



UNIVERSITI PUTRA MALAYSIA

***ROLE OF BIOFILM FORMATION ON THE PATHOGENESIS OF
Streptococcus agalactiae INFECTION IN RED HYBRID TILAPIA
Oreochromis niloticus x Oreochromis mossambicus***

ISIAKU ABDULSALAM IBRAHIM

FPV 2016 31



**ROLE OF BIOFILM FORMATION ON THE PATHOGENESIS OF
Streptococcus agalactiae INFECTION IN RED HYBRID TILAPIA
Oreochromis niloticus x *Oreochromis mossambicus***

By

ISIAKU ABDULSALAM IBRAHIM

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of
Master of Veterinary Science**

December 2016

COPYRIGHT

All materials contained within the thesis, including and without limitation to text, logos, icons, photographs and all other artwork, is a copyright material of the Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with express, prior or written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This work is dedicated to my lovely parents



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Veterinary Science

**ROLE OF BIOFILM FORMATION ON THE PATHOGENESIS OF
Streptococcus agalactiae INFECTION IN RED HYBRID TILAPIA
Oreochromis niloticus x *Oreochromis mossambicus***

By

ISIAKU ABDULSALAM IBRAHIM

December 2016

Chairman : Assoc. Prof. Sabri Md. Yusoff, PhD
Faculty : Veterinary Medicine

Streptococcus agalactiae (group B *Streptococcus*, GBS) is an important pathogen of aquatic animals that has led to significant economic loss due to frequent disease outbreak and mortalities in fish worldwide. Tilapia has shown an unusual susceptibility to GBS infection, which is often characterised by acute septicaemia or chronic meningoencephalitis. While acute septicaemia is a sequel to an invasive infection, the mechanism of chronic meningoencephalitis in fish is not fully understood. However, most pathogens that induce acute invasive diseases are capable of developing biofilm associated chronic lesions. Biofilm is an assemblage of irreversibly attached microbial organisms within generated matrix of extracellular polymeric substances. The aim of this research was to investigate the role of biofilm in piscine GBS infection in red hybrid tilapia. A putative piscine GBS biofilm strain was selected and analysed for biofilm formation *in vitro*. The piscine GBS strain exhibited a weak attachment to polystyrene plates by standard crystal violet assay. Furthermore, fluorescent *in situ* hybridization and confocal laser scanning microscopy revealed discrete aggregates of attached piscine GBS around the brain meningeal surface of the orally exposed experimental tilapia. Importantly, these organised aggregates were first detected at a time point corresponding to the transition from an acute to chronic infection. The aggregates were embedded in a polysaccharide containing matrix and became intractable to antibiotic treatment despite earlier *in vitro* susceptibility on sensitivity test. The eye and stomach had no aggregates suggestive of the sessile life style. Intracellular bacterial aggregates, such as within erythrocytes and ventricular ependyma of the brain were also observed. Leukocytic infiltrates predominantly macrophages were readily seen around biofilms, while

erythrocytes appeared often coagulated and severely injured as shown by increased expression of heat shock protein 70. Moreover, an effective adaptive immune response was not detected during the period of study. The erythrocytes may have facilitated invasion of GBS into the brain of tilapia. The present research demonstrates for the first time, that biofilm is associated with persistence of piscine GBS and development of chronic meningoencephalitis in the experimental tilapia. It provides a foundation for further investigation and the development of a holistic framework to prevent GBS infection in fish. Current approaches including vaccine strategies need to be reviewed to account for the biofilm phenotype.

Keywords: Tilapia, *Streptococcus agalactiae*, meningoencephalitis, biofilm, *in situ* hybridization

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Master Sains Veterinar

**PERANAN PEMBENTUKAN BIOFILEM KE ATAS PATOGENESIS
Streptococcus agalactiae PADA TILAPIA HYBRID MERAH
Oreochromis niloticus x *Oreochromis mossambicus***

Oleh

ISIAKU ABDULSALAM IBRAHIM

Disember 2016

Pengerusi : Prof. Madya Sabri Md. Yusoff, PhD
Fakulti : Perubatan Veterinar

Streptococcus agalactiae (Streptokokus kumpulan B, GBS) merupakan patogen penting dalam haiwan akuatik yang membawa kepada kerugian ekonomi yang ketara disebabkan oleh kekerapan wabak penyakit dan kematian ikan di seluruh dunia. Ikan tilapia telah menunjukkan kecenderungan jangkitan GBS yang selalunya dikategorikan sebagai septisemia akut atau meningoensefalitis kronik. Oleh kerana septisemia akut merupakan ekoran dari jangkitan invasif, mekanisme bagi penyakit meningoensefalitis kronik pada ikan masih belum difahami sepenuhnya. Walau bagaimanapun, kebanyakan patogen yang menggalakkan penyakit invasif akut ini mampu membentuk biofilem yang berkaitan lesi kronik. Biofilem adalah gabungan organisma mikrob yang berkeadaan kekal di antara matrik janaan dari bahan-bahan polimerik luar sel. Kajian ini bertujuan untuk mengkaji peranan biofilem terhadap jangkitan GBS dalam tilapia hibrid merah. Biofilem jenis *piscine* putatif telah dipilih dan dianalisa untuk formasi biofilem secara *in vitro*. *Piscine* GBS telah menunjukkan pelekatan yang lemah kepada plat polisterin oleh asai standard kristal violet. Tambahan, hibridisasi pendarflour *in situ* dan mikroskopi imbasan laser sefokus menunjukkan agregat diskret oleh GBS di sekitar permukaan meningeal otak yang terdedah secara oral dalam tilapia. Pentingnya, agregat yang teratur telah dikesan awal pada titik masa yang sepadan dengan transisi daripada jangkitan akut kepada jangkitan kronik. Agregat tersebut yang terbenam dalam polisakarida yang mengandungi matrik dan menjadi sukar untuk dirawat dengan rawatan walaupun pada awalnya lebih cenderung dengan ujian sensitiviti. Mata dan perut yang tiada agregat menandakan gaya hidup yang sesil. Bakteria intrasel beragregat seperti di antara sel eritrosit dan ventrikel endodima otak juga turut diperhatikan. Penyusupan leukosit kebanyakannya

ialah makrofaj dapat dilihat di sekitar biofilem sementara kemunculan eritrosit kerap dalam keadaan membeku dan luka teruk seperti yang ditunjukkan dengan peningkatan ekspresi kejutan haba protein 70. Disamping itu, tindak balas imun mudah suai efektif tidak dapat dikesan sepanjang tempoh kajian. Eritrosit mungkin memudahkan GBS untuk menyerang bahagian otak tilapia. Kajian terkini menunjukkan buat pertama kalinya, biofilem yang berkaitan dengan GBS berterusan dan pembentukan kronik meningoensefalitis dalam eksperimen tilapia. Ia turut menyediakan asas kepada penyelidikan lanjutan dan pembangunan satu rangka kerja yang menyeluruh untuk mencegah jangkitan GBS dalam ikan. Pendekatan semasa termasuk strategi vaksin perlu dikaji semula untuk diskripsi fenotip biofilem.

