



UNIVERSITI PUTRA MALAYSIA

***ORAL ADMINISTRATION ASSESSMENT OF OUTER MEMBRANE
PROTEINS OF *Vibrio alginolyticus* ON THE GROWTH OF GIANT
FRESHWATER PRAWNS (*Macrobrachium rosenbergii* De Man)***

AJADI ABDULLATEEF ABIODUN

FPV 2016 27



**ORAL ADMINISTRATION ASSESSMENT OF OUTER MEMBRANE
PROTEINS OF *Vibrio alginolyticus* ON THE GROWTH OF GIANT
FRESHWATER PRAWNS (*Macrobrachium rosenbergii* De Man)**

By

AJADI ABDULLATEEF ABIODUN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Master of Science**

December 2016

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the expression, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

**ORAL ADMINISTRATION ASSESSMENT OF OUTER MEMBRANE
PROTEINS OF *Vibrio alginolyticus* ON THE GROWTH OF GIANT
FRESHWATER PRAWNS (*Macrobrachium rosenbergii* De Man)**

By

AJADI ABDULLATEEF ABIODUN

December 2016

Chairman : Associate Professor Sabri Bin Mohd Yusoff, PhD
Faculty : Veterinary Medicine

The production of prawns is unarguably a fast-growing global aquaculture. This is evident in the culture of freshwater prawns which is rapidly gaining momentum in terms of productions and values in Malaysia and the world at large. *Vibrio alginolyticus* is one of the most pathogenic species of *Vibrio* that cause high mortality in freshwater prawns. The conventional use of antibiotics in the treatment of this disease has remained ineffective and resulted in an exponential increase in virulence and pathogenicity of the microbes. Hence, the need for better and practicable measures of disease prevention and treatment. This was the first study to investigate the effects of oral administration of outer membrane protein of vibrio in *Macrobrachium rosenbergii* experimentally infected with *V. alginolyticus*. Prawns were divided into three groups A, B, and C of 10 prawns each with replicates in 6 (150 L) glass aquaria. Group A was fed with OMPs-mixed diet, group B with OMPs-FIA (Freund's incomplete adjuvants) mixed diet while group C was fed with OMPs or adjuvant-free diet (control diet). Groups A and B were fed for seven days, alternated with control diet for seven days and a booster dose for another seven days. All prawns were weighed weekly, and haemolymph was collected to examine the total haemocytes counts (THC), phenoloxidase activity (PO) and the presence of OMPs in the haemolymph. All prawns were challenged intramuscularly with 50 µL of 10⁷ CFU of *V. alginolyticus*. The results of the analysis revealed significance difference in mean weight gain and THC ($P < 0.05$) between the treated groups and the control but not with PO activity. Although there was no significant difference ($P > 0.05$) in the level of mortality in all the groups after 24 h, this was not unconnected to pains from the injection coupled with stress, as this was also observed in blank control (not challenged with *V. alginolyticus*). In the second experiment, 45 prawns were divided into three different groups of 15 prawns each. Group A was treated with formalin killed *Vibrio* cell by immersion only, B with OMPs by intramuscular injection and boosted by immersion and C with PBS by immersion only at days 0, 3, 5 and 7. A bacterial challenge was carried out by immersion on day 9 and observed for mortality

for seven days. The total haemocyte count (THC) increased in the treatment groups more than the control but no significant difference in the level of THC increment between the treatment groups. There was no mortality in the treatment groups, but the mortality rate in the control group was 55% over the period of seven days. Haemolymph (both coagulated and non-coagulated) that was also collected to detect the presence of OMPs in the system using SDS-PAGE revealed no bands of OMPs but only those of the plasma proteins, this could be as a result of natural clearance activity of the prawns to get rid of foreign agents. Gross examination of the experimentally challenged prawns was carried out following the mortality and tissues were processed for histopathological lesions and immuno-histochemical reaction. The untreated group showed more pronounced lesions than the treatment groups. This study, however, concluded that oral administration of OMPs with or without Freund's incomplete adjuvant is a good growth promoter and has the potential for protection against vibriosis in *Macrobrachium rosenbergii* when administered with unique antigen protection vehicle and at appropriate dosages, but the protection may be for a short period of time.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**PENILAIAN PENGAMBILAN PROTEIN MELALUI MULUT MEMBRAN
LUAR *Vibrio alginolyticus* TERHADAP PERTUMBUHAN UDANG
GALAH (*Macrobrachium rosenbergii* DE MAN)**

Oleh

AJADI ABDULLATEEF ABIODUN

Disember 2016

Pengerusi : Profesor Madya Sabri Bin Mohd Yusoff, PhD
Fakulti : Perubatan Veterinar

Pengeluaran udang dengan jelasnya merupakan global akuakultur yang berkembang dengan pesat. Ini dibuktikan dengan kultur udang air tawar di mana ianya mendapat momentum yang besar dari segi pengeluaran dan harga di Malaysia dan di dunia secara amnya. *Vibrio alginolyticus* ialah salah satu daripada sepsis *Vibrio* yang paling patogenik yang menyebabkan kematian yang banyak di kalangan udang air tawar. Penggunaan antibiotik secara konvensional dalam rawatan penyakit ini masih kekal tidak berkesan dan menyebabkan peningkatan eksponen dalam kemudaratan dan kepatogenan mikrob ini. Oleh itu, langkah yang lebih baik dan praktikal untuk menangani penyakit ini dan rawatannya sangat diperlukan. Kajian ini adalah menjadi yang pertama mengkaji kesan-kesan pemberian selaput luar protein *Vibrio* secara oral dalam *Macrobrachium rosenbergii* yang dijangkiti oleh *V. alginolyticus*. udang dibahagikan kepada tiga kumpulan A, B, dan C yang terdiri daripada 10 udang setiap satu dengan 6 replikasi dalam (150 L) akuarium kaca. Kumpulan A diberi makan dengan diet campuran-OMPs, Kumpulan B dengan diet campuran OMPs-FIA (adjuvant tak lengkap Freund), manakala Kumpulan C diberi makan OMPs atau diet bebas adjuvant (diet kawalan). Kumpulan A dan B diberi makan selama 7 hari, diselang dengan diet kawalan selama 7 hari dan satu dos penggalak untuk 7 hari lagi. Kesemua udang ditimbang setiap minggu dan hemolimfa diambil untuk pemeriksaan jumlah kiraan hemosit (THC), aktiviti fenoloksidase (PO) dan kehadiran OMPs di dalam hemolimfa. Kesemua udang dicabar secara intraotot dengan $50 \mu\text{L } 10^7 \text{ CFU } V. alginolyticus$. Keputusan analisa menunjukkan perbezaan yang signifikan bagi min penambahan berat badan dan THC ($P < 0.05$) di antara kumpulan rawatan dan kumpulan kawalan tetapi tidak dengan aktiviti PO. Walaupun tiada perbezaan yang signifikan ($P > 0.05$) bagi tahap kematian dalam semua kumpulan selepas 24 jam, ini bukan berkaitan dengan kesakitan disebabkan suntikan beserta tekanan, kerana ini turut diperhatikan di dalam kawalan blank (tidak dicabar dengan *V. alginolyticus*). Dalam percubaan kedua, 45 udang telah dibahagikan kepada tiga kumpulan yang berbeza daripada 15 udang setiap satu. Kumpulan A telah dirawat dengan formalin

membunuh sel *Vibrio* oleh rendaman sahaja, B dengan Omps melalui suntikan intramuskular, dirangsang oleh keasyikan dan C dengan PBS oleh rendaman hanya pada hari 0, 3, 5 dan 7. Cabaran bakteria telah dijalankan oleh rendaman pada hari 9 dan diperhatikan untuk kematian selama tujuh hari. Total perkiraan haemocyte (THC) meningkat dalam kumpulan rawatan lebih daripada kawalan tetapi tiada perbezaan yang signifikan dalam tahap THC kenaikan di antara kumpulan rawatan. Tidak ada kematian dalam kumpulan rawatan, tetapi kadar kematian dalam kumpulan kawalan adalah 55% sepanjang tempoh tujuh hari.

Hemolimfa (bergumpal dan tidak bergumpal, kedua-duanya) yang turut diekstrak untuk mengesan kehadiran OMPs di dalam sistem menggunakan SDS-PAGE menunjukkan ketiadaan jalur OMPs tetapi jalur yang dihasilkan oleh protein plasma, ini mungkin disebabkan oleh aktiviti pemugaran semulajadi udang bagi menghapuskan agen-agen asing. Pemeriksaan secara kasar terhadap udang-udang yang dicabar secara eksperimen dilakukan selepas kematian dan tisu diproses untuk lesi-lesi histopatologi dan reaksi imunohistokimia. Kumpulan yang tidak dirawat menunjukkan lesi yang lebih ketara berbanding dengan kumpulan yang dirawat. Kajian ini, bagaimanapun, menyimpulkan bahawa pemberian OMPs secara oral dengan atau tanpa adjuvan tidak lengkap Freund merupakan penggalak pertumbuhan yang bagus dan mempunyai potensi untuk perlindungan terhadap vibriosis di dalam *Macrobrachium rosenbergii*, apabila diberi dengan perantara perlindungan antigen yang unik dan pada dos yang sesuai tetapi perlindungan mungkin untuk jangka masa yang pendek.

ACKNOWLEDGEMENTS

All praises and adorations are due to Almighty Allah, the most Gracious and most Merciful. Verily, when He intends a thing, His command is, “be” and it is. In other words without Allah everything is nothing and nothing is everything.

