



**UNIVERSITI PUTRA MALAYSIA**

***BIOREMEDIATION OF RAW PALM OIL MILL EFFLUENT USING MIX  
MICROALGAE IN MALAYSIA***

**INAS JASIM ALWAN**

**FS 2016 29**



## **BIOREMEDIATION OF RAW PALM OIL MILL EFFLUENT USING MIX MICROALGAE IN MALAYSIA**

By

**INAS JASIM ALWAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Master of Science**

**May 2016**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in  
fulfillment of the requirement for the Degree of Master of Science

## **BIOREMEDIATION OF RAW PALM OIL MILL EFFLUENT USING MIX MICROALGAE IN MALAYSIA**

By

**INAS JASIM ALWAN**

**May 2016**

**Chairman : Hishamuddin Omar, PhD**  
**Faculty : Science**

Palm oil industry in Malaysia is one of the key players in the contribution to the economy of the country. Nonetheless, it also produces a massive amount of palm oil mill effluent (POME). It has been known that microalgae, the unicellular photosynthetic microorganisms possess the ability to reduce the number of pollutants of organic waste. Therefore, this study aims to grow mixed microalgae cultured in different concentrations of raw POME (0.5, 1, 1.5 and 2) % v/v as a biological treatment. This study is conducted on open pond cultivation system (glass tanks) in outdoor conditions. First, it is necessary to allow the mixed microalgae make an adaptation in the raw POME before the start of the actual experiment. The results showed positive effects on the biomass growth of mixed microalgae cultured in the concentration of 1% v/v raw POME. The concentration 1% v/v recorded productivity  $0.09 \pm 0.02$  g/L. day, which significantly higher  $p < 0.05$  compared with the controls and other concentrations. The findings showed that the mixed microalgae could reduce BOD, TN and TP to 78.79, 2.176, and 1.591 respectively. Finally, as many as 10 species of mixed microalgae have been identified from all the experiments, namely *Dictyosphaerium* sp., *Scenedesmus* sp., *Oocystis* sp., *Monoraphidium* sp., *Nostoc* sp., *Cosmarium* sp., *Aulacoseira* sp., *Crucigenia* sp., *Cyclotella* sp., and *Asterococcus* sp.. Since, it is proven that the mixed microalgae able to grow in raw POME, it can be concluded that raw POME is a suitable medium for the cultivation of mixed microalgae without adverse effects.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk Ijazah Master Sains

**BIOREMEDIASI EFLUEN KILANG MINYAK SAWIT MENTAH DENGAN  
MENGUNAKAN CAMPURAN MIKROALGA  
DI MALAYSIA**

Oleh

**INAS JASIM ALWAN**

**Mei 2016**

**Pengerusi : Hishamuddin Omar, PhD**  
**Fakulti : Sains**

Industri minyak sawit di Malaysia adalah salah satu penyumbang utama kepada ekonomi negara. Namun begitu, ia juga menghasilkan sejumlah besar efluen kilang minyak sawit (POME). Adalah terdahul merdapat bahawa mikroalga, iaitu mikroorganisma fotosintesis berunisel, mempunyai keupayaan untuk mengurangkan jumlah pencemar sisa organik. Oleh itu, kajian ini bertujuan meng kultur mikroalga campuran dalam kepekatan POME mentah yang berlainan (0.5, 1, 1.5 dan 2) % v/v sebagai rawatan biologi. Kajian ini dijalankan ke atas sistem kolam terbuka (tangki kaca) system terbuka. Pertama, mikroalga campuran itu perlu diaklimitasikan penyesuaian untuk POME mentah sebelum memulakan eksperimen. Hasil kajian menunjukkan kesan positif ke atas pertumbuhan biojisim pada mikroalga campuran yang telah dikultur dalam kepekatan 1% v/v POME mentah. Kepekatan 1% v/v mencatatkan produktiviti  $0.09 \pm 0.02$  g/L /hari, yang jauh lebih tinggi ( $p < 0.05$ ) berbanding dengan kawalan dan kepekatan lain. Kajian juga menunjukkan bahawa mikroalga campuran boleh mengurangkan BOD, TN dan TP hingga 78.79, 2.176, dan 1.591 masing-masing. Sebanyak 10 spesies alga campuran telah dikenal pasti dari semua eksperimen, iaitu *Dictyosphaerium* sp., *Scendesmus* sp., *Oocystis* sp., *Monoraphidium* sp., *Nostoc* sp., *Cosmarium* sp., *Aulacoseira* sp., *Crucigenia* sp., *Cyclotella* sp., dan *Asterococcus* sp. Kajian ini dapat membuktikan mikroalga campuran dapat bahawa membiak dan tumbuh dalam POME mentah. Sebagai kesimpulan, POME mentah adalah medium yang sesuai untuk perkembunan mikroalga campuran tanpa memberikan kesan sampingan

## ACKNOWLEDGEMENTS

First praise is to Almighty Allah for all his blessings for giving me patience, guidance, knowledge and good health throughout the duration of this master research.

I would like to express my deepest appreciation and gratitude to my wonderful supervisor Dr. Hishamuddin Bin Omar and Prof. Ahmad Ismail for their excellent caring, patience, invaluable supervision, guidance, and support me till finishing this research. As well, I would like to thank the department of Biology and Faculty of Science at the Universiti Putra Malaysia.

Finally, I would also like to thank my father and mother. They were always supporting me and encouraging me with their best wishes. I would never have been able to finish my thesis without the help and support of my husband, and support from my family, my lovely sons and my daughter. They were always there cheering me up and stood by me through the good times and bad.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master Science. The members of the Supervisory Committee were as follows:

**Hishamuddin Omar, PhD**

Senior Lecturer  
Faculty of Science  
Universiti Putra Malaysia  
(Chairman)

**Ahmad Ismail, PhD**

Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

---

**BUJANG KIM HUAT, PhD**

Professor and dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_

Date: \_\_\_\_\_

Name and Matric No.: Inas Jasim Alwan GS40427



## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature : \_\_\_\_\_  
Name of  
Chairman of  
Supervisory : Senior Lecturer  
Committee : Dr. Hishamuddin Omar

Signature : \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee : Dr. Ahmad Ismail

