



UNIVERSITI PUTRA MALAYSIA

***MOLECULAR DETECTION AND CHARACTERIZATION OF TICK-BORNE
HEMOPATHOGENS IN STRAY DOGS IN MALAYSIA***

KONTO MOHAMMED

FPV 2016 33



**MOLECULAR DETECTION AND CHARACTERIZATION OF TICK-BORNE
HEMOPATHOGENS IN STRAY DOGS IN MALAYSIA**

By

KONTO MOHAMMED

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the
Degree of Doctor of Philosophy**

October 2016

COPYRIGHT

All material contained within this thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to my beloved mother Late Hajiya Hadiza Mohammed, my father and teachers for their guidance, devotion and relentless patience.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

MOLECULAR DETECTION AND CHARACTERIZATION OF TICK-BORNE HEMOPATHOGENS IN STRAY DOGS IN MALAYSIA

By

KONTO MOHAMMED

October 2016

Chairman : Associate Professor Malaika Watanabe, PhD
Faculty : Veterinary Medicine

Tick-borne hemopathogens like *Babesia*, *Ehrlichia*, *Anaplasma* and *Hepatozoon* species pose significant health threats to dogs worldwide. Southeast Asia is one of the few regions of the world with inadequate records concerning vector-borne diseases of dogs and cats, yet it happens to be among the most heavily populated with stray dogs and cats and a climate that is ideal for the tick vectors. In an attempt to achieve a comprehensive scenario for Malaysia, molecular screening and characterization of five major canine tick-borne hemopathogens (namely: *Anaplasma platys*, *Babesia vogeli*, *Babesia gibsoni*, *Ehrlichia canis* and *Hepatozoon canis*) was carried out.

West (Peninsular) Malaysia was divided into four quadrants; North, East, South and West Peninsular, while the East (Island) Malaysia was divided into North and South. Eight out of the 12 States of Malaysia were selected for this study which comprises: Penang, Kedah, Pahang, Selangor, Kuala Lumpur and Johor from West (Peninsular) Malaysia and Sarawak and Sabah from East (Island) Malaysia. A total of 436 stray dogs from 9 animal shelters were selected for this study comprising seven animal shelters from West Malaysia and two animal shelters from East Malaysia. Age-groups were stratified into young and adult, while the breeds were broadly classified into pedigree and local. Whole blood was collected from each dog and DNA extraction was carried out. All DNA samples were screened via standard PCR using family-specific primers that amplify the 360bp fragment of the 16S rRNA gene for *Anaplasma* and *Ehrlichia* and a genus specific primer that amplify the 350bp fragment of the 18S rRNA gene for *Babesia* and *Hepatozoon*. Amplicons obtained were extracted, purified and confirmed by sequencing. For characterization, DNA from the purified PCR product from each representative samples were cloned into a suitable bacterial cell (*Escherichia coli*). Relations between categorical outcomes were compared using the chi-square test and prevalence rates were calculated at 95% confidence interval. For the characterization study, sequences were aligned and edited using the BioEdit

ClustalW program. For comparing and analyzing the nucleotide sequences, the BLAST program was used. The phylogenetic trees were constructed using the Neighbor-Joining method.

This study confirmed the molecular prevalence of five tick-borne hemopathogens namely: *A. platys*, *B. vogeli*, *B. gibsoni*, *E. canis* and *H. canis* in Malaysia with *A. platys* being the most prevalent hemopathogen (36.5%), followed by *B. gibsoni* (27.6%), *B. vogeli* (22.5%), *E. canis* (18.8%) and *H. canis* (15.8%). Presence of co-infection was also established with a concurrent infection with *A. platys* and *Babesia* being the most prevalent (18.8%); followed by *Babesia* and *E. canis* (10.8%), *A. platys* and *E. canis* (8.4%), *H. canis* and *Babesia* (6.4%), *H. canis* and *E. canis* (4.6%) and *H. canis* and *A. platys* (2.7%). Triple infection combination of *Babesia-Anaplasma-Ehrlichia* predominated (61.3%, 19 dogs). However, local breeds showed a significantly ($p > 0.05$) higher infection rate than the pedigree breed for *H. canis*, *Babesia* and *A. platys* ($p = 0.03$, 0.001 and 0.04 respectively).

Spatial distribution based on individual pathogens per region showed that Northern East Malaysia (Sabah) had the highest prevalence for *H. canis* (37%); *B. vogeli* (35.7%) and *A. platys* (64.9%) predominated in West Peninsular, while *B. gibsoni* (76%) and *E. canis* (46%) predominated in Southern East Malaysia (Sarawak). Distribution of hemopathogens based on geographic locations in Malaysia revealed that Southern East Malaysia (Sarawak) had the highest prevalence for tick-borne hemopathogens in Malaysia.

Infectivity status to hemopathogens among stray dogs revealed that the Eastern Peninsular predominated for infectivity with at least one hemopathogen (35.4%), while the Southern East Malaysia (Sarawak) predominated for dogs infected with 2, 3, 4, and all the 5 pathogens under investigation with prevalence of 44%, 20%, 6% and 2% respectively.

This study reported for the first time the genetic diversity of tick-borne hemopathogens among stray dogs in Malaysia and indicated the presence of both indigenous and foreign genotypes in Malaysia. West Peninsular Malaysia showed the highest number of genotypes, followed by the Southern East Malaysia (Sarawak).

The findings from this study provided a comprehensive data on the etiologies of canine tick-borne hemopathogens, their distribution and genetic diversities in Malaysia. This study can also be used for future epidemiological studies and or intervention programme. However, the zoonotic aspects of this hemopathogens as well as the reservoirs of infection have not been looked into in this study; thus, there is the need to further expand our knowledge on that aspect.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**PENGESANAN MOLEKULAR DAN PENCIRIAN HEMOPATOGEN
BAWAAN SENGKENIT PADA ANJING TERBIAR DI MALAYSIA**

Oleh

KONTO MOHAMMED

Oktober 2016

Pengerusi : Profesor Madya Malaika Watanabe, PhD
Fakulti : Perubatan Veterinar

Beberapa patogen darah bawaan hama kecil contohnya sengkent, seperti spesies *Babesia*, *Ehrlichia*, *Anaplasma* dan *Hepatozoon* telah menimbulkan ancaman kesihatan yang ketara kepada anjing seluruh dunia. Asia Tenggara adalah antara beberapa kawasan di dunia yang tidak mempunyai rekod mencukupi mengenai penyakit bawaan vektor bagi anjing dan kucing. Namun, kawasan ini adalah antara kawasan yang mempunyai taburan anjing dan kucing terbiar yang tinggi dan cuaca yang paling ideal untuk kelansungan hidup vektor seperti sengkent. Dalam usaha untuk mengetahui senario yang lebih menyeluruh di Malaysia, saringan molekular dan penjujukan lima patogen darah bawaan sengkent (seperti: *Anaplasma platys*, *Babesia vogeli*, *Babesia gibsoni*, *Ehrlichia canis* dan *Hepatozoon canis*) telah dijalankan.

Semenanjung Malaysia telah dibahagikan kepada empat kuadran; Utara, Timur, Selatan dan Barat, manakala Malaysia Timur terbahagi kepada Utara dan Selatan. Lapan daripada 12 negeri di Malaysia telah dipilih untuk kajian ini, dimana meliputi: Pulau Pinang, Pahang, Selangor, Kuala Lumpur, dan Johor untuk bahagian Semenanjung Malaysia dan termasuk juga Sabah dan Sarawak daripada Malaysia Timur. Sebanyak 436 anjing terbiar dari 9 tempat perlindungan haiwan di Semenanjung Malaysia dan 2 pusat perlindungan dari Malaysia Timur telah dipilih untuk menjalankan kajian ini. Stratifikasi dikelaskan mengikut kumpulan umur dan baka. Kumpulan umur distratakan kepada muda dan dewasa manakalah kumpulan baka telah dikelaskan secara umum kepada baka tulen dan tempatan. Sampel darah diambil dari setiap anjing dan pengestrakan DNA dilakukan. Semua sampel DNA disaring melalui 'Standard PCR' menggunakan primer dari famili-spesifik yang mengamplifikasi 360 pasangan bes daripada gen 16S rRNA untuk *Anaplasma* dan *Ehrlichia* dan pimer gen-spesifik yang mengamplifikasi 350 pasangan bes daripada gen 18s rRNA untuk *Babesia* dan *Hepatozoon*. Amplikon yang terhasil diestruk, dituliskan dan disahkan oleh penjujukan. Untuk pencirian, DNA daripada produk PCR yang tulen dari setiap sampel wakil telah diklon ke

dalam sel bakteria yang sesuai (*Escherichia coli*). Perhubungan antara hasil mutlak dibandingkan menggunakan ujian dan kelaziman kadar chi-square dikira pada 95% selang keyakinan. Untuk kajian pencirian, urutan jujukan telah disusun dan disunting menggunakan program BioEdit ClustalW. Bagi proses perbandingan dan analisa urutan jujukan, program BLAST telah digunakan. Pokok filogenetik dibangunkan menggunakan kaedah Neighbor-Joining.

Kajian ini mengesahkan prevalen molekular bagi 5 patogen darah bawaan sengkenit iaitu: *A. platys*, *B. vogeli*, *B. gibsoni*, *E. canis* dan *H. canis* di Malaysia dan *A. platys* (36.5%) mencatat prevalen tertinggi, diikuti *B. gibsoni* (27.6%), *B. vogeli* (22.5%), *E. canis* (18.8%) dan *H. canis* (15.8%). Kehadiran ko-infeksi antara *A. platys* dan *Babesia* juga dapat dirungkai dengan jumlah prevalen tinggi sebanyak 18.8%, diikuti ko-infeksi antara *Babesia* dan *E. canis* (10.8%), *A. platys* dan *E. canis* (8.4%), *H. canis* dan *Babesia* sp. (6.4%), *H. canis* dan *E. canis* (4.6%), dan *H. canis* dan *A. platys* (2.7%). Kombinasi 3 infeksi serentak antara *Babesia-Anaplasma-Ehrlichia* mendominasi (61.3%, n=9). Walau bagaimanapun, baka anjing tempatan menunjukkan kadar jangkitan yang ketara ($p > 0.05$) lebih tinggi daripada baka anjing asli bagi *H. canis*, *Babesia* dan *A. platys* ($p = 0.03$, 0.001 dan 0.04) masing-masing.

Distribusi spatial berdasarkan patogen individual mengikut setiap rantau menunjukkan bahawa bahagian Utara Malaysia Timur (Sabah) mempunyai prevalen tertinggi bagi *H. canis* (37%); *B. vogeli* (35.7%) dan *A. platys* (64.9%) mendominasi Barat Semenanjung, manakala *B. gibsoni* (76%) dan *E. canis* (46%) mendominasi Selatan Malaysia Timur (Sarawak). Distribusi patogen darah berdasarkan faktor lokasi geografi di Malaysia menunjukkan bahawa Selatan Malaysia Timur (Sarawak) mencatat prevalen tertinggi untuk patogen darah bawaan sengkenit di Malaysia.

Status jangkitan untuk patogen darah dalam kalangan anjing terbiar mendedahkan bahawa Timur Semenanjung mendominasi sekurang-kurangnya jangkitan satu patogen darah (35.4%), manakala Selatan Malaysia Timur (Sarawak) didominasi oleh anjing yang dijangkiti dengan 2, 3, 4, dan kesemua 5 patogen dengan prevalen sebanyak 44%, 20%, 6% dan 2% masing-masing.

Kajian ini adalah yang pertama melaporkan kepelbagaian genetik patogen darah dalam kalangan anjing terbiar di Malaysia dan menunjukkan kehadiran kedua-dua genotip asli dan asing di Malaysia. Barat Semenanjung Malaysia menunjukkan jumlah tertinggi genotip, diikuti oleh Selatan Malaysia Timur (Sarawak).

Penemuan daripada kajian ini menyumbangkan data yang komprehensif mengenai etiologi patogen darah anjing bawaan sengkenit, distribusinya dan kepelbagaian genetiknya di Malaysia. Kajian ini juga boleh digunakan untuk kajian epidemiologi masa depan dan atau program intervensi. Walau bagaimanapun, aspek-aspek zoonotik patogen darah ini serta reservoir jangkitan masih belum diteliti dalam kajian ini; dengan itu, terdapat keperluan untuk mengembangkan lagi pengetahuan kita mengenai aspek tersebut.

ACKNOWLEDGEMENTS

All praises are due to Almighty Allah (s.w.t) for helping me to successfully pass through this rare and most challenging academic adventure. May the peace and blessing of almighty Allah be upon our beloved prophet Muhammad (SAW), his household, companions and followers (Ameen)! This work will not have been conceptualized without the sacrifice, courage and support of our beloved father Alhaji Mohammed Shettima Kyari who sacrificed his own comfort to give us the best in life. May Allah reward him abundantly and grant him good health and long life to reap the fruit of his labour.

My utmost gratitude and sincere appreciation goes to my supervisory committee members in the persons of Associate Prof. Dr. Malaika Watanabe, Dr. Puteri Azaziah Megat Abd-Rani, Dr. Reuben Sunil Kumar Sharma and Dr. Lau Seng Fong for not only being a source of guidance and support to me; but, more like parents throughout my stay in a strange land far from home. Your patience, commitments, sacrifices, collective efforts, supports and thoroughness were the keys to the success of this work.

I would specially like to acknowledge with thanks the tireless assistance, technical support and guidance rendered to me by Dr. Mahira Watanabe throughout the course of my research. I will also like to express my appreciations to all my friends and colleagues who helped me in one way or the other; to mention but a few, Drs. Salamatu, Gimba , Shola, Asinamai and Nur Shahirah for helping me with various aspects of my research, Drs. Goni, Abatcha, Jalo, Alhassan and M.D. Usman for helping me during sample collection despite their own personal schedules. Drs. Baaji, Kachallah, Bakachallah, Bularafa, Abba, Yannabe, Abdulnasir, Lawan, Ilyasu, Sadiq, Hassan, Abba Nuhu (Bobby), Desmond, and Bande for their words of advices during the course of this research. My appreciations also goes to Drs. Adamu Abdul, Kareem, Khan and Abdinasir, Bizi, Alibe, Abbas, Abiso, Talha, Zaks, Muye, to mention but a few for the good time we have as friends.

My profound gratitude to Dr. Jesse F.F.A., Prof. Ong, B.L. and Dr. Ooi, P.T. for their numerous assistance with equipments, reagents and moral supports. My special thanks to Mr. Jefri and Ms Krish for all their assistance, and the staff and students of Parasitology Laboratory, Faculty of Veterinary Medicine, UPM, Mrs. Maizatul Akmal, Mr. Rashid, Misses Amlizawati, Aida and Donea, Shahid, Ruvi, Vish, Winter, Zarith, Dilaila and Adila for accommodating me as a family.

I would personally like to acknowledge the efforts of the Deputy Dean (Research & Postgraduate), Faculty of Veterinary Medicine Associate Professor Dr. Zunita Zakaria and the entire staff of the Deputy Dean's Office for their kindness and support.

My appreciation goes to those who shared the labour and fatigue of this work with me, more especially my wife, kids and siblings.

Finally, I would like to extend my profound gratitude to the School of Graduate Studies (SGS), Universiti Putra Malaysia for granting me three semesters partial scholarship under its International Graduate Research Scholarship Fund (IGRF) scheme to make my stay and study in Malaysia lively.



