

UNIVERSITI PUTRA MALAYSIA

ASSOCIATION BETWEEN HUMAN CYTOMEGALOVIRUS RELATED FACTORS AND DEVELOPMENT OF THE DISEASE IN RENAL AND BONE MARROW TRANSPLANT RECIPIENTS IN A TERTIARY HOSPITAL, MALAYSIA

MOHD FAHMI BIN MASTUKI

FPSK(M) 2014 18



ASSOCIATION BETWEEN HUMAN CYTOMEGALOVIRUS RELATED FACTORS AND DEVELOPMENT OF THE DISEASE IN RENAL AND BONE MARROW TRANSPLANT RECIPIENTS IN A TERTIARY

HOSPITAL, MALAYSIA

By MOHD FAHMI BIN MASTUKI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

May 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Dedicated with love and gratitude to:

Supervisor: Dr. Niazlin Mohd Taib

Father: Mastuki bin Damar

Mother: Laili binti Gupar

Brother and Sisters: Mohd Azhari, Mohd Azizi, Mohd Fahrul Razi, Mohd Farid, Mohd Hafizuddin, Norhafizah, Mohd Faidz.

Wife and Children: Siti Fairuz Abdul Rashid and Muhammad Hanif bin Mohd Fahmi and Naurah Hani binti Mohd Fahmi.

"The love of a family is life's greatest blessing"

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

ASSOCIATION BETWEEN HUMAN CYTOMEGALOVIRUS RELATED FACTORS AND DEVELOPMENT OF THE DISEASE IN RENAL AND BONE MARROW TRANSPLANT RECIPIENTS IN A TERTIARY HOSPITAL, MALAYSIA

By

MOHD FAHMI BIN MASTUKI

May 2014

Chair: Niazlin Mohd Taib, PhD Faculty: Medicine and Health Sciences

Human cytomegalovirus (HCMV) infection is known to be a major infectious complication after transplantation which associated with significant morbidity and mortality in solid organ and bone marrow transplant recipients. We studied the viral factors of HCMV and correlate results with the development of HCMV disease. This aim of this study is to detect HCMV, their genotypes and co-infection with other herpesviruses namely Epstein-Barr virus (EBV), Human herpesvirus 6 (HHV-6) and Human herpesvirus 7 (HHV-7) in post-solid organ and bone marrow transplant recipients and to correlate them with the clinical presentation and outcome of HCMV disease. In this study, 100 blood samples from renal transplant recipients and 100 bone marrow transplant recipients in Kuala Lumpur Hospital were included. All tests were carried out by real time polymerase chain reaction (qPCR). HCMV were detected in higher incidence compared to other herpes virus indicating that the virus was the most common virus infecting the immunosuppressed patients. The results revealed that the incidence of HCMV infection were 78% and 63% in renal and bone marrow transplant recipients respectively. We found that patients with high viral load show symptoms of HCMV disease, whereby fever being most common symptom. As Malaysia has multi-races citizens, we also demonstrate the incidence of HCMV infection among renal and bone marrow transplant recipients by ethnicity namely Malay, Chinese, Indian and other minority races as 'others'. In renal transplant recipients, there was no significant difference between the ethnic. Nevertheless, we found that there was a significant HCMV positivity among races in bone marrow transplant recipients where Malays were the most infected. One of the pathogenesis of HCMV depends on the genes encoding envelope glycoprotein that associated with different clinical outcomes. Reactivation of latent viral infection by HCMV and other herpesviruses results in active viral infection after organ transplantation and may cause complications. HCMV genotyping analysis revealed that all three HCMV gB, gH and gN genotypes were presence in the population where gB1 strain being the most common gene detected in both renal (100%) and bone marrow (100%) transplant recipients. Mix infection by more than one HCMV genotypes was also detected with various percentages with the gB+gH+gN combination was the least type of

mix infection. We also found that recipient with high HCMV viral load (>5,000 copies/mL) has increased risk of developing HCMV disease. No statistically significant difference was found between type of genotypes and the manifestation of HCMV disease (p>0.05). Co-infection with other herpesviruses with HCMV disease was significant in bone marrow transplant recipients but not significant in renal transplant recipient.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

HUBUNGKAIT ANTARA FAKTOR-FAKTOR BERKAITAN SITOMEGALOVIRUS MANUSIA DENGAN PEMBENTUKAN PENYAKITNYA DIKALANGAN PENERIMA ORGAN BUAH PINGGANG DAN SUM-SUM TULANG DI SEBUAH HOSPITAL TERTIARY, MALAYSIA