Kata kunci: Tilapia, *Streptococcus agalactiae*, biofilem, meningoensefalitis, hibridasi *in situ*

ACKNOWLEDGEMENTS

All praise and glorifications are due to the Almighty, Omnipotent God, Who by His grace we saw to the end of this program. I wish to firstly thank my supervisory committee, especially Assoc. Prof. Dr. Md. Sabri Bin Mohd Yusoff, who painstakingly motivated and guided me through this sojourn. I am most indebted for his humility and understanding. I had at many times gone for consultations to Assoc. Prof. Dr. Hassan Mohd Daud and Dr. Ina-Salwany Mohd Yasin without prior appointments. They were very supportive with their warmth welcome, constructive criticism and encouragements.

I am most grateful to my parents, wife, daughters and siblings for their prayers and support during the research period. The world means nothing without you. I will not easily forget the immense support by my father-in-law Alhaji Umar Ambursa and his family, and that of my extended family.

The list of contributors to this achievement cannot be enumerated here, but it is pertinent to acknowledge Kebbi State Government of Nigeria, all my lectures and teachers, senior colleagues and bosom friends. I also had a lot of friends in UPM who made my stay memorable. And to all staffs and students of the Faculty of Veterinary Medicine, UPM for their contributions too numerous to mention. May all those who have contributed to this achievement be rewarded by The Almighty God, Ameen.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Veterinary Science in Pathology. The members of the Supervisory Committee were as follows:

Sabri Md. Yusoff, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Hassan Md. Daud, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Ina Salwany Md. Yasin, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institution;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, in accordance with the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld in accordance with the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection using turnitin software.

Signature: _____ Date: _____

Name and Matric No.: Isiaku Abdulsalam Ibrahim, GS 40272

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDEgements	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xv
 CHAPTER	
 1 INTRODUCTION	 1
1.1 Background of study	1
1.2 Objectives of the study	2
1.3 Hypothesis	2
1.4 Statement of problems	2
1.5 Justification of the study	2
1.6 Significance of the study	3
 2 LITERATURE REVIEW	 4
2.1 Introduction	4
2.2 The biofilm lifestyle of microbial organisms	4
2.2.1 Definition of biofilm	4
2.2.2 Structure and function of biofilm	5
2.2.3 Formation of biofilm	6
2.3 Biofilm in fish disease pathogenesis	8
2.3.1 Introduction to biofilm associated disease	8
2.3.2 Biofilm formation in aquaculture environment	9
2.3.3 Bacterial biofilm in some diseases of aquaculture	10
2.4 Biofilm formation by piscine GBS and associate factors	10
2.5 Methods and applications for bacterial biofilms	11
2.5.1 Indirect techniques	11
2.5.2 Direct techniques	11
2.6 Biofilm and pathogenesis of GBS infection in tilapia	12
2.6.1 Introduction to GBS infection in tilapia	12
2.6.2 Epidemiology and clinical signs of GBS infection in tilapia	13
2.6.3 Pathogenesis of GBS infection in tilapia	14
2.6.4 Pathology of GBS infection in tilapia	18
2.7 Immune responses to biofilm and control of GBS infection	18
2.7.1 Immunity against GBS pathogen in tilapia	18
2.7.2 Suppression of immune response in tilapia by	19

	temperature stress	
2.7.3	Evasion of immune response by the GBS pathogen	20
2.7.4	Vaccines for the prevention and control of GBS pathogen	20
2.7.5	The need for novel adjuvants and delivery systems	22
3	IN VITRO DEVELOPMENT AND QUANTIFICATION OF PISCINE GBS BIOFILM	23
3.1	Introduction	23
3.2	Materials and Methods	24
3.2.1	Polystyrene plate biofilm formation assay	24
3.2.2	Statistical analysis	24
3.2.3	Detection of sortase A, sortase C1 and minor ancilliary protein	24
3.2.3.1	Genomic DNA extraction	24
3.2.3.2	PCR analysis	25
3.2.3.3	PCR product purification	26
3.2.3.4	Bioinformatics	26
3.3	Results and discussion	27
3.3.1	<i>In vitro</i> biofilm formation by piscine GBS	27
3.3.2	Genomic features of GBS biofilm strains	29
3.4	Conclusion	30
4	INVESTIGATION OF GBS BIOFILM FORMATION IN THE BRAIN, EYE AND STOMACH OF RED HYBRID TILAPIA	31
4.1	Introduction	31
4.2	Materials and Methods	32
4.2.1	Experimental design	32
4.2.2	Antibiotic sensitivity test	32
4.2.3	Preparation of inoculum, oral infection and drug treatment	32
4.2.4	Post-mortem procedure and tissue processing	33
4.2.5	Bacterial isolation	34
4.2.6	Cryo-staining and confocal microscopy	34
4.2.7	Fluorescent in situ hybridization	34
4.2.8	Alcian blue periodic acid special staining	35
4.3	Results and discussion	35
4.3.1	Antibiotic sensitivity test profile of the piscine GBS isolate	35
4.3.2	Induction of subclinical infection by piscine GBS	36
4.3.3	Piscine GBS biofilm formation on meningeal surfaces of the brain	39
4.3.4	Absence of biofilm formation on surfaces of the eye and stomach	40
4.4	Conclusion	43