I cannot but graciously and specially appreciate my amiable supervisor, Associate Professor Dr. Md Sabri Mohd Yusoff for his invaluable guidance, relentless supervision and genuine encouragement throughout the course of this study. My profound gratitude also goes to my co-supervisors, Dr. Ina Salwany Md. Yasin and Dr. Hasliza Abu Hassim for their unwavering support, candid advice and tangible contributions towards the success of this project. Special recognition and appreciation also go to the members of my research team; Dr. Polycarp Tanko, Dr. Abdulsalam Isiaku and Mr. Mohd Jamil Samad for their precious times, ideas and encouragement.

I am also indebted to the students and staff of Histopathology Laboratory, Faculty of Veterinary Medicine, Universiti Putra Malaysia; Nadirah Abu Nor, Noraini Omar, Syafiqah Adilah, Mrs. Jamilah Jahari, Mrs. Latifah Hanan, Dr. Annas Salleh, Dr. Mazlina Mazlan and Mr. Salleh Muritadoh of Aquaculture Nutrition Laboratory, Faculty of Agriculture, Universiti Putra Malaysia.

I wish to express my warmest appreciations to my dear mother, Alhaja Serifat Ajadi for her unalloyed love, understanding, patience and prayers. With a heavy heart of grief but gratitude to Allah, I pray for the repose of my late step mother Alhaja Sidikat Ajadi who died during the course of this study. This woman raised me from primary school until her death. My prayer also goes to my late father Alhaji Muhammad Nuhu Ajadi, may Allah be pleased with his soul. Special thanks also to my uncle Alhaji Ahmed Jimoh, my aunty Alhaja Batuli Amuda and Honourable Olumuyiwa Jimoh for their financial supports.

Finally, my acknowledgement is extended to my family and siblings, most especially Muinat Ajadi, Hamdalat Ajadi, Abdulraheem Ajadi and Ibrahim Ajadi for their unconditional love, encouragement, and support. Lastly, I sincerely appreciate everyone who has contributed one way or the other to the success of this study. God bless you all.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Sabri Bin Mohd Yusoff, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Ina Salwany Md Yasin, PhD

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Hasliza Abu Hassim, PhD

Senior lecturer
Faculty of Veterinary Medicine
Universiti Malaysia Kelantan
(Member)

ROBIAH BINTI YUNUS, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:  Date: _____

Name and Matric No: Ajadi Abdullateef Abiodun / GS40347

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: _____

Name of Chairman
of Supervisory
Committee:

Associate Professor Dr. Sabri Bin Mohd Yusoff

Signature: _____

Name of Member
of Supervisory
Committee:

Dr. Ina Salwany Md Yasin

Signature: _____

Name of Member
of Supervisory
Committee:

Dr. Hasliza Abu Hassim

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF ABBREVIATIONS	xvii
 CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	5
2.1 Aquaculture	5
2.1.1 Historical Background	5
2.1.2 Global Trend in Aquaculture	6
2.1.3 Aquaculture in Malaysia	7
2.2 Similarities and Differences between Shrimp and Prawn	8
2.3 <i>Macrobrachium rosenbergii</i>	10
2.3.1 Life cycle of <i>Macrobrachium rosenbergii</i>	10
2.4 Bacterial Diseases of Prawns	11
2.4.1 Larval Bacterial Necrosis	12
2.4.2 Filamentous Bacterial Disease	12
2.4.3 Shell Disease	12
2.4.4 Black Gill Disease	13
2.4.5 Red Discolouration Disease	13
2.4.6 Vibriosis	14
2.4.6.1 Clinical Signs of Vibriosis	15
2.4.6.2 Gross Pathology	16
2.4.6.3 Histopathology	16
2.4.6.4 Diagnosis	16
2.4.6.5 Treatment	17
2.5 Conventional Use of Antibiotics	17
2.6 Immunostimulants	19
2.6.1 Outer Membrane Proteins (OMPs)	20
2.7 Measurement of Immune Parameters in Shrimps/Prawns	22
2.7.1 Haemocyte Count	22
2.7.2 Phenoloxidase (PO) Activity	23
2.7.3 Superoxide Dismutase (SOD)	24
2.7.4 Total Plasma Protein	25
2.7.5 Phagocytic Activity	25
2.8 Demerits of Immunostimulants	25
 3 THE PROFILES AND ANTIGENICITY ANALYSIS OF OUTER MEMBRANE PROTEINS OF <i>Vibrio alginolyticus</i>	 27

3.1	Introduction	27
3.2	Materials and Methods	27
3.2.1	Culture of Putative <i>Vibrio alginolyticus</i>	28
3.2.2	Phenotypic Identification of <i>V. alginolyticus</i>	28
3.2.3	Extraction of Outer Membrane Proteins	28
3.2.4	Preparation of Hyper Immune Serum against <i>Vibrio alginolyticus</i>	29
3.2.5	Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis	29
3.2.6	Immunoblotting of the Outer Membrane Proteins	30
3.3	Results	30
3.3.1	Phenotypic Identification of <i>V. alginolyticus</i>	30
3.3.2	SDS-PAGE Analysis	31
3.3.3	Immunoblotting	32
3.4	Discussion	33
4	EFFECTS OF <i>VIBRIO</i> OUTER MEMBRANE PROTEINS (OMPs) ON WEIGHT GAIN AND POTENTIAL PROTECTION AGAINST <i>Vibrio alginolyticus</i> IN GIANT FRESHWATER PRAWN (<i>Macrobrachium rosenbergii</i>)	35
4.1	Introduction	35
4.2	Materials and Methods	36
4.2.1	Preparation of OMPs Incorporated in feeds	36
4.2.2	Experimental Design for Oral Stimulation	36
4.2.3	Measurement of Body Weight	37
4.2.4	Total Haemocyte Count (THC)	37
4.2.5	Phenoloxidase (PO) Activity	38
4.2.6	Per-enteral Stimulations	38
4.2.7	Statistical Analysis	39
4.3	Results	39
4.3.1	Oral Stimulation	39
4.3.1.1	Water Parameters	39
4.3.1.2	Average Weight Gain	40
4.3.1.3	Total Haemocyte Count (THC)	41
4.3.1.4	Phenoloxidase (PO) Activity	41
4.3.1.5	Mortality Rate	42
4.3.2	Per-enteral Stimulations	42
4.3.2.1	Detection of OMPs in the Haemolymph of <i>Macrobrachium rosenbergii</i>	42
4.3.2.2	Total Haemocyte Count and Mortality Rate	45
4.4	Discussion	47
5	HISTOPATHOLOGICAL CHANGES of <i>Macrobrachium rosenbergii</i> EXPERIMENTALLY EXPOSED TO <i>Vibrio alginolyticus</i>	51
5.1	Introduction	51
5.2	Materials and Methods	52
5.2.1	Animals	52

5.2.2	Isolation and culture of bacteria from the challenged prawns	52
5.2.3	DNA extraction	52
5.2.4	Polymerase chain reactions	52
5.2.5	Detection of PCR product	53
5.2.6	Gross Examinations	53
5.2.7	Histopathology	53
5.2.8	Immunohistochemistry (IHC)	53
5.3	Results	54
5.3.1	Bacteriological Examination and Molecular Identification	54
5.3.2	Gross Signs	55
5.3.3	Histopathology	55
5.3.4	Immunohistochemistry	62
5.4	Discussion	65
6.	SUMMARY, GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE STUDIES	67
	REFERENCES	70
	APPENDICES	85
	BIODATA OF STUDENT	94
	LIST OF PUBLICATIONS	95

LIST OF TABLES

Table	Page
2.1 Continental World Production of Fish Food from Inland Aquaculture and Mariculture	7
2.2 Aquaculture Production and Value in Malaysia, 2011-2013	8
2.3 Similarities between prawn and shrimp	9
2.4 Differences between prawn and shrimp	9
3.1 Identification of Gram-Negative Bacteria using API 20 E Identification System after 24 hours Incubation	31
4.1 Experimental Design for Oral Stimulation	37
4.2 Experimental Design for Per – enteral Stimulation	39
4.3 Average Water Parameters	40

LIST OF FIGURES

Figure		Page
3.1	Coomassie brilliant blue stained SDS-PAGE profile of sonicated OMPs; Lane M- molecular weight marker; lanes 1 and 2- OMPs of <i>V. alginolyticus</i> .	32
3.2	Immunoblot analysis of OMPs of <i>V. alginolyticus</i> with intensified immunoreactivity of the polypeptides at molecular weight of 42 kDa and 32 kDa; light immunoreactivity reaction at 20 kDa. Lane 1 and 2-immunoreactive bands; lane M – molecular mass marker.	33
4.1	Weekly pattern of weight gain in the three different groups.	40
4.2	THC of the three groups obtained weekly	41
4.3	Phenoloxidase activity of the three groups obtained weekly	42
4.4	Analysis of plasma proteins obtained at different days and OMPs mixed-haemolymph	43
4.5	Analysis of OMPs profile from <i>V. alginolyticus</i> and serum proteins obtained from <i>Macrobrachium rosenbergii</i> at day 0	44
4.6	Immunoblot analysis of OMPs and plasma protein obtained at day 3	45
4.7	THC obtained in the three groups at different days	46
4.8	OMPs group showing progressive increase in the level of THC across different days	46
4.9	Mortality rate of the three groups seven days post-challenge	47
5.1	Agarose gel (1.0%) electrophoresis of Polymerase Chain Reaction (PCR) amplified DNA product of collagenase gene of <i>V. alginolyticus</i>	54
5.2	photomicrograph of different organs of a positive control group of prawns 24 hours post-injection.	55
5.3	photomicrograph of hepatopancreas from positive control group showing vacuolations (black arrow); infiltration of haemocytes; obliteration of tubular lumen and necrosis of the epithelium (blue arrow) 24 hours post infection. H&E, 200x	56

