

UNIVERSITI PUTRA MALAYSIA

PATHOGENESIS OF Corynebacterium pseudotuberculosis INFECTION AND VACCINATION TRIAL AGAINST CASEOUS LYMPHADENITIS IN GOATS

NUR ADZA RINA BINTI MOHD NORDI

FPV 2016 36



PATHOGENESIS OF Corynebacterium pseudotuberculosis INFECTION AND VACCINATION TRIAL AGAINST CASEOUS LYMPHADENITIS IN GOATS



By

NUR ADZA RINA BINTI MOHD NORDI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, In Fulfillments of the Requirements for the Degree of Doctor of Philosophy

November 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes for copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

PATHOGENESIS OF Corynebacterium pseudotuberculosis INFECTION AND VACCINATION TRIAL AGAINST CASEOUS LYMPHADENITIS IN GOATS

By

NUR ADZA RINA MOHD NORDI

November 2016

Chairman: Prof. Mohd Zamri Saad, PhD Faculty: Veterinary Medicine

Corynebacterium pseudotuberculosis is a Gram-positive bacterium that is responsible for a disease called caseous lymphadenitis (CLA) in goats and sheep. This disease has worldwide distributions and the bacterium can remain in the environment for months. It is difficult to eradicate and can be easily transmitted to naive animals. Furthermore, transmission and the pathology of the disease are not fully understood. Therefore, this experiment was conducted to determine the best route of infection of the disease, the clinical and pathological changes in goats following experimental infection, the humoral immune response via antibody titers shown by the infected goats, and the efficacy of a commercial vaccine in preventing CLA in goats in Malaysia.

Twenty adult healthy goats were selected and divided into 4 groups. All goats of the first 3 groups were infected with 10^7 cfu/mL of live *C. pseudotuberculosis* via three different routes; the intradermal, the intranasal and the oral routes. The last group served as the uninfected control group. The goats were observed daily for clinical signs related to CLA for 30 days experimental period. Rectal temperatures and blood samples were taken periodically. The infected goats from all infected groups were depressed, showed lack of appetite, and increased in body temperature in the first week post-inoculation. The intradermal group had swelling with pus at the site of infection. Blood profile of the goats revealed significant decreased in haemoglobin for the intradermal and intranasal groups. Only the intradermal group showed significantly (p<0.05) high total WBC counts, with the increased neutrophils and monocyte concentrations. Generally, goats infected intradermally showed most severe clinical signs and haematology changes.

At the end of 30-day experimental period, all goats were sacrificed. *C. pseudotuberculosis* was re-isolated from most of the intradermally infected goats with 40% of the goats had the bacteria in the liver, 80% in prescapular and 40% in submandibular lymph nodes. Only 20% of goats in the intranasally infected group had the bacteria in the liver. No bacteria were isolated from any organ or lymph nodes from the oral and control goats. Abscessation was the most commonly observed gross lesions, particularly within the lymph nodes of infected goats. Other less common lesions included consolidation of lung lobes, congestion of kidneys and lymph nodes. Histopathological lesions scores revealed significantly (p<0.05) much severe overall lesions among goats exposed intradermally.

Following to the initial 30-day experimental trial, a chronic study was eventually conducted. Nine adult goats were similarly infected intradermally with 10⁷ cfu/mL of live *C. pseudotuberculosis* and were observed for 3 months. Similarly, all infected goats were less active in week 1 post-infection, developed swelling at the injection site with enlargement of submandibular lymph nodes. The haemoglobin, however, remained within normal value and decresed insignificantly (p>0.05) throughout the experimental period. Total white blood cell (WBC) counts were consistently high until day 39 post-infection due to the increased neutrophilic and monocytic counts. The serum IgG increased and exceeded the cut-off value on day 10 post-infection but most significant (p<0.05) increase was observed on day 14 post-infection before it gradually decreased until day 53 post-infection and approaching the cut-off value at the end of the 90-day experimental period.

Three goats were sacrificed by slaughtering monthly. *C. pseudotuberculosis* was successfully isolated from all lymph nodes with abscessation, and from the lungs of a goat that was sacrificed 2 months post-infection. Prescapular node abscessation was observed in 2 goats at 1 month post-inoculation, in all 3 goats at 2 and 3 months. There was also abscessation in the submandibular lymph node of a goat at 3 month post-inoculation. The lungs of infected goats showed thickening of interalveolar septa, mild to moderate congestion of the liver with inflammatory cells found scattered in between hepatocytes and presence of mild fatty degeneration of the hepatocytes. The kidneys showed congestion of blood vessels and presence of uses in which the diameter of the necrotic centre was significantly (p<0.05) larger with increase of time post-infection. These results show that the disease progressed with time after infection.

Currently, there is only one commercial vaccine for CLA in Malaysia. However, the efficacy was uncertain. Twenty-seven goats of different serological status were selected from a farm with endemic CLA. Group A consisted of 10 sero-positive goats, group B with 10 sero-negative goats while group C with 7 sero-negative goats that served as control-unvaccinated group. All goats of groups A and B were vaccinated using Glanvac 6^{TM} vaccine twice at 1-month apart. One month after the second vaccination, all goats were challenged with 10^9

cfu/mL of live *C. pseudotuberculosis*. The goats were observed for clinical signs and were killed a month post-infection. The bacterium was most frequently isolated from lymph nodes of goats of group A but the rate of isolation showed no significant (p>0.05) difference among all groups. Gross lesion was observed in the prescapular lymph nodes of all groups (p>0.05). The goats vaccinated with Glanvac 6^{TM} either with sero-positive or sero-negative still developed signs and lesions of abscessation similar to the unvaccinated goats. Thus, the vaccine was unable to prevent goats from developing CLA.

Keywords: caseous lymphadenitis, *Corynebacterium pseudotuberculosis*, goats



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PATOGENESIS JANGKITAN Corynebacterium pseudotuberculosis DAN PERCUBAAN VAKSIN TERHADAP BENGKAK NODUS LIMFA PADA KAMBING

Oleh

NUR ADZA RINA MOHD NORDI

November 2016

Pengerusi: Prof. Mohd Zamri Saad Fakulti: Perubatan Veterinar

pseudotuberculosis adalah seienis Corynebacterium bakteria vand bertanggungjawab dalam menyebabkan penyakit bengkak nodus limfa pada kambing dan biri-biri.Penyakit ini tersebar meluas di seluruh dunia dan bakteria tersebut boleh berada di persekitaran selama beberapa bulan. Penghapusan penyakit ini sukar dilakukan dan ianya mudah merebak kepada haiwan-haiwan yang naïf. Penyebaran dan patologi penyakit in tidak difahami sepenuhnya. Penyelidikan ini dijalankan bagi mengenalpasti kaedah terlazim С. pseudotuberculosis menjangkiti kambing, kesan terhadap kambing secara klinikal dan patologikal selepas jangkitan serta tahap antibodi yang terbentuk dalam darah kambing, serta keberkesanan vaksin yang berada di pasaran ke atas sebaran dan kawalan penyakit ini pada kambing di Malaysia.

Dua puluh ekor kambing dewasa telah dipilih untuk penyelidikan ini dan dibahagikan kepada 4 kumpulan. Kambing-kambing tersebut telah dijangkiti dengan 10⁷ cfu/mL bakteria C. Pseudotuberculosis hidup melalui tiga kaedah. Kaedah-kaedah tersebut adalah; melalui kulit, saluran pernafasan dan mulut. Satu kumpulan kambing bertindak sebagai kawalan dan tidak dijangkiti bakteria. Kambing-kambing tersebut diperhatikan selama 30 hari untuk sebarang perubahan yang berkaitan dengan penyakit bengkak nodus limfa. Sepanjang masa itu, suhu badan dan sampel darah diambil mengikut selang masa vang ditetapkan. Pada minggu pertama jangkitan, semua kambing terjangkit kelihatan murung, kurang selera makan, dan suhu badan meningkat. Kambing-kambing yang dijangkiti pada kulit mempunyai bengkak yang mengandungi nanah pada kawasan terjangkit. Kambing-kambing terjangkit melalui kulit dan saluran pernafasan mengalami pengurangan dalam jumlah hemoglobin dalam darah. Hanya kambing terjangkit melalui kulit yg menunjukkan kenaikan bererti (p<0.05) dalam jumlah sel darah putih, dengan kenaikan jumlah sel neutrofil dan monosit. Kambing-kambing terjangkit melalui kulit menunjukkan tanda klinikal dan perubahan pada darah yang paling parah.