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	ii
<b>ACKNOWLEDGEMENTS</b>	iii
<b>APPROVAL</b>	iv
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	xi
<b>LIST OF FIGURES</b>	xii
<b>LIST OF APPENDICES</b>	xiv
<b>LIST OF ABBREVIATIONS</b>	xv
 <b>CHAPTER</b>	
 <b>1 INTRODUCTION</b>	 <b>1</b>
1.1 Background	1
1.2 Problem Statement	3
1.3 Objectives	3
 <b>2 LITERATURE REVIEW</b>	 <b>5</b>
2.1 Status of Palm Oil in Malaysia	5
2.2 Palm Oil Plantation contributes to Global Warming	5
2.3 Palm Oil Mill Effluent (POME)	7
2.3.1 POME Production	7
2.3.2 Characteristics and Composition of POME	8
2.3.3 POME as waste	10
2.3.4 POME as a resource	11
2.3.5 Treatment methods used for Treatment of POME	12
2.4 Microalgae	14
2.4.1 Types of microalgae growth process	15
2.4.2 Factors that Affect Algal Growth	16
2.4.3 Nutritional Requirements for Microalgae Culture	21
2.4.4 Growth Monitoring and Biochemical Composition	24
2.4.5 Cultivation System of microalgae	26
2.4.6 Water Remediation Using Microalgae	29
2.4.7 Using of Microalgae for Treatment of POME	30
 <b>3 MATERIALS AND METHODS</b>	 <b>33</b>
3.1 Experimental Design	33
3.1.1 Raw POME collection	34
3.1.2 Mixed Microalgae source	34
3.1.3 Acclimatization and Maintenance of Mixed Microalgae	35
3.1.4 Culture Condition for Mixed Microalgae	36
3.1.5 Aeration and Mixing in Cultivation of Mixed Microalgae	39

3.2	Measurement of water quality of raw POME before treatment	39
3.2.1	Measurements of Physical Parameters	39
3.2.2	Measurements of Chemical Parameters	40
3.3	Measurement of the Water Quality of Mixed Microalgae in Raw POME culture	44
3.3.1	Physical Parameters	44
3.3.2	Chemical Parameters	44
3.4	Measurement of the biomass Productivity of Mixed Microalgae	44
3.4.1	Optical Density (OD)	44
3.4.2	Cells Dry Weight of (DW)	44
3.4.3	Chlorophyll a Content	45
3.5	Diversity of Mixed Microalgae	46
3.5.1	Identification and Quantification of Mixed Microalgae	46
3.5.2	Cells Counting of Mixed Microalgae by Using Hemacytometer and Microscope	46
3.5.3	Diversity of Mixed Microalgae	46
3.6	Data Analysis	48
<b>4</b>	<b>RESULTS</b>	49
4.1	The Adaptation for The Cultivation of Mixed Microalgae	49
4.2	Water Quality Before and After Treatment	49
4.2.1	Physical Parameters	50
4.2.1.1	Weather condition	50
4.2.1.2	Water Quality	53
4.2.2	Chemical Parameters	60
4.3	Productivity	65
4.3.1	Optical Density of Mixed Microalgae in Different Concentration of Raw POME Culture	65
4.3.2	Cell Dry Weight (DW) for Mixed Microalgae in Different Concentration of Raw POME Culture	66
4.3.3	Chlorophyll a Content in Mixed Microalgae in Different Concentration of Raw POME	68
4.4	Diversity of Mixed Microalgae Species	69
4.4.1	List of Species	70
4.4.2	Dominant Species	75
4.4.3	Diversity Index	75
<b>5</b>	<b>DISCUSSION</b>	84
5.1	Adaptation of Mixed Microalgae in Raw POME	84
5.1.1	Adaptation of Mixed Microalgae	84
5.1.2	Raw POME as Growth Medium	84
5.1.3	The Improvement of Water Quality	85
5.2	Possible Contribution of Other Microbes in Raw POME Remediation	85
5.3	Effect of Weather Conditions	85
5.4	Effect of Raw POME Concentration on the Growth of	86

	Mixed Microalgae	
5.5	Effect of Mixed Microalgae Growth of Water Quality	87
5.6	The interaction between Weather Conditions and Raw POME Concentration	88
<b>6</b>	<b>CONCLUSION AND RECOMMENDATIONS FOR FUTURE WORKS</b>	<b>89</b>
6.1	Introduction	89
6.2	Recommendations and Future works	90
	<b>REFERENCES</b>	<b>91</b>
	<b>APPENDICES</b>	<b>102</b>
	<b>BIODATA OF STUDENT</b>	<b>129</b>



## LIST OF TABLES

Table	Page
2.1	9
2.2	29
4.1	53
4.13a	76
4.13b	77
4.13c	78
4.13d	79
4.14a	80
4.14b	81
4.14c	82
4.14d	83

## LIST OF FIGURES

Figure		Page
2.1	Flow chart of the process in Palm Oil Mill (Lam and Lee,2011)	8
2.2	Components of typical microalgae (Singh and Gu, 2010)	26
2.3	Overview design of POME treatment in corporation with ,microalgae culture (Lam and Lee,2011)	31
3.1	Summary of the experimental design	33
3.2	Fresh Palm Oil Mill effluent(POME) is stored in 20L plastic tank	34
3.3	Hatchery of Fisheries in Universiti Putra Malaysia (UPM)	35
3.4	Stages of acclimatization and maintenance for mixed microalgae	36
3.5	Concentration for each treatment	37
3.6	Randomized Complete Block Design (RCBD) arrangement	38
3.7	Preparation of aquarium glass tank	38
3.8	Procedure for measurement of BOD <sub>5</sub>	42
4.1	Adaptation stages of mixed microalgae	49
4.2 a and b	Mean values of air temperature during the cultivation period of cycles 1and 2	51
4.3 a and b	Mean values of the light intensity during the cultivation period of cycles 1 and 2	52
4.4 a and b	Mean values of pH of the cultivation in different concentration of raw POME from culture cycles 1 and 2	55
4.5 a and b	Mean values of water temperature of the cultivation in different concentration of raw POME from culture cycles 1 and 2	56
4.6 a and b	Mean values of water conductivity of cultivation in different concentrations of raw POME from culture cycles 1 and 2	57
4.7 a and b	Means values of dissolved oxygen of cultivation in different concentrations of raw POME from culture cycles 1and 2	58
4.8 a and b	Means values of total dissolved solids of cultivation in different concentrations of raw POME from culture cycles 1and 2	59
4.9 a and b	Mean values of total nitrogen (mg/L) of cultures in different concentration of raw POME from culture cycles 1and 2	61
4.10 a and b	Mean values of total phosphorus (mg/L) of cultures in different concentration of raw POME from culture cycles 1and 2	62

4.11 a and b	Mean values of biochemical oxygen demand of cultures in different concentration of raw POME from culture cycles 1 and 2	63
4.12 a and b	Mean values of CO <sub>2</sub> fixation of cultures in different concentration of raw POME from culture cycles 1 and 2	64
4.13 a and b	Mean values of optical density of mixed microalgae cultures in different concentration of raw POME from culture cycles 1 and 2	66
4.14 a and b	Mean values of dry weight of mixed microalgae cultures in different concentration of raw POME from culture cycles 1 and 2	67
4.15 a and b	Mean values of chlorophyll <i>a</i> content of mixed microalgae cultures in different concentration of raw POME from culture cycles 1 and 2	69
4.16	Mixed microalgae species obtained from culture cycles 1 and 2	74