5.4	Representative photomicrograph of hepatopancreas from OMPs group showing normal cellular structure with mild vacuolations. H&E, 200x.	57
5.5	Representative photomicrograph of hepatopancreas from negative control group showing normal cellular structure. H&E, 200x.	57
5.6	photomicrograph of muscle from positive control group showing diffuse areas of necrosis (arrows) 24 hours post infection. H&E, 200x.	58
5.7	photomicrograph of muscle from OMPs group showing mild to moderate haemocytic infiltration and mild loss of myofibril (arrow) 24 hours post infection. H&E, 200x.	58
5.8	photomicrograph of muscle from negative control group showing normal muscular tissue cell structures. H&E, 200x.	59
5.9	photomicrograph of gill from positive control group showing severe swelling and deformed architecture of the lamellae; separation of lamellar epithelium (black arrow) and gill necrosis (red arrow) 24 hours post infection. H&E, 200x.	59
5.10	photomicrograph of gill from OMPs group showing deformed lamellae with club tip (arrow) 24 hours post infection. H&E, 200x.	60
5.11	photomicrograph of gill from negative control group showing the normal architecture of the gill. H&E, 200x	60
5.12	photomicrograph of heart from positive control group showing moderate haemocytic infiltration, mild nodular haemocytic reaction (arrows) 24 hours post infection. H&E, 200x.	61
5.13	photomicrograph of heart muscle from negative control group showing the normal structure of cardiac muscle cells. H&E, 200x.	61
5.14	Photomicrograph of hepatopancreas from positive control 24 hours post infection showing <i>V. alginolyticus</i> with moderate immunoreactivity to polyclonal antibody (arrows). Haematoxylin counter stain, 200x.	62
5.15	Photomicrograph of immuno-staining of muscle from positive control 24 hours post infection showing <i>V. alginolyticus</i> with moderate immunoreactivity to polyclonal antibody (arrows). Haematoxylin counter stain, 200x.	63
5.16	Photomicrograph of muscle from positive control group 24 hours post infection showing <i>V. alginolyticus</i> with severe immunoreactivity to polyclonal antibody (arrows). Haematoxylin counter stain, 200x.	63

5.17	Photomicrograph of gill from OMPs group 24 hours post infection showing <i>V. alginolyticus</i> with severe immunoreactivity to polyclonal antibody (arrow). Haematoxylin counter stain, 200x.	64
5.18	Photomicrograph of immuno-staining of heart from positive control group 24 hours post infection showing <i>V. alginolyticus</i> with moderate immunoreactivity to polyclonal antibody (arrows). Haematoxylin counter stain, 200x.	64
6.1	Schematic illustration of the merits of immunostimulants over the use of antibiotics.	68



LIST OF ABBREVIATIONS

°C	Degree Celsius
µg	Microgram
µL	Micro litre
%	Percentage
ADJ	Adjuvant
AFA	Alcohol Formalin Acetic-acid
AHPS	Acute Hepatopancreatic Necrosis Syndrome
ANOVA	Analysis of Variance
API	Analytical Profile Index
BCA	Bicinchoninic Acid
BSA	Bovine Serum Albumin
BHIB	Brain Heart Infusion Broth
BW	Body Weight
CFU	Colony Forming Unit
CX	Control
DAB	Diaminobenzidine
DHC	Differential Haemocyte Count
DNA	Deoxyribonucleic acid
DO	Dissolved Oxygen
DoF	Department of Fisheries
EDTA	Ethylene Diamine Tetra Acetic acid
ELISA	Enzyme Linked Immunosorbent Assay
EMS	Early Mortality Syndrome
ETC	Etcetera

EU	European Union
FAO	Food and Agriculture Organization
FCA	Freund's Complete Adjuvant
FIA	Freund's Incomplete Adjuvant
FKC	Formalin Killed Cell
$g (\times g)$	Gravitational acceleration
G	Gram
H _A	Alternative Hypothesis
H & E	Hematoxylin and Eosin
H ₀	Null Hypothesis
HP	Hepatopaneas
IHC	Immunohistochemistry
Kb	Kilobase pair
kDa	Kilodalton
Kg	Kilogram
L-DOPA	L-3,4-Dihydrophenylalanine
LGH	Large Granular Haemocyte
LPS	Lipopolysaccharides
MT	Metric tonnes
Mg	Milligram
mL	Millilitre
Mm	Millimetre
mM	Millimolar
NaCl	Sodium Chloride
NaFisH	National Fish Health Research Centre

OMPs	Outer Membrane Proteins
PBS	Phosphate Buffer Saline
PBST	Phosphate Buffer Saline Tween 20
PCR	Polymerase Chain Reactions
PG	Peptidoglycan
PPM	Part Per Million
PPT	Part Per Thousand
PO	Phenoloxidase
ProPO	Prophenoloxidase
RM	Malaysian Ringgit
ROIs	Reactive Oxygen Intermediates
ROS	Reactive Oxygen Species
RPS	Relative Percentage Survival
SDS-PAGE	Sodium Dodecyl Sulfate Polyacrylamide Gel Electrophoresis
SE	Standard Error
SEM	Scanning Electron Microscopy
SGH	Small Granular Haemocyte
SOD	Superoxide Dismutase
Sp.	Specie
Spp.	Species
TBE	Tris Base Electrophoresis
TBS	Tris Buffer Saline
TCBS	Thiosulfate Citrate Bile salts Sucrose
THC	Total Haemocyte Count
TSB	Tryptic Soy Broth

USD	United States Dollar
USFDA	United States Food and Drug Administration
UN	United Nations
v/v	Volume/Volume
w/v	Weight/Volume
YSI	Yellow Spring Inc.



CHAPTER 1

INTRODUCTION

The growth of global production of seafood from aquaculture has been very consistent and tremendously high in the last decade, with the record of 52.5 million tonnes in 2008, which was 61.7% higher than 32.4 million tonnes in 2000. With the exclusion of aquatic plants, the value of the global harvest of aquaculture was estimated at US\$98.4 billion in 2008 (FAO, 2011). The production rose to 62.7 million tonnes in 2011 at an estimated value of USD130 billion (FAO Fisheries and Aquaculture Department, 2013) and 90.43 million tonnes in 2012, and the estimate of an average global supply of food fish per person by aquaculture was 9.41 kg (FAO, 2014). The essential role of aquaculture in global hunger elimination, health promotion, poverty reduction and to some extent environmental protection cannot be over emphasized. The global production of freshwater prawns has equally gathered momentum in terms of increment in metric tons and values, with a total estimate of USD2.2 billion annually (FAO 2012). Oriental river prawn (*Macrobrachium nipponense*) was valued at USD1.13 billion with 237, 431 metric tons, giant river prawn (*M. rosenbergii*) valued at USD593.6 million with 124,713 metric tons and the rest of the species in the same genus completed the rest of the statistics (FAO, 2012). The discussion on *M. rosenbergii* would not be complete, without reference to Malaysia where it derives one of its other nomenclature (Malaysian prawn). This may be related to the discovery, of Shao Wen Ling of the FAO in the 1960s, at Penang, Malaysia, that the larval stages of *M. rosenbergii* required brackish water for development to post-larvae and survival (Wowor and Ng, 2007). This marked the beginning of modern aquaculture of this species.

Malaysia is one of the forces to reckon with, in terms of global aquaculture (FAO Fisheries and Aquaculture Department, 2013). In 2009, the total annual production of all species of freshwater prawns was 440,000 ton at the value of USD2.2 billion. Out of this total global figure, the production of farmed *M. rosenbergii* contributed 51.7%, while the oriental river prawn *M. nipponense* (exclusively reared in China) constituted 47.2% (New & Nair, 2012). However, for more than two decades, aquaculture has embraced tremendous growth in Malaysia, for instance, in 1992, the total production was estimated to be 79,699 tons of the value of RM 207.4 million, these figures surpassed that of the previous year by 23% and 25% respectively (Kechik, 1995).

Macrobrachium rosenbergii is one of the most important reared and fished crustaceans with high commercial value. For some years now, there had been a heightened interest in the culture of giant freshwater prawn (*M. rosenbergii*), due to its lower risks involvement and abundant market demands (Hameed *et al.*, 2003). The value of the average unit product of both *M. rosenbergii* and all species of freshwater prawns surpassed that of various major aquaculture products such as the two-major species of marine shrimp (*Litopenaeus vannamei* and *Penaeus monodon*) and Atlantic salmon (*Salmo salar*). *M. rosenbergii* possesses unique characteristics such as large size attainment, omnivorous nature of feeding on unconventional feeds and tolerance

to changes in water quality and handling stress (El Sayed, 1997). It can thrive and grow well in both fresh and low salinity water and wide temperature range (FAO, 1975). When the penaeid shrimps culture was increasingly plagued with disease outbreaks, a global economically important cultured species, *Macrobrachium rosenbergii* gained more considerations by farmers due to the perception of its relatively less susceptibility to diseases (Pillai & Bonami, 2012). Meanwhile, the rise in demands, pronounced culture intensification and increase in world trade of this farmed species, have but come with attendant challenges such as disease outbreaks which bring about serious setbacks in management and production. Major diseases that affect the freshwater industry are of bacterial and viral origins. Apart from idiopathic diseases, others that are less common or cause serious detrimental effects than the two formers, include fungi, yeast, parasites and nutritional deficiencies have been reported (Pillai and Bonami, 2012; Bondad-Reantaso *et al.*, 2005; Eshraghi *et al.*, 2005; Michael, 2002; Karunasagar *et al.*, 1998; Bower *et al.*, 1994; Johnson, 1975). The major genera of bacteria which are incriminated in deleterious effects and economic loss in freshwater prawns are *Vibrio*, *Aeromonas*, and *Pseudomonas* (Pillai and Bonami, 2012).

