



UNIVERSITI PUTRA MALAYSIA

***EFFICACY OF A PROTOTYPE VACCINE FOR CASEOUS
LYMPHADENITIS DISEASE IN GOATS***

WESSAM MONTHER MOHAMMED SALEH

FPV 2016 38



**EFFICACY OF A PROTOTYPE VACCINE FOR CASEOUS
LYMPHADENITIS DISEASE IN GOATS**

By

WESSAM MONTHER MOHAMMED SALEH

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia in Fulfillment of the Requirements of the Degree of Doctor of
Philosophy**

November 2016

All material contained in this thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the expression, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

To my parents: for them fearless spirits; thank you for unconditional support with my life, I am honored to have you as my parents, thank you for giving me a chance to prove and improve myself through all my walk of life. I love you.

To my beloved wife, wonderful children: thank you for believing in me, for allowing me; for allowing me to further my studies. Please do not ever doubt my dedication and love for you.

To my brothers and sisters: Hoping that with this research I have proven to you that there is no mountain higher as long as God is on our side. Hoping that, you will walk again and be able to fulfill your dreams.

To my teachers, instructors and mentors: my sincere thanks to make it reality, and utmost respect to all my teachers past, present and future, and to all teachers everywhere.

To my friends: I also dedicate this dissertation and give special thanks to my many friends who have supported me throughout this process.

I would like to conclude by again expressing my deepest gratitude and love to all.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

EFFICACY OF A PROTOTYPE VACCINE FOR CASEOUS LYMPHADENITIS DISEASE IN GOATS

By

WESSAM MONTHER MOHAMMED SALEH

November 2016

Chairman : Associate Professor Faez Firdaus Jesse Abdullah, PhD
Faculty : Veterinary Medicine

Corynebacterium pseudotuberculosis is bacterium responsible for caseous lymphadenitis (CLA), a disease characterized by the formation of suppurative abscesses, particularly in superficial and internal lymph nodes within almost all internal organs of sheep, goats and other species including human. Due to the chronic and subclinical nature of CLA, its control is difficult and hence records a high worldwide prevalence with significant economic impact. In Malaysia, CLA is a potential threat to the livestock industry, details of the immune responses against *C. pseudotuberculosis* infection are not completely understood and effective vaccine is still lacking.

This study was conducted using a local Malaysian *C. pseudotuberculosis* isolate, a murine and caprine experimental model testing the capacity of two concentrations of a prototype vaccine to protect mice and goats against experimental challenge with *C. pseudotuberculosis* were determined. Forty healthy female mice, aged between 3-4 weeks and weight between 15.5-20g were randomly divided in to four equal groups consisting of 10 mice each, designated groups A, B, C and D. Mice in groups A and B were inoculated intraperitoneally with 200µl of the prepared 0.5%, 1% formalin-killed vaccine of local *C. pseudotuberculosis* strain respectively. Groups C and D were kept as positive and negative control respectively, and inoculated intraperitoneally only with 200µl of PBS pH 7.4. Booster doses were inoculated to the mice in groups A and B after 21 days of the initial doses. Week 6th of post inoculation of the first vaccination dose; mice in all groups except negative control group were inoculated intraperitoneally with 200µl of virulent type of *C. pseudotuberculosis* 10⁶ CFU per mice and were observed. All mice were euthanized at day 10 post challenged, and post mortem examination were carried on the liver, lungs, heart, kidneys, spleen and lymph nodes, while tissue samples of same organs were fixed for histopathological examination and further cultured on blood agar for bacterial isolation and identification.

For goat model, 24 adult female goats aged between 11-13 months, weighing 23-30 kg, with no history of vaccination against CLA were randomly divided into 4 groups consisting of 6 goats each as A, B, C and D. Goats in groups A and B were vaccinated, respectively, with 1 ml intramuscular injection of the 0.5% and 1% formalin-killed *C. pseudotuberculosis* vaccine. Groups C and D were kept, respectively, as positive and negative control, and inoculated intramuscularly only with 1 ml of oil-in-water emulsion adjuvant. Booster dose was inoculated after 21 of the initial treatment. 2 weeks of post-booster dose, the goats (except in group D) were challenged with 2ml of 10^6 CFU of wild isolate of *C. pseudotuberculosis* subcutaneously. They were monitored for the entire experiment for clinical signs, and blood samples were collected at predetermined intervals into plain and EDTA tubes for IgM, IgG, IgA, Haptoglobin, Serum Amyloid A and hematology analysis. At the end of the study, the goats were sacrificed and post mortem were carried out and samples such as external and internal lymph nodes, liver, lungs, heart, kidneys, spleen, intestine, spinal cord and brain were collected for histopathological examination and for bacterial isolation and identification.

Mice in group C which were inoculated only with *C. pseudotuberculosis* showed significant ($P < 0.05$) changes of clinical signs. Moderate to severe ruffled fur, reduced rate of movement, decreased responsiveness and ocular discharge were observed. Signs such as huddling together, defection, loss of appetite, pasty feces and respiratory distress were also recorded. In post mortem examination, congestion and abscesses in the site of inoculation, spleen, liver and kidneys were recorded in group C only. Microscopic examination of infected group showed significant ($P < 0.05$) moderate to severe hemorrhage and congestion in all organs, mild to moderate inflammatory cell infiltration and degeneration, multiple abscesses with higher scoring of necrosis and less to mild edema were observed in all organs. In contrast, the results of the vaccinated mice with CLA vaccine showed normal tissues with non-significant histopathological changes. *Corynebacterium pseudotuberculosis* was detected by PCR from most visceral organs and lymph nodes of 9 mice (90%) in control positive group; one mouse was found clear of infection, while it was detected only from 30% and 20% of mice in groups A and B respectively.

For the goat experiment, positive control animals (group C) showed significant clinical symptoms compared to vaccinated groups that observed one week post-bacterial inoculation such as significant raise in body temperature, respiratory and heart rates and sharply decreased in rumen motility. Poor hair coat condition, decrease in body condition scoring and significant enlargement and ruptured of injection sites after 2 to 3 weeks of challenges were also observed. Moreover, a marked enlargement and occasional rupture of the affected lymph nodes were recorded in positive control group 5 to 6 weeks post challenged with *C. pseudotuberculosis*. Slight enlargement of regional lymph nodes near the site bacterial challenged of only 1 goat (16%) in each group A and B was observed.

Both groups exposed to our formulated vaccines (groups A and B) induced significant (for both vaccines) elevations in serum Haptoglobin (Hp) and plasma serum amyloid A (SAA) concentrations which increased after 2, 5, 6 and 7 weeks when compared to non-vaccinated healthy animals of the same conditions. Hp and SAA levels in goats, which had never been exposed to vaccine, were elevated significantly one week after exposure to wild bacteria and maintained sharp increase for the interval of the evaluation. The concentration of serum IgM was found to be significantly increased at week 3 of the exposure to the first dose of vaccination, where 7 and 9-fold increases were observed in groups A and B respectively. Sharp increase up to 18 and 17-fold were observed one week after challenged with viable bacteria in groups A and B respectively.

The levels for the IgM response in group C were significantly higher than the uninfected control goats (group D). The highest concentrations of IgG in group A and B (156.63 ± 2.17 ng/mL) and (164.03 ± 2.92 ng/mL) were observed after two weeks post-exposure to whole viable bacteria, respectively. For Group C, IgG mean concentration reached the maximum level (138.58 ± 1.34 ng/mL) one month after the exposure to whole viable bacteria. However the mean concentration of IgG in the vaccinated groups continued to rise following the primary and secondary vaccination. The goats vaccinated by both the formulated vaccines had no significant elevation of IgA concentration throughout the study period. In contrast, a marked increase of serum IgA levels was observed in group C during the period between 7 to 14 days post infection with viable *C. pseudotuberculosis*. However, only 1.2 and 1.3-fold increases were recorded in group C respectively at weeks 6 and 7.

Post mortem examination of vaccinated groups revealed no significant gross pathological changes in visceral organs and also in the internal and external lymph nodes. On the other hand, high rate of gross pathological changes were recorded in all goats of positive control group. No significant gross pathological changes were observed in the brain and spinal cord of goats in group C. In the visceral organs such as lungs, liver, kidneys, spleen and heart the main gross pathological lesions observed were congestion, hemorrhage, focal areas of necrosis, fatty atrophy and abscessation. Abscess formation was rarely observed in group C except in the few organs such as the liver and lungs. In the case of lymph nodes, high percentages of abscess formation of different sizes were observed in all goats in group C. In contrast, both vaccinated groups did not develop any abscess in the lymph nodes or the visceral organs even though slight enlargements of the lymph nodes were recorded in two goats (16%) only. The histopathological changes which were observed in the positive group include edema, congestion, infiltration of inflammatory cells mainly lymphocytes and macrophages, hemorrhages, degeneration, granulomatous inflammation and caseous necrosis unlike vaccinated goats which showed significant functional hyperplasia in lymphoid tissues without histopathological changes.

In the hematology, there was a statistical significant decline ($P < 0.05$) in Hb concentration, RBC count, MCV value and PCV in group C after exposure to viable bacteria cells. Parallel to that, statistically non-significant elevations ($P > 0.05$) have been observed MCHC in the same group. The WBC counts were the highest during the experimental infection of non-vaccinated goats (group C) compared with those pre-vaccinated groups (A and B). The highest stimulation in segmented neutrophil was reached 7 days post-challenge to live bacteria, when the numbers were ($12.55 \pm 1.05 \times 10^9/L$) in positive control group, ($10.93 \pm 1.55 \times 10^9/L$) and ($10.14 \pm 1.38 \times 10^9/L$) in groups A and B respectively, when compared to ($8.23 \pm 0.46 \times 10^9$) the untreated group (group D). A significant increase ($p < 0.05$) of bands neutrophils count ($0.83 \pm 0.17 \times 10^9/L$) was detected in positive control group after inoculation of *C. pseudotuberculosis*, whereas no significant change was recorded in groups A ($0.49 \pm 0.09 \times 10^9/L$), B ($0.51 \pm 0.13 \times 10^9/L$) and D ($0.30 \pm 0.01 \times 10^9/L$) in the same period. The monocytes, eosinophil and basophiles counts and plasma proteins levels in all blood samples were within the normal range. Nevertheless, no significant change was recorded on the CBC values of both vaccinated groups.

The findings of the PCR showed that five goats (84%) of the six animals in groups A and B pre-exposed to 0.5 % and 1.0% formalin-killed *C. pseudotuberculosis* remained negative. Significantly lower spreading of *C. pseudotuberculosis* DNA was detected in the organs and lymph nodes of the vaccinated groups. The percentage of detection in investigated tissues was found to be 100% in group C.

In conclusion, following experimental challenge with wild type *C. pseudotuberculosis* local isolate, the formalin-killed vaccines were observed to confer statistically significant protection against infection and appeared to significantly restrict the dissemination of challenged bacteria beyond the inoculation site in the 84% of goats. Vaccinated mice (groups A, B) developed humoral immunity and produced 70% and 80% protection respectively against the disease. Moreover, only 16% of the goats immunized with this vaccine manifested mild clinical and pathological infection thus, a potentially important route of disease transmission was eliminated. The results of this study provide information pertinent to the development of an effective caseous lymphadenitis eradication vaccination strategy in Malaysia.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

KEBERKESANAN VAKSIN PROTOTAIP UNTUK PENYALUT NODUS LIMFA PENYAKIT PADA KAMBING

Oleh

WEWSSAM MONTHER MOHAMMED SALEH

November 2016

Pengerusi : Profesor Madya Faez Firdaus Jesse Bin Abdullah, PhD
Fakulti : Perubatan Veterinar

Corynebacterium pseudotuberculosis, bakteria yang bertanggungjawab untuk penyalut nodus limfa (CLA), penyakit yang dicirikan oleh pembentukan abses bernanah, terutamanya dalam nodus limfa luar dan dalaman di dalam hampir semua organ dalaman kambing biri-biri, kambing dan spesies lain termasuk manusia. Oleh kerana sifat kronik dan subklinikal CLA, kawalan adalah sukar dan dengan itu merekodkan kelaziman di seluruh dunia yang tinggi dengan kesan ekonomi yang penting. Di Malaysia, CLA adalah satu ancaman yang berpotensi untuk industri ternakan, terperinci tindak balas imun terhadap jangkitan *C. pseudotuberculosis* tidak difahami sepenuhnya dan vaksin yang berkesan masih kurang.

Kajian ini dijalankan dengan menggunakan *C. pseudotuberculosis* isolate Malaysia yang getir, murine dan berhubung dgn kambing, model eksperimen menguji keupayaan dua vaksin prototaip untuk melindungi tikus dan kambing terhadap cabaran eksperimen dengan *C. pseudotuberculosis* ditentukan. Empat puluh tikus betina yang sihat, berumur antara 3-4 minggu dan berat badan antara 15.5-20g telah dibahagikan secara rawak kepada empat kumpulan yang sama yang terdiri daripada 10 tikus setiap satu, kumpulan ditetapkan A, B, C dan D. Tikus dalam kumpulan A dan B telah disuntik intraperitoneum dengan 200µl daripada bersedia 0.5%, 1% formalin dibunuh vaksin ketegangan *C. pseudotuberculosis* tempatan masing-masing. Kumpulan C dan D merupakan kumpulan kawalan positif dan negatif masing-masing, dan disuntik intraperitoneum hanya dengan 200µl PBS pH 7.4. Dos booster telah disuntik kepada tikus dalam kumpulan A dan B selepas 21 hari dari dos awal. Minggu ke-6 selepas inokulasi dos vaksin pertama; tikus dalam semua kumpulan kecuali kumpulan kawalan negatif telah disuntik intraperitoneally dengan 200µl jenis getir *C. pseudotuberculosis* 10⁶ CFU setiap tikus dan diperhatikan. Semua tikus telah dieutanasia pada akhir kajian, dan pemeriksaan bedah siasat telah dijalankan ke atas hati, paru-paru, jantung, buah pinggang, limpa dan limfa nodus, manakala sampel tisu organ

yang sama telah ditetapkan untuk pemeriksaan histopatologi dan seterusnya diletakkan pada agar darah untuk pengasingan bakteria dan pengenalan.

Untuk model kambing, 24 ekor kambing betina baka yang berusia antara 11-13 bulan, berat 23-30 kg, yang tidak mempunyai sejarah pelalian CLA secara rawak dibahagikan kepada 4 kumpulan yang terdiri 6 ekor kambing setiap kumpulan terdiri daripada A, B, C dan D. Kambing dalam kumpulan A dan B telah disuntik masing-masing dengan 1 ml suntikan intramuskular vaksin *C. pseudotuberculosis* 0.5% dan 1% formalin-membunuh. Kumpulan C dan D disimpan masing-masing sebagai kawalan positif dan negatif, dan disuntik secara intraotot hanya dengan 1 ml minyak dalam air emulsi. Booster dos yang disuntik selepas 21 rawatan awal. 2 minggu selepas booster dos, kambing (kecuali dalam kumpulan D) telah dicabar dengan 2ml 10^6 CFU mengasingkan liar *C. pseudotuberculosis* suntikan secara subcutaneously. Kesemua kambing telah dipantau untuk keseluruhan eksperimen tanda-tanda klinikal, dan sampel darah telah diambil pada jangka masa yang telah ditetapkan ke dalam tiub kosong dan EDTA untuk analisis IgM, IgG, IgA, Haptoglobin, Serum Amyloid A dan analisis hematologi. Pada akhir kajian, kambing dikorbankan dan bedah siasat telah dijalankan dan sampel seperti nodus limfa dalaman dan luaran, hati, paru-paru, jantung, buah pinggang, limpa, usus, saraf tunjang dan otak telah dikumpulkan untuk pemeriksaan histopatologi dan untuk pengasingan bakteria dan pengenalan.

Tikus dalam kumpulan C yang telah disuntik hanya dengan *C. pseudotuberculosis* menunjukkan signifikan perubahan tanda-tanda klinikal. Sederhana kepada bulu bergolak teruk, kadar pergerakan dikurangkan, mengurangkan responsif dan pelepasan okular diperhatikan. Tanda-tanda seperti "huddling" bersama-sama, hilang selera makan, najis pucat dan masalah pernafasan juga telah direkodkan. Dalam pemeriksaan bedah siasat, kesesakan dan abses di tapak inokulasi, limpa, hati dan buah pinggang telah direkodkan dalam kumpulan C sahaja. Pemeriksaan mikroskopik kumpulan dijangkiti menunjukkan signifikan sederhana kepada pendarahan yang teruk dan kesesakan di semua organ, ringan kepada sederhana penyusutan sel radang dan degenerasi, pelbagai abses dengan pemarkahan yang lebih tinggi nekrosis dan kurang kepada edema ringan diperhatikan dalam semua organ-organ. Sebaliknya, keputusan tikus disuntik dengan vaksin CLA menunjukkan tisu normal dengan perubahan histopatologi bukan ketara. *Corynebacterium pseudotuberculosis* dikesan oleh PCR daripada organ-organ dan nodus limfa 9 ekor tikus (90%) dalam kumpulan positif kawalan yang paling mendalam, satu tikus ditemui jelas jangkitan, semasa ia dikesan hanya dari 30% dan 20% daripada tikus dalam kumpulan A dan B masing-masing.

Untuk percubaan kambing, haiwan kawalan positif (Kumpulan C) menunjukkan signifikan gejala klinikal berbanding kumpulan vaksin yang diperhatikan inokulasi satu minggu selepas bakteria seperti kenaikan besar nodus limfa; suhu badan, pernafasan dan kadar jantung dan mendadak berkurangan dalam pergerakan rumen. Teruk keadaan kot rambut, penurunan dalam keadaan

badan pemarkahan dan pembesaran yang ketara dan pecah nodus limfa suntikan selepas 2 hingga 3 minggu cabaran juga diperhatikan. Selain itu, pembesaran ketara dan pecah sesekali nodus limfa yang terjejas telah direkodkan dalam kumpulan kawalan positif 5 hingga 6 minggu selepas cabaran dengan *C. pseudotuberculosis*. Walaupun sedikit pembesaran nodus limfa berhampiran tapak cabaran bakteria hanya sekor kambing (16%) dalam setiap kumpulan A dan B diperhatikan.

Kedua-dua kumpulan terdedah kepada vaksin kami rumuskan (kumpulan A dan B) disebabkan signifikan (untuk kedua-dua vaksin) ketinggian dalam serum Haptoglobin (Hp) dan tahap plasma serum amiloid A (SAA) yang meningkat dengan ketara selepas 2, 5, 6 dan 7 minggu berbanding dengan haiwan yang sihat bukan vaksin pada keadaan yang sama. Hp dan SAA peringkat dalam kambing, yang tidak pernah didedahkan kepada vaksin, telah dinaikkan dengan ketara seminggu selepas pendedahan kepada bakteria liar dan mengekalkan peningkatan mendadak bagi selang penilaian. Kepekatan serum IgM didapati signifikan meningkat pada minggu 3 daripada pendedahan kepada dos pertama suntikan, di mana 7 dan 9 kali ganda kenaikan diperhatikan masing-masing dalam kumpulan A dan B. Walaupun meningkat secara mendadak sehingga 18 dan 17 kali ganda diperhatikan seminggu selepas dicabar dengan bakteria berdaya maju dalam masing-masing kumpulan A dan B.

Tahap respons IgM dalam kumpulan C adalah lebih tinggi berbanding kambing kawalan yang tidak dijangkiti (kumpulan D). Kepekatan tertinggi IgG dalam kumpulan A dan B (156.63 ± 2.17 ng / mL) dan (164.03 ± 2.92 ng / mL) dikesan selepas dua minggu selepas pendedahan kepada bakteria berdaya maju keseluruhan, masing-masing. Bagi Kumpulan C, IgG bermakna kepekatan mencapai tahap maksimum (138.58 ± 1.34 ng / mL) satu bulan selepas pendedahan kepada bakteria berdaya maju keseluruhan. Walau bagaimanapun kepekatan minimum IgG dalam kumpulan vaksin kami terus meningkat berikutan vaksinasi dos rendah dan tirggi. Kambing-kambing yang telah divaksin oleh kedua-dua vaksin dirumuskan tidak mempunyai ketinggian yang signifikan kepekatan IgA sepanjang tempoh kajian. Sebaliknya, bertanda meningkat tahap IgA serum diperhatikan dalam kumpulan C dalam tempoh antara 7 hingga 14 hari jangkitan pos dengan berdaya maju *C. pseudotuberculosis*. Walau bagaimanapun, hanya 1.2 dan 1.3 kali ganda meningkat dicatatkan dalam kumpulan C masing-masing pada minggu 6 dan 7

Post mortem kumpulan vaksin menurunkan sebarang perubahan patologi kasar ketara dalam organ dalaman dan juga dalam nodus limfa dalaman dan luaran. Sebaliknya, kadar yang tinggi perubahan patologi kasar dicatatkan dalam semua kambing daripada kumpulan kawalan positif. Signifikan perubahan patologi kasar diperhatikan dalam saraf otak dan tulang belakang kambing dalam kumpulan C. Dalam organ dalaman seperti paru-paru, hati, buah pinggang, limpa dan hati; lesi patologi kasar utama diperhatikan adalah kesesakan, pendarahan, kawasan tumpuan nekrosis, atrofi lemak dan

abscessation. Pembentukan nanah jarang diperhatikan dalam kumpulan C kecuali dalam beberapa organ-organ seperti hati dan paru-paru. Sekiranya nodus limfa, peratusan yang tinggi pembentukan nanah dalam saiz yang berbeza diperhatikan dalam semua kambing dalam kumpulan C. Sebaliknya, kedua-dua kumpulan vaksin tidak membangunkan apa-apa abses dalam nodus limfa atau organ-organ dalaman walaupun pembesaran sedikit nodus limfa telah direkodkan dalam dua ekor kambing (16%) sahaja. Perubahan histopatologi yang diperhatikan dalam kumpulan positif termasuk edema, kesesakan, penyusupan sel-sel radang terutamanya limfosit dan makrofaj, pendarahan, degenerasi, keradangan granulomatous dan nekrosis caseous tidak seperti kambing vaksin yang menunjukkan hiperplasia berfungsi penting dalam tisu limfoid tanpa perubahan histopatologi.

Dalam hematologi, terdapat penurunan statistik yang signifikan dalam kepekatan Hb, kiraan RBC, nilai MCV dan PCV dalam kumpulan C selepas terdedah kepada sel-sel bakteria berdaya maju. Selari dengan itu, ketinggian statistik tidak signifikan telah diperhatikan MCHC dalam kumpulan yang sama. Kiraan WBC adalah yang tertinggi semasa uji kaji jangkitan kambing bukan vaksin (Kumpulan C) berbanding dengan kumpulan-kumpulan pra-vaksin (A dan B). Rangsangan tertinggi di neutrophil berseghmen telah mencapai 7 hari selepas cabaran untuk hidup bakteria, apabila nombor itu ($12.55 \pm 1.05 \times 10^9/L$) dalam kumpulan kawalan positif, ($10.93 \pm 1.55 \times 10^9/L$) dan ($10.14 \pm 1.38 \times 10^9/L$) dalam kumpulan A dan B masing-masing, jika dibandingkan dengan ($8.23 \pm 0.46 \times 10^9/L$) yang dilihat dalam kumpulan yang tidak dirawat (kumpulan D). Peniingkatan ketara band neutrofil mengira ($0.83 \pm 0.17 \times 10^9 /L$) dikesan dalam kumpulan kawalan positif selepas disuntik *C. pseudotuberculosis*, manakala tiada perubahan ketara dicatatkan dalam kumpulan A ($0.49 \pm 0.09 \times 10^9 /L$), B ($0.51 \pm 0.13 \times 10^9 /L$) dan D ($0.30 \pm 0.01 \times 10^9 /L$) dalam tempoh yang sama. The monosit, eosonofil basofil tuduhan dan protein plasma berada dalam julat yang normal. Walau bagaimanapun, perubahan tidak signifikan telah direkodkan pada nilai CBC pada kedua-dua kumpulan vaksin.