## LIST OF APPENDICES

Appendix	Page
4.2 Mean values of pH for all treatments for the duration of 10 days (Treatment $\pm$ SD, n=3 cycle 1 and 2).	102
4.3 Mean values of water conductivity for all the treatments for duration of 10 days (Treatment $\pm$ SD, n=3 cycle 1 and 2)	103
4.4 Mean values of Dissolved Oxygen for all the treatments for duration of 10 days (Treatment $\pm$ SD, n=3 cycle 1 and 2).	104
4.5 Mean values of total dissolved solids for all the treatment for duration of 10 days (Treatment $\pm$ SD, n=3 cycle 1 and 2)	105
4.6 Mean values of total nitrogen for all the treatments duration of 10 days (Treatment $\pm$ SD, n=3, cycle 1 and 2)	106
4.7 Mean values of total phosphorus for all the treatments for the duration of 10 days (Treatment $\pm$ SD, n=3, cycles 1 and 2)	107
4.8 Mean value of biochemical oxygen demand for all treatments in 5 and 10 days (Treatment $\pm$ SD, n=3, cycles 1 and 2)	108
4.9 Mean values of CO <sub>2</sub> fixation for all the treatments of mixed microalgae for the duration of day 5 and 10 (Treatment $\pm$ SD, n=3, cycles 1 and 2)	109
4.10 Mean values of optical density for all the treatments for the duration of 10 days (Treatment $\pm$ SD, n=3 cycle 1 and 2)	110
4.11 Mean values of cell dry weight for all the treatments for the duration of 10 days (Treatment $\pm$ SD, n=3, cycles 1 and 2)	111
4.12 Mean value of Chlorophyll a content of mixed microalgae for all the treatments for the duration of 10 days (Treatment $\pm$ SD, n=3, cycle 1 and 2)	112



## LIST OF ABBREVIATIONS

ANOVA	Analysis of Variance
ATP	Adenosine triphosphate
BOD <sub>5</sub>	Biochemical oxygen demand
C	Carbon
Ca	Calcium
CH <sub>4</sub>	Methane
Co	Cobalt
CO <sub>2</sub>	Carbon dioxide
COD	Chemical oxygen demand
CPO	Crude Palm oil
Chll-a	Chlorophyll <i>a</i>
Cu	Copper
D	Simpsons index
DDH <sub>2</sub> O	Deionized water
DO	Dissolved Oxygen
DW	Dry weight
E	Evenness
EFB	Empty fruit bunches
Fe	Iron
FFB	Fresh fruit bunch
FEP	Teflon
FFB	Fresh fruit bunches
g	Gram
GHG	Green houses gases
HCO <sub>3</sub>	Bicarbonate
HCl	Hydrochloric acid
H <sub>2</sub> SO <sub>4</sub>	Sulphuric acid
K	Potassium
L	Litter
Mg	magnesium
Mn	manganese
ml	mili litter
MPOB	Malaysian Palm Oil Board
mg	Milligram
mgL <sup>-1</sup>	Milligram per litre
µm	Micro meter
µl	Micro litre
ln	Natural logarithm
MUFA	Mono unsaturated fatty acid
N	Nitrogen
N	Total number of individual
n	Total number of species
Na	Sodium
NADPH	nicotinamide adenine dinucleotide phosphate- oxidase
nm	Nanometre
NH <sub>3</sub> -N	Ammoniac nitrogen
NaOH	Sodium Hydroxide

O <sub>2</sub>	Oxygen
P	Phosphorous
PAR	Photosynthetic active radiation
PBR	Photo- bioreactor
Pi	Proportion of individuals
POME	Palm Oil Mill Effluent
ppt.	Parts per thousand
PUFA	Poly unsaturated fatty acid
rpm	Revolution per minute
R	Richness
RCBD	Randomized Complete Block Design
Sp.	Species
TN	Total Nitrogen
TP	Total Phosphorus
OD	Optical Density
UPM	Universiti Putra Malaysia
UNIDO	United Nation Industrial Development Organization
UV	Ultra violet
WHO	World Health Organisation
YSI	Yellow spring instrument
Zn	Zinc

## CHAPTER 1

### INTRODUCTION

#### 1.1 Background

Nowadays, the world is facing very serious global issues related to the environment. The main problem is global warming, climate change, environmental degradation, severe drought, acid rain and haze (Lee *et al.*, 2013; Naidu *et al.*, 2015). Many of these problems originate from a single source, namely global warming. This phenomenon of global warming results from excessive release of CO<sub>2</sub> and CH<sub>4</sub>, which form greenhouse gases that insulate heat from escaping the earth atmosphere, leading to more global warming (de Richter and Caillol, 2011; Yun *et al.*, 2012; Lee *et al.*, 2013). Mitigating of global warming requires multiple approaches ranging from activities in reducing CO<sub>2</sub> and CH<sub>4</sub> emission, to improving agriculture practice and agroforestry systems (Dhillon and von Wuehlisch, 2013; Crank and Jacoby, 2015). With respect to global warming, Malaysia is also in the spotlight due to palm oil industry (Singh *et al.*, 2010; Madaki and Seng, 2013).

In Malaysia, one fast growing agriculture-based sector is the palm oil industry. It is revealed that this industry controls about 45 % of total palm oil production in the world (Saifuddin and Dinara, 2011; Sukumaran *et al.*, 2014). In the past decade, Malaysia has risen to become a leading global producer and exporter of palm oil products. The Malaysian Palm Oil Board (MPOB) revealed that the production of crude palm oil (CPO) has recorded an increase of 2.3 % from 18.79 million tons in 2012 to 19.22 million tons in 2013 (MPOB, 2015).

In Malaysia, the most practical and feasible method for large-scale production in palm oil is the wet process in palm oil milling. As suggested in the name, this method requires a large quantity of water resources in the production of crude palm oil (CPO) from processing the fresh fruit bunches to become crude oil. This method has produced palm oil mill effluent (POME) which is considered as wastewater (Singh *et al.*, 2010; Wu *et al.*, 2010). According to Ahmad *et al.* (2003), about 5 - 5.7 tons of water is required to produce 1 ton of CPO. This process involves the sterilization of the palm fruit bunches and clarifying the extracted oil. However, more than 50 % of the water ends up as fresh POME. This means that for one ton of CPO obtained in the extraction process, a number of POME beings produced is 2.5 - 3.0 tons. This suggests that the amount of POME will increase as long as the production and processing of palm oil continue the upward trend to meet both domestic and global demand (Madaki and Seng, 2013).