5	MORPHOLOGICAL FEATURES OF SURFACE AND GBS BIOFILM ASSOCIATED LESIONS IN THE BRAIN OF RED HYBRID TILAPIA	44
5.1	Introduction	44
5.2	Materials and Methods	44
5.2.1	Histopathology	44
5.2.2	Immunohistochemistry	45
5.2.3	Enzyme-linked immunosorbent assay	45
5.2.4	Statistical analysis	45
5.3	Results and discussion	46
5.3.1	Morphology of surface and biofilm associated lesions	46
5.3.2	Involvement of erythrocytes in the development of lesion	49
5.3.3	Ineffective adaptive immune response to GBS infection	50
5.4	Conclusion	51
6	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION	52
6.1	Summary	52
6.2	Conclusion	52
6.3	Recommendations	53
	REFERENCES	54
	APPENDICES	65
	BIODATA OF STUDENT	77
	PUBLICATION	78

LIST OF TABLES

Table	Page
3.1 Sequences of established primers and probe used for GBS	25
4.1 Antibiotic sensitivity test result for piscine GBS	36
C.1 Mean biofilm thickness of GBS in the polystyrene plates	73
C.2 IgM response of tilapia experimentally exposed to GBS	73

LIST OF FIGURES

Figure	Page
2.1 Stages of biofilm formation	7
2.2 Aquatic biofilm model of piscine GBS infection in the tilapia host	17
3.1 <i>In vitro</i> characterisation of piscine GBS biofilm	28
4.1 Agarose gel electrophoresis to confirm GBS infection in tilapia	37
4.2 Pie chart representing clinical pattern of piscine GBS chronic infection model	38
4.3 Gross changes in experimental GBS infection	38
4.4 Confocal images of GBS biofilms on the meningeal surface of the brain	41
4.5 Meningeal surface showing piscine GBS exopolysaccharide substance	42
5.1 Histopathology of the meningeal surface of GBS infected tilapia	47
5.2 Microscopic images of the meninges showing increased HSP70 expression by erythrocytes of GBS infected tilapia	48
5.3 Box and leaf plots for IgM antibody response to GBS exposure in tilapia at various time points	50
E.1 Confocal images of the retinal layers of the eye and epithelial layers of the stomach	76

LIST OF ABBREVIATIONS

AB-PAS	Alcian blue and periodic acid Schiff
ANOVA	Analysis of variance
AOPI	Acridine orange and propidium iodide
AP	Ancillary protein
BBB	Blood brain barrier
BHIB	Brain heart infusion broth
BKD	Branched chained alpha-keto acid dehydrogenase
BSA	Bovine serum albumin
CaCl ₂	Calcium chloride
CC	Clonal complex
CFU	Colony forming units
CLSM	Confocal laser scanning microscopy
COX	Cyclooxygenase
Cy3	Cyanine 3
DAB	Diaminobenzidine
DAPI	Diamidino-phenylindone
DB	Denaturation Buffer
ddH ₂ O	Deionised distilled water
DNA	Deoxyribonucleic acid
DVM	Doctor of veterinary medicine
eDNA	Extracellular DNA
ELISA	Enzyme-linked immunosorbent assay
EPS	Exopolymeric substances
FISH	Fluorescent <i>in situ</i> hybridisation
FITC	Fluorescein isothiocyanate

GALT	Gut associated lymphoid tissues
GAPDH	Glyceraldehyde-phosphate dehydrogenase
GAS	General adaptive syndrome
GBS	Group b streptococcus
H&E	Haematoxylin and eosin
HB	Hybridization Buffer
HRP	Horseradish peroxidase
HSP70	Heat shock protein 70
IgM	Immunoglobulin M
IHC	Immunohistochemistry
IL	Interleukin
iTRAQ	Isobaric tagging for relative and absolute quantification
MS 222	Tricaine methanesulfonate
NaCl	Sodium chloride
NYSC	National youth service corp
OD	Optical density
PAMPs	Pathogen associated membrane patterns
PBS	Phosphate buffered saline
PBST	Phosphate buffered saline Tween 20
PCR	Polymerase chain reaction
PDE	Professional diploma in education
PFA	Paraformaldehyde
PI	Pilus island
PK	Pyruvate kinase
PRR	Pathogen recognition receptors
PVC	Polyvinyl chloride
QS	Quorum sensing

RAS	Recirculating aquaculture system
SD	Standard deviation
SEM	Scanning electron microscopy
Srt	Sortase
SSC	Saline sodium citrate
ST	Sequence type
TCR	T-cell receptor
TMB	Tetramethylbenzidine
TNF	Tumour necrosis factor
Tris-HCl	Tris hydrochloric acid

CHAPTER 1

INTRODUCTION

1.1 Background of study

The existence of bacterial organisms in nature as biofilm is recently well acknowledged, and when present on tissues it is often associated with disease chronicity (Hall-Stoodley & Stoodley 2009). Biofilm is an assemblage of surface associated, irreversibly attached microbes within generated matrix of extracellular polymeric substance (Donlan 2002). Unlike freely suspended or planktonic cells, they are characterised by a reduce growth rate and the regulation of specific genes (Donlan 2002, Cheng et al., 2010).

Many species of *Streptococcus* naturally exist within these organised surface associated microbial communities (Nobbs et al., 2009). Importantly, *Streptococcus agalactiae* also known as group B *Streptococcus* (GBS) was shown to exhibit the biofilm phenotype *in vitro* (Rinaudo et al., 2010, Borges et al., 2012). This pathogen affects humans, ruminants and fish. In fish, it has led to a significant economic loss due to disease outbreaks and high mortalities in global aquaculture (Delamare-Deboutteville et al., 2014, Guo et al., 2014).

Piscine GBS infection in Malaysia was first reported in the late 1990s and have since been a serious burden in tilapia culture. Outbreak was earlier confined to Pahang (1997), Terengganu and Kelantan (2002) at Pahang River, Kenyir Lake and Pergau lake respectively, but it is currently widespread in Peninsular Malaysia (Zamri-Saad et al., 2014). The annual revenue from tilapia production has therefore reduced concomitantly. In part, this is because, current treatments and vaccines developed for use against disease by piscine GBS have been ineffective (Sudheesh et al., 2012, Zamri-Saad et al., 2014).

