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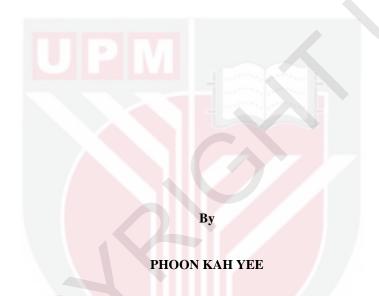
ENRICHMENT OF BIOACTIVE MINOR COMPONENTS FROM CPO AND PPMFO BY SEQUENTIAL ADSORPTION-DESORPTION TECHNIQUE

PHOON KAH YEE

FK 2018 116



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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

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Dedicated to my family

For your endless love, support and encouragement

ENRICHMENT OF BIOACTIVE MINOR COMPONENTS FROM CPO AND PPFMO BY SEQUENTIAL ADSORPTION-DESORPTION TECHNIQUE

By

PHOON KAH YEE

April 2018

Chair : Mohd Noriznan Mokhtar, PhD

Faculty : Engineering

The effort to separate or enrich bioactive compounds such as Vitamin E, phytosterols, squalene, and carotenoids from natural resources has been made to fulfil the increasing demands of the global market. Crude palm oil (CPO) which extracts from the palm oil fruit mesocarp fibre, is the product of palm oil mill, while, palm-pressed mesocarp fibre oil (PPMFO) is the residual oil of palm-pressed mesocarp fibre, a solid biomass which produced after CPO extraction in palm oil mill. Numerous studies reported on the presence of bioactive compounds as the minor components in CPO and PPMFO. In order to fulfil the market demand of these minor components, attempts such as supercritical fluid extraction and molecular distillation were develop to separate the minor components from plant source. However, limitation are present in current separation process. Therefore, in this study, interest is focusing on the enrichment efficiency of the minor components from CPO and PPMFO through a proposed sequential adsorptiondesorption technique. Initially, static adsorption-desorption test was carry out to evaluate the adsorption and desorption efficiency using six commercial mesopores adsorbents which represented different polarity towards the minor components that obtained from CPO. Non-polar adsorbents (Diaion HP20 and Sepabeads SP850) showed better adsorption-desorption efficiency than polar adsorbent (silica gel and Florisil) and weak polar adsorbent (Diaion HP2MG and Amberlite XAD-7HP) due to the similar polarity between the minor components and adsorbent. Diaion HP20 was selected as the best adsorbent resulting of the economic price and it can be stored at room temperature compared with Sepabeads SP850. Then, CPO was adsorbed by Diaion HP20 and placed into a Soxhlet extraction system to perform sequential adsorption-desorption process study. Three different organic solvent were used in the sequential adsorption-desorption process study to investigate the effect of desorption solvent and desorption time towards the enrichment efficiency of minor components. After obtaining the best process parameters, the sequential adsorption-desorption process were repeated by subjecting PPMFO as the feedstock. Under the same process parameters, Vitamin E, phytosterols, and squalene from CPO were obtained in the 1st fraction using methanol with desorption time of 4 h that gave enrichment factor (EF) of 3.4, 3.9, and 1.8, respectively, which slightly higher than those minor components obtained from PPMFO, 1.2, 1.8, and 1.4,

respectively. Meanwhile, the carotene obtained from both CPO and PPMFO was enriched in the 3^{rd} fraction by using *n*-hexane with an enrichment factor of 1.1 and 1.5, respectively. In conclusion, the obtained result revealed the efficiency of the proposed sequential adsorption-desorption technique to enrich the minor components from CPO and PPMFO.



PENGAYAAN SEBATIAN BIOAKTIF KOMPONEN MINOR DARIPADA CPO AND PPMFO OLEH TEKNIK PENJERAPAN-NYAHJERAPAN BERTURUTAN

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Usaha giat untuk memisah atau memperkaya sebatian bioaktif daripada sumber berasaskan semulajadi dilaksanakan untuk memenuhi permintaan pasaran dunia yang semakin meningkat. Minyak kelapa sawit (CPO) diperoleh dari sabut mesokarpa buah kelapa sawit dan merupakan produk kilang minyak kelapa sawit, manakala minyak perahan sabut mesokarpa kelapa sawit (PPMFO) adalah minyak sisa dari perahan sabut mesokarpa kelapa sawit, di mana ia merupakan biojisim pepejal selepas hasilan CPO di kilang minyak kelapa sawit. Kebanyakan kajian menyatakan kehadiran sebatian bioaktif sebagai komponen kecil dalam CPO dan PPMFO. Percubaan seperti esktraksi cecair superkritikal dan penyulingan molekul dilakukan untuk memisah komponent kecil daripada tumbuh-tumbuhan bagi memenuhi permintaan pasaran terhadap komponen kecil. Namun demikian, batasan wujub dalam proses pemisahan semasa. Oleh itu, kajian ini menumpukan terhadap kecekapan pengayaan komponen minor dalam CPO dan PPMFO melalui teknik jujukan jerapan-nyahjerapan yang dicadangkan. Pada permulaan, ujian jerapan-nyahjerapan statik dijalankan untuk menilai kecekapan jerapan dan nyahjerapan terhadap komponen kecil dalam CPO menggunakan enam mesopori penyerap komersial di mananya mewakili kekutuban yang berbeza. Penyerap tak berkutub (Diaion HP20 and Sepabeads SP850) menunjukkan kecekapan jerapannyahjerapan yang lebih baik berbanding dengan penyerap berkutub (silica gel dan Florisil) dan berkutub lemah (Diaion HP2MG and Amberlite XAD-7HP) kerana kekutuban yang sama antara komponen kecil dengan penyerap. Diaion HP20 dipilih sebagai penjerap terbaik berdasarkan harga yang rendah dan boleh disimpan pada suhu bilik berbanding dengan Sepabeads SP850. Selepas itu, CPO dijerap oleh Diaion HP20 dan letak dalam sistem ekstraksi Soxhlet untuk mengkaji proses jerapan-nyahjerapan berturutan. Proses jerapan-nyahjerapan berturutan menggunakan tiga pelarut organik berbeza dan seterusnya mengenalpasti kesan pelarut dan masa terhadap kecekapan pengayaan komponen minor tersebut. Proses jerapan-nyahjerapan berturutan diulangi dengan menggunakan PPMFO sebagai bahan mentah selepas mendapati proses parameter yang sesuai. Dalam proses parameter yang sama, Vitamin E, phytosterols, dan squalene daripada CPO diperolehi dalam pecahan pertama menggunakan metanol dengan masa nyahjerapan, 4 jam dan memberi faktor pengayaan (EF) masing-masing sebanyak 3.4, 3.9 dan 1.8, dimana lebih tinggi berbanding dengan komponen minor yang diperolehi daripada PPMFO, masing-masingnya ialah 1.2, 1.8 dan 1.4. Sementara itu, karotenoid daripada CPO dan PPMFO diperkaya pada pecahan yang ketiga dengan mengguna *n*-heksana dengan masing-masing memberi faktor pengayaan sebanyak 1.1 dan 1.5. Kesimpulannya, keputusan yang diperolehi menunjukkan kecekapan teknik jerapan-nyahjerapan yang dicadangkan untuk memperkayakan komponen kecil daripada CPO dan PPMFO.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