The quality of the raw materials used and the processes of palm oil production in the mills affect the characteristics of POME (Eze *et al.*, 2013). POME as a thick brownish colloidal mixture. This composition of the mixture includes water, oil, and fine suspended solids. It has 80 - 90 °C; the levels of both biochemical oxygen demand

(BOD) and chemical oxygen demand (COD) are very high. The presence of organic acids in complex forms makes it acidic with a pH of around 4.5. This makes the decomposition process of POME difficult to achieve due to their cost effective processing (Habib *et al.*, 1998; Igwe and Onyegbado, 2007; Wu *et al.*, 2010). While POME is considered a waste, it is considered a load rather than a part of the production process of palm oil. Nowadays, the biggest challenge faced by the palm oil industry is related to the disposal of large quantities of POME (Igwe and Onyegbado, 2007; Madaki and Seng, 2013). One of the most common and environmentally friendly methods for the disposal of wastes palm produced from the oil mill industry is the discharge of raw or partially treated POME into the waterways. Nevertheless, this method may cause serious environmental threats since discharge of POME not only pollutes fresh water bodies, but its retention can also be harmful to the environment (Wu *et al.*, 2010). This is because during the retention period in a series of oxidation ponds take up to 155 days before it is being discharged into the rivers. During the oxidation process, POME undergoes fermentation that releases CO<sub>2</sub> and CH<sub>4</sub>, which contribute to the increase in greenhouse gasses (Igwe and Onyegbado, 2007; Singh *et al.*, 2010; Madaki and Seng, 2013). Due to exceptionally high BOD levels in POME, it has been recognized as one of the largest source of pollutants into the rivers in Malaysia (Habib *et al.*, 1998; Singh *et al.*, 2010; Madaki and Seng, 2013). On the other hand, the high organic matter present in POME, which includes carbohydrates, protein, phosphorus, potassium, magnesium and calcium make it as a vital nutrient resource (Habib *et al.*, 1998; Madaki and Seng, 2013).

Based on these findings, many research studies have started the search for alternative solutions on the POME conversion into useful forms such as animal feed, biofertilizer and a source of green energy, for the benefits of mankind including a decrease in environmental pollution Foo and Hameed, (2010);Rupani *et al.*, (2010); Lam and Lee, (2011); Chin *et al.*,(2013).

There are many suggestions to utilize POME such as convert this waste product into beneficial substance namely feedstock and organic fertilizers using the biological method of vermicomposting (Rupani *et al.* 2010). Also, POME can be treated by using the physical method that uses membrane ultrafiltration. It is an excellent way to treat wastewater into the best sample quality (Wah *et al.* 2002). Furthermore, through the use of anaerobic thermophilic digestion process POME can be converted to release biogas(Madaki and Seng, 2013). The other method relies on the use of chemical substances. For example, the flocculation process involves the removed of a substance such as oil, alum, aluminum chlorohydrin, aluminum sulfate and chitosan (Saifuddin and Dinara, 2011).

Recently, microalgae and unicellular photosynthetic microorganisms have been highlighted as attractive sources of renewable energy due to their incredible abilities; in rapid production of biomass and a significant reduction for pollutants (Lam and Lee, 2011). In general, microalgae absorb solar energy, while consuming organic compounds to fix carbon substances, besides producing biomass (Vairappan and Yen, 2008; Wolkers *et al.*, 2011; Sukumaran *et al.*, 2014). It is reported that microalgae grow 100 times faster than land-based plants, so they have the ability to double

biomass production in less than a day (Lam and Lee, 2011). Another feature of microalgae is their ability to utilize organic pollutants, and subsequently improve water quality (Priyadarshani *et al.*, 2011; Othman *et al.*, 2014).

Therefore, many research has adopted the suggestion of converting the treatment of POME, from disposed waste to beneficial activities (Azimatun Nur and Hadiyanto, 2014; Nwuche *et al.*, 2014; Shaha *et al.*, 2014; Sukumaran *et al.*, 2014). In line with this suggestion, many research works used microalgae in the bioremediation of POME. The researchers utilized POME as a low cost. However, most of these research works focused on specific types of microalgae, resulting in various shortcomings. One of the reasons is variation in the composition of POME from different sources which be favorable the growth of certain species of microalgae. Another reason is the pre-determination of the optimum concentration of POME and the cultivation environment for that particular species (Ponraj and Din, 2013; Azimatun Nur and Hadiyanto, 2014; Nwuche *et al.*, 2014). Therefore, the use of more than one specific species of microalgae in the form of mixed microalgae would be a more efficient bioremediation method for the treatment of POME (Pandey and Tiwari, 2010; Kshirsagar, 2013; Shaha *et al.*, 2014).

Hence, this research study proposes the use of different species of microalgae in a form of mixed microalgae that might offer the better ability to remediate the raw POME than the use of a single species of microalgae. Also, the use of microalgae could reduce environmental pollution with the potential to be a great source for many biofuels and renewable green energy. As such, this research study attempts to use raw POME, which is considered a waste pollutant, as a cheap and vital source of nutrient to grow the mixed microalgae in order to determine its physical, chemical and productivity characteristics as an important source of green energy.

## **1.2 Problem Statement**

While POME is considered a waste, it is considered a load rather than a part of the production process of palm oil. Therefore, the choice for the cheapest method of disposal is the continuous discharge of POME into nearby rivers. But, this of method of disposal increases the risk of environmental pollution. Many research used microalgae in the bioremediation of POME. However, most of the research works focused on specific types of microalgae. This research study processes the use of mixed microalgae to remediate the raw POME rather than the use of single species microalgae. This Proposal not only reduces environmental pollution, but it is also a great source for many biofuels and renewable green energy.

## **1.3 Objectives**

The main objective of this research study is to determine the extent of the ability of the use of mixed microalgae to remediate raw POME and consequently to show a decrease in pollution and an enhancement of water quality. Thus, the main objectives of this study are :

1. To characterize raw POME at different concentrations and compare it before and after the cultivation of mixed microalgae.
2. To determine the productivity and species diversity of mixed microalgae in different concentrations of raw POME.





## REFERENCES

- Abdurahman, N. H., Rosli, Y. M., and Azhari, N. H. (2013). The performance evaluation of anaerobic methods for palm oil mill effluent (POME) treatment: a review. In W. T. Q. Nigel (Ed.), International perspectives on water quality management and pollutant control.
- Abedini Najafabadi, H., Malekzadeh, M., Jalilian, F., Vossoughi, M., and Pazuki, G. (2015). Effect of various carbon sources on biomass and lipid production of *Chlorella vulgaris* during nutrient sufficient and nitrogen starvation conditions. *Bioresource technology*, 180(0), 311-317.
- Ahmad, A. L., Ismail, S., and Bhatia, S. (2003). Water recycling from palm oil mill effluent (POME) using membrane technology. *Desalination*, 157(1-3), 87-95.
- Ahmad, F., Khan, A. U., and Yasar, A. (2012). Uptake of nutrients from municipal wastewater and biodiesel production by mixed algae culture. *Pak J Nutr*, 11(7), 648-652.
- Al-Qasbi, M., Raut, N., Talebi, S., Al-Rajhi, S., and Al-Barwani, T. (2012). A review of effect of light on microalgae growth. Paper presented at the Proceedings of the world congress on engineering.
- Asulabh, K., Supriya, G., and Ramachandra, T. (2012, 6th-9th November 2012). Effect of Salinity Concentrations on Growth Rate and Lipid Concentration in *Microcystis* Sp., *Chlorococcum* Sp. and *Chaetoceros* Sp. Paper presented at the National Conference on Conservation and Management of Wetland Ecosystems. School of Environmental Sciences, Mahatma Gandhi University, Kottayam, Kerala, India.
- Atekwana, E. A., Atekwana, E. A., Rowe, R. S., Werkema, D. D., & Legall, F. D. (2004). The relationship of total dissolved solids measurements to bulk electrical conductivity in an aquifer contaminated with hydrocarbon. *Journal of Applied Geophysics*, 56(4), 281-294.
- Azarpira, H., Behdarvand, P., Dhumal, K., & Pondhe, G. (2014). Potential use of cyanobacteria species in phycoremediation of municipal wastewater. *International Journal of Biosciences (IJB)*, 4(4), 105-111.
- Azimatus Nur, M. M., and Hadiyanto. (2014). Evaluation of carbon, nitrogen and phosphorus ratio of palm oil mill effluent digested (POMED) wastewater as replacement synthetic medium for *Spirulina* Sp growth. *American-Eurasian Journal of Agricultural and Environmental Sciences*, 14(6), 536-540.
- Benemann, J. (2013). Microalgae for biofuels and animal feeds. *Energies*, 6(11), 5869-5886.