Selepas 30 hari jangkitan, semua kambing disembelih, Lesi-lesi kasar diperhatikan dan sampel organ-organ dalaman beserta nodus limfa diambil bagi pengasingan semula bacteria dan pemeriksaan histopatologi. Kebanyakkan pengasingan semula bacteria didapati dari kambing terjangkit melalui kulit. 40% kambing terdapat bacteria pada hati, 80% bacteria pada nodus limfa preskapular, dan 40% pada nodus limfa submandibular. Hanya 20% kambing terjangkit melalui saluran pernafasan mempunyai bacteria pada hati. Tiada pengasingan semula bakteria didapati dari kambing terjangkit melalui mulut dan kambing tidak terjangkit. Antara lesi kasar yang dapat dilihat adalah; pembentukan nanah pada nodus limfa, kemerahan pada paru-paru. buah pinggang dan nodus limfa. Lesi histopatologi menunjukkan kambing terjangkit melalui kulit mempunyai lesi yang lebih parah. Melalui penyelidikan ini, didapati nodus limfa yang paling banyak dijangkiti adalah nodus limfa preskapular, Jangkitan oleh C. pseudotuberculosis melalui kulit mempunyai lesi keseluruhan yang paling parah.

Berdasarkan keputusan penyelidikan di atas, 9 ekor kambing dewasa telah dipilih untuk penyelidikan seterusnya. Semua kambing dijangkiti melalui kulit dengan 107 cfu/mL C. pseudotuberculosis. Kambing-kambing tersebut diperhatikan selama 3 bulan untuk sebarang tanda berkaitan bengkak nodus limfa. Suhu badan dan sampel darah diambil mengikut selang masa yang ditetapkan. Semua kambing terjangkit kelihatan tidak aktif pada minggu pertama jangkitan, mempunyai bengkak pada kawasan jangkitan, dan mengalami kenaikan suhu badan. Terdapat juga pembesaran pada nodus limfa submandibular sebesar 3 cm. Pembesaran tersebut dikesan pada seekor kambing pada minggu ke 8 jangkitan. Dalam penyelidikan ini, jumlah haemoglobin dalam darah adalah normal sepanjang 3 bulan penyelidikan, kecuali beberapa hari di penghujung penyelidikan. Jumlah sel darah putih dalam darah adalah tinggi selama 39 hari selepas jangkitan, tetapi kembali ke paras normal selepas itu. Jumlah neutrofil dan monosit dalam darah juga mengalami kenaikan yang sama. Paras IgG dalam serum kambing terjangkit juga di analisis. Paras IgG dalam serum meningkat dan melepasi titik potong pada hari kesepuluh selepas jangkitan. Paras IgG meningkat paling tinggi pada hari ke-14 selepas jangkitan. Bacaan ELISA mula menurun pada hari ke-53 selepas jangkitan dan menghampiri titik potong pada penghujung tempoh penyelidikan.

Tiga ekor kambing disembelih pada setiap bulan. Semasa post-mortem, lesi kasar diperhatikan dan sampel organ dalaman dan nodus limfa diambil untuk pengasingan semula bacteria dan pemeriksaan histopatologi. С. pseudotuberculosis berjaya diasingkan dari semua nodus limfa yang bernanah dan dari paru-paru seekor kambing yang disembelih pada bulan kedua selepas jangkitan. Terdapat nanah pada nodus limfa preskapular 2 ekor kambing yang disembelih pada bulan pertama selepas jangkitan. Pada bulan kedua selepas jangkitan, terdapat pendarahan pada paru-paru seekor kambing dan nanah pada nodus limfa preskapular semua kambing. Pada bulan ketiga selepas jangkitan, semua kambing terjangkit mempunyai organ dalaman yang kelihatan normal pada mata kasar. Tetapi, terdapat nanah pada nodus limfa preskapular pada semua kambing terjangkit dan nanah pada nodus limfa submandibular dalam seekor kambing terjangkit. Pemeriksaan histologi menunjukkan penebalan pada septa interalveolar, kemerahan pada hati, kewujudan sel-sel radang pada hati, degenerasi lemak pada sel hati, kesesakan pada sel darah dalam buah pinggang, dan kast di dalam tubul buah pinggang. Nodus limfa yang bernanah menunjukkan keadaan nanah yang biasa. Nanah tersebut diukur dan didapati bahawa diameter bahagian tengah nanah tersebut meningkat dengan peningkatan masa selepas jangkitan. Keputusan ini menunjukkan penyakit ini bertambah teruk dengan peningkatan masa jangkitan.

Bengkak nodus limfa adalah penyakit yang penting dalam industri kambing di Malaysia, malahan di serata dunia. Oleh itu, pengawalan penyakit ini amat penting. Di Malavsia, hanva terdapat satu vaksin komersial terhadap penvakit ini. Oleh itu, keberkesanan vaksin ini harus dikaji. 27 ekor kambing dengan status serologi yang berbeza telah dipilih untuk penyelidikan ini. Kumpulan A mengandungi 10 ekor kambing sero-positif, kumpulan B mempunyai 10 sekor kambing sero-negatif dan kumpulan C terdiri daripada 7 ekor kambing seronegatif yang bertindak sebagai kawalan. Semua kambing dalam kumpulan A dan B divaksin menggunakan vaksin komersial tersebut sebanyak 2 kali, dalam jarak 1 bulan. Sebulan selepas vaksinasi kedua, semua kambing dijangkitkan dengan 10⁹ cfu/ml C. pseudotuberculosis. Kambing-kambing tersebut diperhatikan untuk sebarang perubahan dan disembelih sebulan selepas jangkitan. Kebanyakkan pengasingan semula bakteria didapati dari kambing dalam kumpulan A. Tetapi, tiada perbezaan bererti (p>0.05) antara semua kumpulan kambing. Kebanyakkan lesi kasar didapati pada nodus limfa preskapular, dan tiada perbezaan bererti (p>0.05) antara semua kumpulan kambing. Lesi kasar menunjukkan bahawa kambing yang divaksin dengan Glanvac 6[™] juga akan terjangkit dengan penyakit ini dan mempunyai nanah pada nodus linfa. Oleh itu, yaksin ini tidak berkesan sepenuhnya dalam menghalang penyakit ini kerana keberadaan penyakit ini tidak mempunyai perbezaan yang bererti (p>0.05) antara kambing yang divaksin dan kambing yang tidak divaksin.

Kata kunci: bengkak nodus limfa, *Corynebacterium pseudotuberculosis*, kambing

ACKNOWLEDGEMENTS

Firstly, I want to express my appreciation and gratitude to my supervisor, Prof. Dr. Mohd Zamri Saad for his time, guidance, support and knowledge. The appreciation also goes to my co-supervisors, Assoc. Prof. Dr. Siti Khairani Bejo and Assoc. Prof. Dr. Faez Firdaus Jesse Abdullah for their guidance, ideas, knowledge and support.

Special thanks to staffs of Faculty of Veterinary Medicine, particularly staffs of the histopathology laboratories, large animals wards, ruminant research centre, clinical pathology laboratory and theriogenology laboratory, as well as the staff of Kota Kinabalu Veterinary Laboratory for their help and knowledge that they share.

I also want to express my appreciations to other post-graduates students that willing to help me in doing the research and interpretating the data obtained.

My uttermost gratitude and love goes to my parents, family and friends who were endlessly and tirelessly supporting and helping me whenever I needed them.