While it has been suggested that juvenile tilapia is an important source of infection to fish farms (Anshary et al., 2014), persistence of GBS in brood stock and its possible acquisition by either egg, larvae, fry or fingerlings require investigation. Indeed, the closely related *Streptococcus pyogenes* and *Streptococcus suis* have been involved in biofilm associated persistence (Grenier et al., 2009, Oliver-Kozup et al., 2011). Biofilm is also very common in aquatic environments, providing a niche for potentially pathogenic bacteria (Parsek & Singh 2003) such as piscine GBS.

The pathogenicity of GBS in tilapia has been ascertained, however, knowledge of its interaction with the respective host is still not fully understood (Abuseliana et al., 2011, Guo et al., 2014, Su et al., 2015). Indeed, it is unknown if piscine GBS has the ability to form biofilm *in vitro* such as the aquatic environment or within the fish host. Therefore, the main aim of this study was to determine biofilm formation by piscine GBS and its role in the pathogenesis of infection in red hybrid tilapia.

1.2 Objectives of the study

The specific objectives set for the study were:

1. to develop and quantify piscine GBS biofilm using a microtitre plate assay.
2. to investigate *in vivo* GBS biofilm formation in the brain, eye and stomach of red hybrid tilapia.
3. to determine the microscopic changes associated with GBS biofilm formation in the brain, eye and stomach of red hybrid tilapia.

1.3 Hypothesis

1. **Null:** piscine GBS does not exhibit biofilm phenotypic characteristics.
2. **Alternate:** piscine GBS exhibit biofilm phenotypic characteristics.

1.4 Statement of problems

1. Piscine GBS is presently a cause of persistent infection, disease outbreak, mortality and high economic loss for tilapia culture in Malaysia and other parts of the world.
2. The pathogenesis of GBS in tilapia is still not fully understood, and the role of biofilm is currently unknown.
3. Biofilm formation is a potential challenge for current attempts to develop effective vaccines and other control strategies against GBS in aquaculture.

1.5 Justification of the study

Piscine GBS is the most important bacterial pathogen in tilapia culture. However, the interaction of GBS with the tilapia host is still not fully understood. In humans, GBS is thought to evade tissue responses and persist through the mechanism of biofilm formation. This underscores the need for a comparative investigation of the biofilm mechanism in the lower vertebrate whose defence system has also completely evolved – with components of both

innate and adaptive immunity. In addition, persistence of GBS in tilapia has been observed in aquaculture and biofilm formation may be involved.

1.6 Significance of the study

The findings of this study further explains the pathogenesis of GBS in tilapia. Accordingly, it provides an additional basis for testing the most effective strategy for prevention, and adoption of better measures in the control of disease by GBS in aquaculture.



REFERENCES

- Abdallah F Ben, Chaieb K, Zmantar T, Kallel H, Bakhrouf A (2009) Adherence assays and slime production of *Vibrio alginolyticus* and *Vibrio parahaemolyticus*. *Brazilian J Microbiol* 40:394–398
- Abdullah S, Omar N, Yusoff SM, Obukwho EB, Nwunuji TP, Hanan L, Samad J (2013) Clinicopathological features and immunohistochemical detection of antigens in acute experimental *Streptococcus agalactiae* infection in red tilapia (*Oreochromis spp.*). *Springerplus* 2:1–7
- Abee T, Kovács ÁT, Kuipers OP, Veen S van der (2011) Biofilm formation and dispersal in Gram-positive bacteria. *Curr Opin Biotechnol* 22:172–179
- Absalon C, Ymele-leki P, Watnick PI (2012) The bacterial biofilm matrix as a platform for protein delivery. *MBio* 3
- Abuseliana AF, Daud HHM, Aziz SA, Bejo SK, Alsaid M (2011) Pathogenicity of *Streptococcus agalactiae* isolated from a fish farm in Selangor to juvenile red tilapia (*Oreochromis sp.*). *J Anim Vet Adv* 10:914–919
- Aisyhah MAS, Amal MNA, Shaqinah NN (2015) *Streptococcus agalactiae* isolates from cultured fishes in Malaysia manifesting low resistance pattern towards selected antibiotics. *J Fish Dis*:1093–1098
- Al-Ahmad A, Wunder A, Auschill TM, Follo M, Braun G, Hellwig E, Arweiler NB (2007) The in vivo dynamics of *Streptococcus spp.*, *Actinomyces naeslundii*, *Fusobacterium nucleatum* and *Veillonella spp.* in dental plaque biofilm as analysed by five-colour multiplex fluorescence *in situ* hybridization. *J Med Microbiol* 56:681–687
- Allan RN, Skipp P, Jefferies J, Clarke SC, Faust SN, Hall-Stoodley L, Webb J (2014) Pronounced metabolic changes in adaptation to biofilm growth by *Streptococcus pneumoniae*. *PLoS One* 9:e107015
- Altinok I, Kurt I (2003) Molecular diagnosis of fish diseases: a review. *Turkish J Fish Aquat Sci* 138:131–138
- Amal MNA, Saad MZ, Zahrah AS, Zulkafli AR (2015) Water quality influences the presence of *Streptococcus agalactiae* in cage cultured red hybrid tilapia, *Oreochromis niloticus* × *Oreochromis mossambicus*. *Aquac Res* 46:313–323
- Amal MNA, Zamri-Saad M, Siti-Zahrah A, Zulkafli AR (2013) Transmission of *Streptococcus agalactiae* from a hatchery into a newly established red hybrid tilapia, *Oreochromis niloticus* (L.) × *Oreochromis mossambicus*