Hasil PCR menunjukkan lima kambing (84% daripada jumlah) daripada enam haiwan dalam kumpulan A dan B sebelum terdedah kepada 0.5% dan 1.0% formalin-membunuh *C. pseudotuberculosis* kekal negatif. Jauh lebih rendah menyebarkan ($P < 0.05$) *C. pseudotuberculosis* DNA dikesan pada organ-organ dan kelenjar limfa kumpulan vaksin. Peratusan pengesanan dalam tisu disiasat didapati 100% dalam kumpulan C.

Kesimpulannya, berikutan cabaran eksperimen dengan liar jenis *C. pseudotuberculosis* isolat tempatan, vaksin formalin dibunuh diperhatikan untuk memberikan perlindungan yang signifikan terhadap jangkitan dan kelihatan ketara menyekat penyebaran bakteria dicabar di luar tapak inokulasi dalam kebanyakan haiwan. Tikus disuntik (kumpulan A, B) membentuk imuniti dan masing-masing menghasilkan 70% dan perlindungan 80% terhadap penyakit ini. Walau bagaimanapun. Selain itu, hanya 16% kambing imunisasi dengan vaksin ini dimanifestasikan jangkitan klinikal dan patologi ringan

dengan itu, laluan penting yang berpotensi untuk penghantaran penyakit telah dihapuskan. Hasil kajian ini memberi maklumat penting kepada pembangunan caseous strategi suntikan pembasmian lymphadenitis yang berkesan di Malaysia.



ACKNOWLEDGEMENTS

I would give the praise to Allah almighty, the most beneficent the most merciful who keep inspiring me, guiding me and always looking after me and directing me toward the utmost goodness.

I also would like to express my sincere gratitude and appreciations to my supervisor Associate Professor Dr. Faez Firdaus Jesse Abdullah for the priceless guidance, supervision and the encouragement throughout the research period. Many thanks and gratitude also goes to the supervisory committee for their guidance, advice and supervision starting with Professor Dr. Abd Wahid Haron, Professor Dr. Abdul Rahman Omar and last but never least Professor Dr. Arbakariya Bin Ariff, for their continuous support and help. Many thanks also go to Professor Dr. Saleh Majeed, Mr. Yap Keng Chee, Mr. Ganesanmurthi Perumal, Mr. Mohd Jefri Norsidin, Mr. Mohd Fahmi Mashuri, Dr. Muhammed Umar, Dr. Kareem Obayes Handool, Dr. Mohammed Najj Auda, Dr. Ahmed Khalaf Ali, Dr. Konto Mohammed and Dr Idris Umar Hanbali. I would like to express my utmost appreciation and gratitude to Universiti Putra Malaysia, School of Graduate Studies and Ministry of Higher Education for giving me the opportunity to pursue this study.

My thanks and appreciations also go to Taman Pertanian Universiti, UPM, Faculty of Veterinary Medicine, UPM, Department of Veterinary Clinical Studies and Department of Pathology, Faculty of Veterinary Medicine and also to Fermentation Technology Unit, Putra Infoport, UPM for providing me with all the facilities pertaining to my research. This study was funded by the Research University Grant Scheme (RUGS), Universiti Putra Malaysia.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of Supervisory Committee were as follows:

Faez Firdaus Jesse Abdullah, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Abd Wahid Haron, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Abdul Rahman Omar, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Arbakariya Bin Ariff, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No.: Wessam Monther Mohammed Saleh, GS 39379

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Associate Professor Dr. Faez Firdaus Jesse Abdullah

Signature: _____

Name of Member of
Supervisory
Committee:

Professor Dr. Abd Wahid Haron

Signature: _____

Name of Member of
Supervisory
Committee:

Professor Dr. Abdul Rahman Omar

Signature: _____

Name of Member of
Supervisory
Committee:

Professor Dr. Arbakariya Bin Ariff

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	v
ACKNOWLEDGEMENTS	x
APPROVAL	xi
DECLARATION	xiii
LIST OF TABLES	xx
LIST OF FIGURES	xxii
LIST OF APPENDICES	xxx
LIST OF ABBREVIATIONS	xxxi
 CHAPTER	
1 INTRODUCTION	1
1.1 Background of the study	1
1.2 Problem statement	4
1.3 Hypothesis	4
1.4 Objectives	5
 2 LITERATURE REVIEW	6
2.1 General Introduction of <i>Corynebacterium pseudotuberculosis</i>	6
2.1.1 Corynebacteria	6
2.1.2 <i>Corynebacterium pseudotuberculosis</i>	7
2.1.3 Biochemical Characteristics	7
2.1.4 Antimicrobial Therapy	8
2.2 Virulence Factors	9
2.2.1 Mycolic acid	9
2.2.2 Phospholipase D (PLD)	10
2.3 Caseous Lymphadenitis	11
2.3.1 Symptoms	11
2.3.2 Prevalence of Caseous Lymphadenitis	12
2.3.3 Epidemiology and Economic Impact	13
2.3.4 Caseous lymphadenitis status in Malaysia	14
2.4 Pathogenicity and Clinical Signs	15
2.4.1 Primary Infection and the Associated Signs	15
2.4.2 Dissemination and the Associated Signs	17
2.5 <i>Corynebacterium pseudotuberculosis</i> infection in other species	18
2.5.1 <i>Corynebacterium pseudotuberculosis</i> Infection in Horses	18
2.5.2 <i>Corynebacterium pseudotuberculosis</i> Infection in Cattle	18
2.6 Zoonosis of <i>Corynebacterium pseudotuberculosis</i>	19

2.7	Immune Response	20
2.8	Acute Phase proteins	22
2.9	Vaccination against caseous lymphadenitis	24
2.10	Histology of caseous lymphadenitis	36
2.11	Effect of Caseous lymphadenitis on Reproduction	30
3	METHODOLOGY	32
3.1	Study Approval	32
3.2	Bacteria and Growth Condition:	32
3.3	Vaccination Procedures	32
3.3.1	Preparation of the Formalin-Killed Vaccine	32
3.3.2	Adjuvant Description	33
3.3.3	Adjuvant mixing procedure	33
3.3.4	Recommended Injection Protocol	34
3.4	Experiment Design and Management of Mice	34
3.5	Experiment Design and Management of Goats	36
3.6	Clinical Signs and histopathological Scoring of Mice Experiment	38
3.7	Clinical Examination of Goats	39
3.7.1	Rumen Motility	39
3.7.2	Body Condition Scoring (BCS)	39
3.7.3	Hair Coat Condition Score	39
3.8	Histopathological Examination of Goats	40
3.8.1	Post Mortem Examination	40
3.8.2	Histopathology Procedures	40
3.8.3	Lesion Scoring	40
3.9	Immune Response Analyses	41
3.9.1	Goat Serum Amyloid A (SAA) ELISA Kit Assay	41
3.9.2	Caprine Haptoglobin (Hp) Assay	42
3.9.3	Goat Immunoglobulin M (IGM) ELISA Kit Assay	43
3.9.4	Goat Immunoglobulin G (IgG) ELISA Kit Assay	44
3.9.5	Goat Immunoglobulin A (IgA) ELISA Kit Assay	45
3.9.6	Calculation Protocole of the Immune Response Data Analysis	46
3.10	Hematological analysis	46
3.11	Polymerase Chain Reaction (PCR) Technique Procedure	47
3.11.1	Bacterial Isolation and Identification	47
3.11.2	Primer Design	47
3.13	Statistical Analysis	47
4	PRELIMINARY STUDY ON EFFICACY OF A PROTOTYPE VACCINE OF <i>Corynebacterium pseudotuberculosis</i> IN MICE MODEL	48
4.1	Introduction	48
4.2	Materials and methods	48
4.3	Results	49
4.3.1	Clinical Observation	49

4.3.2	Post Mortem Examination	49
4.3.3	Histopathological Examination	53
4.3.4	PCR Results	62
4.4	Discussion	64
5	EVALUATION OF THE CLINICAL RESPONSE TO EXPERIMENTALLY INFECTION OF <i>Corynebacterium pseudotuberculosis</i> IN GOATS VACCINATED WITH PROTOTYPE FORMALIN-KILLED WHOLE CELL CASEOUS LYMPHADENITIS VACCINES	67
5.1	Introduction	67
5.2	Materials and methods	68
5.3	Results	68
5.3.1	Inspection of superficial lymph nodes	69
5.3.2	Rumen Motility	73
5.3.3	Body Condition Scoring (BCS)	75
5.3.4	Hair Coat Condition Score	76
5.3.5	Body Temperature	77
5.3.6	Heart Rate	79
5.3.7	Respiratory Rate	81
5.4	Discussion	82
6	ASSESSMENT THE LEVELS OF ACUTE PHASE PROTEINS AGAINST VIABLE <i>Corynebacterium pseudotuberculosis</i> IN GOATS VACCINATED BY PROTOTYPE WHOLE CELL FORMALIN-KILLED CASEOUS LYMPHADENITIS VACCINES	86
6.1	Introduction	86
6.2	Materials and methods	86
6.3	Results	87
6.3.1	Haptoglobin (Hp)	87
6.3.2	Serum Amyloid A (SAA)	88
6.4	Discussion	90
7	ESTIMATION THE LEVELS ANTIBODIES AGAINST VIABLE <i>Corynebacterium pseudotuberculosis</i> IN GOATS VACCINATED BY PROTOTYPE WHOLE CELL FORMALIN-KILLED CASEOUS LYMPHADENITIS VACCINES	93
7.1	Introduction	93
7.2	Materials and Methods	94
7.3	Results	94
7.3.1	Immunoglobulin M (IgM)	94
7.3.2	Immunoglobulin G (IgG)	95
7.3.3	Immunoglobulin A (IgA)	97
7.4	Discussion	98

8	EVALUATION OF THE GROSS AND HISTOPATHOLOGICAL EFFECTS OF <i>Corynebacterium pseudotuberculosis</i> IN VITAL ORGANS AND LYMPH NODES OF GOATS VACCINATED BY WHOLE CELL FORMALIN-KILLED CASEOUS LYMPHADENITIS VACCINES	101
8.1	Introduction	101
8.2	Materials and methods	102
8.3	Results	102
8.3.1	Gross lesion	102
8.3.2	Histopathology evaluation of Lymph Nodes	109
8.3.3	Histopathological Results of the Visceral Organs	124
8.3.4	Histopathological evaluation of the central nervous system	131
8.4	Discussion	133
9	EVALUATION OF CHANGES IN COMPLETE BLOOD COUNT (CBC) TO EXPERIMENTALLY CHALLENGED WITH <i>Corynebacterium pseudotuberculosis</i> IN GOATS VACCINATED WITH PROTOTYPE FORMALIN-KILLED CASEOUS LYMPHADENITIS VACCINES	139
9.1	Introduction	139
9.2	Materials and methods	139
9.3	Results	140
9.3.1	Red Blood Cells (RBC)	140
9.3.2	Hemoglobin (Hb) Concentration	141
9.3.3	Packed Cell Volume (PCV)	142
9.3.4	Mean Corpuscular Volume (MCV)	144
9.3.5	Mean Corpuscular Hemoglobin Concentration (MCHC)	145
9.3.6	White Blood Cells (WBC) Count	146
9.3.7	Band Neutrophils Count	147
9.3.8	Segmented Neutrophils Count	148
9.3.9	Lymphocytes Count	150
9.3.10	Monocytes Count	151
9.3.11	Eosinophil Count	152
9.3.12	Basophiles Count	153
9.3.13	Plasma Proteins Concentration	154
9.4	Discussion	155
10	ISOLATION AND IDENTIFICATION <i>Corynebacterium pseudotuberculosis</i> IN VISCERAL ORGANS AND LYMPH NODES OF GOATS VACCINATED WITH PROTOTYPE FORMALIN-KILLED CASEOUS LYMPHADENITIS VACCINES	159
10.1	Introduction	159
10.2	Materials and methods	160
10.3	Results	160

10.3.1	Bacterial Isolation and Identification	160
10.3.2	PCR results	163
10.3.3	<i>Corynebacterium pseudotuberculosis</i> Sequencing	166
10.4	Discussion	168
11	GENERAL DISCUSSION, CONCLUSION AND RECOMMENDATION FOR FUTURE STUDY	172
11.1	General Discussion	172
11.2	Conclusion and Recommendation	178
	REFERENCES	180
	APPENDICES	222
	BIODATA OF STUDENT	241

LIST OF TABLES

Table	Page
3.1 Clinical signs and lesion scoring in mice vaccinated and inoculated by <i>Corynebacterium pseudotuberculosis</i>	38
3.2 Lesion parameters determined in the tissues of vaccinated and control goats	41
4.1 The scoring of clinical signs for vaccinated and control groups of mice	49
4.2 The percentage of gross lesions detection of vaccinated and control groups of mice	50
4.3 The scoring of histopathological changes in vaccinated and control groups of mice	54
4.4 The overall results for PCR detection of <i>C. pseudotuberculosis</i> in visceral organs and lymph nodes of vaccinated and control groups of mice	62
5.1 Percentage of clinical examination of superficial lymph nodes in vaccinated groups performed in every handling time (per weeks) expressed as a visible enlargement of involved lymph node (swollen LN)	71
5.2 Rumen motility examination scoring of all groups performed at each handling time (per weeks)	74
5.3 Body Condition scoring mean of all groups performed at each handling time (per weeks)	75
5.4 Hair coat condition score percentage of all groups performed at each handling time (per weeks)	77
5.5 Body Temperature Mean of all groups performed at each handling time (per weeks)	78
5.6 Heart rate Mean of all groups performed at each handling time (per weeks)	80
5.7 Respiratory rate of all groups performed at each handling time (per weeks)	82
6.1 Mean of Haptoglobin (Hp) concentration of all groups during the early stage after vaccination and post infection periods	87
6.2 Mean of Serum amyloid A (SAA) concentration of all groups during the early stage after vaccination and post infection periods	89
7.1 Mean of IgM concentration of all groups during the early stage after vaccination and post infection period	94
7.2 Mean of IgG concentration of all groups during the study period	96
7.3 Mean of IgA concentration of all groups during the early stage after vaccination and post infection period	97
8.1 The frequency of gross lesion observed on the harvested organs in both vaccinated and unvaccinated goats	103
8.2 Histopathological scoring of lymph nodes (1)	110
8.3 Histopathological scoring of lymph nodes (2)	111

8.4	Histopathological scoring of visceral organs	125
8.5	Histopathological scoring of central nervous system	132
9.1	Mean of red blood cells count of all groups during the study period	140
9.2	Mean of hemoglobin concentration of all groups during the study period	142
9.3	Mean of packed cell volume (PCV) of all groups during the study period	142
9.4	Mean corpuscular volume (MCV) of all groups during the study period	144
9.5	Mean corpuscular hemoglobin concentration (MCHC) of all groups during the study period	145
9.6	Mean of total white blood cells (WBC) count of all groups during the study period	146
9.7	Mean of band neutrophils count of all groups during the study period	148
9.8	Mean of segmented neutrophils count of all groups during the study period	149
9.9	Mean of Lymphocyte count of all groups during the study period	150
9.10	Mean of monocyte count of all groups during the study period	151
9.11	Mean of eosinophil count of all groups during the study period	152
9.12	Mean of basophiles count of all groups during the study period	153
9.13	Mean of plasma protein concentration of all groups during the study period	154
10.1	The percentage of <i>Corynebacterium pseudotuberculosis</i> isolation from all experimental groups depending on the characteristic morphological feature of the bacterial culture of the selected organs	162
10.2	The percentage of <i>Corynebacterium pseudotuberculosis</i> identification using PCR technique to the harvested organs of all experimental groups	165
10.3	Evolutionary distance between the Malaysian genotypes with genotypes from other parts of the world	168

LIST OF FIGURES

Figure	Page
3.1 The experiment design of the preliminary study of efficacy of CLA vaccination on mice model	35
3.2 The experiment design of Goats	37
3.3 The diagram of standard curve of serum amyloid A (SAA) concentration	42
3.4 The diagram of standard curve of Haptoglobin concentration	43
3.5 The standard curve of IgM concentration	44
3.6 The diagram of standard curve of IgG concentration	45
3.7 The diagram of standard curve of IgA concentration	46
4.1 A post mortem carcass of vaccinated mouse with normal viscera	50
4.2 A post mortem carcass of infected mouse in group C which characterized by generalized mild congestion especially in heart, liver and spleen combined with abscesses in site of injection (A)	51
4.3 A post mortem carcass of infected mouse in group C with a characteristic abscess formation in site of inoculation (A) combined micro-abscess in kidney (B)	51
4.4 Post mortem carcass of infected mouse in group C which characterized by moderate congestion (A) especially in liver and spleen combined with micro-abscesses in spleen (B)	52
4.5 Post mortem carcass of infected mouse in group C which characterized by general congestion especially in liver and spleen combined with micro-abscesses in spleen (arrow)	52
4.6 Photomicrograph of heart of mouse in group C showing congestion (black arrow) (H&E, 200X)	55
4.7 Photomicrograph of mouse kidney tissue from group C showing congestion (black arrow) and protein casts (red arrows) (H & E , 200X)	55
4.8 Photomicrograph of group C mouse liver showing congestions (black arrow) and periportal linflammatory cell infiltration (red arrow) (H&E, 200X)	56
4.9 Photomicrograph of group C mouse lungs showing congestions (black arrows), edema(yellow arrows) and thickening of alveolar septa (blue arrow head) (H&E, 200X)	56
4.10 Photomicrograph of group C mouse lymph node showing moderate congestions (H&E, 200X)	57
4.11 Photomicrograph of group C mouse heart showing mild congestion (black arrow) and hemorrhage (red arrow) (H&E, 200X)	57
4.12 Photomicrograph of group C mouse kidney showing congestions (black arrows), protein casts (red arrow) and degeneration of tubular epithelial cells (yellow arrow) (H&E, 200X)	58

4.13	Photomicrograph of group C mouse liver showing congestion (black arrow) and hemorrhages (red arrows) (H&E, 200X)	58
4.14	Photomicrograph of group C mouse lymph node showing moderate necrosis (black arrow) and mild hemorrhage (red arrow) (H&E, 200X)	59
4.15	Photomicrograph of group C mouse lymph node showing moderate hemorrhage (arrow) (H&E, 200X)	59
4.16	Photomicrograph of group C mouse heart showing mild necrosis (arrow) and degeneration (H&E, 200X)	60
4.17	Photomicrograph of group C mouse kidney showing moderate congestion (black arrows), mild necrosis (red arrow) and moderate hemorrhage (blue arrow) (H&E, 200X)	60
4.18	Photomicrograph of group C mouse liver showing mild necrosis (black arrow), mild infiltration of inflammatory cells (red arrows) and moderate hemorrhages (yellow arrows) (H&E, 200X)	61
4.19	Photomicrograph of group C mouse lung showing mild congestion (black arrow), mild inflammatory cell infiltration (red arrows) and moderate hemorrhage (yellow arrows) (H&E, 200X)	61
4.20	Agarose gel electrophoresis figure showing amplification of 816 bp bands specific for <i>C. pseudotuberculosis</i> isolated from positive control group (group C)	83
4.21	Agarose gel electrophoresis figure showing amplification of 816 bp bands specific for <i>C. pseudotuberculosis</i> isolated from positive control group (group C)	83
5.1	Photo of the goats showing control positive goat with visible poor hair coat condition and significant decrease in body condition scoring (A) compared with normal goats in vaccinated group (B)	69
5.2	Photo of the goats showing control positive goats characterized by marked enlargement of site of <i>C. pseudotuberculosis</i> inoculation (A) combined with formation of characteristic yellowish- green pus (B)	70
5.3	Photo of goat showing a ruptured supra mammary lymph node of control positive female goat with noticeable pus spreading on perineal and mammary gland areas	72
5.4	Photo of two goats at week 12 post inoculation with <i>Corynebacterium pseudotuberculosis</i> showing marked enlargement of submandibular lymph node of goat in positive control group (A), compared to goat in group A (0.5% vaccinated group) with less enlarged submandibular lymph node (B)	72
5.5	Photo showing an enlargement of prescapular lymph node (red circle) of goat in control positive group	73
5.6	Diagram of rumen motility manifestation showing a marked decrease in positive control group (group C) following challenge with <i>C. pseudotuberculosis</i> compared with vaccinated (groups A and B) and negative control group (group D)	74

5.7	Diagram of body condition scoring showing a significant decrease in positive control group (group C) following challenge with <i>C. pseudotuberculosis</i> in contrast with vaccinated (groups A and B) and negative control group (group D)	76
5.8	Diagram of body temperature showing a significant increase in control positive group (group C) following challenge with <i>C. pseudotuberculosis</i> in contrast with vaccinated and negative groups.	79
5.9	Diagram of heart rate showing a significant increase in control positive group (group C) following challenge with <i>C. pseudotuberculosis</i> in contrast with vaccinated and negative groups	80
5.10	Diagram of respiratory rate showing a significant increase in control positive group (group C) following challenge with <i>C. pseudotuberculosis</i> in contrast with vaccinated and negative groups	81
6.1	Diagram of Haptoglobin concentration showing a significant increased after first and boosted doses of vaccine in vaccinated goats (groups A and B), with marked increased of its values during post infection period of <i>C. pseudotuberculosis</i> in positive control group (group C) compared with negative control group (group D)	88
6.2	Diagram of Serum amyloid A concentration showing a significant increase after first and boosted doses of vaccine in vaccinated goats (groups A and B), with marked increased of its values during post infection period of <i>C. pseudotuberculosis</i> in positive control group (group C) compared with negative control group (group D)	89
7.1	Diagram of IgM concentration showing a significant increased up to two weeks follows first and booster doses of vaccine both vaccinated groups (groups A and B), in contrast with control positive (group C) and negative groups (group D)	95
7.2	Diagram of IgG concentration showing a significant increased up to 18 week after first and booster doses of vaccine in both vaccinated groups (groups A and B), at variance with control positive group (group C) in which the IgG concentration was increased follow infection with <i>C. pseudotuberculosis</i>	96
7.3	Diagram of IgA concentration showing a markedly increase in control positive group (group C) in post infection period contrast to vaccinated goats (groups A and B) and positive control group (group C)	97
8.1	Photo of gross lesion of infected goat in group C showing the characteristic yellow-green viscous pus with tooth paste like consistency of <i>C. pseudotuberculosis</i> infection in prescapular lymph node	104
8.2	Photo of gross lesion of infected goat in group C showing the characteristic yellow-green viscous pus with tooth paste like consistency of <i>C. pseudotuberculosis</i> infection in pre-femoral lymph node	104

8.3	Photo of gross lesion of infected goat in group C showing significant enlargement of prefemoral lymph node	105
8.4	Cross-section of prescapular lymph node of control positive goat showing "0.7 × 1 cm in diameter" (A) and "0.8 × 1.4 cm in diameter" (B) abscesses in the medulla and cortex of the node	105
8.5	Longitudinal section of submandibular lymph node of goat in positive control group showing "1.4 × 2 cm in diameter" abscess in the medulla and hilum region of the node	106
8.6	Longitudinal section of prefemoral lymph node of control positive goat showing "0.7 cm in diameter" abscess in the cortex region of the node	106
8.7	Cross-section of prescapular lymph node of control positive goat showing severe congestion in the subcapsular region of the node	107
8.8	Cross section of affected liver of goat in group C showing abscess with lamellar onion-ring appearance revealing typical feature of CLA infection	107
8.9	Gross view of affected lungs of control positive goat showing moderate to severe generalized congestion (A) combined with multifocal areas of hemorrhage (B) and zones of emphysema (C)	108
8.10	Gross view of lungs of control positive goat showing local affected area with interstitial abscess [A], gross and transverse section showing marked enlargement of Mediastinal lymph node revealing typical feature of CLA infection (lamellar onion-ring appearance containing yellow-greenish viscous pus) [B]	109
8.11	Photomicrograph of goat submaxillary lymph node in group C showing suppuration of lymphoid tissue and granuloma (H&E stain 200X)	112
8.12	Photomicrograph of goat submandibular lymph node in vaccinated group showing an active lymphoid tissue (hyperplasia of lymphoid tissue in which the recirculation of T lymphocyte occur to build a new generation in case of exposure to antigen) (H&E 200X)	112
8.13	Photomicrograph of goat submandibular lymph node in group C showing the medullary cord rich in cellularity (lymphadenitis) including macrophage, lymphocyte and plasma cells and slight hemorrhage (H&E stain 200X)	113
8.14	Photomicrograph of goat submandibular lymph node in group C showing a circumscribe area of macrophages. (H&E stain 200X)	113
8.15	Photomicrograph of the medulla of submaxillary lymph node of goats in group C showing congestion (black arrow) with poor cellularity and macrophages vacuolation in sinusoid. (H&E stain 200X)	114
8.16	Photomicrograph of goat submaxillary lymph node of vaccinated goat showing lymphoid hyperplasia with fibrosis (black arrows) (H&E stain 200X)	114