My sincere gratitude for the concerns and encouragements given by all during the accomplishment of the project and my years of study in Universiti Putra Malaysia. Thank you to those who have contributed directly and indirectly to this project. Thank you very much from the bottom of my heart. This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Mohd Zamri Saad, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Siti Khairani Bejo, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

Faez Firdaus Jesse Abdullah, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

ROBIAH BINTI YUNUS, PhD Professor and Dean

School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of DeputyVice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matri	c No.:

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: ______ Name of Chairman of Supervisory Committee: Professor Dr. Mohd Zamri Saad

Signature: Name of Member of Supervisory Committee:

Associate Professor Dr. Siti Khairani Bejo

Signature: Name of Member of Supervisory Committee:

Associate Professor Dr.Faez Firdaus Jesse Abdullah

TABLE OF CONTENTS

ABS ACK APP DEC LIST LIST	ABSTRACT ABSTRAK ACKNOWLEDGEMENTS APPROVALS DECLARATIONS LIST OF TABLES LIST OF FIGURES LIST OF ABBREVIATIONS						
СНА	CHAPTER						
1		1					
2	 LITERATURE REVIEW 2.1 Corynebacterium pseudotuberculosis 2.2 Virulence factor of Corynebacterium pseudotuberculosis 2.2.1 Phospholipase D (PLD) 2.2.2 Cell wall lipids 2.3 Caseous Lymphadenitis (CLA) 2.4 Mode of transmission for CLA 2.5 Risk Factors of CLA 2.6 Pathogenesis of CLA 2.7 Pathology of CLA 2.8 Control and prevention of CLA in goats and sheep 2.9 Diagnosis of CLA in goats 2.9.1 Direct method 2.9.2 Indirect method 2.10 CLA status in Malaysia 2.11 Zoonotic potential of CLA 	3 4 4 5 6 7 8 9 11 14 14 14 16 17					
3	MATERIALS AND METHODS3.1General Materials and Methods 3.1.13.1.1Inoculum Preparation 3.1.23.1.2Clinical Observations 3.1.33.1.3Blood Sampling 3.1.43.1.4Post-mortem Examination 3.1.53.1.5Bacterial Isolation 3.1.63.1.6Histopathological Examination3.2Experimental Infection of Goats with Corynebacterium pseudotuberculosis 3.2.13.2.1The Animals 3.2.23.2.2The Inoculums 3.2.33.2.4Clinical Parameters 3.2.53.2.5Post-mortem Examination 3.2.6	19 19 20 21 22 23 24 25 25 25 25 26 26 26 26					

 \bigcirc

	3.3	Efficacy of a Commercially Available Vaccine against	30
		Caseous Lymphadenitis in Malaysia	
		3.3.1 The Animals	30
		3.3.2 The Inoculums	30
		3.3.3 The Vaccine	30
		3.3.4 Experimental Designs	30
		3.3.5 Sampling and Sample Processing	31
		3.3.6 Statistical Analysis	31
			01
4	RES	ULTS AND DISCUSSION	
-	4.1	Experimental 1: Short Infection	32
	4.1	4.1.1 Clinical Signs	32
		4.1.2 Body Temperature	33
		4.1.3 Complete Blood Count (CBC)	34
		4.1.4 Serum Biochemistry	45
			57
		4.1.6 Gross Lesions	60
		4.1.7 Histological Lesions	62
		4.1.8 Size of Abscesses	66
	4.2	Experiment 2: Prolonged Infection	67
		4.2.1 Clinical Signs	67
		4.2.2 Body Temperature	68
		4.2.3 Complete Blood Count (CBC)	68
		4.2.4 Serum Biochemistry	72
		4.2.5 Antibody Pattern	74
		4.2.6 Bacterial Re-isolation	74
		4.2.7 Gross Lesions	77
		4.2.8 Histological Lesions	79
		4.2.9 Size of Abscesses	86
	4.3	Efficacy of a Commercially Available Vaccine against	89
		Caseous Lymphadenitis in Malaysia	
		4.3.1 Serological status	89
		4.3.2 Bacterial re-isolation	90
		4.3.3 Gross Lesions	90
5	GEN	ERAL DISCUSSION	93
6		MARY, CONCLUSIONS AND RECOMMENDATIONS FUTURE RESEARCH	101
REFE		-8	100
			102
APPE		S F STUDENT	113 115
		BLICATIONS	
LIST		SLICATIONS	116

LIST OF TABLES

Table		Page
3.1	The gross lesions scored for organs and lymph nodes examined.	21
3.2	The histological lesions scored for organs and lymph nodes examined.	24
4.1	Mean score for clinical signs observed in goats for 1 week post-inoculation with <i>C. pseudotuberculosis</i> .	32
4.2	Weekly mean values of RBC, PCV and Hb of different group of goats.	35
4.3	Weekly mean values for total WBC, lymphocytes and monocytes count in different group of goats.	38
4.4	Weekly mean values for band neutrophils and segmented neutrophils count in different group of goats	38
4.5	Weekly mean values for eosinophils and basophils count in diffe <mark>rent</mark> group of goats	39
4.6	Week <mark>ly mean values</mark> for plasma protein concentration in blood of goats in different groups.	43
4.7	Weekly mean values for PT and APTT in different group of goats.	44
4.8	Weekl <mark>y mean values for sodium (Na) and pota</mark> ssium (K) in different group of goats.	46
4.9	Weekly mean values for calcium (Ca) and chloride (Cl) in different group of goats.	46
4.10	Weekly mean values for AST, ALP and GGT in different group of goats.	49
4.11	Weekly mean values for urea and creatinine in different group of goats.	52
4.12	Weekly mean values for albumin, globulin and A:G ratio in different group of goats.	54
4.13	Weekly mean values for CK and total protein in different group of goats.	56
4.14	Isolation of <i>C. pseudotuberculosis</i> from organs and lymph nodes of all groups of goats.	59

6

	4.15	Incidence of <i>C. pseudotuberculosis</i> isolation from organs and lymph nodes of all groups of goats.	59
	4.16	Mean gross lesions scoring for organs and lymph nodes in all groups of goats.	61
	4.17	Histological lesions score of the lymph nodes of all groups.	64
	4.18	Histological lesions score of the organs of all infected goats	64
	4.19	Average thickness of different layers of abscess in the affected lymph nodes (x10 ³ μ m).	66
	4.20	Mean score for clinical signs observed in goats for 1 week post-inoculation with <i>C. pseudotuberculosis</i> .	67
	4.21	Isolation of <i>C. pseudotuberculosis</i> from organs and lymph nodes of all groups of goats.	76
	4.22	Mean gross lesions scoring for organs and lymph nodes in all groups of goats.	78
	4.23	Average histological lesions score of the organs in all groups of goats.	79
	4.24	Average measurements of the different layers of the abscesses (x10 ³ μm) in different groups of goats.	87
	4.25	Serological status of goats before and after vaccination with $Glanvac6^{TM}$.	89
	4.26	Percentages of successful isolation of <i>C. pseudotuberculosis</i> in lymph nodes of infected goats.	90
	4.27	Percentages of incidence of gross lesions observed in all groups of goats.	91
	4.28	Mean gross lesions scoring for organs and lymph nodes in all groups of goats.	92
(\mathbf{G})			

LIST OF FIGURES Figure 3.1 Flowchart of the experimental infection with C. *pseudotuberculosis* in goats 4.1 Average rectal temperature of different groups of goats following exposure to *C. pseudotuberculosis*. The intradermal group showed significantly higher body temperature during the first 4 days on infection. 4.2 The average packed cell volume (PCV)in all goats of all groups following exposure to C. pseudotuberculosis. The normal range for RBC count in goats is between 0.22-0.38 L/L. 4.3 The average haemoglobin levels in all goats of all groups following exposure to C. pseudotuberculosis. The readings below 80 g/L were abnormal, which was observed mainly among goats of groups 1 (intradermal) and 2 (intranasal). 4.4 The white blood cells (WBC) counts for goats of all groups following exposure to *C. pseudotuberculosis*. The normal range for WBC count in goats is between 4-13x10⁹/L. Note the intradermally exposed goats showed consistently high WBC counts throughout the study period 4.5 Band neutrophil concentrations in blood of goats following various routes of infection by C. pseudotuberculosis. The intradermally exposed group showed significantly (p<0.05) high neutrophilic counts, especially in later days in the experiment. 4.6 Segmented neutrophil concentrations in blood of goats following various routes of exposures to С. pseudotuberculosis. Readings above 7.2x10⁹/L are abnormal for goats. Again, intradermally exposed goats showed high counts of segmented neutrophils throughout the study period.

- 4.7 Monocytes concentration in blood of goats following 42 exposure to live *C. pseudotuberculosis.* Readings above 0.55x10⁹/L are abnormal. Again, the intradermal exposure leads to significantly (p<0.05) high monocyte counts throughout the study period.
- 4.8 Calcium (Ca) concentration in serum of goats following 47 exposure to live *C. pseudotuberculosis.* Normal referral range for sodium concentration in goats is between 2.2-3.2 mmol/L.