Oleh

MOHD FAHMI BIN MASTUKI

Mei 2014

Pengerusi: Niazlin Mohd Taib, PhD Fakulti: Perubatan dan Sains Kesihatan

Jangkitan sitomegalovirus manusia (HCMV) telah diketahui sebagai komplikasi jangkitan yang utama yang berlaku selepas pemindahan organ dimana ia dikaitkan dengan kadar morbiditi (kejadian) dan mortaliti (kematian) di kalangan penerima pemindahan organ dan sumsum tulang. Kami mengkaji faktor-faktor virus HCMV dan menghubungkaitkan dengan penyakit HCMV. Tujuan kajian ini dijalankan adalah untuk mengesan HCMV, genotip-genotipnya, dan jangkitan bersama dengan herpesvirus-herpesvirus yang lain iaitu virus Epstein-Barr (EBV), herpesvirus manusia jenis 6 (HHV-6) dan herpesvirus manusia jenis 7 (HHV-7) di kalangan penerima organ dan sumsum tulang dan mengaitkan faktor-faktor ini dengan tanda-tanda klinikal dan akibat oleh penyakit HCMV. Sebanyak 100 sampel darah dari penerima buah pinggang dan 100 sampel darah dari penerima sumsum tulang daripada penerima-penerima organ di Hospital Kuala Lumpur telah dimasukkan di dalam kajian ini. Semua ujian dijalankan dengan menggunakan teknik real time polymerase reaction (qPCR). HCMV dikesan pada kadar yang lebih tinggi berbanding dengan herpesvirus lain. Ini menunjukkan bahawa ia adalah virus yang biasa menjangkiti pesakit-pesakit berimuniti rendah. Dalam populasi kajian ini sebanyak 78% dan 63% jangkitan HCMV masing-masing terhadap penerima buah pinggang dan sumsum tulang. Kami mendapati bahawa pesakit yang mempunyai jumlah viral load yang tinggi menunjukkan gejala-gejala HCMV dimana demam menjadi gejala yang paling banyak didapati. Oleh kerana Malaysia mempunyai pelbagai kaum iaitu Melayu, Cina, India dan lain-lain, kami mengkaji kadar jangkita HCMV dikalangan etnik tetapi tiada perbezaan yang ketara secara statistik yang tentang jangkitan HCMV dengan jenis etnik. Walaubagaimanapun, kami mendapati kadar jangkitan HCMV yang ketara di kalangan pesakit yang menerima sumsum tulang bangsa Melayu. Salah satu pathogenesis HCMV adalah bergantung kepada gen-gen yang mengawalatur sampul glycoprotein yang dikaitkan dengan pelbagai konsekuensi klinikal. Pengaktifan semula jangkitan virus latent oleh HCMV dan herpesvirus-herpesvirus yang lain mengakibatkan jangkitan virus aktif selepas pemindahan organ dan boleh menyebabkan komplikasi. Analisis genotip HCMV menunjukkan bahawa ketiga-tiga genotip hadir didalam populasi kajian ini dimana jenis gB1 adalah gen yang paling banyak dikesan didalam kedua-



dua jenis pemindahan organ iaitu 100% setiap jenis. Jangkitan campuran oleh lebih daripada satu genotip HCMV juga dikenalpasti dengan peratusan yang berbezabeza dimana kombinasi jangkitan gB+gH+gN menjadi kombinasi yang paling sedikit menyebabkan jangkitan campuran Kami juga mendapati bahawa penerima organ yang mepunyai viral load HCMV yang tinggi (>5,000 *copies/mL*) meningkatkan risiko untuk mempunyai penyakit HCMV. Tiada perbezaan statistik yang jelas didapati antara jenis-jenis genotip dengan manifestasi penyakit HCMV. Kaitan jangkitan bersama antara herpesvirus-herpesvirus yang lain dengan penyakit HCMV didapati signifikan di kalangan penerima sumsum tulang (p<0.05) manakala tidak signifikan di kalangan penerima organ buah pinggang.



I certify that a Thesis Examination Committee has met on 23rd May 2014 to conduct the final examination of Mohd Fahmi bin Mastuki on his thesis entitled "Association Between Human Cytomegalovirus Related Factors and Development of the Disease in Renal and Bone Marrow Transplant Recipients in a Tertiary Hospital, Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Mohd Nasir Mohd Desa, PhD

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Zamberi Sekawi, PhD

Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Internal Examiner)

Shuhaimi Mustafa, PhD

Professor Halal Products Research Institute Universiti Putra Malaysia (Internal Examiner)

Mostafizur Rahman, PhD

Professor Universiti Kebangsaan Malaysia Malaysia (External Examiner)

NORITAH OMAR, PhD Associate Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 21 July 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Niazlin Mohd Taib

Medical Lecturer Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairman)

Siti Norbaya Masri

Medical Lecturer Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Zuridah Hassan, PhD

Associate Professor Faculty of Health Sciences Universiti Teknologi MARA (Member)

Mangalam Sinniah

Medical Doctor and Consultant in Virology Head of Department Virology Unit Kuala Lumpur Hospital (Member)

BUJANG BIN KIM HUAT, Ph.D.

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date: 29 th August 2014	
0		

Name and Matric No.: Mohd Fahmi bin Mastuki GS29777

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	
Name of	
Chairman of	
Supervisory	
Committee: Dr. Niazlin Mohd Taib	

Signature: _____ Name of Member of Supervisory Committee: Dr. Mangalam Sinniah

Signature: _____ Name of Member of Supervisory Committee: Dr. Siti Norbaya Masri Signature: _____ Name of Member of Supervisory Committee: Assoc. Prof. Dr. Hjh Zuridah Hassan

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRACT	iv
ACKNOWLEDGEMENTS	vi
APPROVAL	viii
DECLARATION	Х
LIST OF TABLES	xvi
LIST OF FIGURES	xviii
LIST OF ABBREVIATIONS	xxi

CHAPTER

 \bigcirc

1	INT	RODUCTION	1
	1.1	Problem Statement	3
	1.2	Significance of Research	3
	1.3	Objectives of Research	3
		1.3.1 General Objective	3
		1.3.2 Specific Objectives	3
2	LIT	ERATURE REVIEW	4
	2.1	General Information	4
		2.1.1 Viral Infection in Organ Transplant Recipients	4
		2.1.2 The family <i>Herpesviridae</i>	6
		2.1.3 Herpesvirus Infections in Immunocompromised	
		Patients.	9
	2.2	Biology of human cytomegalovirus (HCMV)	9
		2.2.1 Background	9
		2.2.2 HCMV Genome Organization and Structure	10
		2.2.3 HCMV Replication	11
		2.2.4 Clinical Manifestation of HCMV	13
		2.2.5 Transmission of HCMV	16
		2.2.6 Risk Factor in Transplantation	16
	2.3	Different Strains of HCMV	17
		2.3.1 HCMV Genotypes	17
		2.3.2 Glycoprotein B (gB)	17
		2.3.3 Glycoprotein N (gN)	18
		2.3.4 Glycoprotein H (gH)	19
	2.4	Definitions of HCMV Infection and Disease in Transplant	
		Recipients	20
	2.5	Laboratory Diagnosis of HCMV Infection	20
	2.6	Prevention of HCMV Infection	21
	2.7	Treatment of HCMV Infection	22
	2.8	Co-infection with Other Herpesviruses	23
		2.8.1 Human Herpesvirus Type-6 (HHV-6)	23
		2.8.2 Human Herpesvirus Type- 7 (HHV-7)	23
		2.8.3 Epstein-Barr Virus (EBV)	24