However, vibriosis is one of the most important diseases reported in farmed freshwater prawns with grievous devastating effects and huge reduction or loss of production (Khuntia *et al.*, 2008; Jayaprakash *et al.*, 2006; Kennedy *et al.*, 2006; Poupard, 1976). *Vibrio alginolyticus* is one of the highly pathogenic species affecting fish and shell fish farming (Hsieh *et al.*, 2008; Jayaprakash *et al.*, 2006; Liu *et al.*, 2004). Vibriosis causes a high rate of morbidity and mortality especially under stress and other environmental conditions. Many hatcheries and grow out ponds have been plagued by the malady and farmers have sought various means to get out of this situation, the commonest among all is the use of antibiotics.

The incessant and uncontrollable application of antibiotics in aquaculture both prophylactically and chemotherapeutically have resulted to ineffective disease combatant, many species of bacteria have developed resistance, and more virulence strains have been birthed. Subsequent drug residues also resulted from this traditional practice of disease management and eradication which pose a threat to the health of humans through consumption. Many isolates (more than 90%) of bacteria isolated from larvae and post larvae of *M. rosenbergii* showed resistance to some antibiotics including oxytetracycline, erythromycin, and furazolidone (Hameed *et al.*, 2003). Strains of *V. alginolyticus* and other vibrios were reported to carry R plasmid which is responsible for transferable drug resistance (Gomathi *et al.*, 2013). Many hatcheries and grow out ponds of farmed fish and shell fish have gone bankrupt due to outbreaks of highly antimicrobial resistant bacteria and humongous economic losses have been incurred (D. Pillai *et al.*, 2005).

In lieu of this bugging situation of antibiotic resistance, there is an urgent need for better and safer alternatives. Many studies have been carried out in the light of finding lasting solutions to this malady and to some great extents, tremendous successes have been achieved in the area of vaccination and use of probiotics. Meanwhile, researchers have explored various means to boost the non- specific immunity of crustaceans that

are of economic and health importance in order to raise protection against intending disease outbreaks through the concept of immunostimulation.

An immunostimulant in aquaculture of shellfish is said to be any substance that is used to improve immune responses and enhance disease resistance against pathogenic organisms (Smith *et al.*, 2003). Various agents such as microbial components, compounds of animal and plant origins and synthetic substances have been applied as immunostimulants with varying degree of effectiveness. Components of microorganisms cell wall such as beta-glucan (Bai *et al.*, 2014; Chang *et al.*, 2003), lipopolysaccharides (Abbass *et al.*, 2010), peptidoglycan (Purivirojkul *et al.*, 2006; Itami *et al.*, 1998) and outer membrane proteins (Maftuch *et al.*, 2013) have been studied and reported to stimulate immune responses and enhance survival of crustaceans against infections by pathogenic organisms.

Various routes of administrations including injection, immersion and oral have been tested. Injection route is the most effective but costly, labour intensive and cause additional stress to the host species (Smith *et al.*, 2003). However, the two latter are the preferred and most practicable routes (Smith *et al.*, 2003).

In comparison to other components of the cell wall of bacteria, including lipopolysaccharides and peptidoglycan, little studies have been carried out on the enteral route of administration of outer membrane proteins. Hence, this study took into consideration the practicability of administration and compliance by users in investigating the effect of oral administration of outer membrane proteins (OMPs) of *Vibrio* in giant freshwater water prawns (*Macrobrachium rosenbergii*) on weight gain and protective potential against infection by *Vibrio alginolyticus*.

The objectives of this study were:

1. To determine the profiles and immunogenicity of OMPs of *Vibrio alginolyticus* using SDS-PAGE and Western blot techniques.
2. To prepare outer membrane proteins incorporated in feed and evaluate its effects on weight gain and infection by *V. alginolyticus*.
3. To examine the histopathological and immuno-histochemical reactions of prawns which have been experimentally infected with *V. alginolyticus*.

The hypotheses of this study were:

1. **H₀**: OMPs of *V. alginolyticus* do not contain several minor and major polypeptides that are immunogenic
2. **H_A**: OMPs of *V. alginolyticus* do contain several minors and major polypeptides that are immunogenic
3. **H₀**: oral administration of OMPs does not have any effect on weight gain and infection caused by *V. alginolyticus* in freshwater prawns.

4. **H_A**: oral administration of OMPs does have any effect on weight gain and infection caused by *V. alginolyticus* in freshwater prawns.



REFERENCES

- Abbass, A., Sharifuzzaman, S. M., & Austin, B. (2010). Cellular components of probiotics control *Yersinia ruckeri* infection in rainbow trout, *Oncorhynchus mykiss* (Walbaum). *Journal of Fish Diseases*, 33(1), 31–37.
- Abraham, T.J., Manley, R., Palaniappan, R., Dhevendaran, K., 1997. Pathogenicity and antibiotic sensitivity of luminous *Vibrio harveyi* isolated from diseased penaeid shrimp. *Journal of Aquaculture in the Tropics*. 121 , 1–8.
- Adams, A. (1991). Response of penaeid shrimp to exposure to *Vibrio* species. *Fish & Shellfish Immunology*, 1(1), 59–70.
- Ai, Q., Mai, K., Zhang, L., Tan, B., Zhang, W., Xu, W., & Li, H. (2007). Effects of dietary β -1, 3 glucan on innate immune response of large yellow croaker, *Pseudosciaena crocea*. *Fish and Shellfish Immunology*, 22(4), 394–402.
- Alday-Sanz, V., Roque, A. & Turnbull, J. (2002). Clearing mechanisms of *Vibrio vulnificus* biotype I in the black tiger shrimp *Penaeus monodon*. *Diseases of Aquatic Organisms*, 48, 91–99.
- Alippi, A. M. (1999). Bacterial diseases. *Bee Disease Diagnosis*. (Eds. Colin ME, Ball BV, Kilani M). *Options Méditerranéennes, Serie B: Etudes et Recherches*, (25), 1999.
- Anderson, I. G., Shamsudin, M. N., & Nash, G. (1989). A preliminary study on the aerobic heterotrophic bacterial flora in giant freshwater prawn, *Macrobrachium rosenbergii*, hatcheries in Malaysia. *Aquaculture*, 81(3), 213–223.
- Asche, F., Roll, K. H., & Tveteras, R. (2009). Economic inefficiency and environmental impact: An application to aquaculture production. *Journal of Environmental Economics and Management*, 58(1), 93–105.
- Azad, I. S., Panigrahi, A., Gopal, C., Paulpandi, S., Mahima, C., & Ravichandran, P. (2005). Routes of immunostimulation vis-à-vis survival and growth of *Penaeus monodon* postlarvae. *Aquaculture*, 248(1-4), 227–234.
- Bachère, E., Mialhe, E., Noël, D., Boulo, V., Morvan, A., & Rodriguez, J. (1995). Knowledge and research prospects in marine mollusc and crustacean immunology. *Aquaculture*, 132(1-2), 17–32.
- Bai, N., Gu, M., Zhang, W., Xu, W., & Mai, K. (2014). Effects of β -glucan derivatives on the immunity of white shrimp *Litopenaeus vannamei* and its resistance against white spot syndrome virus infection. *Aquaculture*, 426–427, 66–73.
- Barman, D., Nen, P., Mandal, S. C., & Kumar, V. (2013). Immunostimulants for Aquaculture Health Management. *Journal of Marine Science Research and Development*, 3(3).

- Barman, P., Banerjee, A., Bandyopadhyay, P., Chandra, K., Kumar, P., & Mohapatra, D. (2011). Isolation, identification and molecular characterization of potential probiotic bacterium, *Bacillus subtilis* PPP 13 from *Penaeus monodon*. *Biotechnology. Bioinformatic and. Bioengineering.*, 1(4), 473–482.
- Baticados, M. C. L., Lavilla-Pitogo, C. R., Cruz-Lacierda, E. R., De La Pena, L. D., & Sunaz, N. A. (1990). Studies on the chemical control of luminous bacteria *Vibrio harveyi* and *Vibrio splendidus* isolated from diseased *Penaeus monodon* larvae and rearing water. *Disease of Aquatic Organisms*, 9(2), 133-139.
- Bell, T. A. L., Bell, D. V. T. A., & Lightner, D. V. (1988). *A handbook of normal penaeid shrimp histology* (No. 595.3843 B4).
- Beveridge, M. C., & Little, D. C. (2002). The history of aquaculture in traditional societies. *Ecological aquaculture. The evolution of the Blue Revolution*, 3-29.
- Bhavan, P. S., & Geraldine, P. (2000). Histopathology of the hepatopancreas and gills of the prawn *Macrobrachium malcolmsonii* exposed to endosulfan. *Aquatic Toxicology*, 50, 331–339.
- Bondad-Reantaso, M.G., Subasinghe, R.P., Arthur, J.R., Ogawa, K., Chinabut, S., Adlard, R., Tan, Z. and Shariff, M. (2005). Disease and health management in Asian aquaculture. *Veterinary parasitology*, 132(3), 249-272.
- Bower, S. M., McGladdery, S. E., & Price, I. M. (1994). Synopsis of infectious diseases and parasites of commercially exploited shellfish. *Annual Review of Fish Diseases*, 4, 1–199.
- Brock, J. A., & Lightner, D. V. (1990). Diseases of crustacea. Diseases caused by microorganisms. *Diseases of marine animals*, 3, 245-349.
- Cai, S., Yao, S., Lu, Y., Wu, Z., Jian, J., & Wang, B. (2010). Immune response in *Lutjanus erythropterus* induced by the major outer membrane protein (OmpU) of *Vibrio alginolyticus*. *Diseases of Aquatic Organisms*, 90, 63–68.
- Cerenius, L., Jiravanichpaisal, P., Liu, H. P., & Söderhäll, I. (2010). Crustacean immunity. *Advances in Experimental Medicine and Biology*, 708(12), 239–259.
- Cerenius, L., Lee, B. L., & Söderhäll, K. (2008). The proPO-system: pros and cons for its role in invertebrate immunity. *Trends in Immunology*, 29(6), 263–271.
- Chang, C. F., Su, M. Sen, Chen, H. Y., & Liao, I. C. (2003). Dietary β -1,3-glucan effectively improves immunity and survival of *Penaeus monodon* challenged with white spot syndrome virus. *Fish and Shellfish Immunology*, 15(4), 297–310.
- Chen, F. R., Liu, P. C., & Lee, K. K. (2000). Lethal attribute of serine protease secreted by *Vibrio alginolyticus* strains in kuruma prawn *Penaeus japonicus*. *Zeitschrift für Naturforschung C*, 55(1-2), 94-99.