8.17	Photomicrograph of goat submaxillary lymph node in group C showing granulomatous inflammatory reaction and macrophages infiltration enclosed by thin fibrous capsule and congested blood vessels (arrows) (H&E stain 200X)	115
8.18	Photomicrograph of goat pre scapular lymph node in group C showing area of caseous necrosis in lymphoid tissue (arrow) (H&E stain 200X)	115
8.19	Photomicrograph of goat pre scapular lymph node in group C showing areas of granulomatous inflammation and caseous necrosis enclosed with lymphoid tissue and congestion (black arrows) (H&E stain 200X)	116
8.20	Photomicrograph of goat pre scapular lymph node in vaccinated group showing apoptosis in the center of active lymphoid tissue (germinal center) (H&E stain 200X)	116
8.21	Photomicrograph of goat pre femoral lymph node in group C showing area of suppuration of lymphoid tissue (H&E stain 200X)	117
8.22	Photomicrograph of goat pre femoral lymph node in group C showing large granuloma enclosed by thin fibrous capsule (black arrows), medullary sinusoid infiltrated by macrophages, neutrophils and plasma cells and also medullary cord filled with lymphocyte and plasma cells (H&E stain 200X)	117
8.23	Photomicrograph of goat pre femoral lymph node in group C showing large granuloma in the center of the medullary cord (black arrow), sinusoid with poor cellularity (H&E stain 200X)	118
8.24	Photomicrograph of goat popliteal lymph node in group C showing large granuloma in the center of the medullary cord, sinusoid rich in cellularity (lymphocyte and plasma cells) and medullary cord infiltrated by lymphocyte and macrophage (H&E stain 200X)	118
8.25	Photomicrograph of goat supra mammary lymph node in group C showing dilated sinusoid (due to edema) in medulla with some macrophages and neutrophils (H&E stain 200X)	119
8.26	Photomicrograph of goat supra mammary lymph node in group C showing medulla area with lymphadenitis, dilated and high cellularity sinusoid and congestion (arrows) (H&E stain 200X)	119
8.27	Photomicrograph of goat inguinal lymph node in group C showing medulla area in which the sinusoid was rich in neutrophils and macrophages while the medullary cord was poor in cellularity. Severe congestion (black arrows) and edema (red arrow) (H&E stain 200X)	120
8.28	Photomicrograph of goat inguinal lymph node in group C showing sub-capsular sinusoid with early granulomatous inflammatory reaction and congestion (black arrows) (H&E stain 200X)	120
8.29	Photomicrograph of goat mediastinal lymph node in group C showing granulomatous inflammation of lymphoid tissue in the cortico-medullary junction surrounded by fibrous tissue, dilated medullary sinusoid which filled with fluids and rich in cellularity	121

	(plasma cells, macrophages and neutrophils) (H&E stain 200X)	
8.30	Photomicrograph of goat mediastinal lymph node in group C showing large granuloma with sub-capsular sinusoid rich in lymphocytes and macrophages (H&E stain 200X)	121
8.31	Photomicrograph of goat mediastinal lymph node in group C showing dilated sinusoid with vacuolated macrophages, edema (black arrows) and microgranuloma (red arrow) (H&E stain 200X)	122
8.32	Photomicrograph of goat mesenteric lymph node in group C showing granuloma in cortex which was enclosed by fibrous tissue capsule (H&E stain 200X)	122
8.33	Photomicrograph of goat mesenteric lymph node in group C showing granuloma in the cortico-medullary junction, macrophages vacuolation in sinusoid (black arrows) and congestion (red arrow) (H&E stain 200X)	123
8.34	Photomicrograph of goat mesenteric lymph node in group C showing large granuloma in the cortico-medullary junction, with macrophages vacuolation (H&E stain 200X)	123
8.35	Photomicrograph of transverse section of intestine in group C showing increased cellularity of lamina propria, prominent number of mucous glands and congestion of the mucous gland blood vessels. (H&E stain 200X)	126
8.36	Photomicrograph of transverse section of intestine in group C showing edema in submucosa (black arrows), mild congestion and hemorrhage (red arrow). (H&E stain 200X)	126
8.37	Photomicrograph of goat spleen red pulp in vaccinated group showing functional hyperplastic active lymphoid tissue. (H&E stain 200X)	127
8.38	Photomicrograph of goat spleen in group C showing granuloma with signs of degeneration in the center of lymphoid tissue combined by microhemorrhage. (H&E 200X)	127
8.39	Photomicrograph of goat spleen in group C showing focal area of necrotic tissue with degeneration and microhemorrhage. (H&E stain 200X)	128
8.40	Photomicrograph of goat liver in vaccinated group showing normal hepatocellular architecture of portal area. (H&E stain 200X)	128
8.41	Photomicrograph of goat kidney in group C showing vacuolation of mesangial cells and proximal convoluted tubules, accumulation of fluid in bowman space (red arrow), congestion of glomerular capillaries and in the interstitium (black arrows)(H&E 200X)	129
8.42	Photomicrograph of goat kidney (distal part of medulla) in group C showing severe interstitial congestion (black arrows), vacuolation of tubular epithelial lining cell (red arrows) (H&E stain 200X)	129
8.43	Photomicrograph of goat lung in group C showing area of collapse lung which contains congested blood vessels (black arrows), inflammatory cell infiltration (red arrows) and pulmonary edema (yellow arrows) (H&E stain 200X)	130

8.44	Photomicrograph of goat heart in group C showing interstitial edema (red arrows) with hemorrhages and congestion (black arrows) (H&E stain 200X)	130
8.45	Photomicrograph of goat heart in group C showing interstitial edema (red arrows) with hemorrhages and congestion (black arrows) (H&E stain 200X)	131
8.46	Photomicrograph of gray matter (neuron) of goat brain in group C showing a pre-neural vacuolation due to degeneration combined by gliosis and local area of hemorrhage (black arrow). (H&E stain 200X)	132
9.1	Diagram of RBC count showing a significant decreased in positive control group (group C) after challenged with <i>C. pseudotuberculosis</i> at variance with vaccinated (groups A and B) and negative control group (group D)	141
9.2	Diagram of Hb concentration showing a significant decreased in positive control group (group C) after challenged with <i>C. pseudotuberculosis</i> compared with vaccinated (groups A and B) and negative control group (group D)	142
9.3	Diagram of PCV values showing a sharp decreased in positive control group (group C) after challenged with <i>C. pseudotuberculosis</i> compared with vaccinated (groups A and B) and negative control group (group D)	143
9.4	Figure 9.4: Diagram of MCV showing a noticeable decreased in positive control group (group C) after challenged with <i>Corynebacterium pseudotuberculosis</i> compared with vaccinated (groups A and B) and negative control group (group D)	144
9.5	Diagram of MCHC showing a slightly increased in mean positive control group (group C) after challenged with <i>C. pseudotuberculosis</i> compared with vaccinated (groups A and B) and negative control group (group D)	145
9.6	Diagram of WBC count showing a slightly increased in positive control group (group C) after challenged with <i>Corynebacterium pseudotuberculosis</i> compared with vaccinated (groups A and B) and negative control group (group D)	147
9.7	Diagram of band neutrophils count showing a significant increase in positive control group (group C) after challenged with <i>C. pseudotuberculosis</i> compared with vaccinated groups (groups A and B) and negative control group (group D)	148
9.8	Diagram of segmented neutrophils count showing a significant increase in positive control group (group C) after challenged with <i>C. pseudotuberculosis</i> compared with vaccinated groups (groups A and B) and negative control group (group D)	149
9.9	Figure 9.9: Diagram of lymphocytes count showing a gradual decreased in positive control (group C), vaccinated groups (groups A and B) and negative control (group D) during the study period	150

9.10	Diagram of monocytes count showing a slightly increased in positive control (group C) and vaccinated groups (groups A and B) after challenged with <i>C. pseudotuberculosis</i> compared with negative control (group D)	151
9.11	Diagram of eosinophil count showing a significant increase in vaccinated groups (groups A and D) after two months of the experiment compared to positive (group C) and negative (group D) control groups	152
9.12	Diagram of basophiles count in vaccinated groups (groups A and B), positive (group C) and after negative (group D) control groups	153
9.13	Diagram of plasma proteins concentration in vaccinated groups (groups A and B), positive (group C) and after negative (group D) control groups	154
10.1	A characteristic morphology of <i>Corynebacterium pseudotuberculosis</i> culture colonies on blood agar plate	161
10.2	Agarose gel electrophoresis figure (i) [7% Agarose Bench Top 1Kb DNA Ladder, Cat # G7541, Load 6µl/lane] showing amplification of 816 bp bands specific for <i>Corynebacterium pseudotuberculosis</i> isolated from spleen (2), prescapular LN (4) and prefemoral LN (5) of affected vaccinated goats in groups A, B	164
10.3	Agarose gel electrophoresis figure (ii) showing amplification of 816 bp bands specific for <i>Corynebacterium pseudotuberculosis</i> isolated from visceral organs and lymph nodes of positive group goats (Group C)	166
10.4	Agarose gel electrophoresis figure (iii) showing amplification of 816 bp bands specific for <i>Corynebacterium pseudotuberculosis</i> isolated from visceral organs and lymph nodes of positive group goats (Group C)	166
10.5	Phylogenetic tree of the 16S rRNA gene sequences of 2 Malaysian <i>C. pseudotuberculosis</i> isolates identified in this study and 4 other isolates from other regions obtained from GenBank. The tree was inferred using the Neighbor-Joining method, with 1000 bootstrap replicates	167
10.6	Bootstrap consensus tree of the 18S rRNA gene sequences of the 2 Malaysian <i>C. pseudotuberculosis</i> isolates identified in this study and 4 other isolates from other regions obtained from GenBank. The tree was inferred using the Neighbor-Joining method, with 1000 bootstrap replicates	168

LIST OF APPENDICES

Appendix	Page
A The approval letter of the experimental procedures by the "Institutional Animal Care and Use Committee" Universiti Putra Malaysia	222
B General observation sheet	223
C Histopathology Procedure	224
D Goat Serum Amyloid A (SAA) ELISA Kit Assay (Cat. No. : E0056GO)	226
E Estimation of Caprine Haptoglobin (Hp) (Cat. No. TP-801)	228
F Goat Immunoglobulin M (IGM) ELISA Kit Assay (Cat. No. : E0002GO)	230
G Goat Immunoglobulin G (IgG) ELISA Kit Assay (Cat No. QY-E140013)	232
H Goat Immunoglobulin A (IgA) ELISA Kit Assay (Cat No. QY-E140014)	234
I Polymerase Chain Reaction (PCR) Technique Procedure	236

LIST OF ABBREVIATIONS

AGID	Agar gel immunodiffusion
APP	Acute phase proteins
CBC	Complete Blood Count
CFU	Colony forming unit
CLA	Caseous lymphadenitis
DVS	Department of Veterinary Service
ELISA	Enzyme Linked Immunosorbent Assay
H&E	Hematoxylin and Eosin
Hb	Hemoglobin
Hp	Haptoglobin
I.V	Intravenous
IACUC	Institutional Animal Care and Use Committee
IgA	Immunoglobulin A
IgG	Immunoglobulin G
IgM	Immunoglobulin M
IP	Intraperitoneal
MCHC	Mean Corpuscular Hemoglobin Concentration
MCV	Mean Corpuscular Volume
µg	Microgram
mg	Milligram
ng	Nanogram
ml	Milliliter
OD	Optical Density
OIE	Office of International Epizootic
PBS	Phosphate Buffered Saline
PCR	Polymerase Chain Reaction
PCV	Packed Cell Volume
PLD	Phospholipase D
RBC	Red Blood Cell
S.C	Subcutaneous

SAA	Serum Amyloid A
TPU	Taman Pertanian Universiti
UPM	Universiti Putra Malaysia
VLSU	Veterinary Laboratories Service Unit
WBC	White Blood Cell



CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Corynebacterium pseudotuberculosis is the causative agent of caseous lymphadenitis (CLA) or cheesy gland disease which has been found in all major global areas of goats and sheep production (Williamson, 2001; Paton et al., 2003). Caseous lymphadenitis is characterized by the formation of suppurative abscesses particularly in superficial and internal lymph nodes, and also in internal organs of small ruminants. The disease is associated with considerable economic losses to sheep and goat farmers worldwide, mainly due to the reduction of wool, meat and milk yields, decreased reproductive efficiencies of affected animals and condemnation of carcasses and skins (Paton et al., 1994; Arsenault et al., 2003). Due to the chronic and generally subclinical nature of the disease, control is usually difficult and prevalence in animals and herds is high (de Sá Guimarães et al., 2011).

Caseous lymphadenitis has a global distribution and it is endemic in countries such as North and South America, Australia, New Zealand, Europe, Asia (including Malaysia) and Africa (Stanford et al., 1998a; Arsenault et al., 2003; Paton et al., 2003).

Malaysia has a small ruminant population, with the estimated number of sheep and goats as at 2012 to be about 129,850 and 458,646 heads respectively (Khadijah et al., 2014). However, caseous lymphadenitis has been considered as one of the major diseases confronting small ruminants' productivity in Malaysian (Chandrawathani, 1998).

The average prevalence of CLA in Malaysian small ruminant (sheep and goat), was found to be 30% using two (AGPT and ELISA) combined diagnostics tests (Komala et al., 2008). In Malaysia, the disease has been identified as potential threat to the livestock industry causing several negative impacts on both productivity and fertility of small ruminant herds (Jesse et al., 2013).

Once established in a herd or flock; the eradication of CLA is pretty difficult, owing to the inefficacy of antimicrobial therapy (Piontkowski and Shivvers, 1998; Stanford et al., 1998a; Williamson, 2001). Therefore, the most reliable control strategy for this disease involves vaccinating livestock and identifying and removing infected animals from the flock (Brown et al., 1986b; Paton et al., 2003).

Despite the known significant economic effect of CLA, details of the immune responses against *C. pseudotuberculosis* infection are not completely understood and effective vaccine is still lacking (Dorella et al., 2009). It is generally agreed that the best strategy to control the disease is vaccination of healthy animals, along with the identification/removal of infected animals (Paton et al., 1995; Williamson, 2001; Menzies et al., 2004). However, the difficulties associated with the early clinical identification of infected animals have been a great tricky problem to such a strategy. Several serodiagnostic tests have been developed to overcome the problem of clinical identification of CLA, but most have been reported to lack either sensitivity or specificity (Brown and Olander, 1987; Menzies and Muckle, 1989; Williamson, 2001; Menzies et al., 2004).

The diagnosis of CLA is mainly based on the characteristic clinical symptoms and on isolation of the agent from discharging abscesses. Identification of the cultured organisms as *C. pseudotuberculosis* is usually achieved by biochemical tests but is often problematic due to extensive variability in biochemical characteristics of the pathogen (Muckle and Gyles, 1982; Songer et al., 1988). The disease could be detected directly and rapidly from infected sheep and goats with a high diagnostic sensitivity by PCR technique (Pacheco et al., 2007).

Serological diagnosis is also of importance in the diagnosis of this disease, since subclinically infected animals play major roles in introducing CLA into a healthy flock. Serological tests, such as ELISA (Schreuder et al., 1994; Dercksen et al., 2000), complement fixation test (Shigidi, 1979), immunodiffusion test (Burrell, 1980) and the hemolysis inhibition test (Kuria and Holstad, 1988) have been employed for the diagnosis of CLA, and though most of these tests lack either sensitivity or specificity, the ELISA particularly proved to be a versatile tool in control and eradication programs (Dercksen et al., 2000).

Acute phase proteins (APPs) are proteins synthesized during an acute phase response (APR) against several stimuli like infection, inflammation, stress, trauma or tissue change (Petersen et al., 2004; Ceron et al., 2005). The positive APP may show up to 100-1000-fold increase in serum concentration in 1-2 days post inducing of the stimuli; a moderate APP displays a 5-10 fold increase in 2-3 days; and a minor APP increase between 50% and 100%. Negative APPs are those which decrease after a specific stimulus (Petersen et al., 2004; Cray et al., 2009; Eckersall and Bell, 2010). Hp and SAA are considered as major APPs in both ovine and caprine APPs. (Bastos et al., 2011) stated that serum Haptoglobin concentration and monocyte count may be the potential markers for progression of CLA in sheep.

In studies involving sheep challenged subcutaneously, vast numbers of neutrophils were observed at the site of inoculation within the first few hours following infection and from there began to move to the local drainage lymph node within 24 hours. Neutrophils population was found to decrease 3 days following inoculation, while the numbers of macrophages at the inoculation site were reported to increase significantly (Pépin et al., 1992). A period of generalized inflammation of the lymph node has been demonstrated to follow infection and micro-abscesses developed within the cortical region within 24 hours of inoculation (Pépin et al., 1991).

Infection of small ruminants with *C. pseudotuberculosis* usually results in the development of pyogranulomatous lesions (Baird and Fontaine, 2007), which manifested in two different forms. The external which is also known as cutaneous or superficial, form disease is characterized by the development of abscesses within the superficial lymph nodes or within subcutaneous tissue while the visceral form involve the development of abscesses in the internal organs such as liver, lungs and kidneys. In either case, abscesses may develop over a protracted period, becoming swollen and encased within fibrous capsules, subsequently resulting to the loss of overlying hair and often rupturing of the abscess (Radostitis et al., 2000). The purulent contents of ruptured lesion are unfortunately a significant source of cross-infection between animals (Brown and Olander, 1987). In naturally observed infections, the main portal of bacterial entry has been generally accepted to be through the skin, normally due to the presence of minor wounds and abrasions (Batey, 1986b; Brown and Olander, 1987; Davis, 1990; Collett et al., 1994).

Vaccination is the primary means of prevention of the diseases in several countries, whereby immunization reduces the spread of infection, leading to a gradual decline in disease prevalence (Paton et al., 2003). Most of the currently-available commercial vaccines for caseous lymphadenitis are polyvalent vaccines (combined with vaccines against other pathogens). This includes polyvalent vaccines consisting of *Clostridium tetani*, *Clostridium perfringens*, *Clostridium septicum*, *Clostridium novyi* and *Clostridium chauvoei* antigens (Piontkowski and Shivvers, 1998; Paton et al., 2003). Formalin-killed vaccine of local isolate of *C. pseudotuberculosis* has been documented to induce up to 70% protection in sheep challenged with local isolate of *C. pseudotuberculosis* (Selim et al., 2010).

Although *C. pseudotuberculosis* infection has been documented to have worldwide distribution and the disease is associated with huge economic losses, there is currently no vaccine that induced 100% protection against the disease. Additionally, information on the detail clinical manifestation of the disease and identification of possible biomarkers of the disease are still scanty despite the economic significance of the disease. In view of the difficulties and challenges of eradicating CLA in endemic areas or farms through treatment, thrust for the development of an efficient CLA vaccine is a global necessity.

In this study, potential prototype vaccine candidate for CLA based on local Malaysian isolate was developed. It is the authors' view that the development of local isolate prototype vaccine may help in reducing the prevalence of this disease among small ruminants in Malaysia.

1.2 Problem Statement

1. Malaysian small ruminants (sheep and goat) have high average prevalence of CLA which was found to be about 30%. Currently in Malaysia, there is no any available local vaccine against CLA in small ruminant. Therefore this study is designed to develop prototype vaccine of *C. pseudotuberculosis* using local isolates.
2. Caseous lymphadenitis is a chronic disease and difficult to diagnose at an early stage. Acute phase proteins and hematological values have a significant, sensitive and valid role as diagnostic biomarkers for subclinical, acute and chronic conditions. APPs and hematological values were used as indicator of disease progression after vaccination.
3. Caseous lymphadenitis also has a cellular effect on all visceral organs and lymph nodes which eventually affects the activity of the goat. Histopathological changes of vital organs were used as an indicator of disease progression in challenged animals after vaccination.

1.3 Hypothesis

- The development of prototype vaccine from local isolates of *C. pseudotuberculosis* will help in reducing the prevalence of the disease among small ruminants in Malaysia with non-significant side effects on the host.
- Clinical response, complete blood count (CBC), biochemistry, acute phase proteins and histopathological changes of the tested animals will be used as an indicator for disease progression of challenged animals after vaccination in vitro work.

1.4 The Objectives of this study were:

1. To develop a prototype vaccine of *C. pseudotuberculosis* from local Malaysian isolate and to study the efficacy of the vaccine in goats and mice models.
2. To evaluate the clinical response between the infected goat by *C. pseudotuberculosis* and the vaccinated ones.
3. To measure the changes in the haemogram between the infected goats by *C. pseudotuberculosis* and the vaccinated ones.
4. To assess the acute phase proteins responses between infected goats by *C. pseudotuberculosis* and the vaccinated ones.
5. To estimate the antibodies response (IgM, IgG, IgA) between the infected goats by *C. pseudotuberculosis* and the vaccinated ones.
6. To evaluate the histopathological changes of visceral organs and lymph nodes between the infected goats by *C. pseudotuberculosis* and the vaccinated ones.
7. To detect *C. pseudotuberculosis* from vital organs and lymph nodes using bacterial culture, biochemical tests and PCR techniques from the infected goats by *C. pseudotuberculosis* and the vaccinated ones.

REFERENCES

- Abadie, V., Badell, E., Douillard, P., Ensergueix, D., Leenen, P. J., Tanguy, M., Fiette, L., Saeland, S., Gicquel, B. & Winter, N. 2005. Neutrophils rapidly migrate via lymphatics after *Mycobacterium bovis* BCG intradermal vaccination and shuttle live bacilli to the draining lymph nodes. *Blood*, 106, 1843-1850.
- Abdinasir, Y., Jesse, F., Saharee, A., Jasni, S., Khairani-Bejo, S. & Haron, A. 2012. Clinico-Pathological Changes in Mice Following Experimental Infection with Whole Cell and Exotoxin (PLD) Extracted from *C. pseudotuberculosis*. *Journal of Animal and Veterinary Advances*, 11, 4064-4072.
- Abdullah, F. F. J., Adamu, L., Tijjani, A., Mohammed, K., Abba, Y., Sadiq, M. A., Zainal, R., Sabu, M. J. B., Saharee, A. A. & Haron, A. W. 2015a. Hormonal and Histopathological Alterations in Pituitary Glands and Reproductive Organs of Male and Female Mice Orally Inoculated with *Pasteurella multocida* Type B: 2 and its Lipopolysaccharides. *American Journal of Animal and Veterinary Sciences*, 10, 23.
- Abdullah, F. F. J., Latif, N. a. A., Chung, E. L. T., Aimi, S., Sarah, M. Z.-S., Haron, A. W., Lila, M. a. M., Zakaria, Z. & Norsidin, M. J. 2015b. Changes in the Reproductive Hormones of Non-Pregnant Does Infected Intradermally with *Corynebacterium pseudotuberculosis* in Chronic Form. *International Journal of Livestock Research*, 5, 33-40.
- Abdullah, F. F. J., Osman, A. Y., Adamu, L., Azri, N. A. & Wahid, A. 2013. Caseous Lymphadenitis in a Goat. *Veterinary Journal*, 52, 513-514.
- Abdullah, J. F. F., Lau, S. S., Abdul Aziz, S. & Shamsudin, S. 2011. Pathological changes in the organs of mice model inoculated with *Corynebacterium pseudotuberculosis* organism. *Pertanika Journal of Tropical Agricultural Science*, 34, 145-149.
- Adams, D. 1976. The granulomatous inflammatory response. A review. *The American Journal of Pathology*, 84, 164-192.
- Adekeye, J., Shannon, D. & Addo, P. 1980. Mastitis in a cow caused by *Corynebacterium pseudotuberculosis* (C ovis). *Veterinary Record*, 106, 270-270.
- Aderem, A. & Underhill, D. M. 1999. Mechanisms of phagocytosis in macrophages. *Annual review of immunology*, 17, 593-623.
- Ahmed, D. 2015. Association between fatty acids profile of *Corynebacterium pseudotuberculosis* with pathogenicity in mice. UOFK.
- Ahmed, I. M. & El-Tahawy, A. S. 2012. Prevalence of So-called Oedematous Skin Disease in Egyptian buffaloes with particular study on its economic influence. *ALEX. J. VET. SCIENCE*, 37, 129-133.

