology*. https://doi.org/10.1007/s00128-017-2111-9
- Nowakowski, A. B., Wobig, W. J., & Petering, D. H. (2014). Native SDS-PAGE: High resolution electrophoretic separation of proteins with retention of native properties including bound metal ions. *Metallomics:Integrated Biometal Science*, 6(5), 1068–1078. https://doi.org/10.1039/c4mt00033a
- Nunes, B. (2011). The use of cholinesterases in ecotoxicology. *Reviews of Environmental Contamination and Toxicology*, 212, 29–59.
- Nussey, G., Van Vuren, J. H. J., & Preez, H. H. Du. (1995). Effect of copper on blood coagulation of Oreochromis mossambicus (Cichlidae). Comparative Biochemistry and Physiology Part C: Pharmacology, Toxicology and Endocrinology, 111(3), 359–367.

- Oliveira, V. M., Assis, C. R. D., Costa, H. M. S., Silva, R. P. F., Santos, J. F., Carvalho Jr, L. B., & Bezerra, R. S. (2017). Aluminium sulfate exposure: A set of effects on hydrolases from brain, muscle and digestive tract of juvenile Nile tilapia (*Oreochromis niloticus*). Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology, 191, 101–108.
- Olojo, E. A. A., Olurin, K. B., Mbaka, G., & Oluwe-mimo, A. D. (2004). Histopathology of the gill and liver tissues of the African catfish *Clarias* gariepinus exposed to lead. African Journal of Biotechnology, 4(1), 117 – 122.
- Oronsaye, J. A. O. (1997). Ultrastructural changes in the gills of the stickleback, *Gasterosteus aculeatus (L.)* exposed to dissolved cadmium in hard and soft water. *Journal of Aquatic Sciences*, 12, 59 66.
- Osredkar, J. (2011). Copper and Zinc, Biological Role and Significance of Copper/Zinc Imbalance. *Journal of Clinical Toxicology*, 3(1). https://doi.org/10.4172/2161-0495.S3-001
- Ostaszewska, T., Chojnacki, M., Kamaszewski, M., & Sawosz-Chwalibóg, E. (2016). Histopathological effects of silver and copper nanoparticles on the epidermis, gills, and liver of Siberian sturgeon. *Environmental Science and Pollution Research*, 23(2), 1621–1633.
- Ott, P., Hope, M. J., Verkleij, A. J., Roelofsen, B., Brodbeck, U., & Van Deenen. (1981). Effect of dimyristol phosphotidyl choline on intact erythrocyte; release of spectrin-free vesicle without ATP depletion. *Biochimica et Biophysica Acta*, 641, 79–87.
- Paruruckumani, P. S., Maharajan, A., Ganapiriya, V., Narayanaswamy, Y., & Jeyasekar, R. R. (2015). Surface ultrastructural changes in the gill and liver tissue of asian sea bass *Lates calcarifer* (Bloch) exposed to copper. *Biological Trace Element Research*, 168(2), 500–507.
- Parveen, M., & Kumar, S. (2005). *Recent Trends in the Acetylcholinesterase System*. The Netherlands: IOS Press.
- Pelgrom, S. M. G. J., Lock, R. A. C., Balm, P. H. M., & Bonga, S. E. W. (1995). Integrated physiological response of tilapia, *Oreochromis mossambicus*, to sublethal copper exposure. *Aquatic Toxicology*, 32(4), 303–320.
- Perić, L., Ribarić, L., & Nerlović, V. (2013). Cholinesterase activity in the tissues of bivalves Noah's ark shell (Arca noae) and warty venus (Venus verrucosa): Characterisation and in vitro sensitivity to organophosphorous pesticide trichlorfon. Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology, 165(4), 243–249.
- Perrie, Y., Ali, H., Kirby, D. J., Mohammed, A. U. R., McNeil, S. E., & Vangala, A. (2017). Environmental scanning electron microscope imaging of vesicle systems. In *Liposomes* (pp. 131–143). Humana Press, New York, NY. Retrieved from <u>https://link.springer.com/protocol/10.1007/978-1-4939-6591-5_11</u>
- Pfeifer, S., Schiedek, D., & Dippner, J. W. (2005). Effect of temperature and salinity on acetylcholinesterase activity, a common pollution biomarker, in *Mytilus* sp. from the south-western Baltic Sea. *Journal of Experimental Marine Biology and Ecology*, 320, 93-103.

- Ploch, C. C., Mansi, C. S. S. A., Jayamohan, J., & Kuhl, E. (2016). Using 3D printing to create personalized brain models for neurosurgical training and preoperative planning. *World Neurosurgery*, 90, 668–674.