CPO Crude palm oil EF **Enrichment Factor** EFB Empty fruit brunch **FFAs** Free fatty acids FFB Fresh fruit brunch IPA Isopropyl ethanol MAGs Monoacylglycerols **DAGs** Diacylglycerols **TAGs** Triacylglycerols \mathbb{R}^2

Correlation coefficients

HPLC High performance liquid chromatography

GC Gas Chromatography

Low density lipoprotein cholesterol LDL-C

MSTFA N-methyl-N-trimethylsilyl-trifluoroacetamide

PΙ Polarity index

PPMFO Palm-pressed mesocarp fibre oil

ROS Reactive oxygen species RNS Reactive nitrogen species SFE Supercritical fluid exraction

CHAPTER 1

INTRODUCTION

1.1 Background

The demand of bioactive compounds, especially those extracted from natural resources is increasing globally due to the rise of safety concern, higher efficacies, and lower production cost (Ofori-Boateng & Lee, 2013; Puah, Choo, Ma, & Chuah, 2005). Compared with the petroleum synthetic, natural resources bioactive compounds are present in a scarce amount. It is advisable to select ready available natural resources which contain a high concentration of bioactive compounds to increase the separation yield and further reduced the separation cost.

African oil palm (*Elaeis guoneensis*) fruits produces two types of distinctive vegetable oils: crude palm oil (CPO) and palm kernel oil (PKO). CPO, a reddish orange colour oil which extracted from the fruit mesocarp through mechanical screw pressing technique (Mba, Dumont, & Ngadi, 2015). CPO represent the major product in oil palm industry by occupies about 88% of the total palm oil production (Abdullah & Sulaiman, 2013). CPO can be used for edible purposes such as cooking oils and margarine. (Mba et al., 2015) While, PKO, a light yellow colour oil which extracted from the kernel can be applied in oleochemicals such as detergents and soap (Abdullah & Sulaiman, 2013; Basiron, 2007; Sambanthamurthi, Sundram, & Tan, 2000).

Palm oil becomes the major source by contributing 33% (66 million tonnes) of 200 million tonnes of world oils and fats production in 2014 (Malaysian Palm Oil Board (MPOB), 2015). While, in Malaysia, oil palm industry served as the fourth largest economy contributor to give over RM 80 billion (about 8%) to gross national income (Malaysian Innovation Agency, n.d.). Since the first commercial oil palm estate setup at Tennamaran estate, Selangor in 1917, the oil palm plantation area in Malaysia increased drastically. Until 2014, the oil palm plantation area in Malaysia reached to 5.39 million hectares, which recorded a production of 19.67 million tonnes of palm oil (Abdullah & Sulaiman, 2013; Awalludin, Sulaiman, Hashim, & Nadhari, 2015). In addition, Malaysia becomes the world's second palm oil producer and exporter after Indonesia since 2008 (Abdullah & Sulaiman, 2013).

Oil palm industry generates substantial income to Malaysia's economy but in returns, it generates the largest amounts of biomass waste within agriculture sector. A palm tree produces 10% of oil yield and the remaining 90% is the biomass (Ofori-Boateng & Lee, 2013). Oil palm industry generated 83 million tonnes dry biomass in the year 2010 and is estimated will reach to 110 million by 2020 (Malaysian Innovation Agency, n.d.). Palm-pressed mesocarp fibre is one of the solid biomass waste which produced after mechanical screw-pressing CPO extraction process in palm oil mill. It accounts 7% (about 5.8 million tonnes) in the solid biomass waste of Malaysia palm oil industry in the year 2010. This amount is expected re

ach to 8.8 million tonnes in the year 2020 (Malaysian Innovation Agency, n.d.). This creates an issue for oil palm industry on how to valorise the palm-pressed mesocarp fibre for value added end-uses. Currently, palm-pressed mesocarp fibre is utilized as the fuel for sterilization and electricity generation in palm oil mill operation (Abdullah & Sulaiman, 2013). Lau, Choo, Ma, and Chuah (2008) commented that approximately 5-7% residual oil was trapped in the palm-pressed mesocarp fibre after the mechanical screw-pressing. This residual fibre oil is known as palm-pressed mesocarp fibre oil (PPMFO).

