



UNIVERSITI PUTRA MALAYSIA

**MOLECULAR EPIDEMIOLOGY AND ANTIMICROBIAL RESISTANCE
PROFILE OF ESBL-PRODUCING *Escherichia coli* ISOLATED AT
POULTRY FARMS AND POULTRY MEAT WET-MARKETS
IN SELANGOR, MALAYSIA**

ALIYU ABDULRASHEED BELLO

FPV 2016 39



**MOLECULAR EPIDEMIOLOGY AND ANTIMICROBIAL RESISTANCE
PROFILE OF ESBL-PRODUCING *Escherichia coli* ISOLATED AT
POULTRY FARMS AND POULTRY MEAT WET-MARKETS
IN SELANGOR, MALAYSIA**

By

ALIYU ABDURASHEED BELLO



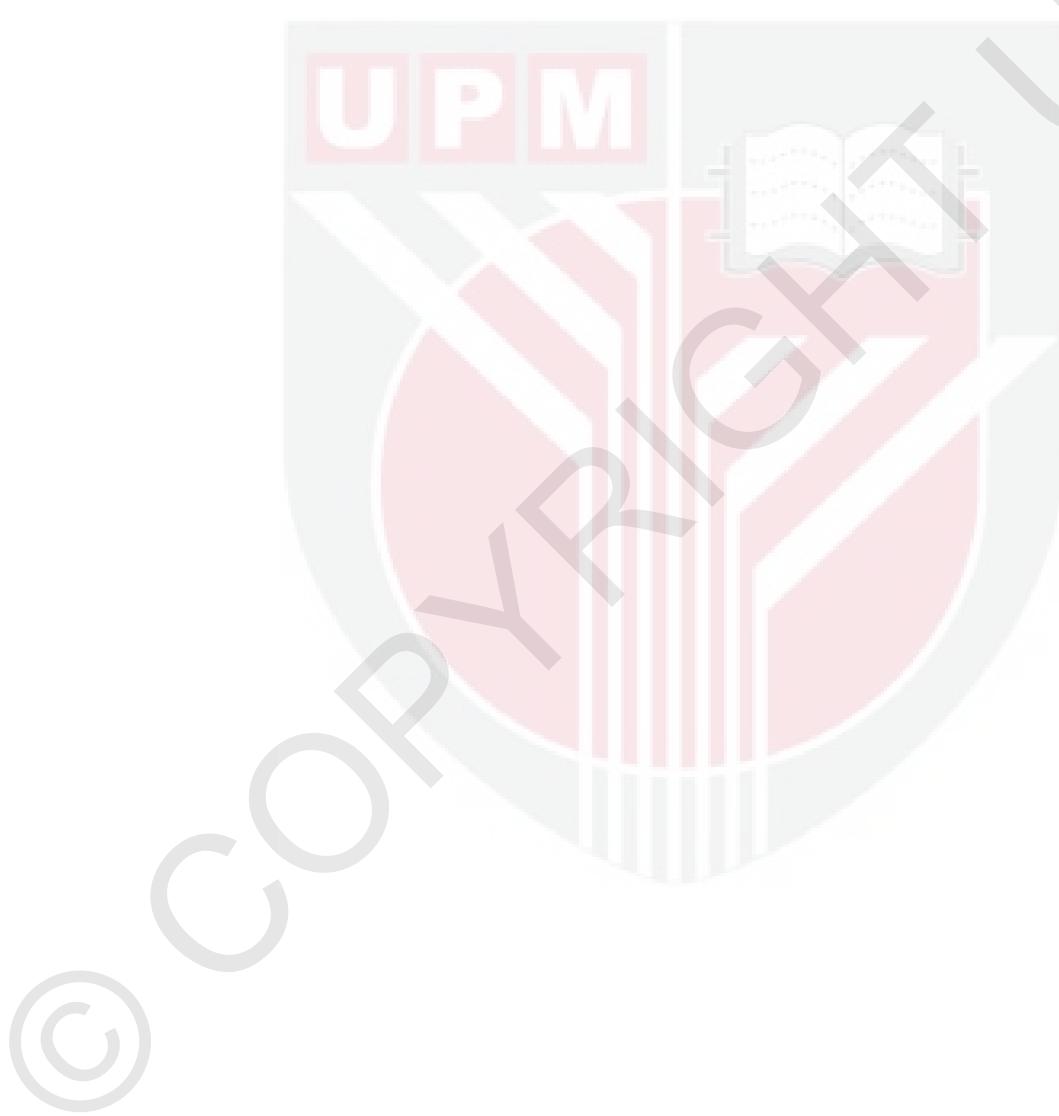
**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

September 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This work is dedicated to:

My late grandparents

Alh. Abubakar Sidiku Tunga (Mutawallen Gwandu)

Haj. Gado (Fadimatu)

Haj. Yaya (Rakiya)

My parents

Late Engr. Alh. Bello Aliyu

Haj. Aisha Abubakar Sidiku Tunga

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment
of the requirement for the Degree of Doctor of Philosophy

**MOLECULAR EPIDEMIOLOGY AND ANTIMICROBIAL RESISTANCE
PROFILE OF ESBL-PRODUCING *Escherichia coli* ISOLATED AT
POULTRY FARMS AND POULTRY MEAT WET-MARKETS
IN SELANGOR, MALAYSIA**

By

ALIYU ABDULRASHEED BELLO

September 2016

Chairman : Professor Saleha Abdul Aziz, PhD
Faculty : Veterinary Medicine

Extended spectrum β -lactamase-producing- *Escherichia coli* (ESBL-EC) are emerging multidrug resistant zoonotic bacteria that threaten animal production, food safety and global antimicrobial therapy. It confers resistance to first line and most widely used antibiotic in both human and veterinary chemotherapeutics. ESBL-EC is a major public health problem challenging worldwide health care facilities including Malaysia. Poultry has been suggested to serve as a major pathway for human exposure to ESBL-EC. To date, there has been little to no research on the occurrence of ESBL-EC within the Malaysian poultry food chain. Hence, the objectives of the study were to determine the occurrence and distribution of ESBL-EC in broiler chickens and chicken meat, to assess their antimicrobial resistance profiles, to identify the potential risk factors associated with the occurrence of ESBL-EC, and to conduct molecular characterization and phylogenetic analysis on the ESBL-EC isolates. A cross-sectional study was conducted with 640 samples collected to determine the presence of ESBL-EC from broiler poultry farms (400 samples) and wet-markets (240 samples) in Selangor, Malaysia using culture and disk combination methods and polymerase chain reaction assays. The overall study was carried out between July-2012 to February 2015.

The findings demonstrated the wide distribution of ESBL-EC across the eight district areas of Selangor, Malaysia. At poultry farms, the overall proportion of ESBL-EC occurrence was 37.2%. There were significant differences in the proportions of ESBL-EC that occurred among poultry farms as well as within the type of study samples. Of the eight farms, Farm A (Gombak), Farm B (Klang) and Farm C (Hulu Selangor), had the highest occurrence rate at 62%, 50%, and 50% respectively, followed by Farm D (Petaling) 38%, Farm E (Sepang) 34%, Farm F (Kuala Langat) 26% and Farm G (Kuala Selangor) 24%, and the lowest was in Farm H (Hulu Langat) 14%. Among the study samples, chicken had the highest occurrence rate at 45.4%, followed by the chicken house floor at 40%, and flies 30%, while feed and water samples were at

17.5%, and 12.5% respectively. Two hundred and forty samples (240) were collected from 40 broiler poultry stalls at wet-markets, with five stalls representing each district area. The overall prevalence was 48.8%. Among the district areas, Hulu Selangor had the highest occurrence rate at 66.7%. Moderate proportion was seen in Hulu Langat 56.7%, and Kuala Selangor 50%. The lowest proportions were found in Klang 46.7%, Sepang 46.7%, Petaling 43.3%, Gombak 40%, and Kuala Langat 40%.

To determine possible factors associated with the presence of ESBL-EC at poultry farms and poultry meat wet-markets, a questionnaire was used as an instrument to obtain information and the data were analyzed using Chi-square test and logistic regression analysis at p-value of less than 0.05. At poultry farms, ten factors were found to be significantly associated in the occurrence of ESBL-EC, which were broadly classified into farm management, biosecurity, and medical history such as antibiotic usage. At the poultry meat wet-markets, only four factors had the significant association with ESBL-EC contamination, which included stall sanitation, type of counter top, source of cleaning water, and type of cutting board.

The antimicrobial resistant profiles of ESBL-EC isolated from poultry farm and wet-markets were determined using the standard disc diffusion method, according to CLSI guidelines. The ESBL-EC isolates were screened for susceptibility against 11 panels of antibiotics and 98% of the isolates showed resistance to tetracycline, 91.9% to cefotaxime, 85.2% to sulfamethoxazole/trimethoprim, 78.5% to nalidixic acid, 73.8% to chloramphenicol, 65.1% to cefpodoxime, 64.4% to ciprofloxacin, 61.1% to ceftriaxone, 59.7% to gentamicin, 53.7% to ceftazidime, and the least resistance was to aztreonam at 30.2%. At the wet-markets, ESBL-EC isolates were also tested against the same panels of 11 antibiotics and 99.1% showed resistance to tetracycline, 94.9% to cefotaxime, 93.2% to nalidixic acid, 82.1% to sulfamethoxazole/trimethoprim, 80.3% to chloramphenicol, 78.6% to ciprofloxacin, 59.8% to cefpodoxime, 41.0% to ceftriaxone, 23.9% to gentamicin, and the least resistance was to aztreonam at 21.4%.

For the molecular characterization and phylogenetic analysis of ESBL-EC, representative isolates from the poultry farms and wet-markets, pulsed-field gel electrophoresis (PFGE) were used. To analyse the PFGE results, 85% similarity was used as the study cut-off point. Although there is wide genomic diversity between, within and across the poultry farms and poultry meat wet- markets isolates. However, the PFGE molecular epidemiological results had evidently demonstrated genomic relationship within the selected study isolates. At the poultry farms, PFGE molecular fingerprinting revealed four major phylogroupings, with 16 isolates demonstrating genomic relationship that ranges 100% to 85% genetic similarity, between and within the poultry farms. Similarly, at the poultry meat wet-markets, four major phylogroups were detected, with 11 isolates demonstrating genomic relationship that ranges from 100% to 86% genetic similarity, between and within the poultry farms. Phylogenetic analysis for the combination of poultry farms and poultry meat wet-markets isolates revealed six major phylogroups, with eight isolates demonstrating genomic relationship across the poultry farms and poultry meat wet-markets, which range from 92% - 85% genetic similarity.

In conclusion, the present study indicated the high occurrence and wide dissemination of multidrug resistant ESBL-EC at the poultry farms and wet-markets. The ESBL-EC occurrence was associated with several factors that can be broadly classified under

imprudent use of antibiotics, poor husbandry, management and biosecurity practices at the farms; while the poultry meat wet-market factors included poor personal hygiene and sanitary practices, poor environmental sanitation and lack of good manufacturing practices. Thus, the findings of the study can assist to serve as a useful guide to Veterinary/Public Health authority in designing evidence-based mitigation strategies for effective control of ESBL-EC at poultry farms and poultry meat wet-markets.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**EPIDEMIOLOGI MOLEKUL DAN PROFIL RINTANGAN
ANTIMIKROBIA ESBL-*Escherichia coli* TERASING DI LADANG
AYAM DAN PASAR BASAH DAGING AYAM DI NEGERI
SELANGOR, MALAYSIA**

Oleh

ALIYU ABDULRASHEED BELLO

September 2016

Pengerusi : Profesor Saleha Abdul Aziz, PhD
Fakulti : Perubatan Veterinar

Spektrum terluas pengeluaran β -laktamase- *Escherichia coli* (ESBL-EC) merupakan bakteria zoonotik rintangan berbilang ubatan yang muncul yang mengancam pengeluaran haiwan, keselamatan makanan, dan terapi antimikrobia global. Ubatan tersebut memberikan rintangan kepada antibiotik barisan pertama dan antibiotik yang paling banyak digunakan dalam kemoterapi manusia dan veterinar. ESBL-EC merupakan masalah utama kesihatan awam yang mencabar kemudahan penjagaan kesihatan sejagat, termasuk Malaysia. Poltri merupakan laluan utama bagi pendedahan manusia kepada ESBL-EC. Sehingga kini, kurang terdapat atau boleh dikatakan tiada penyelidikan mengenai kehadiran ESBL-EC dalam lingkungan jaringan makanan poltri terutama ayam di Malaysia. Oleh sebab itu, objektif kajian ini adalah untuk menentukan kadar kehadiran ESBL-EC pada ayam dan daging ayam, untuk menilai profil rintangan antimikrobia, mengenalpasti faktor risiko potensi yang berkaitan dengan kehadiran ESBL-EC, dan untuk menjalankan pencirian molekul serta analisis filogenetik terhadap isolat ESBL-EC. Kajian keratanrentas telah dilaksanakan ke atas 640 sampel yang dikumpul untuk menentukan kehadiran ESBL-EC di ladang ayam pedaging (400 sampel) dan daging ayam di pasar basah (240 sampel) di Selangor, Malaysia dengan menggunakan kaedah kombinasi kultur dan disk dan assai reaksi jarring polymerase (PCR). Keseluruhan kajian telah dijalankan antara Julai 2012 hingga Februari 2015.

Hasil kajian menunjukkan bahawa sebaran ESBL-EC yang meluas merentasi lapan kawasan daerah di Selangor, Malaysia. Di ladang ayam, keseluruhan kadar kehadiran ESBL-EC ialah 37.2%. Terdapat perbezaan yang signifikan daripada segi kadar ESBL-EC yang hadir dalam kalangan ladang ayam dan dalam lingkungan jenis sampel kajian. Antara lapan ladang, Ladang A (Gombak), Ladang B (Klang) dan Ladang C (Hulu Selangor), mempunyai kehadiran yang paling tinggi, iaitu masing-masing 62%, 50% dan 50%, diikuti oleh Ladang D (Petaling) 38%, Ladang E (Sepang) 34%, Ladang F (Kuala Langat) 26% dan Ladang G (Kuala Selangor) 24%, dan yang paling rendah ialah Ladang H (Hulu Langat) 14%. Antara sampel kajian, ayam mempunyai

kadar kehadiran paling tinggi, iaitu 45.4%, diikuti oleh lantai rumah ayam iaitu 40%, dan lalat 30%, manakala sampel makanan dan air adalah masing-masing 17.5%, dan 12.5%. Dua ratus dan empat puluh sampel (240) telah dikumpulkan daripada 40 gerai daging ayam di pasar basah, dengan lima gerai mewakili setiap kawasan daerah. Keseluruhan kehadiran ialah 48.8%. Antara kawasan daerah, Hulu Selangor mempunyai kadar yang paling tinggi, iaitu 66.7%. Kadar yang sederhana didapati di daerah Hulu Langat, iaitu 56.7%, dan Kuala Selangor 50%. Kadar yang paling rendah ialah daerah Klang 46.7%, Sepang 46.7%, Petaling 43.3%, Gombak 40%, dan Kuala Langat 40%.