- Pohanka, M. (2014). Copper, aluminum, iron and calcium inhibit human acetylcholinesterase *in vitro*. *Environmental Toxicology and Pharmacology*, 37(1), 455–459.
- Price, M. (2013). Sub-lethal metal toxicity effects on salmonids: a review [SkeenaWild Conservation Trust]. Retrieved 26 October 2016, from www.skeenawild.org
- Privitera, G., & Meli, G. (2016). An unusual cause of anemia in cirrhosis: spur cell anemia, a case report with review of literature. *Gastroenterology and Hepatology From Bed to Bench*, 9(4), 335–339.
- Quintaneiro, C., Ranville, J., & Nogueira, A. J. A. (2015). Effects of the essential metals copper and zinc in two freshwater detritivores species: Biochemical approach. *Ecotoxicology and Environmental Safety*, 118, 37–46.
- Rai, A. N., Ullah, A., & Haider, J. (2015). Determination of acute toxicity of copper and cobalt for *Tilapia nilotica*. *Journal of Bioresource Management*, 2(1), 16– 25.
- Rakhi, S. F., Reza, A. H. M. M., Hossen, M. S., & Hossain, Z. (2013). Alterations in histopathological features and brain acetylcholinesterase activity in stinging catfish *Heteropneustes fossilis* exposed to polluted river water. *International Aquatic Research*, 5(1), 5–7.
- Riba, I., Blasco, J., Jiménez-Tenorio, N., González de Canales, M. L., & Ángel DelValls, T. (2005). Heavy metal bioavailability and effects: II. Histopathology-bioaccumulation relationships caused by mining activities in the Gulf of Cádiz (SW, Spain). *Chemosphere*, 58(5), 671–682. https://doi.org/10.1016/j.chemosphere.2004.02.016
- Roberts, R. J. (2012). *Fish Pathology* (4th Edition, pp. 15–17). New York: John Wiley & Sons.
- Rodríguez-Fuentes, G., & Gold-Bouchot, G. (2004). Characterization of cholinesterase activity from different tissues of Nile tilapia (*Oreochromis niloticus*). Marine *Environmental Research*, 58(2-5), 505–509.
- Roméo, M., Siau, Y., Sidoumou, Z., & Gnassia-Barelli, M. (1999). Heavy metal distribution in different fish species from the Mauritania coast. Science of The Total Environment, 232(3), 169–175.
- Rudneva, I. I., & Kovyrshina, T. B. (2015). Comparative study of the cholinesterase activities in tissues of the round goby *Neogobius melanostomus* (Gobiidae) from different locations of the Black and Azov seas. *Journal of Ichthyology*, 55(5), 734–738.
- Sabullah, M. K., Abd. Shukor, M. Y., Shamaan, N. A., Khalid, A., Ganzau, A. J., Sulaiman, M. R., Jirangon, H., & Ahmad, S. A. (2015d). Purification and anticholinesterase sensitivity of cholinesterase extracted from liver tissue of *Puntius javanicus. International Journal of Agriculture and Biology*, 17(5), 1025–1030.
- Sabullah, M. K., Ahmad, S. A., Shukor, M. Y., Gansau, A. J., Syed, M. A., Sulaiman, M. R., & Shamaan, N. A. (2015a). Heavy metal biomarker: Fish behavior, cellular alteration, enzymatic reaction and proteomics approaches. *International Food Research Journal*, 22(2), 435–454.

- Sabullah, M. K., Ahmad, S. A., Shukor, M. Y., Shamaan, N. A., Khalid, A., Gansau, A. J., Dahalan, F. A., & Sulaiman, M. R. (2015b). Acetylcholinesterase from *Puntius javanicus* for the detection of carbamates and organophosphates. *Journal of Chemical and Pharmaceutical Sciences*, 8(2), 348–353.
- Sabullah, M. K., Shukor, M. Y., Sulaiman, M. R., Shamaan, N. A., Syed, M. A., Khalid, A., & Ahmad, S. A. (2014b). The effect of copper on the ultrastructure of *Puntius javanicus* hepatocyte. *Australian Journal of Basic and Applied Sciences*, 8(15), 245–251.
- Sabullah, M. K., Sulaiman, M. R., Shukor, M. Y. A., Shamaan, N. A., Khalid, A., & Ahmad, S. A. (2015c). In vitro and in vivo effects of *Puntius javanicus* cholinesterase by copper. *Fresenius Environmental Bulletin*, 24(12B), 4615– 4621
- Sabullah, M. K., Sulaiman, M. R., Shukor, M. Y. A., Syed, M. A., Shamaan, N. A., Khalid, A., & Ahmad, S. A. (2014a). The assessment of cholinesterase from the liver of *Puntius javanicus* as detection of metal ions. *Scientific World Journal*.