29

33

36

36

40

41

41

- 4.9 Aspartate Aminotransferase (AST) values in blood in all 50 groups of goats infected with *C. pseudotuberculosis.* Readings above 100U/L are abnormal, which was observed late in the infection.
- 4.10 Alkaline phosphatase (ALP) levels in blood of goats 50 exposed to *C. pseudotuberculosis.* Readings above 200U/L are abnormal. The patterns indicate no or mild liver injury.
- 4.11 Urea concentration in serum of goats exposed to *C*. 53 *pseudotuberculosis*. The normal value for goats is between 3.5-7.1mmol/L.
- 4.12 Total protein concentration in serum of goats exposed to *C.* 57 *pseudotuberculosis.* The normal value for goats is between 55-70 g/L.
- 4.13 Abscess at the prescapular lymph node of a goat following 61 intradermal exposure to *C. pseudotuberculosis*. Notice the thick fibrous capsule surrounding the pasty, whitish pus.
- 4.14 Multifocal abscesses in the liver of a goat following 62 intranasal exposure to *C. pseudotuberculosis*
- 4.15 Photomicrograph of kidney section showing congested 65 blood vessels and necrotic tubular cells. HE x400.
- 4.16 Photomicrograph of liver section showing congestion of 65 veins at the portal triad and necrotic hepatocytes. HE x100.
- 4.17 Photomicrograph of prescapular lymph node showing 67 capsulated abscess. The innermost layer (N) is the necrotic cells layer, surrounded by inflammatory cells layer (I) that consists of neutrophils and macrophages, and the outermost fibrous capsule layer (F). HE x40.
- 4.18 Average haemoglobin level in all goats exposed 69 intradermally to *C. pseudotuberculosis*. The readings below 80 g/L were abnormal. The levels are generally within the normal range.
- 4.19: Average white blood cells (WBC) count of the goats 70 following prolonged intradermal exposure to *C. pseudotuberculosis.* The values above 13x10⁹/L are abnormal. There were high WBC counts from day 3 until day 60 before fluctuating until the end of study period on day 90.
- 4.20 Average concentrations of band neutrophils in blood of 71 goats following intradermal exposure to *C. pseudotuberculosis*. They experienced severe left-shift on

the 10th days post-inoculation.

- 4 21 Average concentrations of segmented neutrophils in blood 71 doats followina intradermal exposure to of C. Readings 7.2x10⁹/L are pseudotuberculosis. above abnormal. Abnormal concentrations were in the early stage of the infection.
- 4.22 Average concentrations of monocytes in blood of goats 72 following intradermal exposure to *C. pseudotuberculosis*. Readings above 0.55x10⁹/L are abnormal. The concentration remained high throughout the experimental period that reflects the severe infection.
- 4.23 Average alkaline phosphatase (ALP) values in blood of 73 goats following chronic intradermal exposure to *C. pseudotuberculosis*. Readings above 200U/L are abnormal. Although the levels were high during most of the study period, the changes do not reflect the health status of the goats.
- 4.24 Average IgG level in the goats' serum against days of 74 infection. The cutoff point is 0.2521. The red line is the cutoff value for the ELISA
- 4.25 PCR image of bacterial isolation from abscessed lymph 75 nodes of goats slaughtered at 3 month p.i.; Lane 1: Positive control, Lane 2-4: Prescapular lymph nodes, Lane 5: Submandibular lymph node, Lane 6: Negative control.
- 4.26 Abscess in the prescapular lymph nodes of one of the 78 goats slaughtered at 1 month post-inoculation with *C. pseudotuberculosis.*
- 4.27 Petechiation in the lungs of one of the goats killed at 2 79 months post-inoculation with *C. pseudotuberculosis*
- 4.28 Photomicrograph of the lung section of an infected goat 80 killed after 2 months of infection showing infiltration of inflammatory cells (circle) consisted of alveolar macrophage and polymorphonuclear cells and congestion of alveolar blood vessels (arrows). HE x40.
- 4.29 Photomicrograph of the lung section of an infected goat 81 killed after 2 months of infection showing pulmonary edema and haemorrhages. HE x40.
- 4.30 Photomicrograph of lung section of an infected goat killed 81 after 3 months of infection. Notice the large bronchus associated lymphoid tissue (BALT) consisted of mainly the lymphocytes. HE x40.

- 4.31 Photomicrograph of the liver section of an infected goat 82 killed after 1 month of infection showing individual necrosis scattered throughout the section (arrows), and mild sinusoids congestion.HE x200.
- 4.32 Photomicrograph of the liver section of an infected goat 83 killed after 2 months of infection showing the presence of individual neutrophils (arrows). HE x200.
- 4.33 Photomicrograph of a liver with fatty degeneration in a goat 83 slaughtered at 3 months post-infection. HE x100
- 4.34 Photomicrograph of a kidney showing infiltration of 84 inflammatory cells in the interstitial area of the kidney of a goat killed after 1 month of infection. HE x40
- 4.35 Photomicrograph of a kidney showing congested blood 85 vessels in between renal tubules of a goat killed after 1 month of infection. HE x40.
- 4.36 Photomicrograph of a kidney showing urinary casts in 85 some collecting tubules of a goat killed after 3 months of infection. HE x40.
- 4.37 Photomicrograph of a prescapular lymph node showing 86 infiltration of neutrophils (arrows) in a goat killed after 3 months of infection. HE x100.
- 4.38 Photomicrograph of a lymph node with abscess. Note the 88 distinct different layers of the abscess. HE x40.
- 4.39 Photomicrograph of a lymph node with abscess at higher 88 magnification. Note the distinct different layers of the abscess. HE x100

LIST OF ABBREVIATIONS

CLA:	caseous lymphadenitis
AGPT:	agar gel precipitation test
ELISA:	enzyme-linked immunosorbent assay
PBS:	phosphate buffered saline
BHI:	brain-heart infusion
CFU:	colony forming unit
SD:	standard deviation
CMN:	Corynebacterium, Mycobacterium, Nocardia
CBC:	complete blood count
WBC:	white blood cells
RBC:	red blood cells
PCV:	packed cells volume
PCR:	polymerase chain reaction
ANOVA:	analysis of variance
PLD:	phospholipase D
LN:	lymph node
lgG:	immunoglobulin G
bp:	base pairs
°C:	degrees celcius
μL:	microlitre
g:	gram
p.i.:	post-infection
HE:	Haematoxylin and Eosin stain
rpm:	round per minute

6

- APTT: activated partial thromboplastin time
- PT: prothrombin time
- BUN: blood urea nitrogen
- TP: total protein
- AST: aspartate aminotransferase
- ALP: alkaline phosphatise
- CK: creatinine kinase
- Alb: albumin
- GGT: gamma-glutamyl transferase
- A:G: albumin and globulin ratio
- OD: optical density

CHAPTER 1

INTRODUCTION

Corynebacterium pseudotuberculosis is the aetiological agent of a disease called caseous lymphadenitis (CLA) or "cheesy gland disease" that affects goats and sheep worldwide. It is a non-spore forming, facultative, intracellular Gram positive bacterium. In stained smears, the rods appear isolated and have pleomorphic form, from coccoids to filamentous rods that grouped in parallel cells or in a format similar to Chinese letters. In sheep blood agar incubated at 37°C, the organism appears as cream-colored colonies with a β -hemolysis zone at 48 h. It has a broad spectrum of hosts and causes clinical disease in sheep, goats, cattle, horses, pigs, deer, camels and laboratory animals, as well as in human (Moore *et al.*, 2010).

CLA is characterized by chronic abscess formation in superficial lymph nodes such as the submandibular, parotid, pre-scapular, subiliac, popliteal and supramammary lymph nodes. The internal lymph nodes are also affected such the mediastinal, bronchial and mesenteric lymph nodes. Occasionally, visceral organs like liver, lung and spleen might have the same abscessation (O'Reilly *et al.*, 2008).CLA has a severe economic impact on the sheep and goat industries due to reduction in wool, meat and milk production and condemnation of carcass and skins (Fontaine and Baird, 2008).Thus, it is important to understand more about the disease and to determine whether the current control and prevention measure is efficient in controlling the disease.