- Chen, S. N., Huang, S. L., & Kou, G. H. (1992). Studies on the epizootiology and pathogenicity of bacterial infections in cultured giant tiger prawns, *Penaeus monodon* in Taiwan. *Diseases of cultured penaeid shrimp in Asia and United States. The Oceanic Institute, Hawaii*, 195-205.
- Cheng, W., & Chen, J. C. (1998). Isolation and characterization of an Enterococcus-like bacterium causing muscle necrosis and mortality in *Macrobrachium rosenbergii* in Taiwan. *Diseases of Aquatic Organisms*, 34(2), 93-101.
- Cheng, W., & Wang, C. H. (2001). The susceptibility of the giant freshwater prawn *Macrobrachium rosenbergii* to *Lactococcus garvieae* and its resistance under copper sulfate stress. *Diseases of aquatic organisms*, 47(2), 137-144.
- Cheng, W., Liu, C. H., Kuo, C. M., & Chen, J. C. (2005). Dietary administration of sodium alginate enhances the immune ability of white shrimp *Litopenaeus vannamei* and its resistance against *Vibrio alginolyticus*. *Fish and Shellfish Immunology*, 18(1), 1-12.
- Chiu, S. T., Hsieh, S. L., Yeh, S. P., Jian, S. J., Cheng, W., & Liu, C. H. (2010). The increase of immunity and disease resistance of the giant freshwater prawn, *Macrobrachium rosenbergii* by feeding with selenium enriched-diet. *Fish and Shellfish Immunology*, 29(4), 623-629.
- Chu, S., Cavaignac, S., Feutrier, J., Phipps, B. M., Kostrzynska, M., Kay, W. W., & Trust, T. J. (1991). Structure of the tetragonal surface virulence array protein and gene of *Aeromonas salmonicida*. *Journal of Biological Chemistry*, 266(23), 15258-15265.
- Collins, P. (2010). Environmental stress upon hepatopancreatic cells of freshwater prawns (Decapoda: Caridea) from the floodplain of Paraná River. *Natural Science*, 2(7), 748-759.
- Das, N. G., Monwar, M. M., & Siddique, M. L. (2013). Rematuration of hatchery used wild spawners of *Macrobrachium rosenbergii* (De Man 1879) in captive condition. *Animal Biology & Animal Husbandry*, 5(1).
- Department of Fisheries Malaysia. (2012). *Annual Fisheries Statistics 2012*. Ministry of Agriculture and Agro-based Industry Malaysia. retrieved 15 September 2015 from <http://www.dof.gov.my/en/fishery-statistics>.
- Department of Fisheries Malaysia. (2011). *Annual Fisheries Statistics Book Volume 1*. Paper presented at the annual meeting of Department of Fisheries Malaysia. 18-20.
- Department of Fisheries Malaysia. (2013). *Annual Fisheries Statistics 2013*. Ministry of Agriculture and Agro-based Industry Malaysia. retrieved 15 September 2015 from <http://www.dof.gov.my/en/fishery-statistics>.

- Dirienzo, J. M., Nakamura, K., & Inouye, M. (1978). The outer membrane proteins of gram-negative bacteria: biosynthesis, assembly, and functions. *Annual Review of Biochemistry*, 47, 481–532.
- Doñate, C., Balasch, J. C., Callol, A., Bobe, J., Tort, L., & MacKenzie, S. (2010). The effects of immunostimulation through dietary manipulation in the rainbow Trout; Evaluation of mucosal immunity. *Marine Biotechnology*, 12(1), 88–99.
- Ebanks, R., Goguen, M., McKinnon, S., Pinto, D., & Ross, N. (2005). Identification of the major outer membrane proteins of *Aeromonas salmonicida*. *Diseases of Aquatic Organisms*, 68, 29–38.
- El-Sayed, A. F. M. (1997). Growth rates and feed efficiency of the freshwater prawn *Macrobrachium rosenbergii* fed varying protein and energy levels. *Bulletin of the National Institute of Oceanography and Fishery Egypt*, 23, 439–448.
- Eshraghi, L., You, M. P., & Barbetti, M. J. (2005). Diseases of cultured prawns in australia. *Plant Disease*, 89(10), 1131–1131.
- Evensenl, O., Espelid, S., & Tore, A. (1991). Immunohistochemical identification of *Vibrio salmonicida* in stored tissues of Atlantic salmon *Salmo salar* from the first known outbreak of cold-water vibriosis (Hitra disease). *Diseases of Aquatic Organisms*, 10, 185–189.
- FAO Fisheries and Aquaculture Department. (2014). the Global Aquaculture Production Statistics Database updated to 2011. *Fao*, 2014(March).
- FAO Fisheries and Aquaculture Department. (2013). the Global Aquaculture Production Statistics for the year 2011. *Fao*, 2011(March), 3p. Retrieved from www.fao.org/fishery/topic/16140/en
- FAO (Food & Agriculture Organisation). (2012). *The State of World Fisheries and Aquaculture 2012*. Sofia.
- FAO. (2011). *World aquaculture 2010*. FAO Fisheries and Aquaculture Department. *Technical paper* (Vol. No. 500/1.). Retrieved from <http://www.fao.org/docrep/014/ba0132e/ba0132e.pdf>
- FAO (Food and Agriculture Organization). (2007). *The State of Food and Agriculture*. Rome, 2007.
- FAO (Food and Agriculture Organisation). (2003). *The State of Food Insecurity in the World*. Rome 2003.
- FAO (Food & Agriculture Organisation). (2002). *The State of World Fisheries and Aquaculture 2002*. Sofia.
- FAO (2000). *The State of World Fisheries and Aquaculture 2000*. FAO, Rome, Italy.

FAO. 1995. Code of Conduct for Responsible Fisheries. FAO, Rome.

FAO. (1975). *Macrobrachium rosenbergii*, 6, 103–114.

Flores-Miranda, M. D. C., Luna-González, A., Campa-Córdova, Á. I., González-Ocampo, H. a., Fierro-Coronado, J. a., & Partida-Arangure, B. O. (2011). Microbial immunostimulants reduce mortality in whiteleg shrimp (*Litopenaeus vannamei*) challenged with *Vibrio sinaloensis* strains. *Aquaculture*, 320(1-2), 51–55.

Glauert, A. M., & Thornley, M. J. (1968). The topography of the bacterial cell wall. *Annual review of microbiology*, 23, 159-198.

Gomathi, R. S., Vinothkumar, R., & Arunagiri, K. (2013). Isolation and Identification *Vibrios* from marine Seafood Samples. *Internationa Journal of Current Microbiology and Applied Scinces*, 2(2), 36–43.

Gomez-Gil, B., Tron-Mayen, L., Roque, A., Turnbull, J. F., Inglis, V., & Guerra-Flores, A. L. (1998). Species of *Vibrio* isolated from hepatopancreas, haemolymph and digestive tract of a population of healthy juvenile *Penaeus vannamei*. *Aquaculture*, 163(1), 1-9.

Hameed, A. S., Rahaman, K. H., Alagan, A., & Yoganandhan, K. (2003). Antibiotic resistance in bacteria isolated from hatchery-reared larvae and post-larvae of *Macrobrachium rosenbergii*. *Aquaculture*, 217(1), 39-48.

Harikrishnan, R., Balasundaram, C., Jawahar, S., & Heo, M.-S. (2012). Immunomodulatory effect of *Withania somnifera* supplementation diet in the giant freshwater prawn *Macrobrachium rosenbergii* (de Man) against *Aeromonas hydrophila*. *Fish & Shellfish Immunology*, 32(1), 94–100.

Holmblad, T., & Söderhäll, K. (1999). Cell adhesion molecules and antioxidative enzymes in a crustacean, possible role in immunity. *Aquaculture*, 172(1-2), 111–123.

Hose, J. E., Martin, G. G., Tiu, S., & McKrell, N. (1992). Patterns of hemocyte production and release throughout the molt cycle in the penaeid shrimp *Sicyonia ingentis*. *Biological Bulletin*, 183, 185–199.

Hsieh, S.-L., Ruan, Y.-H., Li, Y.-C., Hsieh, P.-S., Hu, C.-H., & Kuo, C.-M. (2008). Immune and physiological responses in Pacific white shrimp (*Penaeus vannamei*) to *Vibrio alginolyticus*. *Aquaculture*, 275(1-4), 335–341.

Huang, M. T., Eble, A. F., & Hammen, C. S. (1981). Immune response of the prawn, *Macrobrachium rosenbergii* to bacterial infection. *Journal of Invertebrate Pathology*, 38(2), 213-219.