Bagi menentukan faktor risiko yang mungkin berkaitan dengan kewujudan ESBL-EC di ladang ayam dan pasar basah daging ayam, soalselidik telah digunakan sebagai instrument untuk memperoleh maklumat dan data telah dianalisis menggunakan ujian khi kuasa dua (*chi-square test*) dan analisis regresi logistik pada nilai $p < 0.05$. Di ladang ayam, sepuluh faktor risiko telah dikenalpasti secara signifikan yang berkaitan dengan kehadiran ESBL-EC, yang telah diklasifikasikan secara kasar kepada pengurusan ladang, biokeselamatan, dan sejarah perubatan seperti penggunaan antibiotic. Di pasar basah daging ayam, hanya empat faktor mempunyai kemungkinan secara signifikan sebagai penyebab kontaminasi dengan ESBL-EC, termasuk sanitasi gerai, jenis permukaan kaunter, sumber air pembersihan, dan jenis papan memotong daging.

Profil rintangan antimikrobia isolat ESBL-EC daripada ladang ayam dan pasar basah telah ditentukan menggunakan kaedah difusi disk standard, berdasarkan garispanduan CLSI. Isolat ESBL-EC telah uji kepekaan terhadap 11 panel antibiotik dan 98% isolat daripada ladang ayam menunjukkan rintangan terhadap *tetracycline*, 91.9% *cefotaxime*, 85.2% *sulfamethoxazole/trimethoprim*, 78.5% *nalidixic acid*, 73.8% *chloramphenicol*, 65.1% *cefpodoxime*, 64.4% *ciprofloxacin*, 61.1% *ceftriaxone*, 59.7% *gentamicin*, 53.7% *ceftazidime*, dan rintangan paling rendah adalah terhadap *aztreonam*, 30.2%. Isolat ESBL-EC daripada pasar basah menunjukkan rintangan antibiotik seperti berikut: *tetracycline* 94.9%, *cefotaxime* 93.2%, *nalidixic acid* 82.1%, *sulfamethoxazole/trimethoprim* 80.3%, *chloramphenicol* 78.6%, *ciprofloxacin* 59.8%, *cefpodoxime* 41.0%, *ceftriaxone* 23.9% *gentamicin* dan rintangan paling rendah adalah terhadap *aztreonam* 21.4%.

Bagi pencirian molekul dan analisis filogenik isolat ESBL-EC, isolat daripada ladang ayam dan pasar basah, elektrophoresis gel medan denyut (PFGE) telah digunakan. Bagi menganalisis keputusan PFGE, 85% persamaan telah digunakan sebagai titik pemisah. Terdapat kepelbagaiannya genomik yang merentasi isolat ladang ayam dan pasar basah daging ayam. Walaubagai mana pun, keputusan epidemiologi molekul PFGE telah terbukti menunjukkan hubungan genomik dalam lingkungan isolat terpilih yang dikaji. Di ladang ayam, pencapjarian molekul PFGE memperlihatkan empat filokumpulan utama, dengan 16 isolat mempamerkan hubungan genomik yang berjulat 100% hingga 85% persamaan genetik, antara dan dalam ladang ayam. Begitu juga di pasar basah daging ayam, empat filokumpulan utama telah dikesan, dengan 11 isolat mempamerkan hubungan genomik yang berjulat daripada 100% hingga 86% persamaan genetik, antara dan dalam pasar basah. Analisis filogenetik bagikombinasi isolat ladang ayam dan pasar basah daging ayam menunjukkan enam filokumpulan utama, dengan lapan isolat mempamerkan hubungan genomic merentasi ladang ayam dan pasar basah daging ayam, dengan persamaan genetik berkisar 92% - 85%.

Sebagai kesimpulan, kajian ini memperlihatkan kehadiran ESBL-EC yang tinggi dan sebaran ESBL-EC rintangan berbilang ubatan (MDR) yang luas di ladang ayam dan di pasar basah. Kehadiran ESBL-EC telah dikaitkan dengan pelbagai faktor risiko yang secara kasar dapat diklasifikasikan sebagai penggunaan antibiotik yang tak berhemah, amalan penternakan, amalan pengurusan dan amalan biokeselamatan yang lemah di ladang ayam; mana kala faktor di pasar basah daging ayam pula adalah termasuk amalan kebersihan yang lemah dan juga sanitasi persekitaran yang kurang. Oleh sebab itu, daripada hasil kajian ini diharapkan dapat membantu pihak berkuasa Kesihatan Awam / Veterinar dalam merekabentuk strategi mitigasi kawalan ESBL-EC yang berkesan di ladang ayam dan di pasar basah daging ayam.



ACKNOWLEDGEMENTS

All praise is due to Allah with whose favours all good can be accomplished. I wish to express my profound sincere gratitude to Almighty Allah for giving me these great opportunity and wisdom in the successful completion of my thesis. I am most grateful to the Chairperson of my PhD supervisory committee, Prof Dr. Saleha Abdul Aziz for her continuous coaching and support throughout period of my study. She indeed endured the pains of reading, re-reading and revising to improve the quality of work, with insightful comments, constructive criticisms, and professorial guidance, may Allah continue to give her more strength and wisdom for the benefit of entire humanity.

I am equally grateful to my co-supervisors, namely Associate Professor Dr. Zunita Zakaria, and Associate Professor Dr. Jalila Abu for their invaluable guidance, advice, encouragement through the course of the study and in preparation of this thesis. Without their patience, support and contribution, I would not have accomplished the goal of finishing this thesis. Special appreciation also goes to Prof. Kwai Lin Thong, her research assistance Wing Sze and students of Laboratory of Biomedical Science and Molecular Microbiology, UMBIO Research Cluster, University of Malaya, Kuala Lumpur for providing me the guidance materials and workplace to accomplish my pulsed-field gel electrophoresis (PFGE) work, their mentorship will never be forgotten. I am especially grateful to Puan Fauziah Nordin, staff of Veterinary Public Health Lab and those of Bacteriology Lab especially Azri, Krish, Hajar, Haj Razak and Hafiz for their cooperation and unreserved assistances throughout my laboratory work. and all of my lab mates, Soe Soe, Atta, Teguh, Goni, Abdelrahman, Mohd Yousif, Jalo, Shah, Wint, and Emelia, for their generosity and kind assistance toward successful accomplishment my work.

A special gratitude also goes to all the staff and management of the Ministry of Animal Health, Husbandry and Fishery, Birnin-kebbi, Kebbi State, Nigeria especially Dr. Usman D/Gari, Dr. Bala Kakale, Dr. Ambursa, Dr. Alheri, Dr. Dominic, Dr. Ali, Dr. Gwandu, Dr. Maryam, D. Idris, Mal. Bashir, Mal. Ibrahim, Garus and Garus, for their love, care and support.

To my colleagues especially Dr. Kabiru Goje, Dr. Mukhtar Salihu Anka, Dr. Kabir Sahabi Kalgo, Mal. Faruk ambursa, Mal. Jaafar Bunza, Dr. Hassan Ismail Musa, Dr. Adamu Yarima, Dr. Khumran, Dr. Musa (Alawu), Dr. Kani, Dr. Ado Makama, Dr. Aminu Ishak, Dr. Usman Gusau, Dr. Kaikabo, late Dr. Shafiu, Dr. Babane, Ahmad Nasiha, Basheer Somali for their guidance from the inception to the completion of this study, thank you. They have been instrumental in helping me shape my thesis and providing me with insight to the joys (and stress) of academic research. My thanks are due to my friends and mentors Ibrahim Bagudo, Mal. Aliyu (Sufi), Mal. Faruk (Muftee), Mal. Usman (Sheikhy), Sheikh Sadiq, Ali Abbas, Brrs. Aminu Shamaki, Mal. Shuaibu, Mal. Nura, Mal. Hassan, Mal. Adam Otono, Abdulkareem kalgo for their moral support, dua and encouragement during my study. Special thanks also go to my relatives Alh. Bawa Tunga, Kw. Alu AT, Kw. Sule, Kw. Danbawa, Kw. Adamu Tune Waziri, Haj. Mariya, Haj. Amo, Mm. Mabera, Haj. Yarbaba, Mm. Jamila, Mm. Ladi, Mm. Fadimatu, Mm. Aminatu, Mm. Maimuna, Mm. Lantana, Mm. Azumi Bb

Gusau, Kabiru I/kauna, Nura Galadanchi and Uncle Zee, May Allah bless all of you for the motivation and Dua.

I am especially gratitude to my parents specifically my mother Hajiya Aisha Sidiku Tunga and my late Father Engr. Alh. Bello Aliyu for giving me guidance, support and prayers. My heartfelt appreciation goes to my wife Khadeejatul-Kubrah, whose dedication, love and persistent confidence in me has taken the load off my shoulder. To my son Muh'd Bello, you are the source of strength and perseverance. To my siblings Hj. Rabi, Hj. Balki, Hj. Amina, Aunt. Talatu, Late Ahmad, Zulai, Jimmai, Zainab, Bushrah (Malama), BB, Aminu, Ibrahim, Asiyah, Mayam, Abu, Abubakr, Fatima and Ismael. It was you who motivated me when times were difficult and I felt like there was no light at the end of the tunnel.

Finally, I would like to thank every single person who in one way or the other has help towards successful realization of this great achievement of mine, whom I couldn't mention due to space limit, the forgetfulness of my human nature, I am unable to mention their names in persons. I sincerely apologized for my inadequacy as a human being. May God bless all of you and your contribution truly appreciated and will never be forgotten.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Saleha Abdul Aziz, PhD
Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Zunita Zakaria, PhD
Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Jalila Abu, PhD
Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD
Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No.: Aliyu Abdulrasheed Bello / GS27295

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature:

Name of Chairman
of Supervisory
Committee:

Professor Dr. Saleha Abdul Aziz

Signature:

Name of Member
of Supervisory
Committee:

Associate Professor Dr. Zunita Zakaria

Signature:

Name of Member
of Supervisory
Committee:

Associate Professor Dr. Jalila Abu

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	ix
DECLARATION	xi
LIST OF TABLES	xvi
LIST OF FIGURES	xviii
LIST OF ABBREVIATIONS	xx
 CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	4
2.1 Escherichia coli	4
2.2 β-lactam Antibiotics	5
2.2.1 Introduction	5
2.2.2 Mechanism of action	7
2.2.3 Clinical significance in human and veterinary medicine	7
2.2.4 Resistance to β-lactams	8
2.3 Extended-spectrum β-lactamases	12
2.3.1 Introduction and Evolution	12
2.3.2 Types of Extended-spectrum β-lactamases	13
2.4 Global distribution of ESBL-EC	18
2.5 Factors influencing occurrence of ESBL-EC in humans	23
2.6 Factors influencing occurrence of ESBL-EC in poultry farms and retail poultry meat markets	24
2.7 Method of ESBL-EC detection	24
2.7.1 Phenotypic methods	24
2.7.2 Molecular detection method	26
2.8 Antimicrobial resistance in ESBL-EC	27
2.9 Public health and economic implication of ESBL-EC in animals and humans	27
2.9.1 Animals	27
2.9.2 Humans	27
2.10 Control and Prevention of ESBL-EC	28
3 OCCURRENCE OF ESBL-EC AT POULTRY FARMS AND POULTRY MEAT MARKETS	30
3.1 Introduction	30
3.2 Materials and methods	31
3.2.1 Study areas	31
3.2.2 Study design and sampling technique	33
3.2.3 Samples collection and preparation	34

3.2.4	Identification of <i>E. coli</i>	35
3.2.5	Screening and phenotypic confirmation of ESBL-EC	35
3.2.6	Extraction of genomic DNA and Multiplex-PCR assay for <i>E. coli</i> , TEM, SHV, CTX-M, and OXA	36
3.3	Data analysis	37
3.4	Results	38
3.4.1	Poultry farms	38
3.4.2	Poultry meat wet-markets	43
3.5	Discussion	47
3.6	Conclusion	50
4	FACTORS ASSOCIATED WITH OCCURRENCE OF ESBL-EC AT POULTRY FARMS AND POULTRY MEAT WET-MARKETS	51
4.1	Introduction	51
4.2	Materials and methods	51
4.2.1	Data collection	51
4.2.2	Study instrument validation	52
4.2.3	Definition of terms	52
4.3	Data analysis	54
4.4	Results	54
4.4.1	Poultry farms	54
4.4.2	Poultry meat wet-markets	59
4.5	Discussion	62
4.6	Conclusion	67
5	ANTIBIOTIC RESISTANCE PROFILE OF ESBL-EC ISOLATES FROM POULTRY FARMS AND POULTRY MEAT MARKETS	68
5.1	Introduction	68
5.2	Materials and methods	69
5.2.1	Antibiotic susceptibility test (Disc diffusion method)	69
5.3	Data analysis and interpretation	70
5.4	Results	71
5.4.1	Isolates from poultry farm	71
5.4.2	Poultry meat markets	77
5.5	Discussion	86
5.6	Conclusion	88
6	MOLECULAR CHARACTERIZATION AND PHYLOGENETIC ANALYSIS OF ESBL-EC ISOLATES	89
6.1	Introduction	89
6.2	Materials and methods	89
6.2.1	Bacterial Strains and Culture Conditions	89
6.2.2	PFGE protocol	90
6.3	Results	91
6.3.1	ESBL-EC isolates from poultry farms	91
6.3.2	ESBL-EC isolated from poultry meat wet-markets	95
6.4	Discussion	99

6.5	Conclusion	101
7	SUMMARY, CONCLUSION AND RECOMMENDATIONS	102
7.1	General discussion	102
7.2	Conclusion	107
7.3	Recommendations for future research	107
BIBLIOGRAPHY		109
APPENDICES		133
BIODATA OF STUDENT		144
LIST OF PUBLICATIONS		145



