



UNIVERSITI PUTRA MALAYSIA

***HEPATOPROTECTIVE EFFECT OF BAUHINIA PURPUREA L.
METHANOLIC LEAVES EXTRACT***

FARHANA BTE YAHYA

FPSK(M) 2014 26



**HEPATOPROTECTIVE EFFECT OF *BAUHINIA PURPUREA* L.
METHANOLIC LEAVES EXTRACT**

By

FARHANA BTE YAHYA

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Science**

December, 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

HEPATOPROTECTIVE EFFECT OF *BAUHINIA PURPUREA* L. METHANOLIC LEAVES EXTRACT

By

FARHANA BTE YAHYA

December, 2014

Chairman: Associate Professor Zainul Amiruddin Zakaria, PhD
Faculty: Medicine and Health Sciences

The objective of this study was to determine the hepatoprotective activity of methanolic extract of *Bauhinia purpurea* (Fabaceae) leaves (MEBP) and its partitions using rat models, i.e., by evaluating the prophylactic effect of the plant extracts administered prior to the induction of liver toxicity using a hepatotoxic agent. The study was designed as a preventive method, as the hepatoprotective potential of MEBP has never been reported. In an attempt to establish the pharmacological properties of *B. purpurea*, the hepatoprotective potential of MEBP was investigated using paracetamol (PCM)- and carbon tetrachloride (CCl₄)-induced hepatotoxicity in rats. Throughout this study, the animals were divided into 22 groups containing 6 rats per group. For the first stage of the *in vivo* study, rats were divided into groups and administered orally once daily with 10% dimethyl sulfoxide (DMSO) (negative control), 200 mg/kg silymarin (positive control), or MEBP (50, 250, 500 mg/kg) for 7 days, followed by hepatotoxicity induction using PCM or CCl₄. In the second stage of the *in vivo* study, MEBP was partitioned into 3 fractions: petroleum ether extract (PEBP), ethyl acetate extract (EABP), and aqueous extract (AQBP). PEBP, EABP, and AQBP activities were tested on PCM-induced hepatotoxicity in rats. Blood samples underwent biochemical analysis to evaluate alanine transaminase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), lactate dehydrogenase (LDH), and total protein (TP) levels; the livers were subjected to microscopic analysis. All extracts (MEBP, PEBP, EABP, AQBP) underwent antioxidant study using the 2, 2-diphenyl-1-picrylhydrazyl radical scavenging assay (DPPH), superoxide dismutase scavenging assay (SOD), and oxygen radical absorbance capacity assay (ORAC), and anti-inflammatory study using lipooxygenase (LOX) and xanthine oxidase (XO) assays. Total phenolic content (TPC), phytochemical screening, and high-performance liquid chromatography (HPLC) analysis were also performed. From the histological observation, lymphocyte infiltration and marked necrosis were observed in the DMSO-treated

groups (negative control). MEBP showed encouraging activity for reducing the toxic effect of CCl_4 and PCM on the liver by reducing the weight of the liver in a dose-dependent manner; histological observation demonstrated normalization of the histopathological changes, preserving hepatocyte structure, causing a significant decline in ALT and AST levels ($p < 0.05$) and escalation of TP level. PEBP, which contains non-polar compounds, reduced the liver enzyme levels in a dose-dependent manner and increased the production of TP. EABP and AQBP, which contain intermediate compounds and polar compounds, respectively, attenuated the liver enzyme and LDH levels (concentration-independent). Among the extracts, EABP had the best activity for attenuating the liver enzymes. MEBP had the highest TPC value, followed by EABP, AQBP, and PEBP. EABP and MEBP demonstrated potential free radical scavenging activity in the SOD assay. The trend for the ORAC assay was slightly different from that of the DPPH and SOD assays. AQBP and EABP had high ORAC value, which determines the capacity of an extract to act as an antioxidant. All extracts in the present study had weak anti-inflammatory activity in the inhibition of LOX and XO. Phytochemical screening of the extracts showed that MEBP, PEBP, and EABP contained flavonoids, tannins, polyphenolic compounds, and steroids. However, the phytochemical screening showed that AQBP contained fewer compounds. HPLC analysis demonstrated several peaks detected at different wavelengths of the chromatogram of MEBP, EABP and AQBP, which were suggested to be flavonoid-based compounds. In conclusion, MEBP exerted potential hepatoprotective activity that can be partly attributed to its antioxidant activity, and EABP was considered to have the best activity among the fractions, which warrants further investigation.

Abstrak thesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN HEPATOPROTEKTIF OLEH EKSTRAK METANOL DARI DAUN
BAUHINIA PURPUREA L.**

Oleh

FARHANA BTE YAHYA

December, 2014

Pengerusi: Profesor Madya Zainul Amiruddin Zakaria, PhD
Fakulti: Perubatan dan Sains Kesihatan

Objektif kajian ini adalah untuk menentukan aktiviti hepatoprotektif ekstrak metanol daripada daun *Bauhinia purpurea* dan pecahannya dengan menggunakan model tikus dengan menilai kesan profilaksis ekstrak tumbuhan yang diambil sebelum induksi ketoksikan hati menggunakan ejen hepatotoksik. Kajian ini berdasarkan kaedah pencegahan kerana potensi hepatoprotektif daripada MEBP tidak pernah didakwa terbukti lagi. Dalam usaha untuk mengenal pasti sifat-sifat farmakologi *Bauhinia purpurea* (Fabaceae), potensi hepatoprotektif dari ekstrak metanol daun *B. purpurea* (MEBP) telah diuji menggunakan rangsangan hepatotoksik paracetamol (PCM) - dan karbon tetraklorida (CCl₄) pada tikus. Sepanjang kajian ini, haiwan telah dibahagikan kepada 22 kumpulan dengan 6 tikus setiap kumpulan. Untuk bahagian pertama kajian *in vivo*, tikus ($n = 6$ bagi setiap kumpulan) dibahagikan kepada beberapa kumpulan dan diberi makan secara oral sekali sehari dengan 10% dimetil sulfoxide (DMSO) (kawalan negatif), 200 mg / kg silymarin (kawalan positif), atau MEBP (50, 250, dan 500 mg / kg) selama 7 hari, diikuti dengan proses rangsangan hepatotoksik menggunakan PCM atau CCl₄. Kemudian MEBP di ekstrak kepada 3 pecahan: ekstrak petroleum eter (PEBP), ekstrak etil asetat (EABP), dan ekstrak akueus (AQBP). Dalam bahagian kedua kajian *in vivo*, aktiviti PEBP, EABP dan AQBP telah diuji ke atas rangsangan PCM ke atas hati tikus. Sampel darah yang telah diambil dibuat kajian biokimia untuk menganalisis paras enzim seperti alanine transaminase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), lactate dehydrogenase (LDH), dan total protein (TP). Manakala, sampel hati pula diuji secara mikroskopik. Semua ekstrak (MEBP, PEBP, EABP dan AQBP) juga diuji untuk kajian antioksidan menggunakan cerakin 2, 2-difenil-1-picrylhydrazyl radikal (DPPH), pengujian perangkap aktiviti superoxide dismutase (SOD), dan cerakin penyerapan oksigen radikal kapasiti (ORAC), dan kajian anti-radang menggunakan analisis aktiviti lipoxigenase (LOX) dan xanthine oxidase (XO). Kandungan jumlah fenol (TPC), pemeriksaan fitokimia, dan kromatografi cecair berprestasi tinggi (HPLC) analisis juga telah dilaksanakan. Dari segi pemerhatian histologi,

penyusupan limfosit dan nekrosis diperhatikan dalam kumpulan rawatan DMSO (kawalan negatif). MEBP menunjukkan aktiviti yang bagus dalam usaha mengurangkan kesan toksik daripada CCL₄ dan PCM ke atas hati, dengan menyebabkan penurunan berat hati secara kebergantungan pada peningkatan dos, pemantauan histologi menunjukkan pemulihan struktur sel-sel hati, dan menyebabkan penurunan paras ALT dan AST secara signifikan ($P < 0.05$), dan peningkatan paras TP. PEBP yang mengandungi sebatian tak berkutub, mengurangkan paras enzim hati secara kebergantungan terhadap dos dan menyebabkan peningkatan TP. EABP dan AQBP yang mengandungi sebatian pertengahan dan sebatian berkutub, masing-masing menyebabkan penurunan paras enzim hati dan LDH (tidak bergantung kepada dos). Antara semua ekstrak, EABP mempunyai aktiviti terbaik dalam penurunan enzim hati. MEBP mempunyai nilai TPC paling tinggi diikuti oleh EABP, AQBP dan PEBP. EABP dan MEBP sekali lagi menunjukkan potensi di dalam aktiviti SOD. Daripada penemuan, trend ORAC sedikit berbeza daripada aktiviti pengujian perangkap DPPH dan superoxide. AQBP dan EABP mempunyai nilai ORAC yang tinggi, ini menunjukkan kebolehan ekstrak di dalam aktiviti antioksidan. Manakala, semua ekstrak mempunyai kadar anti radang yang rendah dalam menghalang aktiviti LOX dan XO. Pemeriksaan fitokimia ekstrak menunjukkan MEBP, PEBP dan EABP mempunyai flavonoid, tannin, sebatian polifenolik, dan steroid. Manakala AQBP yang menunjukkan lebih sedikit sebatian yang di ekstrak. HPLC analisis menunjukkan beberapa puncak yang di kenal pasti pada gelombang yang berbeza di dalam kromatogram MEBP, EABP dan AQBP boleh dikategorikan sebagai jenis-jenis sebatian flavonoid. Kesimpulannya, MEBP mempunyai potensi sebagai agen hepatoprotektif yang juga sebahagiannya mungkin bergantung kepada aktiviti antioksidan, dan EABP dianggap mempunyai aktiviti yang terbaik di antara pecahan ekstrak, yang memerlukan siasatan lanjut.

ACKNOWLEDGEMENT

Alhamdulillah, All Praise and Thanks is for Allah subhanahu wa ta'ala for helping me in coordinating my life.

I wish to extend my deepest appreciation to my supervisor, Associate Professor Dr. Zainul Amiruddin Zakaria for his guidance, valuable advice, patience, support and continuous supervision throughout the course of this project. I truly thank him for giving me opportunity to be his postgraduate student.

I would like to express my gratitude and appreciation to my co-supervisor, Dr Norhafizah Mohtarrudin for her kindness to teach and guide me in Histological study that is definitely important in order to complete this project.

My sincere appreciation dedicates to the Faculty of Medicine and Health Sciences, Universiti Putra Malaysia for giving me opportunity to carry out this project.