Inside Malaysia, July 2012. retrieved from <http://etp.pemandu.gov.my/upload/Inside%20Investor%20%20Agriculture%20and%20Aquaculture.pdf>

- Itami, T., Asano, M., Tokushige, K., Kubono, K., Nakagawa, A., Takeno, N., ... Takahashi, Y. (1998). Enhancement of disease resistance of kuruma shrimp, *Penaeus japonicus*, after oral administration of peptidoglycan derived from *Bifidobacterium thermophilum*. *Aquaculture*, 164, 277–288.
- Jayabalan, N., Chandran, R., Sivakumar, V., & Ramamoorthi, K. (1982). Occurrence of luminescent bacteria in sediments. *Current Science*, 51(14), 710–711.
- Jayaprakash, N. S., Pai, S. S., Philip, R., & Singh, I. S. B. (2006). Isolation of a pathogenic strain of *Vibrio alginolyticus* from necrotic larvae of *Macrobrachium rosenbergii* (de Man). *Journal of Fish Diseases*, 29(3), 187–191.
- Jayasree, L., Janakiram, P., & Madhavi, R. (2006). Characterization of *Vibrio* spp. associated with diseased shrimp from culture ponds of Andhra Pradesh (India). *Journal of the World Aquaculture Society*, 37(4), 523–532.
- Jayasree, S. (2009). Identification of immune cells interacting with *Vibrio* spp. and its in vitro post-phagocytic killing mechanism of haemocytes in the penaeid shrimp, *Penaeus indicus* H. Milne Edwards. *Journal of Fish Diseases*, 32(4), 359–365.
- Jiravanichpaisal, P., Lee, B. L., & Söderhäll, K. (2006). Cell-mediated immunity in arthropods: Hematopoiesis, coagulation, melanization and opsonization. *Immunobiology*, 211(4), 213–236.
- Jiravanichpaisal, P and Miyazaki, T. 1994. Histopathology, biochemistry and pathogenicity of *Vibrio harveyi* infecting black tiger shrimp *Penaeus monodon*. *Journal of Aquatic Animal Health*, 6: 27–35.
- Johansson, M. W., & Soderhall, K. (1989). Cellular immunity in crustaceans and the proPO system. *Parasitology Today (Personal Ed.)*, 5(6), 171–176.
- Johansson, M. W., Keyser, P., Sritunyalucksana, K., & Söderhäll, K. (2000). Crustacean haemocytes and haematopoiesis. *Aquaculture*, 191(1-3), 45–52.
- Johnson, S. K. (1975). *Handbook of shrimp diseases*. Texas AM Univ., Coll. Stn., Sea Grant Publ. (Vol. 601).
- Karunasagar, I., Otta, S. K., & Karunasagar, I. (1998). disease problem affecting cultured penaeid shrimp in india. *Fish Pathology*, 33(4), 413–419.
- Karunasagar, I., Pai, R., Malathi, G. R., & Karunasagar, I. (1994). Mass mortality of *Penaeus monodon* larvae due to antibiotic-resistant *Vibrio harveyi* infection. *Aquaculture*, 128(3-4), 203–209.
- Kechik, I. A. (1995). Aquaculture in Malaysia. *ADSEA '94 Proceedings*, (1995), 125–134.

- Kennedy, B., Venugopal, M. N., Karunasagar, I., & Karunasagar, I. (2006). Bacterial flora associated with the giant freshwater prawn *Macrobrachium rosenbergii*, in the hatchery system. *Aquaculture*, 261(4), 1156-1167.
- Keysami, M. A., & Mohammadpour, M. (2013). Effect of *Bacillus subtilis* on *Aeromonas hydrophila* infection resistance in juvenile freshwater prawn,. *Springer*, 553–562
- Keysami, M. A., Saad, C. R., Sijam, K., Daud, H. M., & Alimon, A. R. (2007). Effect of *Bacillus subtilis* on growth development and survival of larvae *Macrobrachium rosenbergii* (de Man). *Aquaculture nutrition*, 13(2), 131-136.
- Khuntia, C. P., Das, B. K., Samantaray, B. R., Samal, S. K., & Mishra, B. K. (2008). Characterization and pathogenicity studies of *Vibrio parahaemolyticus* isolated from diseased freshwater prawn, *Macrobrachium rosenbergii* (de Man). *Aquaculture Research*, 39(3), 301–310.
- Khushiramani, R., Girisha, S. K., Bhowmick, P. P., Karunasagar, I., & Karunasagar, I. (2008). Prevalence of different outer membrane proteins in isolates of *Aeromonas* spp. *World Journal of Microbiology and Biotechnology*, 24(10), 2263–2268.
- Kumaresan, V., Palanisamy, R., Pasupuleti, M., & Arockiaraj, J. (2016). Impacts of environmental and biological stressors on immune system of *Macrobrachium rosenbergii*. *Reviews in Aquaculture*.
- Laemmli, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature*, 227, 680-685.
- Landau, M. (1992). *Introduction to aquaculture* (Vol. 639). Nueva York: Wiley.
- Lavilla-Pitogo, Celia R., Lio-Po, Gilda D., Cruz-Lacierda Erlinda R., Alapide-Tendencia, Eleonor V., De la Pena, L. D. (2000). Diseases of Penaeid Shrimps in the Philippines. *Aquaculture Extension Manual* No. 16. Second Edition. July 2000.
- Lavilla-Pitogo, C. R., Leaño, E. M., & Paner, M. G. (1998). Mortalities of pond-cultured juvenile shrimp, *Penaeus monodon*, associated with dominance of luminescent vibrios in the rearing environment. *Aquaculture*, 164(1-4), 337–349.
- Lavilla-Pitogo, C. R., Leano, E. M., & Paner, M. G. (1996, May). Mortalities of pond-cultured juvenile shrimp, *Penaeus monodon*, associated with dominance of luminescent bacteria, *Vibrio harveyi* in the rearing environment. In *SICCPS book of abstracts, SEAFDEC, Iloilo City, Philippines* (Vol. 40).
- Lavilla-Pitogo, C. R., Baticados, M. C. L., Cruz-Lacierda, E. R., & de la Pena, L. D. (1990). Occurrence of luminous bacterial disease of *Penaeus monodon* larvae in the Philippines. *Aquaculture*, 91(1-2), 1–13.

- Le Moullac, G., Soyeux, C., Saulnier, D., Ansquer, D., Avarre, J. C., & Levy, P. (1998). Effect of hypoxic stress on the immune response and the resistance to vibriosis of the shrimp *Penaeus stylirostris*. *Fish & Shellfish Immunology*, 8(8), 621–629.
- Le Moullac, G., Le Groumellec, M., Ansquer, D., Froissard, S., & Levy, P. (1997). Haematological and phenoloxidase activity changes in the shrimp *Penaeus stylirostris* in relation with the moult cycle: protection against vibriosis. *Fish & Shellfish Immunology*, 7(4), 227–234.
- Lee, S. Y., & Söderhäll, K. (2002). Early events in crustacean innate immunity. *Fish & Shellfish Immunology*, 12(5), 421–437.
- Lem, A. L. and Shehadeh, Z., 1998. International trade in aquaculture products. *INFOFISH International* 4/98. pp.25–29.
- Li, C. C., Yeh, S. T., & Chen, J. C. (2010). Innate immunity of the white shrimp *Litopenaeus vannamei* weakened by the combination of a *Vibrio alginolyticus* injection and low-salinity stress. *Fish and Shellfish Immunology*, 28(1), 121–127.
- Li, N., Yang, Z., Bai, J., Fu, X., Liu, L., Shi, C., & Wu, S. (2010). A shared antigen among *Vibrio* species: Outer membrane protein-OmpK as a versatile Vibriosis vaccine candidate in Orange-spotted grouper (*Epinephelus coioides*). *Fish & Shellfish Immunology*, 28(5–6), 952–956.
- Lightner, D. V. (2012). Biology and pathology of early mortality syndrome of shrimp. *Global outlook for aquaculture leadership*, Bangkok, 40.
- Lightner, D. V. (2005). Biosecurity in shrimp farming: pathogen exclusion through use of SPF stock and routine surveillance. *Journal of the World Aquaculture Society*, 36(3), 229–248.
- Lightner, D. V., & Redman, R. M. (1998). Shrimp diseases and current diagnostic methods. *Aquaculture*, 164, 201–220.
- Lightner, D. V. (1996). *A handbook of shrimp pathology and diagnostic procedures for diseases of cultured penaeid shrimp*.
- Lightner, D.V. 1993. Diseases of cultured penaeid shrimp. In: J.P. McVey (ed.) *CRC Handbook of Mariculture*, Second edition, Volume 1, Crustacean Aquaculture. CRC Press Inc., Boca Raton, FL. p. 393–486.
- Lightner, D. V., Brock, J., Le Bitoux, J. F., Johnson, P., Sindermann, C., Overstreet, R., Newman, M. W., Armstrong, D. A., Fisher W. S., Wickham D. E. & Rosemark, R. (1988). Disease diagnosis and control in North American marine aquaculture: crustacean diseases. *Developments in aquaculture and fisheries science*, 17, 6–412.