3	MATERIALS AND METHODS	26
	3.1 Determination of Human Cytomegalovirus Infection in	26
	Renal and Bone Marrow Transplant Recipients by	
	Ouantitative Real Time PCR.	
	3.1.1 Clinical Samples	26
	2.1.2 Paggants and Chamicals	26
	3.1.2 Reagents and Chemicals	20
	3.1.3 Instruments and equipment	26
	3.1.4 HCMV Detection and Quantitation by Real-Time	
	PCR	26
	3.2 Determination of genotypic distribution of HCMV	45
	envelope glycoprotein B, N and H in renal and bone	
	marrow transplant recipients by using SYBR Green-based	
	real time PCR.	
	3.2.1 Reagents and Chemical	27
	3.2.2 Instruments and equipment	27
	2.2.2 Clinical Samples	27
	2.2.4 List of Driver and Laternal Controls	27
	3.2.4 List of Primers and Internal Controls	27
	3.2.5 Real-time PCR for Genotyping of Glycoprotein B	
	(gB), Glycoprotein N (gN) and Glycoprotein H	
	(gH)	27
	3.3 Identification of human herpesvirus type-6 (HHV-6),	31
	human herpesvirus type-7 (HHV-7) and Epstein-Barr virus	
	(EBV) by using SYBR Green-based real time PCR.	
	3 3 1 Reagents and Chemical	31
	3.3.2 Instruments and equipment	31
	2.2.2 Clinical Samples	21
	2.2.4 List of Drivers and Internal Controls	21
	5.5.4 List of Primers and Internal Controls	31
	3.3.5 Real-time PCR for detection of HHV-6, HHV-7	31
	and EBV	22
	3.4 Correlation between HCMV viral loads, patient's	32
	demographic data, the occurrence of various HCMV	
	glycoproteins, co-infection with EBV, HHV-6 and HHV-7	
	and development of HCMV disease in the same study	
	population by statistical analysis.	
	3.4.1 Statistical Analysis	32
		-
4	RESULTS	34
	4.1 Qualitative and Quantitative analysis of HCMV	34
	4.1.1 Detection of HCMV DNA and quantification of	-
	HCMV viral load	34
	4.1.2 Incidence of HCMV Discoss	29
	4.1.2 Includence of inclusive Disease	30
	4.2 ACMIN Genotyping	42
	4.3 Co-infection with other human herpesvirus by using	10
	SYBR Green-based real time PCR.	43
	4.3.1 Detection of human herpesvirus Type-6 (HHV-6)	43
	4.3.2 Detection of human herpesvirus Type-7 (HHV-7)	44

		4.3.3 Detection of Epstein-Barr Virus (EBV)	45
	4.4	Correlation Studies on the occurrence of various HCMV	47
		genotypes, co-infection with EBV, HHV-6 and HHV-7	
		and development of HCMV disease.	
		4.4.1 Correlation between HCMV Genotypes and	
		HCMV Disease	47
		4.4.2 Correlation between co-infection with other	
		Herpersviruses and HCMV Disease	49
_	DIG		
5	DIS		51
	5.1	HCMV as the major viral infection in renal and bone	-1
	5.0	marrow transplant recipients.	51
	5.2	disease	51
	5.3	Co-infection with other herpesviruses as risk factors for developing HCMV disease	53
6	CO	NCLUSION AND RECOMMENDATION	54
LIMITAT	ION	OF THE STUDY	55
REFEREN	NCES		56
APPENDI	CES		64
BIODATA	OF	STUDENT	80

C

C

LIST OF TABLES

Table		Page
2.1	Classification of Herpesvirus subfamily.	7
3.1	Primers to amplify regions of gB, gH and gN for genotypic identification of HCMV.	28
3.2	Housekeeping genes primers used as internal control	30
3.3	Primers to amplify HHV-6, HHV-7 and EBV for co- infection detection	33
4.1	Characteristics of renal and bone marrow transplant recipients (January 2011 - June 2012)	35
4.2	Indications of renal and bone marrow transplantation from January 2011 - June 2012	36
4.3	HCMV positivity of renal and bone marrow transplant recipients according to gender and ethnicity (January 2011 - June 2012)	37
4.4	Percentage of symptoms present in renal and bone marrow transplant recipients.	38
4.5	Viral load quantification of 100 renal transplant recipients.	64
4.6	Viral load quantification of 100 bone marrow transplant recipients.	67
4.7	Incidence of HCMV infections in renal and bone marrow transplant recipients in Malaysia from January 2011 June 2012	39
4.8	Genotypes gB1, gB2, gH and gN with the HCMV viral load and HCMV-related symptoms appeared in the renal transplant reginigate	74
4.9	Genotypes gB1, gB2, gH and gN with the HCMV viral load and HCMV-related symptoms appeared in the bone	77
4.10	The percentages of HCMV genotyping in single and mixed infection with more than one genotypes in renal and bone	42
4.11	marrow transplant recipients. Summary of HCMV, HHV-6, HHV-7 and EBV infection in renal transplant recipients.	47
4.12	Summary of HCMV, HHV-6, HHV-7 and EBV infection in bone marrow transplant recipients.	47
4.13(a)	The distribution of HCMV genotypes among symptomatic and asymptomatic renal transplant recipients.	48