- Akenzua, G. I., Hui, Y. T., Milner, R. & Zipursky, A. 1974. Neutrophil and band counts in the diagnosis of neonatal infections. *Pediatrics*, 54, 38-42.
- Al-Gaabary, M. H., Osman, S. A., Ahmed, M. S. & Oreiby, A. F. 2010. Abattoir survey on caseous lymphadenitis in sheep and goats in Tanta, Egypt. *Small Ruminant Research*, 94, 117-124.
- Al-Gaabary, M. H., Osman, S. A. & Oreiby, A. F. 2009. Caseous lymphadenitis in sheep and goats: clinical, epidemiological and preventive studies. *Small Ruminant Research*, 87, 116-121.
- Aleman, M., Spier, S. J., Wilson, W. D. & Doherr, M. 1996. *Corynebacterium pseudotuberculosis* infection in horses: 538 cases (1982-1993). *Journal of the American Veterinary Medical Association*, 209, 804-809.
- Alharbi, K. 2011. Control of abscess disease of sheep by concurrent vaccination, zinc injection and antiseptic washing. *Res J Vet Sci*, 4, 9-13.
- Alharbi, K. B. & Mahmoud, O. M. 2013. Abscess disease of sheep and goats: A disease of major concern in Saudi Arabia that urges production of an effective vaccine. *Journal of Agricultural and Veterinary Sciences*, 5.
- Alonso, S., Pethe, K., Russell, D. G. & Purdy, G. E. 2007. Lysosomal killing of Mycobacterium mediated by ubiquitin-derived peptides is enhanced by autophagy. *Proceedings of the National Academy of Sciences*, 104, 6031-6036.
- Alsemgeest, S., Horadagoda, A., Hulskamp-Koc, C., Tootend, P., Kim, H., Niewold, T. A. & Gruys, E. 1995. First Evidence for the Existence of Multiple Isoforms of Bovine Serum Amyloid-A (apoSAA). *Scandinavian journal of immunology*, 41, 407-413.
- Alshamaony, L., Goodfellow, M., Minnikin, D., Bowden, G. & Hardie, J. 1977. Fatty and mycolic acid composition of *Bacterionema matruchotii* and related organisms. *Microbiology*, 98, 205-213.
- Anderson, D. E., Rings, D. M. & Kowalski, J. 2004. Infection with *Corynebacterium pseudotuberculosis* in five alpacas. *Journal of the American Veterinary Medical Association*, 225, 1743-1747.
- Andoh, A., Zhang, Z., Inatomi, O., Fujino, S., Deguchi, Y., Araki, Y., Tsujikawa, T., Kitoh, K., Kim-Mitsuyama, S. & Takayanagi, A. 2005. Interleukin-22, a member of the IL-10 subfamily, induces inflammatory responses in colonic subepithelial myofibroblasts. *Gastroenterology*, 129, 969-984.
- Antunes, C. A., Dos Santos, L. S., Hacker, E., Köhler, S., Bösl, K., Ott, L., De Luna, M. D. G., Hirata Jr, R., De Carvalho Azevedo, V. A. & Mattos-Guaraldi, A.-L. 2015. Characterization of DIP0733, a multi-functional virulence factor of *Corynebacterium diphtheriae*. *Microbiology*, 161, 639-647.

- Armstrong, J. & Hart, P. A. 1971. Response of cultured macrophages to *Mycobacterium tuberculosis*, with observations on fusion of lysosomes with phagosomes. *The Journal of experimental medicine*, 134, 713-740.
- Aroch, I., Harmelin, A., Saran, A., Levin, D. & Shpigel, N. 2003. Experimental *Corynebacterium pseudotuberculosis* mastitis in dairy cows. *The Veterinary Record*, 153, 746-750.
- Arpin, C., Dechanet, J., Van Kooten, C., Merville, P., Grouard, G., Briere, F., Banchereau, J. & Liu, Y.-J. 1995. Generation of memory B cells and plasma cells in vitro. *Science*, 268, 720-722.
- Arredouani, M. S., Kasran, A., Vanoirbeek, J. A., Berger, F. G., Baumann, H. & Ceuppens, J. L. 2005. Haptoglobin dampens endotoxin-induced inflammatory effects both in vitro and in vivo. *Immunology*, 114, 263-271.
- Arsenault, J., Girard, C., Dubreuil, P., Daignault, D., Galarneau, J.-R., Boisclair, J., Simard, C. & Bélanger, D. 2003. Prevalence of and carcass condemnation from maedi-visna, paratuberculosis and caseous lymphadenitis in culled sheep from Quebec, Canada. *Preventive veterinary medicine*, 59, 67-81.
- Asif, M. 2012. Study of clinically used and recently developed antimycobacterial agents. *Oriental Pharmacy and Experimental Medicine*, 12, 15-34.
- Augustine, J. & Renshaw, H. *Corynebacterium pseudotuberculosis* survival in soil samples amended with water. *Proceedings*, 1982.
- Augustine, J. & Renshaw, H. 1986. Survival of *Corynebacterium pseudotuberculosis* in axenic purulent exudate on common barnyard fomites. *American journal of veterinary research*, 47, 713-715.
- Ayers, J. 1977. Caseous lymphadenitis in goats and sheep: a review of diagnosis, pathogenesis, and immunity. *Journal of the American Veterinary Medical Association*, 171, 1251.
- Badolato, R., Wang, J. M., Murphy, W. J., Lloyd, A. R., Michiel, D. F., Bausserman, L. L., Kelvin, D. J. & Oppenheim, J. J. 1994. Serum amyloid A is a chemoattractant: induction of migration, adhesion, and tissue infiltration of monocytes and polymorphonuclear leukocytes. *The Journal of experimental medicine*, 180, 203-209.
- Bain, A. 1963. *Corynebacterium equi* infections in the equine. *Australian Veterinary Journal*, 39, 116-121.
- Baird, G. 2003. Current perspectives on caseous lymphadenitis. *In Practice* (0263841X), 25.
- Baird, G. 2006. Treatment of ovine caseous lymphadenitis. *The Veterinary Record*, 159, 500.
- Baird, G. 2007. *Caseous lymphadenitis In Aitkin ID (ed.) Diseases of sheep*, Blackwell Science, Oxford.

- Baird, G. & Fontaine, M. 2007. *Corynebacterium pseudotuberculosis* and its role in ovine caseous lymphadenitis. *Journal of comparative pathology*, 137, 179-210.
- Baird, G. & Malone, F. 2010. Control of caseous lymphadenitis in six sheep flocks using clinical examination and regular ELISA testing. *Veterinary Record: Journal of the British Veterinary Association*, 166.
- Bakkaloglu, A., Duzova, A., Ozen, S., Balci, B., Besbas, N., Topaloglu, R., Ozaltin, F. & Yilmaz, E. 2004. Influence of Serum Amyloid A (SAA1) and SAA2 gene polymorphisms on renal amyloidosis, and on SAA/C-reactive protein values in patients with familial mediterranean fever in the Turkish population. *The Journal of rheumatology*, 31, 1139-1142.
- Baldassarri, L., Puliti, M. & Tissi, L. 2006. *Corynebacterium diphtheriae*: Insight Into New Virulence Properties. *Trends in Diphtheria Research*, 135.
- Bankaitis, V. A., Aitken, J. R., Cleves, A. E. & Dowhan, W. 1990. An essential role for a phospholipid transfer protein in yeast Golgi function.
- Barakat, A., Selim, S., Atef, A., Saber, M., Nafie, E. & El-Ebeedy, A. 1984. Two serotypes of *Corynebacterium pseudotuberculosis* isolated from different animal species. *Rev. Sci. Tech. Off. Int. Epiz*, 3, 151-168.
- Barba, M., Stewart, A., Passler, T., Wooldridge, A., Santen, E., Chamorro, M., Cattley, R., Hathcock, T., Hogsette, J. & Hu, X. 2015. Experimental transmission of *Corynebacterium pseudotuberculosis* biovar equi in horses by house flies. *Journal of veterinary internal medicine*, 29, 636-643.
- Barta, O. & Oyekan, P. P. 1981. Lymphocyte transformation test in veterinary clinical immunology. *Comparative immunology, microbiology and infectious diseases*, 4, 209-221.
- Baskerville, A., Hambleton, P. & Dowsett, A. 1978. The pathology of untreated and antibiotic-treated experimental tularaemia in monkeys. *British journal of experimental pathology*, 59, 615.
- Bastos, B. L., Loureiro, D., Raynal, J. T., Guedes, M. T., Vale, V. L., Moura-Costa, L. F., Guimarães, J. E., Azevedo, V., Portela, R. W. & Meyer, R. 2013. Association between haptoglobin and IgM levels and the clinical progression of caseous lymphadenitis in sheep. *BMC veterinary research*, 9, 254.
- Bastos, B. L., Meyer, R., Guimarães, J. E., Ayres, M. C., Guedes, M. T., Moura-Costa, L. F., Burghgrave, U. S., Sena, L., Azevedo, V. & Portela, R. W. 2011. Haptoglobin and fibrinogen concentrations and leukocyte counts in the clinical investigation of caseous lymphadenitis in sheep. *Veterinary Clinical Pathology*, 40, 496-503.
- Bastos, B. L., Portela, R. W., Dorella, F., Ribeiro, D., Seyffert, N., De Paula Castro, T., Miyoshi, A., Oliveira, S., Meyer, R. & Vasco, A. 2012. *Corynebacterium pseudotuberculosis*: immunological responses in animal models and zoonotic potential. *Journal of Clinical & Cellular Immunology*, S4, 1-15.

- Batey, R. 1986a. © Aspects Of Pathogenesis In A Mouse Model Of Infection By *Corynebacterium pseudotuberculosis*. *Australian Journal of Experimental Biology & Medical Science*, 64.
- Batey, R. 1986b. Frequency and consequence of caseous lymphadenitis in sheep and lambs slaughtered at a Western Australian abattoir. *American journal of veterinary research*, 47, 482-485.
- Batey, R. 1986c. Pathogenesis of caseous lymphadenitis in sheep and goats. *Australian Veterinary Journal*, 63, 269-272.
- Batey, R., Speed, C. & Kobes, C. 1986. Prevalence and distribution of caseous lymphadenitis in feral goats. *Australian veterinary journal*, 63, 33-36.
- Battini, M., Barbieri, S., Fioni, L. & Mattiello, S. 2015. Feasibility and validity of animal-based indicators for on-farm welfare assessment of thermal stress in dairy goats. *International journal of biometeorology*, 1-8.
- Battini, M., Vieira, A., Barbieri, S., Ajuda, I., Stilwell, G. & Mattiello, S. 2014. Invited review: Animal-based indicators for on-farm welfare assessment for dairy goats. *Journal of dairy science*, 97, 6625-6648.
- Beauregard, K. E., Lee, K.-D., Collier, R. J. & Swanson, J. A. 1997. pH-dependent perforation of macrophage phagosomes by listeriolysin O from *Listeria monocytogenes*. *The Journal of experimental medicine*, 186, 1159-1163.
- Beckmen, K. B. 2002. CRC Handbook of Marine Mammal Medicine. *Journal of Zoo and Wildlife Medicine*, 33, 178-179.
- Ben-Dov, E., Yosef, D. Z. B., Pavlov, V. & Kushmaro, A. 2009. *Corynebacterium maris* sp. nov., a marine bacterium isolated from the mucus of the coral *Fungia granulosa*. *International journal of systematic and evolutionary microbiology*, 59, 2458-2463.
- Ben, S. M., Ben, M. H., Benzarti, M., Messadi, L., Rejeb, A. & Amara, A. 2001. [Epidemiological and clinical studies of ovine caseous lymphadenitis]. *Archives de l'Institut Pasteur de Tunis*, 79, 51-57.
- Berger, A., Boschert, V., Konrad, R., Schmidt-Wieland, T., Hörmansdorfer, S., Eddicks, M. & Sing, A. 2013. Two cases of cutaneous diphtheria associated with occupational pig contact in Germany. *Zoonoses and public health*, 60, 539-542.
- Bergman, R. A., Afifi, A. K., Jew, J. & Reimann, P. 2004. Anatomy atlases. *Atlas of Microscopic Anatomy- A Functional Approach: section 9. Lymphatic System*
- Bermudez, L. E. & Goodman, J. 1996. *Mycobacterium tuberculosis* invades and replicates within type II alveolar cells. *Infection and immunity*, 64, 1400-1406.

- Bernard, K. A., Wiebe, D., Burdz, T., Reimer, A., Ng, B., Singh, C., Schindle, S. & Pacheco, A. L. 2010. Assignment of *Brevibacterium stationis* (ZoBell and Upham 1944) Breed 1953 to the genus *Corynebacterium*, as *Corynebacterium stationis* comb. nov., and emended description of the genus *Corynebacterium* to include isolates that can alkalize citrate. *International journal of systematic and evolutionary microbiology*, 60, 874-879.
- Binns, S., Bailey, M. & Green, L. 2002. Postal survey of ovine caseous lymphadenitis in the United Kingdom between 1990 and 1999. *The Veterinary record*, 150, 263-268.
- Binns, S. H., Green, L. E. & Bailey, M. 2007. Development and validation of an ELISA to detect antibodies to *Corynebacterium pseudotuberculosis* in ovine sera. *Veterinary microbiology*, 123, 169-179.
- Björck, L., Collin, M., Olsén, A., Holmdahl, R., Nandakumar, K. S. & Shannon, O. 2014. Use of the endoglycosidase endos for treating immunoglobulin G mediated diseases. Google Patents.
- Blood, D., Radostits, O., Gay, C., Hinchcliff, K. & Constable, P. 2007. *Veterinary Medicine: A textbook of the diseases of cattle, horses, sheep, pigs and goats*. Saunders Ltd.
- Blowey, R. & Weaver, A. D. 2011. *Color atlas of diseases and disorders of cattle*, Elsevier Health Sciences.
- Bode, J. G., Albrecht, U., Häussinger, D., Heinrich, P. C. & Schaper, F. 2012. Hepatic acute phase proteins—regulation by IL-6-and IL-1-type cytokines involving STAT3 and its crosstalk with NF-κB-dependent signaling. *European journal of cell biology*, 91, 496-505.
- Bogdan, J., Newlands-Monteith, C. & Ellis, J. 1997. Nitric oxide production following in vitro stimulation of ovine pulmonary alveolar macrophages. *Veterinary immunology and immunopathology*, 56, 299-310.
- Bouisset, L., Breuillaud, J. & Michel, G. [Study of DNA in Actinomycetales. Comparison between the values of the A&T/G&C relationship and the bacteriological propof *Corynebacterium*.]. *Annales de l'Institut Pasteur*, 1963. 756-770.
- Braga, W., Schul, S., Nuñez, A., Pezo, D. & Franco, E. 2007. A primary *Corynebacterium pseudotuberculosis* low dose infection in alpacas (*Lama pacos*) protects against a lethal challenge exposure. *Small Ruminant Research*, 72, 81-86.
- Bregenzer, T., Frei, R., Ohnacker, H. & Zimmerli, W. 1997. *Corynebacterium pseudotuberculosis* infection in a butcher. *Clinical Microbiology and Infection*, 3, 696-698.
- Brennan, P. 1988. Mycobacterium and other actinomycetes. *Microbial lipids*, 1, 203-298.

- Brogden, K. A., Chedid, L., Cutlip, R., Lehmkuhl, H. & Sacks, J. 1990. Effect of muramyl dipeptide on immunogenicity of *Corynebacterium pseudotuberculosis* whole-cell vaccines in mice and lambs. *American journal of veterinary research*, 51, 200-202.
- Brown, C. & Olander, H. 1987. Caseous lymphadenitis of goats and sheep: a review. *Veterinary Bulletin*, 57, 1-11.
- Brown, C., Olander, H., Biberstein, E. & Morse, S. 1986a. Use of a toxoid vaccine to protect goats against intradermal challenge exposure to *Corynebacterium pseudotuberculosis*. *American journal of veterinary research*, 47, 1116-1119.
- Brown, C. C., Olander, H. J. & Alves, S. F. 1987. Synergistic hemolysis-inhibition titers associated with caseous lymphadenitis in a slaughterhouse survey of goats and sheep in Northeastern Brazil. *Canadian Journal of Veterinary Research*, 51, 46.
- Brown, C. C., Olander, H. J., Zometa, C. & Alves, S. F. 1986b. Serodiagnosis of inapparent caseous lymphadenitis in goats and sheep, using the synergistic hemolysis-inhibition test. *American Journal of Veterinary Research*, 47, 1461-1463.
- Burdon, K. L. 1946. Fatty material in bacteria and fungi revealed by staining dried, fixed slide preparations. *Journal of bacteriology*, 52, 665.
- Burgess, G. 1982. Ovine contagious epididymitis: a review. *Veterinary Microbiology*, 7, 551-575.
- Burkovski, A. 2013. Cell envelope of corynebacteria: structure and influence on pathogenicity. *ISRN microbiology*, 2013.
- Burrell, D. 1980. A simplified double immunodiffusion technique for detection of *Corynebacterium ovis* antitoxin. *Research in Veterinary Science*, 28, 234-237.
- Burrell, D. 1981. Caseous lymphadenitis in goats. *Australian Veterinary Journal*, 57, 105-110.
- Bush, R., Barnett, R., Links, I. & Windsor, P. 2012. Using abattoir surveillance and producer surveys to investigate the prevalence and current preventative management of Caseous lymphadenitis in Merino flocks in Australia. *Animal production science*, 52, 675-679.
- Butler, W., Ahearn, D. & Kilburn, J. 1986. High-performance liquid chromatography of mycolic acids as a tool in the identification of *Corynebacterium*, *Nocardia*, *Rhodococcus*, and *Mycobacterium* species. *Journal of clinical microbiology*, 23, 182-185.
- Buttenschoen, K., Buttenschoen, D. C., Berger, D., Vasilescu, C., Schafheutle, S., Goeltenboth, B., Seidelmann, M. & Beger, H. G. 2001. Endotoxemia and acute-phase proteins in major abdominal surgery. *The American journal of surgery*, 181, 36-43.

- Byrne, M. L., O'Brien-Simpson, N. M., Reynolds, E. C., Walsh, K. A., Laughton, K., Waloszek, J. M., Woods, M. J., Trinder, J. & Allen, N. B. 2013. Acute phase protein and cytokine levels in serum and saliva: a comparison of detectable levels and correlations in a depressed and healthy adolescent sample. *Brain, behavior, and immunity*, 34, 164-175.
- Calabró, P., Willerson, J. T. & Yeh, E. T. 2003. Inflammatory cytokines stimulated C-reactive protein production by human coronary artery smooth muscle cells. *Circulation*, 108, 1930-1932.
- Cameron, C. M. & Engelbrecht, M. M. 1971. Mechanism of immunity to *Corynebacterium pseudotuberculosis* (Buchanan, 1911) in mice using inactivated vaccine.
- Carne, H. 1939. A bacteriological study of 134 strains of *Corynebacterium ovis*. *The Journal of Pathology and Bacteriology*, 49, 313-328.
- Carrington, C. B., Addington, W. W., Goff, A. M., Madoff, I. M., Marks, A., Schwaber, J. R. & Gaensler, E. A. 1969. Chronic eosinophilic pneumonia. *New England journal of medicine*, 280, 787-798.
- Carter, G. R. & Cole Jr, J. R. 2012. *Diagnostic procedure in veterinary bacteriology and mycology*, Academic Press.
- Casadevall, A. & Pirofski, L.-A. 1999. Host-pathogen interactions: redefining the basic concepts of virulence and pathogenicity. *Infection and immunity*, 67, 3703-3713.
- Cerdeño-Tárraga, A., Efstratiou, A., Dover, L., Holden, M., Pallen, M., Bentley, S., Besra, G., Churcher, C., James, K. & De Zoysa, A. 2003. The complete genome sequence and analysis of *Corynebacterium diphtheriae* NCTC13129. *Nucleic acids research*, 31, 6516-6523.
- Cerón, J. J., Eckersall, P. D. & Martínez-Subiela, S. 2005. Acute phase proteins in dogs and cats: current knowledge and future perspectives. *Veterinary Clinical Pathology*, 34, 85-99.
- Çetinkaya, B., Karahan, M., Atil, E., Kalin, R., De Baere, T. & Vaneechoutte, M. 2002. Identification of *Corynebacterium pseudotuberculosis* isolates from sheep and goats by PCR. *Veterinary microbiology*, 88, 75-83.
- Chandrawathani, P. 1998. Control of gastrointestinal helminths in small ruminants the Malaysian perspective. *FAO Animal Production and Health Paper*, 78-81.
- Chandrawathani, P., Adnan, M. & Waller, P. 1999. Anthelmintic resistance in sheep and goat farms on Peninsular Malaysia. *Veterinary Parasitology*, 82, 305-310.
- Chandrawathani, P., Waller, P., Adnan, M. & Höglund, J. 2003. Evolution of high-level, multiple anthelmintic resistance on a sheep farm in Malaysia. *Tropical animal health and production*, 35, 17-25.

- Chaplin, P. J., De Rose, R., Boyle, J. S., Mcwaters, P., Kelly, J., Tennent, J. M., Lew, A. M. & Scheerlinck, J.-P. Y. 1999. Targeting improves the efficacy of a DNA vaccine against *Corynebacterium pseudotuberculosis* in sheep. *Infection and immunity*, 67, 6434-6438.
- Chirino-Zárraga, C., Scaramelli, A. & Rey-Valeirón, C. 2006. Bacteriological characterization of *Corynebacterium pseudotuberculosis* in Venezuelan goat flocks. *Small Ruminant Research*, 65, 170-175.
- Cocco, E., Bellone, S., El-Sahwi, K., Cargnelutti, M., Buza, N., Tavassoli, F. A., Schwartz, P. E., Rutherford, T. J., Pecorelli, S. & Santin, A. D. 2010. Serum amyloid A. *Cancer*, 116, 843-851.
- Cole, S. 1996. Rifamycin resistance in mycobacteria. *Research in microbiology*, 147, 48-52.
- Collett, M., Bath, G. & Cameron, C. 1994. *Corynebacterium pseudotuberculosis* infections. *Infections diseases of livestock with special reference to Southern Africa*. Oxford University Press, 2, 1387-1395.
- Collins, F. & Campbell, S. 1982. Immunity to intracellular bacteria. *Veterinary immunology and immunopathology*, 3, 5-66.
- Collins, M., Goodfellow, M. & Minnikin, D. 1982. A survey of the structures of mycolic acids in *Corynebacterium* and related taxa. *Microbiology*, 128, 129-149.
- Collins, M. D., Burton, R. A. & Jones, D. 1988. *Corynebacterium amycolatum* sp. nov. a new mycolic acid-less *Corynebacterium* species from human skin. *FEMS microbiology letters*, 49, 349-352.
- Colom-Cadena, A., Velarde, R., Salinas, J., Borge, C., García-Bocanegra, I., Serrano, E., Gassó, D., Bach, E., Casas-Díaz, E. & López-Olvera, J. R. 2014. Management of a caseous lymphadenitis outbreak in a new Iberian ibex (*Capra pyrenaica*) stock reservoir. *Acta Veterinaria Scandinavica*, 56, 1-11.
- Concha-Bermejillo, A. D. L., Singh, B., Whitney, M. S. & Bazer, F. W. 2000. Acute-phase proteins and hematologic values in ovine lentivirus-infected lambs treated with recombinant ovine IFN-tau. *Journal of Interferon & Cytokine Research*, 20, 41-53.
- Conn, H. & Dimmick, I. 1947. Soil bacteria similar in morphology to *Mycobacterium* and *Corynebacterium*. *Journal of bacteriology*, 54, 291.
- Connor, K. M., Fontaine, M. C., Rudge, K., Baird, G. J. & Donachie, W. 2007. Molecular genotyping of multinational ovine and caprine *Corynebacterium pseudotuberculosis* isolates using pulsed-field gel electrophoresis. *Veterinary research*, 38, 613-623.
- Connor, K. M., Quirie, M. M., Baird, G. & Donachie, W. 2000. Characterization of United Kingdom Isolates of *Corynebacterium pseudotuberculosis* Using Pulsed-Field Gel Electrophoresis. *Journal of clinical microbiology*, 38, 2633-2637.