- Sabullah, M., Ahmad, S., Hussain, J., Gansau, A., & Sulaiman, M. (2014c). Acute effect of copper on *Puntius javanicus* survival and a current opinion for future biomarker development. *Journal of Environmental Bioremediation and Toxicology*, 2(1), 28–32.
- Sabullah, M., Ahmad, S., Shukor, M., Syed, M., & Shamaan, N. (2013). The evaluation of Periophtalmodon schlosseri as a source of acetylcholinesterase for the detection of insecticides. *Bulletin of Environmental Science and Management*, *1*(1), 20–24.
- Sandrini, J. Z., Rola, R. C., Lopes, F. M., Buffon, H. F., Freitas, M. M., Martins, C. de M. G., & da Rosa, C. E. (2013). Effects of glyphosate on cholinesterase activity of the mussel *Perna perna* and the fish *Danio rerio* and *Jenynsia multidentata*: in vitro studies. *Aquatic Toxicology*, 130, 171–173.
- Santos, C. S. A., Monteiro, M. S., Soares, A. M. V. M., & Loureiro, S. (2016). Brain cholinesterase reactivation as a marker of exposure to anticholinesterase pesticides: a case study in a population of yellow-legged gull *Larus michahellis* (Naumann, 1840) along the northern coast of Portugal. *Environmental Science and Pollution Research*, 23(1), 266–272.
- Sepúlveda, M. S., Gallagher, E. P., & Gross, T. S. (2004). Physiological changes in largemouth bass exposed to paper mill effluents under laboratory and field conditions. *Ecotoxicology*, 13(4), 291–301.
- Sevcikova, M., Modra, H., Slaninova, A., & Svobodova, Z. (2011). Metals as a cause of oxidative stress in fish: A review. *Veterinarni Medicina*, *56*(11), 537–546.
- Sharif, M. S. A., Halmi, M. I. E., Syahir, A., Johari, W. L. W., & Shukor, M. Y. (2014). Assessment of acetylcholinesterase from Channa micropeltes as a source of enzyme for insecticides detection. *International Journal Agricultural Biology*, 16(2), 389–94.
- Shazili, N. A. M., Yunus, K., Ahmad, A. S., Abdullah, N., & Rashid, M. K. A. (2006). Heavy metal pollution status in the Malaysian aquatic environment. *Aquatic Ecosystem Health & Management*, 9(2), 137–145.

- Shi, Y., Jiang, J., Shan, Z., Bu, Y., Deng, Z., & Cheng, Y. (2015). Oxidative stress and histopathological alterations in liver of *Cyprinus carpio L*. induced by intraperitoneal injection of microcystin-LR. *Ecotoxicology*, 24(3), 511–519.
- Shukor, M. Y., & Sulaiman, M. R. (2013). Assessment of acetylcholinesterase (AChE) from silver catfish (*Pangasius* sp.) as an assay for organophosphates and carbamates. *Biosciences Biotechnology Research Asia*. 10(1), 213–218.
- Silva, K. C. C., Assis, C. R. D., Oliveira, V. M., Carvalho, L. B., & Bezerra, R. S. (2013). Kinetic and physicochemical properties of brain acetylcholinesterase from the peacock bass (*Cichla ocellaris*) and in vitro effect of pesticides and metal ions. *Aquatic Toxicology*, 126, 191–197.
- Soorya, S. R., Aruna Devi, C., Binitha, R. N., Amrutha, B. V., Jeyalekshmi, G., & Francis, S. (2012). Oxidative stress experienced by freshwater fish Anabas testudineus exposed to sewage effluents of Parvathyputhenar, Kerala. Biju Kumar A (ed) Biodiversity Utilization and Threats. New Delhi, India: Narendra Publishing House, 769–780.
- Stankevičiūtė, M., Sauliutė, G., Svecevičius, G., Kazlauskienė, N., & Baršienė, J. (2017). Genotoxicity and cytotoxicity response to environmentally relevant complex metal mixture (Zn, Cu, Ni, Cr, Pb, Cd) accumulated in Atlantic salmon (*Salmo salar*). Part I: importance of exposure time and tissue dependence. *Ecotoxicology*, 1–14.
- Strzyzewska, E., Szarek, J., & Babinska, I. (2016). Morphologic evaluation of the gills as a tool in the diagnostics of pathological conditions in fish and pollution in the aquatic environment: a review. *Veterinarni Medicina*, 61(3), 123–132.