4.13(b)	Statistical significance of the distribution of HCMV genotypes among symptomatic and asymptomatic renal transplant recipients.	48
4.14(a)	The distribution of HCMV genotypes among symptomatic and asymptomatic bone marrow transplant recipients.c	48
4.14(b)	4.14(b) : Statistical significance of the distribution of HCMV genotypes among symptomatic and asymptomatic bone marrow transplant recipients.	49
4.15	Statistical significance of the distribution of herpesviruses co-infection among symptomatic and asymptomatic renal transplant recipients.	49
4.16	Statistical significance of the distribution herpesviruses co- infection among symptomatic and asymptomatic bone	50

marrow transplant recipients

C

LIST OF FIGURES

Figure		Page
2.1	Time of presentation of common viral illnesses post-transplant	5
2.2	Composite phylogenetic tree for herpes viruses based on amino acid sequence alignments of eight sets of homologous genes. Human herpesviruses are boxed, to emphasize their distribution throughout the genera.	8
2.3	Schematic map of a HCMV genome	10
2.4	A schematic representation of human cytomegalovirus virion.	12
2.5	Replication cycle of HCMV	15
4.1	Incidence of human cytomegalovirus by gender in renal transplantation.	40
4.2	Incidence of human cytomegalovirus by gender in bone marrow transplantation.	40
4.3	Incidence of human cytomegalovirus by races in renal transplantation.	41
4.4	Incidence of human cytomegalovirus by races in bone marrow transplantation.	41
4.5	HCMV gB1 amplification by using real-time PCR assay. The flat lines show that there are no amplification occurs in negative samples and no template control (NTC).	70
4.6	Specificity of HCMV gB1 genotypes demonstrated by melting curve analysis.	70
4.7	HCMV gB2 amplification by using real-time PCR assay.	71
4.8	Specificity of HCMV gB2 genotypes demonstrated by melting curve analysis.	71
4.9	HCMV gH amplification by using real-time PCR assay.	72
4.10	Specificity of HCMV gH genotypes demonstrated by melting curve analysis.	72

4.11	HCMV gN genotype	73
4.12	Specificity of HCMV gN genotypes demonstrated by melting curve analysis.	73
4.13	HHV-6 amplification by using real-time PCR assay.	43
4.14	Specificity of HHV-6 demonstrated by melting curve analysis.	44
4.15	HHV-7 amplification by using real-time PCR assay.	44
4.16	Specificity of HHV-7 demonstrated by melting curve analysis.	45
4.17	EBV amplification by using real-time PCR assay.	46
4.18	Specificity of EBV demonstrated by melting curve analysis.	46

4.11

C

LIST OF ABBREVIATIONS

°C	Degree Celcius
μL	Micro Liter
μΜ	Micro Moles
cop/µL	Copies per Micro Liter
DNA	Deox yribonucleic Acid
L	Liter
mL	Milliliter
NC	Negative Control
PC	Positive Control
IC	Internal Control
PCR	Polymerase Chain Reaction
qPCR	Quantitative Polymerase Chain Reaction
HCMV	Human Cytomegalovirus
HHV-6	Human Herpervirus- Type 6
HHV-7	Human Herpesvirus- Type 7
EBV	Epstein Barr virus

CHAPTER 1

INTRODUCTION

Organ transplantation such as liver, kidney, heart, lung and bone marrow is a wellknown therapeutic option for many human diseases. It has become a standard therapy for selected end-stage diseases such as renal and bone marrow disease (Cukuranovic *et. al.*, 2012). However, complications may arise after transplantation which includes infection, allograft rejection and side effects of immunosuppressive therapy which remain major causes of morbidity and mortality following solid organ transplantation (Cukuranovic *et. al.*, 2012). According to Kotton & Fishman (2005), viruses are the most common cause of opportunistic infection in post transplantation which can leads to several complications.

Many viral infections after renal and bone marrow transplantation result from reactivation of "latent" viral infection in the host or from the graft. Latency means the virus exist in the host after primary infection then remain in inactive stage. Some of the virus "awakes" due to some reasons including the nature of the virus, host immune response and infection of tissues (Kotton & Fishman, 2005). While other viruses are constantly replicating at low levels, some latent viruses are metabolically inactive, determined by the effectiveness of the hosts immune response. Few factors contribute to viral activation after transplantation, including immune suppression (especially reduction of cytotoxic immunity), graft rejection therapy, inflammation (cytokines), and tissue injury (Kotton & Fishman, 2005; Cukuranovic *et. al.*, 2012).

The risk of infection in transplant recipients is determined by the intensity of exposure to potential viruses (epidemiologic exposure) and factors that cause patient susceptible to infection (immunosuppression) (Fishman & Rubin, 1998). Some incidence of viral infections result from community exposure such as influenza and adenovirus, while some are commonly transmitted within the allograft such as human cytomegalovirus (HCMV) and Epstein-Barr virus, and other are reactivated in the setting of immunosuppression such as varicella zoster virus (Kotton & Fishman, 2005). Moreover, the use of immunosuppressive agents to prevent the rejection of transplanted organs has increased the patients' susceptibility to opportunistic infection (Fishman, 2007).

Neonatal and immunocompromised patients usually associated with a wide range of diseases caused by HCMV infection (Xia & Zhang, 2010). HCMV infections can cause morbidity and mortality in neonatal and immunocompromised patients although advanced antiviral therapy has been introduced after renal and bone marrow transplantation to prevent the disease. It usually happened because of mutations associated with antiviral resistance (Xia & Zhang, 2010). Besides that, combination of viral factors and host immune responses also contribute to the pathogenesis of HCMV. Genomic polymorphism is one of the viral factors that usually associated with different clinical outcomes. Various genomic polymorphism that have been demonstrated among clinical isolates in the HCMV genes encoding envelope glycoprotein includes glycoprotein B (gB), glycoprotein N (gN),

glycoprotein O (gO), glycoprotein H (gH), and glycoprotein L (gL) (Xia & Zhang, 2010).

Besides in transplantation and hospitalised patients, the seroprevalence of HCMV is currently demonstrated all over the world, which also presence in the community, both in under developing and developing countries. Prevalence of HCMV disease varies from 80 to 100% in Africa and Latin America whereas lower in the northern hemisphere countries, which were 40 to 60% (Thomasini *et al.*, 2012). In the Malaysian states of Selangor and Wilayah Persekutuan 92% seroprevalence of HCMV were reported from blinded study performed in 2012 among healthy blood donors (Camalxaman *et al.*, 2012). In other studies done in Hospital Universiti Kebangsaan Malaysia (Jamal *et. al.*, 1998) and Hospital Universiti Sains Malaysia (Ahmad *et. al.*, 2006), it has been documented that both transfused thalassemic patients and regular blood donors have equally high seroprevalence rates. Thus, HCMV infection is expansive, universal and relevant to be studied intensely (Camalxaman *et al.*, 2012).