My special dedication to Encik Ramli, staffs of Pharmacology laboratory, and staffs of Histology laboratory for their kind, cooperation, excellent facilities and support which help to perform the test in laboratory unit and analyses smoothly. I would like to thank to all my fellow friends, Farah Hidayah, Mohammad Fauzi Fahmi, Siti Syariah, Roihana, and Tavamani, for their kindness, patience and cooperation in completing this project successfully.

Words are not enough to express to my beloved family, especially my dearest parents, Yahya bin Md Aziz, and Sopiah binti Yusof for their love, comfort, encouragement, support and advice that truly motivated me to accomplish my master successfully. I owe a depth gratitude to them which can never be repaid. May Allah shower His countless blessings upon them.

Thank you so much.

I certify that a Thesis Examination Committee has met on 16 December 2014 to conduct the final examination of Farhana Bte Yahya on her thesis entitled “Hepatoprotective Effect of *Bauhinia purpurea* L. Methanolic Leaves Extract” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Amin bin Ismail, PhD

Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Chairman)

Sabrina bt Sukardi, PhD

Associate Professor
Faculty of Medicine and Health Sciences
Universiti Putra Malaysia
(Internal Examiner)

Teh Lay Kek, PhD

Professor
Faculty of Pharmacy
Universiti Teknologi Mara
(External Examiner)



ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 26 February 2015

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia (Research Rules 2012);
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: _____

Name and Matric No: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of
Supervisory
Committee: _____

Signature: _____

Name of
Member of
Supervisory
Committee: _____

Signature: _____

Name of
Member of
Supervisory
Committee: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of
Supervisory
Committee: _____

Signature: _____

Name of
Member of
Supervisory
Committee: _____

Signature: _____

Name of
Chairman of
Supervisory
Committee: _____



TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENT	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xx
 CHAPTER	
 1. INTRODUCTION	 1
 2. LITERATURE REVIEW	
2.1 Natural products	5
2.1.1 Plant-based natural products	5
2.1.2 Hepatoprotective-related natural product	6
2.2 <i>Bauhinia purpurea</i>	
2.2.1 Geographical distribution	6
2.2.2 Botanic description	6
2.2.3 Traditional practices	8
2.2.4 Scientific findings	9
2.7.5 Phytochemistry	11
2.3 Gross anatomy of the liver	12
2.4 Microanatomy of the liver	13
2.5 Physiology of the liver	14
2.6 Role of drug metabolism and detoxification	15
2.7 Enzyme liver functions	16
2.8 Diseases of liver	17
2.9 Side effects of available drug	18

2.10	Biomarkers of liver injury	18
2.11	Liver anatomy of rat	19
2.12	Hepatotoxicity models	21
2.13	Free radical formation, oxidative stress and antioxidant	24
2.14	Chromatography in the pharmaceutical area	26

3. EVALUATION OF HEPATOPROTECTIVE ACTIVITY OF METHANOLIC EXTRACT OF *BAUHINIA PURPUREA* LEAVES AND ITS PARTITIONS

3.1	Introduction	28
3.2	Methodology	29
3.2.1	Collection of plant material	29
3.2.2	Preparation of crude methanolic extract	29
3.2.3	Preparation of petroleum ether, ethyl acetate and aqueous plant extract	30
3.2.4	Chemicals	30
3.2.5	Animals	30
3.2.6	Hepatotoxicity assays	32
3.2.7	Biochemical studies	34
3.2.8	Histopathology	36
3.2.9	Statistical analysis	36
3.3	Results	
3.3.1	Effect of MEBP on the liver weight after induction with CCl ₄	37
3.3.2	Histopathological study of liver pre-treated with and without MEBP followed by treatment with CCl ₄	37
3.3.3	Biochemical study of liver pre-treated with and without MEBP followed by treatment with CCl ₄	37
3.3.4	Effect of MEBP on the liver weight after induction with PCM	45
3.3.5	Histopathological study of the liver pre-treated with and without MEBP followed by treatment with PCM	45

3.3.6	Biochemical study of the liver pre-treated with and without MEBP followed by treatment with PCM	45
3.3.7	Effect of PEBP, EABP and AQBP on the liver weight after induction with PCM	53
3.3.8	Histopathological study of the liver pre-treated with PEBP, EABP and AQBP followed by treatment with PCM	53
3.3.9	Biochemical study of the liver pre-treated with PEBP, EABP and AQBP followed by treatment with PCM	53
3.4	Discussion	64

4. EVALUATION OF THE ANTIOXIDANT ANTI-INFLAMMATORY ACTIVITIES OF THE METHANOLIC EXTRACT OF *BAUHINIA PURPUREA* LEAVES AND ITS PARTITIONS

4.1	Introduction	67
4.2	Methodology	
4.2.1	Total phenolic content	68
4.2.2	2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity	69
4.2.3	Superoxide scavenging assay	69
4.2.4	Oxygen radical absorbance capacity (ORAC) test	69
4.2.5	Lipoxygenase assay	70
4.2.6	Xanthine oxidase assay	70
4.3	Results	
4.3.1	Total phenolic content	71
4.3.2	DPPH scavenging activity	71
4.3.3	Superoxide scavenging activity	71
4.3.4	ORAC	71
4.3.4	LOX and XO activities	72
4.4	Discussion	79

**5. PHYTOCHEMICAL SCREENING AND HIGH-PERFORMANCE
LIQUID CHROMATOGRAPHY ANALYSIS OF THE METHANOLIC
EXTRACT OF *BAUHINIA PURPUREA* LEAVES AND ITS PARTITIONS**

5.1	Introduction	82
5.2	Methodology	
5.2.1	Phytochemical screening	83
5.2.2	HPLC analysis	84
5.2.3	Identification of flavonoids in MEBP via HPLC analysis	84
5.3	Results	
5.3.1	Phytochemical screening	85
5.3.2	HPLC chromatogram and UV profile of the MEBP	86
5.3.3	Identification of flavonoids in MEBP via HPLC analysis	86
5.3.4	HPLC chromatogram and UV profile of the partitions of MEBP	91
5.4	Discussion	98
6.	GENERAL DISCUSSION	100
7.	SUMMARY, CONCLUSION AND RECOMMENDATION FOR FUTURE STUDY	103
	REFERENCES	105
	APPENDICES	121
	BIODATA OF STUDENT	126
	LIST OF PUBLICATIONS AND PROCEEDINGS	127

LIST OF TABLES

Table	Page
Table 1: Ethnomedicinal uses of <i>B. purpurea</i>	8
Table 2: Pharmacological activities of <i>B. purpurea</i>	9
Table 3: Compounds of <i>B. purpurea</i>	10
Table 4: CCl ₄ -induced hepatotoxicity treatment groups	32
Table 5: PCM-induced hepatotoxicity treatment groups	32
Table 6: PCM-induced hepatotoxicity treatment groups of partition extracts	33
Table 7: Liver histological scoring	36
Table 8a: Effect of MEBP on percentage change of body and liver weight in CCl ₄ treated rats	38
Table 8b: Histopathological evaluation of effect of various doses of MEBP against CCl ₄ induced hepatic injury in rats	39
Table 8c: Effect of CCl ₄ and protective treatments at ALT, AST, ALP (U/L), total protein (g/dL) and LDH (U/L)	44
Table 9a: Effect of MEBP on percentage change of body and liver weight in PCM- treated rats	46
Table 9b: Histopathological evaluation of effect of various doses of MEBP against PCM-induced hepatic injury in rats	47
Table 9c: Effect of PCM and protective treatments at ALT, AST, ALP (U/L), total protein (g/dL) and LDH (U/L)	52
Table 10a: Effect of PEBP, EABP and AQBP on percentage change of liver weight in PCM- treated rats	55
Table 10b: Histopathological evaluation of effects of PEBP, EABP and AQBP against PCM-induced hepatic injury in rats	56
Table 10c: Effect of PEBP, EABP and AQBP on liver function enzymes, ALT, AST, ALP (U/L), total protein (g/dL) and LDH (U/L)	57

Table 11:	Total phenolic content of different extracts	73
Table 12:	Superoxide scavenging activity of different extracts	76
Table 13:	Oxygen radical absorbance capacity of different extracts	77
Table 14:	Effect of extracts on the anti-inflammatory mediators using the in vitro lipoxygenase and xantine oxidase assays	78
Table 15:	The solvent system used for the HPLC profiling	84
Table 16:	Comparison on the phytochemical constituents of the leaves in different extracts of <i>B. purpurea</i> leaves	85

LIST OF FIGURES

Figure	Page
Figure 1: <i>Bauhinia purpurea</i> flower	7
Figure 2: <i>Bauhinia purpurea</i> leaves	7
Figure 3: Location of the liver in the body	12
Figure 4: Anatomy of liver	13
Figure 5: Zonation of hepatocytes	15
Figure 6: Location of the liver in rat	20
Figure 7: Rat liver (Visceral surface)	20
Figure 8: Hepatic lobes, hepatic portal (black) and hepatic veins (red) in rats liver	21
Figure 9: CCl ₄ metabolism	23
Figure 10: PCM metabolism	24
Figure 11: A chromatogram of single sample showing void time (t_M), retention time (t_R), peak height (h), and peak width (w_b).	27
Figure 12: The partitioning scheme of extracts	31
Figure 13: Timeline of hepatotoxicity assay	35
Figure 14: 14A) normal liver, 14B) liver intoxicated with 1mL/kg CCl ₄ : gross image showing color changes of liver lobes from dark maroon to brownish and coarse surface, 14C) liver pre-treated with 200mg/kg Silymarin and induced with CCl ₄ : showing normal liver color, 14D) liver pre-treated with 50mg/kg MEBP and induced by CCl ₄ , 14E) liver pre-treated with 250mg/kg MEBP and induced by CCl ₄ , 14F) liver pre-treated with 500mg/kg MEBP and induced by CCl ₄ .	40
Figure 15: 15A) Normal liver parenchyma, 15B) Section of liver tissue treated with 1mL/kg CCl ₄ (i.p) showing massive necrosis, inflammation and steatosis. (H & E, X100). NH) normal hepatocyte, CV) central vein. N) necrosis. I) inflammation, S) steatosis.	41