1.2 Problem statements

The demand of bioactive compounds, especially those extracted from natural resources is increasing globally due to the rise of safety concern, higher efficacies, and lower production cost (Ofori-Boateng & Lee, 2013; Puah et al., 2005). Compared with the petroleum synthetic, natural resources bioactive compounds are present in a scarce amount. It is advisable to select ready available natural resources which contain a high concentration of bioactive compounds to increase the separation yield and further reduced the separation cost.

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1.3 Hypothesis

Based on review previous studies, hypothesis could be made:

- 1) The performance of an adsorbent in adsorb and desorb of Vitamin E, phytosterols, carotenoids, and squalene can be characterize based on the its surface polarity (Li & Chase, 2010; Soto, Moure, Domínguez, & Parajó, 2011).
- 2) The concentration of Vitamin E, phytosterols, carotenoids, and squalene that obtained from CPO and PPMFO can be increased through the proposed sequential adsorption-desorption process (Gunawan, Kasim, & Ju, 2008; Latip, Baharin, Man, & Rahman, 2001).

1.4 Objectives

- 1) To characterize different adsorbents in adsorb and desorb Vitamin E, phytosterols, carotenoids, and squalene that obtained from CPO.
- 2) To evaluate the proposed sequential adsorption-desorption process based on the enrichment efficiency of Vitamin E, phytosterols, carotenoids, and squalene that obtained from CPO an+++d PPMFO.

1.5 Scope of work

This thesis concerned on the strategy to enrich the concentration of the Vitamin E, phytosterols, carotenoids, and squalene using the proposed sequential adsorption-desorption process in a Soxhlet extraction system. Vitamin E, phytosterols, carotenoids, and squalene were obtained from CPO source. Static adsorption-desorption test was used to compare the adsorption and desorption efficiency of minor components among six selected commercial adsorbents. The best adsorbent was further used in the sequential adsorption-desorption process study. Three oil fractions were produced after the sequential adsorption-desorption process. Different desorption solvents and times of the 1st fraction were tested to increase the enrichment factor of Vitamin E, phytosterols, and squalene. While different desorption solvents of the 2nd fraction was also done to increase the enrichment factor of carotenoids. Then, study was repeated with PPMFO as the feedstock, performed at the best condition which obtained from the CPO study.

REFERENCES

- Abdullah, N., & Sulaiman, F. (2013). The Oil Palm Wastes in Malaysia. *Biomass Now Sustainable Growth and Use*, 75–100.
- Ahmad, A. L., Chan, C. Y., Abd Shukor, S. R., & Mashitah, M. D. (2008). Recovery of oil and carotenes from palm oil mill effluent (POME). *Chemical Engineering Journal*, 141(1–3), 383–386.
- Ahsan, H., Ahad, A., & Siddiqui, W. a. (2015). A review of characterization of tocotrienols from plant oils and foods, 45–59.
- Akanda, M. J. H., Sarker, M. Z. I., Ferdosh, S., Manap, M. Y. A., Rahman, N. N. N. A., & Kadir, M. O. A. (2012). Applications of supercritical fluid extraction (SFE) of palm oil and oil from natural sources. *Molecules*, *17*(2), 1764–1794.
- Akgün, N. A. (2011). Separation of squalene from olive oil deodorizer distillate using supercritical fluids. *European Journal of Lipid Science and Technology*, 113(12), 1558–1565.
- Amorim-Carrilho, K. T., Cepeda, a., Fente, C., & Regal, P. (2014). Review of methods for analysis of carotenoids. *TrAC Trends in Analytical Chemistry*, *56*, 49–73.
- AOCS Official Method Cd 11d-91. (1998). Determination of Mono- and Diglycerides by Capillary Gas Chromatography. In D. Firestone (Ed.), *Official Methods and Recommended Practices of the AOCS* (5th ed.). Champaign, Illinois, USA.
- Awalludin, M. F., Sulaiman, O., Hashim, R., & Nadhari, W. N. A. W. (2015). An overview of the oil palm industry in Malaysia and its waste utilization through thermochemical conversion, specifically via liquefaction. *Renewable and Sustainable Energy Reviews*, 50, 1469–1484.
- Azzi, A., & Stocker, A. (2000). Vitamin E: non-antioxidant roles. *Progress in Lipid Research*, 39(3), 231–255.
- Baharin, B. S., Abdul Rahman, K., Abdul Karim, M. I., Oyaizu, T., Tanaka, K., Tanaka, Y., & Takagi, S. (1998). Separation of palm carotene from crude palm oil by adsorption chromatography with a synthetic polymer adsorbent. *Journal of the American Oil Chemists' Society*, 75(3), 399–404.
- Basiron, Y. (2007). Palm oil production through sustainable plantations. *European Journal Lipid Science Technology*, 109(4), 289–295.
- BBC Research Report. (2015). The Global Market for Carotenoids (Vol. FOD025E).
- Bermudez, Y., Ahmadi, S., Lowell, N. E., & Kruk, P. A. (2007). Vitamin E suppresses telomerase activity in ovarian cancer cells. *Cancer Detection and Prevention*, 31(2), 119–128.