There are few postulated and confirmed routes of infection for the disease. It can occur through direct or indirect contact or through wounds that come into contact with pus from the sick animals. In naturally observed infections, the main portal of bacterial entry is generally accepted to be through the skin, normally as a result of the presence of minor wounds and abrasions (Collett *et al.*, 1994). A respiratory route for transfer of *C. pseudotuberculosis* has been postulated and some researchers suggest that animals with pulmonary lesions may present the major source of exposure to naive animals within a flock (Ellis *et al.*, 1987). In addition, head and neck lesions are though to arise from bacterial entry via the oral cavity (Ashfaq and Campbell, 1979). To date, no study had been conducted to confirm the best route of transmission of the disease. Therefore, the most efficient route of infection should be determined to better understand the mechanism of the infection.

Vaccination against CLA is one of the ways to prevent infection by *C.* psedotuberculosis. Glanvac 6^{TM} is a vaccine against several important small ruminant diseases that is currently available in Malaysia. It is a multicomponent adjuvanted vaccine containing *C.* pseudotuberculosis and

5 Clostridium sp., which are Clostridium perfringens type D, Clostridium tetani, Clostridium novyi type B, Clostridium septicum and Clostridium chauvoei. It has been evaluated in many countries such as in Australia, Canada, and Saudi Arabia but not in Malaysia.

This study was conducted to evaluate *C. pseudotuberculosis* infection through different routes of infection. The viscerals organs and lymph nodes were examined for lesions and the commercial vaccine was assessed for efficiency against CLA in goats in Malaysia. The objectives of the present study are:

- 1. To determine the most efficient route of infection in producing CLA in goats.
- 2. To assess the clinical and pathological changes in goats following acute and chronic experimental infection by *C. pseudotuberculosis.*
- 3. To determine the efficiency of Glanvac 6[™] vaccine against CLA in Malaysia.

Based on the objectives of the study, the hypotheses are:

- 1. The most efficient route of infection is via dermal route.
- 2. Following infection, lymph nodes abscessation is most frequently developed, with involvement of the visceral organs in which the severity increased with increasing time of infection.
- 3. The Glanvac 6[™] vaccine is efficient in preventing caseous lymphadenitis among goatsin Malaysia.

REFERENCES

- Abdinasir, Y.O., Jesse, F.F.A., Saharee, A.A., (2012). Sero-prevalence of Caseous Lymphadenitis Evaluated by Agar Gel Precipitation Test among Small Ruminant Flocks in East Coast Economic Regions in Peninsular Malaysia. Journal of Animal and Veterinary Advances 11, 3474-3480.
- Abdinasir, Y.O., Jesse, F.F.A., Saharee, A.A., Haron, A.W., Jasni, S., Rasedee, A., (2012). Haematological and Biochemical Alterations in Mice Following Experimental Infection with Whole Cell and Exotoxin (PLD) Extracted from *C. Pseudotuberculosis*. Journal of Animal and Veterinary Advances 11(24), 4660-4667.
- Alloui, M.N., Kaba, J., Alloui, N., (2011). Prevalence and Risk Factors of CLA in Sheep and Goats of Batna Area (Algeria). Research Opinion in Animal and Veterinary Sciences 1(3), 162-164.
- Al-Rawashdeh, O.F., Al-Qudah, K.M., (2000). Effect of Shearing on the Incidence of Caseous Lymphadenitis in Awassi Sheep in Jordan. Journal of Veterinary Medicine Series B 47, 287-293.
- Annas, S., Zamri-Saad, M., Jesse, F.F., Zunita, Z., (2015). Comparative Clinicopathological Changes in Buffalo and Cattle Following Infection by *Pasteurella multocida* B: 2. Microbial Pathogenesis 88, 94-102.
- Arsenault, J., Dubreuil, P., Girard, C., Simard, C., Bélanger, D., Maedi-Visna, (2003). Impact on Productivity in Quebec Sheep Flocks (Canada). Preventive Veterinary Medicine 59, 125–137.
- Ashfaq, M.K., Campbell, S.G., (1979). A Survey of Caseous Lymphadenitis and Its Etiology in Goats in the United States. Veterinary Medicine, Small Animal Clinician 74, 1161-1165.
- Ashfaq, M.K., Campbell, S.G., (1980). Experimentally induced caseous lymphadenitis in goats. American Journal of Veterinary Research 41(11), 1789-1792.
- Bain, P.J., (2011). Liver, In: Duncan & Prasse's Veterinary Laboratory Medicine, Clinical Pathology, 211-229.
- Baird, G.(2006). Treatment of Ovine Caseous Lymphadenitis. Veterinary Record 159, 500
- Baird, G., Synge, B., Dercksend, (2004). Survey of Caseous Lymphadenitis Seroprevalence in British Terminal Sire Sheep Breeds. Veterinary Record 154, 505-506.
- Baird, G.J., (2007). Caseous Lymphadenitis. In: Diseases of Sheep. 4th Edition, Blackwell Publishing, 306-311.

- Baired, G.J., Fontaine, M.C., (2007). *Corynebacterium pseudotuberculosis* and Its Role in Ovine Caseous Lymphadenitis. Journal of Comparative Pathology 137, 179-210.
- Barksdale, L., Linder, R., Sulea, I.T., Pollice, M., (1981). Phospholipase D Activity of *Corynebacterium pseudotuberculosis* (*Corynebacterium ovis*) and *Corynebacterium ulcerans*, a Distinctive Marker within the Genus *Corynebacterium*. Journal of Clinical Microbiology 13, 335-343.
- Bastos, B.L., Dias Portela, R.W., Dorella, F.A., Ribeiro, D., Seyffert, N., (2012) Corynebacterium pseudotuberculosis: Immunological Responses in Animal Models and Zoonotic Potential. Journal of Clinical and Cellular Immunology S4:005.
- Bastos, B.L., Dias Portela, R.W., Dorella, F.A., Ribeiro, D., Seyffert, N., Castro, T.L.P., Miyoshi, A., Oliveira, S.C., Meyer, R., Azevedo, V., (2012). *Corynebacterium pseudotuberculosis*: Immunological Responses in Animal Models and Zoonotic Potential. Journal of Clinical and Cellular Immunology S4:005.
- Batey, R.G. (1985). Aspects of Pathogenesis in a Mice Model of Infection by *Corynebacterium pseudotuberculosis*. Australia Journal of Experimental Medical Science 64, 237-249.
- Batey, R.G. (1986). Pathogenesis of Caseous Lymphadenitis in Sheep and Goats. Australian Veterinary Journal 63, 269-273
- Biberstein, E.L., Knight, H.D., Jang, S., (1971). Two Biotypes of *Corynebacterium pseudotuberculosis*. Veterinary Record 89, 691-692.
- Binns, S.H., Bairley, M., Green, L.E., (2002). Postal Survey of Ovine Caseous Lymphadenitis in the United Kingdom between 1990 and 1999. Veterinary Record 150: 263–268
- Binns, S.H., Green, L.E., Bailey, M. (2007). Development and Validation of an ELISA to Detect Antibodies to *Corynebacterium pseudotuberculosis* in Ovine Sera. Veterinary Microbiology 123, 169-179.
- Braverman, Y., Chizov-Ginzburg, A., Saran, A., (1999). The Role of Houseflies (*Musca domestica*) in Harbouring *Corynebacterium pseudotuberculosis* in Dairy Herds in Israel.Scientific and Technical Review of OIE (International Office of Epizootics) 18, 681–690.
- Brown, C.C., Olander, H.J., (1987). Caseous Lymphadenitis of Goats and Sheep: a Review. Veterinary Bulletin 57, 1–11.
- Brown, C.C., Olander, H.J., Biberstein, E.L.,Morse, S.M., (1986).Use of a Toxoid Vaccine to Protect Goats Against Intradermal Challenge Exposure to *Corynebacterium pseudotuberculosis*. American Journal of Veterinary Research 47, 1116–1119.