I certify that a Thesis Examination Committee has met on 14 October 2016 to conduct the final examination of Konto Mohammed on his thesis entitled "Molecular Detection and Characterization of Tick-Borne Hemopathogens in Stray Dogs in Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Latiffah binti Hassan, PhD
Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Siti Suri binti Arshad, PhD
Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Ngah Zasmy a/l Unyah, PhD
Senior Lecturer
Faculty of Medicine and Health Science
Universiti Putra Malaysia
(Internal Examiner)

Hisashi Inokuma, PhD
Professor
Obihiro University of Agriculture and Veterinary Medicine
Japan
(External Examiner)



NOR AINI AB. SHUKOR, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 22 November 2016

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Malaika Watanabe, PhD

Associate Professor
Faculty of Veterinary Medicine,
Universiti Putra Malaysia,
(Chairman)

Puteri Azaziah Megat Abd Rani, PhD

Senior Lecturer
Faculty of Veterinary Medicine,
Universiti Putra Malaysia,
(Member)

Reuben Sunil Kumar Sharma, PhD

Senior Lecturer
Faculty of Veterinary Medicine,
Universiti Putra Malaysia,
(Member)

Lau Seng Fong, PhD

Senior Lecturer
Faculty of Veterinary Medicine,
Universiti Putra Malaysia,
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature: _____ Date: _____

Name and Matric No: Konto Mohammed / GS36666

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____
Name of
Chairman
of Supervisory
Committee: Associate Professor Dr. Malaika Watanabe

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Puteri Azaziah Megat Abd Rani

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Reuben Sunil Kumar Sharma

Signature: _____
Name of Member
of Supervisory
Committee: Dr. Lau Seng Fong

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xv
LIST OF FIGURES	xvii
LIST OF APPENDICES	xx
LIST OF SYMBOLS AND ABBREVIATIONS	xxi
CHAPTER	
1 INTRODUCTION	1
1.1 Brief Overview	1
1.2 Diseases induced and threat posed by tick-borne hemopathogens	1
1.3 Factors predisposing stray dogs to tick-borne diseases	2
1.4 Prevention and control of tick-borne diseases	2
1.5 Efforts toward tackling companion animal's diseases in Malaysia	3
1.6 Justification of the study	3
1.7 Aim and objectives of the study	4
1.8 Research questions and Hypothesis	5
1.8.1 Research questions	5
1.8.2 Hypothesis	5
2 LITERATURE REVIEW	6
2.1 Dogs As Companion Animals	6
2.2 Canine tick-borne diseases	6
2.3 Tick-borne hemopathogens of dogs	7
2.3.1 <i>Ehrlichia</i>	7
2.3.2 <i>Babesia</i> species	16
2.3.3 <i>Anaplasma</i> species	23
2.3.4 Hepatozoon species	31
3 MOLECULAR DETECTION OF TICK-BORNE HEMOPATHOGENS IN STRAY DOGS IN MALAYSIA	38
3.1 Introduction	38
3.2 Materials And Methods	39
3.2.1 Ethics statement	39
3.2.2 Sampling method	40
3.2.3 Study area	40
3.2.4 Sampling site inclusion criteria	41
3.2.5 Animal inclusion criteria	41
3.2.6 Collection and identification of ticks	42
3.2.7 Collection of blood samples	42

3.2.8	DNA extraction and PCR amplification	42
3.2.9	Sequence and similarity analysis	44
3.2.10	Statistical analysis	44
3.3	Results	44
3.4	Discussion	54
3.4.1	Conclusion	57
4	SPATIAL DISTRIBUTION OF TICK-BORNE HEMOPATHOGENS AMONG STRAY DOGS IN MALAYSIA	58
4.1	Introduction	58
4.2	Materials And Methods	59
4.2.1	Study area demarcation and Sampling distribution method	59
4.2.2	Statistical analysis	60
4.3	Results	61
4.4	Discussion	69
4.4.1	Conclusion	71
5	MOLECULAR CHARACTERIZATION AND PHYLOGENETIC ANALYSIS OF Hepatozoon canis IN MALAYSIA	72
5.1	Introduction	72
5.1.1	Transmission occurs by ingestion of the whole vector containing the oocyst of the parasite	72
5.2	Materials And Methods	74
5.2.1	Blood sampling, DNA extraction and PCR amplification	74
5.2.2	Gel extraction and purification	75
5.2.3	Sub cloning	76
5.2.4	Transformation	77
5.2.5	Plasmid propagation	78
5.2.6	Plasmid extraction/purification	78
5.2.7	Sequencing and data analysis	79
5.3	Results	81
5.3.1	Phylogenetic analysis of Malaysian H. canis strain	81
5.3.2	Phylogenetic relationship with foreign genotypes on the databases	88
5.4	Discussion	90
5.4.1	Conclusion	91
6	MOLECULAR CHARACTERIZATION AND PHYLOGENETIC ANALYSIS OF Anaplasma platys IN MALAYSIA	92
6.1	Introduction	92
6.2	Materials And Methods	93
6.2.1	Blood sampling, DNA extraction and PCR amplification	93
6.2.2	Sequencing and data analysis	94

6.3	Results	96
6.3.1	Molecular characterization of <i>A. platys</i> strains from Malaysia	96
6.3.2	Phylogenetic diversity of Malaysian <i>A. platys</i> strains from other geographically dispersed strains	98
6.4	Discussion	102
6.4.1	Conclusion	103
7	MOLECULAR CHARACTERIZATION AND PHYLOGENETIC ANALYSIS OF <i>Babesia vogeli</i> AND <i>Babesia gibsoni</i> IN MALAYSIA BASED ON THE INTERNAL TRANSCRIBED SPACER GENE (ITS-2)	104
7.1	Introduction	104
7.2	Materials And Methods	105
7.2.1	Blood sampling, DNA extraction and PCR amplification	105
7.2.2	Gel extraction, purification, sub-cloning and plasmid extraction	106
7.2.3	Sequencing and data analysis	106
7.3	Results	109
7.3.1	Molecular characterization of <i>Babesia</i> strains from Malaysia	109
7.3.2	Phylogenetic relationships of <i>Babesia</i> strains from Malaysia with other geographically dispersed strains obtained from the GenBank	112
7.4	Discussion	116
7.4.1	Conclusion	117
8	MOLECULAR CHARACTERIZATION AND GENETIC DIVERSITY OF <i>Ehrlichia canis</i> IN MALAYSIA BASED ON THE TRP36 GENES	118
8.1	Introduction	118
8.2	Materials And Methods	119
8.2.1	Blood sampling, DNA extraction and PCR amplification	119
8.2.2	Gel extraction, purification and sub-cloning	121
8.2.3	Sequencing and data analysis	121
8.3	Results	123
8.3.1	Molecular characterization of <i>E. canis</i> strains from Malaysia	123
8.3.2	Genetic diversity of Malaysian <i>E. canis</i> TRP36 gene isolates from other geographically dispersed isolates	125
8.4	Discussion	133
8.4.1	Conclusion	136
9	SUMMARY, CONCLUSION, LIMITATIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	137
9.1	Summary	137

9.1.1	Conceptual framework of the thesis revisited	137
9.1.2	Diagnostic implications	138
9.1.3	Molecular detection of tick-borne hemopathogens in stray dogs in Malaysia	139
9.1.4	Demographic distribution of tick-borne hemopathogens among stray dogs in Malaysia	139
9.1.5	Molecular characterization of the various hemopathogens from Malaysian stray dogs and their phylogenetic relationship with other geographically dispersed strains	142
9.2	Conclusion	144
9.3	Limitations Of This Investigation	144
9.4	Recommendations For Future Research	145
	REFERENCES	146
	APPENDICES	178
	BIODATA OF STUDENT	185
	LIST OF PUBLICATIONS	186

LIST OF TABLES

Table		Page
2.1	Prevalence of tick-borne hemopathogens of dogs in Malaysia using light microscopy	37
3.1	Primer sets used for PCR amplification	43
3.2	Thermal cycling parameters set for each primer set	44
3.3	Frequency distribution, independent t-test and One-way ANOVA to compare the mean difference of the number of ticks collected based on sex and age group from dogs during the study.	45
3.4	Multiple Comparisons test between the age groups using Dunnett T3	46
3.5	Frequency distribution table for the various hemopathogens infecting dogs in Malaysia	47
3.6	Frequency distribution table for the infectivity status of stray dogs to the various hemopathogens in Malaysia	51
3.7	Prevalence of tick-borne hemopathogens in Malaysia based on sex, age and breed.	54
4.1	Frequency distribution and prevalence (%) of tick-borne hemopathogens per region in Malaysia.	63
4.2	Frequency and spatial distribution of infectivity (%) status among stray dogs per region	66
4.3	Comparative regional prevalence of tick-borne hemopathogens in Malaysia using West Malaysia as reference group	68
5.1	<i>Hepatozoon canis</i> genotypes obtained from this study and those obtained from GenBank for comparative studies.	80
5.2	Prevalence and demographic distribution of the various <i>Hepatozoon canis</i> genotypes identified from this study	82
5.3	Pairwise distance estimation between the 15 partial 18SrRNA sequences of <i>H. canis</i> from Malaysia	86

6.1	<i>A. platys</i> genotypes obtained from this study and those obtained from GenBank from other countries for comparative studies	95
6.2	Prevalence and demographic distribution of the various <i>A. platys</i> 16S rDNA genotypes identified from this study	96
6.3	Nucleotide polymorphisms within the 16S rDNA fragments of <i>A. platys</i> among Malaysian isolates	98
6.4	Nucleotide sequence differences among 16S rDNA from different strains of <i>A. platys</i>	100
7.1	<i>Babesia</i> sp. ITS-2 genotypes obtained from this study and those obtained from the GenBank from other countries for comparative studies	108
7.2	Prevalence and demographic distribution of the various <i>Babesia</i> sp. ITS-2 rDNA genotypes identified from this study	109
7.3	Pairwise distance estimation between 24 partial ITS-2 sequences of <i>Babesia</i> sp. from Malaysia and 8 other sequences obtained from the GenBank	115
8.1	<i>E. canis</i> TRP36 genotypes obtained from this study and those obtained from GenBank from other countries for comparative studies	122
8.2	Demographic distribution of the various <i>E. canis</i> TRP 36 genotypes and number of tandem repeats identified from this study	123
8.3	Pairwise distance estimation between the 7 TRP 36 sequences of <i>E. canis</i> from Malaysia and with other 12 geographically dispersed strains obtained from the GenBank	132
8.4	Summary of different tandem repeat sequences identified in TRP36 gene from different <i>E. canis</i> strains	133

LIST OF FIGURES

Figure		Page
2.1	Morula of <i>E. canis</i> inside monocyte	9
2.2	Morula of <i>E. ewingii</i> inside neutrophil	9
2.3	Life cycle of <i>Ehrlichia canis</i>	10
2.4	Photomicrograph of <i>Babesia canis</i>	16
2.5	Photomicrograph of <i>Babesia gibsoni</i>	17
2.6	Life cycle of <i>Babesia Sp.</i>	19
2.7	Photomicrograph of <i>A. platys</i> inside platelet	24
2.8	Photomicrograph of <i>A. phagocytophilum</i> in neutrophil	24
2.9	Life cycle of canine Anaplasmosis	25
2.10	<i>Hepatozoon canis</i> gamont inside the cell	32
2.11	<i>Hepatozoon americanum</i> gamont inside the cell	33
2.12	Life cycle of <i>Hepatozoon canis</i>	34
3.1	Map of Malaysia showing locations (in black diamond) where samples were taken	41
3.2	Prevalence of the various tick-borne hemopathogens in stray dogs in Malaysia	47
3.3	<i>A. platys</i> positive band amplified at approximately 720bp fragment of 16S rRNA gene	48
3.4	<i>B. gibsoni</i> positive bands amplified at 92bp fragment of the 18S rRNA gene	48
3.5	<i>B. vogeli</i> positive bands amplified at approximately 455bp fragment of 18S rRNA gene	49
3.6	<i>E. canis</i> positive band amplified at approximately 409bp fragment of 16S rRNA gene	49
3.7	<i>H. canis</i> positive bands at approximately 665bp fragment of the 18S rRNA gene	50

3.8	Prevalence of <i>Babesia</i> sp. infection among stray dogs in Malaysia	50
3.9	Infectivity status based on number of hemopathogens per stray dog in Malaysia	51
3.10	Co-infectivity status for tick-borne hemopathogens among stray dogs in Malaysia	52
3.11	Prevalence and frequency distribution of number of triple infection among dogs in Malaysia	53
4.1	Map of Malaysia showing the six arbitrarily stratified zones representing the major geographical regions of the West (Peninsular) and East Malaysia separated by the South China Sea	60
4.2	Prevalence of tick-borne hemopathogens of stray dogs in the various zones in Malaysia	62
4.3	Spatial distribution of infectivity status among stray dogs per region	65
4.4	Infection statuses for at least one hemopathogen per stray dog per region in Malaysia.	67
5.1	Transmission of <i>H. canis</i> among dogs	73
5.2	<i>Hepatozoon canis</i> gamont in a Giemsa stained thin blood smear (x100)	81
5.3	<i>H. canis</i> positive bands at approximately 665bp fragment	82
5.4	Phylogenetic tree of 15 <i>Hepatozoon canis</i> partial 18S rRNA gene sequences obtained from stray dog blood samples collected in West and East Malaysia	84
5.5	Spatial distribution of the various <i>Hepatozoon canis</i> genotypes in Malaysia	87
5.6	Phylogenetic tree of the 18S rRNA gene sequences of 15 Malaysian <i>Hepatozoon canis</i> isolates identified in this study and 7 other isolates from other regions obtained from GenBank	89
6.1	<i>A. platys</i> positive band amplified at approximately 720bp fragment of the 16S rRNA gene	94

6.2	Phylogenetic tree of 15 <i>Anaplasma platys</i> partial 16S rDNA consensus sequences obtained from stray dog blood samples collected in Malaysia	97
6.3	Phylogenetic tree of the 16S rRNA gene sequences of 15 <i>Anaplasma platys</i> isolates identified in this study and 8 other isolates from other regions obtained from GenBank	101
7.1	Agarose gel showing the amplified 453bp region of <i>Babesia</i> sp. ITS-2 region	106
7.2	Phylogenetic tree of 24 <i>Babesia</i> sp. partial ITS-2 gene consensus sequences obtained from stray dog blood samples collected in Malaysia	111
7.3	Phylogenetic tree of the ITS-2 gene sequences of 24 <i>Babesia</i> sp. isolates identified in this study and 8 other isolates from other regions obtained from GenBank	113
8.1	<i>E. canis</i> 18S rDNA positive bands at approximately 420bp fragments	120
8.2	<i>E. canis</i> gp36 positive bands ranging between 800bp to 1000bp fragments	121
8.3	Alignment of the deduced amino acid sequences of <i>E. canis</i> TRP36 from Malaysia	124
8.4	Alignment of the deduced amino acid sequences of <i>E. canis</i> TRP36 from Malaysia and sequences from other geographical locations	128
8.5	Phylogenetic tree for <i>E. canis</i> TRP36 amino acid sequences from geographically dispersed <i>E. canis</i> strains	131
9.1	shows the demographic distribution for the various tick-borne hemopathogens in Malaysia	141
9.2	Shows the demographic distribution of geno-groups (number of genotypes) for the various tick-borne hemopathogens in Malaysia	143

LIST OF APPENDICES

Appendix		Page
A	Sampling and other relevant photos	178
A1	Blood sampling at SPCA, Sabah	178
A2	Blood sampling at SPCA George Town, Pinang	178
A3	Blood sampling at Bentong sanctuary (SPCA), Pahang	179
A4	Sampling record book	179
A5	LB/ampicillin/IPTG/X-gal plates plated with transformation culture showing white colonies that indicate recombinant plasmids and blue colonies indicating non-recombinant plasmids.	180
A6	Sequence alignments using MEGA 6	180
A7	Multiple sequence alignment of the deduced amino acid sequences of gp36 of 7 <i>E. canis</i> isolates from different geographical locations of Malaysia	181
B	Composition of buffers and solutions	182
B1	ITPG	182
B2	X-Gal (2ml)	182
B3	Luria Bertani (LB) medium (per liter)	182
B4	(Chapter 5, page 13) SOC medium (100ml)	182
B5	2M Mg ²⁺	182
B6	2X Rapid Ligation Buffer, T4 DNA Ligase	182
B7	Ampicillin stock (50mg/ml)	183
B8	(Chapter 5, page 11) CaCl ₂	183
C	Sample size estimation	183

LIST OF SYMBOLS AND ABBREVIATIONS

α	Alpha
μm	Micro meter
μg	Microgram
μl	Microliter
Cm^2	Square centimetre
ml	Millilitre
ANOVA	Analysis of variance
CO_2	Carbon dioxide
dH_2O	Deionized distilled water
DMSO	Dimethyl sulphoxide
DNA	Deoxyribonucleic acid

CHAPTER 1

INTRODUCTION

1.1 Brief Overview

Tick-borne hemopathogens are aetiological agents of a spectrum of emerging and re-emerging vector borne diseases of dogs that include Babesia, Theileria, Ehrlichia, Anaplasma and Hepatozoon species. These pathogens have been reported worldwide and exert impact in the tropics and subtropics (Watanabe et al., 2004; Matjila et al., 2008; Cardoso et al., 2010; Chomel, 2011; Chandrawathani et al., 2014) where they pose a significant health threat to dogs (Irwin and Jefferies, 2004; Mariana et al., 2011).

1.2 Diseases induced and threat posed by tick-borne hemopathogens

A wide variety of tick-borne pathogens have been reported to infect dogs in Malaysia, notable among them are protozoa, bacteria and rickettsiae species that cause diseases such as babesiosis, theileriosis, hepatozoonosis, anaplasmosis, ehrlichiosis and rickettsiosis (Rajamanickam, 1985; Watanabe 2012; Nazari *et al.*, 2013; Mokhtar 2013, Chandrawathani, 2014).