- (Peters), farm. J Fish Dis 36:735–739
- Ammann TW, Belibasakis GN, Thurnheer T (2013) Impact of early colonizers on *in vitro* subgingival biofilm formation. PLoS One 8:e83090
- Anshary H, Kurniawan RA, Sriwulan S, Ramli R, Baxa D V (2014) Isolation and molecular identification of the etiological agents of streptococcosis in Nile tilapia (*Oreochromis niloticus*) cultured in net cages in Lake Sentani, Papua, Indonesia. Springerplus 3:627
- Artz LA, Kempf VAJ, Autenrieth IB (2003) Rapid screening for *Streptococcus agalactiae* in vaginal specimens of pregnant women by fluorescent *in situ* hybridization. J Clin Microbiol 41:2170–2173
- Bales PM, Renke EM, May SL, Shen Y, Nelson DC (2013) Purification and characterization of biofilm-associated EPS exopolysaccharides from ESKAPE organisms and other pathogens. PLoS One 8:e67950
- Barato P, Martins ER, Melo-Cristino J, Iregui CA, Ramirez M (2015) Persistence of a single clone of *Streptococcus agalactiae* causing disease in tilapia (*Oreochromis sp.*) cultured in Colombia over 8 years. J Fish Dis:1–5
- Beloin C, Ghigo JM (2005) Finding gene-expression patterns in bacterial biofilms. Trends Microbiol 13:16–19
- Bonifait L, Grignon L, Grenier D (2008) Fibrinogen induces biofilm formation by *Streptococcus suis* and enhances its antibiotic resistance. Appl Environ Microbiol 74:4969–4972
- Bordi C, Bentzmann S de (2011) Hacking into bacterial biofilms: a new therapeutic challenge. Ann Intensive Care 1:19
- Borges S, Silva J, Teixeira P (2012) Survival and biofilm formation by group B streptococci in simulated vaginal fluid at different pHs. Antonie van Leeuwenhoek, Int J Gen Mol Microbiol 101:677–682
- Bowater RO, Forbes-Faulkner J, Anderson IG, Condon K, Robinson B, Kong F, Gilbert GL, Reynolds A, Hyland S, Mcpherson G, Brien JO, Blyde D (2012) Natural outbreak of *Streptococcus agalactiae* (GBS) infection in wild giant Queensland grouper, *Epinephelus lanceolatus* (Bloch), and other wild fish in northern Queensland, Australia. J Fish Dis 35:173–186
- Cai W, La Fuente L De, Arias CR (2013) Biofilm formation by the fish pathogen *Flavobacterium columnare*: development and parameters affecting surface attachment. Appl Environ Microbiol 79:5633–5642

- Cerca N, Gomes F, Pereira S, Teixeira P, Oliveira R (2012) Confocal laser scanning microscopy analysis of *S. epidermidis* biofilms exposed to farnesol, vancomycin and rifampicin. BMC Res Notes 5:244
- Chen M, Li L-P, Wang R, Liang W-W, Huang Y, Li J, Lei A-Y, Huang W-Y, Gan X (2012) PCR detection and PFGE genotype analyses of streptococcal clinical isolates from tilapia in China. Vet Microbiol 159:526–30
- Cheng K, Demirci A, Catchmark JM (2010) Advances in biofilm reactors for production of value-added products. Appl Microbiol Biotechnol 87:445–456
- Chiba A, Sugimoto S, Sato F, Hori S, Mizunoe Y (2014) A refined technique for extraction of extracellular matrices from bacterial biofilms and its applicability. Microb Biotechnol:1–12
- Coffey BM, Anderson GG (2014) Biofilm formation in the 96-well microtiter plate. Alain Fill Juan-Luis Ramos (Eds), Pseudomonas Methods Protoc Methods Mol Biol 1149:631–641. New York. Humana Press
- Connolly KL, Roberts AL, Holder RC, Reid SD (2011) Dispersal of group A streptococcal biofilms by the cysteine protease SpeB leads to increased disease severity in a murine model. PLoS One 6:1–10
- Convert M, Lucchini GM, Dolina M, Piffaretti JC (2005) Comparison of LightCycler PCR and culture for detection of group B streptococci from vaginal swabs. Clin Microbiol Infect 11:1022–1026
- Dalton T, Dowd SE, Wolcott RD, Sun Y, Watters C, Griswold JA, Rumbaugh KP (2011) An *in vivo* polymicrobial biofilm wound infection model to study interspecies interactions. PLoS One 6
- Davey ME, O'toole GA (2000) Microbial biofilms: from ecology to molecular genetics. Microbiol Mol Biol Rev 64:847–867
- Delamare-Deboutteville J, Bowater R, Condon K, Reynolds A, Fisk A, Aviles F, Barnes AC (2014) Infection and pathology in Queensland grouper, *Epinephelus lanceolatus*, (Bloch), caused by exposure to *Streptococcus agalactiae* via different routes. J Fish Dis:1–15
- Delannoy CMJ, Crumlish M, Fontaine MC, Pollock J, Foster G, Dagleish MP, Turnbull JF, Zadoks RN (2013) Human *Streptococcus agalactiae* strains in aquatic mammals and fish. BMC Microbiol 13:41
- Delannoy C. MJ, Zadoks RN, Crumlish M, Rodgers D, Lainson FA, Ferguson HW, Turnbull J, Fontaine MC (2014) Genomic comparison of virulent and non-virulent *Streptococcus agalactiae* in fish. J Fish Dis:1–17

- Delannoy CMJ, Zadoks RN, Lainson FA, Ferguson HW, Crumlish M, Turnbull JF, Fontaine MC (2012) Draft genome sequence of a nonhemolytic fish-pathogenic *Streptococcus agalactiae* strain. J Bacteriol 194:6341–2
- Donlan RM (2002) Biofilms: microbial life on surfaces. Emerg Infect Dis 8:881–890
- Doran KS, Nizet V (2004) Molecular pathogenesis of neonatal group B streptococcal infection: no longer in its infancy. Mol Microbiol 54:23–31
- Eldar A, Bejerano Y, Livoff A, Horovitz A, Bercovier H (1995) Experimental streptococcal meningo-encephalitis in cultured fish. Vet Microbiol 43:33–40
- Erriu M, Genta G, Tuveri E, Orrù G, Barbato G, Levi R (2012) Microtiter spectrophotometric biofilm production assay analyzed with metrological methods and uncertainty evaluation. Measurement 45:1083–1088
- Evans JJ, Klesius PH, Gilbert PM, Shoemaker CA, Sarawi MA Al, Landsberg J, Duremdez R, Marzouk A Al, Zenki S Al (2002) Characterization of β -haemolytic group B *Streptococcus agalactiae* in cultured seabream, *Sparus auratus* L., and wild mullet, *Liza klunzingeri* (Day), in Kuwait. J Fish Dis 25:505–513
- Evans JJ, Klesius PH, Pasnik DJ, Bohnsack JF (2009) Human *Streptococcus agalactiae* isolate in Nile tilapia (*Oreochromis niloticus*). Emerg Infect Dis 15:774–776
- Evans JJ, Wiedenmayer AA, Klesius PH, Shoemaker CA (2004) Survival of *Streptococcus agalactiae* from frozen fish following natural and experimental infections. Aquaculture 233:15–21
- Fettucciari K, Rosati E, Scaringi L, Cornacchione P, Migliorati G, Sabatini R, Petriconi I, Rossi R, Marconi P (2000) Group B *Streptococcus* induces apoptosis in macrophages. J Immunol 165:3923–3933
- Firdaus-Nawi M, Yusoff SM, Yusof H, Abdullah SZ, Zamri-Saad M (2013) Efficacy of feed-based adjuvant vaccine against *Streptococcus agalactiae* in *Oreochromis spp.* in Malaysia. Aquac Res 45:87–96
- Flemming H, Wingender J (2010) The biofilm matrix. Nat Rev Microbiol 8:623–33
- Flemming H-C, Wingender J, Szewzyk U, Steinberg P, Rice SA, Kjelleberg S (2016) Biofilms: an emergent form of bacterial life. Nat Rev Microbiol 14:563–575