- Burrel, D.H., (1980). A Haemolysis Inhibition Test for Detection of Antibody to Corynebacterium ovis Exotoxin. Researchin Veterinary Science 28 (2), 190-194.
- Burtis, C.A., Ashwood, R.A., Bruns, E., (2000). Tiefz Fundamentals of Clinical Chemistry. Saunders. St. Louis.
- Campbell, S.G., Ashfaq, M.K., Tashjian, J.J., (1982) Caseous Lymphadenitis in Goats in the USA. In: Proceedings 3rd International Conference on Goat Production and Disease. Tucson. Arizona 449–454
- Cardenas, L., Clements, J.D., (1992). Oral Immunization Using Live Attenuated *Salmonella* spp. as Carriers of Foreign Antigens. Clinical Microbiology Reviews 5, 328-342.
- Cetinkaya, B., Karahan, M., Atil, E., (2002). Identification of *Corynebacterium* pseudotuberculosis Isolates From Sheep and Goats by PCR. Veterinary Microbiology88, 75–83.
- Chaplin, P.J., De Rose, R., Boyle, J.S., Mcwaters, P., (1999). Targeting Improves the Efficacy of a DNA Vaccine against *Corynebacterium pseudotuberculosis* in Sheep. Infection and Immunity67, 6434-6438
- Collett, M.G., Bath, G.F., Cameron, C.M., (1994). Corynebacterium pseudotuberculosis Infections. In: Infectious Diseases of Livestock With Special Reference To Southern Africe, 2nd Edition, Coetzer,J., Thomson, G.R., Tustin,R.C., 2nd Edition, Oxford University Press, Cape Town, 1387-1395.
- Connor, K.M., Quirie, M.M., Baird, G., Donachie, W., (2000). Characterization of United Kingdom Isolates of *Corynebacterium pseudotuberculosis* Using Pulsed-Field Gel Electrophoresis. Journal of Clinical Microbiology 38, 2633-2637.
- Davis, H.L., Mancini, M., Michel, M.L., Whalen, R.G., (1996). DNA-Mediated Immunization to Hepatitis B Surface Antigen: Longevity of Primary Response and Effect of Boost. Vaccine14, 910-915.
- Dercksen, D.P., Brinkhof, J.M.A., Dekker-Nooren, T., van Maanen,K., Bode, C.F., Baird, G., Kamp, E.M., (2000). A Comparison of Four Serological Tests for the Diagnosis of Caseous Lymphadenitis in Sheep and Goats. Veterinary Microbiology 75, 167–175.
- Dorella, F.A., Pacheco, L.G., Oliveira, S.C., Miyoshi, A., Azavedo, V., (2006). *Corynebacterium pseudotuberculosis*: Microbiology, Biochemical Properties, Pathogenesis and Molecular Studies of Virulence. Veterinary Research37, 201-218.

- Dorella, F.A., Pacheco, L.G., Seyffert, N., Portela, R.W., Meyer, R., (1998). *Corynebacterium pseudotuberculosis* For Use in Sheep. Journal of American Veterinary Medicine Association212, 1765–1768.
- Dowling, A., Hodgson, J.C., Schock, A., Donachie, W., Eckersall, P.D., Mckendrick, I.J., (2002). Experimental Induction of Pneumonic Pasteurellosis in Calves by Intrtracheal Infection with *Pasteurella multocida* Biotype A: 3. Research in Veterinary Science 73, 37-44.
- Egen, N.B., Cuevas, W.A., Mcnamara, P.J., Sammons, D.W., Humphreys, R., Songer, J.G., (1989). Purification of the Phopholipase D of *Corynebacterium pseudotuberculosis* by Recycling Isoelectric Focusing. American Journal of Veterinary Research 50, 1319-1322.
- Eggleton, D.G., Doidge, C.V., Middleton, H.D., Minty, D.W., (1991). Immunization Against Ovine Caseous Lymphadenitis: Efficacy of the Monocomponent *Corynebacterium pseudotuberculosis* Toxoid Vaccine and Combined Clostridial-Corynebacterial Vaccines. Australian Veterinary Journal 68, 320-321.
- Eggleton, D.G., Middleton, H.D., Doidge, C.V., Minty D.W., (1991). Immunization Against Ovine Caseous Lymphadenitis: Comparison of *Corynebacterium pseudotuberculosis* Vaccines With and Without Bacterial Cells. Australian Veterinary Journal 68, 317–319
- Ellis, T.M., Sutherland, S.S., Wilkinson, F.C., (1987). The Role of *Corynebacterium pseudotuberculosis* Lung Lesions in the Transmission of This Bacterium to Other Sheep. Australian Veterinary Journal 64, 261-263.
- Ferreira, R., Fonseca, L.S., Lilenbaum, W., (2002). Agar Gel Immunodiffusion Test (AGID) Evaluation for Detection of Bovine Paratuberculosis in Rio de Janeiro, Brazil. Letters in Applies Microbiology 35, 173-175.
- Fontaine, M.C., Baird, G., Connor, K.M., (2006). Vaccination Confers Significant Protection of Sheep against Infection with a Virulent United Kingdom Strain of *Corynebacterium pseudotuberculosis*. Vaccine 24, 5986–5996.
- Fontaine, M.C., Baird, G.J., (2008).CaseousLymphadenitis. Small Ruminant Research 76, 42–48
- Goldberg, A.C., Lipsky, B.A., Plorde, J.J., (1981). Suppurative Granulomatous Lymphadenitis Caused by *Corynebacterium ovis (Pseudotuberculosis)*. American Society of Clinical Pathologists 76, 486-490.
- Hard, G.C., (1969). Electron Microscopic Examination of *Corynebacterium ovis*. Journal of Bacteriology 97(3), 1480-1485.

- Hassan, N.A., Al-Humiany, A.A., Bahobail, A.S., Mansour, A.M.A., (2011). Bacteriological and Pathological Studies on Caseous Lymphadenitis in Sheep in Saudi Arabia. International Journal of Microbiological Research 2(1), 28-37.
- Hawari, A.D. (2008). *Corynebacterium pseudotuberculosis* Infection (CLA) in Camels (*Camelus diomedarius*) in Jordon. American Journal of Animal and Veterinary Sciences 3(2), 68-72.
- Hodgson, J.C., Dagleish, M.P., Gibbard, L., Bayne, C.W., Finlayson, J., Moon, G.M., Nath, M., (2013). Seven Strains of Mice as Potential Models of Bovine Pasteurellosis Following Intranasal Challenge with a Bovine Pneumonic Strain of *Pasteurella multocida* A: 3, Comparisons of Disease and Pathological Outcomes. Research in Veterinary Science 94, 634-640.
- Hodgson, A.L., Carter, K., Tacedjian, M., Krywult, J., Coener, L.A., Mccoll, M., Cameron, A., (1999). Efficacy of an Ovine Caseous Lymphadenitis Vaccine Formulated Using a Genetically Inactive Form of the *Corynebacterium pseudotuberculosis* Phospholipase D. Vaccine 17, 802-808.
- Hodgson, A.L., Tachedjian, M., Corner, L.A., Radford, A.J., (1994). Protection of Sheep against Caseous Lymphadenitis by Use of a Single Oral Dose of Live Recombinant *Corynebacterium pseudotuberculosis*. Infection and Immunity 62, 5275-5280.
- Ibtisam, M.A. (2008). Some Clinicopathological and Pathological Studies of *Corynebacterium ovis* Infection in Sheep. Egypt Journal of Comparative Pathology of Clinical Pathology 21(1), 327-343.
- Ismail, A.A., Hamid, Y.M.A., (1972). Studies on the Effect of Some Chemical Disinfectants Used in Veterinary Practice in *Corynebacterium ovis*. Journal of Egyptian Veterinary Medicine Association 32, 195–202.
- Jesse, F.F.A., Randolf, P.S.S., Saharee, A.A., Wahid, A.H., Zamri-Saad, M., Jasni, S., Omar, A.R., Adamu, L., Abdinasir, Y.O., (2013). Clinicopathological response of mice following oral route infection of *Corynebacterium pseudotuberculosis*. Journal of Agriculture and Veterinary Science 2, 38-42.
- Jiskoot, W., Kersten, G.F.A., Beuvery, E.C., (2002). Vaccine. In: Crommelin, D.J.A., Sindelar, R.D., Pharmaceutical Biotechnology – An introduction for Pharmacists and Pharmaceutical Scientists, 2nd Edition. London: Taylor and Francis Group, 259–282.
- Johnson, E.H., Vidal, C.E.S., Santa Rosa, J., Kass, P.H., (1993). Observations on Goats Experimentally Infected with *Corynebacterium pseudotuberculosis*. Small Ruminant Research 12, 357-369.