- Suganthi, P., Murali, M., Sadiq Bukhari, A., Syed Mohamed, Basu, & Singhal, R. (2015). Morphological and liver histological effects of ZnO nanoparticles on Mozambique tilapia. *Journal of Advanced Applied Scientific Research*, 1(1), 68–83.
- Sun, P. L., Hawkins, W. E., Overstreet, R. M., & Brown-Peterson, N. J. (2009). Morphological deformities as biomarkers in fish from contaminated rivers in Taiwan. *International Journal of Environmental Research and Public Health*, 6(8), 2307–2331.
- Talesa, V., Grauso, M., Principato, G. B., Giovannini, E., Norton, S. J., & Rosi, G. (1994). Presence of soluble tetrameric (blood) and membrane-bound dimeric forms of cholinesterase in the mollusk *Murex brandaris* (Gastropoda: Neogastropoda). *Journal of Experimental Zoology*, 270(3), 233–244.
- Taweel, A., Shuhaimi-Othman, M., & Ahmad, A. K. (2013). Assessment of heavy metals in tilapia fish (*Oreochromis niloticus*) from the Langat River and Engineering Lake in Bangi, Malaysia, and evaluation of the health risk from tilapia consumption. *Ecotoxicology and Environmental Safety*, 93, 45–51.
- Tecles, F., & Cerón, J. J. (2001). Determination of whole blood cholinesterase in different animal species using specific substrates. *Research in Veterinary Science*. 70(3), 233-238.
- Tilton, F. A., Bammler, T. K., & Gallagher, E. P. (2011). Swimming impairment and acetylcholinesterase inhibition in zebrafish exposed to copper or chlorpyrifos separately, or as mixtures. *Comparative Biochemistry and Physiology. Toxicology & Pharmacology : CBP*, 153(1), 9–16.

- Topal, A., Oruç, E., Altun, S., Ceyhun, S. B., & Atamanalp, M. (2016). The effects of acute boric acid treatment on gill, kidney and muscle tissues in juvenile rainbow trout. *Journal of Applied Animal Research*, 44(1), 297–302.
- Topal, A., Şişecioğlu, M., Atamanalp, M., Işık, A., & Yılmaz, B. (2016). The in vitro and in vivo effects of chlorpyrifos on acetylcholinesterase activity of rainbow trout brain. *Journal of Applied Animal Research*, 44(1), 243–247.
- Tripathi, A., & Srivastava, U. C. (2008). Acetylcholinesterase : A versatile enzyme of nervous system. Annals of Neurosciences, 15(4), 106–111.
- Tymoshenko, A., Tkach, G., Sikora, V., Bumeister, V., Shpetnyi, I., Lyndin, M., Maksymova, O., & Maslenko, A. (2016). The microscopic and ultramicroscopic changes in the skeletal muscles, caused by heavy metal salts. *Interventional Medicine & Applied Science*, 8(2), 82–88.
- Ullah, R., Zuberi, A., Naeem, M., & Ullah, S. (2015). Toxicity to hematology and morphology of liver, brain and gills during acute exposure of Mahseer (*Tor putitora*) to cypermethrin. *International Journal of Agricultural and Biology*, 17(1), 199–204
- Van Der Ent, A., & Edraki, M. (2016). Environmental geochemistry of the abandoned Mamut Copper Mine (Sabah) Malaysia. *Environmental Geochemistry and Health*. doi.org/10.1007/s10653-016-9892-3
- Velcheva, I., Tomova, E., Arnaudova, D., & Arnaudov, A. (2010). Morphological investigation on gills and liver of freshwater fish from dam lake 'Studen kladenets'. *Bulgarian Journal of Agricultural Science*, 16(3), 364–368.
- Velmurugan, B., Selvanayagam, M., Cengiz, E. I., & Unlu, E. (2009). Histopathological changes in the gill and liver tissues of freshwater fish, *Cirrhinus mrigala* exposed to dichlorvos. *Brazilian Archives of Biology and Technology*, 52(5), 1291–1296.
- Wallach, D., Kang, T. B., Dillon, C. P., & Green, D. R. (2016). Programmed necrosis in inflammation: Toward identification of the effector molecules. *Science*, 352(6281), 51–60.
- Wang, D., Zhang, M., Deng, S., Xu, W., Liu, Y., Geng, Z., & Liu, F. (2016). Postmortem changes in actomyosin dissociation, myofibril fragmentation and endogenous enzyme activities of grass carp (*Ctenopharyngodon idellus*) muscle. *Food Chemistry*, 197, 340–344.