Tick and other vector-borne diseases pose an increasingly big threat to dogs and other animals in Malaysia. The recent increase in pet ownership, a high influx of tourists and other exotic pet varieties into the country necessitates for an urgent control measures due to the health and economic impact associated with these pathogens.

International standards for animal health and hygiene for the quality control of pets and other animals' disease prevention and control are essential to sustain the Malaysian expanding economy. The large populations of stray dogs and the increasing popularity of pet ownership in Southeast Asian countries including Malaysia coupled with the ideal environment for the sustainability of vectors helps in transmission of vector-borne disease to susceptible hosts (Irwin and Jefferies, 2004).

In similitude with other developing countries, stray dogs remain a serious public health issue. Priority for controlling stray animal population includes an effective, sustainable and practical approach, but this has always been a problem due to the limitation in financial support and lack of public awareness. With the overpopulation of the stray animals, animal shelters are normally operating with insufficient funding, staffs, and expertise in managing the shelters.

1.3 Factors predisposing stray dogs to tick-borne diseases

Increased vector population densities due to ecological changes is one of the most important factor that leads to the emergence of new or undiscovered diseases and the resurgence of the old quiescent ones. Additionally, unrestricted movements of infected animals from one location to another have also led to the spread of vector-borne pathogens from an endemic region to non-endemic areas (Wilson, 1995; Gratz, 1999; Irwin and Jefferies, 2004; Watanabe, 2012).

Climate, immune status, availability of vertebrate animals and vector populations' densities, and abundance of suitable reservoir hosts, among other factors, exerts influence on disease transmission and establishment (Irwin and Jefferies, 2004; Salman, 2012). However, availability of suitable tick populations is the most important factors that determine the establishment of infection in a new environment (Emmons, 1988).

The three most important factors that moderate the transmission and infection intensity of tick-borne diseases worldwide are climate change, changes in land use or ecological influence and movements of animals through importation of exotic and wildlife species, migratory birds and movement of domestic animals (Gratz, 1999; Robson and Allen, 2000; Gubler *et al.*, 2001; Cumming and Van-Vuuren, 2006; Ergönül, 2006; Nijhof *et al.*, 2007; Tack *et al.*, 2010; Molin *et al.*, 2011; Salman 2012; Madder and Pascucci, 2012; Low *et al.*, 2014). These factors influence the prevalence and proliferation of both the ticks and the pathogens they transmit by moderating the survival and fecundity rate of the ticks, time of the year and level of tick activity; specifically the blood sucking rate and time taken for a complete lifecycle of the tick-borne pathogens within the ticks (Gratz, 1999; Salman, 2012).

Interactions between these three factors are important for forecasting how the prevalence and distribution of ticks and tick-borne diseases may appear in an area. Risk assessments should focus on looking for combinations of factors that may directly or indirectly affect these three factors. A risk assessment module can be proposed based on these factors and a framework designed for this purpose could be used to screen for the emergence of unexpected disease events (Gale *et al.*, 2009).

1.4 Prevention and control of tick-borne diseases

Good quality and proper prevention and control measures can best be achieved by an accurate disease diagnosis method. Newer disease diagnosis method like polymerase chain reaction (PCR) are more accurate and reliable than other conventional techniques like Serology and microscopy that are less accurate especially when there is a case of low level infection and lack of morphological distinguishing features during microscopy and in a case of

cross-reactivity and latent infection as in the case of serology, making detection and identification to specie and sub-specie levels very difficult.

With the advent of newer molecular diagnostic techniques, all anomalies and difficulties associated with accurate pathogen detection and identification of tick-borne hemopathogens were solved; and many new species of pathogens were discovered and many were re-classified.

1.5 Efforts toward tackling companion animal's diseases in Malaysia

The current joint efforts by the Division of Veterinary Services Malaysia and various non-governmental organisations (NGO's) in providing veterinary services for companion animals have yielded tremendous results by creating awareness among animal owners, preventing cruelty to animals by establishing animal welfare facilities, especially in rural areas, and infrastructure for diagnostic support (such as regional veterinary pathology laboratories) in small animal practices. Limited surveillance for the accumulation of information on canine and feline diseases has been put in place in each region. The pet owners on their part have also started to maintain high standards of hygiene in the cities. In recent years, researchers in the field of Veterinary and Animal Sciences have achieved tremendously in their research findings towards providing other specialist in the field with updated and relevant information.

Modern diagnostic tests and epidemiological surveys suitable for diagnosis of infections have been going on to differentiate between a new and previous infections, and or diagnosis in individuals with high antibody levels living in endemic areas. To overcome these limitations, and those associated with traditional pathogen-detection systems such as microscopy, highly sensitive DNA-based methods have been developed. Unfortunately, this new technology is not cost effective and will take many years before it is readily available in all localities. However, it is now apparent from the studies in South East Asia which have applied these molecular tools to companion animal blood or to their arthropod parasites that there are intriguing, and maybe even surprising, discoveries to be made soon.

1.6 Justification of the study

Southeast Asia is one of the few regions of the world with inadequate records concerning diseases of pet animals, yet they happen to be among the most heavily populated with stray dogs and cats (Irwin and Jefferies, 2004). The combination of the tropical climate, large population of stray dogs and cats, and an increase in pet ownership in the region provides favorable conditions for the survival and maintenance of both the tick vector and the pathogens that they harbor. The stray animals not only serve as reservoirs of infection to other animals but also increase the risk of zoonotic disease transmission. Basic

demographics for pet populations are scarce to non-existent in most countries of the region (Peters and Pasvol, 2002; Batson, 2008).

Despite the large stray dog population and an ever increasing pet population in Malaysia, there is limited published data on the molecular prevalence, to non concerning thier epidemiology and phylogenetics of canine vector-borne diseases. Since early 80s, researchers have been striving to report the prevalence of ticks and tick-borne pathogens of dogs in Malaysia and its neighboring countries (Rajamanickan *et al.*, 1985, Inokuma *et al.*, 2003, Irwin and Jefferies, 2004, Rahman *et al.*, 2010, Mariana *et al.*, 2011, Nazari *et al.*, 2013; Mokhtar *et al.*, 2013, Chandrawathani *et al.*, 2014). However, there is no adequate, specific and detailed information regarding the prevalence, spatial distribution and phylogenetic analyses of tick-borne hemopathogens of dogs in the country. Therefore, the needs to further investigate, using more sensitive techniques to determine the prevalence, spatial distribution and phylogenetics of these pathogens infecting dogs in Malaysia.

1. Early reports suggested that tick-borne transmitted infections associated with protozoa, rickettsia and bacteria are prevalent in Southeast Asia including Malaysia (Rajamanickan *et al.*, 1985, Suksawat *et al.*, 2001a; Irwin and Jefferies, 2004, Rahman *et al.*, 2010, Mariana *et al.*, 2011, Chandrawathani *et al.*, 2014). Therefore, for further studies are essential for us to obtain a clear picture of their diversity and distribution.
2. Currently, there is a paucity of information regarding the molecular prevalence, spatial distribution and phylogenetic of the various tick-borne hemopathogens of dogs in Malaysia.
3. The potential risk posed by the increasing number of stray dogs in the country and the pathogens that they harbor requires investigation.

1.7 Aim and objectives of the study

Aim : The aim of this study was to determine the prevalence and spatial distribution as well as to carryout molecular characterization of the various tick-borne hemopathogens of stray dogs in Malaysia to provide a more accurate picture of the current situation of these etiological agents of disease in the country.

Objectives : The objectives of this study are to:

1. Determine the molecular prevalence of common tick-borne hemopathogens of stray dogs in Malaysia.
2. Determine the spatial distribution of the various hemopathogens in Malaysia.
3. Further characterize the hemopathogens identified during the course of the study by polymerase chain reaction.

1.8 Research questions and Hypothesis

1.8.1 Research questions

1. What are the prevalences of the common tick-borne pathogens of stray dogs in Malaysia?
2. What is their distribution pattern among region, gender, age and breeds?
3. What are the genetic similarities of the Malaysian strains of the hemopathogens identified from this study from other geographically dispersed strains?

1.8.2 Hypothesis

H_1 = Prevalence of tick-borne hemopathogens among stray dogs in Malaysia is high at $\alpha \leq 0.05$ ($H_0 = \mu_{\text{prev}} > \mu_0$).

H_1 = Prevalence of tick-borne hemopathogens among stray dogs in Malaysia is low at $\alpha \leq 0.05$ ($H_A = \mu_{\text{prev}} < \mu_0$).

H_2 = There is equal distribution of canine tick-borne hemopathogens among region, gender, age and breeds in Malaysia at $\alpha \leq 0.05$ ($H_0 = \mu_{\text{distribution (West, male, young, locals)}} = \mu_{\text{distribution (East, female, adult, pedigree)}}$).

H_2 = There is no equal distribution of canine tick-borne hemopathogens among region, gender, age and breeds in Malaysia at $\alpha \leq 0.05$ ($H_A = \mu_{\text{distribution (West, male, young, locals)}} \neq \mu_{\text{distribution (East, female, adult, pedigree)}}$).

H_3 = The Malaysian isolates for the various hemopathogens are genetically similar to other geographically dispersed strains at $\alpha \leq 0.05$ ($H_0 = \mu_{\text{Malaysian strain}} = \mu_{\text{other strains}}$).

H_3 = The Malaysian isolates for the various hemopathogens are not genetically similar to other geographically dispersed strains at $\alpha \leq 0.05$ ($H_0 = \mu_{\text{Malaysian strain}} \neq \mu_{\text{other strains}}$).

REFERENCES

- Aguero-Rosenfeld, M. E. (2002). Diagnosis of human granulocytic ehrlichiosis: state of the art. *Vector-Borne and Zoonotic Diseases*, 2(4), 233-239.
- Aguiar, D. M., Hagiwara, M. K., & Labruna, M. B. (2008). In vitro isolation and molecular characterization of an *Ehrlichia canis* strain from São Paulo, Brazil. *Brazilian Journal of Microbiology*, 39(3), 489-493.
- Aguiar, D.M., Zhang, X., Melo, A.L.T., Pacheco, T.A., Meneses, A.M.C., Zanutto, M.S., Horta, M.C., Santarém, V.A., Camargo, L.M.A., McBride, J.W. & Labruna, M.B. (2013). Genetic diversity of *Ehrlichia canis* in Brazil. *Veterinary Microbiology*, 164(3), 315-321.
- Aguiar, D. M., & Melo, A. L. (2015). Divergence of the TRP36 protein (gp36) in *Ehrlichia canis* strains found in Brazil. *Ticks and Tick-borne Diseases*, 6(2), 103-105.
- Aguirre, E., Tesouro, M. A., Amusatogui, I., Rodríguez-Franco, F., & Sainz, A. (2004). Assessment of feline ehrlichiosis in central Spain using serology and a polymerase chain reaction technique. *Annals of the New York Academy of Sciences*, 1026(1), 103-105.
- Ahantarig, A., Trinachartvanit, W., & Milne, J. R. (2008). Tick-borne pathogens and diseases of animals and humans in Thailand. *Southeast Asian Journal of Tropical Medicine and Public Health*, 39(6), 1015.
- Aktas, M., Bendele, K.G., Altay, K., Dumanli, N., Tsuji, M. & Holman, P.J. (2007). Sequence polymorphism in the ribosomal DNA internal transcribed spacers differs among *Theileria* species. *Veterinary Parasitology* 147, 221–230.
- Aktas, M., Özübek, S., & Ipek, D. N. S. (2013). Molecular investigations of *Hepatozoon* species in dogs and developmental stages of *Rhipicephalus sanguineus*. *Parasitology Research*, 112(6), 2381-2385.
- Al-Adhami, B., Scandrett, W. B., Lobanov, V. A., & Gajadhar, A. A. (2011). Serological cross-reactivity between *Anaplasma marginale* and an *Ehrlichia* species in naturally and experimentally infected cattle. *Journal of Veterinary Diagnostic Investigation*, 23(6), 1181-1188.
- Albanese, B., Smiraglia, C. & Lavagnino, A. (1971). Notizie sulle zecche di sicilia con segnalazione di *hyalomma detritum* e *Amblyomma variegatum*. *Rivivsta di Parassitologia* 32, 273-276

- Allen, K. E., Li, Y., Kaltenboeck, B., Johnson, E. M., Reichard, M. V., Panciera, R. J., & Little, S. E. (2008). Diversity of *Hepatozoon* species in naturally infected dogs in the southern United States. *Veterinary Parasitology*, 154(3), 220-225.
- Allsopp, M.T.E.P., Cavalier-Smith, T., de Waal, D.T. & Allsopp, B.A. (1994). Phylogeny and evolution of the piroplasms. *Parasitology* 108, 147–152
- Amusategui, I., Tesouro, M. A., Kakoma, I., & Sainz, Á. (2008). Serological reactivity to *Ehrlichia canis*, *Anaplasma phagocytophilum*, *Neorickettsia risticii*, *Borrelia burgdorferi* and *Rickettsia conorii* in dogs from northwestern Spain. *Vector-Borne and Zoonotic Diseases*, 8(6), 797-804.
- Ambrosio, R. E., & Potgieter, F. T. (1987). The genome of *Anaplasma*: DNA base composition and DNA/DNA hybridization. *Onderstepoort Journal of Veterinary Research*, 54, 63–65.
- Amyx, H.L., Huxsoll, D.L., Zeiler, D.C. & Hildebrandt, P.K. (1971). Therapeutic and prophylactic value of tetracycline in dogs infected with the agent of tropical canine pancytopenia. *Journal of American Veterinary Medicine Association* 159: 428-1432.
- Anderson, B. E., Dawson, J. E., Jones, D. C., & Wilson, K. H. (1991). *Ehrlichia chaffeensis*, a new species associated with human ehrlichiosis. *Journal of Clinical Microbiology*, 29(12), 2838-2842.
- Anderson, B.E., Greene, C.E., Jones, D.C. & Dawson, J.E. (1992a). *Ehrlichia ewingii* spp. Nov., the etiologic agent of canine granulocytic ehrlichiosis. *International Journal of Systemic Bacteriology* 42, 299-302.
- André, M. R., Adania, C. H., Teixeira, R. H. F., Vargas, G. H., Falcade, M., Sousa, L., ... & Machado, R. Z. (2010). Molecular detection of *Hepatozoon* spp. in Brazilian and exotic wild carnivores. *Veterinary Parasitology*, 173(1), 134-138.
- Ariyawutthiphan O, Shokshai-utsaha K, Sananmuang T, Chungpivat S, Sarikaputi M. & Viseshakul N. (2008). The microscopic and molecular detections of canine ehrlichiosis. *Thailand Journal of Veterinary Medicine*, 38: 29- 36.
- Audy, J.R. (1957). *Malaysian Parasites*. Institute for medical research, Federation of Malaya. Vol. XVI-XXXIV 1957, No. 28
- Baneth, G., Waner, T., Koplak, A., Weinstein, S. & Keysary, A. (1996). Survey of *Ehrlichia canis* antibodies among dogs in Israel. *Veterinary Records* 138: 275-295.

- Baneth, G., & Weigler, B. (1997). Retrospective Case-Control Study of Hepatozoonosis in Dogs in Israel. *Journal of Veterinary Internal Medicine*, 11(6), 365-370.
- Baneth, G., Barta, J.R., Shkap, V., Martin, D.S., Macintire, D.K. & Vincent-Johnson, N. (2000). Genetic and antigenic evidence supports the separation of *Hepatozoon canis* and *Hepatozoon americanum* at the species level. *Journal of Clinical Microbiology*; 38: 1298-301.
- Baneth, G. (2001). Transmission of *Hepatozoon canis* to dogs by naturally-fed or percutaneously-injected *Rhipicephalus sanguineus* ticks. *Journal of Parasitology*, 87: 606-611.
- Baneth, G. A. D., Samish, M., Alekseev, E., Aroch, I., & Shkap, V. (2001). Transmission of *Hepatozoon canis* to dogs by naturally-fed or percutaneously-injected *Rhipicephalus sanguineus* ticks. *Journal of Parasitology*, 87(3), 606-611.
- Baneth, G., Samish, M. & Shkap, V. (2007). Life cycle of *Hepatozoon canis* Apicomplexa: Adeleorina: *Hepatozoidae*) in the tick *Rhipicephalus sanguineus* and domestic dog (*Canis familiaris*). *Journal of Parasitology* 93, 283–299.
- Baneth, G. (2011). Perspectives on canine and feline hepatozoonosis. *Veterinary parasitology*, 181(1), 3-11.
- Baker, D. C., Simpson, M., Gaunt, S. D., & Corstvet, R. E. (1987). Acute *Ehrlichia platys* infection in the dog. *Veterinary Pathology Online*, 24(5), 449-453.
- Barbour, A. G., & Fish, D. (1993). The biological and social phenomenon of Lyme disease. *Science*, 260(5114), 1610-1616.
- Barker, S.C. (1998). Distinguishing species and populations of rhipicephaline ticks with its 2 ribosomal RNA. *Journal of Parasitology* 84, 887–892.
- Barker, S.C. (1998). Distinguishing species and populations of rhipicephaline ticks with its 2 ribosomal RNA. *Journal of Parasitology* 84, 887–892.
- Barker, S. B., Rogers, C. S., Turner, J. W., Karpf, A. S., & Suthers-McCabe, H. M. (2003). Benefits of interacting with companion animals a bibliography of articles published in refereed journals during the past 5 years. *American Behavioral Scientist*, 47(1), 94-99.
- Batson, A. (2008). Global Companion Animal Ownership and Trade: Project Summary, June 2008. World Society for the Protection of Animals, Companion & Working Animals unit. www.wspa-international.org.