In solid organ transplant (SOT) recipients, HCMV has become a preventable cause of mortality and morbidity. If prevention strategy is not taken HCMV disease usually occurs during the first 3 months after SOT. However, this onset has been delayed in patients who received HCMV prophylaxis (Razonable & Humar, 2013). Due to aberrant immune response within the allograft, HCMV has the affinity to invade the allograft. Not only that, due to its ability to modulate the immune system, it also has numerous indirect effects. Bacteraemia, invasive fungal and Epstein–Barr virus-associated post-transplant lymphoproliferative disease are the types of infection that usually related to HCMV. Chronic allograft nephropathy (or tubulointerstitial fibrosis in kidney recipients), bronchiolitis obliterans (lung recipients) and coronary vasculopathy (heart recipients) are the types of acute and chronic allograft injury contributed by HCMV infection (Razonable & Humar, 2013).

This study is carried out to identify the HCMV genotype populations that present in the blood sample of transplant recipients. The different genotype populations of HCMV which are glycoprotein B, N and H in bone marrow and renal transplant recipients were identified by using the real-time polymerase chain reaction technique. In addition, this study also identify whether there are co-infection with multiple strains in HCMV infected organ transplant recipients. The identification of HCMV genotyping is very important in order to indicate various HCMV strains presence in transplant recipients and improve the therapeutic interventions. Although HCMV is the most common opportunistic pathogen seen in transplant recipients, many other viruses have also affected outcomes (Weikert & Blumberg, 2008). Co-infection with other herpesviruses, such as Epstein-Barr virus (EBV) human herpesvirus-6 (HHV-6) and human herpesvirus-7 (HHV-7) has been implicated as risk factors for progression from active HCMV infection to HCMV disease. Thus the presence of these viruses need to be identified in the same study population in order to elucidate whether their presence impose a threat on the transplant recipients.

Finally, we analysed the correlation between HCMV viral load, patient's demographic data, different HCMV genotypes, co-infection with other herpesviruses (HHV-6, 7 and EBV) as well as their contribution in the development of HCMV diseases in both the renal and bone marrow transplant recipients in Malaysia.

1.1 **Problem Statement**

Human cytomegalovirus (HCMV) often associated with a wide range of diseases, particularly in transplant recipients undergoing immunosuppressive therapy. Little is known about the clinical manifestation associated with HCMV viral load, specific viral genotypes and host factors among renal and bone marrow transplant recipients especially in Malaysia where data about HCMV is still lacking

1.2 Significance of Research

Solid organ and bone marrow transplant has increased worldwide since there was an improvement of immunosuppressive agents and graft survival. However, the administration of immunosuppressive drugs to prevent graft rejection cause the patient immune system becomes suppressed, thus resulting in an increased incidence of viral infection of post-organ transplantation. Viral infection such as human cytomegalovirus (HCMV) is associated with particular syndromes and morbidity in the immunocompromised patients.

1.3 Objectives of Research

1.3.1 General Objective

The aim of this study is to identify the correlation between HCMV viral load, HCMV genotypes, EBV, HHV-6 and HHV-7 DNAemia and HCMV disease after renal and bone marrow transplantation.

1.3.2 Specific Objectives

- 1) To determine the percentage of human cytomegalovirus DNA in renal and bone marrow transplant recipients by quantitative real time PCR.
- 2) To determine the genotypic distribution of HCMV envelope glycoprotein B, N and H in renal and bone marrow transplant recipients by using SYBR Green-based real time PCR.
- 3) To identify the presence of human herpesvirus type-6 (HHV-6), human herpesvirus type-7 (HHV-7) and Epstein-Barr virus (EBV) by using SYBR Green-based real time PCR.
- 4) To determine the correlation between HCMV viral loads, the occurrence of various HCMV glycoproteins, co-infection with EBV, HHV-6 and HHV-7 and development of HCMV disease in the same study population by statistical analysis.

REFERENCES

- Bagheri, K., Karimi, M. H., Yaghobi, R., Mohammadi, B., Dehghani, M., & Ebadi, P. (2012). DNA viral infections and transient bone marrow failure in southern Iran. African *J Microbiol Res*, 6(36), 6551-6557.
- Burkhardt, C., Himmelein, S., Britt, W., Winkler, T., & Mach, M. (2004). Glycoprotein N subtypes of human cytomegalovirus induce a strainspecific antibody response during natural infection. J Gen. Virol, 90, 1951-1961
- Buser, C., Walther, P., Mertens, T & Michel, D. (2007). Cytomegalovirus Primary Envelopment Occurs at Large Infoldings of the Inner Nuclear Membrane[down-pointing small open triangle. J Virol.: 81(6): 3042– 3048.
- Chan P. K. S., Ng H.K., Cheng A. F. B. (1999). Detection of human herpesviruses 6 and 7 genomic sequences in brain tumours. *J Clin Pathol*, 52, 620–623.
- Cukuranovic J., Ugrenovic S, Jovanovic I., Visnjic M., and Stefanovic V. (2012). Viral Infection in Renal Transplant Recipients. *The Scientific World Journal*, 2012(2012), 1-18.
- Cobo, F. (2012). Application of molecular diagnostic techniques for viral testing. *The Open Virol J*, 6, 104-114.
- Coen, D.M. & Schaffer, P.A., Nat Re Drug Discov 2003: 2: 278–28.
 Deckers, M., Hofmann, J., Kreuzer, K. A., Reinhard, H., Edubio, A., Hengel, H., Voigt, S., & Ehlers, B. (2009). High genotypic diversity and a novel variant of human cytomegalovirus revealed by combined UL33/UL55 genotyping with broad-range PCR. J Virol, 6 (210).
- Cohen, J. I. (2009), Optimal treatment for chronic active Epstein–Barr virus disease. Pediatric Transplantation, 13: 393–396
- Compton T., Nepomuceno, R.R, & Nowlin, D.M. (1992) Human cytomegalovirus penetrates host cells by pH-independent fusion at the cell surface. Virology. 1992 Nov;191(1):387-95.
- Compston, L. I., Li, C., Sarkodie, F., Owusu-Ofori, S., Opare-Sem, O. and Allain, J.-P. (2009), Prevalence of persistent and latent viruses in untreated patients infected with HIV-1 from Ghana, West Africa. J. Med. Virol., 81: 1860–1868.
- Compton T. (2004). Receptors and immune sensors: the complex entry path of human cytomegalovirus. Trends Cell Biol. 2004 Jan;14(1):5-8.