- Geng Y, Wang KY, Huang XL, Chen DF, Li CW, Ren SY, Liao YT, Zhou ZY, Liu QF, Du ZJ, Lai WM (2012) *Streptococcus agalactiae*, an emerging pathogen for cultured ya-fish, *Schizothorax prenanti*, in China. *Transbound Emerg Dis* 59:369–375
- Gil C, Solano C, Burgui S, Latasa C, García B, Toledo-Arana A, Lasa I, Valle J (2014) Biofilm matrix exoproteins induce a protective immune response against *Staphylococcus aureus* biofilm infection. *Infect Immun* 82:1017–1029
- Grenier D, Grignon L, Gottschalk M (2009) Characterisation of biofilm formation by a *Streptococcus suis* meningitis isolate. *Vet J* 179:292–295
- Guo C-M, Chen R-R, Kalhor DH, Wang Z-F, Liu G-J, Lu C-P, Liu Y-J (2014) Identification of genes preferentially expressed by highly virulent piscine *Streptococcus agalactiae* upon interaction with macrophages. *PLoS One* 9:e87980
- Hall-Stoodley L, Nistico L, Sambanthamoorthy K, Dice B, Nguyen D, Mershon WJ, Johnson C, Hu FZ, Stoodley P, Ehrlich GD, Post JC (2008) Characterization of biofilm matrix, degradation by DNase treatment and evidence of capsule downregulation in *Streptococcus pneumoniae* clinical isolates. *BMC Microbiol* 8:173
- Hall-Stoodley L, Stoodley P (2009) Evolving concepts in biofilm infections. *Cell Microbiol* 11:1034–1043
- Harro JM, Peters BM, O'May G a., Archer N, Kerns P, Prabhakara R, Shirliff ME (2010) Vaccine development in *Staphylococcus aureus*: taking the biofilm phenotype into consideration. *FEMS Immunol Med Microbiol* 59:306–323
- Hochstim CJ, Choi JY, Lowe D, Masood R, Rice DH (2010) Biofilm detection with hematoxylin-eosin staining. *Arch Otolaryngol Head Neck Surg* 136:453–456
- Hori TS, Gamperl a K, Afonso LO, Johnson SC, Hubert S, Kimball J, Bowman S, Rise ML (2010) Heat-shock responsive genes identified and validated in Atlantic cod (*Gadus morhua*) liver, head kidney and skeletal muscle using genomic techniques. *BMC Genomics* 11:72
- Houdt R Van, Michiels CW (2010) Biofilm formation and the food industry, a focus on the bacterial outer surface. *J Appl Microbiol* 109:1117–1131
- Huang BF, Zou LL, Xie JG, Huang ZC, Li YW, Li a. X (2013) Immune responses of different species of tilapia infected with *Streptococcus agalactiae*. *J Fish Dis* 36:747–752

- Iregui CA, Comas J, Vasquez G, Verjan N (2015) Experimental early pathogenesis of *Streptococcus agalactiae* infection in red tilapia *Oreochromis spp.* J Fish Dis:1-11
- Kayansamruaj P, Pirarat N, Hirano I, Rodkhum C (2014) Increasing of temperature induces pathogenicity of *Streptococcus agalactiae* and the up-regulation of inflammatory related genes in infected Nile tilapia (*Oreochromis niloticus*). Vet Microbiol 172:265-271
- Khare B, Samal A, Vengadesan K, Rajashankar KR, Ma X, Huang IH, Ton-That H, Narayana SVL (2010) Preliminary crystallographic study of the *Streptococcus agalactiae* sortases, sortase A and sortase C1. Acta Crystallogr Sect F Struct Biol Cryst Commun 66:1096-1100
- Kim BJ, Hancock BM, Cid N Del, Bermudez A, Traver D, Doran KS (2015) *Streptococcus agalactiae* infection in zebrafish larvae. Microb Pathog 79:57-60
- King RK, Flick GJ, Pierson MD, Smith SA, Boardman GD, Coale CW (2004) Identification of bacterial pathogens in biofilms of recirculating aquaculture systems. J Aquat Food Prod Technol 13:125-133
- Lee OO, Wang Y, Tian R, Zhang W, Shek CS, Bougouffa S, Al-Suwailem A, Batang ZB, Xu W, Wang GC, Zhang X, Lafi FF, Bajic VB, Qian P-Y (2014) *In situ* environment rather than substrate type dictates microbial community structure of biofilms in a cold seep system. Sci Rep 4:3587
- Leeuwen LM Van, Kuip M van der, Youssef SA, Bruin A de, Bitter W, Furth AM van, Sar AM van der (2014) Modeling tuberculous meningitis in zebrafish using *Mycobacterium marinum*. Dis Model Mech 7:1111-22
- Liu GY, Doran KS, Lawrence T, Turkson N, Puliti M, Tissi L, Nizet V (2004) Sword and shield: linked group B streptococcal beta-hemolysin/cytolysin and carotenoid pigment function to subvert host phagocyte defense. Proc Natl Acad Sci U S A 101:14491-14496
- Liu L, Li YW, He RZ, Xiao XX, Zhang X, Su YL, Wang J, Li AX (2013) Outbreak of *Streptococcus agalactiae* infection in barcoo grunter, *Scortum barcoo* (McCulloch & Waite), in an intensive fish farm in China. J Fish Dis:1-6
- Liu G, Zhang W, Lu C (2012) Complete genome sequence of *Streptococcus agalactiae* GD201008-001, isolated in China from tilapia with meningoencephalitis. J Bacteriol 194:6653-6653
- Liu G, Zhang W, Lu C (2013) Identification of immunoreactive proteins of *Streptococcus agalactiae* isolated from cultured tilapia in China. Pathog Dis