- Ling, S.W. 1969. Methods of rearing and culturing *Macrobrachium rosenbergii*. *FAO Fisheries Reports* No. 57 Vol. 3:607-619.
- Liu, C., & Chen, J. C. (2004). Effect of ammonia on the immune response of white shrimp *Litopenaeus vannamei* and its susceptibility to *Vibrio alginolyticus*. *Fish and Shellfish Immunology*, 16(3), 321–334.
- Liu, C. H., Cheng, W., Hsu, J. P., & Chen, J. C. (2004). *Vibrio alginolyticus* infection in the white shrimp *Litopenaeus vannamei* confirmed by polymerase chain reaction and 16S rDNA sequencing. *Diseases of Aquatic Organisms*, 61(1-2), 169–174.
- Longshaw, M. (2011). Diseases of crayfish: A review. *Journal of Invertebrate Pathology*, 106(1), 54–70.
- Maftuch, Prasetyo, E., Sudianto, A., Rozik, M., Nurdiani, R., Sanusi, E., Murachman. (2013). Improvement of innate immune responses and defense activity in tiger shrimp (*Penaeus monodon* Fab.) by intramuscular administration of the outer membrane protein *Vibrio alginolyticus*. *SpringerPlus*, 2(1), 432.
- Maji, S., Mali, P., & Joardar, S. N. (2006). Immunoreactive antigens of the outer membrane protein of *Aeromonas hydrophila*, isolated from goldfish, *Carassius auratus* (Linn.). *Fish and Shellfish Immunology*, 20(4), 462–473.
- Man, M. De. (2013). Effect of *Bacillus subtilis* on *Aeromonas hydrophila* infection resistance in juvenile freshwater prawn. *Springer*, 553–562.
- Mao, Z., Yu, L., You, Z., Wei, Y., & Liu, Y. (2007). Cloning, expression and immunogenicity analysis of five outer membrane proteins of *Vibrio parahaemolyticus* zj2003. *Fish & Shellfish Immunology*, 23(3), 567–575.
- Mehdi Raissy. (2011). Molecular detection of *Vibrio* spp. in lobster hemolymph. *African Journal of Microbiology Research*, 5(13), 1697–1700.
- Michael, N. B. (2002). *Farming freshwater prawns*. A manual for the culture of the giant river prawn (*Macrobrachium rosenbergii*). FAO Fisheries Technical Paper, 428.
- Minh, N. P., Lam, T. B., Giao, N. T., & Quan, N. C. (2010). Tissue distribution and elimination of erythromycin in giant freshwater prawn (*Macrobrachium rosenbergii*) depletion. *African Journal of Food Science*, 4(September), 578–584.
- Miyamoto, G., Brock, J., Nakamura, R., Nakagawa, L., Shimojo, R., Sato, V., & Akita, G. (1983). A preliminary microbiological and water quality survey of two Hawaiian prawn (*Macrobrachium rosenbergii*) hatcheries. In *Proceedings of First International Conference on Warm Water Aquaculture-Crustacea* (pp. 429-458). Brigham Young University Hawaii Campus Laie, HI.

- Moriarty, D. J. (1999, August). Disease control in shrimp aquaculture with probiotic bacteria. In *Proceedings of the 8th international symposium on microbial ecology* (pp. 237-243). Atlantic Canada Society for Microbial Ecology, Halifax, Canada.
- Moriarty, D. J. W. (1998). Control of luminous *Vibrio* spp. in penaeid aquaculture ponds. *Aquaculture*, 164(1), 351-358.
- Morton, R. J., René Simons, K., & Confer, A. W. (1996). Major outer membrane proteins of *Pasteurella haemolytica* serovars 1-15: Comparison of separation techniques and surface-exposed proteins on selected serovars. *Veterinary Microbiology*, 51(3-4), 319-330.
- Moullac, G. Le, & Groumellec, M. Le. (1997). Haematological and phenoloxidase activity changes in the shrimp *Penaeus stylirostris* in relation with the moult cycle: protection against vibriosis. *Fish & Shellfish Immunology*, 7(4), 227-234.
- Moullac, G. Le, & Haffner, P. (2000). Environmental factors affecting immune responses in Crustacea, 121-131.
- Nash, C. (2010). *The history of aquaculture*. John Wiley & Sons.
- New, M. B., & Nair, C. M. (2012). Global scale of freshwater prawn farming. *Aquaculture Research*, 43(7), 960-969.
- New, M. B. (2005). Freshwater prawn farming: Global status, recent research and a glance at the future. *Aquaculture Research*, 36(3), 210-230.
- New, M. B. (1990). Freshwater prawn culture: a review. *Aquaculture*, 88(2), 99-143.
- Parker, N. (1989). History, status, and future of aquaculture in the United States. *Critical Reviews in Aquatic Sciences*, 1, 97-109.
- Pati, U. S., Srivastava, S. K., Roy, S. C., & More, T. (1996). Innmunogenicity of outer membrane protein of *Pasteurella multocida* in buffalo calves. *Veterinary Microbiology*, 52, 301-311.
- Pena, L. D. D. La, Momoyama, K., Nakai, T., & Muroga, K. (1992). Detection of the Causative Bacterium of Vibriosis in Kuruma Prawn, *Penaeus japonicus*. *Fish Pathology*, 27(4), 223-228.
- Perazzolo, L. M., & Barracco, M. A. (1997). The prophenoloxidase activating system of the shrimp *Penaeus paulensis* and associated factors. *Developmental and Comparative Immunology*, 21(5), 385-395.
- Persson, M., Vey, A., & Söderhäll, K. (1987). Encapsulation of foreign particles in vitro by separated blood cells from crayfish, *Astacus leptodactylus*. *Cell and tissue research*, 247(2), 409-415.

- Pillai, D., & Bonami, J. R. (2012). A review on the diseases of freshwater prawns with special focus on white tail disease of *Macrobrachium rosenbergii*. *Aquaculture Research*, 43(7), 1029–1037.
- Pillai, D., Nair, C. M., Salin, K. R., Marques, A., Widada, J. S., & Bonami, J. R. (2005). Gross signs and histopathology of branchiostegal blister disease (balloon disease): an idiopathic disease of farmed *Macrobrachium rosenbergii* (De Man). *Journal of fish diseases*, 28(8), 473–478.
- Pope, E. C., Powell, A., Roberts, E. C., Shields, R. J., Wardle, R., & Rowley, A. F. (2011). “Enhanced cellular immunity in shrimp (*Litopenaeus vannamei*) after ‘vaccination’.” *PLoS ONE*, 6(6), 1–7.
- Poupard, J. D.-B. and C. W. (1976). disease problems of prawns in recirculation systems in the U.K. *Aquaculture*, 7, 201–217.
- Powell, A., Pope, E. C., Eddy, F. E., Roberts, E. C., Shields, R. J., Francis, M. J., ... Rowley, A. F. (2011). Enhanced immune defences in Pacific white shrimp (*Litopenaeus vannamei*) post-exposure to a *Vibrio* vaccine. *Journal of Invertebrate Pathology*, 107(2), 95–99.
- Purivirojkul, W., Areechon, N., & Srisapoome, P. (2006). The Effect of Peptidoglycan on Immune Response in Black Tiger Shrimp (*Penaeus monodon* Fabricius). *Kasetsart Journal of Natural Science*. 187, 181–187.
- Qian, R. H., Xiao, Z. H., Zhang, C. W., Chu, W. Y., Wang, L. S., Zhou, H. H., ... Yu, L. (2008). A conserved outer membrane protein as an effective vaccine candidate from *Vibrio alginolyticus*. *Aquaculture*, 278(1-4), 5–9.
- Rengpipata, S., & Rukpratanpornb, S. (2000). Immunity enhancement in black tiger shrimp (*Penaeus monodon*) by a probiont bacterium (*Bacillus* SPP.). *Aquaculture*, 191, 271–288.
- Rinaldo, G., & Yevich, P. (1974). Spot Gill Syndrome P andalus. *Journal of Invertebrate Pathology*, 233, 224–233.
- Rodríguez, J., & Le Moullac, G. (2000). State of the art of immunological tools and health control of penaeid shrimp. *Aquaculture*, 191(1-3), 109–119.
- Rosenberry, B. (2015, September 15). USAdcFDArefusalsSetRecordAugust2015. Retrieved 16 September, 2015 from <http://www.shrimpnews.com/FreeReportsFolder/NewsReportsFolder/>
- Rosenberry B. (2004) World shrimp farming 2004. In: Shrimp News International (ed. by B. Rosenberry), pp. 4. San Diego, CA.
- Rosenberry, B., 1996. World shrimp farming 1996. In: Rosenberry, R. Ed. , Shrimp News International. 167 pp.

- Rowley, A. F., & Pope, E. C. (2012). Vaccines and crustacean aquaculture—A mechanistic exploration. *Aquaculture*, 334-337, 1–11.
- Sabri, M. Y. (1999). Immunology of the outer membrane proteins of *Pasturella multocida* A2, A7 and A9 in sheep. *Master of Science Thesis*, Faculty of Veterinary Medicine, Universiti Putra Malaysia.
- Sabri, M. Y., Zamri-Saad, M., Mutalib, a. R., Israf, D. a., & Muniandy, N. (2000). Efficacy of an outer membrane protein of *Pasteurella haemolytica* A2, A7 or A9-enriched vaccine against intratracheal challenge exposure in sheep. *Veterinary Microbiology*, 73(1), 13–23.
- Saeed Ziaei-Nejad, Gholamreza Rafiee & Mehdi Shakouri, Culture and Breeding of Freshwater Prawn *Macrobrachium rosenbergii* as an Exotic Species in Iran, Present Status and Future Perspective. *Paper presented at the meeting of Asian Pacific Aquaculture*, Kuala Lumpur, 2009.
- Saejung, C., Hatai, K., Wada, S., Kurata, O., & Sanoamuang, L. (2011). Clinical observations of black disease in fairy shrimps, *Streptocephalus sirindhornae* and *Branchinella thailandensis*, from Thailand and pathogen verification. *Journal of Fish Diseases*, 34(12), 911–920.
- Sahoo, P. K., Pillai, B. R., Mohanty, J., Kumari, J., Mohanty, S., & Mishra, B. K. (2007). In vivo humoral and cellular reactions, and fate of injected bacteria *Aeromonas hydrophila* in freshwater prawn *Macrobrachium rosenbergii*. *Fish and Shellfish Immunology*, 23(2), 327–340.
- Sajeevan, T. P., Philip, R., & Bright Singh, I. S. (2009). Dose/frequency: A critical factor in the administration of glucan as immunostimulant to Indian white shrimp *Fenneropenaeus indicus*. *Aquaculture*, 287(3-4), 248–252.
- Sakai, M. (1999). Current research status of fish immunostimulants. *Aquaculture*, 172(1-2), 63–92.
- Sano, T., & Fukuda, H. (1987). Principal microbial diseases of mariculture in Japan. *Aquaculture*, 67(1-2), 59–69.
- Saulnier, D., Haffner, P., Goarant, C., Levy, P., & Ansquer, D. (2000). Experimental infection models for shrimp vibriosis studies: A review. *Aquaculture*, 191(1-3), 133–144.
- Sayuthi, S. (1993). Fish diseases in Malaysia : status and problems . *Proceedings of the Aquaculture Workshop for SEAFDEC/AQD Training Alumni*, 8- 11 September 1992, 56–61.
- Schnaitman, C. a. (1974). Outer membrane proteins of *Escherichia coli*. 3. Evidence that the major protein of *Escherichia coli* O111 outer membrane consists of four distinct polypeptide species. *Journal of Bacteriology*, 118(2), 442–453.