- Bhatnagar, A. S., & Kulshrestha, R. (2017). Natural Antioxidants: Ocurrence and Their Role in Food Preservation. In R. Banerjee, A. K. Verma, & M. W. Siddiqui (Eds.), *Natural Antioxidants Applications in Foods of Animal Origin* (pp. 41–85). CRC Press.
- Burwell, R. . (1975). Manual of Symbols and Terminology for Physicochemical Quantities and Units—Appendix II Part I. *Pure and Applied Chemistry*, 46(July), 71–90.
- Camera, E., Ottaviani, M., & Picardo, M. (2015). Squalene chemistry and biology. In A. Pappas (Ed.), *Lipids and Skin Health* (pp. 1–359). Springer International Publishing.
- Cermak, S. C., Evangelista, R. L., & Kenar, J. A. (2012). Distillation of Natural Fatty Acids and Their Chemical Derivatives. In S. Zereshki (Ed.), *Distillation Advances from Modeling to Applications* (pp. 109–140). InTech.
- Chandrasekar, G., Vinu, A., Murugesan, V., & Hartmann, M. (2005). Adsorption of vitamin E on mesoporous silica molecular sieves. *Studies in Surface Science and Catalysis*, 158(12), 1169–1176.
- Chen, Y., Zhang, W., Zhao, T., Li, F., Zhang, M., Li, J., ... Yang, L. (2016). Adsorption properties of macroporous adsorbent resins for separation of anthocyanins from mulberry. *Food Chemistry*, 194, 712–722.
- Choo, Y. M., Lau, H. L. N., Ma, A. N., & Yusof, B. (2009). U.S. Patent No. 7,575,767 B2. Washington, DC: U.S. Patent and Trademark Office.
- Choo, Y. M., Ma, A. N., Yahaya, H., Yamauchi, Y., Bounoshita, M., & Saito, M. (1996). Separation of crude palm oil components by semipreparative supercritical fluid chromatography. *Journal of the American Oil Chemists' Society*, 73(4), 523–525.
- Choo, Y. M., Yap, S. C., Ooi, C. K., Ma, A. N., Goh, S. H., & Ong, A. S. H. (1996). Recovered oil from palm-pressed fiber: a good source of natural carotenoids, vitamin E, and sterols. *Journal of the American Oil Chemists' Society*, 73(5), 599–602.
- Chu, B. S., Baharin, B. S., Man, Y. B. C., & Quek, S. Y. (2005). Comparison of selected adsorbents for adsorption and desorption of vitamin E from palm fatty acid distillate. *Journal of Food Lipids*, 12(1), 23–33.
- Chu, B. S., Baharin, B. S., & Quek, S. Y. (2002). Factors affecting pre-concentration of tocopherols and tocotrienols from palm fatty acid distillate by lipase-catalysed hydrolysis. *Food Chemistry*, 79(1), 55–59.
- Chu, B. S., Baharin, B. S., Quek, S. Y., & Man, Y. B. C. (2003). Separation of tocopherols and tocotrienols from palm fatty acid distillate using hydrolysis-neutralization-adsorption chromatography method. *Journal of Food Lipids*, 10(2), 141–152.
- Chu, B. S., Quek, S. Y., & Baharin, B. S. (2003). Optimisation of enzymatic hydrolysis

- for concentration of Vitamin E in palm fatty acid distillate. *Food Chemistry*, 80(3), 295–302.
- Chua, C. S. L., Baharin, B. S., Man, Y. B. C., & Tan, C. P. (2007). Separation of squalene from palm fatty acid distillate using adsorption chromatography. *European Journal of Lipid Science and Technology*, 109(11), 1083–1087.
- Chuang, M.-H., & Brunner, G. (2006). Concentration of minor components in crude palm oil. *The Journal of Supercritical Fluids*, 37(2), 151–156.
- Conde, E., Moure, A., & Dominguez, H. (2017). Recovery of phenols from autohydrolysis liquors of barley husks: Kinetic and equilibrium studies. *Industrial Crops and Products*, 103(April), 175–184.
- Corro, G., Bañuelos, F., Vidal, E., & Cebada, S. (2014). Measurements of surface acidity of solid catalysts for free fatty acids esterification in Jatropha curcas crude oil for biodiesel production. *Fuel*, *115*, 625–628.
- da Silva, R. P. F. F., Rocha-Santos, T. A. P., & Duarte, A. C. (2016). Supercritical fluid extraction of bioactive compounds. *TrAC Trends in Analytical Chemistry*, 76, 40–51.
- Dabrowski, A. (2001). Adsorption-from theory to practice, 93, 135–224.
- Dal Prá, V., Lunelli, F. C., Vendruscolo, R. G., Martins, R., Wagner, R., Lazzaretti, A. P., ... da Rosa, M. B. (2017). Ultrasound-assisted extraction of bioactive compounds from palm pressed fiber with high antioxidant and photoprotective activities. *Ultrasonics Sonochemistry*, 36, 362–366.
- Dhawan, V. (2014). Reactive pxygen and nitrogen species: general considerations. In N. K. Ganguly, S. K. Jindal, S. Biswal, P. J. Barnes, & R. Pawankar (Eds.), *Studies on Respiratory Disorders* (pp. 27–47). New York, NY: Springer New York.
- Du, X., Yuan, Q., Li, Y., & Zhou, H. (2008). Preparative purification of solanesol from tobacco leaf extracts by macroporous resins. *Chemical Engineering and Technology*, 31(1), 87–94.
- Eitenmiller, R. R., Landen Jr., W. O., & Ye, L. (1998). *Vitamin Analysis for the Health and Food Sciences* (Second). CRC Press. Retrieved from http://197.14.51.10:81/pmb/AGROALIMENTAIRE/0849397715Vitamin.pdf
- Fabian, C., Gunawan, S., Kasim, N. S., Chiang, C.-L., & Ju, Y.-H. (2009). Separation of nonpolar lipid from soybean oil deodorizer distillate by stirred batch-wise silica gel adsorption-desorption. *Separation Science and Technology*, 44(7), 1621–1637.
- Fernandes, P., & Cabral, J. M. S. (2007). Phytosterols: Applications and recovery methods. *Bioresource Technology*, 98(12), 2335–2350.
- Fiametti, K. G., Rovani, S., de Oliveira, D., Corazza, M. L., Treichel, H., & Oliveira, J.