- Blair, M. F., Kokabian, B., and Gude, V. G. (2014). Light and growth medium effect on *Chlorella vulgaris* biomass production. *Journal of Environmental Chemical Engineering*, 2(1), 665-674.
- Blinová L., Bartošová, A. & Gerulová, K. 2015. Cultivation of Microalgae (*Chlorella Vulgaris*) for Biodiesel Production. *Research Papers Faculty of Materials Science and Technology Slovak University of Technology* 23(36): 87-95.
- Brennan, L., and Owende, P. (2010). Biofuels from microalgae—A review of technologies for production, processing, and extractions of biofuels and co-products. *Renewable and Sustainable Energy Reviews*, 14(2), 557-577.
- Carvalho, A. P., Monteiro, C. M., and Malcata, F. X. (2009). Simultaneous effect of irradiance and temperature on biochemical composition of the microalga *Pavlova lutheri*. *Journal of applied phycology*, 21(5), 543-552.
- Carvalho, A. P., Silva, S. O., Baptista, J. M., and Malcata, F. X. (2011). Light requirements in microalgal photobioreactors: an overview of biophotonic aspects. *Applied microbiology and biotechnology*, 89(5), 1275-1288.
- Cheah, W. Y., Show, P. L., Chang, J.-S., Ling, T. C. & Juan, J. C. 2015. Biosequestration of Atmospheric CO<sub>2</sub> and Flue Gas-Containing CO<sub>2</sub> by Microalgae. *Bioresource technology* 184(190-201).
- Chekroun, K. B., Sánchez, E., and Baghour, M. (2014). The role of algae in bioremediation of organic pollutants. *International Research Journal of Public and Environmental Health*, 1(2), 19-32.
- Chin, M. J., Poh, P. E., Tey, B. T., Chan, E. S., and Chin, K. L. (2013). Biogas from palm oil mill effluent (POME): opportunities and challenges from Malaysia's perspective. *Renewable and Sustainable Energy Reviews*, 26(0), 717-726.
- Chinnasamy, S., Ramakrishnan, B., Bhatnagar, A., and Das, K. C. (2009). Biomass production potential of a wastewater alga *Chlorella vulgaris* ARC 1 under elevated levels of CO<sub>2</sub> and temperature. *International Journal of Molecular Sciences*, 10(2), 518-532.
- Choi, S.-L., Suh, I. S., and Lee, C.-G. (2003). Lumostatic operation of bubble column photobioreactors for *Haematococcus pluvialis* cultures using a specific light uptake rate as a control parameter. *Enzyme and microbial Technology*, 33(4), 403-409.
- Chu, F.-F., Chu, P.-N., Cai, P.-J., Li, W.-W., Lam, P. K. S., and Zeng, R. J. (2013). Phosphorus plays an important role in enhancing biodiesel productivity of *Chlorella vulgaris* under nitrogen deficiency. *Bioresource technology*, 134(0), 341-346.



- Converti, A., Casazza, A. A., Ortiz, E. Y., Perego, P., and Del Borghi, M. (2009). Effect of temperature and nitrogen concentration on the growth and lipid content of *Nannochloropsis oculata* and *Chlorella vulgaris* for biodiesel production. *Chemical Engineering and Processing: Process Intensification*, 48(6), 1146-1151.
- Creswell, L. (2010). Phytoplankton culture for aquaculture feed. Southern Regional Aquaculture Center.
- Crank, J. P., and Jacoby, L. S. (2015). 1 - The Challenges of Global Warming Research. In J. P. C. S. Jacoby (Ed.), *Crime, Violence, and Global Warming* (pp. 3-21): Anderson Publishing, Ltd.
- Cui, X., Gao, F., Song, J., Sang, Y., Sun, J., and Di, X. (2014). Changes in soil total organic carbon after an experimental fire in a cold temperate coniferous forest: A sequenced monitoring approach. *Geoderma*, 226–227(0), 260-269.
- de Richter, R., and Caillol, S. (2011). Fighting global warming: The potential of photocatalysis against CO<sub>2</sub>, CH<sub>4</sub>, N<sub>2</sub>O, CFCs, tropospheric O<sub>3</sub>, BC and other major contributors to climate change. *Journal of Photochemistry and Photobiology C: Photochemistry Reviews*, 12(1), 1-19.
- Dhillon, R. S., and von Wuehlisch, G. (2013). Mitigation of global warming through renewable biomass. *Biomass and Bioenergy*, 48(0), 75-89.
- Dominic, V., Murali, S., and Nisha, M. (2009). Phycoremediation efficiency of three microalgae *Chlorella vulgaris*, *Synechocystis salina* and *Gloeocapsa gelatinosa*. *Academic Review*, 16, 138-146.
- Dubey, S. K., Dubey, J., Mehra, S., Tiwari, P., and Bishwas, A. (2011). Potential use of cyanobacterial species in bioremediation of industrial effluents. *African journal of Biotechnology*, 10(7), 1125-1132.
- Eze, V., Owunna, N., and Avoaja, D. (2013). Microbiological and Physicochemical Characteristics of Soil Receiving Palm Oil Mill Effluent in Umuahia, Abia State, Nigeria. *Journal of Natural Sciences Research*, 3(7), 163-169.
- Fernandes, B. D., Mota, A., Teixeira, J. A., and Vicente, A. A. (2015). Continuous cultivation of photosynthetic microorganisms: Approaches, applications and future trends. *Biotechnology Advances*(0), <http://dx.doi.org/10.1016/j.biotechadv.2015.1003.1004>.
- Foo, K. Y., and Hameed, B. H. (2010). Insight into the applications of palm oil mill effluent: A renewable utilization of the industrial agricultural waste. *Renewable and Sustainable Energy Reviews*, 14(5), 1445-1452.
- Gacheva, G., and Gigova, L. (2014). Biological activity of microalgae can be enhanced by manipulating the cultivation temperature and irradiance. *Central European Journal of Biology*, 9(12), 1168-1181.