- Cooke, M. M. 2000. Pathogenesis of tuberculosis in the brushtail possum *Trichosurus vulpecula*: a thesis presented in partial fulfilment of the requirements for the degree of Doctor of Philosophy at Massey University, Palmerston North, New Zealand.
- Cooper, G. M. & Hausman, R. E. 2000. *The cell*, Sinauer Associates Sunderland.
- Corteel, J., Ricordeau, G. G., Mason, I., Gall, C. G., Huirne, R., Dijkhuizen, A., Renkena, J., Gargamo, A., Adúriz, M. & Saldungaray, M. 1981. Collection, processing and artificial insemination of goat semen. Goat production. Universidad Central de Venezuela, Caracas (Venezuela). Facultad de Agronomía. Instituto de Producción Animal. Programa de Postgrado en Producción Animal.
- Costa, L. R., Spier, S. J. & Hirsh, D. C. 1998. Comparative molecular characterization of *Corynebacterium pseudotuberculosis* of different origin. *Veterinary Microbiology*, 62, 135-143.
- Cray, C., Zaias, J. & Altman, N. H. 2009. Acute phase response in animals: a review. *Comparative medicine*, 59, 517-526.
- Cristobal, G. & Deepak, C. 2010. Actin-Binding Proteins and Disease.
- Da Ca Sá, M., Veschi, J. L., Santos, G. B., Amanso, E. S., Oliveira, S. A., Mota, R. A., Veneroni-Gouveia, G. & Costa, M. M. 2013. Atividade de desinfetantes e produção de biofilme em *Corynebacterium pseudotuberculosis*. *Pesquisa Veterinária Brasileira*, 33, 1319-1324.
- Da Silva, G. N. & Holt, J. G. 1965. Numerical taxonomy of certain coryneform bacteria. *Journal of bacteriology*, 90, 921-927.
- Damron, W. S. 2013. *Introduction to animal science: global, biological, social, and industry perspectives*.
- Daniel, A. 2014. Isolation and Identification of *Corynebacterium pseudotuberculosis* From Superficial Lymph nodes of Sheep and Goat at Organic Export Abattoir, Modjo Ethiopia. PhD Dessertation, Addis Ababa University, Ethiopia.
- Davenport, R., Pickering, R., Goodhead, A. & Curtis, T. 2008. A universal threshold concept for hydrophobic mycolata in activated sludge foaming. *Water research*, 42, 3446-3454.
- Davis, E. 1990. *Corynebacterium pseudotuberculosis* infections in animals.
- Dawson, D. R., Nydam, D. V., Price, C. T., Graham, J. E., Cynamon, M. H., Divers, T. J. & Felipe, M. J. B. 2011. Effects of opsonization of *Rhodococcus equi* on bacterial viability and phagocyte activation. *American journal of veterinary research*, 72, 1465-1475.
- De Briel, D., Couderc, F., Riegel, P., Jehl, F. & Minck, R. 1992. High-performance liquid chromatography of corynomycolic acids as a tool in identification of *Corynebacterium* species and related organisms. *Journal of clinical microbiology*, 30, 1407-1417.

- De Castro Demoner, L., Ramos, J. N., Baio, P. V. P., Simpson-Louredo, L., Santos, C. S., Hirata Jr, R., Ferioli, R. B., Romera, A. R. C., Vieira, V. V. & Mattos-Guaraldi, A. L. 2015. Cutaneous abscess caused by *Corynebacterium lactis* in a companion dog.
- De Rose, R., Tennent, J., Mcwaters, P., Chaplin, P. J., Wood, P. R., Kimpton, W., Cahill, R. & Scheerlinck, J.-P. Y. 2002. Efficacy of DNA vaccination by different routes of immunisation in sheep. *Veterinary immunology and immunopathology*, 90, 55-63.
- De Sá Guimarães, A., Do Carmo, F. B., Pauletti, R. B., Seyffert, N., Ribeiro, D., Lage, A. P., Heinemann, M. B., Miyoshi, A., Azevedo, V. & Gouveia, A. M. G. 2011. Caseous lymphadenitis: epidemiology, diagnosis, and control. *IIOAB Journal*, 2, 33-43.
- Degousee, N., Ghomashchi, F., Stefanski, E., Singer, A., Smart, B. P., Borregaard, N., Reithmeier, R., Lindsay, T. F., Lichtenberger, C. & Reinisch, W. 2002. Groups IV, V, and X Phospholipases A2s in Human Neutrophils ROLE IN EICOSANOID PRODUCTION AND GRAM-NEGATIVE BACTERIAL PHOSPHOLIPID HYDROLYSIS. *Journal of Biological Chemistry*, 277, 5061-5073.
- Der Valk, V. & Meijer, C. 1987. The histology of reactive lymph nodes. *The American journal of surgical pathology*, 11, 866-882.
- Dercksen, D. P., Brinkhof, J. M., Dekker-Nooren, T., Van Maanen, K., Bode, C. F., Baird, G. & Kamp, E. M. 2000. A comparison of four serological tests for the diagnosis of caseous lymphadenitis in sheep and goats. *Veterinary Microbiology*, 75, 167-175.
- Devaney, K. O., Travis, W. D., Hoffman, G., Leavitt, R., Lebovics, R. & Fauci, A. S. 1990. Interpretation of head and neck biopsies in Wegener's granulomatosis: a pathologic study of 126 biopsies in 70 patients. *The American journal of surgical pathology*, 14, 555-564.
- Devendra, C. 1999. Goats: Challenges for increased productivity and improved livelihoods. *Outlook on Agriculture*, 28, 215-226.
- Dierauf, L. & Gulland, F. M. 2001. *CRC handbook of marine mammal medicine: health, disease, and rehabilitation*, CRC press.
- Doherr, M., Carpenter, T., Hanson, K., Wilson, W. & Gardner, I. 1998a. Risk factors associated with *Corynebacterium pseudotuberculosis* infection in California horses. *Preventive veterinary medicine*, 35, 229-239.
- Doherr, M., Carpenter, T., Wilson, W. & Gardner, I. 1998b. Application and evaluation of a mailed questionnaire for an epidemiologic study of *Corynebacterium pseudotuberculosis* infection in horses. *Preventive veterinary medicine*, 35, 241-253.
- Doherr, M., Carpenter, T., Wilson, W. & Gardner, I. 1999. Evaluation of temporal and spatial clustering of horses with *Corynebacterium pseudotuberculosis* infection. *American journal of veterinary research*, 60, 284-291.

- Dorella, F., Estevam, E., Cardoso, P., Savassi, B., Oliveira, S., Azevedo, V. & Miyoshi, A. 2006a. An improved protocol for electrotransformation of *Corynebacterium pseudotuberculosis*. *Veterinary Microbiology*, 114, 298-303.
- Dorella, F. A., Pacheco, L. G., Seyffert, N., Portela, R. W., Meyer, R., Miyoshi, A. & Azevedo, V. 2009. Antigens of *Corynebacterium pseudotuberculosis* and prospects for vaccine development. *Expert Review of Vaccines*, 8, 205-213.
- Dorella, F. A., Pacheco, L. G. C., Oliveira, S. C., Miyoshi, A. & Azevedo, V. 2006b. *Corynebacterium pseudotuberculosis*: microbiology, biochemical properties, pathogenesis and molecular studies of virulence. *Veterinary research*, 37, 201-218.
- Dorny, P., Claerebout, E., Vercruysse, J., Sani, R. & Jalila, A. 1994. Anthelmintic resistance in goats in peninsular Malaysia. *Veterinary parasitology*, 55, 327-342.
- Dorny, P., Symoens, C., Jalila, A., Vercruysse, J. & Sani, R. 1995. Strongyle infections in sheep and goats under the traditional husbandry system in peninsular Malaysia. *Veterinary Parasitology*, 56, 121-136.
- Dos Remedios, C. & Chhabra, D. 2008. *Actin-binding proteins and disease*, Springer Science & Business Media.
- Dover, L. G., Cerdeño-Tárraga, A. M., Pallen, M. J., Parkhill, J. & Besra, G. S. 2004. Comparative cell wall core biosynthesis in the mycolated pathogens, *Mycobacterium tuberculosis* and *Corynebacterium diphtheriae*. *FEMS microbiology reviews*, 28, 225-250.
- Dowling, A., Hodgson, J., Dagleish, M., Eckersall, P. & Sales, J. 2004. Pathophysiological and immune cell responses in calves prior to and following lung challenge with formalin-killed *Pasteurella multocida* biotype A: 3 and protection studies involving subsequent homologous live challenge. *Veterinary Immunology and Immunopathology*, 100, 197-207.
- Dowling, A., Hodgson, J., Schock, A., Donachie, W., Eckersall, P. & Mckendrick, I. 2002. Experimental induction of pneumonic pasteurellosis in calves by intratracheal infection with *Pasteurella multocida* biotype A: 3. *Research in veterinary science*, 73, 37-44.
- Dowling, A. J., Wilkinson, P. A., Holden, M. T., Quail, M. A., Bentley, S. D., Reger, J., Waterfield, N. R. & Titball, R. W. 2010. Genome-wide analysis reveals loci encoding anti-macrophage factors in the human pathogen *Burkholderia pseudomallei* K96243. *PLoS ONE*, 5, e15693.
- Dubnau, E., Chan, J., Raynaud, C., Mohan, V. P., Lanéelle, M. A., Yu, K., Quémard, A., Smith, I. & Daffé, M. 2000. Oxygenated mycolic acids are necessary for virulence of *Mycobacterium tuberculosis* in mice. *Molecular microbiology*, 36, 630-637.

- Dvs 2011. "Penyakit Bisul Nodus Limfa (Cla)" In Protokol Veterinar Malaysia. In: Ministry Of Agriculture And Agro-Based Industry Department Of Veterinary Services, M. (Ed.). Malaysia.
- Eckersall, P. 2000. Recent advances and future prospects for the use of acute phase proteins as markers of disease in animals. *Revue de medecine veterinaire*, 151, 577-584.
- Eckersall, P. & Bell, R. 2010. Acute phase proteins: Biomarkers of infection and inflammation in veterinary medicine. *The Veterinary Journal*, 185, 23-27.
- Eckersall, P. D., Gow, J. W., McComb, C., Bradley, B., Rodgers, J., Murray, M. & Kennedy, P. G. 2001. Cytokines and the acute phase response in post-treatment reactive encephalopathy of *Trypanosoma brucei* brucei infected mice. *Parasitology international*, 50, 15-26.
- Eckersall, P. D., Lawson, F. P., Bence, L., Waterston, M. M., Lang, T. L., Donachie, W. & Fontaine, M. C. 2007. Acute phase protein response in an experimental model of ovine caseous lymphadenitis. *BMC Veterinary Research*, 3, 35-40.
- Efstathiou, A. & George, R. 1999. Laboratory guidelines for the diagnosis of infections caused by *Corynebacterium diphtheriae* and *C. ulcerans*. *Communicable Disease and Public Health*, 2, 250-257.
- Eggleton, D., Doidge, C., Middleton, H. & Minty, D. 1991a. Immunisation against ovine caseous lymphadenitis: efficacy of monocomponent *Corynebacterium pseudotuberculosis* toxoid vaccine and combined clostridial-corynebacterial vaccines. *Australian veterinary journal*, 68, 320-321.
- Eggleton, D., Haynes, J., Middleton, H. & Cox, J. 1991b. Immunisation against ovine caseous lymphadenitis: correlation between *Corynebacterium pseudotuberculosis* toxoid content and protective efficacy in combined clostridial-corynebacterial vaccines. *Australian veterinary journal*, 68, 322-325.
- Eggleton, D., Middleton, H., Doidge, C. & Minty, D. 1991c. Immunisation against ovine caseous lymphadenitis: comparison of *Corynebacterium pseudotuberculosis* vaccines with and without bacterial cells. *Australian veterinary journal*, 68, 317-319.
- Ellis, J. 1988. Immunophenotype of pulmonary cellular infiltrates in sheep with visceral caseous lymphadenitis. *Veterinary Pathology Online*, 25, 362-368.
- Ellis, J., Hawk, D., Holler, L., Mills, K. & Pratt, D. 1990. Differential antibody responses to *Corynebacterium pseudotuberculosis* in sheep with naturally acquired caseous lymphadenitis. *Journal of the American Veterinary Medical Association*, 196, 1609-1613.

- Ellis, J., Russell, H. & Du, C. 1994. Effect of selected cytokines on the replication of *Corynebacterium pseudotuberculosis* and ovine lentiviruses in pulmonary macrophages. *Veterinary immunology and immunopathology*, 40, 31-47.
- Ellis, T., Sutherland, S., Wilkinson, F., Mercy, A. & Paton, M. 1987. The role of *Corynebacterium pseudotuberculosis* lung lesions in the transmission of this bacterium to other sheep. *Australian veterinary journal*, 64, 261-263.
- Elmore, S. A. 2006. Histopathology of the lymph nodes. *Toxicologic pathology*, 34, 425-454.
- Enwonwu, C. O. & Ritchie, C. S. 2007. Nutrition and inflammatory markers. *The Journal of the American Dental Association*, 138, 70-73.
- Eppleston, J. & Windsor, P. 2007. Lesions attributed to vaccination of sheep with Gudair™ for the control of ovine paratuberculosis: post farm economic impacts at slaughter. *Australian veterinary journal*, 85, 129-133.
- Epstein, F. H., Gabay, C. & Kushner, I. 1999. Acute-phase proteins and other systemic responses to inflammation. *New England Journal of Medicine*, 340, 448-454.
- Ernst, J. D. 1998. Macrophage receptors for *Mycobacterium tuberculosis*. *Infection and immunity*, 66, 1277-1281.
- Facchetti, F., Agostini, C., Grigolato, P., Chilosi, M., Mombello, A. & Van Den Oord, J. J. 1992. Suppurative Granulomatous Lymphadenitis: Immunohistochemical Evidence for a B-cell-Associated Granuloma. *The American journal of surgical pathology*, 16, 955-956.
- Farooqui, A. A. & Horrocks, L. A. 2006. *Glycerophospholipids in the brain: phospholipases A2 in neurological disorders*, Springer Science & Business Media.
- Farstvedt, E. G., Hendrickson, D. A., Dickenson, C. E. & Spier, S. J. 2004. Treatment of suppurative facial cellulitis and panniculitis caused by *Corynebacterium pseudotuberculosis* in two horses. *Journal of the American Veterinary Medical Association*, 224, 1139-1142.
- Fearon, D. T. & Locksley, R. M. 1996. The instructive role of innate immunity in the acquired immune response. *Science*, 272, 50-54.
- Foley, J. E., Spier, S. J., Mihalyi, J., Drazenovich, N. & Leutenegger, C. M. 2004. Molecular epidemiologic features of *Corynebacterium pseudotuberculosis* isolated from horses. *American journal of veterinary research*, 65, 1734-1737.
- Fontaine, M. & Baird, G. 2008. Caseous lymphadenitis. *Small Ruminant Research*, 76, 42-48.

- Fontaine, M. C., Baird, G., Connor, K. M., Rudge, K., Sales, J. & Donachie, W. 2006. Vaccination confers significant protection of sheep against infection with a virulent United Kingdom strain of *Corynebacterium pseudotuberculosis*. *Vaccine*, 24, 5986-5996.
- Foreman-Wykert, A. K., Weinrauch, Y., Elsbach, P. & Weiss, J. 1999. Cell-wall determinants of the bactericidal action of group IIA phospholipase A 2 against Gram-positive bacteria. *The Journal of clinical investigation*, 103, 715-721.
- Fournier, D., Campbell, J. R. & Middleton, D. M. 2006. Prevalence of maedi-visna infection in culled ewes in Alberta. *Canadian veterinary journal*, 47, 460.
- Frehel, C., De Chastellier, C., Lang, T. & Rastogi, N. 1986. Evidence for inhibition of fusion of lysosomal and prelysosomal compartments with phagosomes in macrophages infected with pathogenic *Mycobacterium avium*. *Infection and immunity*, 52, 252-262.
- Fudou, R., Jojima, Y., Seto, A., Yamada, K., Kimura, E., Nakamatsu, T., Hiraishi, A. & Yamanaka, S. 2002. *Corynebacterium efficiens* sp. nov., a glutamic-acid-producing species from soil and vegetables. *International journal of systematic and evolutionary microbiology*, 52, 1127-1131.
- Gaillard, J.-L., Berche, P., Mounier, J., Richard, S. & Sansonetti, P. 1987. In vitro model of penetration and intracellular growth of *Listeria monocytogenes* in the human enterocyte-like cell line Caco-2. *Infection and immunity*, 55, 2822-2829.
- Galili, U. & Schlesinger, M. 1974. The formation of stable E rosettes after neuraminidase treatment of either human peripheral blood lymphocytes or of sheep red blood cells. *The Journal of Immunology*, 112, 1628-1634.
- Gameel, A. & Tartour, G. 1974. Haematological and plasma protein changes in sheep experimentally infected with *Corynebacterium pseudotuberculosis*. *Journal of comparative pathology*, 84, 477-484.
- Garçon, N., Leroux-Roels, G. & Cheng, W.-F. 2011. Vaccine adjuvants. *Perspectives in vaccinology*, 1, 89-113.
- Gates, N., Everson, D. & Hulet, C. 1977. Effects of thin ewe syndrome on reproductive efficiency. *Journal of the American Veterinary Medical Association*, 171, 1266-1267.
- Gauldie, J., Richards, C., Harnish, D., Lansdorp, P. & Baumann, H. 1987. Interferon beta 2/B-cell stimulatory factor type 2 shares identity with monocyte-derived hepatocyte-stimulating factor and regulates the major acute phase protein response in liver cells. *Proceedings of the National Academy of Sciences*, 84, 7251-7255.
- George-Gay, B. & Parker, K. 2003. Understanding the complete blood count with differential. *Journal of PeriAnesthesia Nursing*, 18, 96-117.

- Godson, D. L., Campos, M., Attah-Poku, S. K., Redmond, M. J., Cordeiro, D. M., Sethi, M. S., Harland, R. J. & Babiuk, L. A. 1996. Serum haptoglobin as an indicator of the acute phase response in bovine respiratory disease. *Veterinary Immunology and Immunopathology*, 51, 277-292.
- Goldberger, A., Lipsky, B. & Plorde, J. 1981. Suppurative granulomatous lymphadenitis caused by *Corynebacterium ovis* (pseudotuberculosis). *American journal of clinical pathology*, 76, 486-490.
- Goldstein, E. J., Citron, D. M., Merriam, C. V., Warren, Y., Tyrrell, K. & Fernandez, H. T. 2003. In vitro activities of dalbavancin and nine comparator agents against anaerobic gram-positive species and corynebacteria. *Antimicrobial agents and chemotherapy*, 47, 1968-1971.
- Goldstein, E. J., Citron, D. M., Merriam, C. V., Warren, Y. A., Tyrrell, K. L. & Fernandez, H. T. 2004. In vitro activities of the new semisynthetic glycopeptide telavancin (TD-6424), vancomycin, daptomycin, linezolid, and four comparator agents against anaerobic gram-positive species and *Corynebacterium* spp. *Antimicrobial agents and chemotherapy*, 48, 2149-2152.
- Goldstein, E. J., Citron, D. M., Warren, Y. A., Tyrrell, K. L., Merriam, C. V. & Fernandez, H. T. 2006. In vitro activities of dalbavancin and 12 other agents against 329 aerobic and anaerobic gram-positive isolates recovered from diabetic foot infections. *Antimicrobial agents and chemotherapy*, 50, 2875-2879.
- Gomes, M. G. M., White, L. J. & Medley, G. F. 2004. Infection, reinfection, and vaccination under suboptimal immune protection: epidemiological perspectives. *Journal of Theoretical Biology*, 228, 539-549.
- Gómez-Muñoz, A., Kong, J. Y., Salh, B. & Steinbrecher, U. P. 2004. Ceramide-1-phosphate blocks apoptosis through inhibition of acid sphingomyelinase in macrophages. *Journal of lipid research*, 45, 99-105.
- Gonzalez-Suarez, M. L., Salinas-Carmona, M. C. & Pérez-Rivera, I. 2009. IgM but not IgG monoclonal anti-*Nocardia brasiliensis* antibodies confer protection against experimental actinomycetoma in BALB/c mice. *FEMS Immunology & Medical Microbiology*, 57, 17-24.
- González, F. H., Tecles, F., Martínez-Subiela, S., Tvarijonaviciute, A., Soler, L. & Cerón, J. J. 2008. Acute phase protein response in goats. *Journal of Veterinary Diagnostic Investigation*, 20, 580-584.
- Gordon, E. D. Progressive Wasting Diseases of Ewes. Dairy Sheep Association of North America Symposium, 2012. 64.
- Gorman, J. K., Gabriel, M., Maclachlan, N. J., Nieto, N. C., Foley, J. E. & Spier, S. J. 2010. Pilot immunization of mice infected with an equine strain of *Corynebacterium pseudotuberculosis*. *Vet Ther*, 11, 1-8.

- Gotoh, K., Mitsuyama, M., Imaizumi, S., Kawamura, I. & Yano, I. 1991. Mycolic Acid-Containing Glycolipid as a Possible Virulence Factor of *Rhodococcus equi* for Mice. *Microbiology and immunology*, 35, 175-185.
- Groman, N., Schiller, J. & Russell, J. 1984. *Corynebacterium ulcerans* and *Corynebacterium pseudotuberculosis* responses to DNA probes derived from coryneophage beta and *Corynebacterium diphtheriae*. *Infection and immunity*, 45, 511-517.
- Gruys, E., Toussaint, M., Niewold, T. & Koopmans, S. 2005. Acute phase reaction and acute phase proteins. *J Zhejiang Univ Sci B*, 6, 1045-1056.
- Gutierrez, A., Ceron, J., Fuentes, P., Montes, A. & Martinez-Subiela, S. 2012. Longitudinal analysis of acute-phase proteins in saliva in pig farms with different health status. *animal*, 6, 321-326.
- Gyles, C. L., Prescott, J. F., Songer, J. G. & Thoen, C. O. 2011. *Pathogenesis of bacterial infections in animals*, John Wiley & Sons.
- Hall, K., McCluskey, B. J. & Cunningham, W. 2001. *Corynebacterium pseudotuberculosis* infections (Pigeon Fever) in horses in Western Colorado: An epidemiological investigation. *Journal of Equine Veterinary Science*, 21, 284-286.
- Hall, T. A. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, 1999. 95-98.
- Hard, G. 1972. Examination by electron microscopy of the interaction between peritoneal phagocytes and *Corynebacterium ovis*. *Journal of Medical Microbiology*, 5, 483-491.
- Hard, G. C. 1975. Comparative toxic effect of the surface lipid of *Corynebacterium ovis* on peritoneal macrophages. *Infection and immunity*, 12, 1439-1449.
- Hassan, S. S., Guimarães, L. C., Pereira, U. D. P., Islam, A., Ali, A., Bakhtiar, S. M., Ribeiro, D., Rodrigues Dos Santos, A., Soares, S. D. C. & Dorella, F. 2012. Complete genome sequence of *Corynebacterium pseudotuberculosis* biovar ovis strain P54B96 isolated from antelope in South Africa obtained by Rapid Next Generation Sequencing Technology. *Standards in genomic sciences*, 7.
- Hatanaka, E., Ribeiro, F. P. & Campa, A. 2003. The acute phase protein serum amyloid A primes neutrophils. *FEMS Immunology & Medical Microbiology*, 38, 81-84.
- Hauser, D., Popoff, M., Kiredjian, M., Boquet, P. & Bimet, F. 1993. Polymerase chain reaction assay for diagnosis of potentially toxinogenic *Corynebacterium diphtheriae* strains: correlation with ADP-ribosylation activity assay. *Journal of clinical microbiology*, 31, 2720-2723.