- V. (2008). Assessment of variable effects on solvent-free monoacylglycerol enzymatic production in AOT surfactant. *European Journal of Lipid Science and Technology*, 110(6), 510–515.
- Fregolente, L. V, Moraes, E. B., Martins, P. F., Batistella, C. B., Wolf-Maciel, M. R., Afonso, A. P., & Reis, M. H. M. (2006). Enrichment of natural products using an integrated solvent-free process: molecular distillation. *Institution of Chemical Engineers Symposium Series*, (152), 648–656.
- Fu, Y., Zu, Y., Liu, W., & Efferth, T. (2006). Optimization of luteolin separation from pigeonpea [Cajanus cajan (L.) Millsp.] leaves by macroporous resins. *Journal of Chromatography A*, 1137, 145–152.
- Ghosh, R. (2006). Adsorption. In *Principles of Bioseparations Engineering* (pp. 121–150). World Scientific Publishing Co. Pte. Ltd., Singapore.
- Gilbert, J. C., & Martin, S. F. (2010). Chromatography. In *Experimental organic chemistry: A miniscale and microscale approach* (5th ed., pp. 179–212). Brooks Cole.
- Gómez-Coca, R., Pérez-Camino, M., & Moreda, W. (2015). Neutral Lipids: Unsaponifiable. In L. M. L. Nollet & F. Toldra (Eds.), *Handbook of Food Analysis, Third Edition Two Volume Set* (pp. 459–489). CRC Press.
- Gopalan, Y., Shuaib, I. L., Magosso, E., Ansari, M. A., Abu Bakar, M. R., Wong, J. W., ... Yuen, K. H. (2014). Clinical Investigation of the Protective Effects of Palm Vitamin E Tocotrienols on Brain White Matter. *Stroke*, 45(5), 1422–1428.
- Gunawan, S., Kasim, N. S., & Ju, Y. H. (2008). Separation and purification of squalene from soybean oil deodorizer distillate. *Separation and Purification Technology*, 60(2), 128–135.
- Ho, D. S. S. (2009). U.S. Patent No. 7,544,822 B2. Washington, DC: US Patent and Trademark Office.
- Huang, X., Bi, J., Wang, J., Ouyang, J., Xiao, Y., Hao, H., ... Yin, Q. (2015). Liquid—liquid equilibrium of binary and ternary systems composed by palm oil or palm oil fractions with methanol/ethanol and water. *Fluid Phase Equilibria*, 404, 17—25.
- Jiang, Q. (2014). Natural forms of vitamin E: metabolism, antioxidant, and antiinflammatory activities and their role in disease prevention and therapy. *Free Radical Biology & Medicine*, 72C, 76–90.
- Jiang, S. T., Shao, P., Pan, L. J., & Zhao, Y. Y. (2006). Molecular distillation for recovering tocopherol and fatty acid methyl ssters from rapeseed oil deodoriser distillate. *Biosystems Engineering*, 93(4), 383–391.
- Kasim, N. S., Gunawan, S., Yuliana, M., & Ju, Y.-H. (2010). A simple two-step method for simultaneous isolation of tocopherols and free phytosterols from soybean oil

- deodorizer distillate with high purity and recovery. Separation Science and Technology, 45(16), 2437–2446.
- Kier, L. B. (1981). Quantitation of solvent polarity based on molecular structure. *Journal of Pharmaceutical Sciences*, 70(8), 930–3.
- Kim, S. K., & Karadeniz, F. (2012). Biological Importance and Applications of Squalene and Squalane. In *Advances in Food and Nutrition Research* (1st ed., Vol. 65, pp. 223–233). Elsevier Inc.
- Kohno, Y., Egawa, Y., Itoh, S., Nagaoka, S. ichi, Takahashi, M., & Mukai, K. (1995). Kinetic study of quenching reaction of singlet oxygen and scavenging reaction of free radical by squalene in n-butanol. *Biochimica et Biophysica Acta (BBA)/Lipids and Lipid Metabolism*, 1256(1), 52–56.
- Latip, R. A., Baharin, B. S., Man, Y. B. C., & Rahman, A. R. (2000). Evaluation of different types of synthetic adsorbents for carotene extraction from crude palm oil. *Journal of the American Oil Chemists' Society*, 77(12), 1277–1282.
- Latip, R. A., Baharin, B. S., Man, Y. B. C., & Rahman, R. A. (2001). Effect of adsorption and solvent extraction process on the percentage of carotene extracted from crude palm oil. *Journal of the American Oil Chemists' Society*, 78, 83–87.
- Lau, H. L. N., Choo, Y. M., Ma, A. N., & Chuah, C. H. (2006). Quality of residual oil from palm-pressed mesocarp fiber (Elaeis guineensis) using supercritical CO2 with and without ethanol. *Journal of the American Oil Chemists' Society*, 83(10), 893–898.
- Lau, H. L. N., Choo, Y. M., Ma, A. N., & Chuah, C. H. (2008). Selective extraction of palm carotene and vitamin E from fresh palm-pressed mesocarp fiber (Elaeis guineensis) using supercritical CO2. *Journal of Food Engineering*, 84(2), 289–296.
- Lau, H. L. N., Puah, C. W., Choo, Y. M., Ma, A. N., & Chuah, C. H. (2005). Simultaneous quantification of free fatty acids, free sterols, squalene, and acylglycerol molecular species in palm oil by high-temperature gas chromatography-flame ionization detection. *Lipids*, 40(5), 523–528.
- Leyton, A., Vergara-Salinas, J. R., Pérez-Correa, J. R., & Lienqueo, M. E. (2017). Purification of phlorotannins from Macrocystis pyrifera using macroporous resins. *Food Chemistry*, 237, 312–319.
- Li, J., & Chase, H. a. (2010). Development of adsorptive (non-ionic) macroporous resins and their uses in the purification of pharmacologically-active natural products from plant sources. *Natural Product Reports*, 27(10), 1493–1510.
- Lin, K.-M., & Koseoglu, S. S. (2003). Separation of sterols from deodorizer distillate by crystallization. *Journal of Food Lipids*, *10*(2), 107–127.
- Liu, D., Shi, J., Posada, L. R., Kakuda, Y., & Xue, S. J. (2008). Separating tocotrienols