- Crumpacker, C.S., & Zhang, J. L. (2010) Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 7th ed. Churchill Livingstone. California. Elsevier.
- Dignani, M.C., Mykietiuk, A., Michelet, M., Intile, D., Mammana, L., Desmery, P., Milone, G., & Pavlovsky, S. (2002). Valacyclovir prophylaxis for the prevention of Herpes simplex virus reactivation in recipients of progenitor cells transplantation. Bone Marrow Transplant. 29(3):263-7.
- Dolan, A., Cunningham, C., Hector, R. D., Hassan-Walker, A. F., Lee, L., Addison, C., Dargan, D.J., McGeoch, D.J., Gatherer, D., Emery, V.C., Griffiths, P.D., Sinzger, C., McSharry, B.P., Wilkinson, G.W. and Davison, A.J. (2004). Genetic content of wild-type human cytomegalovirus. *Journal of General Virology*. 85(5), 1301-1312.
- Emery V. C. & Clark D. A. (2007). HHV-6A, 6B and 7: Persistence in the population, epidemiology and transmission. *Department of Virology, Royal Free and University College Medical School of UCL*. London, UK
- Fan, J., Zhang, X., Chen, X. M., Gao, H. N., Yang, M. F., Zhao, H., Hu, J. H., & Ma, W.H. (2009). Monitoring of human cytomegalovirus glycoprotein B genotypes using real-time quantitative PCR in immunocompromised Chinese patients. *J Virol. Methods*, 160, 74-77.
- Fauquet, C. M., Mayo, M. A., Maniloff, J., Desselberger, U., and Ball, L. A. (Eds.) (2005). Virus Taxonomy: Eighth Report of the International Committee on Taxonomy of Viruses. Burlington, MA, Elsevier Academic Press.
- Fishman J. A & Rubin R. H. (1998). Infections in Organ Transplant Recipients.Program in Transplantation Infectious Disease, Massachusetts General Hospital, Boston.
- Fishman J. A. (2007). Infection in Solid Organ Transplant Recipients. N Engl J Med, 14, 357-2601.
- Gibson, W., Clopper, K. S., Britt, W. J. and Baxter, M. K. (1996). Human cytomegalovirus (HCMV) smallest capsid protein identified as product of short open reading frame located between HCMV UL48 and UL49. *Journal of Virology*. 70(8), 5680-5683.

Griffiths, P.D. and Grundy, J.E. (1987). Biochem. J. 241: 313–324.

Gorzer, I., Kerschner, H., Fritz, M. R., & Stockl, E. P. (2010).Human cytomegalovirus (HCMV) genotype populations in immunocompetent individuals during primary HCMV infection. *J ClinVirol*, 38, 100-103.

- Hahn, G., Revello, M.G., Patrone, M., Percivalle, E., Campanini, G., Sarasini,
 A., Wagner, M., Gallina, A., Milanesi, G., Koszinowski, U., Baldanti,
 F., Gerna, G. (2004). Human cytomegalovirus UL131-128 genes are indispensable for virus growth in endothelial cells and virus transfer to leukocytes. J Virol. 2004 Sep;78(18):10023-33.
- Irene G. S. & Patel R. (2000). New Strategies for Prevention and Therapy of Cytomegalovirus Infection and Disease in Solid Organ Transplant Recipients. *Clin. Microbiol. Rev.* 13(1):83.
- Kalejta, R. F. (2008). Tegument proteins of human cytomegalovirus. *Microbiology and Molecular Biology Reviews*. 72(2), 249-265.
- Kamalkumar B.S., Agarwal S.K., Garg P., Dinda A. & Tiwani S.C. (2009). Acute pancreatitis with CMV papillitis and cholangiopathyin a renal transplant recipient. *Clin Exp Nephrol*, *13*, 389–391.
- Kapila, N., Kishore, A., Sodhi, M., Sharma, A., Kumar, P., Mohanty, A.K., Jerath, T., & Mukesh, M. (2012). Identification of Appropriate Reference Genes for qRT-PCR Analysis of Heat-Stressed Mammary Epithelial Cells in Riverine Buffaloes (*Bubalus bubalis*). ISRN Biotechnol, 2013, 1-9.
- Keil, G.M., Ebeling-Keil, A., & Koszinowski, U.H. (1987). Immediate-early genes of murine cytomegalovirus: location, transcripts, and translation products. J Virol. Feb 1987; 61(2): 526–533.
- Klemola E. (1970). Hypersensitivity reactions to ampicillin in cytomegalovirus mononucleosis. Scand J Infect Dis. 1970;2(1):29-31.
- Kotton C.N. & Fishman J.A. (2005). Viral Infection in the Renal Transplant Recipient. J Am Soc Nephrol, 16, 1758–1774.
- Krance R. A., Helen E Heslop& Brenner M. K. (1999). Transplantation and Virus Infections.Stem Cell Transplantation Program, Shell Centre for Cell and Gene Therapy, Bylor College of Medicine Houston, Texas, USA.
- Kropff, B., Burkhardt, C., Schott, J., Nentwich, J., Fisch, T., Britt, W., & Mach, M. (2012). Glycoprotein N of human cytomegalovirus protects the virus from neutralizing antibodies. PLoS Pathog 8(10): e1002999.
- Kurath, S., Halwachs-Baumann, G., Müller, W. and Resch, B. (2010), Transmission of cytomegalovirus via breast milk to the prematurely born infant: a systematic review. Clinical Microbiology and Infection, 16: 1172–1178.