- Hawkins, P. N., Lavender, J. P. & Pepys, M. B. 1990. Evaluation of systemic amyloidosis by scintigraphy with 123I-labeled serum amyloid P component. *New England journal of medicine*, 323, 508-513.
- Heggelund, L., Gaustad, P., Håvelsrud, O. E., Blom, J., Borgen, L., Sundset, A., Sørum, H. & Frøland, S. S. *Corynebacterium pseudotuberculosis* pneumonia in a veterinary student infected during laboratory work. Open forum infectious diseases, 2015. Oxford University Press, ofv053.
- Henderson, A. 1979. Pseudotuberculous adenitis caused by *Corynebacterium pseudotuberculosis*. *Journal of Medical Microbiology*, 12, 147-150.
- Henkels, K. M., Boivin, G. P., Dudley, E. S., Berberich, S. J. & Gomez-Cambronero, J. 2013. Phospholipase D (PLD) drives cell invasion, tumor growth and metastasis in a human breast cancer xenograph model. *Oncogene*, 32, 5551-5562.
- Hinnebusch, J., Cherepanov, P., Du, Y., Rudolph, A., Dixon, J., Schwan, T. & Forsberg, Å. 2000. Murine toxin of *Yersinia pestis* shows phospholipase D activity but is not required for virulence in mice. *International journal of medical microbiology*, 290, 483-487.
- Hodgson, A., Krywult, J., Corner, L., Rothel, J. & Radford, A. 1992. Rational attenuation of *Corynebacterium pseudotuberculosis*: potential cheesy gland vaccine and live delivery vehicle. *Infection and immunity*, 60, 2900-2905.
- Hodgson, A., Tachedjian, M., Corner, L. A. & Radford, A. J. 1994. Protection of sheep against caseous lymphadenitis by use of a single oral dose of live recombinant *Corynebacterium pseudotuberculosis*. *Infection and immunity*, 62, 5275-5280.
- Hodgson, A. L., Carter, K., Tachedjian, M., Krywult, J., Corner, L. A., Mccoll, M. & Cameron, A. 1999. Efficacy of an ovine caseous lymphadenitis vaccine formulated using a genetically inactive form of the *Corynebacterium pseudotuberculosis* phospholipase D. *Vaccine*, 17, 802-808.
- Holliman, J. H. 1995. Respiratory Tract. *Pathology*. Springer.
- Hollowood, K. & Goodlad, J. R. 1998. Germinal centre cell kinetics. *The Journal of pathology*, 185, 229-233.
- Hommeze, J., Devriese, L. A., Vaneechoutte, M., Riegel, P., Butaye, P. & Haesebrouck, F. 1999. Identification of nonlipophilic corynebacteria isolated from dairy cows with mastitis. *Journal of clinical microbiology*, 37, 954-957.
- Hsu, T., Renshaw, H., Livingston Jr, C., Augustine, J., Zink, D. & Gauer, B. 1985. *Corynebacterium pseudotuberculosis* exotoxin: fatal hemolytic anemia induced in gnotobiotic neonatal small ruminants by parenteral administration of preparations containing exotoxin. *American journal of veterinary research*, 46, 1206-1211.

- Hucklebridge, F., Lambert, S., Clow, A., Warburton, D., Evans, P. D. & Sherwood, N. 2000. Modulation of secretory immunoglobulin A in saliva; response to manipulation of mood. *Biological Psychology*, 53, 25-35.
- Ibtisam, M. A. 2008. Some clinicopathological and pathological studies of *C. ovis* infection in sheep. *Egyptian Journal of Comparative Pathology and Clinical Pathology*, 21.
- Ionedá, T. 1989. Separation of homologues of methyl ester and 3-O-acetyl methyl ester derivatives of the corynomycolic acid fraction from *Corynebacterium pseudotuberculosis*. *Journal of Chromatography A*, 481, 411-415.
- Ionedá, T. & Silva, C. L. 1979. Purification of 1-monoacylglycerols containing α -branched- β -hydroxylated fatty acids from lipids of *Corynebacterium pseudotuberculosis*. *Chemistry and physics of lipids*, 25, 85-91.
- Istivan, T. S. & Coloe, P. J. 2006. Phospholipase A in Gram-negative bacteria and its role in pathogenesis. *Microbiology*, 152, 1263-1274.
- İzgür, M., Akan, M., İlhan, Z. & Yazıcıoğlu, N. 2010. Studies on vaccine development for ovine caseous lymphadenitis. *Vet J Ankara University*, 57, 161-165.
- Jack, R. W., Tagg, J. R. & Ray, B. 1995. Bacteriocins of gram-positive bacteria. *Microbiological reviews*, 59, 171-200.
- Jain, S., Gautam, V. & Naseem, S. 2011. Acute-phase proteins: As diagnostic tool. *Journal of Pharmacy and Bioallied Sciences*, 3, 118.
- Jain, S. K., Paul-Satyaseela, M., Lamichhane, G., Kim, K. S. & Bishai, W. R. 2006. *Mycobacterium tuberculosis* invasion and traversal across an in vitro human blood-brain barrier as a pathogenic mechanism for central nervous system tuberculosis. *Journal of Infectious Diseases*, 193, 1287-1295.
- Jalila, A., Dorny, P., Sani, R., Salim, N. & Vercruysse, J. 1998. Coccidial infections of goats in Selangor, peninsular Malaysia. *Veterinary parasitology*, 74, 165-172.
- Janeway Jr, C. A. & Medzhitov, R. 2002. Innate immune recognition. *Annual review of immunology*, 20, 197-216.
- Jeber, Z., Mohdjini, Z., Jesse, F., Saharee, A., Sabri, J., Yusoff, R. & Wahid, H. 2016. Influence of *Corynebacterium pseudotuberculosis* infection on level of acute phase proteins in goats. *BMC veterinary research*, 12, 1.
- Jelinek, P., Ellis, T., Wroth, R., Sutherland, S., Masters, H. & Petterson, D. 1988. The effect of selenium supplementation on immunity, and the establishment of an experimental *Haemonchus contortus* infection, in weaner Merino sheep fed a low selenium diet. *Australian veterinary journal*, 65, 214-217.

- Jensen, L. E., Hiney, M. P., Shields, D. C., Uhlar, C. M., Lindsay, A. J. & Whitehead, A. S. 1997. Acute phase proteins in salmonids: evolutionary analyses and acute phase response. *The Journal of Immunology*, 158, 384-392.
- Jensen, R., Maki, L., Lauerman, L., Rath, W., Swift, B., Flack, D., Hoff, R., Hancock, H., Tucker, J. & Horton, D. 1983. Cause and pathogenesis of middle ear infection in young feedlot cattle. *Journal of the American Veterinary Medical Association*, 182, 967-972.
- Jeong, Y. J. & Lee, K. S. 2008. Pulmonary tuberculosis: up-to-date imaging and management. *American Journal of Roentgenology*, 191, 834-844.
- Jeske, J. M., Spier, S. J., Whitcomb, M. B., Pusterla, N. & Gardner, I. A. 2013. Use of antibody titers measured via serum synergistic hemolysis inhibition testing to predict internal *Corynebacterium pseudotuberculosis* infection in horses. *Journal of the American Veterinary Medical Association*, 242, 86-92.
- Jesse, F., Randolph, P., Saharee, A., Wahid, A., Zamri-Saad, M., Jasni, S., Omar, A., Adamu, L. & Abdinasir, Y. 2013. Clinico-pathological response of mice following oral route infection of *C. pseudotuberculosis*. *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)*, 2, 38-42.
- Johnson, E., Al-Busaidy, R. & Hameed, M. 2003. An outbreak of lymphadenitis associated with *Stenotrophomonas* (*Xanthomonas*) *maltophilia* in Omani goats. *Journal of Veterinary Medicine, Series B*, 50, 102-104.
- Join-Lambert, O. F., Ouache, M., Canioni, D., Beretti, J.-L., Blanche, S., Berche, P. & Kayal, S. 2006. *Corynebacterium pseudotuberculosis* necrotizing lymphadenitis in a twelve-year-old patient. *The Pediatric infectious disease journal*, 25, 848-851.
- Jones, M. L. & Allison, R. W. 2007. Evaluation of the ruminant complete blood cell count. *Veterinary Clinics of North America: Food Animal Practice*, 23, 377-402.
- Jordan, D., Sentance, C., Spooner, W., Balan, J. & Morris, S. 2012. Inspection of lymph nodes for caseous lymphadenitis and its effect on the density of microbes on sheep carcasses. *Meat science*, 92, 837-840.
- Jost, B. & Billington, S. 2004. *Corynebacterium* and *Arcanobacterium*. *Pathogenesis of Bacterial Infections in Animals*, 77.
- Judson, R. & Songer, J. G. 1991. *Corynebacterium pseudotuberculosis*: in vitro susceptibility to 39 antimicrobial agents. *Veterinary Microbiology*, 27, 145-150.
- Jukes, T. H. & Cantor, C. R. 1969. Evolution of protein molecules. *Mammalian Protein Metabolism*, 3, 132.

- Junior, J. P., Oliveira, A., Alves, F., Silva, L., Rabelo, S. & Mota, R. 2006. *Corynebacterium pseudotuberculosis* Experimental Infection of Goats Mammary Gland. *Arquivos do Instituto Biologico, Sao Paulo*, 73, 395-400.
- Kahn, C. 2005. Merck veterinary manual, 9th edn. Whitehouse Station, NJ, USA: Merck and Co. Inc.
- Kasapis, C. & Thompson, P. D. 2005. The effects of physical activity on serum C-reactive protein and inflammatory markers: a systematic review. *Journal of the American College of Cardiology*, 45, 1563-1569.
- Kaur, B. 2010. Consumer Preference for Goat Meat in Malaysia: Market Opportunities and Potential. *Journal of Agribusiness Marketing*, Vol. 3, December 2010, p. 40-55.
- Kawata, K. 1958. Histo-Pathological Studies Of Caseous Pneumonia. *Pathology International*, 8, 153-175.
- Kay, A. 1970. Studies on eosinophil leucocyte migration. II. Factors specifically chemotactic for eosinophils and neutrophils generated from guinea-pig serum by antigen-antibody complexes. *Clinical and experimental immunology*, 7, 723.
- Kelsoe, G. 1995. In situ studies of the germinal center reaction. *Advances in immunology*, 60, 267-288.
- Kemp, T. J., Ludwig, A. T., Earel, J. K., Moore, J. M., Vanoosten, R. L., Moses, B., Leidal, K., Nauseef, W. M. & Griffith, T. S. 2005. Neutrophil stimulation with *Mycobacterium bovis* bacillus Calmette-Guerin (BCG) results in the release of functional soluble TRAIL/Apo-2L. *Blood*, 106, 3474-3482.
- Kent, C. 1995. Eukaryotic phospholipid biosynthesis. *Annual review of biochemistry*, 64, 315-343.
- Kent, J. 1992. Acute phase proteins: their use in veterinary diagnosis. *British Veterinary Journal*, 148, 279-282.
- Khadijah, S., Andy, T. F. H., Khadijah, S. S. a. K. & Khairi, A. K. M. 2014. Parasite Infection in Two Goat Farms Located in Kuala Terengganu, Peninsular Malaysia. *Asian Journal of Agriculture and Food Sciences*, 2, 463-468.
- Khuder, Z., Osman, A. Y., Jesse, F. F., Haron, A., Saharee, A., Sabri, J., Yusoff, R. & Abdullah, R. 2012. Sex hormone profiles and cellular changes of reproductive organs of mice experimentally infected with *C. pseudotuberculosis* and its exotoxin phospholipase D (PLD). *IOSR-JAVS*, 1, 24-9.
- Kimberling, C. V. & Parsons, G. 2008. *Raising healthy sheep*, Christian Veterinary Mission.
- Knights, M. & Garcia, G. 1997. The status and characteristics of the goat (*Capra hircus*) and its potential role as a significant milk producer in the tropics: a review. *Small Ruminant Research*, 26, 203-215.

- Komala, T., Ramlan, M., Yeoh, N., Surayani, A. & Sharifah Hamidah, S. 2008. A survey of caseous lymphadenitis in small ruminant farms from two districts in Perak, Malaysia-Kinta and Hilir Perak. *Trop Biomed*, 25, 196-201.
- Kramer, A., Schwebke, I. & Kampf, G. 2006. How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. *BMC infectious diseases*, 6, 1.
- Kristensen, F., Kristensen, B. & Lazary, S. 1982. The lymphocyte stimulation test in veterinary immunology. *Veterinary immunology and immunopathology*, 3, 203-277.
- Kroese, F., Timens, W. & Nieuwenhuis, P. 1990. Germinal center reaction and B lymphocytes: morphology and function. *Reaction Patterns of the lymph node*. Springer.
- Kubera, M., Basta-Kaim, A., Holan, V., Simbirtsev, A., Roman, A., Pigareva, N., Prokopieva, E. & Sham, J. 1998. Effect of mild chronic stress, as a model of depression, on the immunoreactivity of C57BL/6 mice. *International journal of immunopharmacology*, 20, 781-789.
- Kumar, J., Tripathi, B. N., Kumar, R., Sonawane, G. G. & Dixit, S. K. 2013. Rapid detection of *Corynebacterium pseudotuberculosis* in clinical samples from sheep. *Tropical animal health and production*, 45, 1429-1435.
- Kuria, J. & Holstad, G. 1988. A seroepidemiological investigation of *Corynebacterium pseudotuberculosis* infection in sheep flocks in southern Norway. *Acta Veterinaria Scandinavica*, 30, 107-108.
- Kuria, J., Mbuthia, P., Kang'ethe, E. & Wahome, R. 2001. Caseous lymphadenitis in goats: the pathogenesis, incubation period and serological response after experimental infection. *Veterinary research communications*, 25, 89-97.
- Kushner, I. 1993. Regulation of the acute phase response by cytokines. *Perspectives in biology and medicine*, 36, 611-622.
- Lagier, J.-C., Edouard, S., Pagnier, I., Mediannikov, O., Drancourt, M. & Raoult, D. 2015. Current and past strategies for bacterial culture in clinical microbiology. *Clinical microbiology reviews*, 28, 208-236.
- Langlois, M. R. & Delanghe, J. R. 1996. Biological and clinical significance of haptoglobin polymorphism in humans. *Clinical chemistry*, 42, 1589-1600.
- Larkin, J. A., Shashy, R. G. & Gonzalez, C. A. 1997. Difficulties in differentiating a rapidly growing *Mycobacterium* species from diphtheroids in an immunocompromised patient. *Clinical Microbiology Newsletter*, 19, 109-111.
- Lazar, T. 2002. Hematology: Clinical Principles and Applications. *JAMA: The Journal of the American Medical Association*, 288, 2750-2750.

- Leite-Browning, M. 2012. Caseous Lymphadenitis (CL) in Goats and Sheep. Alabama Cooperative Extension System. Retrieved from <http://www.aces.edu/pubs/docs/U/UNP-0085/UNP-0085.pdf>.
- Lengarite, M., Mbugua, P., Gachui, C. & Kabuage, L. 2012. Herders' knowledge on mineral nutrition and implication on sheep and goat productivity in Marsabit South District, Kenya. *Livest. Res. Rural. Dev.*, 24.
- Levine, G. J., Bissett, W. T., Cole, R. C., Janke, J. J., Nunes, J., Porter, B. & Levine, J. M. 2006. Imaging diagnosis—bacterial diskospondylitis in a goat. *Veterinary Radiology & Ultrasound*, 47, 585-588.
- Lewis, K. H. & Rettger, L. F. 1940. Non-Sporulating Anaerobic Bacteria of the Intestinal Tract: I. Occurrence and Taxonomic Relationships. *Journal of bacteriology*, 40, 287.
- Litt, M. 1964. Eosinophils And Antigen-Antibody Reactions*. *Annals of the New York Academy of Sciences*, 116, 964-985.
- Liu, D., Chan, W., Fan, D. & Lam, D. 2005. An infected hydrogel buckle with *Corynebacterium pseudotuberculosis*. *British journal of ophthalmology*, 89, 245-246.
- Liuzzi, J. P., Lichten, L. A., Rivera, S., Blanchard, R. K., Aydemir, T. B., Knutson, M. D., Ganz, T. & Cousins, R. J. 2005. Interleukin-6 regulates the zinc transporter Zip14 in liver and contributes to the hypozincemia of the acute-phase response. *Proceedings of the National Academy of Sciences of the United States of America*, 102, 6843-6848.
- Lloyd, S., Lindsay, H., Slater, J. & Jackson, P. 1990. Caseous lymphadenitis in goats in England. *Veterinary Record*, 127.
- Lopez, J., Wong, F. & Quesada, J. 1966. *Corynebacterium pseudotuberculosis*. First case of human infection. *American journal of clinical pathology*, 46, 562.
- Lowenstine, L. J. 2008. Endocrine and Immune. *Biology, Medicine, and Surgery of Elephants*, 309.
- Luna, L. G. 1968. *Manual of histologic staining methods of the Armed Forces Institute of Pathology*, <http://agris.fao.org/aos/records/US201300459165>.
- Lutz, H. U. 2007. Homeostatic roles of naturally occurring antibodies: an overview. *Journal of autoimmunity*, 29, 287-294.
- Macdiarmid, S. C. & Authority, M. B. 1991. The importation into New Zealand of meat and meat products: a review of the risks to animal health. Ministry of Agriculture and Fisheries.
- MacLennan, I. C. 1994. Germinal centers. *Annual review of immunology*, 12, 117-139.

- Mahmood, Z., Jesse, F., Saharee, A., Jasni, S., Yusoff, R. & Wahid, H. 2015. Assessment of blood changes post-challenge with *Corynebacterium pseudotuberculosis* and its exotoxin (phospholipase D): A comprehensive study in goat. *Veterinary World*, 8, 1105-1117.
- Mahmood, Z. K. 2015. *Etiopathogenesis Of Caseous Lymphadenitis In Goats*. PhD thesis, Universiti Putra Malaysia.
- Maki, L., Shen, S., Bergstrom, R. & Stetzenbach, L. 1985. Diagnosis of *Corynebacterium pseudotuberculosis* infections in sheep, using an enzyme-linked immunosorbent assay. *American journal of veterinary research*, 46, 212-214.
- Marples, R. & McGinley, K. 1974. *Corynebacterium acnes* and other anaerobic diphtheroids from human skin. *Journal of Medical Microbiology*, 7, 349-358.
- Marples, R. R., Downing, D. T. & Kligman, A. M. 1971. Control of free fatty acids in human surface lipids by *Corynebacterium acnes*. *Journal of Investigative Dermatology*, 56, 127-131.
- Marrakchi, H., Lan  elle, M.-A. & Daff  , M. 2014. Mycolic acids: structures, biosynthesis, and beyond. *Chemistry & biology*, 21, 67-85.
- Martin, K., Schumann, P., Rainey, F. A., Schuetze, B. & Groth, I. 1997. *Janibacter limosus* gen. nov., sp. nov., a new actinomycete with meso-diaminopimelic acid in the cell wall. *International journal of systematic and evolutionary microbiology*, 47, 529-534.
- Mavromati, J., Cash, P., Restelli, L. & Soler, L. 2014. Proteomics and Protein Analyses of Ovine and Caprine Body Fluids: Current Studies and Future Promises. *Current Protein and Peptide Science*, 15, 45-55.
- Mazanec, M. B., Nedrud, J. G., Kaetzel, C. S. & Lamm, M. E. 1993. A three-tiered view of the role of IgA in mucosal defense. *Immunology today*, 14, 430-435.
- McIntee, M. 2012. *Reproductive pathology of domestic mammals*, Elsevier.
- McKean, S., Davies, J. & Moore, R. 2005. Identification of macrophage induced genes of *Corynebacterium pseudotuberculosis* by differential fluorescence induction. *Microbes and infection*, 7, 1352-1363.
- McKean, S. C., Davies, J. K. & Moore, R. J. 2007a. Expression of phospholipase D, the major virulence factor of *Corynebacterium pseudotuberculosis*, is regulated by multiple environmental factors and plays a role in macrophage death. *Microbiology*, 153, 2203-2211.
- McKean, S. C., Davies, J. K. & Moore, R. J. 2007b. Probing the heat shock response of *Corynebacterium pseudotuberculosis*: the major virulence factor, phospholipase D, is downregulated at 43 C. *Research in microbiology*, 158, 279-286.

- Mcnamara, P. J., Bradley, G. A. & Songer, J. G. 1994. Targeted mutagenesis of the phospholipase D gene results in decreased virulence of *Corynebacterium pseudotuberculosis*. *Molecular microbiology*, 12, 921-930.
- Mcnamara, P. J., Cuevas, W. A. & Songer, J. G. 1995. Toxic phospholipases D of *Corynebacterium pseudotuberculosis*, *C. ulcerans* and *Arcanobacterium haemolyticum*: cloning and sequence homology. *Gene*, 156, 113-118.
- Mearns, R. 2009. Post mortem examination of sheep aged over 12 months. *Livestock*, 14, 42-49.
- Meena, L. S. 2010. Survival mechanisms of pathogenic *Mycobacterium tuberculosis* H37Rv. *FEBS journal*, 277, 2416-2427.
- Meijer, W. G. & Prescott, J. F. 2004. *Rhodococcus equi*. *Veterinary research*, 35, 383-396.
- Ménsua, C., Carrasco, L., Bautista, M., Biescas, E., Fernández, A., Murphy, C., Weiss, D., Solomon, A. & Luján, L. 2003. Pathology of AA amyloidosis in domestic sheep and goats. *Veterinary Pathology Online*, 40, 71-80.
- Menzies, P. 1998. Caseous lymphadenitis of sheep and goats. *The Merck Veterinary Manual*, ed, 8, 55-56.
- Menzies, P., Hwang, Y. & Prescott, J. 2004. Comparison of an interferon- γ to a phospholipase D enzyme-linked immunosorbent assay for diagnosis of *Corynebacterium pseudotuberculosis* infection in experimentally infected goats. *Veterinary Microbiology*, 100, 129-137.
- Menzies, P. I. & Muckle, C. A. 1989. The use of a microagglutination assay for the detection of antibodies to *Corynebacterium pseudotuberculosis* in naturally infected sheep and goat flocks. *Canadian Journal of Veterinary Research*, 53, 313-318.
- Menzies, P. I., Muckle, C. A., Brogden, K. A. & Robinson, L. 1991. A field trial to evaluate a whole cell vaccine for the prevention of caseous lymphadenitis in sheep and goat flocks. *Canadian Journal of Veterinary Research*, 55, 362.
- Michael, W. 2000. Bacterial Adhesion to Host Tissues Mechanisms and Cosequences. *London: Cambridge University Press*, 26,137-154,216.
- Miettinen, T. A. & Kesäniemi, Y. 1989. Cholesterol absorption: regulation of cholesterol synthesis and elimination and within-population variations of serum cholesterol levels. *The American journal of clinical nutrition*, 49, 629-635.
- Mills, A. E., Mitchell, R. D. & Lim, E. K. 1997. *Corynebacterium pseudotuberculosis* is a cause of human necrotising granulomatous lymphadenitis. *Pathology*, 29, 231-233.