LIST OF TABLES

Table		Page
2.1	Classification and diversity β -lactamases (Bush and Jacoby, 2010).	11
3.1	PCR primers used for the detection of <i>E. coli</i> and β -lactamase encoding genes	37
3.2	Occurrence of ESBL- EC in chickens and environmental samples according to farms	40
3.3	Occurrence of ESBL-EC in chickens and different types of environmental samples collected within poultry farms	41
3.4	Occurrence of ESBL- EC in chickens within the poultry farms	42
3.5	Occurrence of ESBL- EC in environmental samples collected within poultry farms	43
3.6	Occurrence of ESBL- EC at poultry meat wet-markets	44
3.7	Occurrence of ESBL- EC among samples collected	46
3.8	Variation in occurrence of ESBL- EC between meat and contact surfaces	47
4.1	Factors associated with ESBL-EC in poultry farms	56
4.2	Univariable factors associated with ESBL-EC in poultry farms	58
4.3	Factors associated with ESBL-EC at poultry meat wet-markets	60
4.4	Univariable factors associated with ESBL-EC at poultry meat wet-markets	61
4.5	Multivariable factors associated with ESBL-EC at poultry meat wet-markets	61
5.1	Zone diameter breakpoint interpretation criterion based on CLSI (2010)	70
5.2	Comparison of antibiotic resistance between ESBL-EC isolates from chicken and from farm environment	73
5.3	Antibiogram of ESBL-EC isolates from Chicken and farm environment	75

5.4	Major antibiogrouping (clusters) with their sources of ESBL-EC isolates from chicken and farm environment	77
5.5	Comparison of antibiotic resistance between ESBL-EC isolates from chicken meat and from contact surfaces	80
5.6	Antibiogram of ESBL-EC isolates from chicken meat and contact surfaces.	82
5.7	Major antibiogrouping (clusters) with their sources of ESBL-EC isolates from chicken meat and its contact surfaces	83
5.8	Comparison of antibiotic resistance between ESBL-EC isolates from poultry farms and from poultry meat wet-markets	85

LIST OF FIGURES

Figure		Page
2.1	TEM-ESBLs derivatives of TEM-1 β -lactamases encoding gene (Bradford, 2001).	15
2.2	SHV-ESBLs derivatives of SHV-1 β -lactamases encoding gene (Bradford, 2001).	16
3.1	Map of Selangor indicating its nine districts area and Kuala Lumpur as the federal capital territory.	32
3.2	Colonies of <i>E. coli</i> on Chromocult Coliform agar.	38
3.3	Colonies of ESBL-EC on CHROMagar ESBL.	39
3.4	Combination Disk Method for phenotypic confirmation of ESBL-EC Using Ceftazidime (CAZ): ceftazidime/clavulanate (CAZ/CV) & Cefotaxime (CTX): cefotaxime/clavulanate (CTX/CV)	39
3.5	m-PCR for <i>bla</i> -encoding gene ESBL-EC	40
3.6	Comparison between the occurrence of ESBL- <i>E. coli</i> in chickens and environmental samples.	42
3.7	Choropleth map showing the distribution of ESBL-EC at poultry meat wet-market	45
5.1	Percentages of antibiotic resistant ESBL-EC isolates from chicken and farm environment	71
5.2	Number of antibiotics and their percentage resistance among ESBL-EC isolates from chicken and farm environment.	72
5.3	Comparison of percentages of antibiotic resistance between ESBL-EC isolates from chicken and from farm environment.	74
5.4	Antibiotic resistance patterns of ESBL-EC isolates from chicken meat and contact surface	78
5.5	Number and percentage of antibiotic resistance of ESBL-EC isolates from chicken meat and contact surfaces	79
5.6	Comparison of percentages of antibiotic resistance between ESBL-EC isolates from chicken meat and from contact surfaces	81

5.7	Comparison of antibiotic resistance patterns between ESBL-EC isolates from poultry farms and from poultry meat wet-markets.	86
6.1	PFGE of ESBL-EC DNA digested with <i>Xba</i> 1.	93
6.2	Dendrogram resulting from the analysis of poultry farm ESBL-ECPFGE profiles digested with <i>Xba</i> 1.	94
6.3	Dendrogram resulting from the analysis of poultry meat wet-market ESBL-EC PFGE profiles digested with <i>Xba</i> 1.	96
6.4	Dendrogram resulting from the analysis of poultry farm and poultry meat wet-market ESBL-EC PFGE profiles digested with <i>Xba</i> 1.	98



LIST OF ABBREVIATIONS

AMR	Antimicrobial resistance
APEC	Avian pathogenic <i>E. coli</i>
CDC	Centers for Disease Control and Prevention
CI	Confidence Interval
CIPARS	Canadian Integrated Program for Antimicrobial Resistance Surveillance
CLB	Cell lysis buffer
CLSI	Clinical and Laboratory Standards Institute
DAEC	Diffusely adherent <i>E. coli</i>
DNA	Deoxyribonucleic acid
Dntp	Deoxyribonucleotide triphosphate
DSC	Decreased susceptibility to cephalosporins
DSM	Department of Statistics Malaysia
DVS	Department of Veterinary Services
EAEC	Enteroaggregative <i>E. coli</i>
EDTA	Ethylenediaminetetraacetic acid
EHEC	Enterohaemorrhagic <i>E. coli</i>
EIEC	Enteroinvasive <i>E. coli</i>
ELISA	Enzyme-linked immunosorbent assay
EPEC	Enteropathogenic <i>E. coli</i>
ESBL-EC	Extended-spectrum beta-lactamase producing <i>Escherichia coli</i>
ESBLs	Extended-spectrum beta-lactamases
ESC	Extended-spectrum cephalosporins
ETEC	Enterotoxigenic <i>E. coli</i>

FAO	Food and Agricultural Organization of United Nation
HPAI	Highly Pathogenic Avian Influenza
IDSA	Infectious Diseases Society of America
MDR	Multidrug resistant
MHA	Mueller-Hinton agar
MIC	Minimum Inhibitory Concentration
MMT	Million Metric Tons
MRSA	Methicillin resistance <i>Staphylococcus aureus</i>
MT	Metric Tons
NAG	N-acetylglucosamine
NAM	N-acetylmuramic acid
NARMS	Antimicrobial Resistance Monitoring System
NMEC	Neonatal meningitis <i>E. coli</i>
OIE	International Organization for Animal Health
OR	Odds ratio
PBPs	Penicillin-binding proteins
PCR	Polymerase chain reaction
PFGE	Pulsed field gel electrophoresis
PPE	Personal Protective Equipment
REF	Reference variable
RM	Malaysian Ringgit
rRNA	Ribosomal ribonucleic acid
SSCmec	<i>Staphylococcus</i> cassette chromosome mec
TBE	Tris-Borate-EDTA
UPEC	Uropathogenic <i>E. coli</i>

UPM	Universiti Putra Malaysia
USDA	United States Department of Agriculture
UTI	Urinary tract infection
VRI	Veterinary Research Institute
WHO	World Health Organization
WPR	World Poultry Report



CHAPTER 1

INTRODUCTION

Extended-spectrum beta-lactamase producing *Escherichia coli* (ESBL-EC) are plasmid mediated emerging zoonotic and multidrug resistant bacteria. ESBL-EC bear resistance to β -lactams, the penicillins first-, second-, third- and fourth-generation cephalosporins and monobactam; with the exception of cephemycins and carbapenems. At the same time they possess co-resistance to several other groups of antibiotics. However, ESBL-EC are inhibited by clavulanic acid and tazobactam (Rawat & Nair, 2010).

Beta-lactams (β -lactams) are first line antibiotics in human and veterinary medicine (Ewers et al., 2011; Liu et al., 2007). Over the last six decades, β -lactams are considered as the most widely used antibiotic (Livermore et al., 2006). WHO (2009) have categorized β -lactams as critically important antimicrobials for veterinary medicine and highly important antimicrobials in human medicine, with high efficacy and safety in clinical drug therapy. Thus resistance to this group of antibiotics serves as a great threat to public and animal health (Sahu et al., 2011).

The continuous exposure of bacterial strains to the first and second generation cephalosporins has induced a dynamic mutation, with an increase selective pressure and dramatic expansion of β -lactamases spectrum of activity and formation of a newly hydrolytic enzyme known as extended-spectrum beta-lactamases (ESBLs) (Shaikh et al., 2015). These are enzymes that confer resistance to the newly developed extended spectrum β -lactams (third- and fourth-generation cephalosporins). ESBLs are primarily produced by Enterobacteriaceae, particularly *Klebsiella pneumoniae* and *E. coli*. *Escherichia coli* has been reported as the most dominant ESBLs producing organism across the world (Kassakian et al., 2014).

The distribution of ESBLs producing organisms showed endemic level in several countries across the globe (Cantón & Coque, 2006), with several studies consistently reporting relatively high prevalence of ESBLs-positive organisms worldwide; however, Asia has been reported to have the highest proportion of ESBL-EC (Paterson et al., 2005; Dhillon and Clark 2011).

The misuse of extended-spectrum cephalosporins (ESC) in human and veterinary medicine has been considered as the major factor facilitating the development and global dissemination of ESBLs (Ewers et al., 2012). This alarming trend is further favored by the horizontal transfer of plasmid mediated ESBLs encoding gene among different bacterial populations, and the ability of *E. coli* to survive and proliferate across human and animal populations, and in their environment (Cantón et al., 2012; Zurfluh et al., 2014).

Food safety and antimicrobial resistance have recently received international attention, over their huge impact on economy, public health, and their potential threat to post-antibiotic era (WHO, 2014). ESBL-EC is the most frequent cause of blood stream infections, community and hospital acquired urinary tract infection, also associated with increased mortality, prolonged hospitalization, and increased cost of hospitalization, limited therapeutic option, high rate of treatment failure, co-resistance to several groups of antibiotics, and the world most leading cause of foodborne infection (Kotapati et al., 2005; Schwaber et al., 2006; Kraker et al., 2011; Rottier et al., 2012; WHO, 2014).

The incidence of outbreaks and nosocomial infections caused by ESBLs-producing *K. pneumoniae* harboring SHV-5 β -lactamase was reported in a Malaysian tertiary hospital in 1995- 1996 (Subramaniam et al., 2006). Two years later, another variant of SHV-5- β -lactamase was detected from the same hospital; thus it was postulated to be acquired via the horizontal transfer of plasmid encoding SHV-5 β -lactamase genes from the predominance threat of SHV-5 β -lactamases in *K. pneumoniae* isolates that were persistently experienced in the hospital (Subramaniam et al., 2006). Since the first identification of ESBL-EC, it has continue to be a major problem affecting the health care settings in Malaysia (Lim et al. 2009; Sekawi et al. 2008), thus threatening the efficacy of clinical drug therapy.

Poultry industry constitutes a greater percentage of human food animal protein source and poultry has been questioned as a major reservoir for the dissemination of ESBL-EC among humans, which were acquired directly or indirectly through poultry production, handling, processing and consumption of contaminated poultry product (Kola et al., 2012; Hille et al., 2013; Sharp et al., 2013). However, little or no research has been conducted in Malaysia in order to understand the potential role of poultry industry in the dynamic rise and dispersal of ESBL- *E. coli* within the community. Thus, there is a need to understand the role of poultry food chain as an important exposure pathway for acquisition and transmission of ESBL-EC into human populations. Hence, this warrants the study on the prevalence and operational and management practices that may significantly contribute to the occurrence of ESBL-EC in poultry farms and poultry-meat retail-markets. Understanding this phenomenon is very crucial in designing an effective and evidence-based veterinary and public health intervention, which will serve as a useful guide for the control and prevention of ESBL-EC along the poultry food chain.

The study was designed to answer the following questions:

1. What is the prevalence of ESBL-EC in poultry farms and poultry-meat retail-markets in Selangor, Malaysia?
2. What types of β -lactamase encoding genes are associated with the phenotypically detected ESBL-EC from poultry farms and poultry meat wet-markets in Selangor, Malaysia?
3. What types of antibiotics were the isolates resistant to?
4. What factors were possibly associated with the occurrence of ESBL-EC in poultry farms and poultry meat retail-markets?

5. Is there any molecular relationship between and within the ESBL-EC isolated from the farms and markets?

The hypotheses were:

1. There is a variation in the occurrence of ESBL-EC between and within the poultry farms and poultry meat wet-markets of Selangor, Malaysia.
2. There is a difference in the level of antibiotic resistance among the isolates of poultry farms and poultry meat wet-markets.
3. Several risk factors play a vital role in the occurrence of ESBL-EC at poultry farms and poultry meat wet-markets.
4. There are genetic similarities between and within the ESBL-EC isolated from the poultry farms and poultry meat wet-markets.

General objective

The study aimed to determine the prevalence, antimicrobial resistant profile, genetic similarities and risk factor associated with ESBL-EC in poultry farms and poultry-meat retail-markets in Selangor, Malaysia.

Specific objectives

1. To determine the prevalence of ESBL-EC in poultry farms and poultry meat wet-markets in Selangor, Malaysia.
2. To determine the level of antibiotic resistance of the ESBL-EC isolates.
3. To determine the risk factors associated with the occurrence of ESBL-EC in the poultry farms and poultry meat wet-markets.
4. To conduct molecular characterization and cluster analysis among the ESBL-EC isolates.