- from palm oil by molecular distillation. *Food Reviews International*, 24(4), 376–391.
- Loganathan, R., Selvaduray, K. R., Nesaretnam, K., & Radhakrishnan, A. K. (2013). Tocotrienols promote apoptosis in human breast cancer cells by inducing poly(ADP-ribose) polymerase cleavage and inhibiting nuclear factor kappa-B activity. *Cell Proliferation*, 46(2), 203–213.
- Lotero, E., Liu, Y., Lopez, D. E., Suwannakarn, K., Bruce, D. A., & Goodwin, J. G. (2005). Synthesis of biodiesel via acid catalysis. *Ind. Eng. Chem. Res.*, 44(14), 5353–5363.
- Luo, X., Su, P., & Zhang, W. (2015). Advances in microalgae-derived phytosterols for functional food and pharmaceutical applications. *Marine Drugs*, 13(7), 4231–4254.
- Luque de Castro, M. D., & Priego-Capote, F. (2010). Soxhlet extraction: Past and present panacea. *Journal of Chromatography A*, 1217(16), 2383–2389.
- Maarasyid, C., Muhammad, I. I., & Supriyanto, E. (2014). Potential source and extractin of Vitamin E from palm-based oils: a review. *Jurnal Teknologi*, 4(69), 43–50.
- Magosso, E., Ansari, M. A., Gopalan, Y., Shuaib, I. L., Wong, J.-W., Khan, N. A. K., ... Yuen, K.-H. (2013). Tocotrienols for normalisation of hepatic echogenic response in nonalcoholic fatty liver: a randomised placebo-controlled clinical trial. *Nutrition Journal*, 12, 166.
- Makpol, S., Jam, F. A., Khor, S. C., Ismail, Z., Mohd Yusof, Y. A., & Wan Ngah, W. Z. (2013). Comparative effects of biodynes, tocotrienol-rich fraction, and tocopherol in enhancing collagen synthesis and inhibiting collagen degradation in stress-induced premature senescence model of human diploid fibroblasts. *Oxidative Medicine and Cellular Longevity*, 2013.
- Malaysian Innovation Agency. (n.d.). National Biomass Strategy 2020: New wealth creation for Malaysia's biomass industry. Retrieved May 24, 2017, from https://biobs.jrc.ec.europa.eu/sites/default/files/generated/files/policy/Biomass Strategy 2013.pdf
- Malaysian Palm Oil Board (MPOB). (2005). Determination of carotene content. *MPOB Test Method*, 194–197.
- Malaysian Palm Oil Board (MPOB). (2015). Value Added Products from PFAD. Retrieved February 29, 2016, from http://storage.unitedwebnetwork.com/files/23/c87c4a245d1b158f2d7b8e83720df 9a5.pdf
- Man, Y. B. C., & Tan, C. (2012). Carotenoids. In F. D. Gunstone (Ed.), *Lipids for Functional Foods and Nutraceuticals* (pp. 25–52). Elsevier.
- May, C. Y., Nang, H. L. L., Han, N. G., Ngan, M. A., & Basiron, Y. (2003). Value-added products from palm-pressed fibre. *MPOB TT 193*.