- Beck, R., Vojta, L., Mrljak, V, Marinculic, A., Beck, A., Zivicnjak, T., & Caccio, S. M. (2009): Diversity of *Babesia* and *Theileria* species in symptomatic and asymptomatic dogs in Croatia. *International Journal for Parasitology* 39: 843-848.
- Begum, R. A., & Pereira, J. J. (2009). Potential for tackling climate change in the building sector: The case of Malaysia. In *Proceeding of International Conference on Energy and the Environment: Reinvention for Developing Countries, June* (pp. 15-20).
- Begum, R. A., Pereira, J. J., Jaafar, A. H., & Al-amin, A. Q. (2009). Ecological footprint calculations for Malaysia: An empirical assessment. *Resource Conservation Recycling*, 53, 582-587.
- Begum, R. A., Siwar, C., Abidin, R. D. Z. R. Z., & Pereira, J. J. (2011). Vulnerability of climate change and hardcore poverty in Malaysia. *Journal of Environmental Science and Technology*, 4(2), 112-117.
- Beaufils, J., Inokuma, H., Martin-Granel, J., Jumelle, P., Barbault-Jumell, M. & Brouqui, P. (2002). *Anaplasma platys* (*Ehrlichia platys*) infection in a dog in France: description of the case and characterization of the agent. *Revue de Médecine Vétérinaire* 153: 85-90
- Beutler, B., Jiang, Z., Georgel, P., Crozat, K., Croker, B., Rutschmann, S., Du, X. & Hoebe, K. (2006). Genetic analysis of host resistance: Toll-like receptor signaling and immunity at large. *Annual Review on Immunology*, 24, pp.353-389.
- Biodiversity Theme Report (2001). Australian Government Department of the environmental water heritage and the arts 2001. Retrieved 24 January 2009.
- Birkenheuer, A. J., Neel, J., Ruslander, D., Levy, M. G., & Breitschwerdt, E. B. (2004). Detection and molecular characterization of a novel large *Babesia* species in a dog. *Veterinary Parasitology*, 124(3), 151-160.
- Blaschitz, M., Narodoslavsky-Gfoller, M., Kanzler, M., Stanek, G., Walochnik, J. (2008). *Babesia* species occurring in Austrian *Ixodes ricinus* ticks. *Applied Environmental Microbiology*, 74:4841– 4846.
- Brandao, L. P., Hasegawa, M. Y., Hagiwara, M. K., & Kohayagawa, A. (2006). Platelet aggregation studies in acute experimental canine ehrlichiosis. *Veterinary Clinical Pathology*, 35(1), 78-81.
- Bradford, J.F., Vore, S.J., Pryor, W.H. (1996). *Ehrlichia platys* infection in dogs. *Laboratory Animal Science* 46, 565–568.

- Breitschwerdt, E.B. (1995). Canine monocytic ehrlichiosis. In Ettinger, S.J., Feldman, E.C. (Eds.) *Textbook of Veterinary Internal Medicine* WB Saunders Co. Philadelphia, pp. 378-380.
- Breitschwerdt, E.B., Hegarty, B.C. & Hancock, S.I. (1998). Sequential evaluation of dogs naturally infected with *Ehrlichia canis*, *Ehrlichia chaffeensis*, *Ehrlichia equi*, *Ehrlichia ewingii*, or *Bartonella vinsonii*. *Journal of Clinical Microbiology* 36, 2645–2651.
- Breitschwerdt, E.B. (2000). The rickettsioses. In Ettinger, S.J. & Feldman, E.C. (eds). *Textbook of Veterinary Internal Medicine* pp. 400-408. WB Saunders, Philadelphia.
- Breitschwerdt, E.B., Abrams-Ogg, A.C., Lappin, M.R., Bienzle, D., Hancock, S.I., Cowan, S.M., Clouten, J.K., Hegarty, B.C. and Hawkins, E.C. (2002). Molecular Evidence Supporting *Ehrlichia canis*-Like Infection in Cats. *Journal of Veterinary Internal Medicine*, 16(6), pp.642-649.
- Breitschwerdt, E. (2010). Experimental infection and co-infection of dogs with *Anaplasma platys* and *Ehrlichia canis*: hematologic, serologic and molecular findings. *Parasite and Vectors*. 3:33.
- Brown, G. K., Martin, A. R., Roberts, T. K., & Aitken, R. J. (2001). Detection of *Ehrlichia platys* in dogs in Australia. *Australian Veterinary Journal*, 79(8), 554-558.
- Brown, H.E., Harrington, L.C., Kaufman, P.E., McKay, T., Bowman, D.D., Nelson, C.T., Wang, D. & Lund, R. (2014). Key factors influencing canine heartworm, *Dirofilaria immitis*, in the United States. *Parasites and Vectors* 2012, 5:245.
- Brouqui, P. (2002). *Anaplasma platys* (*Ehrlichia platys*) infection in a dog in France: description of the case, and characterization of the agent. *Revue Méd Vét*, 153(2), 85-90.
- Burridge, M. J. (2001). Ticks (Acari: Ixodidae) spread by the international trade in reptiles and their potential roles in dissemination of diseases. *Bulletin of Entomological Research*, 91(01), 3-23.
- Burridge, M. J., Simmons, L. A., Peter, T. F., & Mahan, S. M. (2002). Increasing risks of introduction of heartwater onto the American mainland associated with animal movements. *Annals of the New York Academy of Sciences*, 969(1), 269-274.
- Cardenas, A.M., Doyle, C.K., Zhang, X., Nethery, K., Corstvet, R.E., Walker, D.H. & McBride, J.W., (2007). Enzyme-linked immunosorbent assay with conserved immunoreactive glycoproteins gp36 and gp19 has enhanced sensitivity and provides species-specific immunodiagnosis of *Ehrlichia canis* infection. *Clinical and Vaccine Immunology* 14, 123–128.

- Cardoso, L., Tuna, J., Vieira, L., Yisaschar-Mekuzas, Y., Baneth, G. (2010). Molecular detection of *Anaplasma platys* and *Ehrlichia canis* in dogs from the North of Portugal. *The Veterinary Journal*, 183(2), 232-233.
- Carreno, R. A., & Barta, J. R. (1999). An eimeriid origin of isosporoid coccidia with Stieda bodies as shown by phylogenetic analysis of small subunit ribosomal RNA gene sequences. *The Journal of Parasitology*, 77-83.
- Carrade, D. D., Foley, J. E., Borjesson, D. L., & Sykes, J. E. (2009). Canine granulocytic anaplasmosis: a review. *Journal of Veterinary Internal Medicine*, 23(6), 1129-1141.
- Chae, J. S., Yu, D. H., Shringi, S., Klein, T. A., Kim, H. C., Chong, S. T., ... & Foley, J. (2008). Microbial pathogens in ticks, rodents and a shrew in northern Gyeonggi-do near the DMZ, Korea. *Journal of Veterinary Science*, 9(3), 285-293.
- Chandrawathani, P., Tan, T.K., Adnan, M., Erwanas, A.I., Premaalatha, B., Low, K.N. & Ramlan, M. (2014). Significant blood protozoan infections, their host range and trend of infections in domestic animals of Malaysia diagnosed by the department of veterinary services and veterinary research institute (VRI) from 1931 to 2010—a historical preview. *Malaysian Journal of Veterinary Research*, 5(2), 47-61.
- Chang, W. L., & Pan, M. J. (1996). Specific amplification of Ehrlichia platys DNA from blood specimens by two-step PCR. *Journal of Clinical Microbiology*, 34(12), 3142-3146.
- Chang, W. L., Su, W. L., & Pan, M. J. (1997). Two-step PCR in the evaluation of antibiotic treatment for *Ehrlichia platys* infection. *Journal of Veterinary Medical Science*, 59(9), 849-851.
- Chomel, B. (2011). Tick-borne infections in dogs—an emerging infectious threat. *Veterinary Parasitology*, 179(4), 294-301.
- Chua, K. B., Goh, K. J., Wong, K. T., Kamarulzaman, A., Tan, P. S. K., Ksiazek, T. G., ... & Tan, C. T. (1999). Fatal encephalitis due to Nipah virus among pig-farmers in Malaysia. *The Lancet*, 354(9186), 1257-1259.
- CIESIN (Center for International Earth Science Information Network) 2007. Changes in the incidence of vector-borne diseases attributable to climate change, <http://www.ciesin.columbia.edu/TG/HH/veclev2.html> (accessed October 18, 2007).
- Coetzer, J.A.W. & Tustin, R.C. (2004). Infectious diseases of livestock, 2nd edn. Oxford University Press Southern Africa, Cape Town, South Africa.

- Collins, N.E. & Allsopp, B.A. (1999). *Theileria parva* ribosomal internal transcribed spacer sequences exhibit extensive polymorphism and mosaic evolution: application to the characterization of parasites from cattle and buffalo. *Parasitology*, 118, 541–551.
- Cook, C. A., & Smit, N. J. (2014). *Hepatozoon* species (Adeleorina: Hepatozoidae) of African bufonids, with morphological description and molecular diagnosis of *Hepatozoon ixoxo* sp. nov. parasitising three *Amietophrynus* species (Anura: Bufonidae). *Parasites and Vectors*, 7(1), 1-12.
- Cortese, L., Terrazzano, G., Piantedosi, D., Sica, M., Prisco, M., Ruggiero, G., & Ciaramella, P. (2011). Prevalence of anti-platelet antibodies in dogs naturally co-infected by *Leishmania infantum* and *Ehrlichia canis*. *The Veterinary Journal*, 188(1), 118-121.
- Craig, T. M., Smallwood, J. E., Knauer, K. W., & McGrath, J. P. (1978). *Hepatozoon canis* infection in dogs: clinical, radiographic, and hematologic findings. *Journal of the American Veterinary Medical Association*, 173(8), 967-972.
- Criado-Fornelio, A., Martinez-Marcos, A., Buling-Sarana, A., & Barba-Carretero, J. C. (2003). Molecular studies on Babesia, Theileria and Hepatozoon in southern Europe: Part II. Phylogenetic Analysis and Evolutionary History. *Veterinary Parasitology*, 114(3), 173-194.
- Criado-Fornelio, A., Buling, A., Cunha-Filho, N.A., Ruas, J.L., Farias, N.A.R., Rey-Valeiron, C., Pingret, J.L., Etievant, M. & Barba-Carretero, J.C. (2007a). Development and evaluation of a quantitative PCR assay for detection of *Hepatozoon* sp. *Veterinary Parasitology*, 150(4), pp.352-356.
- Criado-Fornelio, A., Rey-Valeiron, C., Buling, A., BarbaCarretero, J.C., Jefferies, R. & Irwin, P. (2007b). New advances in molecular epizootiology of canine hematic protozoa from Venezuela, Thailand and Spain. *Veterinary Parasitology*, 144: 261-9.
- Cumming, G. S., & Van Vuuren, D. P. (2006). Will climate change affect ectoparasite species ranges?. *Global Ecology and Biogeography*, 15(5), 486-497.
- Dantas-Torres, F. (2010). Biology and ecology of the brown dog tick, *Rhipicephalus sanguineus*. *Parasites and Vectors*, 3(2), 26-37.
- Dantas-Torres, F., Figueredo, L. A., & Otranto, D. (2011). Seasonal variation in the effect of climate on the biology of *Rhipicephalus sanguineus* in southern Europe. *Parasitology*, 138(04), 527-536.

- Dantas-Torres, F., Chomel, B. B., & Otranto, D. (2012). Ticks and tick-borne diseases: a One Health perspective. *Trends in Parasitology*, 28(10), 437-446.
- Dantas-Torres, F., Latrofa, M. S., Annoscia, G., Giannelli, A., Parisi, A., & Otranto, D. (2013). Morphological and genetic diversity of *Rhipicephalus sanguineus* sensu lato from the New and Old Worlds. *Parasites and Vectors*, 6, 213.
- Dawson, J. E., Biggie, K. L., Warner, C. K., Cookson, K., Jenkins, S., Levine, J. F., & Olson, J. G. (1996). Polymerase chain reaction evidence of *Ehrlichia chaffeensis*, an etiologic agent of human ehrlichiosis, in dogs from southeast Virginia. *American Journal of Veterinary Research*, 57(8), 1175-1179.
- de Castro, M. B., Machado, R. Z., de Aquino, L. P. C. T., Alessi, A. C., & Costa, M. T. (2004). Experimental acute canine monocytic ehrlichiosis: clinicopathological and immunopathological findings. *Veterinary Parasitology*, 119 (1), 73-86.
- De la Fuente, J., Torina, A., Naranjo, V., Nicosia, S., Alongi, A., La Mantia, F., & Kocan, K. M. (2006). Molecular characterization of *Anaplasma platys* strains from dogs in Sicily, Italy. *BMC Veterinary Research*, 2(1), 24.
- Donatien, A., & Lestoquard, F. (1935). Existence en Algérie d'une Rickettsia du chien. *Bull. Soc. Pathol. Exot*, 28, 418-419.
- Donatien, A., Lestoquard, F. (1937). State of the present knowledge concerning rickettsiosis of animals. Archives de l'Institut Pasteur d'Algérie Institut Pasteur d'Algérie 15, 142-187.
- Doyle, C. K., Cardenas, A. M., Aguiar, D. M., Labruna, M. B., Ndip, L. M., Yu, X., & McBride, J. W. (2005). Molecular characterization of *E. canis* gp36 and *E. chaffeensis* gp47 tandem repeats among isolates from different geographic locations. *Annals-New York Academy of Sciences*, 1063, 433.
- Doyle, C.K., Popov, V.L. & McBride, J.W. (2006). Differentially expressed and secreted major immunoreactive protein orthologs of *Ehrlichia canis* and *E. chaffeensis* elicit early antibody responses to epitopes on glycosylated tandem repeats. *Infection Immunology* 74. In press.
- Durfy, S. J., & Willard, H. F. (1990). Concerted evolution of primate alpha satellite DNA: evidence for an ancestral sequence shared by gorilla and human X chromosome alpha satellite. *Journal of Molecular Biology*, 216(3), 555-566.