BIBLIOGRAPHY

- Aarestrup, F. M. (1999). Association between the consumption of antimicrobial agents in animal husbandry and the occurrence of resistant bacteria among food animals. *International Journal of antimicrobial Agents*, 12(4), 279-285.
- Abdel-Maksoud, M., Abdel-Khalek, R., El-Gendy, A., Gamal, R. F., Abdelhady, H. M., & House, B. L. (2015). Genetic characterisation of multidrug-resistant *Salmonella enterica* serotypes isolated from poultry in Cairo, Egypt. *African Journal of Laboratory Medicine*, 4(1), 1-7.
- Abeylath, S. C., & Turos, E. (2008). Drug delivery approaches to overcome bacterial resistance to β -lactam antibiotics. *Expert Opinion on Drug Delivery*, 5(9), 931-949.
- Abraham, E. P., & Chain, E. (1940). An enzyme from bacteria able to destroy penicillin. *Nature*, 146(3713), 837.
- Abraham, E. P., & Newton, G. G. F. (1961). The structure of cephalosporin C. *Biochemical Journal*, 79(2), 377.
- Ahmad, A., Græsbøll, K., Christiansen, L. E., Toft, N., Matthews, L., & Nielsen, S. S. (2015). Pharmacokinetic-pharmacodynamic model to evaluate intramuscular tetracycline treatment protocols to prevent antimicrobial resistance in pigs. *Antimicrobial Agents and Chemotherapy*, 59(3), 1634-1642.
- Ahmed, O. I., El-Hady, S. A., Ahmed, T. M., & Ahmed, I. Z. (2013). Detection of bla SHV and bla CTX-M genes in ESBL producing *Klebsiella pneumoniae* isolated from Egyptian patients with suspected nosocomial infections. *Egyptian Journal of Medical Human Genetics*, 14(3), 277-283.
- Aidara-Kane, A., Andremont, A., & Collignon, P. (2013). Antimicrobial resistance in the food chain and the AGISAR initiative. *Journal of Infection and Public Health*, 6(3), 162-165.
- Al-Agamy, M. H., Shibli, A. M., & Tawfik, A. F. (2009). Prevalence and molecular characterization of extended-spectrum β -lactamase-producing *Klebsiella pneumoniae* in Riyadh, Saudi Arabia. *Annals of Saudi Medicine*, 29(4), 253.
- Alhetar, K. Y. A., Sekawi, Z., Mariana, N. S., & Neela, V. (2011). Molecular characterization of extended-spectrum beta-lactamase (ESBL) producing extra-intestinal pathogenic *Escherichia coli*. *African Journal of Microbiology Research*, 5(31), 5662-5668.
- Aliyu, A. B., Saleha, A. A., Jalila, A., & Zunita, Z. (2016). Risk factors and spatial distribution of extended spectrum β -lactamase-producing-*Escherichia coli* at retail poultry meat markets in Malaysia: a cross-sectional study. *BMC Public Health*, 16(1), 699.

- Allocati, N., Masulli, M., Alexeyev, M. F., & Di Ilio, C. (2013). *Escherichia coli* in Europe: An Overview. *International Journal of Environmental Research and Public Health*, 10(12), 6235–6254.
- Andersen, V. D., Jensen, V. F., Vigre, H., Andreasen, M., & Agersø, Y. (2015). The use of third and fourth generation cephalosporins affects the occurrence of extended-spectrum cephalosporinase-producing *Escherichia coli* in Danish pig herds. *The Veterinary Journal*, 204(2015), 345–350.
- Arlet, G., & Philippon, A. (1991). Construction by polymerase chain reaction and intragenic DNA probes for three main types of transferable β -lactamases (TEM, SHV, CARB). *FEMS Microbiology Letters*, 82(1), 19-25.
- Arlet, G., Brami, G., Decre, D., Flippo, A., Gaillot, O., Lagrange, P. H., & Philippon, A. (1995). Molecular characterisation by PCR-restriction fragment length polymorphism of TEM β -lactamases. *FEMS Microbiology Letters*, 134(2-3), 203-208.
- Bauernfeind, A., Holley, M., Jungwirth, R., Mangold, P., Röhnisch, T., Schweighart, S., Wilhelm, R., Casellas, J.M., & Goldberg, M. (1992). A new plasmidic cefotaximase from patients infected with *Salmonella typhimurium*. *Infection*, 20(3), 158-163.
- Bauernfeind, A., Stemplinger, I., Jungwirth, R., Mangold, P., Amann, S., Akalin, E., Anđ, O., Bal, C., & Casellas, J. M. (1996). Characterization of beta-lactamase gene blaPER-2, which encodes an extended-spectrum class A beta-lactamase. *Antimicrobial Agents and Chemotherapy*, 40(3), 616–620.
- Ben Sallem, R., Ben Slama, K., Sáenz, Y., Rojo-Bezares, B., Estepa, V., Jouini, A., Gharsa, H., Klibi, N., Boudabous, A., & Torres, C. (2012). Prevalence and Characterization of Extended-Spectrum Beta-Lactamase (ESBL)-and CMY-2-Producing *Escherichia coli* Isolates from Healthy Food-Producing Animals in Tunisia. *Foodborne Pathogens and Disease*, 9(12), 1137-1142.
- Bengtsson, B., & Greko, C. (2014). Antibiotic resistance-consequences for animal health, welfare, and food production. *Upsala Journal of Medical Sciences*, 119(2), 96-102.
- Berglund, B. (2015). Environmental dissemination of antibiotic resistance genes and correlation to anthropogenic contamination with antibiotics. *Infection Ecology & Epidemiology*, 5, 10.3402/iee.v5.28564.
- Blaak, H., Hamidjaja, R. A., van Hoek, A. H., de Heer, L., de Roda Husman, A. M., & Schets, F. M. (2014). Detection of extended-spectrum beta-lactamase (ESBL)-producing *Escherichia coli* on flies at poultry farms. *Applied and Environmental Microbiology*, 80(1), 239-246.

- Blaak, H., van Hoek, A. H., Hamidjaja, R. A., van der Plaats, R. Q., Kerkhof-de Heer, L., de Roda Husman, A. M., & Schets, F. M. (2015). Distribution, numbers, and diversity of ESBL-producing *E. coli* in the poultry farm environment. *PLoS One*, 10(8), e0135402.
- Blanc, D. S., Lugeon, C., Wenger, A., Siegrist, H. H., & Francioli, P. (1994). Quantitative antibiogram typing using inhibition zone diameters compared with ribotyping for epidemiological typing of methicillin-resistant *Staphylococcus aureus*. *Journal of Clinical Microbiology*, 32(10), 2505-2509.
- Blanc, D. S., Petignat, C., Moreillon, P., Wenger, A., Bille, J., & Francioli, P. (1996). Quantitative Antibiogram as a Typing Method for the Prospective Epidemiological Surveillance and Control of MRSA Comparison with Molecular Typing. *Infection Control & Hospital Epidemiology*, 17(10), 654-659.
- Bonnedahl, J., Drobni, M., Gauthier-Clerc, M., Hernandez, J., Granholm, S., Kayser, Y., Melhus, Å., Kahlmeter, G., Waldenström, J., Johansson, A., & Olsen, B. (2009). Dissemination of *Escherichia coli* with CTX-M type ESBL between humans and yellow-legged gulls in the south of France. *PLoS one*, 4(6), e5958.
- Bonnet, C., Diarrassouba, F., Brousseau, R., Masson, L., Topp, E., & Diarra, M. S. (2009). Pathotype and antibiotic resistance gene distributions of *Escherichia coli* isolates from broiler chickens raised on antimicrobial-supplemented diets. *Applied and Environmental Microbiology*, 75(22), 6955-6962.
- Bouamama, L., Sorlozano, A., Laglaoui, A., Lebbadi, M., Aarab, A., & Gutierrez, J. (2010). Antibiotic resistance patterns of bacterial strains isolated from *Periplaneta americana* and *Musca domestica* in Tangier, Morocco. *The Journal of Infection in Developing Countries*, 4(04), 194-201.
- Boyle, D. P., & Zembower, T. R. (2015). Epidemiology and Management of Emerging Drug-Resistant Gram-Negative Bacteria: Extended-Spectrum β -Lactamases and Beyond. *Urologic Clinics of North America*. <http://dx.doi.org/10.1016/j.ucl.2015.05.005>
- Bradford, P. A. (1999). Automated Thermal Cycling Is Superior to Traditional Methods for Nucleotide Sequencing of bla SHV Genes. *Antimicrobial Agents and Chemotherapy*, 43(12), 2960-2963.
- Bradford, P. A. (2001). Extended-spectrum β -lactamases in the 21st century: characterization, epidemiology, and detection of this important resistance threat. *Clinical Microbiology Reviews*, 14(4), 933-951.
- Burgess, F., Little, C. L., Allen, G., Williamson, K., & Mitchell, R. T. (2005). Prevalence of *Campylobacter*, *Salmonella*, and *Escherichia coli* on the external packaging of raw meat. *Journal of Food Protection*, 68(3), 469-475.

- Burwen, D. R., Banerjee, S. N., Gaynes, R. P., & National Nosocomial Infections Surveillance System. (1994). Ceftazidime resistance among selected nosocomial gram-negative bacilli in the United States. *Journal of Infectious Diseases*, 170(6), 1622-1625.
- Bush, K. (1989). Characterization of beta-lactamases. *Antimicrobial Agents and Chemotherapy*, 33(3), 259.
- Bush, K., & Jacoby, G. A. (2010). Updated functional classification of β -lactamases. *Antimicrobial Agents and Chemotherapy*, 54(3), 969-976.
- Bush, K., Jacoby, G. A., & Medeiros, A. A. (1995). A functional classification scheme for beta-lactamases and its correlation with molecular structure. *Antimicrobial Agents and Chemotherapy*, 39(6), 1211.
- Byarugaba, D. K. (2004). Antimicrobial resistance in developing countries and responsible risk factors. *International Journal of Antimicrobial Agents*, 24(2), 105-110.
- Cantón, R., & Coque, T. M. (2006). The CTX-M β -lactamase pandemic. *Current Opinion in Microbiology*, 9(5), 466-475.
- Cantón, R., González-Alba, J. M., & Galán, J. C. (2012). CTX-M Enzymes: Origin and Diffusion. *Frontiers in Microbiology*, 3, 110.
- Carlier, M., Stove, V., Wallis, S. C., De Waele, J. J., Verstraete, A. G., Lipman, J., & Roberts, J. A. (2015). Assays for therapeutic drug monitoring of β -lactam antibiotics: A structured review. *International Journal of Antimicrobial Agents*. (article in press) <http://dx.doi.org/10.1016/j.ijantimicag.2015.06.016>.
- Centers for Disease Control and Prevention (2014). Antimicrobial Resistance Threat Report. 2013. <http://www.cdc.gov/drugresistance/threat-report-2013/>. Accessed 02 Feb 2016.
- Chopra, I., Hodgson, J., Metcalf, B., & Poste, G. (1997). The search for antimicrobial agents effective against bacteria resistant to multiple antibiotics. *Antimicrobial Agents and Chemotherapy*, 41(3), 497.
- Clinical and Laboratory Standards Institute (CLSI) (2010). Clinical and Laboratory Standards Institute: Performance standards for Antimicrobial Disk Susceptibility Tests. Approved standard, 10th ed. Wayne 2010, PA: CLSI document M02-A10.
- Collignon, P. (2009). Resistant *Escherichia coli*—we are what we eat. *Clinical Infectious Diseases*, 49(2), 202-204.
- Cools, I., Uyttendaele, M., Cerpentier, J., D'haese, E., Nelis, H. J., & Debevere, J. (2005). Persistence of *Campylobacter jejuni* on surfaces in a processing environment and on cutting boards. *Letters in Applied Microbiology*, 40(6), 418-423.

- Cooper, K. L. F., Luey, C. K. Y., Bird, M., Terajima, J., Nair, G. B., Kam, K. M., Arakawa, E., Safa, A., Cheung, D.T., Law, C.P., & Watanabe, H. (2006). Development and validation of a PulseNet standardized pulsed-field gel electrophoresis protocol for subtyping of *Vibrio cholerae*. *Foodborne Pathogens & Disease*, 3(1), 51-58.
- Cormican, M. G., Marshall, S. A., & Jones, R. N. (1996). Detection of extended-spectrum beta-lactamase (ESBL)-producing strains by the E-test ESBL screen. *Journal of Clinical Microbiology*, 34(8), 1880-1884.
- Costa, D., Vinué, L., Poeta, P., Coelho, A. C., Matos, M., Sáenz, Y., Somalo, S., Zarazaga, M., Rodrigues, J., & Torres, C. (2009). Prevalence of extended-spectrum beta-lactamase-producing *Escherichia coli* isolates in faecal samples of broilers. *Veterinary Microbiology*, 138(3), 339-344.
- Cowan, S. T. (1954). A review of names for coliform organisms. *International Bulletin of Bacteriological Nomenclature and Taxonomy*, 4(2), 119-124.
- Croxen, M. A., & Finlay, B. B. (2010). Molecular mechanisms of *Escherichia coli* pathogenicity. *Nature Reviews Microbiology*, 8(1), 26-38.
- De Kraker, M. E. A., Wolkewitz, M., Davey, P. G., Koller, W., Berger, J., Nagler, J., & Grundmann, H. (2011). Burden of antimicrobial resistance in European hospitals: excess mortality and length of hospital stay associated with bloodstream infections due to *Escherichia coli* resistant to third-generation cephalosporins. *Journal of Antimicrobial Chemotherapy*, 66(2), 398-407.
- De Waele, J. J., Lipman, J., Carlier, M., & Roberts, J. A. (2015). Subtleties in practical application of prolonged infusion of β -lactam antibiotics. *International Journal of Antimicrobial Agents*, 45(5), 461-463.
- Delattre, I. K., Musuamba, F. T., Verbeeck, R. K., Dugernier, T., Spapen, H., Laterre, P. F., Wittebole, X., Cumps, J., Taccone, F.S., Vincent, J.L., & Jacobs, F. (2010). Empirical models for dosage optimization of four β -lactams in critically ill septic patients based on therapeutic drug monitoring of amikacin. *Clinical Biochemistry*, 43(6), 589-598.
- Department of Statistics, Malaysia (DSM, 2011a). Preliminary Count Report 2010, p. 27. Retrieved 24 January 2011.
- Department of Statistics, Malaysia (DSM, 2011b). Preliminary Count Report 2010, p. iv. Retrieved 24 January 2011.
- Dhillon, R. H.-P., & Clark, J. (2012). ESBLs: A Clear and Present Danger? *Critical Care Research and Practice*, 2012, 625170.
- Diarra, M. S., Rempel, H., Champagne, J., Masson, L., Pritchard, J., & Topp, E. (2010). Distribution of antimicrobial resistance and virulence genes in *Enterococcus* spp. and characterization of isolates from broiler chickens. *Applied and Environmental Microbiology*, 76(24), 8033-8043.