- Figure 15: 15C) Section of liver tissue pre-treated with 200 mg/kg silymarin followed by CCl₄ showing preservation of normal hepatocytes. 15D) Section of liver tissue pre-treated with 50 mg/kg MEBP followed by CCl₄ showing moderate tissue necrosis, inflammation, and mild steatosis. (H & E, X100). CV) central vein. N) necrosis. I) inflammation, S) steatosis, NH) normal hepatocytes. 42
- Figure 15: 15E) Section of liver tissue pre-treated with 250 mg/kg MEBP followed by CCl₄ showing mild inflammation and steatosis. 15F) Section of liver tissue pre-treated with 500 mg/kg MEBP followed by CCl₄ showing normal histology with mild inflammation and steatosis. (H & E, X100). CV) central vein. I) inflammation, S) steatosis. 43
- Figure 16: 16A) Normal liver. 16B) Liver intoxicated with 3 g/kg PCM: gross image shows major color changes of liver lobes (arrow). 16C) Liver pre-treated with 200 mg/kg silymarin and induced with PCM: spot color changes were noted (arrow). 16D) Liver pre-treated with 50 mg/kg MEBP and induced with PCM. 16E) Liver pre-treated with 250 mg/kg MEBP and induced with PCM. 16F) Liver pre-treated with 500 mg/kg MEBP and induced with PCM. 48
- Figure 17: 17A) Normal liver parenchyma, 17B) Section of liver tissue treated with 3g/kg PCM (p.o) showing massive necrosis, haemorrhage and inflammation. (H & E, X100). CV) central vein. NH) normal hepatocytes N) necrosis. I) inflammation. H) haemorrhage. 49
- Figure 17: 17C) Section of liver tissue pre-treated with 200 mg/kg silymarin followed by PCM showing preservation of normal hepatocytes. 17D)) Section of liver tissue pre-treated with 50 mg/kg MEBP followed by PCM showing tissue moderate necrosis and inflammation. (H & E, X100). CV) central vein, N) necrosis, I) inflammation. 50
- Figure 17: 17E) Section of liver tissue pre-treated with 250 mg/kg MEBP followed by PCM showing mild inflammation. 17F) Section of liver tissue pre-treated with 500 mg/kg MEBP followed by PCM showing normal histology with mild inflammation. (H & E, X100). CV) central vein, I) inflammation. 51

Figure 18:	18A) Normal liver parenchyma, 18B) Section of liver tissue treated with 3g/kg PCM (p.o) showing massive necrosis, and inflammation. (H & E, X100). CV) central vein. N) necrosis, I) inflammation, NH) normal hepatocytes.	58
Figure 18:	18C) Section of liver tissue pre-treated with 200 mg/kg silymarin followed by PCM showing preservation of normal hepatocytes. 18D) Section of liver tissue pre-treated with 50 mg/kg PEBP followed by PCM showing massive tissue necrosis and mild inflammation (H & E, X100). CV) central vein. N) necrosis, I) inflammation, NH) normal hepatocytes.	59
Figure 18:	18E) Section of liver tissue pre-treated with 250 mg/kg PEBP followed by PCM showing moderate necrosis and mild inflammation. 18F) Section of liver tissue pre-treated with 500 mg/kg PEBP followed by PCM showing moderate necrosis and inflammation. (H & E, X100). CV) central vein. N) necrosis. I) inflammation.	60
Figure 18:	18G) Section of liver tissue pre-treated with 50 mg/kg EABP followed by PCM showing moderate necrosis. 18H) Section of liver tissue pre-treated with 250 mg/kg EABP followed by PCM showing mild necrosis. (H & E, X100). N) necrosis. CV) central vein.	61
Figure 18:	18I) Section of liver tissue pre-treated with 500 mg/kg EABP followed by PCM showing mild inflammation and necrosis. 18J) Section of liver tissue pre-treated with 50 mg/kg AQBP followed by PCM showing moderate inflammation and necrosis (H & E, X100). CV) central vein. N) necrosis. I) inflammation.	62
Figure 18:	18K) Section of liver tissue pre-treated with 250 mg/kg AQBP followed by PCM showing mild necrosis and inflammation. 18L) Section of liver tissue pre-treated with 500 mg/kg AQBP followed by PCM showing mild inflammation. (H & E, X100). CV) central vein. N) necrosis. I) inflammation	63
Figure 19a:	DPPH scavenging activity of MEBP	74
Figure 19b:	DPPH scavenging activity of PEBP	74
Figure 19c:	DPPH scavenging activity of EABP	75

Figure 19d:	DPPH scavenging activity of AQBP	75
Figure 20a:	The HPLC profile of MEBP at two different wavelengths, namely 254 and 366 nm.	87
Figure 20b:	The UV spectra analysis of peak 4 (RT = 4.94 min), peak 5 (RT = 6.27 min) and peak 6 (RT = 7.12 min) of the MEBP at 254 nm exhibiting the λ max at 254-351 nm, 265-345 nm and 254-352 nm, respectively, suggesting, in part, the presence of flavonoid-based compounds.	88
Figure 20c:	The UV spectra analysis of rutin (RT 20.4 min) and MEBP (RT 20.018 min) were observed at their respective λ max at the region of 255.5-352.9 and 254.3-351.7 nm.	89
Figure 20d:	Comparison between chromatogram of the standard compound rutin with the chromatogram of MEBP at 330 nm showing both peaks are not parallel to each other, indicating that rutin is not present in MEBP.	90
Figure 21a:	The HPLC profile of PEBP at six different wavelengths, namely 210, 254, 280, 300, 330 and 366 nm.	92
Figure 21b:	The UV spectra analysis of peak 1 (RT = 26.85 min) and peak 2 (RT = 29.4 min) of the PEBP at 254 nm and 339nm exhibiting the λ max at 273 nm and 323 nm respectively.	93
Figure 22a:	The HPLC profile of EABP at six different wavelengths, namely 210, 254, 280, 300, 330 and 366 nm.	94
Figure 22b:	The UV spectra analysis of peak 3 (RT = 17.93 min), peak 4 (RT = 19.20 min) and peak 5 (RT = 19.83 min) of the EABP at 366 nm exhibiting the λ max at 256-353 nm, 265-343 nm and 254-351 nm, respectively, suggesting, in part, the presence of flavonoid-based compounds.	95
Figure 23a:	The HPLC profile of AQBP at six different wavelengths, namely 210, 254, 280, 300, 330 and 366 nm.	96
Figure 23b:	The UV spectra analysis of peak 1 (RT = 7.75 min), peak 2 (RT = 3.69 min), and peak 4 (RT = 17.89 min) and peak 7 (RT = 19.83 min) of the AQBP at 210 nm, 254 nm, and 366 nm respectively. The peaks exhibit the λ_{max} at 251-330 nm, 271-354 nm, 256-349 nm and 254-349 nm, respectively, suggesting, in part, the presence of flavonoid-based compounds.	97

LIST OF ABBREVIATIONS

AAPH	2,2'-azobis-2-methyl-propanimidamide, dihydrochloride
ALP	Alkaline phosphatase
ALT	Alanine aminotransferase
ANOVA	Analysis of variance
AQBP	Aqueous extract of Bauhinia purpurea
AST	Aspartate aminotransferase
ATP	Adenosine triphosphate
AUC	Area under the curve
$\text{CCl}_3\cdot$	Trichloromethyl free radical
CCl_4	Carbon tetrachloride
$\text{Cl}_3\text{COO}\cdot$	Trichloromethyl peroxy
COX	Cyclooxygenase
CYP450	Cytochrome P450
CYP450	Cytochrome P450
DMSO	Dimethyl sulfoxide
DPPH	2,2-diphenyl-1-picrylhydrazyl
GSH	Glutathione
H&E	Haematoxylin & Eosin staining
H_2O_2	Hydrogen peroxide
HCV	Chronic viral hepatitis C
$\text{HO}\cdot$	Hydroxyl radical
HPLC	High performance liquid chromatography
i.p	Intraperitoneally
IC50	Median inhibitory concentration
LDH	Lactate dehydrogenase
LOX	Lipoxygenase
MEBP	Methanol extract of Bauhinia purpurea
MeOH	Methanol
NAC	N-acetyl cysteine
NAPQI	N-acetyl-p-benzoquinoneimine
NBT	Nitroblue tetrazolium
o.p	Orally
O_2	Oxygen
$\text{O}_2\cdot^-$	Superoxide anion
ORAC	Oxygen radical absorbance capacity
PCM	Paracetamol
PEBP	Petroleum ether extract of Bauhinia purpurea
PPAR- α	Peroxisome proliferator-activated receptor alpha
PUFA	Polyunsaturated fatty acids
ROS	Reactive oxygen species
SEM	Standard error mean

SOD	Superoxide dismutase
TP	Total protein
TPC	Total phenolic content
WHO	World Health Organization
XO	Xanthine oxidase



CHAPTER 1

INTRODUCTION

Continual damage to the liver by acute liver insult eventually results in the development of hepatic fibrogenesis. Advanced fibrogenesis leads to the development of severe life-threatening complications in patients, which promotes structural changes to the tissue, known as cirrhosis. Cirrhosis is a common and predictable aftermath of irreversible damage to the liver parenchyma triggered by a variety of etiologies. The leading causes of liver cirrhosis in the Western countries are mostly attributed to chronic hepatitis C and alcohol abuse (Qua and Goh, 2011). The increasing incidence of mortality due to liver diseases has been reported to be the tenth leading cause of death in the United States (Liver Center, Saint Louis University). Meanwhile, chronic hepatitis B appears to be the most prevalent cause of liver cirrhosis in the Asian Pacific region. Malaysia is universally acknowledged to have a unique multiracial composition of Asian populations that comprises three main races: Malay, Chinese, and Indian (Qua and Goh, 2011). In the medical field, the diagnosis of liver cirrhosis among these races is no longer a recently discovered, uncommon occurrence. The only issue is that the etiologies of liver cirrhosis of each ethnic group are apparently different according to their religions, cultural beliefs (Qua and Goh, 2011), lifestyle, and environmental factors.

Chronic liver disease encompasses not only the aforementioned factors, but also a broad spectrum of principles: it also involves drug toxicity, autoimmune disease, fatty infiltration, hereditary linkage, and cryptogenic (unidentifiable) causes. It is understood that hepatic fibrogenesis has a potential reversible component; hindrance of liver trauma has become a reliable therapeutic strategy to minimize the progression of advanced liver disease.

Even though the modern drug technologies of high-throughput screening and synthetic chemistry of the 20th Century have been expanded upon greatly, nature, particularly plant-based therapies, has remained the most valued resource in the drug development arena (Helmstädter and Staiger, 2014; Balunas and Kinghorn, 2005; Fabricant and Farnsworth, 2001). Drug discovery of current active agents has to be discussed from a phytopharmaceutical viewpoint. An analysis from 1981 to 2010 by Newman and Cragg (2012) showed that more than two-thirds of the drug active compounds recently introduced are likely derived from natural sources, and only about 30% are of completely synthetic origin (Newman and Cragg, 2012). For decades, therapeutic practices and roles of medicinal herbs for treating disease have been gathered through an array of trials and errors, and have been documented in the history of medicine. Regardless of the abundance of the number of modern drugs in the pharmaceutical market, traditional medicine has been favored as the primary option for alternative medicines, considering its low cost and effectiveness, and cultural, historical, and even religious inclinations (Priya et al., 2010). The World Health Organization (WHO)

estimated that about 80% of developing societies consider natural products their most preferred healthcare option (WHO, 2002).