- Jolly, R.D., (1966). Some Observations on Surface Lipids of Virulent and Attenuated Strains of *Corynebacterium ovis*. Journal of Applied Bacteriology 29, 189-196.
- Jolly, R.D.,(1965). The Pathogenesis of Experimental *Corynebacterium ovis* Infection in Mice. New Zealand Veterinary Journal, 13, 141-142.
- Jones, D., Collins, M.D., (1986). Irregular, Nonsporing Gram-Positive Rods. In: Sneath, P.H.A. Bergey's Manual of Systematic Bacteriology, 2nd edition. Baltimore: Williams and Wilkins, 1261–1282.
- Jubb, K.V.F., Kennedy, P.C., Palmer, N., (1998). Pathology of Domestic Animals, 3rd Edition Academic Press, Orlando, FA.
- Keslin, M.H., Mccoy, E.L., Mccusker, J.J., Lutch, J.S., (1979). *Corynebacterium pseudotuberculosis*. A New Cause of Infectious and Eosinophilic Pneumonia. American Journal of Medicine, 67, 228-231.
- Komala, T.S., Ramlan, M., Yeoh, N.N., Surayani, A.R., Sharifah Hamidah, S.M., (2008).A Survey of Caseous Lymphadenitis in Small Ruminant FarmsFrom Two Districts in Perak, Malaysia – Kinta and Hilir Perak.Tropical Biomedicine25(3), 196–201.
- Kuria, J.K.N., Mbuthia, P.G., Kang'ethe, E.K., Wahome, R.G., (2001). Caseous Lymphadenitis in Goats: The Pathogenesis, Incubation Period and Serological Response after Experimental Infection. Veterinary Research Communication 25, 89-97.
- Liu, D.T.L, Chan, W.M.,Fan, D.S.P,Lam, D.S.C, (2005). An Infected Hydrogel Buckle with *Corynebacterium pseudotuberculosis* British Journal of Ophthalmology, 89:245-246
- Lopez, J.F., Wonc, F.M., Quesada, J., (1966). *Corynebacterium pseudotuberculosis* First Case of Human Infection. American Journal of Clinical Pathology 46, 562.
- Mahmood, Z.K.H., F.F. Jesse, A.A. Saharee, S. Jasni, R. Yusoff and H. Wahid (2015). Clinico-Pathological Changes in Goats Challenged with *Corynebacterium pseudotuberculosis* and its Exotoxin (PLD). American Journal of Animal and Veterinary Sciences 10 (3): 112.132
- Mahmood, Z.K.H., F.F. Jesse, A.A. Saharee, S. Jasni, R. Yusoff and H. Wahid (2015). Assessment of Blood Changes Post-challenge with *Corynebacterium pseudotuberculosis* and Its Exotoxin (Phospholipase D): AComprehensive Study in Goat. Veterinary World 8(9): 1105-1117.
- Maki, L.R., Shen, S.H., Bergstrom, R.C., Stetzenbach, L.D., (1985).Diagnosis of *Corynebacterium pseudotuberculosis* Infections inSheep, Using an Enzyme-Linked Immunosorbent Assay. American Journal of Veterinary Research 46 (1), 212–214.

Malaysian Veterinary Protocol: Caseous Lymphadenitis, (2011). PVM 3(13):1/2011

- Menzies, P.I., Muckle, C.A., Brogden, K.A., Robinson, L., (1991). A Field Trial to Evaluate a Whole Cell Vaccine for the Prevention of Caseous Lymphadenitis in Sheep and Goat Flocks. Canada Journal of Veterinary Research 55, 362-366.
- Menzies, P.I., Muckle, C.A., Hwang, Y.T., Songer, J.G., (1994). Evaluation of an Enzyme Linked Immunosorbent Assay Usingan *Escherichia coli* Recombinant Phospholipase D Antigen for theDiagnosis of *Corynebacterium pseudotuberculosis* Infection. Small Ruminant Research 13, 193–198.
- Mills, A.E., Mitchell, R.D., Lim, E.K., (1997). *Corynebacterium pseudotuberculosis* is a Cause of Human Necrotising Granulomatous Lymphadenitis. Pathology 29(2), 231-233.
- Miyoshi, A., Azevedo, V., (2009). Antigens of *Corynebacterium pseudotuberculosis* and Prospects for Vaccine Development. Expert Review Vaccines8(2), 205–213.
- Moller, K., Agerholm, J.S., Ahrens, P.,(2000). Abscess Disease, Caseous Lymphadenitis and Pulmonary Adenomatosis in Imported Sheep. Journal of Veterinary Medicine B: Infectious Diseases and Veteterinery Public Health47, 55-62.
- Moore, R., Miyoshi, A., Pacheco, L.G.C., Seyffert, N., Azevedo, V., (2010). Corynebacterium and Arcanobacterium. In: Pathogenesis of Bacterial Infection in Animals. 1st Edition, Blackwell Publishing, 133-147.
- Muckle, C.A., Gyles, C.L., (1982). Characterization of Strains of *Corynebacterium pseudotuberculosis*. Canada Journal of Comparative Medicine 46, 206-208.
- Nguyen, D., Diamond, L., (2000). Neutrophilia Pattern in: Diagnostic Haematology, a Pattern Approach. 1st Edition, Butterworth-Heinemann, London.
- O'Reilly, K.M., Green, L.E., Malone, F.E., Medley, G.F., (2008). Parameter Estimation and Simulations of a Mathematical Model of *Corynebacterium pseudotuberculosis* Transmission in Sheep. Preventive Veterinary Medicine83, 242–259.

- Okwor, E.C., Eze, D.C., Okonkwo, K.E., Ibu, J.O., (2011). Comparative Evaluation of Agar Gel Precipitation Test (AGPT) and Indirect Haemaagglutination Test (IHA) for the Detection of Antibodies against Infectious Bursal Disease (IBD) Virus in Village Chickens. African Journal of Biotechnology 10(71), 16024-16027.
- Othman, A.M, Jesse, F.F.A., Adza- Rina, M.N., Ilyasu, Y., Zamri-Saad, M., Wahid, A.H., Saharee, A.A. and Mohd-Azmi. M.L. (2014). Haematological, Biochemical and Serum Electrolyte Changes in Non-Pregnant Boer Does Inoculated With *Corynebacterium pseudotuberculosis* Via Various Routes. Journal of Agriculture and Veterinary Science 7, 05-08
- Othman, A.M., Abba, Y., Jesse, F.F.A., Ilyasu, Y.M., Saharee A.A., Haron, A.W., Zamri-Saad, M., Lila, M.A.M., (2016). Reproductive Pathological Changes Associated with Experimental Subchronic *Corynebacterium pseudotuberculosis* Infection in Non-Pregnant Boer Does. Journal of Pathogens. Hindawi Publishing Corporation.
- Paton, M.W., Walker, S.B., Rose, I.R., Watt, G.F., (2003). Prevalence of Caseous Lymphadenitis and Usage of Caseous Lymphadenitis Vaccines in Sheep Flocks. Australia Veterinary Journal 81, 91-95
- Paule, J.A., Azevedo, V., Regis, L.F., Carminati, R., Bahia, C.R., Vale, V.L.C., Moura-Costa L.F., Freire, S.M., Nascimento, I., Schaer, R., Goes, A.M., Meyer, R., (2003). Experimental Corynebacterium pseudotuberculosis Infection in Goats: Kinetics of IgG and Interferon-y Production, IgG Avidity and Antigen Recognition by Western Blotting. Veterinary Immunology and Immunopathology 96, 129-139.
- Peel, M.M., Palmer, G.G., Stacpoole, A.M., Kerr, T.G., (1997). Human Lymphadenitis Due to Corynebacterium pseudotuberculosis: Report of Ten Cases from Australia and Review. Clinical Infectious Diseases 24, 185-191.
- Pekelder, J.J., (2003). Caseous Lymphadenitis. In: Martin, W.B., Aitken, I.D., Diseases of Sheep. 3rd Edition, Blackwell Science, Oxford.
- Pepin, M., Fontaine, J.J., Pardon, P., Marly, J., Parody, A.L., (1991). Histopathology of the Early Phase during Experimental *Corynebacterium pseudotuberculosis* Infection in Lambs. Veterinary Microbiology 29, 123-134.
- Pepin, M., Pardon, P., Lantier, F., Marly, J., Levieux, D., Lamand, M., (1990). Experimental *Corynebacterium pseudotuberculosis* Infection in Lambs: Kinetics of Bacterial Dissemination and Inflammation. Veterinary Microbiology 26, 381-392.
- Pepin, M., Paton, M., Hodgson, A.L, (1994). Pathogenesis and Epidemiology of *Corynebacterium pseudotuberculosis* Infection in Sheep. Current Topics in Veterinary Research 1, 63-82.