- Dierikx, C., van der Goot, J., Fabri, T., van Essen-Zandbergen, A., Smith, H., & Mevius, D. (2013). Extended-spectrum- β -lactamase-and AmpC- β -lactamase-producing *Escherichia coli* in Dutch broilers and broiler farmers. *Journal of Antimicrobial Chemotherapy*, 68(1), 60-67.
- Doi, Y., Park, Y. S., Rivera, J. I., Adams-Haduch, J. M., Hingwe, A., Sordillo, E. M., Lewis, J.S., Howard, W.J., Johnson, L.E., Polksky, B., & Jorgensen, J.H. (2013). Community-associated extended-spectrum β -lactamase-producing *Escherichia coli* infection in the United States. *Clinical Infectious Diseases*, 56(5), 641-648.
- Drawz, S. M., & Bonomo, R. A. (2010). Three decades of β -lactamase inhibitors. *Clinical Microbiology Reviews*, 23(1), 160-201.
- Egea, P., López-Cerero, L., Torres, E., del Carmen Gómez-Sánchez, M., Serrano, L., Sánchez-Ortiz, M. D. N., Rodríguez-Baño, J., & Pascual, A. (2012). Increased raw poultry meat colonization by extended spectrum beta-lactamase-producing *Escherichia coli* in the south of Spain. *International Journal of Food Microbiology*, 159(2), 69-73.
- Eric Scholar (2007). Cephalosporins. Reference Module in Biomedical Sciences, from xPharm: *The Comprehensive Pharmacology Reference*, Pages 1-4.
- Escherich, T. (1988). The intestinal bacteria of the neonate and breast-fed infant. *Review of Infectious Diseases*, 10(6), 1220-1225.
- European Commission. Action plan against the rising threats from Antimicrobial Resistance. Communication from the Commission to the European Parliament and the Council. COM (2011) 748. http://ec.europa.eu/dgs/health_food-safety/docs/communication Accessed 02 Feb 2016
- Ewers, C., Bethe, A., Semmler, T., Guenther, S., & Wieler, L. H. (2012). Extended-spectrum β -lactamase-producing and AmpC-producing *Escherichia coli* from livestock and companion animals, and their putative impact on public health: a global perspective. *Clinical Microbiology and Infection*, 18(7), 646-655.
- FAO (2012). FAO supports measures to minimize and contain antimicrobial resistance (AMR).
http://www.fao.org/ag/againfo/home/en/news_archive/2011_04_AMR.html
updated: 25 September 2012. Accessed 20-06-2015.
- FAO (2015): FAO's role in addressing antimicrobial resistance.
<http://www.fao.org/antimicrobial-resistance/en/>. Accessed 20-06-2016.
- FDA (2016): U.S. Food and Drug Administration. Antimicrobial Resistance: Animal and Veterinary.
<http://www.fda.gov/AnimalVeterinary/SafetyHealth/AntimicrobialResistance/>. Accessed 17 Feb 2016.

- Fernandes, S. A., Paterson, D. L., Ghilardi-Rodrigues, Â. C., Adams-Haduch, J. M., Tavechio, A. T., & Doi, Y. (2009). CTX-M-2-Producing *Salmonella Typhimurium* Isolated from Pediatric Patients and Poultry in Brazil. *Microbial Drug Resistance*, 15(4), 317-321.
- Folster, J. P., Pecic, G., Singh, A., Duval, B., Rickert, R., Ayers, S., Abbott, J., McGlinchey, B., Bauer-Turpin, J., Haro, J., & Hise, K. (2012). Characterization of extended-spectrum cephalosporin-resistant *Salmonella enterica* serovar Heidelberg isolated from food animals, retail meat, and humans in the United States 2009. *Foodborne Pathogens and Disease*, 9(7), 638-645.
- Franz, E., Schijven, J., de Roda Husman, A. M., & Blaak, H. (2014). Meta-Regression analysis of commensal and pathogenic *Escherichia coli* survival in soil and water. *Environmental Science & Technology*, 48(12), 6763-6771.
- Gallego, L., Umaran, A., Garaizar, J., Colom, K., & Cisterna, R. (1990). Digoxigenin-labeled DNA probe to detect TEM type β -lactamases. *Journal of Microbiological Methods*, 11(3), 261-267.
- Garau, J. (2008). Other antimicrobials of interest in the era of extended-spectrum β -lactamases: fosfomycin, nitrofurantoin and tigecycline. *Clinical Microbiology and Infection*, 14(s1), 198-202.
- Girlich, D., Poirel, L., Carattoli, A., Kempf, I., Lartigue, M. F., Bertini, A., & Nordmann, P. (2007). Extended-spectrum β -lactamase CTX-M-1 in *Escherichia coli* isolates from healthy poultry in France. *Applied and Environmental Microbiology*, 73(14), 4681-4685.
- Goossens, H., & Grabein, B. (2005). Prevalence and antimicrobial susceptibility data for extended-spectrum β -lactamase-and AmpC-producing Enterobacteriaceae from the MYSTIC Program in Europe and the United States (1997–2004). *Diagnostic Microbiology and Infectious Disease*, 53(4), 257-264.
- Gould, I. M., Wang, G. Q., Wu, J. J., Lim, V. K. E., Hutchinson, J., Walsh, T., & Turnidge, J. (2014). MDRO Beijing Consensus Meeting Report: Global burden of multidrug-resistant organisms' current antimicrobial resistance problems in Asia-Pacific. *Journal of Global Antimicrobial Resistance*, 2(1), 7-9.
- Graham, J. P., Evans, S. L., Price, L. B., & Silbergeld, E. K. (2009a). Fate of antimicrobial-resistant enterococci and staphylococci and resistance determinants in stored poultry litter. *Environmental Research*, 109(6), 682-689.
- Graham, J. P., Price, L. B., Evans, S. L., Graczyk, T. K., & Silbergeld, E. K. (2009b). Antibiotic resistant enterococci and staphylococci isolated from flies collected near confined poultry feeding operations. *Science of the Total Environment*, 407(8), 2701-2710.

- Guenther, S., Bethe, A., Fruth, A., Semmler, T., Ulrich, R. G., Wieler, L. H., & Ewers, C. (2012). Frequent combination of antimicrobial multiresistance and extraintestinal pathogenicity in *Escherichia coli* isolates from urban rats (*Rattus norvegicus*) in Berlin, Germany. *PLoS ONE* 2012 7:e50331.
- Guenther, S., Ewers, C., & Wieler, L. H. (2011). Extended-spectrum beta-lactamases producing *E. coli* in wildlife, yet another form of environmental pollution? *Frontiers in Microbiology*, 2:246.
- Guenther, S., Grobbel, M., Beutlich, J., Guerra, B., Ulrich, R. G., Wieler, L. H., & Ewers, C. (2010). Detection of pandemic B2-O25-ST131 *Escherichia coli* harbouring the CTX-M-9 extended-spectrum β-lactamase type in a feral urban brown rat (*Rattus norvegicus*). *Journal of Antimicrobial Chemotherapy*, 65(3), 582-584.
- Gustaferro, C. A., & Steckelberg, J. M. (1991). Cephalosporin antimicrobial agents and related compounds. In *Mayo Clinic Proceedings*, 66(10), 1064-1073.
- Harbarth, S., Balkhy, H. H., Goossens, H., Jarlier, V., Kluytmans, J., Laxminarayan, R., Saam, M., Van Belkum, A., & Pittet, D. (2015). Antimicrobial resistance: one world, one fight!. *Antimicrobial Resistance and Infection Control*, 4(1), 1.
- Harrison, W. A., Griffith, C. J., Tennant, D., & Peters, A. C. (2001). Incidence of *Campylobacter* and *Salmonella* isolated from retail chicken and associated packaging in South Wales. *Letters in Applied Microbiology*, 33(6), 450-454.
- Hasan, B., Sandegren, L., Melhus, Å., Drobni, M., Hernandez, J., Waldenström, J., Alam, M., & Olsen, B. (2012). Antimicrobial drug-resistant *Escherichia coli* in wild birds and free-range poultry, Bangladesh. *Emerging Infectious Diseases*, 18(12), 2055-2058.
- Hashim, R. B., Husin, S., & Rahman, M. M. (2011). Detection of betalactamase producing bacterial genes and their clinical features. *Pakistan Journal of Biological Sciences*, 14(1), 41-46.
- Hassan, L., Wan Mohd Zain, W. N. A., Saleha, A.A., & Ramanoon, S. Z (2006). Epidemiological features of vancomycin-resistant enterococci in broiler farms in Malaysia. Proceedings of the 11th International Symposium on Veterinary Epidemiology and Economics. Available at www.sciquest.org.nz
- Hassan, S. A., & Shobrak, M. Y. (2014). Prevalence and Antimicrobial Resistance Characteristics of Gram-negative Bacteria Associated with Wild Animals Presenting at Live Animal Market, Taif, Western Saudi Arabia. *Journal of Pure and Applied Microbiology*, 8(Spl. Edn. 2), 273-282.
- Hawkey, P. M. (2008). Prevalence and clonality of extended-spectrum β-lactamases in Asia. *Clinical Microbiology and Infection*, 14(s1), 159-165.

- Hawser, S. P., Bouchillon, S. K., Hoban, D. J., Badal, R. E., Hsueh, P. R., & Paterson, D. L. (2009). Emergence of high levels of extended-spectrum- β -lactamase-producing gram-negative bacilli in the Asia-Pacific region: data from the Study for Monitoring Antimicrobial Resistance Trends (SMART) program, 2007. *Antimicrobial Agents and Chemotherapy*, 53(8), 3280-3284.
- Heritage, J., M'Zali, F. H., Gascoyne-Binzi, D., & Hawkey, P. M. (1999). Evolution and spread of SHV extended-spectrum β -lactamases in Gram-negative bacteria. *Journal of Antimicrobial Chemotherapy*, 44(3), 309-318.
- Heuer, H., Schmitt, H., & Smalla, K. (2011). Antibiotic resistance gene spread due to manure application on agricultural fields. *Current Opinion in Microbiology*, 14(3), 236-243.
- Hille, K., Fischer, J., Falgenhauer, L., Sharp, H., Brenner, G. M., Kadlec, K., & Kreienbrock, L. (2013). On the occurrence of extended-spectrum-and AmpC- β -lactamase-producing *Escherichia coli* in livestock: results of selected European studies. *Berliner und Munchener tierarztliche Wochenschrift*, 127(9-10), 403-411.
- Hirakata, Y., Matsuda, J., Miyazaki, Y., Kamihira, S., Kawakami, S., Miyazawa, Y., Ono, Y., Nakazaki, N., Hirata, Y., Inoue, M., & Turnidge, J.D. (2005). Regional variation in the prevalence of extended-spectrum β -lactamase-producing clinical isolates in the Asia-Pacific region (SENTRY 1998–2002). *Diagnostic microbiology and Infectious Disease*, 52(4), 323-329.
- Ho, W. S., Balan, G., Puthucheary, S., Kong, B. H., Lim, K. T., Tan, L. K., Koh, X.P., Yeo, C.C., & Thong, K.L. (2012). Prevalence and characterization of multidrug-resistant and extended-spectrum beta-lactamase-producing *Escherichia coli* from pediatric wards of a Malaysian hospital. *Microbial Drug Resistance*, 18(4), 408-416.
- <http://www.fao.org/antimicrobial-resistance/en/> Accessed 22-02-2016
- Huang, T. D., Bogaerts, P., Berhin, C., Guisset, A., & Glupczynski, Y. (2010). Evaluation of Brilliance ESBL agar, a novel chromogenic medium for detection of extended-spectrum-beta-lactamase-producing Enterobacteriaceae. *Journal of Clinical Microbiology*, 48(6), 2091-2096.
- Huletsky, A., Knox, J. R., & Levesque, R. C. (1993). Role of Ser-238 and Lys-240 in the hydrolysis of third-generation cephalosporins by SHV-type beta-lactamases probed by site-directed mutagenesis and three-dimensional modeling. *Journal of Biological Chemistry*, 268(5), 3690-3697.
- Huovinen, S., Huovinen, P., & Jacoby, G. A. (1988). Detection of plasmid-mediated beta-lactamases with DNA probes. *Antimicrobial Agents and Chemotherapy*, 32(2), 175-179.

- Imada, A., Kitano, K., Kintaka, K., Muroi, M., & Asai, M. (1981). Sulfazecin and isosulfazecin, novel beta-lactam antibiotics of bacterial origin. *Nature*, 289(5798), 590-591.
- Infectious Diseases Society of America (IDSA, 2011). Combating antimicrobial resistance: policy recommendations to save lives. *Clinical Infectious Diseases*, 52(suppl 5), S397-S428.
- Ismaïl, R., Aviat, F., Michel, V., Le Bayon, I., Gay-Perret, P., Kutnik, M., & Féderighi, M. (2013). Methods for recovering microorganisms from solid surfaces used in the food industry: a review of the literature. *International Journal of Environmental Research and Public Health*, 10(11), 6169-6183.
- Itokazu, G. S., Quinn, J. P., Bell-Dixon, C., Kahan, F. M., & Weinstein, R. A. (1996). Antimicrobial resistance rates among aerobic gram-negative bacilli recovered from patients in intensive care units: evaluation of a national post-marketing surveillance program. *Clinical Infectious Diseases*, 23(4), 779-784.
- Jan, J. S., McIntosh, W. A., Dean, W., & Scott, H. M. (2012). Predictors of differences in the perception of antimicrobial resistance risk in the treatment of sick, at-risk, and high-risk feedlot cattle. *Preventive Veterinary Medicine*, 106(1), 24-33.
- Jarlier, V., Nicolas, M. H., Fournier, G., & Philippon, A. (1988). Extended broad-spectrum β-lactamases conferring transferable resistance to newer β-lactam agents in Enterobacteriaceae: hospital prevalence and susceptibility patterns. *Review of Infectious Diseases*, 10(4), 867-878.
- Johnson, T. J., Kariyawasam, S., Wannemuehler, Y., Mangamele, P., Johnson, S. J., Doekkott, C., Skyberg, J.A., Lynne, A.M., Johnson, J.R., & Nolan, L. K. (2007). The genome sequence of avian pathogenic *Escherichia coli* strain O1: K1:H7 shares strong similarities with human extraintestinal pathogenic *E. coli* genomes. *Journal of Bacteriology*, 189(8), 3228-3236.
- Kader, A. A., & Angamuthu, K. (2005). Extended-spectrum beta-lactamases in urinary isolates of *Escherichia coli*, *Klebsiella pneumoniae* and other gram-negative bacteria in a hospital in Eastern Province, Saudi Arabia. *Saudi Medical Journal*, 26(6), 956-959.
- Kam, K. M., Luey, C. K., Parsons, M. B., Cooper, K. L., Nair, G. B., Alam, M., Islam, M.A., Cheung, D.T., Chu, Y.W., Ramamurthy, T., & Pazhani, G. P. (2008). Evaluation and validation of a PulseNet standardized pulsed-field gel electrophoresis protocol for subtyping *Vibrio parahaemolyticus*: an international multicenter collaborative study. *Journal of Clinical Microbiology*, 46(8), 2766-2773.
- Kaper, J. B., Nataro, J. P., & Mobley, H. L. (2004). Pathogenic *Escherichia coli*. *Nature Reviews Microbiology*, 2(2), 123-140.