In retrospect, there has been tremendous drug discovery from natural products since World War I, but surprisingly, less than 10% of the 250,000 species from worldwide biodiversity has been studied for medicinal purposes (Ramasamy et al., 2011; McChesny et al., 2007), leaving many species awaiting therapeutic exploration. Malaysia has been acknowledged as a land of floral and faunal prosperity, and is believed to be a reservoir of a large collection of potential medicinal plants. An increasing trend in Malaysia was the recent swing in interest from synthetic allopathic drugs to herbal medicine. In 1999, the herbal and natural product domestic market was reported to be Malaysian ringgit (RM)4.55 billion, and the current appraisal growth rate is estimated to be worth 15–20% annually (Nordin et al., 2008; Aziz, 2003). Alongside economic factors, the increased interest in the herbal industry in Malaysia has apparently been caused by changes in lifestyle, increased health consciousness, and the costliness of synthetic medicines (Aziz, 2003). From the perspective of the herbal-based market, particularly herbal medicines, the natural herb heritage in Malaysia merits a favorable position in the industry. An in-depth report by the Ministry of Natural Resources and Environment on Biodiversity in Malaysia (2006) showed that Malaysia enjoys the advantage of genetic resource diversity, lush tropical climate, growing demand for specialty natural products, and indigenous knowledge (Biodiversity in Malaysia, 2006).

To exploit these sources for prospective research, particularly hepatoprotective studies, *Bauhinia purpurea* was selected to be investigated on a large scale. *B. purpurea*, from the family Fabaceae and locally known as *tapak kuda*, is a native plant in Malaysia that has been widely tested and documented for its promising pharmacological properties, such as antioxidant (Joshi et al., 2009; Zakaria et al., 2011a; Annegowda et al., 2012), antiulcer (Zakaria et al., 2011b; Zakaria et al., 2012), anti-inflammatory (Boonphong et al., 2007), antinociceptive, antipyretic (Zakaria et al., 2007; Zakaria et al., 2009), antiproliferative (Zakaria et al., 2011a), antimicrobial (Murugan and Mohan, 2011), and wound healing (Ananth et al., 2010). Nevertheless, its hepatoprotective properties in particular have not been explored properly. As such, further research on its hepatoprotective activity is significant for nominating another plant to the list of potential medicinal hepatoprotective plant-based products.

Problem statement

Liver diseases have been acknowledged as one of the major threats to community health. Contributory factors of these problems are mainly attributable to chemicals such as paracetamol (PCM; overdoses), excessive alcohol consumption, autoimmune disorders, and infections. PCM, a mild analgesic and antipyretic drug developed in the past few decades, causes severe liver injuries (necrosis) in humans and experimental

animals following overdose of the drug. Alcoholic liver disease is the second most common reason for liver transplantation (Adewusi and Afolayan, 2010). Marzilawati *et al.* (2012) reported that acute liver failure caused by PCM toxicity is a major problem leading to death worldwide, whereas acute liver failure among Asians is commonly caused by viral hepatitis, and infrequent cases of PCM toxicity are reported. Nevertheless, it has been highlighted that data analysis of N-acetylcysteine (NAC) therapy, currently one of the most dependable drugs for countering PCM toxicity, cannot be taken for granted, which stated that it is not cost-effective in managing Asian patients with accidental PCM overdose. Moreover, toxicology research on NAC documented by the United States National Library of Medicine states that it has several side effects, usually involving anaphylactoid responses.

Justification for studying the hepatoprotective potential of *B. purpurea*

In spite of the advanced development of modern medicine, there are several obstacles faced by the public, such as the high cost of available drugs, the presence of drug side effects that prevent patients with certain health conditions from consuming a certain drug, and lack of drug availability. Therefore, it is highly recommended to search for alternative medicine for treating liver ailments as a substitute for currently used drugs that have fewer or no side effects and are cheaper and widely available. Encouraging research on medicinal plants indicates that phytochemicals can be exploited for treating many health problems. Extensive studies have been conducted on plant natural products, and most of these products have shown potential as new promising hepatoprotective agents; thus, this study, which aimed to discover the potential hepatoprotective activity of *B. purpurea* leaves, might add another candidate to the list. Scientifically, *B. purpurea* is not traditionally known to have hepatoprotective properties. Nevertheless, the factors that might be involved in its cytoprotective effects can be evaluated and further studied for future plant-derived drug development. Previous studies on *B. purpurea* reported the presence of antioxidant and anti-inflammatory activity that is relevant to hepatoprotective activity. Considering these reports, the antioxidant and anti-inflammatory activity indicate different pathways assisting the hepatoprotective effect. In general, free radicals or reactive oxygen species (ROS) generated from drug or chemical metabolism appear to be the fundamental mechanisms underlying most human ailments. The antioxidant and anti-inflammatory properties of plants facilitate the free radical scavenging process and regulate the inflammatory response, respectively, which are believed to initiate their detrimental effects on the liver. Therefore, this study is expected to discover the capacity of *B. purpurea* for hepatoprotective activity.

Hypothesis

Methanolic extract of *B. purpurea* leaves (MEBP) exerts hepatoprotective activity in PCM- and carbon tetrachloride (CCl₄)- induced liver toxicity assays, and one or more of its partitions is expected to have good hepatoprotective activity in PCM-induced liver toxicity.

General objectives:

- To determine hepatoprotective activity of methanol extract of *Bauhinia purpurea* leaves and its partitions using rat models

Specific objectives:

- To determine hepatoprotective effect of methanolic extract of *B.purpurea* leaves (MEBP) against carbon tetrachloride and paracetamol-induced liver toxicity models in rat, and then find out the most effective partition of MEBP; petroleum ether, ethyl acetate and aqueous extracts on liver toxicity study,
- To examine the involvement of antioxidant and anti-inflammatory activities of the extracts as part of the hepatoprotective pathway,
- To screen for the bioactive compounds present in MEBP and its partitions using high performance liquid chromatography (HPLC)

REFERENCES

- Adewusi, E. A. and Afolayan, A. J. (2010). A review of natural products with hepatoprotective activity. *Journal of Medicinal Plants Research*. 4: 1318-1334.
- Ahsan, M. R., Islam, K. M. and Bulbul, I. J. (2009). Hepatoprotective activity of Methanol Extract of some medicinal plants against carbon tetrachloride-induced hepatotoxicity in rats. *European Journal of Scientific Research*. 37: 302-310.
- Ahsan, M.R., Islam, K.M. and Bulbul, I.J. (2009). Hepatoprotective activity of Methanol Extract of some medicinal plants against carbon tetrachloride-induced hepatotoxicity in rats. *European Journal of Scientific Research*. 37: 302-310.
- Akkol, E. K., Tatli, I. I. and Akdemir, Z. S. (2007). Antinociceptive and anti-inflammatory effects of saponin and iridoid glycosides from *Verbascum pterocalycinum* var. *mutense* Hub.-Mor. *Zeitschrift für Naturforschung. C, A Journal of Biosciences*. 62: 813-820.
- Amacher, D. E. (1998). Serum transaminase elevations as indicators of hepatic injury following the administration of drugs. *Regulatory toxicology and pharmacology*. 27: 119-130.
- Amacher, D. E. (1998). Serum transaminase elevations as indicators of hepatic injury following the administration of drugs. *Regulatory Toxicology and Pharmacology*. 27: 119-130.
- Ananth, K. V., Asad, M., Kumar, N. P., Asdaq, S. M. B. and Rao, G. S. (2010). Evaluation of wound healing potential of *Bauhinia purpurea* leaf extracts in rats. *Indian journal of pharmaceutical sciences*. 72: 122.
- Annegowda, H. V., Mordi, M. N., Ramanathan, S., Hamdan, M. R. and Mansor, S. M. (2012). Effect of extraction techniques on phenolic content, antioxidant and antimicrobial activity of *Bauhinia purpurea*: HPTLC determination of antioxidants. *Food analytical methods*. 5:226-233.
- Asolkar, L. V., Kakkar, K. K. and Charke, O. J. (2000). Second supplement to glossary of Indian medicinal plants with active principles. National Institute of Science Communication, New Delhi.
- Aziz, R. A. (2003). *Turning Malaysia Into A Global Herbal Producer: A Personal Perspective*. Universiti Teknologi Malaysia.
- Azlim Almey, A. A., Ahmed Jalal Khan, C., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R. and Kamarul Rahim, K. (2010). Total phenolic content and primary antioxidant activity of methanolic and ethanolic extracts of aromatic plants' leaves. *International Food Research Journal*. 17: 1077-1084.

- Babu, D., Gurumurthy, P., Borra, S. K. and Cherian, K. M. (2013). Antioxidant and free radical scavenging activity of triphala determined by using different in vitro models. *Journal of Medicinal Plants Research*. 7: 2898-2905.
- Balaji, H., Padmaja, T. K., Naidu, P. B., Naik, S. R. and Meriga, B. (2013). Anti-inflammatory and antioxidant activity of ethanolic extract of *Bauhinia purpurea* bark. *International Journal of Drug Delivery*. 4: 507-514.
- Balan, T., Sani, M. H. M., Ahmad, S. H. M., Suppaiah, V., Mohtarrudin, N., Jamaludin, F., and Zakaria, Z. A. (2014). Antioxidant and anti-inflammatory activities contribute to the prophylactic effect of semi-purified fractions obtained from the crude methanol extract of *Muntingia calabura* leaves against gastric ulceration in rats. *Journal of ethnopharmacology*. doi: 10.1016/j.jep.2014.12.017
- Balunas, M.J. and Kinghorn, A.D. (2005). Drug discovery from medicinal plants. *Life Science*. 78: 431–441.
- Betteridge, D. J. (2000). What is oxidative stress?. *Metabolism*. 49:3-8.
- Bhagwat, S., Haytowitz, D. B. and Holden, J. M. (2007). USDA database for the Oxygen Radical Absorbance Capacity (ORAC) of selected foods. In *American Institute for Cancer Research Launch Conference*. Washington, DC, November (pp. 1-2).
- Biodiversity in Malaysia (2006). Ministry of Natural Resources and Environment Malaysia, ISBN 983-42860-1-5
- Blois, M.S. (1958). Antioxidant determinations by the use of a stable free radical. *Nature*. 181: 1199–1200.
- Bodakhe, S. H. and Ram, A. (2007). Hepatoprotective Properties of *Bauhinia variegata* Bark Extract. *Yakugaku Zasshi*. 127: 1503-1507
- Bokov, A., Chaudhuri, A. and Richardson, A. (2004). The role of oxidative damage and stress in aging. *Mechanisms of Ageing and Development*. 125: 811–826.
- Bonini, M. G., Miyamoto, S., Di Mascio, P. and Augusto, O. (2004). Production of the carbonate radical anion during xanthine oxidase turnover in the presence of bicarbonate. *Journal of Biological Chemistry*. 279: 51836-51843.
- Boonphong, S., Puangsombat, P., Baramée, A., Mahidol, C., Ruchirawat, S. and Kittakoop, P. (2007). Bioactive compounds from *Bauhinia purpurea* possessing antimalarial, antimycobacterial, antifungal, anti-inflammatory, and cytotoxic activities. *Journal of Natural Products*. 70: 795-801.
- Brooks, P.M. and Day, R.O. (1991). Nonsteroidal anti-inflammatory drugs: Differences and similarities. *New England Journal of Medicine*. 324: 1716
- Bulger, E. M. and Maier, R. V. (2001). Antioxidants in critical illness. *Archives of Surgery*. 136: 1201.