- Marks LR, Davidson B a., Knight PR, Hakansson AP (2013) Interkingdom signaling induces *Streptococcus pneumoniae* biofilm dispersion and transition from asymptomatic colonization to disease. MBio 4
- Martinez G, Harel J, Gottschalk M (2001) Specific detection by PCR of *Streptococcus agalactiae* in milk. Can J Vet Res 65:68–72
- Morel G, Cavalier A (2001) *In situ* hybridization in light microscopy. CRC Press LLC, Florida
- Morera D, MacKenzie SA (2011) Is there a direct role for erythrocytes in the immune response? Vet Res 42:89
- Morera D, Roher N, Ribas L, Balasch JC, Donate C, Callol A, Boltana S, Roberts S, Goetz G, Goetz FW, Simon A. MacKenzie. (2011) RNA-Seq reveals an integrated immune response in nucleated erythrocytes. PLoS One 6:e26998
- Moter A, Gobel UB (2000) Fluorescence *in situ* hybridization (FISH) for direct visualization of microorganisms. J Microbiol Methods Vol 41, Issue 2 41:85–112
- Nadell CD, Drescher K, Foster KR (2016) Spatial structure, cooperation and competition in biofilms. Nat Rev Microbiol 14:589–600
- Necchi F, Nardi-Dei V, Biagini M, Assfalg M, Nuccitelli A, Cozzi R, Norais N, Telford JL, Rinaudo CD, Grandi G, Maione D (2011) Sortase A substrate specificity in GBS pilus 2a cell wall anchoring. PLoS One 6
- Nithikulworawong N, Yakupitiyage A, Rakshit S, Srisapoom P (2012) Molecular characterization and increased expression of the Nile tilapia, *Oreochromis niloticus* (L.), T-cell receptor beta chain in response to *Streptococcus agalactiae* infection. J Fish Dis 35:343–358
- Nobbs AH, Lamont RJ, Jenkinson HF (2009) *Streptococcus* adherence and colonization. Microbiol Mol Biol Rev 73:407–50
- Nobbs AH, Rosini R, Rinaudo CD, Maione D, Grandi G, Telford JL (2008) Sortase A utilizes an ancillary protein anchor for efficient cell wall anchoring of pili in *Streptococcus agalactiae*. Infect Immun 76:3550–3560
- Noraini O, Jahwarhar N a., Sabri MY, Emikpe BO, Tanko PN, Latifah MH, Jamil S (2013) The effect of heat stress on clinicopathological changes and immunolocalization of antigens in experimental *Streptococcus agalactiae*

- infection in red hybrid tilapia (*Oreochromis spp.*). Vet World 6:997–1003
- Nur-Nazifah M, Sabri MY, Siti-Zahrah A (2014) Development and efficacy of feed-based recombinant vaccine encoding the cell wall surface anchor family protein of *Streptococcus agalactiae* against streptococcosis in *Oreochromis sp.* Fish Shellfish Immunol 37:193–200
- Oliveira NM, Martinez-Garcia E, Xavier J, Durham WM, Kolter R, Kim W, Foster KR (2015) Biofilm formation as a response to ecological competition. PLoS Biol 13:1–23
- Oliver-Kozup HA, Elliott M, Bachert BA, Martin KH, Reid SD, Schwegler-Berry DE, Green BJ, Lukowski S (2011) The streptococcal collagen-like protein-1 (Scl1) is a significant determinant for biofilm formation by group A *Streptococcus*. BMC Microbiol 11:262
- Pandey PK, Vivekanand B, Kundan K (2014) Biofilm in aquaculture production. African J Microbiol Res 8:1434–1443
- Parsek MR, Singh PK (2003) Bacterial biofilms: an emerging link to disease pathogenesis. Annu Rev Microbiol 57:677–701
- Passantino L, Altamura M, Cianciotta A, Patruno R, Tafaro A, Jirillo E, Passantino GF (2002) Fish immunology I. binding and engulfment of *Candida albicans* by erythrocytes of rainbow trout (*Salmo gairdneri* Richardson). Immunopharmacol Immunotoxicol 24:665–78
- Pittman KJ, Robbins CM, Osborn JL, Stubblefield B a., Gilbert ES (2010) Agarose stabilization of fragile biofilms for quantitative structure analysis. J Microbiol Methods 81:101–107
- Pradeep PJ, Suebsing R, Sirthammajak S, Kampeera J, Jitrakorn S, Saksmerprom V, Turner W, Palang I, Vanichviriyakit R, Senapin S, Jeffs A, Kiatpathomchai W, Withyachumanarnkul B (2016) Evidence of vertical transmission and tissue tropism of streptococcosis from naturally infected red tilapia (*Oreochromis spp.*). Aquac Reports 3:58–66
- Ramakrishnan L (2013) Looking within the zebrafish to understand the tuberculous granuloma. Adv Exp Med Biol 783:251–266
- Redmile-Gordon M a., Brookes PC, Evershed RP, Goulding KWT, Hirsch PR (2014) Measuring the soil-microbial interface: extraction of extracellular polymeric substances (EPS) from soil biofilms. Soil Biol Biochem 72:163–171