- Minnikin, D., Patel, P., Alshamaony, L. & Goodfellow, M. 1977. Polar lipid composition in the classification of *Nocardia* and related bacteria. *International journal of systematic and evolutionary microbiology*, 27, 104-117.
- Moghaddam, A., Olszewska, W., Wang, B., Tregoning, J. S., Helson, R., Sattentau, Q. J. & Openshaw, P. J. 2006. A potential molecular mechanism for hypersensitivity caused by formalin-inactivated vaccines. *Nature medicine*, 12, 905-907.
- Møller, K., Agerholm, J., Ahrens, P., Jensen, N. & Nielsen, T. K. 2000. Abscess disease, caseous lymphadenitis, and pulmonary adenomatosis in imported sheep. *Journal of Veterinary Medicine, Series B*, 47, 55-62.
- Morse, E. V. 1949. Criteria for the identification of corynebacteria isolated from animals. *Cornell Vet*, 39, 266-276.
- Moshage, H. 1997. Cytokines and the hepatic acute phase response. *The Journal of pathology*, 181, 257-266.
- Moura-Costa, L., Bahia, R., Carminati, R., Vale, V., Paule, B., Portela, R., Freire, S., Nascimento, I., Schaer, R. & Barreto, L. 2008. Evaluation of the humoral and cellular immune response to different antigens of *Corynebacterium pseudotuberculosis* in Canindé goats and their potential protection against caseous lymphadenitis. *Veterinary immunology and immunopathology*, 126, 131-141.
- Muckle, C. & Gyles, C. 1982. Characterization of strains of *Corynebacterium pseudotuberculosis*. *Canadian Journal of Comparative Medicine*, 46, 206.
- Muckle, C. A. & Gyles, C. 1983. Relation of lipid content and exotoxin production to virulence of *Corynebacterium pseudotuberculosis* in mice. *American journal of veterinary research*, 44, 1149-1153.
- Muckle, C. A., Menzies, P. I., Li, Y., Hwang, Y. T. & Van Wesenbeeck, M. 1992. Analysis of the immunodominant antigens of *Corynebacterium pseudotuberculosis*. *Veterinary Microbiology*, 30, 47-58.
- Müller, B., De Klerk-Lorist, L.-M., Henton, M. M., Lane, E., Parsons, S., Van Pittius, N. C. G., Kotze, A., Van Helden, P. D. & Tanner, M. 2011. Mixed infections of *Corynebacterium pseudotuberculosis* and non-tuberculous mycobacteria in South African antelopes presenting with tuberculosis-like lesions. *Veterinary Microbiology*, 147, 340-345.
- Murata, H., Shimada, N. & Yoshioka, M. 2004. Current research on acute phase proteins in veterinary diagnosis: an overview. *The Veterinary Journal*, 168, 28-40.
- Musa, M. 1998. Lymphadenitis in sheep and goats in the Sudan. *Revue D Elevage Et De Medicine Veterinaire Des Pays Tropicaux*, 51, 109-112.

- Nairn, M. & Robertson, J. 1974. *Corynebacterium pseudotuberculosis* infection of sheep: role of skin lesions and dipping fluids. *Australian veterinary journal*, 50, 537-542.
- Nakamura, M., Yagi, H., Kayaba, S., Ishii, T., Gotoh, T., Ohtsu, S. & Itoh, T. 1996. Death of germinal center B cells without DNA fragmentation. *European journal of immunology*, 26, 1211-1216.
- Nakamura, Y., Nishio, Y., Ikeo, K. & Gojobori, T. 2003. The genome stability in *Corynebacterium* species due to lack of the recombinational repair system. *Gene*, 317, 149-155.
- Nambuya, A., Sewankambo, N., Mugerwa, J., Goodgame, R. & Lucas, S. 1988. Tuberculous lymphadenitis associated with human immunodeficiency virus (HIV) in Uganda. *Journal of clinical pathology*, 41, 93-96.
- Neutra, M. R. & Kozlowski, P. A. 2006. Mucosal vaccines: the promise and the challenge. *Nature Reviews Immunology*, 6, 148-158.
- Nijsten, M., De Groot, E., Ten Duis, H., Klasen, H., Hack, C. & Aarden, L. 1987. Serum levels of interleukin-6 and acute phase responses. *The Lancet*, 330, 921.
- Nogradi, N., Spier, S. J., Toth, B. & Vaughan, B. 2012. Musculoskeletal *Corynebacterium pseudotuberculosis* infection in horses: 35 cases (1999–2009). *Journal of the American Veterinary Medical Association*, 241, 771-777.
- O'reilly, K., Green, L. E., Malone, F. & Medley, G. 2008. Parameter estimation and simulations of a mathematical model of *Corynebacterium pseudotuberculosis* transmission in sheep. *Preventive veterinary medicine*, 83, 242-259.
- Oda, K., Tsukahara, F., Kubota, S., Kida, K., Kitajima, T. & Hashimoto, S. 2006. Emulsifier content and side effects of oil-based adjuvant vaccine in swine. *Research in veterinary science*, 81, 51-57.
- Oelschlaeger, T. A. & Hacker, J. H. 2013. *Bacterial Invasion Into Eukaryotic Cells: Subcellular Biochemistry*, Springer Science & Business Media.
- Olson, M. E., Ceri, H., Morck, D. W., Buret, A. G. & Read, R. R. 2002. Biofilm bacteria: formation and comparative susceptibility to antibiotics. *Canadian Journal of Veterinary Research*, 66, 86.
- Oreiby, A. F. 2015. Diagnosis of caseous lymphadenitis in sheep and goat. *Small Ruminant Research*, 123, 160-166.
- Osada, J. & Nowacki, W. 1989. Elimination of goat haemoglobin and its complexes with goat haptoglobin from goat and rat circulation. *Acta biochimica Polonica*, 36, 365.

- Osman, A. Y., Abdullah, F. F. J., Saharee, A. A., Haron, A. W., Sabri, I. & Abdullah, R. 2012a. Haematological and Biochemical Alterations in Mice Following Experimental Infection with Whole Cell and Exotoxin (PLD) Extracted from *C. pseudotuberculosis*. *Journal of Animal and Veterinary Advances*, 11, 4660-4667.
- Osman, A. Y., Abdullah, F. F. J. B. & B Saharee, A. A. 2012b. Test among Small Ruminant Flocks in East Coast Economic Regions in Peninsular Malaysia.
- Osman, A. Y., Abdullah, J., Firdaus, F., Adamu, L., Abdullah, T. T., Azuan, M. H., Omar, A. R., Haron, A. W. & Saharee, A. A. 2014. The study of caseous lymphadenitis: dose dependent infection of *C. pseudotuberculosis* in mouse model via oral inoculation. *Research Opinions in Animal & Veterinary Sciences*, 4.
- Osman, A. Y., Bin Abdullah, F. F. J. & Saharee, A. a. B. 2012c. Sero-Prevalence of Caseous Lymphadenitis Evaluated by Agar Gel Precipitation Test among Small Ruminant Flocks in East Coast Economic Regions in Peninsular Malaysia. *Journal of Animal and Veterinary Advances*, 11, 3474-3480.
- Ott, L. 2012. Interaction of *Corynebacterium diphtheriae* with host cells. Universitätsbibliothek der Universität Erlangen-Nürnberg.
- Ott, L. & Burkovski, A. 2014. Toxigenic corynebacteria: adhesion, invasion and host response. *Corynebacterium diphtheriae* and Related Toxigenic Species. Springer.
- Pacheco, L. G., Pena, R. R., Castro, T. L., Dorella, F. A., Bahia, R. C., Carminati, R., Frota, M. N., Oliveira, S. C., Meyer, R. & Alves, F. S. 2007. Multiplex PCR assay for identification of *Corynebacterium pseudotuberculosis* from pure cultures and for rapid detection of this pathogen in clinical samples. *Journal of Medical Microbiology*, 56, 480-486.
- Paltrinieri, S. 2008. The feline acute phase reaction. *The veterinary journal*, 177, 26-35.
- Papaioannou, N., Zavlaris, M., Giadinis, N., Petridou, E. & Psychas, V. 2010. Case report: A case of kidney infection by *Corynebacterium pseudotuberculosis* in sheep. *Journal of the Hellenic Veterinary Medical Society*, 61, 29-35.
- Parsek, M. R. & Singh, P. K. 2003. Bacterial biofilms: an emerging link to disease pathogenesis. *Annual Reviews in Microbiology*, 57, 677-701.
- Paton, M. The epidemiology of Caseous Lymphadenitis in Australia and observations on other production systems. Proceedings of the Annual Meeting-United States Animal Health Association, 1997. United States Animal Health Association, 444-452.
- Paton, M., Collett, M., Pepin, M. & Bath, G. 2005. *Corynebacterium pseudotuberculosis* infections. *Infectious Diseases of Livestock*, 3, 1917-30.

- Paton, M., Mercy, A., Wilkinson, F., Gardner, J., Sutherland, S. & Ellis, T. 1988. The effects of caseous lymphadenitis on wool production and bodyweight in young sheep. *Australian Veterinary Journal*, 65, 117-119.
- Paton, M., Rose, I., Hart, R., Sutherland, S., Mercy, A. & Ellis, T. 1996. Post-shearing management affects the seroincidence of *Corynebacterium pseudotuberculosis* infection in sheep flocks. *Preventive Veterinary Medicine*, 26, 275-284.
- Paton, M., Rose, I., Hart, R., Sutherland, S., Mercy, A., Ellis, T. & Dhaliwal, J. 1994. New infection with *Corynebacterium pseudotuberculosis* reduces wool production. *Australian Veterinary Journal*, 71, 47-49.
- Paton, M., Sutherland, S., Rose, I., Hart, R., Mercy, A. & Ellis, T. 1995. The spread of *Corynebacterium pseudotuberculosis* infection to unvaccinated and vaccinated sheep. *Australian veterinary journal*, 72, 266-269.
- Paton, M., Walker, S., Rose, I. & Watt, G. 2003. Prevalence of caseous lymphadenitis and usage of caseous lymphadenitis vaccines in sheep flocks. *Australian Veterinary Journal*, 81, 91-95.
- Paton, M. W. 2010. *The epidemiology and control of caseous lymphadenitis in Australian sheep flocks*. PhD Thesis, Murdoch University, Perth, Australia.
- Paule, B. J. A., Azevedo, V., Regis, L. F., Carminati, R., Bahia, C., Vale, V. L. C., Moura-Costa, L., Freire, S. M., Nascimento, I. & Schaer, R. 2003. Experimental *Corynebacterium pseudotuberculosis* primary infection in goats: kinetics of IgG and interferon- γ production, IgG avidity and antigen recognition by Western blotting. *Veterinary immunology and immunopathology*, 96, 129-139.
- Pavan, M., Robles, C., Cairó, F., Marcellino, R. & Pettinari, M. 2012. Identification of *Corynebacterium pseudotuberculosis* from sheep by PCR-restriction analysis using the RNA polymerase β -subunit gene (rpoB). *Research in veterinary science*, 92, 202-206.
- Peel, M. M., Palmer, G. G., Stacpoole, A. M. & Kerr, T. G. 1997. Human lymphadenitis due to *Corynebacterium pseudotuberculosis*: report of ten cases from Australia and review. *Clinical Infectious Diseases*, 24, 185-191.
- Pegram, R. 1973. An unusual form of lymphadenitis in sheep and goats in the Somali Democratic Republic. *Tropical animal health and production*, 5, 35-39.
- Pekelder, J. 2000. Caseous lymphadenitis. MARTIN, WB; AITEKEN, ID *Diseases of Sheep*, 3, 270-274.
- Pépin, M., Cannella, D., Fontaine, J.-J., Pittet, J.-C. & Le Pape, A. 1992. Ovine mononuclear phagocytes in situ: identification by monoclonal antibodies and involvement in experimental pyogranulomas. *Journal of Leukocyte Biology*, 51, 188-198.

- Pépin, M., Fontaine, J.-J., Pardon, P., Marly, J. & Parodi, A. 1991. Histopathology of the early phase during experimental *Corynebacterium pseudotuberculosis* infection in lambs. *Veterinary Microbiology*, 29, 123-134.
- Pépin, M., Pardon, P., Lantier, F., Marly, J., Levieux, D. & Lamand, M. 1991. Experimental *Corynebacterium pseudotuberculosis* infection in lambs: kinetics of bacterial dissemination and inflammation. *Veterinary microbiology*, 26, 381-392.
- Pépin, M., Pardon, P., Marly, J., Lantier, F. & Arrigo, J. 1993. Acquired immunity after primary caseous lymphadenitis in sheep. *American journal of veterinary research*, 54, 873-877.
- Pépin, M., Paton, M. & Hodgson, A. 1994a. Pathogenesis and epidemiology of *Corynebacterium pseudotuberculosis* infection in sheep. *Curr. Top. Vet. Res*, 1, 63-82.
- Pépin, M., Pittet, J., Olivier, M. & Gohin, I. 1994b. Cellular composition of *Corynebacterium pseudotuberculosis* pyogranulomas in sheep. *Journal of leukocyte biology*, 56, 666-670.
- Pépin, M., Sanchis, R. & Paton, M. 1999. La lymphadénite caséuse des ovins et des caprins. *Point vétérinaire*, 30, 33-40.
- Pépin, M., Seow, H., Corner, L., Rothel, J., Hodgson, A. & Wood, P. 1997. Cytokine gene expression in sheep following experimental infection with various strains of *Corynebacterium pseudotuberculosis* differing in virulence. *Veterinary research*, 28, 149-163.
- Pepys, M. & Baltz, M. L. 1983. Acute phase proteins with special reference to C-reactive protein and related proteins (pentaxins) and serum amyloid A protein. *Advances in immunology*, 34, 141-212.
- Perkins, S. L., Magdesian, K. G., Thomas, W. P. & Spier, S. J. 2004. Pericarditis and pleuritis caused by *Corynebacterium pseudotuberculosis* in a horse. *Journal of the American Veterinary Medical Association*, 224, 1133-1138.
- Permi, H. S., Shetty, J., Padma, S. K., Teerthanath, S., Mathias, M. & Kumar, S. 2012. A histopathological study of granulomatous inflammation. *Nitte Univ J Health Sc*, 2, 15-9.
- Petersen, H. H., Nielsen, J. P. & Heegaard, P. M. H. 2004. Application of acute phase protein measurements in veterinary clinical chemistry. *Veterinary research*, 35, 163-187.
- Petrovsky, N. & Aguilar, J. C. 2004. Vaccine adjuvants: current state and future trends. *Immunology and Cell Biology*, 82, 488-496.
- Peyron, P., Vaubourgeix, J., Poquet, Y., Levillain, F., Botanch, C., Bardou, F., Daffé, M., Emile, J.-F., Marchou, B. & Cardona, P.-J. 2008. Foamy macrophages from tuberculous patients' granulomas constitute a nutrient-rich reservoir for *M. tuberculosis* persistence. *PLoS Pathog*, 4, e1000204.

- Phythian, C., Hughes, D., Michalopoulou, E., Cripps, P. & Duncan, J. 2012. Reliability of body condition scoring of sheep for cross-farm assessments. *Small Ruminant Research*, 104, 156-162.
- Piñeiro, M., Gymnich, S., Knura, S., Piñeiro, C. & Petersen, B. 2009. Meat juice: An alternative matrix for assessing animal health by measuring acute phase proteins. Correlations of pig-MAP and haptoglobin concentrations in pig meat juice and plasma. *Research in veterinary science*, 87, 273-276.
- Piontkowski, M. & Shivvers, D. 1998. Evaluation of a commercially available vaccine against *Corynebacterium pseudotuberculosis* for use in sheep. *Journal of the American Veterinary Medical Association*, 212, 1765-1768.
- Polepalle, T., Srinivas Moogala, S. B., Pesala, D. S. & Palagi, F. B. 2015. Acute Phase Proteins and Their Role in Periodontitis: A Review. *Journal of clinical and diagnostic research: JCDR*, 9, ZE01.
- Pratt, S. M., Spier, S. J., Carroll, S. P., Vaughan, B., Whitcomb, M. B. & Wilson, W. D. 2005. Evaluation of clinical characteristics, diagnostic test results, and outcome in horses with internal infection caused by *Corynebacterium pseudotuberculosis*: 30 cases (1995-2003). *Journal of the American Veterinary Medical Association*, 227, 441-449.
- Prohl, A., Schroedl, W., Rhode, H. & Reinhold, P. 2015. Acute phase proteins as local biomarkers of respiratory infection in calves. *BMC veterinary research*, 11, 1.
- Puech, V., Chami, M., Lemassu, A., Lanéelle, M.-A., Schiffler, B., Gounon, P., Bayan, N., Benz, R. & Daffé, M. 2001. Structure of the cell envelope of corynebacteria: importance of the non-covalently bound lipids in the formation of the cell wall permeability barrier and fracture plane. *Microbiology*, 147, 1365-1382.
- Pugh, D. G. & Baird, N. N. 2012. *Sheep & goat medicine*, Elsevier Health Sciences.
- Qu, X.-D. & Lehrer, R. I. 1998. Secretory phospholipase A2 is the principal bactericide for staphylococci and other gram-positive bacteria in human tears. *Infection and immunity*, 66, 2791-2797.
- Quinn, P. J., Markey, B. K., Leonard, F. C., Fitzpatrick, E. S., Fanning, S. & Hartigan, P. 2011. *Veterinary microbiology and microbial disease*, John Wiley & Sons.
- Quist, C. 2002. Wyoming State Veterinary Laboratory–Department of Veterinary Sciences.
- Rachek, L. I., Tucker, A. M., Winkler, H. H. & Wood, D. O. 1998. Transformation of *Rickettsia prowazekii* to Rifampin Resistance. *Journal of bacteriology*, 180, 2118-2124.
- Radford, A. J. & Hodgson, A. L. 1997. Use of a phospholipase D mutant of *Corynebacterium pseudotuberculosis* for vaccination. Google Patents.

- Radostitis, O., Gay, C., Blood, D. & Hinchcliff, K. 2000. Caseous lymphadenitis in sheep and goats. . In: *Veterinary medicine, 9th ed.* WB Saunders, London.
- Rahman, M. M., Lecchi, C., Fraquelli, C., Sartorelli, P. & Ceciliani, F. 2010. Acute phase protein response in Alpine ibex with sarcoptic mange. *Veterinary parasitology*, 168, 293-298.
- Ramakrishna, C., Stohlman, S. A., Atkinson, R. D., Shlomchik, M. J. & Bergmann, C. C. 2002. Mechanisms of central nervous system viral persistence: the critical role of antibody and B cells. *The Journal of Immunology*, 168, 1204-1211.
- Rees, M. A., Kleifeld, O., Crellin, P. K., Ho, B., Stinear, T. P., Smith, A. I. & Coppel, R. L. 2014. Proteomic Characterization of a Natural Host–Pathogen Interaction: Repertoire of in Vivo Expressed Bacterial and Host Surface-Associated Proteins. *Journal of proteome research*, 14, 120-132.
- Rekate, H. L., Ruch, T. & Nulsen, F. E. 1980. Diphtheroid infections of cerebrospinal fluid shunts: the changing pattern of shunt infection in Cleveland. *Journal of neurosurgery*, 52, 553-556.
- Renshaw, H., Graff, V. & Gates, N. 1979. Visceral caseous lymphadenitis in thin ewe syndrome: isolation of *Corynebacterium*, *Staphylococcus*, and *Moraxella* spp from internal abscesses in emaciated ewes. *American journal of veterinary research*, 40, 1110-1114.
- Richardson, A. 1931. The Possibilities of Scientific Research. *The ANNALS of the American Academy of Political and Social Science*, 158, 251-258.
- Ripeau, J.-S., Aumont, F., Belhumeur, P., Ostrosky-Zeichner, L., Rex, J. H. & De Repentigny, L. 2002. Effect of the echinocandin caspofungin on expression of *Candida albicans* secretory aspartyl proteinases and phospholipase in vitro. *Antimicrobial agents and chemotherapy*, 46, 3096-3100.
- Robertson, J. P. 1980. Studies on the diagnosis, epidemiology and immunity of *Corynebacterium pseudotuberculosis* infection in sheep, Murdoch University.
- Robinette, W. L., Gashwiler, J. S., Jones, D. A. & Crane, H. S. 1955. Fertility of mule deer in Utah. *The Journal of Wildlife Management*, 19, 115-136.
- Rodak, B. F., Fritsma, G. A. & Keohane, E. 2013. *Hematology: clinical principles and applications*, Elsevier Health Sciences.
- Roslindawani, M., Syafiqah, A., Jesse, F., Effendy, A. & Zamri-Saad, M. 2016. Recombinant caseous lymphadenitis vaccine with palm oil as adjuvant enhances the humoral and cell-mediated immune responses in rat model. *J. Anim. Health Prod*, 4, 22-25.
- Rothenberg, M. E. & Hogan, S. P. 2006. The eosinophil. *Immunology*, 24, 147.

- Ruiz, J. C., D'afonseca, V., Silva, A., Ali, A., Pinto, A. C., Santos, A. R., Rocha, A. A., Lopes, D. O., Dorella, F. A. & Pacheco, L. G. 2011. Evidence for reductive genome evolution and lateral acquisition of virulence functions in two *Corynebacterium pseudotuberculosis* strains. *PLoS ONE*, 6, e18551.
- Ruzevick, J., Jackson, C., Pradilla, G., Garzon-Muvdi, T. & Tamargo, R. J. 2013. Aneurysm formation in proinflammatory, transgenic haptoglobin 2-2 mice. *Neurosurgery*, 72, 70-76.
- Saad, M. Z., Abdullah, J. F. F., Saharee, A. A., Haron, A. W. & Shamsudin, S. 2013. Clinical and pathological changes in goats inoculated *Corynebacterium pseudotuberculosis* by intradermal, intranasal and oral routes. *Online Journal of Veterinary Research*, 17, 73-81.
- Saitou, N. & Nei, M. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4, 406-425.
- Samols, D., Agrawal, A. & Kushner, I. 2002. Acute phase proteins. *Cytokine reference on-line*.
- Sargison, N. 2009. *Sheep flock health: a planned approach*, John Wiley & Sons.
- Sasaki, T. 1958. Pathological Studies of Secondary Intestinal Tuberculosis. *Pathology International*, 8, 229-250.
- Schmiel, D. H. & Miller, V. L. 1999. Bacterial phospholipases and pathogenesis. *Microbes and infection*, 1, 1103-1112.
- Schorey, J. S., Carroll, M. C. & Brown, E. J. 1997. A macrophage invasion mechanism of pathogenic mycobacteria. *Science*, 277, 1091-1093.
- Schreuder, B., Ter Laak, E. & Dercksen, D. 1994. Eradication of caseous lymphadenitis in sheep with the help of a newly developed ELISA technique. *The Veterinary Record*, 135, 174-176.
- Schwab, L. 2007. *Eye care in developing nations*, CRC Press.
- Schwartz, L. B. & Austen, K. F. 1980. Enzymes of the mast cell granule. *Journal of Investigative Dermatology*, 74, 349-353.
- Schwartz, Z. & Boyan, B. 1988. The Effects of Vitamin D Metabolites on Phospholipase A2 Activity of Growth Zone and Resting Zone Cartilage Cells in Vitro*. *Endocrinology*, 122, 2191-2198.
- Scott, D. W. 2008. *Color atlas of farm animal dermatology*, John Wiley & Sons.
- Selim, S. 2001. Oedematous skin disease of buffalo in Egypt. *Journal of Veterinary Medicine, Series B*, 48, 241-258.
- Selim, S., Ghoneim, M. & Mohamed, K. 2010. Vaccinal efficacy of genetically inactivated phospholipase D against caseous lymphadenitis in Small Ruminants. *International Journal of Microbiological Research*, 1, 129-136.