- Cao, G., Alessio, H. M., and Cutler, R. G. (1993). Oxygen-radical absorbance capacity assay for antioxidants. *Free Radical Biology and Medicine*. 14: 303-311.
- CarexCanada. http://www.carexcanada.ca/en/carbon_tetrachloride/ accessed on 3rd August 2013
- CBD Country Profile. <http://www.cbd.int/countries/profile/default.shtml?country=my#status> accessed on 9th January 2014
- Chang, W.S., Lin, C.C. and Chiang, H.C. (1996). Superoxide anion scavenging effect of coumarins. *The American Journal of Chinese Medicine*. 24: 11–17.
- Chatterjee, A. and Pakrashi, S. C. (1992). The treatise on Indian medicinal plants: vol. 2. *Council of Scientific and Industrial Research, New Delhi*.
- Chen, J., Zhao, Y., Tao, X. Y., Zhang, M., and Sun, A. D. (2015). Protective effect of blueberry anthocyanins in a CCl₄-induced liver cell model. *LWT-Food Science and Technology*. 60: 1105-1112.
- Cheng, N., Ren, N., Gao, H., Lei, X., Zheng, J. and Cao, W. (2013). Antioxidant and hepatoprotective effects of *Schisandra chinensis* pollen extract on CCl₄ induced acute liver damage in mice. *Food and Chemical Toxicology*. 55: 234-240.
- Choudhari, A. S., Suryavanshi, S. A., Ingle, H. and Kaul-Ghanekar, R. (2011). Evaluating the antioxidant potential of aqueous and alcoholic extracts of *Ficus religiosa* using ORAC assay and assessing their cytotoxic activity in cervical cancer cell lines. *Biotechnology, Bioinformatics and Bioengineering*. 1:443-450.
- Clawson, G. A. (1989). Mechanisms of carbon tetrachloride hepatotoxicity. *Pathology and immunopathology Research*. 8:104-112.
- Colnot, S. and Perret. C. (2011). Liver Zonation. *Molecular pathology of liver diseases*.5: 7-16
- Conforti, F., Sosa, S., Marrelli, M., Menichini, F., Statti, G. A., Uzunov, D., Tubaro, A., Menichini, F. and Loggia, R. D. (2008). *In vivo* anti-inflammatory and *in vitro* antioxidant activities of Mediterranean dietary plants. *Journal of Ethnopharmacology*. 116: 144-151.
- Cragg, G.M. and Newman, D.J. (2005). Biodiversity: A continuing source of novel drug leads. *Pure and Applied Chemistry*. 77: 7-24.
- Dai, J. and Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*.15: 7313-7352.
- Das, A. K., Dutta, B. K. and Sharma, G. D. (2008). Medicinal plants used by different tribes of Cachar district, Assam. *Indian Journal of Traditional Knowledge*. 7: 446-54.

- Devaraj, S., Ismail, S., Ramanathan, S., Marimuthu, S. and Fei, Y. M. (2010). Evaluation of the hepatoprotective activity of standardized ethanolic extract of *Curcuma xanthorrhiza* Roxb. *Journal of Medicinal Plants Research*. 4: 2512-2517.
- Domitrović, R., Jakovac, H., Marchesi, V. V., Vladimir-Knežević, S., Cvijanović, O., Tadić, Ž., Romić, Ž and Rahelić, D. (2012). Differential hepatoprotective mechanisms of rutin and quercetin in CCl₄-intoxicated BALB/cN mice. *Acta Pharmacologica Sinica*. 33: 1260-1270.
- Dong, M. W. (2006). *Modern HPLC for practicing scientists*. John Wiley & Sons.
- Eckardt, K. U., Pugh, C. W., Ratcliffe, P. J. and Kurtz, A. (1993). Oxygen-dependent expression of the erythropoietin gene in rat hepatocytes in vitro. *Pflügers Archiv*. 423: 356-364.
- El-Beshbishy, H. A., Mohamadin, A. M., Nagy, A. A. and Abdel-Naim, A. B. (2010). Amelioration of tamoxifen-induced liver injury in rats by grape seed extract, black seed extract and curcumin. *Indian Journal of Experimental Biology*. 48: 280– 288.
- Elekofehinti, O. O., Adanlawo, I. G., Komolaf, K. and Ejelonu, O. C. (2012). Saponins from *Solanum anguivi* fruits exhibit antioxidant potential in Wistar rats. *Annals of Biological Research*. 3: 3212-3217.
- Fabricant, D.S. and Farnsworth, N.R. (2001). The value of plants used in traditional medicine for drug discovery. *Environmental Health Perspectives*. 109: 69–75.
- Fang, E. F., Bah, C. S. F., Wong, J. H., Pan, W. L., Chan, Y. S., Ye, X. J. and Ng, T. B. (2012). A potential human hepatocellular carcinoma inhibitor from *Bauhinia purpurea* L. seeds: from purification to mechanism exploration. *Archives of toxicology*. 86:293-304.
- Feldstein, A. E., Canbay, A., Angulo, P., Taniai, M., Burgart, L. J., Lindor, K. D. and Gores, G. J. (2003). Hepatocyte apoptosis and fas expression are prominent features of human nonalcoholic steatohepatitis. *Gastroenterology*. 125: 437-443.
- Ferreira, J. F., Luthria, D. L., Sasaki, T. and Heyerick, A. (2010). Flavonoids from *Artemisia annua* L. as antioxidants and their potential synergism with artemisinin against malaria and cancer. *Molecules*. 15: 3135-3170.
- Fong, B. M. W., Siu, T. S. and Tam, S. (2011). Persistently increased acetaminophen concentrations in a patient with acute liver failure. *Clinical chemistry*. 57: 9-11.
- Foster, S. J., McCormick, M. E. and Howarth, A. (1986). The contribution of cyclooxygenase and lipoxygenase products to acute inflammation in the rat. *Inflammation Research*. 17: 358-359.
- Ginès, P., Kamath, P. S. and Arroyo, V. (2011). Chronic Liver Failure: Mechanisms and Management, *Clinical Gastroenterology*. Springer Dordrecht Heidelberg London New York

- Giordano, F.J. (2005). Oxygen, oxidative stress, hypoxia, and heart failure. *The haloalkanes: carbon tetrachloride as a toxicological model. Critical Reviews in Toxicology*. 33: 105–136.
- Giordano, F.J. (2005). Oxygen, oxidative stress, hypoxia, and heart failure. *The Journal of Clinical Investigation*. 115: 500–508.
- Gupta, A. K., Chitme, H., Dass, S. K. and Misra, N. (2006). Hepatoprotective activity of *Rauwolfia serpentina* rhizome in paracetamol intoxicated rats. *Journal of Pharmacology and Toxicology*. 1: 82-88.
- Han, K. L., Jung, M. H., Sohn, J. H. and Hwang, J. K. (2006). Ginsenoside 20 (S)-Protopanaxatriol (PPT) Activates Peroxisome Proliferator-Activated Receptor. GAMMA.(PPAR. GAMMA.) in 3T3-L1 Adipocytes. *Biological and Pharmaceutical Bulletin*. 29: 110-113.
- Hassan, H. M. (2012). Hepatoprotective effect of red grape seed extracts against ethanol-induced cytotoxicity. *Global Journal of Biotechnology and Biochemistry*. 7: 30-37.
- Hazai, E., Vereczkey, L. and Monostory, K. (2002). Reduction of toxic metabolite formation of acetaminophen. *Biochemical and Biophysical Research Communications*. 291: 1089-1094.
- Helmstädter, A. and Staiger, C. (2014). Traditional use of medicinal agents: a valid source of evidence. *Drug Discovery Today*. 19:1.
- Hisam, E. E. A., Zakaria, Z. A., Mohtaruddin, N., Rofiee, M. S., Hamid, H. A. and Othman, F. (2012). Antiulcer activity of the chloroform extract of *Bauhinia purpurea* leaf. *Pharmaceutical Biology*. 50: 1498-1507.
- Houghton, P. J. and Raman, A. (1998). Laboratory Handbook for the fractionation of Natural Extracts. 1998 Edition, Springer.
- http://images.dailytech.com/nimage/11466_Paracetamol_metabolism.png accessed on 18th January 2015
- <http://www.informatics.jax.org/cookbook/images/55.jpg> accessed on 9th January 2015
- <http://www.nature.com/labinvest/journal/v92/n3/images/labinvest2011193f9.jpg> accessed on 18th January 2015
- http://www.niehs.nih.gov/research/atniehs/labs/assets/images/l_m/liversjpg.jpg accessed on 9th January 2015
- <http://www.paradoja7.com/the-human-liver-functions/the-human-liver-functions/> accessed on 23rd August 2014
- http://www2.massgeneral.org/cancerresourceroom/types/gi/illustrations/images/overview_front.jpg accessed on 9th January 2015