- Scholz, U., Garcia Diaz, G., Ricque, D., Cruz Suarez, L. E., Vargas Albores, F., & Latchford, J. (1999). Enhancement of vibriosis resistance in juvenile *Penaeus vannamei* by supplementation of diets with different yeast products. *Aquaculture*, 176, 271–283.
- Sharma, S. R. K., Shankar, K. M., Sathyanarayana, M. L., Sahoo, a K., Patil, R., Narayanaswamy, H. D., ... Shankar, K. M. (2010). Evaluation of immune response and resistance to diseases in tiger shrimp, *Penaeus monodon* fed with biofilm of *Vibrio alginolyticus*. *Fish & Shellfish Immunology*, 29(5), 724–32.
- Smith, V. J., Brown, J. H., & Hauton, C. (2003). Immunostimulation in crustaceans: does it really protect against infection? *Fish & Shellfish Immunology*, 15(1), 71–90.
- Smith, V. J., & Chisholm, J. R. (1992). Non-cellular immunity in crustaceans. *Fish & Shellfish Immunology*, 2(1), 1-31.
- Smith, V. J., & Johnston, P. A. (1992). Differential haemotoxic effect of PCB congeners in the common shrimp, *Crangon crangon*. *Comparative Biochemistry and Physiology Part C: Comparative Pharmacology*, 101(3), 641-649.
- Smith, V. J., & Söderhäll, K. (1986). Cellular immune mechanisms in the Crustacea. In *Symposia of the Zoological Society of London*, (Vol. 56, pp. 59-79).
- Smith, V. J., Söderhäll, K., & Hamilton, M. (1984). β 1, 3-Glucan induced cellular defence reactions in the shore crab, *Carcinus maenas*. *Comparative Biochemistry and Physiology Part A: Physiology*, 77(4), 635-639.
- Smith, V. J., & Söderhäll, K. (1983). Induction of degranulation and lysis of haemocytes in the freshwater crayfish, *Astacus astacus* by components of the prophenoloxidase activating system in vitro. *Cell and tissue research*, 233(2), 295-303.
- Soderhall, K. (1983). Separation of the Haemocyte Populations of. *Developmental and Comparative Immunology*, 7, 229–239.
- Söderhäll, K., & Cerenius, L. (1998). Role of the prophenoloxidase-activating system in invertebrate immunity. *Current Opinion in Immunology*, 10(1), 23–28.
- Song, Y. L., Liu, J. J., Chan, L. C., & Sung, H. H. (1996). Glucan-induced disease resistance in tiger shrimp (*Penaeus monodon*). *Developments in biological standardization*, 90, 413-421.
- Stickney, R. R., & Treece, G. D. (2012). History of aquaculture. *Aquaculture Production Systems*, 15-50.
- Subasinghe, R. P., Barg, U., Phillips, M. J., Bartley, D., & Tacon, A. (1998). Aquatic animal health management: investment opportunities within developing countries. *Journal of Applied Ichthyology*, 14(3-4), 123-129.

- Sung, H. H. (1990). Enhancement of Growth in Tiger Shrimp (*Penaeus monodon*) by bacterin prepared from *Vibrio vulnificus* - *Vibrio vulnificus* Isolation and characterization. *Bulletin of the European Association of Fish Pathologists*, 10(4), 98–99.
- Sung, H. H., Hwang, S. F., & Tasi, F. M. (2000). Responses of giant freshwater prawn (*Macrobrachium rosenbergii*) to challenge by two strains of *Aeromonas* spp. *Journal of Invertebrate Pathology*, 76(4), 278–284.
- Sung, H. H., Yang, Y. L., & Song, Y. L. (1996). Enhancement of microbicidal activity in the tiger shrimp *Penaeus monodon* via immunostimulation. *Journal of Crustacean Biology*, 16(2), 278–284.
- Sung, H.-H., Kuo, P.-A., & Kao, W. (2000). Effect of Lipopolysaccharide on In Vitro Phagocytosis Hemocytes from Giant Freshwater Prawn (*Macrobrachium rosenbergii*). *The Japanese Society of Fish Pathology*, 3(35).
- Taylor, H. H. (1992). Gills and lungs: the exchange of gases and ions. *Microscopic anatomy of invertebrates*, 10, 203–293.
- Tendencia, E. a., & De La Peña, L. D. (2001). Antibiotic resistance of bacteria from shrimp ponds. *Aquaculture*, 195(3–4), 193–204.
- Thakur, A. B., Vaidya, R. B., & Suryawanshi, S. A. (2003). Pathogenicity and antibiotic susceptibility of *Vibrio* species isolated from moribund shrimps. *Indian Journal of Natural Products and Resources*, 32(March), 71–75.
- Thangaviji, V., Michaelbabu, M., Anand, S. B., & Gunasekaran, P. (2012). Microbial & Biochemical Technology Immunization with the *Aeromonas* OMP Provides Protection against *Aeromonas hydrophila* in Goldfish (*Carassius auratus*). *Microbial and Biochemical Technology*, 4(2), 45–49.
- Thörnqvist, P. O., Johansson, M. W., & Söderhäll, K. (1994). Opsonic activity of cell adhesion proteins and β -1, 3-glucan binding proteins from two crustaceans. *Developmental & Comparative Immunology*, 18(1), 3–12.
- Tonguthai, K. (1992). Diseases of the freshwater prawn *Macrobrachium rosenbergii* in Thailand. *Diseases in Asian Aquaculture*, 1, 89–95.
- Towbin, H., Staehelin, T., & Gordon, J. (1979). Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. *Proceedings of the National Academy of Sciences*, 76(9), 4350–4354.
- Trust, T. J., Kostrzynska, M., Emödy, L., & Wadström, T. (1993). High-affinity binding of the basement membrane protein collagen type IV to the crystalline virulence surface protein array of *Aeromonas salmonicida*. *Molecular Microbiology*, 7(4), 593–600.

- Tseng, D. Y., Ho, P. L., Huang, S. Y., Cheng, S. C., Shiu, Y. L., Chiu, C. S., & Liu, C. H. (2009). Enhancement of immunity and disease resistance in the white shrimp, *Litopenaeus vannamei* by the probiotic, *Bacillus subtilis* E20. *Fish and Shellfish Immunology*, 26(2), 339–344.
- Tseng, I. T., & Chen, J. C. (2004). The immune response of white shrimp *Litopenaeus vannamei* and its susceptibility to *Vibrio alginolyticus* under nitrite stress. *Fish and Shellfish Immunology*, 17(4), 325–333.
- Tsing, A., Arcier, J.-M., & Brehélin, M. (1989). Hemocytes of Penaeid and Palaemonid shrimps: Morphology, cytochemistry, and hemograms. *Journal of Invertebrate Pathology*, 53(1), 64–77.
- Turnidge, J. (2004). Antibiotic use in animals--prejudices, perceptions and realities. *The Journal of Antimicrobial Chemotherapy*, 53(1), 26–27.
- Vaseeharan, B., & Ramasamy, P. (2003). Control of pathogenic *Vibrio* spp . by *Bacillus subtilis* BT23 , a possible probiotic treatment for black tiger shrimp *Penaeus monodon*. *Letters in Applied Microbiology*, 83–87.
- Vogan, C. L., Costa-Ramos, C., & Rowley, A. F. (2002). Shell disease syndrome in the edible crab, *Cancer pagurus* - Isolation, characterization and pathogenicity of chitinolytic bacteria. *Microbiology*, 148(3), 743–754.
- Wang, L. U., & Chen, J. C. (2005). The immune response of white shrimp *Litopenaeus vannamei* and its susceptibility to *Vibrio alginolyticus* at different salinity levels. *Fish and Shellfish Immunology*, 18(4), 269–278.
- Wowor, D., & Ng, P. K. L. (2007). The giant freshwater prawns of the *Macrobrachium rosenbergii* species group (Crustacea: Decapoda: Caridea: Palaemonidae). *Raffles Bulletin of Zoology*, 55(2), 321–336.
- Xiong, X., Wang, C., & Ye, M. (2010). Differentially Expressed Outer Membrane Proteins of *Vibrio alginolyticus* in Response to Six Types of Antibiotics. *Marine Biotechnology*, 12, 686–695.
- Xiong, X.-P., Zhang, B.-W., Yang, M.-J., Ye, M.-Z., Peng, X.-X., & Li, H. (2010). Identification of vaccine candidates from differentially expressed outer membrane proteins of *Vibrio alginolyticus* in response to NaCl and iron limitation. *Fish & Shellfish Immunology*, 29(5), 810–6.
- Yeh, S., & Chen, J. (2008). Immunomodulation by carrageenans in the white shrimp *Litopenaeus vannamei* and its resistance against *Vibrio alginolyticus*. *Aquaculture*, 276(1-4), 22–28.
- Zorriehzahra, M. J. R. B. (2014). Review Article. *Advances in Animal and Veterinary Sciences*, 3(2).