- Dumler, J. S., Asanovich, K. M., Bakken, J. S., Richter, P., Kimsey, R., & Madigan, J. E. (1995). Serologic cross-reactions among *Ehrlichia equi*, *Ehrlichia phagocytophila*, and human granulocytic *Ehrlichia*. *Journal of Clinical Microbiology*, 33(5), 1098-1103.
- Dumler, J.S., Barbet, A.F., Bekker, C.P., Dasch, G.A., Palmer, G.H., Ray, S.C., Rikihisa, Y. & Rurangirwa, F.R. (2001). Reorganization of genera in the family Rickettsiaceae and Anaplasmataceae in the order Rickettsiales: unification of some species of *Ehrlichia* with *Anaplasma*, *Cowdria* with *Ehrlichia* and *Ehrlichia* with *Neorickettsia*, descriptions of six new species combinations and designation of *Ehrlichia equi* and 'HGE agent' as subjective synonyms of *Ehrlichia phagocytophila*. *International Journal of Systematic and Evolutionary Microbiology*, 51(6), pp.2145-2165.
- East, M.L., Wibbelt, G., Lieckfeldt, D., Ludwig, A., Goller, K., Wilhelm, K., Schares, G., Thierer, D. & Hofer, H. (2008). A *Hepatozoon* species genetically distinct from *H. canis* infecting spotted hyenas in the Serengeti ecosystem, Tanzania. *Journal of Wildlife Diseases*, 44, 45–52.
- Eberts, M. D., de Paiva Diniz, P. P. V., Beall, M. J., Stillman, B. A., Chandrashekar, R., & Breitschwerdt, E. B. (2011). Typical and atypical manifestations of *Anaplasma phagocytophilum* infection in dogs. *Journal Information*, 47(6).
- El-Dakhly, K. M., Goto, M., Noishiki, K., El-Nahass, E. S., Hirata, A., Sakai, H., ... & Yanai, T. (2013). Prevalence and diversity of *Hepatozoon canis* in naturally infected dogs in Japanese islands and peninsulas. *Parasitology Research*, 112(9), 3267-3274.
- Ellis, J., Luton, K., Baverstock, P. R., Brindley, P. J., Nimmo, K. A., & Johnson, A. M. (1994). The phylogeny of *Neospora caninum*. *Molecular and Biochemical Parasitology*, 64(2), 303-311.
- Ellis, J. T., Luton, K., Baverstock, P. R., Whitworth, G., Tenter, A. M., & Johnson, A. M. (1995). Phylogenetic relationships between *Toxoplasma* and *Sarcocystis* deduced from a comparison of 18S rDNA sequences. *Parasitology*, 110(05), 521-528.
- Emmons, R.W. (1988). Ecology of Colorado tick fever. *Annual Review on Microbiology*, 42:49–64
- Epstein, P. R., Diaz, H.F., Elias, S., Grabherr, G., Graham, N.E., Martens, W.J.M., Mosley-Thompson, E. & Susskind, J. (1998). Biological and physical signs of climate change: focus on mosquito-borne diseases. *Bulletin of the American Meteorological Society* 79(3):409-417.
- Ergönül, Ö. (2006). Crimean-Congo haemorrhagic fever. *The Lancet infectious Diseases*, 6(4), 203-214.

- Estrada-Peña, A., Ortega, C., Sánchez, N., DeSimone, L., Sudre, B., Suk, J. E., & Semenza, J. C. (2011). Correlation of *Borrelia burgdorferi* sensu lato prevalence in questing *Ixodes ricinus* ticks with specific abiotic traits in the western Palearctic. *Applied and Environmental Microbiology*, 77(11), 3838-3845.
- Estrada-Pena, A., Bouattour, A., Camicas, J. L., & Walker, A. R. (2004). *Ticks of domestic animals in the Mediterranean region* (Vol. 131).
- Ettinger, Stephen J.;Feldman, Edward C. (1995). *Textbook of Veterinary Internal Medicine(4th ed.)*. W.B. Saunders Company. ISBN 0-7216-6795-3.
- Ewing, S.A., Roberson, W.R., Buckner, R.G. & Hayat, C.S. (1971). A new strain of *Ehrlichia canis*. *Journal of American Veterinary Medical Association*, 159:1771-1774.
- Fleck, S.L. & Moody, A.H. (1988). Diagnostic techniques in medical parasitology. 3rd ed. John Wright Butterworth Scientific University Press, Cambridge, 135pp.
- Fleischer, S., Sharkey, M., Mealey, K., Ostrander, E. A., & Martinez, M. (2008). Pharmacogenetic and metabolic differences between dog breeds: their impact on canine medicine and the use of the dog as a preclinical animal model. *The AAPS Journal*, 10(1), 110-119.
- Foggie, A. (1951). Studies on the infectious agent of tick-borne fever in sheep. *The Journal of Pathology and Bacteriology*, 63(1), 1-15.
- Forlano, M. D., Teixeira, K. R. S., Scofield, A., Elisei, C., Yotoko, K. S. C., Fernandes, K. R., ... & Massard, C. L. (2007). Molecular characterization of *Hepatozoon* sp. from Brazilian dogs and its phylogenetic relationship with other *Hepatozoon* spp. *Veterinary Parasitology*, 145(1), 21-30.
- French, T.W. & Harvey, J.W. (1983). Serologic diagnosis of infectious cyclic thrombocytopenia in dogs using an indirect fluorescent antibody test. *American Journal of Veterinary Research*; 44(12):2407-11.
- Gal, A., Harrus, S., Arcoh, I., Lavy, E., Aizenberg, I., Mekuzas-Yisaschar, Y. & Baneth, G. (2007). Coinfection with multiple tick-borne and intestinal parasites in a 6-weekold dog. *Canadian Veterinary Journal* 48, 619–622.
- Gale, P., Drew, T., Phipps, L. P., David, G., & Wooldridge, M. (2009). The effect of climate change on the occurrence and prevalence of livestock diseases in Great Britain: a review. *Journal of Applied Microbiology*, 106(5), 1409-1423.

- Garret, J.J., Kocan, A.A., Reichard, M.V., Panciera, R.J. & Bahr, R.J. (2005). Experimental infection of adult and juvenile coyotes with domestic dog and wild coyote isolates of *Hepatozoon americanum* (Apicomplexa:Adeleorina). *Journal of Wildlife Diseases* 41, 588–592.
- Gaunt, S., Beall, M., Stillman, B., Lorentzen, L., Diniz, P. P. V. P., Chandrashekar, R., & Breitschwerdt, E. B. (2010). Experimental infection and co-infection of dogs with *Anaplasma platys* and *Ehrlichia canis*: hematologic, serologic and molecular findings. *Parasites and Vectors*, 3(1), 33.
- Glaze, M. B., & Gaunt, S. D. (1986). Uveitis associated with *Ehrlichia platys* infection in a dog. *Journal of the American Veterinary Medical Association*, 189(8), 916-917.
- Goff, W., Barbet, A., Stiller, D., Palmer, G., Knowles, D., Kocan, K., Gorham, J. and McGuire, T. (1988). Detection of *Anaplasma marginale*-infected tick vectors by using a cloned DNA probe. *Proceedings of the National Academy of Sciences*, 85(3), pp.919-923.
- Gondim, L.F.P., Kohayagawa, A., Alencar, N.X., Biondo, A.W., Takahira, R.K.E., Franco, S.R.V. (1998). Canine hepatozoonosis in Brazil: description of eight naturally occurring cases. *Veterinary Parasitology* 74, 319–323.
- Gokce, H., Genc, O., Akca, A., Vatansever, Z., Unver, A., & Erdogan, H. (2008). Molecular and serological evidence of *Anaplasma phagocytophilum* infection of farm animals in the Black Sea Region of Turkey. *Acta Veterinaria Hungarica*, 56(3), 281-292.
- Götsch, S., Leschnik, M., Duscher, G., Burgstaller, J. P., Wille-Piazzai, W., & Joachim, A. (2009). Ticks and haemoparasites of dogs from Praia, Cape Verde. *Veterinary Parasitology*, 166(1), 171-174.
- Gouy, M., & Li, W. H. (1989). Molecular phylogeny of the kingdoms Animalia, Plantae, and Fungi. *Molecular Biology and Evolution*, 6(2), 109-122.
- Granick, J. L., Armstrong, P. J., & Bender, J. B. (2009). *Anaplasma phagocytophilum* infection in dogs: 34 cases (2000–2007). *Journal of the American Veterinary Medical Association*, 234(12), 1559-1565.
- Greene, C. E. (2013). *Infectious diseases of the dog and cat*. Elsevier Health Sciences, pp 1354.
- Greig, B., Asanovich, K. M., Armstrong, P. J., & Dumler, J. S. (1996). Geographic, clinical, serologic, and molecular evidence of granulocytic ehrlichiosis, a likely zoonotic disease, in Minnesota and Wisconsin dogs. *Journal of Clinical Microbiology*, 34(1), 44-48.

- Gratz, N. G. (1999). Emerging and resurging vector-borne diseases. *Annual Review of Entomology*, 44(1), 51-75
- Gray, J. S., Dautel, H., Estrada-Peña, A., Kahl, O., & Lindgren, E. (2009). Effects of climate change on ticks and tick-borne diseases in Europe. *Interdisciplinary perspectives on infectious diseases*, 2009. Article ID 593232, doi: 10.1155/2009/593232
- Gray, J., Dantas-Torres, F., Estrada-Peña, A., & Levin, M. (2013). Systematics and ecology of the brown dog tick, *Rhipicephalus sanguineus*. *Ticks and Tick-borne Diseases*, 4(3), 171-180.
- Groves, M.G., Dennis, G.L., Amyx, H.L. & Huxsoll, D.L. (1975). Transmission of *Ehrlichia canis* to dogs by ticks (*Rhipicephalus sanguineus*). *American Journal of Veterinary Research* 36, 937–940.
- Grundemann, D., & Schomig, E. (1996). Protection of DNA during preparative agarose gel electrophoresis against damage induced by ultraviolet light. *BioTechniques*, 21(5), 898-903.
- Gubbels, J.M., de Vos, A.P., van der Weide, M., Viseras, J., Schouls, L.M., de Vries, E. & Jongejan, F. (1999). Simultaneous detection of bovine *Theileria* and *Babesia* species by reverse line blot hybridization. *Journal of Clinical Microbiology* 37 (6), 1782–1789.
- Gubler, D. J. (1998). Resurgent vector-borne diseases as a global health problem. *Emerging Infectious Diseases* 4(3):442-450.
- Gubler, D. J., Reiter, P., Ebi, K. L., Yap, W., Nasci, R., & Patz, J. A. (2001). Climate variability and change in the United States: potential impacts on vector-and rodent-borne diseases. *Environmental Health Perspectives*, 109(Suppl 2), 223.
- Hall, T.A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41, 95-98.
- Harvey, J.W., Simpson, C.F. & Gaskin, J.M. (1978). Cyclic thrombocytopenia induced by a Rickettsia-like agent in dogs. *Journal of Infectious Disease* 137: 182-188.
- Harrus, S., Waner, T., Avidar, Y., Bogin, E., Peh, H. & Bark, H. (1996). Serum protein alterations in canine ehrlichiosis. *Veterinary Parasitology* 66, 241–249.
- Harrus, S., Aroch, I., Lavy, E. & Bark, H. (1997). Clinical manifestations of infectious canine cyclic thrombocytopenia. *Veterinary Record* 141, 247–250.

- Harrus, S., Waner, T. & Bark, H. (1997). Canine monocytic ehrlichiosis update. *Compendium for Continuing Education for the Practicing Veterinarian* 19, 431–444.
- Harrus, S., Aroch, I., Lavy, E. & Bark, H. (1997). Clinical manifestations of infectious canine cyclic thrombocytopenia. *Veterinary Record* 1997;141:247-50.
- Harrus, S., Waner, T., Keysary, A., Aroch, I., Voet, H., & Bark, H. (1998). Investigation of splenic functions in canine monocytic ehrlichiosis. *Veterinary Immunology and Immunopathology*, 62(1), 15-27.
- Harrus, S., Waner, T., Bark, H., Jongejan, F., & Cornelissen, A. W. (1999). Recent advances in determining the pathogenesis of canine monocytic ehrlichiosis. *Journal of Clinical Microbiology*, 37(9), 2745-2749.
- Harrus, S. & Waner, T. (2011). Diagnosis of canine monocytotropic ehrlichiosis (*Ehrlichia canis*): an overview. *Vet J.* 187:292–6.
- Harrus, S., Perlman-Avrahami, A., Mumcuoglu, K. Y., Morick, D., Eyal, O., & Baneth, G. (2011). Molecular detection of *Ehrlichia canis*, *Anaplasma bovis*, *Anaplasma platys*, *Candidatus Midichloria mitochondrii* and *Babesia canis vogeli* in ticks from Israel. *Clinical Microbiology and Infection*, 17(3), 459-463.
- Harvey, J. W., Simpson, C. F., & Gaskin, J. M. (1978). Cyclic thrombocytopenia induced by a Rickettsia-like agent in dogs. *Journal of Infectious Diseases*, 137(2), 182-188.
- Harvey, J.W. (2012). *Anaplasma platys* infection. In: Greene CO, ed. *Infectious diseases of dogs and cats*. 4th ed. St. Louis, MO: Elsevier Saunders; pp 256-258
- Hauschild, S., Shayan, P. & Schein, E. (1995). Characterization and comparison of merozoite antigens of different *Babesia canis* isolates by serological and immunological investigations. *Parasitology Research* 81, 638–642.
- Heeb, H.L., Wilkerson, M.J., Chun, R. & Ganta, R.R. (2003). Large granular lymphocytosis, lymphocyte subset inversion, thrombocytopenia, dysproteinemia, and positive *Ehrlichia* serology in a dog. *Journal of the American Animal Hospital Association* 39, 379–384.
- Hegarty, B. C., Maggi, R. G., Koskinen, P., Beall, M. J., Eberts, M., Chandrashekar, R., & Breitschwerdt, E. B. (2012). Ehrlichia muris infection in a dog from Minnesota. *Journal of Veterinary Internal Medicine*, 26(5), 1217-1220.

- Hengen, P. N. (1997). Protecting vector DNA from UV light. *Trends in Biochemical Sciences*, 22(5), 182-183.
- Hildebrandt, P.K., Conroy, J.D., McKee, A.E., Nyindo, M.B. & Huxsoll, D.L. (1973). Ultrastructure of *Ehrlichia canis*. *Infection and Immunity* 7, 265–271.
- Hillis, D.M. & Dixon, M.T. (1991). Ribosomal DNA: molecular evolution and phylogenetic inference. *Q. Rev. Biol.* 66, 411–453.
- Hillis, D. M., & Huelsenbeck, J. P. (1992). Signal, noise, and reliability in molecular phylogenetic analyses. *Journal of Heredity*, 83(3), 189-195.
- Homer, M.J., Aguilar-Delfin, I., Telford III, S.R., Krause, P.J. & Persing, D.H. (2000). Babesiosis. *Clinical Microbiology Review*; 13: 451-69.
- Hoogstraal, H. (1985): 19: Ticks. In: Gaafer, S. M., Howard, W. E., Marsh, R. E. (Eds.): *Parasites, Pests and Predators*. Elsevier, Amsterdam
- Hoogstraal, H. (1956). Ticks of Sudan (with special reference to Equatorial province and with preliminary reviews of the genera *Boophilus*, *Margaropus* and *Hyalomma*). In: *African Ixodoidea I. Research Report NM 005 050.29.07*, Department of the Navy, Bureau of Medicine and Surgery, Washington, D.C., pp.436-451
- Hoogstraal, H., Kaiser, M. N., Traylor, M. A., Guindy, E., & Gaber, S. (1963). Ticks (Ixodidae) on birds migrating from Europe and Asia to Africa, 1959-61. *Bulletin of the World Health Organization*, 28(2), 235.
- Hoogstraal, H., Lim, B-L. & Anastos, G. (1969): *Haemaphysalis* (Kaiseriana) *bispinosa* Neumann (Ixodoidea, Ixodidae): evidence for consideration as an introduced species in the Malay Peninsula and Borneo. *J. Parasitol.* 55, 1075-1077
- Hoogstraal, H. (1971). Identity, hosts and distribution of *Haemaphysalis* (Rhipistoma) *canestrinii* (Supino) (resurrected), the postulated asian progenitor of the african leachi complex (Ixodoidea: Ixodidae). *J. Parasitol.* 57, 161-172.
- Hoogstraal, H., Lim, B-L., Nadchatram, M. & Anastos, G. (1972). The Gunong Benom Expedition 1967. 8. Ticks (Ixodidae) of Gunong Benom and their altitudinal distribution, hosts and medical relationships. *Bulletin of Brittain Museum (Nat. Hist.)* 23, 167-186
- Hoogstraal, H. & Wassef, H. (1973). The *Haemaphysalis* ticks (Ixodoidea: Ixodidae) of birds. 3. H. (*Ornithophysalis*) subgen. n., definition, species, hosts and distribution in the Oriental, Palearctic, Malagasy and Ethiopian faunal regions. *Journal of Parasitology* 59, 1099-1117

- Hoogstraal, H., & Wassef, H. Y. (1981). *Haemaphysalis* (*Gernhamphysalis*) subgen. nov. (Acarina: Ixodidae): candidate tick vectors of Hematozoa in the Oriental Region. *Special publication-Society of Protozoologists (USA)*, 1, 117-124
- Hoogstraal, K. & Wassef, H. (1984). *Dermacentor* (*Indocentor*) *compactus* (Acari: Ixodoidea: Ixodidae): wild pigs and other hosts and distribution in Malaysia, Indonesia and Borneo. *Journal of Medical Entomology* 21, 174-178.
- Hoogstraal, R & Wassef, H. (1985a). *Dermacentor* (*Indocentor*) *auratus* (Acari: Ixodoidea: Ixodidae): Hosts, distribution and medical importance in tropical Asia. *Journal of Medical Entomology* 22, 170-177. 24.
- Hoogstraal, H., Wassef, H. (1985b). *Dermacentor* (*Indocentor*) *atrosignatus* (Acari: Ixodoidea: Ixodidae): hosts and distribution in the Malay Peninsula, Indonesia, Borneo and southern Philippines. *Journal of Medical Entomology* 22, 644-647
- Hornok, S., Tánczos, B., de Mera, I. G. F., de la Fuente, J., Hofmann-Lehmann, R., & Farkas, R. (2013). High prevalence of Hepatozoon infection among shepherd dogs in a region considered to be free of *Rhipicephalus sanguineus*. *Veterinary Parasitology*, 196(1), 189-193.
- Hoskins, J.D. (1991). Ehrlichial diseases of dogs: Diagnosis and treatment. *Canine Practice* 16: 13-21.
- Hossain, M. A., Yamato, O., Yamasaki, M., Jeong, J. R., Chang, H. S., & Maede, Y. (2003). Serum from dogs infected with *Babesia gibsoni* inhibits maturation of reticulocytes and erythrocyte 5'-nucleotidase activity in vitro. *Journal of Veterinary Medical Science*, 65(12), 1281-1286.
- Hsieh, Y. C., Lee, C. C., Tsang, C. L., & Chung, Y. T. (2010). Detection and characterization of four novel genotypes of *Ehrlichia canis* from dogs. *Veterinary Microbiology*, 146(1), 70-75.
- Hua, P., Yuhai, M., Shide, T., Yang, S., Bohai, W., & Xiangrui, C. (2000). Canine ehrlichiosis caused simultaneously by *Ehrlichia canis* and *Ehrlichia platys*. *Microbiology and Immunology*, 44(9), 737-739.
- Huang, H., Unver, A., Perez, M. J., Orellana, N. G., & Rikihisa, Y. (2005). Prevalence and molecular analysis of *Anaplasma platys* in dogs in Lara, Venezuela. *Brazilian Journal of Microbiology*, 36(3), 211-216.
- Huang, C. C., Hsieh, Y. C., Tsang, C. L., & Chung, Y. T. (2010). Sequence and phylogenetic analysis of the gp200 protein of *Ehrlichia canis* from dogs in Taiwan. *Journal of Veterinary Science*, 11(4), 333-340.