- Garofalo, R. (2009). Algae and aquatic biomass for a sustainable production of 2nd generation biofuels. *AquaFUELS-Taxonomy, Biology and Biotechnology*, 1-258.
- Germer, J., and Sauerborn, J. (2008). Estimation of the impact of oil palm plantation establishment on greenhouse gas balance. *Environment, Development and Sustainability*, 10(6), 697-716.
- Gobi, K., and Vadivelu, V. M. (2013). By-products of palm oil mill effluent treatment plant – A step towards sustainability. *Renewable and Sustainable Energy Reviews*, 28(0), 788-803.
- Gonzalez-Perez, J. A., Gonzalez-Vila, F. J., Almendros, G., and Knicker, H. (2004). The effect of fire on soil organic matter—a review. *Environment International*, 30(6), 855-870.
- Habib, M., Yusoff, F., Phang, S., Kamarudin, M., and Mohmed, S. (1998). Chemical characteristics and essential nutrients of agroindustrial effluents in Malaysia. *Asian Fisheries Science. Metro Manila*, 11(3), 279-286.
- Hadiyanto, H., Nur, M. M. A., and Hartanto, G. D. (2012). Cultivation of *Chlorella* sp. as biofuel sources in palm oil mill effluent (POME). *International Journal of Renewable Energy Development (IJRED)*, 1(2), 45-49.
- Hadiyanto, M. C., and Soetrisnanto, D. (2013). Phytoremediations of palm oil mill effluent (POME) by using aquatic plants and microalgae for biomass production. *Journal of Environmental Science and Technology*, 6(2), 79-90.
- Ibrahim, A. H., Dahlan, I., Adlan, M. N., Dasti, A. F., Nirja, G., Prachi, P., ... & Suman, C. (2012). Comparative study on characterization of Malaysian palm oil mill effluent. *Res J Chem Sci*, 2(12), 1-5.
- Igwe, J., and Onyegbado, C. (2007). A review of palm oil mill effluent (POME) water treatment. *Global Journal of Environmental Research*, 1(2), 54-62.
- Ikaran, Z., Suárez-Alvarez, S., Urreta, I., and Castañón, S. (2015). The effect of nitrogen limitation on the physiology and metabolism of *Chlorella vulgaris* var L3. *Algal Research*, 10(0), 134-144.
- Janse, V. V. S., Taylor, J., Gerber, A., & Van Ginkel, C. (2006). Easy Identification of the Most Common Freshwater Algae. A Guide for the Identification of Microscopic Algae in South African Freshwaters. North-West University, Potchefstroom, South Africa. ISBN 0-621-35471-6.
- Job Gopinath.M., Kaveriammal.S and Elayarasi.M, (2014). Phytoremediation of domestic waste water of Gudiyatham town by Microalgae *Chlorella vulgaris*. *International Journal of Environmental Biology*.

- Juneja, A., Ceballos, R. M., & Murthy, G. S. (2013). Effects of environmental factors and nutrient availability on the biochemical composition of algae for biofuels production: a review. *Energies*, 6(9), 4607-4638.
- Kennedy, A. G., & Hishamuddin, O. (2001). Bioremediation of treated and raw POME (Palm Oil Mill Effluent) using *Spirulina plantensis*. *Aquatic resources and environment studies of the straits of Malacca: current research and reviews*, Kuala Lumpur, 203-210.
- Kshirsagar, A. D. (2013). Bioremediation of wastewater by using microalgae: An experimental study. *International Journal of Life Sciences Biotechnology and Pharma Research*, 2(3), 338-346.
- Kumar, M., Kulshreshtha, J., & Singh, G. P. (2011). Growth and biopigment accumulation of cyanobacterium *Spirulina platensis* at different light intensities and temperature. *Brazilian Journal of Microbiology*, 42(3), 1128-1135.
- Lam, M. K., and Lee, K. T. (2011). Renewable and sustainable bioenergies production from palm oil mill effluent (POME): Win-win strategies toward better environmental protection. *Biotechnology Advances*, 29(1), 124-141.
- Larsdotter, K. (2006) Wastewater treatment with microalgae-a literature review. *Vatten*, 62(1), 31.
- Latif Ahmad, A., Ismail, S., and Bhatia, S. (2003). Water recycling from palm oil mill effluent (POME) using membrane technology. *Desalination*, 157(1-3), 87-95.
- Lee, H.-S., Seo, M.-W., Kim, Z.-H., and Lee, C.-G. (2006). Determining the best specific light uptake rates for the lumostatic cultures in bubble column photobioreactors. *Enzyme and microbial technology*, 39(3), 447-452.
- Lee, K.-S., Park, S. R., and Kim, Y. K. (2007). Effects of irradiance, temperature, and nutrients on growth dynamics of seagrasses: A review. *Journal of Experimental Marine Biology and Ecology*, 350(1-2), 144-175.
- Levlin, E. (2007). Conductivity measurements for controlling municipal waste-water treatment. In *Proceedings of a Polish-Swedish-Ukrainian Seminar* (pp. 51-62).
- Lee, Z. H., Sethupathi, S., Lee, K. T., Bhatia, S., and Mohamed, A. R. (2013). An overview on global warming in Southeast Asia: CO<sub>2</sub> emission status, efforts done, and barriers. *Renewable and Sustainable Energy Reviews*, 28(0), 71-81.
- Madaki, Y. S., and Seng, L. (2013). palm oil mill effluent (POME) from Malaysia palm oil mills waste or resource. *International Journal of Science, Environment and Technology*, 2(6), 1138 – 1155.
- Magurran, A. E. (2004). Measuring biological diversity. *African Journal of Aquatic Science*, 29(2), 285-286.