- Huang, D., Ou, B., Hampch-Woodill, M., Flanagan, J.A. and Prior, R.L. (2002). High throughput assay of oxygen radical absorbance capacity (ORAC) using a multichannel liquid handling system coupled with a microplate fluorescence reader in 96-well format. *Journal of Agricultural and Food Chemistry*. 5: 4437–4444.
- Huebert, R. C. and Shah, V. H. (2010). Hepatic sinusoidal endothelial cells. In *Signaling Pathways in Liver Diseases* (pp. 79-91). Springer Berlin Heidelberg.
- Iniaghe, O. M., Malomo, S. O. and Adebayo, J. O. (2008). Hepatoprotective effect of the aqueous extract of leaves of *Acalypha racemosa* in carbon tetrachloride treated rats. *Journal of Medicinal Plant Research*. 2: 301-305.
- Jaeschke, H. (2000). Reactive oxygen and mechanisms of inflammatory liver injury. *Journal of Gastroenterology and Hepatology*. 15: 718–724.
- Jaeschke, H. and Bajt, M. L. (2006). Intracellular signaling mechanisms of acetaminophen-induced liver cell death. *Toxicological Sciences*. 89: 31-41.
- Jancova, P., Anzenbacher, P. and Anzenbacherova, E. (2010). Phase II drug metabolizing enzymes. *Biomedical Papers*. 154: 103-116.
- Jatwa, R. and Kar, A. (2009). Amelioration of metformin-induced hypothyroidism by *Withania somnifera* and *Bauhinia purpurea* extracts in Type 2 diabetic mice. *Phytotherapy Research*. 23: 1140-1145.
- Joshi, A. B., Desai, R. R. and Bhoje, M. P. (2013). Phytochemical investigation of the hexane extract of stem bark of *Bauhinia purpurea* Linn. *Der Pharma Chemica*. 5: 116-121.
- Joshi, V.D., Verma, T. and Shetty, P.R. (2009). Antioxidant potential of *Bauhinia purpurea* Linn. leaves. *International Journal of Pharmaceutical Research*. 1: 51-55.
- K"ahk"onen, M.P., Hopia, A.I., Vuorela, H.J., Rauha, J.P., Pihlaja, K., Kujala, T.S. and Heinonen, M. (1999). Antioxidant activity of plant extracts containing phenolic compounds. *Journal of Agricultural and Food Chemistry*. 47: 3954–3962.
- Kadir, F. A., Kassim, N. M., Abdulla, M. A. and Yehye, W. A. (2013). Hepatoprotective role of ethanolic extract of *Vitex negundo* in thioacetamide-induced liver fibrosis in male rats. *Evidence-Based Complementary and Alternative Medicine*. 2013:9
- Kamarolzaman, M. F. F., Yahya, F., Mamet, S. S., Jakius, K. F., Mahmood, N. D., Shahril, M. S, Mohtarrudin, N., Suhaili, Z. and Zakaria, Z. A. (2014). Gastroprotective activity and mechanisms of action of *Bauhinia purpurea* Linn (Leguminosae) leaf methanol extract. *Tropical Journal of Pharmaceutical Research*. 13: 1889-1898.

- Kamble, S. Y., Patil, S. R., Sawant, P. S., Sawant, S., Pawar, S. G. and Singh, E. A. (2010). Studies on plants used in traditional medicine by Bhilla tribe of Maharashtra. *Indian Journal of Traditional Knowledge*. 9: 591-598.
- Kamisan, F.H., Yahya, F., Ismail, N.A., Din, S.S., Mamat, S.S., Zabidi, Z., Zainulddin, W.N., Mohtarrudin, N., Husain, H., Ahmad, Z. and Zakaria, Z.A. (2013). Hepatoprotective activity of methanol extract of *Melastoma malabathricum* leaf in rats. *Journal of Acupuncture & Meridian Studies*. 6: 52-5.
- Kanel, G. C., Korula, J. and Renshaw, A. (2005). Atlas of Liver Pathology 2nd edition. Elsevier Saunders, Philadelphia.
- Kapoor, M., Shaw, O. and Appleton, I. (2005). Possible anti-inflammatory role of COX-2-derived prostaglandins: implications for inflammation research. *Current opinion in investigational drugs (London, England: 2000)*. 6: 461-466.
- Kazakevich, Y. V., and Lobrutto, R. (2007). *HPLC for pharmaceutical scientists*. John Wiley & Sons.
- Kerksick, C. and Willoughby, D. (2005). The antioxidant role of glutathione and N-acetyl-cysteine supplements and exercise-induced oxidative stress. *Journal of the International Society of Sports Nutrition*. 2: 38-44.
- Khare, C.P. (2004). Encyclopaedia of Indian Medicinal Plant (pp. 95-96). Springer-Verlag, New York.
- Klaunig, J.E. and Kamendulis, L.M. (2004). The role of oxidative stress in carcinogenesis. *Annual Review of Pharmacology and Toxicology*. 44: 239-267.
- Kogure, K., Ishizaki, M., Nemoto, M., Kuwano, H., and Makuuchi, M. (1999). A comparative study of the anatomy of rat and human livers. *Journal of Hepato-Biliary-Pancreatic Surgery*. 6: 171-175.
- Korkina, L. G. and Afanas' Ev, I. B. (1996). Antioxidant and chelating properties of flavonoids. *Advances in Pharmacology*. 38: 151-166.
- Kose, N., Yamamoto, K., Sai, Y., Isawa, M., Suwa, T. and Nakashima, E. (2005). Prediction of theophylline clearance in CCl₄-treated rats using in vivo CYP1A2 and CYP3A2 contents assessed with the PKCYP test. *Drug Metabolism and Pharmacokinetics*. 20: 168-176.
- Krishnakumar, M. N., Latha, P. G., Suja, S. R., Shine, V. J., Shyamal, S., Anuja, G. I., Sini S., Pradeep, S., Shikha, P., Somasekharan Unni, P. K. and Rajasekharan, S. (2008). Hepatoprotective effect of *Hibiscus hispidissimus Griffith*, ethanolic extract in paracetamol and CCl₄ induced hepatotoxicity in Wistar rats. *Indian Journal of Experimental Biology*. 46: 653.
- Kroymann, J. (2011). Natural diversity and adaptation in plant secondary metabolism. *Current Opinion in Plant Biology*. 14: 246-251.

- Kumar, G., Banu, G. S., Pappa, P. V., Sundararajan, M. and Pandian, M. R. (2004). Hepatoprotective activity of *Trianthema portulacastrum* L. against paracetamol and thioacetamide intoxication in albino rats. *Journal of Ethnopharmacology*. 92: 37-40.
- Kumar, K. A., Reddy, T. C., Reddy, G. V., Reddy, D., Mahipal, S. V. K., Sinha, S., Anil. N. Gaikwad. and Reddanna, P. (2011). High-throughput screening assays for cyclooxygenase-2 and 5-lipoxygenase, the targets for inflammatory disorders. *Indian Journal of Biochemistry and Biophysics*. 48: 256-61.
- Kumar, T. and Chandrashekar, K. S. (2011). *Bauhinia purpurea* Linn.: A review of its Ethnobotany, phytochemical and pharmacological profile. *Research Journal of Medicinal Plant*. 5: 420-31.
- Kumaraswamy, M. V. and Satish, S. (2008). Antioxidant and anti-lipoxygenase activity of *Thespesia lampas* Dalz & Gibbs. *Advances in Biological Research*. 2: 56-59.
- Kuo, Y.H., Yeh, M.H. and Huan, S.L. (1998). A novel 6- butyl-3-hydroxyflavanone from heartwood of *Bauhinia purpurea*. *Phytochemistry*. 49: 2529-2530.
- Kurt C. K. and Kathleen A. (2011). Chapter 12 : Biochemical and Metabolic Principles, Goldfrank's Toxicologic Emergencies. *Annals of Internal Medicine*.
- Lakshmi, B. V. S., Neelima, N., Kasthuri, N., Umarani, V. and Sudhakar, M. (2009). Protective effect of *Bauhinia purpurea* on gentamicin-induced nephrotoxicity in rats. *Indian Journal of Pharmaceutical Sciences*. 71: 551.
- Lavhale, M. S. and Mishra, S. H. (2007). Evaluation of free radical scavenging activity of *Butea monosperma* Lam. *Indian Journal of Experimental Biology*. 45: 376.
- Lee, W. M. (2004). Acetaminophen and the U.S. Acute Liver Failure Study Group: Lowering the risks of hepatic failure. *Hepatology*. 40: 6-9.
- Liang, Y. C., Tsai, S. H., Tsai, D. C., Lin-Shiau, S. Y. and Lin, J. K. (2001). Suppression of inducible cyclooxygenase and nitric oxide synthase through activation of peroxisome proliferator-activated receptor- γ by flavonoids in mouse macrophages. *FEBS Letters*. 496: 12-18.
- Lin, C. C., Hsu, Y. F. and Lin, T. C. (2000). Antioxidant and free radical scavenging effects of the tannins of *Terminalia catappa* L. *Anticancer Research*. 21: 237-243.
- Lin, S. C., Lin, C. H., Lin, C. C., Lin, Y. H., Chen, C. F., Chen, I. C. and Wang, L. Y. (2002). Hepatoprotective effects of *Arctium lappa* linne on liver injuries induced by chronic ethanol consumption and potentiated by carbon tetrachloride. *Journal of Biomedical Science*. 9: 401-409.
- Liver Center, Saint Louis University, <http://livercenter.slu.edu/index.php?page=liver-disease-facts> accessed on 29th December 2014

- Loganayaki, N., Suganya, N. and Manian, S. (2012). Evaluation of edible flowers of agathi (*Sesbania grandiflora* L. Fabaceae) for in vivo anti-inflammatory and analgesic, and in vitro antioxidant potential. *Food Science and Biotechnology*. 21: 509-517.
- López-Alarcón, C. and Denicola, A. (2013). Evaluating the antioxidant capacity of natural products: a review on chemical and cellular-based assays. *Analytica Chimica Acta*. 763: 1-10.
- Mahesh, B. U., Shrivastava, S., Pragada, R. R., Naidu, V. G. M. and Sistla, R. (2014). Antioxidant and hepatoprotective effects of *Boswellia ovalifoliolata* bark extracts. *Chinese Journal of Natural Medicines*. 12: 663-671.
- Malhi, H. and Gores, G. J. (2008). Cellular and molecular mechanisms of liver injury. *Gastroenterology*. 134: 1641-1654.
- Malik, A., Anis, I., Khan, S.B., Ahmed, E., Ahmed, Z., Nawaz, S.A. and Choudhary, M.I. (2004). Enzymes inhibiting lignans from *Vitex negundo*. *Chemical and Pharmaceutical Bulletin*. 52: 1269-1272.
- Mamat, S. S., Kamarolzaman, M. F., Yahya, F., Mahmood, N. D., Shahril, M. S., Jakius, K. F., Mohtarrudin, N. and Zakaria, Z. A. (2013). Methanol extract of *Melastoma malabathricum* leaves exerted antioxidant and liver protective activity in rats. *BMC Complementary and Alternative Medicine*. 13: 326.
- Manautou, J. E., Hart, S. G. E., Khairallah, E. A. and Cohen, S. D. (1996). Protection against acetaminophen hepatotoxicity by a single dose of clofibrate: effects on selective protein arylation and glutathione depletion. *Toxicological Sciences*. 29: 229-237.
- Marzilawati, A. R., Ngau, Y. Y. and Mahadeva, S. (2012). Low rates of hepatotoxicity among Asian patients with paracetamol overdose: a review of 1024 cases. *BMC Pharmacology and Toxicology*. 13: 8.
- Masih, N. G. and Singh, B. S. (2012). Phytochemical Screening of Some Plants Used in Herbal Based Cosmetic Preparations. *Chemistry of Phytopotentials: Health, Energy and Environmental Perspectives* Springer Berlin Heidelberg 111-112.
- Matsumura, H., Shimizu, Y., Ohsawa, Y., Kawahara, A., Uchiyama, Y. and Nagata, S. (2000). Necrotic death pathway in Fas receptor signaling. *The Journal of Cell Biology*. 151: 1247-1256.
- McChesney, J.D., Venkataraman, S.K. and Henri, J.T. (2007). Plant natural products: Back to the future or into extinction?. *Phytochemistry*. 68: 2015-2022.
- McEvoy, G.K. (2005). American Hospital Formulary Service- Drug Information 2005. Bethesda, MD: American Society of Health-System Pharmacists, Inc. 2005 (Plus Supplements) [Electronic version] 3564.
- Mehendale, H. M. (2000). PPAR- α : a key to the mechanism of hepatoprotection by clofibrate. *Toxicological Sciences*. 57: 187-190.