- Kassakian, S. Z., & Mermel, L. A. (2014). Changing epidemiology of infections due to extended spectrum beta-lactamase producing bacteria. *Antimicrobial Resistance and Infection Control*, 3(1), 9.
- Keelara, S., Scott, H. M., Morrow, W. M., Gebreyes, W. A., Correa, M., Nayak, R., Stefanova, R., & Thakur, S. (2013). Longitudinal study of distributions of similar antimicrobial-resistant *Salmonella* serovars in pigs and their environment in two distinct swine production systems. *Applied and Environmental Microbiology*, 79(17), 5167-5178.
- Kilani, H., Abbassi, M. S., Ferjani, S., Mansouri, R., Sghaier, S., Salem, R. B., Jaouani, I., Douja, G., Brahim, S., Hammami, S., & Chehida, N. B. (2015). Occurrence of blaCTX-M-1, qnrB1 and virulence genes in avian ESBL-producing *Escherichia coli* isolates from Tunisia. *Frontiers in Cellular and Infection Microbiology*, 5, 38.
- Kilic, A., Bedir, O., Kocak, N., Levent, B., Eyigun, C. P., Tekbas, O. F., Gorenek, L., Baylan, O., & Basustaoglu, A. C. (2010). Analysis of an outbreak of *Salmonella enteritidis* by repetitive-sequence-based PCR and pulsed-field gel electrophoresis. *Internal Medicine*, 49(1), 31-36.
- Kilonzo-Nthenge, A., Rotich, E., & Nahashon, S. N. (2013). Evaluation of drug-resistant Enterobacteriaceae in retail poultry and beef. *Poultry Science*, 92(4), 1098-1107.
- Kliebe, C., Nies, B. A., Meyer, J. F., Tolxdorff-Neutzling, R. M., & Wiedemann, B. (1985). Evolution of plasmid-coded resistance to broad-spectrum cephalosporins. *Antimicrobial Agents and Chemotherapy*, 28(2), 302-307.
- Kluytmans, J. A., Overdevest, I. T., Willemsen, I., Kluytmans-van den Bergh, M. F., van der Zwaluw, K., Heck, M., Rijnsburger, M., Vandebroucke-Grauls, C.M., Savelkoul, P.H., Johnston, B.D., & Gordon, D. (2012). Extended-Spectrum β-lactamase-producing *Escherichia coli* from retail chicken meat and humans: comparison of strains, plasmids, resistance genes, and virulence factors. *Clinical Infectious Diseases*, cis929.
- Knothe, H., Shah, P. D. P., Krcmery, V., Antal, M., & Mitsuhashi, S. (1983). Transferable resistance to cefotaxime, cefoxitin, cefamandole and cefuroxime in clinical isolates of *Klebsiella pneumoniae* and *Serratia marcescens*. *Infection*, 11(6), 315-317.
- Koga, V. L., Rodrigues, G. R., Scandoriero, S., Vespero, E. C., Oba, A., Brito, B. G. D., de Brito, K.C., Nakazato, G., & Kobayashi, R. K. (2015a). Evaluation of the Antibiotic Resistance and Virulence of *Escherichia coli* Strains Isolated from Chicken Carcasses in 2007 and 2013 from Paraná, Brazil. *Foodborne Pathogens and Disease*, 12(6) 479-485.

- Koga, V. L., Scandoriero, S., Vespero, E. C., Oba, A., de Brito, B. G., de Brito, K. C., Nakazato, G., & Kobayashi, R. K. (2015b). Comparison of Antibiotic Resistance and Virulence Factors among *Escherichia coli* Isolated from Conventional and Free-Range Poultry. *BioMed Research International*, 2015: 618752
- Kola, A., Kohler, C., Pfeifer, Y., Schwab, F., Kühn, K., Schulz, K., Balau, V., Breitbach, K., Bast, A., Witte, W., & Gastmeier, P. (2012). High prevalence of extended-spectrum-β-lactamase-producing Enterobacteriaceae in organic and conventional retail chicken meat, Germany. *Journal of Antimicrobial Chemotherapy*, 67(11), 2631-2634.
- Komatsu, M., Aihara, M., Shimakawa, K., Iwasaki, M., Nagasaka, Y., Fukuda, S., Matsuo, S., & Iwatani, Y. (2003). Evaluation of MicroScan ESBL confirmation panel for Enterobacteriaceae-producing, extended-spectrum β-lactamases isolated in Japan. *Diagnostic Microbiology and Infectious Disease*, 46(2), 125-130.
- Kong, K. F., Schneper, L., & Mathee, K. (2010). Beta-lactam antibiotics: from antibiosis to resistance and bacteriology. *Acta Pathologica, Microbiologica, et Immunologica Scandinavica*, 118(1), 1-36.
- Kotapati, S., Kuti, J. L., Nightingale, C. H., & Nicolau, D. P. (2005). Clinical implications of extended spectrum β-lactamase (ESBL) producing *Klebsiella* species and *Escherichia coli* on cefepime effectiveness. *Journal of Infection*, 51(3), 211-217.
- Kotra, L. P., & Mobashery, S. (1998). β-Lactam antibiotics, β-lactamases and bacterial resistance. *Bulletin de l'Institut Pasteur*, 96(3), 139-150.
- Krishnasamy, V., Otte, J., & Silbergeld, E. (2015). Antimicrobial use in Chinese swine and broiler poultry production. *Antimicrobial Resistance and Infection Control*, 4(1), 17.
- Kümmerer, K. (2009). Antibiotics in the aquatic environment—a review—part II. *Chemosphere*, 75(4), 435-441.
- Kusumaningrum, H. D., Riboldi, G., Hazeleger, W. C., & Beumer, R. R. (2003). Survival of foodborne pathogens on stainless steel surfaces and cross-contamination to foods. *International Journal of Food Microbiology*, 85(3), 227-236.
- Lahlaoui, H., Khalifa, A. B. H., & Moussa, M. B. (2014). Epidemiology of Enterobacteriaceae producing CTX-M type extended spectrum β-lactamase (ESBL). *Medecine et Maladies Infectieuses*, 44(9), 400-404.
- Laube, H., Friese, A., von Salviati, C., Guerra, B., & Rösler, U. (2014). Transmission of ESBL/AmpC-producing *Escherichia coli* from broiler chicken farms to surrounding areas. *Veterinary Microbiology*, 172(3), 519-527.

- Laube, H., Friese, A., von Salviati, C., Guerra, B., Käsbohrer, A., Kreienbrock, L., & Roesler, U. (2013). Longitudinal monitoring of extended-spectrum-beta-lactamase/AmpC-producing *Escherichia coli* at German broiler chicken fattening farms. *Applied and Environmental Microbiology*, 79(16), 4815-4820.
- Laupland, K. B., Church, D. L., Vidakovich, J., Mucenski, M., & Pitout, J. D. (2008). Community-onset extended-spectrum β-lactamase (ESBL) producing *Escherichia coli*: importance of international travel. *Journal of Infection*, 57(6), 441-448.
- Le, Q. P., Ueda, S., Nguyen, T. N. H., Dao, T. V. K., Van Hoang, T. A., Tran, T. T. N., Hirai, I., Nakayama, T., Kawahara, R., Do, T.H., & Vien, Q.M. (2015). Characteristics of Extended-Spectrum β-Lactamase-Producing *Escherichia coli* in Retail Meats and Shrimp at a Local Market in Vietnam. *Foodborne Pathogens and Disease*, 12(8), 719-725.
- Leete, Richard. "Selangor's Human Development Progress and Challenges". UN Development Program. Retrieved 9 July 2011.
- Leistner, R., Meyer, E., Gastmeier, P., Pfeifer, Y., Eller, C., Dem, P., & Schwab, F. (2013). Risk factors associated with the community-acquired colonization of extended-spectrum beta-lactamase (ESBL) positive *Escherichia coli*. An exploratory case-control study. *PloS one*, 8(9), e74323.
- Leverstein-van Hall, M. A., Dierikx, C. M., Cohen Stuart, J., Voets, G. M., Van Den Munckhof, M. P., van Essen-Zandbergen, A., Platteel, T., Fluit, A.C., van de Sande-Bruinsma, N., Scharringa, J., & Bonten, M. J. M. (2011). Dutch patients, retail chicken meat and poultry share the same ESBL genes, plasmids and strains. *Clinical Microbiology and Infection*, 17(6), 873-880.
- Li, L., Jiang, Z. G., Xia, L. N., Shen, J. Z., Dai, L., Wang, Y., Huang, S.Y., & Wu, C. M. (2010). Characterization of antimicrobial resistance and molecular determinants of beta-lactamase in *Escherichia coli* isolated from chickens in China during 1970–2007. *Veterinary Microbiology*, 144(3), 505-510.
- Li, S., Zhao, M., Li, Y., Zhang, L., Zhang, X., & Miao, Z. (2013). Detection and source identification of airborne extended-spectrum beta-lactamase-producing *Escherichia coli* isolates in a chicken house. *Aerobiologia*, 29(2), 315-319.
- Lim, K. T., Yasin, R., Yeo, C. C., Puthucheary, S., & Thong, K. L. (2009). Characterization of multidrug resistant ESBL-producing *Escherichia coli* isolates from hospitals in Malaysia. *BioMed Research International*, 2009:165637.
- Little, C. L., Richardson, J. F., Owen, R. J., De Pinna, E., & Threlfall, E. J. (2008). *Campylobacter* and *Salmonella* in raw red meats in the United Kingdom: prevalence, characterization and antimicrobial resistance pattern, 2003–2005. *Food Microbiology*, 25(3), 538-543.

- Livermore, D. M. (1995). β -Lactamases in laboratory and clinical resistance. *Clinical Microbiology Reviews*, 8(4), 557-584.
- Livermore, D. M. (2008). Defining an extended-spectrum β -lactamase. *Clinical Microbiology and Infection*, 14(s1), 3-10.
- Livermore, D. M., & Woodford, N. (2006). The β -lactamase threat in Enterobacteriaceae, Pseudomonas and Acinetobacter. *Trends in Microbiology*, 14(9), 413-420.
- Livermore, D. M., Struelens, M., Amorim, J., Baquero, F., Bille, J., Canton, R., Henning, S., Gatermann, S., Marchese, A., Mittermayer, H., & Nonhoff, C. (2002). Multicentre evaluation of the VITEK 2 Advanced Expert System for interpretive reading of antimicrobial resistance tests. *Journal of Antimicrobial Chemotherapy*, 49(2), 289-300.
- Lu, P. L., Liu, Y. C., Toh, H. S., Lee, Y. L., Liu, Y. M., Ho, C. M., Huang, C.C., Liu, C.E., Ko, W.C., Wang, J.H., & Tang, H.J. (2012). Epidemiology and antimicrobial susceptibility profiles of Gram-negative bacteria causing urinary tract infections in the Asia-Pacific region: 2009–2010 results from the Study for Monitoring Antimicrobial Resistance Trends (SMART). *International Journal of Antimicrobial Agents*, 40, S37-S43.
- Lübbert, C., Straube, L., Stein, C., Makarewicz, O., Schubert, S., Mössner, J., Pletz, M.W., & Rodloff, A. C. (2015). Colonization with extended-spectrum beta-lactamase-producing and carbapenemase-producing Enterobacteriaceae in international travelers returning to Germany. *International Journal of Medical Microbiology*, 305(1), 148-156.
- Lukac, P. J., Bonomo, R. A., & Logan, L. K. (2015). Extended-Spectrum β -Lactamase-Producing Enterobacteriaceae in Children: Old Foe, Emerging Threat. *Clinical Infectious Diseases*, 60(9), 1389-1397.
- Mabilat, C., & Courvalin, P. (1990). Development of " oligotyping" for characterization and molecular epidemiology of TEM beta-lactamases in members of the family Enterobacteriaceae. *Antimicrobial Agents and Chemotherapy*, 34(11), 2210-2216.
- Magiorakos, A. P., Srinivasan, A., Carey, R. B., Carmeli, Y., Falagas, M. E., Giske, C. G., Harbarth, S., Hindler, J.F., Kahlmeter, G., Olsson-Liljequist, B. & Paterson, D. L. (2012). Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clinical Microbiology and Infection*, 18(3), 268-281.
- Manges, A. R., & Johnson, J. R. (2012). Foodborne origins of *Escherichia coli* causing extraintestinal infections. *Clinical Infectious Diseases*, 55(5) 712-719.

- Mather, A. E., Reid, S. W. J., McEwen, S. A., Ternent, H. E., Reid-Smith, R. J., Boerlin, P., Taylor, D.J., Steele, W.B., Gunn, G.J., & Mellor, D. J. (2008). Factors associated with cross-contamination of hides of Scottish cattle by *Escherichia coli* O157. *Applied and Environmental Microbiology*, 74(20), 6313-6319.
- McEwen, S. A., & Fedorka-Cray, P. J. (2002). Antimicrobial use and resistance in animals. *Clinical Infectious Diseases*, 34(Supplement 3), S93-S106.
- Medeiros, A. A. (1997). Evolution and dissemination of β -lactamases accelerated by generations of β -lactam antibiotics. *Clinical Infectious Diseases*, 24(Supplement 1), S19-S45.
- Medeiros, M. A. N., Oliveira, D. C. N. D., Rodrigues, D. D. P., & Freitas, D. R. C. D. (2011). Prevalence and antimicrobial resistance of *Salmonella* in chicken carcasses at retail in 15 Brazilian cities. *Revista Panamericana de Salud Pública*, 30(6), 555-560.
- Menon, T., Bindu, D., Kumar, C., Nalini, S., & Thirunarayan, M. A. (2006). Comparison of double disc and three dimensional methods to screen for ESBL producers in a tertiary care hospital. *Indian Journal of Medical Microbiology*, 24(2), 117.
- Meyer, E., Gastmeier, P., Kola, A., & Schwab, F. (2012). Pet animals and foreign travel are risk factors for colonisation with extended-spectrum β -lactamase-producing *Escherichia coli*. *Infection*, 40(6), 685-687.
- Miranda, J. M., Guarddon, M., Vázquez, B. I., Fente, C. A., Barros-Velazquez, J., Cepeda, A., & Franco, C. M. (2008). Antimicrobial resistance in Enterobacteriaceae strains isolated from organic chicken, conventional chicken and conventional turkey meat: A comparative survey. *Food Control*, 19(4), 412-416.
- Moore, G., Blair, I. S., & McDowell, D. A. (2007). Recovery and transfer of *Salmonella typhimurium* from four different domestic food contact surfaces. *Journal of Food Protection*, 70(10), 2273-2280.
- Morgan, D. J., Okeke, I. N., Laxminarayan, R., Perencevich, E. N., & Weisenberg, S. (2011). Non-prescription antimicrobial use worldwide: a systematic review. *The Lancet Infectious Diseases*, 11(9), 692-701.
- Mugnier, P., Dubroux, P., Casin, I., Arlet, G., & Collatz, E. (1996). A TEM-derived extended-spectrum beta-lactamase in *Pseudomonas aeruginosa*. *Antimicrobial Agents and Chemotherapy*, 40(11), 2488-2493.
- M'Zali, F. H., Heritage, J., Gascoyne-Binzi, D. M., Snelling, A. M., & Hawkey, P. M. (1998). PCR single strand conformational polymorphism can be used to detect the gene encoding SHV-7 extended-spectrum beta-lactamase and to identify different SHV genes within the same strain. *Journal of Antimicrobial Chemotherapy*, 41(1), 123-125.