- Markou, G., Vandamme, D., and Muylaert, K. (2014). Microalgal and cyanobacterial cultivation: The supply of nutrients. *Water Research*, 65(0), 186-202.
- Mastrolonardo, G., Certini, G., Krebs, R., Forte, C., and Egli, M. (2013). Effects of fire on soil organic matter quality along an altitudinal sequence on Mt. Etna, Sicily. *CATENA*, 110(0), 133-145.
- McGinn, P. J., Dickinson, K. E., Bhatti, S., Frigon, J.-C., Guiot, S. R., and O'Leary, S. J. (2011). Integration of microalgae cultivation with industrial waste remediation for biofuel and bioenergy production: opportunities and limitations. *Photosynthesis research*, 109(1-3), 231-247.
- Mekki, A., Arous, F., Aloui, F., & Sayadi, S. (2013). DISPOSAL OF AGRO-INDUSTRIALS WASTES AS SOIL AMENDMENTS. *American Journal of Environmental Sciences*, 9(6), 458.
- Mosley, L., Singh, S., & Aalbersberg, B. (2005). Water quality monitoring in Pacific island countries. SOPAC technical reports, (381), 42.
- MPOB. (2015). Overview of the Malaysian oil palm industry 2014 Retrieved 19.3.2015, from <http://bepi.mpob.gov.my/index.php/publications.html>
- Mujtaba, G., Choi, W., Lee, C.-G., and Lee, K. (2012). Lipid production by *Chlorella vulgaris* after a shift from nutrient-rich to nitrogen starvation conditions. *Bioresource technology*, 123(0), 279-283.
- Munir, N., Imtiaz, A., Sharif, N. & Naz, S. 2015. Optimization of Growth Conditions of Different Algal Strains and Determination of Their Lipid Contents. *JAPS, Journal of Animal and Plant Sciences* 25(2): 546-553.
- Naidu, C. V., Dharma Raju, A., Satyanarayana, G. C., Vinay Kumar, P., Chiranjeevi, G., and Suchitra, P. (2015). An observational evidence of the decrease in Indian summer monsoon rainfall in the recent three decades of global warming era. *Global and Planetary Change*, 127(0), 91-102.
- Nwuche, C. O., Ekpo, D. C., Eze, C. N., Aoyagi, H., and Ogbonna, J. C. (2014). Use of palm oil mill effluent as medium for cultivation of *Chlorella sorokiniana*. *British Biotechnology Journal*, 4(3), 305-316.
- Ohimain, E., Daokoru-Olukole, C., Izah, S., Eke, R. & Okonkwo, A. 2012. Microbiology of Palm Oil Mill Effluents. *J. Microbiol. Biotech. Res* 2(6): 852-857.
- Okwute Ojonoma, L., and Ijah Udeme, J. (2014). Bioremediation of palm oil mill effluent (POME) polluted soil using microorganisms found in organic wastes. *International Journal of Biotechnology*, 3(3), 32-46.

- Othman, M. R., Hassan, M. A., Shirai, Y., Baharuddin, A. S., Ali, A. A. M., and Idris, J. (2014). Treatment of effluents from palm oil mill process to achieve river water quality for reuse as recycled water in a zero emission system. *Journal of Cleaner Production*, 67(0), 58-61.
- Pandey, J. P., and Tiwari, A. (2010). Optimization of biomass production of *Spirulina maxima*. *Journal of Algal Biomass Utilization (JABU)*, 1(2), 20-32.
- Park, J., Craggs, R., and Shilton, A. (2011). Wastewater treatment high rate algal ponds for biofuel production. *Bioresource technology*, 102(1), 35-42.
- Phang, S. M., & Kim-Chong, O. (1988). Algal biomass production in digested palm oil mill effluent. *Biological wastes*, 25(3), 177-191.
- Ponraj, M., and Din, M. F. M. (2013). Effect of light/dark cycle on biomass and lipid productivity by *Chlorella pyrenoidosa* using palm oil mill effluent (POME). *Journal of Scientific and Industrial Research*, 72(11), 703-706.
- Prasertsan, S., and Prasertsan, P. (1996). Biomass residues from palm oil mills in Thailand: an overview on quantity and potential usage. *Biomass and Bioenergy*, 11(5), 387-395.
- Priyadarshani, I., and Rath, B. (2012). Commercial and industrial applications of micro algae—a review. *Journal of Algal Biomass Utilization (JABU)*, 3(4), 89-100.
- Priyadarshani, I., Sahu, D., and Rath, B. (2011). Microalgal bioremediation: current practices and perspectives. *Journal of Biochemistry Technology*, 3(3), 299-304.
- Pruvost, J., Cornet, J. F., Le Borgne, F., Goetz, V., and Legrand, J. (2015). Theoretical investigation of microalgae culture in the light changing conditions of solar photobioreactor production and comparison with cyanobacteria. *Algal Research*, 10(0), 87-99.
- Raesossadati, M. J., Ahmadzadeh, H., McHenry, M. P., and Moheimani, N. R. (2014). CO<sub>2</sub> bioremediation by microalgae in photobioreactors: Impacts of biomass and CO<sub>2</sub> concentrations, light, and temperature. *Algal Research*, 6, Part A(0), 78-85.
- Rahman, A., Ellis, J. T., and Miller, C. D. (2012). Bioremediation of domestic wastewater and production of bioproducts from microalgae using waste stabilization ponds. *Journal of Bioremediation and Biodegradation*.
- Raja, R., Shanmugam, H., Ganesan, V., and Carvalho, I. (2014). Biomass from microalgae: an overview. *Oceanography*, 2(118), 2.
- Rao, A. R., Dayananda, C., Sarada, R., Shamala, T., and Ravishankar, G. (2007). Effect of salinity on growth of green alga *Botryococcus braunii* and its constituents. *Bioresource technology*, 98(3), 560-564.



- Ras, M., Steyer, J. P., & Bernard, O. (2013). Temperature effect on microalgae: a crucial factor for outdoor production. *Reviews in Environmental Science and Bio/Technology*, 12(2), 153-164.
- Reijnders, L., and Huijbregts, M. A. J. (2008). Palm oil and the emission of carbon-based greenhouse gases. *Journal of Cleaner Production*, 16(4), 477-482.
- Renaud, S. M., Thinh, L.-V., Lambrinidis, G., and Parry, D. L. (2002). Effect of temperature on growth, chemical composition and fatty acid composition of tropical Australian microalgae grown in batch cultures. *Aquaculture*, 211(1-4), 195-214.
- Roleda, M. Y., Slocombe, S. P., Leakey, R. J. G., Day, J. G., Bell, E. M., and Stanley, M. S. (2013). Effects of temperature and nutrient regimes on biomass and lipid production by six oleaginous microalgae in batch culture employing a two-phase cultivation strategy. *Bioresource technology*, 129(0), 439-449.
- Ruiz-Martinez, A., Martin Garcia, N., Romero, I., Seco, A., and Ferrer, J. (2012). Microalgae cultivation in wastewater: nutrient removal from anaerobic membrane bioreactor effluent. *Bioresource technology*, 126, 247-253.
- Rupani, P. F., Singh, R. P., Ibrahim, M. H., and Esa, N. (2010). Review of current palm oil mill effluent (POME) treatment methods: vermicomposting as a sustainable practice. *World Applied Sciences Journal*, 11(1), 70-81.
- Saifuddin, N., and Dinara, S. (2011). Pretreatment of palm oil mill effluent (POME) using magnetic chitosan. *Journal of Chemistry*, 8(S1), S67-S78.
- Selmani, N., Mirghani, M. E., and Alam, M. Z. (2013). Study the growth of microalgae in palm oil mill effluent waste water. Paper presented at the IOP Conference Series: Earth and Environmental Science.
- Sevrin-Reyssac, J. (1998). Biotreatment of swine manure by production of aquatic valuable biomasses. *Agriculture, ecosystems & environment*, 68(3), 177-186.
- Shaha, S. M., Ahmada, A., Othmanb, M. F., and Abdullah, M. A. (2014). Enhancement of lipid content in *Isochrysis galbana* and *Pavlova lutheri* using palm oil mill effluent as an alternative medium. *Chemical Engineering Transactions*, 37, 733-738.
- Sharma, G. K. & Khan, S. A. Bioremediation of Sewage Wastewater Using Selective Algae for Manure Production.
- Singh, J., and Gu, S. (2010). Commercialization potential of microalgae for biofuels production. *Renewable and Sustainable Energy Reviews*, 14(9), 2596-2610.
- Singh, R., Ibrahim, M. H., Esa, N., and Iliyana, M. (2010). Composting of waste from palm oil mill: a sustainable waste management practice. *Reviews in Environmental Science and Biotechnology*, 9(4), 331-344.