- Miesel, R. and Zuber, M. (1993). Elevated levels of xanthine oxidase in serum of patients with inflammatory and autoimmune rheumatic diseases. *Inflammation*. 17: 551-561.
- Mukherjee, M., Bhaskaran, N., Srinath, R., Shivaprasad, H.N., Allan, J.J., Shekhar, D. and Agarwal, A. (2010). Anti-ulcer and antioxidant activity of GutGard. *Indian Journal of Experimental Biology*. 48: 269–274.
- Mukherjee, P. K., Gopal, T. K. and Subburaju, T. (1998). Studies on the Anti-diarrheal Profiles of *Bauhinia purpurea* Linn Leaves (Fam. Caesalpiniaceae) Extract. *Natural Product Sciences*. 4: 234-237.
- Muralikrishna, K. S., Latha, K. P., Shreedhara, C. S., Vaidya, V. P. and Krupanidhi, A. M. (2008). Effect of *Bauhinia purpurea* Linn. on Alloxan-induced diabetic rats and isolated Frog's heart. *International Journal of Green Pharmacy*. 2: 83.
- Murugan, M. and Mohan, V. R. (2011). Evaluation of phytochemical analysis and antibacterial activity of *Bauhinia purpurea* L. and *Hiptage benghalensis* L.Kurz. *Journal of Applied Pharmaceutical Science*. 1: 157–160.
- Natori, S., Rust, C., Stadheim, L. M., Srinivasan, A., Burgart, L. J. and Gores, G. J. (2001). Hepatocyte apoptosis is a pathologic feature of human alcoholic hepatitis. *Journal of Hepatology*. 34: 248-253.
- Negi, B. S., Dave, B. P. and Agarwal, Y. K. (2012). Evaluation of antimicrobial activity of *Bauhinia purpurea* leaves under in vitro conditions. *Indian Journal of Microbiology*. 52: 360-365.
- Newman, D.J. and Cragg, G.M. (2012). Natural products as sources of new drugs over the 30 years from 1981 to 2010. *Journal of Natural Products*. 75: 311–335.
- Nijveldt, R. J., Van Nood, E., Van Hoorn, D. E., Boelens, P. G., Van Norren, K. and Van Leeuwen, P. A. (2001). Flavonoids: a review of probable mechanisms of action and potential applications. *The American Journal of Clinical Nutrition*. 74: 418-425.
- Niki, E., Yoshida, Y., Saito, Y. and Noguchi, N. (2005). Lipid peroxidation: mechanisms, inhibition, and biological effects. *Biochemical and Biophysical Research Communications*. 338: 668-676.
- Nithianantham, K., Ping, K. Y., Latha, L. Y., Jothy, S. L., Darah, I., Chen, Y., Chew, A.L. and Sasidharan, S. (2013). Evaluation of hepatoprotective effect of methanolic extract of *Clitoria ternatea* a (Linn.) flower against acetaminophen-induced liver damage. *Asian Pacific Journal of Tropical Medicine*. 3: 314-319
- Nithianantham, K., Shyamala, M., Chen, Y., Latha, L. Y., Jothy, S. L. and Sasidharan, S. (2011). Hepatoprotective potential of *Clitoria ternatea* Leaf extract against paracetamol induced damage in mice. *Molecules*. 16: 10134-10145.

- Nordin, N., Othman, S. N., and Che Mat, R. (2008). Technology implementation barriers in the Malaysian herbal industry: A case study. *Malaysian Management Journal*. 12:, 79-88.
- Noro, T., Miyase, T. and Kuroyanagi, M. (1983). Monoamine oxidase inhibitor from the rhizomes of *Kaempferia galanga* L. *Chemical and Pharmaceutical Bulletin*. 31: 2708–2711.
- Ogasawara, J., Watanabe-Fukunaga, R., Adachi, M., Matsuzawa, A., Kasugai, T., Kitamura, Y., Itoh, N., Suda, T. and Nagata, S. (1993). Lethal effect of the anti-Fas antibody in mice. *Nature*. 364: 806-809.
- Otsuka, H. (2005). Purification by solvent extraction using partition coefficient. *Natural Products Isolation Humana Press* 269-273.
- Panda, S. and Kar, A. (1999). *Withania somnifera* and *Bauhinia purpurea* in the regulation of circulating thyroid hormone concentrations in female mice. *Journal of Ethnopharmacology*. 67: 233–239.
- Patil, G. G., Mali, P. Y. and Bhadane, V. V. (2008). Folk remedies used against respiratory disorders in Jalgaon district, Maharashtra. *Natural Product Radiance*. 7: 354-358.
- Pawar, S. and Patil, D.A. (2007). Ethnomedicinal uses of barks in Jalgaon district. *Natural Product Radiance*. 6: 341-346.
- Pérez, M. B., Calderón, N. L. and Croci, C. A. (2007). Radiation-induced enhancement of antioxidant activity in extracts of rosemary *Rosmarinus officinalis* L. *Food Chemistry*. 104: 585-592.
- Pettit, G. R., Numata, A., Iwamoto, C., Usami, Y., Yamada, T., Ohishi, H. and Cragg, G. M. (2006). Antineoplastic Agents. 551. Isolation and Structures of Bauhiniastatins 1-4 from *Bauhinia purpurea*. *Journal of Natural Products*. 69:323-327.
- Phillipson, J. D. (2001). Phytochemistry and medicinal plants. *Phytochemistry*. 56: 237–243.
- Pithayanukul, P., Nithitanakool, S. and Bavovada, R. (2009). Hepatoprotective potential of extracts from seeds of *Areca catechu* and nutgalls of *Quercus infectoria*. *Molecules*. 14: 4987-5000.
- Porchezian, E. and Ansari, S. H. (2005). Hepatoprotective activity of *Abutilon indicum* on experimental liver damage in rats. *Phytomedicine*. 12: 62-64.
- Prior, R. L., Wu, X. and Schaich, K. (2005). Standardised Methods for the Determination of Antioxidant Capacity and Phenolics in Foods and Dietary Supplements. *Journal of Agricultural and Food Chemistry*. 53:4290–4302.
- Prior, R.L., Gu, L., Wu, X., Jacob, R.A., Sotoudeh, G., Kader, A.A. and Cook, R.A. (2007). Plasma antioxidant capacity changes following a meal as a measure of

- the ability of a food to alter *in vivo* antioxidant status. *The Journal of the American College of Nutrition*. 26: 170–181.
- Priya, V., Niveda S., Pratiksha G. and Gayathri R. (2010). A review of hepatoprotective natural products. *Recent Research in Science and Technology*. 2: 49-52.
- Qua, C. S., and Goh, K. L. (2011). Liver cirrhosis in Malaysia: peculiar epidemiology in a multiracial Asian country. *Journal of Gastroenterology and Hepatology*. 26: 1333-1337.
- Ragasa, C. Y., Hofileña, J. and Rideout, J. A. (2004). Secondary metabolite from *Bauhinia purpurea*. *Indian Journal of Microbiology*. 133: 1-5.
- Raj Kapoor, B., Venugopal, Y., Anbu, J., Harikrishnan, N., Gobinath, M. and Ravichandran, V. (2008). Protective effect of *Phyllanthus polyphyllus* on acetaminophen induced hepatotoxicity in rats. *Pakistan Journal of Pharmaceutical Sciences*. 21: 57-62.
- Raj Kapoor, B., Venugopal, Y., Anbu, J., Harikrishnan, N., Gobinath, M. and Ravichandran, V. (2008). Protective effect of *Phyllanthus polyphyllus* on acetaminophen induced hepatotoxicity in rats. *Pakistan Journal of Pharmaceutical Sciences*. 21: 57-62.
- Ramadori G. and Ramadori, P. (2010). Chapter 1; Hepatocytes. *Signaling Pathways in Liver Diseases*, Ed. J.-F. Dufour, P.-A. Clavien. Springer-Verlag Berlin Heidelberg.
- Ramadori, G., Rieder, H., Sipe, J., Shirahama, T. and Büschenfelde, K. H. (1989). Murine tissue macrophages synthesize and secrete amyloid proteins different to amyloid A (AA). *European Journal of Clinical Investigation*. 19: 316-322.
- Ramasamy, S., Wahab, N. A., Abidin, N. Z. and Manickam, S. (2011). Cytotoxicity evaluation of five selected Malaysian Phyllanthaceae species on various human cancer cell lines. *Journal of Medicinal Plants Research*. 5: 2267-2273.
- Ramchandra, R. and Joshi, B.C. (1967). Chemical examination of *Bauhinia purpurea* flower. *Current Science*. 36: 574-575.
- Ranawat, L., Bhatt, J. and Patel, J. (2010). Hepatoprotective activity of ethanolic extracts of bark of *Zanthoxylum armatum* DC in CCl₄ induced hepatic damage in rats. *Journal of Ethnopharmacology*. 127: 777-780.
- Rao Gururaj, A. and Balasubramaniam, N. A. (1999). Derivatives of *Bauhinia purpurea* lectin and their use as larvicides. United States Pioneer Hi-Bred International, Inc.(Des Moines, IA), 5945589.
- Ribeiro, P.S., Cortez-Pinto, H., Solá, S., Castro, R.E., Ramalho, R.M., Baptista, A., Moura, M.C., Camilo, M.E. and Rodrigues, C.M. (2004). Hepatocyte apoptosis, expression of death receptors, and activation of NF-kappaB in the liver of nonalcoholic and alcoholic steatohepatitis patients. *The American Journal of Gastroenterology*. 99:1708–1717