- Lee, C.-P. and Chen, M.-R. (2010), Escape of herpesviruses from the nucleus. Rev. Med. Virol., 20: 214–230.
- Ljungman, P., Griffiths, P., and Paya, C. (2002). Definitions of Cytomegalovirus Infection and Disease in Transplant Recipients. Clin Infectios Disease, 34: 1094-1097.
- Limaye A.P., Jerome K.R., Kuhr C.S., Ferrenberg J., Davis C.L., Corey L. & Marsh C.L. (2001). Quantitation of BK Virus Load in Serum for the Diagnosis of BK Virus–Associated Nephropathy in Renal Transplant Recipients. *The Journal of Infectious Diseases*, 183, 1669–72.
- Liu, F. and Zhou, Z.H. (2007). Comparative virion structures of human herpesvirus. In: *Human herpesviruses: Biology, Therapy and Immunoprophylaxis.* Eds. Arvin, A.,Campadelli-Fiume, G., Mocarski., Moore, P.S., Roizman, B., Whitley, R. And Yamanishi, K. Cambridge University Press, Cambridge, U.K. pp: 27-43.
- Messerle, M., Buhhler, B., Keil, G. M. & Koszinowski, U. H. (1992). Structural organization, expression, and functional characterization of the murine cytomegalovirus immediate-early gene 3. Journal of Virology 66, 27-36.
- McGavran, M. H. & Smith. M. G., (1965). Ultrastructural, cytochemical and microchemical observations on cytomegalovirus (salivary gland virus) infection of human cells in tissue culture, Exp. Mol. Pathol. 4:1-10.
- Mifflin T.E (2013). Setting Up a PCR Laboratory. Department of Pathology, University of Virginia, Charlottesville, Virginia.
- Mocarski, E.S., and Courcelle, C.T. (2001). Cytomegaloviruses and their replication. In: *Fields Virology*. Eds. Knipe, D,M., Howley, D.E., Lamb, R.A., Martin, M.A., Roizman, B. and Straus, S.E. Lippincott Williams and Wilkins, Philadelphia, USA, pp: 2629–2673.
- Monica F. A. & Michaels M. G (2006). Infections in pediatric solid organ transplant recipients. Organ Transplantation in Pediatric Patients15(3), 153–161
- Nichols, W.G. (2003) Combating infections in hematopoietic stem cell transplant recipients. Expert Rev Anti Infect Ther. 2003 Jun;1(1):57-73.
- Nogueira, E., Ozaki, K.S., Tomiyama, H., Camara, N.O.S., & Granato, C.F.H. (2008). Clinical correlations of human cytomegalovirus strains and viral load in kidney transplant recipients. *J Int. Immunopharmacology*, 9, 26-31.

- Pass, R.F. (2001). Cytomegalovirus. In: *Fields Virology*. Eds. Knipe, D,M., Howley, D.E., Lamb, R.A., Martin, M.A., Roizman, B. and Straus, S.E. Lippincott Williams and Wilkins, Philadelphia, USA, pp: 2575-2705.
- Pellet P. E., Roizman B. (2007). "The family Herpesviridae: a brief introduction," in Field' Virology, 5th Edn, eds Knipe D. M. P. H., Griffin D. E., Lamb R. A., Martin M. A., Roizman B., Straus S. E., editors. New York, NY: Lippincott-Williams and Wilkins. 2479–2499.
- Pignatelli, S., Maurizio, D., Ladini, M. P., & Monte, P. D. (2010).Development of a Multiplex PCR for the simultaneous amplification and genotyping of glycoprotein N among human cytomegalovirus strains. *New Microbiol*, 33, 257-262.
- Pignatelli, S., & Monte, P. D. (2009). Epidemiology of human cytomegalovirus strains through comparison of methodological approaches to explore gN variants. *New Microbiol*, 32, 1-10.
- Pignatelli, S., Monte, P. D., Rossini, G., & Ladini, M. P. (2004). Genetic polymorphisms among human cytomegalovirus (HCMV) wild-type strains. *J Rev. Med. Virol*, 14, 383-410.
- Qian, H. L., Cai, T., & Jin H. M. (2009). Human cytomegalovirus glycoprotein genotypes in the genital tract tissue of tubal pregnancy patients. *J Int Med Res*, 37, 385-391.
- Rafailidis P., Mourtzoukou E. G., Varbobitis I. C. & Falagas M. E (2008). Severe Cytomegalovirus Infection in Apparently Immunocompetent Patients: a systematic review. *Virology Journal* 5(47).
- Ratcliff R. M., Chang G., Kok T. W. & Sloots T. P (2011). Molecular Diagnosis of Medical Viruses. *Molecular Virology* 9, 87–102.
- Rawlinson, W. D., Farrell, H. E.,and Barrell, B. G. (1996). Analysis of the complete DNA sequence of murine cytomegalovirus. *Journal of Virology*. 70(12), 8833-8849.
- Razonable R. R. & Zerr D. M. (2009). HHV-6, HHV-7 and HHV-8 in Solid Organ Transplant Recipients. *American Journal of Transplantation* 9(4), 97–S10.
- Razonable, R. R., Humar, A. and the AST Infectious Diseases Community of Practice (2013), Cytomegalovirus in Solid Organ Transplantation. American Journal of Transplantation, 13: 93–106.
- Reddehase MJ, Podlech J, Grzimek NK (2002) Mouse models of cytomegalovirus latency: overview. J Clin Virol 25: S23-S36.