- Mba, O. I., Dumont, M.-J., & Ngadi, M. (2015). Palm oil: Processing, characterization and utilization in the food industry A review. *Food Bioscience*, *10*, 26–41.
- Mendes, M. F., Pessoa, F. L. P., Coelho, G. V., & Uller, A. M. C. (2005). Recovery of the high aggregated compounds present in the deodorizer distillate of the vegetable oils using supercritical fluids. *Journal of Supercritical Fluids*, *34*(2 SPEC. ISS.), 157–162.
- Mendes, M. F., Pessoa, F. L. P., & Uller, A. M. C. (2005). Optimization of the process of concentration of vitamin E from DDSO using supercritical CO2. *Brazilian Journal of Chemical Engineering*, 22(1), 83–91.
- Merck KGaA. (n.d.). Florisil specification. Retrieved February 12, 2016, from https://www.merckmillipore.com/MY/en/product/Florisil-0.150-0.250-mm,MDA_CHEM-112518#anchor_BRO
- Mezzomo, N., & Ferreira, S. R. S. (2016). Carotenoids functionality, sources, and processing by supercritical technology: A review, 2016.
- Mitsubishi Chemical Corporation. (2013). Production line brochure. Retrieved January 21, 2016, from https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=9&cad=rja&uact=8&ved=0ahUKEwiml7iLs-nXAhUC208KHfYUAZcQFghaMAg&url=https%3A%2F%2Fwww.lenntech.com%2FD ata-sheets%2FMitsubishi-DIAION-Cataloque.pdf&usg=AOvVaw37tnbreOVMWEVIWkyhIxTd
- Mitsubishi Chemical Corporation. (2015). Purification of medicine with Mitsubishi Chemical's separation medias (DIAION/SEPABEADS). Retrieved February 28, 2016, from http://lab-comp.hu/wp-content/uploads/2015/06/Purification-of-medicine-with-Mitsubishi-Chemical's-separation-medias.pdf
- Mustapa, A. N., Manan, Z. A., Azizi, C. Y. M., Setianto, W. B., & Omar, A. K. M. (2011). Extraction of β-carotenes from palm oil mesocarp using sub-critical R134a. *Food Chemistry*, *125*(1), 262–267.
- Naziri, E., Mantzouridou, F., & Tsimidou, M. Z. (2011). Squalene resources and uses point to the potential of biotechnology. *Lipid Technology*, *23*(12), 270–273.
- Neoh, B. K., Thang, Y. M., Zain, M. Z. M., & Junaidi, A. (2011). Palm pressed fibre oil: A new opportunity for premium hardstock? *International Food Research Journal*, 18(2), 769–773.
- Nesaretnam, K., Guthrie, N., Chambers, A. F., & Carroll, K. K. (1995). Effect of tocotrienols on the growth of a human breast cancer cell line in culture. *Lipids*, 30(12), 1139–1143.
- Nitsche, M., Johannisbauer, & Jordan. (1999). U.S. Patent No. 5,902,890. Washington, DC: U.S. Patent and Trademark Office.

- Ofori-Boateng, C., & Lee, K. T. (2013). Sustainable utilization of oil palm wastes for bioactive phytochemicals for the benefit of the oil palm and nutraceutical industries. *Phytochemistry Reviews*, 12(1), 173–190.
- Othman, N., Manan, Z. A., Wan Alwi, S. R., & Sarmidi, M. R. (2010). A review of extraction technology for carotenoids and vitamin e recovery from palm oil. *Journal of Applied Sciences*.
- Peh, H. Y., Tan, W. S. D., Liao, W., & Wong, W. S. F. (2016). Vitamin E therapy beyond cancer: Tocopherol versus tocotrienol. *Pharmacology & Therapeutics*, *162*, 152–69. https://doi.org/10.1016/j.pharmthera.2015.12.003
- Ping, B. T. Y. (2007). Palm Carotene Concentrates From Crude Palm Oil Using Vacuum Liquid Chromatography on Silica Gel. *Journal of Oil Palm Research*, 19(December), 421–427. Retrieved from http://jopr.mpob.gov.my/wp-content/uploads/2013/09/joprv19dec2007-bonnie1.pdf
- Poku, K. (2002). *Small-scale palm oil processing in Africa*. Rome: Food and Agricultural Organization (FAO).
- Popa, O., Babeanu, N. E., Popa, I., Nita, S., & Dinu-Parvu, C. E. (2015). Methods for obtaining and determination of squalene from natural sources. *BioMed Research International*, 2015.
- Pope Science Inc. (n.d.). Wiped film evaporator and vacuum distillate. Retrieved August 23, 2017, from http://www.popeinc.com/
- Posada, L. R., Shi, J., Kakuda, Y., & Xue, S. J. (2007). Extraction of tocotrienols from palm fatty acid distillates using molecular distillation. *Separation and Purification Technology*, 57(2), 220–229.
- Puah, C. W., Choo, Y. M., Ma, A. N., & Chuah, C. H. (2005). Supercritical Fluid Extraction of Palm Carotenoids. *American Journal of Environmental Sciences*, 1(4), 264–269.
- Puah, C. W., Choo, Y. M., Ma, A. N., & Chuah, C. H. (2007). Solubility of tocopherol and tocotrienols from palm oil in supercritical carbon dioxide. *Journal of Food Lipids*, 14(4), 377–385.
- Puah, C. W., Choo, Y. M., Ma, A. N., & Chuah, C. H. (2008). Production of Carotenoids and Tocols Concentrates From Palm Oil Using Supercritical Carbon Dioxide. *Journal of Oil Palm Research*, 12–15.
- Quek, S., Chu, B., & Baharin, B. S. (2007). Commercial Extraction of Vitamin E from Food Sources. In V. R. Preedy & R. R. Watson (Eds.), *The Encyclopedia of Vitamin E* (pp. 140–152). CABI International.
- Ramamurthi, S., & McCurdy, A. R. (1993). Enzymatic pretreatment of deodorizer distillate for concentration of sterols and tocopherols. *Journal of the American Oil Chemists' Society*, 70(3), 287–295.