- Rolo, A.P. and Palmeira, C.M. (2006). Diabetes and mitochondrial function: role of hyperglycemia and oxidative stress. *Toxicology and Applied Pharmacology*. 212: 167–178.
- Rubinstein, D. (1962). Epinephrine release and liver glycogen levels after carbon tetrachloride administration. *American Journal of Physiology*. 203: 1033-1037.
- Salama, S. M., Abdulla, M. A., AlRashdi, A. S., Ismail, S., Alkiyumi, S. S. and Golbabapour, S. (2013). Hepatoprotective effect of ethanolic extract of *Curcuma longa* on thioacetamide induced liver cirrhosis in rats. *BMC Complementary and Alternative Medicine*. 13: 56.
- Salatino, A., Blatt, C. T., Santos, D. Y. D. and Vaz, A. M. (1999). Foliar flavonoids of nine species of *Bauhinia*. *Brazilian Journal of Botany*. 22: 17-20.
- Sambasivam, H., Rassouli, M., Murray, R. K., Nagpurkar, A., Mookerjee, S., Azadi, P., Dell, A. and Morris, H. R. (1993). Studies on the carbohydrate moiety and on the biosynthesis of rat C-reactive protein. *Journal of Biological Chemistry*. 268: 10007-10016.
- Sandhar, H. K., Kumar, B., Prasher, S., Tiwari, P., Salhan, M. and Sharma, P. (2011). A review of phytochemistry and pharmacology of flavonoids. *Internationale Pharmaceutica Scientia*. 1: 25-41.
- Schmidt, E., and Schmidt, F. W. (1990). Progress in the enzyme diagnosis of liver disease: reality or illusion?. *Clinical Biochemistry*. 23: 375-382.
- Shahidi, F. and Marian, N. (2003). Phenolics in Food and Nutraceuticals, vol. 1. CRS Press LLC, Boca Raton, FL 144–150.
- Shanani S (1999). Evaluation of hepatoprotective efficacy of APCL-A polyherbal formulation *in vivo* in rats. *Indian Drugs*. 36: 628-631.
- Sheikh, N., Tron, K., Dudas, J. and Ramadori, G. (2006). Cytokine-induced neutrophil chemoattractant-1 is released by the noninjured liver in a rat acute-phase model. *Laboratory Investigation*. 86: 800-814.
- Shiddamallayya, N., Yasmeen, A. and Gopakumar, K. (2010). Medico-botanical survey of Kumar parvatha Kukke Subramanya, Mangalore, Karnataka. *Indian Journal of Traditional Knowledge*. 9:96-99.
- Shimada, Y., Kato, T., Ogami, K., Horie, K., Kokubo, A., Kudo, Y., Maeda, E., Sohma, Y., Akahori, H. and Kawamura, K. (1995). Production of thrombopoietin (TPO) by rat hepatocytes and hepatoma cell lines. *Experimental Hematology*. 23: 1388-1396.
- Shnoy, A.K., Somayaji, S.N. and Bairy, K.L. (2002). Hepatoprotective activity of ethanolic extract of *Ginkgo biloba* against carbon tetrachloride induced hepatic injury in rat. *Indian Journal Pharmacology*. 46:167-174.
- Shreedhara, C. S., Vaidya, V. P., Vagdevi, H. M., Latha, K. P., Muralikrishna, K. S. and Krupanidhi, A. M. (2009). Screening of *Bauhinia purpurea* Linn. for

- analgesic and anti-inflammatory activities. *Indian Journal of Pharmacology*. 41: 75.
- Singleton, V. L. and Rossi J. A. Jr. (1965). Colorimetry of total phenolics with phosphomolybdicphosphotungstic acid reagents. *American Journal of Enology and Viticulture*. 16: 144–158.
- Snyder, L. R., Kirkland, J. J., & Glajch, J. L. (2012). *Practical HPLC method development*. John Wiley & Sons.
- Somchit, M. N., Zuraini, A., Bustamam, A. A., Somchit, N., Sulaiman, M. R. and Noratunlina, R. (2005). Protective activity of turmeric (*Curcuma longa*) in paracetamol-induced hepatotoxicity in rats. *International Journal of Pharmacology*. 1: 252–256
- Soobrattee, M.A., Neergheen, V.S., Luximon-Ramma, A., Aruoma, O.I. and Bahorun, T. (2005). Phenolics as potential antioxidant therapeutic agents: mechanism and actions. *Mutation Research*. 579: 200–213.
- Sowndhararajan, K. and Kang, S. C. (2013). Protective effect of ethyl acetate fraction of *Acacia ferruginea* DC. Against ethanol-induced gastric ulcer in rats. *Journal of Ethnopharmacology*. 148: 175-181.
- Stickel, F. and Schuppan, D. (2007). Herbal medicine in the treatment of liver diseases. *Digestive and Liver Disease*. 39: 293-304
- Tanna, A., Nair, R. and Chanda, S. (2009). Assessment of anti-inflammatory and hepatoprotective potency of *Polyalthia longifolia* var. pendula leaf in Wistar albino rats. *Journal of Natural Medicines* . 63: 80-85.
- Tapas, A. R., Sakarkar, D. M. and Kakde, R. B. (2008). Flavonoids as nutraceuticals: a review. *Tropical Journal of Pharmaceutical Research*. 7: 1089-1099.
- Tolonen, A. (2003). *Analysis of Secondary Metabolites in Plant and Cell Culture Tissue of Hypericum Perforatum L. and Rhodiola Rosea L.* Oulun yliopisto.
- Tripathi, K.D. (2001). *Essentials of Medical Pharmacology*, 4th ed. Jaypee Brothers Medical Publishers, New Delhi, pp. 52–53.
- Tsimogiannis, D., Samiotaki, M., Panayotou, G. and Oreopoulou, V. (2007). Characterization of flavonoid subgroups and hydroxy substitution by HPLC-MS/MS. *Molecules*. 12: 593–606.
- U.S National Library of Medicine, <http://toxnet.nlm.nih.gov/cgi-bin/sis/search2/r?db=hsdb:@term+@rn+616-91-1> accessed on 28th December 2014
- Uma Jr, N., Fakurazi, S. and Hairuszah, I. (2010). *Moringa oleifera* enhances liver antioxidant status via elevation of antioxidant enzymes activity and counteracts paracetamol-induced hepatotoxicity. *Malaysian Journal of Nutrition*. 16: 293-307

- Vilgrain, V., Ronot, M., Abdel-Rehim, M., Zappa, M., d'Assignies, G., Bruno, O. and Vullierme, M. P. (2013). Hepatic steatosis: A major trap in liver imaging. *Diagnostic and Interventional Imaging*. 94: 713-727.
- Vimala, S., Rohana, S., Rashih, A. A. and Juliza, M. (2012). Antioxidant Evaluation in Malaysian Medicinal Plant: *Persicaria minor* (Huds.) Leaf. *Science Journal of Medicine and Clinical Trials*, 2012.
- Vishnu Priya. V., Niveda ,S., Pratiksha, G. and Gayathri, R. (2010). A Review of Hepatoprotective Natural Products. *Recent Research in Science and Technology*. 2: 49-52
- Walgren, J.L., Mitchell, M.D. and Thompson, D.C. (2005). Role of metabolism in drug-induced idiosyncratic hepatotoxicity. *Critical Reviews in Toxicology*. 35: 325-61.
- Walter F. B. and Emile L. B. (2011). Functional Anatomy of the Liver and Biliary Tree. *Medical Physiology*. Second Edition.
- Watz, B. (2008). Anti-inflammatory effects of plant-based foods and of their constituents. *International Journal for Vitamin and Nutrition Research*. 78: 293-298.
- Weber, L. W., Boll, M. and Stampfl, A. (2003). Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model. *CRC Critical Reviews in Toxicology*. 33:105-136.
- World Health Organization. (1993). *Research guidelines for evaluating the safety and efficacy of herbal medicines*. World Health Organization, Regional Office for the Western Pacific.
- World Health Organization. (2002) Traditional medicine strategy launched. *Bulletin of the World Health Organization* 80: 7
- Yadav, R.N. and P. Tripathi. (2000). A novel flavone glycoside from the stem of *Bauhinia purpurea*. *Fitoterapia*. 71: 88-90.
- Yadav, S. and Bhadoria, B. K. (2005). Two dimeric flavonoids from *Bauhinia purpurea*. *Indian Journal of Chemistry B*. 44: 2604-2607.
- Yanpallewar, S. U., Sen, S., Tapas, S., Kumar, M., Raju, S. S. and Acharya, S. B. (2003). Effect of *Azadirachta indica* on paracetamol-induced hepatic damage in albino rats. *Phytomedicine*. 10: 391-396.
- Zabidi, Z., Wan Zainulddin, W. N., Mamat, S. S., Shamsah Din, S., Kamisan, F. H., Yahya, F., Ismail, N.A. and Zakaria, Z. A. (2012). Antiulcer activity of methanol extract of *Melastoma malabathricum* leaves in rats. *Medical Principles and Practice*. 21: 501-503.
- Zakaria, Z. A., Abdul Hisam, E. E., Norhafizah, M., Rofiee, M. S., Othman, F., Hasiah, A. H. and Vasudevan, M. (2012). Methanol extract of *Bauhinia*

purpurea leaf possesses Anti-Ulcer activity. *Medical Principles and Practice*. 21:476-482.

Zakaria, Z. A., Abdul Hisam, E. E., Rofiee, M. S., Norhafizah, M., Somchit, M. N., Teh, L. K. and Salleh, M. Z. (2011b). *In vivo* antiulcer activity of the aqueous extract of *Bauhinia purpurea* leaf. *Journal of Ethnopharmacology*. 137: 1047-1054.

Zakaria, Z. A., Rahman, N. A., Loo, Y. W., Ayub, A. A., Sulaiman, M. R., Jais, A. M., Gopalan, H.K. and Fatimah, C. A. (2009). Antinociceptive and anti-inflammatory activities of the chloroform extract of *Bauhinia purpurea* L.(Leguminosae) leaves in animal models. *International Journal of Tropical Medicine*. 4: 140-145.

Zakaria, Z. A., Rofiee, M. S., Teh, L. K., Salleh, M. Z., Sulaiman, M. R. and Somchit, M. N. (2011a). *Bauhinia purpurea* leaves' extracts exhibited in vitro antiproliferative and antioxidant activities. *African journal of Biotechnology*. 10: 65-74.

Zakaria, Z. A., Wen, L. Y., Abdul Rahman, N. I., Abdul Ayub, A. H., Sulaiman, M. R. and Gopalan, H. K. (2007). Antinociceptive, anti-inflammatory and antipyretic properties of the aqueous extract of *Bauhinia purpurea* leaves in experimental animals. *Medical Principles and Practice*. 16: 443-449.