- Ryckman, B.J., Jarvis, M.A., Drummond, D.D., Nelson, J.A. & Johnson, D.C. (2006). Human cytomegalovirus entry into epithelial and endothelial cells depends on genes UL128 to UL150 and occurs by endocytosis and low-pH fusion. J Virol. 2006 Jan;80(2):710-22.
- Samarbasf-Zadeh A. R., Makvandi M., Kayedani G., Shahbazian H., Poorfarziani V., Sia I. G. & Patel R. (2000). New Strategies for Prevention and Therapy of Cytomegalovirus Infection and Disease in Solid-Organ Transplant Recipients. *Clinical Microbiology Reviews* 13(1), 83-110.
- Schleiss, M. R. (2007). Prospects for development and potential impact of a vaccine against congenital cytomegalovirus (CMV) infection. *The Journal of Pediatrics*. 151(6), 564-570.
- Sia I. G. & Patel R. (2000).New Strategies for Prevention and Therapy of Cytomegalovirus Infection and Disease in Solid-Organ Transplant Recipients.*Clin. Microbiol. Rev, 13*(1), 83.
- Sharma, S., Wisner, T.D., Johnson, D.C., & Heldwein, E.E. (2012). HCMV gB shares structural and functional properties with gB proteins from other herpesviruses. J Virol, 435, 239-249.
- Sinzger, C., Digel, M. & Jahn G. (2008). Cytomegalovirus cell tropism. Curr Top Microbiol Immunol. 2008;325:63-83.
- Snydman D. R. & Emery V. C. (2001). Human Herpesviruses 6 and 7 in Solid Organ Transplant Recipients. *Clinical Infectious Disease* 32(9), 1357-1360
- Snydman, D. R. (2013). Epidemiology of Infections after Solid-Organ Transplanta-tion. Clinical Infectious Diseases 33(Suppl 1), 5–8.
- Souza, Marli Adelina, Passos, Ana Maria, Treitinger, Arício, & Spada, Celso. (2010). Seroprevalence of cytomegalovirus antibodies in blood donors in southern, Brazil. Revista da Sociedade Brasileira de Medicina Tropical, 43(4).
- Stephanie A. S. Staras, Sheila C. Dollard, Kay W. Radford, W. Dana Flanders, Robert F. Pass, and Michael J. Cannon Seroprevalence of Cytomegalovirus Infection in the United States, 1988–1994 Clin Infect Dis. (2006) 43 (9): 1143-1151.
- Strauss, H. J and Strauss E. G. (2008). Viruses and Human Diseases. California, Elsevier.
- Thomasini, R. L., Costa, F., Sampaio, A. M., Bonon, S. H. A., Durante, P., Boin, I. F. S. F., Pereira, F. S. M., and Costa, S. C. B. (2012). Betaherpesviruses in Adult Liver Transplant Recipients. University of Campinas Laboratory of Clinical Pathology.

- Vries J. J. C. (2012). Congenital cytomegalovirus infection: disease burden and screening tools: towards newborn screening. University of Leiden.
- Vries, J. J. C., Wessels, E., Korver, A. M. H., Eijk, A. A. V., Rusman, L. G., Kroes, A. C. M., & Vossena, A. C. T. M. (2012). Rapid genotyping of cytomegalovirus in dried blood spots by multiplex real-time PCR assays targeting the envelope glycoprotein gB and gH genes. J *ClinMicrobiol*, 232(7), 114-129.
- Wald A, Huang M, Carrell C.,Selke S. &Corey L. (2003).Polymerase Chain Reaction for Detection of Herpes Simplex Virus (HSV) DNA on Mucosal Surfaces: Comparison with HSV Isolation in Cell Culture.Journal of Infectious Diseases 51 (188), 1345.
- Weikert B. C & Blumberg E. A. (2008). Viral Infection after Renal Transplantation: Surveillance and Management. Clinical Journal of the American Society of Nephrology 3, 76-86.
- Wood, L.J., Baxter, M.K., Plafker, S.M., & Gibson, W. (1997). Human cytomegalovirus capsid assembly protein precursor (pUL80.5) interacts with itself and with the major capsid protein (pUL86) through two different domains. J Virol. Jan 1997; 71(1): 179–190.
- Wu, X., Wang, Y., Xu, Y., Wu, D., Sun, A., & Zhu, Z. (2009). Cytomegalovirus glycoprotein B genotype in hematopoietic stem cell transplant patients from China. *Biol Blood Marrow Transplant*, 16, 647-652.
- Xia, C. S., & Zhang, Z. (2010). Analysis of human cytomegalovirus glycoprotein N genotypes in Chinese hematopoietic stem cell transplant recipients. J Arch Virol, 156, 17-23.
- Yaghoobi R. & Seyed M.L. (2009). Prevalence of BK virus in renal allograft recipients pre and post transplantation in Iran. *Jundishapur Journal of Microbiology*, 2(2), 47-52.
- Ye J., Colulouris G., Zaretskaya I., Cutcutache I., Rozen S. & Madden T. L (2012).

Primer-BLAST: A tool to design target-specific primers for polymerase chain reaction. US National Library of Medicine, National Institute of Health 13:134.

Young, J.H., Weisdorf, D.J..(2010). Infections in Recipients of Hematopoietic Cell Transplantation. In: Mandell GL, Bennett JE, Dolin R, editors. Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases. 7th ed. Philadelphia, USA: Churchill Livingstone; 3809-3820.

- Zhang S., ZhouY. H.,LiL. & Hu Y. (2010). Monitoring human cytomegalovirus infection with nested PCR: comparison of positive rates in plasma and leukocytes and with quantitative PCR. Virology Journal 7,73.
- Zhao, X. T., Zhou, D. Q., Wu, S., Chen, Y. W., Shao, Y., Zhang, J., Xia, C. S., Wang, K.P., Yang, H., Wan, J., Yu, B., Zhang, Z., & Zhang, W. (2012). Genotyping cytomegalovirus UL97 mutations by high-resolution melting analysis with unlabeled probe. J Arch Virol, 157, 475-481.