- Senturk, S. & Temizel, M. 2006. Clinical efficacy of rifamycin SV combined with oxytetracycline in the treatment of caseous lymphadenitis in sheep. *Veterinary record*, 159, 216-217.
- Seyffert, N., Guimarães, A., Pacheco, L., Portela, R., Bastos, B., Dorella, F., Heinemann, M., Lage, A., Gouveia, A. & Meyer, R. 2010. High seroprevalence of caseous lymphadenitis in Brazilian goat herds revealed by *Corynebacterium pseudotuberculosis* secreted proteins-based ELISA. *Research in veterinary science*, 88, 50-55.
- Seyrek, A., Kocyigit, A. & Erel, O. 2005. Essential trace elements selenium, zinc, copper, and iron concentrations and their related acute-phase proteins in patients with vivax malaria. *Biological trace element research*, 106, 107-115.
- Sheikh-Omar, A. & Shah, M. 1984. Caseous lymphadenitis in sheep imported from Australia for slaughter in Malaysia Faculty of Veterinary Medicine. *Australian Veterinary Journal*, 61, 410-410.
- Shekh-Omar, A. & Shah, M. 1984. Caseous lymphadenitis in sheep imported from Australia for slaughter in Malaysia Faculty of Veterinary Medicine. *Australian veterinary journal*, 61, 410-410.
- Shigidi, M. 1979. comparison of five serological tests for the diagnosis of experimental *Corynebacterium ovis* infection in sheep. *British veterinary journal*.
- Shin, A.-R., Lee, K.-S., Lee, J.-S., Kim, S.-Y., Song, C.-H., Jung, S.-B., Yang, C.-S., Jo, E.-K., Park, J.-K. & Paik, T.-H. 2006. *Mycobacterium tuberculosis* HBHA protein reacts strongly with the serum immunoglobulin M of tuberculosis patients. *Clinical and vaccine immunology*, 13, 869-875.
- Shkreta, L., Talbot, B. G., Diarra, M. S. & Lacasse, P. 2004. Immune responses to a DNA/protein vaccination strategy against *Staphylococcus aureus* induced mastitis in dairy cows. *Vaccine*, 23, 114-126.
- Sibille, Y. & Reynolds, H. Y. 1990. Macrophages and polymorphonuclear neutrophils in lung defense and injury. *American Review of Respiratory Disease*, 141, 471-501.
- Silva, A., Schneider, M. P. C., Cerdeira, L., Barbosa, M. S., Ramos, R. T. J., Carneiro, A. R., Santos, R., Lima, M., D'afonseca, V. & Almeida, S. S. 2011. Complete genome sequence of *Corynebacterium pseudotuberculosis* I19, a strain isolated from a cow in Israel with bovine mastitis. *Journal of bacteriology*, 193, 323-324.
- Simmons, C. P., Hodgson, A. & Strugnell, R. A. 1997. Attenuation and vaccine potential of aroQ mutants of *Corynebacterium pseudotuberculosis*. *Infection and immunity*, 65, 3048-3056.

- Simon, L., Gauvin, F., Amre, D. K., Saint-Louis, P. & Lacroix, J. 2004. Serum procalcitonin and C-reactive protein levels as markers of bacterial infection: a systematic review and meta-analysis. *Clinical Infectious Diseases*, 39, 206-217.
- Singer, W. D., Brown, A., H Alex & Sternweis, P. C. 1997. Regulation of eukaryotic phosphatidylinositol-specific phospholipase C and phospholipase D. *Annual review of biochemistry*, 66, 475-509.
- Sivaraj, S., Dorny, P., Vercruysse, J. & Pandey, V. 1994. Multiple and multigeneric anthelmintic resistance on a sheep farm in Malaysia. *Veterinary Parasitology*, 55, 159-165.
- Sivasupramaniam, G. 2008. Goat farming in Malaysia. *Goats-Undervalued Assets In Asia*, 45.
- Slavin, R. E., Walsh, T. J. & Pollack, A. D. 1980. Late generalized tuberculosis: a clinical pathologic analysis and comparison of 100 cases in the preantibiotic and antibiotic eras. *Medicine*, 59, 352-366.
- Slifka, M. K., Antia, R., Whitmire, J. K. & Ahmed, R. 1998. Humoral immunity due to long-lived plasma cells. *Immunity*, 8, 363-372.
- Smith, M. C. & Sherman, D. M. 2011. *Goat medicine*, John Wiley & Sons.
- Smole, I., Thomann, A., Frey, J. & Perreten, V. 2010. Repression of common bull sperm flora and in vitro impairment of sperm motility with *Pseudomonas aeruginosa* introduced by contaminated lubricant. *Reproduction in domestic animals*, 45, 737-742.
- Soares, S. C., Silva, A., Trost, E., Blom, J., Ramos, R., Carneiro, A., Ali, A., Santos, A. R., Pinto, A. C. & Diniz, C. 2013. The pan-genome of the animal pathogen *Corynebacterium pseudotuberculosis* reveals differences in genome plasticity between the biovar ovis and equi strains. *PLoS ONE*, 8, e53818.
- Songer, J. G., Beckenbach, K., Marshall, M. M., Olson, G. B. & Kelley, L. 1988. Biochemical and genetic characterization of *Corynebacterium pseudotuberculosis*. *American journal of veterinary research*, 49, 223-226.
- Sood, N., Sandhu, B., Gupta, K., Narang, D., Vasudeva, K. & Singh, N. 2012. Mesenteric caseous lymphadenitis in a cow calf caused by *Corynebacterium pseudotuberculosis*: a case report. *Vet Med*, 57, 371-375.
- Souček, A., Michalec, Č. & Součková, A. 1971. Identification and characterization of a new enzyme of the group "phospholipase D" isolated from *Corynebacterium ovis*. *Biochimica et Biophysica Acta (BBA)-Enzymology*, 227, 116-128.
- Spencer, H. & Salfelder, K. 1973. Tropical mycotic diseases. *Tropical Pathology*. Springer.

- Spickler, A. R. & Roth, J. A. 2003. Adjuvants in veterinary vaccines: modes of action and adverse effects. *Journal of Veterinary Internal Medicine*, 17, 273-281.
- Spiegel, I. B., White, S. D., Foley, J. E., Drazenovich, N. L., Ihrke, P. J. & Affolter, V. K. 2006. A retrospective study of cutaneous equine sarcoidosis and its potential infectious aetiological agents. *Veterinary dermatology*, 17, 51-62.
- Stanford, K., Brogden, K., McClelland, L., Kozub, G. & Audibert, F. 1998a. The incidence of caseous lymphadenitis in Alberta sheep and assessment of impact by vaccination with commercial and experimental vaccines. *Canadian Journal of Veterinary Research*, 62, 38.
- Stanford, K., Brogden, K., McClelland, L., Kozub, G. & Audibert, F. 1998b. The incidence of caseous lymphadenitis in Alberta sheep and assessment of impact by vaccination with commercial and experimental vaccines. *Canadian Journal of Veterinary Research*, 62, 38-43.
- Stefńska, I., Gieryńska, M., Rzewuska, M. & Binek, M. 2010. Survival of *Corynebacterium pseudotuberculosis* within macrophages and induction of phagocytes death. *Polish journal of veterinary sciences*, 13, 143.
- Sting, R., Steng, G. & Spengler, D. 1998. Serological Studies on *Corynebacterium pseudotuberculosis* Infections in Goats using Enzyme-linked Immunosorbent Assay. *Journal of Veterinary Medicine, Series B*, 45, 209-216.
- Stoll, S., Delon, J., Brotz, T. M. & Germain, R. N. 2002. Dynamic imaging of T cell-dendritic cell interactions in lymph nodes. *Science*, 296, 1873-1876.
- Straw, B., MacLachlan, N., Corbett, W., Carter, P. & Schey, H. 1985. Comparison of tissue reactions produced by *Haemophilus pleuropneumoniae* vaccines made with six different adjuvants in swine. *Canadian journal of comparative medicine*, 49, 149.
- Streetz, K., Wüstefeld, T., Klein, C., Manns, M. & Trautwein, C. 2001. Mediators of inflammation and acute phase response in the liver. *Cellular and molecular biology (Noisy-le-Grand, France)*, 47, 661-673.
- Suter, E. 1956. Interaction between phagocytes and pathogenic microorganisms. *Bacteriological Reviews*, 20, 94.
- Sutherland, S., Ellis, T., Mercy, A., Paton, M. & Middleton, H. 1987. Evaluation of an enzyme-linked immunosorbent assay for the detection of *Corynebacterium pseudotuberculosis* infection in sheep. *Australian veterinary journal*, 64, 263-266.
- Sutherland, S., Hart, R. & Buller, N. 1996. Genetic differences between nitrate-negative and nitrate-positive *C. pseudotuberculosis* strains using restriction fragment length polymorphisms. *Veterinary Microbiology*, 49, 1-9.

- Syame, S. M., El-Hewairy, H. & Selim, S. 2008. Protection of buffaloes against oedematous skin disease by recombinant-bacterin and toxoid-bacterin vaccines. *Global Vet*, 2, 151-156.
- Szalus-Jordanow, O., Kaba, J., Czopowicz, M., Witkowski, L., Nowicki, M., Nowicka, D., Stefanska, I., Rzewuska, M., Sobczak-Filipiak, M. & Binek, M. 2010. Epidemiological features of Morel's disease in goats. *Polish journal of veterinary sciences*, 13, 437.
- Szonyi, B., Swinford, A., Clavijo, A. & Ivanek, R. 2014. Re-emergence of pigeon fever (*Corynebacterium pseudotuberculosis*) infection in TEXAS horses: Epidemiologic investigation of laboratory-diagnosed cases. *Journal of Equine Veterinary Science*, 34, 281-287.
- Szwako, A., Ortiz, N. & López, D. 2014. Caseous Lymphadenitis (*Corynebacterium pseudotuberculosis*) Prevalence In Caprine Dairy Farms, Central Department-Paraguay, Year 2012. *Compendio de Ciencias Veterinarias*, 4, 24-29.
- Tachedjian, M., Krywult, J., Moore, R. J. & Hodgson, A. L. 1995. Caseous lymphadenitis vaccine development: site-specific inactivation of the *Corynebacterium pseudotuberculosis* phospholipase D gene. *Vaccine*, 13, 1785-1792.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. 2013. MEGA6: molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, 30, 2725-2729.
- Tashjian, J. J. 1982. The interaction between caprine macrophages and I. *Corynebacterium pseudotuberculosis*, II. *Mycobacterium paratuberculosis*: an electron microscopic study, Cornell University, Jan.
- Ter Laak, E., Bosch, J., Bijl, G. & Schreuder, B. 1992. Double-antibody sandwich enzyme-linked immunosorbent assay and immunoblot analysis used for control of caseous lymphadenitis in goats and sheep. *American journal of veterinary research*, 53, 1125-1132.
- Tham, K., Rahman, A. & Omar, S. 1981. A study on causes of condemnation of carcass and organs at Shah Alam abattoir. *Pertanika*, 4, 43-46.
- Timoney, J. F., Gillespie, J. H., Scott, F. W. & Barlough, J. E. 1988. *Hagan and Bruner's microbiology and infectious diseases of domestic animals*, Cornell University Press.
- Tîrziu, E. 2009. Acute-phase proteins in immune response. *Lucr. St. Med. Vet. Timisoara*, 42, 329-339.
- Tizard, I. R. 2013. *Veterinary immunology*, Elsevier Health Sciences.
- Tothova, C., Nagy, O. & Kovac, G. 2014. Acute phase proteins and their use in the diagnosis of diseases in ruminants: a review. *Vet Med-Czech*, 59, 163-180.

- Tourlomoussis, P., Eckersall, P., Waterson, M. & Buncic, S. 2004. A comparison of acute phase protein measurements and meat inspection findings in cattle. *Foodborne Pathogens & Disease*, 1, 281-290.
- Trost, E., Ott, L., Schneider, J., Schröder, J., Jaenicke, S., Goesmann, A., Husemann, P., Stoye, J., Dorella, F. A. & Rocha, F. S. 2010. The complete genome sequence of *Corynebacterium pseudotuberculosis* FRC41 isolated from a 12-year-old girl with necrotizing lymphadenitis reveals insights into gene-regulatory networks contributing to virulence. *BMC genomics*, 11, 728.
- Ugochukwu, N. H. & Figgers, C. L. 2007. Caloric restriction inhibits up-regulation of inflammatory cytokines and TNF- α , and activates IL-10 and haptoglobin in the plasma of streptozotocin-induced diabetic rats. *The Journal of nutritional biochemistry*, 18, 120-126.
- Unanian, M., Silva, A. F. & Pant, K. 1985. Abscesses and caseous lymphadenitis in goats in tropical semi-arid north-east Brazil. *Tropical animal health and production*, 17, 57-62.
- Ural, K., Alic, D., Haydardedeoglu, A., Cedden, F., Guzel, M., Özyıldız, Z. & Cantekin, Z. 2008. *Corynebacterium pseudotuberculosis* infection in Saanen× Kilis crossbred (White) goats in Ankara, Turkey and effective kanamycin treatment: A prospective, randomized, double-blinded, placebo-controlled clinical trial. *Small Ruminant Research*, 77, 84-88.
- Urieli-Shoval, S., Linke, R. P. & Matzner, Y. 2000. Expression and function of serum amyloid A, a major acute-phase protein, in normal and disease states. *Current opinion in hematology*, 7, 64-69.
- Vadas, P. 1984. Elevated plasma phospholipase A2 levels: correlation with the hemodynamic and pulmonary changes in gram-negative septic shock. *The Journal of laboratory and clinical medicine*, 104, 873-881.
- Valdivia, J., Real, F., Acosta, F., Acosta, B., Déniz, S., Ramos-Vivas, J., Elaamri, F. & Padilla, D. 2013. Interaction of *Corynebacterium pseudotuberculosis* with ovine cells in vitro. *Veterinary Pathology Online*, 50, 318-323.
- Valli, V. T., Kiupel, M. & Bienzle, D. 2015. Hematopoietic system. *Jubb, Kennedy & Palmer's Pathology of Domestic Animals-E-BOOK*, 3, 102.
- Vasco, A. 2012. *Corynebacterium pseudotuberculosis*: immunological responses in animal models and zoonotic potential. *Journal of Clinical & Cellular Immunology*.
- Vasudevan, D. M., Sreekumari, S. & Vaidyanathan, K. 2010. *Textbook of biochemistry for medical students*, Jaypee Brothers Publishers.
- Vergne, I., Chua, J., Singh, S. B. & Deretic, V. 2004. Cell biology of Mycobacterium tuberculosis phagosome. *Annu. Rev. Cell Dev. Biol.*, 20, 367-394.

- Villaquiran, M., Gipson, T., Merkel, R., Goetsch, A. & Sahl, T. 2004. Body condition scores in goats. *American Institute for Goat Research, Langston University*.
- Voigt, K., Baird, G. J., Munro, F., Murray, F. & Brülisauer, F. 2012. Eradication of caseous lymphadenitis under extensive management conditions on a Scottish hill farm. *Small Ruminant Research*, 106, 21-24.
- Wain, J., Diep, T. S., Ho, V. A., Walsh, A. M., Hoa, N. T. T., Parry, C. M. & White, N. J. 1998. Quantitation of bacteria in blood of typhoid fever patients and relationship between counts and clinical features, transmissibility, and antibiotic resistance. *Journal of clinical microbiology*, 36, 1683-1687.
- Walker, C. A. 2009. Iron-dependent regulation of gene expression in *Corynebacterium pseudotuberculosis*. University of Glasgow.
- Walker, H. L., Chowdhury, K. A., Thaler, A. M., Petersen, K. E., Ragland, R. D. & James, W. O. 2000. Relevance of carcass palpation in lambs to protecting public health. *Journal of Food Protection®*, 63, 1287-1290.
- Walker, J., Jackson, H., Brandon, M. & Meeusen, E. 1991. Lymphocyte subpopulations in pyogranulomas of caseous lymphadenitis. *Clinical & Experimental Immunology*, 86, 13-18.
- Walker, J., Jackson, H., Eggleton, D., Meeusen, E., Wilson, M. & Brandon, M. 1994. Identification of a novel antigen from *Corynebacterium pseudotuberculosis* that protects sheep against caseous lymphadenitis. *Infection and immunity*, 62, 2562-2567.
- Wallach, D. P. & Brown, V. J. 1981. Studies on the arachidonic acid cascade—I: Inhibition of phospholipase A₂ in vitro and in vivo by several novel series of inhibitor compounds. *Biochemical pharmacology*, 30, 1315-1324.
- Washburn, K. E., Bissett, W. T., Fajt, V. R., Libal, M. C., Fosgate, G. T., Miga, J. A. & Rockey, K. M. 2009. Comparison of three treatment regimens for sheep and goats with caseous lymphadenitis. *Journal of the American Veterinary Medical Association*, 234, 1162-1166.
- Watts, J. L. 1988. Etiological agents of bovine mastitis. *Veterinary Microbiology*, 16, 41-66.
- Webber, J., Dobrenov, B., Lloyd, J. & Jordan, D. 2012. Meat inspection in the Australian red-meat industries: past, present and future. *Australian veterinary journal*, 90, 363-369.
- Weibel, J. C. 2011. *Corynebacterium Pseudotuberculosis*: Cell Invasion and Intracellular Survival.
- Weisbart, R., Kacena, A., Schuh, A. & Golde, D. 1988. GM-CSF induces human neutrophil IgA-mediated phagocytosis by an IgA Fc receptor activation mechanism. *Nature*, 332, 647-648.
- Willard-Mack, C. L. 2006. Normal structure, function, and histology of lymph nodes. *Toxicologic Pathology*, 34, 409-424.

- Willems, H., Thiele, D. & Krauss, H. 1993. Plasmid based differentiation and detection of *Coxiella burnetii* in clinical samples. *European journal of epidemiology*, 9, 411-418.
- Williams, D., Spring, L., Collins, L., Miller, L., Heifets, L., Gangadharam, P. & Gillis, T. 1998. Contribution of *rpoB* mutations to development of rifamycin cross-resistance in *Mycobacterium tuberculosis*. *Antimicrobial agents and chemotherapy*, 42, 1853-1857.
- Williamson, L. H. 2001. Caseous lymphadenitis in small ruminants. *Veterinary Clinics of North America: Food Animal Practice*, 17, 359-371.
- Williamson, P. & Nairn, M. 1980. Lesions caused by *Corynebacterium pseudotuberculosis* in the scrotum of rams. *Australian veterinary journal*, 56, 496-498.
- Wilson, B. A., Salyers, A. A., Whitt, D. D. & Winkler, M. E. 2011. *Bacterial pathogenesis: a molecular approach*, American Society for Microbiology (ASM).
- Wilson, J., Schurr, M., Leblanc, C., Ramamurthy, R., Buchanan, K. & Nickerson, C. 2002. Mechanisms of bacterial pathogenicity. *Postgraduate medical journal*, 78, 216-224.
- Wilson, M. 2002. *Bacterial adhesion to host tissues: mechanisms and consequences*, Cambridge University Press.
- Wilson, W. D. Rational selection of antimicrobials for use in horses. *Proc AAEP*, 2001. 75-93.
- Windsor, P. A. 2011. Control of caseous lymphadenitis. *Veterinary Clinics of North America: Food Animal Practice*, 27, 193-202.
- Winter, A. 2009. Vaccination programmes for sheep flocks. *In Practice* (0263841X), 31.
- Winter, P., Fuchs, K., Walshe, K. & Colditz, I. 2003. Serum amyloid A in the serum and milk of ewes with mastitis induced experimentally with *Staphylococcus epidermidis*. *The Veterinary Record*, 152, 558-562.
- Winter, P., Miny, M., Fuchs, K. & Baumgartner, W. 2006. The potential of measuring serum amyloid A in individual ewe milk and in farm bulk milk for monitoring udder health on sheep dairy farms. *Research in veterinary science*, 81, 321-326.
- Wolf, R. A. & Gross, R. 1985. Identification of neutral active phospholipase C which hydrolyzes choline glycerophospholipids and plasmalogen selective phospholipase A2 in canine myocardium. *Journal of Biological Chemistry*, 260, 7295-7303.
- Woods, G. L. & Walker, D. H. 1996. Detection of infection or infectious agents by use of cytologic and histologic stains. *Clinical microbiology reviews*, 9, 382-404.
- Wright, D., Addis, B. J. & Leong, A. S. 2011. *Diagnostic lymph node pathology*, CRC Press.

- Xavier, M. N., Silva, T. M., Costa, É. A., Paixão, T. A., Moustacas, V. S., Júnior, C. a. C., Sant'anna, F. M., Robles, C. A., Gouveia, A. M. & Lage, A. P. 2010. Development and evaluation of a species-specific PCR assay for the detection of *Brucella ovis* infection in rams. *Veterinary Microbiology*, 145, 158-164.
- Yeruham, I., Braverman, Y., Shpigel, N., Chizov-Ginzburg, A., Saran, A. & Winkler, M. 1996. Mastitis in Dairy Cattle Caused by *Corynebacterium pseudotuberculosis* and the Feasibility Of Transmission by Houseflies I. *Veterinary quarterly*, 18, 87-89.
- Yeruham, I., Elad, D., Friedman, S. & Perl, S. 2003. *Corynebacterium pseudotuberculosis* infection in Israeli dairy cattle. *Epidemiology and infection*, 131, 947-955.
- Yeruham, I., Friedman, S., Elad, D. & Perl, S. 2000. Association between milk production, somatic cell count and bacterial dermatoses in three dairy cattle herds. *Australian veterinary journal*, 78, 250-253.
- Yeruham, I., Friedman, S., Perl, S., Elad, D., Berkovich, Y. & Kalgard, Y. 2004. A herd level analysis of a *Corynebacterium pseudotuberculosis* outbreak in a dairy cattle herd. *Veterinary dermatology*, 15, 315-320.
- Yokota, A., Tamura, T., Nishii, T. & Hasegawa, T. 1993. *Kineococcus aurantiacus* gen. nov., sp. nov., a new aerobic, gram-positive, motile coccus with meso-diaminopimelic acid and arabinogalactan in the cell wall. *International journal of systematic and evolutionary microbiology*, 43, 52-57.
- Young, A. B., Ott, L. G., Beard, D., Dempsey, R. J., Tibbs, P. A. & McClain, C. J. 1988. The acute-phase response of the brain-injured patient. *Journal of neurosurgery*, 69, 375-380.
- Yozwiak, M. & Songer, J. G. 1993. Effect of *Corynebacterium pseudotuberculosis* phospholipase D on viability and chemotactic responses of ovine neutrophils. *American journal of veterinary research*, 54, 392-397.
- Yun, C.-H., Wynn, P. & Ha, J. K. 2014. Stress, acute phase proteins and immune modulation in calves. *Animal Production Science*, 54, 1561-1568.
- Zahorec, R. 2000. Ratio of neutrophil to lymphocyte counts--rapid and simple parameter of systemic inflammation and stress in critically ill. *Bratislavské lekárske listy*, 102, 5-14.
- Zeru, F. & Kahsay, A. G. 2014. Caseous lymphadenitis in goats from Borena Range Land South Ethiopia slaughtered at Luna Export Abattoir. *Journal of Veterinary Medicine and Animal Health*, 6, 168-173.
- Zimlichman, S., Danon, A., Nathan, I., Mozes, G. & Shainkin-Kestenbaum, R. 1991. Serum amyloid A, an acute phase protein, inhibits platelet activation. *Amyloid and Amyloidosis 1990*. Springer.

Zink, M., Yager, J., Prescott, J. & Fernando, M. 1987. Electron microscopic investigation of intracellular events after ingestion of *Rhodococcus equi* by foal alveolar macrophages. *Veterinary Microbiology*, 14, 295-305.