- Naas, T., Poirel, L., & Nordmann, P. (2008). Minor extended-spectrum β -lactamases. *Clinical Microbiology and Infection*, 14(s1), 42-52.
- Naas, T., Poirel, L., Karim, A., & Nordmann, P. (1999). Molecular characterization of In50, a class 1 integron encoding the gene for the extended-spectrum β -lactamase VEB-1 in *Pseudomonas aeruginosa*. *FEMS Microbiology Letters*, 176(2), 411-419.
- Nagarajan, R., Boeck, L. D., Gorman, M., Hamill, R. L., Higgens, C. E., Hoehn, M. M., Stark, W.M., & Whitney, J. G. (1971). beta-Lactam antibiotics from Streptomyces. *Journal of the American Chemical Society*, 93(9), 2308-2310.
- Nedbalcova, K., Nechvatalova, K., Pokludova, L., Bures, J., Kucerova, Z., Koutecka, L., & Hera, A. (2014). Resistance to selected beta-lactam antibiotics. *Veterinary Microbiology*, 171(3), 328-336.
- Nordmann, P., Ronco, E., Naas, T., Duport, C., Michel-Briand, Y., & Labia, R. (1993). Characterization of a novel extended-spectrum beta-lactamase from *Pseudomonas aeruginosa*. *Antimicrobial Agents and Chemotherapy*, 37(5), 962-969.
- Nüesch-Inderbinen, M. T., Hächler, H., & Kayser, F. H. (1996). Detection of genes coding for extended-spectrum SHV beta-lactamases in clinical isolates by a molecular genetic method, and comparison with the E test. *European Journal of Clinical Microbiology and Infectious Diseases*, 15(5), 398-402.
- Nugent, M. E., & Hedges, R. W. (1979). The nature of the genetic determinant for the SHV-1 β -lactamase. *Molecular and General Genetics MGG*, 175(3), 239-243.
- Ohnishi, M., Golparian, D., Shimuta, K., Saika, T., Hoshina, S., Iwasaku, K., Nakayama, S.I., Kitawaki, J., & Unemo, M. (2011). Is *Neisseria gonorrhoeae* initiating a future era of untreatable gonorrhea? Detailed characterization of the first strain with high-level resistance to ceftriaxone. *Antimicrobial Agents and Chemotherapy*, 55(7), 3538-3545.
- Ouellette, M., Paul, G. C., Philippon, A. M., & Roy, P. H. (1988). Oligonucleotide probes (TEM-1, OXA-1) versus isoelectric focusing in beta-lactamase characterization of 114 resistant strains. *Antimicrobial Agents and Chemotherapy*, 32(3), 397-399.
- Overdevest, I., Willemsen, I., Rijnsburger, M., Eustace, A., Xu, L., Hawkey, P., Heck, M., Savelkoul, P., Vandebroucke-Grauls, C., van der Zwaluw, K., & Huijsdens, X. (2011). Extended-spectrum β -lactamase genes of *Escherichia coli* in chicken meat and humans, The Netherlands. *Emerging Infectious Diseases*, 17(7), 1216-1222.
- Paphitou, N. I. (2013). Antimicrobial resistance: action to combat the rising microbial challenges. *International Journal of Antimicrobial Agents*, 42, S25-S28.

- Paterson, D. L., & Bonomo, R. A. (2005). Extended-spectrum β -lactamases: a clinical update. *Clinical Microbiology Reviews*, 18(4), 657-686.
- Paterson, D. L., Egea, P., Pascual, A., López-Cerero, L., Navarro, M. D., Adams-Haduch, J. M., Qureshi, Z.A., Sidjabat, H.E., & Rodríguez-Baño, J. (2010). Extended-spectrum and CMY-type β -lactamase-producing *Escherichia coli* in clinical samples and retail meat from Pittsburgh, USA and Seville, Spain. *Clinical Microbiology and Infection*, 16(1), 33-38.
- Paterson, D. L., Ko, W. C., Von Gottberg, A., Mohapatra, S., Casellas, J. M., Goossens, H., Mulazimoglu, L., Trenholme, G., Klugman, K.P., Bonomo, R.A., & Victor, L. Y. (2004). Antibiotic therapy for *Klebsiella pneumoniae* bacteremia: implications of production of extended-spectrum β -lactamases. *Clinical Infectious Diseases*, 39(1), 31-37.
- Peirano, G., & Pitout, J. D. (2010). Molecular epidemiology of *Escherichia coli* producing CTX-M β -lactamases: the worldwide emergence of clone ST131 O25: H4. *International Journal of Antimicrobial Agents*, 35(4), 316-321.
- Perez, F., Endimiani, A., Hujer, K. M., & Bonomo, R. A. (2007). The continuing challenge of ESBLs. *Current Opinion in Pharmacology*, 7(5), 459-469.
- Pérez-Rodríguez, F., Castro, R., Posada-Izquierdo, G. D., Valero, A., Carrasco, E., García-Gimeno, R. M., & Zurera, G. (2010). Evaluation of hygiene practices and microbiological quality of cooked meat products during slicing and handling at retail. *Meat Science*, 86(2), 479-485.
- Pfaller, M. A., & Segreti, J. (2006). Overview of the epidemiological profile and laboratory detection of extended-spectrum β -Lactamases. *Clinical Infectious Diseases*, 42(Supplement 4), S153-S163.
- Philippe, L., Franil, T., Patricia, B., George, G. Z., & Daryl, J. H. (2010) Evaluation of a chromogenic medium for extended-spectrum beta-lactamase (ESBL)-producing Enterobacteriaceae. University of Manitoba, Canada. ECCMID, Vienna, Austria 10 – 13 April 2010.
- Poirel, L., Naas, T., & Nordmann, P. (2008). Genetic support of extended-spectrum β -lactamases. *Clinical Microbiology and Infection*, 14(s1), 75-81.
- Poirel, L., Naas, T., Guibert, M., Chaibi, E. B., Labia, R., & Nordmann, P. (1999). Molecular and biochemical characterization of VEB-1, a novel class A extended-spectrum β -lactamase encoded by an *Escherichia coli* integron gene. *Antimicrobial Agents and Chemotherapy*, 43(3), 573-581.
- Poole, K. (2004). Resistance to β -lactam antibiotics. *Cellular and Molecular Life Sciences CMLS*, 61(17), 2200-2223.
- Popham, D. L., & Young, K. D. (2003). Role of penicillin-binding proteins in bacterial cell morphogenesis. *Current Opinion in Microbiology*, 6(6), 594-599.

- Rahme, C., Butterfield, J. M., Nicasio, A. M., & Lodise, T. P. (2014). Dual beta-lactam therapy for serious Gram-negative infections: is it time to revisit? *Diagnostic Microbiology and Infectious Disease*, 80(4), 239-259.
- Rammelkamp, C. H., & Maxon, T. (1942). Resistance of *Staphylococcus aureus* to the Action of Penicillin. *Experimental Biology and Medicine*, 51(3), 386-389.
- Rawat, D., & Nair, D. (2010). Extended-spectrum β -lactamases in Gram negative bacteria. *Journal of Global Infectious Diseases*, 2(3), 263.
- Reich, F., Schill, F., Atanassova, V., & Klein, G. (2016). Quantification of ESBL-*Escherichia coli* on broiler carcasses after slaughtering in Germany. *Food Microbiology*, 54, 1-5.
- Reuland, E. A., Overdevest, I. T. M. A., Al Naiemi, N., Kalpoe, J. S., Rijnsburger, M. C., Raadsen, S. A., Ligtenberg-Burgman, I., van der Zwaluw, K.W., Heck, M., Savelkoul, P.H.M., & Vandebroucke-Grauls, C. M. J. E. (2013). High prevalence of ESBL-producing Enterobacteriaceae carriage in Dutch community patients with gastrointestinal complaints. *Clinical Microbiology and Infection*, 19(6), 542-549.
- Roca, I., Akova, M., Baquero, F., Carlet, J., Cavaleri, M., Coenen, S., Cohen, J., Findlay, D., Gyssens, I., Heure, O.E. and Kahlmeter, G., & Vila, J. (2015). The global threat of antimicrobial resistance: science for intervention. *New Microbes and New Infections*, 6: 22–29.
- Roess, A. A., Winch, P. J., Ali, N. A., Akhter, A., Afroz, D., El Arifeen, S., Darmstadt, G.L., Baqui, A.H., & Bangladesh PROJAHNMO Study Group. (2013). Animal husbandry practices in rural Bangladesh: potential risk factors for antimicrobial drug resistance and emerging diseases. *The American Journal of Tropical Medicine and Hygiene*, 89(5), 965-970.
- Rosenau, A., Cattier, B., Gousset, N., Harria, P., Philippon, A., & Quentin, R. (2000). *Capnocytophaga ochracea*: characterization of a plasmid-encoded extended-spectrum TEM-17 β -lactamase in the phylum Flavobacter-Bacteroides. *Antimicrobial Agents and Chemotherapy*, 44(3), 760-762.
- Rossolini, G. M., Franceschini, N., Lauretti, L., Caravelli, B., Riccio, M. L., Galleni, M., Frère, J.M., & Amicosante, G. (1999). Cloning of a *Chryseobacterium* (*Flavobacterium*) *meningosepticum* Chromosomal Gene (blaA CME) Encoding an Extended-Spectrum Class A β -Lactamase Related to the Bacteroides Cephalosporinases and the VEB-1 and PER β -Lactamases. *Antimicrobial agents and Chemotherapy*, 43(9), 2193-2199.
- Rottier, W. C., Ammerlaan, H. S., & Bonten, M. J. (2012). Effects of confounders and intermediates on the association of bacteraemia caused by extended-spectrum β -lactamase-producing Enterobacteriaceae and patient outcome: a meta-analysis. *Journal of antimicrobial chemotherapy*, 67(6), 1311-1320.

- Rupp, M. E., & Fey, P. D. (2003). Extended spectrum β -lactamase (ESBL)-producing Enterobacteriaceae. *Drugs*, 63(4), 353-365.
- Samanta, I., Joardar, S. N., Das, P. K., & Sar, T. K. (2015). Comparative possession of Shiga toxin, intimin, enterohaemolysin and major extended spectrum beta lactamase (ESBL) genes in *Escherichia coli* isolated from backyard and farmed poultry. *Iranian Journal of Veterinary Research*, 16(1), 90-93.
- Sanders, C. C., Barry, A. L., Washington, J. A., Shubert, C., Moland, E. S., Traczewski, M. M., Knapp, C., & Mulder, R. (1996). Detection of extended-spectrum-beta-lactamase-producing members of the family Enterobacteriaceae with Vitek ESBL test. *Journal of Clinical Microbiology*, 34(12), 2997-3001.
- Sanders, C. C., Peyret, M., Moland, E. S., Shubert, C., Thomson, K. S., Boeufgras, J. M., & Sanders, W. E. (2000). Ability of the VITEK 2 Advanced Expert System to Identify β -Lactam Phenotypes in Isolates of Enterobacteriaceae and *Pseudomonas aeruginosa*. *Journal of Clinical Microbiology*, 38(2), 570-574.
- Sanderson, M. W., Sargeant, J. M., Renter, D. G., Griffin, D. D., & Smith, R. A. (2005). Factors associated with the presence of coliforms in the feed and water of feedlot cattle. *Applied and Environmental Microbiology*, 71(10), 6026-6032.
- Sanguinetti, M., Posteraro, B., Spanu, T., Ciccaglione, D., Romano, L., Fiori, B., Nicoletti, G., Zanetti, S., & Fadda, G. (2003). Characterization of clinical isolates of Enterobacteriaceae from Italy by the BD Phoenix extended-spectrum β -lactamase detection method. *Journal of Clinical Microbiology*, 41(4), 1463-1468.
- Schwaber, M. J., Navon-Venezia, S., Kaye, K. S., Ben-Ami, R., Schwartz, D., & Carmeli, Y. (2006). Clinical and economic impact of bacteremia with extended-spectrum- β -lactamase-producing Enterobacteriaceae. *Antimicrobial agents and chemotherapy*, 50(4), 1257-1262.
- Schwarz, S., & Chaslus-Dancla, E. (2001). Use of antimicrobials in veterinary medicine and mechanisms of resistance. *Veterinary Research*, 32(3-4), 201-225.
- Sekawi, Z., Yusof, R., & Shamsudin, M. N. (2008). Extended-Spectrum β -Lactamases-Producing *Escherichia coli* from a Tertiary Hospital in Malaysia: Emergence of CTX-M-Type β -Lactamases Variation. *Research Journal of Microbiology*, 3(6), 489-493.
- Selangor: www.2malaysia.com/selangor. Accessed 04-06-2015.
- Shaikh, S., Fatima, J., Shakil, S., Rizvi, S. M. D., & Kamal, M. A. (2015). Antibiotic resistance and extended spectrum beta-lactamases: Types, epidemiology and treatment. *Saudi Journal of Biological Sciences*, 22(1), 90-101.