- Hutcheon, D. (1893). Diseases amongst dogs. Malignant jaundice or bilious fever of the dog. *Agric Journal Cape Good Hope*, 6, 476-477.
- Inokuma, H., Ohno, K., Onishi, T., Raoult, D., & Brouqui, P. (2001). Detection of ehrlichial infection by PCR in dogs from Yamaguchi and Okinawa Prefectures, Japan. *Journal of Veterinary Medical Science*, 63(7), 815-817.
- Inokuma, H., Terada, Y., Kamio, T., Raoult, D., & Brouqui, P. (2001b). Analysis of the 16S rRNA gene sequence of *Anaplasma centrale* and its phylogenetic relatedness to other ehrlichiae. *Clinical and diagnostic laboratory immunology*, 8(2), 241-244.
- Inokuma, H., Fujii, K., Okuda, M., Onishi, T., Beaufils, J. P., Raoult, D., & Brouqui, P. (2002). Determination of the nucleotide sequences of heat shock operon groESL and the citrate synthase gene (gltA) of *Anaplasma (Ehrlichia) platys* for phylogenetic and diagnostic studies. *Clinical and Diagnostic Laboratory Immunology*, 9(5), 1132-1136.
- Inokuma, H., Beppu, T., Okuda, M., Shimada, Y., & Sakata, Y. (2003). Epidemiological survey of *Anaplasma platys* and *Ehrlichia canis* using ticks collected from dogs in Japan. *Veterinary Parasitology*, 115(4), 343-348.
- Inokuma, H., Yoshizaki, Y., Matsumoto, K., Okuda, M., Onishi, T., Nakagome, K., Kosugi, R. and Hirakawa, M. (2004). Molecular survey of Babesia infection in dogs in Okinawa, Japan. *Veterinary Parasitology*, 121(3), pp. 341-346.
- Irwin, P.J. & Jeffries, R. (2004). Arthropod-transmitted diseases of companion animals in Southeast Asia. *Trends Parasitology*, 20:27-34.
- Irwin, P.J. (2005). Babesiosis and Cytauxzoonosis. In *Arthropod-borne infectious diseases of the dog and cat*. Edited by Shaw SE, Day MJ. London: Manson Publishing/The Veterinary Press; pp 63-77.
- Irwin, P. J. (2009). Canine babesiosis: from molecular taxonomy to control. *Parasites and Vectors*, 2(Suppl 1), S4.
- Irwin, P. J. (2010). Canine babesiosis. *Veterinary Clinics of North America: Small Animal Practice*, 40(6), 1141-1156.
- Iqbal, Z., Chaichanasiriwithaya, W., Rikihisa, Y. (1994). Comparison of PCR with other tests for early diagnosis of canine ehrlichiosis. *Journal of Clinical Microbiology* 32, 1658–1662.
- Jacobson, L. S. (2006). The South African form of severe and complicated canine babesiosis: clinical advances 1994–2004. *Veterinary Parasitology*, 138(1), 126-139.

- Jaenson, T. G., Tälleklint, L., Lundqvist, L., Olsen, B., Chirico, J., & Mejlom, H. (1994). Geographical distribution, host associations, and vector roles of ticks (Acari: Ixodidae, Argasidae) in Sweden. *Journal of Medical Entomology*, 31 (2), 240-256.
- Jaenson, T. G., Jaenson, D. G., Eisen, L., Petersson, E., & Lindgren, E. (2012). Changes in the geographical distribution and abundance of the tick *Ixodes ricinus* during the past 30 years in Sweden. *Parasites & Vectors*, 5(1), 1-15.
- Jafar, A. H., Al-Amin, A. Q., & Siwar, C. (2008). Environmental impact of alternative fuel mix in electricity generation in Malaysia. *Renewable Energy*, 33(10), 2229-2235.
- Jamaludin, S., Suhaila, S., Wan Zin, W. Z., & Jemain, A. A. (2008). Tracing trends in the sequences of dry and wet days over peninsular Malaysia. *Journal of Environmental Science and Technology*, 1(3), 97-110.
- James, A. (2006). Malaysia, Brunei and Singapore. New Holland Publisher. Pp 46-50. ISBN 1-86011-309-5.
- John, J. K., Kattoor, J. J., Nair, A. R., Bharathan, A. P., Valsala, R., & Sadanandan, G. V. (2014). Kyasanur Forest Disease: a status update. *Advances in Animal and Veterinary Sciences*, 2(6), 329-336.
- Johnson, E. M., Ewing, S. A., Barker, R. W., Fox, J. C., Crow, D. W. & Kocan, K. M. (1998). Experimental transmission of *Ehrlichia canis* (Rickettsiales: Ehrlichieae) by *Dermacentor variabilis* (Acari: Ixodidae). *Veterinary Parasitology*. 74: 277-288.
- Jongejan, F. (2001). Teken en door teken overgedragen ziekten. *Diergeneeskundig Memorandum* 48, 1-51.
- Jongejan, F., & Uilenberg, G. (2004). The global importance of ticks. *Parasitology*, 129(S1), S3-S14.
- Jukes T.H. & Cantor C.R. (1969). Evolution of protein molecules. In Munro HN, editor, *Mammalian Protein Metabolism*, pp. 21-132, Academic Press, New York.
- Kingsbury, A. N. (1925). On the occurrence of *Piroplasma (Babesia canis)* in Malaya. *Parasitology* 27, 190-191.
- Kamani, J., Sannusi, A.A., Dogo, G., Tanko, J. T., Egwu, K. O., Tafarki, A. E., Ogo, I. N., Kemza, S., Onovoh, E., Shamakim, D., Lombin, L. H., Catto, V. and Birkenheuer, A. J. (2010). *Babesia canis* and *Babesia rossi* co-infection in an untraveled Nigerian dog. *Veterinary Parasitology* 173 :334-335.

- Kamani, J., Baneth, G., Mumcuoglu, K. Y., Waziri, N. E., Eyal, O., Guthmann, Y., & Harrus, S. (2013a). Molecular detection and characterization of tick-borne pathogens in dogs and ticks from Nigeria. *PLoS Neglected Tropical Diseases*, 7(3), e2108.
- Kamani, J., Lee, C. C., Haruna, A. M., Chung, P. J., Weka, P. R., & Chung, Y. T. (2013b). First detection and molecular characterization of *Ehrlichia canis* from dogs in Nigeria. *Research in veterinary science*, 94(1), 27-32.
- Kamau, J., Salim, B., Yokoyama, N., Kinyanjui, P. & Sugimoto, C. (2011). Rapid discrimination and quantification of *Theileria orientalis* types using ribosomal DNA internal transcribed spacers. *Infection Genetics Evolution* 11, 407–414.
- Karesh, W.B. (2005). "Wildlife Trade and Global Disease Emergence," *Emerging Infectious Diseases*, vol. 11, no. 7, pp. 1000-1002.
- Karlin, S., & Brocchieri, L. (2000). Heat shock protein 60 sequence comparisons: duplications, lateral transfer, and mitochondrial evolution. *Proceedings of the National Academy of Sciences*, 97(21), 11348-11353.
- Kocan, A.A., Cummings, C.A., Panciera, R.J., Mathew, J.S., Ewingii, S.A. & Barker, R.W. (2000). Naturally occurring and experimentally transmitted Hepatozoon americanum in coyotes from Oklahoma. *Journal of Wildlife Disease* 36, 149–153.
- Koh, F. X., Panchadcharam, C., & Tay, S. T. (2015). Vector-Borne Diseases in Stray Dogs in Peninsular Malaysia and Molecular Detection of *Anaplasma* and *Ehrlichia* spp. from *Rhipicephalus sanguineus* (Acari: Ixodidae) Ticks. *Journal of Medical Entomology*, tjv153.
- Kohn, B., Galke, D., Beelitz, P., & Pfister, K. (2008). Clinical features of canine granulocytic anaplasmosis in 18 naturally infected dogs. *Journal of Veterinary Internal Medicine*, 22(6), 1289-1295.
- Komnenou, A. A., Mylonakis, M. E., Kouti, V., Tendoma, L., Leontides, L., Skountzou, E., ... & Ofri, R. (2007). Ocular manifestations of natural canine monocytic ehrlichiosis (*Ehrlichia canis*): a retrospective study of 90 cases. *Veterinary ophthalmology*, 10(3), 137-142.
- Konto, M. (2013). Ticks and tick-borne hemopathogens of dogs in Maiduguri, Nigeria. Masters Thesis, University of Maiduguri, Nigeria. Pp 201.
- Konto, M., Biu, A. A., Ahmed, M. I., & Charles, S. (2014a). Prevalence and seasonal abundance of ticks on dogs and the role of *Rhipicephalus sanguineus* in transmitting *Babesia* species in Maiduguri, North-Eastern Nigeria. *Veterinary World*, 7(3), 119-124.

- Konto, M., Biu, A. A., Ahmed, M. I., Mbaya, A. W., & Luka, J. (2014b). Clinico-biochemical responses of dogs to experimental infection with *Babesia canis*. *Veterinary World*, 7(3), 113-118.
- Konto, M., Fufa, G. I., Zakaria, A., Tukur, S. M., Watanabe, M., Ola-Fadunsin, S. D., ... & Babjee, S. M. A. (2015). Tick fauna of Malaysian red jungle fowl (*Gallus gallus*) in Bangi, Malaysia. *Veterinary World*, 8(10), 1167-1171.
- Kuttler, K.L. (1988). World-wide impact of Babesiosis. In: Ristic, M. (Ed.), *Babesiosis of Domestic Animals and Man*. CRC Press, Boca Raton, FL, pp. 1–22.
- Lam, S. K., & Chua, K. B. (2002). Nipah virus encephalitis outbreak in Malaysia. *Clinical Infectious Diseases*, 34(Supplement 2), S48-S51.
- Lang-Unnasch, N., Reith, M. E., Munholland, J., & Barta, J. R. (1998). Plastids are widespread and ancient in parasites of the phylum Apicomplexa. *International Journal for Parasitology*, 28(11), 1743-1754.
- Leiva, M., Naranjo, C., & Pena, M. T. (2005). Ocular signs of canine monocytic ehrlichiosis: a retrospective study in dogs from Barcelona, Spain. *Veterinary Ophthalmology*, 8(6), 387-393.
- Lew, A. E., Gale, K. R., Minchin, C. M., Shkap, V., & de Waal, D. T. (2003). Phylogenetic analysis of the erythrocytic *Anaplasma* species based on 16S rDNA and GroEL (HSP60) sequences of *A. marginale*, *A. centrale*, and *A. ovis* and the specific detection of *A. centrale* vaccine strain. *Veterinary Microbiology*, 92(1), 145-160.
- Lewis Jr, G. E., Ristic, M., Smith, R. D., Lincoln, T., & Stephenson, E. H. (1977). The brown dog tick *Rhipicephalus sanguineus* and the dog as experimental hosts of *Ehrlichia canis*. *American Journal of Veterinary Research*, 38(12), 1953-1955.
- Lim, J. T., & Samah, A. A. (2004). *Weather and climate of Malaysia*. University of Malaya Press.
- Lim, C.W. (2007). Prevalence of blood parasites in canine pet and stray populations: prevalence and effect on hematological parameters. DVM Thesis. Faculty of Veterinary Medicine, Universiti Putra Malaysia, Serdang. Unpublished data.
- Little, S.E., Allen, K.E., Johnson, E.M., Panciera, R.J., Reichard, M.V. & Ewing, S.A. (2009). New development in canine hepatozoonosis in North America: a review. *Parasites and Vectors*, 2(1): S5

- Liu, A.H., Yin, H., Guan, G.Q., Schnittger, L., Liu, Z.J., Ma, M.L., Dang, Z.S., Liu, J.L., Ren, Q.Y., Bai, Q., Ahmed, J.S. & Luo, J.X. (2007). At least two genetically distinct large *Babesia* species infective to sheep and goats in China. *Veterinary Parasitology*, 147, 246– 251.
- Lobetti, R. G., & Reyers, F. (1996). Met-haemoglobinuria in naturally occurring *Babesia canis* infection. *Journal of the South African Veterinary Association*, 67(2), 88-90.
- Lobetti, R.G., (1998). Canine babesiosis. *Compendium on Continuing Education for the Practicing Veterinarian (USA)*, 20: 418-431.
- Low, K., Moo, K., Soon, X., Yvonne, A., Ramlan, M., Chandrawathani, P., Tan, T.K. 2, Adnan, M., Erwanas, Ai, Premaalatha, B. (2014). Significant blood protozoan infections, their host range and trend of infections in domestic animals of Malaysia diagnosed by the Department of Veterinary Services and Veterinary Research Institute (VRI) from 1931 to 2010 – a historical preview. *Malaysian Journal of Veterinary Research* Volume 5 No. 2 July 2014 pages 47-61
- Luo, J., Chen, F., Lu, W., Guan, G., Ma, M. & Yin, H. (2003). Experimental transmission of an unnamed bovine *Babesia* by *Hyalomma* spp., *Haemaphysalis longicornis* and *Boophilus microplus*. *Veterinary Parasitology* 116, 115–124.
- Luo, J., Yin, H., Liu, Z., Yang, D., Guan, G., Liu, A., Ma, M., Dang, S., Lu, B., Sun, C., Bai, Q., Lu, W. & Chen, P. (2005a). Molecular phylogenetic studies on an unnamed bovine *Babesia* sp. based on small subunit ribosomal RNA gene sequences. *Veterinary Parasitology* 133, 1–6.
- Luo, J., Yin, H., Guan, G., Yang, D., Liu, A., Ma, M., Liu, Z., Dang, Z., Liu, G., Bai, Q., Lu, W. & Chen, P. (2005b). A comparison of small-subunit ribosomal RNA gene sequences of bovine *Babesia* species transmitted by *Haemaphysalis* spp. in China. *Parasitology Research* 95, 145–149.
- Macintire, D. K., Vincent-Johnson, N. A., Kane, C. W., Lindsay, D. S., Blagburn, B. L., & Dillon, A. R. (2001). Treatment of dogs infected with *Hepatozoon americanum*: 53 cases (1989-1998). *Journal of the American Veterinary Medical Association*, 218(1), 77-82.
- Madder, M., Thys, E., Geysen, D., Baudoux, C., & Horak, I. (2007). *Boophilus microplus* ticks found in West Africa. *Experimental and Applied Acarology*, 43(3), 233-234.
- Madder, M., Thys, E., Achi, L., Touré, A., & De Deken, R. (2011). *Rhipicephalus (Boophilus) microplus*: a most successful invasive tick species in West-Africa. *Experimental and Applied Acarology*, 53(2), 139-145.