- Rieger AM, Konowalchuk JD, Grayfer L, Katzenback BA, Havixbeck JJ, Kiemele MD, Belosevic M, Barreda DR (2012) Fish and mammalian phagocytes differentially regulate pro-inflammatory and homeostatic responses *in vivo*. PLoS One 7:e47070
- Rinaudo CD, Rosini R, Galeotti CL, Berti F, Necchi F, Reguzzi V, Ghezzi C, Telford JL, Grandi G, Maione D (2010) Specific involvement of pilus type 2a in biofilm formation in group B *Streptococcus*. PLoS One 5:e9216
- Roberts RJ, Agius C, Saliba C, Bossier P, Sung YY (2010) Heat shock proteins (chaperones) in fish and shellfish and their potential role in relation to fish health: A review. J Fish Dis 33:789–801
- Rodkhum C, Kayansamruaj P, Pirarat N, Wongtawatchai J (2012) Duplex PCR for simultaneous and unambiguous detection of *Streptococcus iniae* and *Streptococcus agalactiae* associated with streptococcosis of cultured tilapia in Thailand. Thai J Vet Med 42:153–158
- Rombout JHWM, Yang G, Kiron V (2014) Adaptive immune responses at mucosal surfaces of teleost fish. Fish Shellfish Immunol 40:634–643
- Rosini R, Margarit I (2015) Biofilm formation by *Streptococcus agalactiae*: influence of environmental conditions and implicated virulence factors. Front Cell Infect Microbiol 5:6
- Rosini R, Rinaudo CD, Soriani M, Lauer P, Mora M, Maione D, Taddei A, Santi I, Ghezzi C, Brettoni C, Buccato S, Margarit I, Grandi G, Telford JL (2006) Identification of novel genomic islands coding for antigenic pilus-like structures in *Streptococcus agalactiae*. Mol Microbiol 61:126–141
- Schreck CB, Contreras-sanchez W, Fitzpatrick MS (2001) Effects of stress on fish reproduction, gamete quality, and progeny. Aquaculture 197:3–24
- Sebastião FDA, Pilarski F, Victor M, Lemos F, Pós-graduação P De, Aplicada M (2013) Composition of extracellular polymeric substances (EPS) produced by *Flavobacterium columnare* isolated from tropical fish in Brazil. Brazilian J Microbiol 864:861–864
- Shapiro K, Krusor C, Mazzillo FFM, Conrad PA, Largier JL, Mazet JAK, Silver MW (2014) Aquatic polymers can drive pathogen transmission in coastal ecosystems. Proc R Soc B 281:1287
- Siti-Zahrah A, Padilah B, Azila A, Rimatulhana R, Shahidan H (2008) Multiple streptococcal species infection in cage-cultured red tilapia but showing similar clinical signs. Dis Asian Aquac VI VI:313–320

- Soto E, Halliday-Simmonds I, Francis S, Kearney MT, Hansen JD (2015) Biofilm formation of *Francisella noatunensis* subsp. *orientalis*. Vet Microbiol
- Su Y-L, Feng J, Li Y-W, Bai J-S, Li A-X (2015) Development of a quantitative PCR assay for monitoring *Streptococcus agalactiae* colonization and tissue tropism in experimentally infected tilapia. J Fish Dis:1–10
- Sudheesh PS, Al-Ghabshi A, Al-Mazrooei N, Al-Habsi S (2012) Comparative pathogenomics of bacteria causing infectious diseases in fish. Int J Evol Biol 2012:457264
- Suebsing R, Kampeera J, Tookdee B, Withyachumnarnkul B, Turner W, Kiatpathomchai W (2013) Evaluation of colorimetric loop-mediated isothermal amplification assay for visual detection of *Streptococcus agalactiae* and *Streptococcus iniae* in tilapia. Lett Appl Microbiol 57:317–324
- Thurnheer T, Gmür R, Guggenheim B (2004) Multiplex FISH analysis of a six-species bacterial biofilm. J Microbiol Methods 56:37–47
- Trebesius K, Leitritz L, Adler K, Schubert S, Autenrieth IB, Heesemann J (2000) Culture independent and rapid identification of bacterial pathogens in necrotising fasciitis and streptococcal toxic shock syndrome by fluorescence *in situ* hybridisation. Med Microbiol Immunol 188:169–175
- Volkman HE, Clay H, Beery D, Chang JCW, Sherman DR, Ramakrishnan L (2004) Tuberculous granuloma formation is enhanced by a mycobacterium virulence determinant. PLoS Biol 2
- Wang B, Jian J, Lu Y, Cai S, Huang Y, Tang J, Wu Z (2012) Complete genome sequence of *Streptococcus agalactiae* ZQ0910, a pathogen causing meningoencephalitis in the GIFT strain of Nile tilapia (*Oreochromis niloticus*). J Bacteriol 194:5132–5133
- Wang Y, Yi L, Wu Z, Shao J, Liu G, Fan H, Zhang W, Lu C (2012) Comparative proteomic analysis of *Streptococcus suis* biofilms and planktonic cells that identified biofilm infection-related immunogenic proteins. PLoS One 7
- Wingender J, Flemming HC (2011) Biofilms in drinking water and their role as reservoir for pathogens. Int J Hyg Environ Health 214:417–423
- Wu J, Xi C (2009) Evaluation of different methods for extracting extracellular DNA from the biofilm matrix. Appl Environ Microbiol 75:5390–5395
- Yadav MK, Chae SW, Park K, Song JJ (2013) Hyaluronic acid derived from other streptococci supports *Streptococcus pneumoniae* *in vitro* biofilm formation. Biomed Res Int 2013

- Yi L, Wang Y, Ma Z, Zhang H, Li Y, Zheng JX, Yang YC, Fan HJ, Lu CP (2014) Biofilm formation of *Streptococcus equi* ssp. *zooepidemicus* and comparative proteomic analysis of biofilm and planktonic cells. *Curr Microbiol* 69:227–233
- Yuasa K, Kamaishi T, Hatai K, Bahnnan M, Borisutpeth P (2008) Two cases of streptococcal infections of cultured tilapia in Asia. *Dis Asian Aquac* VI:259–268
- Zamri-Saad M, Amal MN a, Siti-Zahrah A (2010) Pathological changes in red tilapias (*Oreochromis spp.*) naturally infected by *Streptococcus agalactiae*. *J Comp Pathol* 143:227–229
- Zamri-Saad M, Amal MNA, Siti-Zahrah A, Zulkafli AR (2014) Control and prevention of streptococcosis in cultured tilapia in Malaysia: A Review. *Pertanika J Trop Agric Sci* 37:389–410