- Wang, L., Lu, J., Sun, W., Gu, Y., Zhang, C., Jin, R., Li, L., Zhang, Z., & Tian, X. (2017). Hepatotoxicity induced by radix *Sophorae tonkinensis* in mice and increased serum cholinesterase as a potential supplemental biomarker for liver injury. *Experimental and Toxicologic Pathology*, 69(4), 193–202.
- Wang, T., Chen, X., Long, X., Liu, Z., & Yan, S. (2016). Copper nanoparticles and copper sulphate induced cytotoxicity in hepatocyte primary cultures of *Epinephelus coioides*. *PLoS ONE*, 11(2), https://doi.org/10.1371/journal.pone.0149484
- Wang, Z., Zhang, K., Zhao, J., Liu, X., & Xing, B. (2010). Adsorption and inhibition of butyrylcholinesterase by different engineered nanoparticles. *Chemosphere*, 79(1), 86 – 92.
- Wei, K., & Yang, J. (2015). Oxidative damage induced by copper and betacypermethrin in gill of the freshwater crayfish *Procambarus clarkii*. *Ecotoxicology and Environmental Safety*, 113, 446–453.

- Wong, S. K., Zhang, X. H., & Woo, N. Y. S. (2012). Vibrio alginolyticus thermolabile hemolysin (TLH) induces apoptosis, membrane vesiculation and necrosis in sea bream erythrocytes. *Aquaculture*, 330-333, 29–36.
- Xuereb, B., Noury, P., Felten, V., Garric, J., & Geffard, O. (2007). Cholinesterase activity in *Gammarus pulex* (Crustacea Amphipoda): Characterization and effects of chlorpyrifos. *Toxicology*, 236(3), 178–189.
- Yanong, R. (2009). Use of copper in marine aquaculture and aquarium systems [Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences]. Retrieved 28 October 2016, from http://edis.ifas.ufl.edu.
- Yap, C. K. (2014). A review on contamination of heavy metals, linear alkylbenzenes, polycyclic aromatic hydrocarbons, phenolic endocrine disrupting chemicals and organochlorine compounds in *Perna viridis* from the Coastal Waters of Malaysia: A compilation of 1998 Data. *International Journal of Advances in Applied Sciences*, 3(1), 1–10.
- Yaqin, K., & Hansen, P. D. (2010). The use of cholinergic biomarker, cholinesterase activity of blue mussel Mytilus edulis to detect the effects of organophosphorous pesticides. African Journal of Biochemistry Research, 4(12), 265–272.
- Younis, E. M., Abdel-Warith, A., Al-Asgah, N. A., Ebaid, H., & Mubarak, M. (2016). Histological changes in the liver and intestine of Nile tilapia, *Oreochromis niloticus*, exposed to sublethal concentrations of cadmium. *Pakistan Journal of Zoology*, 45(3), 833–841.
- Yousafzai, A. M., Ullah, F., Bari, F., Raziq, S., Riaz, M., Khan, K., Nishan, U., Sthanadar, I. A., Shaheen, B., Shaheen, M., & Ahmad, H. (2017). Bioaccumulation of some heavy metals: analysis and comparison of *Cyprinus* carpio and Labeo rohita from Sardaryab, Khyber Pakhtunkhwa. BioMed Research International, 2017. doi.org/10.1155/2017/5801432
- Yuan, S. S., Xu, H. Z., Liu, L. Q., & Zheng, J. L. (2017). Different effects of blue and red light-emitting diodes on antioxidant responses in the liver and ovary of zebrafish *Danio rerio*. Fish Physiology and Biochemistry, 43(2), 411–419. https://doi.org/10.1007/s10695-016-0296-1.
- Zhang, T. T., Li, W., Meng, G., Wang, P., & Liao, W. (2016). Strategies for transporting nanoparticles across the blood-brain barrier. *Biomaterials Science*, 4(2), 219–229.
- Zhang, T., Yang, M., Pan, H., Li, S., Ren, B., Ren, Z., Xing, N., Qi, L., Ren, Q., Xu, S., Song, J., & Ma, J. (2017). Does time difference of the acetylcholinesterase (AChE) inhibition in different tissues exist? A case study of zebra fish (*Danio rerio*) exposed to cadmium chloride and deltamethrin. *Chemosphere*, 168, 908–916.
- Zhou, Q., Zhang, J., Fu, J., Shi, J., & Jiang, G. (2008). Biomonitoring: An appealing tool for assessment of metal pollution in the aquatic ecosystem. *Analytica Chimica Acta*, 606(2), 135 – 150.