- Madder, M & Pascucci, I. (2012). *Ticks and tick-borne diseases: geographical distribution and control strategies in the Euro-Asia region*. CABI. Chapter 3: 27-32.
- Makinde, M. O., & Bobade, P. A. (1994). Osmotic fragility of erythrocytes in clinically normal dogs and dogs infected with parasites. *Research in Veterinary Science*, 57(3), 343-348.
- Malaysia & Malaysia. Unit Perancang Ekonomi (2006). *Ninth Malaysia Plan, 2006-2010*. Economic Planning Unit, Prime Minister's Department.
- Marchette, N. J. (1966). Rickettsioses (Tick typhus, Q-fever, Urban typhus) in Malaya. *Journal of Medical Entomology*, 2, 339-371
- Mariana, A., Kulaini, B.M., Halimatun, I., Suhaili, Z.A., Shahrul-Anuar, M.S., Zalipah, N.N. & Ho, T.M. (2011) Acarine ectoparasites of Panti Forest Reserve in Johore Malaysia. *Asian Pacific Journal of Tropical Biomedicine*, 1(1): 1-5.
- Matijatko, V., Kiš, I., Torti, M., Brkljačić, M., Rafaj, R. B., Žvorc, Z., & Mrljak, V. (2010). Systemic inflammatory response syndrome and multiple organ dysfunction syndrome in canine babesiosis. *Veterinarski Arhiv*, 80(5), 611-626.
- Matijatko, V., Torti, M., & Schetters, T. P. (2012). Canine babesiosis in Europe: how many diseases?. *Trends in Parasitology*, 28(3), 99-105.
- Martin, A. R., Brown, G. K., Dunstan, R. H., & Roberts, T. K. (2005). *Anaplasma platys*: an improved PCR for its detection in dogs. *Experimental Parasitology*, 109(3), 176-180.
- Marston, E. L., Sumner, J. W., & Regnery, R. L. (1999). Evaluation of intraspecies genetic variation within the 60 kDa heat-shock protein gene (groEL) of Bartonella species. *International Journal of Systematic and Evolutionary Microbiology*, 49(3), 1015-1023.
- Masala, G., Chisu, V., Foxi, C., Socolovschi, C., Raoult, D., & Parola, P. (2012). First detection of *Ehrlichia canis* in *Rhipicephalus bursa* ticks in Sardinia, Italy. *Ticks and Tick-borne Diseases*, 3(5), 396-397.
- Mathew, J. S., Ewing, S. A., Murphy, G. L., Kocan, K. M., Corstvet, R. E., & Fox, J. C. (1997). Characterization of a new isolate of *Ehrlichia platys* (Order Rickettsiales) using electron microscopy and polymerase chain reaction. *Veterinary Parasitology*, 68(1), 1-10.
- Mathew, J. S., Van Den Bussche, R. A., Ewing, S. A., Malayer, J. R., Latha, B. R., & Panciera, R. J. (2000). Phylogenetic relationships of Hepatozoon (Apicomplexa: Adeleorina) based on molecular, morphologic, and life-cycle characters. *Journal of Parasitology*, 86(2), 366-372.

- Mathew, J.S., Saliki, J.T., Ewing, S.A., Lehenbauer, T.W., Panciera, R.J., Malayer, J.R., Cummings, C.A. & Kocan, A.A. (2001). An indirect enzyme-linked immunosorbent assay for diagnosis of American canine hepatozoonosis. *Journal of Veterinary Diagnostic Investigation*, 13(1), pp.17-21.
- Matjila, P.T., Leisewitz, A.L., Jongejan, F. & Penzhorn, B.L. (2008). Molecular detection of tick-borne protozoal and ehrlichial infections in domestic dogs in South Africa. *Veterinary Parasitology* 2008, 155(1), 152-157.
- Matjila, P T., Carcy, B., Leisewitz, A. L., Schetters, T., Jongejan, E., Gorenflot, A., & Penzhon, B. (2009). Preliminary evaluation of the BrEMA1 gene as a tool for associating *Babesia rossi* genotypes and clinical manifestation of Canine babesiosis. *Journal of Clinical Microbiology*. 3586-3592.
- Mavromatis, K., Doyle, C. K., Lykidis, A., Ivanova, N., Francino, M. P., Chain, P., ... & Kyripides, N. C. (2006). The genome of the obligately intracellular bacterium *Ehrlichia canis* reveals themes of complex membrane structure and immune evasion strategies. *Journal of Bacteriology*, 188(11), 4015-4023.
- McBride, J.W., Corstvet, R.E., Gaunt, S.D., Boudreaux, C., Guedry, T. & Walker, D.H. (2003). Kinetics of antibody response to *Ehrlichia canis* immunoreactive proteins. *Infection and Immunity* 71, 2516–2524.
- McBride, J. W., & Walker, D. H. (2011). Molecular and cellular pathobiology of *Ehrlichia* infection: targets for new therapeutics and immunomodulation strategies. *Expert Review on Molecular Medicine*, 13(3).
- McCall, J.W., Baker, C.F., Mather, T.N., Chester, S.T., McCall, S.D., Irwin, J.P., Young, S.L., Cramer, L.G. & Pollmeier, M.G. (2011). The ability of a topical novel combination of fipronil, amitraz and (S)-methoprene to protect dogs from *Borrelia burgdorferi* and *Anaplasma phagocytophilum* infections transmitted by *Ixodes scapularis*. *Veterinary Parasitology*, 179(4), pp.335-342.
- McGuire, T. C., Palmer, G. H., Goff, W. L., Johnson, M. I., & Davis, W. C. (1984). Common and isolate-restricted antigens of *Anaplasma marginale* detected with monoclonal antibodies. *Infection and Immunity*, 45(3), 697-700.
- Minjauw, B. & Mcleod, A. (2003). Tick-Borne Diseases and Poverty. The Impact Of Ticks And Tick-Borne Diseases On The Livelihoods Of Small-Scale And Marginal Livestock Owners In India And Eastern And Southern Africa. Research Report, Dfid Animal Health Programme, Centre For Tropical Veterinary Medicine, University Of Edinburgh, Uk.
- Mohan, R. N. (1968): Diseases and parasites of buffaloes. III. Parasitic and miscellaneous diseases. *Veterinary Bulletin*, 38, 735-756

- Mokhtar, A.S., Lim, S.F. & Tay, S.T. (2013). Molecular detection of *Anaplasma platys* and *Babesia gibsoni* in dogs in Malaysia. *Tropical Biomedicine* 30(2): 345-348.
- Molin, Y., Lindeborg, M., Nyström, F., Madder, M., Hjelm, E., Olsen, B., & Ehrenborg, C. (2011). Migratory birds, ticks, and *Bartonella*. *Infection Ecology & Epidemiology*, 1.doi: 10.3402/iee.v1i0.5997
- Molloy, J. B., Bock, R. E., Templeton, J. M., Bruyeres, A. G., Bowles, P. M., Blight, G. W., & Jorgensen, W. K. (2001). Identification of antigenic differences that discriminate between cattle vaccinated with *Anaplasma centrale* and cattle naturally infected with *Anaplasma marginale*. *International Journal for Parasitology*, 31(2), 179-186.
- Mourya, D. T., Yadav, P. D., & Patil, D. Y. (2014). Highly infectious tick borne viral diseases: Kyasanur forest disease and Crimean-Congo hemorrhagic fever in India. *WHO South-East Asia Journal of Public Health*, 3(1), 8-21.
- Morel, P. (1989). *Ticks identification using keys*. *Manual for Tropical Veterinary Parasitology*, 9: 301-463pp.
- Murase, T., Ueda, T. & Yamato, O. (1996). Oxidative damage and enhanced erythrophagocytosis in canine erythrocytes infected with *Babesia gibsoni*. *Journal of Veterinary Medical Science*, 58(3), 259-261.
- Mylonakis, M.E., Koutinas, A.F., Breitschwerdt, E.B., Hegarty, B.C., Billinis, C.D., Leontides, L.S. & Kontos, V.S. (2004). Chronic canine ehrlichiosis (*Ehrlichia canis*): a retrospective study of 19 natural cases. *Journal of the American Animal Hospital Association* 40, 174–184.
- Mylonakis, M. E., Leontides, L., Gonen, L., Billinis, C., Koutinas, A. F., & Baneth, G. (2005). Anti-*Hepatozoon canis* serum antibodies and gamonts in naturally-occurring canine monocytic ehrlichiosis. *Veterinary Parasitology*, 129(3), 229-233.
- Mylonakis, M.E., Siarkou, V.I., Leontides, L., Bourtzi-Hatzopoulou, E., Kontos, V.I., Koutinas, A.F. (2009). Evaluation of a serum-based PCR assay for the diagnosis of canine monocytic ehrlichiosis. *Veterinary Microbiology* 138, 390–393.
- Mylonakis, M. E., Ceron, J. J., Leontides, L., Siarkou, V. I., Martinez, S., Tvarijonaviciute, A., ... & Harrus, S. (2011). Serum acute phase proteins as clinical phase indicators and outcome predictors in naturally occurring canine monocytic ehrlichiosis. *Journal of Veterinary Internal Medicine*, 25(4), 811-817.
- Nazari, M., Lim, S.Y., Watanabe, M., Sharma, R.S., Cheng, N.A. & Watanabe, M. (2013). Molecular detection of *Ehrlichia canis* in dogs in Malaysia. *PLoS Neglected Tropical Diseases*, 2013, 7(1), e1982.

- Neer, T. M., & Harrus, S. (2006). Canine monocytotropic ehrlichiosis and neorickettsiosis. *E. canis*, *E. chaffeensis*, *E. ruminantium*, *N. sennetsu*, 203-216.
- Nijhof, A.M., Penzhorn, B.L., Lynen, G., Mollel, J.O., Morkel, P., Bekker, C.P. & Jongejan, F. (2003). *Babesia bicornis* sp. nov. and *Theileria bicornis* sp. nov.: tick-borne parasites associated with mortality in the black rhinoceros (*Diceros bicornis*). *Journal of Clinical Microbiology*, 41:2249– 2254.
- Nijhof, A. M., Bodaan, C., Postigo, M., Nieuwenhuijs, H., Opsteegh, M., Franssen, L., & Jongejan, F. (2007). Ticks and associated pathogens collected from domestic animals in the Netherlands. *Vector-borne and Zoonotic Diseases*, 7(4), 585-596.
- Nowak, M. (2010). Parasitisation and localisation of ticks (Acari: Ixodidae) on exotic reptiles imported into Poland. *Annals of Agricultural and Environmental Medicine* 17, 237-242
- Nyindo, M., Huxsoll, D.L., Ristic, M., Kakoma, I., Brown, J.L., Carson, C.A. & Stephenson, E.H. (1980). Cell-mediated and humoral immune responses of German Shepherd Dogs and Beagles to experimental infection with Ehrlichia canis. *American Journal of Veterinary Research*, 41, 250–254.
- Ogden, N. H., Bigras-Poulin, M., O'callaghan, C. J., Barker, I. K., Lindsay, L. R., Maarouf, A. & Charron, D. (2005). A dynamic population model to investigate effects of climate on geographic range and seasonality of the tick *Ixodes scapularis*. *International Journal for Parasitology*, 35(4), 375-389.
- Ogo, N. I., Lawal, A. I., Okubanjo, O.O., Kamani, J., & Ajayi, O.O. (2013). Current Status of Canine Babesiosis and the Situation in Nigeria: A Review . *Nigerian Veterinary Journal* Vol. 32(2): 2011 ; 69 – 78
- Oinesa, O., Storli, K., & Brun-Hansen, H. (2010). First case of babesiosis caused by *Babesia canis canis* in a dog from Norway. *Veterinary Parasitology* 171: 350-353
- Otsuka, Y., Yamasaki, M., Yamato, O., & Maede, Y. (2001). Increased Generation of Superoxide in Erythrocytes infected with *Babesia gibsoni*. *Journal of Veterinary Medical Science*, 63(10), 1077-1081.
- Oyamada, M., Davoust, B., Boni, M., Dereure, J., Bucheton, B., Hammad, A., Itamoto, K., Okuda, M. & Inokuma, H. (2005). Detection of *Babesia canis rossi*, *B. canis vogeli*, and *Hepatozoon canis* in dogs in a village of eastern Sudan by using a screening PCR and sequencing methodologies. *Clinical and Diagnostic Laboratory Immunology*, 12(11), 1343-1346.

- Palmer, G. H., Abbott, J. R., French, D. M., & McElwain, T. F. (1998). Persistence of *Anaplasma ovis* infection and conservation of the msp-2 and msp-3 multigene families within the genus *Anaplasma*. *Infection and immunity*, 66(12), 6035-6039.
- Papadopoulos, B., Morel, P. C., & Aeschlimann, A. (1996). Ticks of domestic animals in the Macedonia region of Greece. *Veterinary Parasitology*, 63(1), 25-40.
- Parola, P., Cornet, J. P., Sanogo, Y. O., Miller, R. S., Van Thien, H., Gonzalez, J. P., ... & Wongsrichanalai, C. (2003). Detection of *Ehrlichia* spp., *Anaplasma* spp., *Rickettsia* spp., and other eubacteria in ticks from the Thai-Myanmar border and Vietnam. *Journal of Clinical Microbiology*, 41(4), 1600-1608.
- Patz, J. A., Olson, S. H., Uejio, C. K., & Gibbs, H. K. (2008). Disease emergence from global climate and land use change. *Medical Clinics of North America*, 92(6), 1473-1491.
- Pawar, R. M., Poornachandar, A., Srinivas, P., Rao, K. R., Lakshmikantan, U., & Shivaji, S. (2012). Molecular characterization of *Hepatozoon* spp. infection in endangered Indian wild felids and canids. *Veterinary Parasitology*, 186(3), 475-479.
- Penzhorn, B. L., Lewis, B. D., De Waal, D. T., & Rebollar, L. L. (1995). Sterilisation of *Babesia canis* infections by imidocarb alone or in combination with diminazene. *Journal of the South African Veterinary Association*, 66(3), 157-159.
- Perez, M., Rikihisa, Y., & Wen, B. (1996). *Ehrlichia canis*-like agent isolated from a man in Venezuela: antigenic and genetic characterization. *Journal of Clinical Microbiology*, 34(9), 2133-2139.
- Perry, B. D., Randolph, T. F., Mcdermott, J. J., Sones, K. R. & Thornton, P. K. (2002). Investing in animal health research to alleviate poverty. International Livestock Research Institute, Nairobi, Kenya.
- Peters, W., & Pasvol, G. (2002). Arthropod-borne infections. *Tropical Medicine and Parasitology* (Peters, W. and Pasvol, G., eds), 1-107.
- Petney, T. N. (1993). A preliminary study of the significance of ticks and tick-borne diseases in South-east Asia. *Mitt Österr Ges Tropenmed Parasitol*, 15, 33-42.
- Piesman, J. (1987): Emerging tick-borne diseases in temperate climates. *Parasitology Today*, 3, 197-199

- Pinyoowong D, Jittapalapong S, Suksawat F, Stich R. & Thamchaipenet A (2008). Molecular characterization of Thai *Ehrlichia canis* and *Anaplasma platys* strains detected in dogs. *Infection Genetics and Evolution* 2008: 433-8.
- Podberscek, A. L., Paul, E. S., & Serpell, J. A. (2005). *Companion animals and us: Exploring the relationships between people and pets*. Cambridge University Press.
- Poitout, F. M., Shinozaki, J. K., Stockwell, P. J., Holland, C. J. & Shukla, S. K. (2005). Genetic variants of *Anaplasma phagocytophilum* infecting dogs in Western Washington State. *Journal of Clinical Microbiology*, 43(2), 796-801.
- Purnell, R. E. (1981). Babesiosis in various hosts. In: Ristic, M., Kreier, J. P. (Eds.): *Babesiosis*. Academic Press, New York.
- Rahman W.A., Lye YP. & Chandrawathani, P. (2010). The seroprevalence of bovine babesiosis in Malaysia. *Tropical Biomedicine* 27(2): 301-307.
- Rajamanickam, C., Wiesenhutter, E., Zin, F. M., & Hamid, J. (1985). The incidence of canine haematozoa in Peninsular Malaysia. *Veterinary Parasitology*, 17(2), 151-157.
- Ramananickam, C. (1977). *Babesia* infection in a ten-day old calf. *South-east Asian Journal of Tropical Medicine and Public Health*. 8, 132
- Rani, P. A., Irwin, P. J., Coleman, G. T., Gatne, M., & Traub, R. J. (2011). A survey of canine tick-borne diseases in India. *Parasites and Vectors*, 4(article 141).
- Richmond, Simon (2010). Malaysia, Singapore & Brunei. *Lonely Planet*. pp. 74–75. ISBN 978-1-74104-887-2.
- Rikihisa, Y., Perry, B.D. & Cordes, D.O. (1985). Ultrastructural study of ehrlichial organisms in the large colons of ponies infected with Potomac horse fever. *Infection and Immunity* 49, 505–512.
- Rikihisa, Y., Zhi, N., Wormser, G.P., Wen, B., Horowitz, H.W., Hechemy, K.E. (1997). Ultrastructural and antigenic characterization of a granulocytic ehrlichiosis agent directly isolated and stably cultivated from a patient in New York State. *Journal of Infectious Diseases* 175, 210–213.
- Ristic, M. & Holland, C.J. (1993). Canine Ehrlichiosis. In: Woldehiwet, Z., Ristic, M. (Eds): *Rickettsial and Chlamydial Diseases of Domestic Animals*. Pergamon Press. New York, pp. 169-186. 1993
- Robson, C. & Allen, R. (2000). *A field guide to the birds of South-East Asia* (p. 504). London: New Holland.