- Rao, C. V., Newmark, H. L., & Reddy, B. S. (1998). Chemopreventive effect of squalene on colon cancer. *Carcinogenesis*, 19(2), 287–290.
- Rodriguez-Amaya, D. B. (2015). Carotenes and xanthophylls as antioxidants. In F. Shahidi (Ed.), Handbook of Antioxidants for Food Preservation (pp. 17–50). Elsevier Ltd.
- Rohm and Haas. (n.d.). AmberliteTM XAD7hp. Retrieved March 18, 2016, from http://www.dow.com/assets/attachments/business/process_chemicals/amberlite_xad/amberlite_xad7_hp/tds/amberlite_xad7hp.pdf
- Ruthven, D. M. (1984). Principles of adsorption and adsorption processes. Wiley & Sons Ltd.
- Sabliov, C. M., Fronczek, C., Astete, C. E., Khachaturyan, M., Khachatryan, L., & Leonardi, C. (2009). Effects of temperature and UV light on degradation of α-tocopherol in free and dissolved form. *Journal of the American Oil Chemists' Society*, 86(9), 895–902.
- Saini, R. K., Nile, S. H., & Park, S. W. (2015). Carotenoids from fruits and vegetables: Chemistry, analysis, occurrence, bioavailability and biological activities. *Food Research International*, 76, 735–750.
- Salo, P., Wester, I., & Hopia, A. (2012). Phytosterols. In *Lipids for Functional Foods* and *Nutraceuticals* (pp. 183–224).
- Sambanthamurthi, R., Sundram, K., & Tan, Y. (2000). Chemistry and biochemistry of palm oil. *Progress in Lipid Research*, 39(6), 507–558.
- Scharlau Chemicals. (n.d.). Scharlau Silica gel. Retrieved March 17, 2016, from http://www.scharlab.com/catalogo-productos-detalle-articulo.php?c=40&sc=261&p=7323
- Schmitt, D., Beiser, N., Regenbrecht, C., Zirbes, M., & Waldvogel, S. R. (2017). Adsorption and separation of black liquor-derived phenol derivatives using anion exchange resins. *Separation and Purification Technology*, 181, 8–17.
- Silva, S. M., Sampaio, K. A., Ceriani, R., Verhe, R., Stevens, C., De Greyt, W., & Meirelles, A. J. A. (2014). Effect of type of bleaching earth on the final color of refined palm oil. *LWT Food Science and Technology*, *59*(2P2), 1258–1264.
- Song, E.-S., Lim, J., Lee, H.-S., & Lee, Y.-W. (2008). Transesterification of RBD palm oil using supercritical methanol. *The Journal of Supercritical Fluids*, 44(3), 356–363.
- Soto, M. L., Moure, A., Domínguez, H., & Parajó, J. C. (2011). Recovery, concentration and purification of phenolic compounds by adsorption: A review. *Journal of Food Engineering*, 105(1), 1–27.
- Spanova, M., Zweytick, D., Lohner, K., Klug, L., Leitner, E., Hermetter, A., & Daum,

- G. (2012). Influence of squalene on lipid particle/droplet and membrane organization in the yeast Saccharomyces cerevisiae. *Biochimica et Biophysica Acta Molecular and Cell Biology of Lipids*, 1821(4), 647–653.
- Stephen, N. M., R., G., Niranjana, K., Y. P., Das, A. K., V, B., & P., G. (2017).
 Carotenoids: Types, Sources, and Biosynthesis. In M. W. Siddiqui, V. Bansal, & K. Prasad (Eds.), *Plant Secondary Metabolites*, *Volume Two: Stimulation*, *Extraction, and Utilization* (pp. 77–106). CRC Press.
- Stone, W. L., & Papas, A. (2012). Tocopherols, tocotrienols and vitamin E. In *Lipids for Functional Foods and Nutraceuticals* (pp. 53–72). Elsevier.
- Sundram, K., Sambanthamurthi, R., & Tan, Y. A. (2003). Palm fruit chemistry and nutrition. *Asia Pacific Journal of Clinical Nutrition*, 12(3), 355–362.
- Verleyen, T., Forcades, M., Verhe, R., Dewettinck, K., Huyghebaert, a., & Greyt, W. (2002). Analysis of free and esterified sterols in vegetable oils. *Journal of the American Oil Chemists' Society*, 79(2), 117–122.
- Vincent, C. J., Shamsudin, R., & Baharuddin, A. S. (2014). Pre-treatment of oil palm fruits: A review. *Journal of Food Engineering*, 143, 123–131.
- Visioli, L. J., de Castilhos, F., Cardozo-Filho, L., de Mello, B. T. F., & da Silva, C. (2016). Production of esters from soybean oil deodorizer distillate in pressurized ethanol. *Fuel Processing Technology*, 149, 326–331.
- Wang, T., Lu, S., Xia, Q., Fang, Z., & Johnson, S. (2015). Separation and purification of amygdalin from thinned bayberry kernels by macroporous adsorption resins. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*, 975, 52–58.
- Watanabe, Y., Nagao, T., Hirota, Y., Kitano, M., & Shimada, Y. (2004). Purification of tocopherols and phytosterols by a two-step in situ enzymatic reaction. *Journal of the American Oil Chemists' Society*, 81(4), 339–345.
- Woyengo, T. a, Ramprasath, V. R., & Jones, P. J. H. (2009). Anticancer effects of phytosterols. *European Journal of Clinical Nutrition*, 63(7), 813–20.
- Xu, Z., Harvey, K. A., Pavlina, T., Dutot, G., Hise, M., Zaloga, G. P., & Siddiqui, R. A. (2012). Steroidal compounds in commercial parenteral lipid emulsions. *Nutrients*, *4*(8), 904–921.
- Yunos, N. S. H. M., Baharuddin, A. S., Md Yunos, K. F., Hafid, H. S., Busu, Z., Mokhtar, M. N., ... Som, A. M. (2015). The physicochemical characteristics of residual oil and fibers from oil palm empty fruit bunches. *BioResources*, *10*(1), 14–29.
- Zawistowski, J. (2010). Tangible health benefits of phytosterol functional foods. In J. Smith & E. Charter (Eds.), *Functional Food Product Development* (pp. 362–381). Oxford, UK: Wiley-Blackwell.