Pepin, M., Seow, H.F., Corner, L., Rothel, J.S., Hodgson, A.L., (1997). Cytokine Gene Expression in Sheep Following Experimental Infection with Various Strains of *Corynebacterium pseudotuberculosis* differing in Virulence. Veterinary Research 28, 149-163.

Pfizer Australia Pty Ltd (2010). Glanvac[™] 6 pamphlet, Pfizer Animal Health.

- Pinder, A.G., Rogers, S.C., Morris, K., James, P.E., (2009). Haemoglobin Saturation Controls the Red Blood Cells Mediated Hypoxic Vasorelaxation, In: Oxygen Transport in Tissue. Springer U.S, 13-20.
- Quinn, P.J., Carter, M.E., Markey, B., Carter, G.R., (1994) *Corynebacterium* species and *Rhodococcus equi* In: Clinical Veterinary Microbiology. Wolfe PublishingCompany, London.
- Radostits, N.M., Gay, C.C., Hinchcliff, W.K., Constable, P.D., (2007). Veterinary Medicine: A Textbook of the Disease of Cattle, Horses, Sheep, Pigs and Goats. 10th Edition. Saunders Publisher, USA, 314-325.
- Radostits, O.M., Gay, C.C., Blood, D.C., Hinchcliff, K.W., (2000). Caseous Lymphadenitis in Sheep and Goats. Veterinary Medicine, 9th Edition, W.B. Saunders, London, 727-730.
- Rhyan. J.C., Saari, D.A., Williams, E.S., (1992). Gross and Microscopic Lesions of Naturally Occurring Tuberculosis in a Captive Herd of Wapiti (*Cervus elaphus nelson*) in Colorado. Journal of Veterinary Diagnosis and Investigation 4, 428-433.
- Saragea, A., Maximescu, P., Meitert, E., Stuparu, I., (1966). Incidence and Geographical Distribution of Phage Types of *Corynebacterium diphtheriae* in the Dynamics of the Epidemic Process of Diphtheria in the Rumanian Socialist Republic. Microbiology, Parasitology and Epidemiology11, 351-362
- Schreuder, B.E., Ter Laak E.A., Dercksen, D.P., (1994). Eradication of Caseous Lymphadenitis in Sheep with the Help of a Newly Developed ELISA Technique. Veterinary Record135, 174-176.
- Senturk, S., Temizel, M., (2006). Clinical Efficacy of Rifamycin SV Combined with Oxytetracycline in the Treatment of Caseous Lymphadenitis in Sheep. Veterinary Record 159, 216–217
- Serres E., Hehenberger E., Allen A.L., (2011). Multiple Pyogranulomas in a Katahdin Ewe. Canadian Veterinary Journal 52, 555-560
- Seyffert, N., Guimaraes, A.S., Pacheco, L.G.C., Portela, R.W., Bastos, B.L., Dorella, F.A., Heinemann, M.B., Lage, A.P., Gouveia, A.M.G., Meyer, R., Miyoshi, A., Azevedo, V.,(2010). High Seroprevalence of Caseous Lymphadenitis in Brazilian Goat Herds Revealed by *Corynebacterium*

pseudotuberculosis Secreted Protein-Based ELISA. Research in Veterinary Science 88, 50-55

- Smith, M.C., Sherman, D., (1994). Caseous Lymphadenitis. Goat Medicine, 1st Edition, Lea and Febier, Iowa.
- Songer, J.G. (1997). Bacterial Phospholipases and Their Role in Virulence. Trendsin Microbiology 5, 156-160.
- Songer, J.G., Beckenbach, K., Marshall, M.M., Olson, G.B., Kelly, L., (1988). Biochemical and Genetic Characterization of *Corynebacterium peudotuberculosis*. American Journal of Veterinary Research 49, 221-226.
- Stoops, S.G., Renshaw, H.W., Thilsted, J.P., (1984). Ovine Caseous Lymphadenitis: Disease Prevalence, Lesion Distribution and Thoracic Menifestations in a Population of Mature Culled Sheep from Western United States. American Journal of Veteterinary Research 45, 557-561.
- Sutherland, S.S., Eillis, T.M., Mercy, A.R., Paton, M.W. & Middleton, H., (1987). Evaluation of an Enzyme-Linked Immunosorbent Assay for the Detection of *Corynebacterium pseudotuberculosis* Infection in Sheep. Australian Veterinary Journal 64 (9), 263-266.
- Tashjian, J.J., Campbell, S.G., (1983). Interaction between Caprine Macrophages and *Corynebacterium pseudotuberculosis*: An Electron Microscopic Study. American Journal of Veterinary Research 44, 690-693.
- Ter Laak, E.A., Bosch, J., Bijl, G.C., Screuder, B.E.C., (1992). DoubleAntibody Sandwich Enzyme-Linked Immunosorbent Assay andImmunoblot Analysis Used for Control of Caseous Lymphadenitis in Goats and Sheep. American Journal of Veterinary Research 53 (7), 1125–1132.
- Toshach, S., Valentine, A., Sigurdson, S., (1977). Bacteriophage Typing of *Corynebacterium diphtheriae*. Jornal of Infectious Diseases 136, 655-660
- Tunkel A.R., (2012). Fever, In: MSD Manual Professional Version. Merck & Co., Kenilworth, New Jersey, United States of America.
- Ural, K., Alic, D., Haydardedeoglu, A.E., (2008). Corynebacterium pseudotuberculosis Infection in Saanen×Kilis Crossbred (White) Goats in Ankara, Turkey and Effective Kanamycin Treatment: A Prospective, Randomized, Double-Blinded, Placebo-Controlled Clinical Trial. Small Ruminant Research77, 84–88
- Wagner, K.S., White, J.M., Crowcroft, N.S., Mann, G., Efstratiou, A., (2010). Diphtheriain the United Kingdom, 1986-2008: the Increasing Role of *Corynebacteriumulcerans*. Epidemiology and Infection 138, 1519-1530.

- Walker, J., Jackson, H., Brandon, M.R., Meeusen, E., (1991). Lymphocyte Subpopulations in Pyogranulomas of Caseous Lymphadenitis. Clinical and Experimental Immunology 86(1), 13-18.
- Wawrzkiewicz, J., Dziedzic, B., Koziol, T., (1989). Sensitivity and Specificity of a Modified Agar Gel Precipitation Test and Its Application to the Diagnosis of Enzootic Bovine Leukosis. Acta Virologica 33(2), 143-50.
- West, D.M., Bruere, A.N., Ridler, A.L., (2002). Caseous Lymphadenitis. In: The Sheep: Health, Disease and Production. Foundation for Veterinary Continuing Education, Massey University, New Zealand.
- Williamson, L.H., (2001). Caseous Lymphadenitis in Small Ruminants. Veterinary Clinics of North America: Food Animals Practice 17, 359-371.
- Wolf, C., (2007). North America. In: Diseases of Sheep. 4th Edition, Blackwell Publishing, 511.
- Yeruham, I., Braverman, Y., Shpigel, N.Y., (1996). Mastitis in Dairy Cattle Caused by *Corynebacterium pseudotuberculosis* and the Feasibility of Transmission by Houseflies. Veterinary Quarterly18, 87–89.
- Zavoshti, F.R., Khoojine, A.B.S., Helan, J.A., Hassanzadeh, B., Heydari, A.A., (2012). Frequency of caseous lymphadenitis (CLA) in Sheep Slaughtered in an Abbatoir in Tabriz: Comparison of Bacterial Culture and Pathological Study. Comparative Clinical Pathology 21, 667-671.