- Singh, S. P., and Singh, P. (2015). Effect of temperature and light on the growth of algae species: A review. *Renewable and Sustainable Energy Reviews*, 50(0), 431-444.
- Soleimaninanadegani, M. & Manshad, S. 2014. Enhancement of Biodegradation of Palm Oil Mill Effluents by Local Isolated Microorganisms. *International Scholarly Research Notices* 2014.
- Spellerberg, I. F., & Fedor, P. J. (2003). A tribute to Claude Shannon (1916–2001) and a plea for more rigorous use of species richness, species diversity and the ‘Shannon–Wiener’ Index. *Global ecology and biogeography*, 12(3), 177-179.
- Sukumaran, P., Nulit, R., Zulkifly, S., Halimoon, N., Omar, H., and Ismail, A. (2014). Potential of fresh POME as a growth medium in mass production of *Arthrospira platensis*. *International Journal of Current Microbiology and Applied Sciences*, 3(4), 235-250.
- Sumi, Y. (2009). Microalgae pioneering the future-application and utilization. *Life Science Research Unit, quarterly review*(34).
- Suresh, A., Praveen Kumar, R., Dhanasekaran, D., & Thajuddin, N. (2012). Biodiversity of microalgae in western and eastern ghats, India. *Pakistan Journal of Biological Sciences*, 15(19), 919-928.
- Sutherland, D. L., Howard-Williams, C., Turnbull, M. H., Broady, P. A., and Craggs, R. J. (2015). Enhancing microalgal photosynthesis and productivity in wastewater treatment high rate algal ponds for biofuel production. *Bioresource technology*, 184(0), 222-229.
- Takache, H., Pruvost, J., and Marec, H. (2015). Investigation of light/dark cycles effects on the photosynthetic growth of *Chlamydomonas reinhardtii* in conditions representative of photobioreactor cultivation. *Algal Research*, 8(0), 192-204.
- Tong, S., and Jaafar, A. B. (2004). Waste to energy: methane recovery from anaerobic digestion of palm oil mill effluent. *Energy Smart*, 4, 1-8.
- Vairappan, C. S., and Yen, A. M. (2008). Palm oil mill effluent (POME) cultured marine microalgae as supplementary diet for rotifer culture. *Journal of applied phycology*, 20(5), 603-608.
- Wahidin, S., Idris, A., and Shaleh, S. R. M. (2013). The influence of light intensity and photoperiod on the growth and lipid content of microalgae *Nannochloropsis* sp. *Bioresource technology*, 129(0), 7-11.
- Wah, W. P., Sulaiman, N. M., Nachiappan, M., & Varadaraj, B. (2002). Pre-treatment and membrane ultrafiltration using treated palm oil mill effluent (POME). *Songklanakarin J. Sci. Technol*, 24, 891-898.

- Wang, Y., Yan, X., and Wang, Z. (2015). Global warming caused by afforestation in the Southern Hemisphere. *Ecological Indicators*, 52(0), 371-378.
- Wolkers, H., Barbosa, M., Kleinegris, D., Bosma, R., and Wijffels, R. H. (2011). Microalgae: the green gold of the future. Large-scale sustainable cultivation of microalgae for the production of bulk commodities Retrieved 23.1.2015, from [http://www.groenegrondstoffen.nl/downloads/Boekjes/12Microalgae\\_UK.pdf](http://www.groenegrondstoffen.nl/downloads/Boekjes/12Microalgae_UK.pdf)
- Wu, L. F., Chen, P. C., and Lee, C. M. (2013). The effects of nitrogen sources and temperature on cell growth and lipid accumulation of microalgae. *International Biodeterioration & Biodegradation*, 85(0), 506-510.
- Wu, T. Y., Mohammad, A. W., Jahim, J. M., and Anuar, N. (2010). Pollution control technologies for the treatment of palm oil mill effluent (POME) through end-of-pipe processes. *Journal of Environmental Management*, 91(7), 1467-1490.
- Xia, S., Wan, L., Li, A., Sang, M., and Zhang, C. (2013). Effects of nutrients and light intensity on the growth and biochemical composition of a marine microalga *Odontella aurita*. *Chinese Journal of Oceanology and Limnology*, 31(6), 1163-1173.
- Xiao, R., Chen, R., Zhang, H.-Y. & Li, H. 2011. Microalgae *Scenedesmus Quadricauda* Grown in Digested Wastewater for Simultaneous CO<sub>2</sub> Fixation and Nutrient Removal. *Journal of Biobased Materials and Bioenergy* 5(2): 234-240.
- Xin, L., Hong-ying, H., Ke, G., and Ying-xue, S. (2010). Effects of different nitrogen and phosphorus concentrations on the growth, nutrient uptake, and lipid accumulation of a freshwater microalga *Scenedesmus* sp. *Bioresource technology*, 101(14), 5494-5500.
- Xin, L., Hong-ying, H., and Yu-ping, Z. (2011). Growth and lipid accumulation properties of a freshwater microalga *Scenedesmus* sp. under different cultivation temperature. *Bioresource technology*, 102(3), 3098-3102.
- Yaakob, Z., Kamarudin, K. F., Rajkumar, R., Takriff, M. S., and Badar, S. N. (2014). The current methods for the biomass production of the microalgae from wastewaters: an overview. *World Applied Sciences Journal*, 31(10), 1744-1758.
- Yun, S.-I., Kang, B.-M., Lim, S.-S., Choi, W.-J., Ko, J., Yoon, S., et al. (2012). Further understanding CH<sub>4</sub> emissions from a flooded rice field exposed to experimental warming with elevated CO<sub>2</sub>. *Agricultural and Forest Meteorology*, 154–155(0), 75-83.
- Zhu, J., Rong, J., and Zong, B. (2013). Factors in mass cultivation of microalgae for biodiesel. *Chinese Journal of Catalysis*, 34(1), 80-100.



Zhu, S., Huang, W., Xu, J., Wang, Z., Xu, J., and Yuan, Z. (2014). Metabolic changes of starch and lipid triggered by nitrogen starvation in the microalga *Chlorella zofingiensis*. *Bioresource technology*, 152(0), 292-298.

Zhu, S., Wang, Y., Shang, C., Wang, Z., Xu, J., and Yuan, Z. (2015). Characterization of lipid and fatty acids composition of *Chlorella zofingiensis* in response to nitrogen starvation. *Journal of Bioscience and Bioengineering*(0), <http://dx.doi.org/10.1016/j.jbiosc.2014.1012.1018>.