- Sharp, H., Valentin, L., Fischer, J., Guerra, B., Appel, B., & Kaesbohrer, A. (2013). Estimation of the transfer of ESBL-producing *Escherichia coli* to humans in Germany. *Berliner und Munchener Tierarztliche Wochenschrift*, 127(11-12), 464-477.
- Shi, H., Yang, Y., Liu, M., Yan, C., Yue, H., & Zhou, J. (2014). Occurrence and distribution of antibiotics in the surface sediments of the Yangtze Estuary and nearby coastal areas. *Marine Pollution Bulletin*, 83(1), 317-323.
- Shorr, A. F. (2007). Epidemiology of staphylococcal resistance. *Clinical Infectious Diseases*, 45(Supplement 3), S171-S176
- Silbergeld, E. K., Graham, J., & Price, L. B. (2008). Industrial food animal production, antimicrobial resistance, and human health. *Annu. Rev. Public Health*, 29, 151-169.
- Singh, A., Goering, R. V., Simjee, S., Foley, S. L., & Zervos, M. J. (2006). Application of molecular techniques to the study of hospital infection. *Clinical Microbiology Reviews*, 19(3), 512-530.
- Sjölund-Karlsson, M., Howie, R. L., Blickenstaff, K., Boerlin, P., Ball, T., Chalmers, G., Duval, B., Haro, J., Rickert, R., Zhao, S., & Fedorka-Cray, P.J. (2013). Occurrence of β -Lactamase Genes among Non-Typhi *Salmonella enterica* Isolated from Humans, Food Animals, and Retail Meats in the United States and Canada. *Microbial Drug Resistance*, 19(3), 191-197.
- Soilleux, M. J., Morand, A. M., Arlet, G. J., Scavizzi, M. R., & Labia, R. (1996). Survey of *Klebsiella pneumoniae* producing extended-spectrum beta-lactamases: prevalence of TEM-3 and first identification of TEM-26 in France. *Antimicrobial Agents and Chemotherapy*, 40(4), 1027-1029.
- Solà-Ginés, M., González-López, J. J., Cameron-Veas, K., Piedra-Carrasco, N., Cerdà-Cuéllar, M., & Migura-Garcia, L. (2015). Houseflies (*Musca domestica*) as Vectors for Extended-Spectrum β -Lactamase-Producing *Escherichia coli* on Spanish Broiler Farms. *Applied and Environmental Microbiology*, 81(11), 3604-3611.
- Songserm, T., Jam-on, R., Sae-Heng, N., Meemak, N., Hulse-Post, D. J., Sturm-Ramirez, K. M., & Webster, R. G. (2006). Domestic ducks and H5N1 influenza epidemic, Thailand. *Emerging Infectious Diseases*, 12(4), 575.
- Spink, W. W., Ferris, V., & Vivino, J. J. (1944). Comparative in vitro resistance of staphylococci to penicillin and to sodium sulfathiazole. *Experimental Biology and Medicine*, 55(3), 207-210.
- Stuart, J. C., van den Munckhof, T., Voets, G., Scharringa, J., Fluit, A., & Leverstein-Van Hall, M. (2012). Comparison of ESBL contamination in organic and conventional retail chicken meat. *International Journal of Food Microbiology*, 154(3), 212-214.

- Subramaniam, G., Palasubramaniam, S., & Navaratnam, P. (2006). SHV-5 extended-spectrum beta-lactamases in clinical isolates of *Escherichia coli* in Malaysia. *Indian Journal of Medical Microbiology*, 24(3), 205-207.
- Suzuki, S., Horinouchi, T., & Furusawa, C. (2015). Suppression of antibiotic resistance acquisition by combined use of antibiotics. *Journal of Bioscience and Bioengineering*, 120 (4), 467-469.
- Sykes, R. B., Cimarusti, C. M., Bonner, D. P., Bush, K., Floyd, D. M., Georgopapadakou, N. H., Koster, W.H., Liu, W.C., Parker, W.L., Principe, P.A., & Rathnum, M.L. (1981). Monocyclic β -lactam antibiotics produced by bacteria. *Nature*, 291 (5815), 489-491.
- Tacão, M., Moura, A., Correia, A., & Henriques, I. (2014). Co-resistance to different classes of antibiotics among ESBL-producers from aquatic systems. *Water Research*, 48, 100-107.
- Tang, J. Y. H., Nishibuchi, M., Nakaguchi, Y., Ghazali, F. M., Saleha, A. A., & Son, R. (2011). Transfer of *Campylobacter jejuni* from raw to cooked chicken via wood and plastic cutting boards. *Letters in Applied Microbiology*, 52(6), 581-588.
- Tawfik, A. F., Alswailem, A. M., Shibli, A. M., & Al-Agamy, M. H. (2011). Prevalence and genetic characteristics of TEM, SHV, and CTX-M in clinical *Klebsiella pneumoniae* isolates from Saudi Arabia. *Microbial Drug Resistance*, 17(3), 383-388.
- Tenover, F. C. (2006). Mechanisms of antimicrobial resistance in bacteria. *The American Journal of Medicine*, 119(6), S3-S10.
- Teuber, M. (2001). Veterinary use and antibiotic resistance. *Current Opinion in Microbiology*, 4(5), 493-499.
- Thomson, K. S., & Sanders, C. C. (1992). Detection of extended-spectrum beta-lactamases in members of the family Enterobacteriaceae: comparison of the double-disk and three-dimensional tests. *Antimicrobial Agents and Chemotherapy*, 36(9), 1877-1882.
- Thong K. L., Lai K. S., & Goh, Y. L. (2003). Technical improvement in DNA preparation in pulsed-field gel electrophoresis. *Malaysian Journal of Science*, 22:55-9.
- Tzelepi, E., Giakkoupi, P., Sofianou, D., Loukova, V., Kemeroglou, A., & Tsakris, A. (2000). Detection of Extended-Spectrum β -Lactamases in Clinical Isolates of *Enterobacter cloacae* and *Enterobacter aerogenes*. *Journal of Clinical Microbiology*, 38(2), 542-546.
- Tzouvelekis, L. S., & Bonomo, R. A. (1999). SHV-type β -lactamases. *Current Pharmaceutical Design*, 5(11), 847-864.

- Tzouvelekis, L. S., Tzelepi, E., Tassios, P. T., & Legakis, N. J. (2000). CTX-M-type β -lactamases: an emerging group of extended-spectrum enzymes. *International Journal of Antimicrobial Agents*, 14(2), 137-142.
- Umgelter, A., Reindl, W., Miedaner, M., Schmid, R. M., & Huber, W. (2009). Failure of current antibiotic first-line regimens and mortality in hospitalized patients with spontaneous bacterial peritonitis. *Infection*, 37(1), 2-8.
- USDA | International Egg and Poultry Review (2014). 17 (14) ISSN 1522-5100 <http://search.ams.usda.gov/MNDMS/2014/04/PY20140408WIntlPoultryandEgg.pdf>. updated: April 08, 2014. Accessed 20-01-2016.
- Usha, M. R., Maarof, F., Robin, T., Chai, L. C., Cheah, Y. K., Mohamad Ghazali, F., & Radu, S. (2010). Occurrence and antibiotic resistance of *Campylobacter jejuni* and *Campylobacter coli* in retail broiler chicken. *International Food Research Journal*, 17(2), 247-255.
- Vahaboglu, H., Oztürk, R., Aygün, G., Coşkunkan, F., Yaman, A., Kaygusuz, A., Leblebicioglu, H., Balik, I., Aydin, K., & Otkun, M. (1997). Widespread detection of PER-1-type extended-spectrum beta-lactamases among nosocomial *Acinetobacter* and *Pseudomonas aeruginosa* isolates in Turkey: a nationwide multicenter study. *Antimicrobial Agents and Chemotherapy*, 41(10), 2265–2269.
- Van Boeckel, T. P., Brower, C., Gilbert, M., Grenfell, B. T., Levin, S. A., Robinson, T. P., Teillant, A., & Laxminarayan, R. (2015). Global trends in antimicrobial use in food animals. *Proceedings of the National Academy of Sciences*, 112(18), 5649-5654.
- Van den Bogaard, A. E., London, N., Driessen, C., & Stobberingh, E. E. (2001). Antibiotic resistance of faecal *Escherichia coli* in poultry, poultry farmers and poultry slaughterers. *Journal of Antimicrobial Chemotherapy*, 47(6), 763-771.
- Vercauteren, E., Descheemaeker, P., Ieven, M., Sanders, C. C., & Goossens, H. (1997). Comparison of screening methods for detection of extended-spectrum beta-lactamases and their prevalence among blood isolates of *Escherichia coli* and *Klebsiella* spp. in a Belgian teaching hospital. *Journal of Clinical Microbiology*, 35(9), 2191-2197.
- Villegas, M. V., Kattan, J. N., Quinteros, M. G., & Casellas, J. M. (2008). Prevalence of extended-spectrum β -lactamases in South America. *Clinical Microbiology and Infection*, 14(s1), 154-158.
- Visschers, V. H., Iten, D. M., Riklin, A., Hartmann, S., Sidler, X., & Siegrist, M. (2014). Swiss pig farmers' perception and usage of antibiotics during the fattening period. *Livestock Science*, 162, 223-232.
- von Salviati, C., Laube, H., Guerra, B., Roesler, U., & Friese, A. (2015). Emission of ESBL/AmpC-producing *Escherichia coli* from pig fattening farms to surrounding areas. *Veterinary Microbiology*, 175(1), 77-84.

- Vondruskova, H., Slamova, R., Trckova, M., Zraly, Z., & Pavlik, I. (2010). Alternatives to antibiotic growth promoters in prevention of diarrhoea in weaned piglets: a review. *Veterinarni Medicina*, 55(5), 199-224.
- Warren, R. E., Ensor, V. M., O'neill, P., Butler, V., Taylor, J., Nye, K., Harvey, M., Livermore, D.M., Woodford, N., & Hawkey, P.M. (2008). Imported chicken meat as a potential source of quinolone-resistant *Escherichia coli* producing extended-spectrum β-lactamases in the UK. *Journal of Antimicrobial Chemotherapy*, 61(3), 504-508.
- Wayne PA: Clinical and Laboratory Standards Institute. 2010. Performance standards for antimicrobial disk susceptibility tests. Approved standard, 10th ed. CLSI document M02-A10.
- Wells, J. S., Trejo, W. H., Principe, P. A., Bush, K., Georgopapadakou, N., Bonner, D. P., & Sykes, R. B. (1982). EM5400, a family of monobactam antibiotics produced by *Agrobacterium radiobacter*. I. Taxonomy, fermentation and biological properties. *The Journal of Antibiotics*, 35(3), 295-299.
- Winokur, P. L., Canton, R., Casellas, J. M., & Legakis, N. (2001). Variations in the prevalence of strains expressing an extended-spectrum β-lactamase phenotype and characterization of isolates from Europe, the Americas, and the Western Pacific region. *Clinical Infectious Diseases*, 32(Supplement 2), S94-S103.
- Wittum, T. E. (2012). The challenge of regulating agricultural ceftiofur use to slow the emergence of resistance to extended-spectrum cephalosporins. *Applied and Environmental Microbiology*, 78(22), 7819-7821.
- World Health Organization (2009) Report of the 1st Meeting of the WHO Advisory Group on Integrated Surveillance of Antimicrobial Resistance. Copenhagen, 15-19 June 2009. (AGISAR) (Part I: 3rd Edition WHO List on Critically Important Antimicrobials For Human Health Part II: Strategic Framework for WHO Activities on Integrated Antimicrobial Resistance Surveillance). <http://apps.who.int/medicinedocs/en/d/Js16735e/>. Last updated: April 14, 2016. Accessed 20-06-2016.
- World Health Organization (2014) Antimicrobial resistance: global report on surveillance. <http://www.who.int/drugresistance/documents/surveillancereport/en/> Last updated: April 14, 2014. ISBN: 978 92 4 156474 8, Accessed 10-06-2016.
- WPR: World Poultry Report (2014). Malaysian poultry industry meeting domestic demand <http://www.worldpoultry.net/Broilers/Markets--Trade/2014/3/Malaysian-poultry-industry-meeting-domestic-demand-1486503W> updated:Mar 9, 2016 Accessed 10-04-2016.
- Yuan, L., Liu, J. H., Hu, G. Z., Pan, Y. S., Liu, Z. M., Mo, J., & Wei, Y. J. (2009). Molecular characterization of extended-spectrum β-lactamase-producing *Escherichia coli* isolates from chickens in Henan Province, China. *Journal of Medical Microbiology*, 58(11), 1449-1453.

- Zamberi Sekawi, Rusmah Yusof and Mariana Nor Shamsudin. (2008) Extended-spectrum β -lactamases-producing *Escherichia coli* from a tertiary hospital in Malaysia: Emergence of CTX-M-Type β -lactamases variation. *Research Journal of Microbiology*. 3(6): 489-493.
- Zhang, X. X., Zhang, T., & Fang, H. H. (2009). Antibiotic resistance genes in water environment. *Applied Microbiology and Biotechnology*, 82(3), 397-414.
- Zhao, L., Dong, Y. H., & Wang, H. (2010). Residues of veterinary antibiotics in manures from feedlot livestock in eight provinces of China. *Science of the Total Environment*, 408(5), 1069-1075.
- Zheng, H., Zeng, Z., Chen, S., Liu, Y., Yao, Q., Deng, Y., Chen, X., Lv, L., Zhuo, C., Chen, Z., & Liu, J. H. (2012). Prevalence and characterisation of CTX-M β -lactamases amongst *Escherichia coli* isolates from healthy food animals in China. *International Journal of Antimicrobial Agents*, 39(4), 305-310.
- Zhu, J., Wang, Y., Song, X., Cui, S., Xu, H., Yang, B., Huang, J., Liu, G., Chen, Q., Zhou, G., & Chen, Q. (2014). Prevalence and quantification of *Salmonella* contamination in raw chicken carcasses at the retail in China. *Food Control*, 44, 198-202.
- Zou, W., Chen, H. C., Hise, K. B., Tang, H., Foley, S. L., Meehan, J., Lin, W.J., Nayak, R., Xu, J., Fang, H., & Chen, J. J. (2013). Meta-analysis of pulsed-field gel electrophoresis fingerprints based on a constructed *Salmonella* database. *PloS one*, 8(3), e59224.
- Zurfluh, K., Jakobi, G., Stephan, R., Hächler, H., & Nüesch-Inderbinen, M. (2014). Replicon typing of plasmids carrying blaCTX-M-1 in Enterobacteriaceae of animal, environmental and human origin. *Frontiers in Microbiology*, 5, 555. doi:10.3389/fmicb.2014.00555.