- Rubini, A. S., dos Santos Paduan, K., Cavalcante, G. G., Ribolla, P. E. M., & O'Dwyer, L. H. (2005). Molecular identification and characterization of canine Hepatozoon species from Brazil. *Parasitology Research*, 97(2), 91-93.
- Rudoler, N., Harrus, S., Martinez-Subiela, S., Tvarijonaviciute, A., van Straten, M., Cerón, J. J., & Baneth, G. (2015). Comparison of the acute phase protein and antioxidant responses in dogs vaccinated against canine monocytic ehrlichiosis and naive-challenged dogs. *Parasites & Vectors*, 8(1), 175.
- Saitou N. & Nei M. (1987). The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4:406-425.
- Salman, M. D. (2012). Ticks and tick-borne diseases: geographical distribution and control strategies in the Euro-Asia region. *CABI*. Pp 292.
- Sasaki, M., Omobowale, O., Tozuka, M., Ohta, K., Matsuu, A., Nottidge, H. O., Hirata, H., Ikadai, H., & Oyamada, T. (2007). Molecular survey of *Babesia canis* in dogs in Nigeria. *Journal of Veterinary Medical Science*, 69(11), 1191-1193.
- Schaffner, W. & Standaert S.M. (1996). Ehrlichiosis—in pursuit of an emerging infection. *New England Journal of Medicine*, 334(4):262– 63
- Schlegel, M. (1991). Protist evolution and phylogeny as discerned from small subunit ribosomal RNA sequence comparisons. *European Journal of Protistology*, 27(3), 207-219.
- Serpell, J. (1996). In the company of animals: A study of Human-Animal relationships Cambridge: *Cambridge University Press*.
- Shaw, S., Day, M., Birtles, R. & Breitschwerdt, E.B. (2001a). Tickborne infectious diseases of dogs. *Trends Parasitology*; 17: 74-80.
- Smith, R.D., Sells, D.M., Stephenson, E.H., Rictic, M.R. & Huxsoll, D.L. (1976). Development of Ehrlichia canis, causative agent of canine ehrlichiosis, in the tick Rhipicephalus sanguineus and its differentiation from a symbiotic rickettsia. *American Journal of Veterinary Research*, 37: 119-126.
- Smith, R. D., & Kakoma, I. (1989). A reappraisal of vector control strategies for babesiosis. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 83, 43-52.
- Smith, T.G. (1996). The genus *Hepatozoon* (Apicomplexa: Adeleina). *Journal of Parasitology*, 82, 565–585.

- Smith, T. G., & Desser, S. S. (1997). Phylogenetic analysis of the genus Hepatozoon Miller, 1908 (Apicomplexa: Adeleorina). *Systematic Parasitology*, 36(3), 213-221.
- Sobo, E., Eng, B. & Kassity-Krich, N. (2006). Canine visitation (pet) therapy: pilot data on decreases in child pain perception. *Journal of Holistic Nursing*, 24(1):51-57.
- Solano-Gallego, L., Llull, J., Osso, M., Hegarty, B., & Breitschwerdt, E. (2006). A serological study of exposure to arthropod-borne pathogens in dogs from northeastern Spain. *Veterinary Research*, 37(2), 231-244.
- Solano-Gallego, L., & Baneth, G. (2011). Babesiosis in dogs and cats—expanding parasitological and clinical spectra. *Veterinary Parasitology*, 181(1), 48-60.
- Soulsby E.J.L. (1982). Helminthes, Arthropods and Protozoa of Domestic Animals, 7th ed, Bailliere Tindall, London, Pp. 765-777.
- Stich, R.W., Rikihisa, Y., Ewing, S.A., Needham, G.R., Grover, D.L. & Jittapalapong, S. (2002). Detection of *Ehrlichia canis* in canine carrier blood and in individual experimentally infected ticks with a p30-based PCR assay. *Journal of Clinical Microbiology* 40, 540–546.
- Stich, R. W., Blagburn, B. L., Bowman, D. D., Carpenter, C., Cortinas, M. R., Ewing, S. A., Foley, D., Foley, J.E., Gaff, H., Hickling, G.J. and Lash, R. R. (2014). Quantitative factors proposed to influence the prevalence of canine tick-borne disease agents in the United States. *Parasites and Vectors*, 7(1), 1-8.
- Stockham, S.L., Schmidt, D.A. and Tyler, J.W. (1985). Canine granulocytic ehrlichiosis in dogs from central Missouri: A possible cause of polyarthritis. *Veterinary Medical Review*, 6: 3-5.
- Suksawat, J., Xuejie, Y., Hancock, S., Hegarty, B., Nilkumhang, P. & Breitschwerdt, E. (2001a). Serologic and molecular evidence of coinfection with multiple vector-borne pathogens in dogs from Thailand. *Journal of Veterinary Internal Medicine*, 2001a; 15: 453-462.
- Suksawat, J., Pitulle, C., Arraga-Alvarado, C., Madrigal, K., Hancock, S.I. & Breitschwerdt, E.B. (2001b). Coinfection with three *Ehrlichia* species in dogs from Thailand and Venezuela with emphasis on consideration of 16S ribosomal DNA secondary structure. *Journal of Clinical Microbiology*, 39: 90-3.
- Sumner, J. W., Nicholson, W. L., & Massung, R. F. (1997). PCR amplification and comparison of nucleotide sequences from the groESL heat shock operon of Ehrlichia species. *Journal of Clinical Microbiology*, 35(8), 2087-2092.

- Sykes, J.E. (2014). A Text book on Canine and feline infectious diseases: Bacterial diseases. Copyright 2014 by Saunders, an imprint of Elsevier Inc. ISBN 978-1-4377-0795-3. Pp 915.
- Sykes, J.E. & Foley, J.E. (2014). Anaplasmosis: A textbook on canine and feline infectious diseases. Chapter 29: Page 290-299.
- Tack, W., Madder, M., and Verheyen, K. (2010). De impact van mastjaren op knaagdieren, teken en de ziekte van lyme. *Antenna 4* (3), 29-31
- Tamura, K., Stecher, G., Peterson, D., Filipski, A., & Kumar S. (2013). MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Molecular Biology and Evolution* 30: 2725-2729.
- Tanskul, P. (L.) & Inlao, I. (1989). Keys to the adult ticks of *Haemaphysalis* Koch, 1844, in Thailand with notes on changes in taxonomy (Acari: Ixodoidea: Ixodidae). *Journal of Medical Entomology*, 26, 573-601.
- Temeyer, K. B., Pound, J. M., Miller, J. A., Chen, A. C., Pruett, J. H., Guerrero, F. D., ... & George, J.E. (2004). Organophosphate resistance in Mexican strains of *Boophilus microplus*: a major threat to the US cattle industry. *SAAS Bull. Biochemistry and Biotechnology*, 17, 43-51.
- Thompson, J.D., Higgins, D.G., Gibson, T.J. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties, and weight matrix choice. *Nucleic Acids Research*. 22, 4673–4680.
- Tian, Z., Luo, J., Zheng, J., Xie, J., Shen, H., Yin, H., Luo, J., Tian, M., Yuan, X., Wang, F. & Liu, G. (2013). Phylogenetic analysis of *Babesia* species in China based on cytochrome b (COB) gene. *Infection, Genetics and Evolution*, 13, 36-40.
- Tiwary, S. S. (2009). Encyclopaedia of Southeast Asia and Its Tribes (Set Of 3 Vols.). *Anmol Publications Pvt. Ltd.* p. 37. ISBN 978-81-261-3837-1.
- Uilenberg, G. (1992). Veterinary significance of ticks and tick-borne diseases. In *Tick Vector Biology* (pp. 23-33). Springer Berlin Heidelberg.
- Varikoden, H., Samah, A. A., & Babu, C. A. (2010). Spatial and temporal characteristics of rain intensity in the peninsular Malaysia using TRMM rain rate. *Journal of Hydrology*, 387(3), 312-319.
- Varikoden, H., Preethi, B., Samah, A. A., & Babu, C. A. (2011). Seasonal variation of rainfall characteristics in different intensity classes over Peninsular Malaysia. *Journal of Hydrology*, 404(1), 99-108.
- Varshney, D.V.V and Chaudhary, P.S. (2008). Multisystemic effects of canine babesiosis and management of critical cases. *Intas Polivet*. 9:281-287

- Vincent-Johnson, N. A., Macintire, D. K., Lindsay, D. S., Lenz, S. D., Baneth, G., Shkap, V., & Blagburn, B. L. (1997). A new *Hepatozoon* species from dogs: description of the causative agent of canine hepatozoonosis in North America. *The Journal of Parasitology*, 1165-1172.
- Voinov, I.N. & Votyakov, V.I. (1992). Influence of socio-economic and ecological conditions on tickborne encephalitis morbidity in populated areas. *Medical Parasitology*, 1:34–37 (from Russian).
- Unver, A., Rikihisa, Y., Kawahara, M., & Yamamoto, S. (2003). Analysis of 16S rRNA gene sequences of *Ehrlichia canis*, *Anaplasma platys*, and *Wolbachia* species from canine blood in Japan. *Annals of the New York Academy of Sciences*, 990(1), 692-698.
- Unver, A., Huang, H., & Rikihisa, Y. (2006). Cytokine gene expression by peripheral blood leukocytes in dogs experimentally infected with a new virulent strain of *Ehrlichia canis*. *Annals of the New York Academy of Sciences*, 1078(1), 482-486.
- Vargas-Hernández, G., André, M. R., Faria, J. L. M., Munhoz, T. D., Hernandez-Rodriguez, M., Machado, R. Z., & Tinucci-Costa, M. (2012). Molecular and serological detection of *Ehrlichia canis* and *Babesia vogeli* in dogs in Colombia. *Veterinary Parasitology*, 186(3), 254-260.
- Viale, A. M., Arakaki, A. K., Soncini, F. C., & Ferreyra, R. G. (1994). Evolutionary relationships among eubacterial groups as inferred from GroEL (chaperonin) sequence comparisons. *International Journal of Systematic and Evolutionary Microbiology*, 44(3), 527-533.
- Visser, E. S., Ambrosio, R. E., & De Waal, D. T. (1991). An *Anaplasma centrale* DNA probe that differentiates between *Anaplasma ovis* and *Anaplasma marginale* DNA. *Veterinary microbiology*, 28(3), 313-325.
- Waner, T., Harrus, S., Jongejan, F., Bark, H., Keysary, A., & Cornelissen, A. W. (2001). Significance of serological testing for ehrlichial diseases in dogs with special emphasis on the diagnosis of canine monocytic ehrlichiosis caused by *Ehrlichia canis*. *Veterinary Parasitology*, 95(1), 1-15.
- Waner, T., Harrus, S., Bark, H., Bogin, E., Avidar, Y. & Keysary, A. (1997). Characterization of the subclinical phase of canine ehrlichiosis in experimentally infected beagle dogs. *Veterinary Parasitology* 69, 307–317.
- Wang, D., Bowman, D.D., Brown, H.E., Harrington, L.C., Kaufman, P.E., McKay, T., Nelson, C.T., Sharp, J.L., Lund, R. (2014). Factors influencing U.S. canine heartworm (*Dirofilaria immitis*) prevalence. *Parasite and Vectors* 2014, 7(1):264.

- Watanabe, M., Okuda, M., Tsuji, M., Inokuma, H. (2004). Seroepidemiological study of canine ehrlichial infections in Yamaguchi prefecture and surrounding areas of Japan. *Veterinary Parasitology*, 124(1), 101-107.
- Watanabe, M. (2012). Main pet Arthropod-borne diseases in Asia. *9th Merial symposium on parasitosis and arthropod-borne diseases*. Tokyo 6th – 9th November 2012. Pp 18-21.
- Wilson, M.E. (1995). Travel and the emergence of infectious diseases. *Emerging Infectious Diseases*, 1(2):39–46
- Woody, B.J., Hoskins, J.D. (1991). Ehrlichial diseases of dogs. *Veterinary Clinics of North America: Small Animal Practice* 21, 75–98.
- Yadav, P. D., Vincent, M. J., Khristova, M., Kale, C., Nichol, S. T., Mishra, A. C., & Mourya, D. T. (2011). Genomic analysis reveals Nairobi sheep disease virus to be highly diverse and present in both Africa, and in India in the form of the Ganjam virus variant. *Infection, Genetics and Evolution*, 11(5), 1111-1120.
- Yap, B.K. (2004). Survey of blood protozoon and Ehrlichia spp. in dogs in Klang, Selangor. DVM Thesis. Faculty of Veterinary Medicine and Animal Science, Universiti Putra Malaysia, Serdang. Unpublished data.
- Yeoh, W.H. (2009). Blood parasites in dogs in Klang Valley: Prevalence and diagnosis. DVM Thesis. Faculty of Veterinary Medicine, Universiti Putra Malaysia, Serdang. Unpublished data.
- Yu, X. J., Zhang, X. F., McBride, J. W., Zhang, Y., & Walker, D. H. (2001). Phylogenetic relationships of *Anaplasma marginale* and 'Ehrlichia platys' to other Ehrlichia species determined by GroEL amino acid sequences. *International Journal of Systematic and Evolutionary Microbiology*, 51(3), 1143-1146.
- Yu, X. J., McBride, J. W., & Walker, D. H. (2007). Restriction and expansion of *Ehrlichia* strain diversity. *Veterinary Parasitology*, 143(3), 337-346.
- Zahler, M., Schein, E., Rinder, H., Gothe, R. (1998). Characteristic genotypes discriminate between *Babesia canis* isolates of differing vector specificity and pathogenicity to dogs. *Parasitology Research*, 84: 544-54.
- Zhang, X., Luo, T., Keysary, A., Baneth, G., Miyashiro, S., Strenger, C., Waner, T. and McBride, J.W., (2008). Genetic and antigenic diversities of major immunoreactive proteins in globally distributed *Ehrlichia canis* strains. *Clinical and Vaccine Immunology*, 15(7), 1080-1088.
- Zimmermann, M., Veeck, J., & Wolf, K. (1998). Minimizing the exposure to UV light when extracting DNA from agarose gels. *BioTechniques*, 25(4), 586.

- Zulkifli, A.R. (2011) Molecular study of *Babesia* in canine blood and comparison between conventional and molecular diagnostic methods. DVM Thesis. Faculty of Veterinary Medicine, Universiti Putra Malaysia, Serdang. Unpublished data.
- Zahler, M., Schein, E., Rinder, H., & Gothe, R. (1998). Characteristic genotypes discriminate between *Babesia canis* isolates of differing vector specificity and pathogenicity to dogs. *Parasitology Research*, 84(7), 544-548.
- Zahler, M., Rinder, H., Schein, E., & Gothe, R. (2000a). Detection of a new pathogenic *Babesia* microti-like species in dogs. *Veterinary Parasitology*, 89(3), 241-248.
- Zahler, M., Rinder, H., Zwegarth, E., Fukata, T., Maede, Y., Schein, E., & Gothe, R. (2000b). '*Babesia gibsoni*' of dogs from North America and Asia belong to different species. *Parasitology*, 120(04), 365-369.
- Zygner, W. O. J. C. I. E. C. H., Gojska-Zygner, O., Długosz, E., & Wędrychowicz, H. A. L. I. N. A. (2011). Liver enzyme activity in dogs infected with *Babesia canis*. *Bulletin of Veterinary Institute Pulawy*, 55, 423-427.